Electronic Supporting Information

An AI-Engineered Peptidyl Fluorescence Probe for GRP78:

Advancing Renal Cell Carcinoma Diagnosis

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1. Experimental Section

Materials. All chemicals and solvents were purchased as reagent grade and used without further purification unless otherwise noted. Fetal bovine serum and penicillin/streptomycin were obtained from Biological Industries and Beijing Solarbio Science&Technoiogy Co., Ltd. Human GRP78 protein was purchased from Abcam. Fetal bovine serum and penicillin/streptomycin were obtained from Biological Industries and Beijing Solarbio Science&Technoiogy Co., Ltd. respectively. Roswell Park Memorial Institute 1640 (RPMI 1640), Dulbecco's Modified Eagle Medium (DMEM) and Modified Eagle Medium (MEM) were purchased from Beijing Solarbio Science&Technoiogy Co., Ltd.

Instruments. ¹H NMR and ¹³C NMR spectra were recorded on the Bruker Avance-600 MHz and 151 MHz NMR spectrometers, respectively. ESI mass spectrometry was carried out on the AB Triple TOF 5600plus System. UV-visible spectra and steady-state emission experiments were performed on a Hitachi U-3900 spectrometer and a Hitachi F-7000 spectrometer, respectively. Cell and tissue imaging experiments were measured by Zeiss LSM880 Airyscan confocal laser scanning microscope. *In vivo* imaging assays were performed in the Perkin Elmer Lumina LT small animal optical imaging system.

Cell culture. Human colon carcinoma HCT116 cells were cultured and maintained in RPMI 1640 medium (supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin) at 37 °C and 5% CO₂ in a humidified incubator. Human embryonic kidney 293T cells, human breast cancer MCF-7 cells, mouse hepatoma Hepa1-6 cells, human cervical cancer Hela cells were cultured in DMEM medium supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin. Human renal adenocarcinoma ACHN cells were cultured in MEM medium supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin. All cells were provided by the Institute of Biochemistry and Cell Biology, SIBS, CAS (China).

Confocal laser scanning microscope (CLSM) imaging. Cells were cultured overnight 14 mm glass coverslips overnight and washed with PBS (3 times × 2 mL).

After incubation with 10 μ M probes for 120 min at 37 °C, cells were washed with PBS (3 times \times 2 mL) and subjected to CLSM imaging. The green channel (493-593 nm) was used to collect the probes signal under 488 nm as the excitation wavelength.

Model mice and *in vivo* imaging. All the animal experiments were approved by the Radiation Protection Institute of Drug Safety Evaluation Center in China (Production license: SYXK (Jin) 2018-0005). All the animal experiments were performed by following the protocols approved by the Radiation Protection Institute of Drug Safety Evaluation Center in China (Production license: SYXK (Jin) 2018-0005). The studies involving human participants were reviewed and approved by the Ethics Committee of the First Hospital of Shanxi Medical University, China (No. KYLL-2024-197). Female BALB/c nude mice of 6-8 weeks old were purchased from Beijing Viton River Experimental Technology Co., Ltd. The mice were housed under a 12-h light/dark cycle and were allowed free access to food and water. ACHN cells were injected into mice to obtain xenograft tumor bearing mice model. Mice were intratumor injection with 2 mM probes. Fluorescence images were acquired with time change via the following parameters: excitation wavelength: 465 nm; emission filter: 520 nm - 580 nm. Afterward, mice were euthanized by cervical dislocation, and organ were collected and imaged further.

Tissue slice imaging of RCC. All tissue slice samples from patients were harvested from First Hospital of Shanxi Medical University. The studies involving human participants were reviewed and approved by the Ethics Committee of the First Hospital of Shanxi Medical University, China (No. KYLL-2024-197). All patients gave their informed consent to participate in the research. All methods were performed in accordance with the relevant guidelines and regulations. Adjacent tissues were identified for longitudinal assessment of kidney function.

2. Synthesis and Characterization

Synthesis details of probes. The peptidyl probes (designated as **probe2-5**, as shown in Scheme S1), which introduced peptide sequence modified with 5-FAM fluorophore and dabcyl quencher, were custom made from Bankpeptide Biological

Technology Co., Ltd. **Probe1** was developed by Wen and colleagues to target GRP78, as described in their published work S1.

Scheme S1. Chemical structures of probes1-5.

probe5

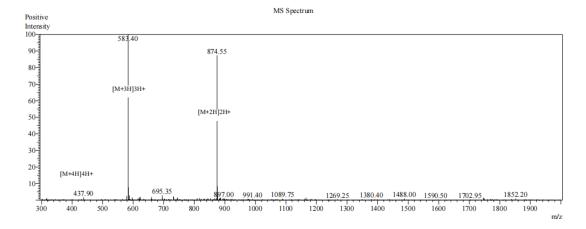
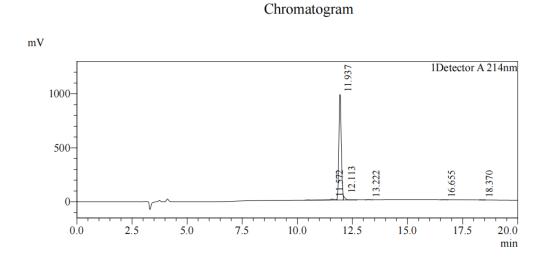


Figure S1. The HR-MS of **probe2**. Calc. for $C_{98}H_{118}N_{13}O_{17}^{2+}$ [M+3H]²⁺ 874.44, found 874.55.



	Peak Table				
Detector A 21	4nm				
Peak#	Ret. Time	Area	Height	Area%	
1	11.572	148733	9540	1.940	
2	11.937	7319228	976575	95.458	
3	12.113	178458	32352	2.327	
4	13.222	13235	1864	0.173	
5	16.655	5164	679	0.067	
6	18.370	2627	417	0.034	
Total		7667445	1021426	100.000	

Figure S2. Typical chromatogram of **probe2**, which presented that the purification of the final probe >95%.

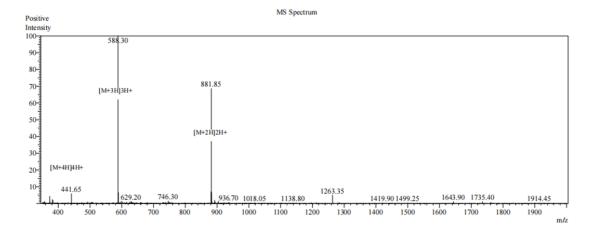


Figure S3. The HR-MS of **probe3**. Calc. for $C_{97}H_{115}N_{14}O_{18}^{2+}$ [M+3H]²⁺ 881.93, found 881.85.

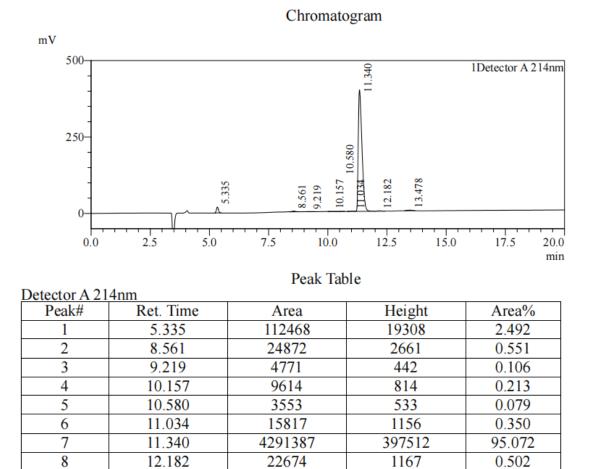


Figure S4. Typical chromatogram of **probe3**, which presented that the purification of the final probe >95%.

28675

4513830

2232

425824

9

Total

13.478.

0.635

100.000

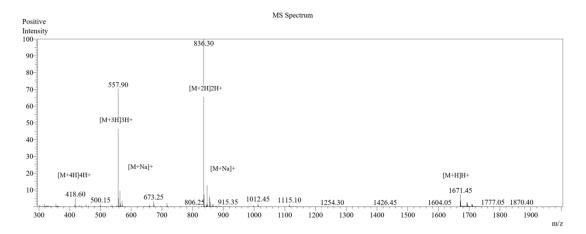
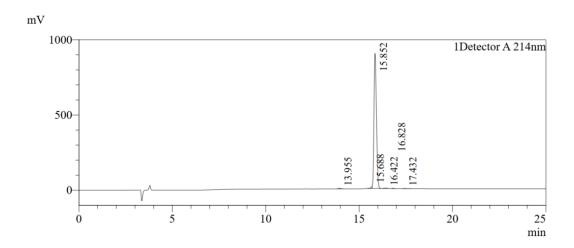


Figure S5. The HR-MS of **probe4**. Calc. for $C_{86}H_{106}N_{13}O_{20}S^{2+}$ [M+3H]²⁺ 836.37, found 836.30.

Chromatogram



Detector A 214nm Ret. Time Height Area% Peak# Area 1 13.955 29762 3766 0.357 2 15.688 63159 11500 0.757 3 15.852 8186517 898250 98.181 4 16.422 24310 2869 0.292 5 16.828 11752 1673 0.141 6 17.432 22723 1670 0.273

Peak Table

919728

Figure S6. Typical chromatogram of **probe4**, which presented that the purification of the final probe >95%.

8338224

Total

100.000

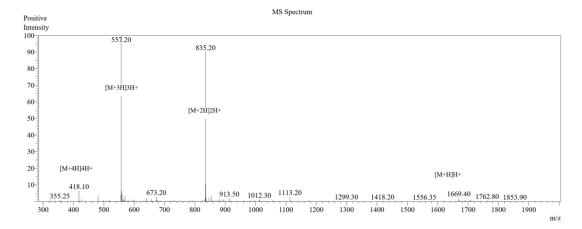
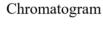
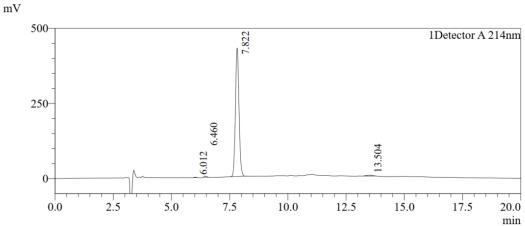


Figure S7. The HR-MS of **probe5**. Calc. for $C_{86}H_{104}N_{13}O_{22}^{2+}$ [M+3H]²⁺ 835.37, found 835.20.





Peak Table

Detector A 21	4nm			
Peak#	Ret. Time	Area	Height	Area%
1	6.012	8014	1497	0.173
2	6.460	25957	3565	0.559
3	7.822	4558341	427459	98.130
4	13.504	52840	2961	1.138
5	21.083	56	19	0.001
Total		4645209	435502	100.000

Figure S8. Typical chromatogram of **probe5**, which presented that the purification of the final probe >95%.

3. Additional spectra data

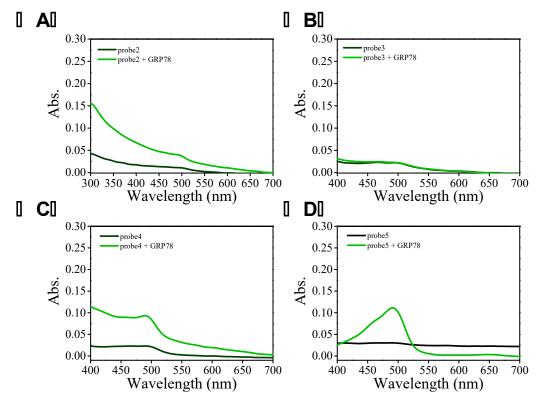


Figure S9. Absorption spectra of 5 μ M probe 2 (A), probe 3 (B), probe 4 (C) and probe 5 (D) upon addition of 50 μ g/mL GRP78 in 10 mM phosphate buffer (DMSO 0.25%, v/v, pH = 7.2).

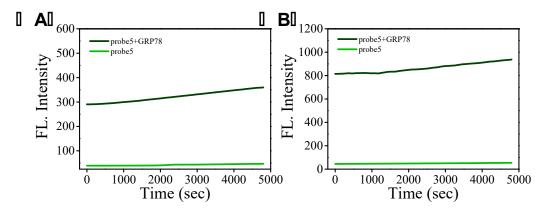


Figure S10. Change of fluorescence intensity at 528 nm of **probe 5** (5 μ M) with/without 50 μ g/mL GRP78 over time (A) in PBS buffer (10 % fetal bovine serum) and (B) in PBS buffer (10 % MEM medium), λ_{ex} = 490 nm.

4. Additional cells imaging data

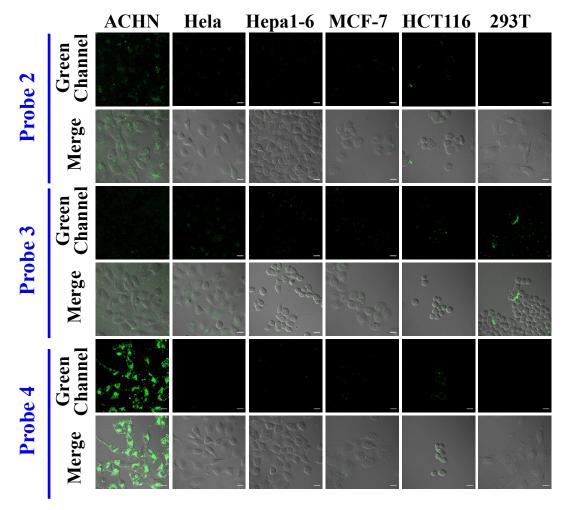


Figure S11. CLSM imaging with probes in ACHN, Hela, Hepa1-6, MCF-7, HCT116, 293T cells. Green channel ($\lambda_{ex} = 488$ nm, $\lambda_{em} = 500\text{-}560$ nm) imaging signal of cells preincubated with **probe2**, **probe3** or **probe4** (10 μ M) were collected for 120 min. Scale bar = 10 μ m.

5. Western blot assays

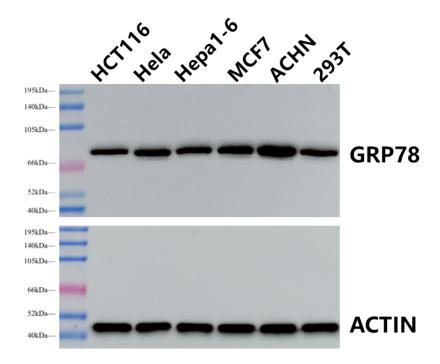


Figure S12. Expression of GRP78 in ACHN, HeLa, Hepa1-6, MCF-7, HCT116, and 293T cells.

6. Cell viability data

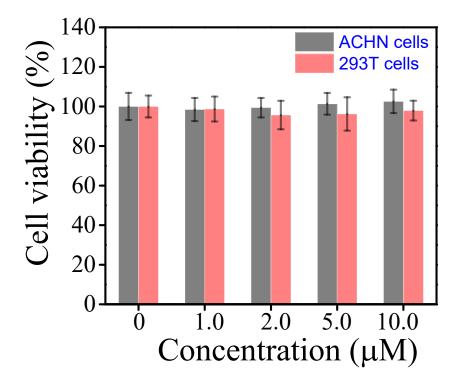


Figure S13. Cell viability values (%) estimated by MTT assay in ACHN cells and 293T cells, which were cultured in the presence of 0-10 μ M **probe 5** for 5 h.

7. Additional in vivo imaging data

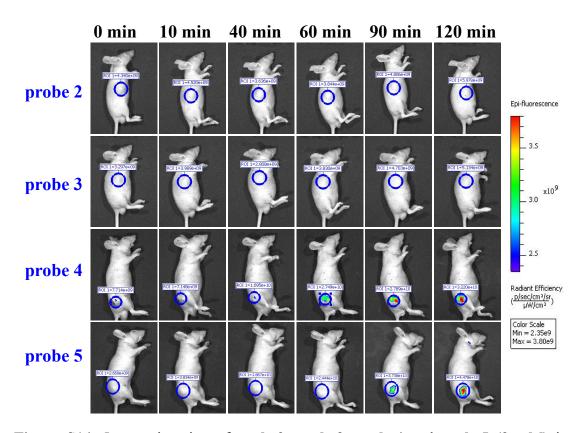


Figure S14. *In vivo* imaging of probe2, probe3, probe4 and probe5 (2 mM) in ACHN-bearing nude mice over time. Green channel: $\lambda_{ex} = 465$ nm, $\lambda_{em} = 520-580$ nm.

8. Additional References

Y. Wen, N. Jing, F. H. Huo, C. X. Yin*, Rational design of a turn-on fluorescent probe for visualization of GRP78 protein in tumor models, *Chin. Chem. Lett.*, 2023, 34, 107604.