# Cu-Catalyzed Selective Coupling of Alkynes with danB-Bpai

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### 1. General Considerations

All manipulations were conducted with Schlenk tube.  $^{1}$ H-NMR spectra were recorded on BrukerAVIII-400 spectrometers, JNM-ECZ400S/L1 and JNM-ECZ600R/S1 spectrometers. Chemical shifts (in ppm) were referenced to tetramethylsilane ( $\delta = 0$  ppm) in CDCl<sub>3</sub> as an internal standard and DMSO- $D_6$  ( $\delta = 2.5$  ppm).  $^{13}$ C-NMR spectra were obtained by using the same NMR spectrometers and were calibrated with CDCl<sub>3</sub> ( $\delta = 77.00$  ppm) and DMSO- $D_6$  ( $\delta = 39.52$  ppm).  $^{11}$ B-NMR spectra were obtained by using the same NMR spectrometers.  $^{19}$ F-NMR spectra were obtained by the same NMR High resolution mass spectrometry (HRMS) data were obtained on a QTOF mass analyzer with electrospray ionization (ESI) through a Waters Acquity UPLC Class I/Xevo G2 Q-Tof. Substrates were purchased from Aldrich, TCI, Acros, Energy, Aladdin, or synthesized according to the procedures outlined below. Unless otherwise noted, materials obtained from commercial suppliers were used without further purification.

# 2. The effect of different reaction conditions

Table S1 The effect of different copper catalyst<sup>a</sup>

Entry	[Cu]	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	CuBr SMe <sub>2</sub>	57	18
2	CuCN	12	2
3	$Cu(OAc)_2$	33	0
4	CuO	0	0
5	$CuF_2$	89	trace
6	$CuCl_2$	70	10
7	$CuBr_2$	56	30
8	Cu(acac) <sub>2</sub>	86	0

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), Copper catalyst (0.02 mmol, 10 mol%), L<sub>1</sub>(0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S2. The effect of different ligands on the formation of alkenyl Bdan<sup>a</sup>

Entry	Ligand	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	$L_1$	89	trace
2	$L_2$	90	0
3	$L_3$	82	trace
4	$L_4$	0	0
5	$L_5$	49	5

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (0.02 mmol, 10 mol%), Ligand (0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S3. The effect of different base on the formation of alkenyl Bdan<sup>a</sup>

Entry	Base	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	MeOLi	90	trace
2	MeONa	93	trace
3	MeOK	0	0
4	<sup>t</sup> BuOLi	65	trace
5	<sup>t</sup> BuONa	30	trace
6	<sup>t</sup> BuOK	0	0

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (0.02 mmol, 10 mol%), PPh<sub>3</sub> (0.02 mmol, 10 mol%), Base (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S4. The effect of different solvent on the formation of alkenyl  $\operatorname{Bdan}^a$ 

Entry	Solvent	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	<sup>i</sup> Pr <sub>2</sub> O	93	trace
2	DME	61	trace
3	Dioxane	25	trace
4	THF	88	trace
5	ClCH <sub>2</sub> CH <sub>2</sub> Cl	82	trace
6	Cyclohexane	96	trace
7	DMA	29	trace
8	CH <sub>3</sub> CN	49	trace

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (0.02 mmol, 10 mol%), PPh<sub>3</sub> (0.02 mmol, 10 mol%), MeONa (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), Solvent (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S5. The effect of different proton source on the formation of alkenyl  $\operatorname{Bdan}^a$ 

Entry	[H]	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	MeOH	96	trace
2	EtOH	96	trace
3	<sup>i</sup> PrOH	92	trace
4	CH <sub>3</sub> COOH	8	11

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (0.02 mmol, 10 mol%), PPh<sub>3</sub> (0.02 mmol, 10 mol%), MeONa (0.3 mmol, 1.5 equiv.), [H] (0.4 mmol, 2.0 equiv.), Cyclohexane (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S6. The effect of different temperature on the formation of alkenyl Bdan<sup>a</sup>

Entry	T °C	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	65	96	trace
2	55	96	trace
3	45	96	trace
4	35	96	trace
5	25	87	trace

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (0.02 mmol, 10 mol%), PPh<sub>3</sub> (0.02 mmol, 10 mol%), MeONa (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), Cyclohexane (1.5 mL),T °C, 12 h. <sup>b</sup> Isolated yield.

Table S7. The effect of different ligands on the formation of alkenyl  $\operatorname{Bpai}^a$ 

Entry	L	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	$L_1$	56	30
2	$L_2$	59	27
3	$L_3$	8	77
4	$L_4$	42	trace
5	$L_5$	57	trace
6	Xantphos	53	47

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (0.02 mmol, 10 mol%), L (0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S8. The effect of different base on the formation of alkenyl  $\operatorname{Bpai}^a$ 

Entry	Base	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	MeOLi	8	77
2	MeONa	70	0
3	MeOK	58	0
4	<sup>t</sup> BuOLi	8	60
5	<sup>t</sup> BuONa	66	0
6	<sup>t</sup> BuOK	28	trace

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (0.02 mmol, 10 mol%), PCy<sub>3</sub> (0.02 mmol, 10 mol%), Base (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S9. The effect of different solvent on the formation of alkenyl  $\operatorname{Bpai}^a$ 

Entry	Solvent	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	<sup>i</sup> Pr <sub>2</sub> O	8	77
2	DME	17	7
3	Dioxane	0	0
4	THF	21	trace
5	ClCH <sub>2</sub> CH <sub>2</sub> Cl	7	49
6	Cyclohexane	17	44
7	DMA	41	0
8	CH <sub>3</sub> CN	12	5

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (0.02 mmol, 10 mol%), PCy<sub>3</sub> (0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), Solvent (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S10. The effect of different proton source on the formation of alkenyl Bpai<sup>a</sup>

Entry	[H]	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	MeOH	8	77
2	$H_2O$	14	13
3	EtOH	18	46
4	<sup>i</sup> PrOH	12	23
5	CH <sub>3</sub> COOH	0	0

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (0.02 mmol, 10 mol%), PCy<sub>3</sub> (0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), [H] (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), 65 °C, 12 h. <sup>b</sup> Isolated yield.

Table S11. The effect of different temperature on the formation of alkenyl  $Bpai^a$ 

Entry	T °C	<b>2a</b> yield% <sup>b</sup>	<b>3a</b> yield% <sup>b</sup>
1	70	5	31
2	65	8	77
3	55	8	73
4	45	10	57
5	35	6	10

<sup>&</sup>lt;sup>a</sup> **1a** (0.2 mmol, 1.0 equiv.), danB-Bpai (0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (0.02 mmol, 10 mol%), PCy<sub>3</sub> (0.02 mmol, 10 mol%), MeOLi (0.3 mmol, 1.5 equiv.), MeOH (0.4 mmol, 2.0 equiv.), <sup>i</sup>Pr<sub>2</sub>O (1.5 mL), T °C, 12 h. <sup>b</sup> Isolated yield.

## 3. General procedure for making products

General procedure A:

In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (83.04 mg, 0.24 mmol, 1.2 equiv.), CuF<sub>2</sub> (2.1 mg, 0.02 mmol, 10 mol%), PPh<sub>3</sub> (5.3 mg, 0.02 mmol, 10 mol%), and MeONa (16.2 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under N<sub>2</sub> flow (this sequence was repeated three times). Anhydrous Cyclohexane (1.5 mL), alkenyl (0.2 mmol, 1.0 equiv.), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under N<sub>2</sub>. The tube was stirred at 35 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE/EA) to afford the product 2.

### General procedure B:

$$R^{1} = R^{2} + danB-Bpai \qquad \begin{array}{c} CuBr_{2} \ (10 \ mol\%), \ PCy_{3} \ (10 \ mol\%) \\ \hline MeOLi \ (1.5 \ equiv.), \ MeOH \ (2.0 \ equiv.) \\ \hline {}^{j}Pr_{2}O \ (1.5 \ mL), \ 65 \ {}^{o}C, \ 12 \ h \\ \hline \\ R^{2} \\ \hline \\ 3 \\ \end{array}$$

In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (83.04 mg, 0.24 mmol, 1.2 equiv.), CuBr<sub>2</sub> (4.5 mg, 0.02 mmol, 10 mol%), PCy<sub>3</sub> (5.6 mg, 0.02 mmol, 10 mol%), and MeOLi (11.4 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under  $N_2$  flow (this sequence was repeated three times). Anhydrous iPr<sub>2</sub>O (1.5 mL), alkenyl (0.2 mmol, 1.0 equiv.), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under  $N_2$ . The tube was stirred at 65 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and  $H_2$ O, then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE/EA) to afford the product 3.

### 4. General Procedure for danB-Bpai

A 100 mL round-bottom flask equipped with a stirring bar was charged with (1S,2S,3R,5S)-(+)-2,3-pinanediol (1.7)10 mmol, g, tetrakis(dimethylamino)diboron (4.0 g, 20 mmol, 2.0 equiv.) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL). The reaction mixture was stirred at room temperature for 4 hours, after which 1,8-diaminonaphthalene (4.7 g, 30 mmol, 3.0 equiv.) was added to the system and stirring was continued at room temperature for 6 hours. The reaction mixture was concentrated by rotary evaporation and purified by silica gel chromatography (PE/EA) to afford the product **danB-Bpai** with 1.8g (53% yield). TLC (PE:EA):  $R_f = 0.7$ ; <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{CDCl}_3) \delta 7.06 \text{ (dd}, J = 8.3, 7.2 \text{ Hz}, 2\text{H}), 6.98 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{H}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{Hz}, 2\text{Hz}, 2\text{Hz}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{Hz}, 2\text{Hz}, 2\text{Hz}), 6.26 \text{ (dd}, J = 8.4, 1.0 \text{ Hz}, 2\text{Hz}, 2\text{$ 7.3, 1.0 Hz, 2H), 6.18 (s, 2H), 4.28 (dd, J = 8.8, 1.9 Hz, 1H), 2.41 – 2.33 (m, 1H), 2.22 (m, 1H), 2.07 (t, J = 5.5 Hz, 1H), 1.96 - 1.85 (m, 2H), 1.42 (s, 3H), 1.31 (s, 3H), 1.10 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H)ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 140.5, 136.4, 127.5, 121.1, 117.6, 105.5, 86.2, 77.6, 51.1, 39.6, 38.0, 35.3, 28.8, 27.1, 26.6, 24.0 ppm; <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 28.3 (large singlets) ppm;  $[\alpha]_D^{25} = +11.2$  (c=1.0, CH<sub>2</sub>Cl<sub>2</sub>), **HRMS** (ESI) m/z calcd for C<sub>20</sub>H<sub>25</sub>B<sub>2</sub>N<sub>2</sub>O<sub>2</sub> (M + H)<sup>+</sup> :347.2102, found 347.2109. This compound can be separated by silica gel column chromatography and does not deteriorate when stored at room temperature.

## 5. Analytical data for compounds

(E)-2-styryl-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2a)<sup>[1]</sup>: The general procedure was followed, using **phenylacetylene** (1a, 20.4 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE: EA = 100: 1) afforded product 2a as a yellow solid (52.0 mg, 96% yield);  $\mathbf{R_f} = 0.2$  (PE: EA = 50: 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.51 – 7.47 (m, 2H), 7.39 – 7.34 (m, 2H), 7.33 – 7.28 (m, 1H), 7.13 (d, J = 1.1 Hz, 1H), 7.12 – 7.08 (m, 2H), 7.02 (dd, J = 8.3, 1.1 Hz, 2H), 6.33 (dd, J = 7.3, 1.1 Hz, 2H), 6.28 (d, J = 18.6 Hz, 1H), 5.80 (s, 2H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 143.6, 141.1, 137.5, 136.3, 128.7, 127.6, 126.8, 119.8, 117.6, 105.8 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>) δ 27.5 (large singlets) ppm.

(E)-2-(2-methylstyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2b)<sup>[2]</sup>: The general procedure was followed, using 2-methylphenylacetylene (1b, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2b as a yellow solid (55.7 mg, 98% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.56 (dd, J = 5.4, 3.7 Hz, 1H), 7.37 (d, J = 18.5 Hz, 1H), 7.23 – 7.16 (m, 3H), 7.12 (dd, J = 8.2, 7.2 Hz, 2H), 7.02 (dd, J = 8.3, 1.0 Hz, 2H), 6.36 (dd, J = 7.3, 1.0 Hz, 2H), 6.20 (d, J = 18.4 Hz, 1H), 5.82 (s, 2H), 2.44 (s, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.2, 141.1, 136.8, 136.3, 135.8, 130.5, 128.4, 127.6, 126.2, 125.6, 119.8, 117.6, 105.8, 19.9 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  27.7 (large singlets) ppm.

(E)-2-(3-methylstyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2c)[: The general procedure was followed, using 3-methylphenylacetylene (1c, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2c as a yellow solid (55.7 mg, 98% yield);  $\mathbf{R_f} = 0.2$  (PE : EA= 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.34 – 7.23 (m, 3H), 7.15 – 7.06 (m, 4H), 7.02 (dd, J = 8.3, 1.0 Hz, 2H), 6.33 (dd, J = 7.3, 1.0 Hz, 2H), 6.27 (d, J = 18.6 Hz, 1H), 5.80 (s, 2H), 2.37 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>) δ 143.7, 141.1, 138.2, 137.4, 136.3, 129.5, 128.5, 127.6, 127.5, 123.9, 119.8, 117.5, 105.7, 21.4 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>) δ 27.6 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C_{19}H_{18}BN_2}$  (M + H)<sup>+</sup> :285.1563, found 285.1566.

(E)-2-(4-methylstyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2d)<sup>[2]</sup>: The general procedure was followed, using 4-methylphenylacetylene (1d, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2d as a yellow solid (54.9 mg, 96% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.44 – 7.34 (m, 2H), 7.17 (d, J = 7.8 Hz, 2H), 7.13 – 7.06 (m, 3H), 7.01 (dd, J = 8.4, 1.0 Hz, 2H), 6.33 (dd, J = 7.2, 1.0 Hz, 2H), 6.22 (d, J = 18.5 Hz, 1H), 5.80 (s, 2H), 2.35 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>) δ 143.5, 141.1, 138.7, 136.3, 134.8, 129.4, 127.6, 126.7, 119.8, 117.5, 105.7, 21.3 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>) δ 27.5 (large singlets) ppm.

(E)-2-(4-(tert-butyl)styryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2e)<sup>[3]</sup>: The general procedure was followed, using 4-(tert-butyl)phenylacetylene (1e, 31.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2e as a yellow solid (63.9 mg, 98% yield);  $\mathbf{R_f} = 0.3$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 (d, J = 8.4 Hz, 2H), 7.40 (d, J = 8.5 Hz, 2H), 7.19 – 7.08 (m, 3H), 7.02 (dd, J = 8.3, 1.0 Hz, 2H), 6.36 (dd, J = 7.3, 1.0 Hz, 2H), 6.28 (d, J = 18.6 Hz, 1H), 5.85 (s, 2H), 1.34 (s, 9H)ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.0, 143.5, 141.2, 136.3,

134.8, 127.6, 126.5, 125.6, 119.8, 117.5, 105.7, 34.7, 31.2 ppm (the carbon next to boron could not be detected);  $^{11}B$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.3 (large singlets) ppm.

(E)-2-(4-(trimethylsilyl)styryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborini (2f): The general procedure was followed, ne using (4-Ethynylphenyl)trimethylsilane (1f, 34.9 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product **2f** as a yellow solid (67.1 mg, 98% yield);  $\mathbf{R_f} = 0.3$  (PE : EA =50 : 1); <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.57 – 7.45 (m, 4H), 7.15 (d, J = 10.4 Hz, 1H), 7.13 - 7.09 (m, 2H), 7.02 (dd, J = 8.3, 1.0 Hz, 2H), 6.38 - 6.31 (m, 3H), 5.85 (s, 2H), 0.28 (s, 9H)ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 143.7, 141.4, 141.1, 137.8, 136.3, 133.7, 127.6, 126.0, 119.8, 117.6, 105.7, -1.2 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 27.4 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $C_{21}H_{24}BN_2Si(M + H)^+$ : 343.1802, found 343.1806.

(E)-2-(4-methoxystyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2g)<sup>[4]</sup>: The general procedure was followed, using 4-Ehynylanisole (1g, 26.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 20 : 1) afforded product 2g as a yellow solid (55.4 mg, 92% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 10 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.49 – 7.40 (m, 2H), 7.16 – 7.05 (m, 3H), 7.01 (dd, J = 8.4, 1.0 Hz, 2H), 6.94 – 6.86 (m, 2H), 6.35 (dd, J = 7.3, 1.0 Hz, 2H), 6.14 (d, J = 18.5 Hz, 1H), 5.82 (s, 2H), 3.82 (s, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.1, 143.2, 141.2, 136.3, 130.4, 128.1, 127.6, 119.8, 117.5, 114.1, 105.7, 55.3 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  28.3 (large singlets) ppm.

(E)-3-(2-(1H-naphtho[1,8-de][1,3,2]diazaborinin-2(3H)-yl)vinyl)aniline (2h): The general procedure was followed, using 4-Ethynylaniline (1h, 23.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 5 : 1) afforded product 2h as a yellow solid (56.0 mg, 98% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 2 : 1); <sup>1</sup>H NMR (400 MHz, DMSO- $D_6$ )  $\delta$  7.99 (s, 2H), 7.33 – 7.21 (m, 3H), 7.05 (t, J = 7.8 Hz, 2H), 6.85 (dd, J = 8.3, 1.0 Hz, 2H), 6.67 – 6.55 (m, 2H), 6.45 (dd, J = 7.4, 1.0 Hz, 2H), 6.00 (d, J = 18.5 Hz, 1H), 5.43 (s, 2H) ppm; <sup>13</sup>C NMR (101 MHz, DMSO- $D_6$ )  $\delta$  149.6, 144.7, 142.7, 136.1, 127.9, 127.7, 125.7, 119.5, 115.8, 113.9, 105.1 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, DMSO- $D_6$ )  $\delta$  31.5 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{18}H_{17}BN_3$  (M + H)<sup>+</sup> :286.1515, found 286.1519.

(E)-2-(2-([1,1'-biphenyl]-3-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazab orinine (2i): The general procedure was followed, using 4-biphenylacetylene (1i, 35.7 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product 2i as a yellow solid (65.2 mg, 94% yield);  $\mathbf{R_f} = 0.1$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.64 – 7.60 (m, 4H), 7.57 (d, J = 8.6 Hz, 2H), 7.48 – 7.42 (m, 2H), 7.39 – 7.33 (m, 1H), 7.21 – 7.09 (m, 3H), 7.03 (dd, J = 8.3, 1.0 Hz, 2H), 6.39 – 6.31 (m, 3H), 5.85 (s, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>) δ 143.1, 141.4, 141.1, 140.5, 136.5, 136.3, 128.8, 127.6, 127.5, 127.4, 127.2, 127.0, 119.8, 117.6, 105.8 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>) δ 28.3 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{24}\mathbf{H}_{20}\mathbf{B}\mathbf{N}_{2}$  (M + H) $^{+}$ :347.1719, found 347.1724.

$$F_3C$$
 $\mathbf{2j}$ 
 $\mathbf{Bdan}$ 

(E)-2-(4-(trifluoromethyl)styryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazabori nine (2j)<sup>[4]</sup>: The general procedure was followed, using 4-(Trifluoromethyl)phenylacetylene (1j, 30.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product 2j as a yellow solid (65.6 mg, 97% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 20 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.60 (d, J = 8.2 Hz, 2H), 7.53 (d, J = 8.2 Hz, 2H), 7.14 – 7.01 (m, 5H), 6.40 – 6.30 (m, 3H), 5.80 (s, 2H) ppm; <sup>13</sup>C NMR (101

MHz, CDCl<sub>3</sub>)  $\delta$  141.9, 140.9, 136.3, 130.3, 130.0, 127.6, 126.8, 125.6 (q, J = 14.7 Hz), 122.7, 119.9, 117.8, 105.9 ppm (the carbon next to boron could not be detected); <sup>19</sup>**F NMR** (376 MHz, CDCl<sub>3</sub>)  $\delta$  -62.5 ppm; <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)  $\delta$  27.7 (large singlets) ppm.

(E)-2-(4-chlorostyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2k)<sup>[4]</sup>: The general procedure was followed, using 4-Chlorophenylacetylene (1k, 27.3 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2k as a yellow solid (50.2 mg, 82% yield);  $\mathbf{R_f} = 0.1$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.45 – 7.40 (m, 2H), 7.36 – 7.31 (m, 2H), 7.14 – 7.01 (m, 5H), 6.36 (dd, J = 7.2, 1.1 Hz, 2H), 6.27 (d, J = 18.6 Hz, 1H), 5.83 (s, 2H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  142.2, 141.0, 136.3, 136.0, 134.3, 128.9, 127.9, 127.6, 119.8, 117.7, 105.8 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  27.6 (large singlets) ppm.

(E)-2-(4-bromostyryl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2l)<sup>[4]</sup>: The general procedure was followed, using 4-Bromophenylacetylene (1l, 36.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2l as a yellow solid (68.4 mg, 98% yield);  $\mathbf{R_f} = 0.1$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.49 (d, J = 8.5 Hz, 2H), 7.37 – 7.34 (m, 2H), 7.12 (dd, J = 8.3, 7.2 Hz, 2H), 7.05 – 7.01 (m, 3H), 6.35 (dd, J = 7.3, 1.1 Hz, 2H), 6.29 (d, J = 18.5 Hz, 1H), 5.82 (s, 2H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  142.3, 141.0, 136.4, 136.3, 131.8, 128.2, 127.6, 122.6, 119.8, 117.7, 105.8 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  27.6 (large singlets) ppm.

# (E)-4-(2-(1H-naphtho[1,8-de][1,3,2]diazaborinin-2(3H)-yl)vinyl)benzaldehyde

(2m): The general procedure was followed, using **4-Ethynylbenzaldehyde** (1m, 26.0 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 10 : 1) afforded product **2m** as a yellow solid (60.6 mg, 97% yield); **R**<sub>f</sub> = 0.2 (PE : EA = 5 : 1); <sup>1</sup>**H NMR** (400 MHz, DMSO- $D_6$ )  $\delta$  10.01 (s, 1H), 8.19 (s, 2H), 7.95 (d, J = 8.0 Hz, 2H), 7.74 (d, J = 8.0 Hz, 2H), 7.52 (d, J = 18.4 Hz, 1H), 7.06 (t, J = 7.8 Hz, 2H), 6.88 (d, J = 8.1 Hz, 2H), 6.57 (d, J = 18.6 Hz, 1H), 6.46 (d, J = 7.3 Hz, 2H) ppm; <sup>13</sup>**C NMR** (101 MHz, DMSO- $D_6$ )  $\delta$  192.6, 143.4, 142.6, 142.3, 136.1, 135.8, 130.2, 127.7, 127.1, 119.8, 116.2, 105.4 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, DMSO- $D_6$ )  $\delta$  30.4 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>19</sub>H<sub>16</sub>BNO<sub>2</sub> (M + H)<sup>+</sup> :299.1355, found 299.1359.

(E)-1-(4-(2-(1H-naphtho[1,8-de][1,3,2]diazaborinin-2(3H)-yl)vinyl)phenyl)ethan-1-one (2n)<sup>[2]</sup>: The general procedure was followed, using 4-Acetylphenylacetylene (1n , 28.8 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 10 : 1) afforded product 2n as a yellow solid (61.2 mg, 98% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 5 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.95 (d, J = 8.2 Hz, 2H), 7.55 (d, J = 8.2 Hz, 2H), 7.18 – 7.09 (m, 3H), 7.07 – 7.00 (m, 2H), 6.43 (d, J = 18.6 Hz, 1H), 6.36 (dd, J = 7.2, 1.0 Hz, 2H), 5.88 (s, 2H), 2.61 (s, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  197.6, 142.3, 141.9, 140.9, 136.7, 136.3, 128.8, 127.6, 126.8, 119.9, 117.8, 105.9, 26.7 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  28.1 (large singlets) ppm.

(E)-4-(2-(1H-naphtho[1,8-de][1,3,2]diazaborinin-2(3H)-yl)vinyl) methyl benzoate (2o): The general procedure was followed, using 4-(Methoxycarbonyl)phenylacetylene (1o, 32.0 mg, 0.2 mmol), danB-Bpai (83.04)

mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 10 : 1) afforded product **20** as a yellow solid (64.3 mg, 98% yield);  $\mathbf{R_f} = 0.3$  (PE : EA = 5 : 1); <sup>1</sup>**H NMR** (400 MHz, DMSO- $D_6$ )  $\delta$  8.17 (s, 2H), 8.04 – 7.97 (m, 2H), 7.69 – 7.62 (m, 2H), 7.49 (d, J = 18.6 Hz, 1H), 7.06 (dd, J = 8.2, 7.4 Hz, 2H), 6.87 (dd, J = 8.3, 1.0 Hz, 2H), 6.52 (d, J = 18.7 Hz, 1H), 6.45 (dd, J = 7.4, 1.0 Hz, 2H), 3.86 (s, 3H) ppm; <sup>13</sup>**C NMR** (101 MHz, DMSO- $D_6$ )  $\delta$  166.0, 142.6, 142.3, 142.2, 136.1, 129.9, 129.1, 127.7, 126.7, 119.8, 116.2, 105.4, 52.2 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, DMSO- $D_6$ )  $\delta$  31.9 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $C_{20}H_{18}BN_2O_2$  (M + H)<sup>+</sup> :329.1461, found 329.1465.

(E)-2-(2-(naphthalen-2-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazabori nine (2p): The general procedure was followed, using 2-Ethynyl-naphthalene (1p, 30.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 20 : 1) afforded product 2p as a yellow solid (56.8 mg, 89% yield);  $\mathbf{R_f} = 0.4$  (PE : EA= 10 : 1);  $^1\mathbf{H}$  NMR (400 MHz, DMSO- $D_6$ )  $\delta$  8.20 (s, 2H), 8.01 – 7.89 (m, 4H), 7.78 (dd, J = 8.6, 1.7 Hz, 1H), 7.63 (d, J = 18.6 Hz, 1H), 7.56 – 7.48 (m, 2H), 7.08 (t, J = 7.8 Hz, 2H), 6.89 (dd, J = 8.3, 1.0 Hz, 2H), 6.57 – 6.46 (m, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, DMSO- $D_6$ )  $\delta$  143.9, 142.5, 136.1, 135.3, 133.2, 133.1, 128.5, 128.3, 127.7, 127.7, 126.9, 126.6, 126.5, 123.5, 123.2, 119.8, 116.1, 105.3 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, DMSO- $D_6$ )  $\delta$  31.3 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{22}\mathbf{H}_{18}\mathbf{B}\mathbf{N}_{2}$  (M + H) $^+$  :321.1563, found 321.1567.

(E)-2-(2-(naphthalen-1-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazabori nine (2q): The general procedure was followed, using 1-Ethynyl-naphthalene (1q, 30.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2q as a yellow solid (58.1 mg, 91% yield);  $\mathbf{R_f} = 0.2$  (PE : EA= 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.22 (dd, J = 8.4, 1.3 Hz, 1H), 7.95 – 7.81 (m, 3H), 7.71 (dt, J = 7.2, 1.0 Hz, 1H), 7.58 – 7.46 (m, 3H), 7.13 (dd, J = 8.3, 7.2 Hz, 2H), 7.04 (dd, J = 8.3, 1.1 Hz, 2H), 6.40 – 6.33 (m, 3H), 5.89 (s, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.1, 140.6, 136.3, 135.5, 133.6, 131.0, 128.8, 128.6, 127.6, 126.3, 125.9, 125.6, 124.0, 123.5, 119.9, 117.6, 105.8 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR

(128 MHz, CDCl<sub>3</sub>)  $\delta$  28.0 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $C_{22}H_{18}BN_2 (M + H)^+$ :321.1563, found 321.1567.

2r

(E)-2-(2-(thiophen-2-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborini ne (2r): The general procedure was followed, using 2-Ethynylthiophene (1r, 21.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product 2r as a yellow solid (44.8 mg, 81% yield);  $\mathbf{R_f} = 0.5$  (PE : EA = 10 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.26 – 7.21 (m, 2H), 7.10 (dd, J = 8.3, 7.2 Hz, 2H), 7.07 – 7.05 (m, 1H), 7.03 – 6.98 (m, 3H), 6.31 (dd, J = 7.2, 1.1 Hz, 2H), 6.04 (d, J = 18.3 Hz, 1H), 5.75 (s, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>) δ 144.0, 141.0, 136.3, 136.1, 127.7, 127.5, 127.1, 125.8, 119.8, 117.6, 105.8 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>) δ 27.6 (large singlets) ppm; HRMS (ESI) m/z calcd for  $\mathbf{C}_{16}\mathbf{H}_{14}\mathbf{B}\mathbf{N}_{2}\mathbf{S}$  (M + H) $^{+}$  :277.0970, found 277.0974.

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(E)-2-(2-(thiophen-3-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborini ne (2s): The general procedure was followed, using 3-Ethynylthiophene (1s, 21.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product 2s as a yellow solid (51.6 mg, 93% yield);  $\mathbf{R_f} = 0.1$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.31 – 7.26 (m, 3H), 7.13 – 7.05 (m, 3H), 7.01 (dd, J = 8.2, 1.1 Hz, 2H), 6.31 (dd, J = 7.2, 1.1 Hz, 2H), 6.04 (d, J = 18.5 Hz, 1H), 5.75 (s, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>) δ 141.1, 141.1, 137.3, 136.3, 127.5, 126.3, 124.8, 124.0, 119.7, 117.5, 105.7 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>) δ 27.9 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{16}\mathbf{H}_{12}\mathbf{B}\mathbf{N}_{2}$  (M - H) $^{-}$  :275.0814, found 275.0817.

2t

(E)-2-(2-(pyridin-3-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2t): The general procedure was followed, using 3-Ethynylpyridine (1t, 20.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by

chromatography on silica gel (PE : EA = 5 : 1) afforded product **2t** as a yellow solid (26.2 mg, 48% yield);  $\mathbf{R_f} = 0.5$  (EA);  $^1\mathbf{H}$  NMR (400 MHz, DMSO- $D_6$ )  $\delta$  8.70 (d, J = 2.2 Hz, 1H), 8.52 (dd, J = 4.7, 1.6 Hz, 1H), 8.17 (s, 2H), 7.96 (d, J = 8.0 Hz, 1H), 7.50 – 7.40 (m, 2H), 7.06 (t, J = 7.8 Hz, 2H), 6.88 (d, J = 8.2 Hz, 2H), 6.54 – 6.42 (m, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, DMSO- $D_6$ )  $\delta$  149.4, 148.3, 142.3, 140.4, 136.1, 133.2, 132.8, 127.7, 124.1, 119.8, 116.1, 105.3 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  30.5 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{17}\mathbf{H}_{15}\mathbf{B}\mathbf{N}_3$  (M + H) $^+$ :272.1359, found 272.1362.

**2**u

(E)-2-(2-(quinolin-3-yl)vinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinin e (2u): The general procedure was followed, using 3-Ethynylquinoline (1u, 30.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 5 : 1) afforded product 2u as a yellow solid (56.0 mg, 87% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 1 : 1);  $^1\mathbf{H}$  NMR (400 MHz, DMSO- $D_6$ ) δ 9.14 (d, J = 2.2 Hz, 1H), 8.41 (d, J = 2.2 Hz, 1H), 8.23 (s, 2H), 8.09 – 8.00 (m, 2H), 7.76 (m, 1H), 7.69 – 7.59 (m, 2H), 7.07 (t, J = 7.8 Hz, 2H), 6.94 – 6.84 (m, 2H), 6.68 (d, J = 18.8 Hz, 1H), 6.49 (dd, J = 7.4, 1.0 Hz, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, DMSO- $D_6$ ) δ 149.2, 147.3, 142.4, 140.6, 136.1, 132.8, 130.6, 129.8, 128.7, 128.6, 127.7, 127.1, 125.9, 119.8, 116.2, 105.4 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, DMSO- $D_6$ ) δ 30.7 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{21}\mathbf{H}_{17}\mathbf{B}\mathbf{N}_3$  (M + H)<sup>+</sup> :322.1515, found 322.1519.

2v

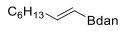
(Z)-2-(1,2-diphenylvinyl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2v)<sup>[1]</sup>: The general procedure was followed, using **Diphenylacetylene** (1v, 35.6 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2v as a yellow solid (43.5 mg, 63% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.41 – 7.30 (m, 3H), 7.18 – 6.99 (m, 13H), 6.29 (dd, J = 7.2, 1.1 Hz, 2H), 5.68 (s, 2H) ppm ; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.0, 140.8, 137.3, 136.7, 136.3, 129.8, 129.0, 128.6, 128.0, 127.6, 127.5, 126.7, 119.7, 117.6, 105.9 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  28.9 (large singlets) ppm.

2w

(Z)-2-(1-phenylprop-1-en-2-yl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborin ine (2w)<sup>[5]</sup>: The general procedure was followed, using 1-Phenylpropyne (1w, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2v as a yellow solid (46.4 mg, 78% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.41 – 7.36 (m, 2H), 7.34 – 7.29 (m, 1H), 7.18 – 7.13 (m, 5H), 7.11 – 7.07 (m, 3H), 7.03 (dd, J = 7.3, 1.8 Hz, 3H), 7.00 (d, J = 1.1 Hz, 1H), 6.29 (dd, J = 7.2, 1.1 Hz, 2H), 5.68 (s, 2H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.1, 137.7, 136.9, 136.3, 129.2, 128.1, 127.5, 127.0, 119.7, 117.5, 105.8, 15.8 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.6 (large singlets) ppm.

2x

(Z)-2-(1-phenylbut-1-en-2-yl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborini ne (2x)<sup>[5]</sup>: The general procedure was followed, using 1-Butynylbenzene (1x, 26.0 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2x as a yellow solid (42.1 mg, 74% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.41 – 7.34 (m, 2H), 7.34 – 7.26 (m, 3H), 7.12 (dd, J = 8.3, 7.2 Hz, 2H), 7.03 (dd, J = 8.3, 1.0 Hz, 2H), 6.95 (s, 1H), 6.37 (dd, J = 7.3, 1.0 Hz, 2H), 5.84 (s, 2H), 2.43 (qd, J = 7.5, 1.0 Hz, 2H), 1.15 (t, J = 7.5 Hz, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.1, 137.8, 136.3, 136.1, 128.7, 128.2, 127.6, 127.0, 119.8, 117.6, 105.8, 22.7, 14.9 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.0 (large singlets) ppm.



(E)-2-(oct-1-en-1-yl)-2,3-dihydro-1H-naphtho[1,8-de][1,3,2]diazaborinine (2y)<sup>[6]</sup>: The general procedure was followed, using 1-Octyne (1y, 22.0 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 2y as a pale yellow oil (49.5 mg, 89% yield);  $\mathbf{R_f} = 0.3$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.13 – 6.94 (m, 4H), 6.38 – 6.23 (m, 3H), 5.66 (s, 2H), 5.52 (dt, J = 18.0, 1.6 Hz, 1H), 2.25 – 2.10 (m, 2H), 1.42 (q, J = 7.5 Hz, 2H), 1.37 – 1.22 (m, 6H), 0.96 – 0.83 (m, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  148.1, 141.2, 136.3, 127.5, 119.7, 117.3, 105.5, 35.9, 31.7,

28.9, 28.6, 22.6, 14.1 ppm (the carbon next to boron could not be detected);  $^{11}B$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  27.3 (large singlets) ppm.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-styryl)hexahydro-4,6-methanobenzo[d][  $(3a)^{[7]}$ : 1,3,2|dioxaborole The general procedure followed, was phenylacetylene (1a, 20.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3a as a pale yellow oil (43.4 mg, 77% yield);  $\mathbf{R_f} = 0.4$  (PE: EA = 50 : 1); <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.53 – 7.47 (m, 2H), 7.41 (d, J = 18.4 Hz, 1H), 7.36 - 7.27 (m, 3H), 6.20 (d, J = 18.5 Hz, 1H), 4.37 (dd, J = 8.8, 1.9 Hz, 1H), 2.38 (m, 1H), 2.24 (m, 1H), 2.11 (dd, J = 6.0, 4.8 Hz, 1H), 1.97 – 1.90 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.20 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  149.4, 137.4, 128.8, 128.5, 127.0, 85.8, 77.8, 51.3, 39.5, 38.1, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>11</sup>B NMR (128) MHz, CDCl<sub>3</sub>) δ 29.5 (large singlets) ppm.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-2-methylstyryl)hexahydro-4,6-methano benzo[d][1,3,2]dioxaborole (3b): The general procedure was followed, using 2-methylphenylacetylene (1b, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3b as a pale yellow oil (43.4 mg, 73% yield);  $\mathbf{R_f} = 0.3$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.66 (d, J = 18.3 Hz, 1H), 7.60 – 7.56 (m, 1H), 7.21 – 7.17 (m, 2H), 7.16 – 7.13 (m, 1H), 6.11 (d, J = 18.3 Hz, 1H), 4.37 (dd, J = 8.8, 1.9 Hz, 1H), 2.42 (s, 3H), 2.40 – 2.34 (m, 1H), 2.29 – 2.21 (m, 1H), 2.11 (dd, J = 6.1, 4.8 Hz, 1H), 1.98 – 1.90 (m, 2H), 1.45 (s, 3H), 1.31 (s, 3H), 1.22 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>) δ 147.0, 136.7, 130.4, 128.6, 126.1, 125.7, 85.8, 77.8, 51.3, 39.5, 38.2, 35.5, 28.7, 27.1, 26.5, 24.0, 19.8 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>) δ 30.1 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>19</sub>H<sub>26</sub>BO<sub>2</sub> (M + H) + :297.2026, found 297.2029.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-3-methylstyryl)hexahydro-4,6-methano benzo[d][1,3,2]dioxaborole (3c): The general procedure was followed, using 3-methylphenylacetylene (1c, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3c as a pale yellow oil (37.1 mg, 63% yield);  $\mathbf{R_f} = 0.3$  (PE : EA= 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 (d, J = 18.4 Hz, 1H), 7.33 – 7.28 (m, 2H), 7.26 – 7.20 (m, 1H), 7.10 (dt, J = 7.3, 1.6 Hz, 1H), 6.17 (d, J = 18.4 Hz, 1H), 4.36 (dd, J = 8.7, 1.9 Hz, 1H), 2.43 – 2.37 (m, 1H), 2.36 – 2.33 (m, 3H), 2.24 (m, 1H), 2.11 (dd, J = 6.0, 4.8 Hz, 1H), 1.96 – 1.89 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.20 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  149.6, 138.0, 137.4, 129.7, 128.4, 127.7, 124.2, 85.7, 77.8, 51.3, 39.5, 38.1, 35.5, 28.6, 27.1, 26.4, 24.0, 21.4 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.4 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{19}H_{26}BO_2$  (M + H) $^+$  :297.2026, found 297.2029.

**3**d

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-4-methylstyryl)hexahydro-4,6-methano benzo[d][1,3,2]dioxaborole (3d): The general procedure was followed, using 4-methylphenylacetylene (1d, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3d as a pale yellow solid (38.4 mg, 64% yield);  $\mathbf{R_f} = 0.3$ (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.42 – 7.35 (m, 3H), 7.15 (d, J = 7.9 Hz, 2H), 6.13 (d, J = 18.4 Hz, 1H), 4.37 (dd, J = 8.7, 1.9 Hz, 1H), 2.43 – 2.36 (m, 1H), 2.35 (s, 3H), 2.28 – 2.21 (m, 1H), 2.11 (dd, J = 6.1, 4.9 Hz, 1H), 1.93 (m, 2H), 1.45 (s, 3H), 1.31 (s, 3H), 1.20 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  149.4, 139.0, 134.8, 129.3, 127.0, 85.7, 77.8, 51.4, 39.5, 38.2, 35.5, 28.7, 27.1, 26.4, 24.0, 21.3 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  30.8 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>19</sub>H<sub>24</sub>BO<sub>2</sub> (M - H) $^{-}$ :295.1870, found 295.1873.

3€

 $(3aR,4R,6R,7aS)-2-((E)-4-(tert-butyl)styryl)-3a,5,5-trimethylhexahydro-4,6-meth \\anobenzo[d][1,3,2]dioxaborole (3e): The general procedure was followed, using$ 

**4-(tert-butyl)phenylacetylene** (**1e**, 31.6 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product **3e** as a colorless oil (42.8 mg, 63% yield); **R**<sub>f</sub> = 0.4 (PE : EA = 50 : 1); <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.43 (dd, J = 8.6, 6.4 Hz, 3H), 7.38 – 7.35 (m, 2H), 6.15 (d, J = 18.4 Hz, 1H), 4.37 (dd, J = 8.7, 1.9 Hz, 1H), 2.43 – 2.34 (m, 1H), 2.28 – 2.20 (m, 1H), 2.11 (dd, J = 6.0, 4.8 Hz, 1H), 1.97 – 1.90 (m, 2H), 1.45 (s, 3H), 1.31 (d, J = 4.0 Hz, 12H), 1.20 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm; <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 152.1, 149.3, 134.8, 126.8, 125.5, 85.7, 77.8, 51.4, 39.5, 38.2, 35.5, 34.7, 31.2, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 29.5 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $C_{22}H_{32}BO_2$  (M + H)<sup>+</sup> :339.2495, found 339.2499.

(4-((E)-2-((3aR,4R,6R,7aS)-3a,5,5-trimethylhexahydro-4,6-methanobenzo[d][1,3, 2]dioxaborol-2-yl)vinyl)phenyl) trimethyl silane (4t): The general procedure was followed, using (4-Ethynylphenyl)trimethylsilane (1f, 34.9 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3f as a pale yellow oil (47.9 mg, 68% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.52 – 7.45 (m, 4H), 7.40 (d, J = 18.4 Hz, 1H), 6.22 (d, J = 18.4 Hz, 1H), 4.37 (dd, J = 8.7, 1.9 Hz, 1H), 2.38 (m, 1H), 2.23 (m, 1H), 2.11 (dd, J = 6.0, 4.8 Hz, 1H), 1.97 – 1.90 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.21 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H), 0.26 (s, 9H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>) δ 149.5, 141.5, 137.8, 133.6, 126.2, 85.8, 77.8, 51.3, 39.5, 38.1, 35.5, 28.6, 27.1, 26.4, 24.0, -1.2 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>) δ 29.2 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{21}H_{32}BO_2Si$  (M + H) $^+$ :355.2264, found 355.2269.

(3aR,4R,6R,7aS)-2-((E)-4-methoxystyryl)-3a,5,5-trimethylhexahydro-4,6-methan obenzo[d][1,3,2]dioxaborole (3g)<sup>[7]</sup>: The general procedure was followed, using **4-Ehynylanisole** (1g, 26.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product 3g as a pale yellow oil (31.2 mg, 50% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 20 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 – 7.42 (m, 2H), 7.36 (d, J = 18.4 Hz, 1H), 6.89 – 6.84 (m, 2H), 6.04 (d, J = 18.4 Hz, 1H), 4.36 (dd, J = 8.7, 1.9 Hz, 1H), 3.80 (s,

3H), 2.43 - 2.34 (m, 1H), 2.28 - 2.20 (m, 1H), 2.10 (dd, J = 6.0, 4.8 Hz, 1H), 1.97 - 1.89 (m, 2H), 1.44 (s, 3H), 1.30 (s, 3H), 1.20 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm; <sup>13</sup>C **NMR** (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.2, 149.0, 130.3, 128.4, 113.9, 85.6, 77.7, 55.2, 51.3, 39.5, 38.1, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>11</sup>B **NMR** (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.3 (large singlets) ppm.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-4-(trifluoromethyl)styryl)hexahydro-4,6 -methanobenzo[d][1,3,2]dioxaborole (3h): The general procedure was followed, using 4-(Trifluoromethyl)phenylacetylene (1j, 30.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3h as a white solid (39.5 mg, 56% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50 : 1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.59 (d, J = 1.6 Hz, 4H), 7.41 (d, J = 18.5 Hz, 1H), 6.29 (d, J = 18.4 Hz, 1H), 4.39 (dd, J = 8.7, 1.8 Hz, 1H), 2.45 – 2.35 (m, 1H), 2.29 – 2.22 (m, 1H), 2.12 (dd, J = 6.1, 4.9 Hz, 1H), 1.98 – 1.90 (m, 2H), 1.46 (s, 3H), 1.31 (s, 3H), 1.19 (d, J = 11.0 Hz, 1H), 0.88 (s, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 147.6, 140.8, 127.1, 125.5 (q, 14.7 Hz), 86.0, 78.0, 51.3, 39.5, 38.2, 35.4, 28.6, 27.0, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -62.5 ppm; <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>) δ 29.2 (large singlets) ppm; HRMS (ESI) m/z calcd for C<sub>19</sub>H<sub>23</sub>BF<sub>3</sub>O<sub>2</sub> (M + H)<sup>+</sup> :351.1743, found 351.1747.

(3aR,4R,6R,7aS)-2-((E)-2-([1,1'-biphenyl]-4-yl)vinyl)-3a,5,5-trimethylhexahydro-4,6-methanobenzo[d][1,3,2]dioxaborole (3i): The general procedure was followed, using 4-biphenylacetylene (1i, 35.7 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3i as a white solid (43.0 mg, 60% yield);  $\mathbf{R_f} = 0.5$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.62 – 7.56 (m, 6H), 7.48 – 7.40 (m, 3H), 7.36 – 7.32 (m, 1H), 6.23 (d, J = 18.4 Hz, 1H), 4.38 (dd, J = 8.7, 1.8 Hz, 1H), 2.45 – 2.35 (m, 1H), 2.30 – 2.21 (m, 1H), 2.12 (dd, J = 6.1, 4.8 Hz, 1H), 1.99 – 1.91 (m, 2H), 1.46 (s, 3H), 1.31 (s, 3H), 1.22 (d, J = 10.9 Hz, 1H), 0.88 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  148.9, 141.5, 140.5, 136.4, 128.8, 127.5, 127.4, 127.2, 127.0, 85.8, 77.8,

51.3, 39.5, 38.2, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected);  ${}^{11}B$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.2 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{24}H_{26}BO_2$  (M - H) $^{-1}$ : 357.2026, found 357.2030.

(3aR,4R,6R,7aS)-2-((E)-4-bromostyryl)-3a,5,5-trimethylhexahydro-4,6-methano benzo[d][1,3,2]dioxaborole (3j): The general procedure was followed, using 4-Bromophenylacetylene (1l, 36.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3j as a pale yellow solid (43.2 mg, 60% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.48 – 7.44 (m, 2H), 7.37 – 7.29 (m, 3H), 6.17 (d, J = 18.4 Hz, 1H), 4.37 (dd, J = 8.7, 1.9 Hz, 1H), 2.38 (m, 1H), 2.24 (m, 1H), 2.13 – 2.08 (m, 1H), 1.97 – 1.88 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.18 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  148.0, 136.3, 131.7, 128.5, 122.8, 85.9, 77.8, 51.3, 39.4, 38.1, 35.4, 28.6, 27.0, 26.4, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.2 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{18}\mathbf{H}_{23}\mathbf{B}\mathbf{B}\mathbf{r}\mathbf{O}_{2}$  (M + H) $^{+}$  :361.0974, found 361.0978.

**1-(4-((E)-2-((3aR,4R,6R,7aS)-3a,5,5-trimethylhexahydro-4,6-methanobenzo[d][1, 3,2]dioxaborol-2-yl)vinyl)phenyl)ethan-1-one (3k):** The general procedure was followed, using **4-Acetylphenylacetylene (1n**, 28.8 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product **3k** as a colorless oil (34.7 mg, 54% yield); **R**<sub>f</sub> = 0.2 (PE : EA = 20 : 1); <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.99 – 7.89 (m, 2H), 7.60 – 7.54 (m, 2H), 7.42 (d, J = 18.4 Hz, 1H), 6.32 (d, J = 18.4 Hz, 1H), 4.39 (dd, J = 8.7, 1.8 Hz, 1H), 2.60 (s, 3H), 2.40 (m, 1H), 2.25 (m, 1H), 2.12 (dd, J = 6.1, 4.9 Hz, 1H), 1.99 – 1.90 (m, 2H), 1.46 (s, 3H), 1.31 (s, 3H), 1.19 (d, J = 11.0 Hz, 1H), 0.88 (s, 3H) ppm; <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 197.57, 147.93, 141.80, 136.93, 128.70, 127.07, 86.02, 77.94, 51.28, 39.45, 38.17, 35.45, 28.61, 27.05, 26.65, 26.42, 24.01 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 29.88 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>20</sub>H<sub>26</sub>BO<sub>3</sub> (M + H)<sup>+</sup> :325.1975, found 325.1979.

**4-((E)-2-((3aR,4R,6R,7aS)-3a,5,5-trimethylhexahydro-4,6-methanobenzo[d][1,3,2**] **dioxaborol-2-yl)vinyl) methyl benzoate (3l):** The general procedure was followed, using **4-(Methoxycarbonyl)phenylacetylene (1o**, 32.0 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 50 : 1) afforded product **3l** as a white solid (41.5 mg, 61% yield); **R**<sub>f</sub> = 0.3 (PE : EA = 20 : 1); <sup>11</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.04 – 7.99 (m, 2H), 7.54 (d, J = 8.4 Hz, 2H), 7.42 (d, J = 18.5 Hz, 1H), 6.30 (d, J = 18.4 Hz, 1H), 4.38 (dd, J = 8.8, 1.8 Hz, 1H), 3.91 (s, 3H), 2.45 – 2.35 (m, 1H), 2.29 – 2.22 (m, 1H), 2.11 (dd, J = 6.1, 4.9 Hz, 1H), 1.97 – 1.90 (m, 2H), 1.46 (s, 3H), 1.31 (s, 3H), 1.19 (d, J = 11.0 Hz, 1H), 0.88 (s, 3H) ppm; <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 166.8, 148.1, 141.7, 130.1, 129.9, 126.9, 86.0, 77.9, 52.1, 51.3, 39.5, 38.2, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 29.8 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>20</sub>H<sub>26</sub>BO<sub>4</sub> (M + H)<sup>+</sup> :341.1924, found 341.1928.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-2-(naphthalen-2-yl)vinyl)hexahydro-4,6 -methanobenzo[d][1,3,2]dioxaborole (3m): The general procedure was followed, using 2-Ethynyl-naphthalene (1p, 30.5 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3m as a white solid (40.9 mg, 62% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>) δ 7.86 – 7.77 (m, 4H), 7.75 – 7.70 (m, 1H), 7.58 (d, J = 18.4 Hz, 1H), 7.49 – 7.41 (m, 2H), 6.32 (d, J = 18.4 Hz, 1H), 4.39 (dd, J = 8.7, 1.9 Hz, 1H), 2.44 – 2.35 (m, 1H), 2.29 – 2.22 (m, 1H), 2.13 (dd, J = 6.0, 4.9 Hz, 1H), 1.99 – 1.92 (m, 2H), 1.46 (s, 3H), 1.30 (s, 3H), 1.23 (d, J = 11.0 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>) δ 149.4, 134.9, 133.7, 133.4, 128.4, 128.2, 128.0, 127.6, 126.4, 126.2, 123.3, 85.8, 77.8, 51.3, 39.5, 38.1, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>) δ 29.2 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{22}H_{24}BO_2$  (M - H) :331.1870, found 331.1873.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-2-(naphthalen-1-yl)vinyl)hexahydro-4,6 -methanobenzo[d][1,3,2]dioxaborole (3n): The general procedure was followed, using 1-Ethynyl-naphthalene (1q, 30.4 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3n as a pale yellow oil (33.2 mg, 50% yield);  $\mathbf{R_f} = 0.2$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.28 – 8.19 (m, 2H), 7.85 – 7.71 (m, 3H), 7.54 – 7.42 (m, 3H), 6.30 (d, J = 18.2 Hz, 1H), 4.40 (dd, J = 8.7, 1.9 Hz, 1H), 2.44 – 2.36 (m, 1H), 2.25 (m, 1H), 2.13 (dd, J = 6.0, 4.9 Hz, 1H), 2.01 – 1.90 (m, 2H), 1.48 (s, 3H), 1.34 – 1.24 (m, 4H), 0.86 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.3, 135.2, 133.5, 131.0, 129.0, 128.4, 126.1, 125.7, 125.5, 124.0, 123.7, 85.8, 77.8, 51.3, 39.5, 38.1, 35.5, 28.7, 27.1, 26.5, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.2 (large singlets) ppm; HRMS (ESI) m/z calcd for  $\mathbf{C}_{22}\mathbf{H}_{26}\mathbf{BO}_{2}$  (M + H) $^{+}$  :333.2026, found 333.2030.

3о

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-2-(thiophen-2-yl)vinyl)hexahydro-4,6-m ethanobenzo[d][1,3,2]dioxaborole (3o): The general procedure was followed, using 2-Ethynylthiophene (1r, 21.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3o as a pale yellow oil (32.4 mg, 56% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.48 (d, J = 18.1 Hz, 1H), 7.23 (dt, J = 5.0, 1.0 Hz, 1H), 7.09 – 7.07 (m, 1H), 6.98 (dd, J = 5.1, 3.6 Hz, 1H), 5.93 (d, J = 18.1 Hz, 1H), 4.35 (dd, J = 8.7, 1.8 Hz, 1H), 2.37 (m, 1H), 2.28 – 2.20 (m, 1H), 2.09 (dd, J = 6.1, 4.9 Hz, 1H), 1.96 – 1.87 (m, 2H), 1.43 (s, 3H), 1.30 (s, 3H), 1.18 (d, J = 11.0 Hz, 1H), 0.86 (s, 3H) ppm;  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.9, 141.7, 127.6, 127.6, 126.2, 85.7, 77.8, 51.3, 39.4, 38.1, 35.5, 28.6, 27.0, 26.4, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.3 (large singlets) ppm; HRMS (ESI) m/z calcd for  $C_{16}$ H<sub>22</sub>BO<sub>2</sub>S (M + H)<sup>+</sup>:289.1433, found 289.1436.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((E)-2-(thiophen-3-yl)vinyl)hexahydro-4,6-m ethanobenzo[d][1,3,2]dioxaborole (3p): The general procedure was followed, using 3-Ethynylthiophene (1s, 21.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3p as a pale yellow oil (33.4 mg, 58% yield);  $\mathbf{R_f} = 0.4$  (PE : EA = 50:1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.39 (d, J=18.4 Hz, 1H), 7.33 – 7.30 (m, 2H), 7.29 – 7.27 (m, 1H), 5.97 (d, J=18.4 Hz, 1H), 4.36 (dd, J=8.7, 1.8 Hz, 1H), 2.38 (m, 1H), 2.28 – 2.20 (m, 1H), 2.10 (dd, J=6.1, 4.9 Hz, 1H), 1.97 – 1.89 (m, 2H), 1.44 (s, 3H), 1.30 (s, 3H), 1.19 (d, J=11.1 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.1, 126.1, 125.0, 124.8, 85.8, 77.8, 51.4, 39.5, 38.2, 35.5, 28.6, 27.1, 26.4, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.8 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>16</sub>H<sub>20</sub>BO<sub>2</sub>S (M - H)<sup>-</sup>:287.1277, found 287.1280.

**3-((E)-2-((3aR,4R,6R,7aS)-3a,5,5-trimethylhexahydro-4,6-methanobenzo[d][1,3,2**] **Idioxaborol-2-yl)vinyl)pyridine** (**3q**): The general procedure was followed, using **3-Ethynylpyridine** (**1t**, 20.6 mg, 0.2 mmol), **danB-Bpai** (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 10 : 1) afforded product **3q** as a colorless oil (31.9 mg, 56% yield); **R**<sub>f</sub> = 0.4 (PE : EA = 2 : 1); **1H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.65 (d, J = 68.8 Hz, 2H), 7.82 (d, J = 7.9 Hz, 1H), 7.43 – 7.28 (m, 2H), 6.28 (d, J = 18.5 Hz, 1H), 4.39 (dd, J = 8.7, 1.8 Hz, 1H), 2.40 (m, 1H), 2.31 – 2.21 (m, 1H), 2.11 (dd, J = 6.1, 4.9 Hz, 1H), 1.99 – 1.86 (m, 2H), 1.46 (s, 3H), 1.31 (s, 3H), 1.19 (d, J = 10.9 Hz, 1H), 0.88 (s, 3H) ppm; <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 149.7, 149.0, 145.7, 133.1, 86.0, 77.9, 51.3, 39.4, 38.2, 35.4, 28.6, 27.0, 26.4, 24.0 ppm (the carbon next to boron could not be detected); <sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>) δ 29.9 (large singlets) ppm; **HRMS** (ESI) m/z calcd for C<sub>17</sub>H<sub>23</sub>BNO<sub>2</sub> (M + H) <sup>+</sup> :284.1822, found 284.1825.

(3aR,4R,6R,7aS)-2-((Z)-1,2-diphenylvinyl)-3a,5,5-trimethylhexahydro-4,6-metha nobenzo[d][1,3,2]dioxaborole (3r): The general procedure was followed, using Diphenylacetylene (1v, 35.6 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE : EA = 100 : 1) afforded product 3r as a pale yellow oil (36.2 mg, 51% yield);  $\mathbf{R_f} = 0.3$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 (s, 1H), 7.31 – 7.25 (m, 2H), 7.24 – 7.17 (m, 3H), 7.14 – 7.03 (m, 5H), 4.38 (dd, J = 8.7, 1.9 Hz, 1H), 2.39 – 2.22 (m, 2H), 2.11 (dd, J = 6.0, 4.8 Hz, 1H) 1.96 – 1.89 (m, 2H), 1.45 (s, 3H), 1.28 (d, J = 12.5 Hz, 4H), 0.85 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.1, 140.5, 136.9, 129.9, 128.6, 128.3, 127.8, 127.6, 126.3, 86.1, 78.3, 51.3, 39.5, 38.1, 35.6, 28.6, 27.1, 26.6, 24.0 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  29.8 (large singlets) ppm; HRMS (ESI) m/z calcd for  $\mathbf{C}_{24}\mathbf{H}_{28}\mathbf{BO}_{2}$  (M + H)  $^+$  :359.2182, found 359.2187.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((*Z*)-1-phenylprop-1-en-2-yl)hexahydro-4,6-methanobenzo[d][1,3,2]dioxaborole (3s): The general procedure was followed, using 1-Phenylpropyne (1w, 23.2 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE) afforded product 3s as a pale yellow oil (38.0 mg, 64% yield);  $\mathbf{R_f} = 0.5$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.41 – 7.31 (m, 4H), 7.27 – 7.21 (m, 2H), 4.38 (dd, J = 8.7, 1.9 Hz, 1H), 2.44 – 2.35 (m, 1H), 2.25 (m, 1H), 2.11 (dd, J = 6.1, 4.8 Hz, 1H), 2.01 (d, J = 1.9 Hz, 3H), 1.94 (m, 2H), 1.45 (s, 3H), 1.30 (s, 3H), 1.22 (d, J = 10.9 Hz, 1H), 0.87 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  142.3, 137.9, 229.3, 128.0, 127.1, 85.9, 78.0, 51.3, 39.5, 38.1, 35.6, 28.7, 27.1, 26.5, 24.0, 16.0 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  23.0 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{19}\mathbf{H}_{26}\mathbf{BO}_{2}$  (M + H)  $^{+}$  :297.2026, found 297.2029.

(3aR,4R,6R,7aS)-3a,5,5-trimethyl-2-((Z)-1-phenylbut-1-en-2-yl)hexahydro-4,6-m ethanobenzo[d][1,3,2]dioxaborole (3t): The general procedure was followed, using 1-Butynylbenzene (1x, 26.0 mg, 0.2 mmol), danB-Bpai (83.04 mg, 0.24 mmol). Purification of this material by chromatography on silica gel (PE) afforded product 3t

as a pale yellow oil (31.6 mg, 51% yield);  $\mathbf{R_f} = 0.5$  (PE : EA = 50 : 1);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (d, J = 4.4 Hz, 4H), 7.23 (d, J = 8.2 Hz, 2H), 4.37 (dd, J = 8.7, 1.8 Hz, 1H), 2.40 (m, 3H), 2.29 – 2.22 (m, 1H), 2.15 – 2.09 (m, 1H), 1.99 – 1.89 (m, 2H), 1.49 – 1.42 (m, 3H), 1.30 (s, 3H), 1.22 (d, J = 10.9 Hz, 1H), 1.12 (t, J = 7.5 Hz, 3H), 0.87 (s, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  141.4, 137.8, 128.9, 128.1, 127.0, 85.8, 77.9, 51.3, 39.5, 38.1, 35.6, 28.7, 27.1, 26.5, 24.0, 22.7, 14.7 ppm (the carbon next to boron could not be detected);  $^{11}\mathbf{B}$  NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  23.0 (large singlets) ppm; **HRMS** (ESI) m/z calcd for  $\mathbf{C}_{20}\mathbf{H}_{28}\mathbf{BO}_{2}$  (M + H)  $^+$  :311.2182, found 311.2186.

#### 6. Product transformations

In a 50 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (415.2 mg, 1.2 mmol, 1.2 equiv.), CuF<sub>2</sub> (10.1 mg, 0.1 mmol, 10 mol%), PPh<sub>3</sub> (26.2 mg, 0.1 mmol, 10 mol%), and MeONa (81.0 mg, 1.5 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under N<sub>2</sub> flow (this sequence was repeated three times). Anhydrous Cyclohexane (7.5 mL), Phenylacetylene (102 mg, 1.0 mmol, 1.0 equiv.), and MeOH (64.1 mg, 2.0 mmol, 2.0 equiv.) were added subsequently under N<sub>2</sub>. The tube was stirred at 35 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography to afford the product **2a** (260.7mg, 96% yield).

In a 50 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (415.2 mg, 1.2 mmol, 1.2 equiv.), CuBr<sub>2</sub> (22.3 mg, 0.1 mmol, 10 mol%), PCy<sub>3</sub> (28.0 mg, 0.1 mmol, 10 mol%), and MeOLi (57.0 mg, 1.5 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under N<sub>2</sub> flow (this sequence was repeated three times). Anhydrous iPr<sub>2</sub>O (7.5 mL), Phenylacetylene (102 mg, 1.0 mmol, 1.0 equiv.), and MeOH (64.1 mg, 2.0 mmol, 2.0 equiv.) were added subsequently under N<sub>2</sub>. The tube was stirred at 65 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography to afford the product 3a (183.3 mg, 65% yield).

#### Step 1:

In a 25 mL Schlenk tube, which contained a stirring bar, was charged with **2a** (81.3 mg, 0.3 mmol, 1.0 equiv.) in THF (3.0 mL). 5M sulfuric acid (0.18 mL, 0.9 mmol, 3.0 equiv.) and pinacol (177.3 mg, 1.5 mmol, 5.0 equiv.) were added .The tube was stirred at room temperature for 18 hours. The reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation to afford the product (E)-4,4,5,5-tetramethyl-2-styryl-1,3,2-dioxaborolane.

#### Step 2:

In a 25 mL Schlenk tube, which contained a stirring bar, was charged with Pd(PPh<sub>3</sub>)<sub>4</sub> (34.7 mg, 0.03 mmol, 10 mol%) and K<sub>2</sub>CO<sub>3</sub> (124.4 mg, 0.9 mmol, 3.0 equiv.). The tube was then evacuated and back-filled under N2 flow (this sequence repeated three times). To solution (E)-4,4,5,5-tetramethyl-2-styryl-1,3,2-dioxaborolane in THF (1.8 mL), H<sub>2</sub>O (0.2 mL), and Ethyl 4-iodobenzoate (248.5 mg, 0.9 mmol, 3.0 equiv.) were added subsequently under N₂. The tube was stirred at 85 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE : EA = 100 : 1) to afford the product  $\mathbf{4}^{[8]}$  (42.4 mg, 56% yield) as a yellow solid;  $\mathbf{R_f} = 0.3 \; (\text{PE} : \text{EA} = 50 : 1); \, ^1\mathbf{H} \; \mathbf{NMR} \; (400 \; \text{MHz}, \; \text{CDCl}_3) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 8.03 \; (\text{d}, \; J = 1.00 \; \text{m}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.00 \; \text{d}) \; \delta \; 6.03 \; (\text{d}, \; J = 1.$ 8.4 Hz, 2H), 7.57 - 7.50 (m, 4H), 7.37 (t, J = 7.5 Hz, 2H), 7.31 - 7.27 (m, 1H), 7.21(d, J = 16.3 Hz, 1H), 7.12 (d, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.3 Hz, 1H), 7.12 (d, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 16.2 Hz, 1H), 1H 7.1 Hz, 3H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 166.4, 141.7, 136.7, 131.1, 123.0, 129.2, 128.7, 128.2, 127.6, 126.7, 126.2, 60.9, 14.3 ppm.

## Step 1:

In a 25 mL Schlenk tube, which contained a stirring bar, was charged with **2a** (108.4 mg, 0.4 mmol, 1.0 equiv.) in THF (4.0 mL). 5M sulfuric acid (0.24 mL, 1.2

mmol, 3.0 equiv.) and pinacol (236.3 mg, 2.0 mmol, 5.0 equiv.) were added .The tube was stirred at room temperature for 18 hours. The reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation to afford the product (E)-4,4,5,5-tetramethyl-2-styryl-1,3,2-dioxaborolane.

## Step 2:

In a 25 mL Schlenk tube, which contained a stirring bar, was charged with  $Pd(PPh_3)_2Cl_2$  (28.1mg, 0.04 mmol, 10 mol%),  $Na_2CO_3$  (84.8 mg, 0.8 mmol, 2.0 equiv.) and TsCl (38.1 mg, 0.2 mmol, 0.5 equiv.). The tube was then evacuated and back-filled under  $N_2$  flow (this sequence was repeated three times). To a solution of (E)-4,4,5,5-tetramethyl-2-styryl-1,3,2-dioxaborolane in tBuOH (1.5 mL) and  $H_2O$  (1.5 mL) were added subsequently under  $N_2$ . The tube was stirred at 35 °C for 12 hours. The reaction mixture was diluted with EA and  $H_2O$ , then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE) to afford the product  $\mathbf{5}^{[9]}$  (19.4 mg, 47% yield) as a white solid;  $\mathbf{R_f} = 0.5$  (PE);  $^1\mathbf{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.44 (dd, J = 8.3, 1.4 Hz, 4H), 7.33 (dd, J = 8.3, 6.9 Hz, 4H), 7.26 – 7.21 (m, 2H), 6.99 – 6.94 (m, 2H), 6.71 – 6.65 (m, 2H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  137.3, 132.8, 129.2, 128.6, 127.5, 126.4 ppm.

In a 25 mL Schlenk tube, which contained a stirring bar, was charged with **3a** (84.7 mg, 0.3 mmol, 1.0 equiv.),  $Cu(OAc)_2$  (108.9 mg, 0.6 mmol, 2.0 equiv.),  $Et_3N$  (121.4 mg, 1.2 mmol, 4.0 equiv.), and EtOH (2.0 mL). The tube was stirred at 25 °C for 12 hours. The reaction mixture was diluted with EA and  $H_2O$ , then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE) to afford the product  $\mathbf{6}^{[9]}$  (37.3 mg, 84% yield) as a colorless oil;  $\mathbf{R_f} = 0.4$  (PE);  $^1\mathbf{H}$  NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.28 – 7.18 (m, 4H), 7.15 – 7.09 (m, 1H), 6.98 (d, J = 12.9 Hz, 1H), 5.83 (d, J = 13.0 Hz, 1H), 3.89 (q, J = 7.1 Hz, 2H), 1.33 (t, J = 7.1 Hz, 3H) ppm;  $^{13}\mathbf{C}$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  147.9, 136.5, 128.5, 125.5, 125.0, 105.8, 65.4, 14.8 ppm.

In a 25 m Schlenk tube, which contained a stirring bar, was charged with **3a** (84.7 mg, 0.3 mmol, 1.0 equiv.), NaBO<sub>3</sub> 4H<sub>2</sub>O (138.5 mg, 0.9 mmol, 3.0 equiv.), THF (1.5 mL), and H<sub>2</sub>O (2.0 mL). The tube was stirred at room temperature for 3 hours. The reaction mixture was diluted with EA and H<sub>2</sub>O. Then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. At 0°C, NaBH<sub>4</sub> (34.0 mg, 0.9 mmol, 3.0 equiv.) and MeOH (1.5 mL) were added. The tube was stirred at room temperature for 3 hours. The reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE : EA = 20 : 1) to afford the product  $7^{[10]}$  (19.4 mg, 53% yield) as a colorless oil;  $\mathbf{R_f} = 0.4$  (PE : EA = 5 : 1); <sup>1</sup>HNMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 – 7.27 (m, 2H), 7.22 (td, J = 6.3, 1.6 Hz, 3H), 3.81 (t, J = 6.6 Hz, 2H), 2.84 (t, J = 6.6 Hz, 2H), 1.83 (s, 1H) ppm; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.4, 129.0, 128.5, 126.4, 63.6 ppm.

# Failed example:

As shown above, using 4-ethynylbenzaldehyde as the starting material afforded the desired product but generated many unknown byproducts; this result suggested that the aldehyde group might not be compatible with the reaction conditions.

### 7. Mechanism experiment

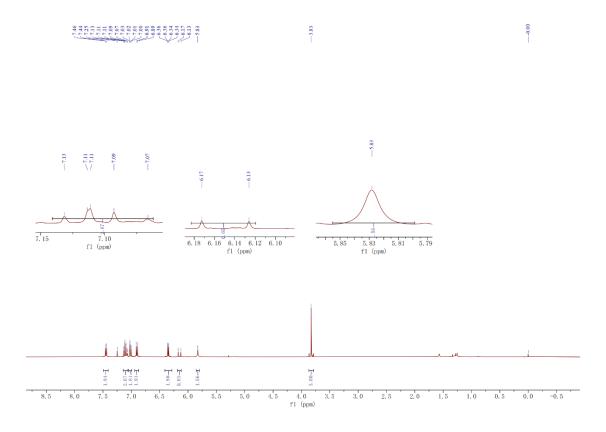
### 7.1 The effect of substituent electricity:

In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (69.2 mg, 0.2 mmol, 1.0 equiv.), 4-Ethynylanisole (79.3 mg, 0.6 mmol, 3.0 equiv.),  $CuF_2$  (2.1 mg, 0.02 mmol, 10 mol%),  $PPh_3$  (5.3 mg, 0.02 mmol, 10 mol%), and MeONa (16.2 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under  $N_2$  flow (this sequence was repeated three times). Anhydrous Cyclohexane (1.5 mL), Phenylacetylene (61.3 mg, 0.6 mmol, 3.0 equiv.), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under  $N_2$ . The tube was stirred at 35 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and  $H_2O$ , then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography to afford the product 2a (21.3 mg, 40% yield) and 2g (13.2 mg, 22% yield), 2a:2g = 1.8:1.

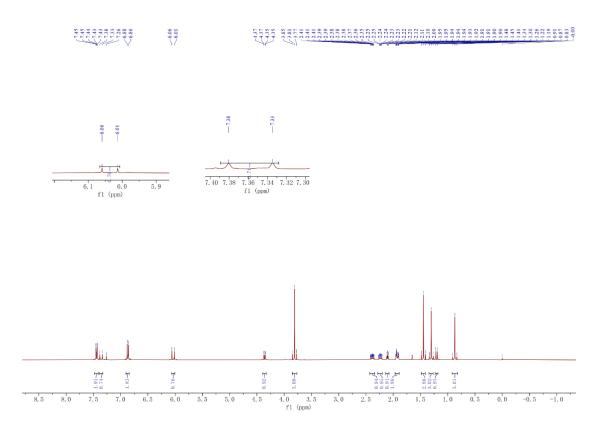
In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (69.2 mg, 0.2 mmol, 1.0 equiv.), 4-(Methoxycarbonyl)phenylacetylene (96.1 mg, 0.6 mmol, 3.0 equiv.),  $CuF_2$  (2.1 mg, 0.02 mmol, 10 mol%),  $PPh_3$  (5.3 mg, 0.02 mmol, 10 mol%), and MeONa (16.2 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under  $N_2$  flow (this sequence was repeated three times). Anhydrous Cyclohexane (1.5 mL), Phenylacetylene (61.3 mg, 0.6 mmol, 3.0 equiv.), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under  $N_2$ . The tube was stirred at 35 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and  $H_2O$ , then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography to afford the product 2a (3.5 mg, 6% yield) and 2o (31.7 mg, 48% yield), 2a:2o = 1:8.

## 7.2 Deuterium experiments:

In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (83.04 mg, 0.2 mmol, 1.2 equiv.), 4-Ethynylanisole (26.4 mg, 0.6 mmol, 3.0 equiv.),  $CuF_2$  (2.1 mg, 0.02 mmol, 10 mol%),  $PPh_3$  (5.3 mg, 0.02 mmol, 10 mol%), and MeONa (16.2 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under  $N_2$  flow (this sequence was repeated three times). Anhydrous Cyclohexane (1.5 mL), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under  $N_2$ . The tube was stirred at 35 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and  $H_2O$ , then extracted with EA. The organic layer was combined and dried over  $Na_2SO_4$ . Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography to afford the product 2g-D (53.4 mg, 89% yield).



In a 25 mL dry Schlenk tube, which contained a stirring bar, was charged with danB-Bpai (83.04 mg, 0.24 mmol, 1.2 equiv.), 4-Ethynylanisole (26.4 mg, 0.6 mmol, 3.0 equiv.), CuBr<sub>2</sub> (4.5 mg, 0.02 mmol, 10 mol%), PCy<sub>3</sub> (5.6 mg, 0.02 mmol, 10 mol%), and MeOLi (11.4 mg, 0.3 mmol, 1.5 equiv.). The tube was then evacuated and back-filled under N<sub>2</sub> flow (this sequence was repeated three times). Anhydrous iPr<sub>2</sub>O (1.5 mL), alkenyl (0.2 mmol, 1.0 equiv.), and MeOH (12.8 mg, 0.4 mmol, 2.0 equiv.) were added subsequently under N<sub>2</sub>. The tube was stirred at 65 °C for 12 hours. After cooling to room temperature, the reaction mixture was diluted with EA and H<sub>2</sub>O, then extracted with EA. The organic layer was combined and dried over Na<sub>2</sub>SO<sub>4</sub>. Then filtered and concentrated by rotary evaporation. The residue was purified by silica gel chromatography (PE/EA) to afford the product **3g-D** (28.1mg, 45% yield).

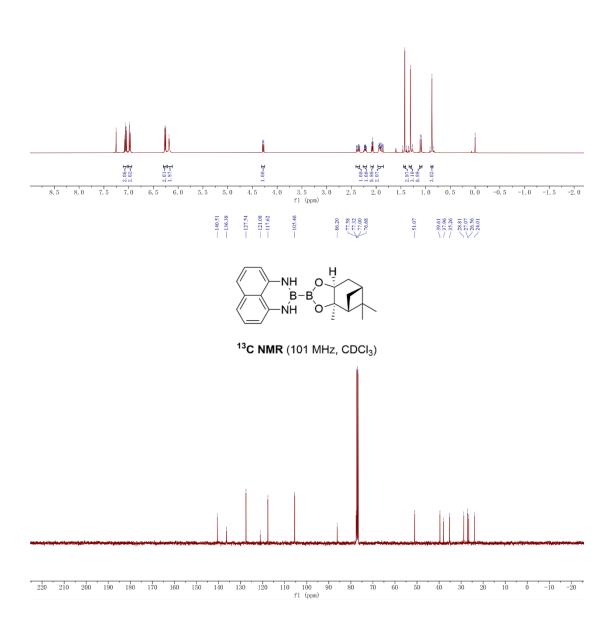


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# 9. NMR Spectra

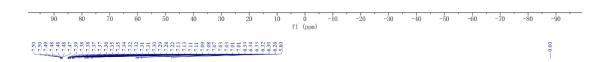
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





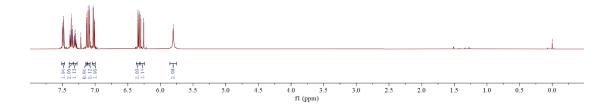
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)





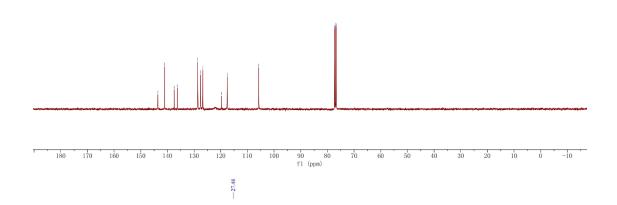
2a

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



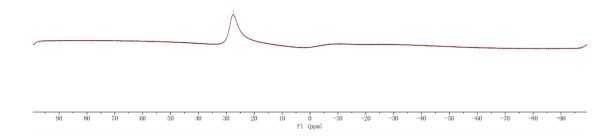
2a

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



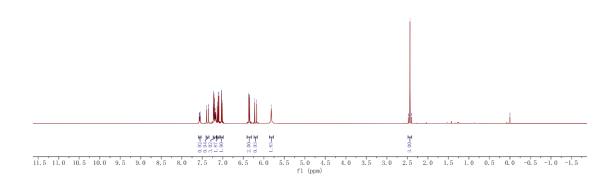
2a

<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)



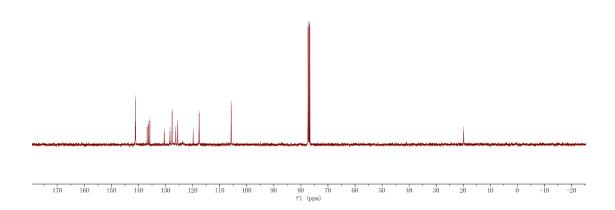
2b

1H NMR (400 MHz, CDCl<sub>3</sub>)



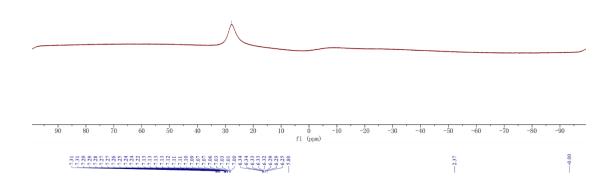


 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



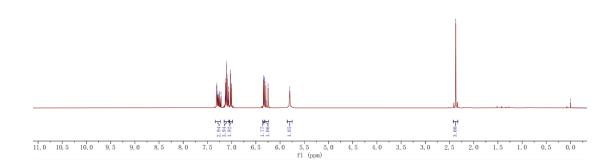


 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)



2c

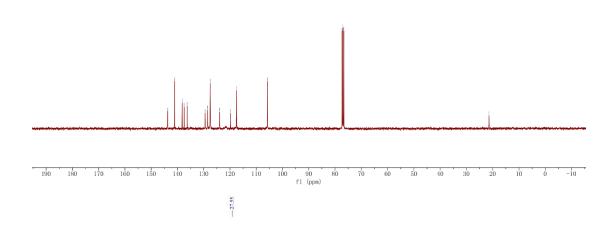
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





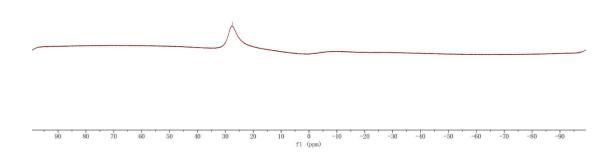
**2**c

 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



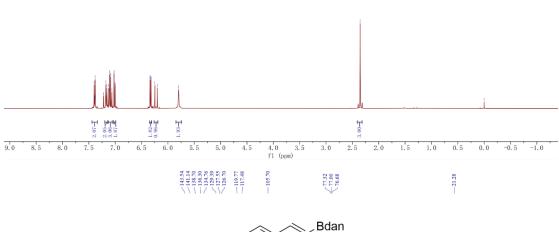
2c

<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)



2d

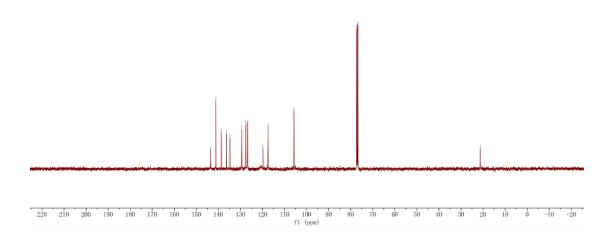
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



Me

2d

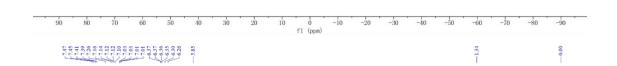
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



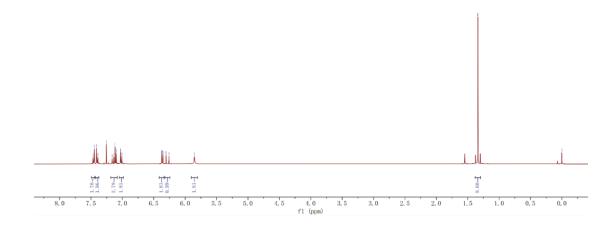


 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)

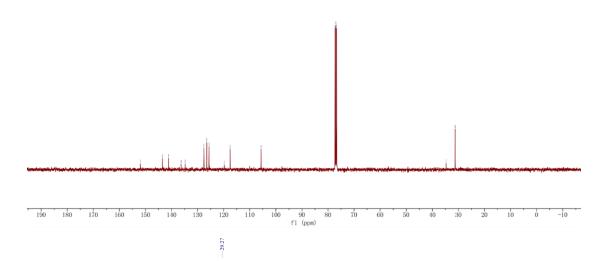




<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

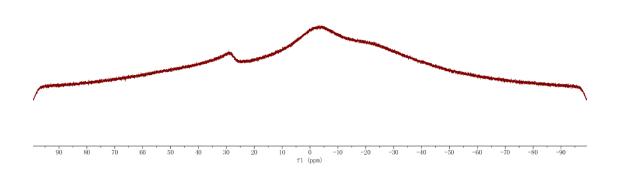


<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

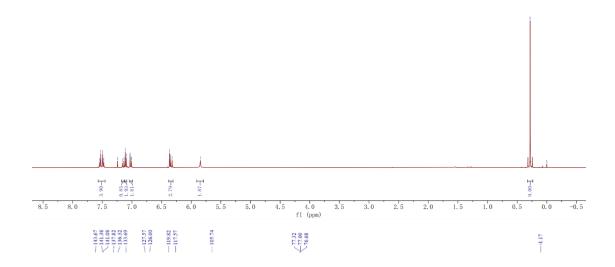


**2**e

<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)

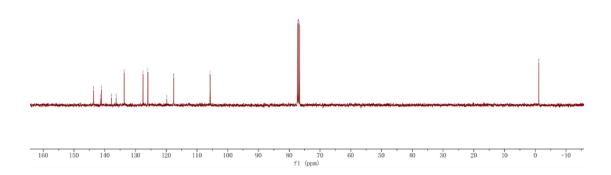


\$2f\$  $^{1}\mbox{H}$  NMR (400 MHz, CDCl\_3)



2f

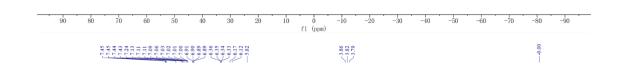
<sup>13</sup>CNMR (101 MHz, CDCl<sub>3</sub>)



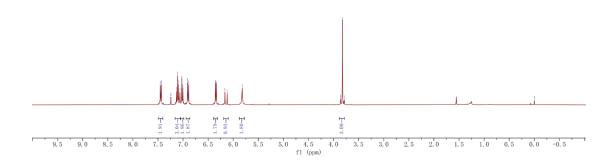


 $^{11}\text{B}$  NMR (128 MHz, CDCl<sub>3</sub>)

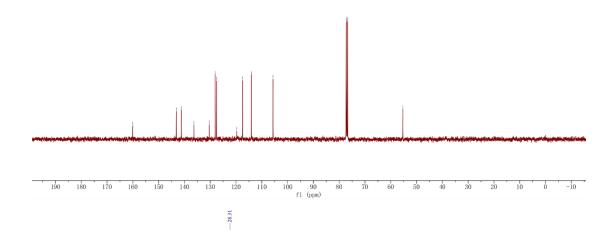




 $^{1}H$  NMR (400 MHz, CDCl $_{3}$ )



 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



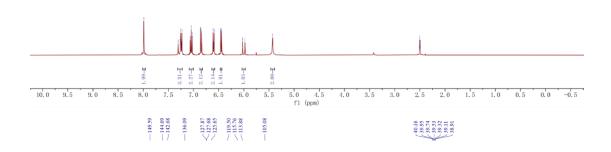
MeO Bdan

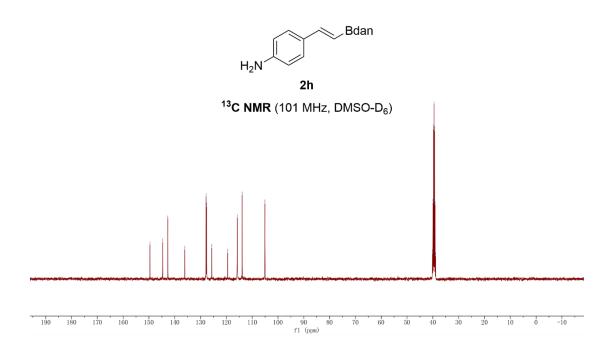
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)





<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)

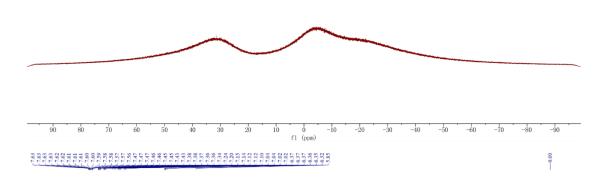




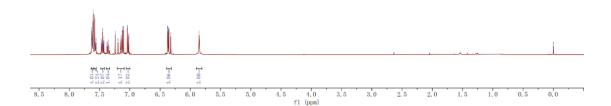


$$\mathsf{H_2N} \qquad \qquad \mathsf{Bdan}$$
 
$$\mathbf{2h}$$

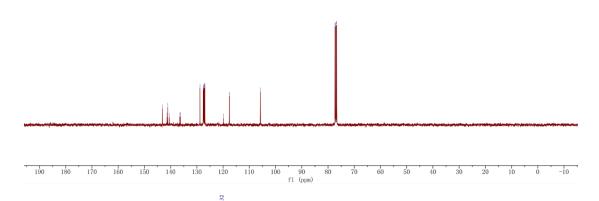
<sup>11</sup>**B NMR** (128 MHz, DMSO-D<sub>6</sub>)



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



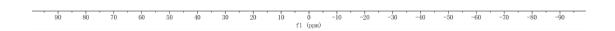
^ ^

Bdan

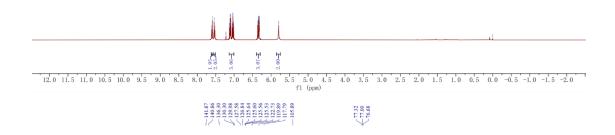
2i

 $^{11}\text{B NMR}$  (128 MHz, CDCl<sub>3</sub>)

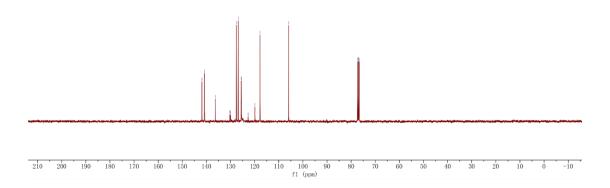




<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



-07.43

$$F_3$$
C  $2j$  Bdan

<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)

0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -110 (ppm)

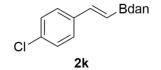
-27.73

2j

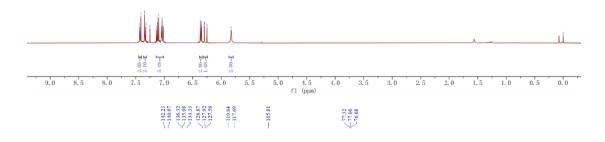
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)



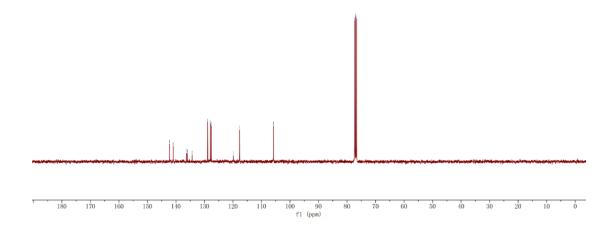
90 80 70 60 50 40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



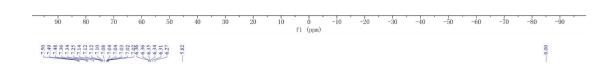
2k <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



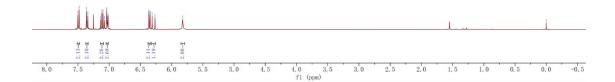


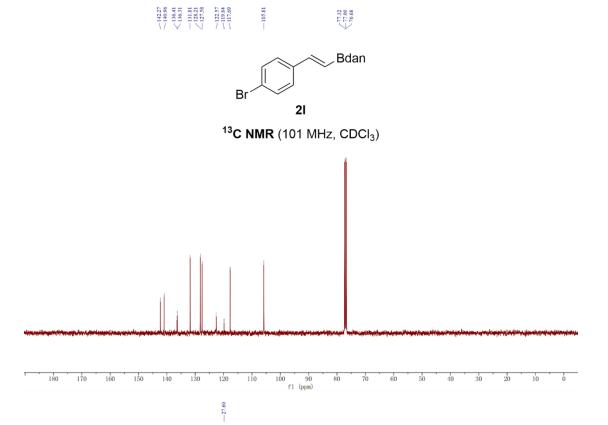
 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)

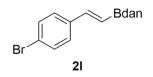




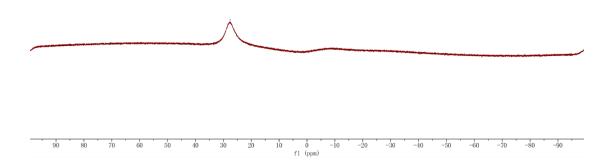
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



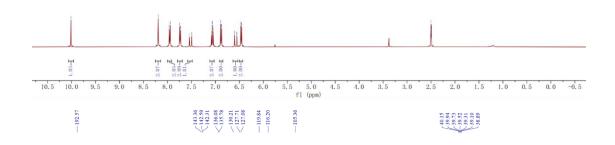




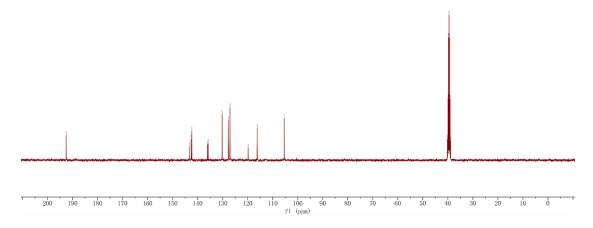
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)

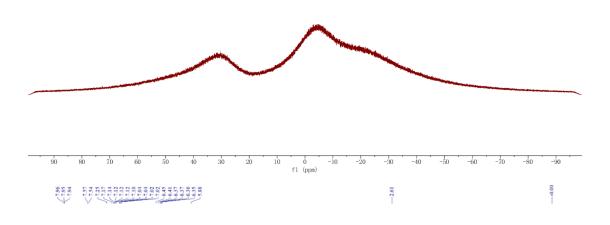


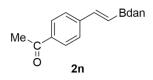
<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)



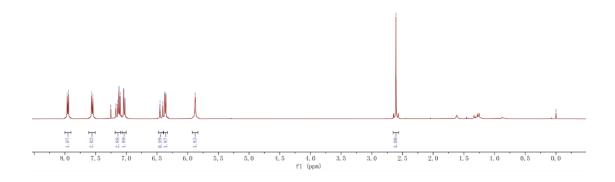
 $^{13}\text{C NMR}$  (101 MHz, DMSO-D<sub>6</sub>)

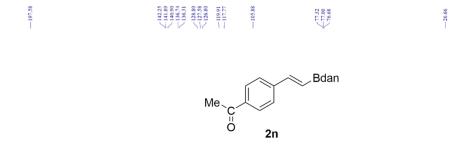




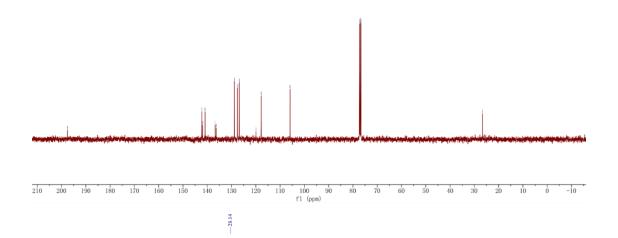


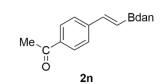
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



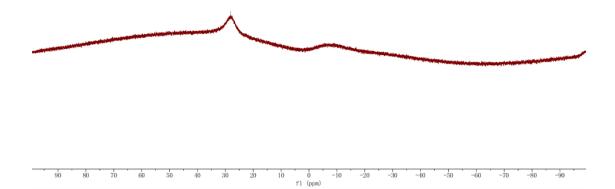


 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)



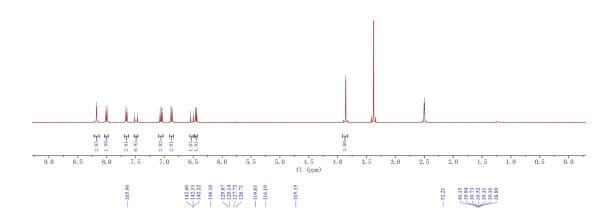


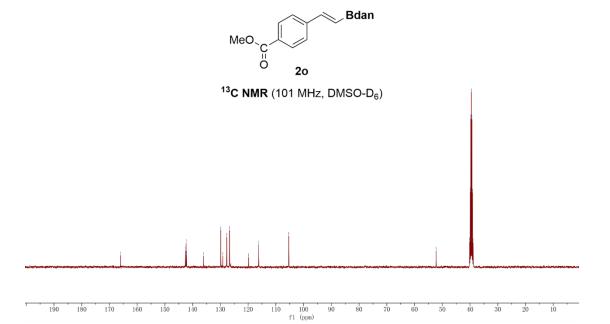
<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)

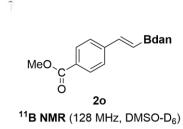


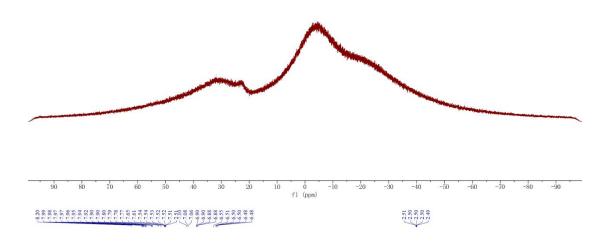


<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)

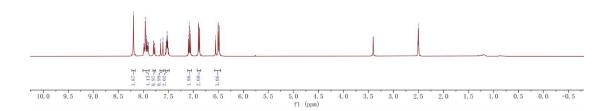






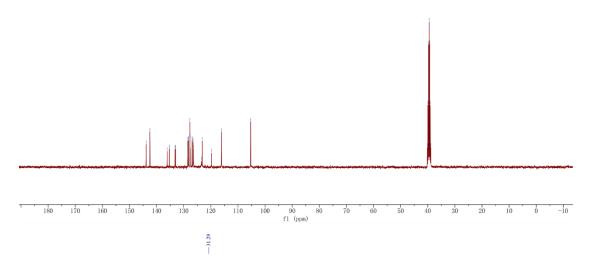


<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)



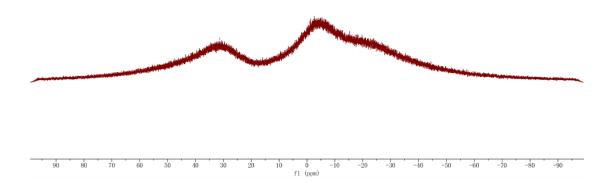


 $^{13}$ C NMR (101 MHz, DMSO-D<sub>6</sub>)

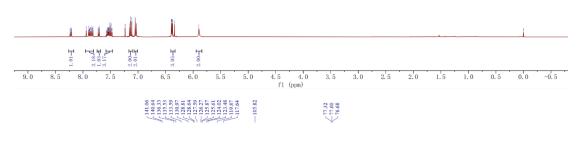


Bdan 2p

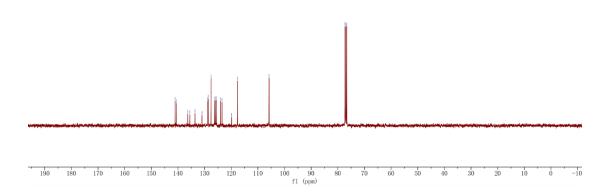
<sup>11</sup>**B NMR** (128 MHz, DMSO-D<sub>6</sub>)



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

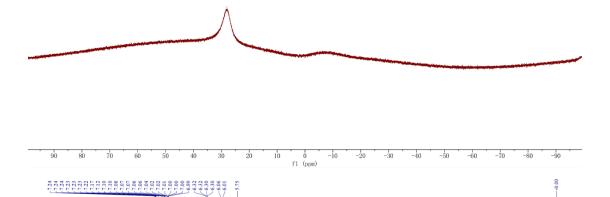


 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



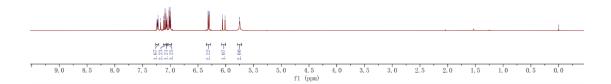


<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)



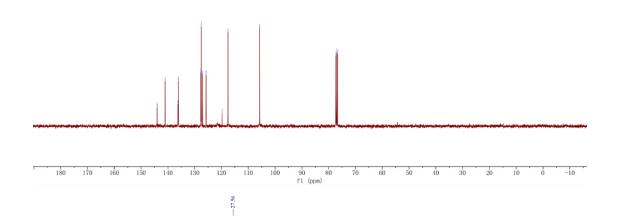
2r

 $^{1}\text{H NMR}$  (400 MHz, CDCl<sub>3</sub>)



2r

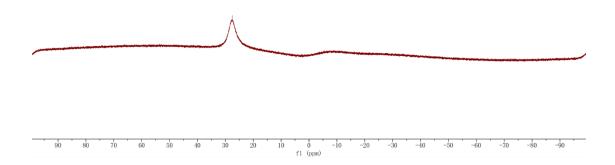
 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)

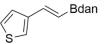


Bdan

2r

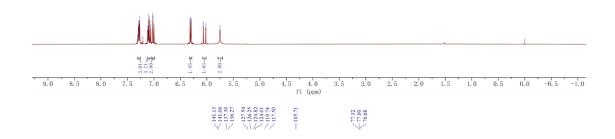
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)

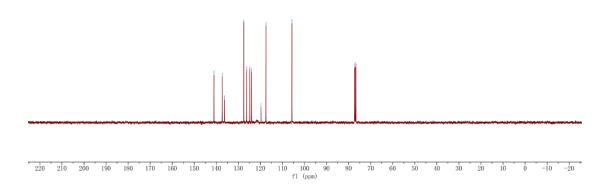




2s

1H NMR (400 MHz, CDCl<sub>3</sub>)

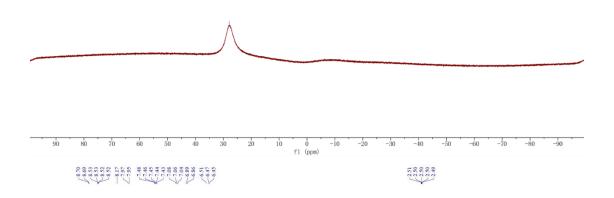




727.87

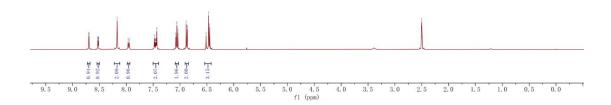
2s

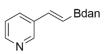
 $^{11}\mathrm{B}~\mathrm{NMR}~\mathrm{(128~MHz,~CDCl_3)}$ 



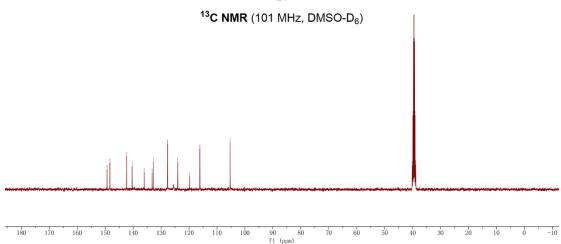
2t

<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)





2t

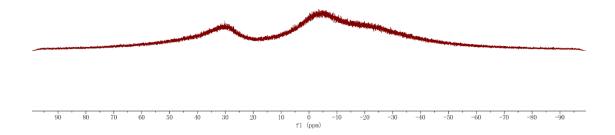


Bdan

-30.45

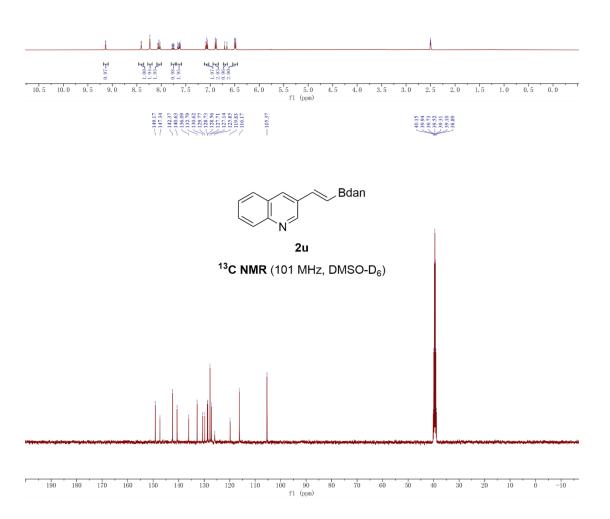
2t

 $^{11}\mathrm{B}\ \mathrm{NMR}\ (128\ \mathrm{MHz},\ \mathrm{DMSO-D_6})$ 



2u

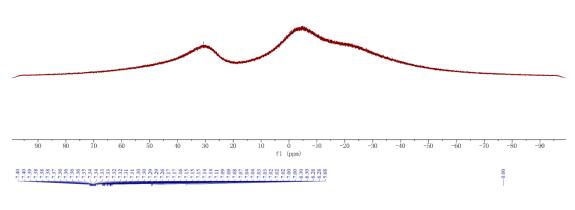
<sup>1</sup>H NMR (400 MHz, DMSO-D<sub>6</sub>)





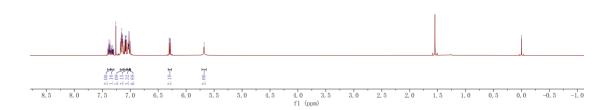
2u

<sup>11</sup>**B NMR** (128 MHz, DMSO-D<sub>6</sub>)

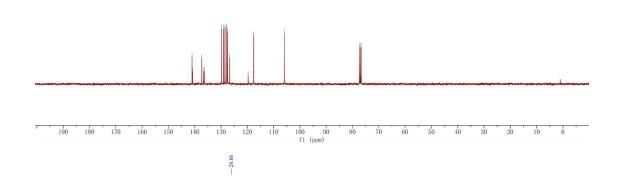


2v

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



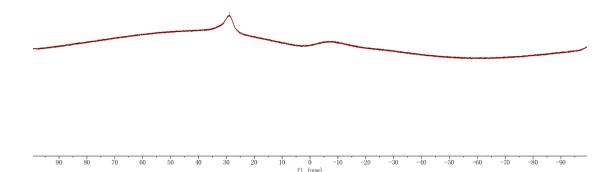
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



Bdan

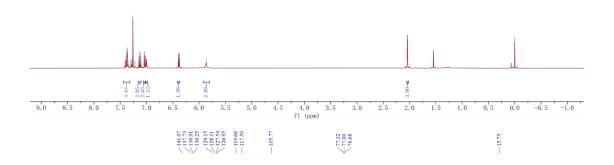
2v

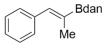
<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)



2w

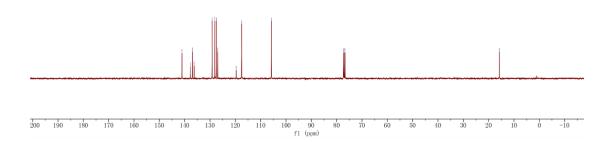
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





2w

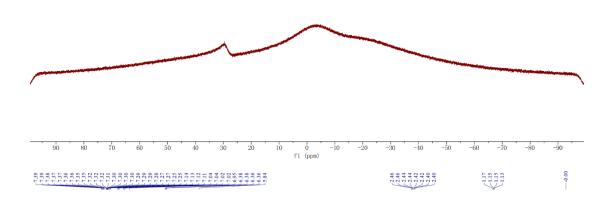
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)





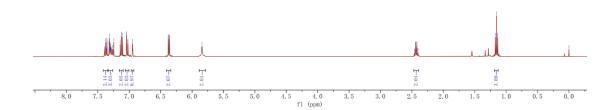
2w

<sup>11</sup>B **NMR** (128 MHz, CDCl<sub>3</sub>)



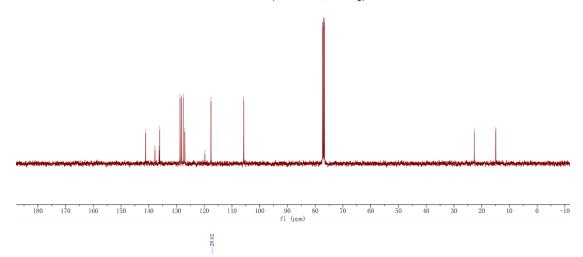
**2**x

 $^{1}H$  NMR (400 MHz, CDCl<sub>3</sub>)



**2**x

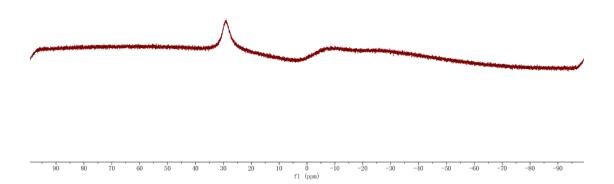
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



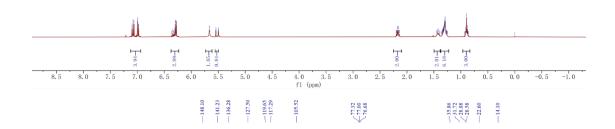
Bdan

2x

 $^{11}\mathrm{B}\ \mathrm{NMR}\ (128\ \mathrm{MHz},\ \mathrm{CDCI_3})$ 



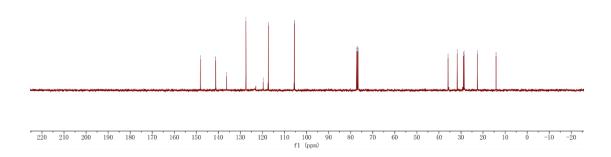
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



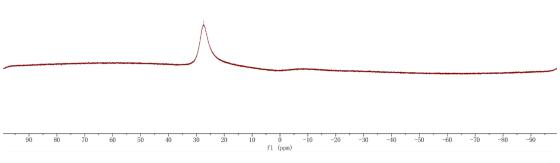
C<sub>6</sub>H<sub>13</sub> Bdan

2у

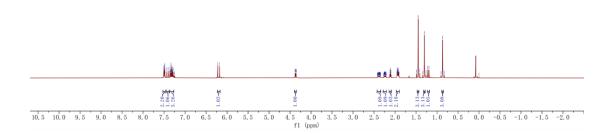
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



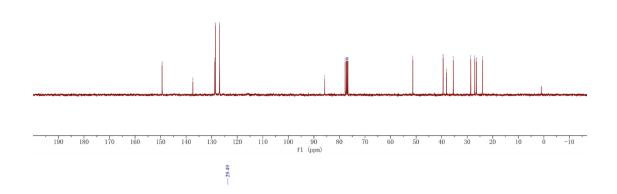
<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)



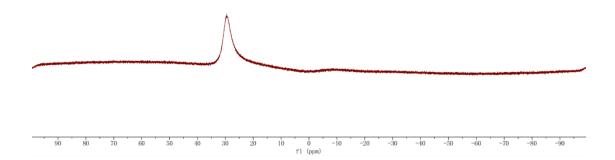
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



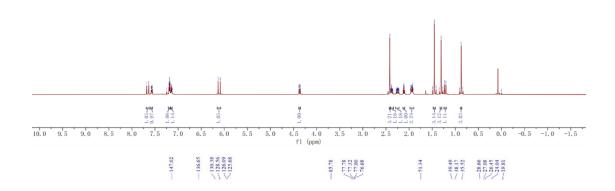
 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



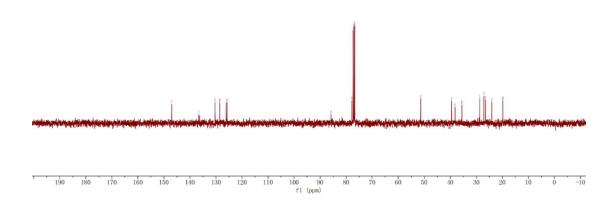
\$3a\$  $$^{11}\mbox{B}$  NMR (128 MHz, CDCl $_{3}$ )



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

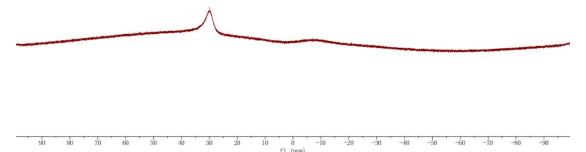


13C NMR (101 MHz, CDCl<sub>3</sub>)

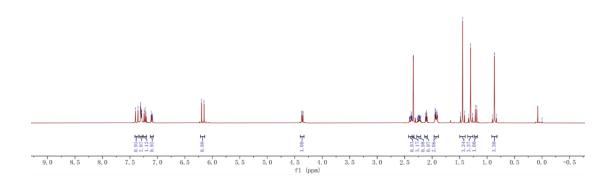




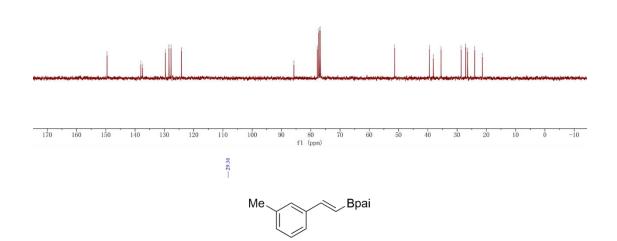
 $^{11}\mathrm{B}\ \mathrm{NMR}\ (128\ \mathrm{MHz},\ \mathrm{CDCI_3})$ 



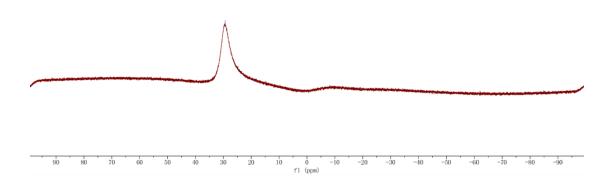
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



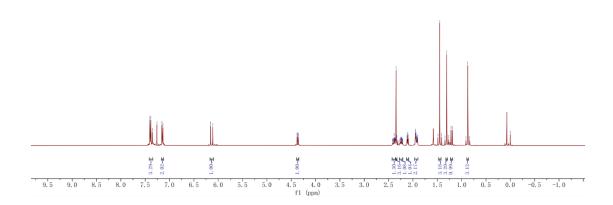
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

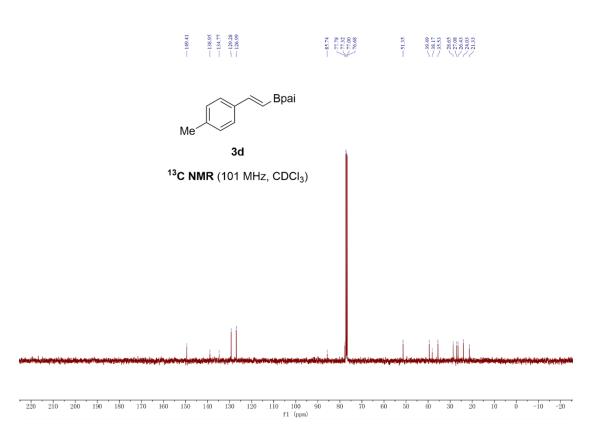


3c  $^{11}B$  NMR (128 MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

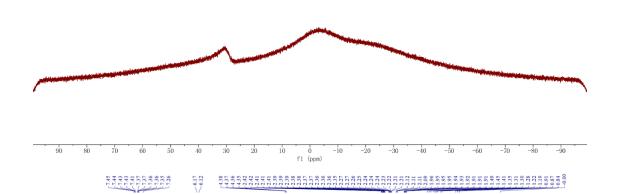


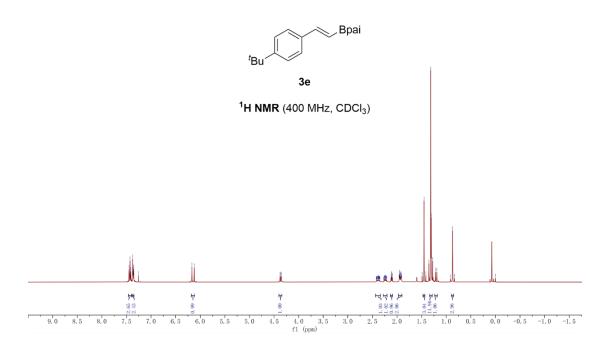




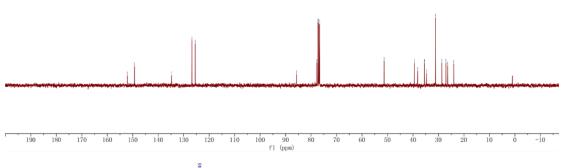
3d

<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)





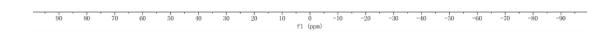
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

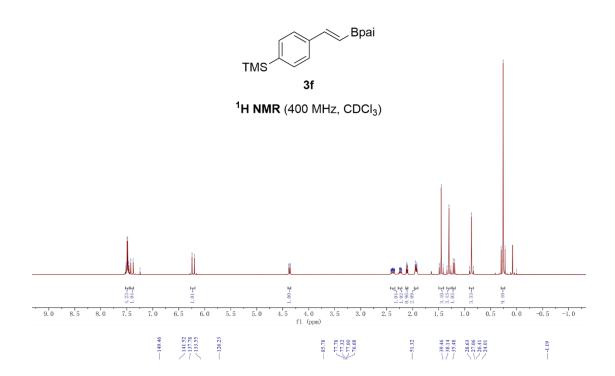


-29.46

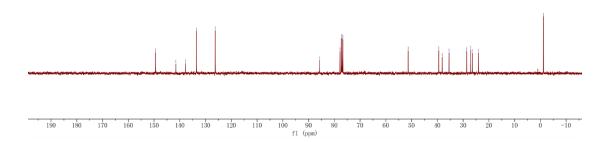
<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)







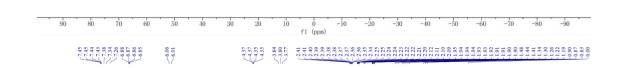
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



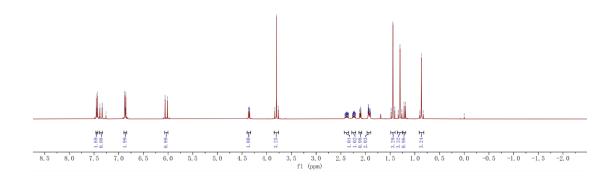


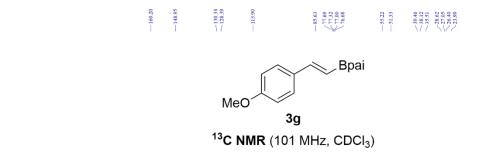
<sup>11</sup>B **NMR** (128 MHz, CDCl<sub>3</sub>)

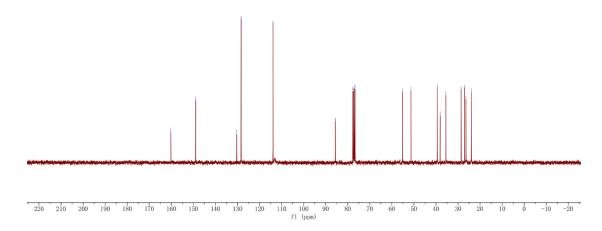




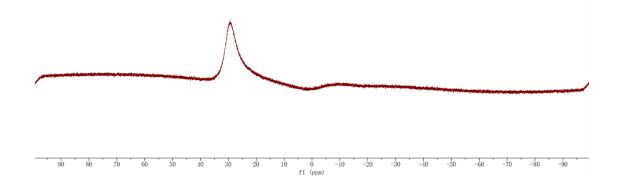
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



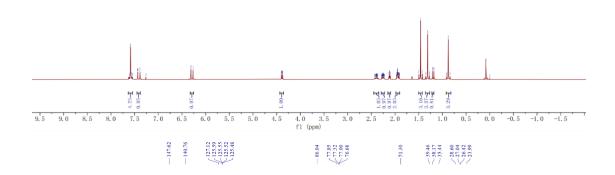




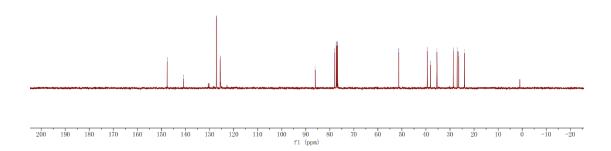




<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

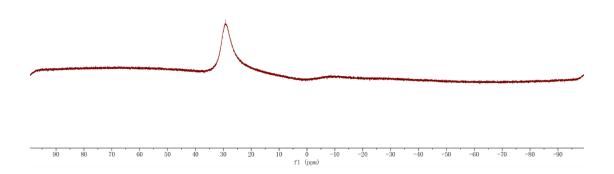




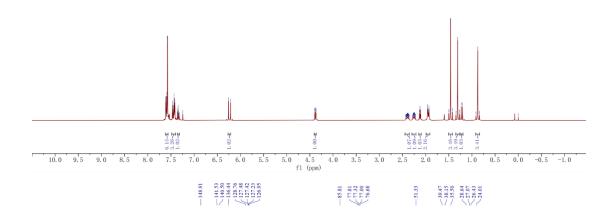
 $^{19}$ F NMR (376 MHz, CDCl<sub>3</sub>)

0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 1 (ppm)

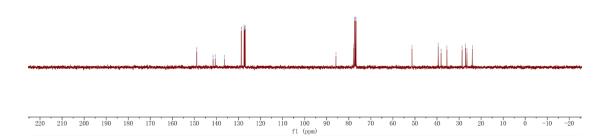
 $^{11}\mathrm{B}\ \mathrm{NMR}\ (128\ \mathrm{MHz},\ \mathrm{CDCI_3})$ 



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



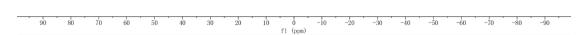
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



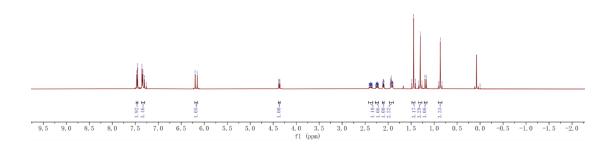


<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)

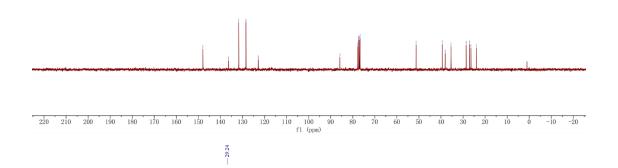




<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

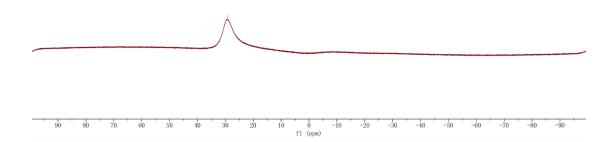


 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)

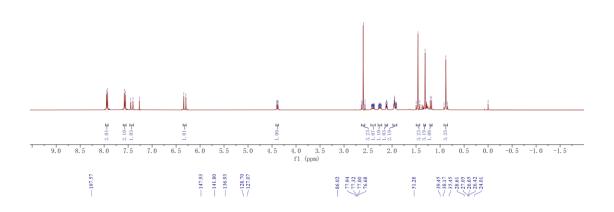


Br Bpai

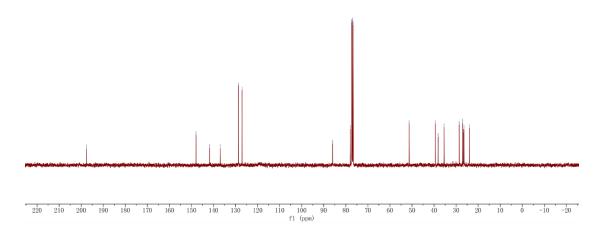
 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



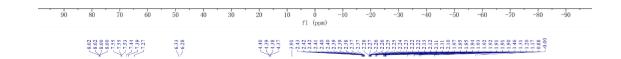
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



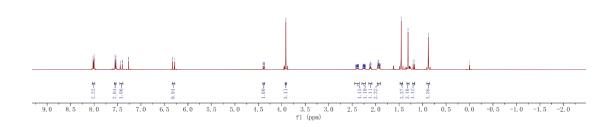


 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)

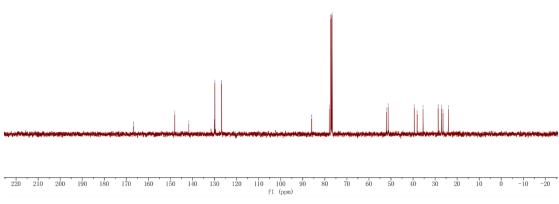




<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

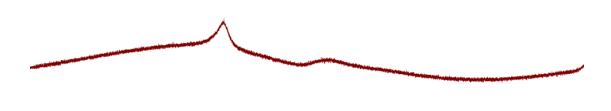


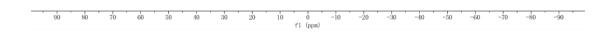
 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)



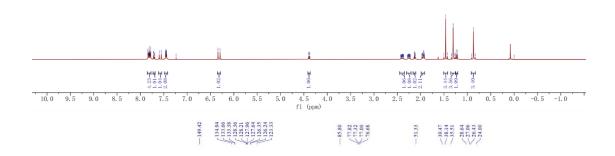
-29.76

 $^{11}\text{B}$  NMR (128 MHz, CDCl<sub>3</sub>)



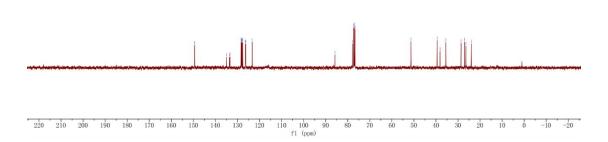


 $^{1}H$  NMR (400 MHz, CDCl $_{3}$ )

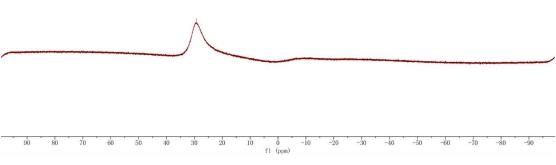


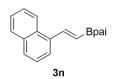
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<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

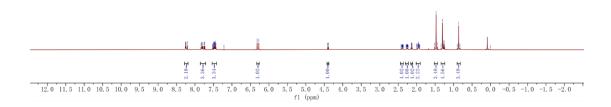


<sup>11</sup>**B NMR** (128 MHz, CDCl<sub>3</sub>)





<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

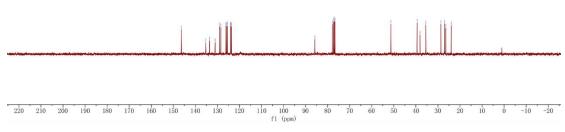




Bpai

3n

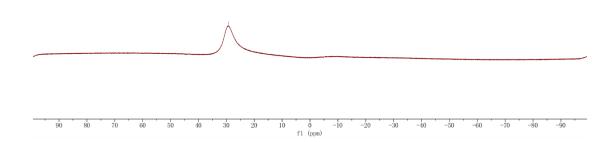
 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



-29.25

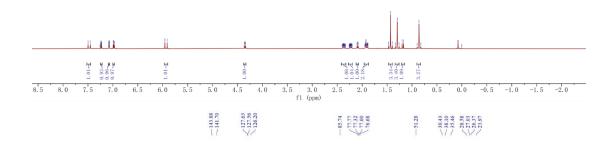
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 $^{11}$ B NMR (128 MHz, CDCl<sub>3</sub>)



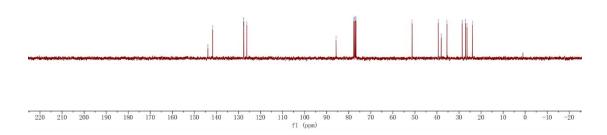
3о

 $^{1}H$  NMR (400 MHz, CDCl<sub>3</sub>)

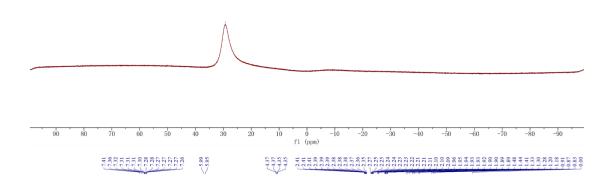


30

 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)

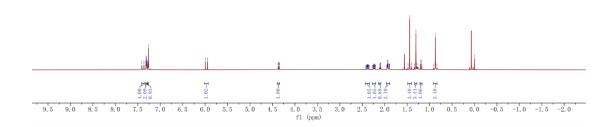


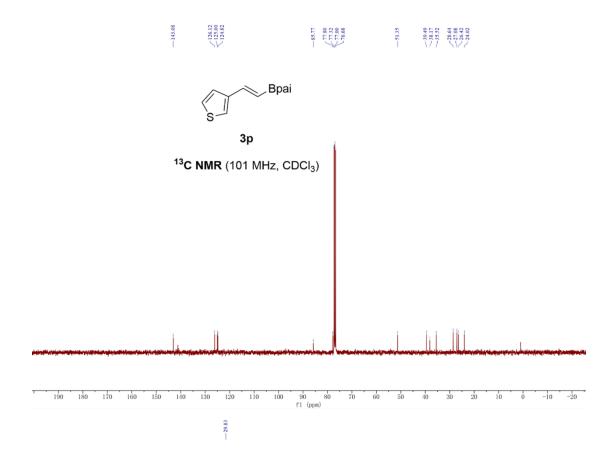
 ${f 3o}$   ${f ^{11}B}$  NMR (128 MHz, CDCl3)

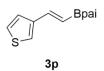


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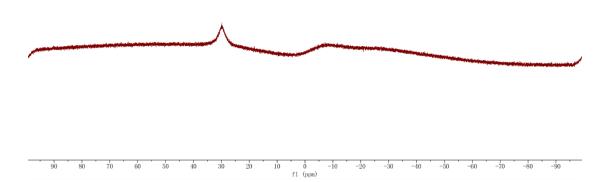
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

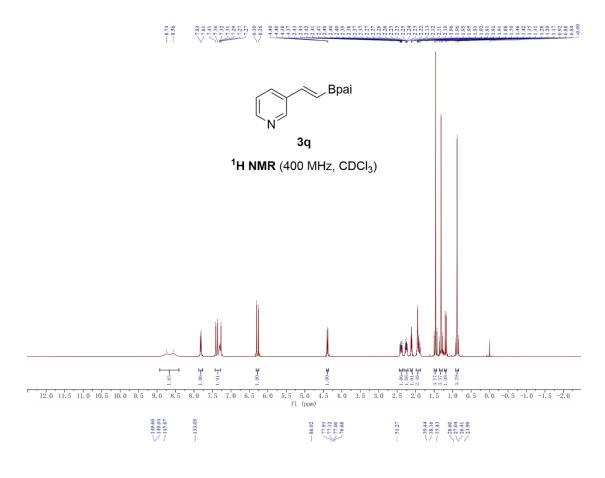


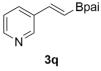




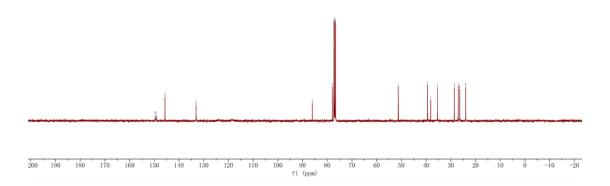
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)







 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)

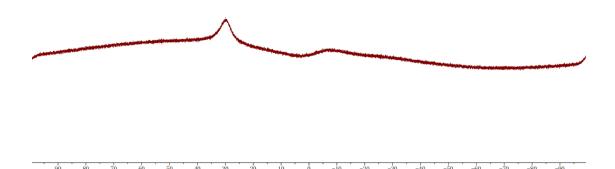




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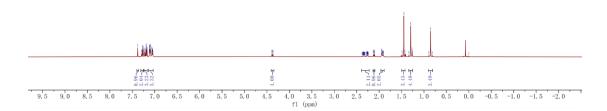
Bpai

<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)

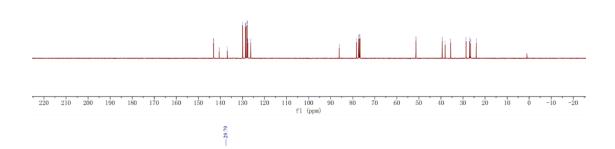


2477888889999

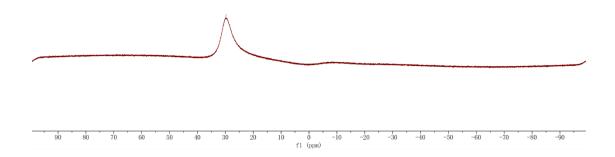
 $^{1}H$  NMR (400 MHz, CDCl<sub>3</sub>)



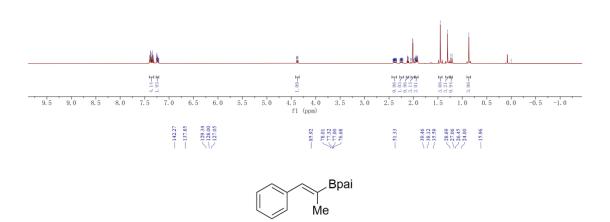
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



<sup>11</sup>B **NMR** (128 MHz, CDCl<sub>3</sub>)

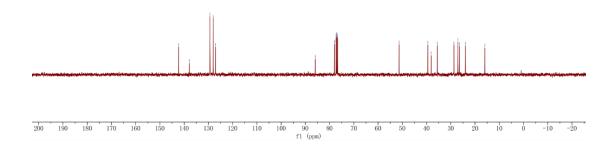


<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



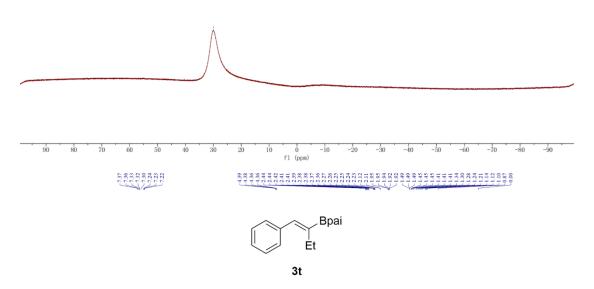
 $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>)

3s

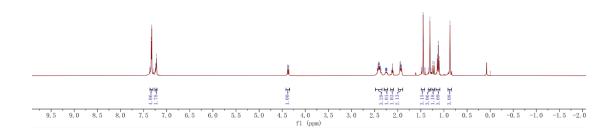




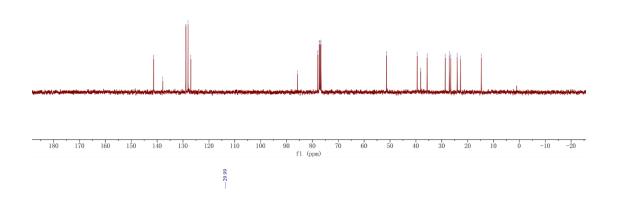
<sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)



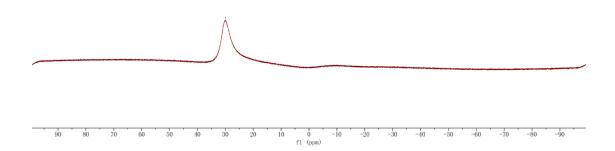
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



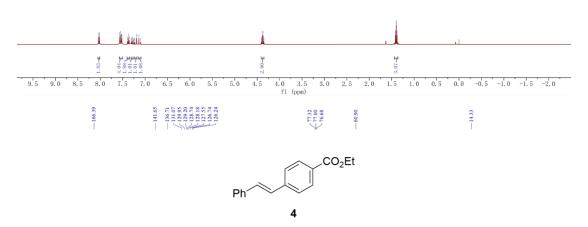
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



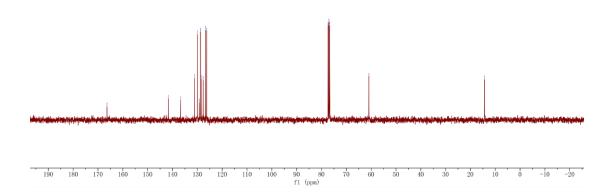
 $^{11}\text{B NMR}$  (128 MHz, CDCl<sub>3</sub>)



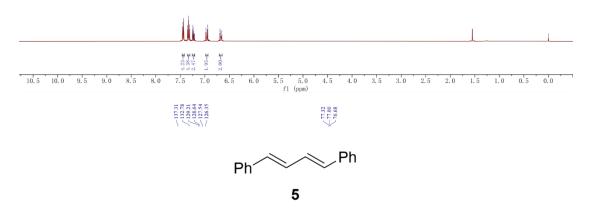
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



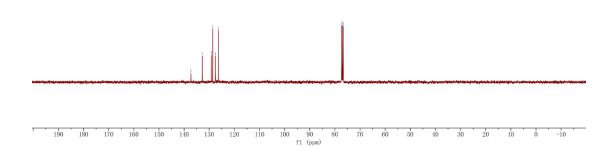
 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)

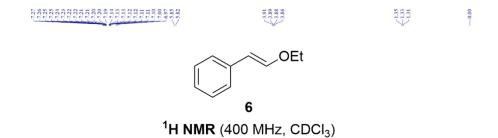


<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

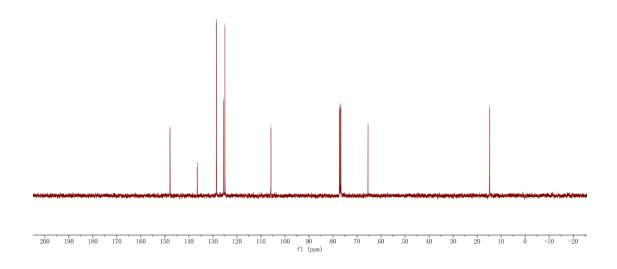


 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)

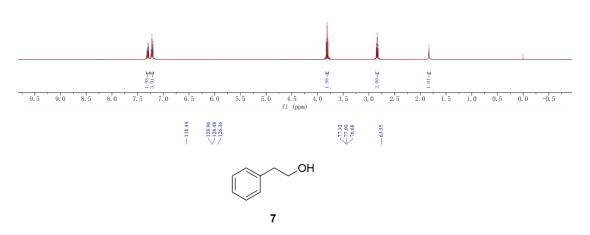




 $^{13}\text{C NMR}$  (101 MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)

