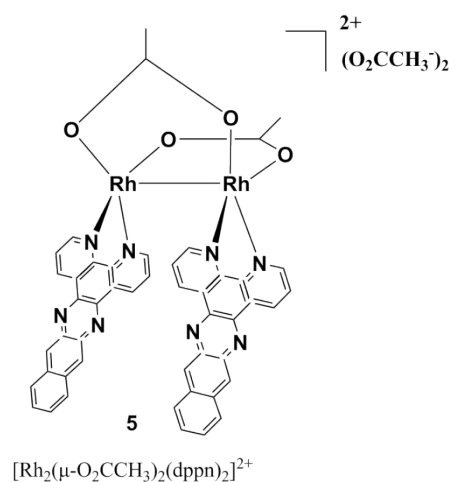


## ELECTRONIC SUPPORTING INFORMATION

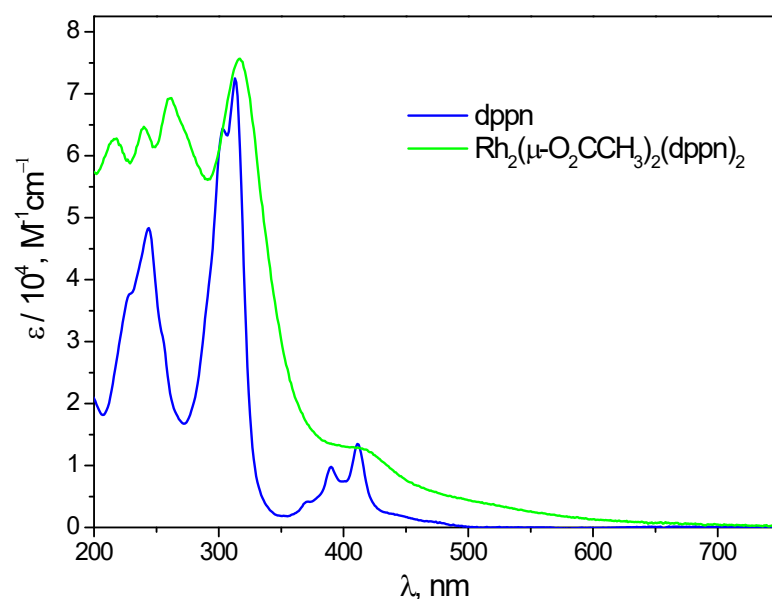
### Photophysics and photochemistry of a prospective light-activated anticancer dirhodium complex

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#### S1. UV spectroscopy of Complex 5



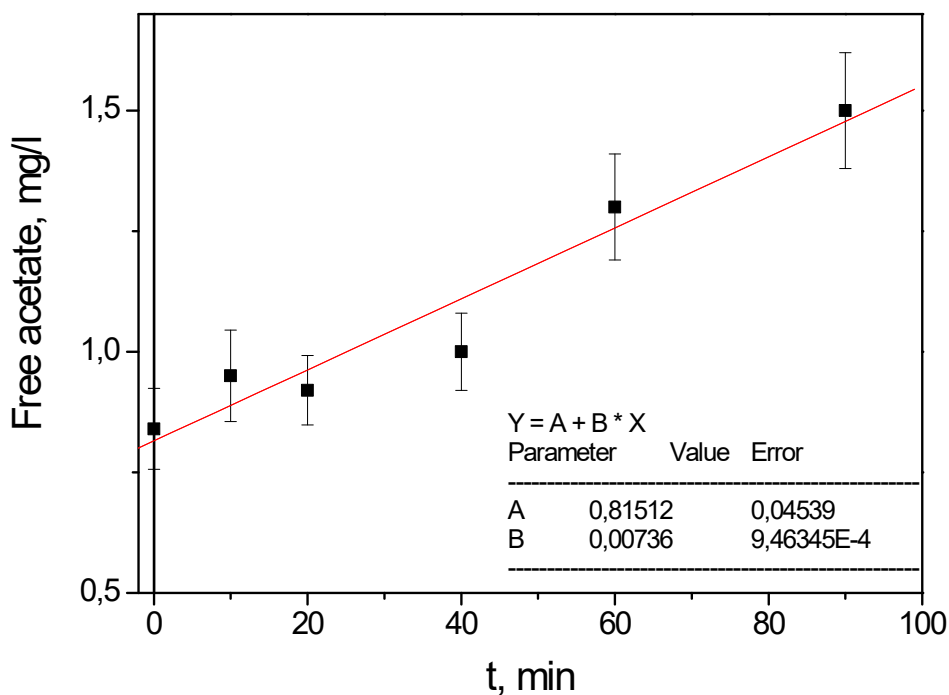
**Scheme S1.** Complex 5.



**Figure S1.** Electronic absorption spectra of dppn in  $\text{CH}_3\text{CN}$  and Complex 5 in  $\text{H}_2\text{O}$ .

## S2. Quantum yield of Complex 5 photoaquation

Quantum yield of reaction (S1) was measured with capillary electrophoresis. The dependence of free acetate ions ( $\text{O}_2\text{CCH}_3^-$ ) released from the first coordination sphere of Rh(II) vs. irradiation time is shown in Fig. S2. The quantum yield was calculated using Eq. SI.



**Figure S2.** Dependence of acetate ions ( $\text{O}_2\text{CCH}_3^-$ ) measured by capillary electrophoresis as a function of irradiation time. Irradiation (282 nm)  $6,87 \times 10^{-6} \text{M}$  of Complex 5 in an 1 cm cell, air-saturated aqueous solution.

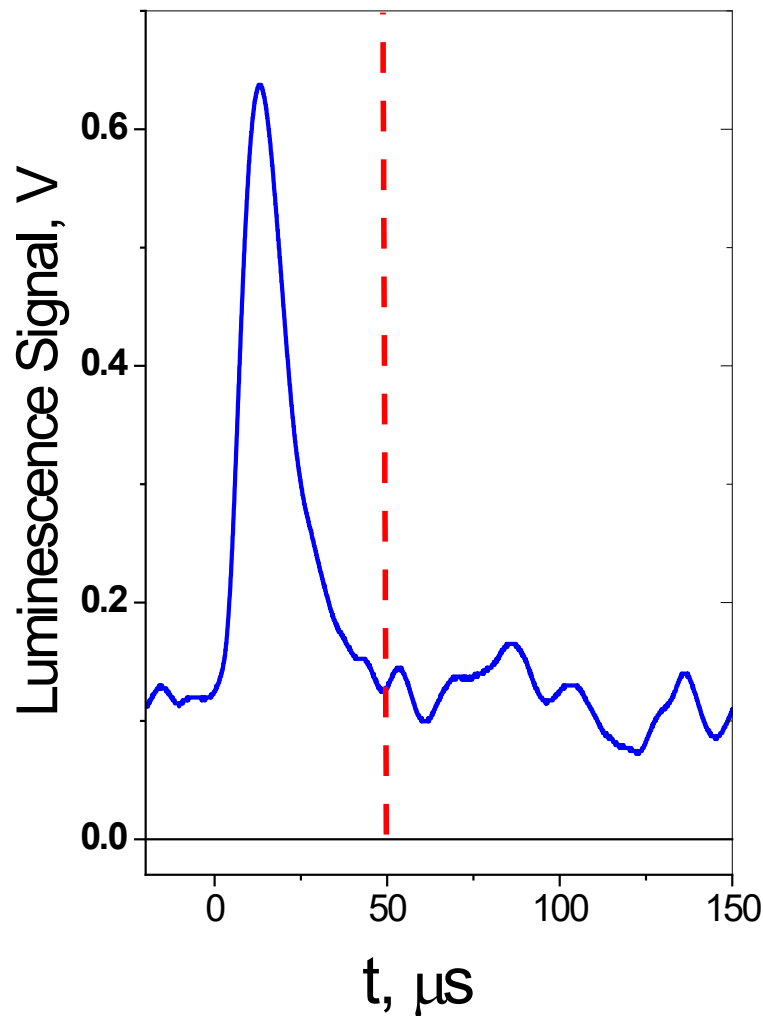


$$\varphi = \frac{dC}{dt} / (I_0 \times (1 - 10^{D(282)})) \quad (\text{SI})$$

$I_0 = 7.6 \times 10^{-4}$  moles of quanta per minute — the concentration (moles per liter) of quanta per one minute provided by the 282 nm excilamp in a 2 ml cell. The light is absorbed completely.

The result is  $\varphi = 0.026$  (relative error ca. 10%).

### S3. Singlet oxygen formation



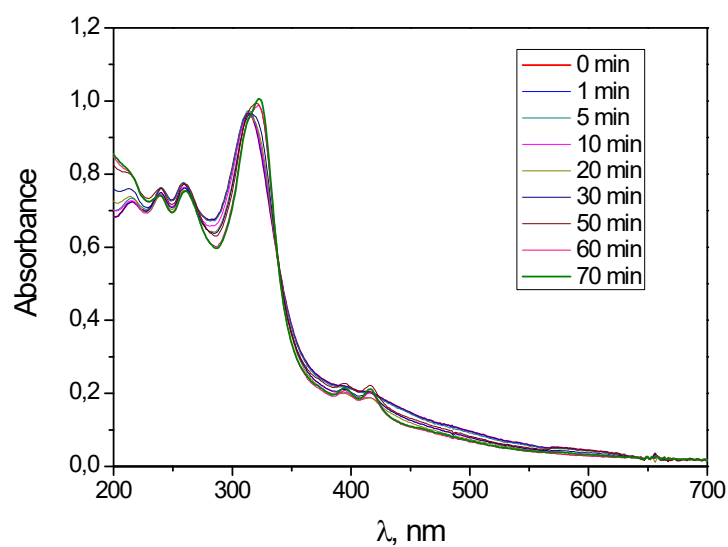
**Figure S3.** Signal of IR photodetector obtained from the quartz cell filled with deionized water. Excitation at 355 nm, laser energy is 5.7 mJ. Red-dash vertical line indicates the short-time edge of temporal interval used for fitting of singlet oxygen luminescence signal.

Quantum yield was calculated with Eq. SII. where  $dA/dE$  and  $dA_{st}/dE$  are the slopes for the target compound and the standard correspondingly,  $D(355)$  and  $D(355)_{st}$  are the absorbancies at the excitation wavelength (355 nm), and  $\varphi_{st}$  is the quantum yield of the singlet oxygen formation for the reference sensitizer phenalenon.

$$\varphi = \varphi_{st} \frac{dA/dE}{dA_{st}/dE} \frac{1 - 10^{-D(355)_{st}}}{1 - 10^{-D(355)}} \quad (\text{SII})$$

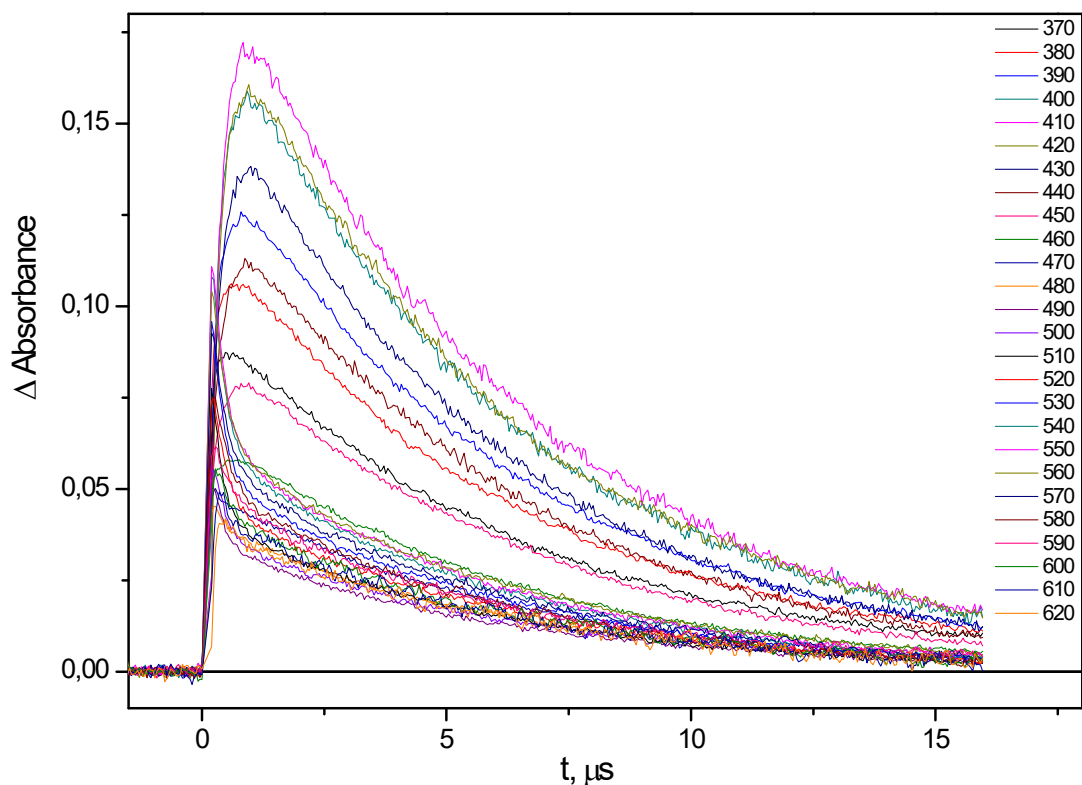
Equation (SII) is satisfied for small values of absorbancies, which didn't overcome 0.3 in our experiments. The slopes  $dA/dE$  and  $dA_{st}/dE$  for pulse energy dependence of the  $^1\text{O}_2$

luminescence amplitude were measured at the fixed absorbance of investigated substances. The example of the determined slope  $dA/dE$  is presented in Fig.4b of Manuscript.



**Figure S4.** Changes of the UV absorption spectra of  $\text{Rh}_2(\mu\text{-O}_2\text{CCH}_3)_2(\text{dppn})_2$  in  $\text{CH}_3\text{CN}$  air-saturated solution during stationary photolysis ( $\lambda = 305 \text{ nm}$ ). 1 cm cell, concentration  $1.35 \times 10^{-5} \text{ M}$ , irradiation time is indicated in the figure.

#### S4. Laser flash photolysis.



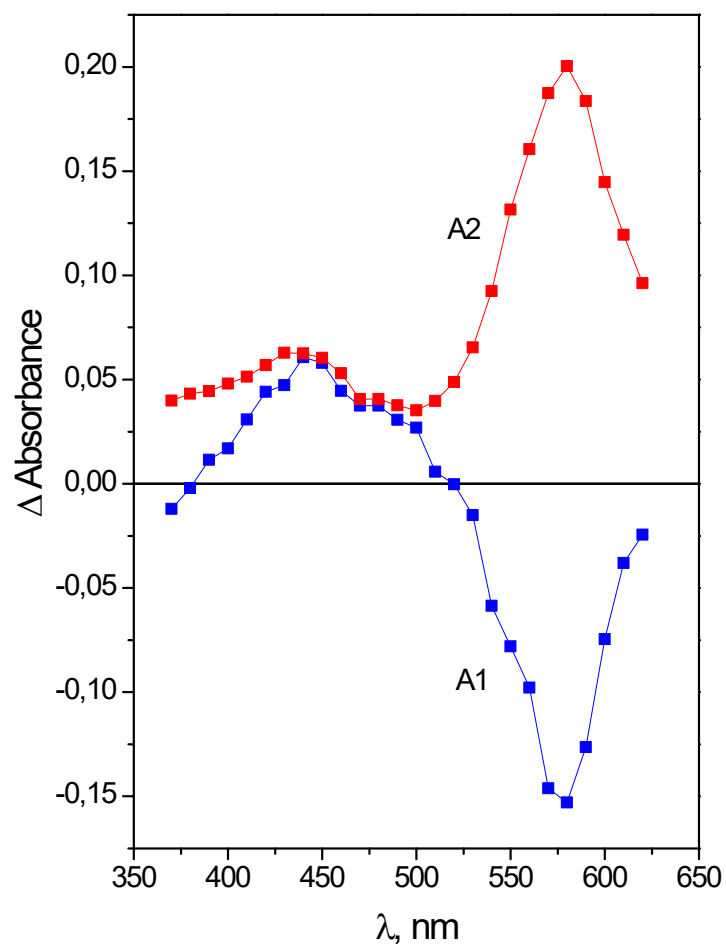
**Figure S5.** Kinetic curves obtained in the course of laser flash photolysis (355 nm) of Complex 5 ( $3.1 \times 10^{-4}$  M in deaerated  $\text{H}_2\text{O}$ ). Wavelengths are depicted.

$$\Delta D(\lambda, t) = A_1(\lambda) \exp\left(-\frac{t}{\tau_1}\right) + A_2(\lambda) \exp\left(-\frac{t}{\tau_2}\right) \quad (\text{SIII})$$

Exact formulae for calculation of Species Associated Difference Spectra (SADS) from the amplitudes of Eq. SIII for the case of sequential transitions of intermediates  $A \rightarrow B \rightarrow (\text{Ground State})$  [S1]:

$$S_A(\lambda) = A_1(\lambda) + A_2(\lambda) \quad (\text{SIV})$$

$$S_B(\lambda) = \left(1 - \frac{\tau_1}{\tau_2}\right) A_2(\lambda) \quad (\text{SV})$$



**Figure S6.** Results of biexponential global fit (Eq. SII) of the kinetic curves (Fig. S5) obtained in the course of laser flash photolysis (355 nm) of Complex **5** ( $3.1 \times 10^{-4}$  M in deaerated  $\text{H}_2\text{O}$ ). Spectra of amplitudes.

## S5. Ultrafast TA spectroscopy

The results of ultrafast TA experiments were globally fitted by 2- or 3-exponential functions with residuals (SVI, SVII). Assuming the successive transitions of the intermediates  $A \rightarrow B \rightarrow C$ , the SADS could be calculated from amplitudes using formulae (SVI, SVIII-SX) [S1]. For the case of big difference in characteristic lifetimes  $\tau_1$  and  $\tau_2$  these formulae could be simplified to the sums of amplitudes (SVIII, SXI, SX).

$$\Delta D(\lambda, t) = A_1(\lambda) \exp(-\frac{t}{\tau_1}) + A_2(\lambda) \exp(-\frac{t}{\tau_2}) + A_3(\lambda) \quad (\text{SVI})$$

$$\Delta D(\lambda, t) = A_1(\lambda) \exp(-\frac{t}{\tau_1}) + A_2(\lambda) \exp(-\frac{t}{\tau_2}) + A_3(\lambda) \exp(-\frac{t}{\tau_3}) + A_4(\lambda) \quad (\text{SVII})$$

$$S_A(\lambda) = A_1(\lambda) + A_2(\lambda) + A_3(\lambda) \quad (\text{SVIII})$$

$$S_B(\lambda) = (1 - \frac{\tau_1}{\tau_2}) A_2(\lambda) + A_3(\lambda) \quad (\text{SIX})$$

$$S_C(\lambda) = A_3(\lambda) \quad (\text{SX})$$

$$S_B(\lambda) = A_2(\lambda) + A_3(\lambda) \quad (\text{SXI})$$

The results of 2-exponential approximation with the residual for the case of Complex **5** ultrafast irradiation at 400 nm are shown in Figs. S7, S8. The characteristic lifetimes are  $\tau_1 = 1.8 \pm 0.7$  ps and  $\tau_2 = 120 \pm 50$  ps.

Assuming the successive transitions of the intermediates  $A \rightarrow B \rightarrow C \rightarrow D$ , the SADS could be calculated from amplitudes using formulae (SVII, SXII-SXV) [S1]. For the case of big difference in characteristic lifetimes  $\tau_1$  and, these formulae could be simplified to the sums of amplitudes (SXII, SXVI, SXVII, SXV).

$$S_A(\lambda) = A_1(\lambda) + A_2(\lambda) + A_3(\lambda) + A_4(\lambda) \quad (\text{SXII})$$

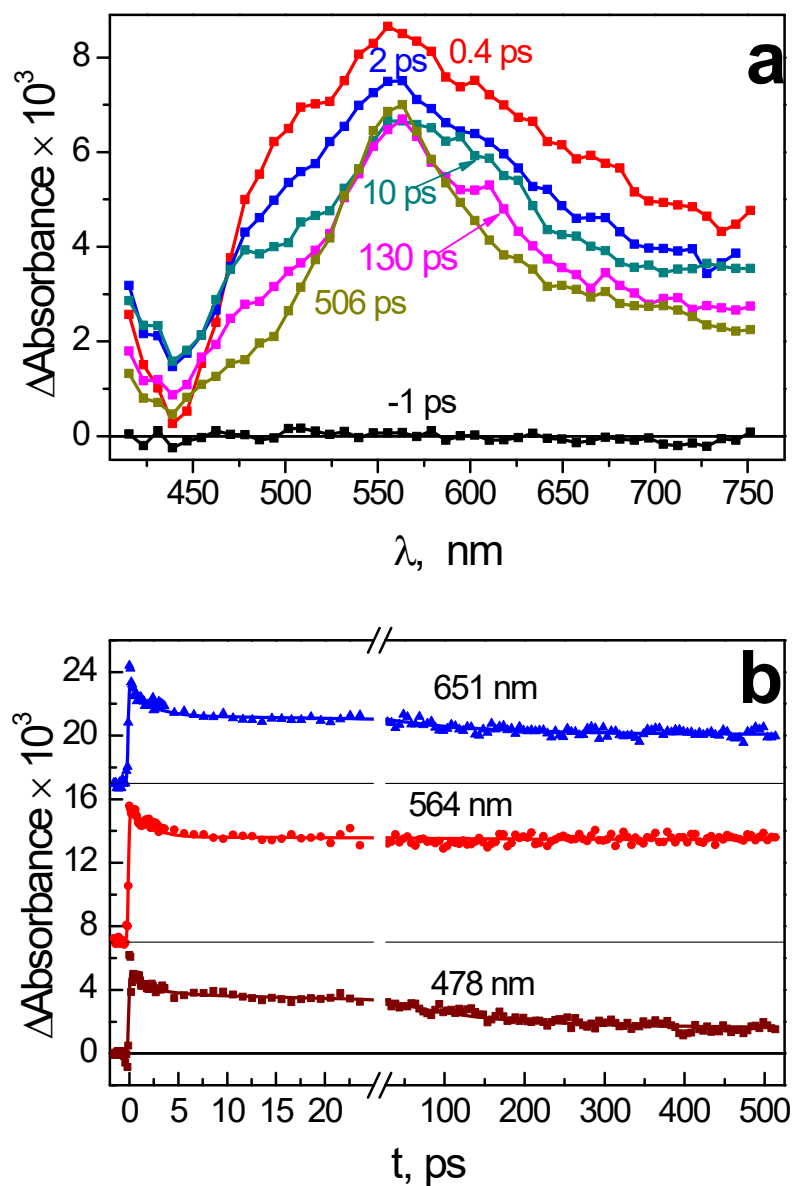
$$S_B(\lambda) = A_2(\lambda) \frac{\tau_2 - \tau_1}{\tau_2} + A_3(\lambda) \frac{\tau_3 - \tau_1}{\tau_3} + A_4(\lambda) \quad (\text{SXIII})$$

$$S_C(\lambda) = A_3(\lambda) \frac{(\tau_3 - \tau_1)(\tau_3 - \tau_2)}{\tau_3^2} + A_4(\lambda) \quad (\text{SXIV})$$

$$S_D(\lambda) = A_4(\lambda) \quad (\text{SXV})$$

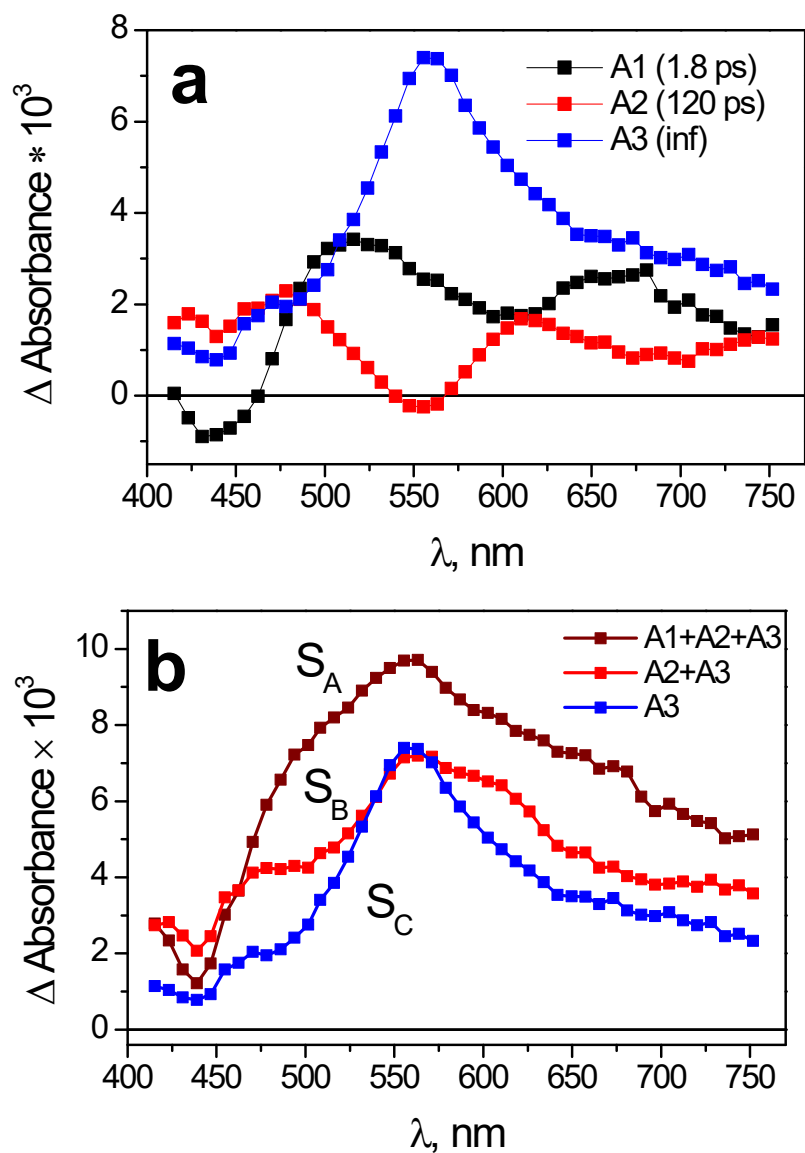
$$S_A(\lambda) = A_2(\lambda) + A_3(\lambda) + A_4(\lambda) \quad (\text{SXVI})$$

$$S_A(\lambda) = A_3(\lambda) + A_4(\lambda) \quad (\text{SXVII})$$

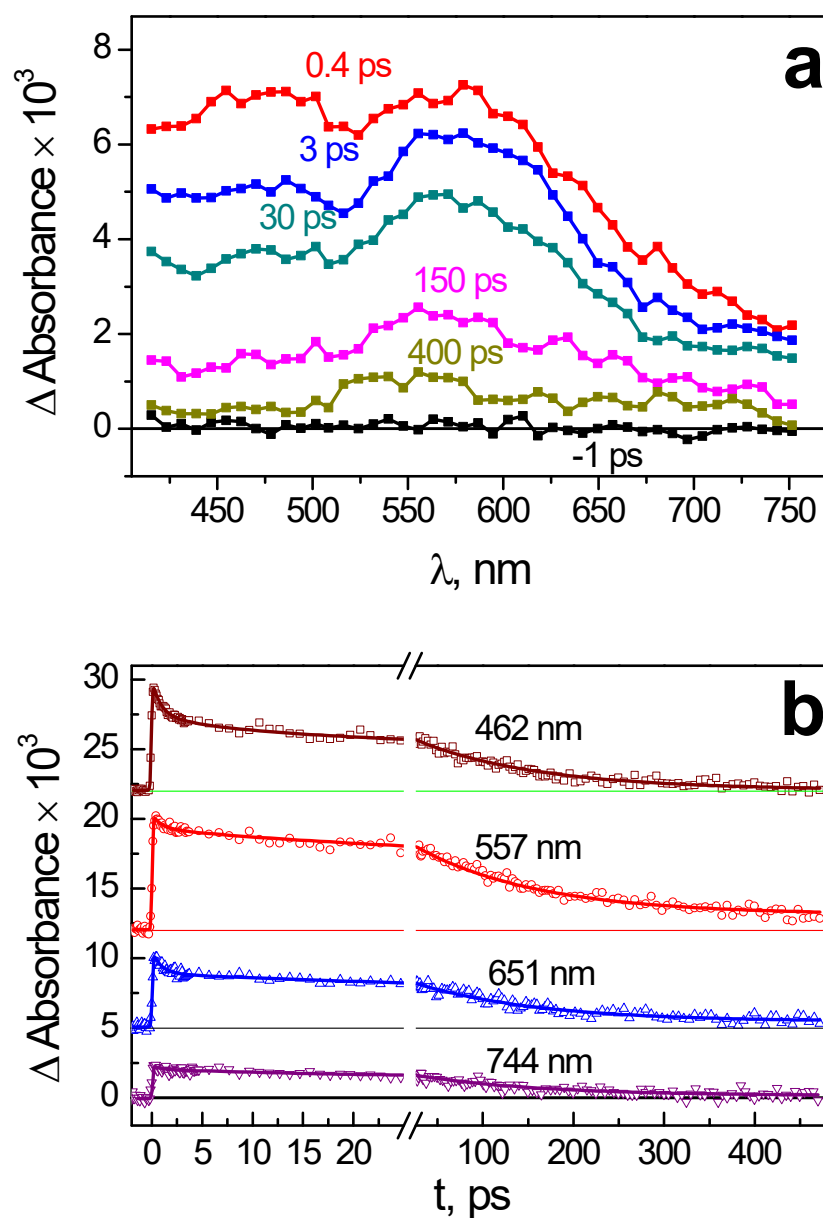


**Figure S7.** **a** - TA spectra obtained in ultrafast pump-probe experiment with Complex **5** in  $\text{H}_2\text{O}$  ( $1.2 \times 10^{-4} \text{ M}$ , 1 mm cell). Excitation at 400 nm. Curves 1-6 correspond to time delays between exciting and probing pulses equal to -1; 0.4; 2; 10; 130; 506 ps. **b** - examples of kinetic curves obtained by processing TA spectra and their 2-exponential global fit by function (SVI).

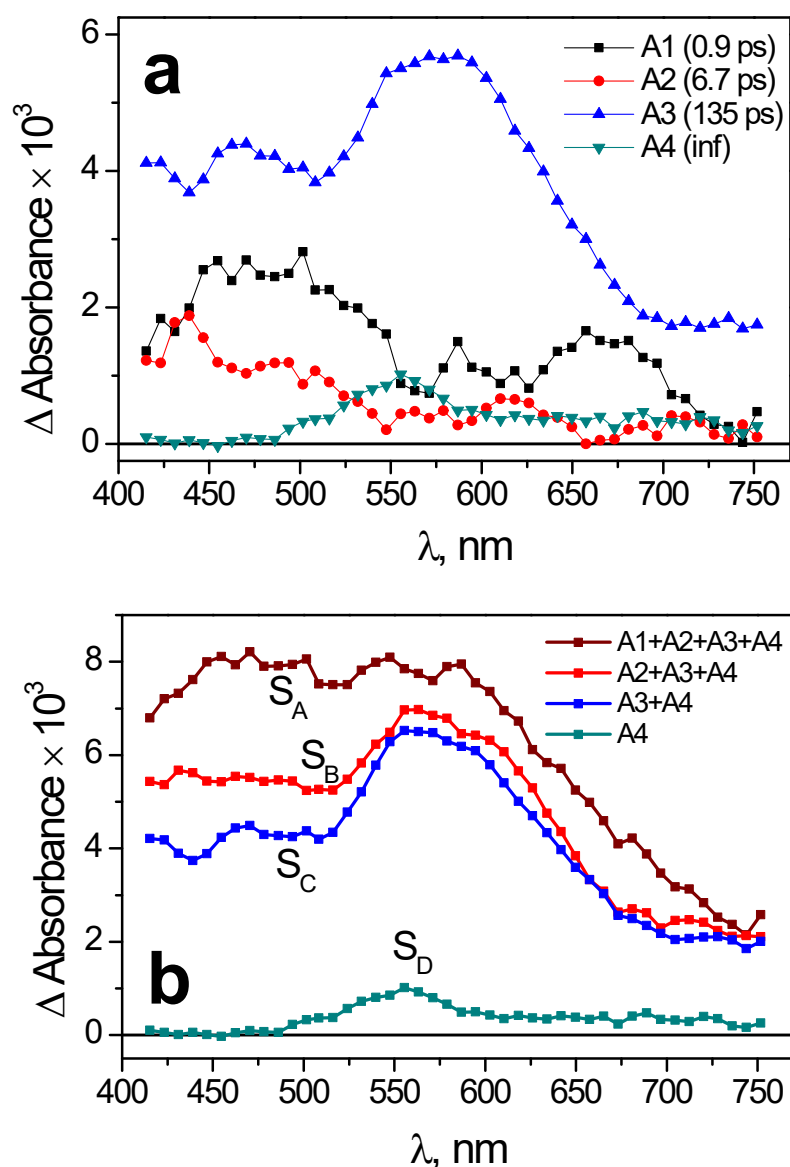




**Figure S8.** Results of processing of the TA spectra obtained in ultrafast pump-probe experiment with Complex **5** in H<sub>2</sub>O ( $1.2 \times 10^{-4}$  M, 1 mm cell, excitation at 400 nm) by formulae (SVI, SVIII, SXI, SX). **a** – amplitudes  $A_i(\lambda)$ , **b** – SADS. The characteristic lifetimes are  $\tau_1 = 1.8 \pm 0.7$  ps;  $\tau_2 = 120 \pm 50$  ps.



**Figure S9.** **a** - TA spectra obtained in ultrafast pump-probe experiment with Complex **5** in H<sub>2</sub>O ( $1.28 \times 10^{-4}$  M, 1 mm cell). Excitation at 320 nm. Curves 1-6 correspond to time delays between exciting and probing pulses equal to -1; 0.4; 2; 10; 130; 506 ps. **b** - examples of kinetic curves obtained by processing TA spectra and their 3-exponential global fit by function (SVII).



**Figure S10.** Results of processing of the TA spectra obtained in ultrafast pump-probe experiment with Complex **5** in H<sub>2</sub>O ( $1.28 \times 10^{-4}$  M, 1 mm cell, excitation at 320 nm) by formulae (SXII, SXVI, SXVII, SXV). **a** – amplitudes  $A_i(\lambda)$ , **b** – SADS. The characteristic lifetimes are  $\tau_1 = 0.9 \pm 0.4$  ps,  $\tau_2 = 6.7 \pm 4.6$  ps and  $\tau_3 = 135 \pm 18$  ps.

## References

- [S1] A.S. Rury and R.J. Sension, *Chem. Phys.*, 2013, **422**, 220-228.  
<https://doi.org/10.1016/j.chemphys.2013.01.025>