

*Supporting Information for*

# Conformational and Environmental Determinants of RNA Solvation Dynamics: Roles of Intrinsic Flexibility, Allostery, and Protein Binding

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**This PDF file includes:**

## **Supporting Results**

- ❖ Bulk salt concentration calculation
- ❖ Comparison of probability distribution of water's residence time around TAR RNA in BIV apo to complex state.
- ❖ Power-law fitting of solvation decay computed for BIV complex state
- ❖ Computing solvation correlation function and timescale prediction at 1ps resolution and timescale prediction
- ❖ Comparison of solvation correlation function,  $C_s(t)$ , decay profiles for BIV TAR-TAT complex with varying trajectories
- ❖ Solvation dynamics calculated for HIV-2 TAR and HIV-2 TAR-TAT complex and respective timescale is predicated upon fitting the plots using triexponential function
- ❖ Periodic energy fluctuation and distance fluctuation happening coherently for HIV-2 Apo

- ❖ Decomposition of solvation energy relaxation into distinct self and cross terms for the HIV-2
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**Bulk salt concentration calculation:**

Time-averaged bulk salt concentrations are calculated by taking the time-averaged ratio between the numbers of bulk ions to that of the number of bulk water molecules multiplied by the molarity of pure water. All species which are present beyond 2 nm of the RNA are considered to be bulk species according to the radial distribution function as depicted in **RDF plots**.

$$[Cl^-]^* = 55.51M \left\langle \frac{B_{Cl^-}}{B_{H_2O}} \right\rangle \quad (\text{eq 1})$$

$$[Na^+]^* = 55.51M \left\langle \frac{B_{Na^+}}{B_{H_2O}} \right\rangle \quad (\text{eq 2})$$

From this raw concentration one has to determine the corrected concentration of the ionic species in the bulk following the relation as:

$$[j] = [j]^* \left\{ 1 - q_j \frac{\sum q_i [i^*]}{\sum q_i^2 [i^*]} \right\} \quad (\text{eq 3})$$

Here,  $q_j$  is the charge of the ionic species,  $[j]^*$  and  $[j]$  denotes the raw concentration and the corrected concentration, respectively. The bulk ionic concentration of NaCl for both the system was found to be within the range of  $100 \pm 10 \text{mM}$ , which is in the physiological ionic range. Details about the value of the ionic concentrations are given in **Table S1, S2, and S3**.

**Table S1: Accounting for total number of surrounding species to the system**

System	No. of Water molecule	No. of Sodium ion	Number of Chloride ion
BIV TAR RNA	56308	116	89
BIV TAR-TAT Complex	55974	116	97
HIV-2 TAR RNA	56003	133	104
HIV-2 TAR-TAT Complex	55920	125	104

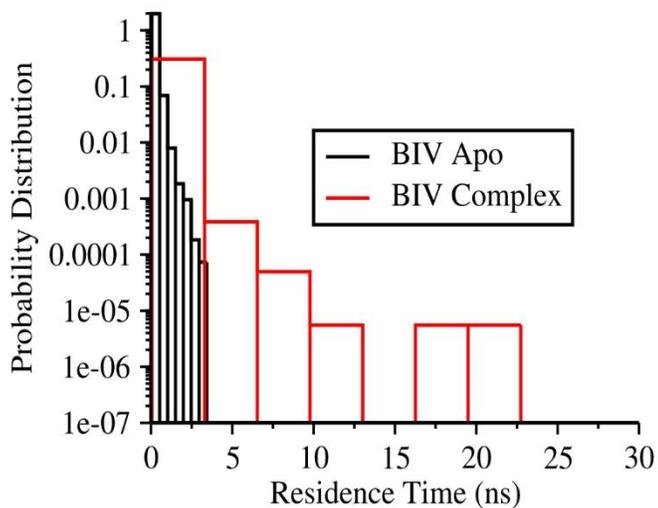
**Table S2: Accounting for bulk number of surrounding species to the system**

System	No. of bulk Water molecule	No. of bulk Sodium ion	No. of bulk Chloride ion
BIV TAR RNA	50306.03	88.35	83.11
BIV TAR-TAT Complex	49911.18	94.24	90
HIV-2 TAR RNA	49868.86	101.11	96.52
HIV-2 TAR-TAT Complex	49688.44	111.93	107.27

**Table S3: Accounting for corrected concentration of ions and preferential interaction coefficient**

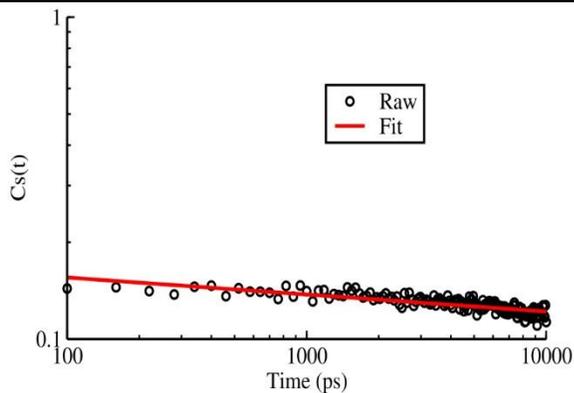
System	Ions	Rough concentration(mM)	Corrected Concentration(mM)	Preferential interaction coefficient
BIV TAR RNA	Na+	97.49	94.51	20.13
	Cl-	91.71	94.51	-6.87
BIV TAR-TAT Complex	Na+	104.81	102.4	12.75
	Cl-	100.1	102.4	-6.25
HIV-2 TAR RNA	Na+	112.55	109.93	22.09
	Cl-	107.44	109.93	-6.91
HIV-2 TAR-TAT Complex	Na+	111.93	109.55	14.64
	Cl-	107.27	109.55	-6.36

**Comparison of probability distribution of water's residence time around TAR RNA in BIV apo to complex state:**



**Figure S1: Comparison of the residence time of water molecules in the first solvation shell of BIV TAR RNA in the apo state versus the TAT-bound complex.**

**Power-law fitting of solvation decay computed for BIV complex state:**

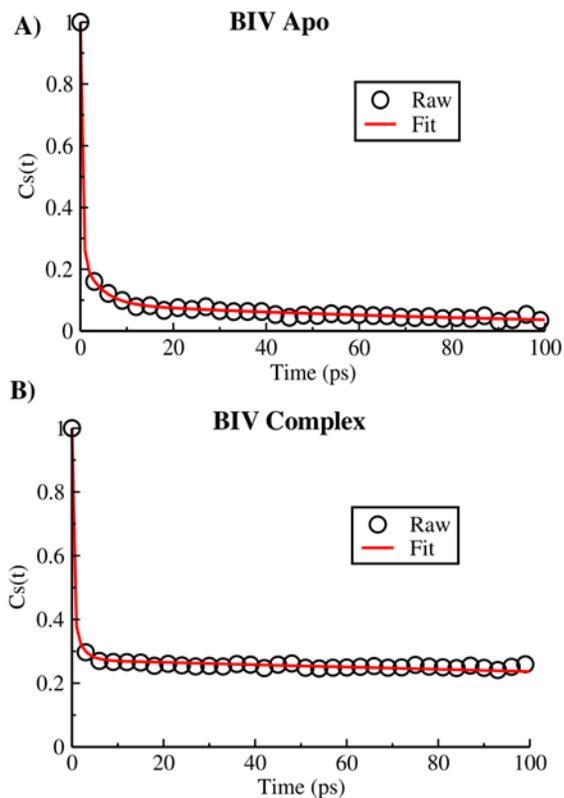


**Figure S2: Power-law<sup>1</sup> fitting of the decay profile of the BIV TAR–TAT complex within the 100 ps to 10 ns time window (shown on a log–log scale).**

**Table S4: Fitting parameter for BIV TAR-TAT Complex (Power-Law)**

<b>a0</b>	<b>a1</b>
<b>0.198</b>	<b>0.0529</b>

**Computing solvation correlation function and timescale prediction at 1ps resolution and timescale prediction:**



**Figure S3: Multi-exponential fitting of BIV TAR solvation dynamics at 1 ps resolution for (A) the apo state and (B) the TAT-bound complex state.**

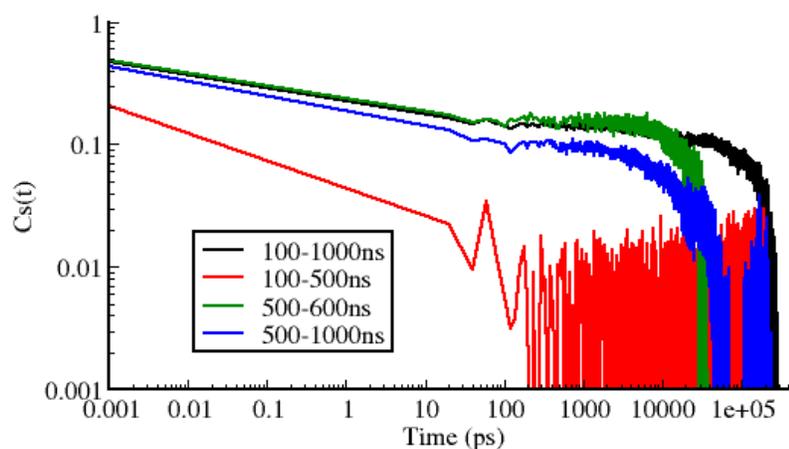
**Table S5: Fitting parameter for BIV Apo (Tri-exponential<sup>2,3</sup>, resolution=1ps)**

<b>a1</b>	<b>a2</b>	<b>a3</b>	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>&lt;T&gt;</b>
<b>0.751</b>	<b>0.162</b>	<b>0.087</b>	<b>0.36 ps</b>	<b>4.02 ps</b>	<b>114.43 ps</b>	<b>10.88 ps</b>

**Table S6: Fitting parameter for BIV Complex (Tri-exponential<sup>2,3</sup>, resolution=1ps)**

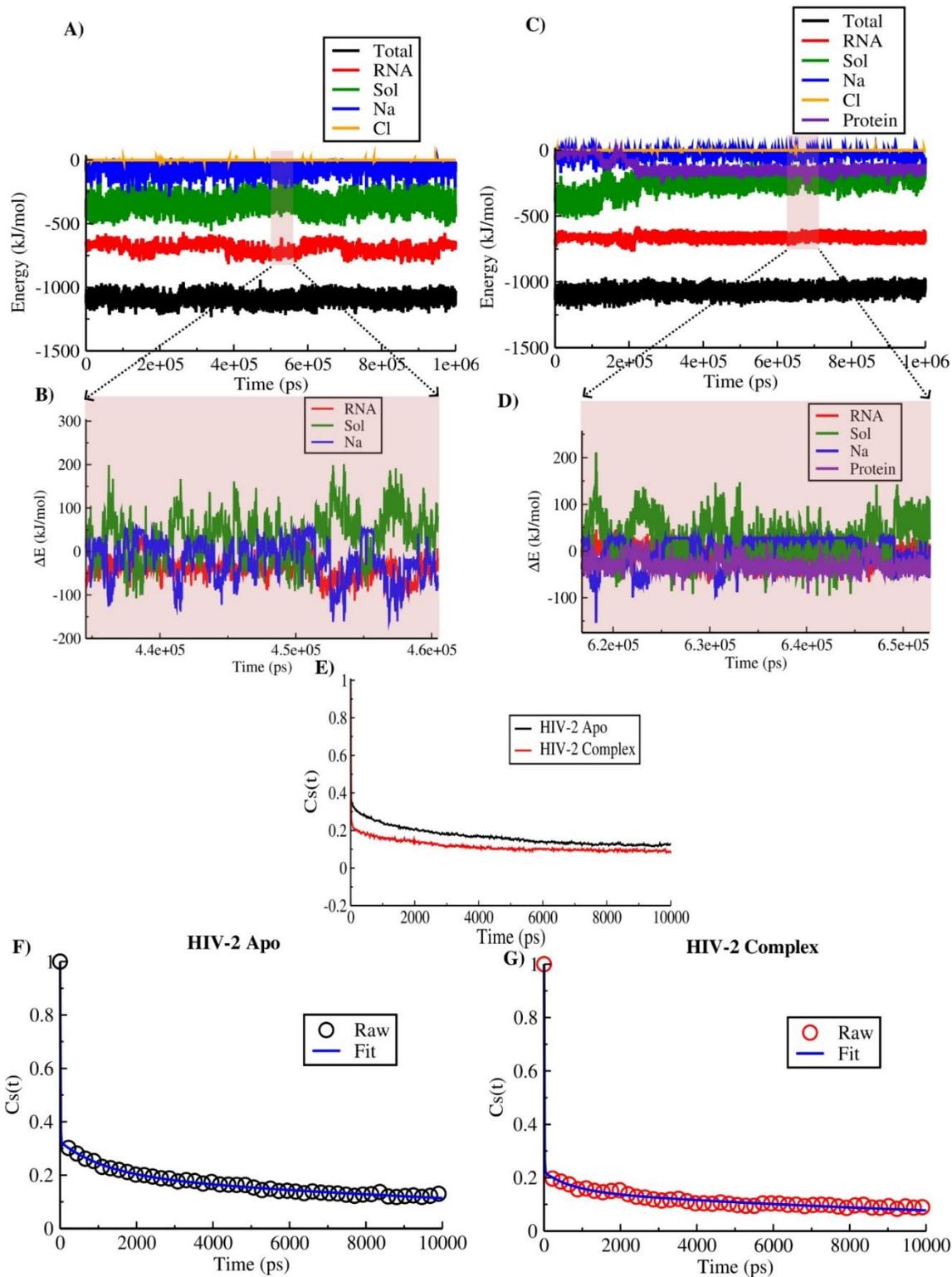
a1	a2	a3	T1	T2	T3	<T>
0.589	0.138	0.273	0.32 ps	1.86 ps	700.0 ps	191.55 ps

**Comparison of decay profiles for BIV TAR-TAT complex with varying trajectories:**



**Figure S4: Comparison of solvation time-correlation function  $C_s(t)$  for the BIV TAR RNA in the TAT-bound complex, evaluated across trajectory segments of varying lengths (100–1000 ns, 100–500 ns, 500–600 ns, and 500–1000 ns).**

**Solvation dynamics calculated for HIV-2 TAR and HIV-2 TAR-TAT complex and respective timescale is predicated upon fitting the plots using triexponential function:**



**Figure S5: Decomposition of solvation energy relaxation into distinct environmental contributions for the HIV-2 TAR RNA in its apo and TAT-bound complex states.**

(A) Probe–rest interaction energy along the trajectory for the HIV-2 apo RNA, shown as both the total contribution and its decomposition into individual components. (B) Enlarged view of panel (A), highlighting fluctuations in interaction energy ( $\Delta E$ ) for the RNA, solvent, and Na<sup>+</sup> ion components. (C) and (D) Same analyses as in panels (A) and (B), respectively, but for the HIV-2 TAR–TAT complex. (E) Comparative decay profiles of the solvation time-correlation function for the apo and complex systems at 20 ps resolution. (F) and (G) Triexponential fits to the solvation decay profiles for the apo and complex states, respectively.

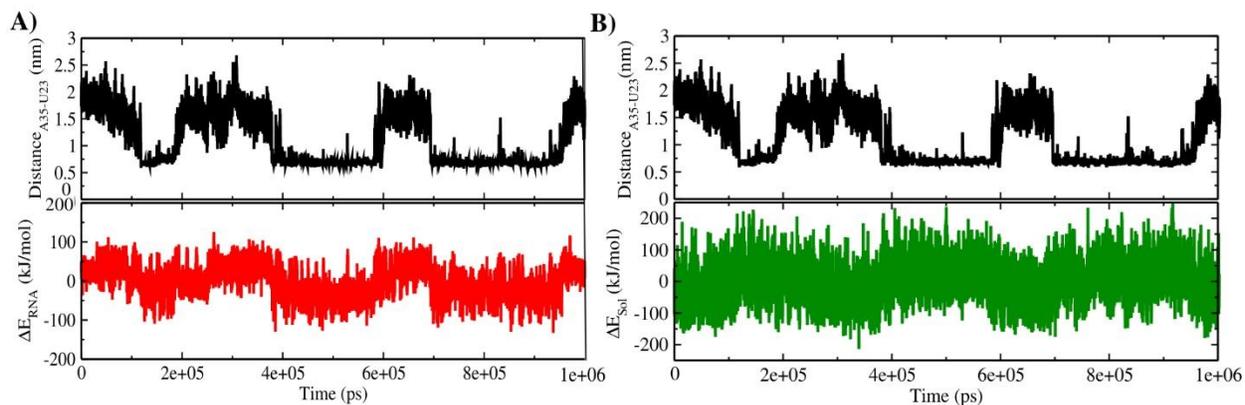
**Table S7: Fitting parameter for HIV-2 Apo (Tri-exponential, resolution=20ps)**

<b>a1</b>	<b>a2</b>	<b>a3</b>	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>&lt;T&gt;</b>
<b>0.675</b>	<b>0.122</b>	<b>0.203</b>	<b>7.38 ps</b>	<b>1184.66 ps</b>	<b>17377.2 ps</b>	<b>3677.08 ps</b>

**Table S8: Fitting parameter for HIV-2 Complex (Tri-exponential, resolution=20ps)**

<b>a1</b>	<b>a2</b>	<b>a3</b>	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>&lt;T&gt;</b>
<b>0.779</b>	<b>0.07</b>	<b>0.151</b>	<b>7.32 ps</b>	<b>746.97ps</b>	<b>15000 ps</b>	<b>2322.99 ps</b>

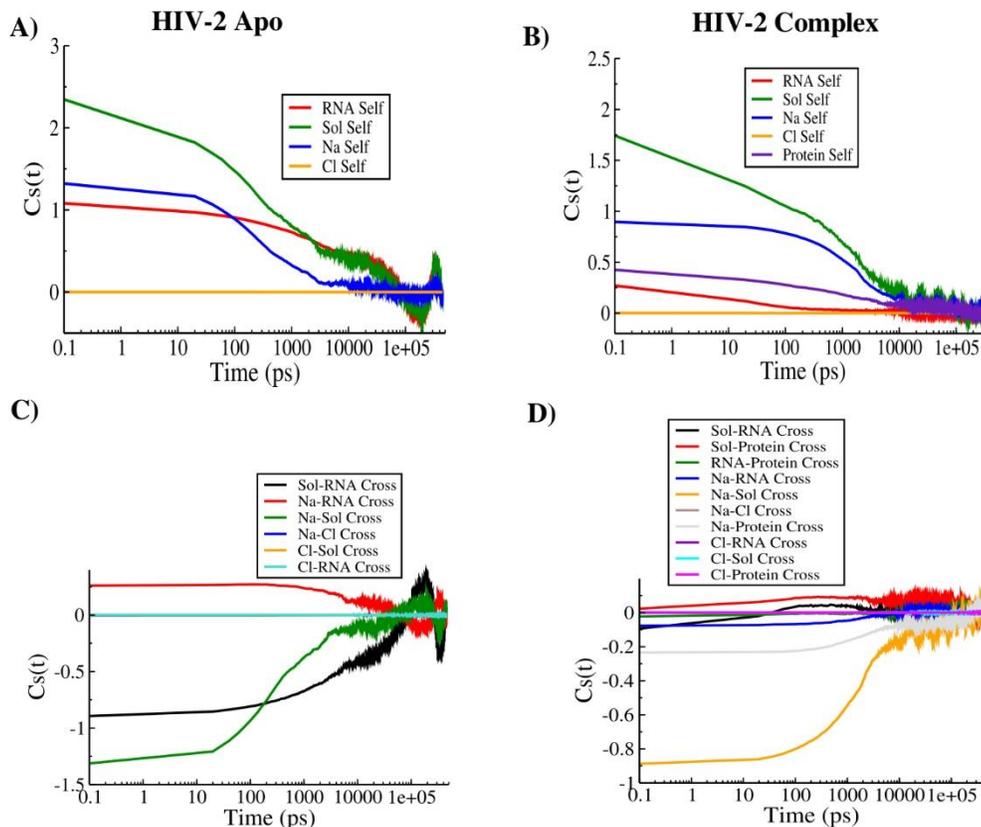
**Periodic energy fluctuation and distance fluctuation happening coherently for HIV-2 Apo:**



**Figure S6: Time evolution of structural energy fluctuations and their coherent periodicity with the A35 distance fluctuation, illustrating a distal conformational response sensed by the probe.**

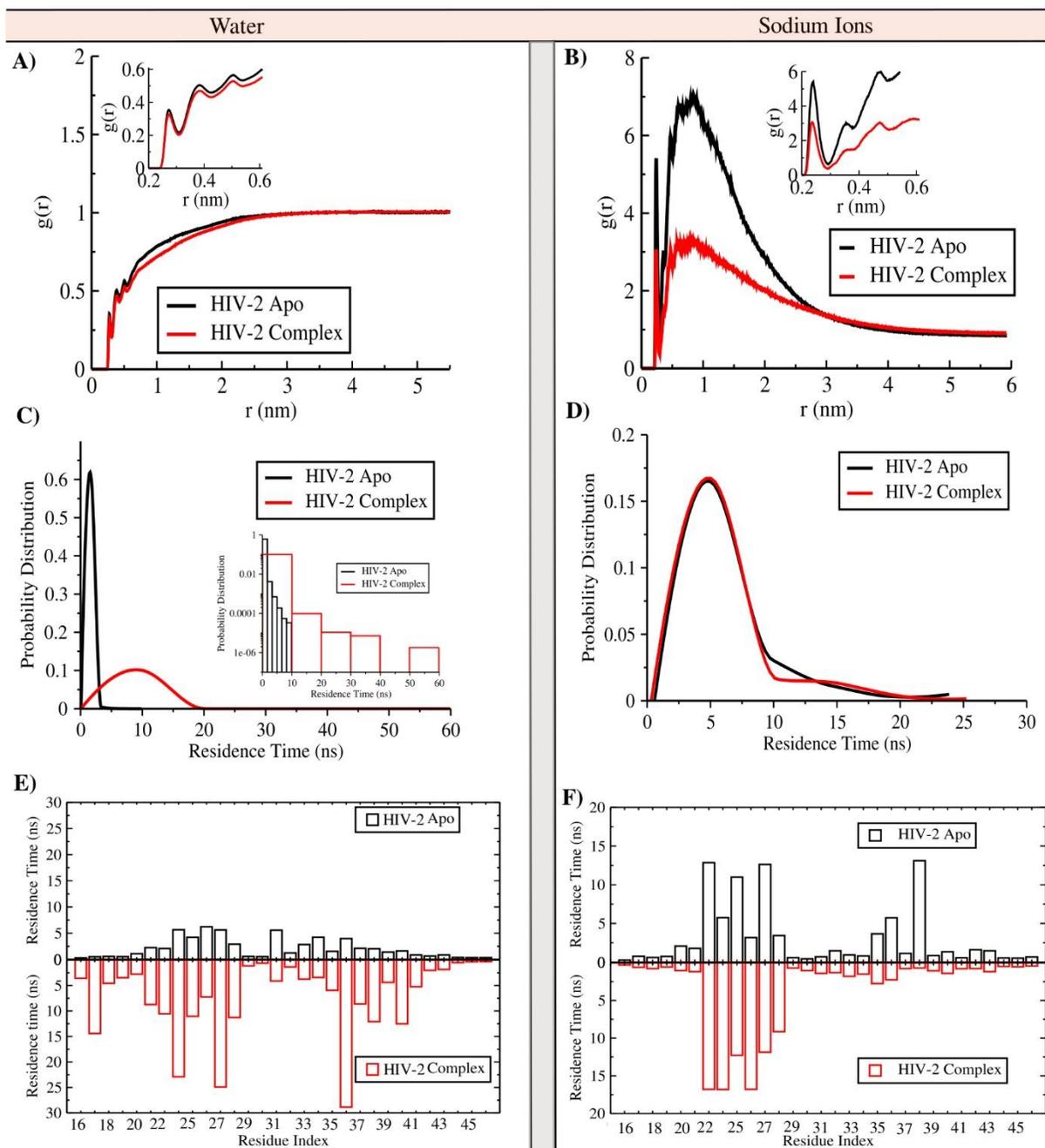
## Decomposition of solvation energy relaxation into distinct self and cross terms for the HIV-

2:



**Figure S7: Linear-response decomposition<sup>4,5</sup> of the decay function of HIV-2 TAR RNA into self and cross terms.** (A) and (B) Self-term and cross-term contributions of the individual components governing the decay in the HIV-2 apo TAR. (C) and (D) Corresponding self-term and cross-term decompositions for the HIV-2 TAR–TAT complex.

## Comparing characterizations of solvation environment for HIV-2 Apo vs complex:



**Figure S8: Characterization of the water and ion ( $\text{Na}^+$ ) environment surrounding the RNA duplex in the apo and TAT-bound HIV-2 TAR states. (A) and (B) Radial distribution functions of water and  $\text{Na}^+$  ions, respectively, around all RNA heavy atoms, shown in a comparative manner for the apo and complex forms. (C) and (D) Distributions of residence times for water and  $\text{Na}^+$  ions within the first hydration shell (up to  $3.5 \text{ \AA}$ ) of the RNA in both states. (E) and (F) Changes**

in the residue-wise maximum residence times of first-shell water and Na<sup>+</sup> ions, respectively, upon transition from the apo state to the TAT-bound complex.

### References:

- 1 D. Andreatta, J. L. Pérez Lustres, S. A. Kovalenko, N. P. Ernsting, C. J. Murphy, R. S. Coleman and M. A. Berg, Power-law solvation dynamics in DNA over six decades in time, *J Am Chem Soc*, DOI:10.1021/ja044177v.
- 2 R. Jimenez, G. R. Fleming, P. V. Kumar and M. Maroncelli, Femtosecond solvation dynamics of water, *Nature*, DOI:10.1038/369471a0.
- 3 B. Bagchi and B. Jana, Solvation dynamics in dipolar liquids, *Chem Soc Rev*, DOI:10.1039/b902048a.
- 4 S. Pal, P. K. Maiti, B. Bagchi and J. T. Hynes, Multiple time scales in solvation dynamics of DNA in aqueous solution: The role of water, counterions, and cross-correlations, *Journal of Physical Chemistry B*, DOI:10.1021/jp065690t.
- 5 K. E. Furse and S. A. Corcelli, Molecular dynamics simulations of DNA solvation dynamics, *Journal of Physical Chemistry Letters*, DOI:10.1021/jz100485e.