

## Supporting information

### **Aggregation polymorphism of photoactivatable Pt(IV) prodrugs as a strategy for the design of theranostic nanomaterials**

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## 1. Materials and methods

Acetic acid, 2-azidoacetic acid, boron trifluoride etherate (BF<sub>3</sub>-Et<sub>2</sub>O), cisplatin, 2,3-dichloride-5,6-dicyano-p-benzoquinone (DDQ), 2,4-dimethyl-1H-pyrrole, N,N'-dicyclohexylcarbodiimide (DCC), hydrogen peroxide (30% w/w in water), 4-hydroxybenzaldehyde, propargyl bromide, potassium carbonate (K<sub>2</sub>CO<sub>3</sub>), sodium azide (NaN<sub>3</sub>), sodium ascorbate, trifluoroacetic acid (TFA), triethylamine (Et<sub>3</sub>N), 4-Nitrobenzaldehyde, pentafluorobenzaldehyde were used without purification. Acetone, acetonitrile (CH<sub>3</sub>CN), dichloromethane (DCM, CH<sub>2</sub>Cl<sub>2</sub>), diethyl ether (Et<sub>2</sub>O), methanol (MeOH), dimethylformamide (DMF), acetone, Pluronic® F127 (F127), DSPE-PEG<sub>2000</sub>-OMe, dimethyl sulfoxide (DMSO) and petroleum ether were purchased from commercial sources and purified following the described procedures.<sup>1</sup>

**Analytical thin-layer chromatography (TLC)** was performed on Merck silica gel aluminum plates with F-254 indicator. Compounds were visualized by irradiation with UV light (254, 365 nm).

**Preparative column chromatography** was performed using Acros brand silica gel (60–200 mesh).

**The NMR spectra** were recorded on a Bruker Avance 400 MHz spectrometer (USA) in DMSO-d<sub>6</sub> and CDCl<sub>3</sub> with TMS as an internal standard for <sup>1</sup>H and <sup>13</sup>C and K<sub>2</sub>PtCl<sub>6</sub> for <sup>195</sup>Pt NMR. All <sup>13</sup>C spectra were <sup>1</sup>H decoupled.

**High-resolution mass spectra (HRMS)** were recorded with a G3 QToF quadrupole-time-of-flight (Waters, USA) equipped with an ESI (ionization: ESI, capillary temperature 300 °C, voltage on the capillary: 5500 V for positive mode, -4500 V for negative mode, declustering potential: 90 V. Mass-spectra were recorded in m/z range 150 – 3000, accumulation time 250 ms.

**Purity** of the compounds was assessed using HPLC-DAD-MS. HPLC-DAD-MS system consisting of a Vanquish liquid chromatograph (Thermo Fisher Scientific, USA) and an Orbitrap Fusion Lumos Tribrid (Thermo Fisher Scientific, USA) mass spectrometer was used. A Shim-pack GIST C18-Aq liquid chromatography reversed-phase column (3 x 150 mm, 3 μm, Shimadzu, Japan) was used.

HPLC-DAD-MS system consisting of a Vanquish liquid chromatograph (Thermo Fisher Scientific, USA) and an Orbitrap Fusion Lumos Tribrid (Thermo Fisher Scientific, USA) mass spectrometer was used. A Shim-pack GIST C18-Aq liquid chromatography reversed-phase column (3 x 150 mm, 3 μm, Shimadzu, Japan) was used.

**HPLC-HRMS** analysis was performed with Vanquish liquid chromatograph (Thermo Fisher Scientific, USA) and a high-resolution mass spectrometer based on the G3 QToF quadrupole-time-of-flight (Waters, USA) equipped with an electrospray ionization source (ESI) was used.

To separate the components of the analyzed solution, a Shim-pack GIST C18-Aq chromatographic column (3 x 150 mm, 3 μm, Shimadzu, Japan) filled with a reverse-phase sorbent with polar-endcapping was used. The column temperature was maintained at 35 °C throughout the analysis using a thermostat. A 0.1% aqueous solution of formic acid (A) and acetonitrile

(B) were used as eluents. An isocratic elution mode was used with a ratio of eluents A and B of 15:85%. The flow rate of the mobile phase was 0.4 ml/min. The injected sample volume was 1.0  $\mu\text{l}$ . The analysis time was 10 minutes. The ESI-MS conditions on the Orbitrap Fusion Lumos Tribrid were as follows: mode for recording positively and negatively charged molecules; the capillary voltage of the ionization source was 3500 V for the positive mode and -2500 V for the negative mode; ion source chamber temperature - 350  $^{\circ}\text{C}$ ; ion transfer interface temperature - 325  $^{\circ}\text{C}$ ; gas pressure for solvent atomization in the ion source (nitrogen) - 50, auxiliary gas pressure - 10, curtain gas pressure - 1. Scanning range  $m/z$ : 200-1700 Da. The resolution of the mass spectrometer for analysis is not less than 15000. The mass spectrometer was calibrated immediately prior to sample analysis using CalMix Pierce<sup>TM</sup> calibration mixtures (Thermo Scientific, USA). Data processing was performed using Xcalibur 4.6 software (Thermo Scientific, USA).

**ICP-MS.** Determination of Pt concentration was carried out with a quadrupole ICP-MS spectrometer PlasmaQuant MS Elit (AnalytikJena, Germany). The data were acquired and processed with the ASpect MS software package (version 4.3, AnalytikJena). ICP-MS procedure included determining of following isotopes:  $^{195}\text{Pt}$  as the analyte and  $^{175}\text{Lu}$  as the internal standard (the concentration of Lu 1  $\mu\text{g L}^{-1}$ ).

The ICP-MS-68A-C multi-element standard solution (High-Purity Standards, USA), containing 10  $\text{mg L}^{-1}$  Ru, Rh, Pd, Os, Ir, Pt and Au, was used to prepare calibration solutions (0.2, 0.5, 1, 2, 5, 10, 20  $\mu\text{g L}^{-1}$ ). A single-element solution of lutetium (1  $\text{g L}^{-1}$ ) was obtained by dissolution of lutetium oxide. All solutions were prepared using deionized water (18.2  $\text{M}\Omega\text{ cm}$ , Millipore, France).

**Temperature monitoring** was conducted using the IR camera Gobi-384-GigE (Envision Xenics, Belgium). The recorded objects were placed at 25 cm from the camera to place them in focus of the camera. Data was recorded in an unaltered configuration at 10 fps, the obtained images were processed in the Xeneth software v.2.6.0.309. For *in vitro* experiments, the unaffected by NIR light part of the surrounding room was used as a reference temperature of the environment.

**Dynamic light scattering (DLS) and zeta-potential measurements** were performed using Zetasizer Nano ZS device (Malvern Pananalytical, Malvern, Worcestershire, UK) at 25  $^{\circ}\text{C}$ . Each measurement was performed in triplicate. DLS analysis was performed in backscatter mode ( $173^{\circ}$ ), and the results are reported as intensity-based distributions.

**Atomic force microscopy (AFM)** measurements for  **$\text{C}_6\text{F}_5\text{-Pt-F127-NPs}$**  and  **$\text{NO}_2\text{-Pt-NPs}$**  were performed in air using a NTEGRA II microscope (NT-MDT SI, Russia) in semi-contact mode with silicon cantilevers (1.74 N/m, 90 kHz). AFM measurements for  **$\text{C}_6\text{F}_5\text{-Pt-DSPE-NPs}$**  and  **$\text{NO}_2\text{-Est-NPs}$**  were performed in air using a NTEGRA Prima II microscope (NT-MDT SI, Russia) in semi-contact mode with HA-CNC cantilevers (1 N/m, 46kHz).

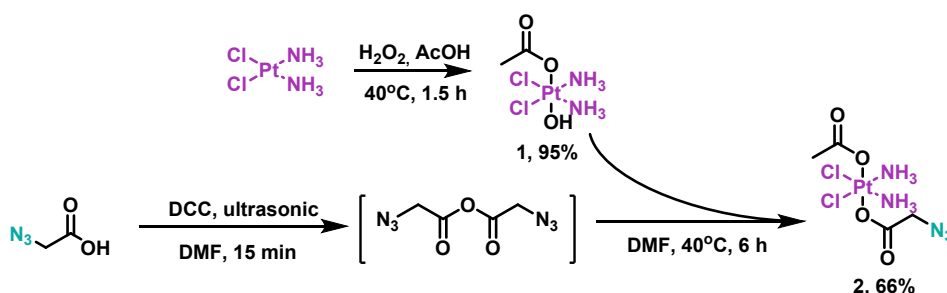
**Transmission electron microscopy (TEM).** The morphology of J-aggregates was studied using a JEOL JEM-1400 (120 kV) transmission electron microscope. 10  $\mu\text{L}$  of colloidal solution of J-aggregates was dropped onto the surface of a Formvar-coated copper grid, and the solvent was subsequently evaporated before analyzing at room temperature. No additional contrasts were used.  **$\text{C}_6\text{F}_5\text{-Pt}$**  (99%  $\text{H}_2\text{O}$ ) was used to study CT-coupled aggregates. To study slipped-stacked J-aggregates,  **$\text{C}_6\text{F}_5\text{-Pt}$**  (1  $\text{mg/ml}$  in acetone) was diluted to 30% acetone, incubated for 1 minute and then diluted with water prior to dropping on the copper grid.

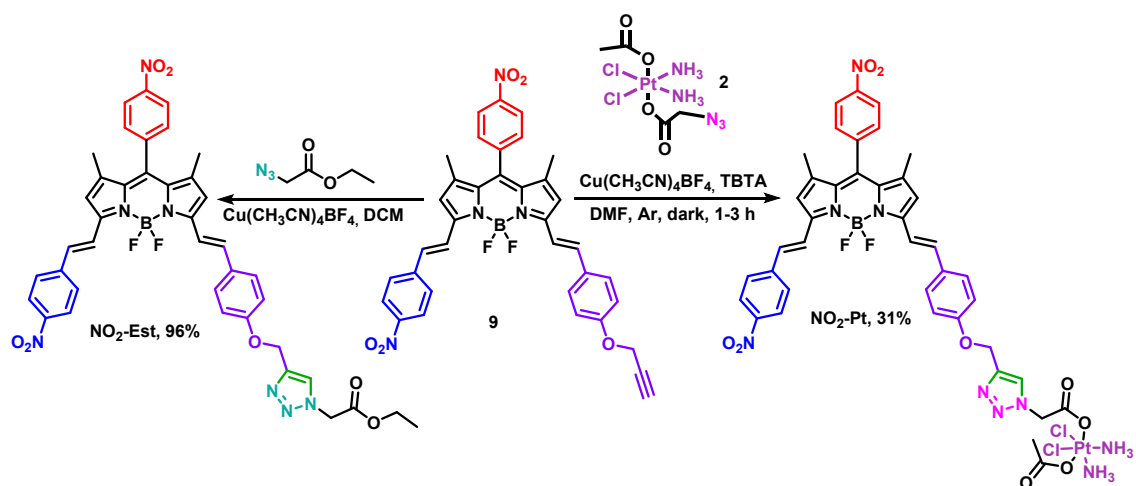
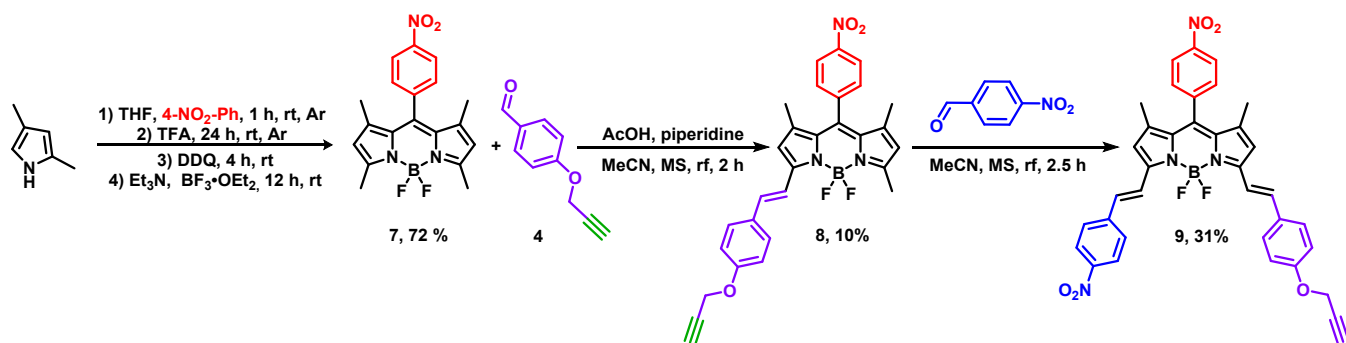
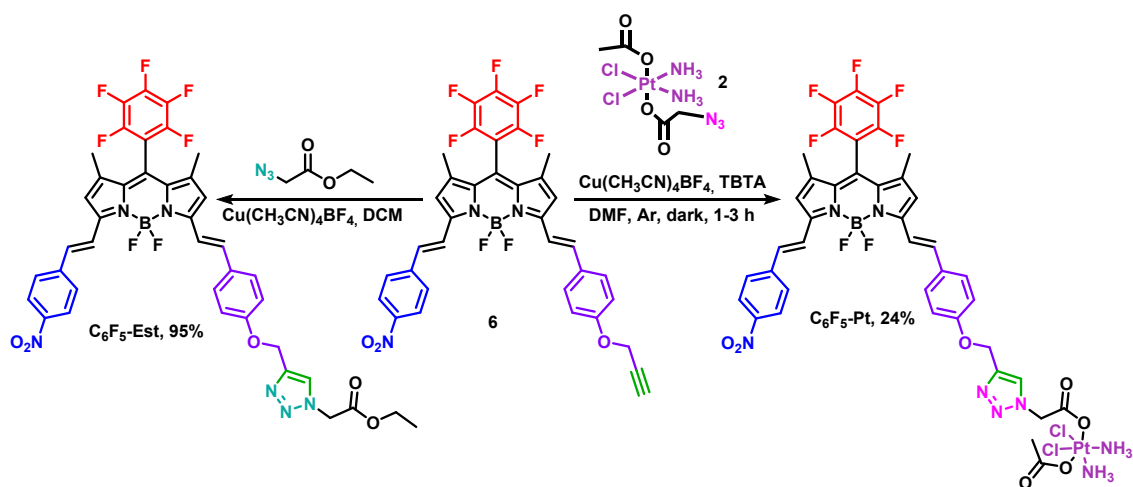
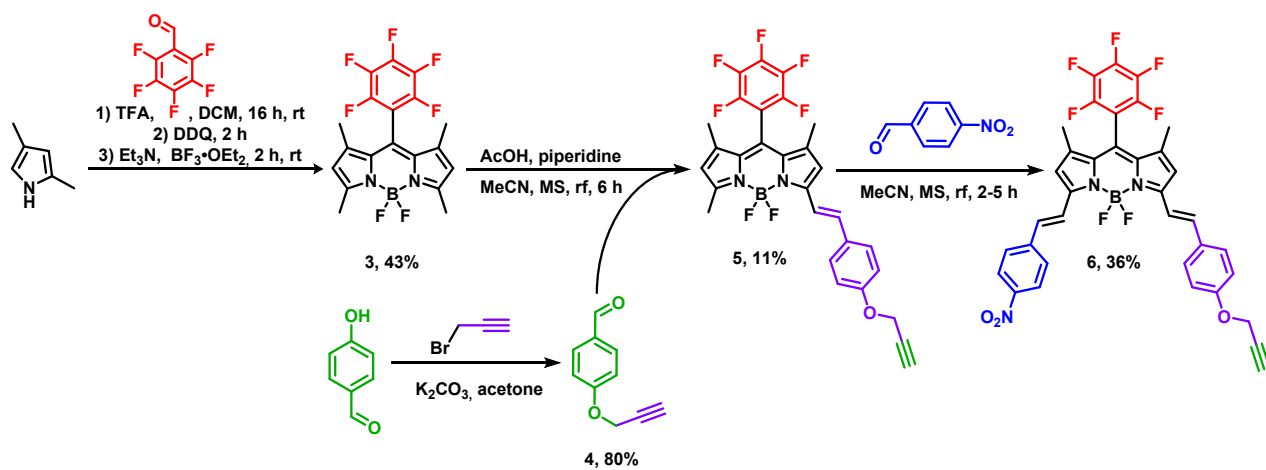
**Photoreduction studies of  $\text{NO}_2\text{-Pt}$  and  $\text{C}_6\text{F}_5\text{-Pt}$ .** All experiments were carried out at room temperature. Pt(IV) prodrugs  $\text{NO}_2\text{-Pt}$  and  $\text{C}_6\text{F}_5\text{-Pt}$  were dissolved in a mixture of  $\text{MeOH:DMSO:H}_2\text{O}$  (6:3:1, v/v/v) to the resulting concentration of 10-3 M. The 1 mM solutions of  $\text{NO}_2\text{-Pt}$  and  $\text{C}_6\text{F}_5\text{-Pt}$  in a transparent vial ( $d = 1\text{ cm}$ ) were placed in front of the light source ( $\lambda = 660\text{ nm}$ ) so that irradiation power density was equal to 13.5  $\text{mW/cm}^2$ .

In all experiments a 30 ml aliquot of the Pt(IV) prodrugs solution was taken and adjusted with MeOH to 500 ml at different time points (0, 2, 5, 10, 20, 40, 60 min). HPLC-MS-analysis of the probes was performed and repeated two times each. An amount of cisplatin-adduct in the probes was calculated from the peak area on the chromatogram. Changes in the levels of the compound of interest within the experiment were investigated by integrating the peaks of the  $\text{Pt}(\text{NH}_3)_2\text{Cl}(\text{C}_2\text{H}_6\text{SO})$  ion in the range  $m/z$  340.5-345.5 Da.

## 2. Synthesis and characterization of monomers

**Scheme S1.** Synthesis of  $\text{C}_6\text{F}_5\text{-Pt}$ ,  $\text{C}_6\text{F}_5\text{-Est}$ ,  $\text{NO}_2\text{-Pt}$  and  $\text{NO}_2\text{-Est}$ .





**[OC-6-44]-Acetatodiamminedichloridohydroxidoplatinum(IV) 1** was synthesized according to previous reports.<sup>2</sup> 100 mg (0.334 mmol) of cisplatin was dissolved in 40 ml of glacial acetic acid and 1.73 ml of H<sub>2</sub>O<sub>2</sub> (30% w/w) was added. The reaction mixture was stirred for 2 hours at 35–40°C until the precipitate dissolved and the solution became transparent. Then, the solution was concentrated under reduced pressure to 3–4 ml. The excess (30 ml) diethyl ether was added and the resulting suspension was centrifuged. The precipitate was separated and washed with diethyl ether, then it was left to dry on air. Compound **1** was obtained as a white powder. Yield: 119 mg (95%).

<sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>, δ, ppm.): 6.13 – 5.78 (m, 6H, NH<sub>3</sub>), 1.87 (s, 3H, CH<sub>3</sub>)

**[Pt(OAc)(2-azidoacetate)(Cl<sub>2</sub>(NH<sub>3</sub>)<sub>2</sub>)] 2** was synthesized according to previous reports.<sup>2</sup> 266 mg (1.29 mmol, 4 equiv.) of DCC was dissolved in 6 ml of DMF and 130 mg (1.29 mmol, 4 equiv.) of 2-azidoacetic acid was added. The reaction mixture was suspended in an ultrasonic bath for 15 min. Then, the formed precipitate was separated by centrifugation. The resulting solution of 2-azidoacetic anhydride in DMF was mixed with suspension of 121 mg (0.32 mmol, 1 equiv.) of compound **1** in 4.5 ml of DMF. The reaction mixture was stirred at r.t. overnight and then at 40°C for 6 hours. After that, the solution was removed under reduced pressure and the crude product was suspended in 0.5 ml of MeOH and precipitated by addition of diethyl ether. The precipitate was separated by centrifugation and was purified by flash chromatography using CH<sub>2</sub>Cl<sub>2</sub>:MeOH = 10:1 as eluent. Compound **2** was obtained as a light yellow powder. Yield: 98 mg (66%).

<sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>, δ, ppm.): 6.88 – 6.20 (m, 6H, NH<sub>3</sub>), 3.89 (s, 2H, C(O)CH<sub>2</sub>N<sub>3</sub>), 1.92 (s, 3H, C(O)-CH<sub>3</sub>).

**5,5-difluoro-1,3,7,9-tetramethyl-10-(perfluorophenyl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 3.** BODIPY **3** was synthesized according to previous reports<sup>3</sup> with slight modifications. Pentafluorobenzaldehyde (429 mg, 2.19 mmol) and 2,4-dimethyl-1H-pyrrole (500 μl, 4.86 mmol) were dissolved in DCM (172 ml) under argon atmosphere, then TFA (18 μl) was added and the mixture was stirred at room temperature (r.t.). After 4 h stirring 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) (1192 mg, 5.25 mmol) was added. The reaction was allowed to proceed for 4 h at r.t. The reaction mixture was then cooled in an ice-water bath, Et<sub>3</sub>N (5.15 ml) and BF<sub>3</sub>·Et<sub>2</sub>O (5.15 ml)

were added dropwise. The resulting mixture was kept stirring at r.t. overnight, then washed with H<sub>2</sub>O and extracted with DCM. The combined organic layers were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure. The crude product was purified by silica gel column chromatography (DCM : Petroleum ether = 1:1). BODIPY **3** was obtained as a red powder. Yield: 390 mg (43 %).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 6.06 (s, 1H, 2,6-BP), 2.57 (s, 6H, 3,5-Me), 1.62 (s, 6H, 1,7-Me).

**4-(prop-2-yn-1-yloxy)benzaldehyde 4.** *p*-hydroxybenzaldehyde (10 g, 1 equiv.), propargyl bromide (14.03 ml, 2 equiv.) and K<sub>2</sub>CO<sub>3</sub> (22.62 g, 2 equiv.) were dissolved in 340 ml of acetone. The reaction mixture was washed with brine and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure. The crude product was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>). Aldehyde **4** was obtained as a white powder. Yield: 13.11 g (quantitative yield).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 9.91 (s, 1H, CHO), 7.88 – 7.84 (m, 2H, 2,6-Ph), 7.13 – 7.05 (m, 2H, 3,5-Ph), 4.78 (d, *J* = 2.4 Hz, 2H, CH<sub>2</sub>), 2.57 (t, *J* = 2.4 Hz, 1H, CH).

**(E)-5,5-difluoro-1,3,9-trimethyl-10-(perfluorophenyl)-7-(4-(prop-2-yn-1-yloxy)styryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 5.** To a stirred solution of BODIPY **3** (278 mg, 0.67 mmol) and aldehyde **4** (131 mg, 0.61 mmol) in 18 ml of MeCN 20 μl of piperidine, 10 μl of acetic acid and 0.7 g of 3 Å molecular sieves were added. The reaction mixture was then refluxed for 2 hours with the progress of the reaction monitored by thin-layer chromatography (TLC). Following this, the solvent was evaporated under reduced pressure, and the residue purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub> : Petroleum ether = 1:1). The BODIPY **5** was obtained as a red powder. Yield: 41 mg (11%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 7.61 – 7.51 (m, 3H, H=2, 5, 1), 7.26 (d, *J* = 16.4 Hz, 1H, H=1'), 7.00 (d, *J* = 8.7 Hz, 2H, H=3, 4), 6.67 (s, 1H, H=2-BP), 6.07 (s, 1H, H=6-BP), 4.74 (d, *J* = 2.4 Hz, 2H, H=CH<sub>2</sub>), 2.61 (s, 3H, H=3-Me), 2.55 (t, *J* = 2.4 Hz, 1H, H=CH), 1.67 (s, 3H, H=1-Me), 1.63 (m, 3H, H=7-Me).

HRMS: calc. for C<sub>29</sub>H<sub>21</sub>BF<sub>7</sub>N<sub>2</sub>O<sup>+</sup> (5+H)<sup>+</sup>, 557.1635; found C<sub>29</sub>H<sub>21</sub>BF<sub>7</sub>N<sub>2</sub>O<sup>+</sup> (5+H)<sup>+</sup>, 557.1622.

**5,5-difluoro-1,9-dimethyl-3-((E)-4-nitrostyryl)-10-(perfluorophenyl)-7-((E)-4-(prop-2-yn-1-yloxy)styryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 6.** To a stirred solution of BODIPY **5** (78 mg, 0.14 mmol) and 4-Nitrobenzaldehyde (21 mg, 0.14 mmol) in 13 ml of MeCN 20 μl of piperidine, 10 μl of acetic acid and 1 g of 3 Å molecular sieves were added. The reaction mixture was then refluxed for 3 hours with the progress of the reaction monitored by thin-layer chromatography (TLC). Following this, the solvent was evaporated under reduced pressure, and the residue purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub> : Petroleum ether = 2:1). The BODIPY **6** was obtained as a dark blue powder. Yield: 35 mg (36 %).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.26 (d, *J* = 8.8 Hz, 2H, H=3, 4), 7.84 (d, *J* = 16.3 Hz, 1H, H=1'), 7.74 (d, *J* = 8.7 Hz, 2H, H=2, 5), 7.67 – 7.59 (m, 3H, H=2', 5', 1), 7.36 (d, *J* = 16.2 Hz, 1H, H=1a), 7.25 (d, *J* = 16.0 Hz, 1H, H=1a'), 7.05 (d, *J* = 8.6 Hz, 2H, H=3', 4'), 6.77 (s, 1H, H=6-BP), 6.72 (s, 1H, H=2-BP), 2.57 (t, *J* = 2.4 Hz, 1H, H=CH), 1.71 (s, 3H, H=7-Me), 1.70 (s, 3H, H=1-Me).

HRMS: calc. for C<sub>36</sub>H<sub>23</sub>BF<sub>7</sub>N<sub>3</sub>NaO<sub>3</sub><sup>+</sup> (6+Na)<sup>+</sup> 712.1618; found C<sub>36</sub>H<sub>23</sub>BF<sub>7</sub>N<sub>3</sub>NaO<sub>3</sub><sup>+</sup> (6+Na)<sup>+</sup> 712.1622.

**ethyl 2-(4-((E)-2-(5,5-difluoro-1,9-dimethyl-7-((E)-4-nitrostyryl)-10-(perfluorophenyl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinin-3-yl)vinyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)acetate C<sub>6</sub>F<sub>5</sub>-Est.** BODIPY **6** (23.4 mg, 0.034 mmol) and ethyl 2-azidoacetate (8.8 mg, 0.068 mmol) were dissolved in 3 ml of CH<sub>2</sub>Cl<sub>2</sub> and Cu(CH<sub>3</sub>CN)<sub>4</sub>·BF<sub>4</sub> (5.3 mg, 0.017 mmol) was added under argon atmosphere. The reaction mixture was stirred for 2 hours at r.t. The solvent was evaporated under reduced pressure, and the residue was purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>). C<sub>6</sub>F<sub>5</sub>-Est was obtained as a dark blue powder. Yield: 27 mg (96%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.26 (d, *J* = 8.7 Hz, 2H, H=3, 4), 7.84 (d, *J* = 16.5 Hz, 1H, H=1'), 7.80 (s, 1H, H=CH), 7.74 (d, *J* = 8.7 Hz, 2H, H=2, 5), 7.65 – 7.57 (m, 3H, H=2', 5', 1), 7.35 (d, *J* = 16.2 Hz, 1H, H=1a), 7.26 (d, 1H, H=1a'), 7.06 (d, *J* = 8.7 Hz, 2H, H=3', 4'), 6.76 (s, 1H, H=6-BP), 6.72 (s, 1H, H=2-BP), 5.32 (s, 2H, H=N-CH<sub>2</sub>), 5.19 (s, 2H, H=CH<sub>2</sub>), 4.29 (q, *J* = 7.2 Hz, 2H, H=O-CH<sub>2</sub>), 1.71 (s, 3H, H=7-Me), 1.69 (s, H=1-Me), 1.31 (t, *J* = 7.1 Hz, 3H, H=CH<sub>3</sub>).  
HRMS: calc. for C<sub>40</sub>H<sub>31</sub>BF<sub>7</sub>N<sub>6</sub>O<sub>5</sub><sup>+</sup> (C<sub>6</sub>F<sub>5</sub>-Est +H)<sup>+</sup> 819.2337; found C<sub>40</sub>H<sub>31</sub>BF<sub>7</sub>N<sub>6</sub>O<sub>5</sub><sup>+</sup> (C<sub>6</sub>F<sub>5</sub>-Est +H)<sup>+</sup>, 819.2330.

**C<sub>6</sub>F<sub>5</sub>-Pt. BODIPY 6** (35 mg, 0.05 mmol) was dissolved in 1.1 ml of DMF and Cu(CH<sub>3</sub>CN)<sub>4</sub>•BF<sub>4</sub> (3.9 mg, 12 μmol), TBTA (8 mg, 15 μmol) were added under argon atmosphere. The reaction mixture was stirred for 15 min at r.t., complex **2** (19 mg, 0.041 mmol) was added and the solution was stirred for another 2 hour at r.t. The solvent was evaporated under reduced pressure. The residue was purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub>:MeOH = 10:1). Complex **C<sub>6</sub>F<sub>5</sub>-Pt** was obtained as a dark green powder. Yield: 11 mg (24%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.31 (d, *J* = 8.8 Hz, 2H, H=3, 4), 8.16 (s, 1H, H=CH), 7.88 (d, *J* = 8.7 Hz, 2H, H=2, 5), 7.81 (d, *J* = 15.9 Hz, 1H, H=1'), 7.74 (d, *J* = 16.2 Hz, 1H, H=1), 7.71 – 7.63 (m, 3H, H=1a, 2', 5'), 7.44 (d, *J* = 16.2 Hz, 1H, H=1a'), 7.24 – 7.17 (m, 3H, H=3', 4', 6-BP), 7.12 (s, 1H, 2-BP), 6.52 (br.s, 6H, H=NH<sub>3</sub>), 5.26 (s, 2H, H=N-CH<sub>2</sub>), 5.24 (s, 2H, H=CH<sub>2</sub>), 1.92 (s, 3H, H=CH<sub>3</sub>), 1.74 (s, 3H, H=7-Me), 1.72 (s, 3H, H=1-Me).  
HRMS: calc. for C<sub>40</sub>H<sub>35</sub>BCl<sub>2</sub>F<sub>7</sub>N<sub>8</sub>O<sub>7</sub>Pt<sup>+</sup> (C<sub>6</sub>F<sub>5</sub>-Pt+H)<sup>+</sup>, 1149.1584; found C<sub>40</sub>H<sub>35</sub>BCl<sub>2</sub>F<sub>7</sub>N<sub>8</sub>O<sub>7</sub>Pt<sup>+</sup> (C<sub>6</sub>F<sub>5</sub>-Pt+H)<sup>+</sup>, 1149.1583.

**5,5-difluoro-1,3,7,9-tetramethyl-10-(4-nitrophenyl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 7.** BODIPY **7** was synthesized according to previous reports<sup>4</sup> with slight modifications. 4-Nitrobenzaldehyde (187 mg, 1.24 mmol) and 2,4-dimethyl-1H-pyrrole (281 μl, 2.74 mmol) were dissolved in THF (37 ml) under argon atmosphere, after 1 h of stirring TFA (20 μl) was added and the mixture was stirred at r.t. After 24 h stirring DDQ (364 mg, 1.6 mmol) in dry THF (41 ml) was added drop by drop. The reaction was allowed to proceed for 5 h at r.t. The reaction mixture was then cooled in an ice-water bath, Et<sub>3</sub>N (7.4 ml) and BF<sub>3</sub>•Et<sub>2</sub>O (7.4 ml) were added dropwise. The resulting mixture was kept stirring at r.t. overnight, then it was filtered. The solid obtained was washed with THF, acetone and DCM. The organic phases were concentrated under reduced pressure. Then the residue was dissolved in DCM, washed with saturated NaHCO<sub>3</sub> and water. The organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated under reduced pressure. The crude product was purified by silica gel column chromatography (DCM : Petroleum ether = 1:1). BODIPY **7** was obtained as an orange-red powder. Yield: 318 mg (70 %).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.40 (m, 2H, H=2,6-Ph), 7.55 (m, 2H, H=3,5-Ph), 6.02 (s, 2H, H=2,6-BP), 2.58 (s, 6H, H=3,5-Me), 1.37 (s, 6H, H=1,7-Me).

**(E)-5,5-difluoro-1,3,9-trimethyl-10-(4-nitrophenyl)-7-(4-(prop-2-yn-1-yloxy)styryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 8.** To a stirred solution of BODIPY **7** (343 mg, 0.93 mmol) and aldehyde **4** (134 mg, 0.84 mmol) in 29 ml of MeCN 20 μl of piperidine, 10 μl of acetic acid and 0.81 g of 3 Å molecular sieves were added. The reaction mixture was then refluxed for 3 hours with the progress of the reaction monitored by thin-layer chromatography (TLC). Following this, the solvent was evaporated under reduced pressure, and the residue purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub> : Petroleum ether = 1:1). The BODIPY **8** was obtained as a violet powder. Yield: 47 mg (10%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.39 (d, *J* = 8.5 Hz, 2H, H=2,6-Ph), 7.56 (m, 5H, H=3,5-Ph, 2, 5, 1), 7.23 (d, *J* = 17.1 Hz, 1H, H=1'), 6.99 (d, *J* = 8.6 Hz, 2H, H=3, 4), 6.62 (s, 1H, H=6-BP), 6.03 (s, 1H, H=2-BP), 4.74 (d, *J* = 2.4 Hz, 2H, H=CH<sub>2</sub>), 2.60 (s, 3H, H=5-Me), 2.55 (t, *J* = 2.2 Hz, 1H, H=CH), 1.41 (s, 3H, H=1-Me), 1.37 (s, 3H, H=7-Me).

**5,5-difluoro-1,9-dimethyl-10-(4-nitrophenyl)-3-((E)-4-nitrostyryl)-7-((E)-4-(prop-2-yn-1-yloxy)styryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 9.** To a stirred solution of BODIPY **8** (53 mg, 0.1 mmol) and 4-Nitrobenzaldehyde (16 mg, 0.1 mmol) in 8 ml of MeCN 20 μl of piperidine, 10 μl of acetic acid and 0.74 g of 3 Å molecular sieves were added. The reaction mixture was then refluxed for 3 hours with the progress of the reaction monitored by thin-layer chromatography (TLC). Following this, the solvent was evaporated under reduced pressure, and the residue purified by silica gel column chromatography (Acetone : Petroleum ether = 1:3). The BODIPY **9** was obtained as a dark blue powder. Yield: 21 mg (31 %).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.42 (d, *J* = 8.3 Hz, 2H, H=2,6-Ph), 8.26 (d, *J* = 8.4 Hz, 2H, H=3,4), 7.85 (d, *J* = 16.4 Hz, 1H, H=1'), 7.74 (d, *J* = 8.6 Hz, 2H, H=3,5-Ph), 7.62 (m, 5H, H=2,5, 2', 5', 1), 7.33 (d, *J* = 16.1 Hz, 1H, H=1a), 7.21 (s, 1H, H=1a'), 7.05 (d, *J* = 8.5 Hz, 2H, H=3',4'), 6.72 (s, 1H, H=6-BP), 6.68 (s, 1H, H=2-BP), 4.77 (d, *J* = 2.5 Hz, 2H, H=CH<sub>2</sub>), 2.60 – 2.55 (m, 1H, H=CH), 1.46 (s, 3H, H=7-Me), 1.44 (s, 3H, H=1-Me).

HRMS: calc. for C<sub>36</sub>H<sub>27</sub>BF<sub>2</sub>N<sub>4</sub>NaO<sub>5</sub><sup>+</sup> (9+Na)<sup>+</sup> 667.1940; found C<sub>36</sub>H<sub>27</sub>BF<sub>2</sub>N<sub>4</sub>NaO<sub>5</sub><sup>+</sup> (9+Na)<sup>+</sup> 667.1946.

**ethyl 2-(4-(((E)-2-(5,5-difluoro-1,9-dimethyl-10-(4-nitrophenyl)-7-((E)-4-nitrostyryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinin-3-yl)vinyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)acetate NO<sub>2</sub>-Est.** BODIPY **9** (26.4 mg, 0.041 mmol) and ethyl 2-azidoacetate (10.6 mg, 0.082 mmol) were dissolved in 4.8 ml of CH<sub>2</sub>Cl<sub>2</sub> and Cu(CH<sub>3</sub>CN)<sub>4</sub>•BF<sub>4</sub> (6.4 mg, 0.021 mmol) was added under argon atmosphere. The reaction mixture was stirred for 2 hours at r.t. The solvent was evaporated under reduced pressure, and the residue was purified by silica gel column chromatography (CH<sub>2</sub>Cl<sub>2</sub>). **NO<sub>2</sub>-Est** was obtained as a dark blue powder. Yield: 30 mg (95%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.42 (d, *J* = 8.6 Hz, 2H, 2,6-Ph), 8.26 (d, *J* = 8.7 Hz, 2H, H=3,4), 7.85 (d, *J* = 16.2 Hz, 1H, H=1'), 7.81 (s, 1H, H=CH), 7.74 (d, *J* = 8.7 Hz, 2H, H=3,5-Ph), 7.61 (m, *J* = 8.8, 3.4 Hz, 5H, H=2,5, 2', 5', 1), 7.32 (d, *J* = 16.2 Hz, 1H, H=1a), 7.23 (d, *J* = 16.5 Hz, 1H, H=1a'), 7.06 (d, *J* = 8.7 Hz, 2H, H=3',4'), 6.71 (s, 1H, H=6-BP), 6.68 (s, 1H, H=2-BP), 4.69 (s, 4H, H=CH<sub>2</sub>, N-CH<sub>2</sub>), 4.29 (q, *J* = 7.2 Hz, 2H, H=O-CH<sub>2</sub>), 1.45 (s, 3H, H=7-Me), 1.44 (s, 3H, H=1-Me), 1.31 (t, *J* = 7.2 Hz, 3H, H=CH<sub>3</sub>).

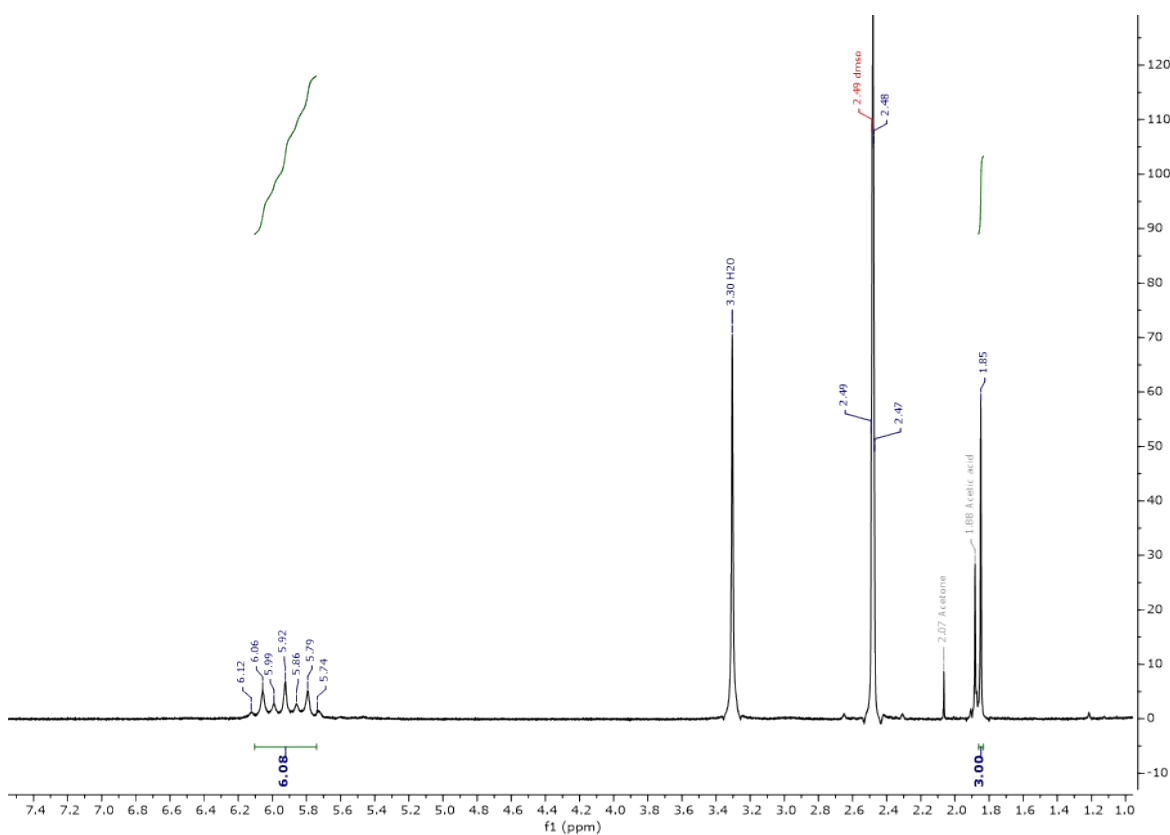
**NO<sub>2</sub>-Pt.** BODIPY **6** (42 mg, 0.065 mmol) was dissolved in 3 ml of DMF and Cu(CH<sub>3</sub>CN)<sub>4</sub>•BF<sub>4</sub> (6.1 mg, 19.6 μmol), TBTA (10.4 mg, 19.6 μmol) were added under argon atmosphere. The reaction mixture was stirred for 15 min at r.t., complex **2** (30 mg, 0.065 mmol) was added and the solution was stirred for another 2 hour at r.t. The solvent was evaporated under reduced pressure. The residue was purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub>:MeOH = 10:1). Complex **NO<sub>2</sub>-Pt** was obtained as a dark green powder. Yield: 22 mg (31%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ, ppm): 8.43 (d, *J* = 8.5 Hz, 2H, 2,6-Ph), 8.31 (d, *J* = 8.6 Hz, 2H, H=3,4), 8.16 (s, 1H, H=CH), 7.85 (m, 4H, H=1, 1', 3,5-Ph), 7.75 – 7.62 (m, 5H, H=2, 5, 2', 5', 1a), 7.45 (d, *J* = 16.2 Hz, 1H, H=1a'), 7.19 (d, *J* = 8.4 Hz, 2H, H=3',4'), 7.11 (s, 1H, H=6-BP), 7.04 (s, 1H, H=2-BP), 6.52 (br. s, 6H, H=NH<sub>3</sub>), 5.25 (s, 2H, N-CH<sub>2</sub>), 5.24 (s, 2H, H=CH<sub>2</sub>), 1.92 (s, 3H, H=CH<sub>3</sub>), 1.45 (s, 3H, H=7-Me), 1.42 (s, 3H, H=1-Me).

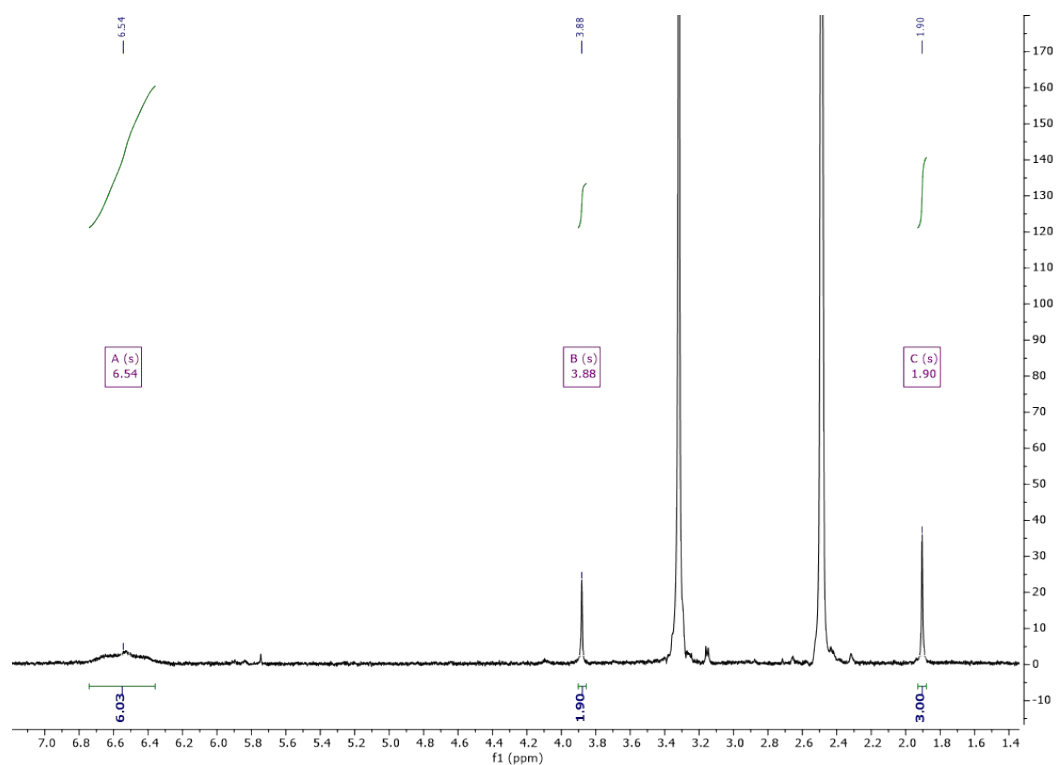
<sup>13</sup>C NMR (101 MHz, DMSO-d<sub>6</sub>, δ, ppm): 78.13, 173.32, 159.78, 155.45, 149.34, 148.14, 146.88, 143.42, 142.76, 142.11, 140.83, 140.06, 139.55, 136.28, 133.24, 133.13, 130.51, 129.38, 127.90, 127.74, 126.03, 124.42, 124.35, 122.11, 119.76, 118.81, 115.54, 61.31, 50.57, 22.66, 14.79, 14.47.

HRMS: calc. for C<sub>40</sub>H<sub>38</sub>BCl<sub>2</sub>F<sub>2</sub>N<sub>9</sub>NaO<sub>9</sub>Pt<sup>+</sup> (**NO<sub>2</sub>-Pt+Na**)<sup>+</sup> 1126.1778; found C<sub>40</sub>H<sub>38</sub>BCl<sub>2</sub>F<sub>2</sub>N<sub>9</sub>NaO<sub>9</sub>Pt<sup>+</sup> (**NO<sub>2</sub>-Pt+Na**)<sup>+</sup> 1126.1773.

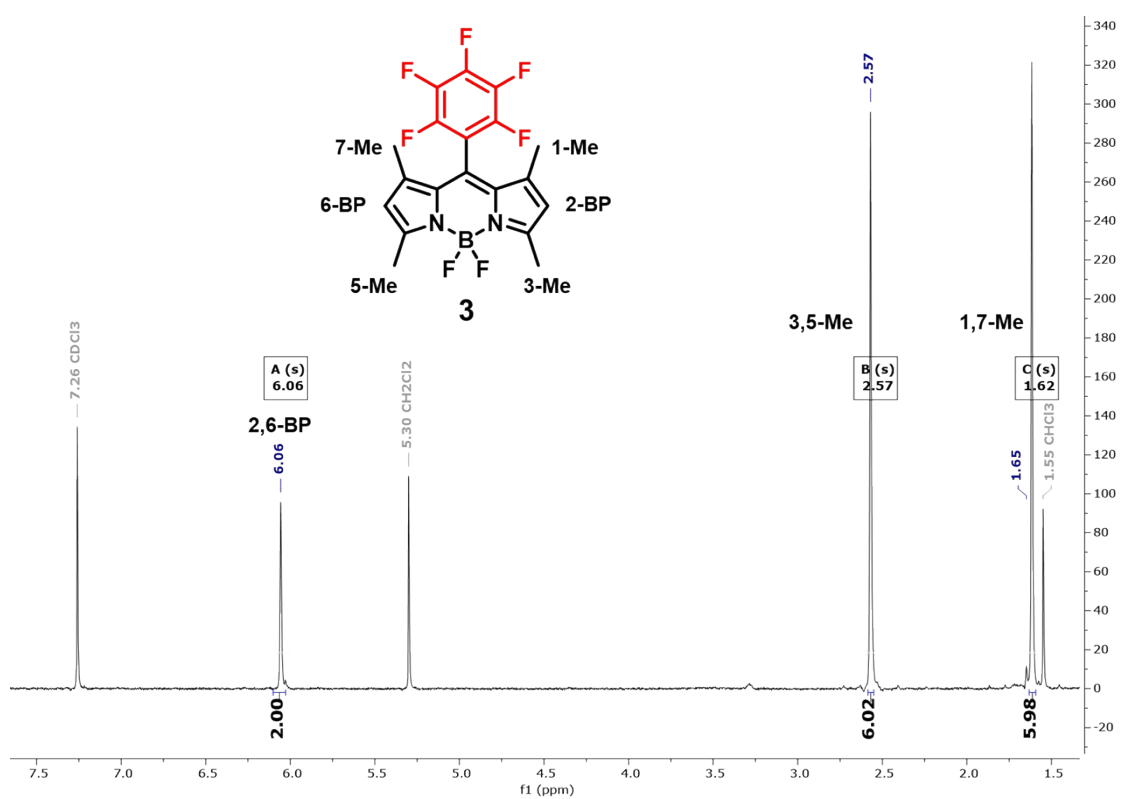
## 2. Structural Characterization and Purity Analysis (NMR, HRMS and RP-HPLC analysis)



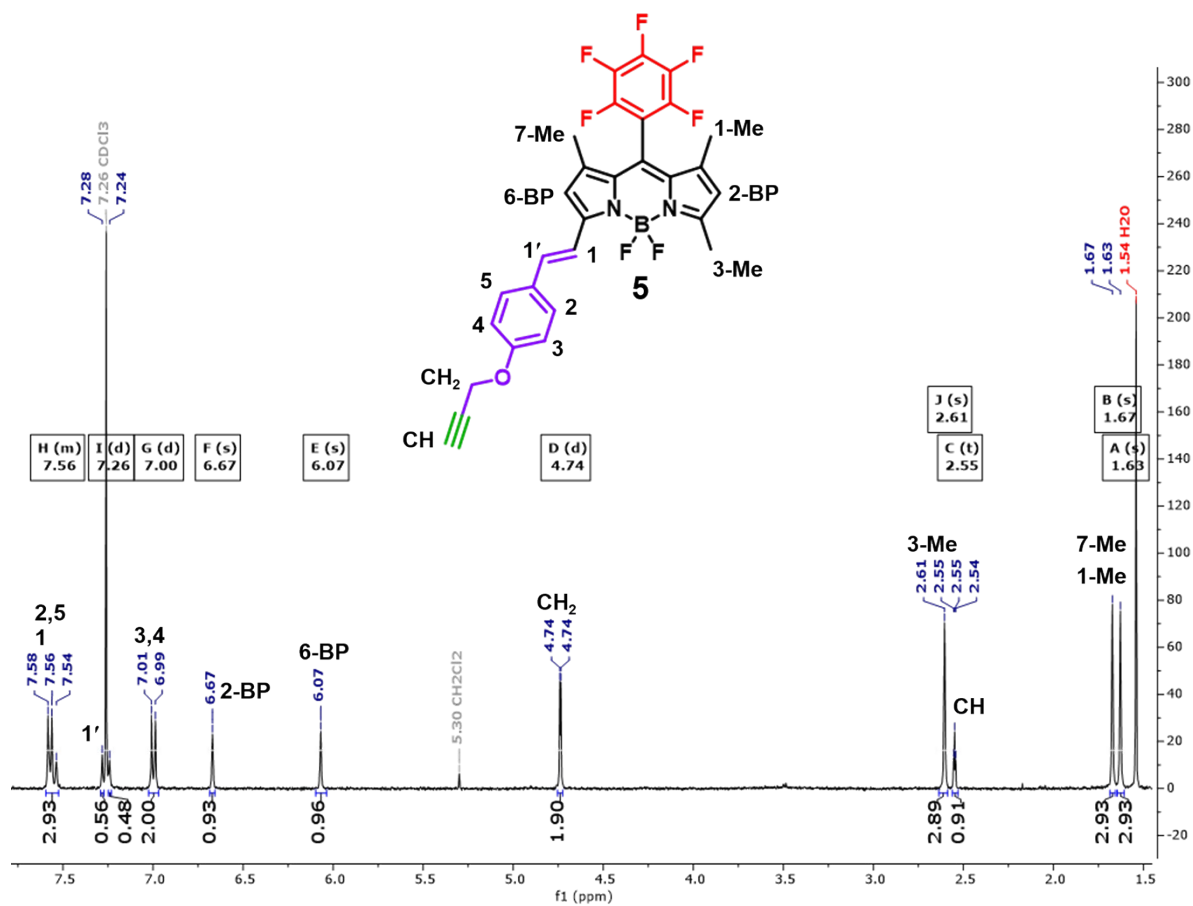
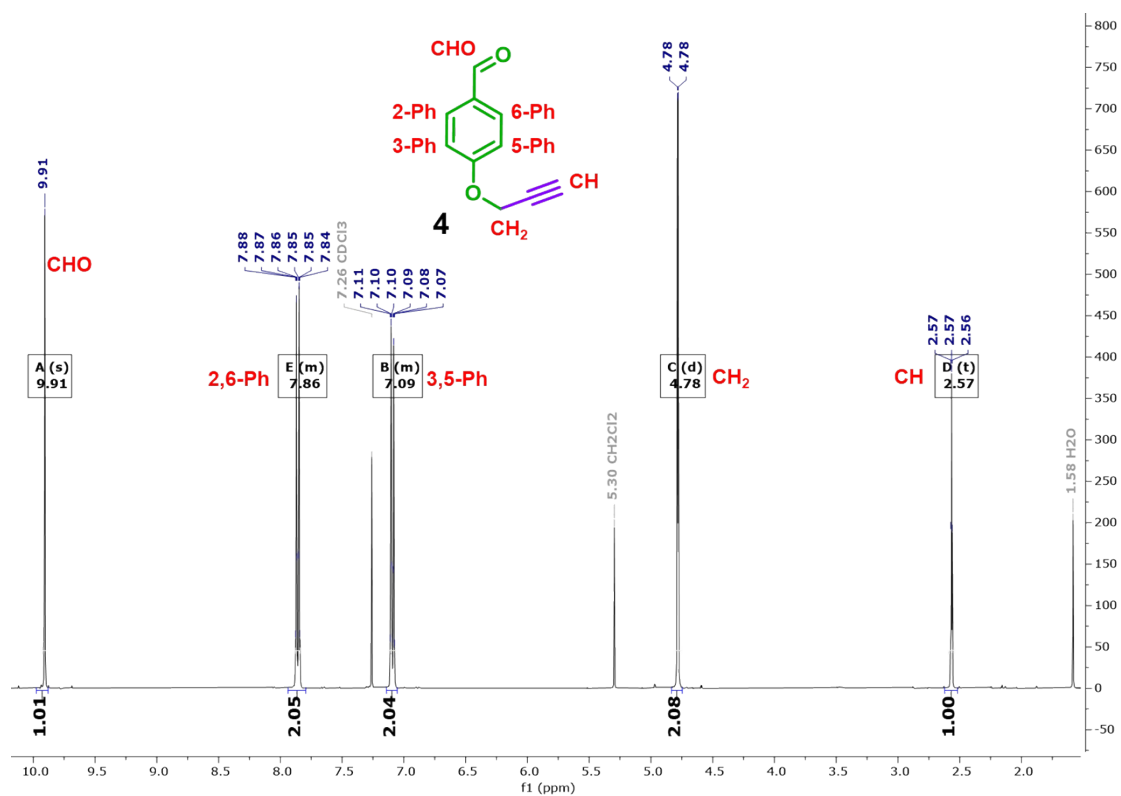
**Figure S1.** <sup>1</sup>H NMR-spectrum of [OC-6-44]-Acetatodiamminedichloridohydroxidoplatinum(IV) **1** in DMSO-d<sub>6</sub>.

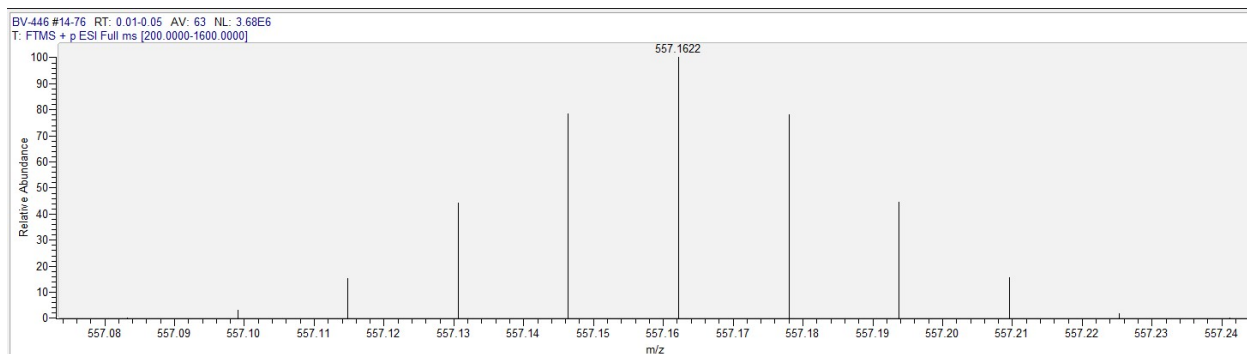


**Figure S2.**  $^1\text{H}$  NMR-spectrum of  $[\text{Pt}(\text{OAc})(2\text{-azidoacetate})(\text{Cl}_2(\text{NH}_3)_2)]$  **2** in  $\text{DMSO-d}_6$ .

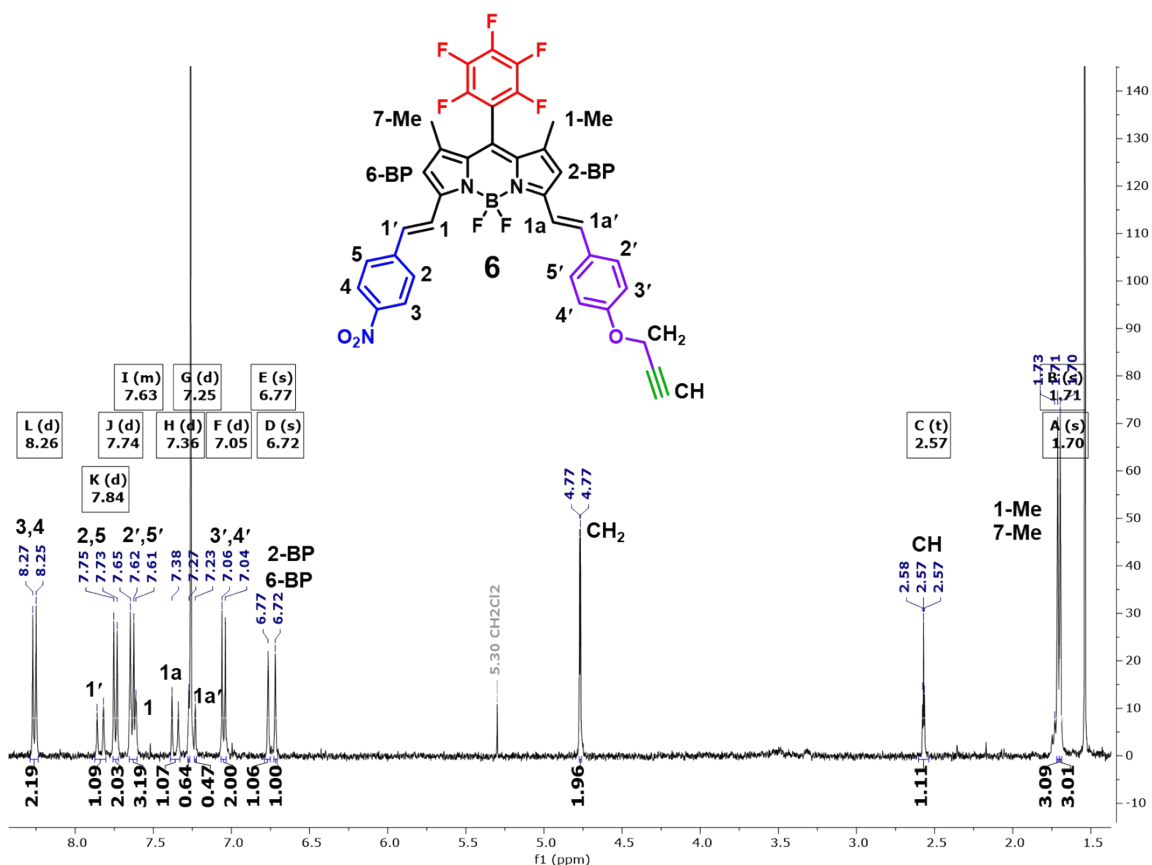


**Figure S3.**  $^1\text{H}$  NMR-spectrum of 5,5-difluoro-1,3,7,9-tetramethyl-10-(perfluorophenyl)-5H-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-c:2',1'- $\beta$ ][1,3,2]diazaborinine **3** in  $\text{CDCl}_3$ .

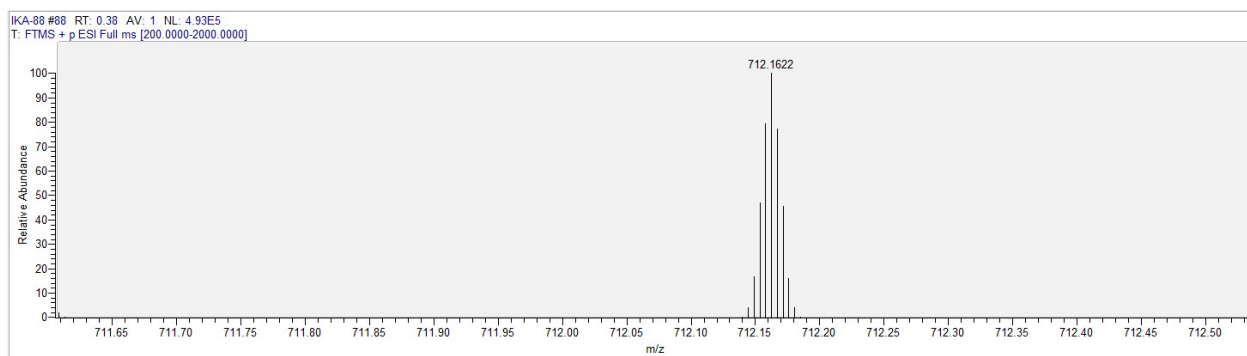




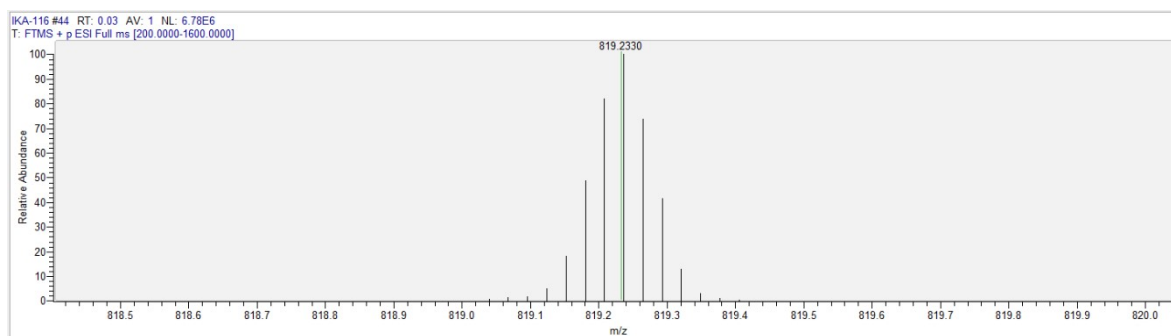
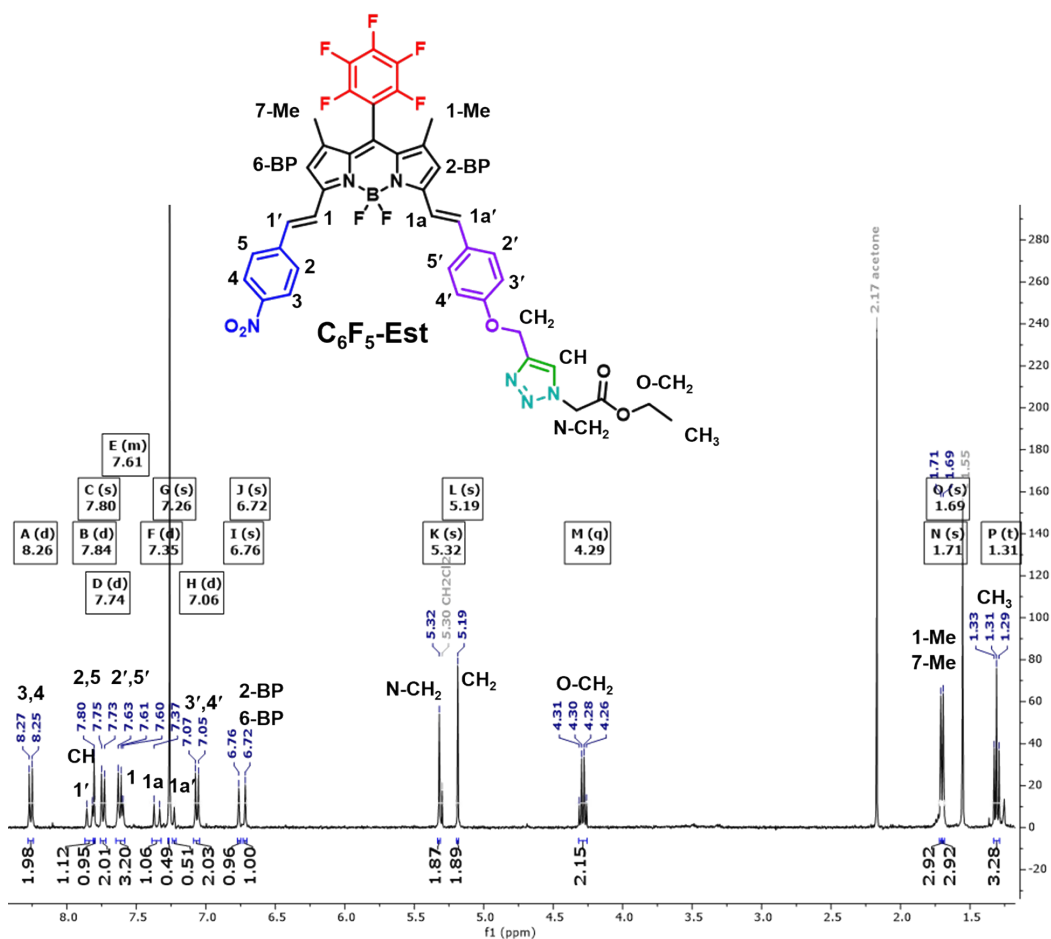
**Figure S8.** ESI-high resolution mass spectrum of (*E*)-5,5-difluoro-1,3,9-trimethyl-10-(perfluorophenyl)-7-(4-(prop-2-yn-1-yloxy)styryl)-5*H*-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinine 5.



**Figure S9.** <sup>1</sup>H NMR-spectrum of 5,5-difluoro-1,9-dimethyl-3-((*E*)-4-nitrostyryl)-10-(perfluorophenyl)-7-((*E*)-4-(prop-2-yn-1-yloxy)styryl)-5*H*-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinine 6 in CDCl<sub>3</sub>.



**Figure S10.** ESI-high resolution mass spectrum of 5,5-difluoro-1,9-dimethyl-3-((*E*)-4-nitrostyryl)-10-(perfluorophenyl)-7-((*E*)-4-(prop-2-yn-1-yloxy)styryl)-5*H*-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinine 6.



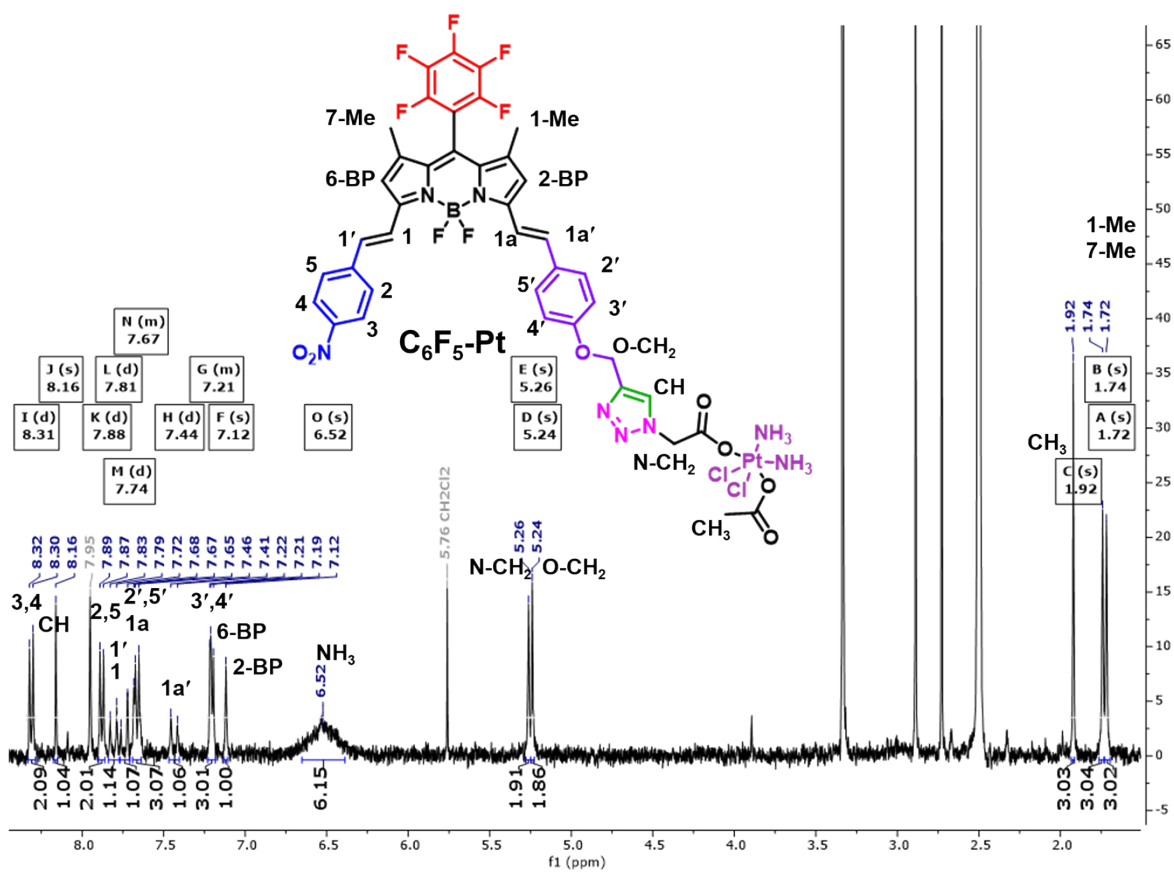


Figure S13.  $^1\text{H}$  NMR-spectrum of  $\text{C}_6\text{F}_5\text{-Pt}$  in  $\text{DMSO-d}_6$ .

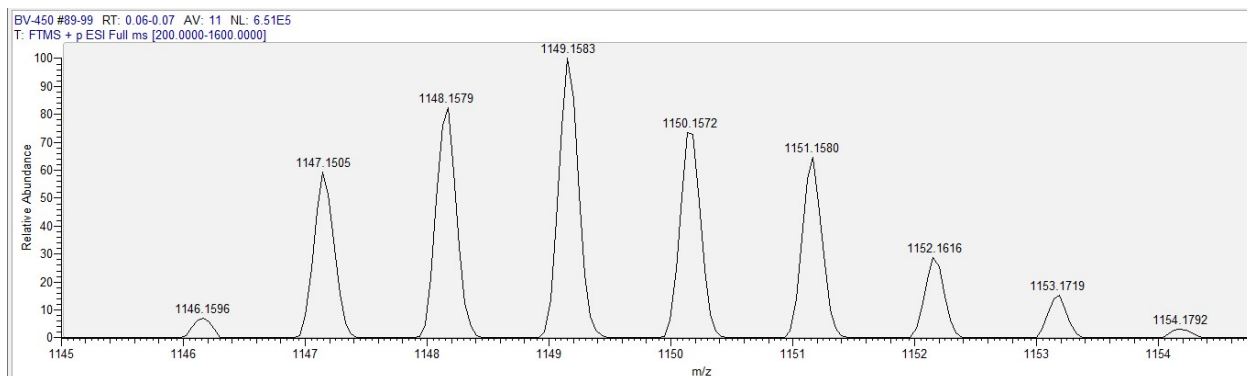


Figure S14. ESI-high resolution mass spectrum of  $\text{C}_6\text{F}_5\text{-Pt}$ .

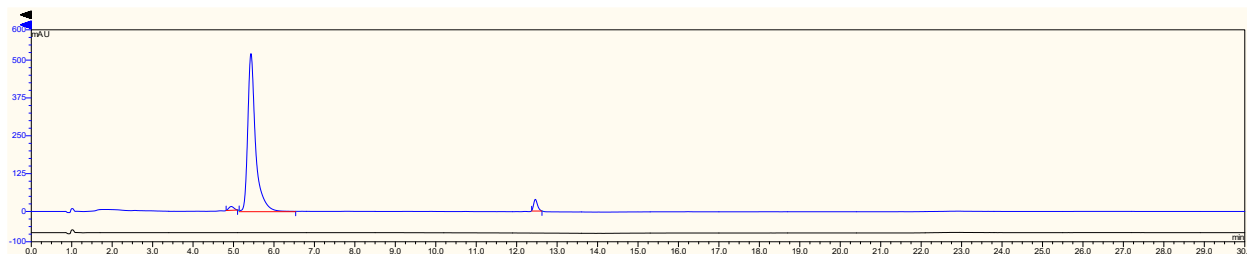


Figure S15. RP-HPLC analysis of  $\text{C}_6\text{F}_5\text{-Pt}$ . UV-VIS detector was set at 650 nm. Main peak area: 95.04%.

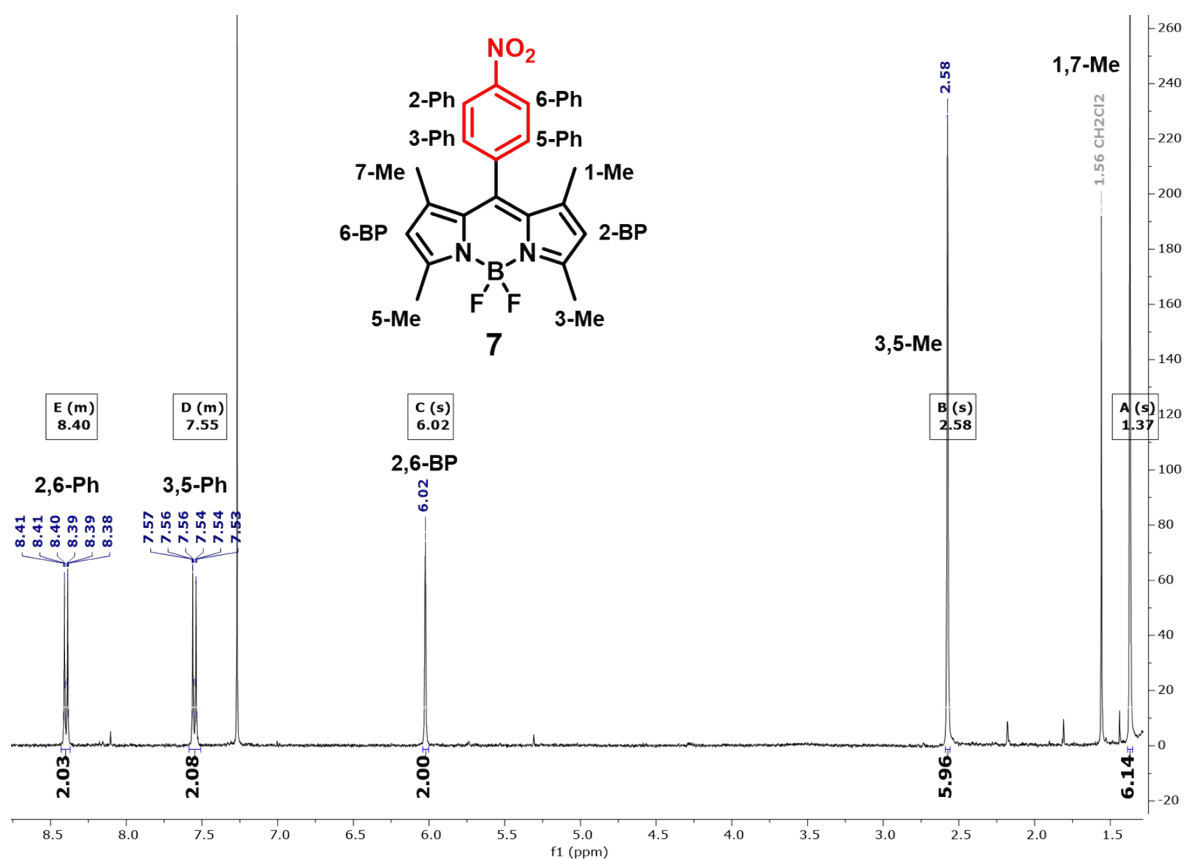


Figure S16. <sup>1</sup>H NMR-spectrum of 5,5-difluoro-1,3,9-trimethyl-10-(4-nitrophenyl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 7 in CDCl<sub>3</sub>.

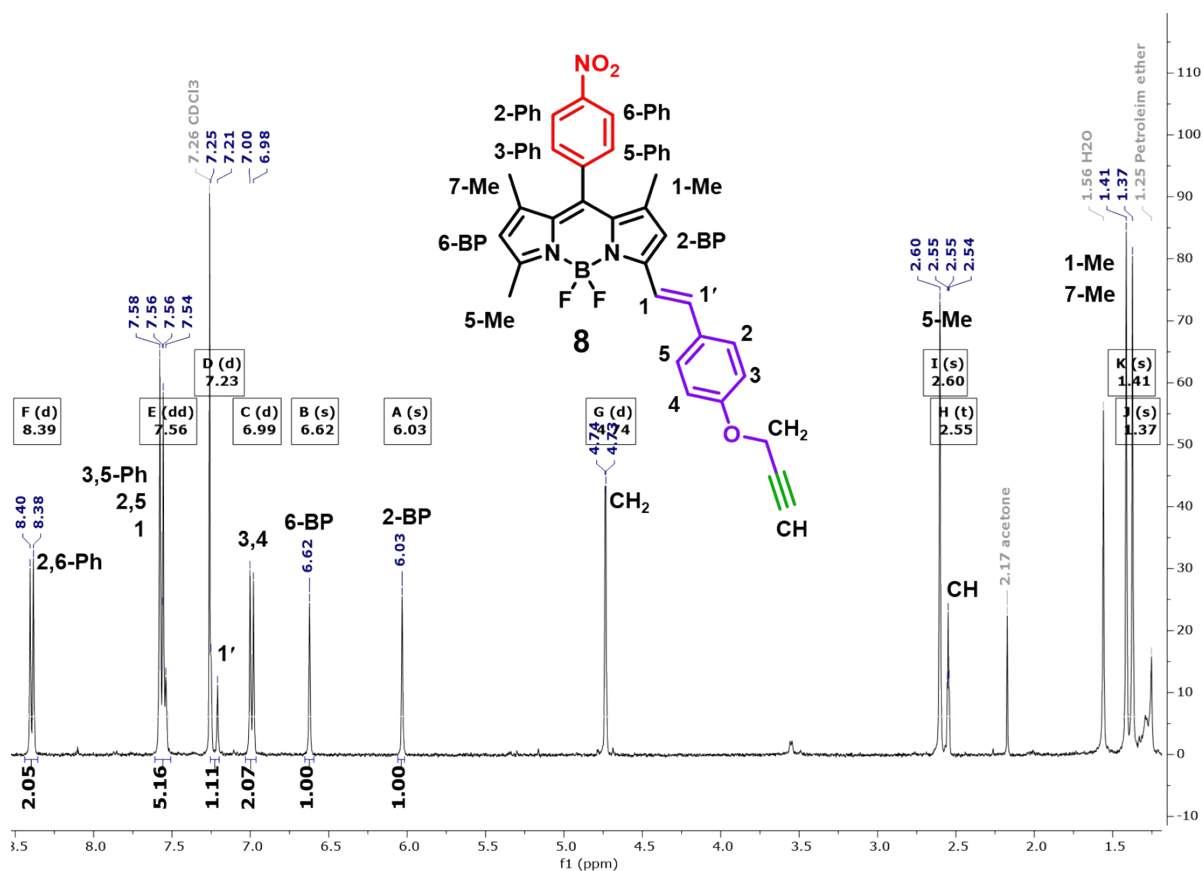
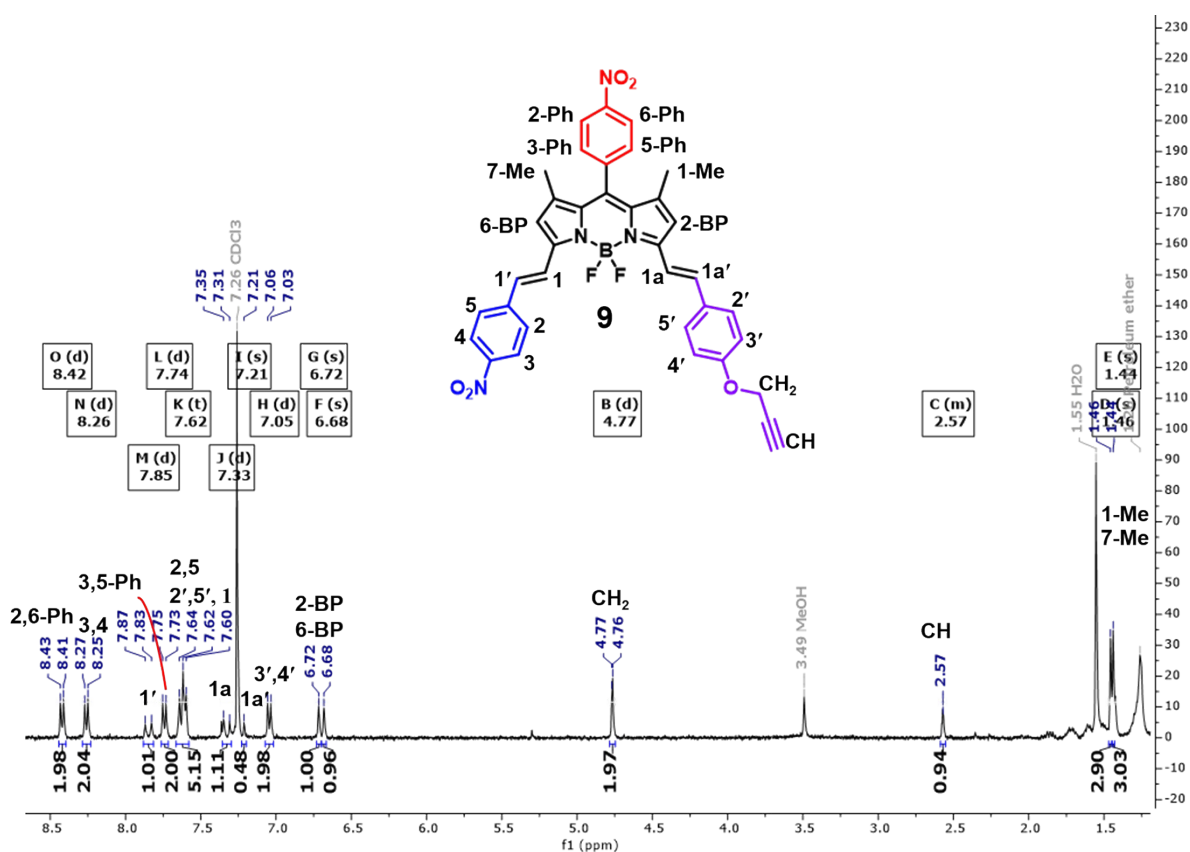
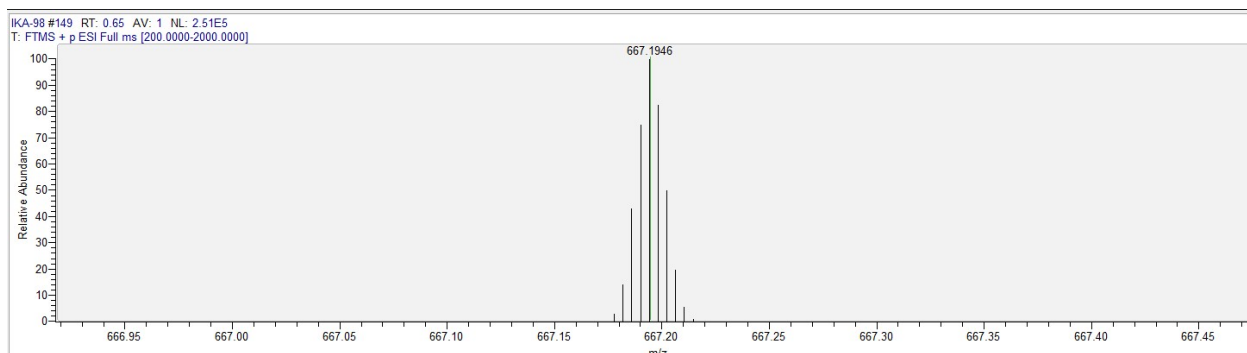


Figure S17. <sup>1</sup>H NMR-spectrum of (E)-5,5-difluoro-1,3,9-trimethyl-10-(4-nitrophenyl)-7-(4-(prop-2-yn-1-yloxy)styryl)-5H-4λ<sup>4</sup>,5λ<sup>4</sup>-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine 8 in CDCl<sub>3</sub>.



**Figure S18.**  $^1\text{H}$  NMR-spectrum of 5,5-difluoro-1,9-dimethyl-10-(4-nitrophenyl)-3-((*E*)-4-nitrostyryl)-7-((*E*)-4-(prop-2-yn-1-yloxy)styryl)-5*H*-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinine **9** in  $\text{CDCl}_3$ .



**Figure S19.** ESI-high resolution mass spectrum of 5,5-difluoro-1,9-dimethyl-10-(4-nitrophenyl)-3-((*E*)-4-nitrostyryl)-7-((*E*)-4-(prop-2-yn-1-yloxy)styryl)-5*H*-4 $\lambda^4$ ,5 $\lambda^4$ -dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinine **9**.

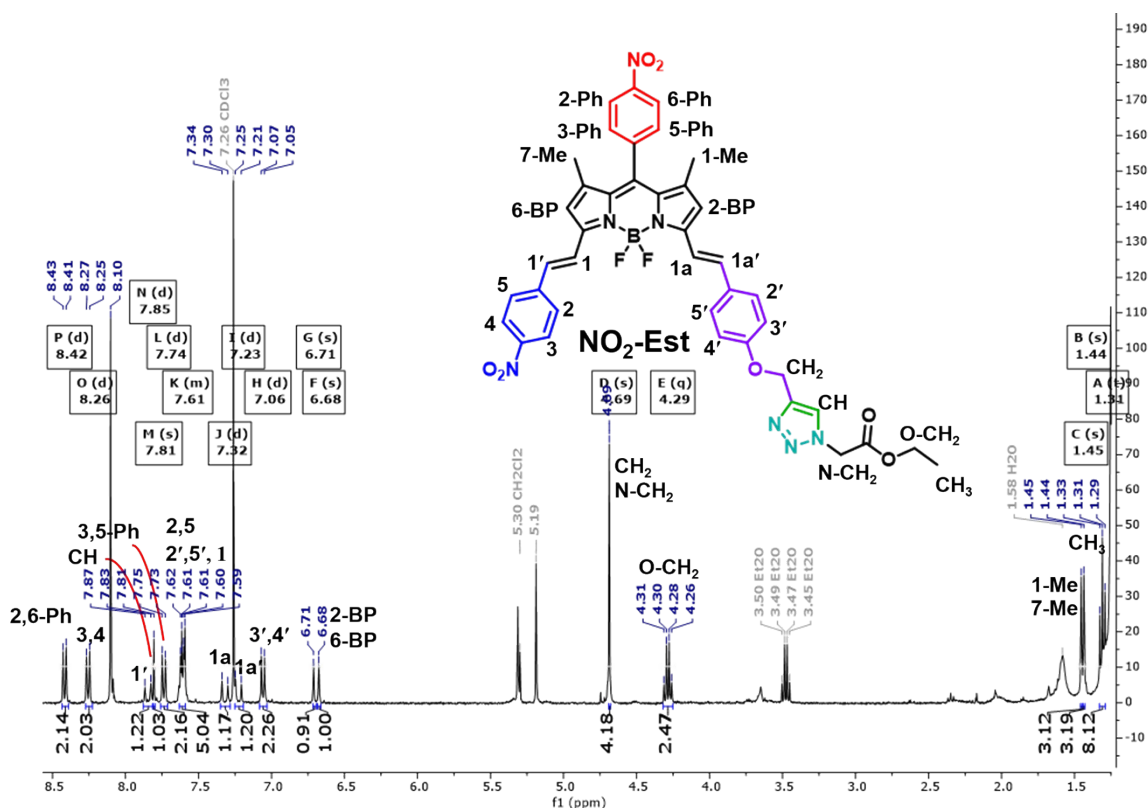


Figure S20.  $^1\text{H}$  NMR-spectrum of ethyl 2-(4-((*E*)-2-(5,5-difluoro-1,9-dimethyl-10-(4-nitrophenyl)-7-((*E*)-4-nitrostyryl)-5*H*-4 $\lambda$ <sup>4</sup>,5 $\lambda$ <sup>4</sup>-dipyrrolo[1,2-*c*:2',1'-*f*][1,3,2]diazaborinin-3-yl)vinyl)phenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)acetate  $\text{NO}_2\text{-Est}$  in  $\text{CDCl}_3$ .

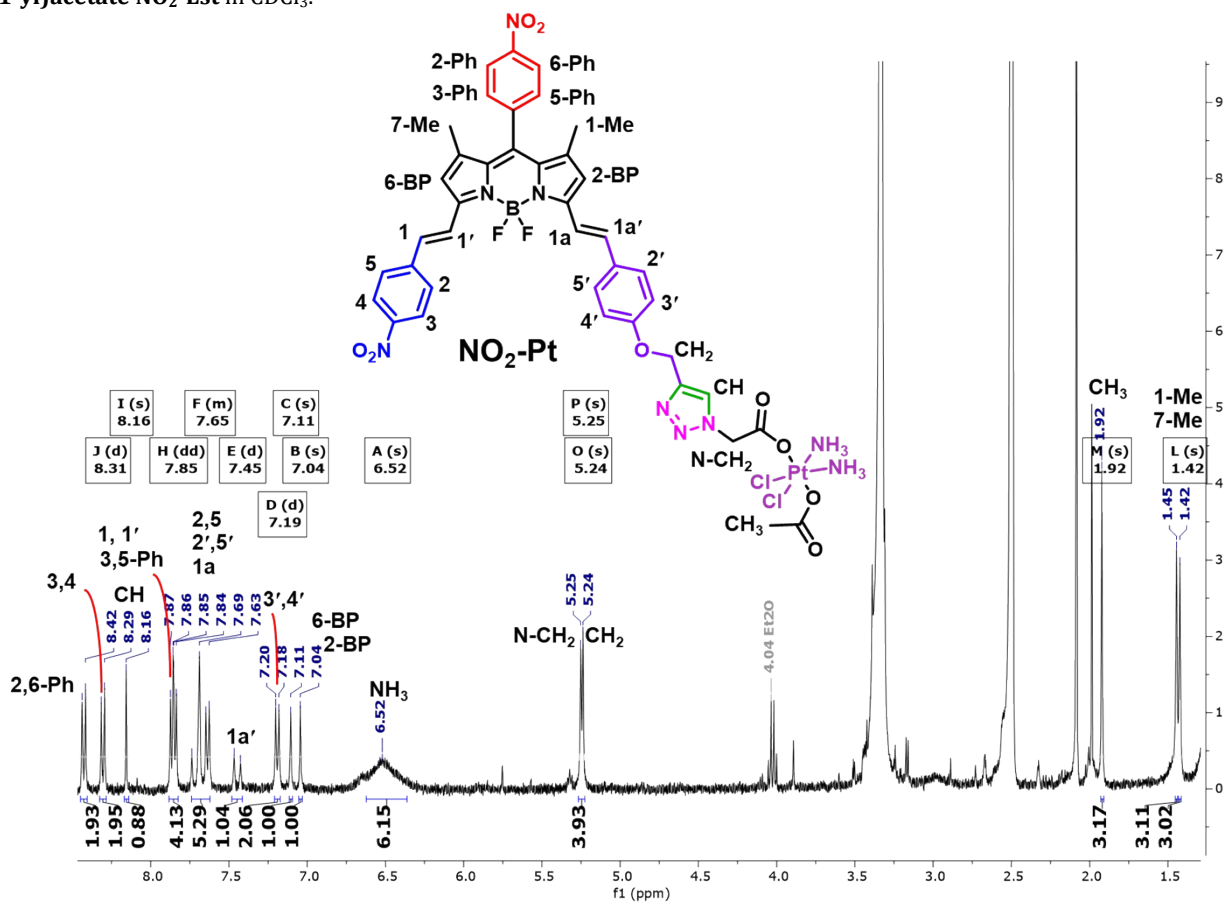


Figure S21.  $^1\text{H}$  NMR-spectrum of  $\text{NO}_2\text{-Pt}$  in  $\text{DMSO-}d_6$ .

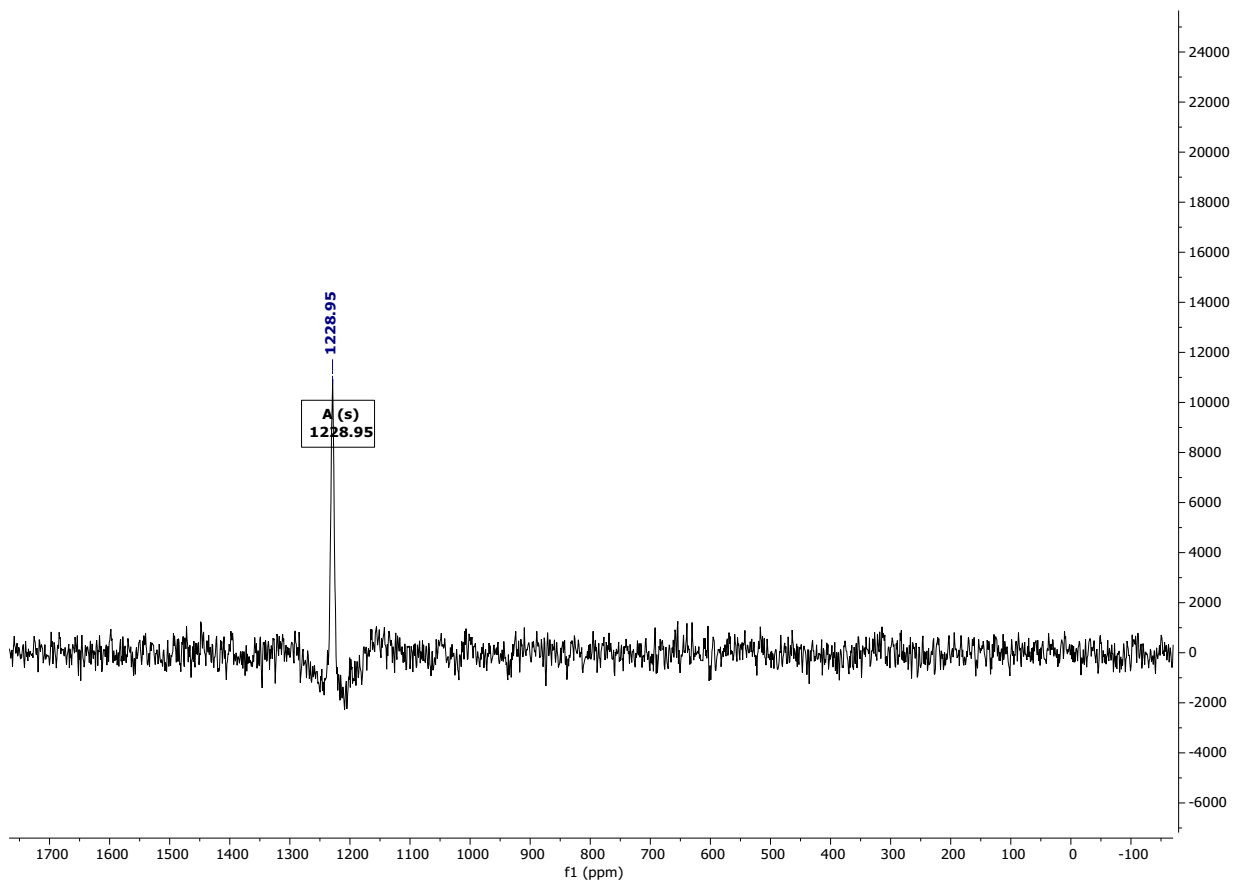


Figure S22.  $^{195}\text{Pt}$  NMR-spectrum of  $\text{NO}_2\text{-Pt}$  in  $\text{DMSO-d}_6$ .

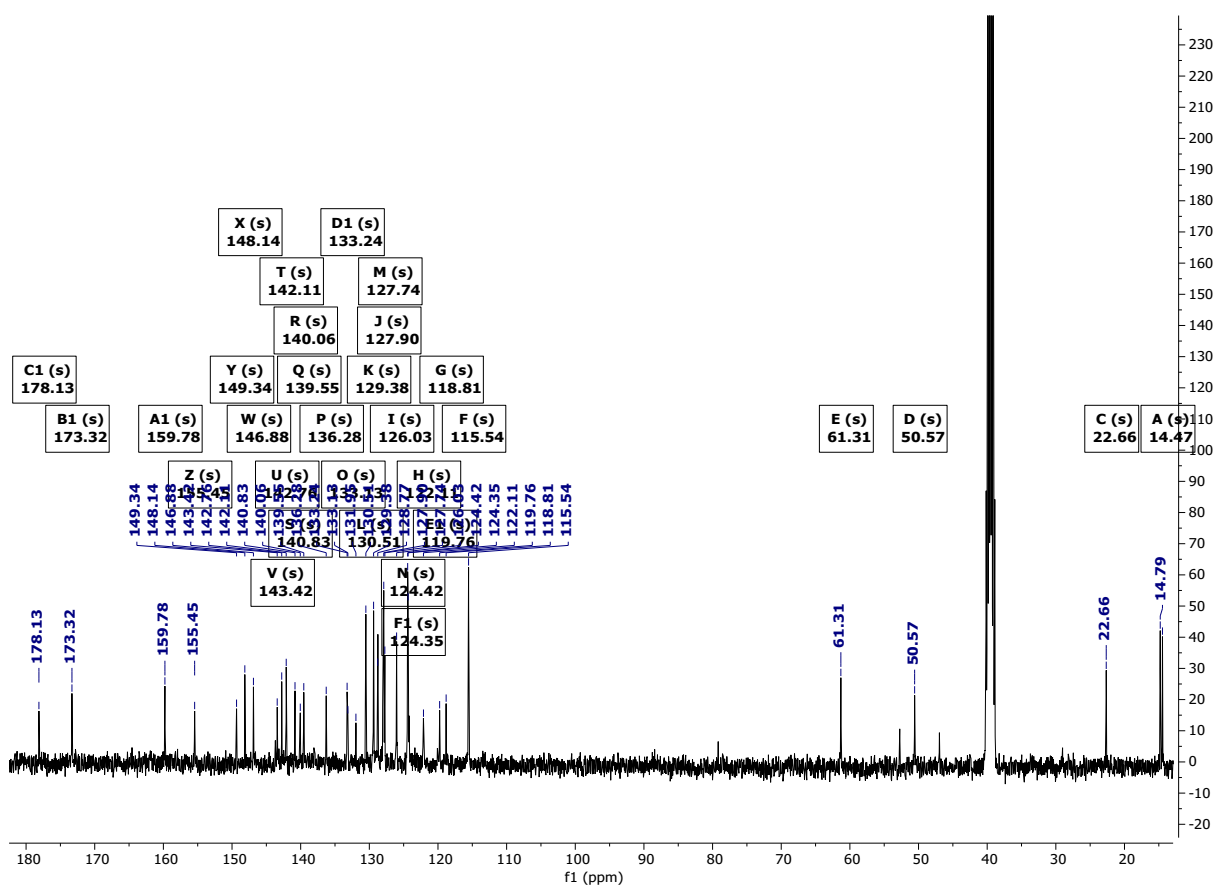
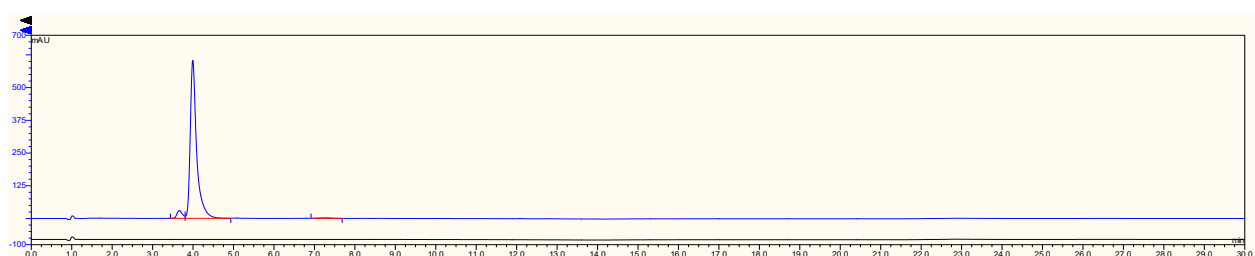
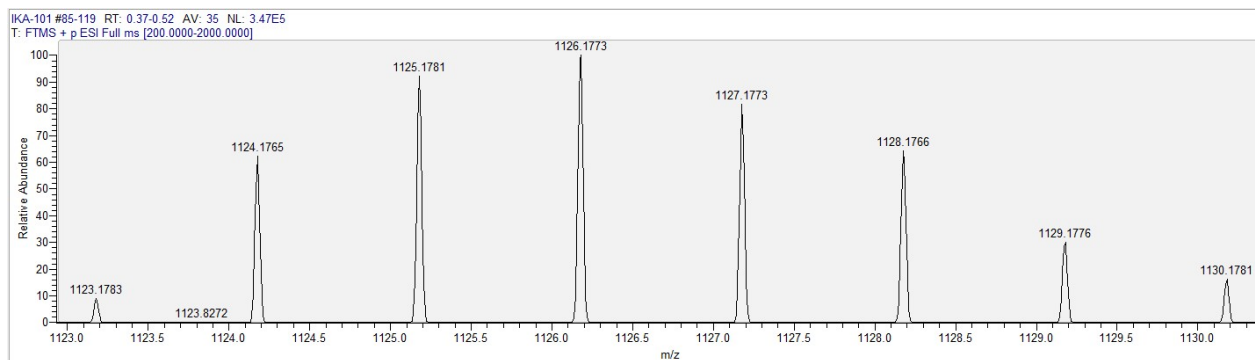


Figure S23.  $^{13}\text{C}$  NMR-spectrum of  $\text{NO}_2\text{-Pt}$  in  $\text{DMSO-d}_6$ .

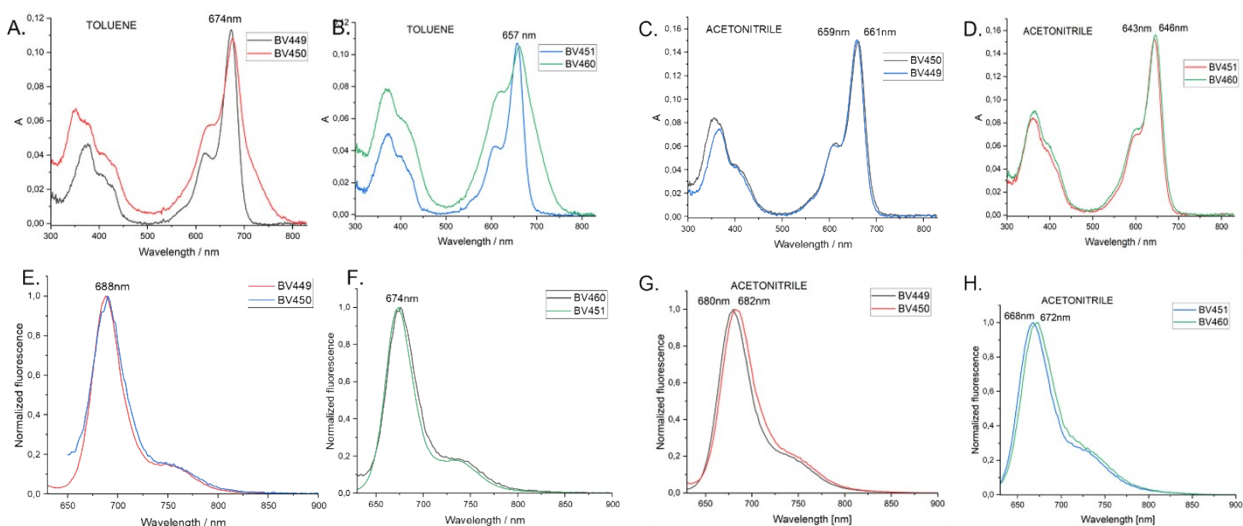


**Figure S24.** ESI-high resolution mass spectrum of  $\text{NO}_2\text{-Pt}$ , and RP-HPLC analysis of  $\text{NO}_2\text{-Pt}$ . UV-VIS detector was set at 650 nm. Main peak area: 95.55%

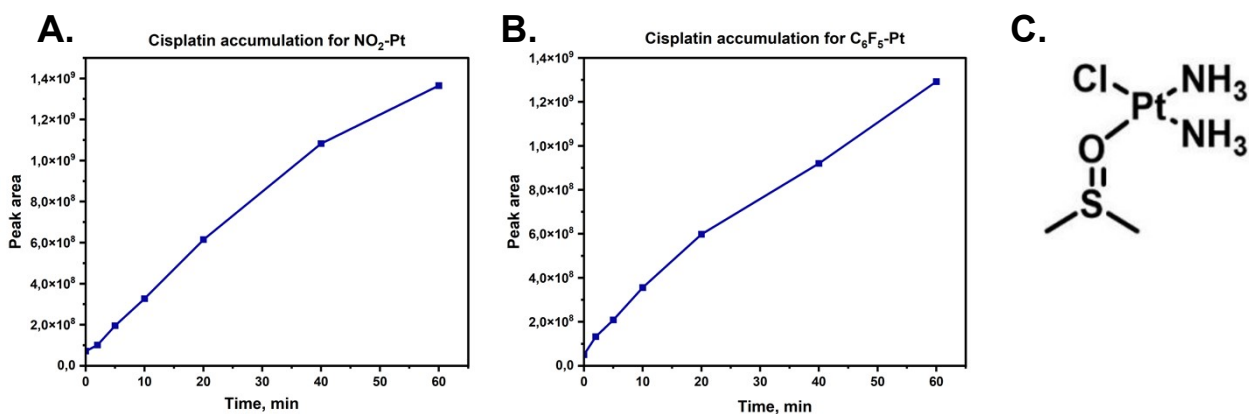
#### 4. Photophysical properties of monomers & cisplatin release

**Table S1.** Quantum yield of singlet oxygen, quantum yield of fluorescence, absorption and emission wavelengths for  $\text{C}_6\text{F}_5\text{-Pt}$ ,  $\text{C}_6\text{F}_5\text{-Est}$ ,  $\text{NO}_2\text{-Pt}$ ,  $\text{NO}_2\text{-Est}$  ( $\text{CH}_3\text{CN}$ ,  $10^{-5}\text{M}$ )

Compound	$\Delta\Phi$	$\Phi_f$	$\lambda_{\text{abs}}$	$\lambda_f$
$\text{C}_6\text{F}_5\text{-Est}$	$\sim 0.001$	0.10	660	680
$\text{C}_6\text{F}_5\text{-Pt}$	$\sim 0.001$	0.09	661	682
$\text{NO}_2\text{-Est}$	$\sim 0.001$	0.04	644	669
$\text{NO}_2\text{-Pt}$	$\sim 0.001$	0.04	646	672



**Figure S25.** Absorption (A-D) and emission (E-H) spectra of  $\text{C}_6\text{F}_5\text{-Pt}$  (BV-450),  $\text{C}_6\text{F}_5\text{-Est}$  (BV-449),  $\text{NO}_2\text{-Pt}$  (BV-460),  $\text{NO}_2\text{-Est}$  (BV-451) in toluene and acetonitrile.



**Figure S26.** HPLC-MS monitoring of the accumulation of the cisplatin adduct  $[\text{Pt}(\text{NH}_3)_2\text{Cl}(\text{DMSO})]^+$  upon irradiation (660 nm, 13.5 mW/cm<sup>2</sup>) of solutions of **NO<sub>2</sub>-Pt** (A) and **C<sub>6</sub>F<sub>5</sub>-Pt** (B). C. Structure of cisplatin-DMSO adduct: one chloride ligand of cisplatin is replaced by DMSO.

## 5. Preparation and characterization of nanoparticles

### 5.1. Nanoparticles preparation

**C<sub>6</sub>F<sub>5</sub>-Pt-F127-NPs**, **C<sub>6</sub>F<sub>5</sub>-Est-F127-NPs**, **NO<sub>2</sub>-Pt-NPs** and **NO<sub>2</sub>-Est-NPs** were synthesized using the similar nanoprecipitation protocol.<sup>5</sup> Briefly, 40 mg of F127 was dissolved in 100 ml of pure distilled water under sonication for 15 minutes. Subsequently, 1000  $\mu\text{L}$  of **BODIPY** solution (1 mg/ml in acetone) was added dropwise slowly to the F127 solution under vigorous stirring. After stirring for 1 hour, the solution was purified and concentrated by centrifugation (1800 rpm, 15 min) using centrifugal filters (Millipore Amicon Ultra-15, MWCO 30 kDa) and washed three times with clean distilled water (1800 rpm, 15 min). The concentrated nanoparticle solution was then passed through a 0.22  $\mu\text{m}$  Millex-HV syringe filter to remove any aggregates. The resulting dark-colored solutions of **NPs** were then characterized via DLS and AFM. The concentration of the nanoparticles was determined spectrophotometrically. To assess the drug loading efficiency (DLE), the BODIPY content within the nanoparticles was quantified using a spectrophotometer (PerkinElmer Lambda 1050). This was done by applying previously obtained calibration curves for **BODIPY** in the concentration range of 0.005–0.0005 mg/ml, prepared in an aqueous solution of F127 (0.1 mg/ml).

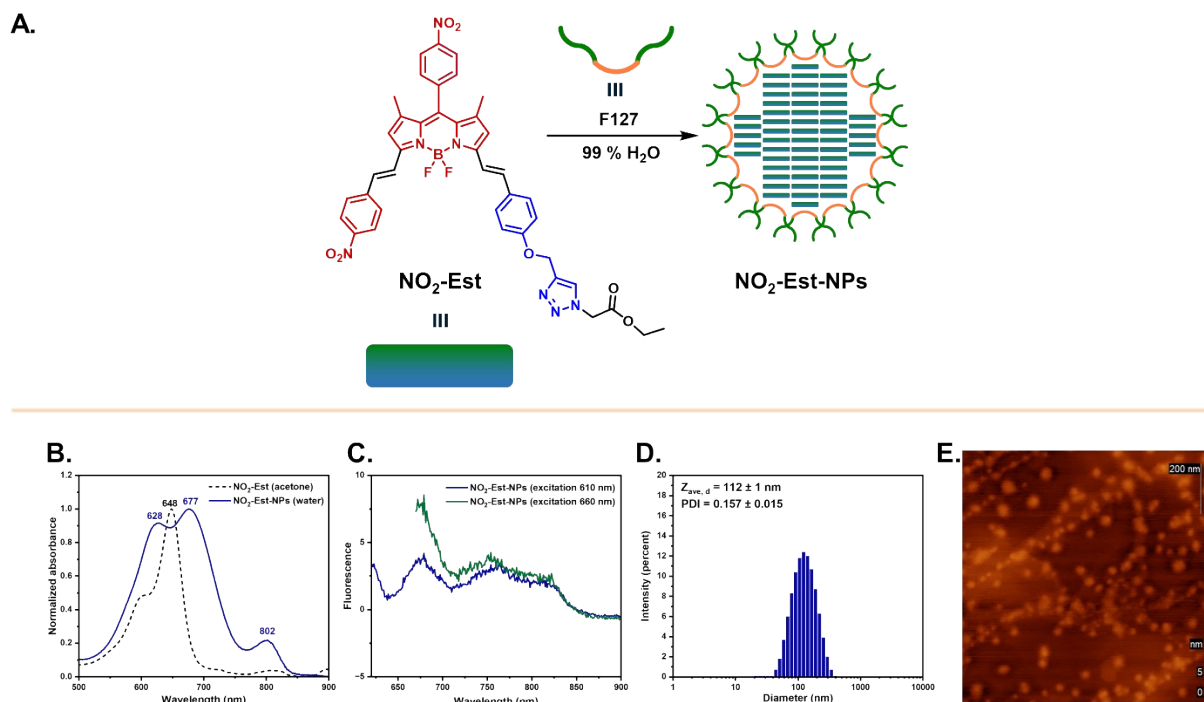
**C<sub>6</sub>F<sub>5</sub>-Pt-TF-NPs** were prepared by thin-film hydration of BODIPY and F127.<sup>2</sup> Briefly, 1 mg of **C<sub>6</sub>F<sub>5</sub>-Pt** and 28 mg of F127 were dissolved by sonication in 6.3 mL of THF. The solvent was slowly removed under reduced pressure. The resulting residue was diluted with 15 mL of distilled water and sonicated in a water bath for 5 minutes. NPs solution was purified and concentrated by centrifugal ultrafiltration (1800 rpm, 15 min) using centrifugal filters (Millipore Amicon Ultra-15, MWCO 30 kDa) and washed four times with clean distilled water (1800 rpm, 15 min). The resulting dark-green nanoparticle suspension was passed through a 0.45  $\mu\text{m}$  Millex-HV syringe filter to remove aggregates. The obtained nanoparticles were characterized via DLS.

An optimized nanoprecipitation protocol was employed for the preparation of **C<sub>6</sub>F<sub>5</sub>-Pt-DSPE-NPs** and **C<sub>6</sub>F<sub>5</sub>-Est-DSPE-NPs**. Briefly, 1000  $\mu\text{L}$  of BODIPY solution (0.2 mg/mL in acetone) was mixed with 1000  $\mu\text{L}$  of an aqueous DSPE-PEG<sub>2000</sub>-OMe solution (1.8 mg/mL). The resulting mixture was subjected to sonication in a water bath maintained at 50 °C. Subsequently, 2.1 mL of ice-cooled pure distilled water (6 °C) was added dropwise over 10 seconds under continuous sonication. The mixture was further sonicated for 1.5 minutes. Thereafter, an additional 10 mL of distilled water (6 °C) was introduced over 40 seconds, followed by extended sonication for 1 minute. Obtained nanoparticle solutions were purified using centrifugal filters (Millipore Amicon Ultra-15, MWCO 30 kDa) to gradually remove residual acetone (1300 rpm, 6 °C, 20 min), and then washed three times with pure distilled water (1300 rpm, 15 °C, 20 min). To eliminate large aggregates, the nanoparticle suspension was passed through 0.45  $\mu\text{m}$  Millex-HV syringe filters. The resulting dark-colored green solution of **C<sub>6</sub>F<sub>5</sub>-Est-DSPE-NPs** and light-colored blue solution **C<sub>6</sub>F<sub>5</sub>-Pt-DSPE-NPs** were then characterized via DLS and AFM. To assess the drug loading efficiency (DLE), the BODIPY content within the nanoparticles was quantified using a spectrophotometer (PerkinElmer Lambda 1050). This was done by applying previously obtained calibration curves for **C<sub>6</sub>F<sub>5</sub>-Est** and **C<sub>6</sub>F<sub>5</sub>-Pt** in the concentration range of 0.005–0.0005 mg/ml, prepared in an acetone:water (1:3) solution of DSPE-PEG<sub>2000</sub>-OMe (0.1 mg/ml).

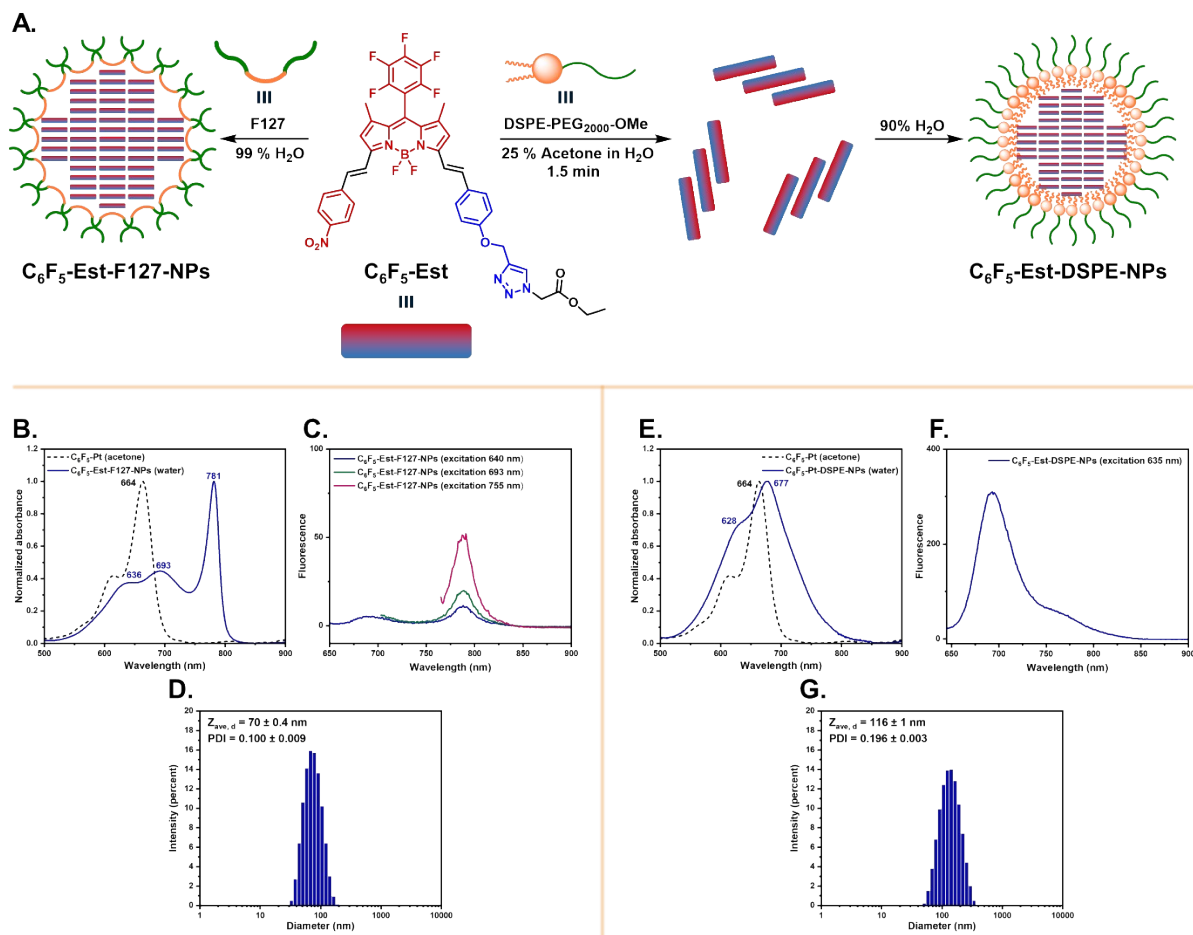
DLE was determined using the equation (1).

$$DLE = \frac{\text{Mass of BODIPY loaded into the nanoparticles (mg)}}{\text{Initial mass of BODIPY (mg)}} \times 100 \quad (1)$$

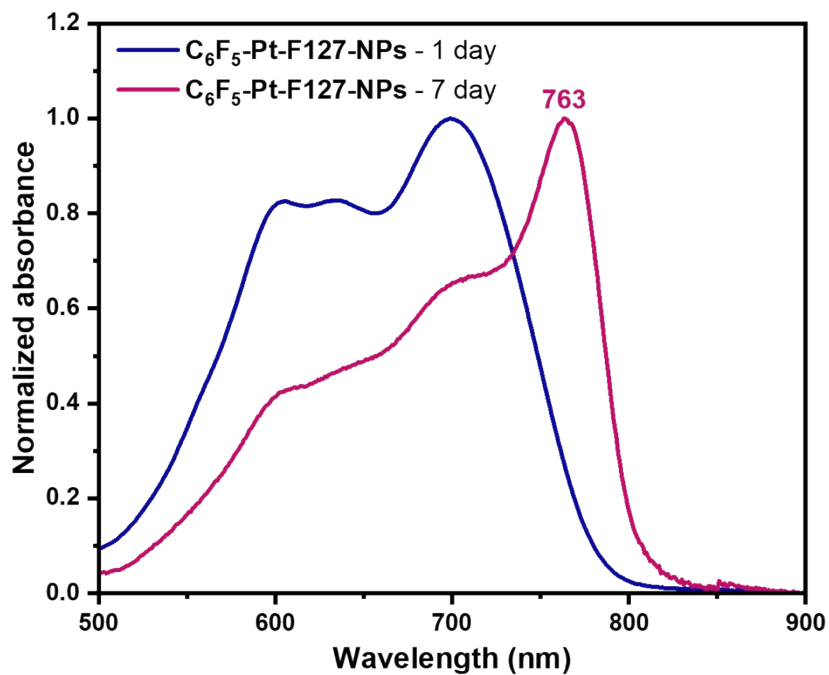
The measured DLE was 43.8% for **C<sub>6</sub>F<sub>5</sub>-Pt-F127-NPs**, 84.5% for **C<sub>6</sub>F<sub>5</sub>-Est-F127-NPs**, 56.0% for **NO<sub>2</sub>-Pt-NPs**, 70.5% for **NO<sub>2</sub>-Est-NPs**, 18.4% for **C<sub>6</sub>F<sub>5</sub>-Pt-DSPE-NPs** and 19.6% for **C<sub>6</sub>F<sub>5</sub>-Est-DSPE-NPs**.



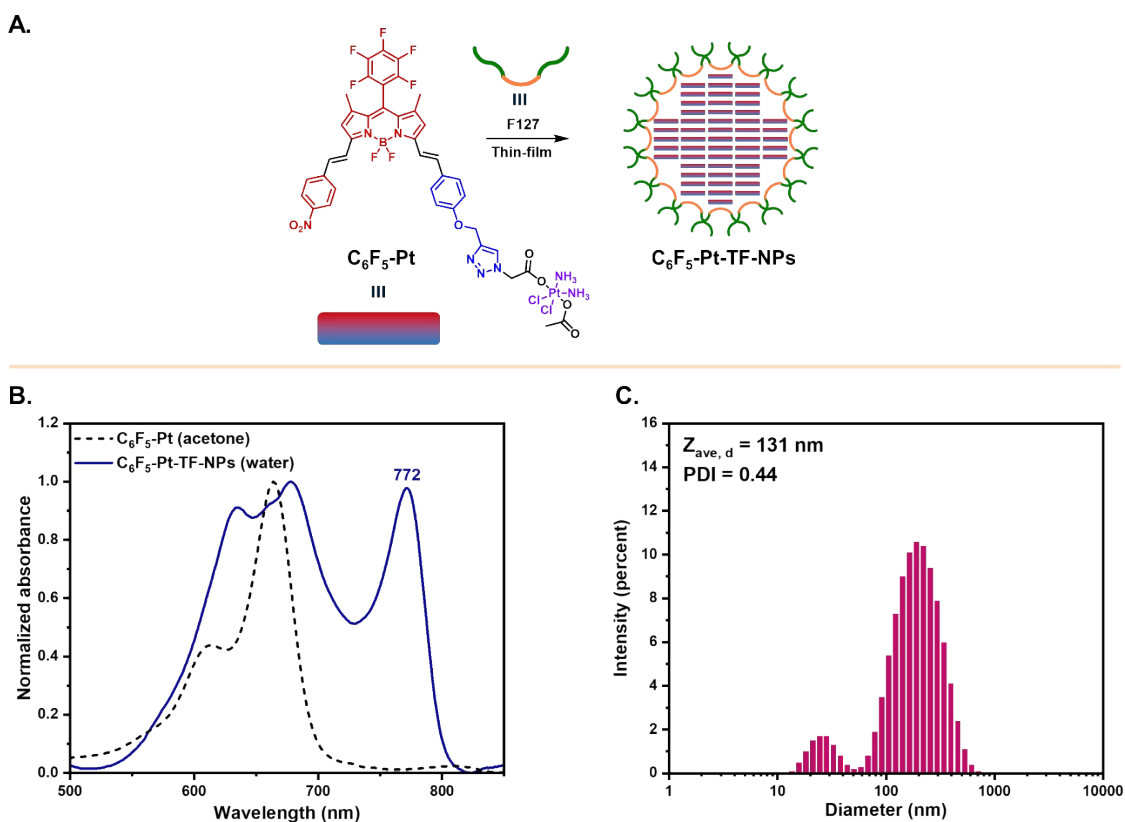
**Figure S27.** A. Synthesis scheme of  $\text{NO}_2$ -Est-NPs. B. Normalized absorption spectra of  $\text{NO}_2$ -Est and  $\text{NO}_2$ -Est-NPs in acetone and water respectively. C. Emission spectra ( $\lambda_{\text{ex}} = 610 \text{ nm}$  and  $660 \text{ nm}$ ) of  $\text{NO}_2$ -Est-NPs in water ( $C = 5 \mu\text{M}$ ). D. Hydrodynamic size distribution of  $\text{NO}_2$ -Est-NPs obtained via DLS. E. AFM image of  $\text{NO}_2$ -Est-NPs. Scale bar: 200 nm



**Figure S28.** A. Synthesis scheme of  $C_6F_5$ -Est-F127-NPs and  $C_6F_5$ -Est-DSPE-NPs. B. Normalized absorption spectra of  $C_6F_5$ -Est and  $C_6F_5$ -Est-F127-NPs in acetone and water, respectively. C. Emission spectra ( $\lambda_{ex}$  = 640 nm, 693 nm and 755 nm) of  $C_6F_5$ -Est-F127-NPs in water ( $C$  = 5  $\mu$ M). D. Hydrodynamic size distribution of  $C_6F_5$ -Est-F127-NPs obtained via DLS. E. Normalized absorption spectra of  $C_6F_5$ -Est and  $C_6F_5$ -Est-F127-NPs in acetone and water, respectively. F. Emission spectra ( $\lambda_{ex}$  = 635 nm) of  $C_6F_5$ -Est-DSPE-NPs in water ( $C$  = 2  $\mu$ M). G. Hydrodynamic size distribution of  $C_6F_5$ -Est-DSPE-NPs obtained via DLS.



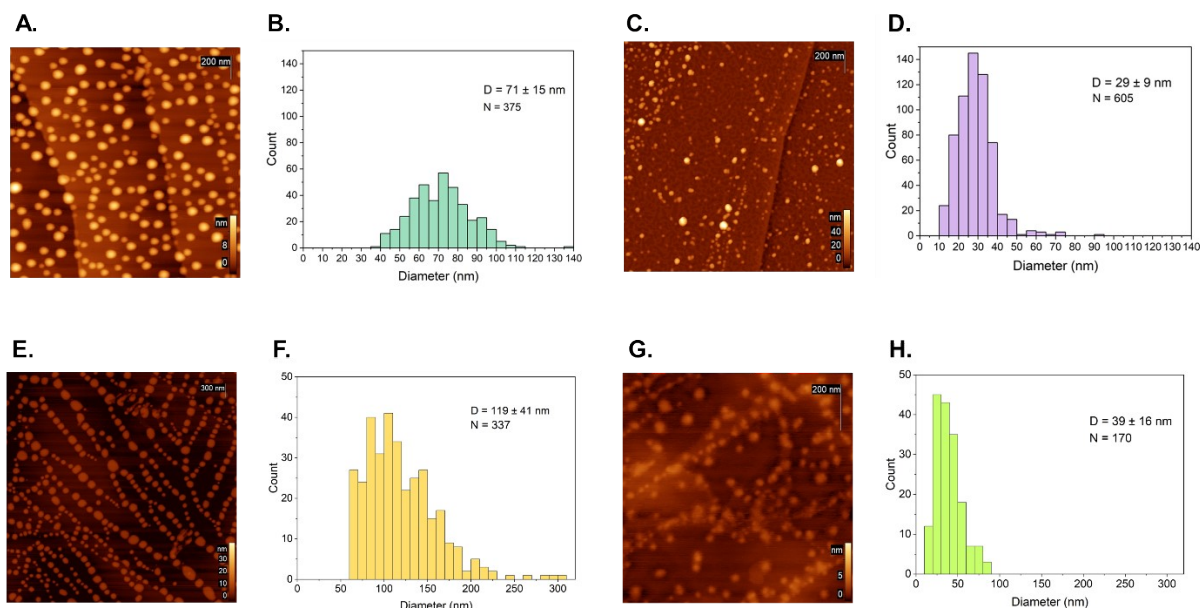
**Figure S29.** Normalized absorption spectra of  $C_6F_5$ -Pt-F127-NPs 1 (blue) and 7 days (pink) after synthesis.



**Figure S30.** A. Synthesis scheme of  $C_6F_5$ -Pt-TF-NPs. B. Normalized absorption spectra of  $C_6F_5$ -Pt and  $C_6F_5$ -Pt-TF-NPs in acetone and water, respectively. C. Hydrodynamic size distribution of  $C_6F_5$ -Pt-TF-NPs obtained via DLS.

## 5.2. AFM characterization of nanoparticles

20  $\mu$ l of each sample solutions were placed on a freshly cleaved highly oriented pyrolytic graphite (ZYH grade, NT-MDT SI, Russia), incubated for 30 min, rinsed with 1 ml of ultrapure water, and dried an argon flow. AFM images were processed in Femtoscan software (Advanced Technologies Center, Russia).<sup>6</sup> For the statistical analysis NPs were traced from the images by circular segments using the special tool in the software. The selected contours were converted into sequences of two-dimensional coordinates and the obtained data were presented as NPs diameters in the form of histograms with bin size 5 or 10.



**Figure S31.** (A) AFM image of  $C_6F_5$ -Pt-F127-NPs. Scale bar: 200 nm. (B) Size distribution histogram via AFM of  $C_6F_5$ -Pt-F127-NPs. Bin size: 5 (C) AFM image of  $NO_2$ -Pt-NPs. Scale bar: 200 nm. (D) Size distribution histogram via AFM of  $NO_2$ -Pt-F127-NPs. Bin size: 5 (E) AFM image of  $C_6F_5$ -Pt-DSPE-NPs. Scale bar: 300 nm. (F) Size distribution histogram via AFM

of  $C_6F_5$ -Pt-DSPE-NPs. Bin size: 10 (G) AFM image of  $NO_2$ -Est-NPs. Scale bar: 200 nm. (H) Size distribution histogram via AFM of  $NO_2$ -Est-NPs. Bin size: 10

### 5.3. Zeta-potential of nanoparticles

For the zeta-potential measurements were used aqueous solution of **BODIPY-NPs** ( $50 \mu\text{M}$ ) 24 hours after their synthesis. Measurements were performed in triplicate. The measured zeta-potential was  $-16.0 \pm 1.5 \text{ mV}$  for  $C_6F_5$ -Pt-F127-NPs,  $-38.0 \pm 2.8 \text{ mV}$  for  $C_6F_5$ -Pt-DSPE-NPs,  $-28.3 \pm 0.7 \text{ mV}$  for  $C_6F_5$ -Est-F127-NPs,  $-21.0 \pm 0.3 \text{ mV}$  for  $C_6F_5$ -Est-DSPE-NPs,  $-20.4 \pm 0.5 \text{ mV}$  for  $NO_2$ -Pt-NPs and  $-32.1 \pm 1.2 \text{ mV}$  for  $NO_2$ -Est-NPs.

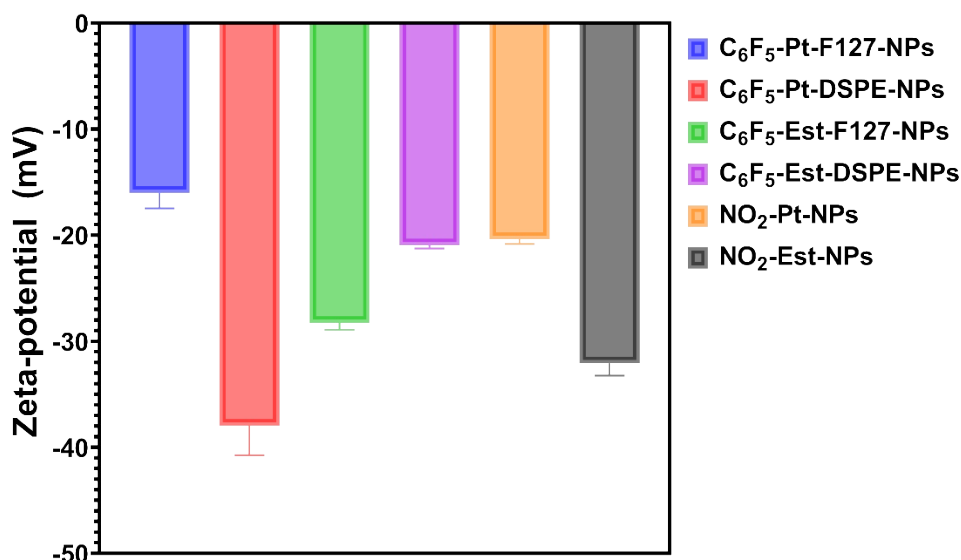


Figure S32. Zeta potential of obtained **BODIPY-NPs** detected by DLS.

### 5.4. Nanoparticles stability assay

Aqueous solutions of **BODIPY-NPs** ( $50 \mu\text{M}$ ) were placed in sealed disposable cuvettes for DLS measurements and incubated at room temperature. At predetermined time intervals the hydrodynamic size of nanoparticles was measured by DLS. Measurements were performed in triplicate.

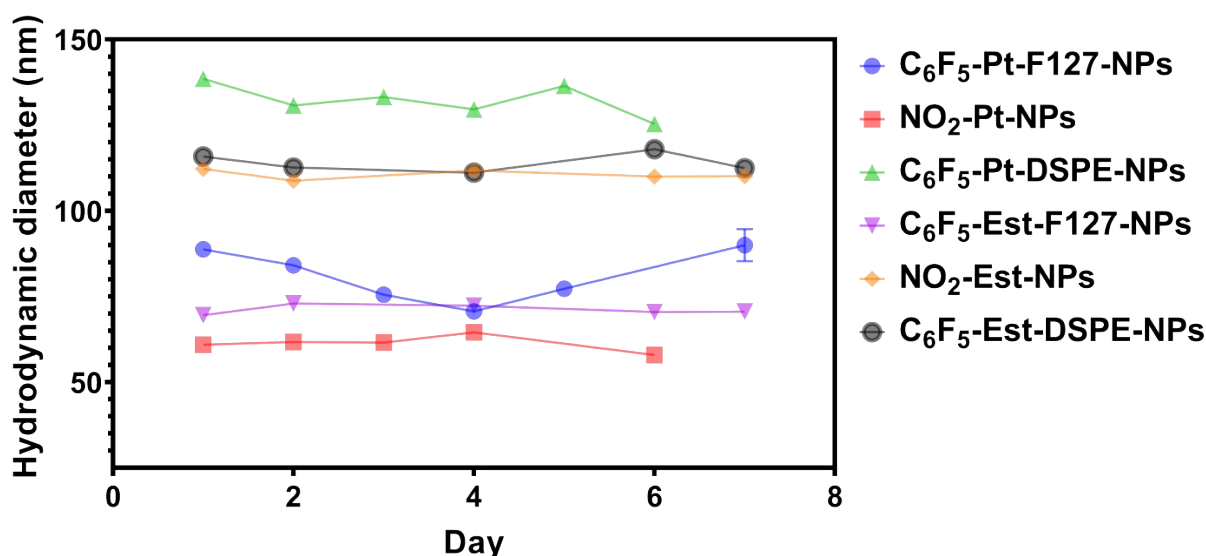
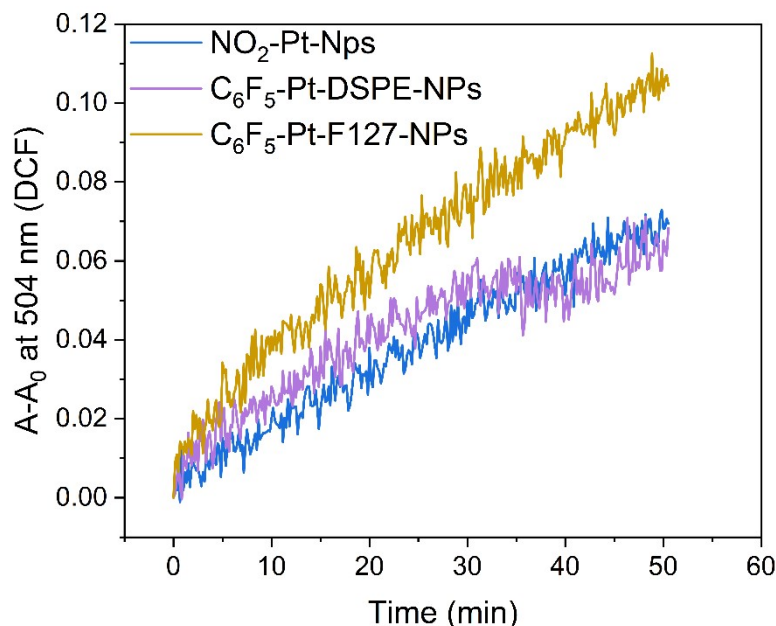


Figure S33.  $Z_{average}$  hydrodynamic diameters of obtained **BODIPY-NPs** in water for 7 or 6 days determined by DLS.

### 5.5. ROS detection using $H_2DCF$ -DA

**ROS detection.** 2',7'-dichlorodihydrofluorescein diacetate ( $H_2DCF$ -DA) was used as a chemical quencher for the detection of total ROS in solution according to generally accepted protocols with slight modifications.<sup>7-9</sup> The non-fluorescent 2',7'-dichlorodihydrofluorescein  $H_2DCF$  was prepared from  $H_2DCF$ -DA by alkaline hydrolysis. An aliquot of  $H_2DCF$ -DA ( $256 \mu\text{L}$ ,  $6269 \mu\text{M}$ ) in methanol was adjusted to 4 mL with aqueous NaOH solution (1 mM) and left for 30 min at  $10^\circ\text{C}$  in the dark. The reaction mixture was then diluted to 40 mL with 10 mM PBS and thoroughly mixed. This resulted in a solution of  $H_2DCF$

in 10 mM PBS with a concentration of 40  $\mu\text{M}$ . This solution was used to prepare mixtures of **NPs** and  $\text{H}_2\text{DCF}$  (40  $\mu\text{M}$ ). These solutions (2 mL) were irradiated at 671 nm with vigorous stirring (2000 rpm) and thermostated at 25°C in cuvettes. The UV-Vis spectra of the solutions were recorded every 10 s during irradiation. Increase in optical density at 504 nm was observed, indicating accumulation of 2',7'-dichlorofluorescein (DCF), the product of the reaction of  $\text{H}_2\text{DCF}$  with ROS.



**Figure S34.** Changes in the UV-Vis spectra of solutions of  $\text{H}_2\text{DCF}$  (40  $\mu\text{M}$ ) and **BODIPY-NPs** in PBS under irradiation at 671 nm.

## 5.6. Photothermal conversion efficiency (PCE) of NPs.

The photothermal conversion efficiency (PCE) of **BODIPY-NPs** was calculated using previously published method.<sup>10</sup> To evaluate photoconversion ability, 100  $\mu\text{L}$  solutions of **BODIPY-NPs** (100  $\mu\text{M}$ ) were irradiated with 745 nm or 660 nm LED at a power density of 120  $\text{mW}/\text{cm}^2$  and 450  $\text{mW}/\text{cm}^2$ , respectively, and then the solution was allowed to cool to room temperature. The temperature was monitored throughout the entire heating and cooling cycle. The PCE was calculated using the Equation (2):

$$\eta = \frac{hA(T_{max} - T_{surr}) - Q_{DIS}}{I(1 - 10^{-A})} \quad (2)$$

where  $h$  represents the heat transfer coefficient,  $A$  is the surface area of the container,  $T_{max}$  is the steady-state temperature (the maximum temperature reached during the heating cycle),  $T_{surr}$  is temperature of the environment,  $Q_{DIS}$  is the heat dissipation property of the solvent and the container (see Equation(6)),  $I$  is the LED irradiation power density ( $\text{W}/\text{cm}^2$ ) and  $A$  is the average absorbance of the sample at FWHM of LED emission.  $hA$  coefficient could be calculated from the Equation (3):

$$\tau_s = \frac{m_D C_D}{hA} \quad (3)$$

where  $m_D$  is the mass of the solution (0.1 g) and  $c_D$  is the heat capacity of  $\text{H}_2\text{O}$  (4.2 J/g). The  $\tau_s$  parameter, which is the sample system time constant, could be calculated from the Equations (4) and (5):

$$t = -\tau_s \ln \Theta \quad (4)$$

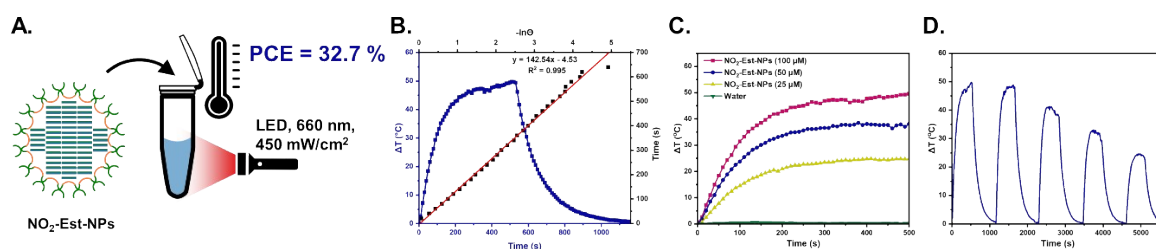
$$\Theta = \frac{T_{RT} - T_{surr}}{T_{max} - T_{surr}} \quad (5)$$

where  $\Theta$  is the dimensionless driving force,  $T_{RT}$  is the real-time temperature at time point  $t$  of the cooling period. Thus,  $\tau_s$  could be obtained from the linear fitting of  $-\ln \Theta$  and time  $t$  of the cooling period of the irradiated solution. The heat dissipation  $Q_{DIS}$  could be calculated from Equation (6):

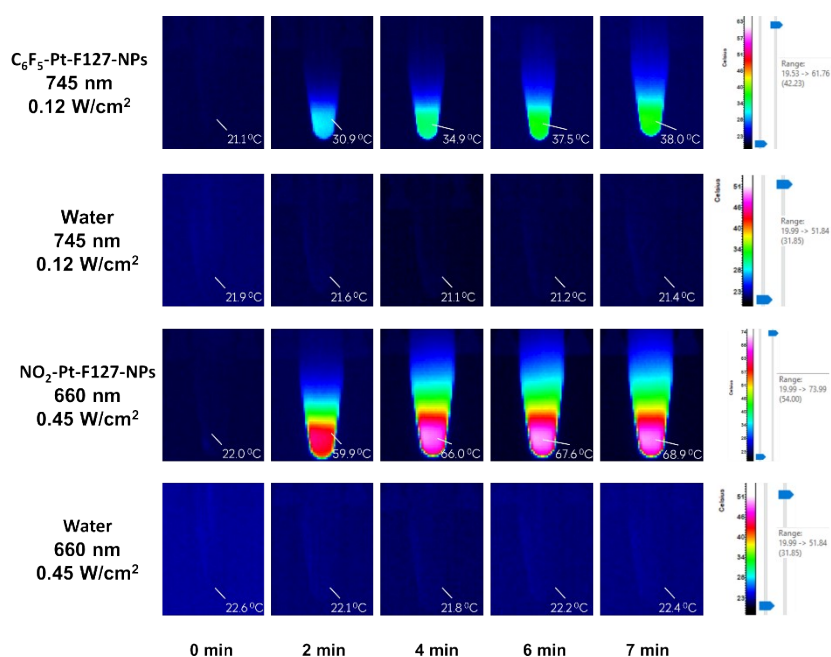
$$Q_{DIS} = \frac{m_D C_D (T_{max(water)} - T_{surr(water)})}{\tau_{S(water)}} \quad (6)$$

where  $T_{max(water)}$  is the steady-state temperature of the solvent heating and cooling cycle and  $\tau_{S(water)}$  is the system time constant of the solvent heating and cooling cycle. According to the obtained data and following the equations (2)-(6)

the photothermal conversion efficiency of  $C_6F_5$ -Pt-F127-NPs,  $NO_2$ -Pt-NPs and  $NO_2$ -Est-NPs was determined to be 31.6 %, 31.2 % and 32.7 % respectively.



**Figure S35.** A. General scheme of photothermal properties evaluation of  $NO_2$ -Est-NPs. B. Temperature elevation and linear fitting of  $-\ln\theta$  and time of the cooling curve of  $100 \mu M$   $NO_2$ -Est-NPs solution under  $660 \text{ nm}$  LED ( $450 \text{ mW/cm}^2$ ). C. Temperature elevation curves of various  $NO_2$ -Est-NPs solutions ( $25\text{--}100 \mu M$ ). D. Temperature elevation curves of 5 heating-cooling cycles of  $100 \text{ mM}$   $NO_2$ -Est-NPs solution.



**Figure S36.** Infrared thermal images of  $100 \mu M$  NPs solutions and water under  $745 \text{ nm}$  ( $0.12 \text{ W/cm}^2$ ) and  $650 \text{ nm}$  ( $0.45 \text{ W/cm}^2$ ).

## 5.7. Light-induced platinum release from $C_6F_5$ -Pt-F127-NPs and $NO_2$ -Pt-NPs

### Release studies protocol

The solutions of  $C_6F_5$ -Pt-F127-NPs and  $NO_2$ -Pt-NPs was washed three times with pure distilled water using centrifugal filters (Millipore Amicon Ultra-0.5, MWCO 30 kDa) at 9000 rpm for 12 minutes to remove any low-molecular weight Pt impurities prior to the start of the experiment. The solution of  $CF_3$ -Pt-NPs was adjusted with pure distilled water to concentrations  $6.5 \text{ mg/L}$  of Pt and was irradiated with  $650 \text{ nm}$  LED ( $400 \text{ mW/cm}^2$ ) under constant stirring. A solution kept in the dark served as a control.

At predetermined time points, an aliquot of either the irradiated solution or the control solution was transferred to a centrifuge filter (Millipore Amicon Ultra-0.5, MWCO 30 kDa), diluted with pure distilled water, and centrifuged at  $10\,000 \text{ rpm}$  for 10 minutes. This process was performed in triplicate for each time point. The platinum concentration in the filtrate and platinum concentration at the start of the experiment were subsequently determined by ICP-MS. Release was determined using Equation (7).

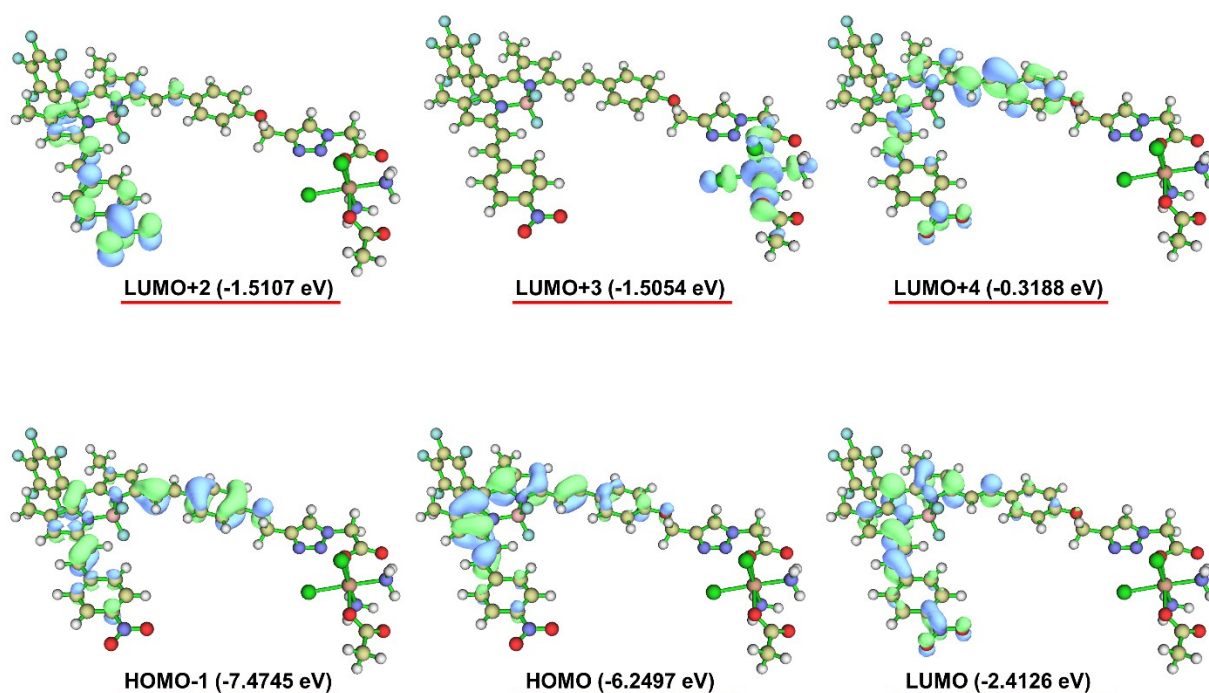
$$\% \text{ of released Pt} = \frac{\text{Concentration of Pt in filtrate}}{\text{Total concentration of Pt in the solution}} \times 100 \quad (7)$$

## 5.8. Quantum-chemical calculations

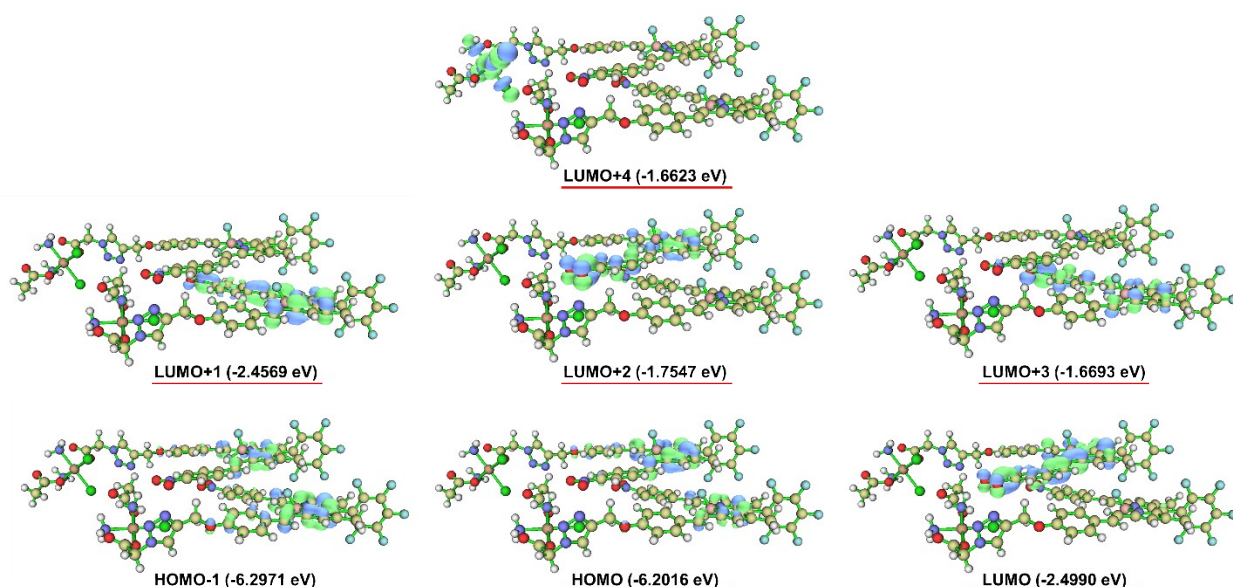
**Quantum-chemical calculations** were performed in ORCA 6.0.1 package.<sup>11,12</sup> The analysis of computational results was performed using the Multiwfn 3.8(dev) package.<sup>13</sup>

**C<sub>6</sub>F<sub>5</sub>-Pt monomer.** Geometry optimization was performed using the composite electronic-structure method r<sup>2</sup>SCAN-3c<sup>14</sup> Implicit solvation was described with the solvation model based on density (SMD)<sup>15</sup> model using parameters for acetone. Single-point and linear-response TDDFT calculations on the optimized monomer geometries were performed with the range-separated CAM-B3LYP functional with relativistic full-electron basis sets (ZORA-def2-TZVP for light elements and SARC-ZORA-TZVP for Pt) within relativistic ZORA approximation.<sup>16,17</sup> Solvent effects were included via SMD (acetone). Excited states were obtained within the Tamm–Dancoff approximation (TDA), requesting 10 singlet roots; triplet states were not computed.

**C<sub>6</sub>F<sub>5</sub>-Pt dimer.** A representative C<sub>6</sub>F<sub>5</sub>-Pt dimer geometry was generated in vacuo using Docker (ORCA's module).<sup>18</sup> The lowest-energy docking pose (prior to any local optimization) was selected and subjected to a short pre-relaxation with r<sup>2</sup>SCAN-3c in the presence of implicit solvent. Specifically, 100 geometry-optimization steps were performed with SMD parameters for water. Single-point and TDDFT calculations on the pre-relaxed dimer were performed with the range-separated CAM-B3LYP functional with relativistic full-electron basis sets (ZORA-def2-TZVP for light elements and SARC-ZORA-TZVP for Pt) within relativistic ZORA approximation. Solvation was described with SMD (water). TDDFT was performed within TDA, requesting 5 singlet roots; triplet states were not computed.



**Figure S37.** Molecular orbitals of C<sub>6</sub>F<sub>5</sub>-Pt involved in S<sub>0</sub>→S<sub>1</sub> transition and lowest Pt-centered molecular orbital (LUMO+3) at an isosurface value of 0.035 a.u.



**Figure S38.** Molecular orbitals of  $C_6F_5$ -Pt-dimer involved in  $S_0 \rightarrow S_{1-3}$  transitions and lowest Pt-centered molecular orbital (LUMO+4) at an isosurface value of 0.035 a.u.

**Table S2.** Calculated electronic excitations energies, oscillator strengths and the related wave functions.

	Excited state	Energy (eV)	$\lambda$ (nm)	$f$ (Oscillator strength)	MOs involved in the transitions
$C_6F_5$ -Pt	$S_1$	2.08	595.6	1.44	H $\rightarrow$ L (93%), H $\rightarrow$ L+2 (1.0%), H-1 $\rightarrow$ L+2 (1.1%), H-1 $\rightarrow$ L+4 (1.3%).
$C_6F_5$ -Pt dimer	$S_1$	1.99	622.7	0.09	H $\rightarrow$ L (25%), H-1 $\rightarrow$ L (29%), H-1 $\rightarrow$ L+1 (6.1%), H $\rightarrow$ L+1 (31%) H $\rightarrow$ L+2 (1.2%)
$C_6F_5$ -Pt dimer	$S_2$	2.07	599.1	2.51	H $\rightarrow$ L (39%), H-1 $\rightarrow$ L+1 (33%), H-1 $\rightarrow$ L+3 (1.9%), H $\rightarrow$ L+1 (17%) H $\rightarrow$ L+2 (1.5%)
$C_6F_5$ -Pt dimer	$S_3$	2.35	527.0	0.06	H $\rightarrow$ L (2.4%), H $\rightarrow$ L+1 (37%), H $\rightarrow$ L+3 (10%), H-1 $\rightarrow$ L (4%) H-1 $\rightarrow$ L+1 (41%) H-1 $\rightarrow$ L+3 (3.6%)

**Neutral  $C_6F_5$ -Pt ( $S_0$ )**

B	-0.92317728706723	-3.03716733098207	-0.07781772734588
N	-2.17222981689864	-3.94935679135754	-0.14359437320949
C	-3.47449975521060	-3.53561219494344	0.06939728922265
C	-3.75488880852913	-2.19047903987495	0.36650843599385
C	-2.73756697645448	-1.24718608768899	0.46642190272182
N	-1.41051896893280	-1.61228689935630	0.27652894434406
C	-0.62888290535088	-0.50260194609875	0.41377254363012
C	-1.47550026140804	0.60530553823273	0.70076531490768
C	-2.77815566076742	0.16699185338213	0.73691558819703
C	-4.33737063986364	-4.67136558595614	-0.06044963938572
C	-3.51942152588250	-5.74610357586621	-0.35521700713033
C	-2.18026713326528	-5.29271507792383	-0.40711243373078
F	-0.02945547533448	-3.50720210851711	0.90615991316219
F	-0.25919585680960	-3.03078465388699	-1.32185128779986
C	-5.16334294218261	-1.77202688002688	0.57021944091818
C	-5.71518158896714	-1.67821984177755	1.84188419566808
C	-7.03071522015834	-1.29047770386705	2.04304088931205

C	-7.82612919157472	-0.98447109168525	0.94754932632431
C	-7.30254032575908	-1.06874565107962	-0.33514860274374
C	-5.98453354663658	-1.46171337638355	-0.50706702903630
F	-9.09300329946923	-0.60863307925916	1.12783734794694
C	-3.96333073796168	1.02759708443842	1.01267998372738
C	-5.82096066693103	-4.74192771434818	0.08122523980248
C	-0.98415489618779	-6.02327501336385	-0.68006514149814
C	0.78769222427275	-0.53674343973869	0.27178983638756
C	1.56870771357770	0.57346694395710	0.37489719875865
C	-0.96168904260422	-7.35157384626035	-0.96135223508769
C	3.00241264541076	0.62978144370271	0.23679549647562
C	0.22204459884566	-8.13065093454414	-1.24779388938480
C	3.64140126231023	1.88524287577175	0.33192990556127
C	5.00727449774492	2.01172291845483	0.19064929989652
C	5.79204248866928	0.87466697749166	-0.05191499220940
C	5.18486079885204	-0.38828331896909	-0.13822019644874
C	3.81222630897405	-0.49837593120443	0.00371941197096
C	0.06780900256888	-9.50543318741316	-1.52464202629049
C	1.15579833011453	-10.31063292005859	-1.80373717663151
C	2.42718432751597	-9.74009563077823	-1.80925008643511
C	2.62065867383050	-8.38285307906942	-1.54235027479521
C	1.52645568362733	-7.58951331326337	-1.26504989724196
O	7.11908896038069	1.09845284010729	-0.18786416791025
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**Neutral C<sub>6</sub>F<sub>5</sub>-Pt-dimer (S0)**

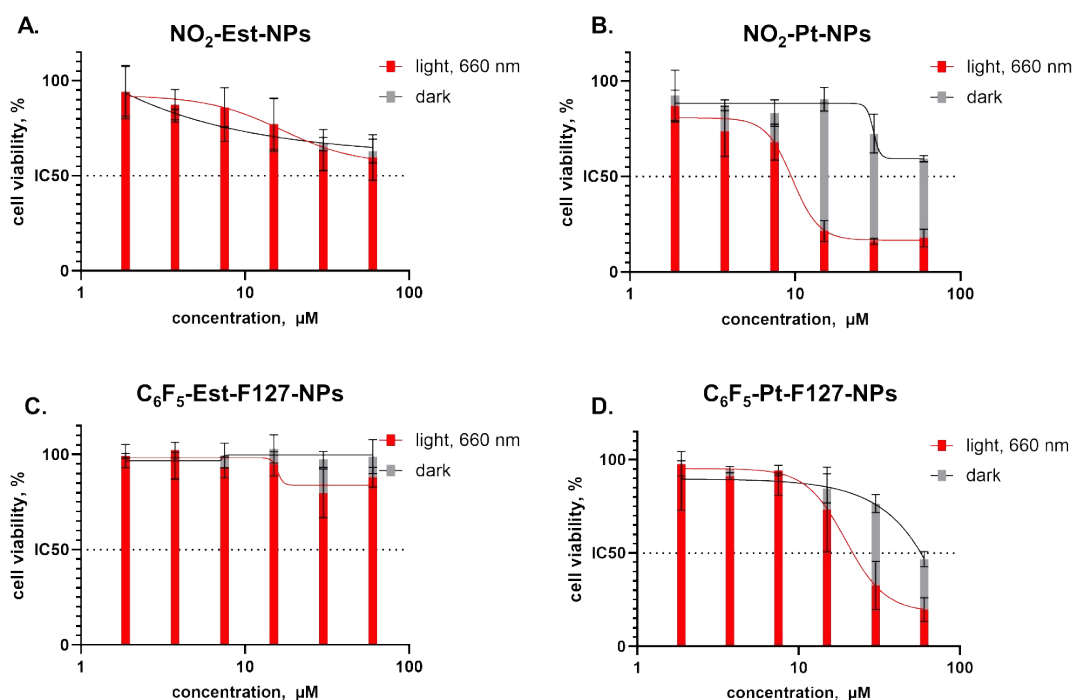
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H	15.26480561156563	-10.37487292254789	0.62463320910079
H	14.22965884296979	-9.11591190386574	1.34168939174669
H	15.78329305017383	-9.57075573704269	2.11448081867876
O	14.90618860433816	-11.92301274986587	2.91108769705928
N	12.95402345810279	-12.37466677781737	4.97863211297915
H	12.84023551893612	-13.28118101149843	5.43665615628273
H	12.69799448211518	-11.62539455640483	5.61997515849659
H	13.92536340185815	-12.26230861299578	4.67750545675844

## 5.9. MTT assay

Human breast adenocarcinoma Sk-Br-3 cells were seeded in 96-well plates ( $10^4$  cells per well) in complete DMEM medium. The following day, the medium in some wells was replaced with 100  $\mu$ l of fresh medium containing **BODIPY-NPs** in the concentration range of 60-1.87  $\mu$ M (including control points and negative control). The cells were incubated with nanoparticles for 1 h, after which they were irradiated at a wavelength of 650 nm ( $90 \text{ mW/cm}^2$ ) for 13 min; non-irradiated cells served as a control. Cell viability was assessed after 72 h using the MTT assay. For this, an MTT solution (0.5 mg/ml) was added to each well. After 2 h, the medium was removed and 100  $\mu$ l of DMSO (99%) was added. Optical density was recorded at 565 nm on an Infinite M Nano reader (Tecan, Switzerland).



**Figure S39.** Cell viability of Sk-Br-3 cells incubated with BODIPY-NPs at various concentrations in the dark and after LED light irradiation (650 nm, 13 min, 90 mW/cm<sup>2</sup>). A. NO<sub>2</sub>-Est-NPs. B. NO<sub>2</sub>-Pt-NPs. C. C<sub>6</sub>F<sub>5</sub>-F127-NPs. D. C<sub>6</sub>F<sub>5</sub>-Pt-F127-NPs.

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