

## Supporting Information

# Rigorous data curation, enrichment and meta-analysis enable autoML prediction of plant length responses to nanoparticles powered by the Enalos Cloud platform

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## Supplementary tables

**Table S1.** List of features included in the original Deng et al.<sup>1</sup> “Length” dataset.

Feature	Description
Carbide	Number of carbon atoms
Metal	Number of metal atoms
MC	Macromolecular compound
Oxide	Number of oxygen atoms
Com1	Component 1 [ $A_r$ of Com1] where $A_r$ is the relative atomic mass
Com2	Component 2 [ $A_r$ of Com2]
Dim0	Granular
Dim1	One-dimensional
Dim2	Two-dimensional
Hollow	Hollow
sizeTEM	Particle size measured by transmission electron microscopy [nm]
Purity	NP purity [%]
sizeDLS	Particle size measured by dynamic light scattering [nm]
Zeta	Zeta potential [mV]
Species	Plant species (cucumber, bean, wheat, rice, tomato, maize)
EP	NP exposure pathway (seeds, root, leaf)
MT	Measured tissue (root, shoot, plant)
Age	Exposure age [day]
GS	Growth stage of plant (germination, seedling, vegetative)

<b>Cultured</b>	Cultivation method (medium, hydroponic, soil)
<b>Category</b>	Plant carbon fixation metabolic pathway (C <sub>3</sub> or C <sub>4</sub> photosynthetic process)
<b>TC</b>	Total content (dose) [mg]
<b>Duration</b>	The time elapsed from the exposure of the plant to NPs to the measurement [day]
<b>Photoperiod</b>	Hours of plant exposure to light per day [hours/day]
<b>Illumination</b>	Illumination intensity [ $\mu\text{mol}\cdot\text{m}^{-2}\text{s}^{-1}$ ]
<b>Humidity</b>	Humidity [%]
<b>DT</b>	Daytime temperature [°C]
<b>NT</b>	Night temperature [°C]

**Table S2.** List of corrections to the original Deng *et al.*<sup>1,2</sup> dataset.

Original dataset rows	Corrections	Referenced study
2-7	Positive labels were normalised based on the control sample.	Cui <i>et al.</i> (2014) <sup>3</sup>
13,17	Positive labels were normalised based on the control sample.	Konate <i>et al.</i> (2018) <sup>4</sup>
18-29	Rows removed because the NP size was not clearly mentioned.	Gopalakrishnan Nair <i>et al.</i> (2015) <sup>5</sup>
30-37	Rows removed because the total NP concentration in mg/L could not be calculated.	Abdel Latef <i>et al.</i> (2018) <sup>6</sup>
38-51	Illumination intensity values for medium cultivations were corrected to 500 $\mu\text{mol}/\text{m}^2\text{s}$ .	Feng <i>et al.</i> (2019) <sup>7</sup>
45-51	“Cultured” column correction. The root length was measured after medium cultivation instead of hydroponic cultivation.	Feng <i>et al.</i> (2019) <sup>7</sup>
52-58	Positive labels were normalized based on the control sample. Negative labels were re-calculated for consistency.	Feng <i>et al.</i> (2019) <sup>7</sup>
67-68, 71-72, 76-77, 80-81	Rows removed because the NP size was not clearly mentioned in the referenced study.	Mahawar <i>et al.</i> (2018) <sup>8</sup>
65-66	Rows removed because atomistic descriptors could not be calculated for large $\text{Fe}_2\text{O}_3$ NPs.	Mahawar <i>et al.</i> (2018) <sup>8</sup>
74-75	Rows removed because atomistic descriptors could not be calculated for large $\text{Fe}_2\text{O}_3$ NPs.	Mahawar <i>et al.</i> (2018) <sup>8</sup>
82-90	Positive labels were normalized based on the control sample. Negative labels were re-calculated for consistency.	Cui <i>et al.</i> (2014) <sup>3</sup>
91-108	Rows removed because the total NP concentration in mg/L could not be calculated.	Wang <i>et al.</i> (2019) <sup>9</sup>
109-110	Rows removed because the NP size was not clearly mentioned in the referenced study.	Zuo <i>et al.</i> (2017) <sup>10</sup>
111-113	Rows removed because the total NP concentration in mg/L could not be calculated.	Du <i>et al.</i> (2015) <sup>11</sup>
114-117	Rows removed because the NP size was not clearly mentioned in the referenced study.	Wang <i>et al.</i> (2020) <sup>12</sup>
118-119	Rows removed because the NP shape was not clearly mentioned in the referenced study.	Iftikhar <i>et al.</i> (2020) <sup>13</sup>
120-129	Rows removed because the NP shape was not clearly mentioned in the referenced study.	Hussain <i>et al.</i> (2018) <sup>14</sup>
130-133	Rows removed because the NP shape was not clearly mentioned in the referenced study.	Khan <i>et al.</i> (2019) <sup>15</sup>
134-137	Rows removed because the NP shape was not clearly mentioned in the referenced study.	Wang <i>et al.</i> (2020) <sup>16</sup>
138-140	Rows removed because the total NP concentration in mg/L could not be calculated.	Li <i>et al.</i> (2020) <sup>17</sup>
141-146	Rows removed because the NP size was not clearly	Adhikari <i>et al.</i> (2015) <sup>18</sup>

	mentioned in the referenced study.	
148-149	Positive labels were normalized based on the control sample.	Lian <i>et al.</i> (2020) <sup>19</sup>
150-151, 155-156	Rows removed because the total NP concentration in mg/L could not be calculated for foliar NP treatments.	Lian <i>et al.</i> (2020) <sup>19</sup>
157-160	Rows removed because the total NP concentration in mg/L could not be calculated.	Toqeer <i>et al.</i> (2020) <sup>20</sup>
161-163	Rows removed because the total NP concentration in mg/L could not be calculated.	Yan <i>et al.</i> (2020) <sup>21</sup>
165-167	Positive labels were normalized based on the control sample.	Rizwan <i>et al.</i> (2019) <sup>22</sup>
175-181	Positive labels were normalized based on the control sample. Negative labels were re-calculated for consistency.	Feng <i>et al.</i> (2019) <sup>7</sup>
189-210	Rows removed because atomistic descriptors could not be calculated for large carbon sheets and tubes.	López-Vargas <i>et al.</i> (2020) <sup>23</sup>
211-215	Rows removed because the total NP concentration in mg/L could not be calculated.	Zadeh <i>et al.</i> (2019) <sup>24</sup>
216-231	Rows removed because the data could not be matched to any referenced study.	No referenced study found
233-234	Positive labels were normalized based on the control sample.	Wang <i>et al.</i> (2020) <sup>25</sup>
239-248	Positive labels were normalized based on the control sample. Negative labels were re-calculated for consistency.	Abbas <i>et al.</i> (2019) <sup>26</sup>
249-260	sizeTEM and purity of NPs were corrected to 30 nm and 97.5% respectively.	Liu <i>et al.</i> (2018) <sup>27</sup>
261-268	Mean illumination intensity, mean humidity and daytime and nighttime temperatures were corrected to 305 $\mu\text{mol}/\text{m}^2\text{s}$ , 65% and 20°C and 28°C respectively.	Ji <i>et al.</i> (2017) <sup>28</sup>
269-272	Rows removed because atomistic descriptors could not be calculated for polymeric NPs.	Zhou <i>et al.</i> (2021) <sup>29</sup>
273-277	Rows removed because atomistic descriptors could not be calculated for large polymeric NPs.	Li <i>et al.</i> (2020) <sup>30</sup>
278-282	Rows removed because atomistic descriptors could not be calculated for large polymeric NPs.	Li <i>et al.</i> (2020) <sup>31</sup>
283-294	Rows removed because atomistic descriptors could not be calculated for large polymeric NPs.	Gong <i>et al.</i> (2021) <sup>32</sup>
295-299	Rows removed because atomistic descriptors could not be calculated for large polymeric NPs.	Lian <i>et al.</i> (2020) <sup>33</sup>
	7 rows were added to include shoot measurements.	Wang <i>et al.</i> (2020) <sup>25</sup>
	5 rows were added to include shoot measurements.	Gopalakrishnan Nair <i>et al.</i> (2015) <sup>34</sup>

## Plot digitisation error analysis

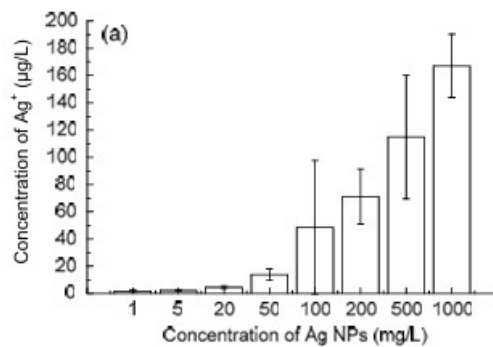
In order to normalise the (root, shoot, overall plant) length labels relative to the control value, it was required to extract the actual experimental measurement values from the bar plots presented in the referenced papers, whenever these were not clearly referred in the text. Each plot was copied with its original aspect ratio and image resolution preserved: Screenshots of plots were taken directly from PDF files, avoiding any external resizing or compression artifacts and then imported into the WebPlotDigitizer<sup>35</sup> tool. As an initial step, two known, distinct points on the y-axis range were manually selected, and their values were entered into the tool to calibrate the bar plot. Subsequently, a point on the top of each bar was manually identified, from which the tool computed the corresponding numerical value.

During this process, both human systematic error and low image quality were recognised as potential sources of inaccuracy in the extracted numerical length values. To minimise this error, all plots were digitised by the same analyst, and a quality control analysis was conducted. To do so, plots for which the original numerical values were explicitly reported in the referenced articles were selected for digitisation, allowing a direct comparison between the extracted and published values. Although numerical values were not provided for the length parameter, similarly styled plots from the same studies depicting experimental parameters not included in the final dataset were used for this comparison. As these plots originate from the same studies (featuring similar aspect ratios, bar dimensions, and image resolutions) and were subjected to an identical digitisation procedure, including the same axis calibration method, they were considered appropriate proxies for assessing the digitisation process error.

Three examples of differently designed bar plots (different axis scaling, bar width and colour, see Figures S1-S3) were analysed, and their extracted, digitised values were compared to the referenced ones within the text. The relative absolute errors (RAE, Eq. 1) were then calculated as reported in Tables S3-S5. In all cases, the RAE was less than 1% which confirms that the plot digitisation method is suitable for extracting numerical values to be used in model development.

$$\%RAE = \frac{|x_{reported} - x_{digitized}|}{x_{reported}} \cdot 100 [1]$$

### Example 1. Digitisation of plots from Cui *et al.* (2014)<sup>3</sup>

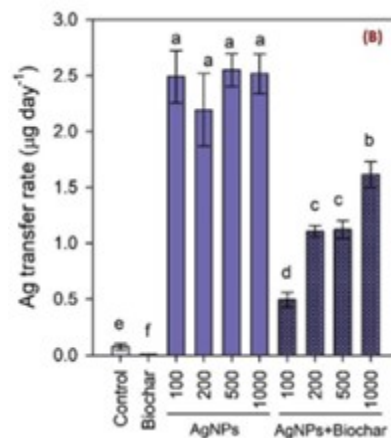


**Figure S1.** Free ion concentrations in exposure solutions of Ag NPs after 5-d treatment of Ag NPs at the germination stage of cucumber. Figure 4(a) in the study of Cui *et al.*<sup>3</sup>

**Table S3.** Reported and Digitiser values comparison for Figure S1 and calculated %RAE.

Concentration of Ag NPs (mg/L)	Reported value	Digitiser value	%RAE
1	2	2.016	0.806
1000	167	167.742	0.444

Example 2. Digitisation of plots from Abbas *et al.* (2019)<sup>26</sup>

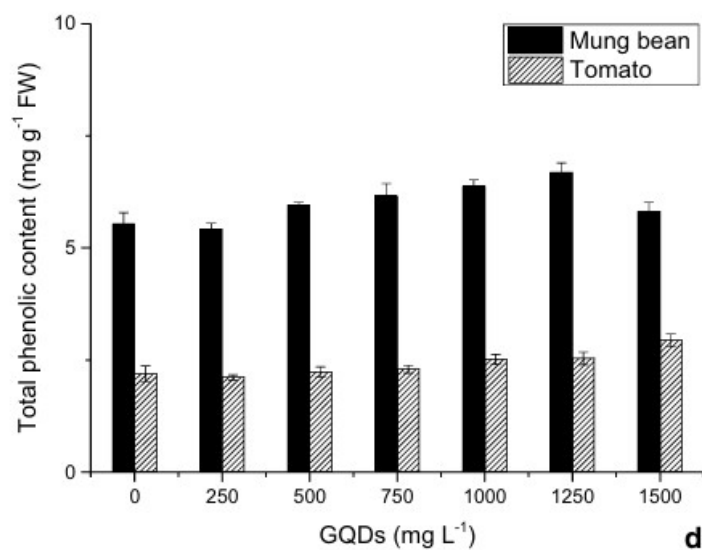


**Figure S2.** Ag transfer rate ( $\mu\text{g day}^{-1}$ ). Figure 4(B) in the study of Abbas *et al.*<sup>26</sup>

**Table S4.** Reported and Digitiser values comparison for Figure S2 and calculated %RAE.

AgNPs+Biochar	Reported value	Digitiser value	%RAE
1000	1.62	1.616	0.225

### Example 3. Digitisation of plots from Feng *et al.* (2019)<sup>7</sup>

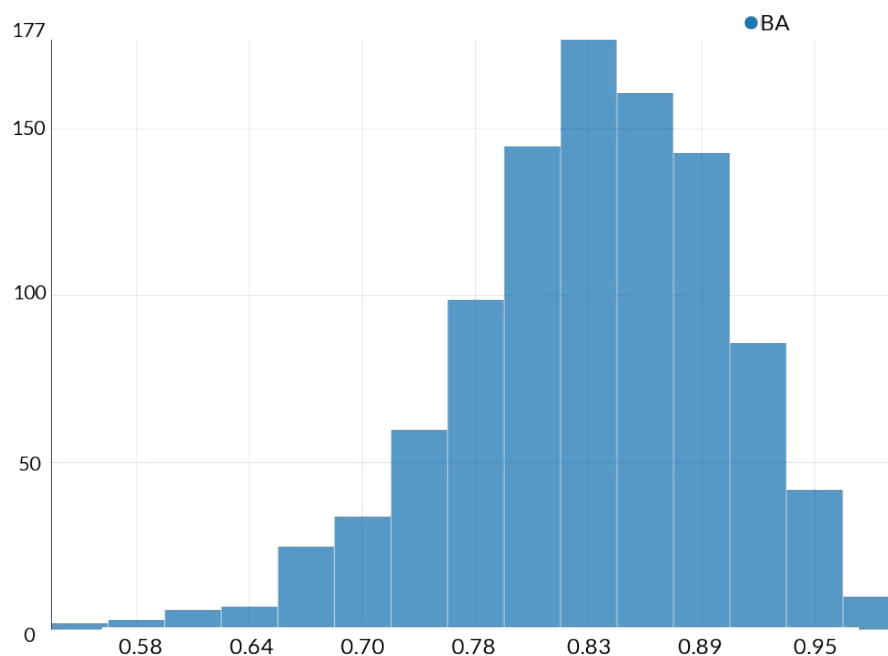


**Figure S3.** Effect of GQDs total phenolic contents in mung bean and tomato seedlings after 2 weeks of culture in hydroponic medium containing different concentrations of GQDs. Figure 5d in the study of Feng *et al.*<sup>7</sup>

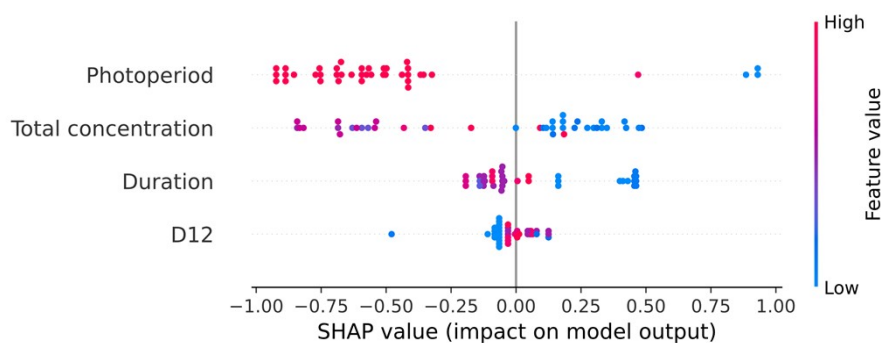
**Table S5.** Reported and Digitiser values comparison for Figure S3 and calculated %RAE.

GQDs (mg L <sup>-1</sup> )	Reported value	Digitiser value	%RAE
1250 Mung Bean	6.69	6.686	0.061
1500 Tomato	2.94	2.948	0.271

## Supplementary figures



**Figure S4:** Histogram of BA values obtained from 1000 bootstrap resamplings of the test set predictions using the XGBoost model.



**Figure S5:** SHAP analysis summary plot for the XGBoost model applied on the test sample. Only the features with numerical values are included in the analysis.

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