

*Supplementary Information for*

**Predicting Skin Irritation, Corrosion, and Sensitization Potential of Nanomaterials Using OECD-Validated New Approach Methodologies (NAMs)**

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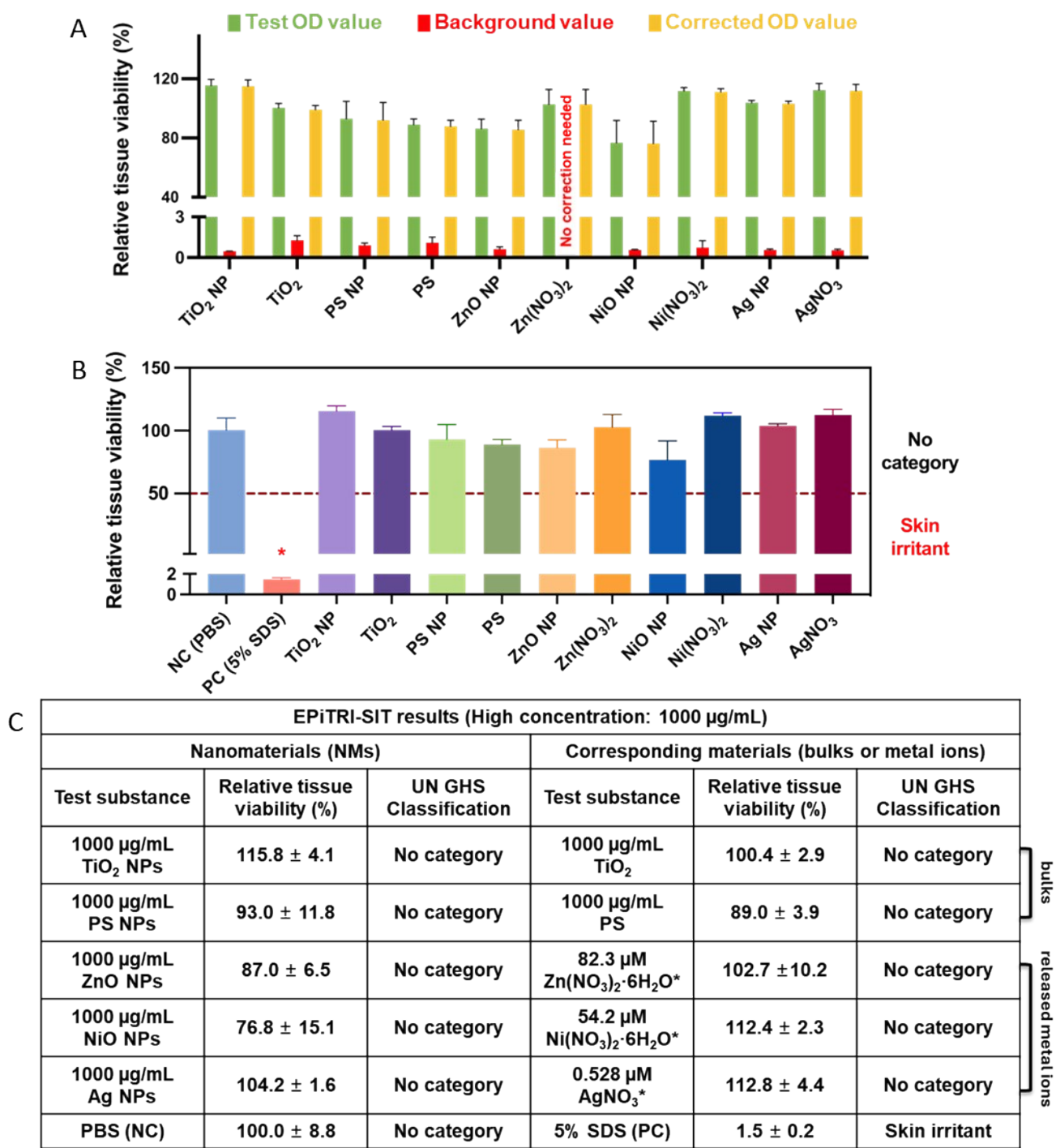
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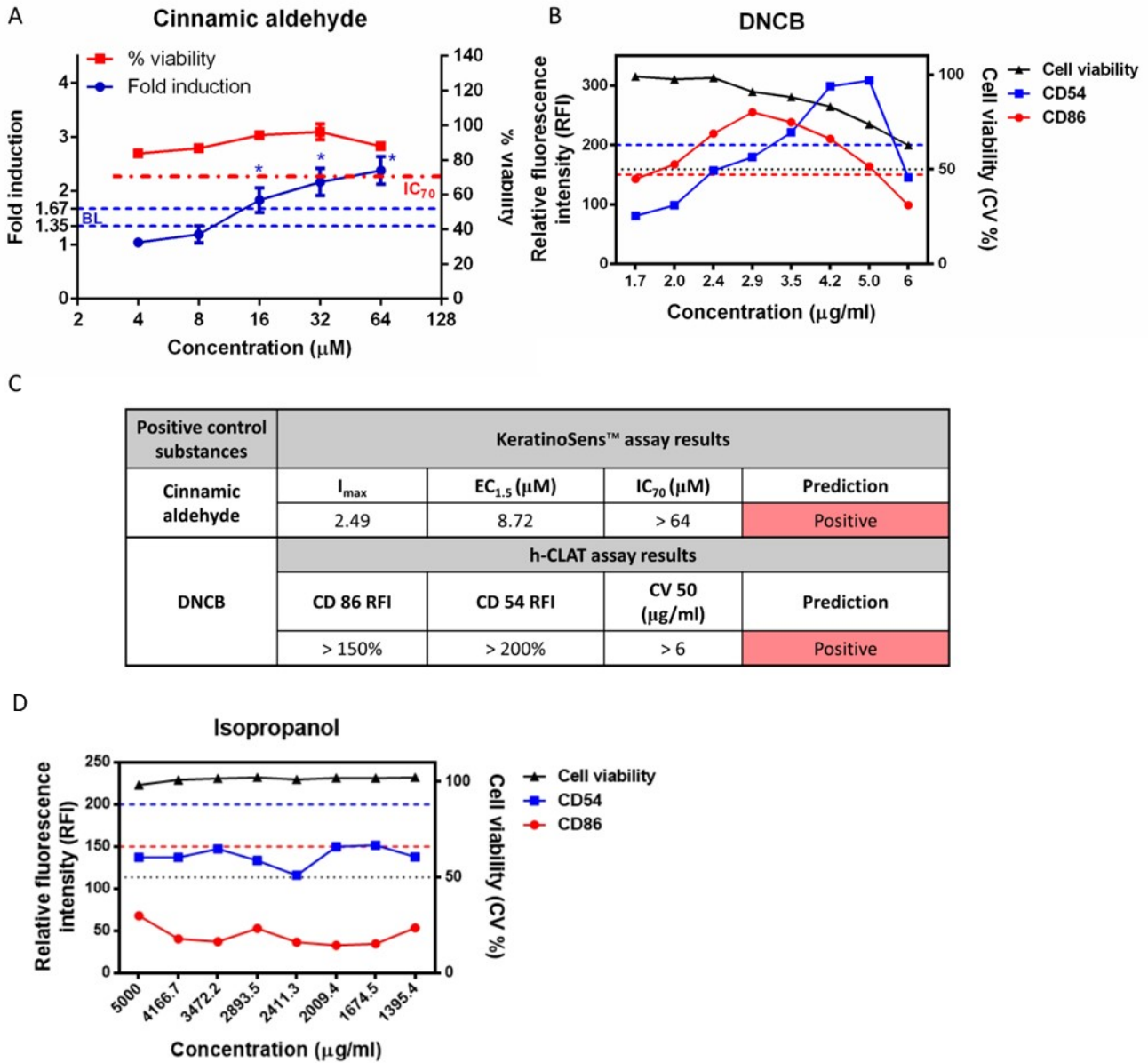
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**Figure S1.** (A) Relative cell viability results from the functional check prior to skin irritation/corrosion tests using the 3D RHE model. Green bars represent the measured OD value, red bars represent the background OD value of each test substance, and yellow bars represent the corrected OD value (green – red). (B) Quantification of relative tissue viability after 30-minute exposure to 1 mg/mL suspensions of the five nanoparticles (NPs) and their corresponding bulk materials or released ions (measured by ICP-MS). (C) Classification of test substances according to OECD TG 439 (bottom-up approach). Mean ± S.D (n ≥ 3); \* *p*-value ≤ 0.05 compared to control.



**Figure S2. KeratinoSens and h-CLAT results of positive control substances.** (A) Dose response curve of luciferase activity induction and cell viability changes of positive control cinnamic aldehyde. (B) Dose response curve of CD86 and CD54 fluorescence induction and cell viability changes of positive control DNCB. (C) Summary results of KeratinoSens™ and h-CLAT assays for positive control substances. (D) Dose response curve of CD86 and CD54 fluorescence induction and cell viability changes of negative control isopropanol.

**Supplementary Table 1. Summary of testing challenges and the corresponding technical modifications in this study.**

| Testing challenges   | Modifications relating to OECD TG   |
|--|---|
| <b>DPRA assay</b>  |   |
| <ul style="list-style-type: none"> <li>• Inorganic particles and poorly soluble metal oxides fall outside the applicability domain of TG 442C.</li> <li>• These materials may not act through classical electrophilic haptentation mechanisms (KE1).</li> <li>• Not expected to form stable covalent adducts with model peptides under DPRA conditions.</li> </ul> | Not used in this study.   |
| <b>KeratiNoSens assay</b>  |   |
| Cell viability interference of NPs in the KeratiNoSens assay   | Calibration for background interference by measuring background absorbance values of NPs            |
| <b>h-CLAT assay</b>  |   |
| Fluorescence interference of NPs in h-CLAT assay   | Applied DiOC18 cell tracer dye in cell staining to separate nanoparticle and cell in flow cytometry |

**Supplementary Table 2. Summary of available *in vivo* and *in vitro* skin sensitization data for the NPs used in this study.**

| Nanoparticles        | Skin sensitization results |  |                       | Ref    |
|----------------------|----------------------------|--|-----------------------|--------|
|                      |                            | Skin sensitization assay   | Prediction            |        |
| PS NPs               | <i>In vivo</i>             | LLNA: SI < 2   | Negative              | [1]    |
| TiO <sub>2</sub> NPs | <i>In vivo</i>             | LLNA: SI < 2   | Negative              | [1]    |
|                      |                            | LLNA: BrdU-FCM assay: SI < 2.7                                     | Negative              | [2]    |
|                      |                            | LLNA: BrdU-FCM assay: SI < 1                                       | Negative              | [3]    |
|                      | <i>In vitro</i>            | KeratinoSens: I <sub>max</sub> = 1.3, IC <sub>50</sub> : >2000 μM  | Negative              | [2, 4] |
|                      |                            | U-SENS: CD86 EC <sub>150</sub> : 183.1 μg/mL                       | Positive              | [5]    |
| ZnO NPs              | <i>In vivo</i>             | LLNA: BrdU-FCM assay: SI < 2                                       | Negative              | [3]    |
|                      |                            | LLNA: BrdU-FCM assay: SI < 2.06                                    | Negative              | [6]    |
|                      | <i>In vitro</i>            | KeratinoSens: I <sub>max</sub> =1.42, IC <sub>50</sub> : 162.64 μM | Negative (borderline) | [4]    |
|                      |                            | h-CLAT: CD86 RFI = 217~360 %, CD54 RFI > 3000%                     | Positive              | [7]    |
| ZnO Z-Cote HP1 NPs   | <i>In vitro</i>            | KeratinoSens: I <sub>max</sub> =5.17                               | Positive              | [8]    |
|                      |                            | h-CLAT: CD86 RFI = 163%, CD54 RFI = 2585%                          | Positive              |        |
| NiO NPs              | <i>In vivo</i>             | LLNA: BrdU-FCM assay: SI < 2.7                                     | Negative              | [2]    |
|                      |                            | LLNA: BrdU-FCM assay: SI < 2                                       | Negative              | [3]    |
|                      | <i>In vitro</i>            | KeratinoSens: I <sub>max</sub> =1.05, IC <sub>50</sub> : >2000 μM  | Negative              | [2, 4] |
| Ag NPs               | <i>In vivo</i>             | OECD TG 406: one animal (1/20) showed patchy erythema              | Positive              | [9]    |
|                      | <i>In vitro</i>            | h-CLAT: CD86 RFI = 156~260 %, CD54 RFI > 234 %                     | Positive              | [7]    |

References:

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