

1 Supplementary material

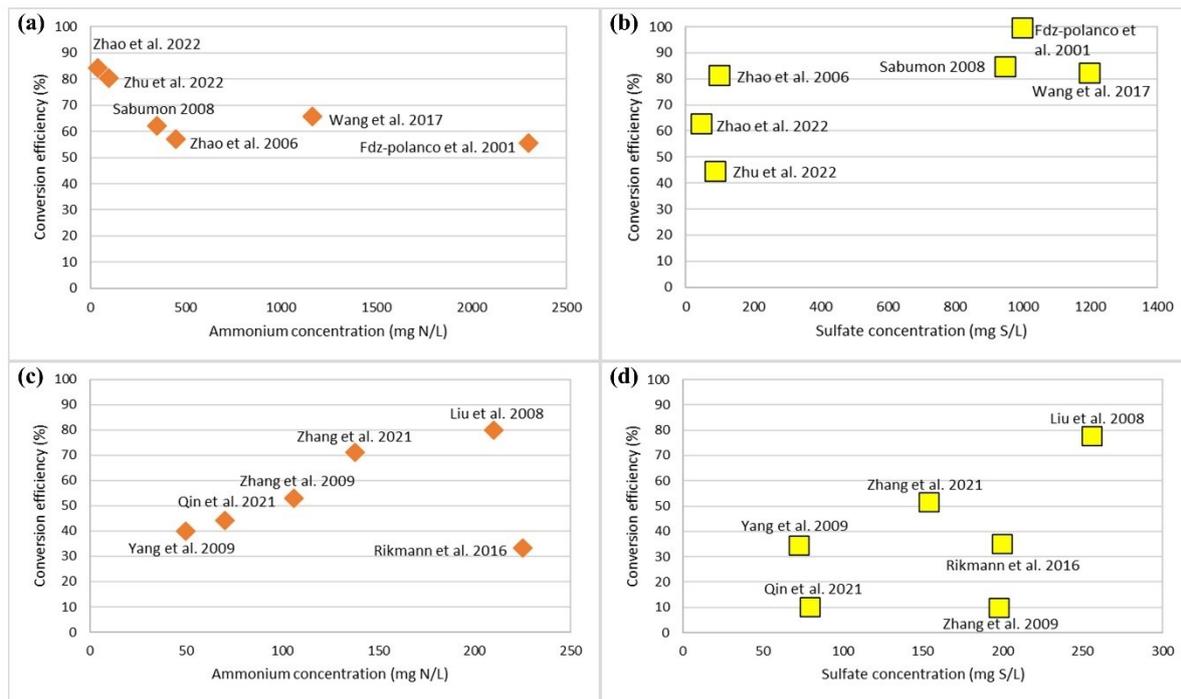
2 S1. Numeric results of the steady-state SRAO reactors

3 Table S1 showed the influent/effluent result of steady-state SRAO reactors, with the end product and conversion parameters. There are totally 12
4 available studies to make this table, with 6 of them under heterotrophic conditions (in orange) and 6 of them under autotrophic conditions (in
5 blue), based on which the figures in Table 1 can be made, also the conversion parameters can be obtained for analysing the stoichiometry and
6 reaction rate of SRAO. The data in green was calculated in this study.

7 Table S1. The steady-state experimental results from long-term SRAO reactors. The cell in green stands for data calculated in this study.

Reference	Influent, mg-N/L, mg-S/L, mg-COD/L				Effluent, mg-N/L, mg-S/L, mg-COD/L								Conversion		
	NH ₄ ⁺	SO ₄ ²⁻	COD	N/S (mol/mol)	NH ₄ ⁺	NO ₃ ⁻	NO ₂ ⁻	N ₂	SO ₄ ²⁻	S ⁰	S ²⁻	COD	□N/□S (mol/mol)	□COD/□S (mg/mgS)	TN removal rate(kg-N/m ³ /d)
(Fdz-Polanco, 2001)	2300	1000	27000	5.26	1025	N.A.	N.A.	1275	5	795	200	1950	2.9	8.4	0.15
(Wang et al., 2017)	1163	1200	9000	2.22	400	100	50	613	216	290	100	2520	1.8	2.2	0.9
(Zhao et al., 2006)	450	100	1750	10.28	193.5	N.A.	N.A.	225	19	71	10	525	7.2	5.0	0.02
(Sabumon, 2008)	350	950	3750	0.81	133	N.A.	N.A.	217	147	N.A.	N.A.	1618	0.6	0.9	0.11
(Zhu et al., 2022)	97	90	400	2.47	19	12	8	57	50	N.A.	N.A.	50	4.5	2.9	0.04
(Rikmann et al., 2016)	225	200	N.A.	2.6	150	N.A.	N.A.	N.A.	130	N.A.	N.A.		2.45	N.A.	0.08
(Liu et al., 2008)	210	256	N.A.	1.9	42	21	0	147	57.6	N.A.	N.A.		1.94	N.A.	0.67
(Madani et al., 2021)	138	154	N.A.	2.1	40	25	10	63	75	N.A.	N.A.		2.8	N.A.	0.1
(Zhang et al., 2009)	106	198	N.A.	1.9	50	<1	<1	56	179	N.A.	<1		0.59	N.A.	0.04
(Qin et al., 2021)	70	80	N.A.	2.0	39	5	2	24	72	N.A.	2		8.9	N.A.	0.05
(Yang et al., 2009)	50	73	N.A.	2.3	30	N.A.	N.A.	16	48	7	N.A.		1.83	N.A.	0.01

9 S2. Steady-state conversion efficiency of ammonium and sulfate



10

11 Fig. S1. Effect of influent substrate concentration on the conversion efficiency of ammonium (left) and sulfate (right), under
 12 heterotrophic conditions (top) and autotrophic condition (bottom) in different studies

13 To investigate the effect of influent concentration on the removal capacity under heterotrophic
 14 condition, Fig. S1 (a)(b) illustrates the relation between influent substrate concentration and
 15 the corresponding conversion efficiency. In the analysed literatures (the environmental
 16 conditions were similar), the ammonium conversion efficiencies were found close to each
 17 other, which seemed irrelevant to the influent ammonium concentration. As for the
 18 conversion amount of sulfate, few conclusions can be drawn due to the coexistence of
 19 heterotrophic sulfate reduction.

20

21 Regarding to autotrophic conditions as shown in Fig. S1(c)(d), it is observed that as the
 22 influent ammonium concentration increases, the conversion efficiency exhibits an increase,
 23 which differs from the case under organic carbon conditions. The reason might be the
 24 enrichment of functional biomass, as autotrophic biomass is more likely to be enriched in

25 high-concentration conditions. Conversely, no distinct pattern is observed for sulfate

26 conversion.

27 S3. Calculation process of the overall pathways

28 Table S2. Detailed calculations for complex autotrophic SRAO reaction 1A (see Figure 1 and Table 3)

Factor	Substrate/product	NH ₄ ⁺	NO ₃ ⁻	NO ₂ ⁻	N ₂	S ²⁻	S ⁰	SO ₄ ²⁻	H ⁺	H ₂ O	Organic-e ⁻	ΔG ₀ (kJ/mol/eq)	ΔG ₀ (kJ/mol/e ⁻)
	Microbial process												
SRAO-N₂-S⁰ reaction, pathway 1: Elementary reaction													
1x	Sulfamox-N ₂ -S ⁰ (1A)	-2			1		1	-1		4		-47.8	-8.0
pathway 2: SRAO -NO₃-S²⁻ + SAD (NO₃; S₂) + SAD (NO₂; S⁰)													
2x	Sulfamox-NO ₃ -S ²⁻ (1D)	-1	1			1		-1	2	1		309.9	38.7
+													
2x	SD (NO ₃ ; S ₂) (2A)		-1	1		-1	1		-2	1		2.2	1.1
+													
1x	SD (NO ₂ ; S ⁰) (2D)			-2	1		-1	1				-670.2	-111.7
pathway 3: SRAO -NO₃-S⁰ + SAD (NO₃; S⁰) + SAD (NO₂; S⁰)													
2/3x	Sulfamox-NO ₃ -S ⁰ (1C)	-3	3				4	-4	-2	7		1302.1	54.3
+													
2/3x	SAD (NO ₃ ; S ⁰) (2B)		-3	3			-1	1	2	-1		-211.6	-35.3
+													
1x	SAD (NO ₂ ; S ⁰) (2D)			-2	1		-1	1				-670.2	-111.7
pathway 4: SRAO -NO₂-S⁰ + SAD (NO₂; S⁰)													
2x	Sulfamox-NO ₂ -S ⁰ (1E)	-1		1			1	-1		2		312.4	52.1
+													
1x	SAD (NO ₂ ; S ⁰) (2D)			-2	1		-1	1				-670.2	-111.7
pathway 5: SRAO -NO₂-S⁰ + nitrite anammox													
1x	Sulfamox-NO ₂ -S ⁰ (1E)	-1		1			1	-1		2		312.4	52.1
+													
1x	Anammox (4)	-1		-1	1					2		-357.5	-119.2
pathway 6: SRAO -NO₃-S²⁻ + SAD (NO₃; S⁰) + SAD (NO₂; S⁰)													

1x	Sulfamox-NO ₃ -S ²⁻ (1D)	-1	1			1		-1	2	1		309.9	38.7
+													
1x	SAD (NO ₃ ; S ₂) (2A)		-1	1		-1	1		-2	1		2.2	1.1
+													
1x	Anammox (4)	-1		-1	1						2	-357.5	-119.2
pathway 7: SRAO -NO₃-S⁰ + SAD (NO₃; S⁰) + nitrite anammox													
1/3x	Sulfamox-NO ₃ -S ⁰ (1C)	-3	3				4	-4	-2	7		1302.1	54.3
+													
1/3x	SAD (NO ₃ ; S ⁰) (2B)		-3	3		-1	1	2	-1			-364.9	-60.8
+													
1x	Anammox (4)	-1		-1	1						2	-357.5	-119.2

29

Factor	Substrate/product	NH ₄ ⁺	NO ₃ ⁻	NO ₂ ⁻	N ₂	S ²⁻	S ⁰	SO ₄ ²⁻	H ⁺	H ₂ O	Organic-e ⁻	ΔG ₀ (kJ/mol/eq)	ΔG ₀ (kJ/mol/e ⁻)
Microbial process													
SRAO -N₂-S²⁻ reaction, pathway 1: Elementary reaction													
1x	Sulfamox-N ₂ -S ²⁻ (1B)	-8			4	3		-3	8	12		-551.9	-23.0
SRAO -N₂-S²⁻ reaction, pathway 2: Sulfamox-NO₂-S²⁻ + Nitrite anammox													
1x	Sulfamox-NO ₂ -S ²⁻ (1F)	-4		4		3		-3	8	4		878.2	36.6
+													
4x	Anammox (4)	-1		-1	1						2	-357.5	-119.2

30 Table S3. Calculation process of 1B

31

Factor	Substrate/product Microbial process	NH ₄ ⁺	NO ₃ ⁻	NO ₂ ⁻	N ₂	S ²⁻	S ⁰	SO ₄ ²⁻	H ⁺	H ₂ O	Organic-c	CO ₂	ΔG ₀ (kJ/mol/eq)	ΔG ₀ (kJ/mol/e ⁻)
		SRAO with organic carbon, pathway 1: SRAO -NO ₃ ⁻ -S ²⁻ + Hetero-denitrification (NO ₃ ⁻) + Hetero-denitrification (NO ₂ ⁻)												
2X	SRAO -NO ₃ ⁻ -S ²⁻ (1D)	-1	1			1		-1	2	1			309.9	38.7
+	Hetero-denitrification (NO ₃ ⁻) (3A)		-2	2						4/3(methanol); 1(acetate); 1(glucose)	-2/3(methanol); -1/2(acetate); -1/6(glucose)	2/3(methanol); 1(acetate); 1/6(glucose)	-314.1(methanol); -298.7(acetate); -330.8(glucose)	-78.5(methanol); -74.7(acetate); -82.7(glucose)
+	Hetero-denitrification (NO ₂ ⁻) (3B)			-2	1				-2	3(methanol); 5/2(acetate); 5/2(glucose)	-1(methanol); -6/8(acetate); -6/24(glucose)	1(methanol); 12/8(acetate); 36/24(glucose)	-776.4(methanol); -753.4(acetate); -801.6(glucose)	-129.4(methanol); -125.6(acetate); -133.6(glucose)
=	Hetero-SRAO-1	-2			1	2		-2	2	19/3(methanol); 11/2(acetate); 11/2(glucose)	5/3(methanol); 20/8(acetate); 10/4(glucose)	-5/3(methanol); 10/8(acetate); 5/12(glucose)	-470.7(methanol); -432.3(acetate); -512.6(glucose)	-47.1(methanol); -43.2(acetate); -51.3(glucose)
SRAO with organic carbon, pathway 2: SRAO -NO ₂ ⁻ -S ²⁻ + Hetero-denitrification (NO ₂ ⁻)														
2X	SRAO -NO ₂ ⁻ -S ²⁻ (1F)	-4		4		3		-3	8	4			878.2	36.6
+	Hetero-denitrification (NO ₂ ⁻) (3B)			-2	1				-2	3(methanol); 5/2(acetate); 5/2(glucose)	-1(methanol); -6/8(acetate); -6/24(glucose)	1(methanol); 12/8(acetate); 36/24(glucose)	-776.4(methanol); -753.4(acetate); -801.6(glucose)	-129.4(methanol); -125.6(acetate); -133.6(glucose)
=	Hetero-SRAO-2	-4			2	3		-3	4	10(methanol); 9(acetate); 9(glucose)	2(methanol); 3(acetate); 3(glucose)	-2(methanol); 6/4(acetate); 1/2(glucose)	-674.6(methanol); -628.6(acetate); -725.0(glucose)	-52.2(methanol); -52.4(acetate); -60.4(glucose)
SRAO with organic carbon, pathway 3: SRAO -NO ₃ ⁻ -S ⁰ + Hetero-denitrification (NO ₃ ⁻) + Hetero-denitrification (NO ₂ ⁻)														
2X	SRAO -NO ₃ ⁻ -S ⁰ (1C)	-3	3				4	-4	-2	7			1302.1	54.3
+	Hetero-denitrification (NO ₃ ⁻) (3A)			-2	2						4/3(methanol);1(acetate);1(glucose)	-2/3(methanol); -1/2(acetate); -1/6(glucose)	2/3(methanol); 1(acetate); 1/6(glucose)	-314.1(methanol); -298.7(acetate); -330.8(glucose)
+	Hetero-denitrification (NO ₂ ⁻) (3B)				-2	1				-2	3(methanol);5/2(acetate);5/2(glucose)	-1(methanol); -6/8(acetate); -6/24(glucose)	1(methanol); 12/8(acetate); 36/24(glucose)	-776.4(methanol); -753.4(acetate); -801.6(glucose)
=	Hetero-SRAO-3	-6			3		8	-8	-10	27(methanol); 49/2(acetate); 49/2(glucose)	5(methanol); 60/8(acetate); 30/4(glucose)	-5(methanol); 30/8(acetate); 5/4(glucose)	-667.3(methanol); -552.1(acetate); -793.0(glucose)	-22.0(methanol); -18.0(acetate); -26.4(glucose)
SRAO with organic carbon, pathway 4: SRAO -NO ₂ ⁻ -S ⁰ + Hetero-denitrification (NO ₂ ⁻)														
2X	SRAO -NO ₂ ⁻ -S ⁰ (1E)	-1		1			1	-1		2			312.4	52.1
+	Hetero-denitrification (NO ₂ ⁻) (3B)				-2	1				-2	3(methanol);5/2(acetate);5/2(glucose)	-1(methanol); -6/8(acetate); -6/24(glucose)	1(methanol); 12/8(acetate); 36/24(glucose)	-776.4(methanol); -753.4(acetate); -801.6(glucose)
=	Hetero-SRAO-4	-2			1		2	-2	-2	7(methanol); 13/2(acetate); 13/2(glucose)	1(methanol); 12/8(acetate); 6/4(glucose)	6/8(acetate); 1/4(glucose)	-151.6(methanol); -128.6(acetate); -176.8(glucose)	-25.2(methanol); -21.4(acetate); -29.5(glucose)

32 Table S4. Calculation process of the complex heterotrophic SRAO pathways, for various organic carbon sources (methanol, acetate, glucose)

34 S4. Operation conditions of the SRAO reactors in the literature

Reactor specifications and initial intention	Acclimation time (days)	Seed sludge	Reference
Heterotrophic			
EGSB, 1.96L, HRT=21h. Anaerobic digestion	52	Anaerobic fermentation sludge	(Wang et al. 2017)
Anaerobic fluidized bed reactor, 1.5L. Anaerobic digestion	40	Anaerobic tank sludge from yeast factory	(Fdz-polanco. et al. 2001)
Up-flow hybrid reactor, 1.75L, HRT=2d Sulfamox with organic carbon	60	Activated sludge from tannery sewage, VSS/TSS=0.58	(Sabumon 2008)
Anaerobic attached-growth reactor, 3.8L Sulfamox with organic carbon	156	Anaerobic sludge from sulfate-rich sewage plant	(Zhao et al. 2006)
Sequencing batch reactor, 250mL, cycle time=48h. Sulfamox with organic carbon	90	Anammox mixed with anaerobic tank sludge, TSS=2.2 g/L	(Zhu et al. 2022a)
Autotrophic			
Non-woven rotating biological contactor, 1.7L, HRT=6h. Anammox	45	Anammox sludge, VSS=0.32 g/L	(Liu et al. 2008)
Sequencing batch reactor, 1.5L, cycle time=108.25h. Sulfamox with inorganic carbon	60	Anaerobic digestion sludge from municipal sewage plant, VSS=15 g/L	(Zhang et al. 2009)
Self-designed mixed reactor, 5L, HRT=1B. Sulfamox with inorganic carbon	61	Activated sludge from municipal sewage plant, VSS=3.5 g/L	(Zhang et al., 2019)
UASB, 0.75L, HRT=1B. Sulfamox with inorganic carbon	50	Anaerobic sludge from yeast factory	(Rikmann et al., 2014)
UASB, 3.93L, HRT=1.5d. Sulfamox with inorganic carbon	60	Nitrifying sludge from municipal plant, VSS=1.56 g/L	(Yang et al. 2009)
UASB, 10L, HRT=16h. Sulfamox with inorganic carbon	158	Anammox sludge, VSS=1.4 g/L	(Qin et al. 2021)
Up-flow anaerobic reactor, 1.7L, HRT=12-24h. Sulfamox in mature leachate treatment	23	Anammox sludge, VSS=1.78 g/L	(Zhan et al. 2023)

35 Table S5. Reactor operation conditions of the literature

37 S5. Gibbs free energy calculation

38

39 The standard Gibbs free energy of formation (ΔG_0) has been widely used for checking the
 40 thermodynamical feasibility of a given chemical reaction. The defined condition was set as
 41 298.15 K (25°C), 1 atm. The involved substances in this study were listed in Table S6.

42

43 Table S6. Standard Gibbs free energy of formation under standard conditions (Rudolf K.
 44 Thauer 1977)

Substrate	NH ₄ ⁺	NO ₃ ⁻	NO ₂ ⁻	N ₂	S ²⁻	S ⁰	SO ₄ ²⁻	H ⁺	H ₂ O	Methanol	Glucose	Acetate	CO ₂
Standard - ΔG_0 (kJ/mol)	79.4	111.3	37.2	0	85.8	0	744.6	0	237.2	175.4	915.4	369.4	394.4

45

$$46 \Delta G_0 = \left[\sum Y_{product} \Delta G_{0_{product}} \right] - \left[\sum Y_{substrate} \Delta G_{0_{substrate}} \right]$$

47 Where Y presents for stoichiometric coefficient of substrate and product; $\Delta G_{0_{product}}$ and
 48 $\Delta G_{0_{substrate}}$ can be found in Table S6. If $\Delta G_0 < 0$, then the equation can be marked as
 49 thermodynamically feasible.

50

51 However, the standard Gibbs free energy of formation considers H⁺ has an activity of 1
 52 mol/kg, which is in equivalence to pH=0. For biochemical reactions, the proton concentration
 53 should be corrected to a proper value of physiological conditions, which is normally taken by
 54 pH=7. the corrected standard Gibbs free energy of formation ΔG_0^* can be calculated by the
 55 following equation (Kleerebezem and Van Loosdrecht, 2010).

$$56 \Delta G_0^* = \Delta G_0 + R \cdot T \cdot m_H \cdot \ln [H^+]$$

57

58 Where ΔG_0 is the standard Gibbs free energy of formation; R is gas constant, 8.314 J/mol/K;
59 T is thermodynamic temperature, K; $[H^+]$ is the concentration of proton, here it is 10^{-7} mol/kg.
60 By correcting the standard Gibbs free energy of formation, the thermodynamical feasibility of
61 microbial reactions can be better checked.

62

63 In addition to the standard Gibbs free energy of formation, it is worth noting that the actual
64 Gibbs free energy is more dynamic, driven by the concentration difference between the
65 substrates and products, as shown in the following equation.

66 Given a reaction equation, $aA + bB \rightleftharpoons cC + dD$, the Gibbs free energy is calculated by

$$\Delta G = \Delta G_0 + R \cdot T \cdot \ln \frac{[C]^c [D]^d}{[A]^a [B]^b}$$

67
68 Where $[A][B]$ and $[C][D]$ are the concentrations of the substrates and products, respectively.

69 It literally means that by introducing more substrates into the system to broaden the gap
70 between the concentrations of substrates and products, the thermodynamical feasibility of the
71 reaction can be improved.

72

73 The actual Gibbs free energy was able to be checked by making the assumption that the
74 concentration of substrates, i.e., $[NH_4^+]$ and $[SO_4^{2-}]$ equals to 100 mM, the concentration of
75 the substrates are 5 times greater than the product. The actual Gibbs free energy can be
76 derived as a function of nitrate/nitrite byproduct rate. As a result, the thermodynamics of the
77 SRAO reactions can be double checked via standard Gibbs free energy and actual Gibbs free
78 energy.

79

80 As shown in Fig. S2, with the increase of nitrate/nitrite byproduct proportion (f), the ΔG and
81 ΔG_0 value increases accordingly. At some point, it will turn from negative to positive, which
82 means it will become thermodynamically unfavourable. The turning point varies among the

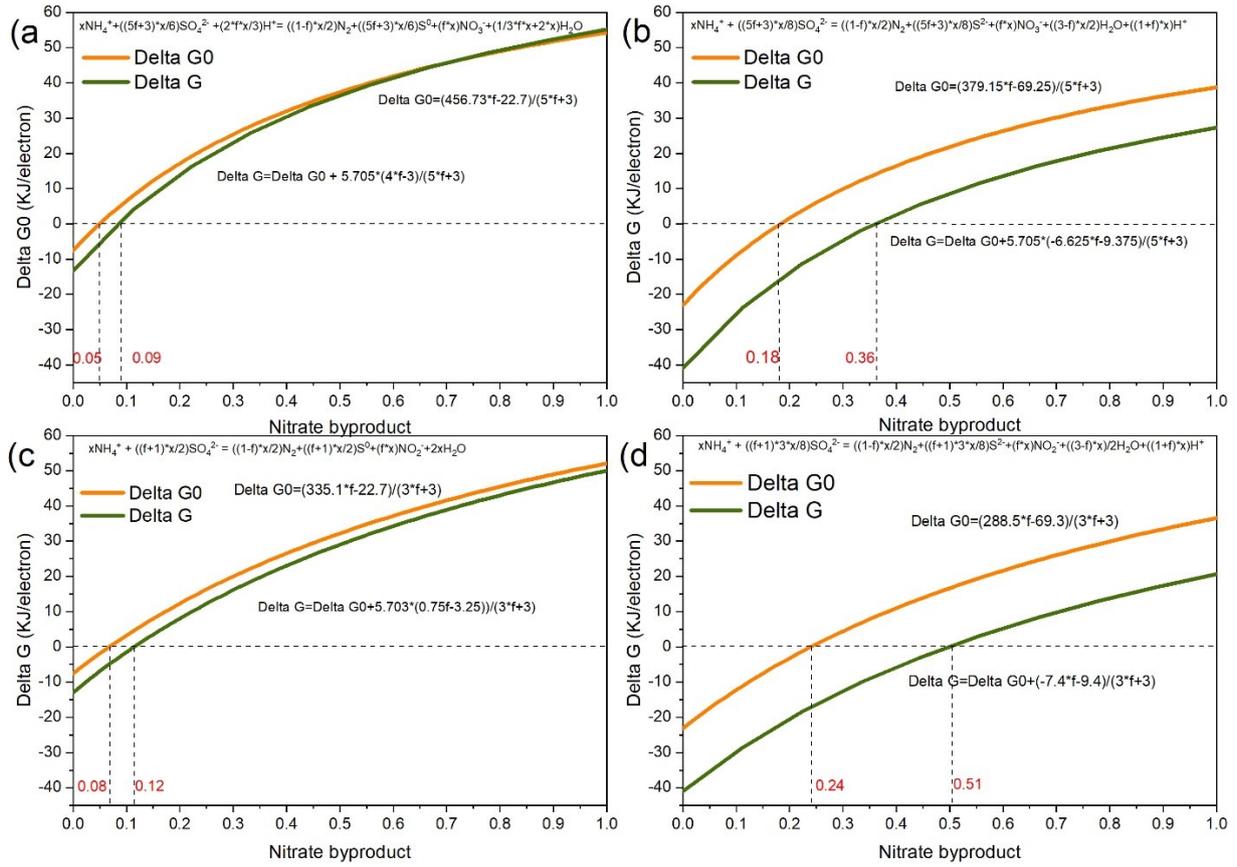
83 reactions. For SRAO reactions with elemental sulfur as end products (Fig. S2a and S2c), it
84 only allows 5-9% of nitrate, or 8-12% of nitrite existed as byproduct, whereas for SRAO
85 reactions with sulfide as end products (Fig. S2b and S2d), 18-36% of nitrate or 24-51% of
86 nitrite can be produced as byproducts. This result is consistent with the observation of nitrite
87 and nitrate in the previous experiments, but it is not able to pick out which reactions are
88 taking place in reality out of the infinite possibilities.

89

90 Overall, SRAO reactions with sulfide as end products tend to be more thermodynamically
91 favourable than that with elemental sulfur as end products, which indicates that from a
92 thermodynamic perspective elemental sulfur might not be the main product of SRAO
93 reactions, as usually stated in previous studies. The ΔG and ΔG_0 of SRAO reactions with
94 sulfide as end products can reach -20 to -40 KJ/electron, while the SRAO reactions with
95 elemental sulfur as end products can barely approach a negative Gibbs free energy change.

96

97 Due to the low efficiency of SRAO reactions, the substrate concentrations were usually 2-5
98 times of the products concentrations (Table S1), making the actual ΔG always more negative
99 than ΔG_0 (Fig.1). Therefore, in reality the thermodynamic potential of SRAO reactions can
100 be considered stronger than the standard value ΔG_0 . All those SRAO reactions that have
101 $\Delta G_0 < 0$ are thereby thermodynamic feasible.



102

103 Fig. S2. Gibbs free energy change (KJ/electron) as a function of nitrite/nitrate byproduct proportion: (a)
 104 SRAO:N₂-NO₃--S₀; (b) SRAO:N₂-NO₃--S₂⁻; (c) SRAO:N₂-NO₂--S₀; (d) SRAO:N₂-NO₂--S₂⁻. The
 105 calculations were performed and adjusted to physiological conditions (298.15 K, 1 atm, pH=7).

106

107 S6. Methodology to obtain biomass concentration in Table 8

108 Generally, the reaction rate should be calculated by following equation:

$$109 \text{ Reaction rate (mg/g VSS/h)} = \frac{\text{Substrate consumption (mg)}}{\text{Time (h)} * \text{Biomass (g VSS/L)}}$$

110 However, the biomass concentration is missing in some of the studies, which needs to be
 111 calculated first. After that the reaction rate can be obtained.

112 For Prachakittikul et al. (2016): the maximum reaction rate was already indicated in the
 113 paper as 0.102 g N/g VSS/d and 0.574 g S/g VSS/d at 8 hour. Therefore, the biomass can be
 114 reversely calculated as 1.47 g.

115 For Zhang et al (2023): the VSS concentration was given as 3173 mg/L and the
116 inoculated volume of the batch was 1.5L. Thus, the biomass can be calculated as
117 $3.173 \times 1.5 = 5.595$ g.

118 For Liu et al. (2008): the VSS concentration was given as 0.0544 g/L, and the inoculated
119 volume of the batch was 100 mL. Thus, the biomass can be calculated as $0.0544 \times 100 \times 10^{-3} = 0.00544$ g.

121 For Lin et al. (2022): the VSS concentration was given as 2.654 g/L, and the inoculated
122 volume of the batch was 250 mL. Thus, the biomass can be calculated as $2.654 \times 250 \times 10^{-3} = 0.6635$ g.

124 For Zhan et al (2023): neither biomass nor reaction rate in the batch test was given.
125 However, the batch test was inoculated by sludge from continuous reactor, where the biomass
126 was 1.78 g VSS/L. The inoculated volume was 200 mL. Thus, biomass in the batch test could
127 be calculated as $1.78 \times 0.2 = 0.356$ g, after which the reaction rate can be calculated.

128 For Zhu et al. (2022b): neither biomass nor reaction rate of sulfamox phenomenon
129 batch test was given. However, according to another experiment with same incubation in the
130 paper, which was given as “NO₃⁻-N dropped rapidly from 80 mg/L to 72 mg/L in 30 min with
131 a consumption rate of 17.53 mg N/(g VSS · h)”, the biomass amount could be reversely
132 calculated as 0.91 g.

133 For Wang et al. (2017): although neither VSS nor reaction rate was given, the dataset of
134 this paper was found from Master Thesis Database of Nanjing University. The VSS of the
135 granular sludge used this study was 21.073 g/L, and the inoculated volume of the batch was
136 0.1L. Thus, the biomass was $21.073 \times 0.1 = 2.1073$ g.

137
138

139 **Reference**

- 140 Fdz-Polanco, F., 2001. New process for simultaneous removal of nitrogen and sulphur under
141 anaerobic conditions. *Water Research* 35, 1111–1114. <https://doi.org/10.1016/S0043->
142 1354(00)00474-7
- 143 Kleerebezem, R., Van Loosdrecht, M.C.M., 2010. A Generalized Method for Thermodynamic
144 State Analysis of Environmental Systems. *Critical Reviews in Environmental Science*
145 and *Technology* 40, 1–54. <https://doi.org/10.1080/10643380802000974>
- 146 Liu, S., Yang, F., Gong, Z., Meng, F., Chen, H., Xue, Y., Furukawa, K., 2008. Application of
147 anaerobic ammonium-oxidizing consortium to achieve completely autotrophic
148 ammonium and sulfate removal. *Bioresource Technology* 99, 6817–6825.
149 <https://doi.org/10.1016/j.biortech.2008.01.054>
- 150 Madani, R.M., Liang, J., Cui, L., Zhang, D., Otitoju, T.A., Elsalahi, R.H., Song, X., 2021.
151 Novel simultaneous anaerobic ammonium and sulfate removal process: A review.
152 *Environmental Technology & Innovation* 23, 101661.
153 <https://doi.org/10.1016/j.eti.2021.101661>
- 154 Qin, Y., Wei, Q., Zhang, Y., Li, H., Jiang, Y., Zheng, J., 2021. Nitrogen removal from
155 ammonium- and sulfate-rich wastewater in an upflow anaerobic sludge bed reactor:
156 performance and microbial community structure. *Ecotoxicology* 30, 1719–1730.
157 <https://doi.org/10.1007/s10646-020-02333-x>
- 158 Rikmann, E., Zekker, I., Tomingas, M., Tenno, T., Loorits, L., Vabamäe, P., Mandel, A.,
159 Raudkivi, M., Daija, L., Kroon, K., Tenno, T., 2016. Sulfate-reducing anammox for
160 sulfate and nitrogen containing wastewaters. *Desalination and Water Treatment* 57,
161 3132–3141. <https://doi.org/10.1080/19443994.2014.984339>
- 162 Rikmann, E., Zekker, I., Tomingas, M., Vabamäe, P., Kroon, K., Saluste, A., Tenno, Taavo,
163 Menert, A., Loorits, L., dC Rubin, S.S.C., Tenno, Toomas, 2014. Comparison of
164 sulfate-reducing and conventional Anammox upflow anaerobic sludge blanket
165 reactors. *Journal of Bioscience and Bioengineering* 118, 426–433.
166 <https://doi.org/10.1016/j.jbiosc.2014.03.012>
- 167 Sabumon, P.C., 2008. Development of a novel process for anoxic ammonia removal with
168 sulphidogenesis. *Process Biochemistry* 43, 984–991.
169 <https://doi.org/10.1016/j.procbio.2008.05.004>
- 170 Wang, D., Liu, B., Ding, X., Sun, X., Liang, Z., Sheng, S., Du, L., 2017. Performance
171 evaluation and microbial community analysis of the function and fate of ammonia in a
172 sulfate-reducing EGSB reactor. *Appl Microbiol Biotechnol* 101, 7729–7739.
173 <https://doi.org/10.1007/s00253-017-8514-z>
- 174 Yang, Z., Zhou, S., Sun, Y., 2009. Start-up of simultaneous removal of ammonium and
175 sulfate from an anaerobic ammonium oxidation (anammox) process in an anaerobic
176 up-flow bioreactor. *Journal of Hazardous Materials* 169, 113–118.
177 <https://doi.org/10.1016/j.jhazmat.2009.03.067>
- 178 Zhang, D., Cui, L., Wang, H., Liang, J., 2019. Study of sulfate-reducing ammonium oxidation
179 process and its microbial community composition. *Water Science and Technology* 79,
180 137–144. <https://doi.org/10.2166/wst.2019.027>
- 181 Zhang, L., Zheng, P., He, Y., Jin, R., 2009. Performance of sulfate-dependent anaerobic
182 ammonium oxidation. *Sci. China Ser. B-Chem.* 52, 86–92.
183 <https://doi.org/10.1007/s11426-008-0161-x>
- 184 Zhao, Q. -l., Li, W., You, S. -j., 2006. Simultaneous removal of ammonium-nitrogen and
185 sulphate from wastewaters with an anaerobic attached-growth bioreactor. *Water*
186 *Science and Technology* 54, 27–35. <https://doi.org/10.2166/wst.2006.762>

187 Zhu, Y., Yang, S., Wang, W., Meng, L., Guo, J., 2022. Applications of Sponge Iron and
188 Effects of Organic Carbon Source on Sulfate-Reducing Ammonium Oxidation
189 Process. IJERPH 19, 2283. <https://doi.org/10.3390/ijerph19042283>
190