Supporting Information

Post-Polymerisation Oxyfunctionalisation of Styrene and Butadiene-Based (Co-)Polymers using a Homogeneous Manganese Catalyst

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1. General

Polybutadiene (100% 1,2-type unsaturated bonds, $M_w = 5.0$ kDa and 80% 1,4-type, and 20% 1,2 unsaturated bonds, $M_w = 10.7/22.8$ kDa), polystyrene ($M_w = 24$ kDa) and styrene-butadiene-styrene block-co-polymer ($M_w = 191/90/16$ kDa, trimodal weight distribution) were acquired from sigma Aldrich. Polystyrene ($M_w = < 364$ kDa, and $M_w = 1.1$ kDa) was acquired from Fischer scientific. The used Mn(TACN) catalyst was kindly supplied by prof.dr. Ronald Hage and CATEXCEL BV. All chemicals were acquired from commercial sources and used as received. *trans*-4-octene, hydrogen peroxide (35% in H₂O) and tetrabutylammonium hydrogensulphate were purchased from Sigma-Aldrich. Dichloromethane, tetrahydrofuran and acetonitrile were acquired from Biosolve. Sodium periodate and sulphuric acid were purchased from VWR Chemicals. All reagents and starting materials were purchased from commercial sources and used from the Cambridge Isotope Laboratory Incorporation (Cambridge, USA) or Sigma-Aldrich and used as received.

NMR spectroscopy (¹H, ¹³C{¹H}, ¹H-¹³C and ¹H-¹⁵N) was conducted using a 400 MHz Varian spectrometer equipped with AutoX probe and Agilent ProTune probe tuning accessory or a 400 MHz Jeol EZCL G spectrometer with a HFX probe. ¹H and ¹³C NMR chemical shift are reported in the standard δ notation of part per million (ppm) and are referenced to a residual peak of the solvent, as determined relative to SiMe4. Infrared spectroscopy was conducted using a PerkinElmer SpectrumTwo FT-IR Spectrometer equipped with an ATR-probe. Peaks are annotated by (w), (m) and (s) to indicate weak, medium and strong signals, respectively.

Thermogravimetric analysis (TGA) measurements were performed on a TA Instruments TGA Q50 Thermographic Analyser under nitrogen atmosphere on a platinum pan and analysed using TA Instruments Universal Analysis 2000 software. The thermal degradation was investigated with approximately 10-20 mg sample, loaded onto the pan and heated from ambient temperature to 700 °C with a rate of 20 °C/min. Modulated Differential Scanning Calorimetry (MDSC) measurements were performed on a TA Instruments Discovery DSC equipped with a TA Instruments Refrigerated Cooling System 90 and analysed with TA Instruments Trios Software (version 5.7.1.74). Tzero low mass aluminium pans were loaded with approximately 3-5 mg of sample and closed with a Tate of 2.00 °C/min to -90 °C. After 10 min at -90 °C, the sample was heated to 80 °C at a rate of 2.00 °C/min. After 10 min at 80 °C, the sample was cooled down to 25 °C. The modulation was set at 1 °C/min.

GPC measurements were recorded on as Waters Alliance e2695 Separations Module equipped with 3 x PLGel mixed-D columns in series, a guard column and a Waters 2489 UV/visible Detector or a Shimadzu (SCI-40 controller module, DGU-203 degassing unit, SIL-40 autosampler, CTO-40C column oven, RID-20A refractive index detector) equipped with an Agilent Technologies PLgel 5µm Guard 50 x 7.5 mm guard column and two Agilent Technologies PLgel 5µm MIXED-D 300 x 7.7 mm columns. All polymer samples were dissolved in the corresponding solvent (THF, chloroform, or DMF) and filtered with a 45 µm PTFE syringe filter prior to the GPC analysis. Refractive index or UV detection at 254 nm was performed and molecular weight determinations were based on calibrations with polystyrene standards (**Section 4**).

2. Oxyfunctionalisation reactions

2.1 Synthesis of mono-oxo-di-acetate Mn(TACN) (2)



This synthesis was based on literature.¹ A 100 mL Schlenk was charged with 1,4,7-trimethyl-1,4,7-triazacyclononane (203.5 mg, 1.12 mmol, 1.7 eq.) whereafter it was dissolved in an ethanol/water (5 mL/0.9 mL) mixture. $Mn(OAc)_2 \cdot 2H_2O$ (180.6 mg, 0.67 mmol, 1 eq.) and NaOAc (400 mg, 4.88 mmol, 7.3 eq.) were added resulting in a dark brown/red solution, which was stirred at ambient temperature for 30 minutes before the addition of approximately 15 drops of $HCIO_4$ using a glass pipette. Then $NaCIO_4$ (600 mg, 4.90 mmol, 7.3 eq.) was added, followed by another 8 drops of $HCIO_4$. The reaction mixture turned more red throughout the addition of the acid. To isolate the compound, the solvent was evaporated in vacuo and a dark red solid was obtained.

¹**H NMR (paramagnetic settings)** (400 MHz, MeCN-d₃) δ 80.61 (s), 70.32 (s), 67.39 (s), 36.89 (s), 35.28 (s), 21.22 (s), -78.18 (s), -90.70 (s), -95.68 (s) ppm.

ATR-IR 621 (m), 1076 (s, br), 1417 (w, br), 1574 (m, br), 1719 (w, br), 2872 (w), 3612 (w) cm⁻¹.

UV-Vis (MeCN): λ_{max} = 487, 524, 723 nm.

ESI-MS (MeCN): (low fragmentation setting) $m/z = 172.1 \{ [tmtacn+H]^+, calc. 172.3 \}, 293.0 \{ [Mn^{III}_2(\mu-O)(\mu-CH_3CO_2)_2(tmtacn)_2]^{2+}, calc 293.3 \}, 285.0 \{ [Mn^{II}(CH_3CO_2)-(tmtacn)]^+, calc. 285.19 \}$



Figure S1: ¹H NMR spectrum with paramagnetic settings of $[Mn_2(\mu-OAc)_2(\mu-O)(tmtacn)_2](ClO_4)_2$ measured in MeCN-d₃ at 25 °C. Peaks were assigned as reported by Hage et al.²



Figure S2: FTIR spectrum of $[Mn_2(\mu-OAc)_2(\mu-O)(tmtacn)_2](ClO_4)_2$.



Figure S3: UV-Vis spectrum of $[Mn_2(\mu-OAc)_2(\mu-O)(tmtacn)_2](ClO_4)_2$ measured in MeCN.



Figure S4: ESI-MS spectrum of $[Mn_2(\mu-OAc)_2(\mu-O)(tmtacn)_2](ClO_4)_2$ measured in MeCN using low fragmentation settings.

2.2 Catalytic oxidation reaction of PS with mono-oxo-di-acetate Mn(TACN) (2)



Oxidation reactions were performed based on literature.³ A 20 mL vial was charged with polystyrene high M_w (48.0 mg, 0.461 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (2.8 mL/1.2 mL) at 30 °C. The $[Mn_2(\mu-OAc)_2(\mu-O)(tmtacn)_2](ClO_4)_2$ (18.8 mg, 0.024 mmol, 5 mol%) was added and the opening of the vial was covered using parafilm. 1 mL of 35% H_2O_2 was added to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour. During the addition, the colour changed to yellow, and the mixture was left to stir for 72 hours at 30 °C. Afterwards, the reaction mixture was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours (Yield = 86%).

¹**H NMR** (400 MHz, 1,1,2,2-tetrachloroethane-d₂) δ 7.13 (s, 1H), 7.07 (s, 1H), 6.98 (s, 1H), 6.63 (s, 0.5H), 6.57 (s, 0.5H), 6.51 (s, 0.5H), 6.42 (s, 0.5H), 3.71 (s), 3.48 (s), 3.12 (s), 2.94 (s), 2.25 (s), 2.18 (s), 2.11 (s), 1.92 (s, 1H), 1.48 (s, 2H) ppm.

ATR-IR 697 (s), 755 (m), 1028 (w), 1452 (m), 1493 (m), 1600 (w), 1744 (w), 1771 (w), 1870 (w), 1947 (w), 2847 (w, br), 2920 (m, br), 3026 (m), 3061 (w), 3083 (w), 3515 (w, br) cm⁻¹.

DSC T_g = 106.85 °C

TGA T_d (90 wt%) = 356.3 °C, $T_{max wt loss}$ = 409.8 °C.



Figure S5: ¹H NMR spectrum of the oxidation of polystyrene using Mn^{III}₂(tmtacn)₂ measured in 1,1,2,2-tetrachloroethane-d2 at 25 °C.



Figure S6: IR spectrum of oxidised polystyrene using Mn^{III}₂(tmtacn)₂.



Figure S7: DSC plot of oxidised polystyrene using $Mn^{III}_2(tmtacn)_2$. Red trace is reversing heat flow. Blue trace is Heat flow. Green trace is non-reversing heat flow.



Figure S8: TGA plot of oxidised polystyrene using Mn^{III}₂(tmtacn)₂.

2.3 Catalytic oxidation reaction of PS with tri-oxo Mn(TACN) (1) 2.3.1 H_2O_2 addition in 1 batch (Table 1, Entry 5)



Oxidation reactions were performed based on literature.³ A 50 mL flask was charged with polystyrene high M_w (480 mg, 4.61 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (1) (187.7 mg, 0.23 mmol, 5 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H₂O₂ was added to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour. During the addition, the colour changed to yellow, and the mixture was left to stir for 72 hours at 30 °C. Afterwards, the reaction mixture was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours. (see table below for functionalisation degree and yields)



Figure S9: ¹H NMR spectrum of the oxidation of polystyrene using Mn(TACN) measured in chloroform-d at 25 °C.



Figure S10: FTIR spectrum of the oxidation of polystyrene using Mn(TACN).

2.3.2 H_2O_2 addition 1 batch 1 mol% and 0.05 mol% catalyst loading (Table 1, Entries 3 & 4)



Figure S11: ¹H NMR spectrum of the oxidation of polystyrene using 1 mol% Mn(TACN) measured in chloroform-d at 25 °C.



Figure S12: ¹H NMR spectrum of the oxidation of polystyrene using 0.05 mol% Mn(TACN) measured in chloroform-d at 25 °C.

2.3.4 H₂O₂ addition portion wise (Table 1, Entries 6, 7 & 8)



A 50 mL flask was charged with polystyrene high M_w (480 mg, 4.61 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (**1**) (187.7 mg, 0.23 mmol, 5 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H_2O_2 was added portion wise (in 2 batches, 3 batches or 10 batches) to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour with 24 or 96 hour time intervals. During the addition, the colour changed to yellow, and the mixture was left to stir at 30 °C. Afterwards, the reaction mixture was washed with brine and DCM, the organics were dried over Na₂SO₄ and the solvent was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours.



Figure S13: ¹H NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 3x addition of H_2O_2 with a 24 h interval measured in chloroform-d at 25 °C.



Figure S14: FTIR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN) and 3x batch wise addition of H_2O_2 with a 24 hour interval.



Figure S15: ¹H NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 10x addition of H_2O_2 with a 24 h interval measured in chloroform-d at 25 °C.



Figure S16: FTIR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN) and 10x batch wise addition of H_2O_2 with a 24 hour interval.





Figure S17: ¹H NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S18: ¹³C NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S19: ${}^{1}H^{13}C$ HMBC NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of $H_{2}O_{2}$ with a 96 h interval measured in chloroform-d at 25 °C.



Figure S20: ${}^{1}H^{13}C$ HSQC NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of $H_{2}O_{2}$ with a 96 h interval measured in chloroform-d at 25 °C.



Figure S21: FTIR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN) and 2x batch wise addition of H_2O_2 with a 96 hour interval.

Table 1, Entry 9



Figure S22: ¹H NMR spectrum of the oxidation of polystyrene using Mn(TACN) measured in chloroform-d at 25 °C.

2.3.5 H₂O₂ addition portion wise (Table 1, Entry 10)



A 50 mL flask was charged with polystyrene low M_w (480 mg, 4.61 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (1) (187.7 mg, 0.23 mmol, 5 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H_2O_2 was added portion wise (in 2 batches) to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour with a 96 hour time interval. During the addition, the colour changed to yellow, and the mixture was left to stir at 30 °C. Afterwards, the reaction mixture was washed with brine and DCM, the organics were dried over Na_2SO_4 and the solvent was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours.



Figure S23: ¹H NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S24: ¹³C NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S25: ¹H¹³C HMBC NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S26: ${}^{1}H^{13}C$ HSQC NMR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN), with 2x addition of H_2O_2 with a 96 h interval measured in chloroform-d at 25 °C.



Figure S27: FTIR spectrum of the oxidation of polystyrene using 5 mol% Mn(TACN) and 2x batch wise addition of H_2O_2 with a 96 hour interval.

2.4 Catalytic oxidation reaction of PBD8020 with tri-oxo Mn(TACN) (1)





A 50 mL flask was charged with polybutadiene (1000 mg, 18.7 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (1) (3.7 mg, 0.005 mmol, 0.03 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H_2O_2 was added to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour. During the addition, the colour changed to yellow, and the mixture was left to stir for 72 hours at 30 °C. Afterwards, the reaction mixture was washed with brine and DCM, the organics were dried over Na_2SO_4 and the solvent was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours. (see table below for functionalisation degree and yields)



Figure S28: ¹H NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), measured in chloroform-d at 25 °C.



Figure S29: ¹³C APT NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), measured in chloroform-d at 25 °C.



Figure S30: ¹H¹³C HMBC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% *Mn*(TACN), measured in chloroform-d at 25 °C.



Figure S31: ¹H¹³C HSQC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% *Mn(TACN), measured in chloroform-d at 25 °C.*

2.4.2 H₂O₂ addition portion wise (Table 4, Entries 4 & 5)



A 50 mL flask was charged with polybutadiene (1000 mg, 18.7 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (1) (3.7 mg, 0.005 mmol, 0.03 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H₂O₂ was added portion wise (in 2 batches or 3 batches) to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour with 24 or 96 hour time intervals. During the addition, the colour changed to yellow, and the mixture was left to stir at 30 °C. Afterwards, the reaction mixture was washed with brine and DCM, the organics were dried over Na₂SO₄ and the solvent was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours.





Figure S32: ¹H NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 3x batchwise addition of H_2O_2 with 24 h time intervals, measured in chloroform-d at 25 °C.



Figure S33: ¹³C NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 3x batchwise addition of H_2O_2 with 24 h time intervals, measured in chloroform-d at 25 °C.



Figure S34: ¹H¹³C HMBC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 3x batchwise addition of H_2O_2 with 24 h time intervals, measured in chloroform-d at 25 °C.



Figure S35: ¹H¹³C HSQC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 3x batchwise addition of H_2O_2 with 24 h time intervals, measured in chloroform-d at 25 °C.



Figure S36: FTIR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 3x batchwise addition of H_2O_2 with 24 h time intervals, measured in chloroform-d at 25 °C.



Figure S37: ¹H NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 2x batchwise addition of H_2O_2 with 96 h time intervals, measured in chloroform-d at 25 °C.



Figure S38: ¹³C NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 2x batchwise addition of H_2O_2 with 96 h time intervals, measured in chloroform-d at 25 °C.



Figure S39: ${}^{1}H^{13}C$ HMBC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 2x batchwise addition of H_2O_2 with 96 h time intervals, measured in chloroform-d at 25 °C.



Figure S40: ¹H¹³C HSQC NMR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 2x batchwise addition of H_2O_2 with 96 h time intervals, measured in chloroform-d at 25 °C.



Figure S41: FTIR spectrum of the oxidation of polybutadiene using 0.03 mol% Mn(TACN), with 2x batchwise addition of H_2O_2 with 96 h time intervals, measured in chloroform-d at 25 °C.





A 50 mL flask was charged with styrene-butadiene-styrene block-co-polymer (1000 mg, 15.8 mmol, 1 eq.) and dissolved in a mixture of DCM/MeCN (8 mL/2 mL) at 30 °C. The MnTACN (1) (128.0 mg, 0.158 mmol, 1 mol%) was added and the opening of the flask was covered using parafilm. 10 mL of 10% H_2O_2 was added to the red-coloured solution containing the catalyst and substrate using a syringe pump in 1 hour. During the addition, the colour changed to yellow, and the mixture was left to stir for 72 hours at 30 °C. Afterwards, the reaction mixture was washed with brine and DCM, the organics were dried over Na_2SO_4 and the solvent was evaporated using a rotary evaporator. The obtained off-white solids were thereafter dried in a vacuum oven for 24 hours (FG_{Total} = 11.4%, Yield recovered material = 91%)



Figure S42: ¹H NMR spectrum of the scaled up oxidation of styrene-butadiene-styrene block-copolymer using 1 mol% Mn(TACN), measured in chloroform-d at 25 °C.



Figure S43: ¹³C NMR spectrum of the scaled up oxidation of styrene-butadiene-styrene block-copolymer using 1 mol% Mn(TACN), measured in chloroform-d at 25 °C.



Figure S44: ¹H¹³C HMBC NMR spectrum of the scaled up oxidation of styrene-butadiene-styrene block-co-polymer using 1 mol% Mn(TACN), measured in chloroform-d at 25 °C.



Figure S45: ¹H¹³C HSQC NMR spectrum of the scaled up oxidation of styrene-butadiene-styrene block-co-polymer using 1 mol% Mn(TACN), measured in chloroform-d at 25 °C.

2.6 Epoxide opening to diol on oxypolybutadiene



To a flask equipped with a stir bar and condenser was added 116.1 mg oxidised polybutadiene in 10 mL THF. To this mixture was added 10 mL 3M H_2SO_4 in H_2O (30 mmol), whereafter the mixture was refluxed at 80 °C for 2 hours. After cooling down the mixture was washed with 10 mL each of DCM and brine, whereafter the organics were dried over Na_2SO_4 . After evaporation of DCM the diol functionalized polybutadiene was isolated as a clear wax (90.5% yield).



Figure S46: ¹H NMR spectrum of the fully diol functionalised polybutadiene, measured in chloroform-d at 25 °C.



Figure S47: ¹³C NMR spectrum of the fully diol functionalised polybutadiene, measured in chloroform-d at 25 °C.



Figure S48: ¹H¹³C HMBC NMR spectrum of the fully diol functionalised polybutadiene, measured in chloroform-d at 25 °C.



Figure S49: ¹H¹³C HSQC NMR spectrum of the fully diol functionalised polybutadiene, measured in chloroform-d at 25 °C.

2.7 Oxidative cleavage of the diol motives on oxypolybutadiene



To a flask equipped with a stir bar was added 46.7 mg diol functionalized polybutadiene in 5 mL DCM. To this mixture was added 57.3 mg (0.27 mmol) NaIO₄ and 11.1 mg (0.03 mmol) tetrabutylammonium hydrogen sulphate in 5 mL H₂O. The resulting mixture was vigorously stirred at ambient temperature for 72 hours, whereafter the mixture was washed several times with DCM and brine. The organics were subsequently dried over Na₂SO₄ and the solvent was evaporated to isolate the oxidatively cleaved aldehyde species as a clear liquid/wax (85.0% yield). For the GPC measurement the solvent was not evaporated prior to the measurement to ensure no low molecular weight fragments would evaporate.



Figure S50: ¹H NMR spectrum of the cleaved diol functionalised polybutadiene to aldehydes, measured in chloroform-d at 25 °C.



Figure S51: ¹³C NMR spectrum of the cleaved diol functionalised polybutadiene to aldehydes, measured in chloroform-d at 25 °C.



Figure S52: ¹H¹³C HMBC NMR spectrum of the cleaved diol functionalised polybutadiene to aldehydes, measured in chloroform-d at 25 °C.



Figure S53: ¹H¹³C HSQC NMR spectrum of the cleaved diol functionalised polybutadiene to aldehydes, measured in chloroform-d at 25 °C.





Figure S54: ¹H NMR spectrum of the reaction mixture after the catalytic oxidation of trans-4octene, measured in chloroform-d at 25 °C.



Figure S55: ¹³C NMR spectrum of the reaction mixture after the catalytic oxidation of trans-4octene, measured in chloroform-d at 25 °C.



Figure S56: ¹H¹³C HMBC NMR spectrum of the reaction mixture after the catalytic oxidation of trans-4-octene, measured in chloroform-d at 25 °C.



Figure S57: ¹H¹³C HSQC NMR spectrum of the reaction mixture after the catalytic oxidation of trans-4-octene, measured in chloroform-d at 25 °C.



Figure S58: ¹H¹H COSY NMR spectrum of the reaction mixture after the catalytic oxidation of trans-4-octene, measured in chloroform-d at 25 °C.

3. DSC and TGA Data



Figure S59: Differential Scanning Calorimetry (DSC) of the unfunctionalised high molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S60: Differential Scanning Calorimetry (DSC) of the unfunctionalised low molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S61: Differential Scanning Calorimetry (DSC) of the unfunctionalised PBD8020. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S62: Differential Scanning Calorimetry (DSC) of the unfunctionalised styrene-butadienestyrene (SBS). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S63: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised high molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S64: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised high molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S65: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised high molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S66: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised low molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S67: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised PBD8020. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S68: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised PBD8020. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S69: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised PBD8020. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S70: Differential Scanning Calorimetry (DSC) of the oxyfunctionalised styrene-butadienestyrene (SBS). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S71: Thermogravimetric Analysis (TGA) of the unfunctionalised high molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S72: Thermogravimetric Analysis (TGA) of the unfunctionalised low molecular weight PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S73: Thermogravimetric Analysis (TGA) of the unfunctionalised PBD8020. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S74: Thermogravimetric Analysis (TGA) of the unfunctionalised styrene-butadiene-styrene (SBS). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S75: Thermogravimetric Analysis (TGA) of oxyfunctionalised high M_w PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S76: Thermogravimetric Analysis (TGA) of oxyfunctionalised high M_w PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S77: Thermogravimetric Analysis (TGA) of oxyfunctionalised high M_w PS (entry 4). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S78: Thermogravimetric Analysis (TGA) of oxyfunctionalised low M_w PS. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S79: Thermogravimetric Analysis (TGA) of oxyfunctionalised PBD820. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S80: Thermogravimetric Analysis (TGA) of oxyfunctionalised PBD820. The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S81: Thermogravimetric Analysis (TGA) of oxyfunctionalised PBD820 (entry 10). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.



Figure S82: Thermogravimetric Analysis (TGA) of oxyfunctionalised styrene-butadiene-styrene (SBS) (entry 12). The Blue trace depicts the normalized total heat flow, the green trace depicts the normalized reversing heat flow, and the red trace depicts the normalized non-reversed heat flow.

4. GPC Data

For the high and mid molecular weight OPS entries and PS starting materials, GPC was measured in DMF using a refractive index detector. All samples were measured in duplo. The data for both starting material and oxidised polymer material is determined using the same calibration, shown below unless shown different.



Figure S83: GPC calibration Plot for the measurements in DMF.

PS High M_w





LabSolutions Analysis Report

<Sample Information>

Sample Name	:PS_High_Mw_ref	Sample ID	:
Data Filename	:PS_High_Mw_ref_MO-1_41020	125_012.lcd	Method Filename :DM F_Defa
Batch Filename Date Acquired	: MO-1.lcb : 10-Apr-25 6:34:02 PM	Date Processed	: 11-Apr-25 10:43:01 AM

<Chromatogram>



Figure S84: GPC traces of the unfunctionalised High M_w PS on two different GPCs running both on DMF.

OxyPS High M_w



Figure S85: GPC trace of the oxyfunctionalised High M_w PS.

OxyPS High M_w



Figure S86: GPC trace of the oxyfunctionalised High M_w PS.

OxyPS High M_w



Figure S87: GPC trace of the oxyfunctionalised High M_w PS.



<Sample Information>

Sample Name Data Filename	:PS_24kDa_Reference_2 :PS_24kDa_Reference_2_MO-2_4282025_004.lcd :MO-21cb		Sample ID Method Filename	: DMF_Defa
Date Acquired	: 28-Apr-25 6:08:22 PM	Date Processed	: 28-Apr-25 6:33:2	3 PM

<Chromatogram>



Figure S88: GPC trace of the unfunctionalised Mid M_w PS.

OxyPS Mid M_w



<Sample Information>

Sample Name	:MOC066	Sample ID	15
Data Filename	:MOC066_MO-2_4282025_002.lcd	Method Filename	: DMF_Default_method.icm
Batch Filename	: MO-2.lcb		
Date Acquired	: 28-Apr-25 5:17:24 PM	Date Processed	: 28-Apr-25 5:42:25 PM



Figure S89: GPC trace of the oxyfunctionalised Mid M_w PS.

For the low molecular weight OPS entry and PS starting material, GPC was measured in THF using a refractive index detector. All samples were measured in duplo. The data for both starting material and oxidised polymer material is determined using the same calibration, shown below.



Figure S90: GPC calibration Plot for the measurements in THF.

PS Low M_w



Figure S91: GPC trace of the unfunctionalised Low M_w PS.

OxyPS Low M_w



Figure S92: GPC trace of the oxyfunctionalised Low M_w PS.

For the OPBD8020 and OSBS entries as well as the PBD8020 and SBS starting material, GPC was measured in chloroform using a refractive index or 254 nm UV detector. All samples were measured in duplo. The data for both starting materials and oxidised polymer materials is determined using the same calibration, shown below.



Figure S93: GPC calibration Plot for the measurements in chloroform.

PBD8020



Figure S94: GPC trace of the unfunctionalised PBD8020.

OPBD8020



Figure S95: GPC trace of the oxyfunctionalised PBD8020.

OPBD8020



Figure S96: GPC trace of the oxyfunctionalised PBD8020.

OPBD8020



Figure S97: GPC trace of the oxyfunctionalised PBD8020.



Figure S98: GPC trace of the unfunctionalised SBS.

OSBS



Figure S99: GPC trace of the oxyfunctionalised SBS.

SBS

Oxidative cleavage of the fully hydroxylated PBD



Figure S100: GPC trace of the oxidatively cleaved fully hydroxylated PBD.

References

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