

Supplementary Methods

***In vitro* digestion and *in vitro* colonic fermentation**

In vitro digestion was performed according to INFOGEST 2.0 method which was described previously^{1, 2}. Ten mL of boiled and cooled tarhana soup was used as the starting material. Cellulose and inulin were also subjected to *in vitro* digestion and colonic fermentation as well as tarhana soups. Five grams of starting material was used for both controls. The entire procedure was performed at a constant temperature of 37°C on a rotator (VWR, Radnor, PA, United States). The oral phase of digestion was performed for 2 minutes at 37°C, during which the sample was mixed with simulated saliva solution containing α -amylase (1,500 U mL⁻¹, Sigma, A0521) and the pH was adjusted to 7. Following the oral phase, the mixture underwent gastric digestion for 2 hours. This phase involved the addition of simulated gastric solution and pepsin enzyme (25,000 U mL⁻¹, Sigma, P6887), maintaining the mixture at 37°C while continuously rotating, with the pH adjusted to 3. The final phase was the intestinal digestion, which included gastric chyme, simulated intestinal fluid electrolyte solution, pancreatin (800 U mL⁻¹, Sigma, P7545), and fresh bile (10 mM), the pH was adjusted to 7.0 and samples were incubated in an incubating rocking platform shaker at 37°C for 2 h. Afterward, the post-intestinal phase liquids were transferred to dialysis tubes and immersed in water as the dialysate for 24 h. The retentate was taken from dialysis tubing for MicroMatrix™ *in vitro* colonic fermentation experiment.

Following simulated salivary, gastric and intestinal digestion, samples were used in triplicate for the *in vitro* colonic fermentation experiment. Cellulose and inulin were used as negative and positive controls, respectively, for *in vitro* digestion and *in vitro* colonic fermentation in triplicate. Digested soups were abbreviated as CFB_SD (Chickpea Flour baker's yeast fermented Soup Digested), CFS_SD (Chickpea Flour Sourdough fermented Soup Digested), EFB_SD (Einkorn Flour baker's yeast fermented Soup Digested), EFS_SD (Einkorn Flour

Sourdough fermented Soup Digested), PPB_SD (Purple Potato flour baker's yeast fermented Soup Digested), PPS_SD (Purple Potato flour Sourdough fermented Soup Digested), WFB_SD (Wheat Flour baker's yeast fermented Soup Digested), WFS_SD (Wheat Flour Sourdough fermented Soup Digested).

Subsequent to *in vitro* digestion, *in vitro* colonic fermentation was conducted based on a previously described protocol ³. Participant recruitment and stool sample collection were approved by the Clinical Research Ethics Committee of the Cork Teaching Hospitals (review reference numbers: ECM 4 (gg) 11/02/20 & ECM 3 (iii) 22/02/2022). Faecal samples were obtained from nine healthy individuals who had no antibiotic usage for last 3 months, no medicine usage and healthy and pooled in equal amounts for use in a MicroMatrix™ *in vitro* benchtop fermentation system, adhering to a protocol outlined previously ⁴. Faecal samples were collected from all individuals in the morning at the Teagasc Moorepark Research Centre. The samples were kept in anaerobic jars in a cold room at 4°C. Within two hours, all samples were transferred to an anaerobic chamber for pooling. Each sample was weighed, and 60 g of each sample was transferred into a stomacher bag and mixed with 400 mL of phosphate buffer. The mixture was then transferred to sterile 50 mL falcon tubes and centrifuged at 4000 g for 25 minutes at 4°C. The supernatant was discarded, and the pellet was washed with phosphate buffer. The falcon tubes were centrifuged again at 4000 g for 25 minutes at 4°C. Following centrifugation, the pellet was dissolved in phosphate buffer containing 25% glycerol. Aliquots were then stored at -80°C until use in MicroMatrix™.

The faecal fermentation medium (FFM) was prepared following the protocol of Fooks and Gibson, 2003 ⁵ with the following composition: tryptone water (2 g/L), yeast extract (2 g/L), cysteine HCl (1 mg/L), bile salts (0.5 g/L), Tween 80 (2 mL/L), hemin (0.05 mg/L, dissolved in three drops of 1 M NaOH), vitamin K1 (10 µL/L), NaCl (100 mg/L), KH₂PO₄ (40 mg/L), K₂HPO₄ (40 mg/L), CaCl₂·6H₂O (40 mg/L), MgSO₄·7H₂O (10 mg/L), and NaHCO₃ (2 mg/L).

The medium was autoclaved at 121°C for 15 minutes and stored at 4°C until use. For the fermentation process, 2 mL of digesta was added to the fecal fermentation medium and fecal slurry mixture (20%). The fermentation was conducted under anaerobic conditions at 37°C, with the pH maintained at 6.8³. The mixture was continuously agitated at 250 rpm using a cassette to ensure homogeneity. Faecal fermentation was performed for 12 hours and samples were collected at baseline (T0) and at the end of fermentation (T12). All samples were collected in 2 mL sterile screw cap tubes. Subsequently, the tubes containing the samples were centrifuged at 16,000 g for 10 minutes at 4°C. Following centrifugation, the pellet was used for DNA extraction to analyse the microbial composition and the supernatant was processed for short-chain fatty acid (SCFA) and branched-chain fatty acid (BCFA) analysis. Following *in vitro* colonic fermentation, faecal samples were labelled with the suffix 'F'; for example, CFB_SDF represents faecal samples fermented with digested chickpea flour baker's yeast fermented soup.

References

1. F. Koc, J. Atzler, A. W. Sahin, E. Arendt, R. P. Ross and C. Stanton, Biscuits for the gut: A symphony of FODMAPs and dietary fibre in gut microbiome for irritable bowel syndrome (IBS) management, *Innovative Food Science & Emerging Technologies*, 2024, **97**.
2. A. Brodkorb, L. Egger, M. Alminger, P. Alvito, R. Assuncao, S. Ballance, T. Bohn, C. Bourlieu-Lacanal, R. Boutrou, F. Carriere, A. Clemente, M. Corredig, D. Dupont, C. Dufour, C. Edwards, M. Golding, S. Karakaya, B. Kirkhus, S. Le Feunteun, U. Lesmes, A. Macierzanka, A. R. Mackie, C. Martins, S. Marze, D. J. McClements, O. Menard, M. Minekus, R. Portmann, C. N. Santos, I. Souchon, R. P. Singh, G. E. Vegarud, M. S. J. Wickham, W. Weitschies and I. Recio, INFOGEST static in vitro simulation of gastrointestinal food digestion, *Nat Protoc*, 2019, **14**, 991-1014.
3. F. Koc, I. Sugrue, K. Murphy, S. Renzetti, M. Noort, R. P. Ross and C. Stanton, The microbiome modulating potential of superheated steam (SHS) treatment of dietary fibres, *Innovative Food Science & Emerging Technologies*, 2022, **80**, 13.
4. M. M. O'Donnell, M. C. Rea, F. Shanahan and R. P. Ross, The Use of a Mini-Bioreactor Fermentation System as a Reproducible, High-Throughput ex vivo Batch Model of the Distal Colon, *Front Microbiol*, 2018, **9**, 1844.
5. L. J. Fooks and G. R. Gibson, Mixed culture fermentation studies on the effects of synbiotics on the human intestinal pathogens *Campylobacter jejuni* and *Escherichia coli*, *Anaerobe*, 2003, **9**, 231-242.