# Supporting Information Selective hydrogenolysis of lignin for sustainable production of 4-

# propylsyringol over hierarchical CoNC catalysts

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#### **1. Experimental section**

#### 1.1 Chemicals and materials

Sodium lignosulfonate was purchased from TCI. Cobalt acetate, zinc acetate dihydrate, potassium acetate and potassium hydroxide were provided by Tianjin Fuchen chemical reagents factory. DMSO-*d6*, Pd/C (5% Pd, contains 40-60% H<sub>2</sub>O) and Ru/C (5% Ru, contains 50% H<sub>2</sub>O) were obtained from Aladdin. Co<sub>3</sub>O<sub>4</sub> and 5,5-dimethyl-1-pyrroline N-oxide (DMPO) were purchased from Macklin. benzyloxybenzene **(a1)**, dibenzyl ether **(a2)** and benzyloxybenzene **(a3)** were provided by Energy Chemical. 2-phenoxy-1-phenethanol **(a4)**, 2-(2-methoxyphenoxy)-1-phenylethanol **(a5)** and 2-(2,6-Dimethoxyphenoxy)-1-phenylethanol **(a6)** were synthesized according to a previously reported method<sup>1</sup> with minor modifications, the synthesis process and structural characterization data are provided in the Supporting Information. The organosolv lignins were separated from the lignocellulosic resources of bagasse, wheat straw, bamboo, corncob, birch, poplar, pine and Chinese fir according to the procedures shown in previous study.<sup>2</sup>

#### 1.2 Characterization of catalyst

The cobalt contents in the CoNC catalysts were determined by an inductively coupled plasma optical emission spectrometer (ICP-OES) Optima 8300. The C, H, N and S contents of CoNC were determined by Vario EL cube (Germany), and the O content was calculated by conservation of mass based on the assumption that the sample only contains Co, C, H, N, S and O.

Nitrogen adsorption-desorption experiments were conducted on the Micromeritics ASAP 2460 instrument to accurately determine the specific surface area and pore properties of catalysts. Prior to testing, the catalysts underwent a degassing process at 120°C for 12 h under a pressure of 133 Pa. Subsequently, the samples were subjected to the static-volumetric method at -196°C on the instrument. The specific surface area was determined utilizing the Brunauer-Emmett-Teller (BET) method, while the pore volume was evaluated through the Barrett-Joyner-Halenda (BJH) method.

The crystal phases of catalysts were examined by the powder X-ray diffraction (XRD) patterns (Bruker D8 Advance) with a Cu Ka radiation. Raman spectra of the CoNC catalysts were recorded on a LabRAM Aramis (HORIBA) Raman Spectroscopy System. Scanning electron microscopy (SEM) images were obtained on a Hitachi FESEM SU 8220 instrument operating at 5 kV. The transmission electron microscope (TEM) and high-resolution transmission electron microscope (HRTEM) images were obtained on the Rigaku JEOL JEM-2100F TEM operated at 200 kV. The aberration-corrected high-angle annular dark-field scanning transmission electron microscopy (AC-HAADF-STEM) tests were performed using JEM-200F (JEOL, Tokyo, Japan). X-ray photoelectron spectroscopy (XPS) was performed on an AXIS SUPRA+ (Kratos) instrument with a monochromate Al K $\alpha$  anode. All binding energy calculations were calibrated using the peak position of C1s at 284.8 eV. The X-ray absorption near edge structure (XANES) and extended X-ray absorption fine structure (EXAFS) data were acquired at RapidXAFS HE Ultra, and energy calibration using Co foil. The raw data were background deducted and Fourier transformed using Athena software, and the data fitted was performed using Demeter software.

NH<sub>3</sub> temperature programmed desorption (NH<sub>3</sub>-TPD) patterns were recorded by a BELCAT II instrument with a thermal conductivity detector (TCD) and a quadrupole mass

spectrometer. In a typical experiment, 100 mg sample was heated at 120°C for 5 h to remove the physisorbed species like water and CO<sub>2</sub>. Then the sample was cooled to 50°C in a He stream, and further flushed with a flow of 5% (v/v) NH<sub>3</sub> diluted with He (total flow rate of 20 mL min<sup>-1</sup>) for 1 h. Afterwards, the sample was flushed with flowing He (20 mL min<sup>-1</sup>) for another 1 h to ensure complete removal of physisorbed species. Then, the samples were heated to 900°C in the He stream at a rate of 10°C min<sup>-1</sup> and the amount of NH<sub>3</sub> desorbed was monitored by a TCD. The mass spectrometer followed the NH<sub>3</sub> desorption by m/z = 17.

Thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC) were performed on a NETZSCH STA 449 F5 instrument. Approximately 5-10 mg of sample was heated from room temperature to 900°C at a rate of 10°C/min in air flow (50 mL/min). The mixtures (CoNC<sub>M4</sub>+Lignin) were prepared by static adsorption experiment method, which will be will be introduced in the following section.

#### 1.3 Lignin depolymerization and product separation

100 mg organosolv lignin, certain amounts of catalyst and 10 mL ethanol were put into a 50 mL stainless autoclave (Labe Scientific Instrument Co., Ltd., Shanghai, China). The air inside the autoclave was replaced with pure hydrogen three times. Then, the autoclave was pressurized to 1.0 MPa using hydrogen and heated to a desired temperature (190-270°C) with a heating rate of 10°C min<sup>-1</sup>. After maintaining at the final temperature for a designated time (2-6 h), the autoclave was rapidly cooled to room temperature and the mixture in reactor was separated by using filtration. The solid fraction, which mainly contained catalyst, was washed with THF three times (4 mL  $\times$  3). Subsequently, the resulting solid fraction was dried at 120°C until no significant weight loss, then it was employed in the next run for investigating its reusability.

The THF wash solution was combined with the filtrate above and diluted to 25 mL for further analysis. 2 mL of the above diluted solution was used for quantitative and qualitative analysis. 200 mL deionized water was then added into the left diluted solution, yielding a flocculent precipitate. This flocculent precipitate composing unconverted or partly-converted lignin, was separated using filtration, so obtained solid was then dried at 80°C under a reduced pressure until constant weight. The recovered lignin was finally obtained and designated as Re-lignin.

#### 1.4 Product analysis

The analysis of monophenols was conducted on a GC-MS-FID instrument (Agilent 7890B/5977A). The NIST Mass Spectral Search Program was employed to conduct qualitatively identification. An FBX-5MS PLUS gas chromatography column (30 m×0.25  $\mu$ m×0.25 mm) was used for product separation with 1.0 mL min<sup>-1</sup> He as the carrier gas. The temperature of injector and detector was set at 280°C. The oven temperature started at 50°C (held for 1 min), then ramped to 250°C at a rate of 10°C min<sup>-1</sup> and held for 6 min. Dimethyl phthalate was used as internal standard for quantitatively analysis. Each experimental was repeated three times and the results were averaged. The conversion of lignin was calculated based on the weight comparison of regenerated lignin (Re-lignin) and raw lignin (eqn.1), the yield of products was determined based on the quality of the volatile products and the raw lignin (eqn.2). The selectivity of 4-PS was calculated through the weight percentage of 4-PS in the total volatile products (eqn.3).

$$Conversion(\%) = \frac{W_O - W_R}{W_O} \times 100\%$$
(1)

$$Yield(wt.\%) = \frac{W_{VP}}{W_O} \times 100\%$$
(2)

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Selectivity(%) = 
$$\frac{W_{4-PS}}{W_{VP}} \times 100\%$$
 (3)

Where  $W_{O}$ ,  $W_{R}$ ,  $W_{4-PS}$  and  $W_{VP}$  were the weights of organosolv lignin, Re-lignin, 4-PS and the volatile products, respectively.

#### 1.5 Lignin characterization

The molecular weight distributions of raw lignin and Re-lignin samples were obtained by a gel permeation chromatography (GPC) using 1.0 mL min<sup>-1</sup> tetrahydrofuran (THF) as the eluent, the oven temperature was set at 35°C. The two-dimensional heteronuclear single quantum correlation nuclear magnetic resonance (2D HSQC NMR) of raw lignin and Re-lignin were recorded on a Bruker NMR spectrometer (Avance III 600 MHz), 80 mg of sample was dissolved in 0.6 mL of DMSO-*d6* (D.99.8%, TMS 0.03%), the spectrometer frequency was 500.19/125.77 Hz, and the spectral width was 5882.4/20752.3 Hz for <sup>1</sup> H/<sup>13</sup> C and the relaxation delay was set at 1.5 s. The <sup>1</sup>H-NMR and <sup>13</sup>C-NMR of samples were recorded on a Bruker NMR spectrometer (Avance III 600 MHz) as well. Fourier transform infrared (FT-IR) analysis of raw lignin and Re-lignin were performed using a Thermo Scientific Nicolet iS10 instrument. All FT-IR spectra were scanned 64 times at a resolution of 4 cm<sup>-1</sup>.

#### 1.6 Static adsorption measurements

The static adsorption experiments were carried out to investigate the diffusion behaviour and absorption properties of lignin over different catalysts according to our previous work<sup>3</sup>. In a typical run, 30 mg lignin and 30 mg catalyst were thoroughly immersed in 30 mL THF, after continuous stirring at room temperature for 24 h, the solid fraction was separated by filtration and washed using fresh THF (20 mL×3). The liquid fraction was combined with that used for solid washing, and then it was concentrated under reduced pressure to remove THF. The

unabsorbed lignin was then obtained as a brown solid. Subsequently, this unabsorbed lignin and a certain amount of internal standard (dimethyl terephthalate) were dissolved into 0.5 mL DMSO-*d6*. The adsorption capacities of different CoNC catalysts for lignin were measured by the areas of 1H NMR at the chemical shift of 10.01 ppm of lignin (phenolic protons)<sup>4</sup> and 8.14 ppm of dimethyl terephthalate (aromatic protons).<sup>5</sup>

#### 1.7 Electron paramagnetic resonance measurements

The electron paramagnetic resonance (EPR) was recorded at 20°C and maintained its operating temperature by circulating water. Spectral scanned with a center field of 3505 G, sweep width of 100 G, modulation amplitude of 2 G, receiver gain of 10 dB. Samples for EPR testing were reacted with water solvent under optimal conditions (230°C, 1 MPa H<sub>2</sub>). 1 mL of reaction solution was collected by real-time sampling and quickly cooled to room temperature in an ice water bath. Subsequently, 0.01 g DMPO was dissolved into the above sample. The resulting mixture was subjected to EPR testing.

#### 1.8 Determination of the adsorption energy of lignin model compounds

Adsorption energy calculations were performed using three model compounds, 2-phenoxy-1phenethanol (**a4**), 2-(2-methoxyphenoxy)-1-phenylethanol (**a5**) and 2-(2,6-Dimethoxyphenoxy)-1-phenylethanol (**a6**), representing the H, G and S units of the lignin structure, respectively. All geometry optimizations were employed using DMol3 program from the DFT calculation via software Materials Studio. The Effective Core Potentials (ECP)<sup>6</sup> were treated in the calculations for Co atom, whereas H, C and O were performed as in the allelectron. The generalized gradient approximation (GGA) with a Perdew-Burke-Ernzerh (PBE) of method as the exchange-correlation functional was achieved.<sup>7</sup> The double numerical plus polarization (DNP) as the basis set was used. The following convergence standard of total energy was 10-5 Ha, the force was 0.002 Ha/Å, and the maximum displacement tolerance was 0.005 Å.

#### 1.9 Catalyst reusability and regeneration

The catalyst after the fourth recycle was regenerated by a water steam activation method. The spent catalyst was heated to 700°C at a speed of 2°C min<sup>-1</sup> in a tube furnace under a flowing N<sub>2</sub>. After holding at the terminal temperature for 30 min, the tube furnace was rapidly heated to 820°C within 12 min. Then, the atmosphere was switched to steam and maintained for 10 min. The regenerated catalyst was thus obtained when the sample cooled down to room temperature under the protection of N<sub>2</sub>.

#### 2. Preparation of model compounds



The 2-bromoacetophenone (10 mmol, 2.0 g) was dissolved in acetone (60 mL) and guaiacol (12 mmol, 1.5 g) was added at RT under vigorous stirring until completely dissolved.  $K_2CO_3$  (70 mmol, 9.67 g) was added to the above solution and refluxed at 60°C. The reaction progression was monitored by thin-layer chromatography (TLC). After the end of the reaction, it was cooled to room temperature, and the organic phase was combined after filtration, and dried over MgSO<sub>4</sub>. The crude product was obtained by concentrating on a rotary evaporator under reduced pressure. 2-(2-Methoxyphenoxy) acetophenone would be obtained by column chromatography (petroleum ether/ethyl acetate 4/1) purification and characterized by NMR spectroscopy. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.01, 8.00, 7.61, 7.60, 7.58, 7.50, 7.49, 7.47, 6.99,

6.98, 6.97, 6.96, 6.95, 6.92, 6.91, 6.85, 6.85, 5.34, 3.89. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 195.53, 149.91, 147.62, 134.74, 133.86, 128.90, 128.20, 122.61, 120.90, 115.02, 112.33, 72.21, 56.01.







2-(2-Methoxyphenoxy)-1-phenylethanol

The 2-(2-Methoxyphenoxy) acetophenone (8 mmol, 1.9 g) was dissolved in THF (100 mL). An aqueous solution containing NaBH<sub>4</sub> (16 mmol, 0.6 g) was slowly added to the THF solution described above in an ice water bath. The mixture was subjected to continuous stirring in the ice water bath for a duration of 50 minutes, followed by subsequent transfer to ambient room temperature conditions for 4 h. A saturated aqueous solution of NH<sub>4</sub>Cl (100 mL) was used for the quenching of the reaction. The organic phase was extracted with EtOAc (3x20 mL), and dried over MgSO<sub>4</sub>. Model 1b was obtained by column chromatography (petroleum ether/ethyl

acetate 4/1) purification and characterized by NMR spectroscopy. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.45, 7.44, 7.39, 7.38, 7.36, 7.33, 7.32, 7.30, 7.01, 7.01, 7.00, 6.99, 6.99, 6.98, 6.98, 6.95, 6.94, 6.93, 6.93, 6.93, 6.92, 6.90, 6.90, 6.89, 6.89, 5.13, 5.13, 5.12, 5.11, 4.19, 4.19, 4.17, 4.17, 4.01, 3.99, 3.97, 3.88. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 150.09, 148.08, 139.68, 128.60, 128.12, 126.41, 122.55, 121.24, 115.81, 112.10, 76.28, 72.43, 55.95.







The synthesis method was similar to Model 1b. The 2-bromoacetophenone (10 mmol, 2.0 g) was dissolved in acetone (60 mL) and Syringol (12 mmol, 1.85 g) was added at RT under vigorous stirring until completely dissolved.  $K_2CO_3$  (70 mmol, 9.67 g) was added to the above solution and refluxed at 60°C. The reaction progression was monitored by thin-layer chromatography (TLC). After the end of the reaction, it was cooled to room temperature, and the organic phase was combined after filtration, and dried over MgSO<sub>4</sub>. The crude product was obtained by concentrating on a rotary evaporator under reduced pressure. 2-(2,6-Dimethoxyphenoxy)-1-phenylethan-1-one would be obtained by column chromatography

(petroleum ether/ethyl acetate 4/1) purification and characterized by NMR spectroscopy. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.05, 8.03, 7.58, 7.57, 7.55, 7.48, 7.46, 7.45, 7.02, 7.00, 6.98, 6.58, 6.56, 5.20, 3.80. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 195.30, 153.27, 136.74, 135.28, 133.38, 128.65, 128.42, 124.18, 105.43, 75.48, 56.16.







The synthesis method was similar to Model 2b. 2-(2,6-Dimethoxyphenoxy)-1-phenylethan-1one (8 mmol, 2.2 g) was dissolved in THF (100 mL). An aqueous solution containing NaBH<sub>4</sub> (16 mmol, 0.6 g) was slowly added to the THF solution described above in an ice water bath. The mixture was subjected to continuous stirring in the ice water bath for a duration of 50 minutes, followed by subsequent transfer to ambient room temperature conditions for a period of 4 h. A saturated aqueous solution of NH<sub>4</sub>Cl (100 mL) was used for the quenching of the reaction. The organic phase was extracted with EtOAc (3x20 mL), and dried over MgSO<sub>4</sub>. Model 2b was obtained by column chromatography (petroleum ether/ethyl acetate 4/1)

purification and characterized by NMR spectroscopy.<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.32, 7.31, 7.27, 7.26, 7.24, 7.21, 7.19, 7.18, 6.98, 6.96, 6.94, 6.55, 6.53, 4.90, 4.89, 4.88, 4.87, 4.47, 4.35, 4.35, 4.33, 4.33, 3.80, 3.67, 3.65, 3.63. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 153.35, 140.12, 136.83, 128.93, 127.44, 126.03, 124.79, 105.25, 80.84, 72.53, 56.20.







Fig. S1 Treating bagasse lignin with  $\text{CoNC}_{M4}$  under the optimized condition forms 4-

propylsyringol with 8.3 wt.% isolated yields.



Fig. S2 MS spectra of 4-PS.



<sup>1</sup>**H NMR (CDCl<sub>3</sub>, 500 MHz) δ (ppm):** 6.39(s, 2H, Ar-*H*), 5.30 (s,1H, Ar-O*H*), 3.87 (s, 6H, Ar-O-C*H*<sub>3</sub>), 2.50-2.53 (t, 2H, Ar-C*H*<sub>2</sub>-CH<sub>2</sub>-), 1.60-1.64 (m, 2H, -CH<sub>2</sub>-C*H*<sub>2</sub>-CH<sub>3</sub>), 0.92-0.95 (t, 3H,-CH<sub>2</sub>-C*H*<sub>3</sub>).



<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz) δ (ppm): 13.95 (-CH<sub>3</sub>), 24.99 (-CH<sub>2</sub>-), 38.42 (Ar-CH<sub>2</sub>-

), 56.35 (Ar-O-CH<sub>3</sub>), 105.9, 132.74, 133.98, 146.94 (ArC).

Fig. S3 <sup>1</sup>H and <sup>13</sup>C NMR of 4-propylsyringol.



Fig. S4 TEM of CoNC<sub>M4</sub>.



Fig. S5 HAADF-STEM images and EDS mappings of (a)  $CoNC_{M1}$ , (b)  $CoNC_{M2}$ , (c)  $CoNC_{M3}$ ,

#### (d) Co/NC<sub>M4</sub>.



Fig. S6 XPS N1s of CoNC<sub>M4-KOH</sub> and NC<sub>M4-KOH</sub>. CoNC<sub>M4-KOH</sub> and NC<sub>M4-KOH</sub>:After stirring in a 1 mol L<sup>-1</sup> KOH solution for 5 h, the catalyst is filtered to obtain a solid, which is then washed three times with deionized water to obtain the catalyst.



Fig. S7 (a) The first derivative curves of Co K-edge XANES for Co foli, CoO,  $CoNC_{M4}$  and

Co<sub>2</sub>O<sub>3</sub>, respectively. (b) Fitted valence states of CoNC<sub>M4</sub>.



Fig. S8 (a) SEM of the reused and regenerated of  $CoNC_{M4}$ . (b)  $N_2$  adsorption-desorption isotherms and (c) comparison of the pore distribution of the fresh, reused and regenerated

# CoNC<sub>M4</sub>.



Fig. S9 TG (a) and DSC (b) of lignin, CoNC<sub>M4</sub>, CoNC<sub>M4</sub>+lignin and spent CoNC<sub>M4</sub>.



Fig. S10 XPS of C1s (a), N1s (b) and Co2p (c) of the fresh, reused and regenerated  $CoNC_{M4}$ .



Fig. S11 FT-IR spectra of the raw-lignin, Re-lignin-blank and Re-lignin-CoNC<sub>M4</sub>.



Scheme S1 Control experiments used to determine the reaction pathway. Conditions: substrate (0.2 mmol), catalyst (20 mg), solvents (10 mL), H<sub>2</sub> (1.0 MPa), 230°C, 4 h. (3) After stirring in a 1 mol L<sup>-1</sup> KOH solution for 5 h, the catalyst is filtered to obtain a solid, which is then washed three times with deionized water to obtain the catalyst.

Biomass							
Catalant	D ti 1'ti	Primary	Yi	eld/ wt.%		D-f	
Catalyst	Reaction conditions	monomers	4-PS	Monomers	4-PS	Monomers	Kel.
CoNC <sub>M4</sub> (Co,	Organsolv bagasse lignin (0.1g),	4 DS	0.0	20 0	6 19	21.20	This most
0.46 wt.%)	Catalyst (0.1g), 230°C, 1MPa H <sub>2</sub> , 4h	4-P5	9.9	28.8	0.48	21.50	I his work
Co1Ti0.5@BC	0.5 g Enzymatic lignin (0.5g), 0.2 g		/	22.7	1	5 10	0
(Co:4.5 wt.%)	Catalyst (0.2g), 250°C, 2MPa H <sub>2</sub> , 4h.	HI, GI, G2	/	23.7	/	5.18	8
Co-Zn/Off-Al	Vroft lignin (0.5g) Catalyst (0.25g)						
H-beta (Co:	$200\%$ C $2MD_{\rm e}$ H $-6h$	G0, G1	/	15.6	/	9.99	9
1.24 wt.%)	300°C, 2MPa H <sub>2</sub> , 6n.						
30Ni-Mg <sub>3</sub> Al-	Organsolv bagasse lignin (0.2g),						
LDOs (29.90	Catalyst (0.15g), 260°C, 1MPa Ar,	H1, G1	2.8	21.3	0.04	0.40	10
wt.%)	3h.						
Ni <sub>5</sub> Fe <sub>5</sub> /Al <sub>2</sub> O <sub>3</sub>	People conduct $(1\alpha)$ Catching $(0,2\alpha)$						
(Ni: 3.65 wt.%,	$250\%$ $23MP_{0}$ $U_{0}$ $2h$	G2, 4-PS	12.2	27.2	2.32	5.37	11
Fe: 4.05 wt.%)	250°C, 3MPa H <sub>2</sub> , 5n						
Ni/C <sub>f</sub> (14.77	Birch sawdust (0.1g), Catalyst (0.1g),		10.2	45.2	0.20	0.04	12
wt.%)	220°C,2MPa H <sub>2</sub> , 3h.	4-P5,4-P50H	19.5	45.2	0.39	0.94	12
CuO/C (Cu	Poplar sawdust (0.05g), Catalyst		11.2	16 7	0.00	0.27	12
82.40 wt.%)	(0.025g), 240°C, 3MPa H <sub>2</sub> , 4h.	4-Р5, 4-Р5ОП	11.5	40.7	0.09	0.57	15
$D_{t}D_{a}/T_{t}O_{a}$ (Dt	Acid extracted birch lignin (0.2g),						
$\frac{100}{2} (\text{Pl})$	Catalyst (0.1g), 240°C, 1.5MPa He,	4-PS	7.5	18.7	7.44	19.95	14
2.00 <i>wt</i> .%)	12h.						
Ru/C (5.00	Birch wood (150g), Catalyst (15g),	4 DS C2	22.05	50.5	60	10.00	15
wt.%)	235°C, 3MPa H <sub>2</sub> ,3h.	4-PS, G2	33.83	30.3	0.8	10.00	15
Pd/C (1.00	Birch wood (1g), Catalyst (0.25g),	C2 4 DSOU	2 42	19 1	1.02	6.00	16
wt.%)	200°C, 4MPa H <sub>2</sub> , 15h.	G2, <b>4-</b> PSOH	2.42	48.4	1.03	0.00	10
Mo <sub>1</sub> Al/MgO	Eucalyptus wood (0.3g), Catalyst	G2-eOH	4	460/	1	22.00	17
(1.20 <i>wt</i> .%)	(0.1g), 200°C, 1MPa N <sub>2</sub> ,8h.	4-PSeOH	trace	40%	/	22.00	1/

# Table S1 Comparison of reductive catalytic fractionation results over different catalysts in

literatures.

TON denotes turnover numbers, calculated based on the total number of moles of active metal

in the catalyst (mol\_{phenols} mol\_{Active metal}^{-1})

Catalant	Suface area $(m^2g^{-1})^a$			Pore v	olume (cr	Mesoporous		
Catalyst	S <sub>BET</sub>	S <sub>micro</sub>	S <sub>ext</sub>	V <sub>total</sub>	V <sub>micro</sub>	V <sub>meso</sub>	ratio (%) °	
CoNC <sub>M1</sub>	380	248	132	0.27	0.13	0.14	51.9	
CoNC <sub>M2</sub>	646	159	487	0.40	0.09	0.31	77.5	
CoNC <sub>M3</sub>	1044	242	803	0.66	0.14	0.52	78.8	
CoNC <sub>M4</sub>	1394	137	1257	0.99	0.07	0.92	92.9	
<sup>5th</sup> CoNC <sub>M4</sub>	52	6	46	0.11	0.003	0.107	97.3	
CoNC <sub>M4</sub>	10/7	120	0.47	0.07	0.06	0.00	02.0	
Regenerate	1067	120	947	0.86	0.06	0.80	93.0	
CoC <sub>M4</sub>	1709	37	1672	1.47	0.01	1.46	99.3	
Co/NC <sub>M4</sub>	1012	168	843	0.86	0.09	0.77	89.5	
Co/AC	555	179	376	0.55	0.1	0.45	81.8	
Ru/C	925	501	424	0.67	0.27	0.4	59.7	
Pd/C	721	359	362	0.59	0.19	0.40	67.8	

 Table S2 Pore structure of catalysts.

<sup>a</sup> The surface area is determined utilizing the BET method. <sup>b</sup> The pore volume is evaluated through the BJH method. <sup>c</sup> The mesoporous ratio pore volume is calculated through  $V_{meso}/V_{Total}$ .

Catalant	Elemental content (wt.%)										
Catalyst	N	С	Н	S	0	Со	K	Na	Zn		
CoNC <sub>M1</sub>	6.90	71.64	2.08	0.27	18.08	1.03	0.11	0.06	0.06		
CoNC <sub>M2</sub>	3.10	59.41	3.07	1.84	30.78	1.80	0.20	0.15	0.03		
CoNC <sub>M3</sub>	5.10	56.42	3.06	0.37	24.17	10.88	0.14	0.09	1.37		
CoNC <sub>M4</sub>	8.61	70.76	2.38	1.95	15.84	0.46	0.12	0.10	0.03		
NC <sub>M4</sub>	/	/	/	/	/	/	0.11	0.06	1.19		
CoC <sub>M4</sub>	0.22	82.04	2.57	2.53	11.20	1.44	/	/	/		
Co/NC <sub>M4</sub>	4.73	68.96	2.82	0.43	25.56	0.50	/	/	/		
CoNC <sub>M4</sub> <sup>b</sup>	9.51	73.30	3.00	0.32	13.52	0.35	/	/	/		

Table S3 Element analysis of the CoNC catalysts <sup>a</sup>.

<sup>a</sup> The Co, K, Na and Zn contents are detected by inductively coupled plasma atomic emission spectroscopy (ICP-OES), C, H, N, S contents are detected by elemental analysis. O content is calculated from the other element content. <sup>b</sup> CoNC<sub>M4</sub> is obtained after five times reuse.

		Relative amount (%)						Acidity (µmol g <sup>-1</sup> )	
Catalyst Pyric	Pyridinic-	Co-N	Pyrrolic-	Graphitic	Oxidized-	Pyrrolic/	Weak	Total	Total
	Ν		Ν	-N	Ν	Pyridinic			(%)
CoNC <sub>M1</sub>	34.01	15.66	22.23	19.76	8.34	0.65	162.98	376.10	43.33
CoNC <sub>M2</sub>	26.29	16.53	22.01	28.78	6.39	0.84	307.31	465.90	65.96
CoNC <sub>M4</sub>	32.69	12.02	21.21	23.21	10.87	0.65	111.71	273.29	40.88
NC <sub>M4</sub>	33.33	/	31.41	24.04	11.22	0.94	200.08	658.61	30.38
NC <sub>M4-KOH</sub>	40.94	/	16.56	28.85	13.65	0.40	/	/	/

Table S4 N atomic relative content ratio and acidity of CoNC.

The amounts of acidic sites were determined according to the temperature and the area of the NH<sub>3</sub> desorption peaks.

samples	path	C. N. <sup>[a]</sup>	R (Å) <sup>[b]</sup>	$\sigma^2 (\times 10^{-3} \text{ Å}^2)^{[c]}$	$\Delta E (eV)^{[d]}$	R factor <sup>[e]</sup>	
Co foil	Co-Co	12*	2.51*	6.2	5.2	0.009	
Co-sample	Co-N	4.3	2.08	6.5	2.0	0.012	
	Co-Co	0.6	2.56	7.9	2.0	0.012	

Table S5 EXAFS fitting parameters at the Co K-edge various samples ( $S_0^2=0.71$ ).

<sup>*a*</sup>C. N.: coordination numbers; <sup>*b*</sup>*R*: bond distance; <sup>*c*</sup> $\sigma^2$ : Debye-Waller factors; <sup>*d*</sup> $\Delta E_0$ : the inner potential correction. <sup>*e*</sup>*R* factor: goodness of fit. \*The experimental EXAFS fit by fixing C. N. as the known crystallographic value.

		GI OH	G2 OH		••							
			Yield wt.% <sup>b</sup>									
	Lignin	Н	G	4-PS	Others	Sel. <sub>4-PS.</sub>	Total					
Hardwood	Birch	/	5.6	8.0	6.0	40.8	19.6					
	Poplar	/	7.1	7.4	2.8	42.8	17.3					
Softwood	Pine	/	11.1	0.6	3.3	4.0	15.0					
	Chinese Fir	/	7.6	1.4	1.2	13.7	10.2					
Grasses	Wheat bran	0.6	5.6	3.4	2.0	29.3	11.6					
	Bamboo	2.5	5.2	7.2	4.1	37.9	19.0					
	Corncob	9.5	6.0	4.8	1.5	22.1	21.7					

Table S6 Catalytic of various lignin samples over  $\text{CoNC}_{M4}$  catalyst<sup>a</sup>.

<sup>a</sup> Conditions: organosolv lignin (100 mg), catalyst (100 mg), EtOH (10 mL), H<sub>2</sub> (1.0 MPa), 230

°C, 4 h. <sup>b</sup> Yields were determined by GC-FID.

#### References

- S. Liu, A. P. van Muyden, L. Bai, X. Cui, Z. Fei, X. Li, X. Hu and P. J. Dyson, *ChemSusChem*, 2019, 12, 3271-3277.
- Z. Li, Z. Cai, Q. Zeng, T. Zhang, L. J. France, C. Song, Y. Zhang, H. He, L. Jiang, J. Long and X. Li, *Green Chem.*, 2018, 20, 3743-3752.
- L. Li, J. Kong, H. Zhang, S. Liu, Q. Zeng, Y. Zhang, H. Ma, H. He, J. Long and X. Li, *Appl. Catal. B-Environ.*, 2020, 279, 119343.
- 4. Y. Wang and S. Liu, J. Agric. Food. Chem., 2024, 72, 13320-13327.
- 5. A. Bahadoor, A. Brinkmann and J. E. Melanson, Anal. Chem., 2021, 93, 851-858.
- 6. M. Dolg, U. Wedig, H. Stoll and H. Preuss, J. Chem. Phys., 1987, 86, 866-872.
- 7. J. P. Perdew, K. Burke and M. Ernzerhof, *Phys. Rev. Lett.*, 1997, **78**, 1396-1396.
- 8. B. Luo, Z. Tian, R. Shu, C. Wang, Y. Chen, J. Liu and Y. Liao, J. Catal., 2025, 442.
- X. Dou, X. Jiang, W. Li, C. Zhu, Q. Liu, Q. Lu, X. Zheng, H.-m. Chang and H. Jameel, *Appl. Catal. B-Environ.*, 2020, 268, 118429.
- 10. Y. Tian, M. Gao, Z. Tang, F. Li, Q. Zeng, J. Long and X. Li, J. Catal., 2024, 440, 115833.
- Z. Shen, W. Wang, L. Pan, Z. Huang, X. Zhang, C. Shi and J.-J. Zou, *Green Chem.*, 2023, 25, 7782-7793.
- K. Wu, B. Luo, K. Yang, S. Wang, M. Li and H. Zhang, ACS Sustainable Chem. Eng., 2024, 12, 1536-1547.
- 13. Q. Wang, L.-P. Xiao, Y.-H. Lv, W.-Z. Yin, C.-J. Hou and R.-C. Sun, *ACS Catal.*, 2022, **12**, 11899-11909.
- J. Hu, S. Zhang, R. Xiao, X. Jiang, Y. Wang, Y. Sun and P. Lu, *Bioresour. Technol.*, 2019, 279, 228-233.
- Y. Liao, S.-F. Koelewijn, G. Van den Bossche, J. Van Aelst, S. Van den Bosch, T. Renders, K. Navare, T. Nicolai, K. Van Aelst, M. Maesen, H. Matsushima, J. M. Thevelein, K. Van Acker, B. Lagrain, D. Verboekend and B. F. Sels, *Science*, 2020, **367**, 1385-1390.
- W. Lan, M. T. Amiri, C. M. Hunston and J. S. Luterbacher, *Angew. Chem. Int. Ed.*, 2018, 57, 1356-1360.
- G. Meng, W. Lan, L. Zhang, S. Wang, T. Zhang, S. Zhang, M. Xu, Y. Wang, J. Zhang, F. Yue, Y. Wu and D. Wang, J. Am. Chem. Soc., 2023, 145, 12884-12893.