Supporting Information for

In situ capture and value-added utilization of CO₂ from flue gas using ionic liquid polymer supported Zn catalyst

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Contents

1. General Information	3
Chemicals and materials	3
Characterization	3
2. Synthetic procedures of the porous organic polymer catalysts	4
3. General procedures	5
Recyclability Test for Zn@PIP-1	6
4. Catalyst characterization	6
5. Performance Comparison of cycloaddition of PO using flue gas	7
6. NMR data of the products	8
7. NMR specture of the products	13

1. General Information

Chemicals and materials

All solvents and chemicals, unless otherwise noted, were obtained commercially and were used as received without further purification. All glassware was dried before use. Flue gas was purchased from Lanzhou Yulon Gas Co., Ltd., where the flue gas is directly supplied from a coal power group. The flue gas composition is as follows: CO₂ (19.4%), O₂ (1.8%), N₂ (78.8%), and NO_x (\leq 20 ppm). Analytical thin layer chromatography (TLC) was performed using pre-coated Jiangyou silica gel HSGF254 (0.2mm \pm 0.03mm). Flash chromatography was performed using silica gel 60, 0.063-0.2 mm, 200-300 mesh (Jiangyou, Yantai) with the indicated solvent system.

Characterization

Gas chromatography analysis was performed on Agilent 7890A GC equipped with a HP-5 capillary column and FID detector. GC-MS analysis was in general recorded on an Agilent 5977A MSD GC-MS. Fourier transform infrared (FT-IR) spectrum were recorded with a Bruker VERTEX 70FTIR spectrometer. The liquid nuclear magnetic resonance spectra (NMR) were recorded on a Bruker Avance TM III 400 MHz in deuterated chloroform unless otherwise noted. Data are reported in parts per million (ppm) as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, m = multiples, dd = doublet of doublet and br = broad signal), coupling constant in Hz and integration. Powder X-ray diffraction (PXRD) measurements were conducted by a STADIP automated transmission diffractometer (STOE) equipped with an incident beam curved germanium monochromator selecting CuKal radiation and a 6° position sensitive detector (PSD) (step size: 0.014, step time: 25.05 s). The XRD patterns were scanned in the 20 range of 0-80°. Nitrogen adsorption-desorption isotherms were measured at 77 K using an American Quantachrome iQ2 automated gas sorption analyzer. The samples were outgassed at 120 °C for 12 h before the measurements. Surface areas were calculated from the adsorption data using Langmuir and Brunauer-Emmett-Teller (BET) methods. The pore-sizedistribution curves were obtained from the adsorption branches using the non-local density functional theory (NLDFT) method. X-ray photoelectron spectroscopy (XPS) measurements were carried out by a VG ESCALAB 210 instrument equipped with a dual Mg/Al anode X-ray source, a hemispherical capacitor analyzer, and a 5 keV Ar⁺ ion gun. All spectra were recorded by using AlKa (1361 eV) radiation. The electron binding energy was referenced to the C1s peak at 284.8 eV. The thermal properties of Zn@PIP-1 catalysts were evaluated using a METTLER TOLEDO simultaneous thermal analyzer over the temperature range from 30 to 800 °C under a nitrogen atmosphere (20 mL/min) with a heating rate of 5 °C/min. Field emission scanning electron microscopy (SEM) observations were performed on a Hitachi S-4800 microscope operated at an accelerating voltage of 5.0 kV. High-resolution transmission electron microscope (HR-TEM) analysis was carried out on a Talos F200S operating at 200 kV. Zn K-edge analysis was performed with Si (111) crystal monochromators at the BL11B beamlines at the Shanghai Synchrotron Radiation Facility (SSRF) (Shanghai, China). Before the analysis at the beamline, samples were pressed into thin sheets with 1 cm in diameter and sealed using Kapton tape film. The XAFS spectra were recorded at room temperature using a 4-channel Silicon Drift Detector (SDD) Bruker 5040. XX-edge extended X-ray absorption fine structure (EXAFS) spectra were recorded in transmission mode. Negligible changes in the line-shape and peak position of Zn Kedge XANES spectra were observed between two scans taken for a specific sample. The XAFS

spectra of these standard samples (Zn Foil, ZnO) were recorded in transmission mode (fluorescence mode). The spectra were processed and analyzed by the software codes Athena and Artemis.

2. Synthetic procedures of the porous organic polymer catalysts

1. Preparation of Zn@2P2Br&*p*-3vPPh₃ porous ionic liquid polymer catalysts (Zn@PIP-1):

a. Synthesis of the quaternary phosphonium salt ionic liquid (2P2Br): Under argon protection, trivinylphenylphosphine (2.1 mmol), *p*-dibromobenzyl (1 mmol), and 4-5 g of TBC (*p*-tert-butylcatechol) were dissolved into a solution of anhydrous dichloromethane (20 mL). The mixture was stirred at 40 °C for 4 h, then washed with methyl tert-butyl ether and dried. A white solid was obtained in quantitative yield, noted as **2P2Br**.



b. Under argon protection, **2P2Br** (0.36 mmol), trivinylphenylphosphine (1 mmol), zinc bromide (0.5 mmol), and 18 mg of AIBN were dissolved into a solution of anhydrous N, N-Dimethylformamide (8 mL). After stirring for 24 h at 100 ° C, the solution was washed with methyl tert-butyl ether and drained at room temperature. The white solid obtained was labeled as Zn@PIP-1.



2. Preparation of Zn@2P2Br&p-3vPPh₃ porous ionic liquid polymer catalysts (Zn@PIP-2):

Synthesise in the same way as above, but replace monomer 1 with monomer 2.



monomer 2

3. Preparation of Zn@2P2Br&p-3vPPh₃ porous ionic liquid polymer catalysts (Zn@PIP-3):

Synthesise in the same way as above, but replace monomer 1 with monomer 3.



monomer 3

3. General procedures



In a typical procedure, a 100 mL stainless steel autoclave was filled with 40 mg of Zn@PIP-1 catalyst, 2.5 mmol of substrates, 3.0 MPa of flue gas, and 2 mL of hexane as solvent. At room temperature, the air in the autoclave was replaced three times with 1 MPa flue gas and then filled with 3.0 MPa flue gas. The autoclave was placed in the reactor and heated and stirred at 110 °C for 12 h. After the reaction, the autoclave was cooled down to room temperature, and the pressure in the autoclave was slowly released. Subsequently, the reaction mixture was diluted with ethyl acetate (6.0 mL), and the selectivity of the product was confirmed by gas chromatography (Agilent 7890A GC, HP-5 capillary column, 5 wt.% phenyl, FID detector). The quantitative yield of target product GC: The yield of the product can be obtained by GC analysis using biphenyl as an internal standard. Target product isolation yield: After the reaction mixture was concentrated in a vacuum to remove the solvent, the crude product was analyzed by GC using PE/EA as eluent. After the reaction mixture was concentrated in a vacuum to remove the solvent, the crude product was analyzed by GC using PE/EA as eluent. After the reaction mixture was concentrated in a vacuum to remove the solvent, the crude product was analyzed by GC using PE/EA as eluent. After the reaction mixture was concentrated in a vacuum to remove the solvent, the crude product was analyzed by GC using PE/EA as eluent.

Recyclability Test for Zn@PIP-1.

After the cycloaddition reaction of epoxides with flue gas was completed, the recycled catalyst was separated by centrifugation, washed with degassed THF (tetrahydrofuran), and used directly for the next run.

4. Catalyst characterization



Figure S1. Thermal gravimetric analysis (TGA) curve of Zn@PIP-1



Figure S2. Metal loading optimization identified an ideal Zn content



Figure S3. N₂ sorption isotherms measured at 77 K.



Figure S4. (A) Effect of reaction solvents on the yield of carbonates.: A. n-hexane. B. dioxane.C. acetonitrile. D. methyl tertbuty ether. E. dichloromethane. F. methylbenzene. G. dimethyl formamide. (B) Effect of reaction pressure on carbonate yield.



Table S1 ICP data for fresh and used catalysts.

Entry	Zn (wt%)
Zn@PIP-1	3.04
Zn@PIP-1-used	3.90

5.Performance Comparison of cycloaddition of PO using flue gas.

Entry	Catalyst	Co- catalyst	Reaction condition	Flue gas composition	Num . of S	Y (%)	Cycle	Ref.
1ª	Zn@PIP-1	-	110°C, 30bar, 12h	19.4% CO ₂ → 78.8 % N ₂ , 1.8% O ₂ , NO _x (≤20ppm)	22	99	5	(1)
2ª	Al-MDC-NC	-	100°С, 1bar, 12h	15% CO ₂ , 85% N ₂ , 0.0200g fly ash, and 95% RH)	-	82	-	(2)
5ª	PBPILs	-	100°C, 20bar, 4h	15% CO ₂ and 85% N ₂	6	95	5	(3)
7 ^a	I _{C2} HCP-5 _b	-	120°C, 30bar, 24h	15% CO ₂ and 85% N ₂	6	92	6	(4)
11 ^b	(3- aminobenzyl alcohol/n Bu ₄ NI	-	60°C, 10bar, 35h	14.5% CO ₂ , 2% H ₂ O, 1.5% O ₂ , 300 ppm CO, 207 ppm SO _x , 300 ppm NO _x and the other component was N ₂	-	99	-	(5)
12 ^b	HBDs	Ammo nium salt	60°C, 1bar, 10h	14.5% CO ₂ , 2% H ₂ O, 1.5% O ₂ , 300 ppm CO, 207 ppm SO _x , 300 ppm NO _x and the other component was N ₂	-	99	-	(6)
13 ^b	PP _{1,4} Br/ZnC l ₂	ZnCl ₂	60°C, 40bar, 18h	19.4% CO ₂ , 78.8 % N ₂ 1.8% O ₂ , No _x (≤20ppm)	19	99	7	(7)

Table S2 Performance Comparison of cycloaddition of PO using flue gas.

^a Heterogeneous, ^b Homogeneous

(1). This work. (2). Ind.Eng.Chem.Res. 2024, 63, 16725–16735. (3). Molecular Catalysis 2023, 544, 113157. (4). ChemSusChem 2020, 13, 341. (5). Journal of Environmental Chemical Engineering 2024, 12, 113614. (6). Catal. Sci. Technol. 2024, 14, 6215 - 6223. (7).Green Chem. 2023, 25, 2293 - 2298.

6. NMR data of the products



1,3-dioxolan-2-one ¹H NMR (400 MHz, CDCl₃) δ 4.51 (s, 1H). **¹³C NMR (101 MHz, CDCl₃)** δ 155.60, 64.74.



4-Ethyl-1,3-Dioxolan-2-one

¹H NMR (400 MHz, CDCl₃) δ 4.78 – 4.64 (m, 1H), 4.56 (t, *J* = 8.2 Hz, 1H), 4.11 (dd, *J* = 8.4, 7.0 Hz, 1H), 1.91 – 1.72 (m, 2H), 1.03 (t, *J* = 7.5 Hz, 3H).
¹³C NMR (101 MHz, CDCl₃) δ 155.19, 78.08, 69.04, 26.82, 8.41.



4-Butyl-1,3-dioxolan-2-one,

¹**H** NMR (400 MHz, CDCl₃) δ 4.72 (dd, J = 7.5, 5.5 Hz, 1H), 4.54 (t, J = 8.1 Hz, 1H), 4.08 (dd, J = 8.3, 7.3 Hz, 1H), 1.74 (dd, J = 23.8, 18.4 Hz, 2H), 1.50 – 1.24 (m, 4H), 0.93 (t, J = 7.0 Hz, 3H). ¹³**C** NMR (101 MHz, CDCl₃) δ 155.21, 77.17,69.50, 33.66, 26.53, 22.35, 13.89.



4-Hexyl-1,3-dioxolan-2-one,

¹**H NMR (400 MHz, CDCl₃)** δ 4.69 (dd, *J* = 7.5, 5.4 Hz, 1H), 4.63 – 3.89 (m, 2H), 1.89 – 1.57 (m, 2H), 1.47 – 1.20 (m, 8H), 0.86 (t, *J* = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 155.19, 77.16, 69.48, 33.92, 31.56, 28.84, 24.38, 22.51, 14.04.



4-Methoxymethyl-1,3-dioxolan-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 4.85 – 4.75 (m, 1H), 4.55 – 4.27 (m, 2H), 3.67 – 3.48 (m, 2H), 3.39 (dd, J = 5.5, 2.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 155.09, 75.17, 71.51, 66.22, 59.64.



4-(isopropoxymethyl)-1,3-dioxolan-2-one

¹**H NMR** (400 MHz, CDCl₃) δ 4.83 – 4.79 (m, 1H), 4.56 – 4.36 (m, 1H), 3.72 – 3.55 (m, 1H), 1.17 (d, *J* = 6.1 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 155.20, 75.31, 72.98, 67.18, 66.48, 21.95, 21.84.



4-tert-butoxymethyl-1,3-dioxolan-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 4.77 – 4.54 (m, 1H), 4.41 – 4.14 (m, 1H), 3.42 (dd, *J* = 44.9, 2.4 Hz, 1H),

1.03 (d, J = 1.0 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 155.11,75.27, 73.30, 66.10, 61.00, 26.92.



4-Chloromethyl-1,3-dioxolan-2-one,

¹**H** NMR (400 MHz, CDCl₃) δ 5.01 – 4.87 (m, 1H), 4.51 (td, *J* = 8.7, 2.6 Hz, 1H), 4.29 (ddd, *J* = 9.3, 5.6, 3.9 Hz, 1H), 3.77 (ddd, *J* = 12.5, 4.1, 1.3 Hz, 1H), 3.70 – 3.62 (m, 1H). ¹³**C** NMR (101 MHz, CDCl₃) δ 154.49, 74.45, 66.75, 44.22.



4-(fluoromethyl)-1,3-dioxolan-2-one

¹H NMR (400 MHz, CDCl₃) δ 4.96 – 4.79 (m, 1H), 4.78 – 4.28 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 154.71, 82.30, 80.55, 74.78, 74.59, 64.98, 64.91.



4-Vinyl-1,3-dioxolan-2-one,

¹H NMR (400 MHz, CDCl₃) δ 5.83 (s, 1H), 5.59 – 5.24 (m, 2H), 5.07 (d, *J* = 7.4 Hz, 1H), 4.54 (t, *J* = 8.4 Hz, 1H), 4.25 – 3.80 (m, 1H).
¹³C NMR (101 MHz, CDCl₃) δ 154.82, 132.15, 120.96, 77.30, 77.16, 69.03.



4-(But-3-en-1-yl)-1,3-dioxolan-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 5.76 (ddd, *J* = 16.9, 7.6, 2.9 Hz, 1H), 5.18 – 4.94 (m, 2H), 4.71 (dt, *J* = 7.8, 4.0 Hz, 1H), 4.58 – 4.46 (m, 1H), 4.11 – 3.99 (m, 1H), 2.19 (ddd, *J* = 9.3, 8.3, 4.2 Hz, 2H), 1.95 – 1.74 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 155.06, 136.18, 116.43, 76.43, 69.40, 33.08, 28.70.



4-((Allyloxy)methyl)-1,3-dioxolan-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 4.88 – 4.79 (m, 1H), 4.48 (t, *J* = 8.5 Hz, 1H), 4.35 (dd, *J* = 8.4, 6.1 Hz, 1H), 4.25 – 4.12 (m, 2H), 3.80 – 3.64 (m, 2H), 2.48 (t, *J* = 2.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 154.97, 78.61, 75.63, 74.86, 68.47, 66.19, 58.78.



4-((Prop-2-yn-1-yloxy) methyl)-1,3-dioxola n-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 4.88 – 4.79 (m, 1H), 4.48 (t, *J* = 8.5 Hz, 1H), 4.35 (dd, *J* = 8.4, 6.1 Hz, 1H), 4.25 – 4.12 (m, 2H), 3.80 – 3.64 (m, 2H), 2.48 (t, *J* = 2.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 154.97, 78.61, 75.63, 74.86, 68.47, 66.19, 58.78.



4-(2-ethylhexyl) oxy) methyl)-1,3-dioxolan-2-one

¹**H** NMR (400 MHz, CDCl₃) δ 4.85 – 4.62 (m, 1H), 4.49 – 4.16 (m, 1H), 3.63 – 3.38 (m, 1H), 3.33 – 3.21 (m, 1H), 1.39 (d, J = 5.8 Hz, 1H), 1.31 – 1.01 (m, 1H), 0.77 (dt, J = 10.1, 6.7 Hz, 1H). ¹³**C** NMR (101 MHz, CDCl₃) δ 155.03, 75.25, 74.44, 74.42, 69.75, 66.10, 39.39, 30.24, 28.86, 23.57, 22.82, 13.84, 10.82.



4-((3-(trimethoxy silyl) proxy) methyl)-1,3-dioxolan-2-one

¹**H NMR** (400 MHz, CDCl₃) δ 4.80 (dd, *J* = 4.0, 2.0 Hz, 1H), 4.53 – 4.26 (m, 5H), 3.72 – 3.41 (m, 29H), 1.73 – 1.59 (m, 5H), 0.68 – 0.58 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 155.07, 77.48, 75.14, 74.05, 69.71, 66.43, 50.67, 22.79, 5.21.



4-((3-(Triethoxysilyl) propoxy) methyl)-1,3-dioxolan-2-one,

¹**H NMR** (400 MHz, CDCl₃) δ 4.73 – 4.66 (m, 1H), 4.41 – 4.21 (m, 1H), 3.67 (q, *J* = 7.0 Hz, 3H), 3.50 (ddd, *J* = 38.7, 11.1, 3.6 Hz, 1H), 3.34 (td, *J* = 6.6, 1.2 Hz, 1H), 1.59 – 1.48 (m, 1H), 1.07 (t, *J* = 7.0 Hz, 5H), 0.54 – 0.42 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 156.94, 154.90, 75.09, 73.73, 69.39, 66.06, 58.11, 22.68, 18.04, 6.10.



4-phenyl-1,3-dioxolan-2-one

¹**H NMR** (400 MHz, CDCl₃) δ 7.43 – 7.37 (m, 3H), 7.36 – 7.31 (m, 2H), 5.65 (t, *J* = 8.0 Hz, 1H), 4.82 – 4.69 (m, 1H), 4.29 (dd, *J* = 8.6, 7.8 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 154.90, 135.78, 129.55, 129.06, 125.87, 77.94, 71.08.



4-benzyl-1,3-dioxolan-2-one

¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.18 (m, 1H), 5.03 – 4.65 (m, 1H), 4.42 (dd, *J* = 8.6, 7.9 Hz, 1H),
4.15 (dd, *J* = 8.6, 6.9 Hz, 1H), 3.05 (ddd, *J* = 55.9, 14.2, 6.4 Hz, 1H).
¹³C NMR (101 MHz, CDCl₃) δ 154.90, 134.05, 129.40, 128.98, 127.55, 76.93, 68.54, 39.54.



4-(4-chlorophenyl)-1,3-dioxolan-2-one

¹H NMR (400 MHz, CDCl₃) δ 7.37 (dd, J = 47.7, 8.5 Hz, 1H), 5.66 (s, 1H), 4.56 (d, J = 199.6 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 154.62, 135.95, 134.42, 129.67, 127.38, 77.35, 77.16, 71.12.



4-((benzyloxy)methyl)-1,3-dioxolan-2-one

¹**H NMR** (400 MHz, CDCl₃) δ 7.29 (s, 1H), 4.75 (dd, *J* = 5.8, 2.6 Hz, 1H), 4.52 (d, *J* = 1.7 Hz, 1H), 4.46 – 4.20 (m, 1H), 3.61 (ddd, *J* = 46.2, 12.9, 7.2 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 155.06, 137.18, 128.56, 128.04, 127.73, 75.13, 68.88, 66.28.



4-(phenoxymethyl)-1,3-dioxolan-2-one

¹**H** NMR (400 MHz, CDCl₃) δ 7.34 – 7.28 (m, 1H), 7.04 – 6.98 (m, 1H), 6.93 – 6.88 (m, 1H), 5.09 – 4.97 (m, 1H), 4.57 (ddd, J = 14.4, 12.7, 7.2 Hz, 1H), 4.19 (ddd, J = 39.3, 10.6, 3.9 Hz, 1H). ¹³**C** NMR (101 MHz, CDCl₃) δ 157.87, 154.80, 129.82, 122.11, 74.24, 66.99, 66.35.



4-((o-tolyloxy) methyl)-1,3-dioxolan-2-one

¹**H NMR** (400 MHz, CDCl₃) δ 7.18 – 7.09 (m, 1H), 6.94 – 6.85 (m, 1H), 6.77 (s, 1H), 5.02 (d, *J* = 3.0 Hz, 1H), 4.56 (dd, *J* = 11.2, 6.9 Hz, 1H), 4.15 (dd, *J* = 45.7, 3.1 Hz, 1H), 2.20 (s, 1H). ¹³**C NMR** (101 MHz, CDCl₃) δ 155.84, 155.01, 131.07, 127.06, 126.94, 121.63, 110.91, 77.16, 74.41, 67.06, 66.30, 15.

7. NMR specture of the products











































