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# Light-driven Newman Kwart Rearrangement under Ambient Conditions with Cysteine Quantum Dots

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Supplementary Information

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#### 1. General considerations:

All chemicals were procured from commercial suppliers and used without further purification: Spectrochem, Avra, SRL, TCI, BLD Pharm, Finar and Sigma Aldrich. The solvents were purchased from SD Fine Chemicals Limited (SDFCL), India. For column chromatography, hexane was distilled to remove the grease. For extraction/workup, standard laboratory grade solvents procured from SDFCL India were used. For the purpose of the reaction, acetonitrile was procured from Finar (HPLC grade) and distilled to dryness using P<sub>2</sub>O<sub>5</sub>. The distilled solvent was collected and stored over activated 4 Å molecular sieves. For column chromatography, silica gel (100-200 mesh) was taken from Isochem and thin layer chromatography (TLC) was carried out on silica gel

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60 F<sub>254</sub> precoated plates (0.25 mm), obtained from Sigma Aldrich (Supelco). The nanomaterial

synthesis was done in distilled water.

All the reactions were carried out in oven-dried glassware (120°C). Dry reaction conditions were

maintained (wherever mentioned) using standard Schlenck line techniques with an Argon

atmosphere. NMR spectra was recorded in CDCl<sub>3</sub> solvent (procured from Cambridge Isotopes

Limited and used as obtained). NMR yields were obtained using Dibromomethane as the

reference.

The LEDs, CFLs and Kessil Lamp (PR160L) for photo-irradiation were procured commercially.

The data was normalized/smoothed/deconvoluted/fit (as required) in GraphPad/OriginPro 9.1.

All the images were processed in Adobe Illustrator CS6 software.

#### 2. Instrumentation:

NMR spectroscopy: Bruker Ultrashield spectrometer (400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C); NMR spectra were processed on TopSpin 3.6.4; chemical shifts have been reported in parts per million (ppm), while coupling constants (*J*) have been reported in Hz. For <sup>1</sup>H-NMR, data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = double doublet, t =

triplet, q = quartet, br = broad, m = multiplet), coupling constants (Hz) and integration.

HRMS: Waters XEVO G2-XS QTof instrument.

UV-visible spectroscopy: UV-3600 Shimadzu UV-Vis-NIR Spectrophotometer from Japan

Analytical Instruments (1 mL quartz cuvette with a 10 mm optical path).

Fluorescence spectroscopy: Varian Cary Eclipse Fluorescence Spectrophotometer.

FT-IR spectroscopy: CaF<sub>2</sub> cell in Bruker alfa FT-IR instrument.

TEM: JOEL JEM-2100F with an accelerating voltage of 200 kV.

<u>XPS</u>: ESCALAB 250 (Thermo Electron) instrument with monochromatic Al K $\alpha$  (1486.6 eV) as the

X-ray source.

3

CV: Electrochemical Analyser (HCH Instruments CHI604E); platinum (glossy carbon) working electrode, platinum wire counter electrode and Ag/AgCl reference electrode were procured from CH Instruments, Inc.

Zeta Potential, Dynamic Light Scattering (DLS) and Molecular Weight calculation: Malvern Zetasizer Nano ZS System.

### 

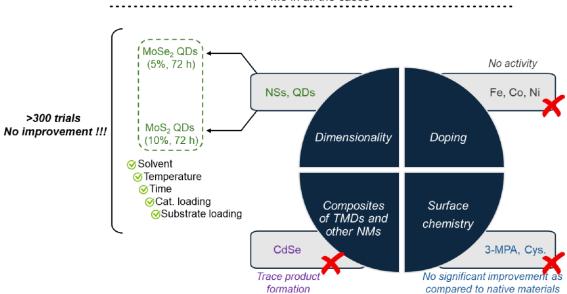
1a: R' = 4-0Me

2a: R' = 4-0Me

 1b: R' = 4-H 2b: R' = 4-H 

 1v:  $R' = 4-NO_2$  2v:  $R' = 4-NO_2$ 

R = Me in all the cases



**Scheme S1:** Initial attempts to mediate NKR using TMDs bearing various dimensionality, functionality, dopant or composition.

The nanosheets (NSs) and quantum dots (QDs) of TMDs, MoS<sub>2</sub> and MoSe<sub>2</sub> were prepared using solvent exfoliation and hydrothermal routes respectively, as per our previous reports.<sup>1-3</sup> The doping with Fe/Co/Ni was done following documented protocols.<sup>4,5</sup> For functionalization on the surface, the nanomaterial was stirred with the ligand 3-mercaptopropionic acid or L-cysteine.<sup>6</sup> CdSe<sup>7</sup> and

CdS<sup>8</sup> QDs were also prepared using bottom-up approaches, with the reaction conditions maintained so as to furnish QDs having absorption maxima in the range 350-420 nm. As NKR could proceed via the anionic or cationic pathway, we chose three different substrates for the trial, such that the substituents on the aryl ring could range from electron-donating (1a: R'

= 4-OMe) or withdrawing (1v: R' = 4-NO<sub>2</sub>) to electronically neutral groups (1b: R' = H) (Scheme S1).

Since the NKR of **1a** with MoS<sub>2</sub> QDs gave a minimal yield of 10% (which could not be improved further by tuning the surface chemistry, dimensionality, composition or incorporation of metal dopant), the reaction parameters were modulated sequentially, *vide infra*:

- a) Solvents: H<sub>2</sub>O, MeCN, EtOH, THF, DMF (individual as well as mixture of solvents in different ratios).
- b) Temperature: 25-70°C.
- c) Time: 1-96 h.
- d) Catalyst loading: 1-20 wt% of the catalyst, relative to the O-arylcarbamothioate.
- e) Substrate loading: 0.1-0.25 mmol.

#### 4. Experimental section:

## 4.1 Procedure for optical/microscopic characterization and sample preparation:

**TEM (Transmission Electron Microscopy):** The samples for TEM imaging were prepared on a Formvar/carbon 200 mesh copper grid loaded with a dilute solution of the sample and dried under vacuum.

**UV-Visible (absorption) and Fluorescence (emission) spectroscopy:** A 1 mL quartz cuvette having a 10 mm optical path was taken and the diluted sample was added to it. Scanning was done

from 200 to 800 nm with a step duration of 0.5 nm. For the fluorescence spectrum, the sample was excited at different wavelengths from 250-450 nm (at intervals of 10 nm) and the emission profile was recorded.

**Zeta Potential and hydrodynamic diameter measurement:** For measuring surface charge and hydrodynamic diameter, the aqueous solution of the sample was used.

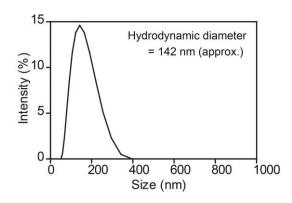
**Molecular weight measurement:** Toluene was used as the standard while water was used as the solvent. The measurements were done at variable concentrations of the nanomaterial.

**FT-IR** (**Fourier transform InfraRed**) **spectroscopy:** The FT-IR spectrum of CQDs was recorded by dissolving the lyophilized sample in MeOH and spreading it on a CaF<sub>2</sub> cell. The residual solvent was removed by drying. The spectrum was recorded from 4000 to 500 cm<sup>-1</sup>.

**XPS** (X-ray Photoelectron Spectroscopy): The samples were prepared by drop-casting a relatively concentrated solution of the nanomaterial on silicon wafers (size less than 1 cm X 1 cm; cleaned thrice by heating at 60°C in isopropanol followed by acetone wash and oven drying). The sample was dried in air, and then in vacuum for at least 24 h.

#### 4.2 Characterization of CQDs:

DLS measurements revealed a large hydrodynamic diameter compared to the size of the individual domains of the CQDs, which confirmed the close proximity of these entities.

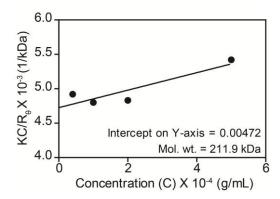


**Figure S1:** DLS measurement, showing the hydrodynamic diameter of the CQDs.

#### 4.3 Calculation of molecular weight for the CQDs (reference given at the end):

The molecular weight was calculated using the Zetasizer equipment by the SLS (Static Light Scattering) method. The refractive index increment (dn/dc) was set as 0.1 mL/g, while the shape correction model was taken as that of a sphere. The temperature was kept as 25°C throughout and the equilibration time was fixed to 100 seconds. First, the dark count measurement (laser turned off) was done for the background light level. Toluene was used as the scattering standard to establish the scattering intensity of the reference. Water was used as the solvent (first, a blank measurement was taken corresponding to zero sample concentration). The CQDs (homogeneous solution) were taken at different concentrations (diluted in water). The scattering intensity and concentration were ensured to be in accordance with the instrument quality criteria.

Finally, a plot of KC/R $_{\theta}$  (1/KDa) vs. concentration C (g/mL) was made (all the values were taken directly from the instrument) and the fitting was done using a linear regression model (K: optical constant; C: concentration; R $_{\theta}$ : Rayleigh ratio or the ratio of scattered light to incident light of the sample). The inverse of the intercept at the Y-axis gives the molecular weight in KDa. From this value, the mol% of the catalyst relative to the substrate 1 can be calculated.



**Figure S2:** Molecular weight determination for the CQDs using the SLS method: plot of  $KC/R_{\theta}$  vs. concentration.

#### 4.4 Procedure for preparation of substrates: 9-18

The substrates *O*-aryl carbamothioates 1 were prepared using one of the following methods (**Scheme S2** and **Scheme S3**), and characterized using <sup>1</sup>H and <sup>13</sup>C NMR as well as ESI-MS techniques:

### 4.4.1 General procedure for the preparation of substrates from *N*,*N*-dimethyl thiocarbamoyl chloride

**Scheme S2:** General scheme to access *O*-aryl carbamothioates from phenols starting from *N*,*N*-dimethyl thiocarbamoyl chloride.

- a) In a clean, dried round-bottom flask equipped with a magnetic stir-bar, *N*,*N*-dimethyl thiocarbamoyl chloride (1.5 equiv.) was dissolved in anhydrous DMF/NMP under Ar atmosphere. The phenol counterpart (1 equiv.) along with a suitable base (DABCO, 1.5 equiv.) was sequentially added. The reaction mixture was stirred for 16-24 h (monitored by TLC). Reaction temperature was maintained at 25-50°C, depending on the substrate. After the completion of the reaction, cold H<sub>2</sub>O was added to the reaction mixture. The work-up and purification was achieved in one of the following ways:
  - i) If addition of water results in the formation of a precipitate, the latter was collected via filtration. If required, recrystallization can be done using EtOH or EtOH/H<sub>2</sub>O mixture. In case the washing with water gives precipitate of phenol and the desired *O*-aryl carbamothioate together, a suitable base (such as 0.3 M NaOH) can be used to remove the excess phenol derivative (provided no base sensitive functionalities are present) by first dissolving the precipitate in Et<sub>2</sub>O prior to workup. The *O*-aryl

- carbamothioate can be retrieved post workup by concentrating the organic phase under reduced pressure.
- ii) If no precipitation was observed upon addition of cold H<sub>2</sub>O, the reaction mixture was extracted with EtOAc (3 × 20 mL). The combined organic phases were washed with brine and dried over anhydrous sodium sulphate. The solvent was removed by rotary evaporation. The *O*-aryl carbamothioate was obtained by purification using column chromatography with silica gel (100-200 mesh) using EtOAc/Hexane solvent. If the phenol derivative and *O*-aryl carbamothioate eluted together under the column conditions, the excess phenol counterpart was removed by dissolving the mixture in in Et<sub>2</sub>O, followed by washing with a suitable base (such as 0.3 M NaOH), provided no base sensitive functionalities are present. The *O*-aryl carbamothioate was retrieved post workup by concentrating the organic phase under reduced pressure.
- b) A clean, dried round-bottom flask equipped with a magnetic stir-bar was purged with Ar. NaH (1.2 equiv.) was taken in the flask. Anhydrous DMF was added slowly with stirring after placing the flask in ice. The phenol derivative (1 equiv.), dissolved in anhydrous DMF, was added dropwise to the flask over 5-10 mins. Then, the *N*,*N*-dimethyl thiocarbamoyl chloride (1.2 equiv.) was added in a single portion. The flask was gradually heated to 50-80°C with stirring for 1-3 h. The setup was allowed to come to room temperature, followed by gradual transfer to a separatory funnel containing Et<sub>2</sub>O. The workup was done with H<sub>2</sub>O and the organic layer was collected over anhydrous sodium sulphate. The solvent was removed under the effect of vacuum. The crude reaction mixture was finally purified through column chromatography on silica gel (100200 mesh) using EtOAc/Hexane eluent. If the phenol analogue and *O*-aryl carbamothioate eluted together under the column conditions, the excess phenol moiety was

removed by dissolving the mixture in in Et<sub>2</sub>O, followed by washing with a suitable base (such as 0.3 M NaOH), provided no base sensitive functionalities are present. The *O*-aryl carbamothioate was retrieved post workup by concentrating the organic phase under reduced pressure.

### **4.4.2** General procedure for the preparation of substrates from *N*,*N*-diethyl thiocarbamoyl chloride

**Scheme S3:** General scheme to access *O*-aryl carbamothioates from phenols starting from *N*,*N*-diethylthiocarbamoyl chloride.

In a clean, dried round-bottom flask equipped with a magnetic stir-bar, the phenol derivative (1 equiv.), *N*,*N*-diethylthiocarbamoyl chloride (1.1 equiv.), DMAP (0.1 equiv.) and triethylamine (2 equiv.) were taken. The flask was purged with Ar and 1,4-dioxane was added. The reaction mixture was refluxed for 18-24 h. The setup was cooled to room temperature, poured on a mixture of 12 N HCl and ice. The extraction was done with Et<sub>2</sub>O and the combined organic phases were washed with 1 N HCl and 1 N NaOH, followed by collection over Na<sub>2</sub>CO<sub>3</sub>. The solvent was removed with a rotary evaporator. If required, further purification was done using column chromatography with silica gel (100-200 mesh) in EtOAc/Hexane solvent system to obtain the *O*-aryl carbamothioate derivative.

#### 4.5 General procedure for the cyclic voltammetry (CV) studies:

The Ag/AgCl electrode was taken as the reference and cleaned with distilled water. Glassy carbon electrode was chosen as the working electrode and cleaned by rubbing on alumina sheet followed by rinsing with water. The Pt wire electrode was selected as the counter electrode and cleaned by

dipping in conc. HCl, followed by heating over a burner flame and finally rinsing with distilled water.

Distilled water was used to prepare the solutions. A total of 40 mL of 5 mM K<sub>4</sub>[Fe(CN)<sub>6</sub>] in 0.1 M KCl was taken with and without the CQDs, and CV studies were carried out. The scan rate was kept constant at 0.05 V/s throughout the experiment. The voltammograms were recorded in static mode, with stirring between successive experiments to prevent deposition of any analyte or material on the disc of the working electrode.

#### 4.6 General procedure for the crossover experiments:

A clean, dried reaction tube equipped with a magnetic stir-bar was taken and cooled under the effect of Ar flow. 0.125 mmol of the *O*-aryl carbamothioates **1t** and **1j** were taken in the reaction tube. The tube was sealed with a septum and degassed under the effect of vacuum, followed by purging with Ar. The CQDs (2.5 wt% relative to **1t** and 2.5 wt% relative to **1j**, dissolved in 50 μL of H<sub>2</sub>O) along with 4 mL of anhydrous MeCN (distilled to dryness with P<sub>2</sub>O<sub>5</sub> and stored over activated 4 Å MS) were added. The reaction mixture was ultra-sonicated (if required) and subsequently irradiated for the stated time using a Kessil Lamp (370 nm, PR160L). Post irradiation, the reaction mixture may be extracted with EtOAc (3 X 10 mL) (removal of MeCN under reduced pressure is sufficient otherwise). The combined organic phases were washed with brine and dried over anhydrous sodium sulphate. The solvent was removed by rotary evaporation. The products formed in the reaction were quantified using <sup>1</sup>H-NMR, taking Dibromomethane as the reference in CDCl<sub>3</sub> solvent.

#### 5. Measurement of quantum yield:1

7-hydroxycoumarin was taken as the reference to measure the quantum yield (QY) of the CQDs. The UV-visible absorption and fluorescence emission spectra was recorded for the reference as well as the sample in H<sub>2</sub>O. QY was determined using the following general formula:

where:

 $QY_{ref.} = QY$  of reference

 $QY_{sample} = QY$  of sample (in this case, CQDs)  $\eta = \eta_{ref.} = 1.33$ 

(refractive index of medium; in this case H<sub>2</sub>O)

 $A_{ref.}$  = Absorbance of reference at  $\lambda_{max}$ 

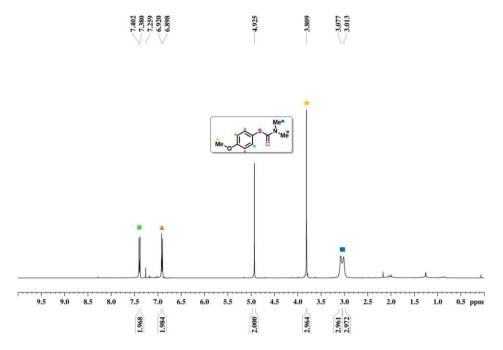
 $A_{\text{sample}} = Absorbance \text{ of sample at } \lambda_{\text{max}}$ 

 $I_{ref.}$  = Integrated intensity of reference

 $I_{sample} = Integrated intensity of sample$ 

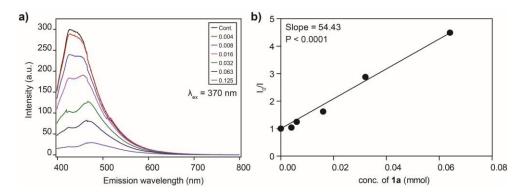
#### 6. Spectra of the crude reaction mixture:

We present the  ${}^{1}$ H-NMR spectrum of the crude reaction mixture, recorded in CDCl<sub>3</sub>, ruling out the formation of any side products. For this, the reaction was done as per the optimal conditions (Primary Manuscript: Table 1, Entry 9) and the reaction mixture may be subjected to EtOAc/H<sub>2</sub>O workup. The organic layer was collected, concentrated and CDCl<sub>3</sub> was added along with Dibromomethane ( $\delta$  (ppm) = 4.93) as the reference. The resultant spectrum clearly indicates the absence of any side product (**Figure S3**).



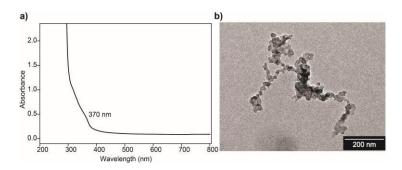
**Figure S3:** NMR spectrum of the crude reaction mixture, recorded in CDCl<sub>3</sub> solvent with Dibromomethane as the reference.

#### 7. Stern-Volmer quenching experiment:<sup>19</sup>



**Figure S4:** Stern-Volmer quenching experiment using CQDs: a) reduction in the fluorescence intensity upon increasing concentration of **1a** (the numbers represent the concentration in mmol) and b) calculation of the Stern-Volmer quenching constant.

#### 8. Characterization of the recycled CQDs:



**Figure S5:** Characterization of the CQD post 10 runs of the NKR: a) UV-visible absorption profile and b) TEM image of the recycled catalyst.

#### 9. Estimation of EcoScale value (considering the model substrate 1a):

Parameter	Penalty	Explanation
	Points	
Yield	1	(100 – % yield)/2
Price of reaction components (to obtain 10 mmol of end product)	0	Inexpensive reagents
Safety	5	Highly flammable
Technical setup	1	Inert gas
Temp./Time	0	Room temp./<1 h
Workup and purification	0	Removal of solvent with bp < 150°C
Workup and purification	10	Classical chromatography
TOTAL	17	
EcoScale (= 100 – Penalty Points)	83	

#### **10.** Characterization data: 9-11, 13-16, 20-24

The substrates prepared by any of the methods from **Section 4.3** were characterized subsequently by <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy, as well as ESI-MS technique. The data was verified from literature reports to confirm the formation of the pure *O*-aryl carbamothioate. The data for those substrates that have not been reported in literature, are presented below:

#### **O-(2,3-dimethylphenyl) dimethylcarbamothioate:** Light orange solid; isolated using silica gel

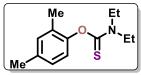
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(100-200 mesh) column chromatography in 3-4% EtOAc/Hexane eluent;

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.14-7.10 (t, J = 7.7 Hz, 1H),

7.07-7.05 (d, J = 7.4 Hz, 1H), 6.86-6.84 (d, J = 7.9 Hz, 1H), 3.47 (s, 3H), 3.37 (s, 3H), 2.31 (s, 3H), 2.08 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 187.96, 152.94, 138.77, 129.91, 127.96, 126.26, 120.96, 43.70, 38.99, 20.53, 13.04; **HRMS (ESI+):** Calculated for C<sub>11</sub>H<sub>15</sub>NOS [M+H]<sup>+</sup>: 210.0953, Found [M+H]<sup>+</sup>: 210.0950.

#### O-(2,4-dimethylphenyl) diethylcarbamothioate: Orange solid; precipitated as per the

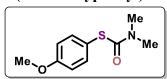


procedure stated in section 6.5.3.2; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.04-7.01 (m, 2H), 6.87-6.85 (m, 1H), 3.94-3.88 (q, J = 7.1 Hz,

2H), 3.73-3.67 (q, J = 7.1 Hz, 2H), 2.32 (s, 3H), 2.17 (s, 3H), 1.35-1.31 (m, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 186.91, 150.72, 135.96, 132.10, 130.89, 127.76, 122.96, 48.71, 44.43, 21.35, 16.80, 13.95, 12.30; **HRMS (ESI+):** Calculated for C<sub>13</sub>H<sub>19</sub>NOS [M+H]<sup>+</sup>: 238.1266, Found [M+H]<sup>+</sup>: 238.1274.

The data for the rearranged products are given below:

#### S-(4-methoxyphenyl) dimethylcarbamothioate: Yellowish-white solid; isolated using silica



gel (100-200 mesh) column chromatography in 3-4% EtOAc/Hexane eluent; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.41-7.38 (d, J = 8.8

Hz, 2H), 6.92-6.90 (d, J = 8.8 Hz, 2H), 3.81 (s, 3H), 3.05 (br s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 168.11, 160.99, 137.75, 119.86, 115.05, 55.77, 37.29; **HRMS (ESI+)**:

Calculated for  $C_{10}H_{13}NO_2S$  [M+H]<sup>+</sup>: 212.0745, Found [M+H]<sup>+</sup>: 212.0741.

S-phenyl dimethylcarbamothioate: Yellow liquid; isolated using silica gel (100-200 mesh)

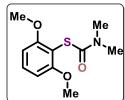
S Me N Me

column chromatography in 3-4% EtOAc/Hexane eluent; <sup>1</sup>H NMR (400

MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.52-7.48 (m, 2H), 7.39-7.37 (m, 3H), 3.08 (s,

3H), 3.04 (s, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 136.27, 129.68, 129.45, 37.45; **HRMS** (**ESI+**): Calculated for C<sub>9</sub>H<sub>11</sub>NOS [M+H]<sup>+</sup>: 182.0640, Found [M+H]<sup>+</sup>: 182.0642.

S-(2,6-dimethoxyphenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100-



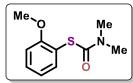
200 mesh) column chromatography in 13-14% EtOAc/Hexane eluent;  $^1\!\mathbf{H}$ 

**NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.38-7.34 (t, J = 8.4 Hz, 1H), 6.63-

6.61 (d, J = 8.4 Hz, 2H), 3.86 (s, 6H), 3.18 (s, 3H), 2.99 (s, 3H); <sup>13</sup>C NMR

(100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.24, 161.90, 132.22, 105.31, 104.73, 56.89, 37.65; **HRMS** (ESI+): Calculated for C<sub>11</sub>H<sub>15</sub>NO<sub>3</sub>S [M+H]<sup>+</sup>: 242.0851, Found [M+H]<sup>+</sup>: 242.0856.

S-(2-methoxyphenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100-200

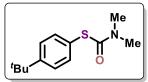


mesh) column chromatography in 4-5% EtOAc/Hexane eluent;  $^1H$  NMR

(400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.47-7.45 (m, 1H), 7.42-7.38 (m, 1H),

6.99-6.95 (m, 2H), 3.87 (s, 3H), 3.13 (s, 3H), 3.01 (s, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.77, 160.55, 138.59, 131.97, 121.42, 117.14, 111.91, 56.54, 37.42; **HRMS (ESI+):** Calculated for C<sub>10</sub>H<sub>13</sub>NO<sub>2</sub>S [M+H]<sup>+</sup>: 212.0745, Found [M+H]<sup>+</sup>: 212.0747.

S-(4-(tert-butyl)phenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100-



200 mesh) column chromatography in 2-3% EtOAc/Hexane eluent; <sup>1</sup>H

**NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.41-7.34 (m, 4H), 3.09 (s, 3H),

3.01 (s, 3H), 1.31 (s, 9H);  ${}^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 152.70, 149.52, 148.27,

135.79, 126.51, 126.47, 125.59, 121.42, 37.27, 35.09, 34.78, 31.82, 31.61, 31.33; **HRMS** 

(ESI+): Calculated for C<sub>13</sub>H<sub>19</sub>NOS [M+H]<sup>+</sup>: 238.1266, Found [M+H]<sup>+</sup>: 238.1264. S-(2,6-dimethylphenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100-200 mesh) Me Me N Me

column chromatography in 2-3% EtOAc/Hexane eluent; <sup>1</sup>H NMR (400 MHz,

CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.22-7.18 (m, 1H), 7.15-7.13 (m, 2H), 3.16 (s, 3H), 3.02 (s, 3H),

2.42 (s, 6H);  ${}^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.48, 144.15, 130.00,

128.59, 37.45, 22.52; **HRMS (ESI+):** Calculated for C<sub>11</sub>H<sub>15</sub>NOS [M+H]<sup>+</sup>: 210.0953, Found [M+H]<sup>+</sup>: 210.0951.

S-(2,4-dimethylphenyl) dimethylcarbamothioate: White solid; isolated using silica gel

(100200 mesh) column chromatography in 2-3% EtOAc/Hexane eluent;

 $^{1}H$ 

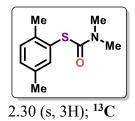
NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.35-7.33 (d, J = 7.8 Hz, 1H), 7.12 (s, 1H), 7.02-7.00 (d, J = 7.8 Hz, 1H), 3.12 (s, 3H), 3.01 (s, 3H), 2.37 (s, 3H), 2.32 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.90, 142.79, 140.06, 137.02, 131.50, 127.29, 124.70, 36.94, 21.25, 20.97; HRMS (ESI+): Calculated for C<sub>11</sub>H<sub>15</sub>NOS [M+H]<sup>+</sup>: 210.0953, Found [M+H]<sup>+</sup>: 210.0952.

S-(2,3-dimethylphenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100200

mesh) column chromatography in 2-3% EtOAc/Hexane eluent; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.36-7.34 (d, J = 7.6 Hz, 1H), 7.21-

7.19 (d, J = 7.6 Hz, 1H), 7.11-7.07 (t, J = 7.6 Hz, 1H), 3.13 (s, 3H), 3.02 (s, 3H), 2.36 (s, 3H), 2.31 (s, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 167.23, 141.92, 138.14, 135.21, 131.95, 128.55, 126.13, 37.30, 21.43, 17.92; **HRMS** (**ESI+**): Calculated for C<sub>11</sub>H<sub>15</sub>NOS [M+H]<sup>+</sup>: 210.0953, Found [M+H]<sup>+</sup>: 210.0949.

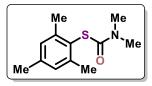
S-(2,5-dimethylphenyl) dimethylcarbamothioate: White solid; isolated using silica gel (100-



200 mesh) column chromatography in 2-3% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.30 (s, 1H), 7.19-7.17 (d, J = 7.8 Hz, 1H), 7.12-7.10 (d, J = 7.6 Hz, 1H), 3.12 (s, 3H), 3.02 (s, 3H), 2.36 (s, 3H),

**NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.78, 139.86, 137.58, 135.97, 130.78, 130.63, 130.40, 127.68, 126.23, 122.78, 36.97, 20.75, 20.56; **HRMS** (**ESI+**): Calculated for C<sub>11</sub>H<sub>15</sub>NOS [M+H]<sup>+</sup>: 210.0953, Found [M+H]<sup>+</sup>: 210.0956.

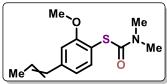
S-mesityl dimethylcarbamothioate: White solid; isolated using silica gel (100-200 mesh)



column chromatography in 7-8% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 6.97 (s, 2H), 3.15 (s, 3H), 3.02 (s, 3H), 2.38 (s,

6H), 2.28 (s, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.79, 143.85, 139.95, 129.51, 125.06, 37.40, 22.36, 21.57; **HRMS (ESI+):** Calculated for  $C_{12}H_{17}NOS [M+H]^+$ : 224.1109, Found  $[M+H]^+$ : 224.1107.

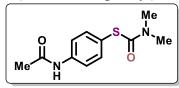
S-(2-methoxy-4-(prop-1-en-1-yl)phenyl) dimethylcarbamothioate: Yellowish-white solid;



isolated using silica gel (100-200 mesh) column chromatography in 1-2% EtOAc/Hexane eluent;  $^1$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm)

= 7.36-7.34 (d, J = 7.9 Hz, 1H), 6.94-6.92 (d, J = 7.6 Hz, 1H), 6.89 (s, 1H), 6.41-6.37 (d, J = 15.5 Hz, 1H), 6.32-6.23 (m, 1H), 3.87 (s, 3H), 3.12 (s, 3H), 3.00 (s, 3H), 1.89-1.88 (d, J = 6.3 Hz, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.79, 160.46, 141.88, 138.40, 131.08, 127.73, 119.01, 114.96, 109.17, 56.38, 37.31, 18.87; **HRMS** (**ESI**+): Calculated for C<sub>13</sub>H<sub>17</sub>NO<sub>2</sub>S [M+H]<sup>+</sup>: 252.1058, Found [M+H]<sup>+</sup>: 252.1053.

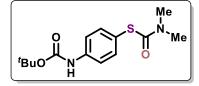
**S-(4-acetamidophenyl) dimethylcarbamothioate:** White solid; isolated using silica gel (100200



mesh) column chromatography in 35-37% EtOAc/Hexane eluent;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.82 (br s, 1H),

7.42-7.40 (d, J = 8.5 Hz, 2H), 7.36-7.34 (d, J = 8.3 Hz, 2H), 3.10 (s, 3H), 3.04 (s, 3H), 2.09 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 168.94, 168.46, 139.85, 136.72, 122.76, 120.67, 37.38, 24.75; **HRMS (ESI+):** Calculated for C<sub>11</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>S [M+Na]<sup>+</sup>: 261.0674, Found [M+Na]<sup>+</sup>: 261.0677.

tert-butyl (4-((dimethylcarbamoyl)thio)phenyl)carbamate: White solid; isolated using silica



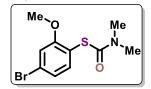
gel (100-200 mesh) column chromatography in 32-33%

EtOAc/Hexane eluent; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) =

7.38 (s, 4H), 6.56 (s, 1H), 3.08 (s, 3H), 3.00 (s, 3H), 1.51 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 139.95, 137.10, 122.63, 119.08, 37.37, 28.77; HRMS (ESI+): Calculated for  $C_{14}H_{20}N_2O_3S$  [M+Na]<sup>+</sup>: 319.1092, Found [M+Na]<sup>+</sup>: 319.1096.

*Product decomposes during the course of purification.* 

S-(4-bromo-2-methoxyphenyl) dimethylcarbamothioate: White solid; isolated using silica gel



(100-200 mesh) column chromatography in 3-4% EtOAc/Hexane eluent;

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.32-7.30 (d, J = 8.0, 1H), 7.12-

7.10 (dd, J = 8.2 Hz, J = 1.5 Hz, 1H), 7.08-7.07 (d, J = 1.8 Hz, 1H), 3.86 (s, 3H), 3.12 (s, 3H),

3.00 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.11, 161.00, 139.36, 125.66, 124.57,

116.55, 115.62, 56.84, 37.45; **HRMS (ESI+):** Calculated for C<sub>10</sub>H<sub>12</sub>BrNO<sub>2</sub>S [M+H]<sup>+</sup>: 289.9850,

Found [M+H]<sup>+</sup>: 289.9856.

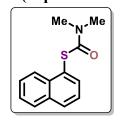
S-(naphthalen-2-yl) dimethylcarbamothioate: Yellowish-white solid; isolated using silica gel



(100-200 mesh) column chromatography in 3-4% EtOAc/Hexane eluent;  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 8.02 (s, 1H), 7.85-7.77

(m, 3H), 7.57-7.43 (m, 3H), 3.16 (s, 3H), 3.05 (s, 3H);  ${}^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 167.50, 135.76, 133.91, 133.72, 132.71 128.90, 128.40, 128.16, 127.37, 126.76, 126.48, 37.39;

**HRMS** (**ESI+**): Calculated for C<sub>13</sub>H<sub>13</sub>NOS [M+H]<sup>+</sup>: 232.0796, Found [M+H]<sup>+</sup>: 232.0797. **S-(naphthalen-1-yl) dimethylcarbamothioate:** Yellowish-white solid; isolated using silica gel



(100-200 mesh) column chromatography in 3-4% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 8.33-8.31 (d, J = 8.2 Hz, 1H), 7.95-7.93 (d, J = 8.2 Hz, 1H), 7.88-7.86 (d, J = 8.2 Hz, 1H), 7.79-7.77 (d, J = 7.2 Hz,

1H), 7.59-7.55 (t, J = 7.5 Hz, 1H), 7.52-7.47 (q, J = 7.8 Hz, 2H), 3.24 (s, 3H), 3.02 (s, 3H); <sup>13</sup>C **NMR** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 167.04, 136.61, 135.65, 134.63, 131.27, 129.01, 127.50, 126.67, 126.57, 126.18, 126.00, 37.59; **HRMS** (**ESI+**): Calculated for C<sub>13</sub>H<sub>13</sub>NOS [M+H]<sup>+</sup>: 232.0796, Found [M+H]<sup>+</sup>: 232.0799.

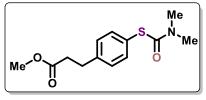
S-(7-bromonaphthalen-2-yl) dimethylcarbamothioate: White solid; isolated using silica gel

(100- 200 mesh) column chromatography in 5-6% EtOAc/Hexane eluent; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.97-7.91 (m, 1H),

7.83-7.78 (m, 1H), 7.72-7.68 (m, 1H), 7.59-7.54 (m, 1H), 7.52-7.48 (m, 1H), 7.30-7.27 (m, 1H), 3.15 (s, 3H), 3.04 (s, 3H); **HRMS (ESI+):** Calculated for C<sub>13</sub>H<sub>12</sub>BrNOS [M+H]<sup>+</sup>: 309.9901, Found [M+H]<sup>+</sup>: 309.9905.

Product decomposes very rapidly, hence <sup>13</sup>C-NMR spectrum could not be recorded.

methyl 3-(4-((dimethylcarbamoyl)thio)phenyl)propanoate: White solid; isolated using silica



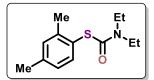
gel (100- 200 mesh) column chromatography in 3-5% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) =

7.41-7.39 (d, J = 8.3 Hz, 2H), 7.23-7.21 (d, J = 8.2 Hz, 2H),

3.67 (s, 3H), 3.08 (s, 3H), 3.03 (s, 3H), 2.98-2.94 (t, J = 7.9 Hz, 2H), 2.65-2.61 (t, J = 7.8 Hz, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 173.58, 142.20, 136.31, 129.38, 126.81, 52.08,

37.31, 35.78, 31.05; **HRMS (ESI+):** Calculated for C<sub>13</sub>H<sub>17</sub>NO<sub>3</sub>S [M+H]<sup>+</sup>: 268.1007, Found [M+H]<sup>+</sup>: 268.1007.

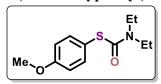
S-(2,4-dimethylphenyl) diethylcarbamothioate: White solid; isolated using silica gel (100-200



mesh) column chromatography in 6-8% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.37-7.35 (d, J = 7.7 Hz, 1H), 7.11

(s, 1H), 7.01-6.99 (d, J = 7.7 Hz, 1H), 3.44 (br s, 4H), 2.36 (s, 3H), 2.31 (s, 3H), 1.28 (s, 3H), 1.15 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.10, 143.22, 140.37, 137.52, 131.92, 127.69, 125.30, 42.95, 42.88, 21.69, 21.42, 14.33, 14.03; **HRMS (ESI+):** Calculated for  $C_{13}H_{19}NOS [M+H]^+$ : 238.1266, Found  $[M+H]^+$ : 238.1268.

S-(4-methoxyphenyl) diethylcarbamothioate: White solid; isolated using silica gel (100-200



mesh) column chromatography in 4-5% EtOAc/Hexane eluent; <sup>1</sup>H

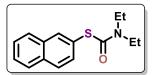
**NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 7.42-7.40 (d, J = 8.8 Hz, 2H),

6.92-6.90 (d, J = 8.7 Hz, 2H), 3.81 (s, 3H), 3.45-3.39 (q, J = 7.3 Hz, 4H), 1.25 (s, 3H), 1.15 (s, 3H);

13C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 166.91, 160.92, 137.84, 119.94, 115.02, 55.80,

42.74, 14.31, 13.61; **HRMS (ESI+):** Calculated for C<sub>12</sub>H<sub>17</sub>NO<sub>2</sub>S [M+H]<sup>+</sup>: 240.1058, Found [M+H]<sup>+</sup>: 240.1060.

S-(naphthalen-2-yl) diethylcarbamothioate: White solid; isolated using silica gel (100-200



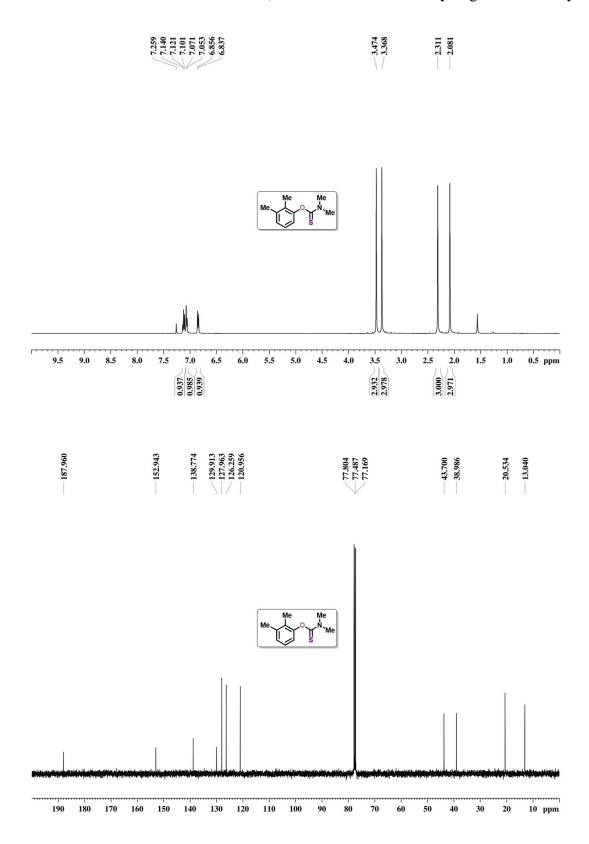
mesh) column chromatography in 4-6% EtOAc/Hexane eluent;  ${}^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 8.03 (s, 1H), 7.85-7.81 (m, 3H),

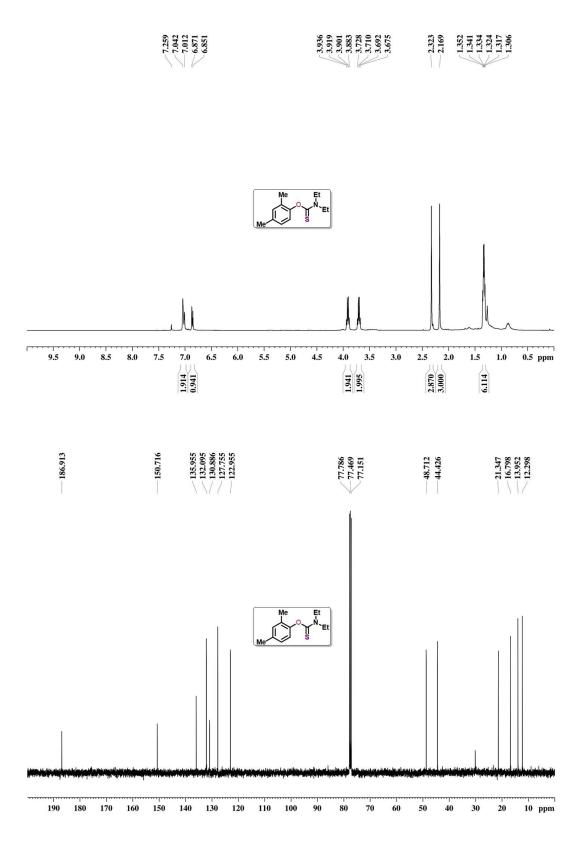
7.57-7.41 (m, 3H), 3.47 (br s, 4H), 1.32-1.28 (t, J = 6.9 Hz, 3H), 1.25-1.21 (m, 3H); <sup>13</sup>C NMR

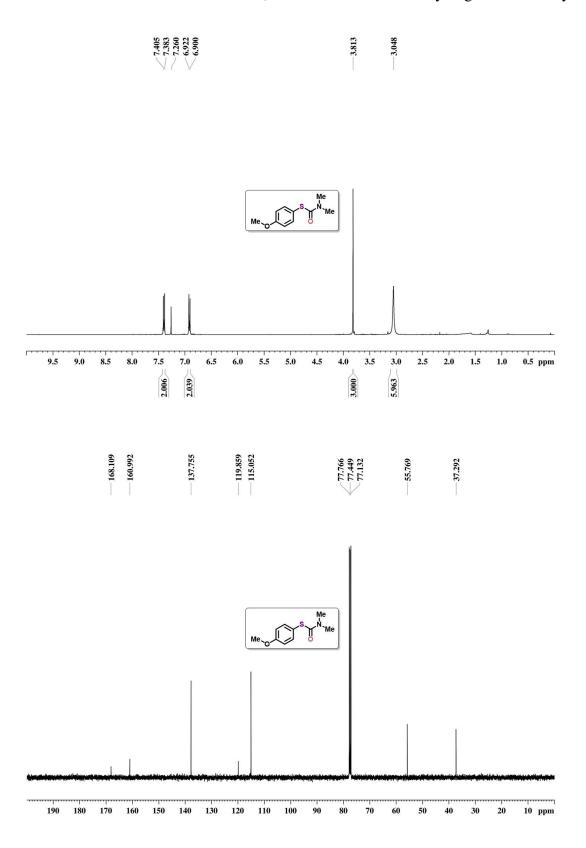
(100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 166.29, 135.76, 133.99, 133.74, 132.87, 128.84, 128.44, 128.18, 127.33, 126.73, 126.69, 42.87, 14.32, 13.90; **DEPT-135** (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) = 135.77, 132.88, 128.85, 128.44, 128.18, 127.34, 126.74, 42.88, 14.34, 13.90; **HRMS (ESI+):** Calculated for C<sub>15</sub>H<sub>17</sub>NOS [M+H]<sup>+</sup>: 260.1109, Found [M+H]<sup>+</sup>: 260.1112.

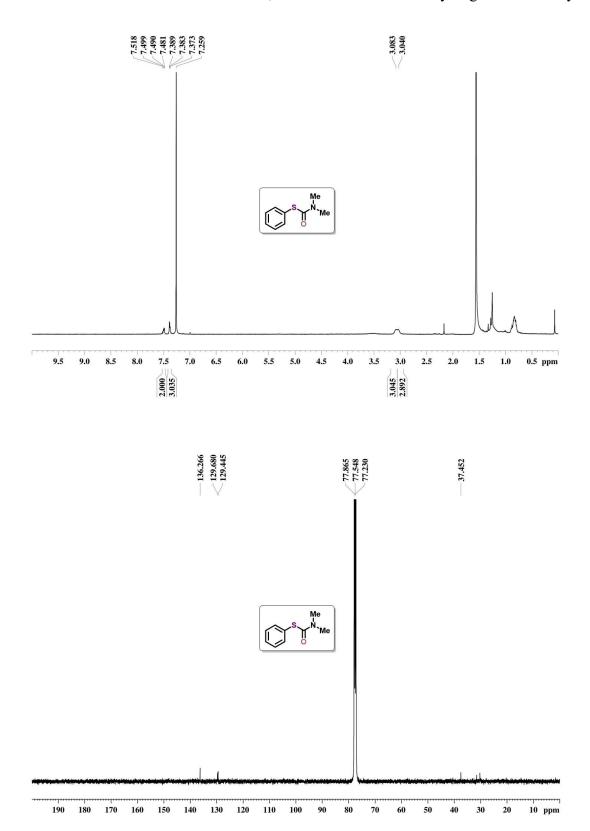
Additional peaks appear in the <sup>13</sup>C-NMR spectrum due to the rapid decomposition of the compound, possibly via oxidation. <sup>25, 26</sup>

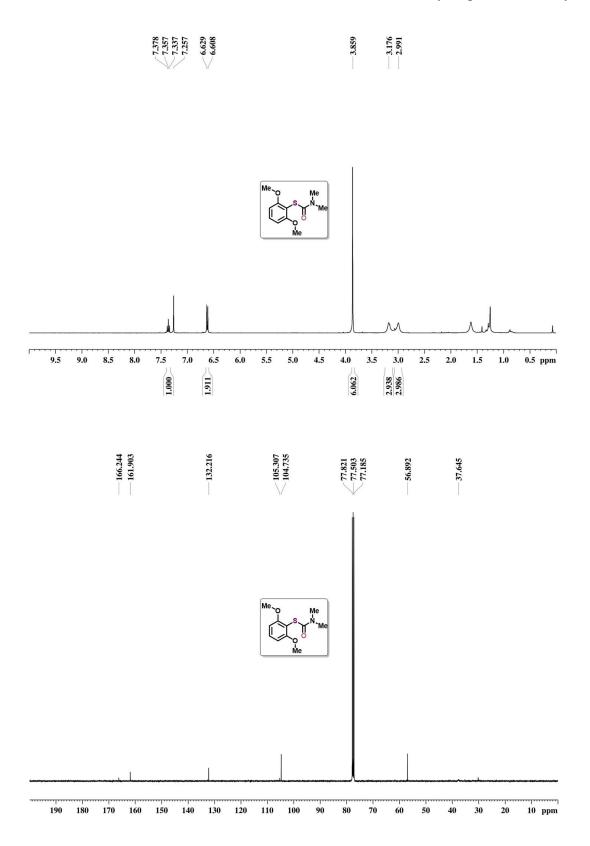
#### 11. NMR Spectra:

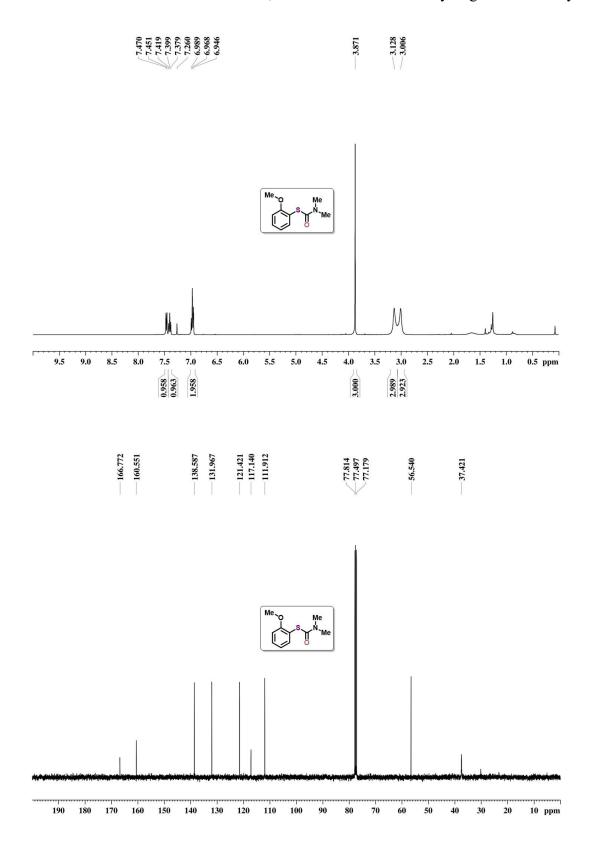


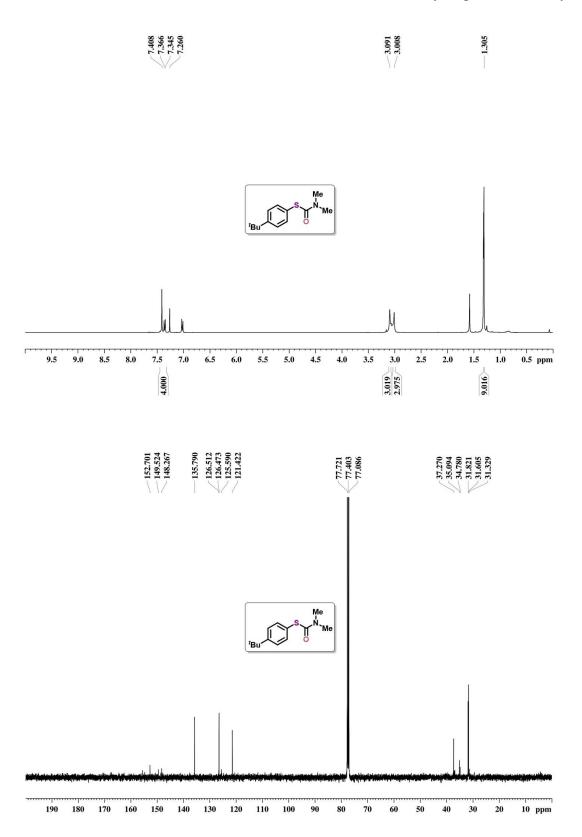


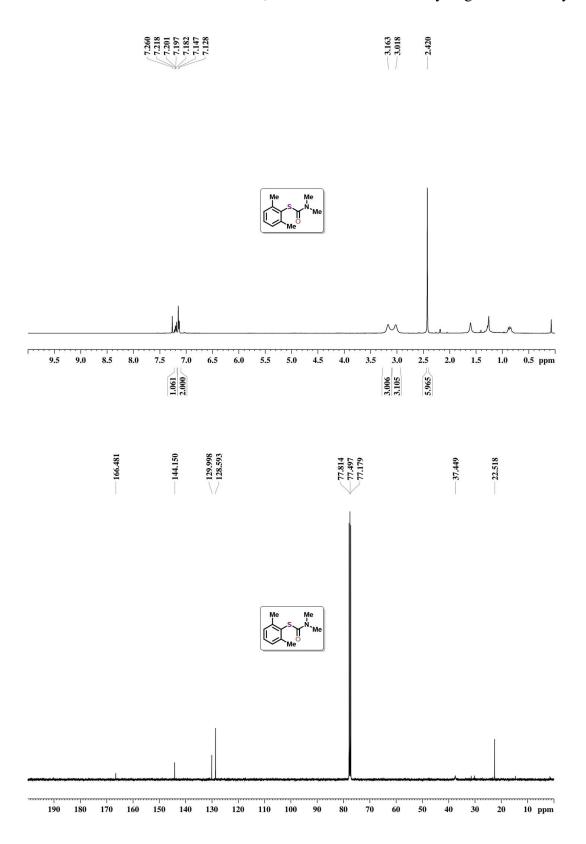


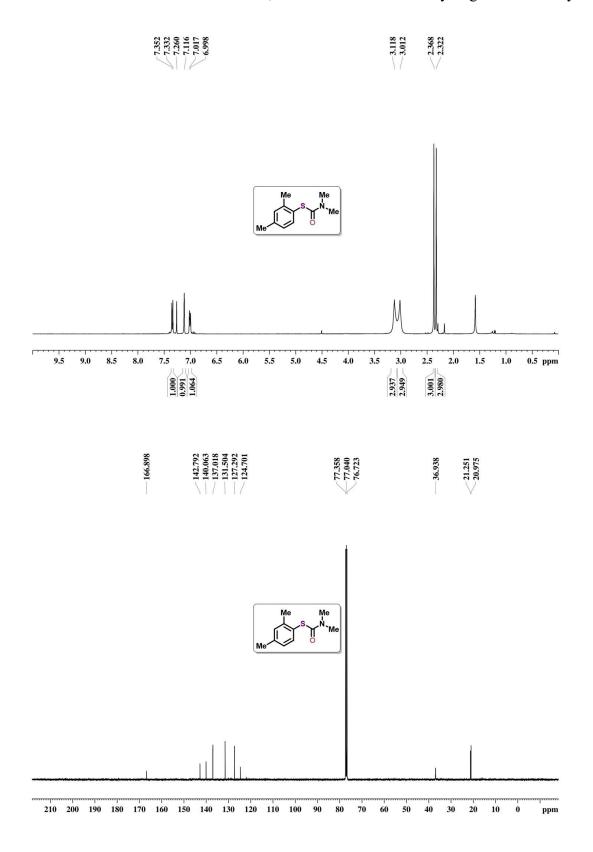


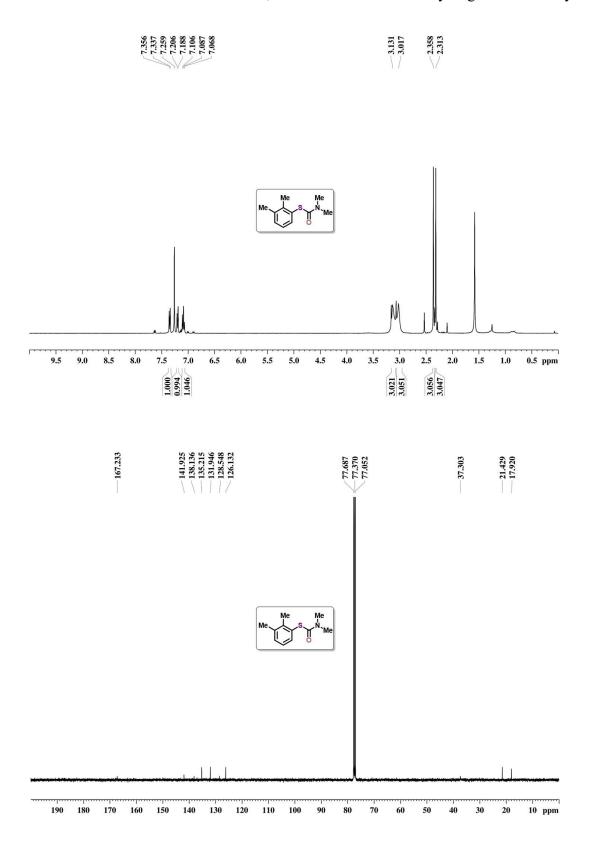


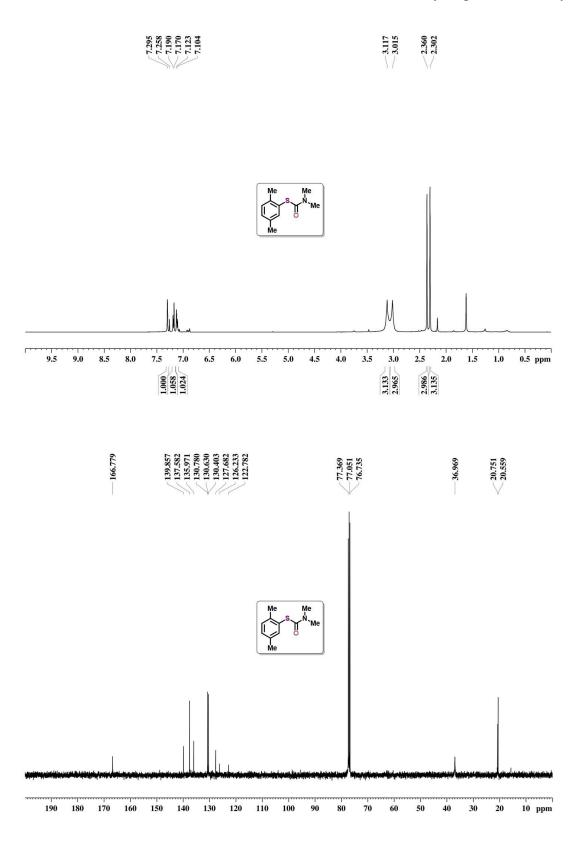


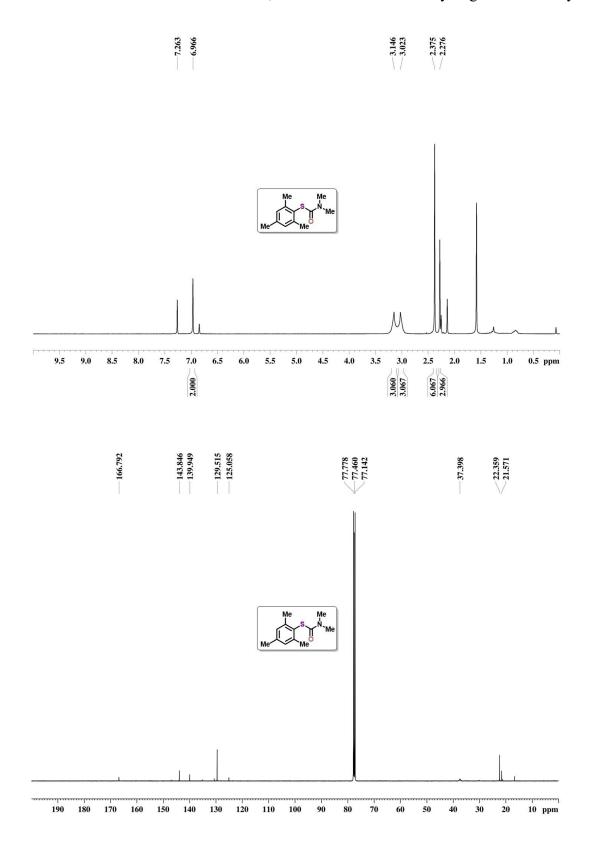


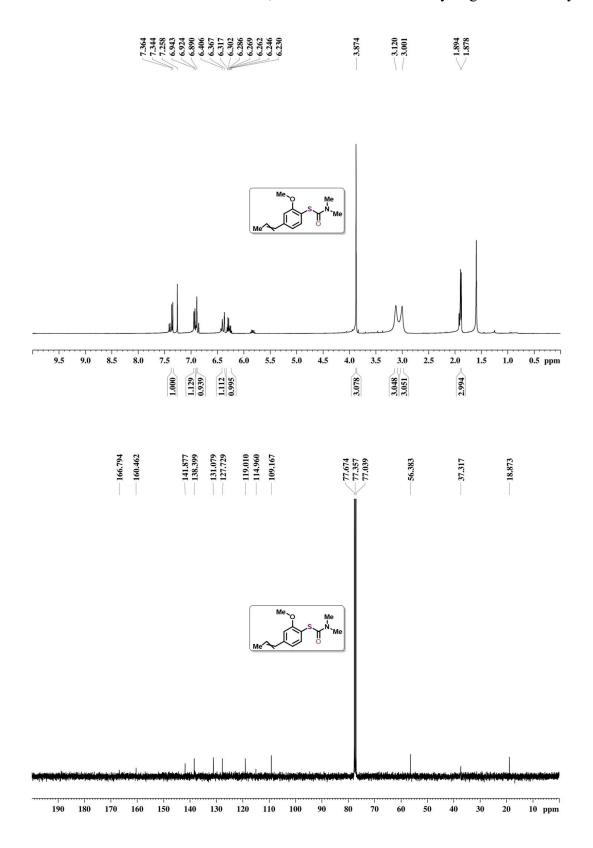


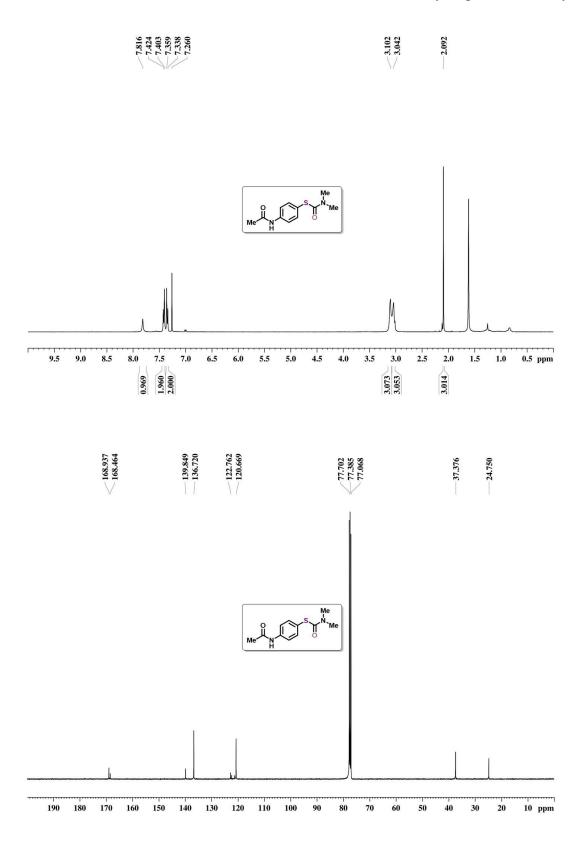


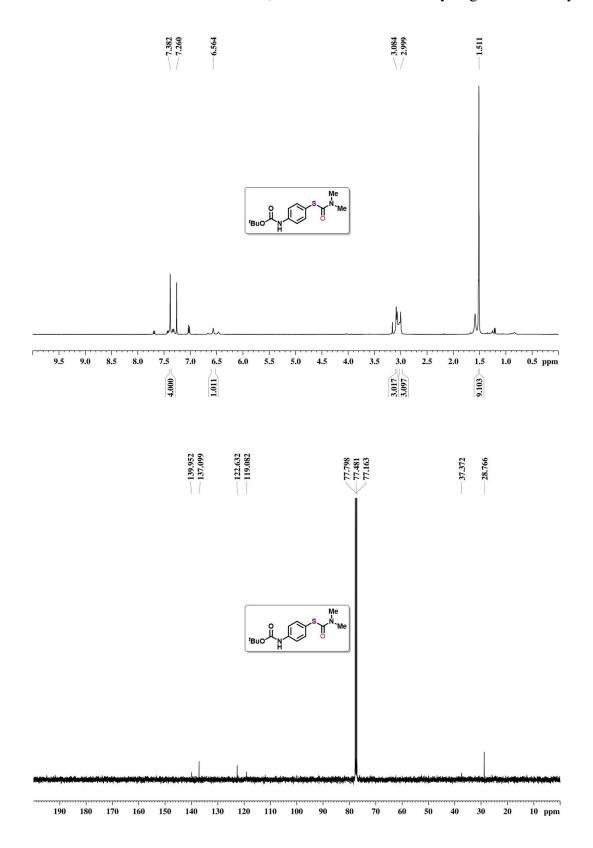


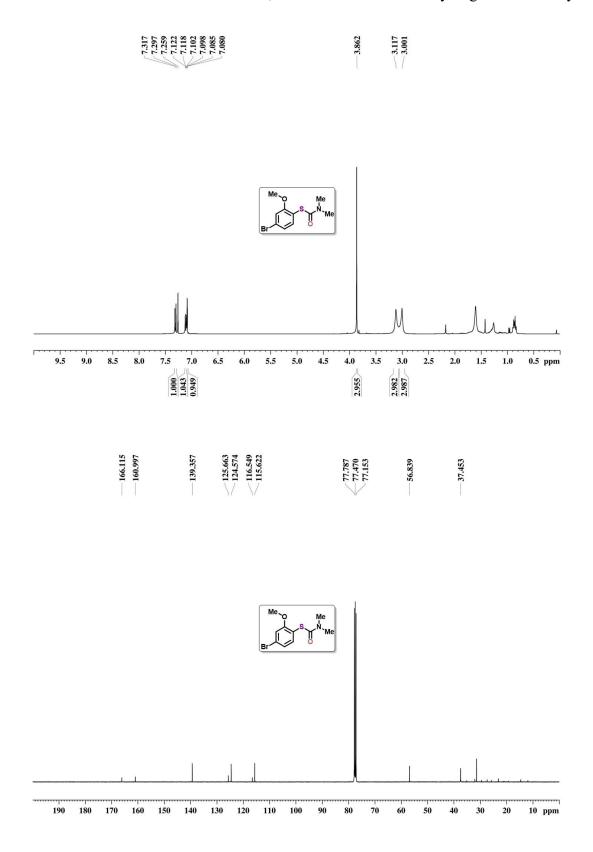


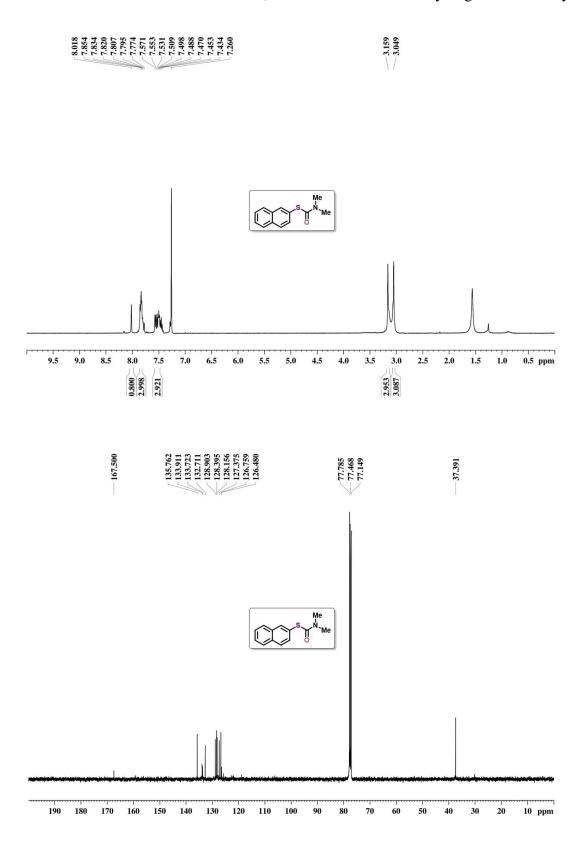


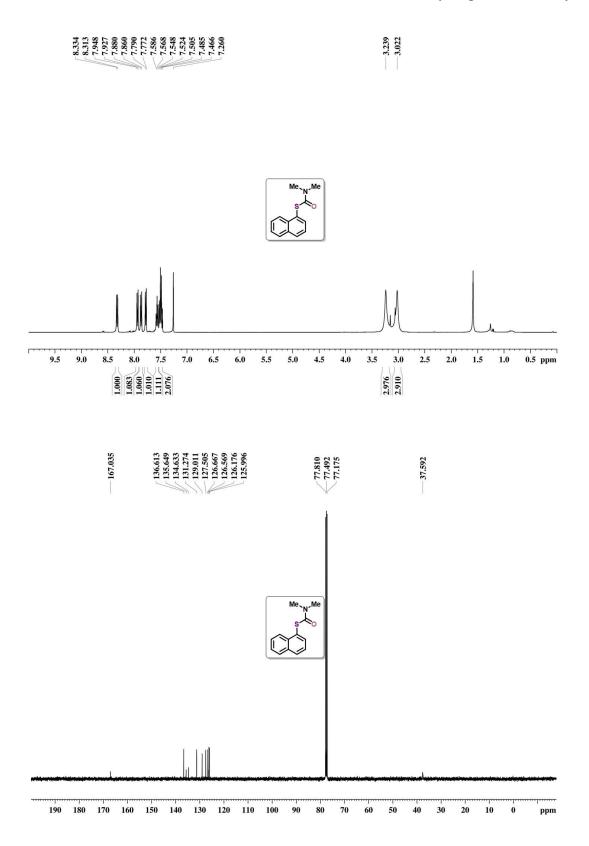


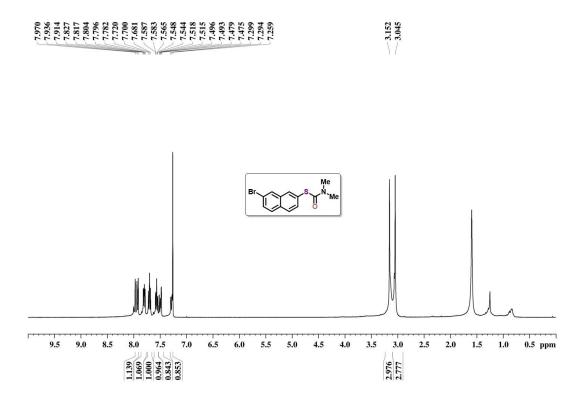




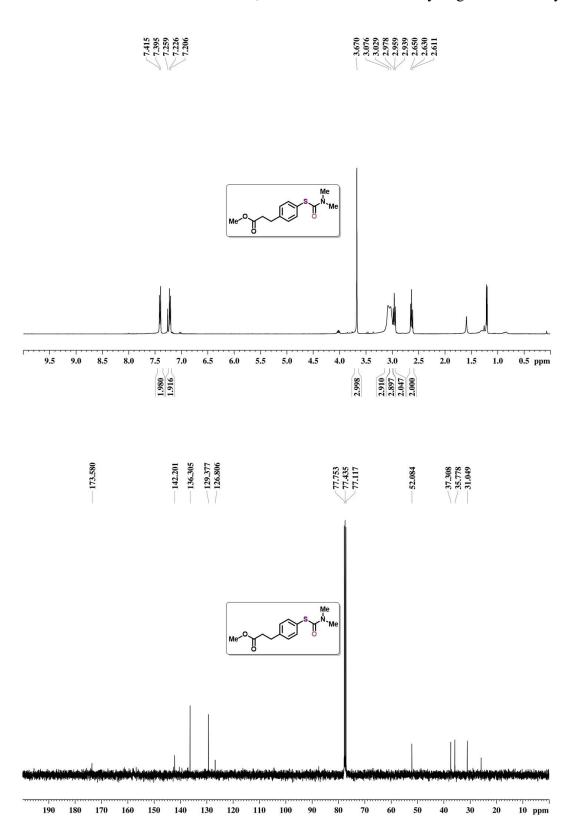


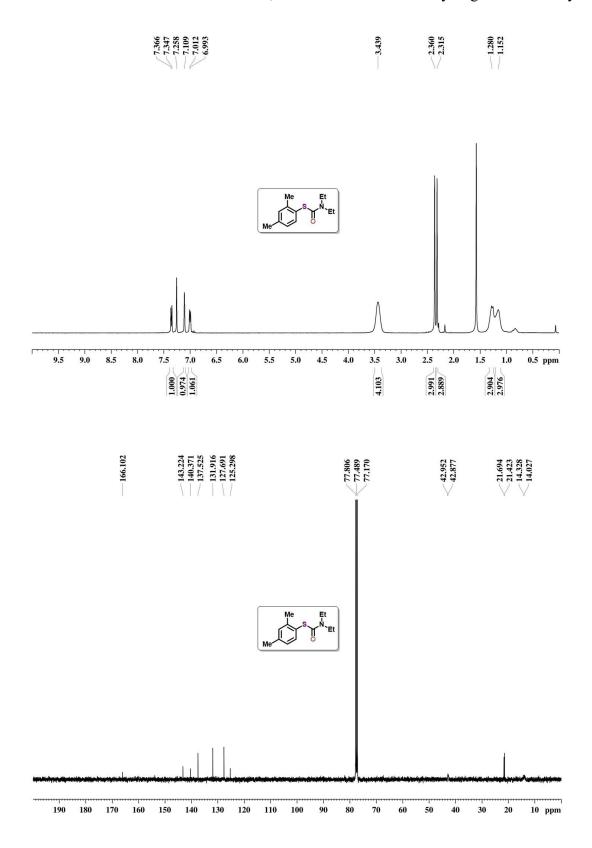


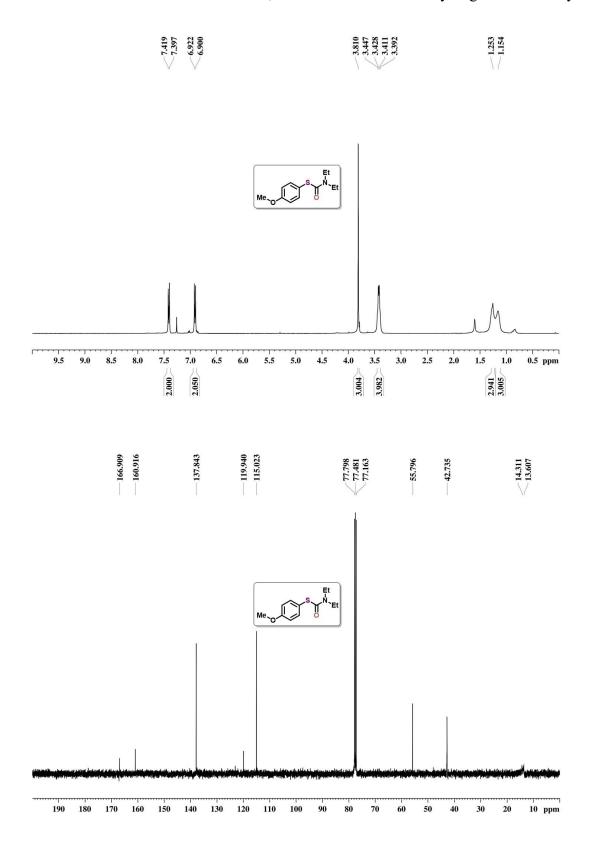


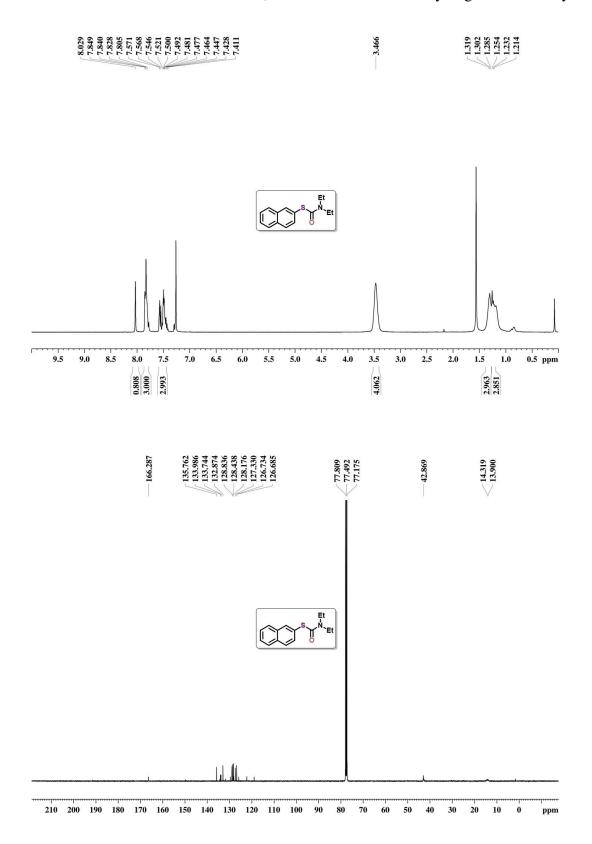


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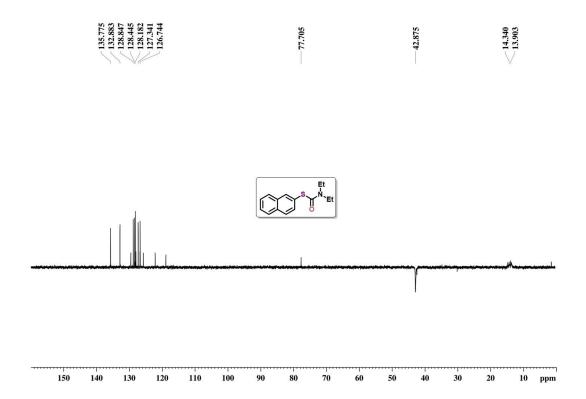








## **DEPT-135:**



## 12. Supplemental references:

- 1. Jaiswal, K.; Girish, Y. R.; Behera, P.; De, M., Dual Role of MoS<sub>2</sub> Quantum Dots in a Cross-Dehydrogenative Coupling Reaction. *ACS Org. Inorg. Au* **2022**, *2*, 205-213.
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