

Supplementary information

High-efficiency CO₂ fixation and single-cell protein production in *Cupriavidus necator* via ARTP-driven metabolic rewiring

Chunling Ma,^{†*ab*} Xiaolei Cheng,^{†*b*} Yuhua Wang,^{†*b*} Ke Chen,^{*b*} Yihua Ma,^{*c*} Yuping Lin,^{*d*} Kun Guo^{*a*} and Zhiguang Zhu^{*be*}

^a School of Chemical Engineering and Technology, Xi'an Jiaotong University, Xi'an 710049, China

^b State Key Laboratory of Engineering Biology for Low-Carbon Manufacturing, Tianjin Institute of Industrial Biotechnology, Chinese Academy of Sciences, Tianjin, 300308, China

^c In vitro Synthetic Biology Center, Tianjin Institute of Industrial Biotechnology, Chinese Academy of Sciences, Tianjin 300308, China

^d Department of Strategic and Integrative Research, Tianjin Institute of Industrial Biotechnology, Chinese Academy of Sciences, Tianjin 300308, China

^e University of Chinese Academy of Sciences, 19A Yuquan Road, Shijingshan District, Beijing 100049, China

* Corresponding Author: Kun Guo (kun.guo@xjtu.edu.cn), Zhiguang Zhu (zhu_zg@tib.cas.cn)

This file includes:

Table S1 Primers for RT-qPCR.

Table S2 Plasmids and primers used in the study.

Table S3 Strains, plasmids and primers used in the study.

Fig. S1. Comparative assessment of mutagenesis strategies for *C. necator*.

Fig. S2. PHB content of H16, R1, R2, R3.

Fig. S3. The codons of *C. necator*.

Fig. S4. The results of strengthening different promoter for A2170 gene or deleting B1682 gene in H16.

Fig. S5. The SDS-PAGE results of engineered strains.

Fig. S6. The growth of *C. necator* H16 and engineered strain HS05.

Fig. S7. The relationship between OD and nitrogen concentration.

Table S1 Primers for RT-qPCR.

Gene	Primer	Sequence (5' → 3')	Gene	Primer	Sequence (5' → 3')
<i>glk</i>	IF- <i>glk</i>	catcctcctgcgctgacttc	<i>cisY</i>	IF- <i>cisY</i>	cctatggcgtacaagtacaac
	IR- <i>glk</i>	catcctcctgcgctgacttc		IR- <i>cisY</i>	ctgcgtgcaggatgaagatg
<i>gap</i>	IF- <i>gap</i>	gccaagaaggtgatcatgtc	<i>odhA</i>	IF- <i>odhA</i>	cggtttcgaattcatgtacg
	IR- <i>gap</i>	cttgatgccccacttgcgt		IR- <i>odhA</i>	cgacgtacttggtgtgcagg
<i>fba</i>	IF- <i>fba</i>	ctacaagttcacgcgcaagc	<i>sdhA</i>	IF- <i>sdhA</i>	tctatgcagcttcaccaac
	IR- <i>fba</i>	atcgcttctggatttctc		IR- <i>sdhA</i>	ggtgaaactgccagaactcc
<i>eno</i>	IF- <i>eno</i>	agaagatcctggccgacaag	<i>mdh</i>	IF- <i>mdh</i>	cctacatcgccatgaagtcg
	IR- <i>eno</i>	cttcgccttcgtgtagaac		IR- <i>mdh</i>	gttgatcaggtccttgacgc
<i>pdhB</i>	IF- <i>pdhB</i>	gaagtcaaggtcaaggtcgg	<i>soxY</i>	IF- <i>soxY</i>	aagagegttgctgacgtgat
	IR- <i>pdhB</i>	cacgttctcctgggtgatac		IR- <i>soxY</i>	gggttctttccaccaggat
<i>pdhA</i>	IF- <i>pdhL</i>	ctacaagaaccaggtggtcg	<i>cysN</i>	IF- <i>cysN</i>	gacaacatcgtgcatgagag
	IR- <i>pdhL</i>	aagcggatcacggttctt		IR- <i>cysN</i>	gtcagcacttcagccaccac
<i>cbbSp</i>	IF- <i>cbbSp</i>	gacgattcacgatgaacgac	<i>hoxF</i>	IF- <i>hoxF</i>	ggacggatttgagatctttg
	IR- <i>cbbSp</i>	gcaatacctactgggagatg		IR- <i>hoxF</i>	cggcgttgacgattacatac
<i>cbbP2</i>	IF- <i>cbbP2</i>	ccgataccgatctgctgttc	<i>hoxH</i>	IF- <i>hoxH</i>	tcaacaacaacctgagcatc
	IR- <i>cbbP2</i>	ccgataccgatctgctgttc		IR- <i>hoxH</i>	ggtgttctgatggaagtcg
			<i>ppc</i>	IF- <i>ppc</i>	ccagcgaagagtattctcg
				IR- <i>ppc</i>	ggaaacaggtgcggtagtag

Table S2 Plasmids and primers used in the study.

Strains	Primer	Sequence (5'→3')
H16/R3-ΔlysS	IF-lys-C	atgacatgattacgaattctggtgcccacacctgctga
	IR-lys-C	aagcgttgatgctctgtcggatcgcgc
	IF-lys-D	acaagacatcgaagcgtttggcaaga
	IR-lys-D	cgacggccagtccaagcttgccgggtgtgctgccc
	F-PK18-lys	acaacccggcaagcttggcactggccgtctgttta
	R-PK18-lys	cggcaccagaattcgaatcatggtcatagctgtt
H16-P_{mbh}-A0443	A0443-Pmbh-up-F	aaacagctatgacatgattacgaattcaagatcctgaccagctgcccgtcgt
	A0443-Pmbh-up-R	tccgagcaatgccacgcagctactcaggcgtcggcgttcgagcagcga
	A0443-mbh-F	tttcgtgctcgaagccggcgacgcctgagtagctgctggcattgctcgga
	A0443-mbh-R	gtgactgaggtcggagtggaggacataacctgtctctaatttctattgg
	A0443-Pmbh-down-F	ccaatacagaattaggagacaggtatgtctcactccgacctcagtcac
	A0443-Pmbh-down-R	ttgtaaacgacggccagtccaagctttgcccccagcgcaaaagctgat
	A0443-Pmbh-V-F	atcagctttgctgctggcgcaaaagcttggcactggccgtctgtttaca
	A0443-Pmbh-V-R	acgacgggcagctggcaggatctgaattcgaatcatggtcatagctgtt
	A0443-Pchr-up-F	aaacagctatgacatgattacgaattcaagatcctgaccagctgcccgtcgt
	A0443-Pchr-up-R	ttgcagatcttcgtcaccgtggccaggcacgcctcaggcgtcggcgttcgagcagca
H16-P_{chr}-A0443	A0443-chr-F	tttcgtgctcgaagccggcgacgcctgaggcgtgcctggccacgggtgacgaaga
	A0443-chr-R	gtgactgaggtcggagtggaggacatgcttctccttgcgtggtgagcgt
	A0443-Pchr-down-F	acgctcaaccacgcaaggagacaagcatgtctcactccgacctcagtcac
	A0443-Pchr-down-R	ttgtaaacgacggccagtccaagctttgcccccagcgcaaaagctgat
	A0443-Pchr-V-F	atcagctttgctgctggcgcaaaagcttggcactggccgtctgtttaca
	A0443-Pchr-V-R	acgacgggcagctggcaggatctgaattcgaatcatggtcatagctgtt
	A2170-Pmbh-up-F	aaacagctatgacatgattacgaattcgataacggcggtcatggtgctg
H16-P_{mbh}-A2170	A2170-Pmbh-up-R	tcgtccgagcaatgccacgcagctacatggaccgttcgacgcgctgcaact
	A2170-mbh-F	agttgcagcgcgtcgaagcggctcatgtagctgctggcattgctcggacga
	A2170-mbh-R	tgaccgcgactcaggaacggcgtcataacctgtctcctaatttctgtatt
	A2170-Pmbh-down-F	aatacagaattaggagacaggttatgagcggcttctgagtcgcggtca
	A2170-Pmbh-down-R	taaacgacggccagtccaagcttagaagatggagacaatgtaccggcaat
	A2170-Pmbh-V-F	attgccgttacattgtctccatcttctaagcttggcactggccgtctgttta
	A2170-Pmbh-V-R	cgcaacctgaccggcttatcgaattcgaatcatggtcatagctgtt
H16-P_{chr}-A2170	A2170-Pchr-up-F	aaacagctatgacatgattacgaattcgataacggcggtcatggtgctg
	A2170-Pchr-up-R	acttctcaccgtggccaggcacgccatggaccgttcgacgcgctgcaact

H16- Δ B1682

A2170-chr-F	agttgcagcgcgtcgaagcgggccatggcgtgcctggccacggtgacgaagat
A2170-chr-R	acccgcgactcaggaacggcgctcatgcttgccttgcgtggtgagcgtct
A2170-Pchr-down-F	agacgctcaaccacgcaaggagacaagcatgagcggcctcctgagtcgggt
A2170-Pchr-down-R	taaaacgacggccagtccaagcttagaagatggagacaaatgtaccggcaat
A2170-Pchr-V-F	attgccggtacattgtctccatcttctaagcttggcactggccgctgttta
A2170-Pchr-V-R	cgcaacctgaccgccgttatcgaatcgaatcatggcatagctgttt
B1682-up-F	aaacagctatgaccatgattacgaattcaagcgtgtcgacggcattgtcgagt
B1682-up-R	tgaactgactccctgtctgtgttggcgcgcgagggcctccagcaggctgttca
B1682-down-F	tgaacgacctgctggaggccctcgcgcgccaacacagacagggagtcagttca
B1682-down-R	acgttgtaaacgacggccagtccaagcttagcacgccaggtcggcatcgagat
B1682-V-R	actcgacaatgccgtcgacacgcttgaatcgaatcatggcatagctgttt
B1682-V-F	atctcgatgccgacctggcgtgctaagcttggcactggccgctgtttacaacgt

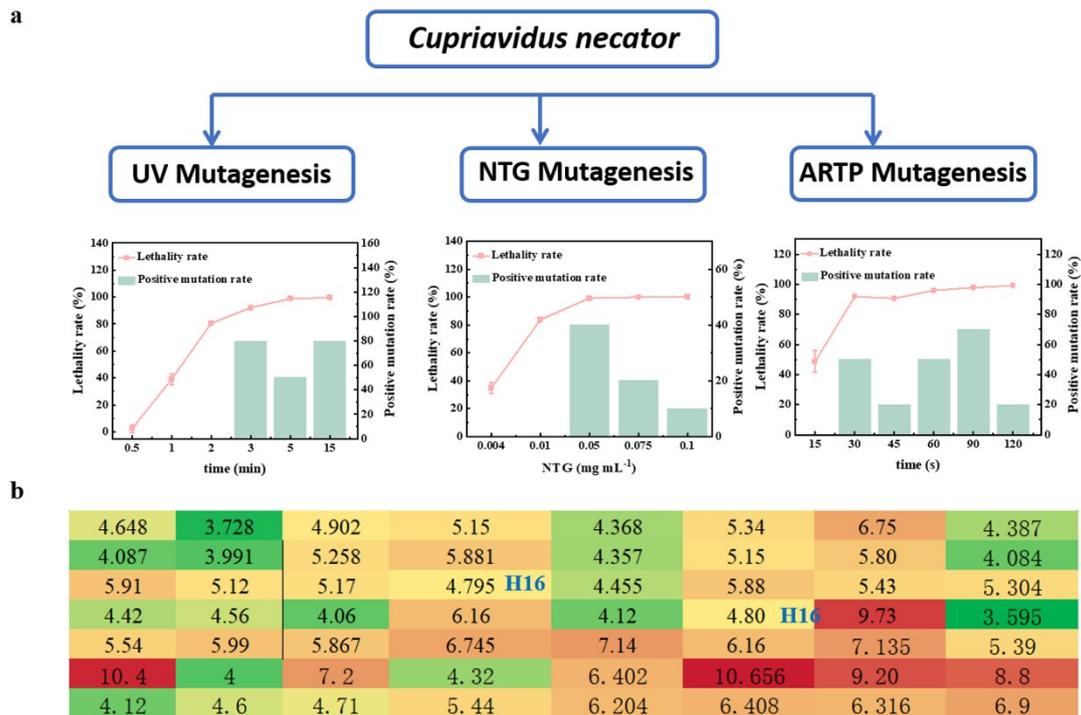


Fig. S1 Comparative assessment of mutagenesis strategies for *C. necator*. (a) Lethality and positive mutation rate induced by UV, NTG and ARTP treatments. A positive mutant is defined as a strain whose autotrophic growth is $\geq 10\%$ higher than that of the wild-type H16 control. Data represent means \pm SD of three independent experiments. (b) Numerical comparison of the screened mutant libraries. Each dot represents the relative autotrophic performance of an individual mutant. Green symbols: mutants performing worse than H16; red symbols: mutants outperforming H16.

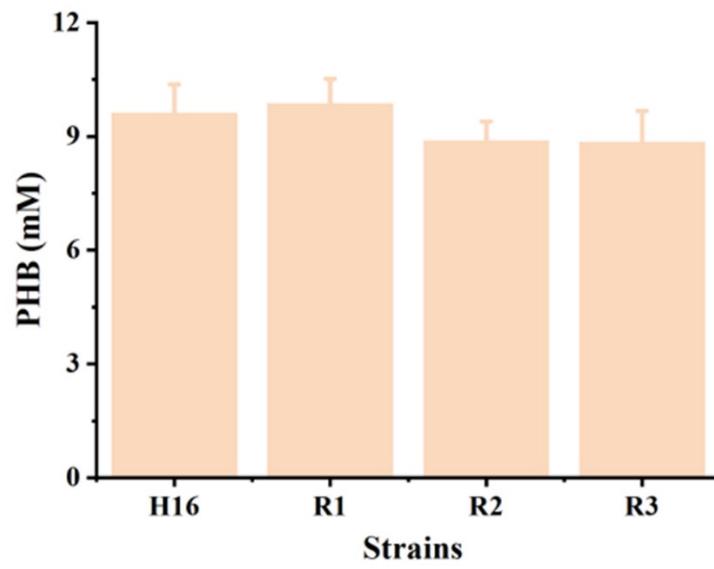


Fig. S2 PHB content of H16, R1, R2, R3.

***Cupriavidus necator* [gbbct]: 74 CDS's (25789 codons)**

fields: [triplet] [frequency: per thousand] ([number])

UUU	4.7 (121)	UCU	1.9 (49)	UAU	6.9 (177)	UGU	0.7 (17)
UUC	28.2 (727)	UCC	10.1 (261)	UAC	19.0 (490)	UGC	8.5 (220)
UUA	0.7 (17)	UCA	1.9 (50)	UAA	0.5 (13)	UGA	2.2 (57)
UUG	6.4 (165)	UCG	19.9 (514)	UAG	0.2 (4)	UGG	13.2 (341)
CUU	4.2 (108)	CCU	2.9 (76)	CAU	7.5 (194)	CGU	7.0 (181)
CUC	16.2 (417)	CCC	14.7 (380)	CAC	17.6 (454)	CGC	45.1 (1164)
CUA	1.1 (28)	CCA	3.4 (87)	CAA	7.2 (185)	CGA	2.9 (75)
CUG	70.9 (1828)	CCG	33.1 (853)	CAG	30.1 (777)	CGG	10.1 (261)
AUU	5.3 (136)	ACU	2.6 (68)	AAU	5.9 (153)	AGU	1.5 (38)
AUC	42.5 (1095)	ACC	29.8 (769)	AAC	23.5 (605)	AGC	16.6 (427)
AUA	1.0 (27)	ACA	2.6 (68)	AAA	3.6 (94)	AGA	0.7 (19)
AUG	24.9 (643)	ACG	17.3 (445)	AAG	35.1 (905)	AGG	2.8 (71)
GUU	3.6 (93)	GCU	6.0 (155)	GAU	11.7 (301)	GGU	8.3 (215)
GUC	25.6 (659)	GCC	61.4 (1583)	GAC	41.1 (1060)	GGC	61.6 (1588)
GUA	1.8 (47)	GCA	11.8 (305)	GAA	25.2 (651)	GGA	3.4 (88)
GUG	41.4 (1068)	GCG	42.4 (1093)	GAG	31.9 (823)	GGG	8.0 (206)

Fig. S3 The codons of *C. necator*.

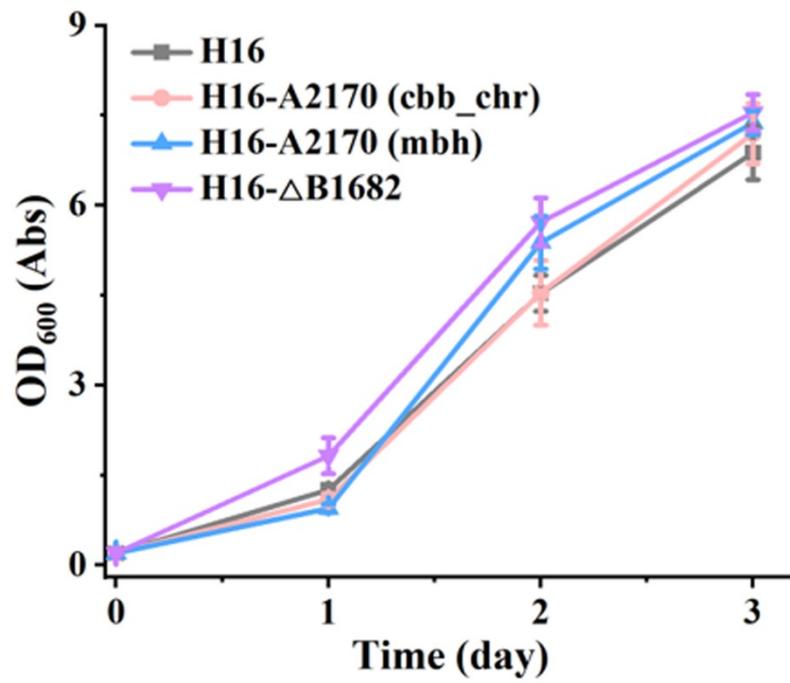


Fig. S4 The results of strengthening different promoter for A2170 gene or deleting B1682 gene in H16.

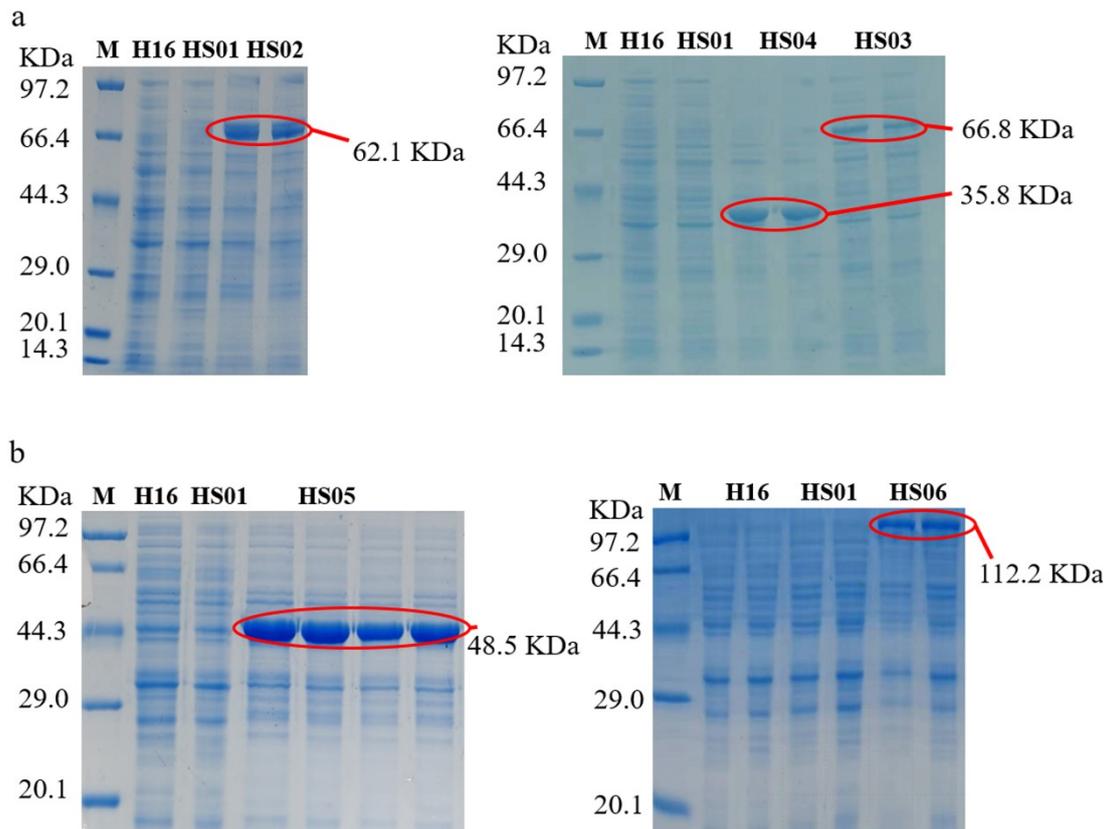


Fig. S5 The SDS-PAGE results of (a) the overexpression of engineered strains HS01, HS02, HS03, HS04, and (b) the overexpression of engineered strains HS01, HS06, HS05 and HS05 with oxalacetate or acetyl-CoA.

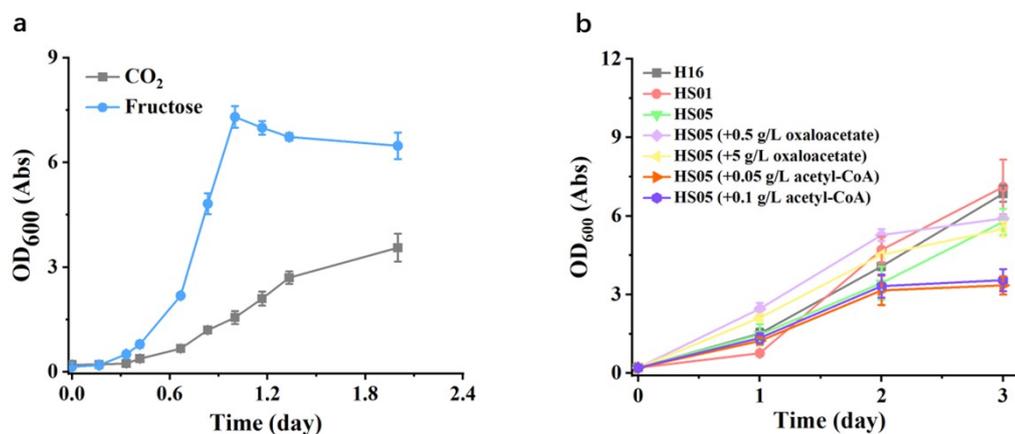


Fig. S6 The growth of *C. necator* H16 and engineered strain HS05. (a) The growth of *C. necator* H16 based on CO₂ or fructose. (b) The engineered strain HS05, and the comparison of the growth of strains HS05 with different concentrations of oxaloacetate or acetyl-CoA added during its cultivation, versus strains H16 and HS01.

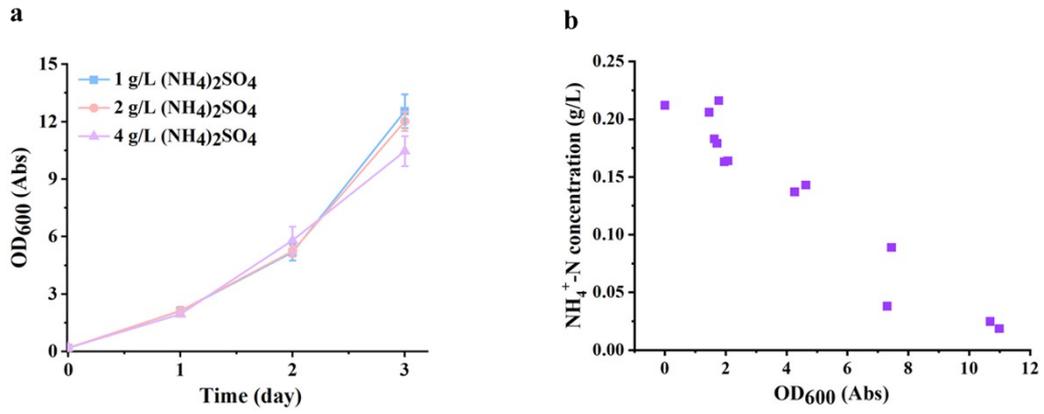


Fig. S7 The relationship between OD and nitrogen concentration. (a) Nitrogen source was verified by cultivating R3 with CO₂ as the carbon source. (b) The relationship between OD and NH₄⁺-N concentration.