Efficient Dual-Phase Visual Detection of Pesticides in Real Samples with Electron-rich Emitters Carrying Multiple Twists

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Materials and Methods

General Experimental Information

All the required chemicals were purchased from various companies and used without purification. All the products were characterized by ¹H, and ¹³C NMR spectroscopy. NMR spectra were recorded on a Bruker 400 MHz instrument (400 MHz for ¹H NMR, and 101 MHz for ¹³C NMR). ¹H NMR experiments are reported in units, parts per million (ppm), and were measured relative to residual DMSO (2.50 ppm) in the deuterated solvent. ¹³C NMR spectra are reported in ppm relative to deuteron DMSO (39.52 ppm) and all were obtained with ¹H decoupling. Coupling constants were reported in Hz. Reactions were monitored by thin-layer chromatography (TLC). liquid chromatography-mass spectrometry (LC-MS) was obtained by the electron spray ionization (ESI) technique using the Q-TOF mass analyzer and was reported as m/z (relative intensity). Melting points of compounds were recorded on a KRUSS Optronic M3000 apparatus.

Synthesis of (E)-4-(2-(10-(4-(diphenylamino) phenyl) anthracen-9-yl) vinyl)-N, Ndiphenylaniline (**TT1**)., 4-((E)-2-(10-(4-(diphenylamino) phenyl) anthracen-9-yl) vinyl)-N-(4-((E)-2-(10-(4-(diphenylamino) phenyl) anthracen-9-yl) vinyl) phenyl)-N-phenylaniline (**TT2**), and 4-(10-((E)-4-(bis(4-((E)-2-(10-(4-(diphenylamino) phenyl) anthracen-9-yl) vinyl) phenyl)amino) styryl) anthracen-9-yl)-N, N-diphenylaniline (**TT3**).

The molecules TT1, TT2 and TT3 were synthesised via the reported protocol¹

Thermogravimetric studies



Figure S1. Thermogravimetric analysis of (a) TT1, (b) TT2 and (c) TT3. PXRD Analysis



Figure S2. Comparative PXRD patterns of TT1, TT2 and TT3.





Figure S3. UV-Vis spectrum of TT1, TT2 and TT3 in different solvents.

Table S1. Tabulated photophysical parameters of TT1, TT2 and TT3 in different solvents.

Entra	Coluonto	TT:	1	тт	2	TT3		
Entry	Solvents	λ _{emi} (nm)	Φ _f (%)*	λ _{emi} (nm)	Ф _f (%)*	λ _{emi} (nm)	Ф _f (%)*	
а	N-Hexane	383	28	516	11	527	16	
b	Carbontetrachloride	443	4	525	6	529	12	
с	Toluene	414	51	531	23	532	28	
d	Dichloromethane	459	53	548	6	556	8	
е	Chloroform	451	51	540	7	543	11	
f	1,4-dioxan	423	61	530	14	533	20	
g	Tetrahydrofuran	427	51	537	22	543	26	
h	N,N-Dimethylacetamide	445	44	558	16	574	22	
i	N,N-Dimethylsulphoxide	458	3	564	3	585	5	
j	Dimethylformamide	452	37	558	3	545	5	
k	Acetonitrile	425	5	539	1	527	4	
I	Methanol	524	1	534	2	510	3	
m	Water	557	1	558	1	549	2	

The relative quantum yield of the probe was measured with respect to quinine sulfate (in 0.1 M H₂SO₄) using the formula,

$$\Phi_f = \Phi_f \times \frac{a_{ref}}{a_{sam}} \times \frac{A_{sam}}{A_{ref}} \times \left(\frac{n_{sam}}{n_{ref}}\right)^2$$

Where, Φ_f = quantum yield of probe Φ_{ref} = quantum yield of quinine sulfate (0.54)

 A_{ref} = area under curve of emission spectra of quinine sulfate in 0.1M H₂SO₄, Asam = area under curve of emission spectra of probe, a_{sam} = Absorbance of probe, a_{ref} = Absorbance of quinine sulfate in 0.1M H₂SO₄, n_{sam} = refractive index of respective solvents, n_{sam} = refractive index of water.



Figure S4. Emission spectra of (a) TT1, (b) TT2, and (c) TT3 in different solvents. Selectivity experiment for TT2 and TT3.



Figure S5. Fl. Emission spectra of (a) TT2, (b) TT3 after adding all the analytes; bar diagram representing the % quenching of (c) TT2 (d) TT3 (inset is photograph of TT2 and TT3 (10 μ M in 1,4-dioxane) after treatment with various analytes): i. Trifluralin, ii. Fenitrothion, iii. Imidacloprid, iv. β -cyfluthrin, v. Chlorpyrifos, vi. Glyphosate, vii. Permethrin, viii. Diphenylamine, ix. Sodium pyrophosphate tetrabasic, x. Dipotassium hydrogen phosphate, xi. Tricalcium phosphate, xii 1. 4-Nitrotoluene, xiii. 1,2-Chloro dinitrobenzene, xiv. 2-Chloro-4-nitroaniline, xv. Diethylbenzyl phosphonate. (e) molecular structure of various pesticides and other analogues used for selectivity.



Titration studies

Figure S6. Change in UV-Vis spectra of **TT1**, **TT2** and **TT3** by gradual addition of TN and FN, respectively; Inset pictures correspond to before and after pesticide addition in ambient light.

NMR Studies



Figure S7. ¹H NMR (partial) spectra of **TT1** in CDCl₃ before (TT1) and after (TT1+TN) the addition of TN (1 equiv.).

Stern Volmer plot for TT1 TT2 and TT3



Figure S8. Stern-Volmer plots for different pesticides with (a) **TT1+TN**, (b) **TT1+FN**, (c) **TT2+TN**, (d) **TT3+TN**, in 1,4-dioxane plotting I₀/I against concentration of pesticide.



Figure S9. Stern-Volmer plots for different pesticides at lower concentration for K_{sv} (Stern-Volmer constant) determination with (a) TT1+TN, (b) TT1+FN, (c) TT2+TN, (d)TT3+TN, in 1,4-dioxane plotting I₀/I against concentration of pesticide.

Lifetime studies



Figure S10. Lifetime decay profile of TT1, TT2 and TT3 (10 μ M in 1,4-dioxane) in solution state.

Table S2. Lifetime decay parameters for TT1, TT2 and TT3 (10 μ M in 1,4-dioxane) in solution state.

Sample	$ au_l$	$ au_2$	α_{l}	α_2	Avg. $<\tau>$ (ns)	χ^2	Ф _f (%)	k_r (10 ⁶ s ⁻¹)	k_{nr} (10 ⁶ s ⁻¹)
10 µM TT1	2.40	4.55	0.03	0.97	4.52	1.3	61	134.9	86.2
10 μM TT2	1.63	6.59	0.99	0.01	1.88	0.9	14	74.4	457.4
10 μM TT3	1.55	2.09	0.65	0.35	1.78	0.9	20	112.3	449.4



Figure S11. Lifetime decay profile of (a)**TT1**, (b)**TT2** and (c)**TT3** before and after the addition of pesticides.

Table S3. Lifetime decay parameters for **TT1**, **TT2** and **TT3** (10 μ M in 1,4-dioxane) before and after the addition of pesticides (TN and FN).

Sample	$ au_{I}$	$ au_2$	α_1	α2	Avg.<τ> (ns)	χ^2
10 μM TT1	2.40	4.55	0.03	0.97	4.52	1.3
10 μM TT1 +5 x 10 ⁻⁶ M TN	1.74	4.31	0.07	0.93	4.23	1.1
10 μM TT1 +5 x 10 ⁻⁶ M FN	3.25	4.48	0.04	0.96	4.43	1.2
10 μM TT2	1.63	6.59	0.99	0.01	1.88	0.9
10 μM TT2 +50 x 10 ⁻⁶ M TN	2.13	1.25	0.39	0.61	1.71	1.5
10 μM TT3	1.55	2.09	0.65	0.35	1.78	0.9
10 μM TT2 +5 x 10 ⁻⁵ M TN	1.92	1.43	0.55	0.45	1.73	0.9



Figure S12. Lifetime decay profile of TT1, upon gradual addition of Trifluralin.

Table S4. Lifetime decay parameters for TT1, TT2 and TT3 (10 μ M in 1,4-dioxane) upon gradual addition of TN.

Sample	$ au_{I}$	$ au_2$	α_{l}	α_2	Avg. $<\tau>$ (ns)	χ^2
10 μM TT1	2.40	4.55	0.03	0.97	4.52	1.3
10 μM TT1 + 0.5 x 10 ⁻⁷ M TN	2.23	4.44	0.06	0.94	4.37	1.0

10 µM TT1 + 1 x 10 ⁻⁷ M TN	1.71	4.39	0.05	0.95	4.35	0.9
10 μM TT1 + 2 x 10 ⁻⁶ M TN	1.83	4.32	0.05	0.95	4.27	1.0
10 μM TT1 +5 x 10 ⁻⁶ M TN	1.74	4.31	0.07	0.93	4.23	1.1

Limit of Detection Plots for TT1, TT2 and TT3



Figure S13. Limit of detection plot of different pesticides with TT1, TT2 and TT3 (10 μ M in 1.4 disease)

1,4-dioxane)

Calculation of the Limit of Detection (LOD):

The standard deviation and slope were obtained by plotting the ratio of Fl intensity at 423, 530,

533 nm for TT1, TT2, TT3 respectively with a concentration of pesticides

 $LOD = 3.3 \sigma/K$

Where, σ = Standard error of the linear fitting and K = Slope

Example: For Trifluralin, $\sigma = 17.46$ and K = 278 x 10⁶ M⁻¹, thus LOD= $\frac{3 x 17.46}{278 x 10^6} = 18 x 10^{-8}$

Μ

JOBS plot for binding ratio



Figure S14. Fluorescence Job's plot of (a) TT1+TN, (b)TT1+FN, (c)TT2+TN and (d)TT3+TN with different pesticides, depicting 1:1 complex.

Comparative E_{HOMO} – E_{LUMO} energy of TT1, TT2 and TT3.



Figure S15. HOMO-LUMO energy diagrams and their respective values of TT1, TT2, and TT3. Note: The E_{HOMO} - E_{LUMO} of TT1 was calculated theoretically as explained below. All the

calculations were performed in Gaussian using B3LYP-D3(BJ)/6-31G** (with dispersion corrections).



Figure S16. Comparative E_{LUMO} levels of all pesticides and **TT1**, **TT2** and **TT3**. The E_{LUMO} of TN, FN, IM, GP, and CF were known in literature² and CP, DP and PN were calculated through DFT.(All the calculations were performed in Gaussian using B3LYP-D3(BJ)/6-31G** (with dispersion corrections).



Figure S17. Schematic representation of the photoinduced electron transfer mechanism for (a) **TT2** and (b) **TT3** in the presence of trifluralin and fenitrothion. (All the calculations were performed in Gaussian using B3LYP-D3(BJ)/6-31G** (with dispersion corrections).



Figure S18. Spectral overlap between absorption and normalised emission bands (a) pesticides and TT1 ($\lambda_{ex} = 310 \text{ nm}$), TT2 ($\lambda_{ex} = 410 \text{ nm}$) and TT3 ($\lambda_{ex} = 410 \text{ nm}$) respectively, (b) TN and FN absorbance overlap with TT1 emission band.

AIE properties



Figure S19. Change in emission of TT1, TT2 and TT3 (10 µM) on increasing water

fraction.



Figure S20. (a) Fl. lifetime decay plot for TT1 ($f_w = 00\%$) and TT1 ($f_w = 50\%$), showing the decay profile for molecular and aggregated states. (b) SEM image for morphology of nanoaggregates and their sizes, and DLS plot for TT1 ($f_w = 50\%$) for the hydrodynamic diameter.

Table S5- Fluorescence lifetime decay parameters

Sample	$ au_1$	$ au_2$	α1	α_2	Avg. <τ>	χ^2
TT1 ($f_w = 00\%$)	10.7	1.7	0.15	0.85	6.41	1.0
TT1 ($f_w = 50\%$)	2.1	11.2	0.67	0.33	8.74	1.1

Absorbance of TT1, TT2 and TT3 in solid state.



Figure S21. Combined UV-Vis spectrum of TT1, TT2, and TT3 in solid state.

Paper strip characterisation



Figure S22. TT1@WP characterisation through (a) IR, and (b) Fluorescence emission spectrum.

Detection of pesticides in the solid state of the probe



Figure S23. TT1@WP, TT2@WP and **TT3@WP** (10μ M) before and after exposing various pesticide solutions (under 365 nm UV lamp).Where, I. Probe II. Trifluralin; III: Fenitrothion IV: Glyphosate V. Imidacloprid VI. β -Cyfluthrin; VII. Permethrin VIII. Chlorpyrifos IX: Diphenylamine X: Control (1,4-dioxane)

Schematic diagram for real-life applicability



Figure S24. Schematic illustration of experiment for real life applicability of sensor spray.

Fabrication of TT1-coated fluorescent cotton buds

Commercial cotton buds were taken and was cleaned with methanol for removing the chemical residues, further it was dried for 30 min in 40 °C in hot air oven, the cleaned buds were dipped in **TT1** solution (10 μ M in acetone) for 40 minutes, to ensure even absorption of dye in cotton buds, further the cotton buds were separated and was hung freely for 20 minutes with paper

clip to rinse extra **TT1** solution, further the buds were dried at 40 °C for 30 minutes, and were ready to use.



Figure S25. Schematic illustration of the fabrication of TT1-coated fluorescent buds.

Table S6: Con	nparative literature	of Trifluralin and	l Fenitrothion	detection
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Sl.No	Reported work	Material	Detection limit (solution)	Detection limit (solid)	Thermal stability (solid)	Response time (solution)	Response time (solid)	Reference
1.	Efficient Dual-State Visual Detection of Pesticides in Real Samples Using Triphenylamine- Enriched Twisted π- Conjugates	Small Molecule	180 nM	100 nM	>300 °C	Rapid	Rapid	This Work
2	Creation of Carbazole- Based Fluorescent Porous Polymers for Recognition and Detection of Various Pesticides in Water	Porous organic polymer (POPs)	NA	0.35 µM	NA	Rapid	Rapid	ACS Sens. 2020 , 5, 162–170
3	Synthesis of Fluorescent Micro- and Mesoporous Polyaminals for Detection of Toxic Pesticides	Porous organic polymer (POPs)	NA	6.8 μM	NA	Rapid	Rapid	Macromo lecules 2018 , 51, 1769– 1776
4	Highly Sensitive and Recognizable Detection for Trifluralin with Alkyl- Decorated Fluorescent Porous Polymers	Porous polymer	6 μΜ	5 μΜ	< 250 °C	4 s	NA	Chem. Eng. J. 2023 , 470, 144123.

5	A Simple AIE Probe to Pesticide Trifluralin Residues in Aqueous Phase: Ultra-Fast Response, High Sensitivity, and Quantitative Detection Utilizing a Portable Platform	Small molecule	6.28 μM	NA	NA	Rapid	NA	Talanta 2024 , 269, 125352.
6	A practical fluorometric and colorimetric dual-mode sensing platform based on two-dimensional porous organic nanosheets for rapid determination of trifluralin	porous organic nanosheets	3.50 nM	0.41 μM	NA	9 s	NA	Anal. Methods, 2025 ,17, 1188- 1195
7	Green Synthesis of Fluorescent Carbon Dots from Cherry Tomatoes for Highly Effective Detection of Trifluralin Herbicide in Soil Samples	carbon quantum dots	0.5 nM	NA	NA	NA	NA	Chemistr y Select 2020 , 5 (6), 1956– 1960

Reference

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Sidearms Elongated C3-Symmetric Electrochromic and Electrofluorochromic π-Conjugate:
Small Molecule in Efficient Transparent to Black Switching. *Next Mater.* 2025, *6*, 100472.
Zhang, B.; Li, B.; Wang, Z. Creation of Carbazole-Based Fluorescent Porous Polymers for
Recognition and Detection of Various Pesticides in Water. *ACS Sens.* 2020, 5, 162–170.