

Design and Evaluation of Novel Hydrazone Derivatives as NS1 Inhibitors for Dengue Virus: Synthesis, Experimental, and Computational Studies

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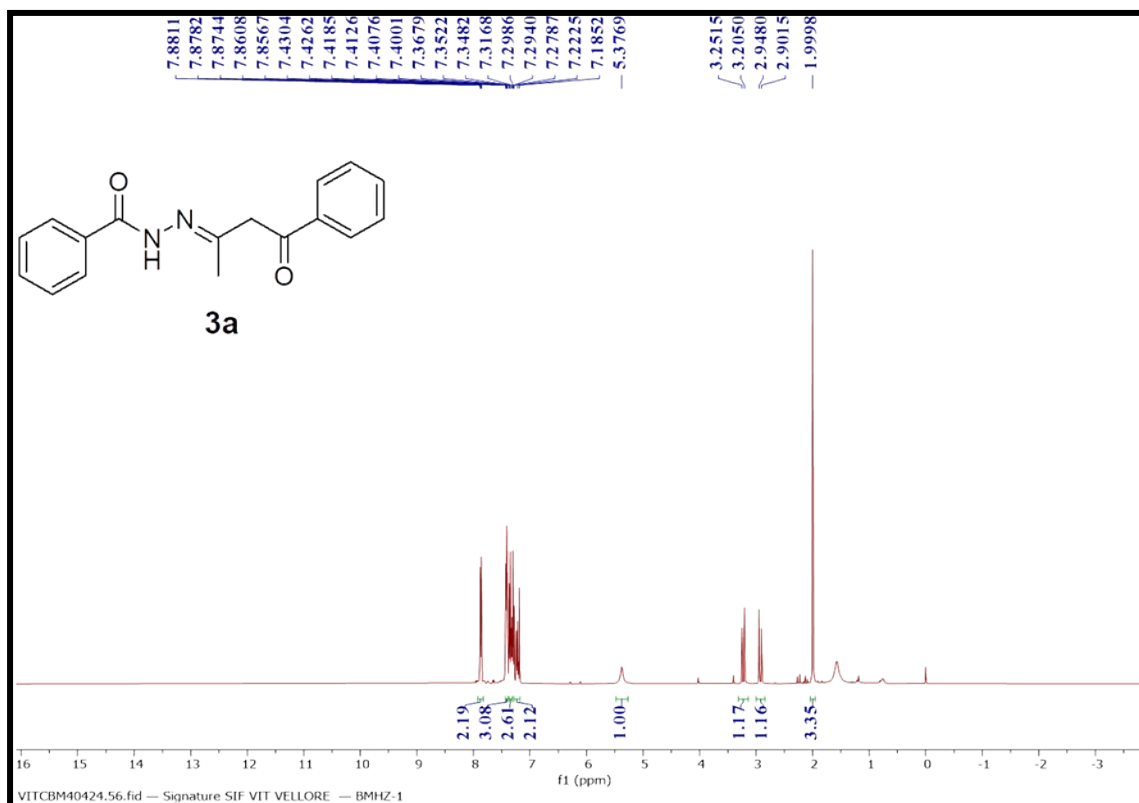
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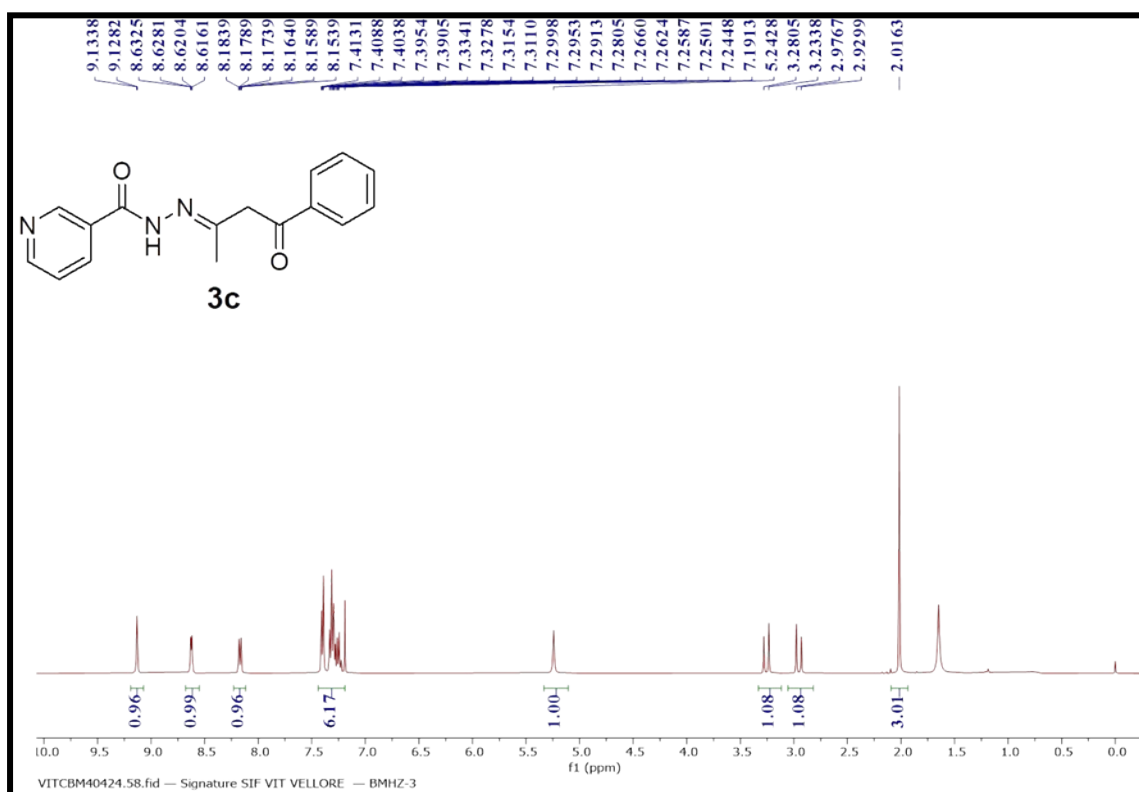
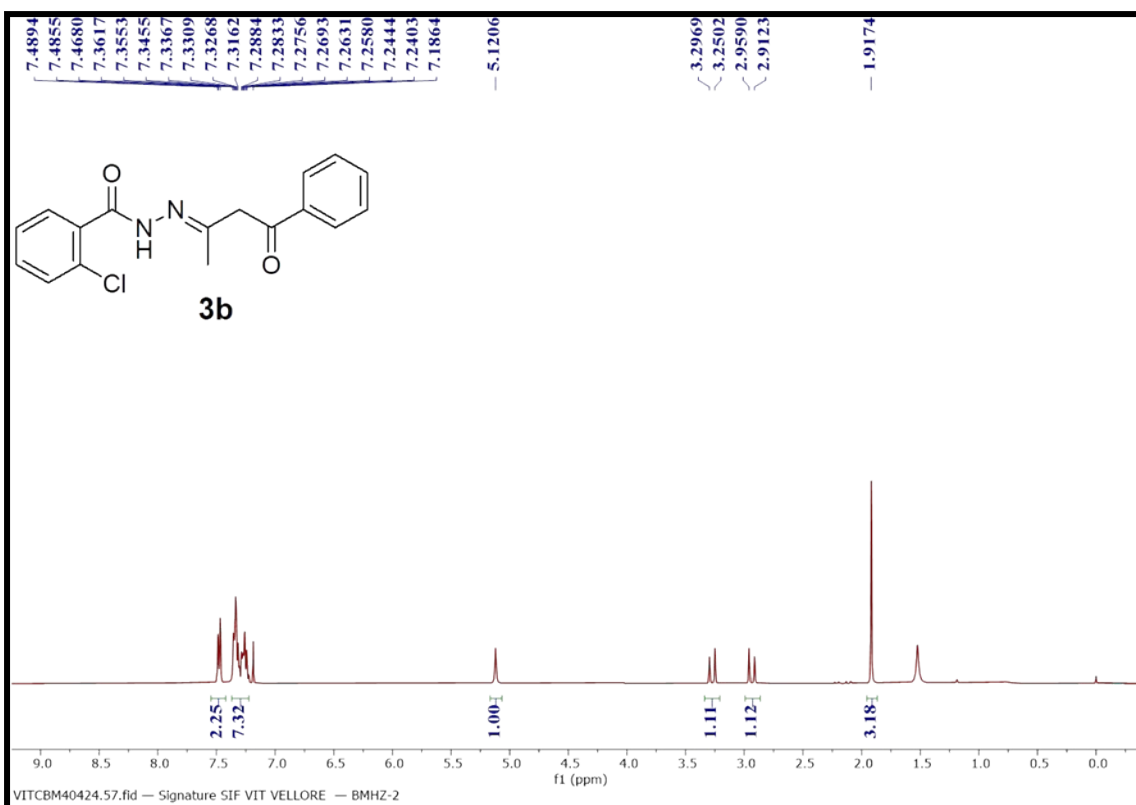
^cDepartment of Chemistry, Rabindranath Tagore University, Hojai 782435, Assam, India

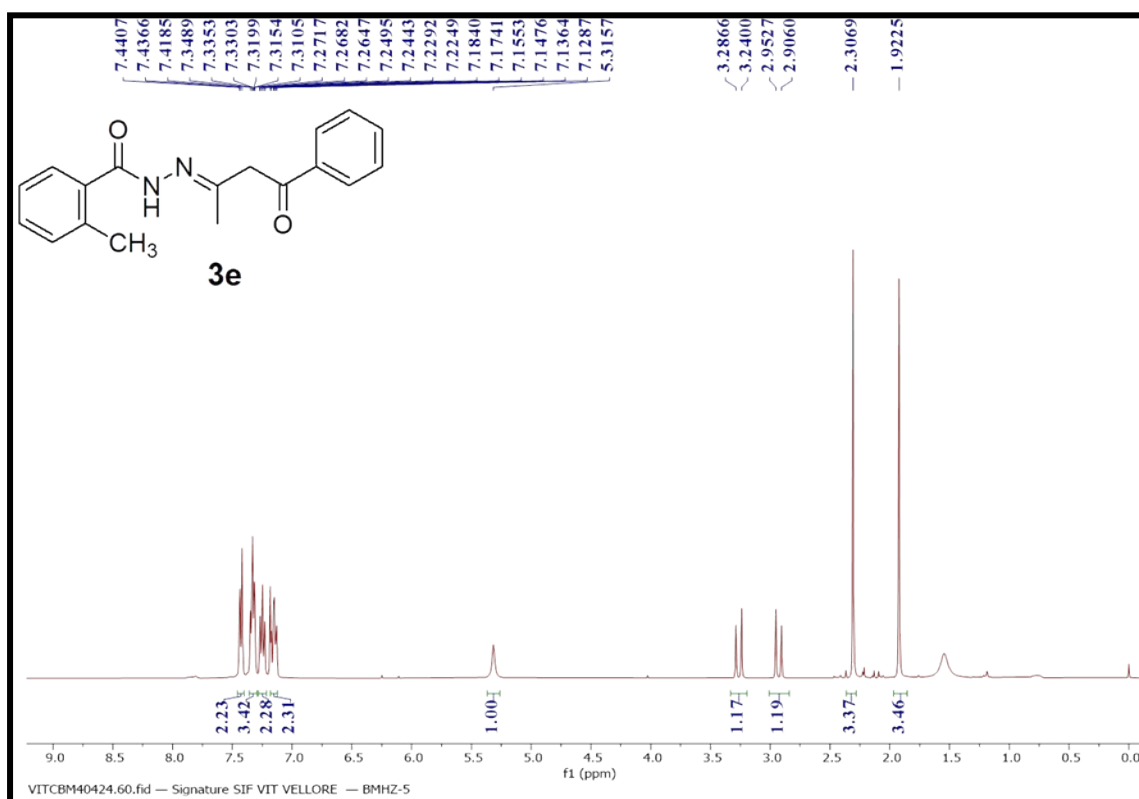
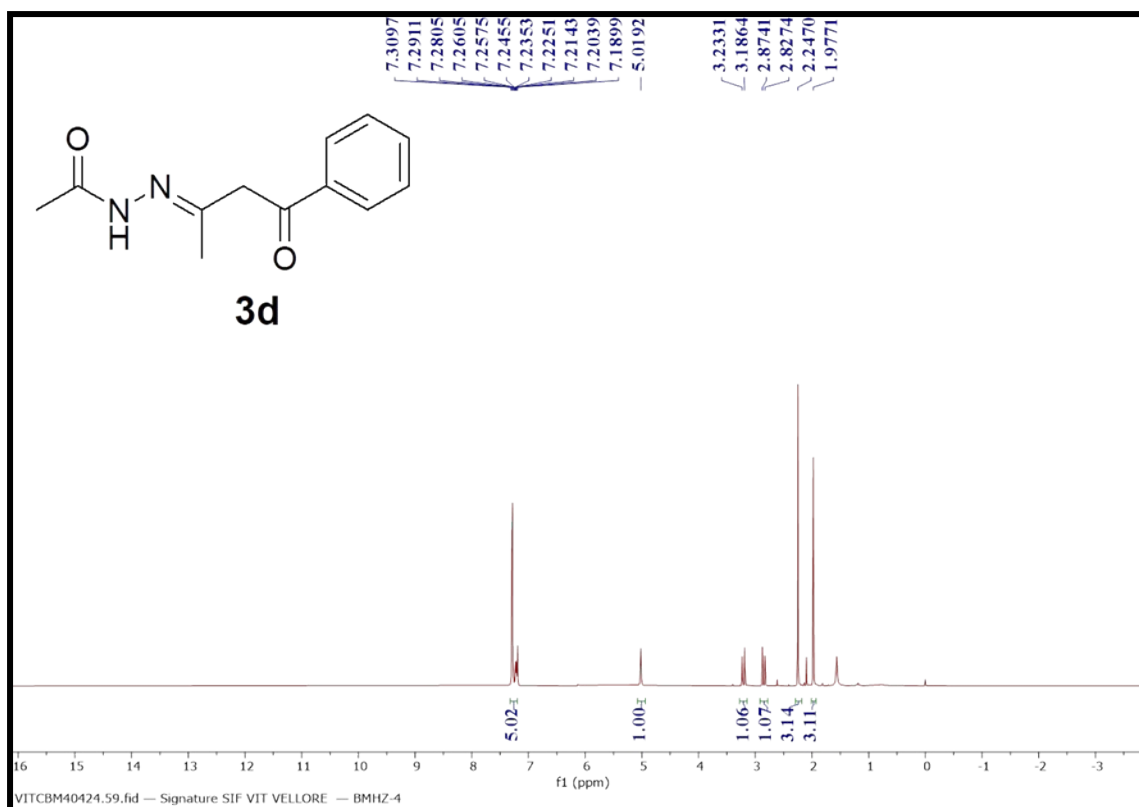
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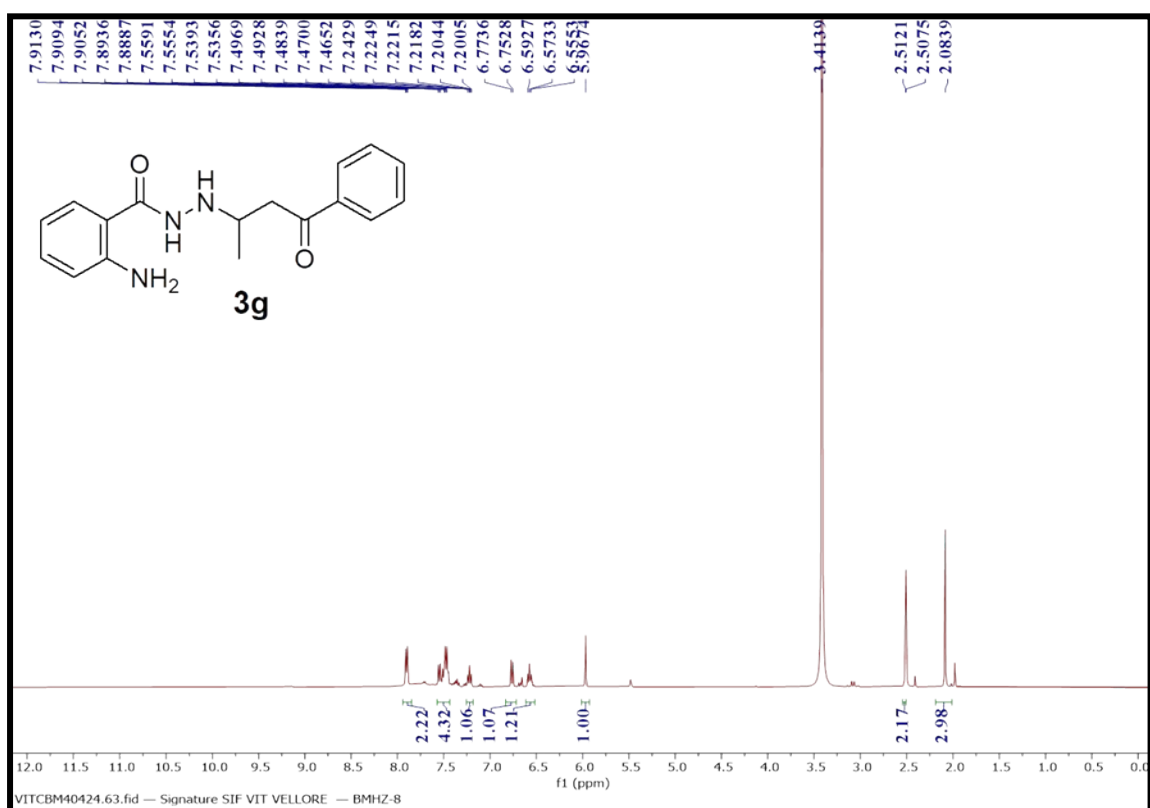
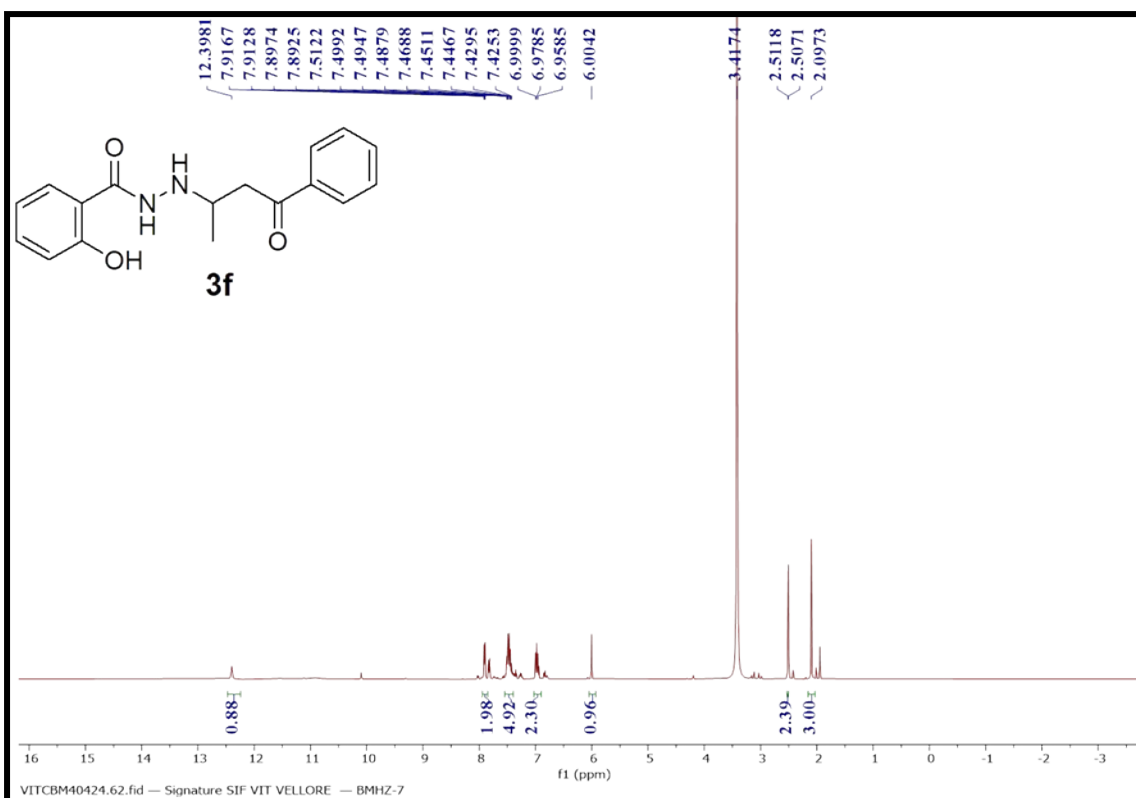
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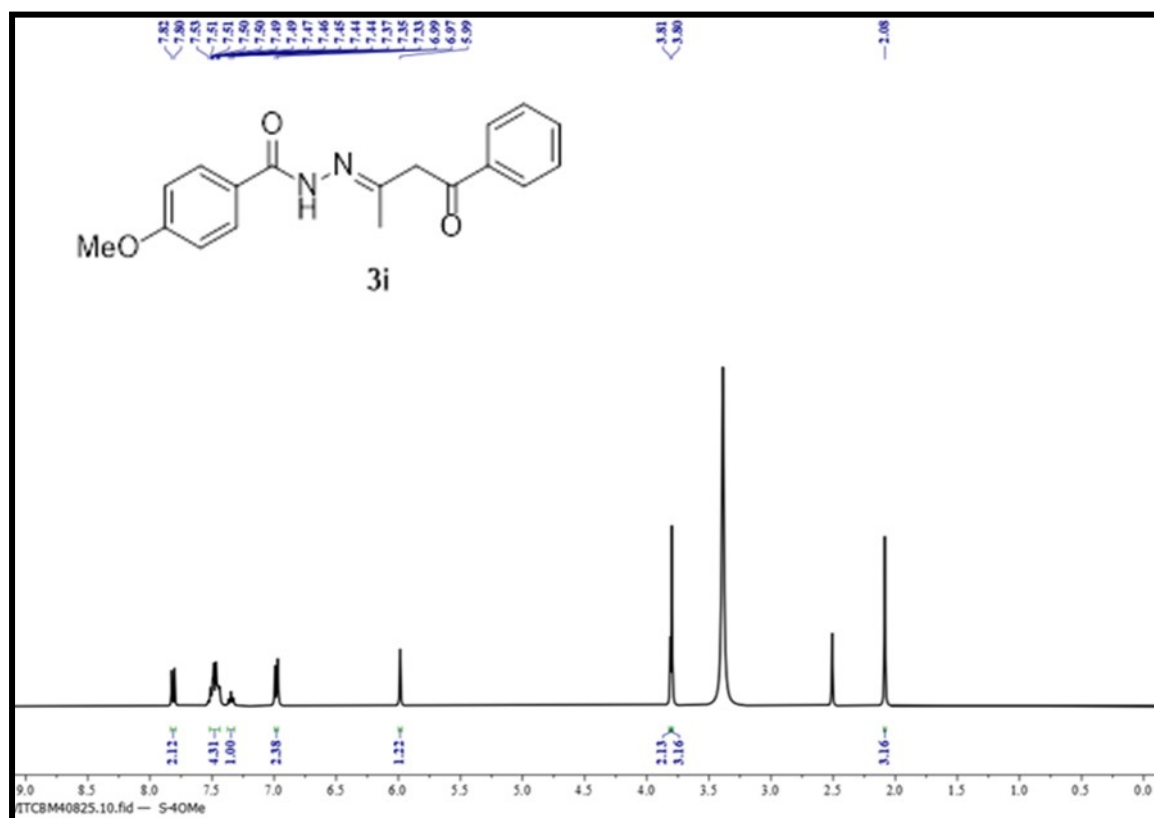
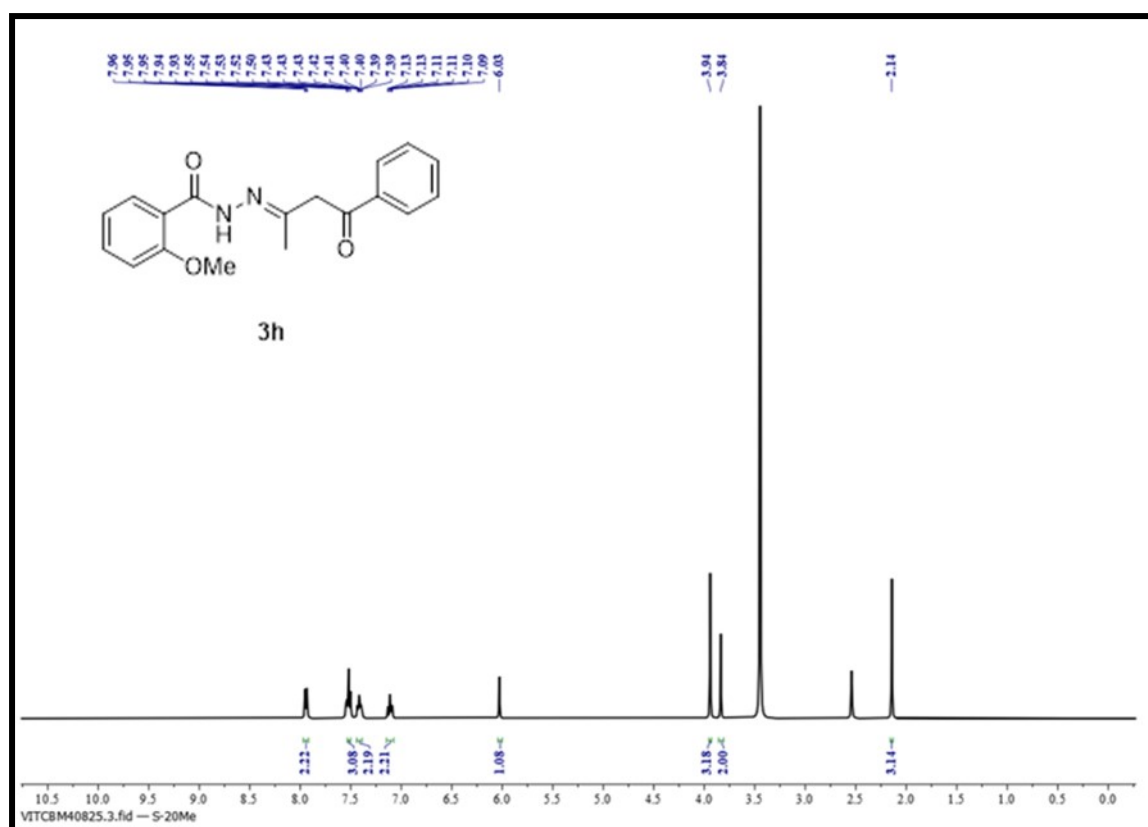
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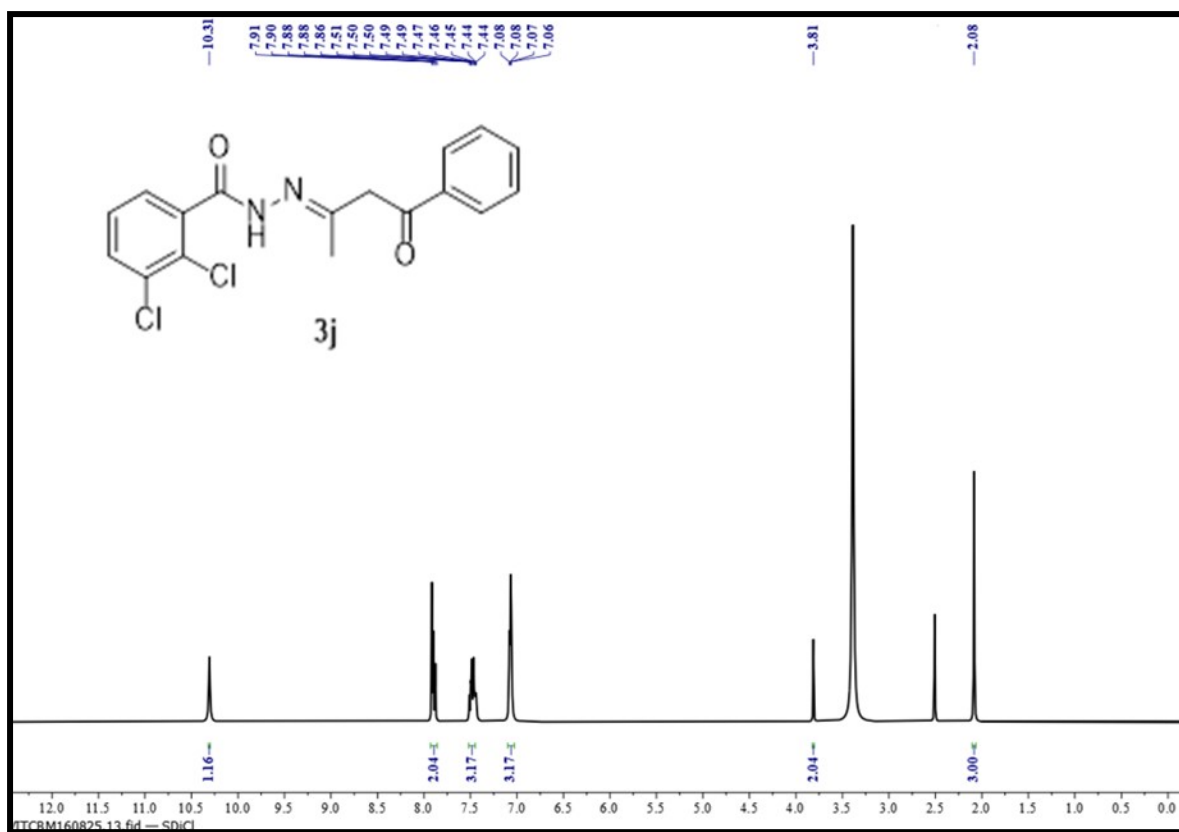




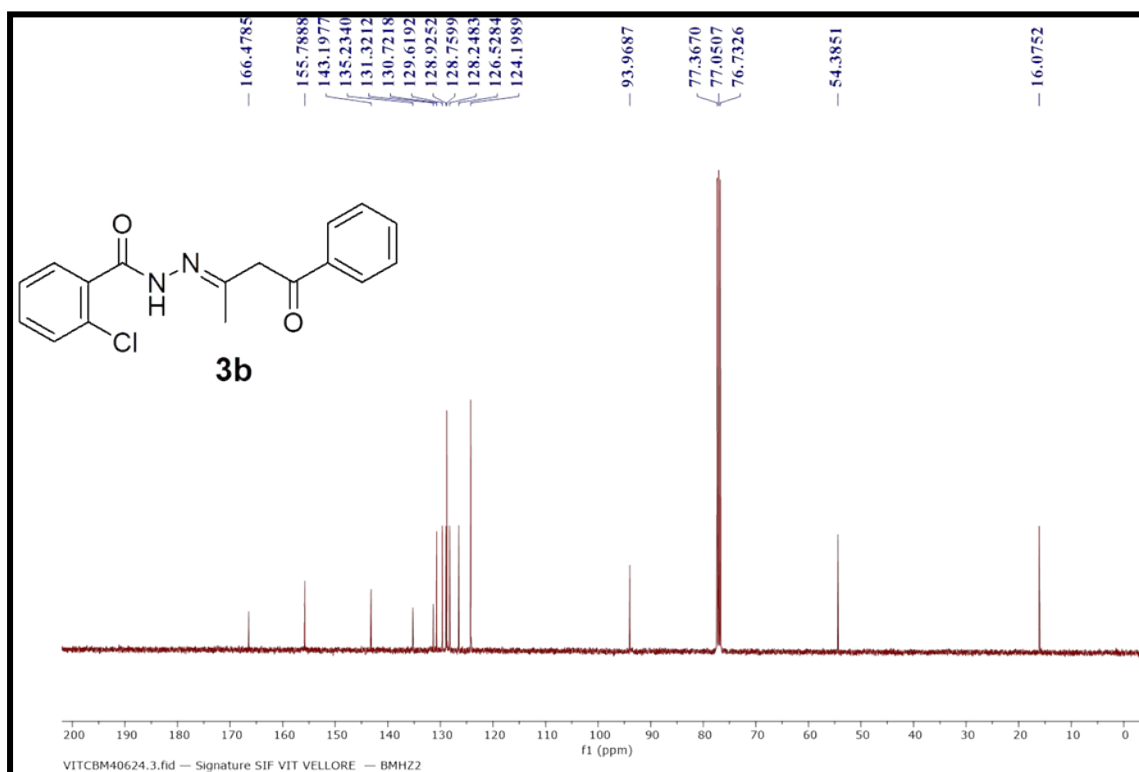
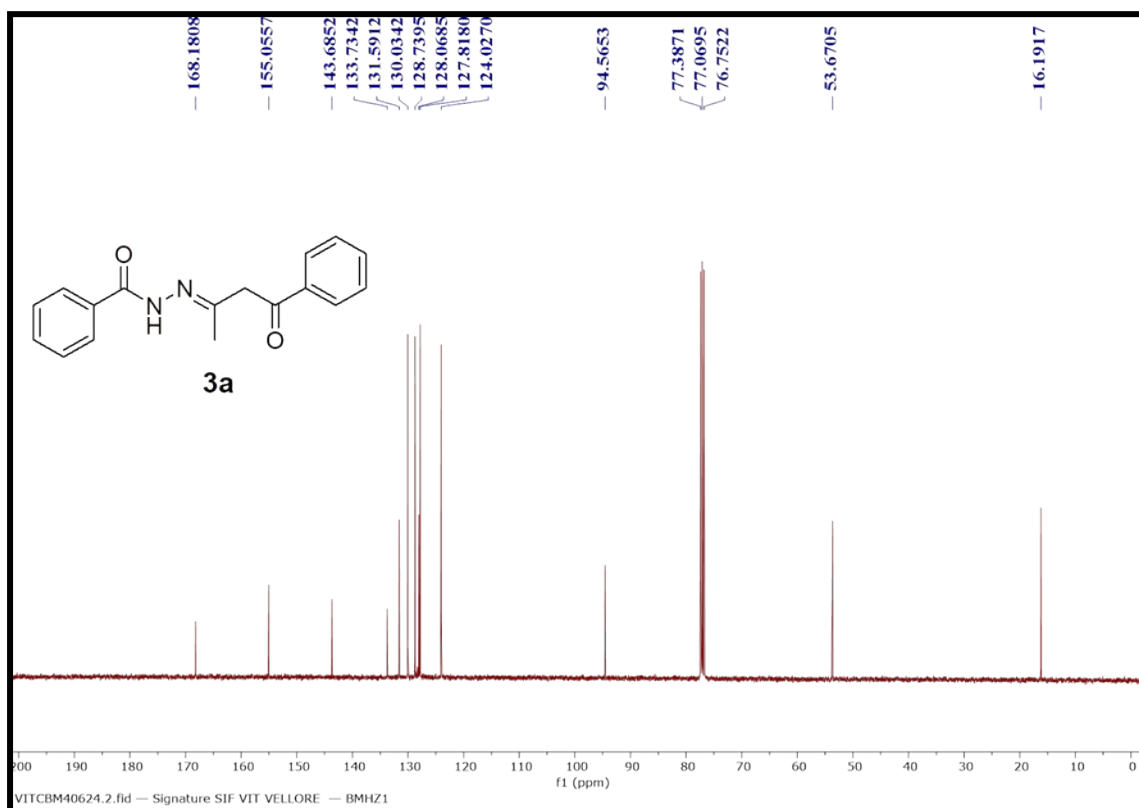


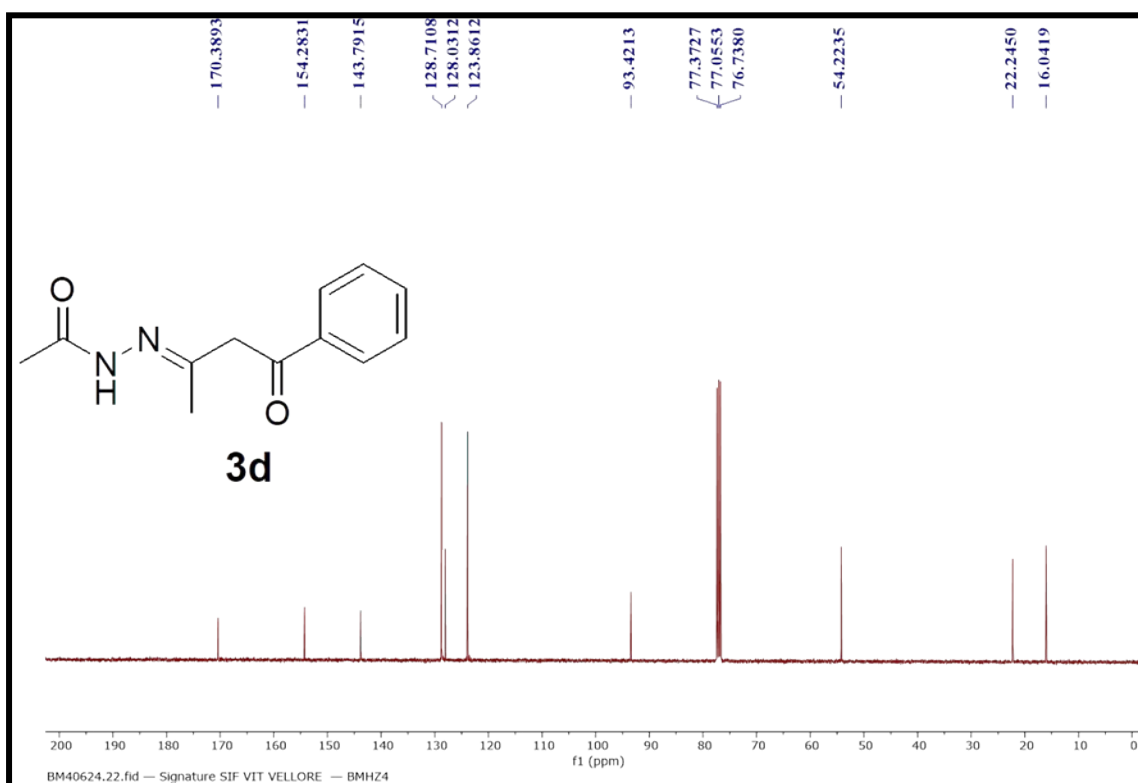
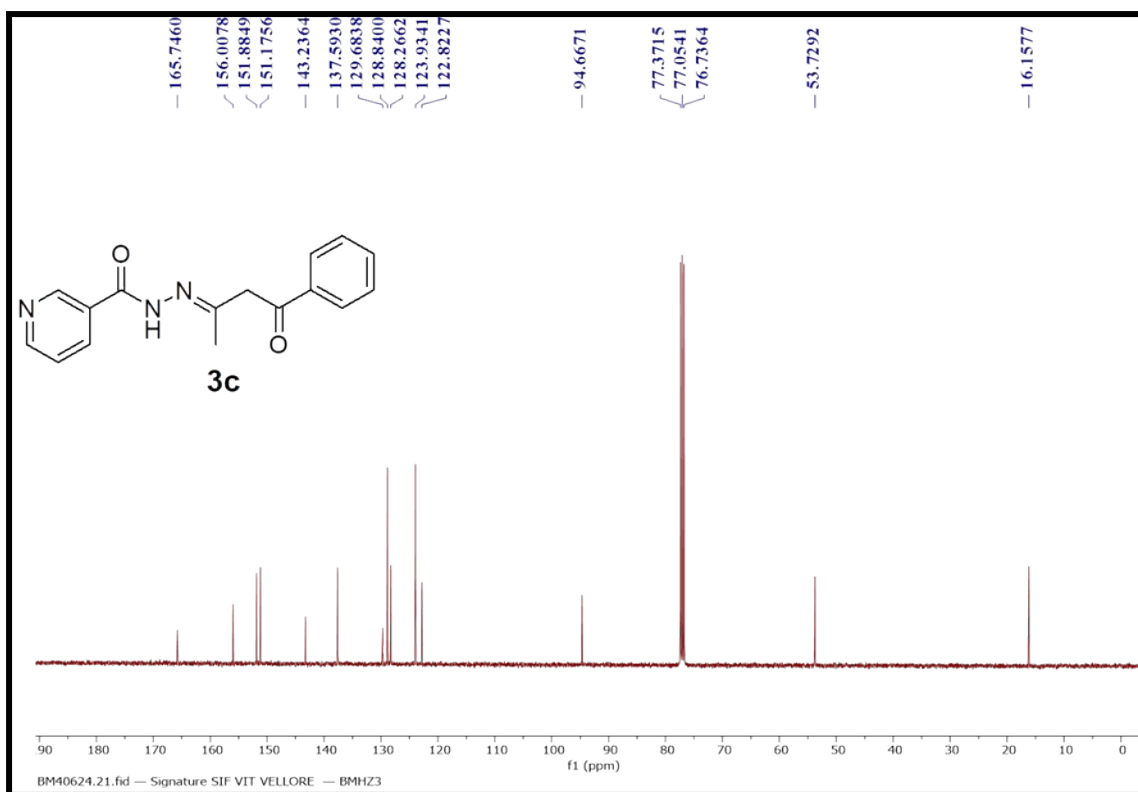


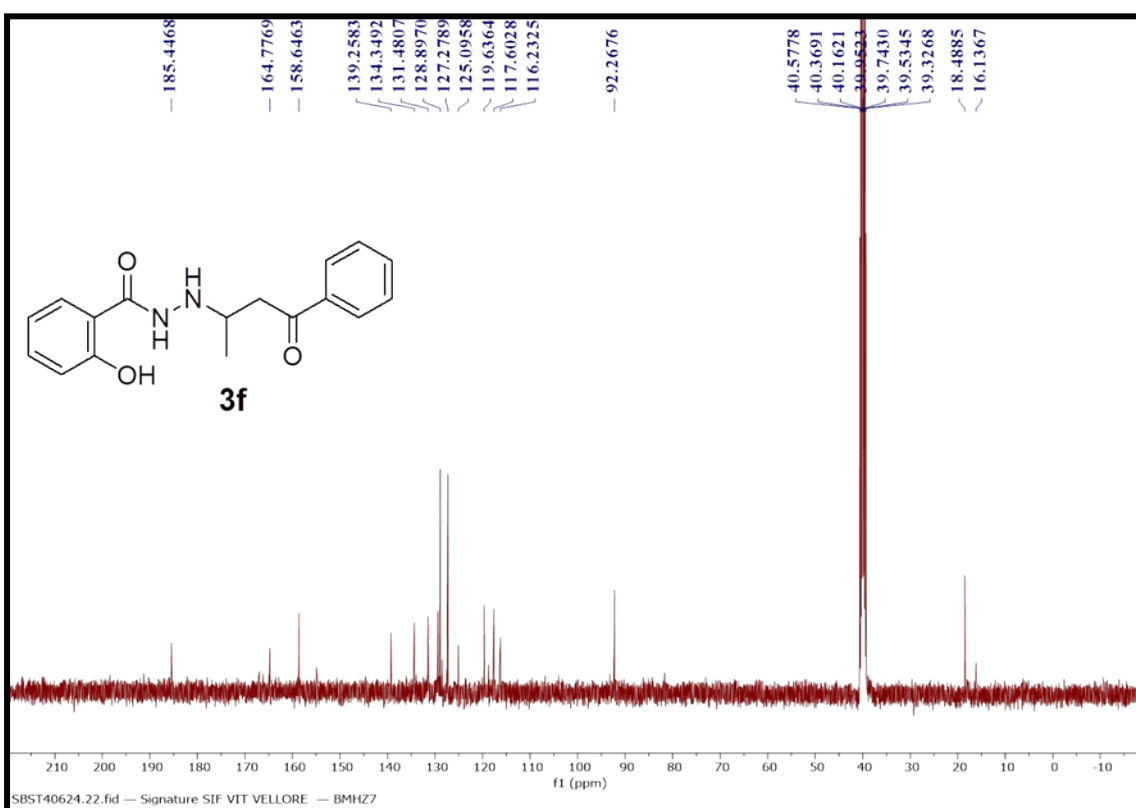
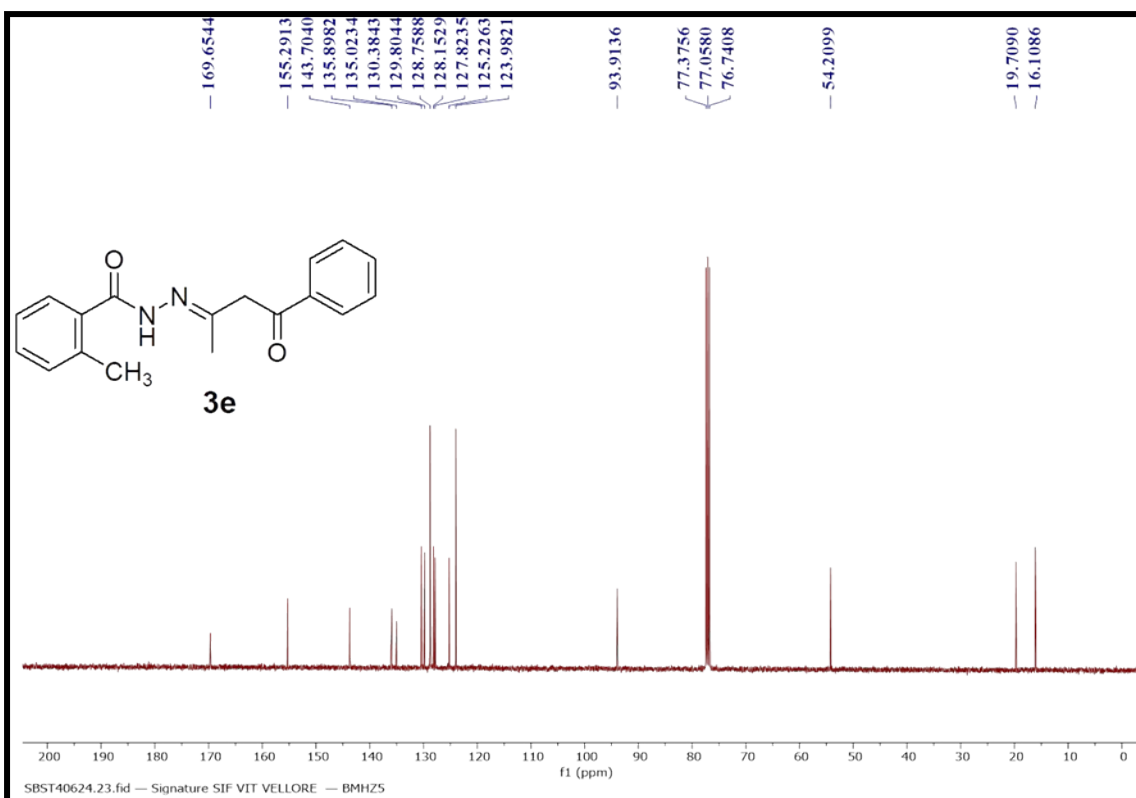


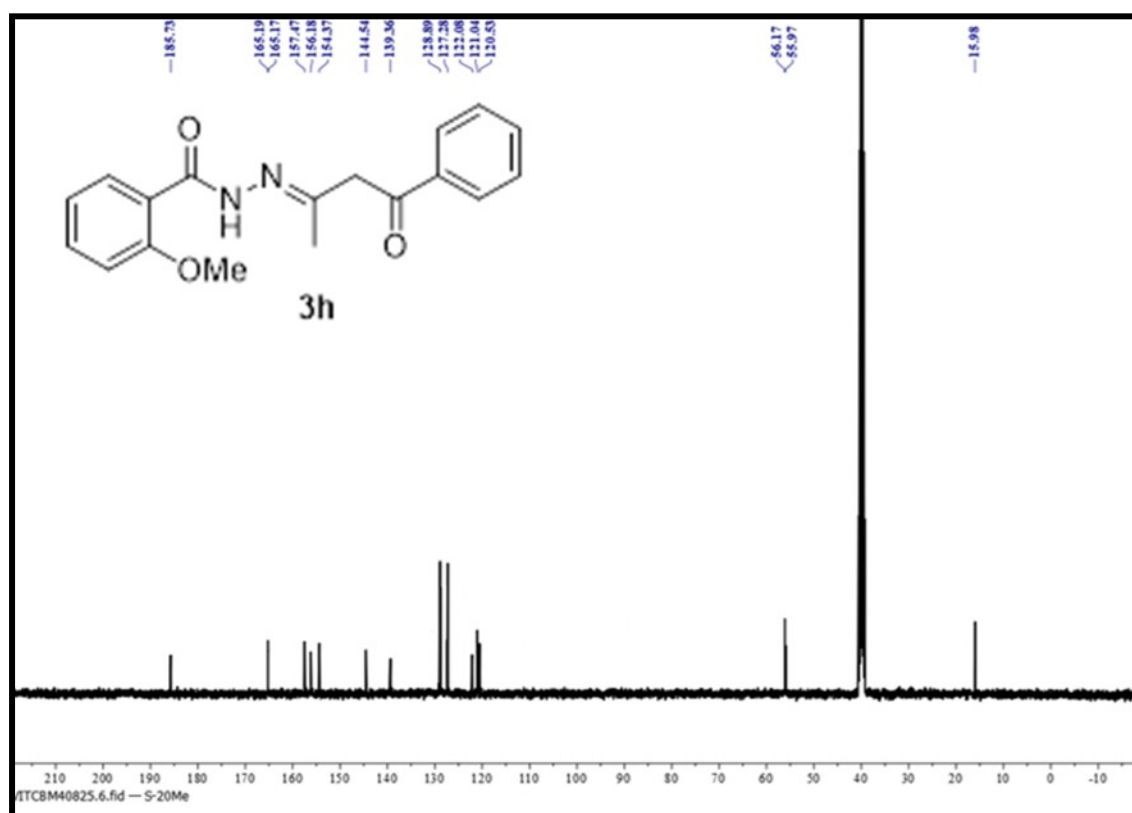
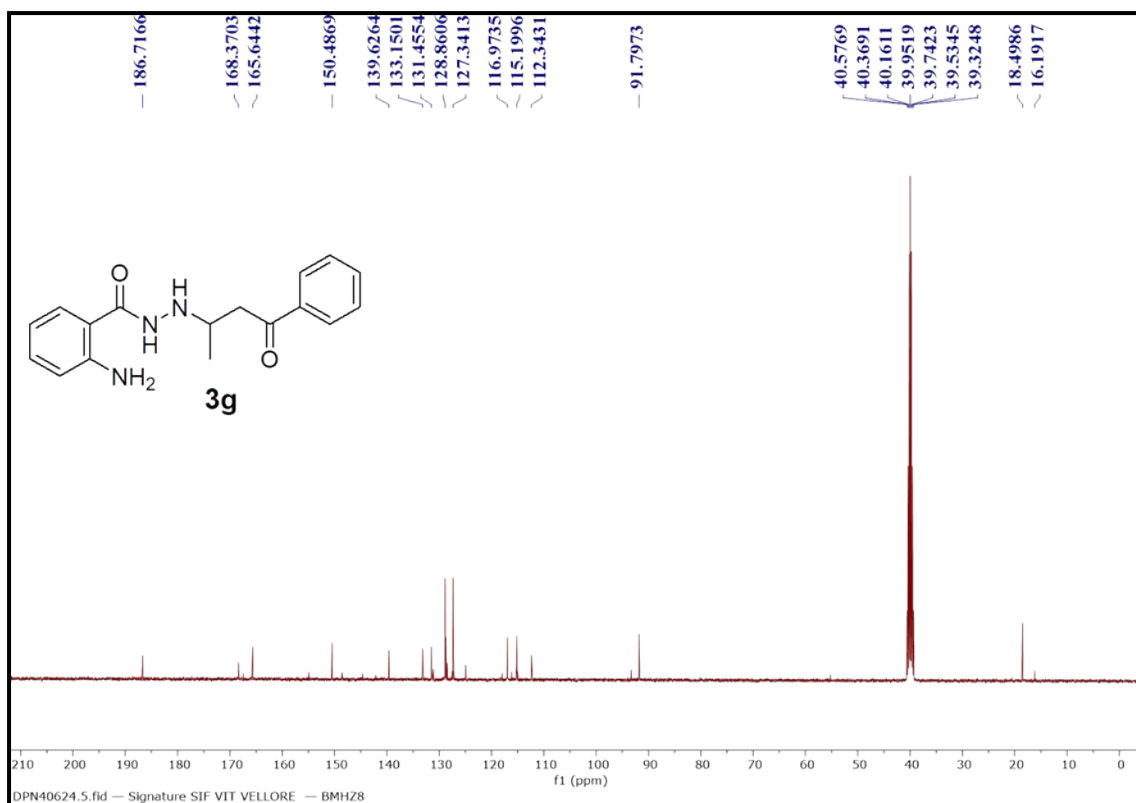


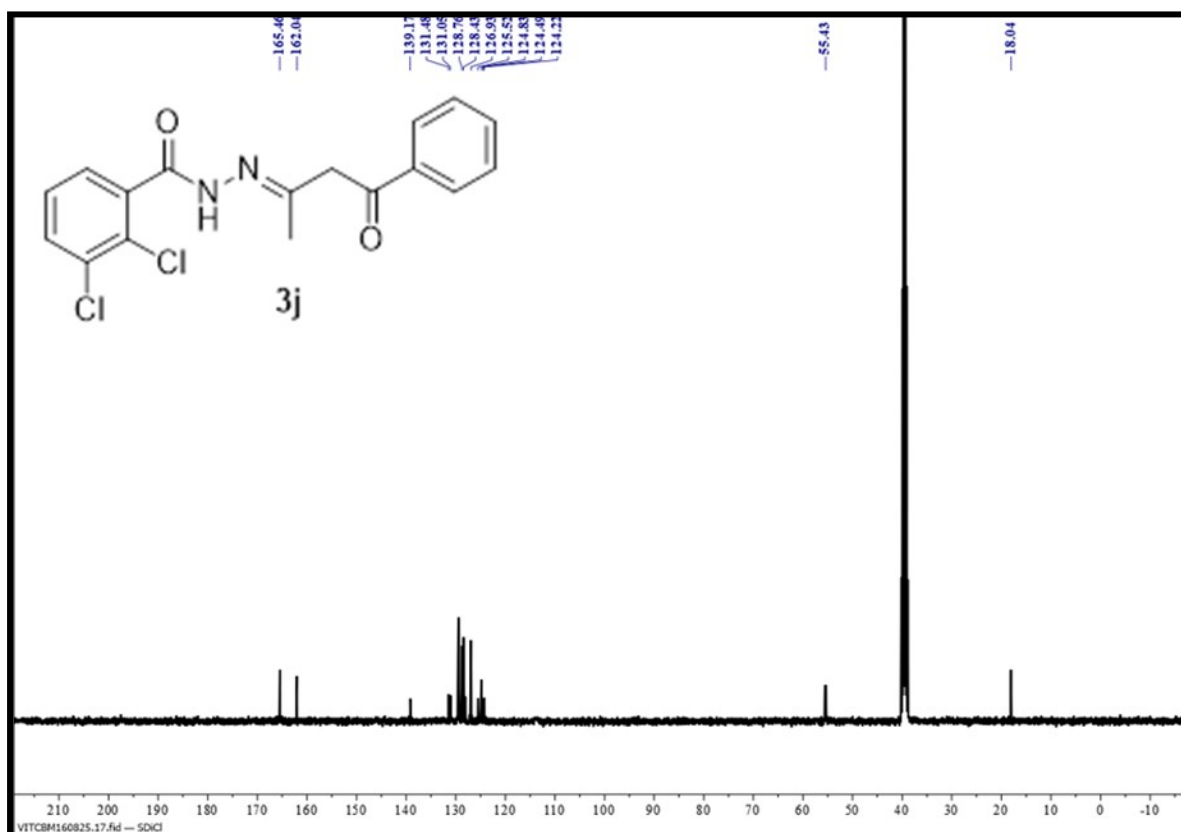
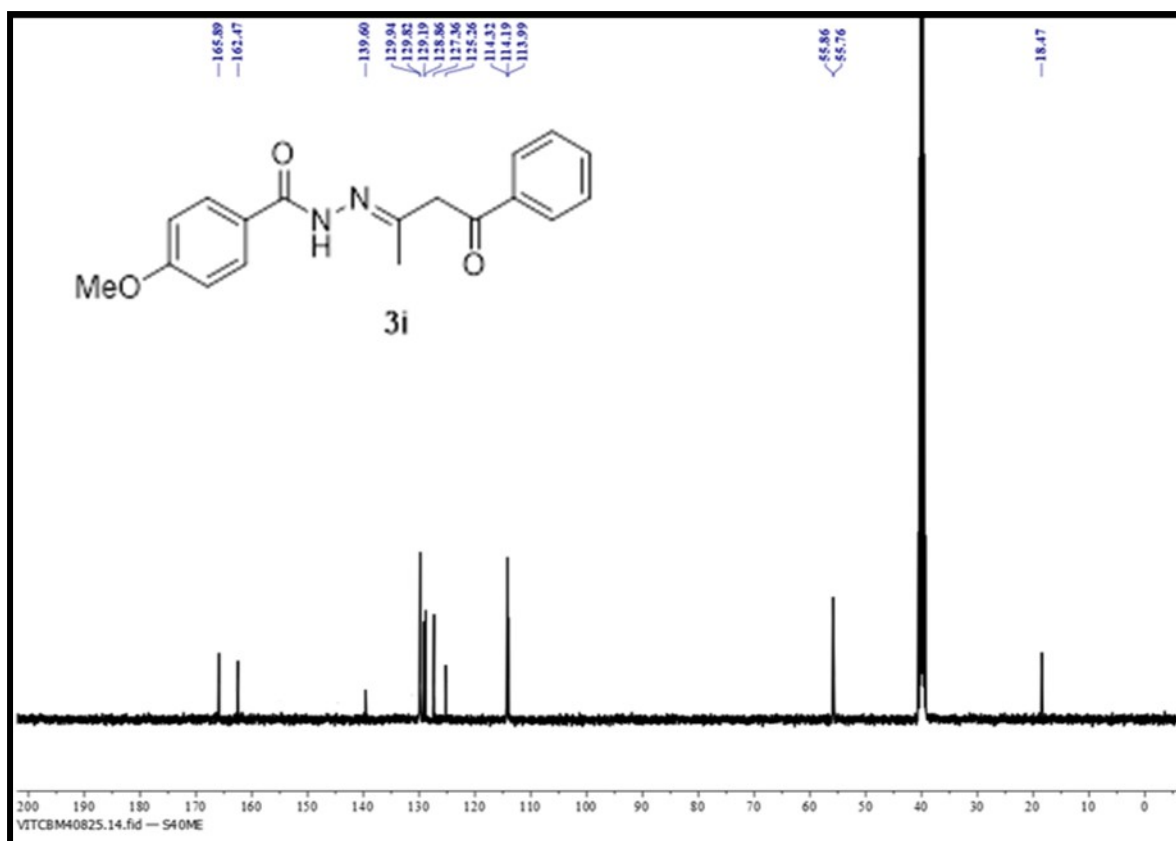
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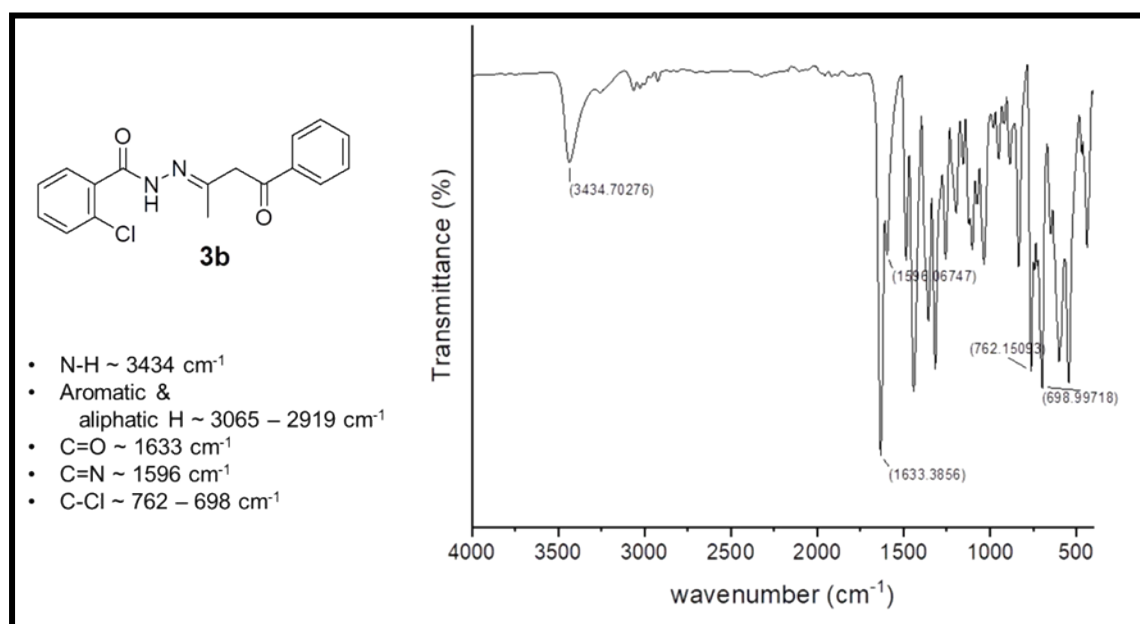
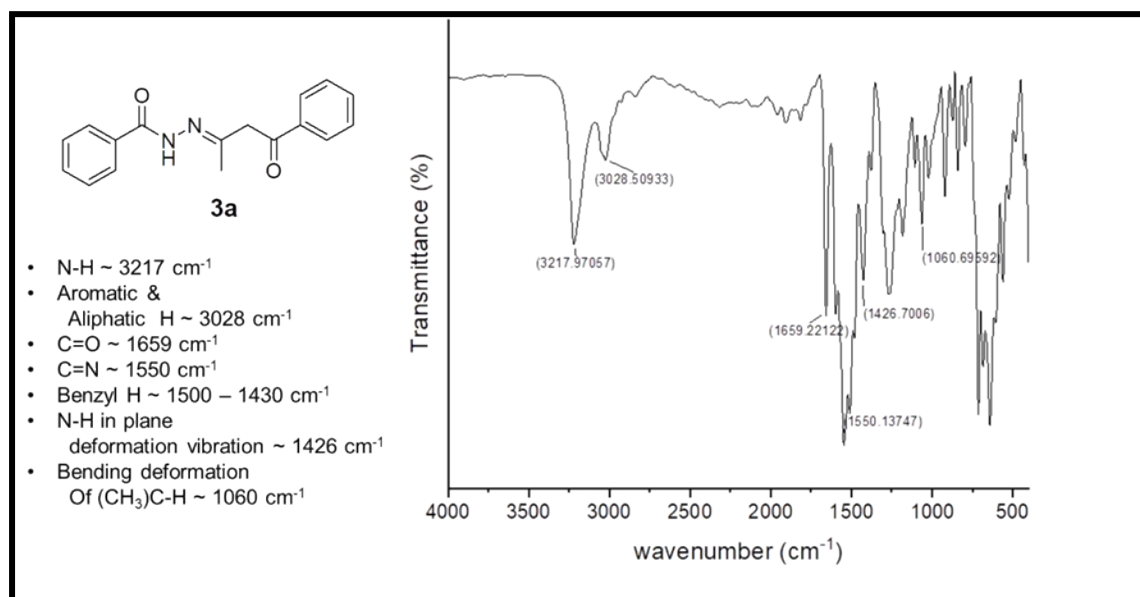


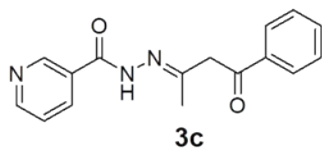




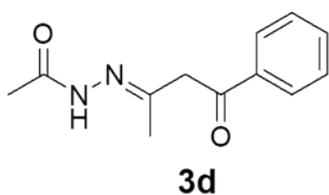
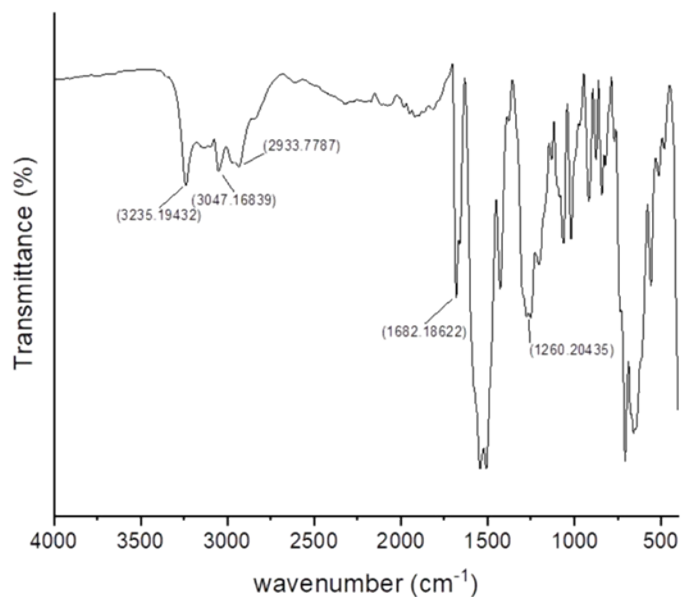



IR Spectra

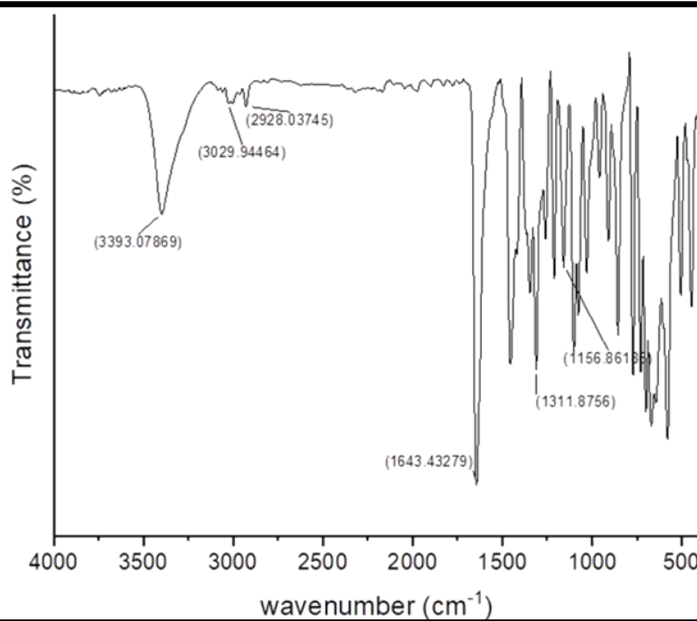


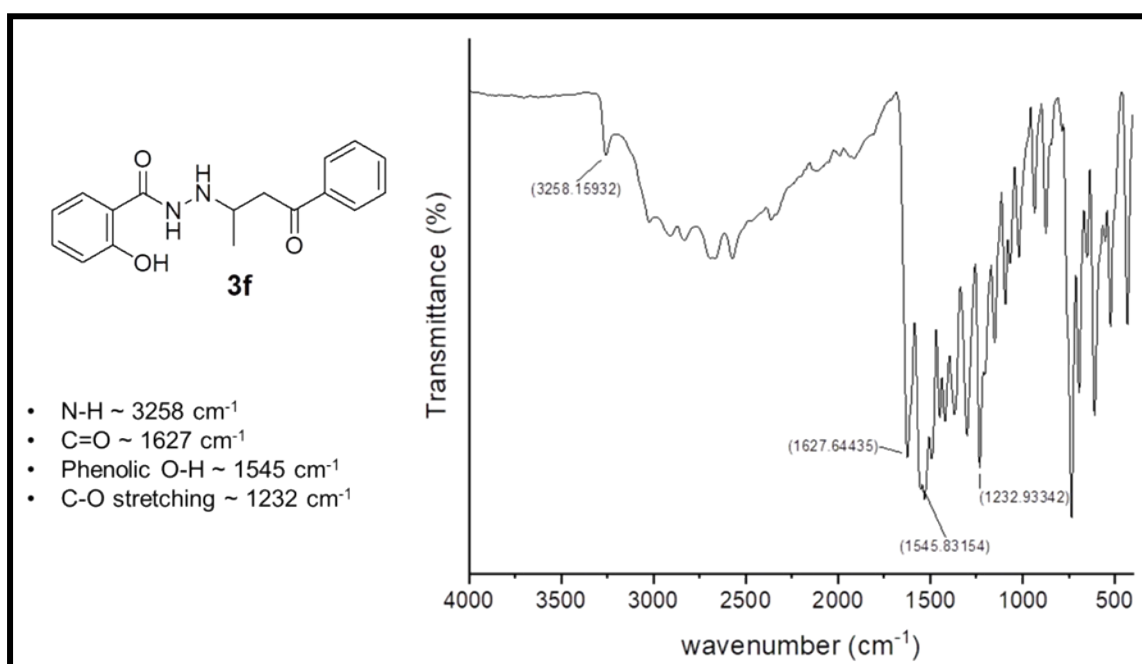
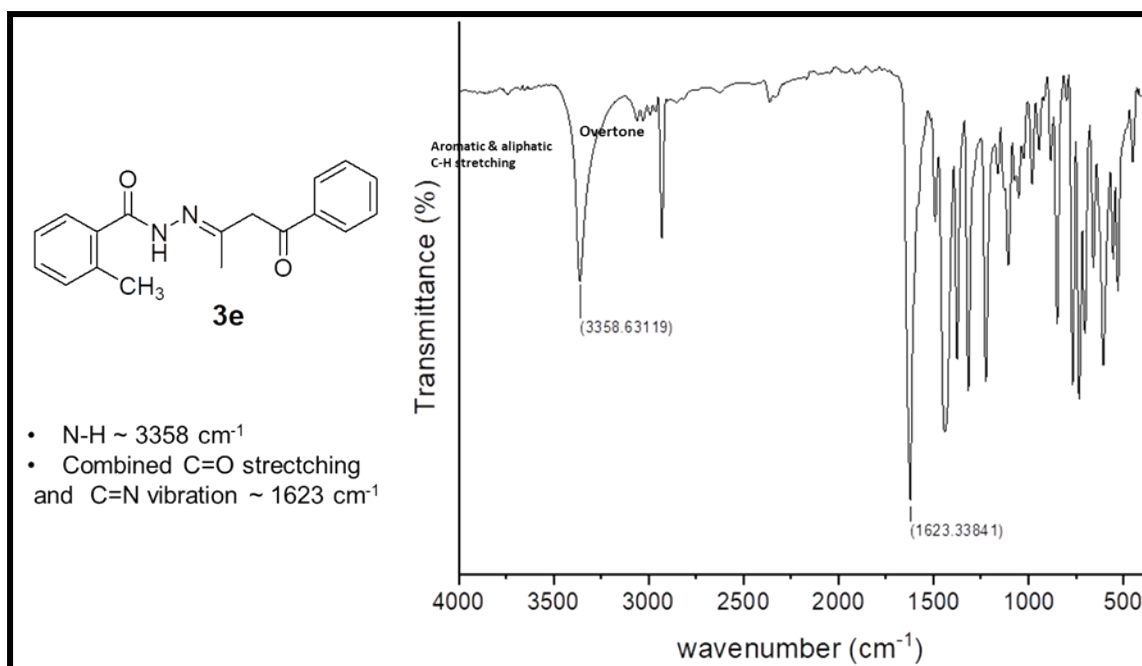


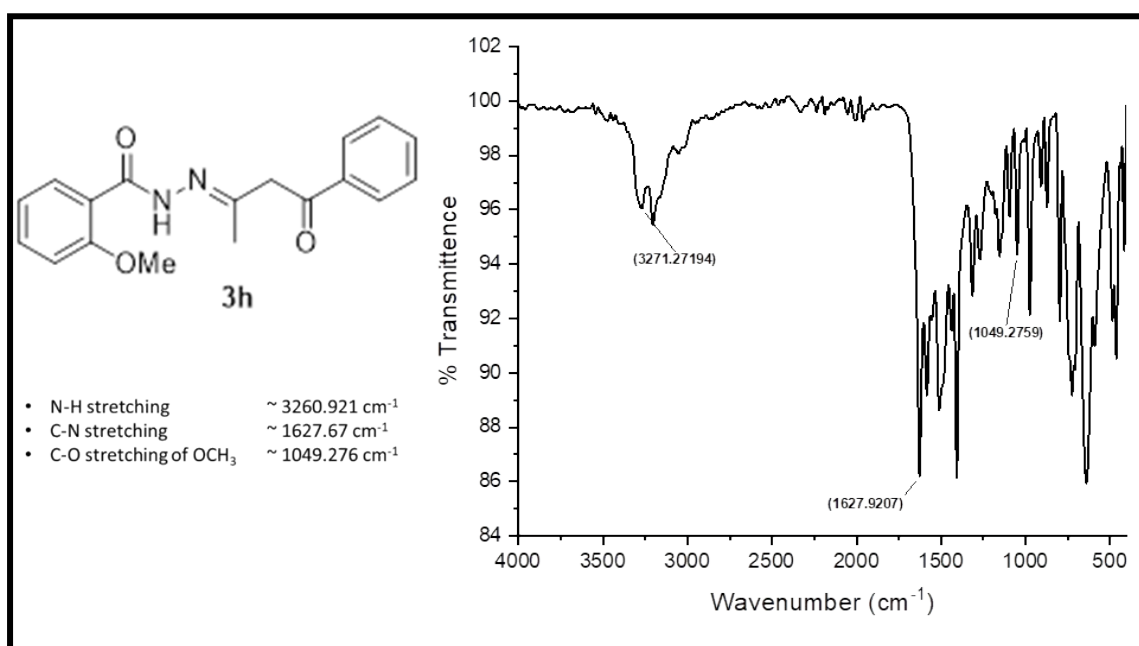
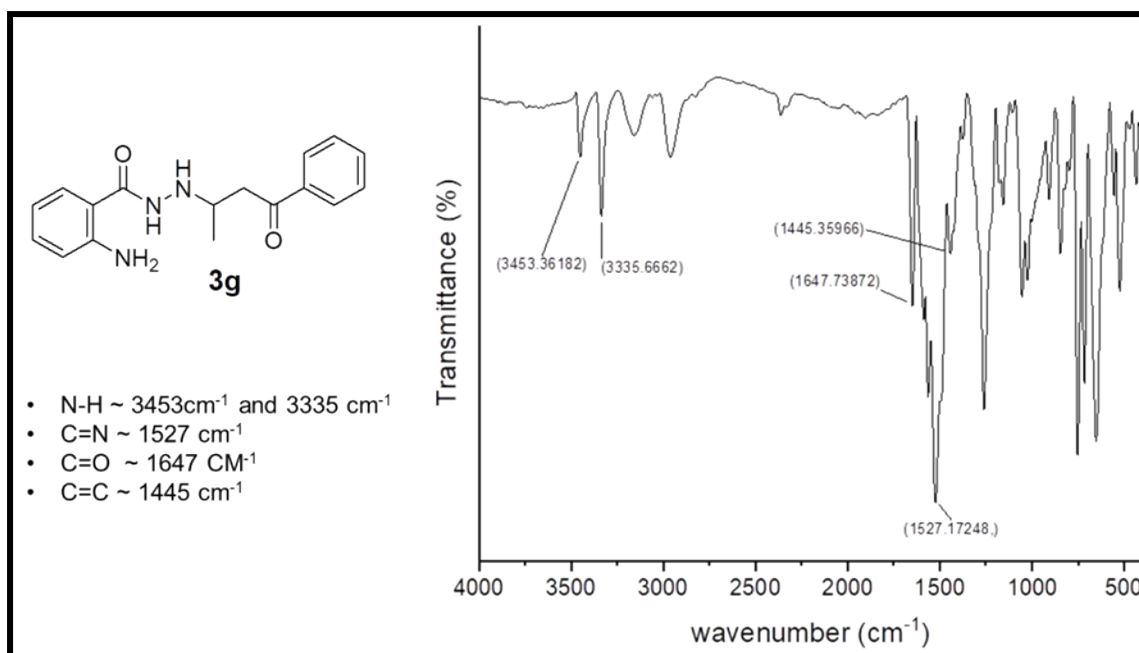
- N-H $\sim 3235\text{ cm}^{-1}$
- Aromatic & aliphatic H $\sim 3056 - 2933\text{ cm}^{-1}$
- C=O $\sim 1682\text{ cm}^{-1}$
- Aromatic C=C $\sim 1500 - 1430\text{ cm}^{-1}$
- N-H in plane deformation vibration $\sim 1428\text{ cm}^{-1}$
- C=O vibration Wrt. Pyridine $\sim 1260\text{ cm}^{-1}$

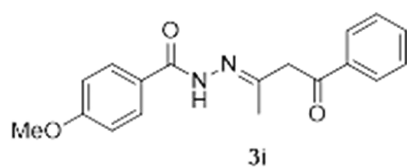


- N-H $\sim 3393\text{ cm}^{-1}$
- Aromatic & aliphatic H $\sim 3029 - 2928\text{ cm}^{-1}$
- C=N $\sim 1643\text{ cm}^{-1}$
-  $\sim 1156\text{ cm}^{-1}$
- $\text{CH}_3\text{C}=\text{O}$ bend $\sim 1311\text{ cm}^{-1}$

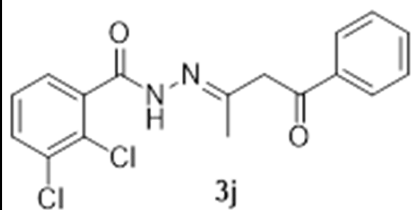
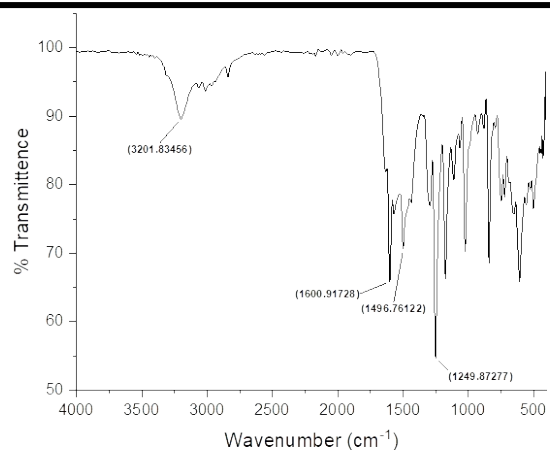




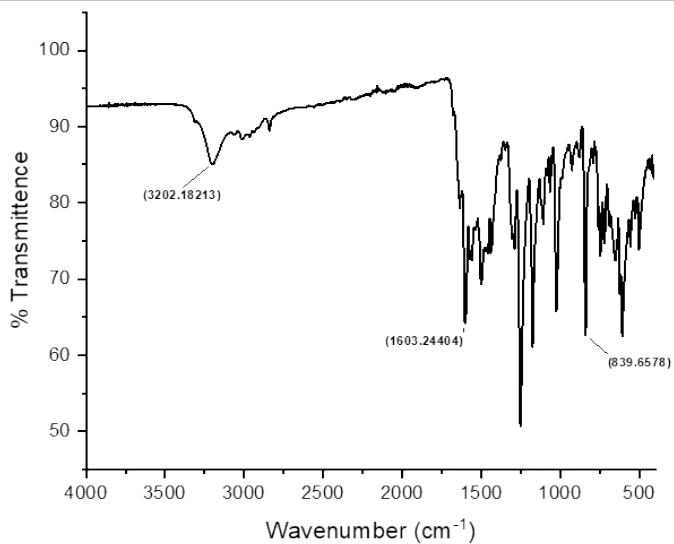




- N-H stretching $\sim 3201.835 \text{ cm}^{-1}$
- C=O stretching $\sim 1600.917 \text{ cm}^{-1}$
- C=C aromatic stretching $\sim 1445 \text{ cm}^{-1}$
- C-O stretching of OCH₃ $\sim 1249.872 \text{ cm}^{-1}$

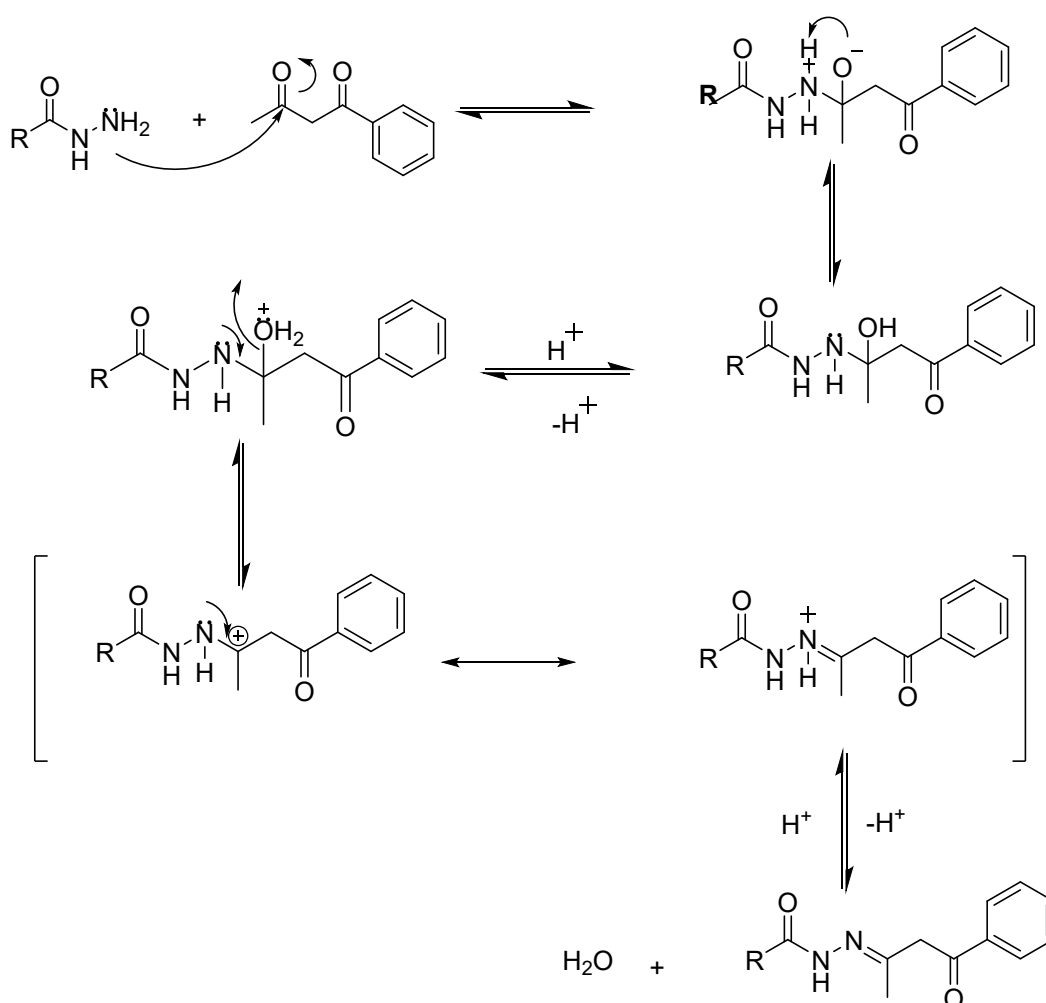


- N-H stretching $\sim 3202.182 \text{ cm}^{-1}$
- C=N stretching $\sim 1603.244 \text{ cm}^{-1}$
- C-Cl stretching $\sim 839.658 \text{ cm}^{-1}$



General mechanism for the synthesis of various hydrazone derivatives

Reaction between hydrazine and 1-phenyl-1,3-butanedione produces hydrazone. Initially, the amine containing a lone pair undergoes electrophilic addition with the carbonyl compound forming semicarbazone intermediate. Then the hydrogen atom in nitrogen will be deprotonated creating carbinolamine intermediate. Secondly, the intermediate hydroxyl group undergoes protonation and introducing a good leaving group – water molecule. Thus the second step involves dehydration of the molecule. Finally, after rearrangement, hydrazone inhibitor is crystallized. The mechanistic scheme for the synthesis is shown in the below scheme.



Evaluation of interaction potential of various hydrazide derivatives to the binding pocket of dNS1 protein.

The ability of various hydrazide derivatives (**3a-j**) to interact with the binding pocket of dNS1 protein was evaluated by following the changes observed in the intrinsic fluorescence of the protein with increasing concentrations of various hydrazides (0 – 500 μ M). The reaction was performed in a volume of 3 ml at room temperature and at a fixed protein concentration of $\sim 10^{-5}$ M in 1x PBS, pH 7.2. The reaction mixture was excited at 280 nm and the emission was monitored in the range of 290 to 550 nm. The quenching of fluorescence was observed for every incremental addition of hydrazide derivative.

Molecular docking

Structure retrieval and preparation: Three dimensional protein structure of Dengue type 4 was retrieved from RSCB protein databank (PDB ID: 8wbb) with a structural resolution of 2.9Å. Using the same as template file, the target protein sequence (dNS1) model was built from Swiss-model. Further the prepared protein macromolecule was optimized by protein preparation wizard of Schrodinger Suite before docking. The 2D ligand structures were drawn using ChemDraw Ultra 8.0. Optimization of the ligand was carried out using Gaussian 09 program by means of density functional theory – B3LYP involving 6-31 ++ G(d,p) basis set to get 3D confrontation . Which was further optimised using LigPrep Module of Schrodinger suite. Here, the ligands were retained in their original state, hydrogen bonds, bond angles, with single conformations after which it was utilized in docking. The protein was pre-processed, followed by minimization and then optimisation in order to retain the protein's functionality. This may prevent any erroneous output and further used for docking. The binding interactions and binding energy were calculated with Glide Module of Schrodinger suite. Appropriate grid was generated around the target protein's binding site for the prepared ligand to precisely interact with the target site. Further, MM-GBSA procedure was followed to ascertain the binding interactions as evaluated by glide docking.

MD-Simulation

DESMOND module of Schrodinger Suite was used for Simulation. The protein-ligand complex was built through system builder panel and localized in an orthorhombic box of TIP3P solvent module. Further physiological salt (NaCl) of 0.15M was added to neutralise

the environment. The complex was simulated using OPLS4 force field under 100ns period using NPT ensemble class at 300k and 1.013 bar atmospheric pressure.

Molecular cloning, Protein expression and purification

The codon optimized (for E.coli) gene sequence of dNS1 was commercially procured and cloned into pET28A vector between NdeI and XhoI restriction sites. The clone was constructed in such a way to possess N-terminal 6X His tag. The confirmation of constructed clone was done by sequencing. Further confirmation was performed by digestion with restriction endonucleases NdeI and XhoI to release a pop out of ~1200 bp.

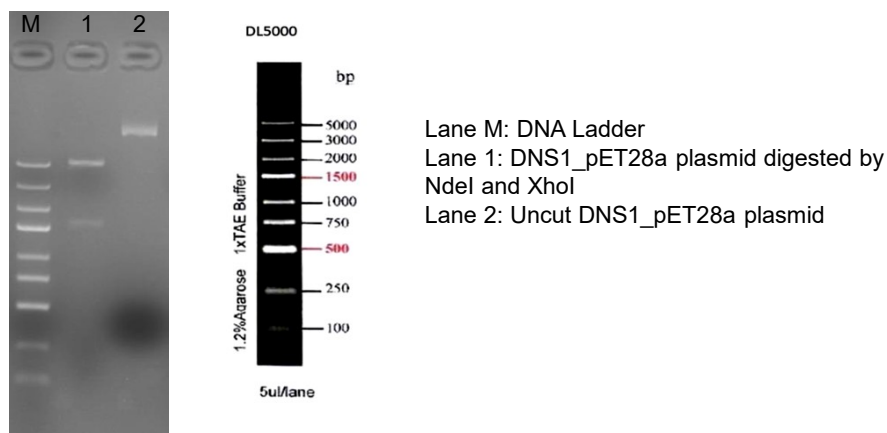


Figure S1. Agarose gel depicting the release of NdeI and XhoI double digested fragment (~1150 bp) and the undigested plasmid (~6 kb)

Protein sequence

MGSSHHHHHHSSGLVPRGSHMDMGCVVSWNGKELKCGSGIFVVDNVHTWTEQYK
FQPESPARLASAILNAHKDGVCGIRSTTRLENIMWKQITNELNYVLWEGGHDLTVVA
GDVKGVLTKGKRALTPPVNDLKYSWKTWGWKAKIFTPEARNSTFLIDGPD TSECPNER
RAWNFFEVEDYGFGMFTTNIWMKFREGSSEVCDHRLMSAAIKDQKAVHADMGYWI
ESSKNQTWQIEKASLIEVKTCLWPKHTLWSNGVLESQMLIPRSYAGPFSQHNYRQG
YATQTAGPWHLGKLEIDFGCEPGTTVTIQEDCDHRGPSLRTTTASGKLVTQWCCRSC
TMPPLRFLGEDGCWYGMEIRPLSEKEENMVKSQVTA

Gene sequence

atgggcagcagccatcatcatcatcacagcagcggcctggcgccgcggcagccatatggatatgggttgctgttagctgga
acggtaaagagctgaaatcggttcgggcatcttcgtgtggataacgttcacacctggaccgaacagtacaaattcagccggaagt

ccggcacgtctggcatctgcaattctgaacgcgcataaagacggcggttgcggtattcgtagtaccacccgcctggaaaacatcatgtg
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gaaattcgtccgctgagcgaaaaagaagaaaacatggtaaaagccaggtcaccgcataactcgag

Further the expression of the above construct was done by transforming the above plasmid into BL21D3 competent cells followed by 0.5 mM IPTG induction at OD600 ~0.5 for 4 hours. Subcellular fractionation was performed by lysing the cells in the presence of 0.5 M PMSF followed by centrifugation at 10000 xg for 30 minutes at 4 oC. The fractionated cell lysates were subjected to SDS-PAGE. The expressed protein was found to be localized as inclusion bodies. In order to solubilize the inclusion bodies protein, the pellet was treated with 6 M guanidinium hydrochloride for 6 hours followed by centrifugation at 10000 xg for 45 minutes. The above supernatant was subject to Ni-NTA affinity purification using 30 mM imidazole in 1x PBS, pH 7.4 as wash buffer and 300 mM imidazole in 1XPBS as elution buffer. Thus eluted protein was extensively dialyzed against phosphate-buffered saline (PBS) to remove imidazole.

Optimization of protein expression

The expression of dNS1 protein in BL21DE3 was optimized under various conditions of temperature (30 and 37 oC), IPTG concentration (0.5 mM and 1 mM) and post induction time (4 hours and overnight). SDS-PAGE results have revealed that best expression was observed at 37 oC with 0.5 mM IPTG for 4 hours of induction. The optimized expression of dNS1 is depicted in figure S1.

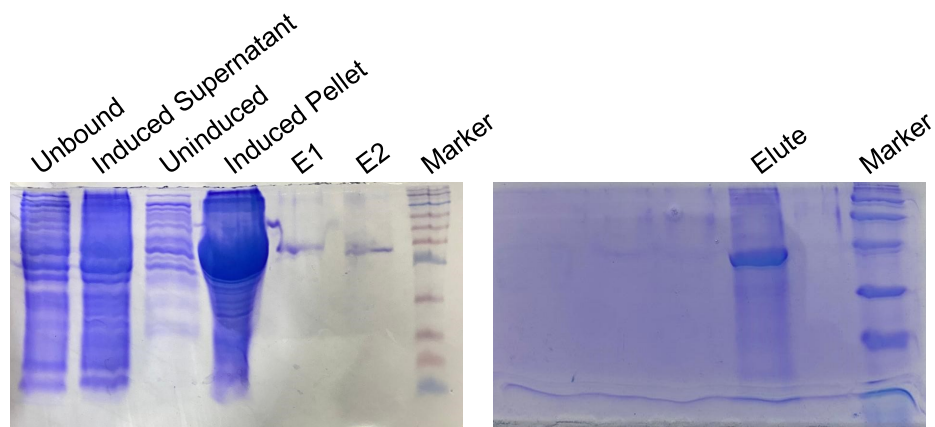


Figure S2. SDS-PAGE depicting the expression, sub-cellular localization and purification of dNS1 protein from pET28a- dNS1 construct.

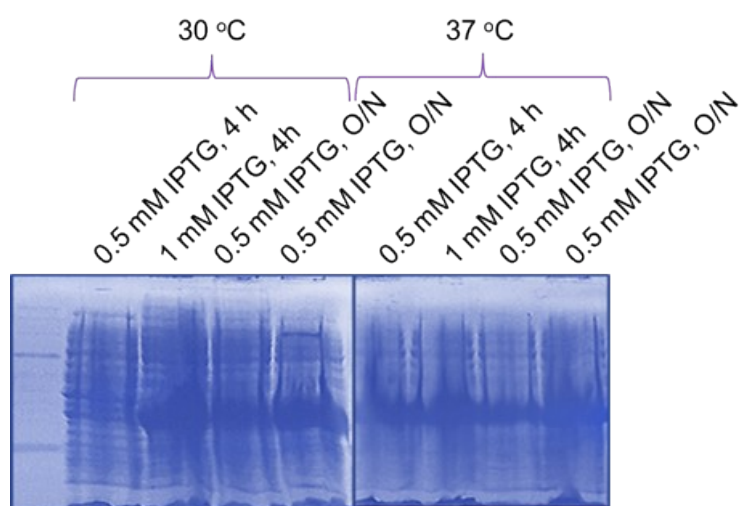


Figure S3. SDS-PAGE depicting the optimized expression of dNS1 in BL21-DE3 cells.

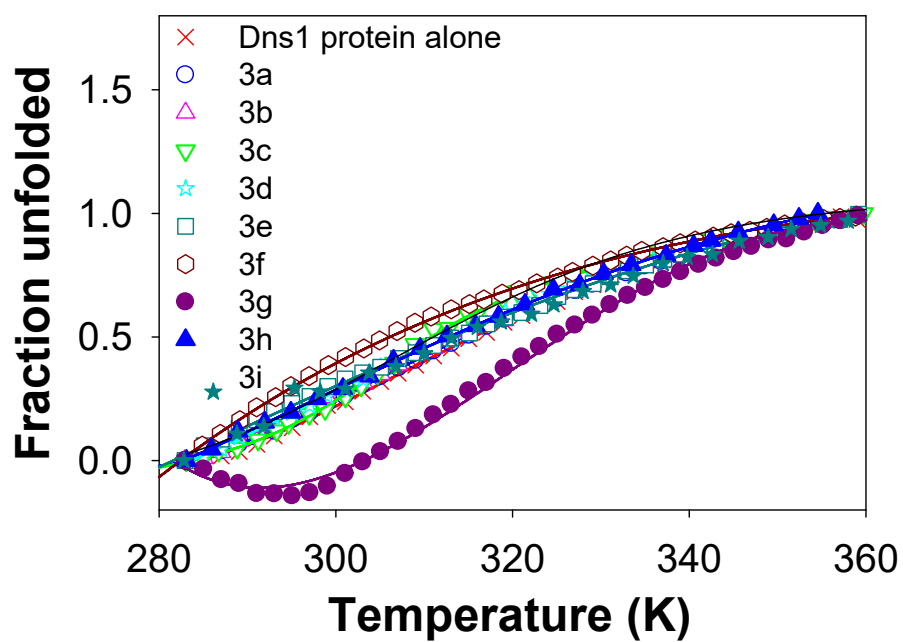
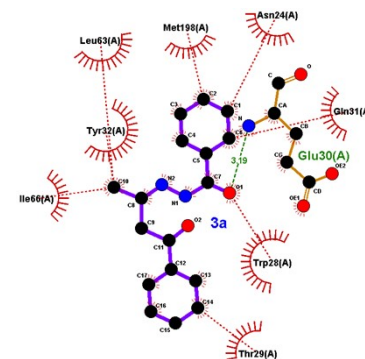
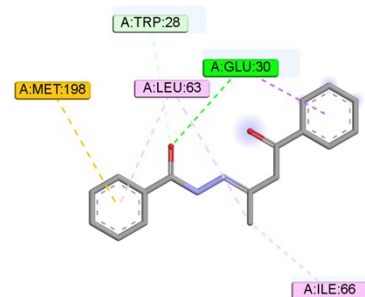
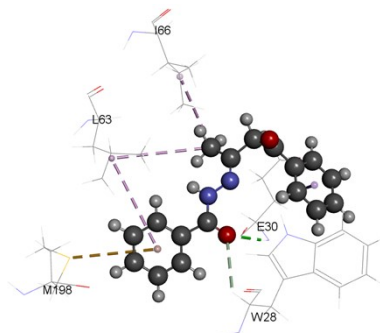
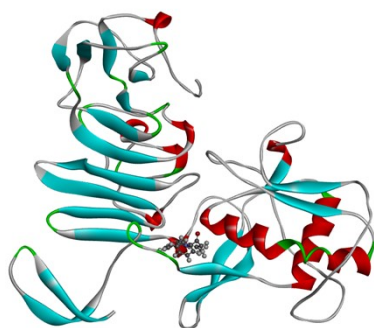
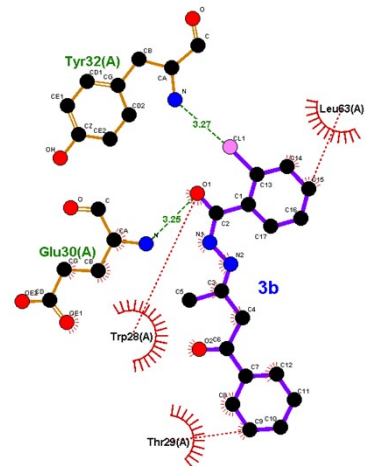
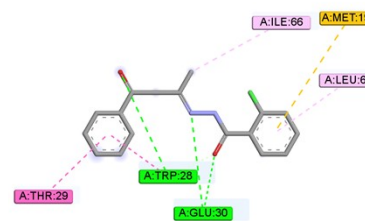
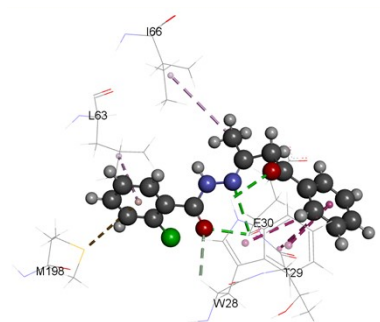
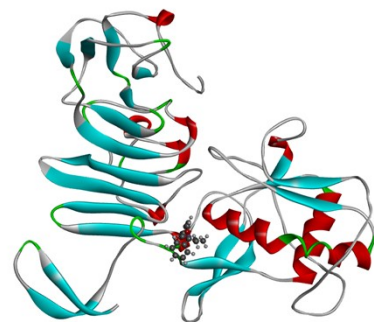


Figure S4. Plot depicting the melting temperature of dNS1 protein in the presence and absence of various hydrazides.

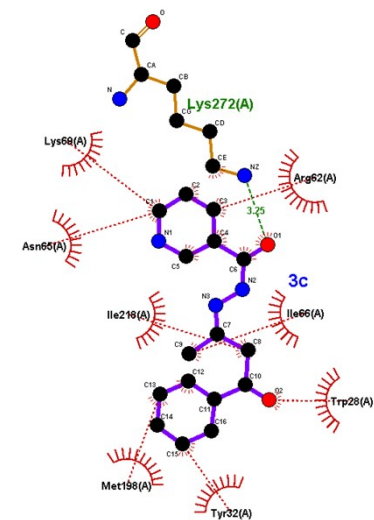
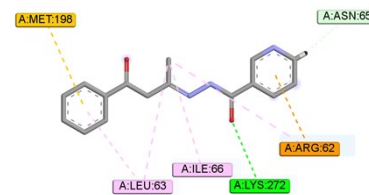
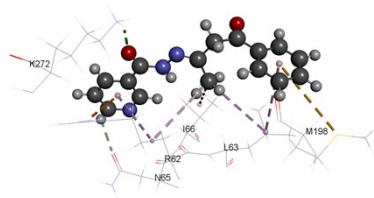
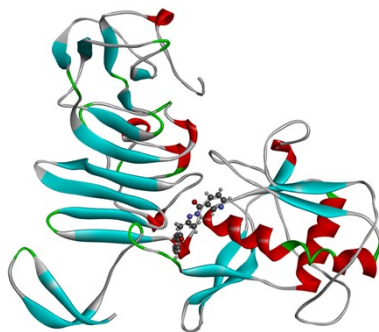
3a



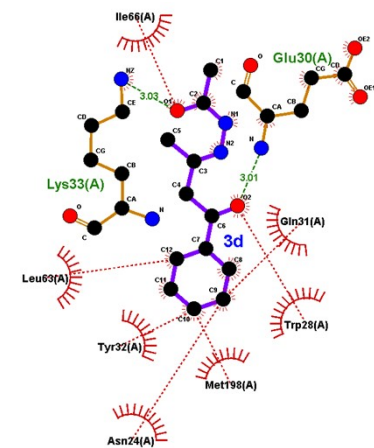
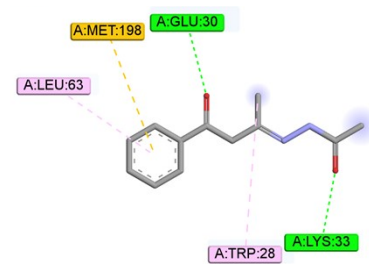
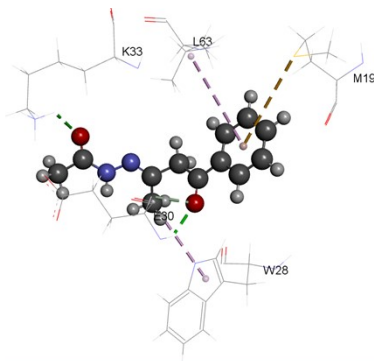
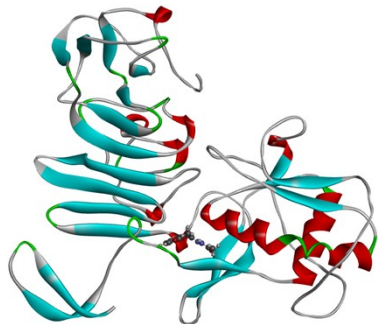
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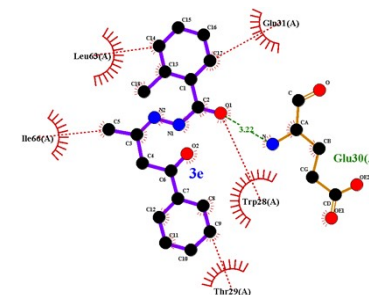
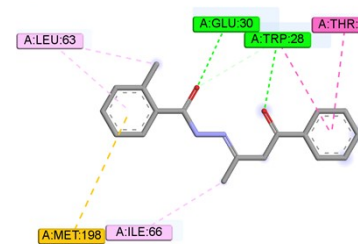
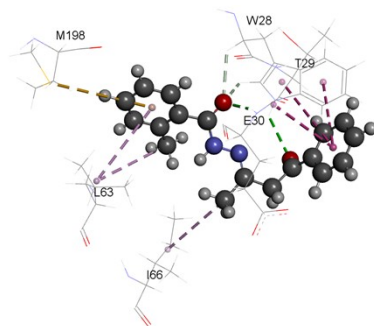
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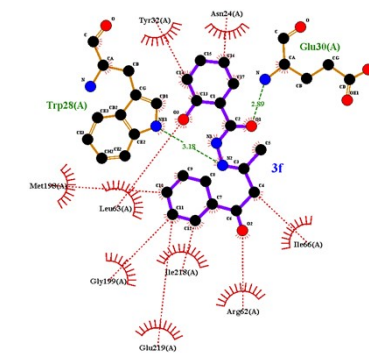
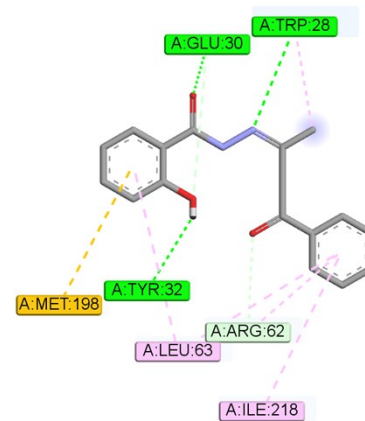
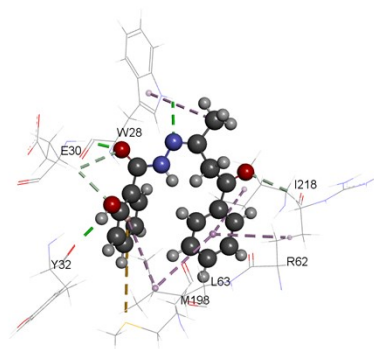
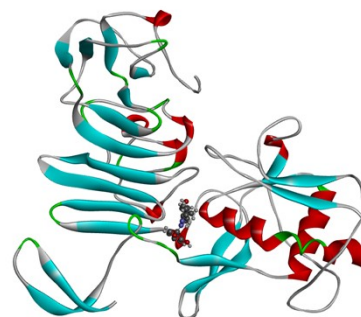
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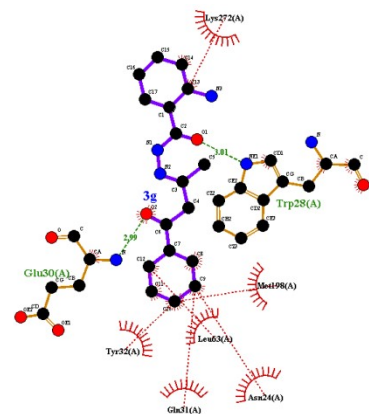
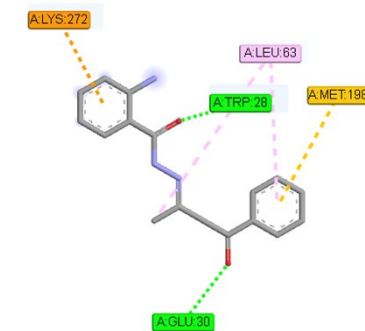
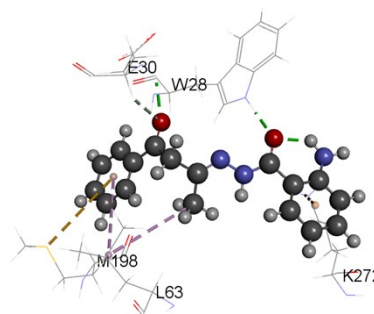
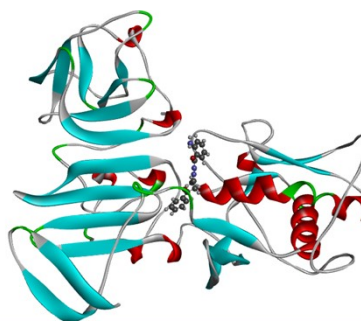
3e



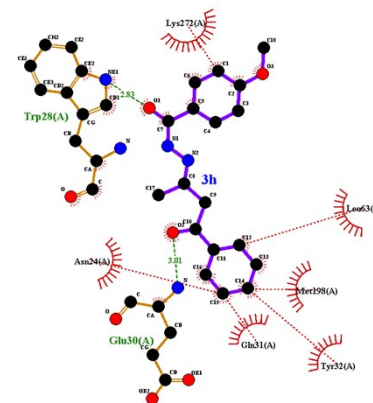
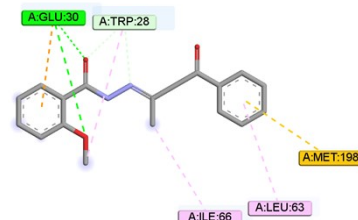
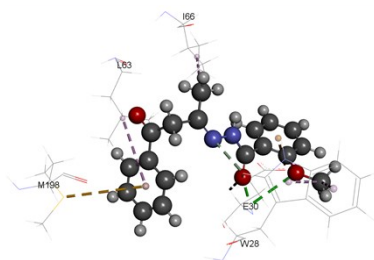
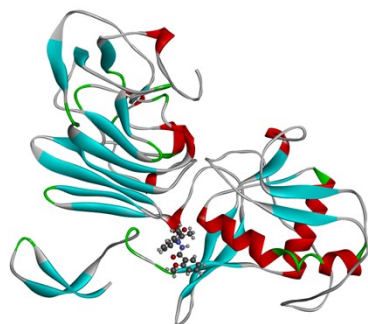
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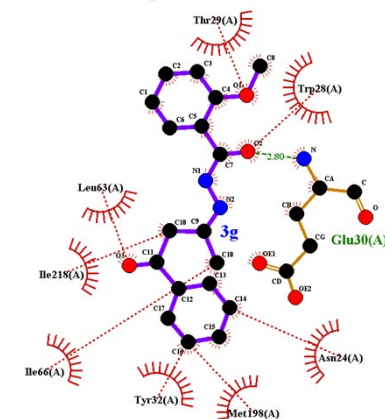
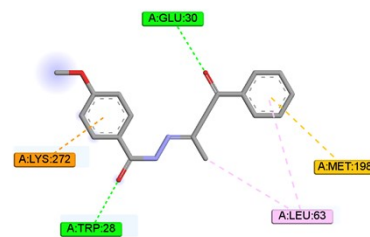
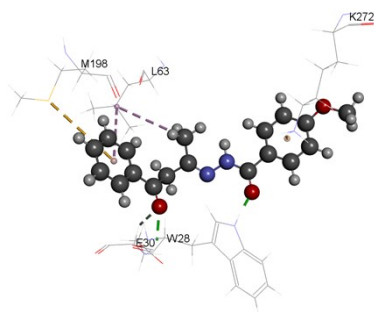
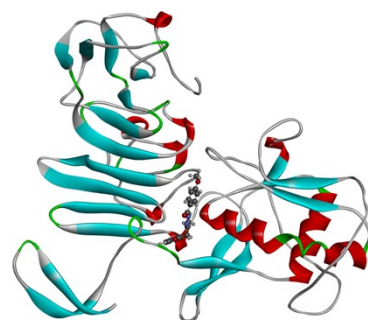
3g



3h



3i



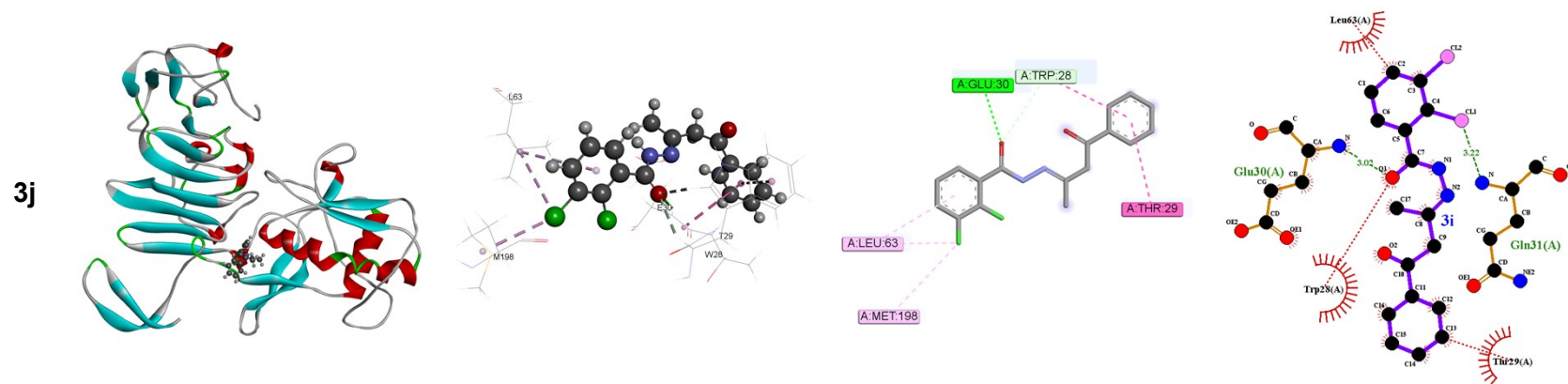


Figure S5. Docked poses of various hydrazide derivatives with dNS1.

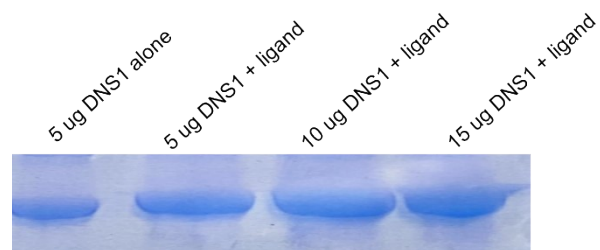


Figure S6. Native gel electrophoresis depicting the migration of DNS1 protein in the presence and absence of ligand **3b**.

Pharmacokinetic properties

Analyzing the Pharmacokinetic parameters of a drug exhibiting their potential treatment towards the target has an important role before synthesis. To understand the target quality such as Absorption, Distribution, Metabolism, Excretion, Toxicity study was performed by QikProp module of Schrödinger suite. So, the descriptors like HOA (Human Oral Absorption), stars, Lipinski rule of five were under study. Human Oral Absorption predicted on 1,2 or 3 scale for low, medium and high, indicates the values greater than 1 can become a potent drug candidate. Star is a number of property or descriptor that dis-obey when compared to 95% of known drugs. Descriptors such as molecular weight, dipole, Ionization Potential, Electron Affinity, Solvent Accessible Surface Area, hydrophobic component of SASA between saturated carbon and hydrogen(FOSA), hydrophilic component of SASA on N,O, and H on heteroatom (FISA), π – component of SASA (PISA), weakly polar component of the SASA of halogens, phosphorous, and sulfur atoms (WPSA), van der waals surface area of polar nitrogen and oxygen atoms (PSA), Globularity descriptor, volume, #rotor, donor hydrogen bond, acceptor hydrogen bond, polarizability in cubic angstroms (QPpolrZ), hexadecane/gas partition coefficient (QPlogPC16), octanol/gas partition coefficient (QPlogP_{oct}), water/gas partition coefficient (qplogPw), QPlogPo/w, aqueous solubility (QlogS), binding to human serum albumin (QPlogKhsa), Blood-Brain partition coefficient (QPlogBB), number of likely metabolic reactions (#metabol) were involved in the determination of star. A large number of stars tell us the molecule is unlikely for a drug. Besides, Lipinski rule of five provides suggestions for a compound's druggability: molecular weight (<500), Donor hydrogen bond (≤ 5), octonal-water coefficient (<5), acceptor hydrogen bond (≤ 10), aqueous solubility (-6.5 to 0.5). Table 3 shows the results of ADME analysis. Except 2-aminohydrazone (**3g**) rest of the compounds **3a – j** are satisfactory and therefore further analysis was preceded. QPlogPW should be from 4 to 45. QPlogPo/W should be

between -2 to 6.5. QPPCaCo is CaCo-2 cell permeability which has to greater than 25 and not beyond 500. From the discussion, except 2-aminohydrazone (**3g**), every other molecule tends to satisfy the parameters and could be considered potential against dNS1.