

Electronic Supplementary Material

Magnetic Hyperthermia Triggered Multi-Functional Thermo-Responsive Lipid Nanoparticles for Enhanced Paclitaxel Release and Cytotoxicity

Preparation of $\gamma\text{-Fe}_2\text{O}_3$ loaded TLN

PTX is soluble in ethanol, acetonitrile, and methanol (50 mg/cm³) ¹. Alternatively, it has low water solubility ², and high hydrolytic potential when exposed to different aqueous and organic media. It is rapidly destroyed in weakly alkaline ³ and strongly acidic aqueous solutions, whereas the lowest amount of degradation in aqueous solutions occurs at weakly acidic pH (3 to 5) ⁴. PTX solution of methanol undergoes hydrolysis and transesterification of ~30% after two weeks of storage at room temperature. Surfactant action on hydrophobic PTX employing Cremophor EL and ethanol blend (1:1) diluted with 5–20 fold (30 mg paclitaxel dissolved in 5 mL paclitaxel; Bristol-Myers Squibb) in aqueous solution ⁵ solubilizes and inhibits hydrolysis. CrEL is FDA approved, and already clinically used for intravenous injection ⁶. Cremophor EL is a thermally stable and hydrophilic ⁷ non-ionic polyoxyethylated surfactant ⁸ with an HLB value of 13.5 ⁹. During the emulsification process, high temperature may destabilize the TLN and change the curvature of the surfactant structure, which is prevented by the incorporation of highly thermally stable Cremophor EL ¹⁰.

The structure of the nanoparticle shell has a significant impact on physicochemical properties, drug release or leakage ¹¹. The leakage of payload is faster in the case of nanoparticles with large pores ¹² which are covered by a layer of larger surfactant molecules ¹³.

Solid saturated fatty acid based lauric acid is responsible for drug release upon degradation at neutral pH. Lauric acid is insoluble in water, but laurate has a solubility of more than 1000 μM in the buffer of pH 7.4 ¹⁴. Conversion of insoluble lauric acid to laurate depends on the pH of the medium i.e., no dissociation at pH < 4, while laurate ions are generated at circum-neutral pH and pH > 9 all lauric acid is in the form of laurate ions ¹⁵.

Factorial Optimization

Design of Experiments (DOE) is a method used to analyze the responses by exposing them to a set of different variables involving the least number of experiments runs to obtain the maximum optimum conditions.

Table S1: Best-fit model validation by examining ANOVA, F-value, lack of fit P-value, R², adjusted-R² - predicted-R², coefficient of variance (%CV), and adequate precision (ADP).

Parameter	Model	P-value	Lack of Fit P-value	F-value	R ²	Adjusted R ² - Predicted R ²	ADP	%C.V.	
Particle size	Linear	< 0.0001	0.1809	14.02	0.7244	0.1449	11.2427	15.24	Suggested
Polydispersity	Linear	0.0038	0.2779	6.74	0.5584	0.2227	8.7747	15.24	Suggested
Zeta potential	Linear	< 0.0001	0.5846	37.65	0.8759	0.0346	20.1769	6.82	Suggested
EE of PTX	2FI	0.0526	0.0823	3.71	0.9668	0.0995	30.1397	5.31	Suggested
EE of $\gamma\text{-Fe}_2\text{O}_3$	2FI	0.0228	0.5767	4.48	0.9236	0.065	18.3324	5.48	Suggested

Table S2: ANOVA results obtained from design expert analysis to validate the accuracy of all the experimental coefficients.

Source	A-Surfactant/Co-Surfactant	P- γ -Fe ₂ O ₃	C-Stirring Speed	AB	BC	AC
Particle size	0.0004	0.0002	0.9018	-	-	-
Polydispersity	0.0207	0.2525	0.003	-	-	-
Zeta potential	< 0.0001	0.0073	0.0359	-	-	-
EE of PTX	< 0.0001	< 0.0001	< 0.0001	0.0453	0.2137	0.0872
EE of Fe₂O₃	< 0.0001	< 0.0001	0.0015	0.0034	0.7025	0.5016

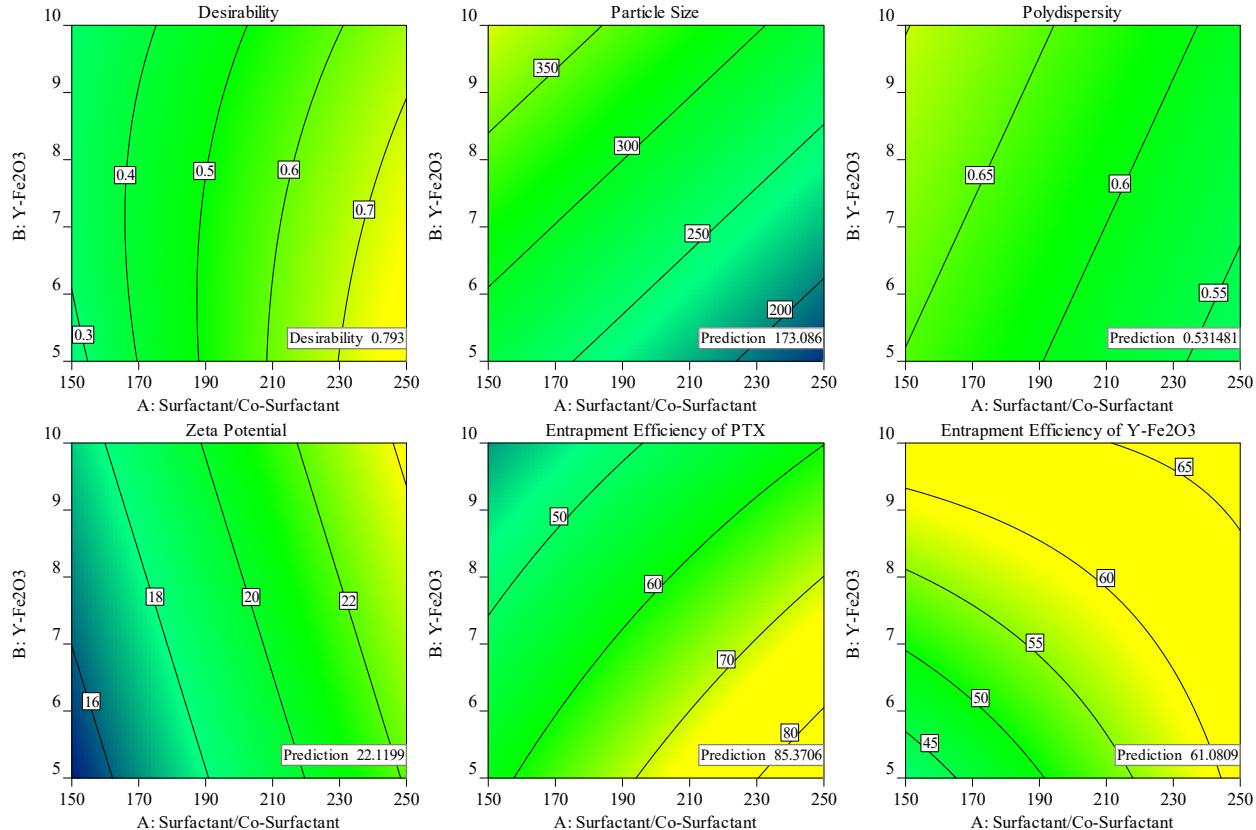


Figure S1: Contour illustration of desirability and interaction factor AB, for all the dependent variables.

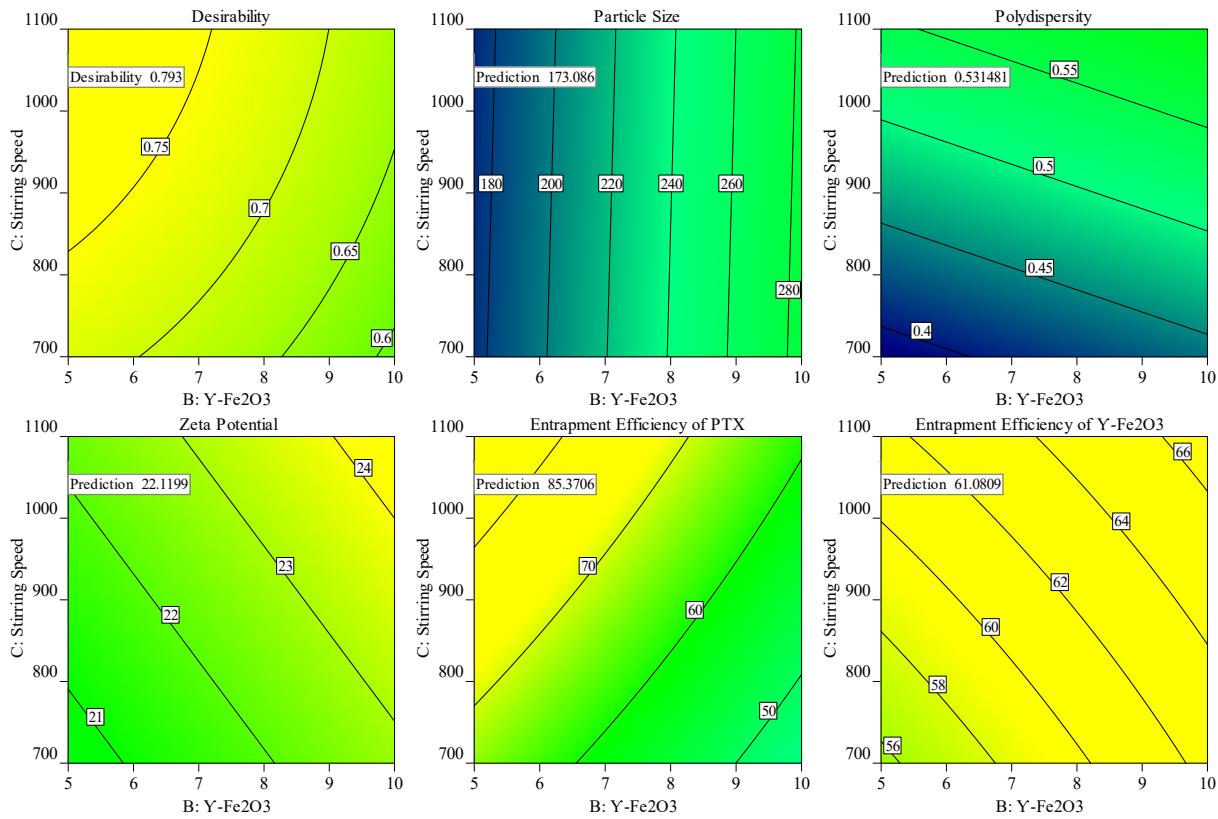


Figure S2: Contour illustration of desirability and interaction factor BC, for all the dependent variables.

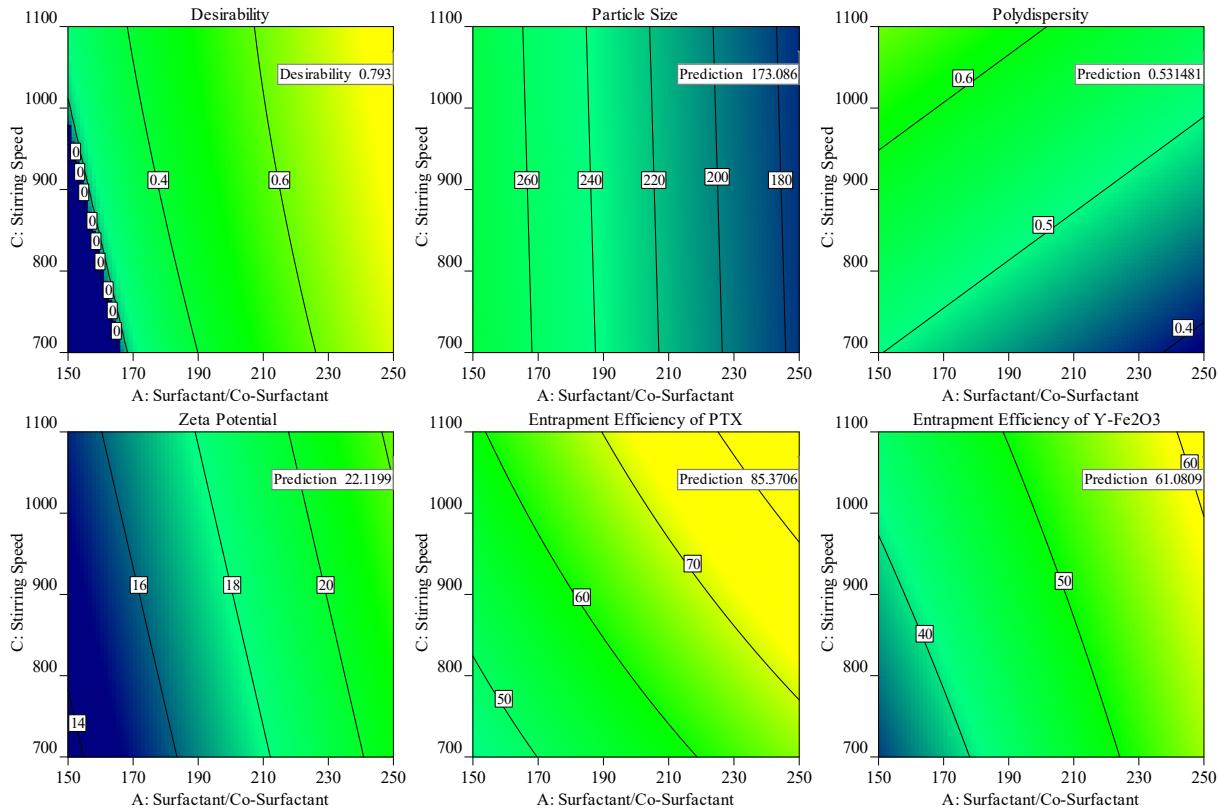


Figure S3: Contour illustration of desirability and interaction factor AC, for all the dependent variables.

Characterizations:

Determinization of hydrodynamic diameter PDI and zeta potential:

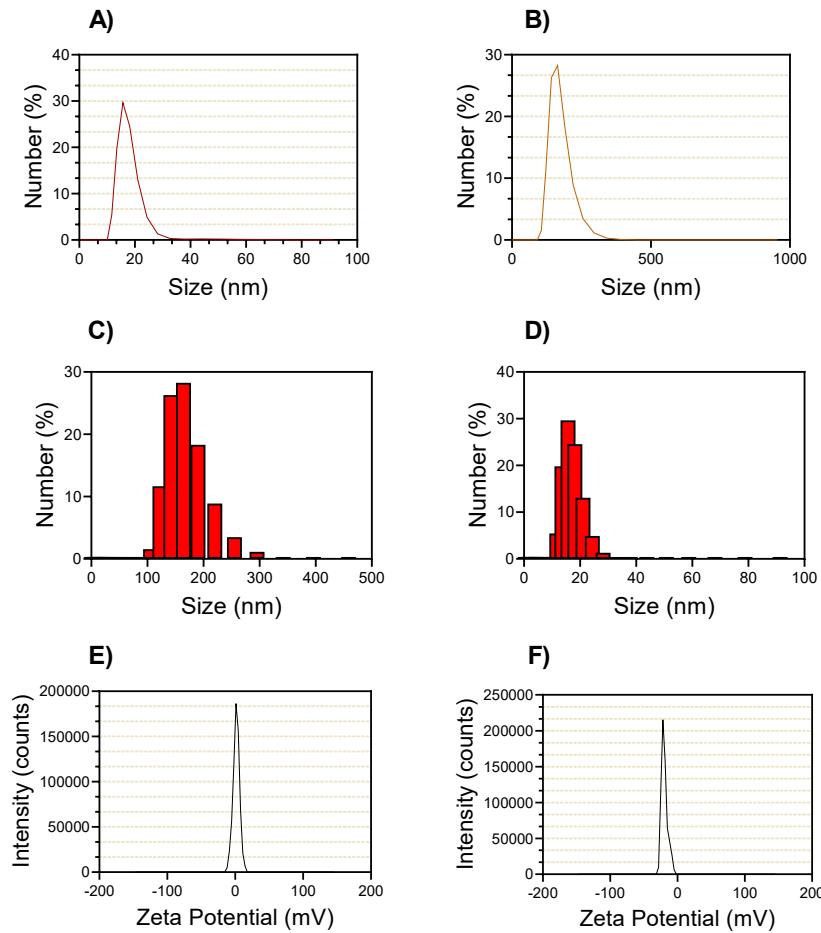


Figure S4: Size measurements for $\gamma\text{-Fe}_2\text{O}_3$ and P- $\gamma\text{-TLN}$ (A and B), size distribution analysis (C and D), and zeta potential measurements (E and F)

Compatibility analysis and Solid-State characterization:

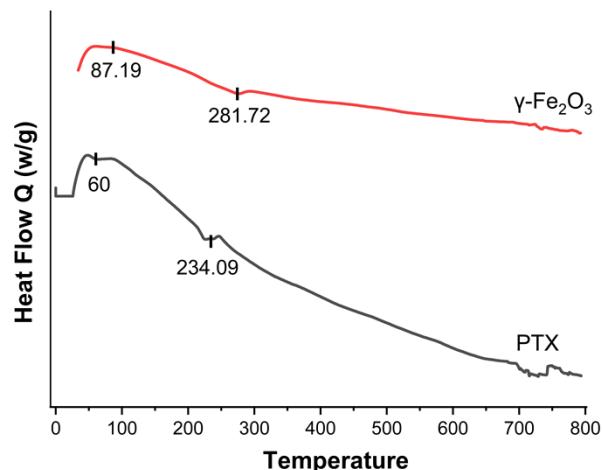


Figure S5: Thermograms for pure PTX and $\gamma\text{-Fe}_2\text{O}_3$ depicting degradations and melting.

Table S3: Calorimetric data depicting temperature variation, thermal stability, and thermal transitions in ingredients and lipid formulations.

Material	Onset (°C)	Melting peak (°C)	Endset (°C)	ΔH Melting enthalpy J/g	Tm-To
Lauric Acid	45.40	48.08	76.12	72.478	2.68
PTX	25.42	234.09	647.84	505.16	-
$\gamma\text{-Fe}_2\text{O}_3$	34.15	275.88	712.2	1129.7	-
P- γ -TLN (10 mg)	23.23	39.52	47.51	7.1695	16.29
P- γ -TLN (15 mg)	42.35	52.96	63.26	306.58	10.61

In vitro magnetic evaluations:

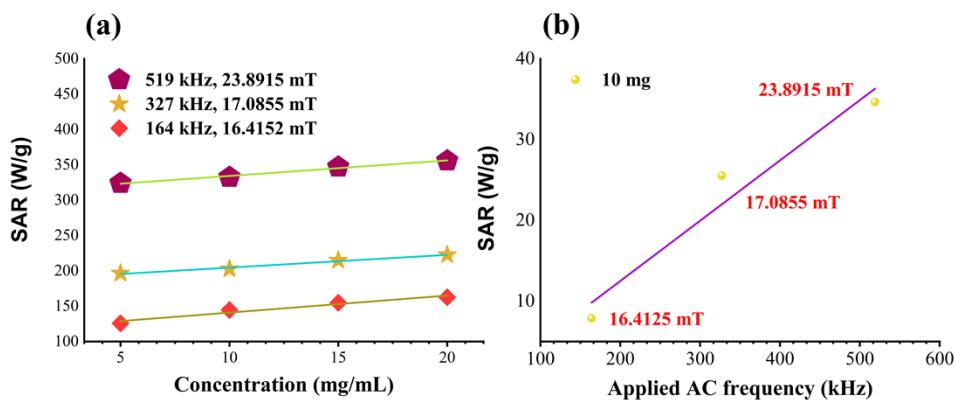


Figure S6: SLP of $\gamma\text{-Fe}_2\text{O}_3$ fluid samples measured in an ac magnetic field of different fields and frequencies as a function of concentration and (b) SLP of $\gamma\text{-Fe}_2\text{O}_3$ encapsulated TLN lyophilized samples measured at different fields and frequencies. The straight lines are a guide to the eye.

In vitro drug release study:

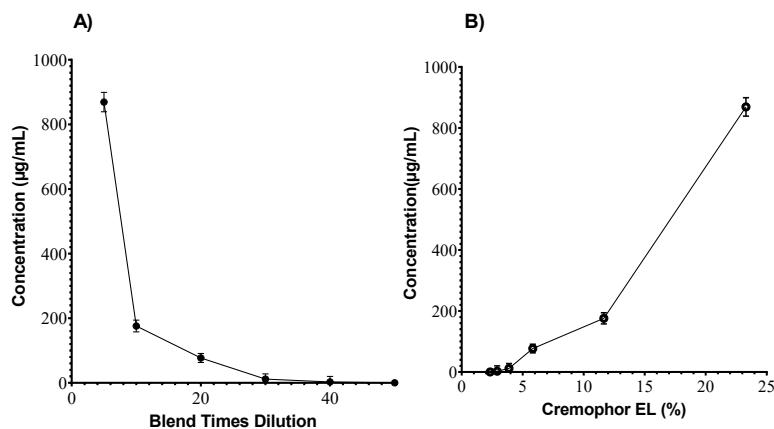


Figure S7: Dissolution estimation using Cremophor EL blend in terms of blend times (A) and percentage (B) corresponding to the PTX concentration ($\mu\text{g/mL}$).

Stability studies:

Table S4: Stability parameters for optimized P- γ -TLN 12 following phase IVa for a period of twelve months.

Type of study	Parameters	Months						Change%
		0	1	3	6	9	12	
Long-term 5°C ± 3°C	Particle size	183.1	182.7	183.9	187.7	193.1	192.7	5.17
	PDI	0.507	0.517	0.522	0.547	0.527	0.585	3.79
	Entrapment of PTX	85.37	83.23	83.35	82.32	80.39	80.47	5.73
	Entrapment of γ -Fe ₂ O ₃	60.49	60.21	58.24	56.11	56.21	56.17	7.69
Accelerated 25°C ± 2°C	Particle size	183.1	182.1	188.2	191.6	-	-	4.43
	PDI	0.507	0.526	0.522	0.536	-	-	13.92
	Entrapment of PTX	85.37	84.23	81.39	81.03	-	-	5.08
	Entrapment of γ -Fe ₂ O ₃	60.49	60.02	57.21	54.29	-	-	11.82

References:

- [1] Vyas, D. M.; & Kadow J. F. Paclitaxel: a unique tubulin interacting anticancer agent. *Prog. Med. Chem.* **1995**, 32, 289-337.
- [2] Ma P.; & Mumper R. J. Paclitaxel nano-delivery systems: a comprehensive review. *J. Nanomed. Nanotech.* **2013**, 4, 1000164.
- [3] Richheimer S. L.; Tinnermeier D. M.; & Timmons D. W. High-performance liquid chromatographic assay of taxol. *Anal. Chem.* **1992**, 64, 2323-2326.
- [4] Xu Q.; Trissel L. A.; & Martinez J. F. Stability of paclitaxel in 5% dextrose injection or 0.9% sodium chloride injection at 4 22 or 32° C. *Am. J. Health-Syst. Pharm.* **1994**, 51, 3058-3060.
- [5] Gelderblom H.; Verweij J.; Nooter K. & Sparreboom A. Cremophor EL: the drawbacks and advantages of vehicle selection for drug formulation. *Eur. J. Cancer.* **2001**, 37, 1590-1598.
- [6] Chen Y.; Yang X.; Zhao L.; Almásy L.; Garamus V. M.; Willumeit R.; & Zou A. Preparation and characterization of a nanostructured lipid carrier for a poorly soluble drug. *Coll. Surf A: Physicochem. Eng. Asp.* **2014**, 455 36-43.
- [7] Adel S.; Fahmy R. H.; Elsayed I.; Mohamed M. I.; & Ibrahim R. R. Exploiting itraconazole-loaded nanomixed micelles in coated capsules as efficient colon-targeted delivery system for improved antifungal and potential anticancer efficacy. *Pharm. Dev. Tech.* **2023**, 28, 333-350.
- [8] Chiang J.-L.; & Yang Y.-W. Modulation of the anticancer activities of paclitaxel by Cremophor micelles. *International J. Pharm.* **2021**, 603, 120699.
- [9] Shakeel F. Criterion for excipients screening in the development of nanoemulsion formulation of three anti-inflammatory drugs. *Pharm. Dev. Tech.* **2010**, 15, 131-138.
- [10] Kaur P.; Mishra V.; Shunmugaperumal T.; Goyal A. K.; Ghosh G.; & Rath G. Inhalable spray dried lipidnanoparticles for the co-delivery of paclitaxel and doxorubicin in lung cancer. *J. Drug Del. Sci. Tech.* **2020**, 56, 101502.
- [11] Huang Z. R.; Hua S. C.; Yang Y. L.; & Fang J. Y. Development and evaluation of lipid nanoparticles for camptothecin delivery: a comparison of solid lipid nanoparticles nanostructured lipid carriers and lipid emulsion. *Acta. Pharmacol. Sin.* **2008**, 29, 1094-1102.
- [12] Gao Y.; Chen Y.; Ji X.; He X.; Yin Q.; Zhang Z.; Shi J.; & Li Y. Controlled intracellular release of doxorubicin in multidrug-resistant cancer cells by tuning the shell-pore sizes of mesoporous silica nanoparticles. *ACS Nano.* **2011**, 5, 9788-9798.

- [13] Souto E.; Almeida A.; & Müller R. Lipid nanoparticles (SLN® NLC®) for cutaneous drug delivery: structure protection and skin effects. *J. Biomed. Nanotech.* **2007**, 3, 317-331.
- [14] Vorum H.; Brodersen R.; Kragh-Hansen U.; & Pedersen A. O. Solubility of long-chain fatty acids in phosphate buffer at pH 7.4. *Biochim Biophys. Acta (BBA)-Lipids Lipid Metab.* **1992**, 1126, 135-142.
- [15] Earnden L.; Marangoni A. G.; Laredo T.; Stobbs J.; Marshall T.; & Pensini E. Decontamination of water co-polluted by copper toluene and tetrahydrofuran using lauric acid. *Sci. Rep.* **2022**, 12, 15832.