

Supplementary Informations

MgFeTi-LDH/g-C₃N₄ promote sodium persulfate activation through Fe³⁺/Fe²⁺ cycle for efficient tetracycline degradation under visible light irradiation

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1. Radical-capturing experiment

A radical-capturing experiment is an effective approach to explain the reaction mechanism. Radical capturing agents were added during photocatalytic testing. chloroform (TCM, 10 mmol L⁻¹), ethylenediaminetetraacetic acid disodium salt (EDTA-2Na, 10 mmol L⁻¹), and methanol (MeOH, 10 mmol L⁻¹) were added as scavengers for superoxide radicals ($\bullet\text{O}_2^-$), photo-generated holes (h^+), hydroxyl radicals ($\bullet\text{OH}$).

2. Text toxicity assessment

Toxicity assessment is one of the key factors in evaluating the safety of wastewater treatment processes, particularly due to the potential formation of highly toxic intermediates during degradation. In order to determine the toxicity of TCH and intermediates, the toxicity estimation software tool (TEST) was applied to assess ecotoxicity according to various mathematical models of QSAR from Environmental Protection Agency database in US. Here, six toxicity parameters such as acute toxicity of fathead minnow (LC₅₀-96 h), daphnia magna 50% lethal dose (LC₅₀-48 h), oral rats (LD₅₀), bioconcentration factor, developmental toxicity, and mutagenicity are employed to evaluate the toxicity of TCH and degradation intermediates.

Furthermore, this work used OD₆₀₀ value to evaluate toxicity of byproducts from TCH degradation. The degradation solution of 20 mg/L TCH was collected for bacterial culturing. *Escherichia coli* DH5 α , a representative model bacterium, was selected to investigate its growth under the presence of TCH degradation solution. This experiment aims to assess the detoxification capacity of the system in treating TCH-contaminated wastewater, particularly focusing on

evaluating the potential of the degradation products to support bacterial growth without causing toxicity. In a typical run, the strain in logarithmic growth stage was inoculated in LB medium for toxicity test. The OD₆₀₀ of *Escherichia coli* DH5a was measured to assess toxic inhibition after 24 h incubation at 25 ± 1 °C. All experiments were repeated three times.

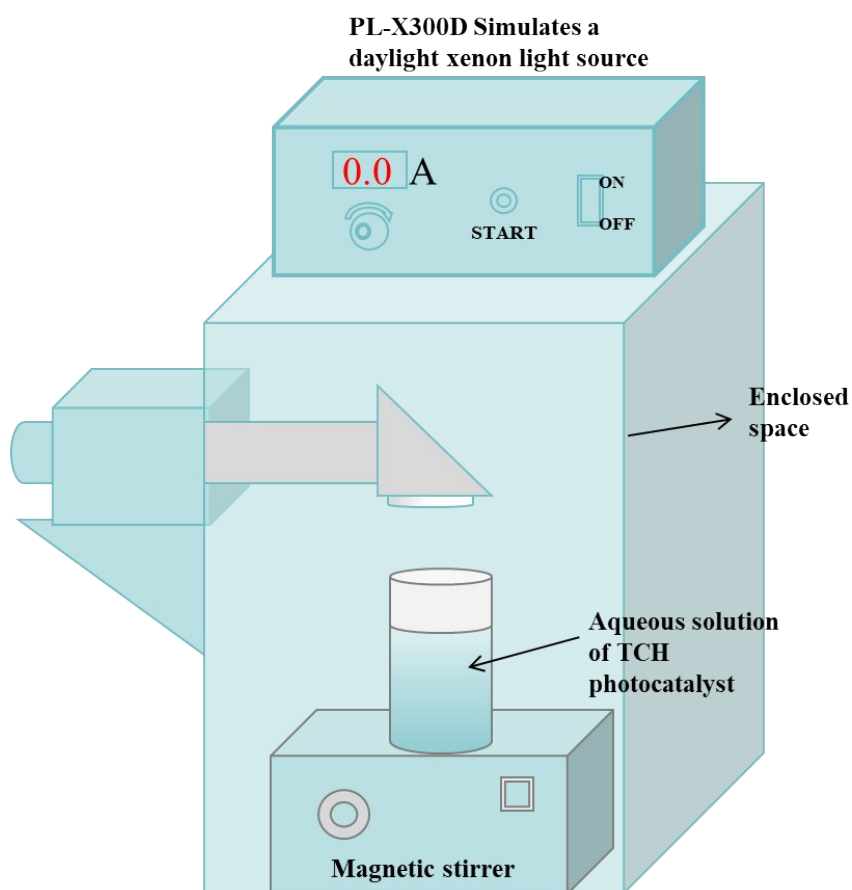


Fig. S1. Photocatalytic reaction device

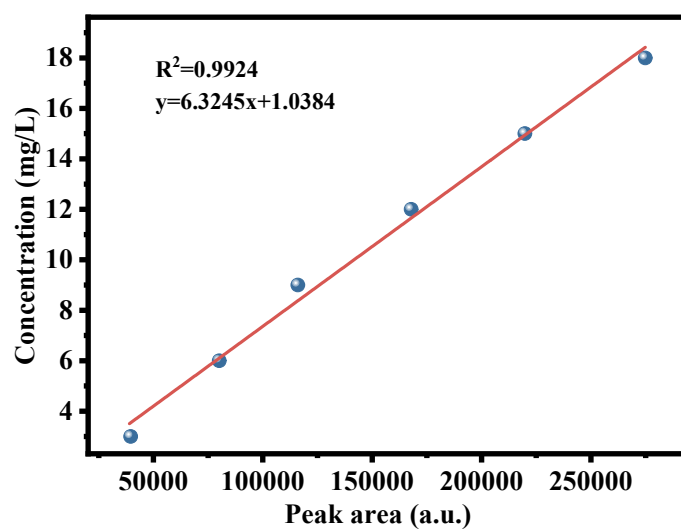


Fig. S2. Standard curve of TCH

Table S1 The specific surface areas, pore volumes, and pore sizes of catalyst

Sample	Specific surface area(m ² /g)	Pore volume(cm ³ /g)	Pore size (nm)
g-C ₃ N ₄	53.8	0.16	16.7
MgFeTi(3:1)-LDH	99.1	0.08	5.2
MgFeTi(3:1)-LDH/g-C ₃ N ₄	83.2	0.42	13.6

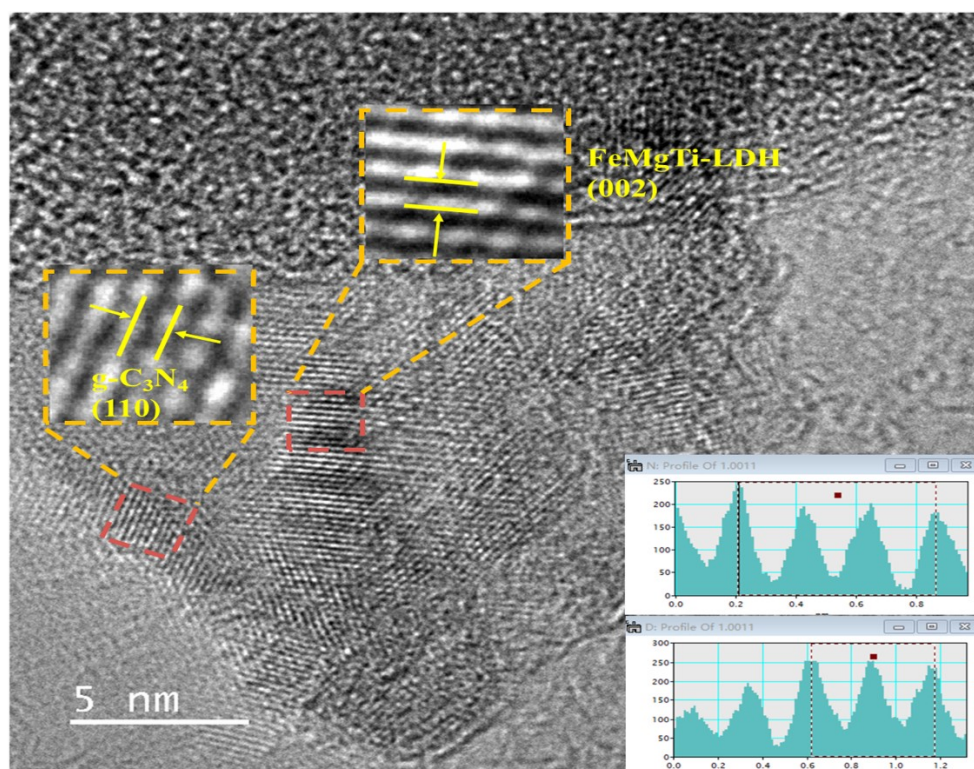


Fig. S3. the HRTEM image of MgFeTi (3:1)-LDH/g-C₃N₄

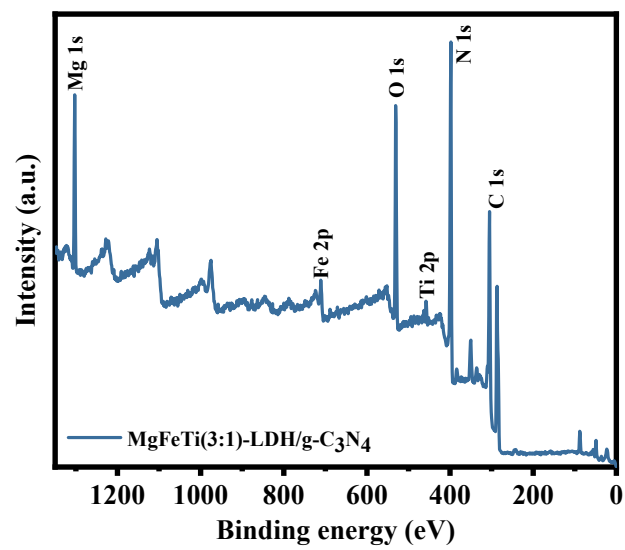


Fig. S4. XPS survey spectra of MgFeTi (3:1)-LDH/g-C₃N₄

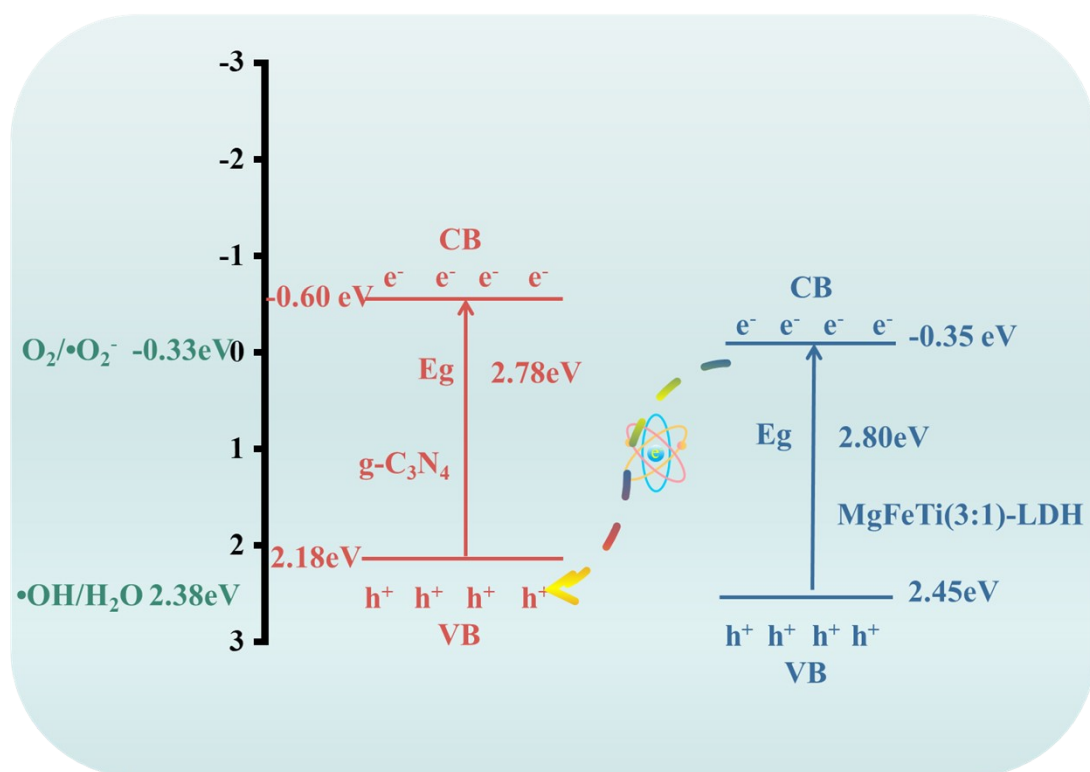


Fig. S5. CB and VB of the MgFeTi (3:1)-LDH/g-C₃N₄

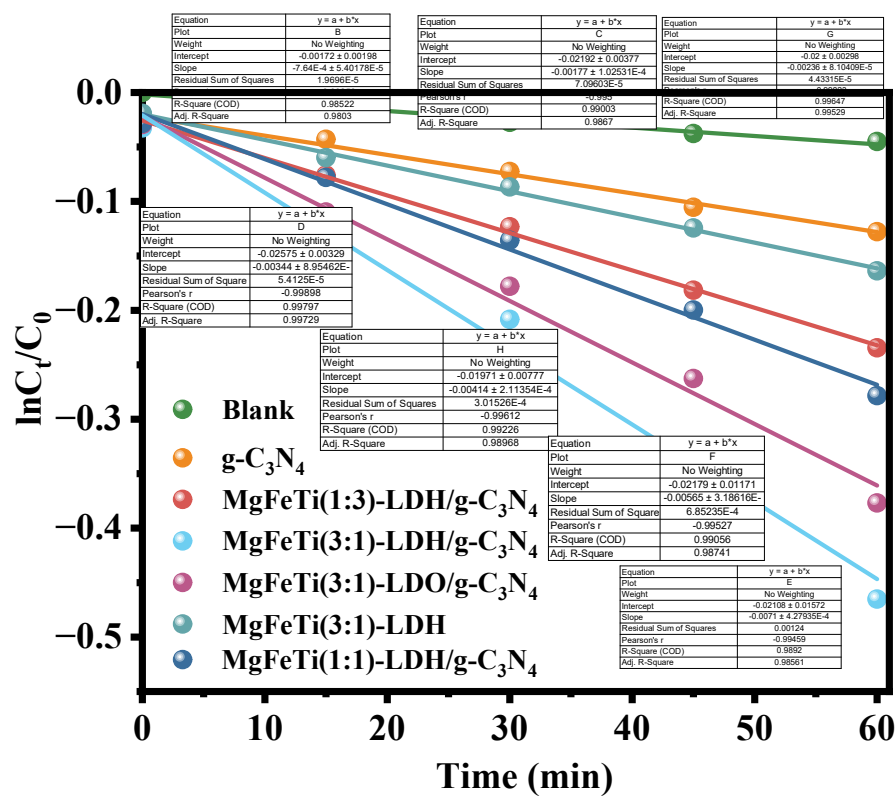


Fig. S6. kinetics of degradation reaction

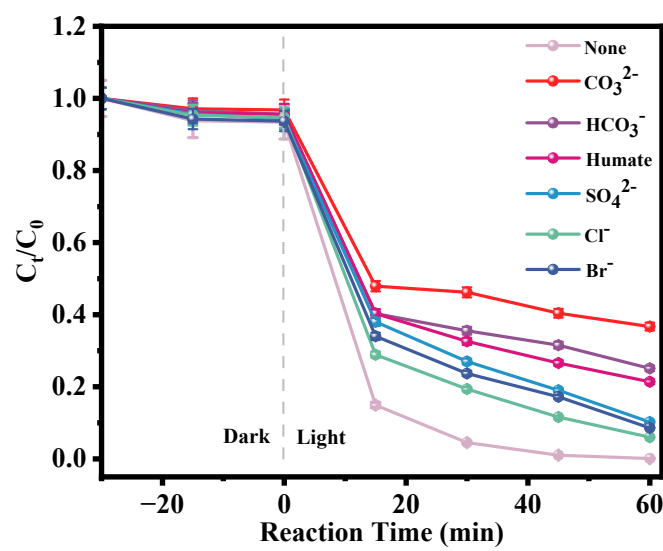


Fig. S7. The effect of various inorganic anions and humate on degradation efficiency

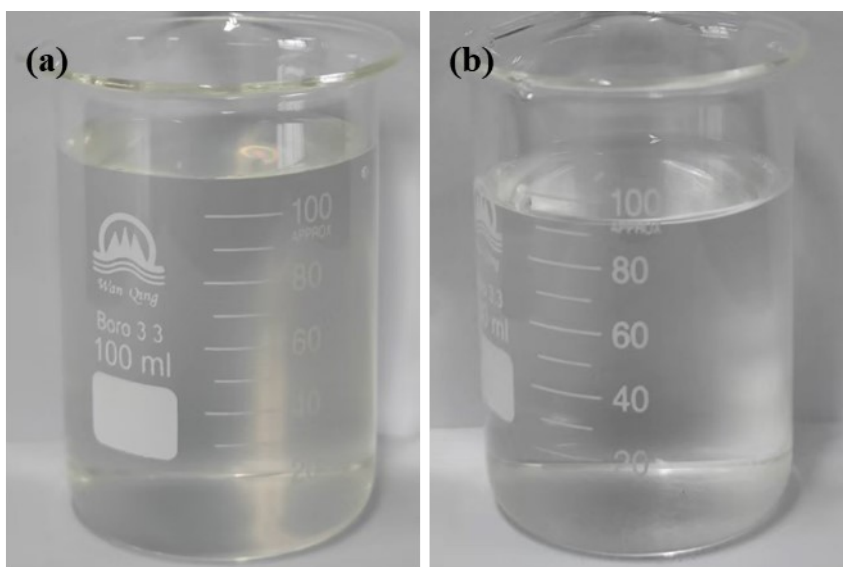


Fig.S8. The decolorization efficiency of MgFeTi-LDH/g-C₃N₄ composites for TCH in actual effluents (a) TCH in actual effluents, (b) Decolorized TCH solution

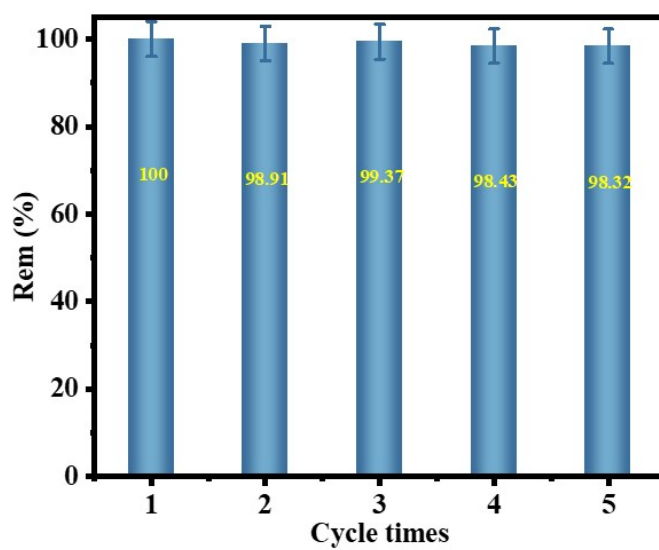


Fig. S9. Reusability performance of MgFeTi(3:1)-LDH/g-C₃N₄

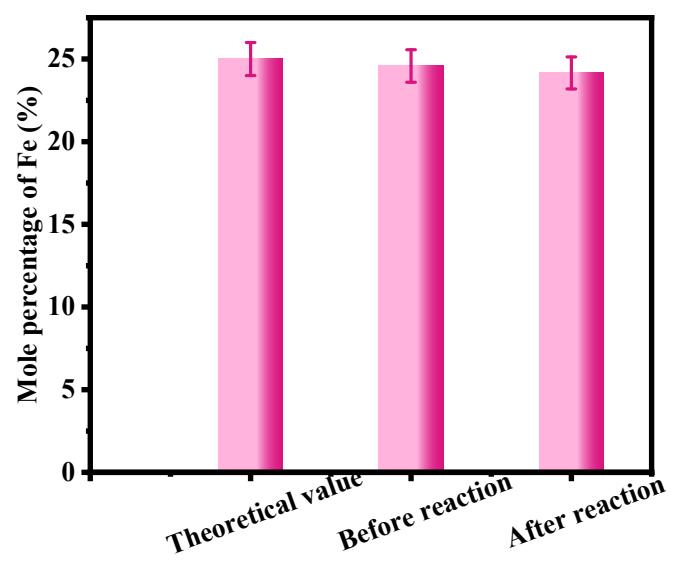


Fig. S10. The Fe mole percentage from ICP

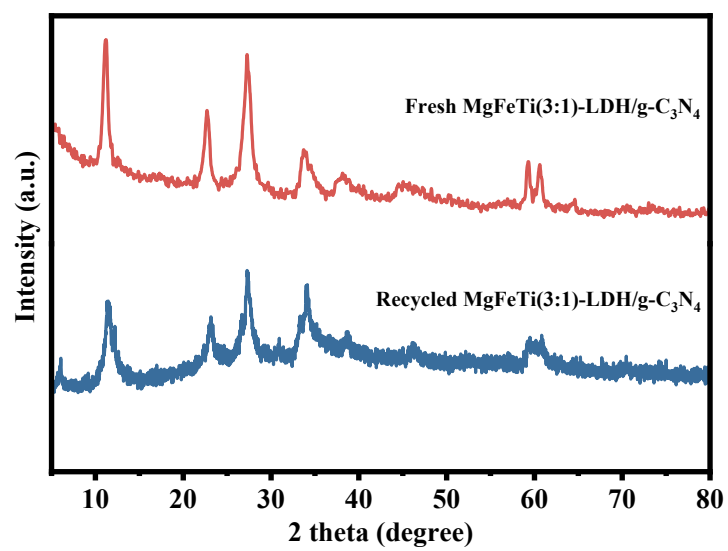


Fig. S11. The XRD patterns of recycled MgFeTi(3:1)-LDH/g-C₃N₄ and fresh MgFeTi(3:1)-LDH/g-C₃N₄

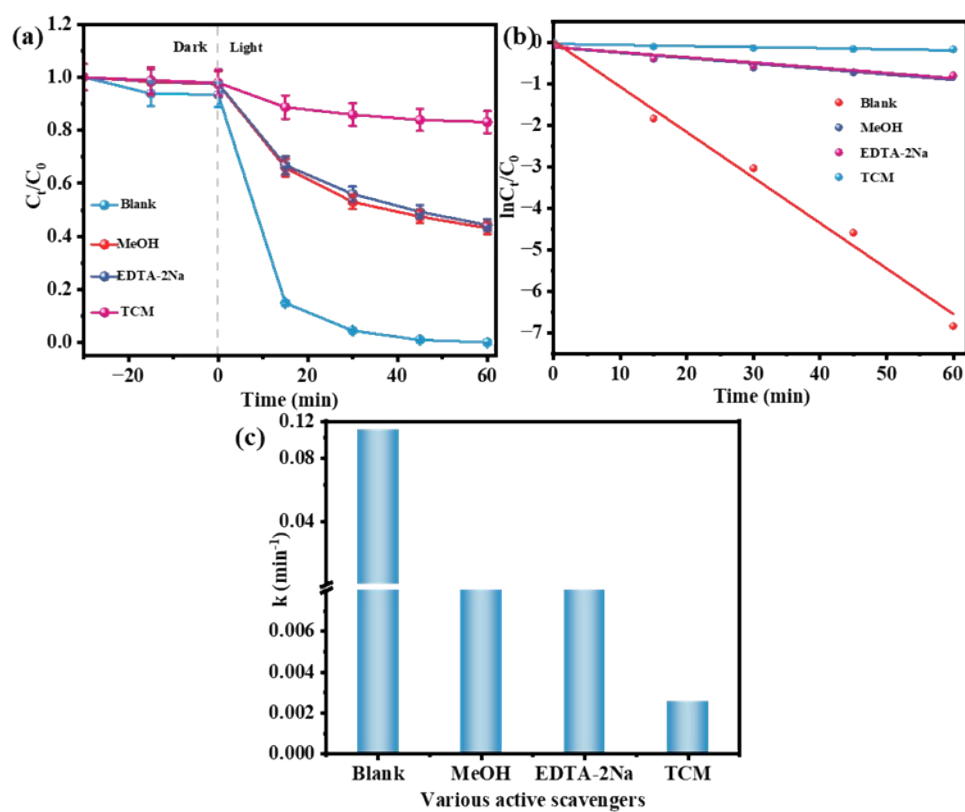


Fig. S12. The effects of various scavengers on (a) C_t/C_0 , (b) $\ln C_t/C_0$, (c) k

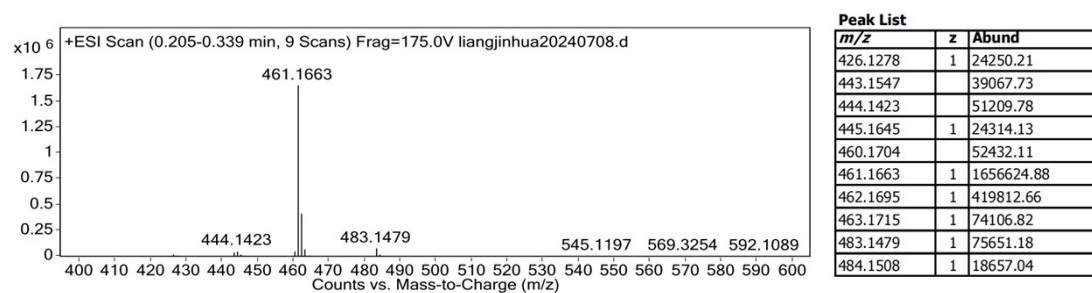


Fig.S13. LC-MS spectra of TCH photocatalytic degradation recorded by MgFeTi(3:1)-LDH/g-C₃N₄ at 0 min

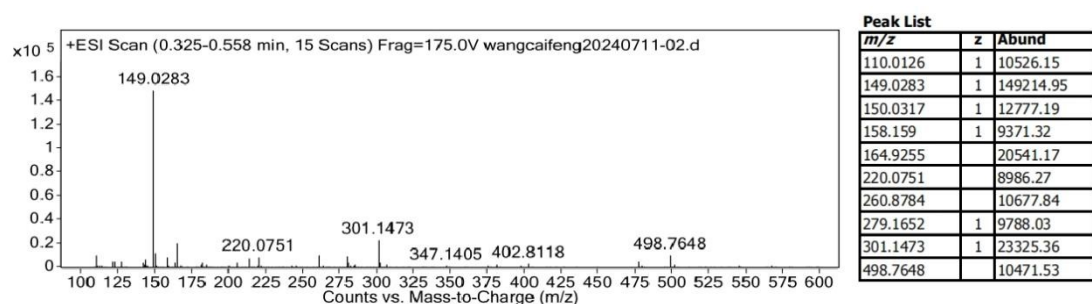


Fig. S14. LC-MS spectra of TCH photocatalytic degradation recorded by MgFeTi(3:1)-LDH/g-C₃N₄ at 30 min

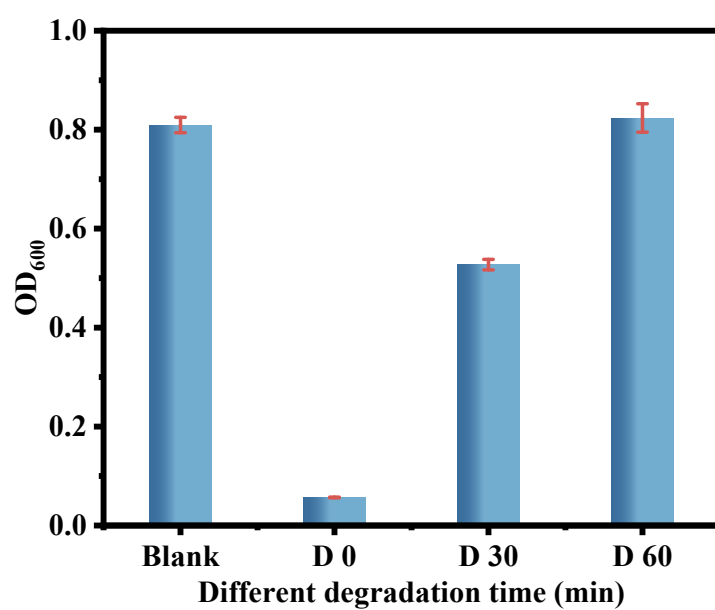


Fig. S15. Optical density (OD₆₀₀) values of degradation TCH at different time periods over MgFeTi (3:1)-LDH/g-C₃N₄