Supplementary Information (SI) for New Journal of Chemistry. This journal is © The Royal Society of Chemistry and the Centre National de la Recherche Scientifique 2025

Supporting Information

A novel internal standard ratio fluorescent probe for nitroreductase

detection in cells

Han Zeng, Haijie Wang, Mingchao Huang, Yuanyuan Wu, Yaping Wang, Yiyi Li, Shicong

Hou^{*}.

College of Science, China Agricultural University, Beijing 100193, P.R. China

Corresponding author: * E-mail: houshc@cau.edu.cn

Table of Contents

1. Fluorescent probes for NTR	S2
2. Supplementary Methods	S2
3. Supplementary Schemes	S3
4. Study on Spectral Properties	S5
5. HRMS, ¹ H-NMR and ¹³ C-NMR	S9
References	

Probe	ratio	$\lambda_{em} (nm)$	DL (ng/mL)	Response time (min)	Reference
T-TPE-NO ₂	NO	626	46.3	40	1
Pro-CN	NO	560	4.4	25	2
PC-NTR	NO	635	13.8	30	3
NORP	Yes	455	30	/	4
		520			
ETH1-NO ₂	NO	603	562	/	5
Ox-NTR	NO	683	90	/	6
CS-CN-NO	Yes	530	70	16	This Work
		700	/0	10	I IIIS VVOĽK

1. Fluorescent probes for NTR

2.Supplementary methods

Cell image

The cells were plated at 1×10^5 cells/mL suspension in a Glass bottomed petri dish and allowed to culture overnight with (normoxia) or without (hypoxia) CoCl₂(100µM). The cells were incubated with CS-CN-NO for 30 min before imaging and then washed with PBS three times. Fluorescent cell images were obtained by confocal imaging microscopy. Unless otherwise specified, the probe concentration is always 20 µM.

Instruments

UV–Vis absorption spectra were recorded using a PerkinElmer Lambda 650S UV/Vis spectrometer. Fluorescence spectra were recorded using a PerkinElmer LS55 fluorescence spectrometer. ¹H NMR and ¹³C NMR spectra were acquired using a Bruker AvanceAVII-500 MHz spectrometer. High-resolution mass spectra were acquired with a Waters Xevo G2-XS QTOF mass spectrometer. All live cell images were obtained using a Nikon A1 confocal microscope.

General procedure for fluorescence detection

Stock solutions of interference ions were prepared in ultra-filter deionized water with the concentration of 1.0×10^{-2} M and then diluted to a desired concentration. The concentrations of interfering ions and enzymes were 10 mM and 5 U/mL, respectively. Probes stock solution was prepared at the concentration of 1.0×10^{-3} M in DMSO and then diluted to 2.0×10^{-5} M for titration experiments. NTR was prepared at the concentration of 200 µg/mL in PBS. NADH was prepared at the concentration of 1.0×10^{-2} M in PBS and then diluted to 1.0×10^{-4} M for titration experiments. UV-vis and fluorescence titration experiments were operated in PBS/CH₃CN = 3:7 (10 mM pH 7.4), (λ_{ex} = 480 nm, excitation slit =15.0 nm, emission slit = 15.0 nm). Because the enzyme requires a milder and more stable environment in the organism, in order to better detect the activity of NTR, all experiments were carried out at 37°C.

Determination of detecting limits

The detecting limits (DL) were calculated according to Eq.

$$DL = 3.29\sigma/k$$

Where σ is the standard derivation of blank solution and k is the slope of calibration curve.

MTT assay for the cell cytotoxicity.

Cell cytotoxicity was evaluated by MTT assay. Cells were cultivated in a 96-well plate until 70% confluence, and incubated with different concentrations of CS-CN-NO(0-30 μ M) or CoCl₂ (0-100 μ M) for 24 h. Then 20 μ L 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyltetrazolium bromide (MTT, 5 mg/mL) was added for 4 h at 37°C. After MTT was washed, 110 μ L DMSO was added. Absorbance was measured at 570 nm with a multi-function microplate reader. All experiments were repeated 3 times, and the data were presented as the percentage of control cells.

3.Supplementary Schemes

Syntheses of CS-CN-NO



Scheme. S1. Reaction scheme for probe and fluorophore synthesis.

Compound H-CHO-Br, CS-CHO, CS-CHO-OH, CS-CN-OH, CS-FNT was synthesized by literature method⁷.

CS-CHO-NO: Compound CS-CHO-OH (301 mg, 0.66 mmol), $K_2CO_3(276 mg, 2mmol)$ and 4-nitrobenzyl bromide (173 mg, 0.80mmol) were dissolved in DMF (8 mL). After stirring overnight at 40 °C and dark environment. After the reaction was completed, it was extracted by DCM. After extraction, dried on Na₂SO₄ and solvent was removed by reducing pressure. The product was purified by silica gel column chromatography (DCM/MeOH = 250:1, v/v) to give the pure compound as a light yellow solid (106 mg, 27%). ¹H NMR (500 MHz, CDCl₃) δ 10.32 (s, 1H), 8.31 – 8.24 (m, 3H), 8.07 – 8.00 (m, 1H), 7.73 – 7.59 (m, 5H), 7.18 – 7.13 (m, 1H), 6.91 (d, *J* = 2.3 Hz, 1H), 6.83 – 6.66 (m, 4H), 5.23 (s, 2H), 2.70 (dd, *J* = 8.0, 5.0 Hz, 2H), 2.48 (t, *J* = 6.1 Hz, 2H), 1.78 (h, *J* = 5.8 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 187.85, 169.08, 159.90, 159.42, 152.96, 152.66, 151.87, 147.80, 146.41, 143.48, 135.28, 130.31, 130.10, 129.51, 128.75, 127.60, 126.60, 125.33, 123.99, 123.80, 119.80, 113.91, 113.84, 112.59, 112.05, 111.55, 110.44, 102.10, 82.17, 68.98, 30.41, 21.50, 20.22.

CS-CN-NO: Compound CS-CHO-NO (80 mg, 0.14 mmol), Acetic acid (0.5 mL), piperidine (1mL) and Compound CS-FNT (44 mg, 0.21mmol) were dissolved in toluene (40 mL). After stirring overnight at 80 °C and dark environment. After the

reaction was completed, removed solvent by reducing pressure. The product was purified by silica gel column chromatography (DCM/MeOH = 250:1, v/v) to give the pure compound as a purple solid (11 mg, 10%). ¹H NMR (500 MHz, CDCl3) δ 8.83 (d, J = 8.2 Hz, 1H), 8.21 (d, J = 8.5 Hz, 2H), 8.04 – 7.95 (m, 2H), 7.67 – 7.50 (m, 5H), 7.45 (d, J = 8.4 Hz, 1H), 7.34 (t, J = 7.6 Hz, 1H), 7.10 (d, J = 7.5 Hz, 1H), 7.00 (s, 1H), 6.84 (s, 1H), 6.77 – 6.64 (m, 4H), 6.57 (d, J = 8.7 Hz, 1H), 6.10 (d, J = 15.4 Hz, 1H), 5.16 (s, 2H), 2.62 (t, J = 5.9 Hz, 2H), 2.46 (t, J = 5.7 Hz, 2H), 1.83 (p, J = 2.8 Hz, 2H). ¹³C NMR (126 MHz, Pyr) δ 175.82, 169.20, 160.38, 159.24, 153.70, 152.98, 152.48, 152.35, 151.49, 147.70, 147.58, 146.06, 144.29, 139.38, 139.19, 134.39, 133.38, 131.79, 131.12, 130.27, 129.57, 128.46, 127.97, 126.87, 125.55, 125.28, 125.19, 124.84, 123.85,119.35, 118.73, 117.80, 116.45, 115.34, 112.36, 112.26, 111.63, 111.31, 105.94, 102.43, 82.37, 69.05, 64.40, 30.21, 24.52, 20.50.HRMS Calc for C₄₈H₂₉N₃O₈ [M+H]⁺ Calcd for 776.2028, found 776.2032.



Scheme. S2. The proposed recognition mechanism of CS-CN-NO toward NTR.

4. Study on Spectral Properties



Fig. S1. UV-vis absorbance of CS-CN-NO(20 $\mu M)$, CS-CN-OH (20 $\mu M)$ and CS-CN-NO (20 $\mu M)$ with NADH (100 $\mu M)$ and NTR (5 $\mu g/mL).$



Fig. S2. Fluorescent intensity of CS-CN-NO (20 μ M), CS-CN-OH (20 μ M) and CS-CN-NO (20 μ M)with NADH (100 μ M) and NTR (5 μ g/mL).



Fig. S3. MTT assay for the survival rate of HeLa cells treated with various concentrations of $CoCl_2$ (from 0 to 100 μ M) for 24 h. Error bars represent the standard deviation (n = 3).



Fig. S4. MTT assay for the survival rate of HeLa cells treated with various concentrations of CS-CN-NO (from 0 to 30 μ M) for 24 h. Error bars represent the standard deviation (n = 3).



5. HRMS,¹H-NMR and ¹³C-NMR

Fig. S5. ¹H-NMR spectrum of CS-CHO-NO in CDCl₃.



Fig. S6. ¹H NMR spectrum of CS-CN-NO in CDCl₃.







Fig. S8. ¹³C NMR spectrum of CS-CN-NO in pyridine-d5.



Fig. S9. HRMS spectrum of CS-CN-NO.



Fig. S10. HRMS spectrum of CS-CN-OH.



Reference

- 1. Y. T. Bao, H. B. Mao, K. W. Lei, J. B. Hu and J. Huang, *Talanta*, 2025, 285.
- 2. Y. Ji, X. Z. Zou, D. G. Chen, S. R. Sun and S. Z. Pu, Dyes and Pigments, 2024, 226.
- 3. L. Wen, M. Q. Shao, Y. H. Li, Y. J. Zhang, C. Peng, H. Yu and K. Zhang, *Talanta*, 2024, 266.
- 4. Y. M. Xu, B. Hu, Y. J. Cui, L. Li, F. Nian, Z. X. Zhang and W. T. Wang, *Chemical Communications*, 2023, **60**, 83-86.
- B. Y. Zhang, H. Chen, L. Shi, R. R. Guo, Y. Wang, Y. H. Zheng, R. Y. Bai, Y. X. Gao, B. Liu and X. F. Zhang, *Acs Sensors*, 2024, 9, 4560-4567.
- S. Q. Zhang, M. Ma, C. Zhao, J. K. Li, L. L. Xu, Z. H. Zhang, Q. P. Diao, P. Y. Ma and D. Q. Song, *Biosensors & Bioelectronics*, 2024, 261.
- L. M. Shi, C. X. Yan, Z. Q. Guo, W. J. Chi, J. L. Wei, W. M. Liu, X. G. Liu, H. Tian and W. H. Zhu, *Nat. Commun.*, 2020, 11, 11.