

Supporting Information

Self-assemblies of cell-penetrating peptides and ferrocifens: design and biological evaluation of an innovative platform for lung cancer treatment

Léna Guyon^{a1}, Abdallah Ladaycia^{a1}, Agnese Bosio^a, Laurent Lemaire^{a,b}, Florence Franconi^{a,b}, Bénédicte Lelièvre^c, Nolwenn Lautram^a, Pascal Pigeon^{d,e}, Gérard Jaouen^{d,e}, Catherine Passirani^a, Elise Lepeltier^{*a,f}

^a Univ Angers, Inserm, CNRS, MINT, SFR ICAT, F-49000 Angers, France

^b Univ Angers, Univ Rennes, INRAE, Inserm, CNRS, PRISM, Biogenouest, F-49000 Angers, Rennes, France
France

^c Centre régional de pharmacovigilance, Laboratoire de pharmacologie-toxicologie, CHU Angers, 4 rue Larrey, F-49100 Angers, France

^d CNRS, Institut Parisien de Chimie Moléculaire (IPCM), Sorbonne Université, 4 Place Jussieu, 75005 Paris, France

^e PSL, Chimie ParisTech, 11 Rue Pierre et Marie Curie, 75005 Paris, France

^f Institut Universitaire de France (IUF)

*E-mail: elise.lepeltier@univ-angers.fr

¹: authors contributed equally to this work

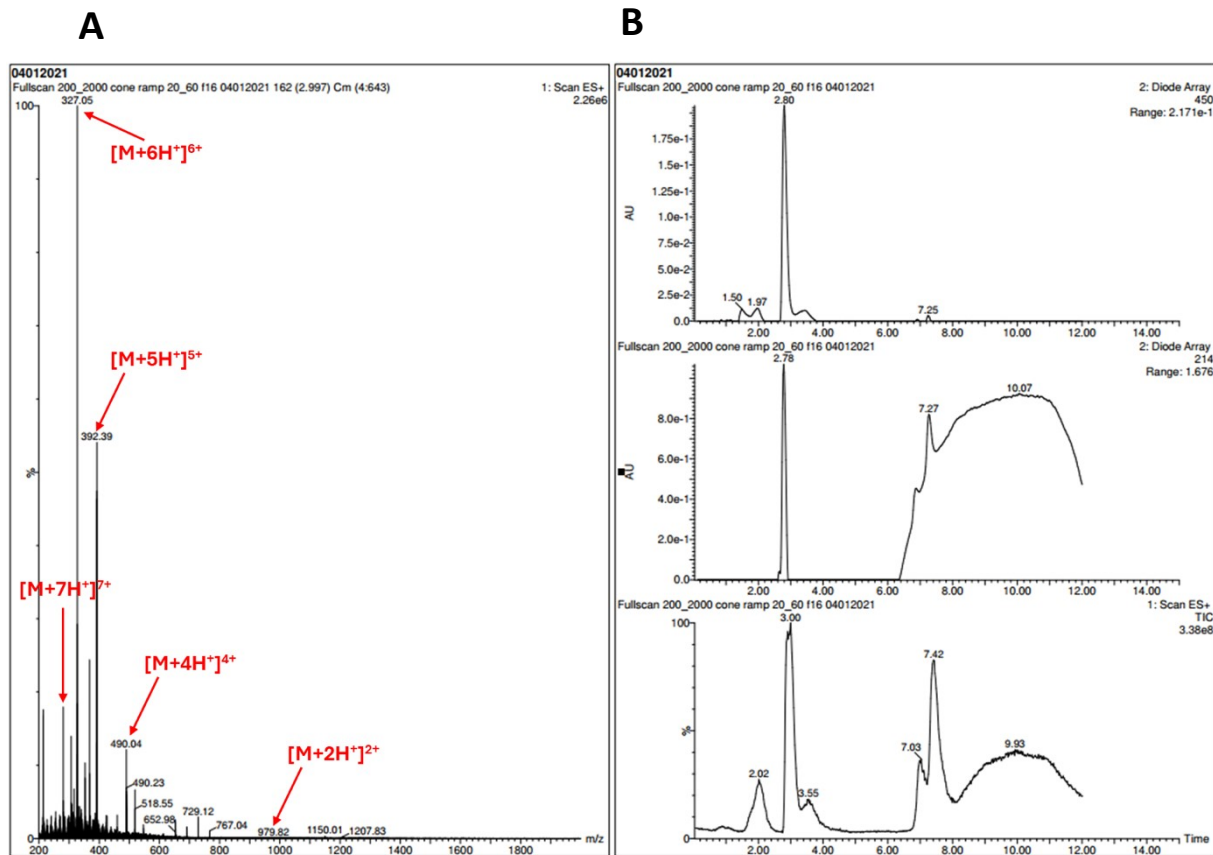


Figure S1. Mass spectrometry analysis of Arg₉-P819. (A) Mass spectra of the purified Arg₉-P819 showing the different pics corresponding to m/z of the pure molecule. (B) Chromatogram of the pure product detected at 214 nm (middle part) for the peptide part and at 450 nm for the ferrocifen part (top part). At the bottom, the Total Ion Chromatogram (TIC) of the conjugate. The baseline shape on the middle and bottom part of the figure are due the chromatography analysis method: a gradient of water and acetonitrile mixture.

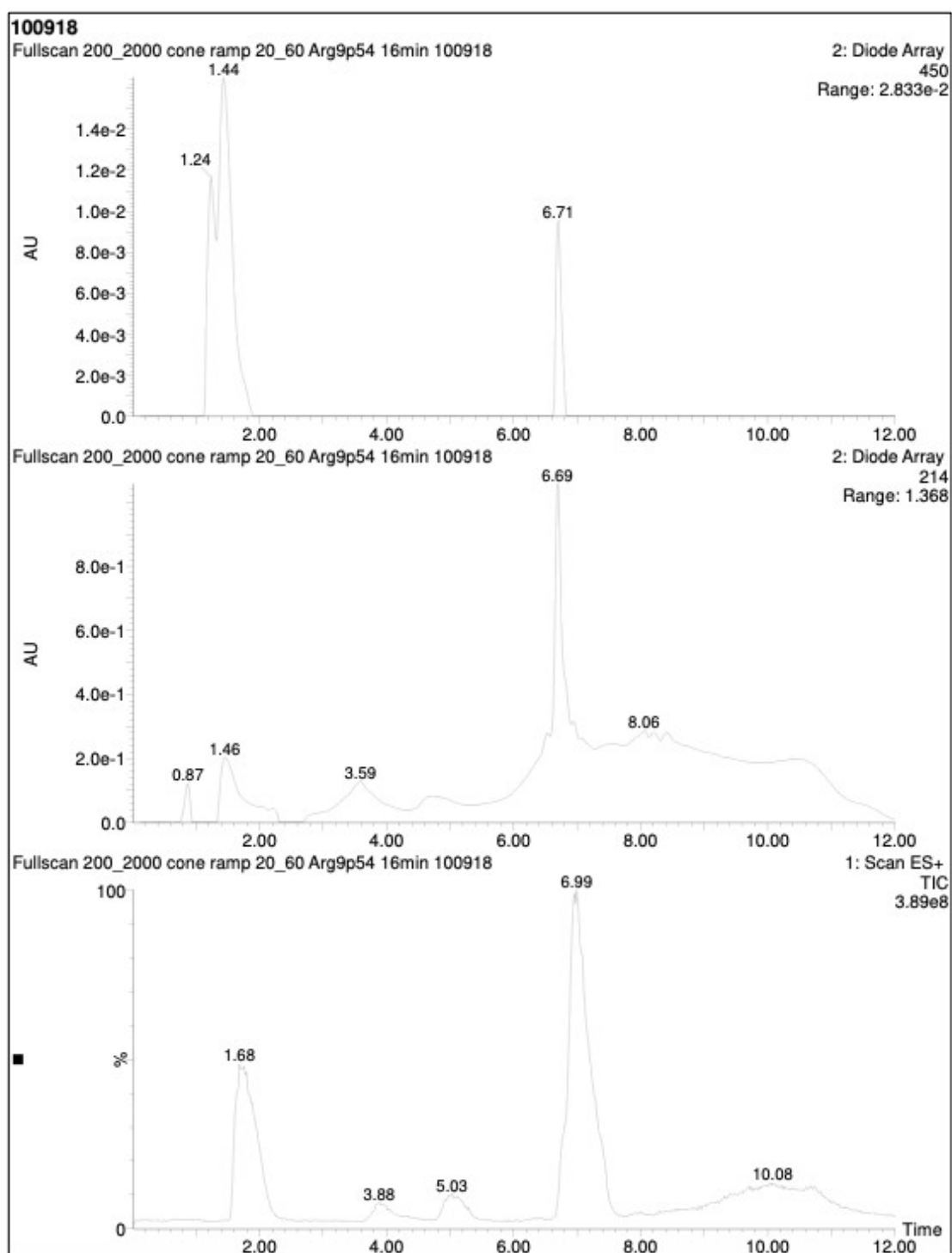


Figure S2. Chromatograms obtained at 214 (middle part) and 450 nm (top part) of the purified conjugate Arg₉-P54: the retention time of the compound is at 6.7 min for the UV detector and at 6.99 min for the MS detector, the last detector being after the UV detector in the channel.

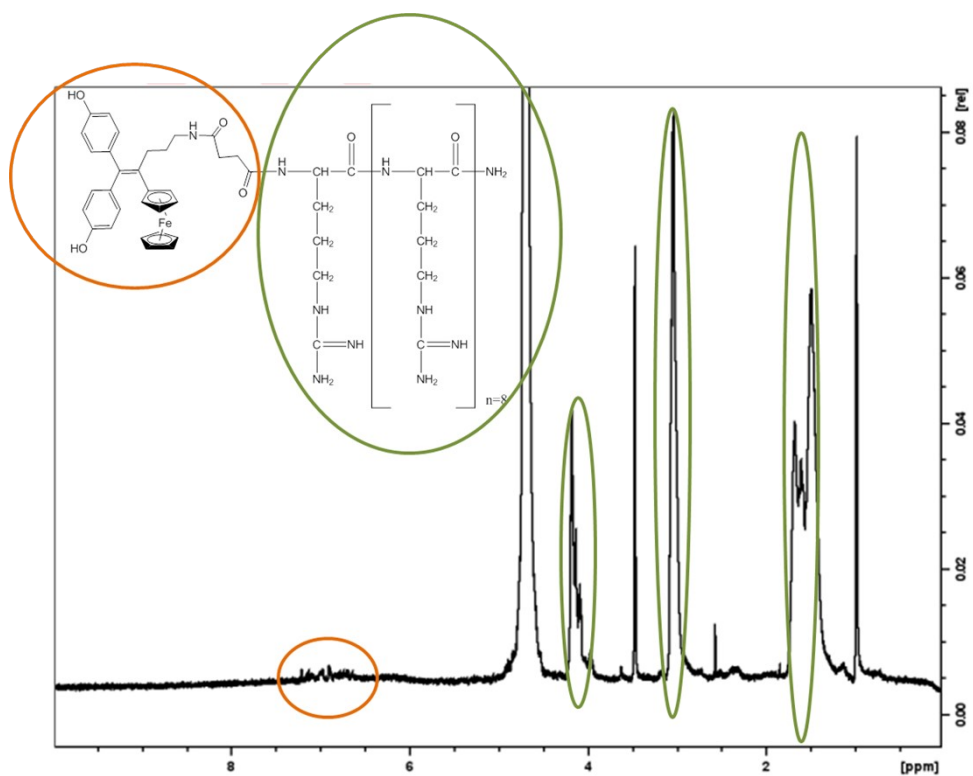


Figure S3. ^1H NMR spectrum of Arg₉-P819 in deuterated DMSO.

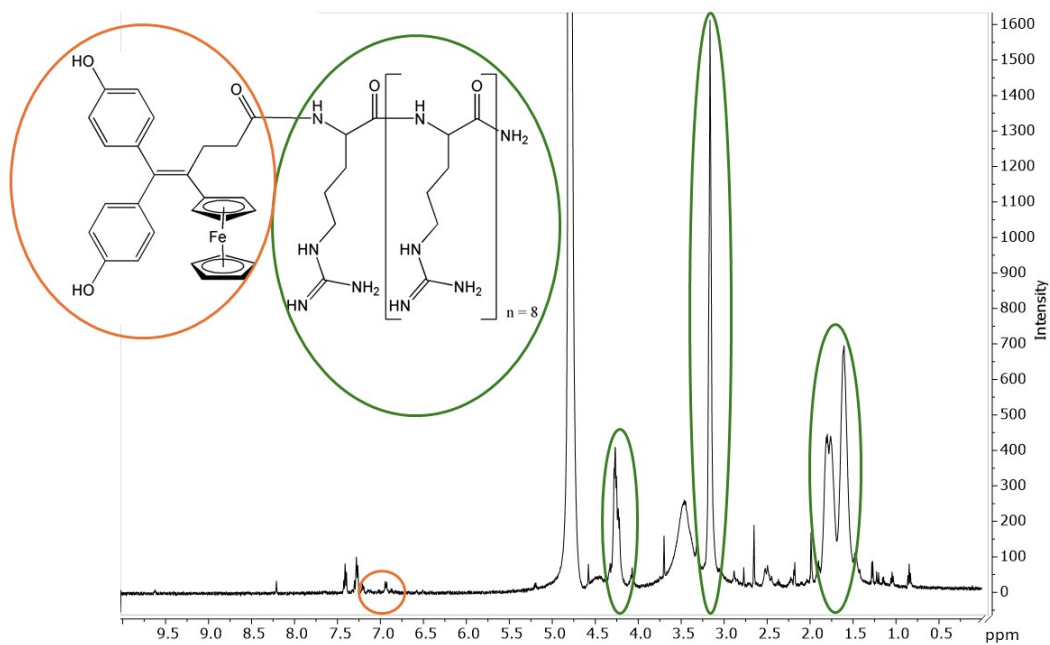


Figure S4. ^1H NMR spectrum of Arg₉-P54 in deuterated DMSO.

Supplementary Section S5: ^1H NMR analysis of PEG-P54

^1H NMR (400 MHz, DMSO- d_6) : δ 2.30 (t, 2 H, CH_2 of P54), 2.73 (t, 2 H, CH_2 of P54), 3.51 (s, 185H, CH_2 of PEG), 3.83 (t, 2 H, C_5H_4 of P54), 4.09 (t, 2 H, C_5H_4 of P54), 4.12 (s, 5H, Cp of P54), 6.61 (d, 2H, C_6H_4 of P54), 6.71 (d, 2H, C_6H_4 of P54), 6.77 (d, 2H, C_6H_4 of P54), 6.97 (d, 2H, C_6H_4 of P54), 9.28 (s, 1H, OH), 9.34 (s, 1H, OH), 12.01 (s broad, 1H, OH).

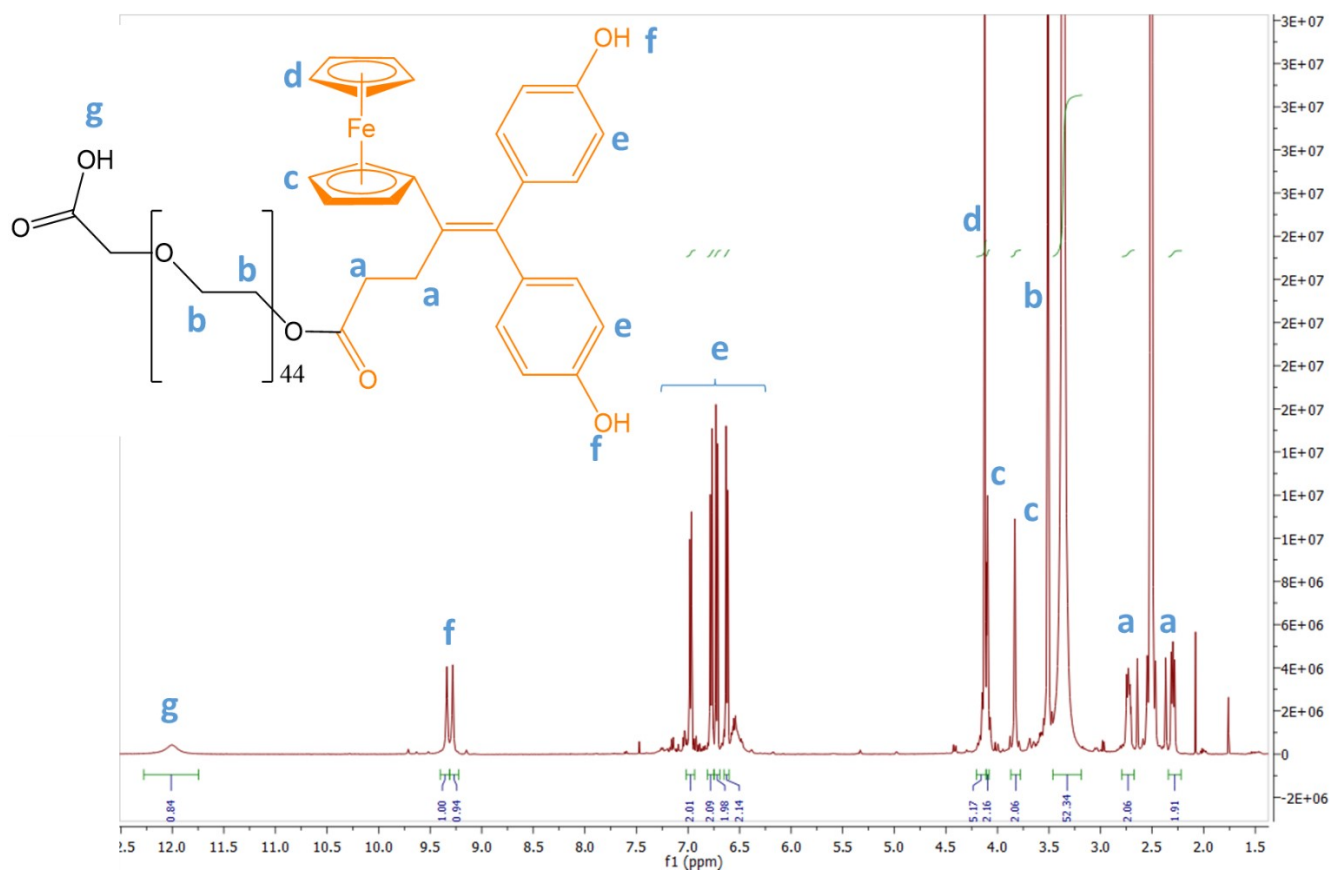
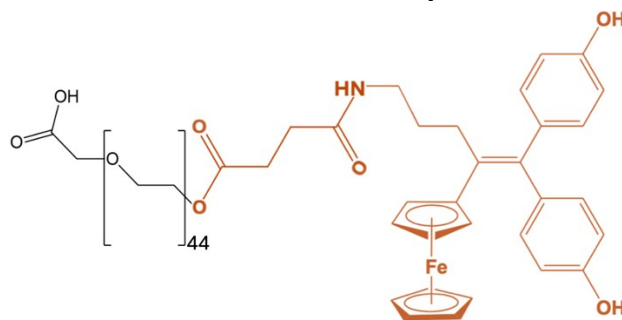
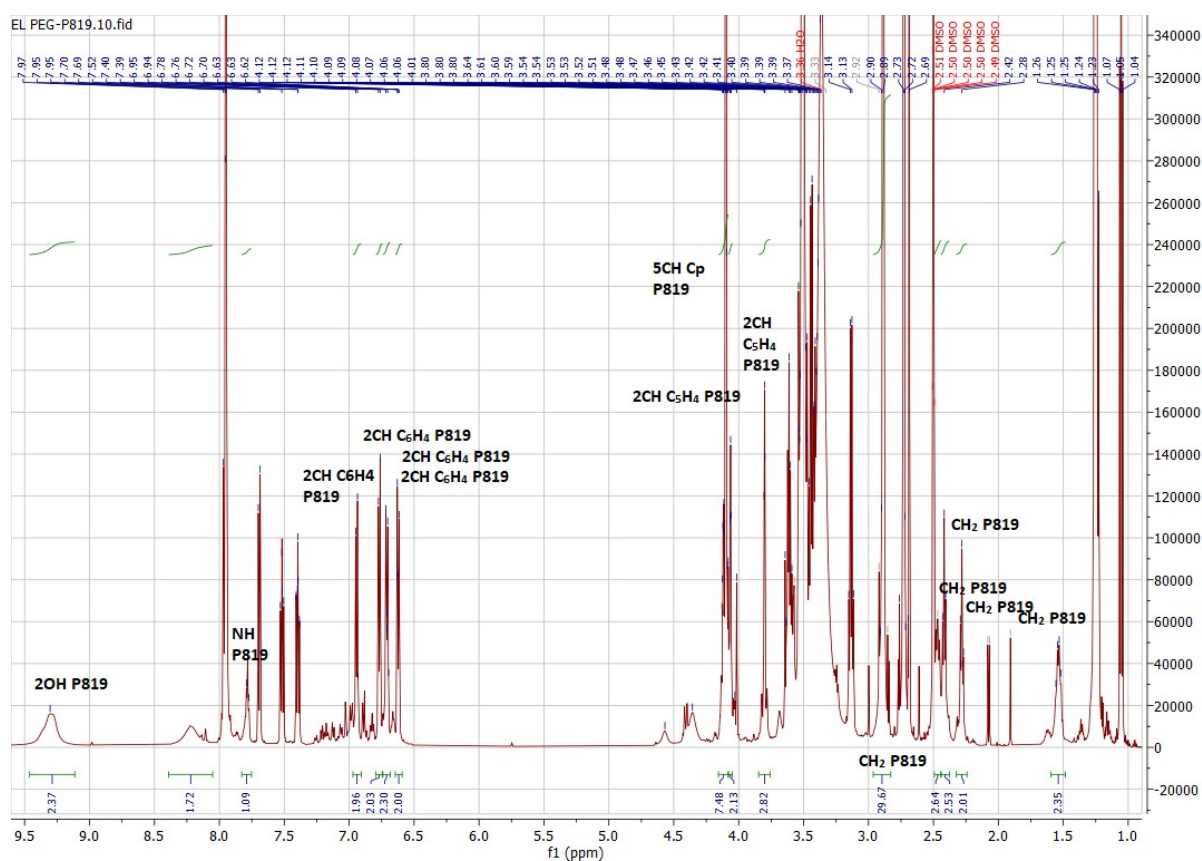


Figure S5. ^1H NMR spectrum of PEG-P54 in deuterated DMSO.

Supplementary Section S6: ^1H NMR and ^{13}C NMR analysis of PEG-P819



^1H NMR (DMSO- d_6): δ 1.48-1.59 (m, 2H, CH_2 P819), 2.28 (t, $J = 7.1$ Hz 2H, CH_2 P819), 2.42 (t, $J = 7.1$ Hz, 2H, CH_2 P819), 2.43-2.50 (m, 2H, $\text{CH}_2\text{-C=C}$ P819), 2.85 (d, $J = 7.1$ Hz, 2H, CH_2N P819), 3.5 (s, CH_2 of PEG), 4.06 (t, $J = 2.0$ Hz, 2H, C_5H_4), 4.09 (t, $J = 2.0$ Hz, 2H, C_5H_4), 4.10 (s, 5H, Cp), 6.62 (d, $J = 8.5$ Hz, 2H, C_6H_4), 6.71 (d, $J = 8.5$ Hz, 2H, C_6H_4), 6.77 (d, $J = 8.5$ Hz, 2H, C_6H_4), 6.94 (d, $J = 8.5$ Hz, 2H, C_6H_4), 7.79 (q, $J = 6.0$ Hz, 1H, NH).



^{13}C NMR (DMSO- d_6): δ 29.7 (CH_2 P819), 30.5 (CH_2 P819), 30.9 (CH_2 P819), 32.2 (CH_2 P819), 39.2 (CH_2 P819), 69.2 (2CH C_5H_4), 69.46 (5CH Cp), 69.53 (2CH C_5H_4), 70.0 (2 CH_2 of PEG), 87.1 (C C_5H_4), 115.5 (2x2CH C_6H_4), 130.4 (2CH C_6H_4), 130.8 (2CH C_6H_4), 133.7 (C), 135.7 (C), 136.1 (C), 138.5 (C), 156.0 (C-OH), 156.1 (C-OH), 171.1 (CO P819), 174.3 (CO P819), 177.9 (COO of the PEG-P819).

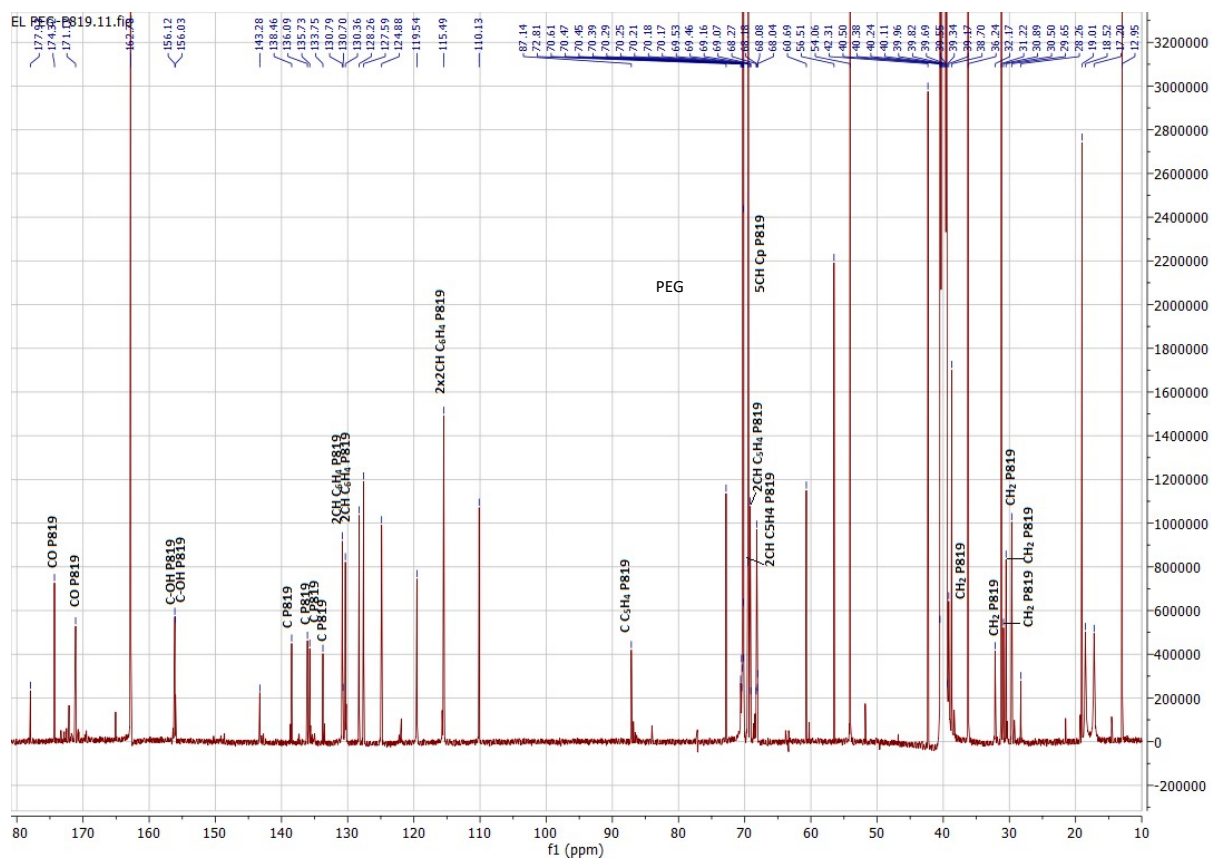


Figure S6. ^1H NMR spectrum of PEG-P819 on the top and ^{13}C NMR spectrum of PEG-P819 in DMSO- d_6 below.

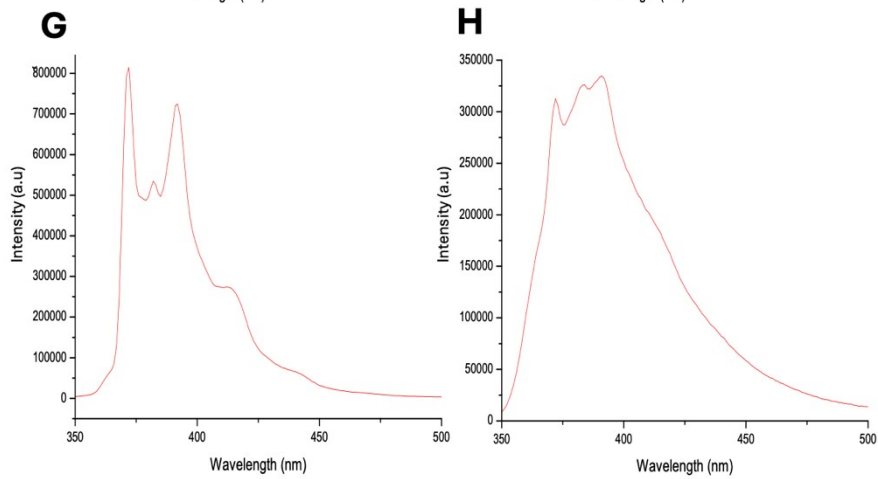
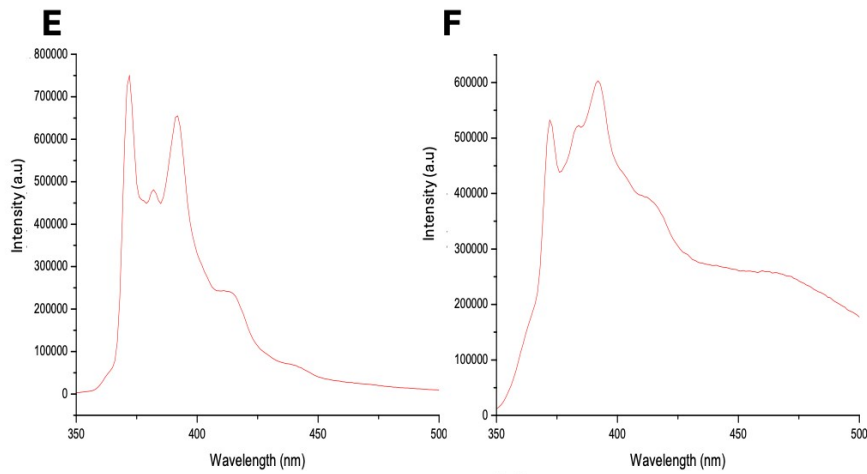
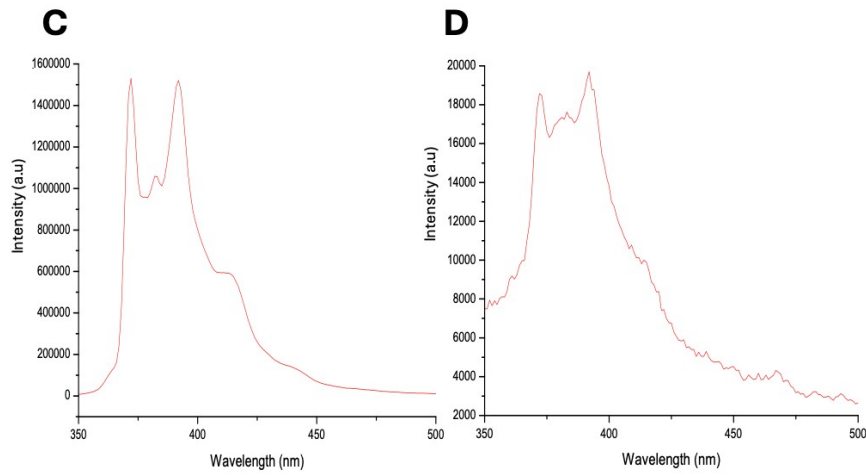
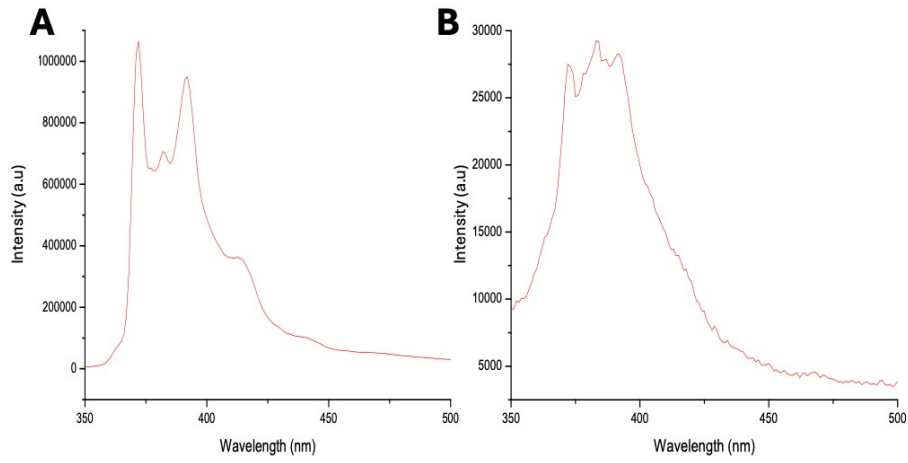


Figure S7. Using the pyrene method, representative fluorescent spectra for calculating the CAC: P54 at 10 μM (A) and at 3.2 mM (B), PEG-P54 at 50 μM (C) and at 600 μM (D), Arg₉-P54 at 6 μM (E) and at 1 mM (F) and Arg₉-P819 at 1 μM (G) and at 1.8 mM (H).

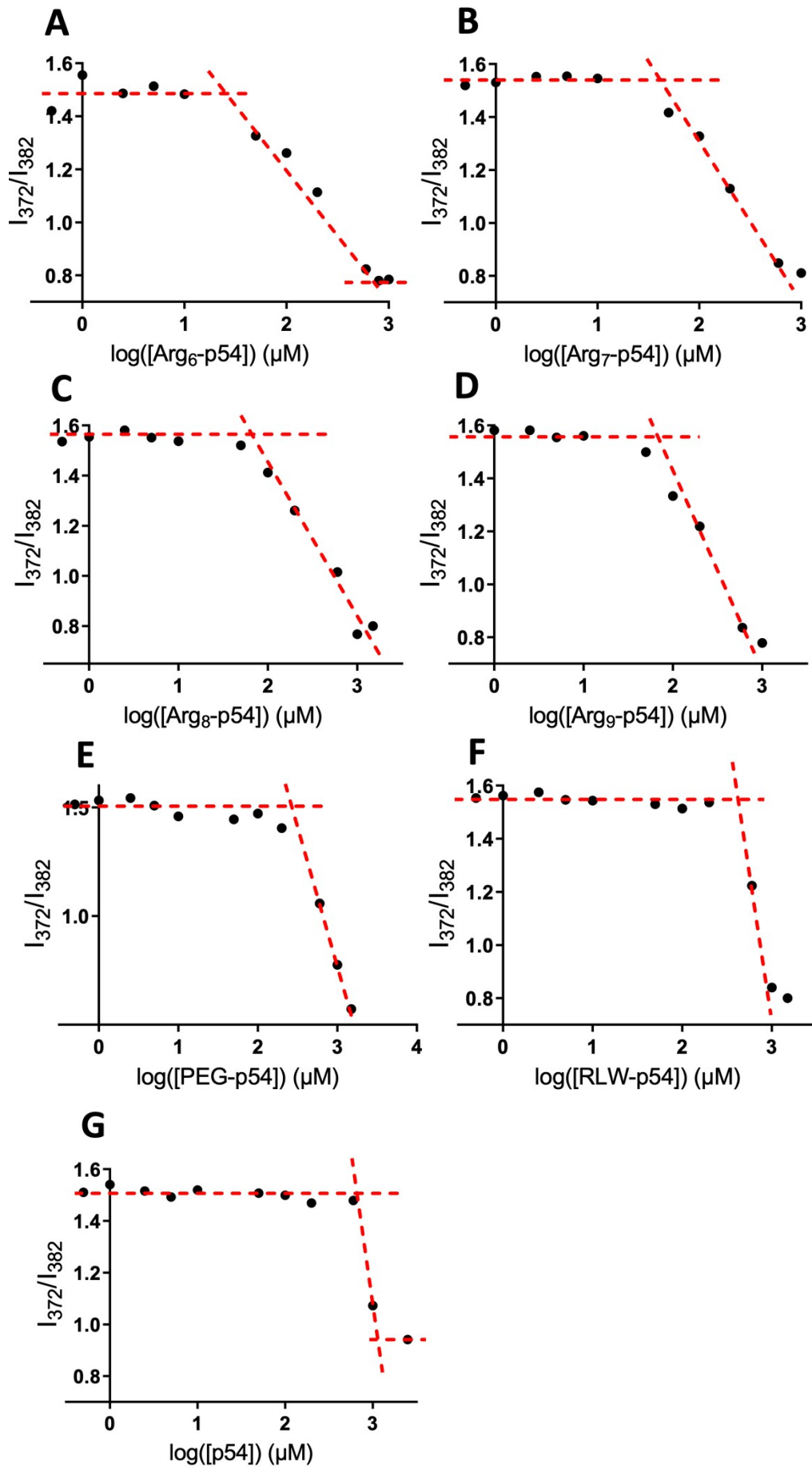


Figure S8. Quotient of vibrational band intensities (I_{11}/I_{13}) as a function of $\log [\text{Arg}_6\text{-P54}]$ (A), $\log [\text{Arg}_7\text{-P54}]$ (B), $\log [\text{Arg}_8\text{-P54}]$ (C), $\log [\text{Arg}_9\text{-P54}]$ (D), $\log [\text{PEG-p54}]$ (E), $\log [\text{RLW-P54}]$ (F) and $[\text{P54}]$ (G).

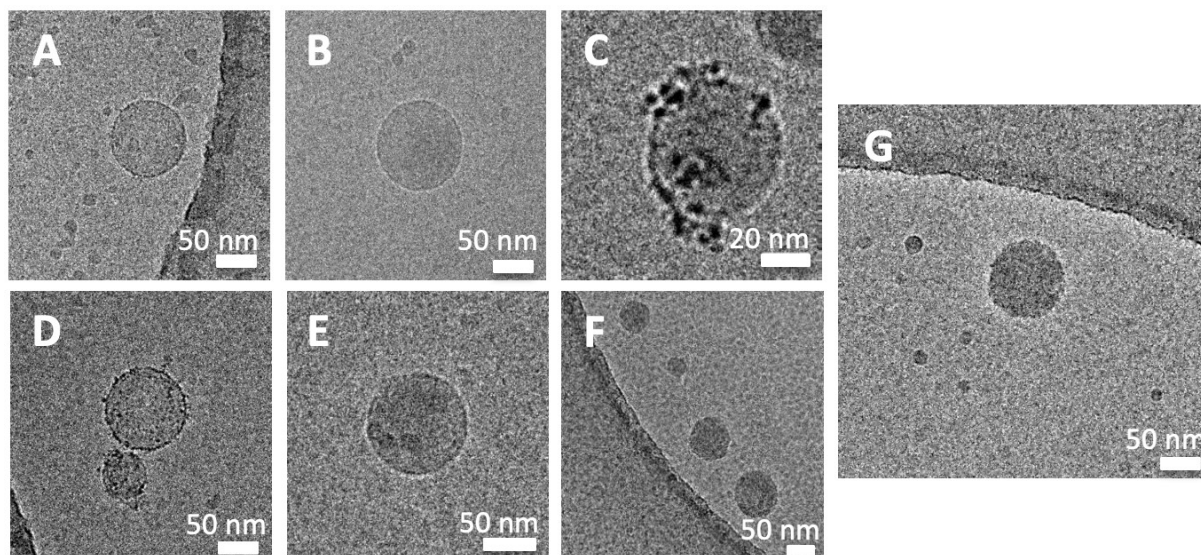


Figure S9. Representative Cryo-TEM images of self-assemblies formed from $\text{Arg}_6\text{-P54}$ (A)¹, $\text{Arg}_7\text{-P54}$ (B)¹, $\text{Arg}_8\text{-P54}$ (C)¹, $\text{Arg}_9\text{-P54}$ (D)¹, RLW-P54 (E)¹, PEG-P54 (F) and $\text{Arg}_9\text{-P54/PEG-P54}$ (G).

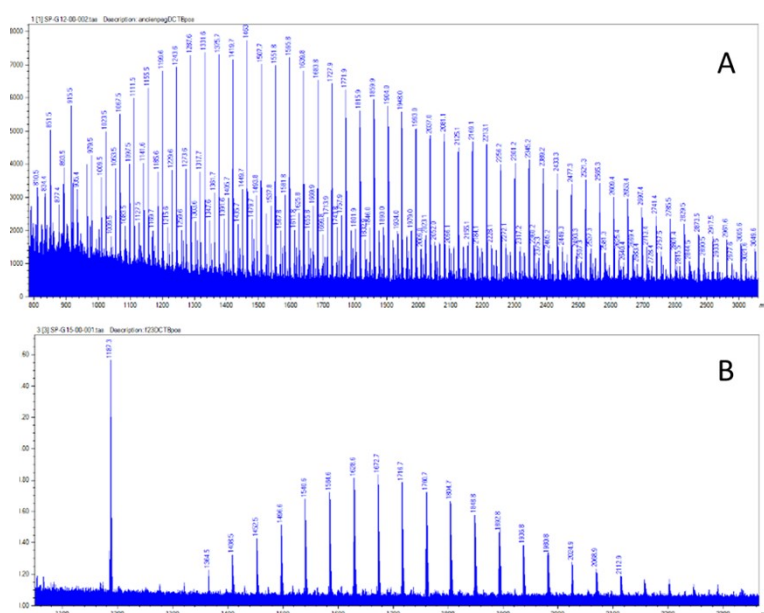


Figure S10. MALDI spectrum of free PEG (A), and of PEG-P819 (B), where a switch of the mass spectrum is observed and corresponds to the addition of P819 ($M_{\text{P819}} = 569.47 \text{ g.mol}^{-1}$).

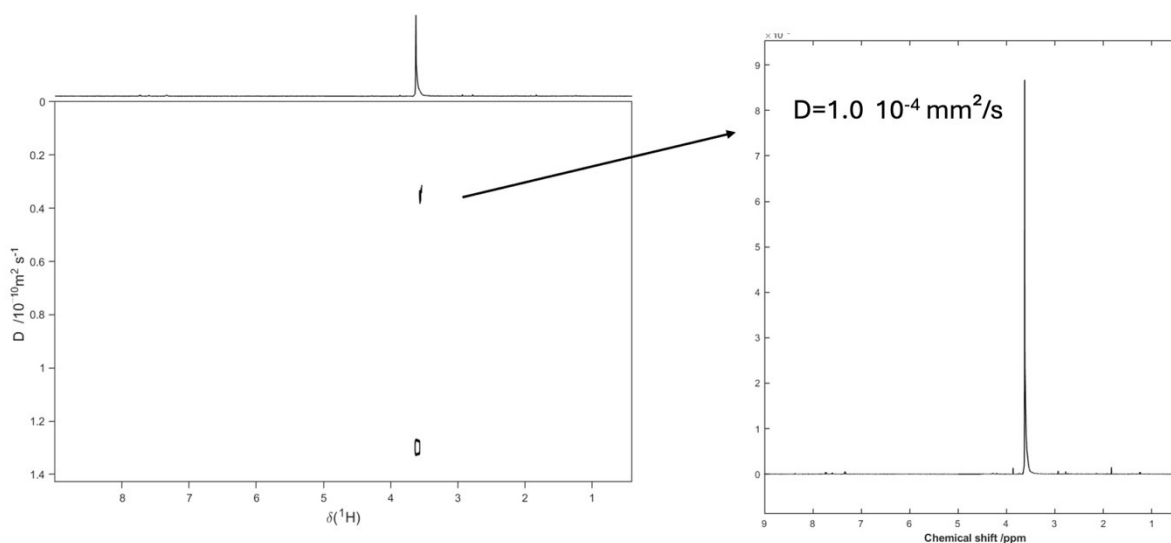


Figure S11. DOSY ^1H spectrum (left) of PEG-P54 suspension in deuterated water at 25 °C after filtration through 0.2 μm and corresponding to one main component SCORE analysis (right).

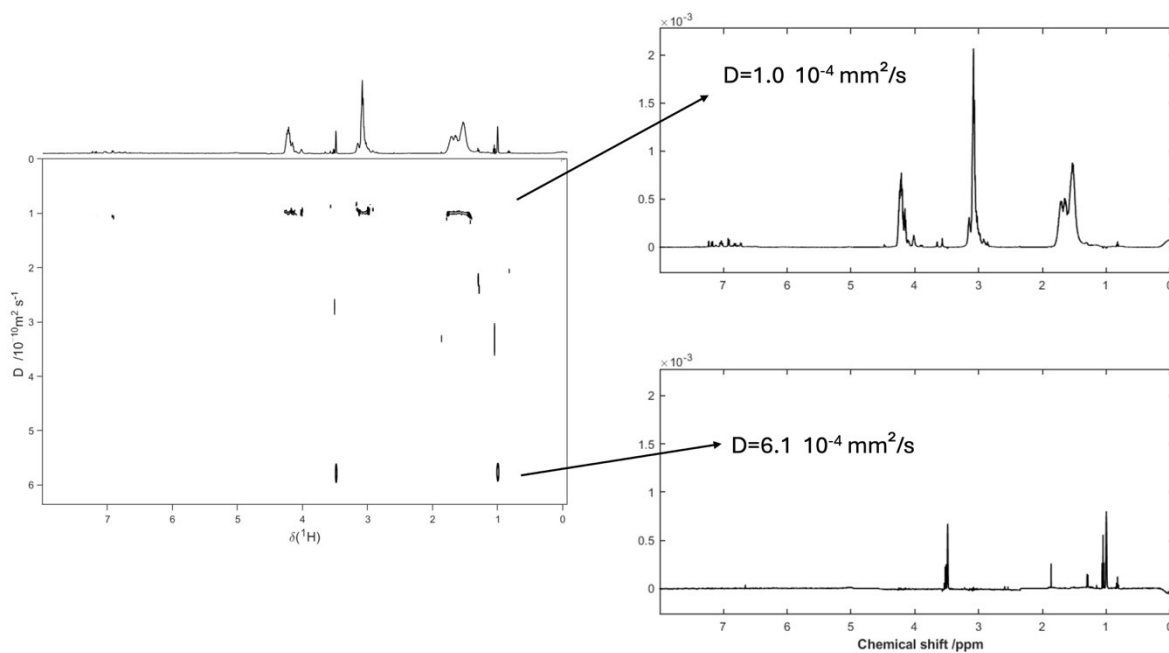


Figure S12. DOSY ^1H spectrum (left) of Arg₉-P819/PEG-P819 suspension in deuterated water at 25 °C after filtration through 0.2 μm and corresponding two components SCORE analysis (right).

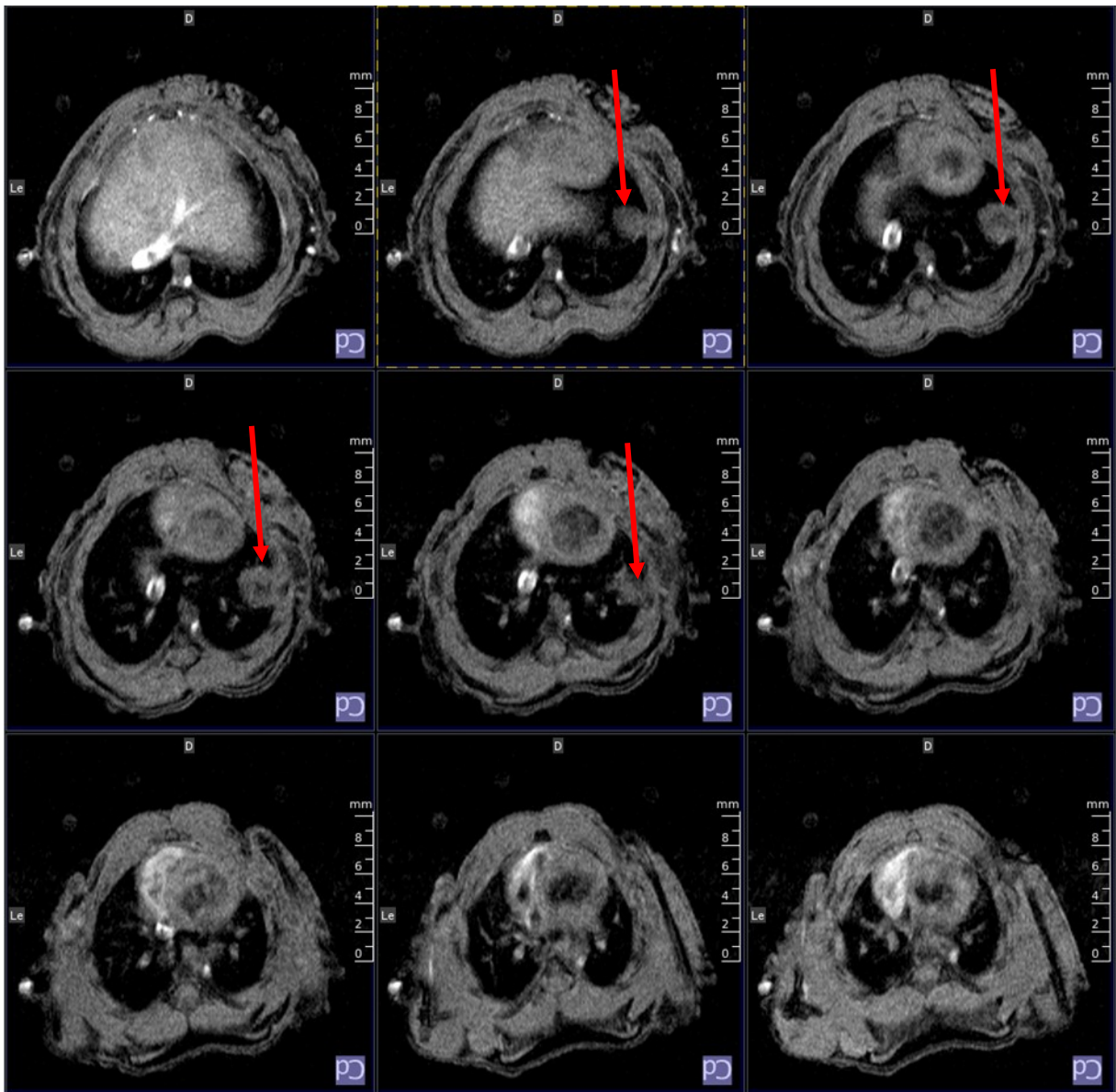
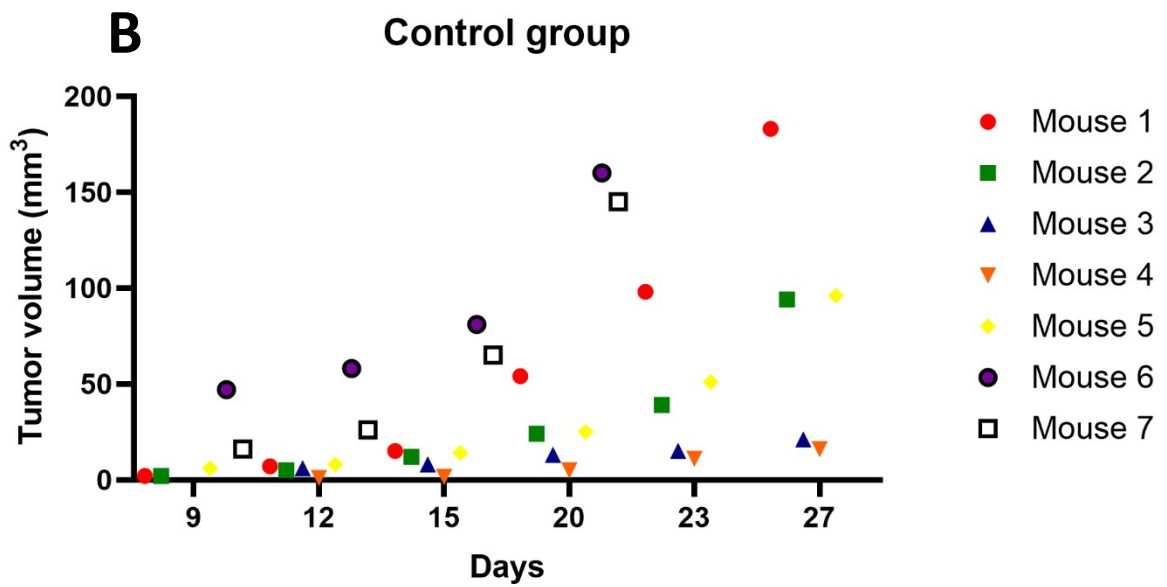
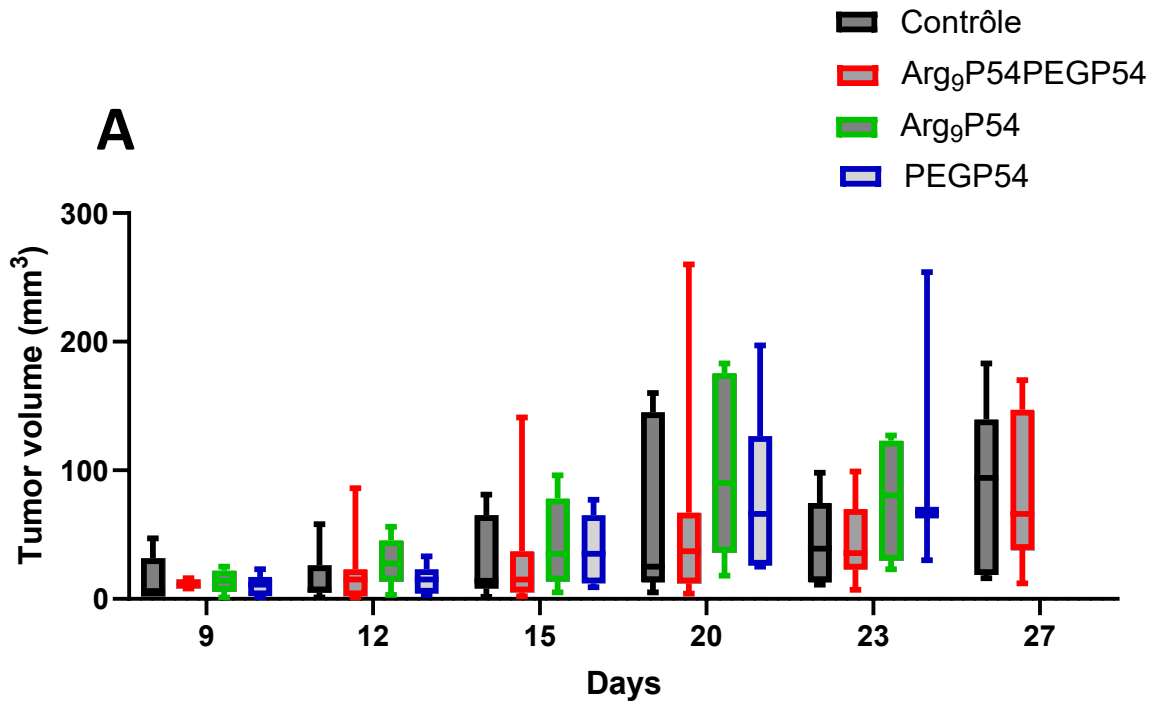


Figure S13. intrapulmonary tumor detected by MRI (the red arrow). This representative picture shows 9 slices of 0.75 millimeters of thickness per slice. Starting from the upper left image to the lower right ones, the slices were taking from the lower part to the upper part of the mouse.



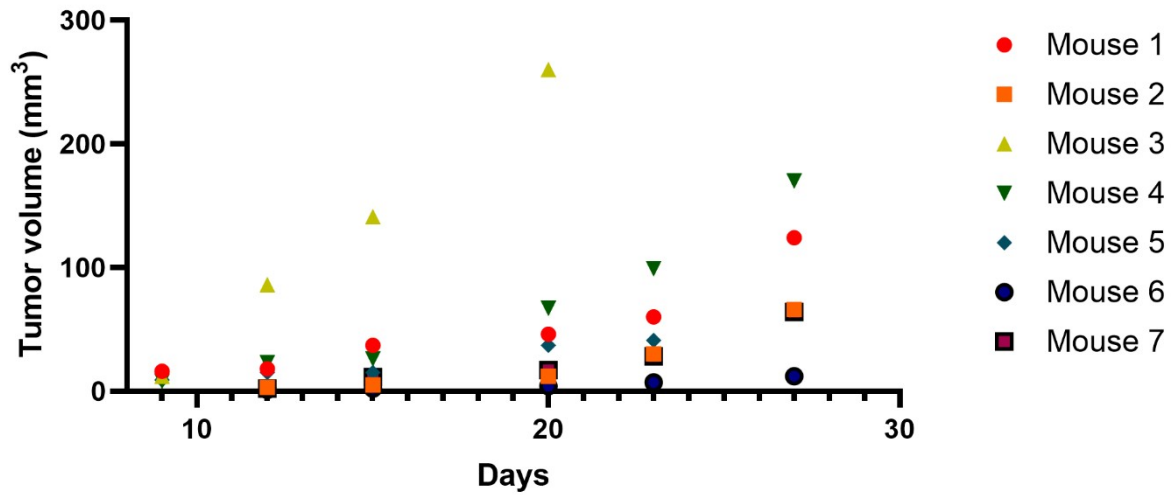
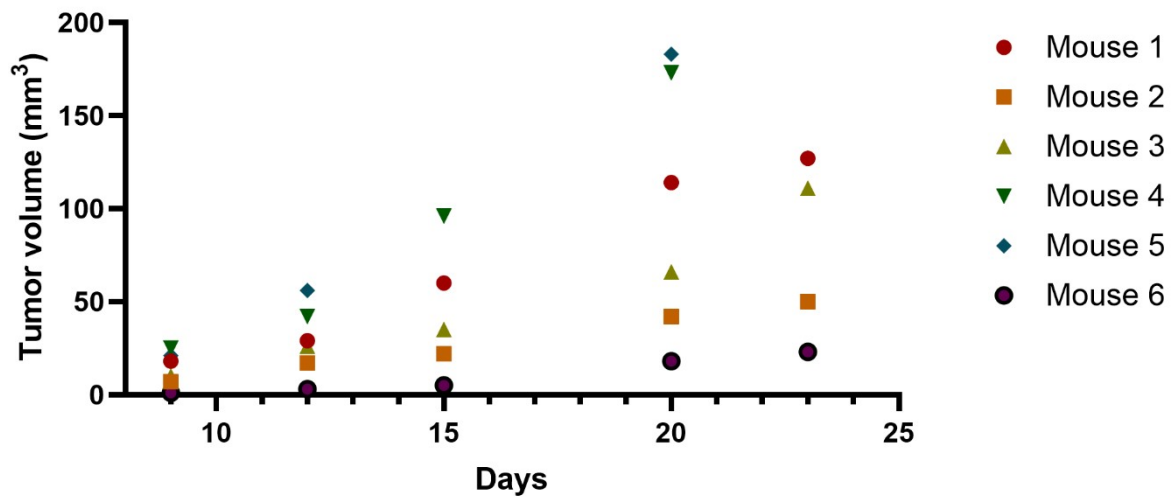
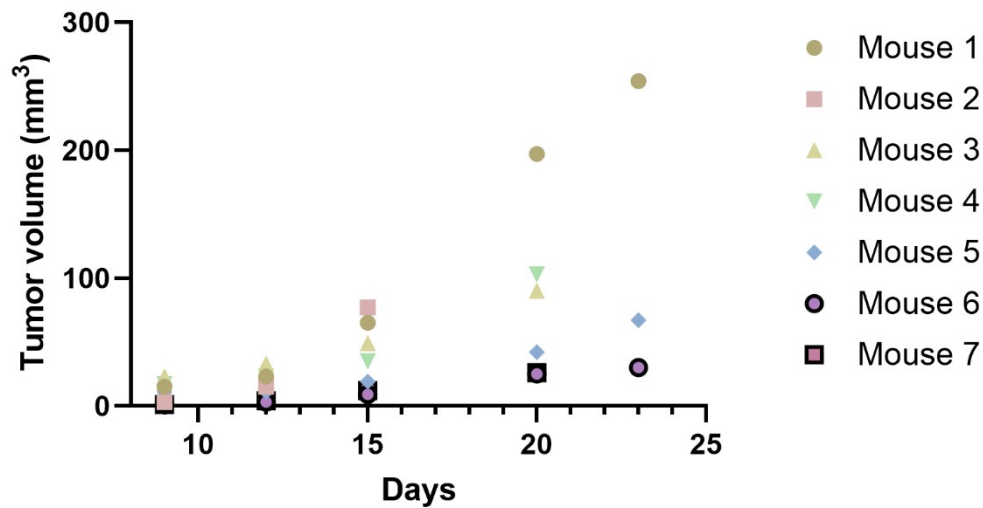
C**Arg₉P54PEGP54****D****Arg₉P54****PEGP54****E**

Figure S14. Global tumor size evolution for 27 days (A) and the detailed description per group of study: Control group (B), Arg₉-P54/PEG-P54 (C), Arg₉-P54 (D) and PEG-P54 (E).

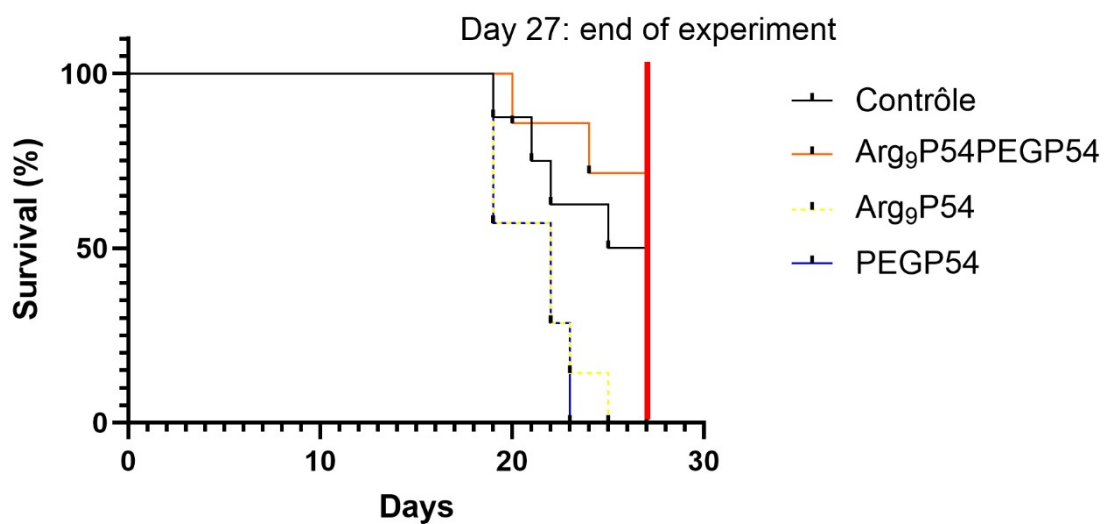


Figure S15. Mice survival during the course of the study, 27 days.

References

1. Guyon, L. *et al.* Importance of Combining Advanced Particle Size Analysis Techniques To Characterize Cell-Penetrating Peptide–Ferrocifen Self-Assemblies. *J. Phys. Chem. Lett.* **10**, 6613–6620 (2019).