

## Supporting Information

### Hybrid lipoplex co-delivering siPD-L1/miR-140-5p reverses T cell exhaustion via post-transcriptional regulation for in situ photothermal vaccination

Qunjie Bi<sup>1</sup>, Huan Yang<sup>1</sup>, Shiyi Li<sup>1</sup>, Haoyue Chen<sup>1</sup>, Yichun Wang<sup>1</sup>, Binbin Yang<sup>1</sup>, Tao Zhang<sup>2</sup>, Xia He<sup>3</sup>, Matthias Barz<sup>4</sup>, Heyang Zhang<sup>4</sup>, Rongrong Jin<sup>1\*</sup>, Yu Nie<sup>1,2,3\*</sup>

1. National Engineering Research Center for Biomaterials, College of Biomedical Engineering, Sichuan University, Chengdu, 610041, China

2. Department of Biochemistry and Molecular Biology, School of Biological Sciences and Technology, Chengdu Medical College, Chengdu 610500, Sichuan, China

3. Department of Pharmacy, Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital, School of Medicine, University of Electronic Science and Technology of China, Chengdu, 610072, China

4. Leiden Academic Center for Drug Research (LACDR), Leiden University, Leiden 2333 CC, the Netherlands

\*Corresponding authors: nie\_yu@scu.edu.cn (Yu Nie); jinrr2015@scu.edu.cn (Rongrong Jin);

#### 1. Acronyms

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|          |  |
|----------|--|
| 3'-UTR   | 3' untranslated region   |
| AR       | AuNF coated RLS lipoplexes   |
| AR/si/mi | AuNF/RLS/siRNA/miRNA   |
| ARC      | AuNF/RLS/CpG   |
| AuNFs    | gold nanoflowers   |
| AuNPs    | gold nanospheres   |
| BMDCs    | bone marrow-derived dendritic cells  |
| CCK-8    | cell counting kit-8  |
| CDS      | coding sequence  |
| CpG ODN  | CpG oligodeoxynucleotide   |
| DAPI     | 4',6-diamidino-2-phenylindole  |
| DCs      | dendritic cells  |
| EG.7.OVA | OVA-expressing EL4 murine thymoma cell line  |
| FITC     | fluorescein isothiocyanate   |
| HeLa-GFP | Green fluorescent protein (GFP) stable-expressing human cervical carcinoma cell line |
| ICD      | immunogenic cell death   |
| ICIs     | immune checkpoint inhibitors   |

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|----------|--|
| Lipo2000 | Lipofectamine 2000                               |
| miRNA    | microRNA   |
| MPTT     | mild photothermal therapy                        |
| NIR      | Near-infrared                                    |
| OS       | overall survival                                 |
| OVA      | Ovalbumin  |
| PD-L1    | programmed cell death ligand 1                   |
| PTT      | Photothermal treatment                           |
| qPCR     | quantitative real-time polymerase chain reaction |
| RIPA     | radioimmunoprecipitation assay                   |
| RNAi     | RNA interference                                 |
| Rr       | pearson correlation coefficient                  |
| siGFP    | silencing GFP siRNA                              |
| siPD-L1  | silencing PD-L1 siRNA                            |
| siRNA    | small interfering RNA                            |
| TAAAs    | tumor-associated antigens                        |
| TEM      | transmission electron microscope                 |

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**Table S1.** The sequences of siRNA

| siRNA     | Sense (5' - 3')         | Antisense (5' - 3')     |
|-----------|-------------------------|-------------------------|
| siPD-L1-1 | CCCACAUA AAAAACAGUUGTT  | CAACUGUUUUUAUGUGGGTT    |
| siPD-L1-2 | UGAGAAAACAAGUGAGAAUTT   | AUUCUCACUUGUUUCUCATT    |
| siPD-L1-3 | GAGGUA AUCUGGACAAACATT  | UGUUUGUCCAGAUUACCUCTT   |
| siPD-L1-4 | AGACGUAAGCAGUGUUGAATT   | UUCAACACUGCUUACGUCUTT   |
| siPD-L1-5 | GCCGAAGUCAUCUGGACAATT   | UUGUCCAGAUGACUUCGGCTT   |
| siPD-L1-6 | ACUUCUGAGCAUGAACUAAUAUG | CAUAUUAGUUCAUGCUCAGAAGU |

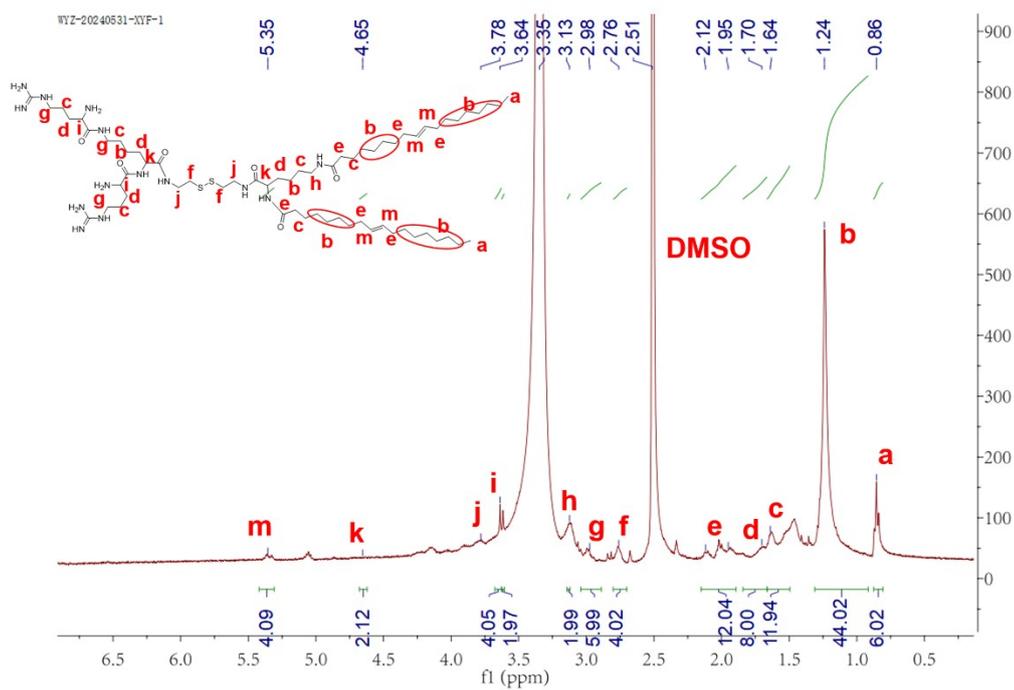
**Table S2.** The sequences of miRNA

| miRNA      | Sense (5' - 3')          | Antisense (5' - 3')     |
|------------|--------------------------|-------------------------|
| miR-17-5p  | CAAAGUGCUUACAGUGCAGGUAG  | ACCUGCACUGUAAGCACUUUGUU |
| miR-155-5p | UUA AUGCUAAUUGUGAUAGGGGU | CCCUAUCACAAUAGCAUUA AUU |
| miR-142-5p | CAUAAAGUAGAAAGCACUACU    | UAGUGCUUUCUACUUUAUGUU   |
| miR-140-5p | CAGUGGUUUUACCCUAUGGUAG   | ACCAUAGGGUAAAACCACUGUU  |

**Table S3.** The primer sequences for various genes.

| Target         | Forward primer          | Reverse primer           |
|----------------|-------------------------|--------------------------|
| IL-1 $\beta$   | TCGCAGCAGCACATCAACAAGAG | TGCTCATGTCCTCATCCTGGAAGG |
| TNF- $\alpha$  | GCGACGTGGA ACTGGCAGAAG  | GCCACAAGCAGGAATGAGAAGAGG |
| IL-6           | AGAGGAGACTTCACAGAGGA    | ATCTCTCTGAAGGACTCTGG     |
| PD-L1          | CGAGGGTTATCCAGAAGCTGAG  | CTGGTCACATTGAGAAGCATCC   |
| $\beta$ -actin | GTGCTATGTTGCTCTAGACTTCG | ATGCCACAGGATTCCATACC     |

A



B

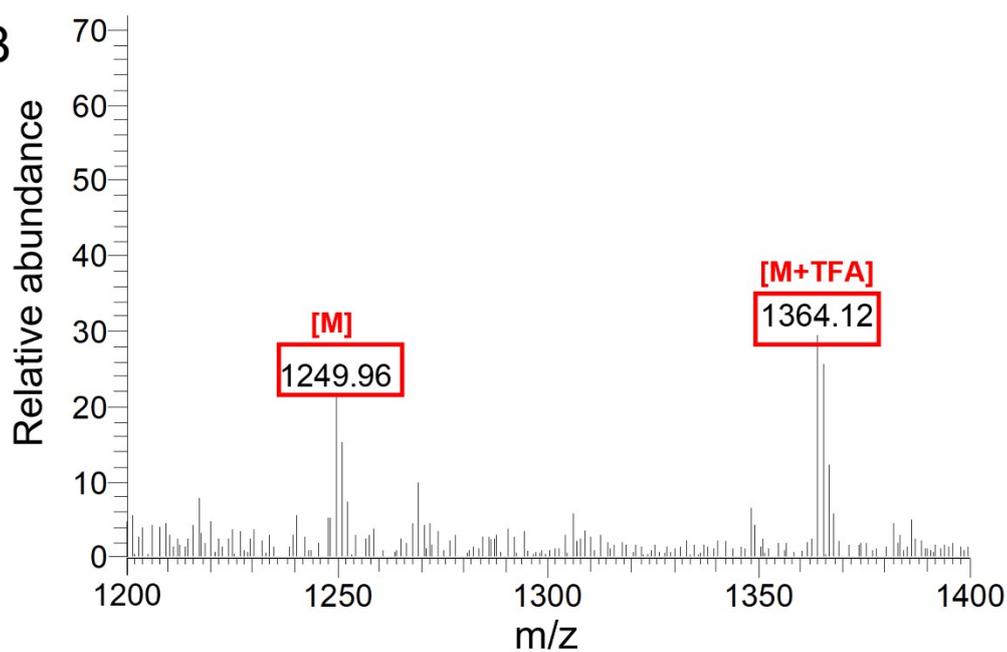
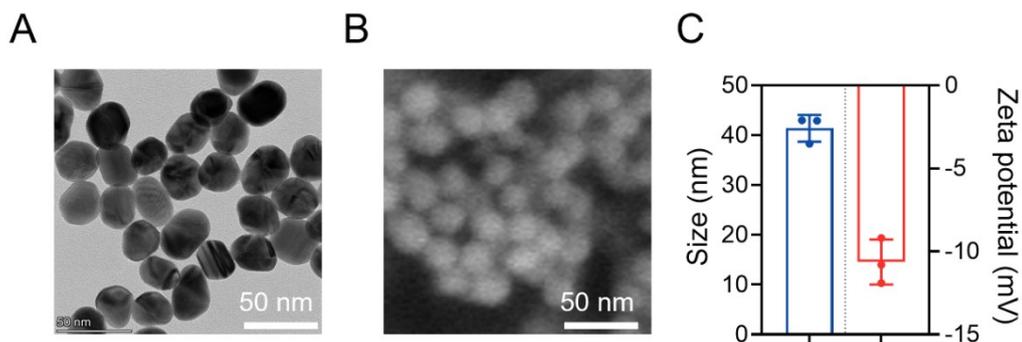
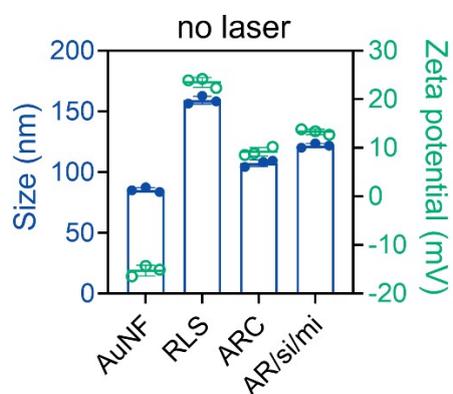


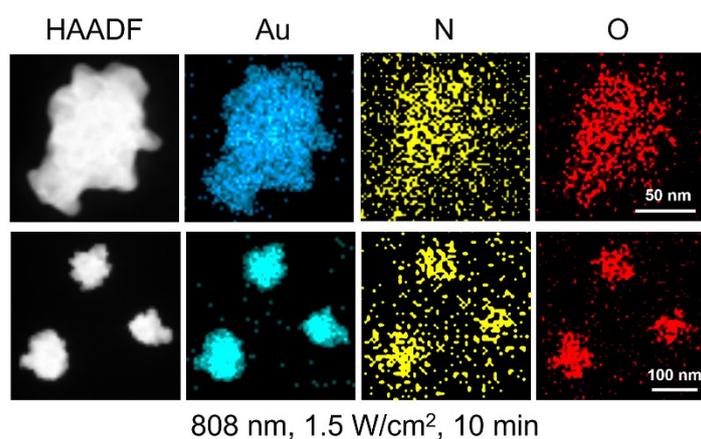
Fig. S1. Chemical characterization (A)  $^1\text{H}$  NMR spectrum and (B) LC-MS of RLS.



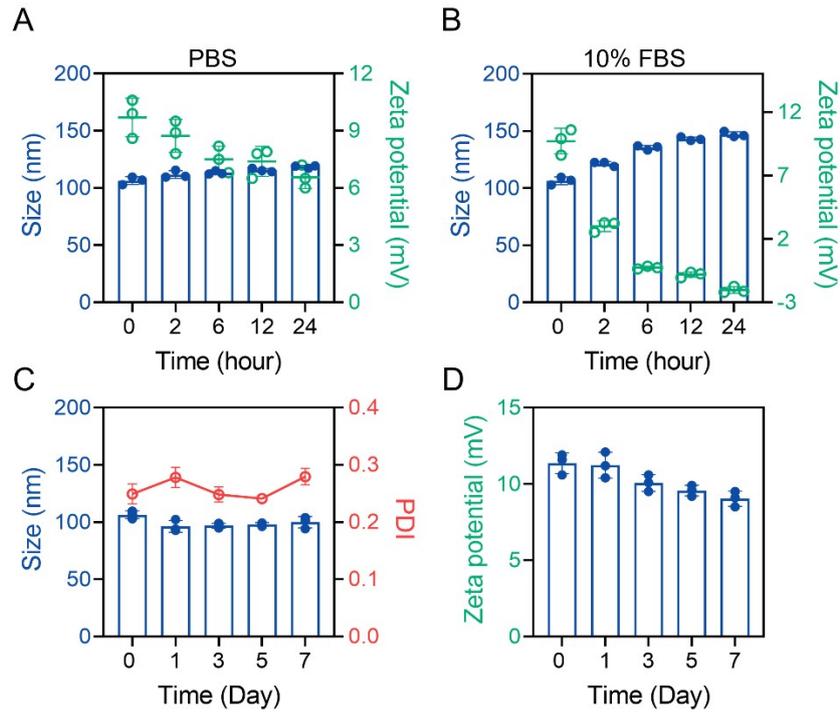
**Fig. S2.** Characterization of gold nanoparticles. (A) The representative (TEM) image of AuNPs. The scale bar is 50 nm. (B) The representative scanning electron microscope (SEM) image of AuNP. The scale bar is 50 nm. (C) Size distribution and  $\zeta$  potential of AuNPs as detected by dynamic light scattering (DLS).



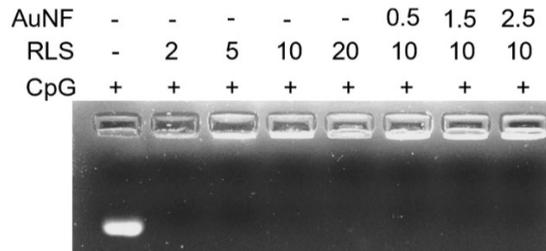
**Fig. S3.** Size distribution and  $\zeta$  potential of AuNF, RLS, and hybrid lipoplexes after a mild irradiation condition (808 nm, 1.5 W/cm<sup>2</sup>, 10 min) as detected by dynamic light scattering (DLS). All values represent the mean  $\pm$  SD (n = 3).



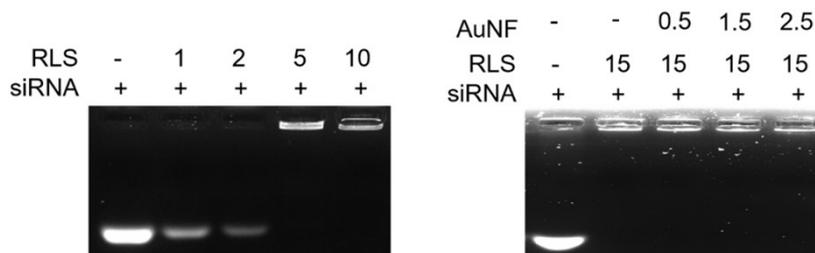
**Fig.S4.** The HAADF-STEM image of ARC after a mild irradiation condition (808 nm, 1.5 W/cm<sup>2</sup>, 10 min). The scale bar is 50 and 100 nm.



**Fig.S5.** Stability evaluation of the hybrid lipoplexes. Size distribution and  $\zeta$  potential of hybrid lipoplexes after incubation in (A) PBS or (B) 10% FBS containing medium at 37 °C for 0, 2, 4, 6, 12 and 24 h as detected by dynamic light scattering (DLS). (C) Size distribution and PDI of hybrid lipoplexes during storage at 4 °C in deionized water for 0, 1, 3, 5 and 7 days. (D)  $\zeta$  potential of hybrid lipoplexes during storage at 4 °C in deionized water for 0, 1, 3, 5 and 7 days. All values represent the mean  $\pm$  SD (n = 3).

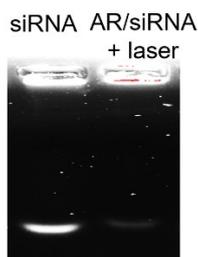


**Fig. S6.** Gene compaction of ARC at different weight ratios. RLS/CPG at N/P ratios of 1, 2, 5, 10, and 20, and AuNF/CpG at weight ratios of 0.5, 1.5, 2.5.

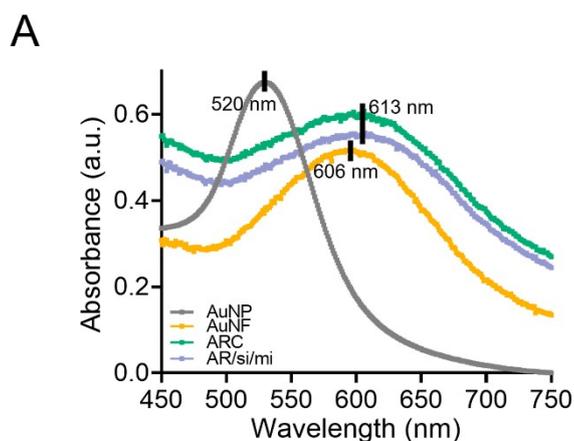


**Fig. S7.** Gene compaction of RLS/siRNA lipoplexes and AR/siRNA with different mass ratios. (RLS/siRNA at N/P ratios of 1, 2, 5, and 10; and AuNF/siRNA at weight ratios of 0.5, 1.5, 2.5,

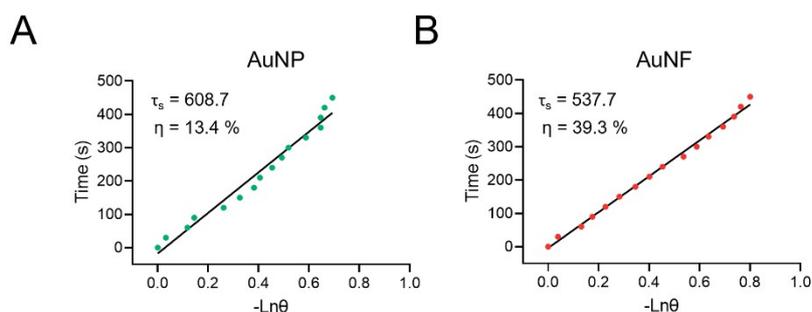
siRNA = 0.2  $\mu\text{g}$ ).



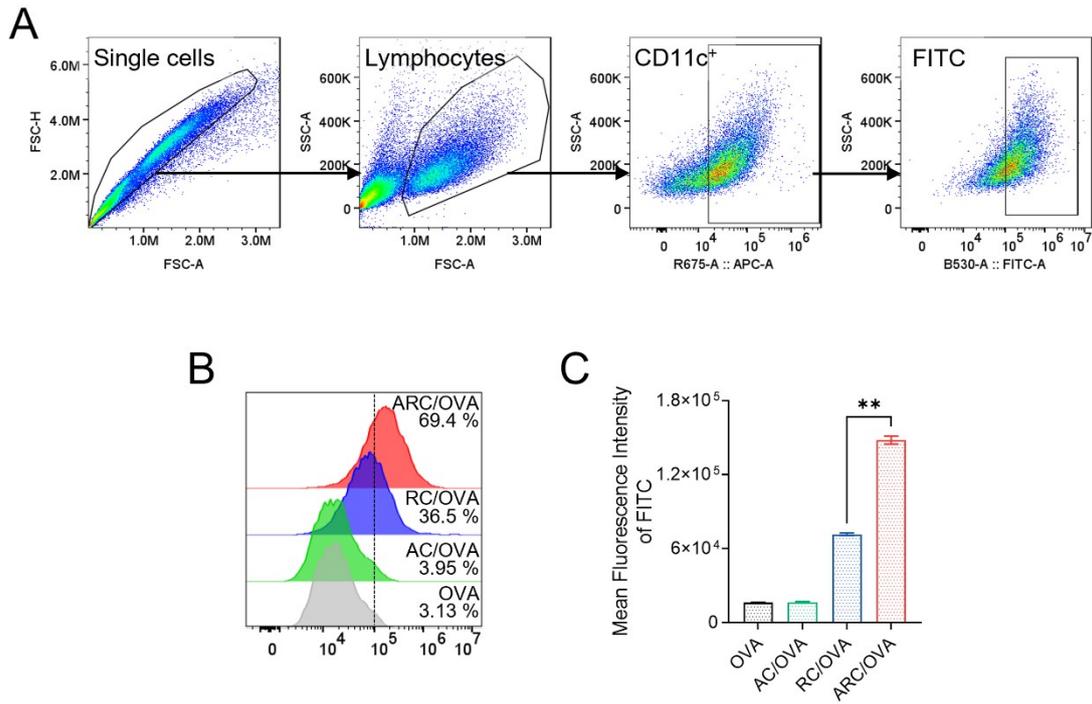
**Fig.S8.** Gene compaction of AR/siRNA after a mild irradiation condition (808 nm, 1.5 W/cm<sup>2</sup>, 10 min). (siRNA = 0.2  $\mu\text{g}$ , RLS/siRNA at N/P ratio of 15; and AuNF/siRNA at weight ratio of 1.5.)



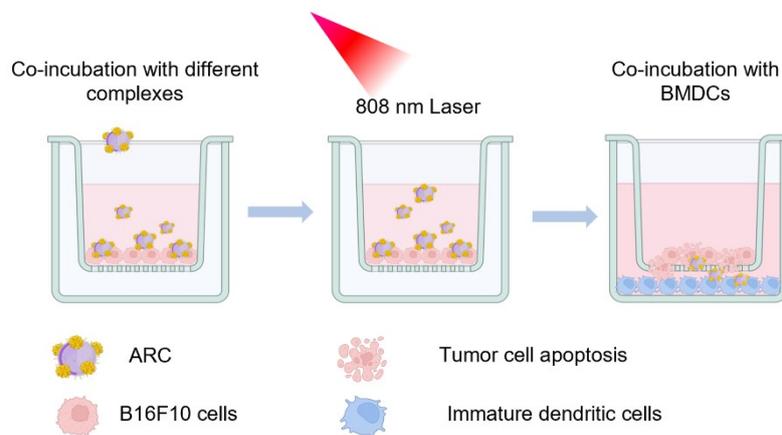
**Fig. S9.** Ultraviolet-visible spectroscopy (UV-vis) absorption spectra of AuNPs, AuNFs, ARC and AR/si/mi.



**Fig. S10.** Evaluation of photothermal conversion efficiency. Calculation time constant ( $\tau_s$ ) and photothermal-conversion efficiency ( $\eta$ ) of (A) AuNPs and (B) AuNFs. Photothermal stability of AuNPs or AuNFs with five cycled irradiations (808 nm, 1.5 W/cm<sup>2</sup>, 10 min) separated by a 20 min cooling phase. Cooling curves used for the determination of the time constant ( $\tau_s$ ).



**Fig. S11.** Cellular uptake of different lipoplexes in BMDCs. (A) Gating strategy for CD11c and FITC-OVA. (B) BMDCs treated with various OVA-loaded lipoplexes for 4 h and the cellular uptake determined by flow cytometer. (C) Quantitative analysis of the cellular uptake of various lipoplexes by MFI. All values represent the mean  $\pm$  SD ( $n = 3$ ) and were analyzed by two-sided Student's  $t$ -test.  $**p < 0.01$ .



**Fig. S12.** Schematic illustration of the transwell system experiment. The B16F10 cells were cultured in the upper chamber and irradiated after the addition of different complexes for 4 h ( $1.5 \text{ W/cm}^2$ , 10 min). Then, the upper chamber was placed into a new lower chamber pre-seeded with BMDCs for a further 20 h. Finally, the BMDCs were harvested for analysis.

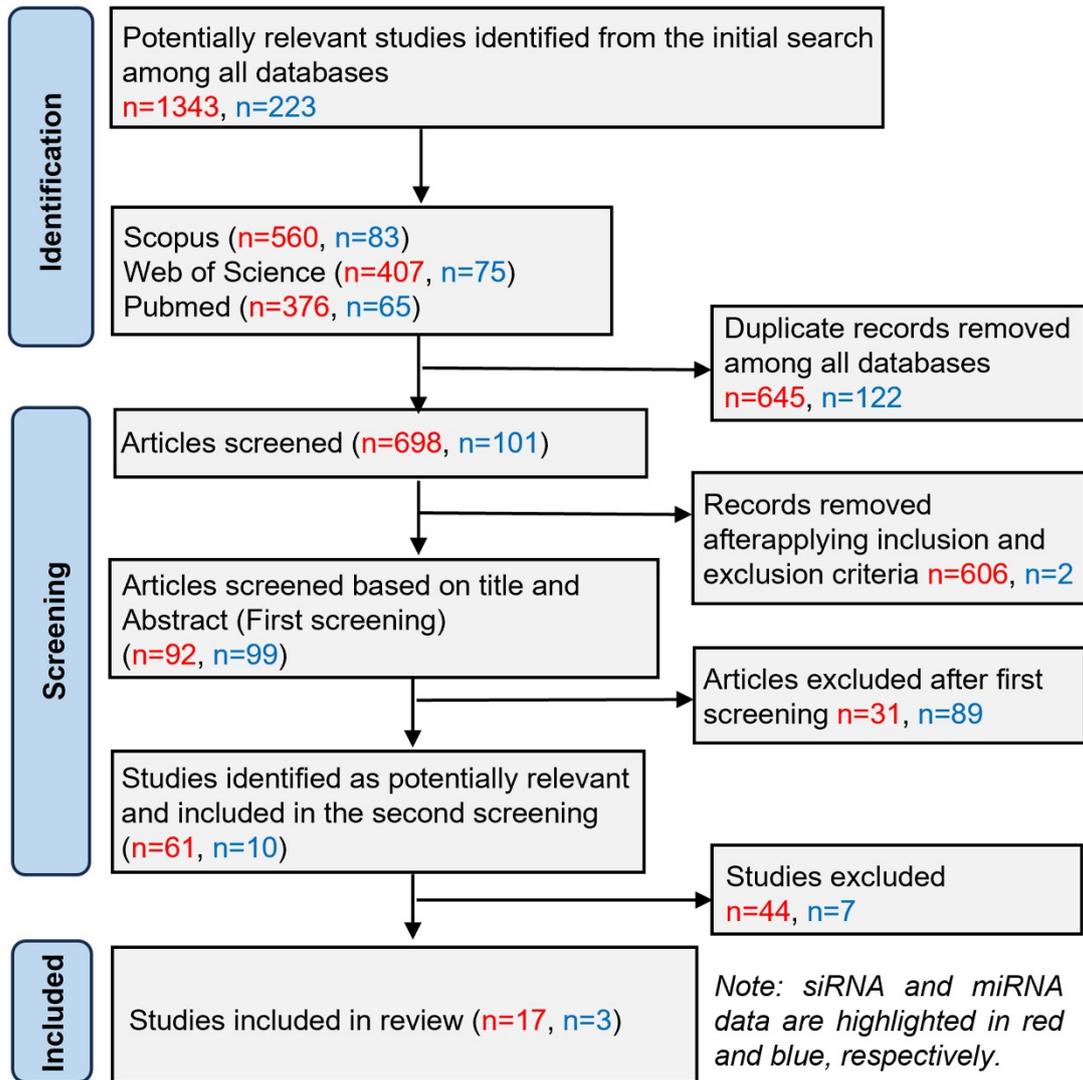
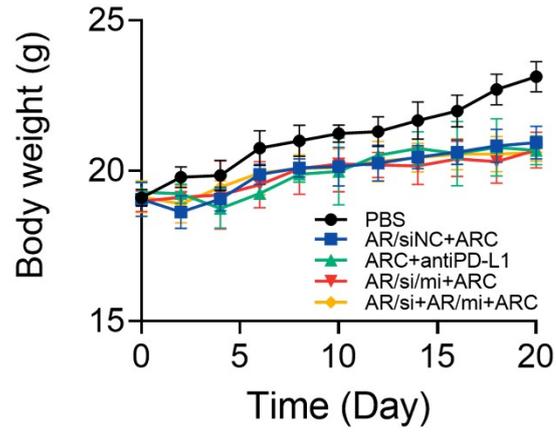
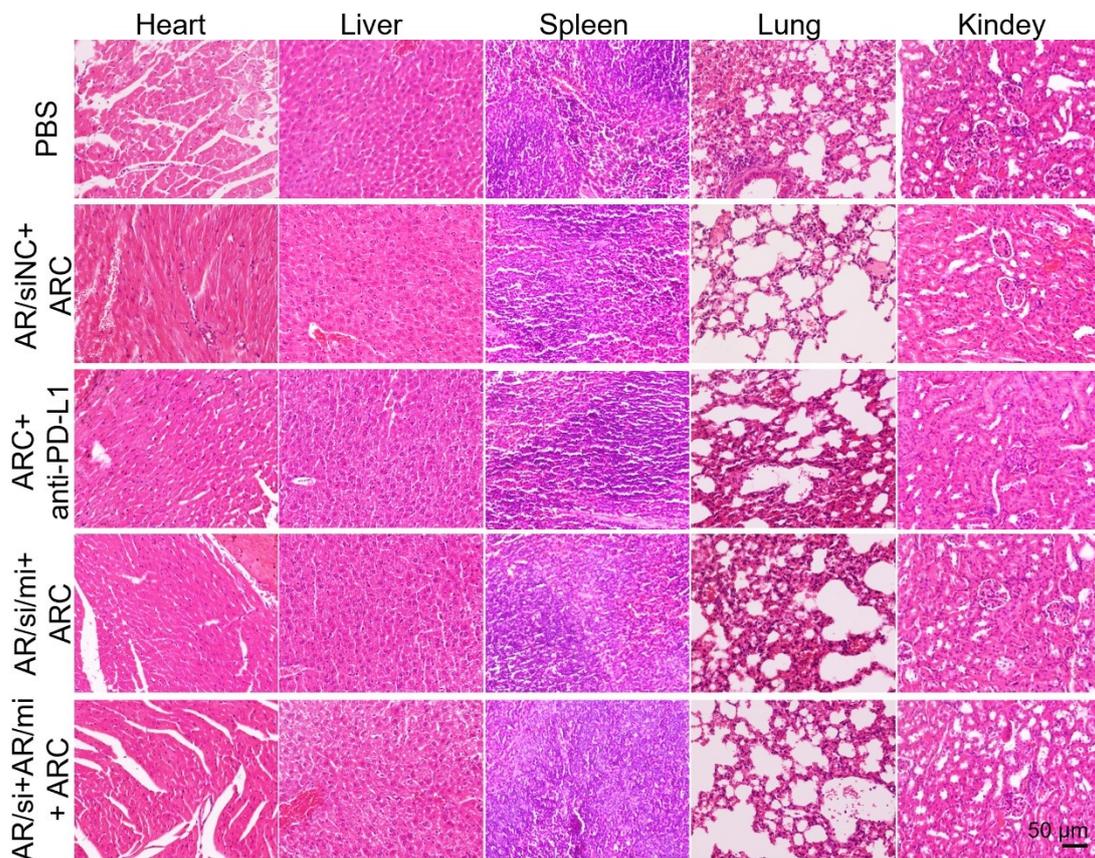


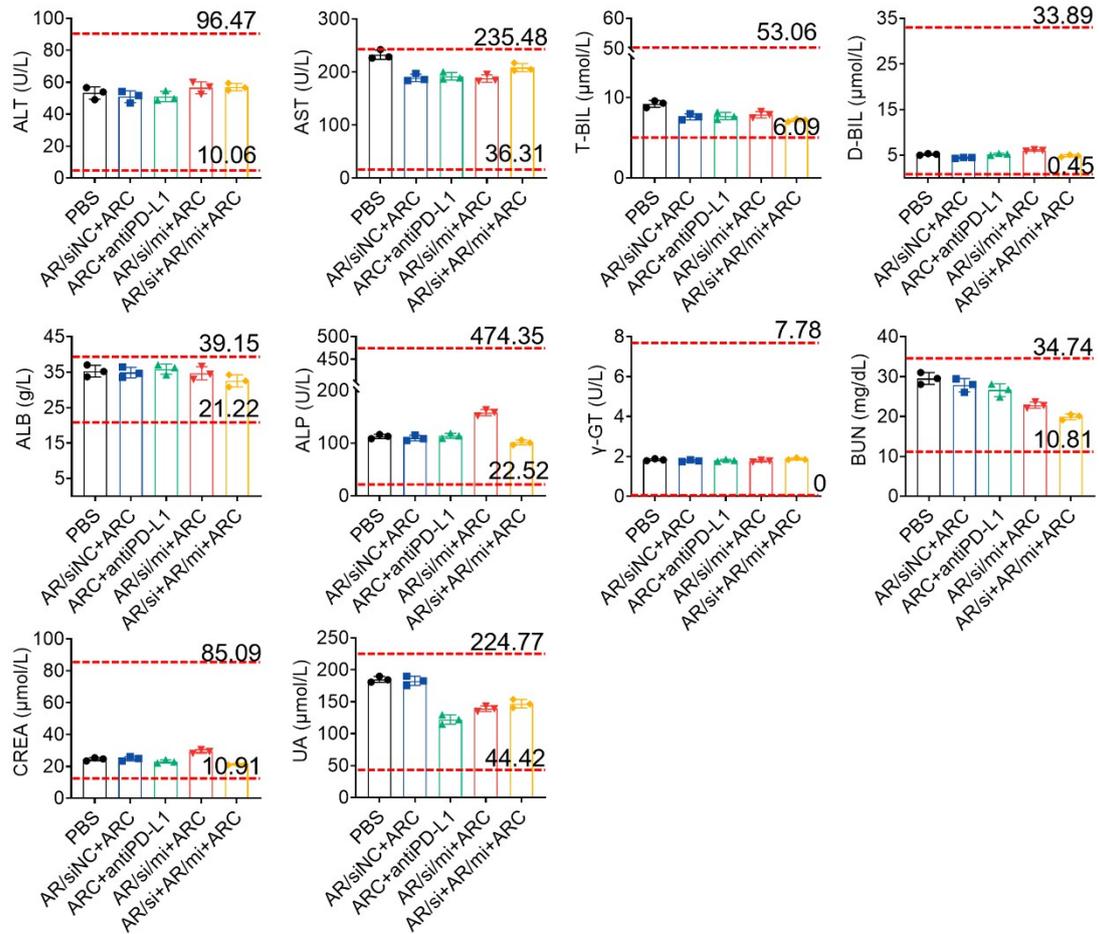
Fig. S13. Flowchart of the literature screening process.



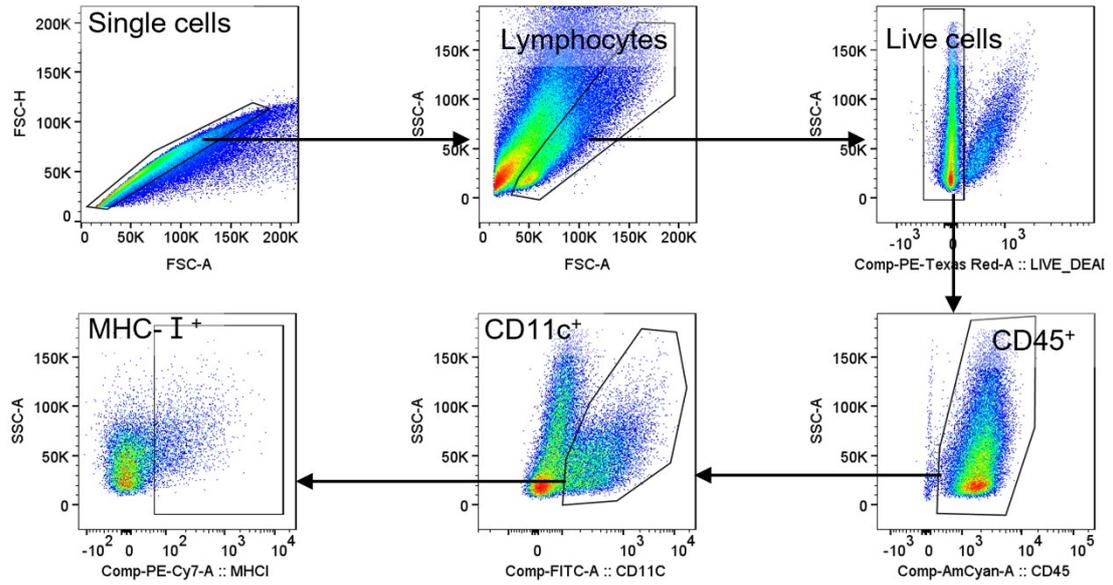
**Fig. S14.** Body weight change of C57BL/6 mice with B16F10 xenografts in different groups. All values represent the mean  $\pm$  SD.  $n = 5$  mice.



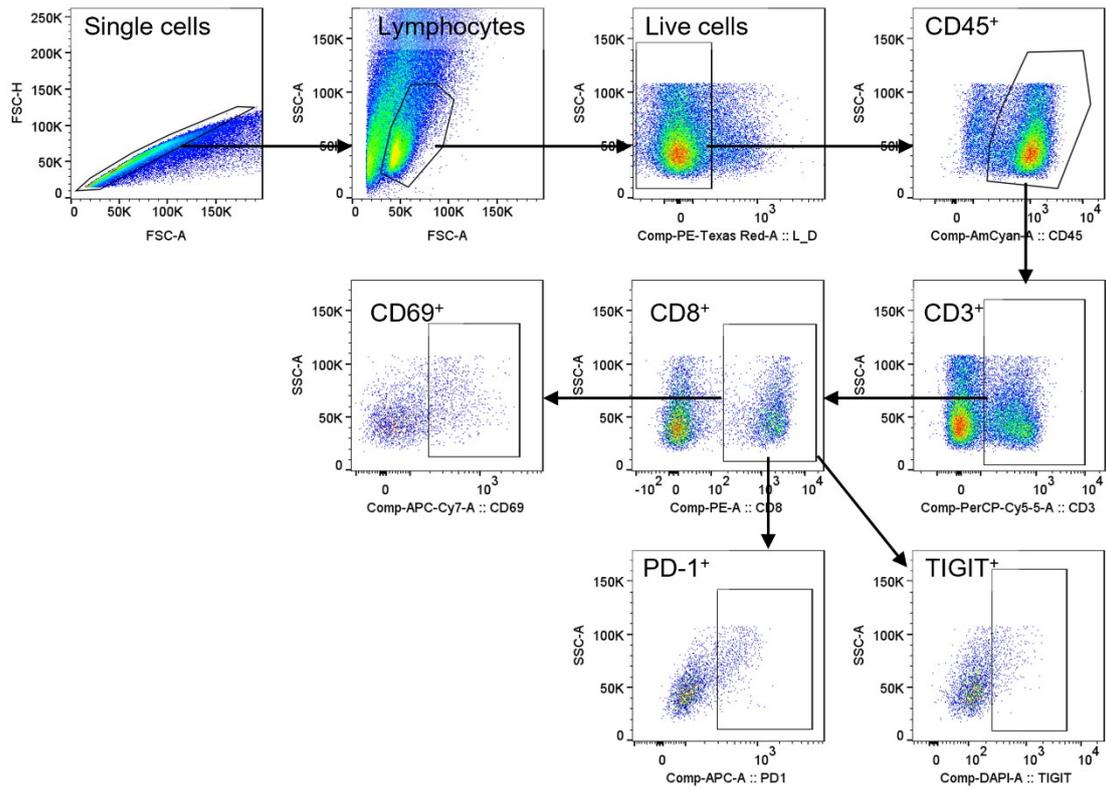
**Fig. S15.** H&E staining images of major organs of B16F10 tumor-bearing mice after treatment with different lipoplexes. Major organs (including heart, liver, spleen, lung, and kidney) were isolated from the sacrificed mice on day 20. Scale bar is 50  $\mu$ m.



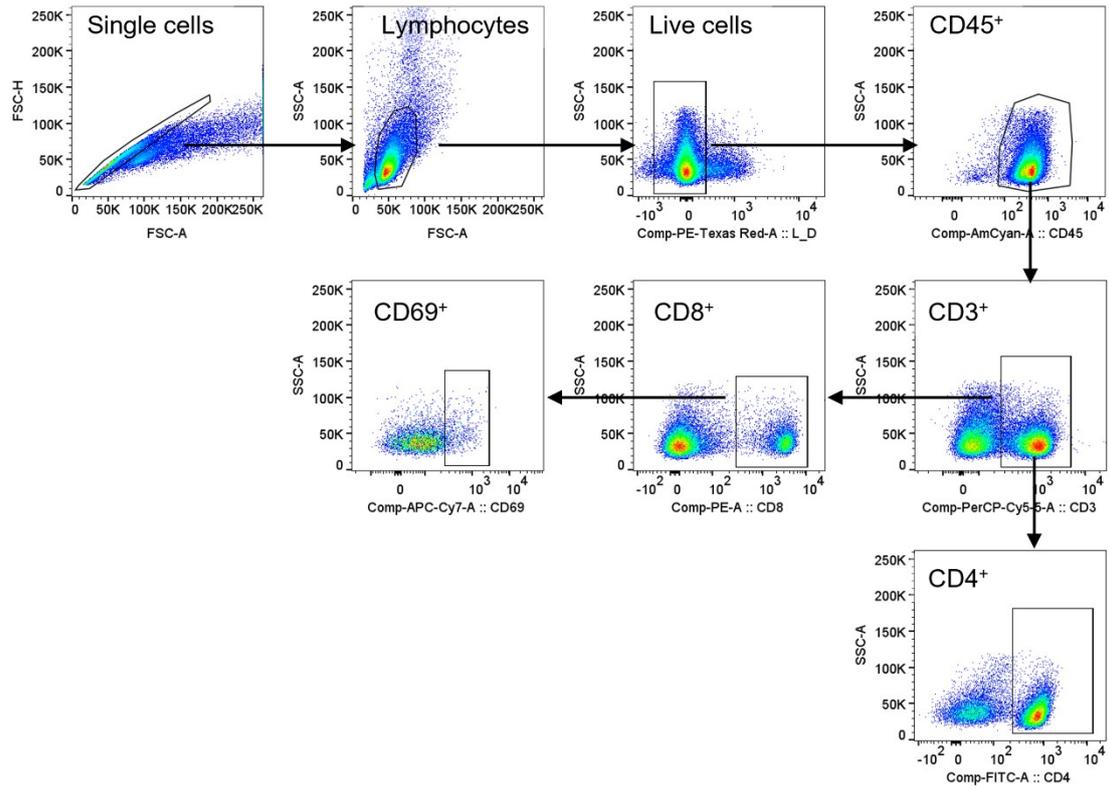
**Fig. S16.** Hepatic and renal function analyses of B16F10 tumor-bearing mice following different lipoplex administrations. Including alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (T-BIL), direct bilirubin (D-BIL), albumin (ALB), alkaline phosphatase (ALP),  $\gamma$ -glutamyl transferase ( $\gamma$ -GT), blood urea nitrogen (BUN), creatinine (CREA), and uric acid (UA). All values represent the mean  $\pm$  SD (n = 3).



**Fig. S17.** Gating strategy for identifying BMDC populations within the tumor microenvironment by flow cytometry.



**Fig. S18.** Gating strategy for identifying T cell populations in the tumor microenvironment by flow cytometry.



**Fig. S19.** Gating strategy for identifying immune cell populations in the spleen by flow cytometry.