Supplementary Information (SI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2025

#### 1. General considerations

All reagents and starting materials were commercially obtained and were used without further purification unless otherwise noted. The 4-arylpyrrolo[1,2-a]quinoxaline substrates were prepared according to the literature.<sup>1</sup>

Gas chromatography (GC) analyses were performed using Shimadzu GC-2010 Plus equipped with a flame ionization detector (FID) and an SPB-5 column (length = 30 m, inner diameter = 0.25 mm, and film thickness =  $0.25 \mu$ m). The GC yield was calculated using diphenyl ether as the internal standard. Gas chromatography – mass spectrometry (GC-MS) analyses were performed using Shimadzu GCMS-QP2010 Ultra with a ZB-5MS column (length = 30 m, inner diameter = 0.25 mm, and film thickness =  $0.25 \mu$ m). Proton and carbon-13 (<sup>1</sup>H NMR, <sup>13</sup>C NMR) spectra were recorded on Bruker AV 600 and 500 spectrometers, respectively, using residual solvent as reference. Splitting is reported with the following symbols: s = singlet, d = doublet, t = triplet, dd = doubletof doublets, and m = multiplet. Coupling constants (*J*) are reported in Hertz. Fourier transform infrared (FT-IR) spectra were obtained from a Shimadzu IRPrestige-21 instrument, with samples being dispersed on potassium bromide pallets. Highresolution mass spectra (HRMS) were recorded by Agilent 6500 Series Q-TOF LC/MS System. The mass spectrometry was performed in the positive electrospray ionization (ESI+) mode. For compound 3ai, mass spectrum was obtained from UPLC Acquity H-Class, Waters.

Analytical thin-layer chromatography (TLC) was performed on silica gel 60 F254 (Merck), visualized by irradiation with UV light (254 nm and 365 nm). Column chromatography was performed on silica gel (Merck,  $15 - 40 \mu$ m). To prepare a caffeinated silica gel, silica gel ( $15 - 40 \mu$ m, 40 g), caffeine (8 g), and CH<sub>2</sub>Cl<sub>2</sub> ( $100 \mu$ ) were added to a round-bottom flask. The ensuing mixture was stirred for 15 min then evaporated by rotovap until a free-flowing powder was obtained.

## 2. Optimization of reaction conditions

## 2.1. Effect of palladium catalyst



Entry <sup>a</sup>	Catalyst	GC yield (%) <sup>b</sup>
1	Pd(OAc) <sub>2</sub>	55
2	$Pd(TFA)_2$	0
3	PdCl <sub>2</sub>	13
4°	$PdCl_2(PPh_3)_2$	19
5°	Pd(PPh <sub>3</sub> ) <sub>4</sub>	16

<sup>a</sup>Reaction conditions: **1a** (0.1 mmol), **2a** (0.15 mmol), catalyst (10 mol%), PPh<sub>3</sub> (20 mol%), K<sub>2</sub>CO<sub>3</sub> (0.2 mmol), toluene (1 mL), 100 °C, 24 h. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard. °No PPh<sub>3</sub>.

## 2.2. Effect of environment



Entry	Environment	GC yield (%) <sup>a</sup>
1	Oxygen	35
2	Argon	87
3	Air	55

<sup>a</sup>Reaction condition: **1a** (0.1 mmol), **2a** (0.15 mmol),  $Pd(OAc)_2$  (10 mol%),  $PPh_3$  (20 mol%),  $K_2CO_3$  (0.2 mmol), toluene (1 mL), 100 °C, 24 h. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard.

## 2.3. Effect of temperature



Entry <sup>a</sup>	Temperature (°C)	GC yield (%) <sup>b</sup>
1	40	0
2	60	11
3	80	27
4	100	87
5	120	78

<sup>a</sup>Reaction condition: **1a** (0.1 mmol), **2a** (0.15 mmol), Pd(OAc)<sub>2</sub> (10 mol%), PPh<sub>3</sub> (20 mol%), K<sub>2</sub>CO<sub>3</sub> (0.2 mmol), toluene (1 mL), 24 h, under argon. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard.

# 2.4. Effect of solvent



Entry <sup>a</sup>	Solvent	GC yield (%) <sup>b</sup>
1	toluene	87
2	DMSO	12
3	NMP	35
4	1,4-dioxane	97
5	PhCl	83
6	DMF	89
7	DMAc	96
8	<i>m</i> -xylene	70
9	$H_2O$	78
10	n-butanol	0
11	diethyl carbonate	79

<sup>a</sup>Reaction condition: **1a** (0.1 mmol), **2a** (0.15 mmol), Pd(OAc)<sub>2</sub> (10 mol%), PPh<sub>3</sub> (20 mol%), K<sub>2</sub>CO<sub>3</sub> (0.2 mmol), solvent (1 mL), 100 °C, 24 h, under argon. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard.

## 2.5. Effect of ligand



Entry <sup>a</sup>	Ligand	GC yield (%) <sup>b</sup>
1	No ligand	23
2	XPhos	0
3	BINAP	0
4	PPh <sub>3</sub>	97
5	1,10-phenanthroline	13

<sup>a</sup>Reaction condition: **1a** (0.1 mmol), **2a** (0.15 mmol), Pd(OAc)<sub>2</sub> (10 mol%), ligand (20 mol%), K<sub>2</sub>CO<sub>3</sub> (0.2 mmol), 1,4-dioxane (1 mL), 100 °C, 24 h, under argon. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard.

## 2.6. Effect of reaction time



Entry <sup>a</sup>	Time (hour)	GC yield (%) <sup>b</sup>
1	2	23
2	4	45
3	6	85
4	16	98
5	24	97

<sup>a</sup>Reaction condition: **1a** (0.1 mmol), **2a** (0.15 mmol), Pd(OAc)<sub>2</sub> (10 mol%), ligand (20 mol%), K<sub>2</sub>CO<sub>3</sub> (0.2 mmol), 1,4-dioxane (1 mL), 100 °C, under argon. <sup>*b*</sup>GC yields using diphenyl ether as an internal standard.

#### 3. General procedure for arylation of 4-arylpyrrolo[1,2-a]quinoxalines



In a typical experiment, a 4-mL vial was charged with a derivative of 4-phenyl pyrrolo[1,2-*a*]quinoxaline (0.1 mmol), an aryl bromide (0.15 mmol), Pd(OAc)<sub>2</sub> (2.3 mg, 10 mol%), PPh<sub>3</sub> (5.4 mg, 20 mol%), K<sub>2</sub>CO<sub>3</sub> (27.8 mg, 0.2 mmol), and 1,4-dioxane (1 mL). The vial was purged with argon, then stirred at 100 °C for 16 h. The mixture was cooled down to room temperature. For GC analysis, diphenyl ether (0.1 mmol) was added at this stage. The obtained mixture was diluted with ethyl acetate (3 mL) and washed with brine (2 x 5 mL). Aqueous phases were further extracted with ethyl acetate (3 x 5 mL). Aliquots of combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered, and analyzed with GC. For isolation, the organic phase was mixed with Celite®, then evaporated by rotovap until a free-flowing powder was obtained. Purification by column chromatography with appropriate eluents afforded the arylation product.

## 4. Characterization data of products

## 1,4-Diphenylpyrrolo[1,2-*a*]quinoxaline (3aa)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/toluene/acid acetic gradient from 115:0:5 to 75:25:20 for 1<sup>st</sup> column, then eluent: hexanes/ diethyl ether 1:1 for 2<sup>nd</sup> column) as a pale yellow solid; isolated yield: 96% (30.7 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (dd, J = 8.1, 1.6 Hz, 1H), 8.02 – 7.99 (m, 2H), 7.58 – 7.53 (m, 5H), 7.54 – 7.51 (m, 3H), 7.44 (dd, J = 8.5, 1.3 Hz, 1H), 7.39 – 7.34 (m, 1H), 7.13 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.04 (d, J = 4.0 Hz, 1H), 6.79 (d, J = 4.0 Hz, 1H). <sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  155.0, 138.7, 137.7, 134.4, 132.9, 130.4, 129.82, 129.80, 128.87, 128.86, 128.73, 128.69, 128.3, 126.9, 126.3, 125.1, 117.0, 116.6, 109.1. HRMS (ESI) m/z: [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>17</sub>N<sub>2</sub><sup>+</sup>: 321.1386; found: 321.1390.

#### 4-(4-Phenylpyrrolo[1,2-*a*]quinoxalin-1-yl)benzonitrile (3ab)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ethyl acetate 10:1) as a yellow solid; isolated yield: 74% (25.5 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.06 (dd, J = 8.0, 1.5 Hz, 1H), 8.00 – 7.96 (m, 2H), 7.80 – 7.77 (m, 2H), 7.70 – 7.65 (m, 2H), 7.59 – 7.52 (m, 3H), 7.41 (ddd, J = 8.0, 7.2, 1.3 Hz, 1H), 7.35 (dd, J = 8.5, 1.3 Hz, 1H), 7.20 (ddd, J = 8.5, 7.2, 1.5 Hz, 1H), 7.06 (d, J = 4.1 Hz, 1H), 6.83 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 155.0, 138.7, 138.3, 137.8, 132.6, 130.7, 130.5, 130.0, 129.9, 128.8, 128.7, 128.0, 127.7, 126.6, 125.6, 118.7, 118.1, 116.5, 112.1, 109.5.

**FT-IR** (ATR, cm<sup>-1</sup>) υ 2224.

**HRMS** (ESI) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>24</sub>H<sub>16</sub>N<sub>3</sub><sup>+</sup>: 346.1338; found: 346.1341.

#### 1-(4-Nitrophenyl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3ac)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ chloroform/ethyl acetate gradient from 40:10:0.25 to 40:10:2) as a yellow solid; isolated yield: 80% (29.1 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.39 – 8.33 (m, 2H), 8.07 (dd, J = 8.0, 1.5 Hz, 1H), 8.01 – 7.95 (m, 2H), 7.77 – 7.71 (m, 2H), 7.60 – 7.52 (m, 3H), 7.42 (ddd, J = 8.0, 7.2, 1.2 Hz, 1H), 7.37 (dd, J = 8.5, 1.2 Hz, 1H), 7.21 (ddd, J = 8.5, 7.2, 1.5 Hz, 1H), 7.08 (d, J = 4.0 Hz, 1H), 6.88 (d, J = 4.0 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 155.0, 147.6, 140.6, 138.3, 137.8, 130.8, 130.2, 130.1, 129.9, 128.8, 128.7, 128.3, 127.6, 126.7, 125.7, 124.1, 118.5, 116.6, 109.7.

**FT-IR** (ATR, cm<sup>-1</sup>) υ 1334, 1523.

**HRMS** (ESI) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>16</sub>N<sub>3</sub>O<sub>2</sub><sup>+</sup>: 366.1237; found: 366.1246.

## 1-(4-Methoxyphenyl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3ad)



Prepared according to the general procedure but extended the reaction time to 24 h. The product was isolated by column chromatography on silica gel (eluent: hexanes/ethyl acetate 20:1) as pale yellow solid; isolated yield: 53% (18.5 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (dd, J = 8.0, 1.6 Hz, 1H), 8.00 – 7.98 (m, 2H), 7.58 – 7.51 (m, 3H), 7.51 – 7.45 (m, 3H), 7.36 (ddd, J = 8.0, 7.2, 1.3 Hz, 1H), 7.14 (ddd, J =

8.6, 7.2, 1.6 Hz, 1H), 7.07 – 7.03 (m, 2H), 7.02 (d, *J* = 4.0 Hz, 1H), 6.74 (d, *J* = 4.0 Hz, 1H), 3.92 (s, 3H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  160.1, 155.0, 138.7, 137.7, 132.9, 131.1, 130.3, 129.8, 128.9, 128.7, 128.5, 126.7, 126.2, 125.0, 116.8, 116.5, 114.3, 108.9, 55.5. One carbon signal could not be located.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{24}H_{19}N_2O^+$ : 351.1492; found: 351.1493.

#### 1-(4-Methylphenyl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3ae)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ethyl acetate/acid acetic 20:1:1) as a pale yellow solid; isolated yield: 74% (24.7 mg); purity: 80%. The product contained 20% of inseparable phenylation product (presumably from PPh<sub>3</sub>), thus the true yield of **3ae** should be 60%.

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (dd, J = 8.0, 1.6 Hz, 1H), 8.01 – 7.98 (m, 2H), 7.57 – 7.52 (m, 3H), 7.49 (dd, J = 8.6, 1.2 Hz, 1H), 7.46 – 7.43 (m, 2H), 7.38 – 7.34 (m, 1H), 7.32 (d, J = 7.2 Hz, 1H), 7.13 (ddd, J = 8.6, 7.2, 1.6 Hz, 1H), 7.02 (d, J = 4.0 Hz, 1H), 6.76 (d, J = 4.0 Hz, 1H), 2.49 (s, 3H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) due to overlapping of signals, possible peaks are listed δ 155.0, 138.7, 138.6, 137.7, 133.1, 131.5, 130.3, 129.82, 129.78, 129.7, 129.6, 128.9, 128.7, 128.4, 126.8, 126.2, 125.1, 116.9, 116.6, 109.0, 21.6.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{24}H_{19}N_2^+$ : 335.1542; found: 335.1542.

The NMR data reported herein matched with those known.<sup>2</sup>

## 4-Phenyl-1-(4-(trifluoromethyl)phenyl)pyrrolo[1,2-*a*]quinoxaline (3af)



Prepared according to the general procedure and isolated by column chromatography on caffeinated silica gel (eluent: hexanes/ ethyl acetate gradient from 100:1 to 10:1) as a yellow solid; isolated yield: 65% (25.2 mg).

<sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.05 (dd, J = 7.9, 1.6 Hz, 1H), 8.01 – 7.98 (m, 2H), 7.78 (d, J = 8.3 Hz, 2H), 7.70 (d, J = 8.3 Hz, 2H), 7.59 – 7.52 (m, 3H), 7.45 – 7.36 (m, 2H), 7.19 (ddd, J = 8.6, 7.1, 1.6 Hz, 1H), 7.06 (d, J = 4.1 Hz, 1H), 6.82 (d, J = 4.1 Hz, 1H). <sup>13</sup>C{<sup>1</sup>H} **NMR** (125 MHz, CDCl<sub>3</sub>)  $\delta$  155.0, 138.5, 137.9, 137.7, 131.1, 130.65, 130.60 (q,  $J_{C-F}$  = 32.9 Hz), 130.0, 129.8, 128.9, 128.8, 127.9, 127.61, 126.60, 125.8 (q,  $J_{C-F}$  =

3.5 Hz), 125.5, 124.2 (q,  $J_{C-F}$  = 272.2 Hz), 117.8, 116.5, 109.4.

<sup>19</sup>F NMR (471 MHz, CDCl<sub>3</sub>)  $\delta$  -62.52.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{24}H_{16}N_2F_3^+$ : 389.1260; found: 389.1263.

## 1-(4-Fluorophenyl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3ag)



Prepared according to the general procedure and isolated by column chromatography on caffeinated silica gel (eluent: hexanes/ethyl acetate 20:1) as a yellow solid; isolated yield: 67% (22.8 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.03 (dd, J = 7.9, 1.6 Hz, 1H), 8.00 – 7.98 (m, 2H), 7.58 – 7.50 (m, 5H), 7.41 – 7.35 (m, 2H), 7.24 – 7.19 (m, 2H), 7.15 (ddd, J = 8.6, 7.2, 1.6 Hz, 1H), 7.03 (d, J = 4.1 Hz, 1H), 6.76 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  163.1 (d,  $J_{C-F} = 249.2$  Hz), 155.0, 138.6, 137.71, 131.72, 131.65 (d,  $J_{C-F} = 8.2$  Hz), 130.49, 130.45, 129.87, 128.85, 128.7, 128.3, 127.0,

126.4, 125.2, 117.1, 116.3, 116.0 (d,  $J_{C-F}$  = 21.9 Hz), 109.0. One coupling constant could not be determined due to overlapping and complexity of peaks.

<sup>19</sup>**F NMR** (471 MHz, CDCl<sub>3</sub>) δ -112.42.

**HRMS** (ESI) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>16</sub>N<sub>2</sub>F<sup>+</sup>: 339.1292; found: 339.1292.

## 4-(4-Phenylpyrrolo[1,2-*a*]quinoxalin-1-yl)benzaldehyde (3ah)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ chloroform 1/1.25) as a yellow solid; isolated yield: 63% (21.8 mg); purity: 84%. The product presumably contained 16% of inseparable C3-arylation product, thus the true yield should be 53%.

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  10.13 (s, 1H), 8.06 (d, J = 8.2 Hz, 1H), 8.02 (d, J = 8.2 Hz, 2H), 8.00 – 7.98 (m, 2H), 7.75 (d, J = 7.7 Hz, 2H), 7.58 – 7.53 (m, 3H), 7.44 – 7.39 (m, 2H), 7.20 – 7.14 (m, 1H), 7.07 (d, J = 4.1 Hz, 1H), 6.86 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) due to mixture of inseparable isomers, possible peaks are listed δ 191.7, 191.6, 155.0, 140.2, 138.4, 137.8, 136.0, 131.3, 130.6, 130.4, 130.1, 130.0, 129.9, 129.4, 128.9, 128.8, 128.2, 127.9, 127.9, 126.6, 125.6, 118.1, 116.7, 109.5.

**FT-IR** (ATR, cm<sup>-1</sup>) υ 1693, 1599.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{24}H_{17}N_2O^+$ : 349.1335; found: 349.1339.

## 1-(3,5-Bis(trifluoromethyl)phenyl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3ai)



Prepared according to the general procedure and isolated by column chromatography on caffeinated silica gel (eluent: hexane/dichloromethane/acetic acid gradient from 100:2.5:1 to 100:20:1 for 1<sup>st</sup> column, then eluent: hexane/ethyl acetate/acetic acid gradient from 200:0.75:0.25 to 200:4.5:1.5 for 2<sup>nd</sup> column) as yellow solid; isolated yield: 59% (27.2 mg); purity: 90%. The product presumably contained 10% of inseparable diarylated product, thus the true yield should be 51%.

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.09 (dd, J = 8.0, 1.5 Hz, 1H), 8.05 (d, J = 1.7 Hz, 2H), 8.02 – 7.95 (m, 3H), 7.61 – 7.52 (m, 3H), 7.44 (ddd, J = 8.0, 7.1, 1.5 Hz, 1H), 7.28 (dd, J = 8.5, 1.5 Hz, 1H), 7.22 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.09 (d, J = 4.1 Hz, 1H), 6.90 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) due to overlapping and complexity of peaks, only possible coupling constants are listed  $\delta$  155.0, 138.3, 137.8, 136.3, 132.3 (q,  $J_{C-F}$  = 33.6 Hz), 131.0, 130.1, 129.4, 129.4, 129.1, 129.1, 128.9, 128.8, 128.1, 128.1, 127.6, 126.8, 125.9, 123.2 (q,  $J_{C-F}$  = 273.2 Hz), 122.1, 122.0, 118.4, 116.1, 109.6.

<sup>19</sup>F NMR (471 MHz, CDCl<sub>3</sub>) δ -62.93.

**HRMS** (ESI) m/z: [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>15</sub>F<sub>6</sub>N<sub>2</sub><sup>+</sup>: 457.11339, found: 457.11417.

## 1-(Naphthalen-2-yl)-4-phenylpyrrolo[1,2-a]quinoxaline (3aj)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/dichloromethane/acetic acid gradient from 50:2.5:10 to 50:10:10 for 1<sup>st</sup> column), then eluent: hexanes/acetone 20:1 for 2<sup>nd</sup> column on caffeinated silica gel, then eluent: toluene 100% for 3<sup>rd</sup> column) as a yellow solid; isolated yield: 32% <sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.09 (s, 1H), 8.08 – 8.02 (m, 3H), 7.96 (dd, J = 9.0, 2.6 Hz, 2H), 7.94 – 7.90 (m, 1H), 7.63 (dd, J = 8.6, 1.7 Hz, 1H), 7.61 – 7.54 (m, 5H), 7.51 (d, J = 8.6 Hz, 1H), 7.39 – 7.33 (m, 1H), 7.09 (d, J = 4.0 Hz, 1H), 7.09 – 7.05 (m, 1H), 6.88 (d, J = 4.0 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  155.0, 138.7, 137.8, 133.5, 133.2, 132.8, 131.8, 130.3, 129.8, 128.9, 128.7, 128.6, 128.37, 128.36, 128.0, 127.4, 127.1, 126.9, 126.3, 125.2, 117.4, 116.8, 109.2. Two carbon signals could not be located.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{27}H_{19}N_2^+$ : 371.1542; found: 371.1552.

## 4-Phenyl-1-(pyridin-3-yl)pyrrolo[1,2-*a*]quinoxaline (3ak)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ ethyl acetate/triethylamine gradient from 40:1:0.8 to 5:1:0.1) as a yellow solid; isolated yield: 40% (12.8 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.86 (dd, J = 2.3, 0.9 Hz, 1H), 8.75 (dd, J = 4.9, 1.6 Hz, 1H), 8.05 (dd, J = 8.0, 1.6 Hz, 1H), 8.01 – 7.97 (m, 2H), 7.87 (dt, J = 8.0, 2.3 Hz, 1H), 7.59 – 7.52 (m, 3H), 7.45 (ddd, J = 7.8, 4.8, 0.9 Hz, 1H), 7.41 – 7.37 (m, 2H), 7.17 (ddd, J = 8.6, 7.1, 1.6 Hz, 1H), 7.07 (d, J = 4.1 Hz, 1H), 6.83 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 155.0, 150.2, 149.8, 138.5, 137.7, 136.9, 130.7, 130.6, 130.0, 128.9, 128.84, 128.75, 128.1, 127.7, 126.7, 125.5, 123.5, 117.9, 116.2, 109.3.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{22}H_{16}N_3^+$ : 322.1339; found: 322.1351.

## 1-(Isoquinolin-4-yl)-4-phenylpyrrolo[1,2-*a*]quinoxaline (3al)



Prepared according to the general procedure and isolated by column chromatography on caffeinated silica gel (eluent: hexanes/ ethyl acetate gradient from 5:1 to 2:1) as a yellow solid; isolated yield: 21% (7.8 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  9.54 (s, 1H), 8.82 (s, 1H), 8.22 (d, J = 8.2 Hz, 1H), 8.16 – 8.12 (m, 2H), 8.11 (dd, J = 8.0, 1.5 Hz, 1H), 7.74 (ddd, J = 8.0, 6.8, 1.1 Hz, 1H), 7.71 – 7.63 (m, 4H), 7.51 (d, J = 8.4 Hz, 1H), 7.37 (ddd, J = 8.2, 7.2, 1.3 Hz, 1H), 7.26 (d, J = 4.0 Hz, 1H), 7.07 (dd, J = 8.6, 1.3 Hz, 1H), 7.00 (d, J = 4.0 Hz, 1H), 6.97 (ddd, J = 8.6, 7.1, 1.5 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 154.9, 153.8, 144.4, 138.5, 137.4, 135.5, 131.6, 130.4, 130.0, 128.9, 128.8, 128.4, 128.3, 128.2, 128.1, 127.5, 127.0, 126.9, 126.1, 125.3, 125.1, 118.5, 115.9, 109.1.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{26}H_{18}N_3^+$ : 372.1495; found: 372.1506.

#### 7-Chloro-1,4-diphenylpyrrolo[1,2-*a*]quinoxaline (3ba)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ dichloromethane 10:1 for 1<sup>st</sup> column, then eluent: hexanes/ acetone 50:1 for 2<sup>nd</sup> column on caffeinated silica gel) as a yellow solid; isolated yield: 70% (24.8 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.01 (d, J = 2.5 Hz, 1H), 8.00 – 7.98 (m, 2H), 7.58 – 7.54 (m, 3H), 7.54 – 7.51 (m, 4H), 7.35 (d, J = 9.1 Hz, 1H), 7.08 – 7.06 (m, 2H), 6.80 (d, J = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 155.9, 138.8, 138.3, 134.0, 133.2, 130.2, 130.1, 129.7, 129.5, 129.03, 129.00, 128.8, 128.7, 126.9, 126.7, 126.2, 117.6, 117.2, 109.7.
HRMS (ESI) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>16</sub>N<sub>2</sub>Cl<sup>+</sup>: 355.0997; found: 355.1008.

## 1,4-Diphenylpyrrolo[1,2-*a*]quinoxaline-7-carbonitrile (3ca)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ toluene/ acid acetic/ ethyl acetate gradient from 20:80:0.5:1 to 100:0:0.5:20) as a yellow solid; isolated yield: 70% (24.2 mg).

<sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) δ 8.30 (d, *J* = 2.0 Hz, 1H), 8.00 – 7.97 (m, 2H), 7.59 – 7.56 (m, 3H), 7.56 – 7.52 (m, 5H), 7.47 (d, *J* = 8.8 Hz, 1H), 7.33 (dd, *J* = 8.8, 2.0 Hz, 1H), 7.13 (d, *J* = 4.0 Hz, 1H), 6.86 (d, *J* = 4.0 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 156.7, 137.9, 137.7, 134.6, 134.0, 133.5, 131.3, 130.4, 129.7, 129.4, 129.3, 128.9, 128.8, 128.7, 126.9, 118.6, 118.2, 117.5, 110.6, 108.5.

**FT-IR** (ATR, cm<sup>-1</sup>) υ 2229.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{24}H_{16}N_3^+$ : 346.1339; found: 346.1351.

## 7-Fluoro-1,4-diphenylpyrrolo[1,2-*a*]quinoxaline (3da)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/toluene = 1:1 for  $1^{st}$  column, then eluent: hexanes/acetic acid 4:1 for  $2^{nd}$  column) as a yellow solid; isolated yield: 76% (25.7 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>) δ 8.01 – 7.97 (m, 2H), 7.69 (dd, *J* = 9.4, 3.0 Hz, 1H), 7.59 – 7.54 (m, 5H), 7.53 – 7.51 (m, 3H), 7.39 (dd, *J* = 9.4, 5.3 Hz, 1H), 7.06 (d, *J* = 4.1 Hz, 1H), 6.86 (ddd, *J* = 9.4, 7.7, 3.0 Hz, 1H), 6.78 (d, *J* = 4.1 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  159.6 (d,  $J_{C-F}$  = 244.4 Hz), 156.0, 139.2, 139.1, 138.3, 134.1, 133.0, 130.0, 129.7, 129.0, 128.9, 128.8, 128.7, 126.6, 124.9 (d,  $J_{C-F}$  = 2.5

Hz), 117.6 (d,  $J_{C-F}$  = 8.8 Hz), 117.0, 115.3 (d,  $J_{C-F}$  = 21.4 Hz), 113.8 (d,  $J_{C-F}$  = 22.7 Hz), 109.5. One coupling signal could not be determined due to overlapping and complexity of peaks.

<sup>19</sup>**F NMR** (471 MHz, CDCl<sub>3</sub>) δ -117.05.

**HRMS** (ESI) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>16</sub>N<sub>2</sub>F<sup>+</sup>: 339.1292; found: 339.1289.

### Methyl 1,4-diphenylpyrrolo[1,2-*a*]quinoxaline-7-carboxylate (3ea)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ ethyl acetate 12:1) as a yellow solid; isolated yield: 52% (19.5 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.71 (d, J = 2.0 Hz, 1H), 8.00 (dd, J = 7.7, 2.0 Hz, 2H), 7.78 (dd, J = 8.9, 2.0 Hz, 1H), 7.58 – 7.51 (m, 8H), 7.45 (d, J = 8.9 Hz, 1H), 7.09 (d, J= 4.1 Hz, 1H), 6.83 (d, J = 4.1 Hz, 1H), 3.93 (s, 3H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 166.6, 155.7, 138.3, 137.3, 133.9, 133.6, 132.1, 131.4, 130.1, 129.8, 129.1, 129.0, 128.85, 128.76, 127.1, 127.0, 126.8, 117.7, 116.6, 109.8, 52.3.

**FT-IR** (ATR, cm<sup>-1</sup>) υ 1715.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{25}H_{19}N_2O_2^+$ : 379.1441; found: 379.1443.

4-(Naphthalen-2-yl)-1-phenylpyrrolo[1,2-a]quinoxaline (3fa)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/acetone 20:1) as a yellow solid; isolated yield: 45% (15.8 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.52 (s, 1H), 8.12 (dd, J = 8.5, 1.6 Hz, 1H), 8.07 (dd, J = 8.0, 1.6 Hz, 1H), 8.03 (d, J = 8.5 Hz, 1H), 8.01 – 7.98 (m, 1H), 7.96 – 7.93 (m, 1H), 7.60 – 7.56 (m, 4H), 7.56 – 7.51 (m, 3H), 7.46 (dd, J = 8.5, 1.3 Hz, 1H), 7.39 (ddd, J = 8.0, 7.2, 1.3 Hz, 1H), 7.14 (ddd, J = 8.5, 7.2, 1.6 Hz, 1H), 7.12 (d, J = 4.0 Hz, 1H), 6.82 (d, J = 4.0 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 154.9, 137.8, 136.1, 134.4, 134.2, 133.3, 133.0, 130.4, 129.8, 128.89, 128.86, 128.8, 128.6, 128.5, 128.3, 127.9, 127.1, 127.0, 126.5, 126.4, 126.3, 125.2, 117.1, 116.7, 109.1.

**HRMS** (ESI) m/z:  $[M+H]^+$  calcd for  $C_{27}H_{19}N_2^+$ : 371.1542; found: 371.1542.

#### 7-Methyl-1-phenyl-4-(o-tolyl)pyrrolo[1,2-a]quinoxaline (3ga)



Prepared according to the general procedure and isolated by column chromatography on silica gel (eluent: hexanes/ toluene/acetic acid 4:4:1) as a yellow solid; isolated yield: 86% (30.4 mg).

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.81 (d, J = 2.1 Hz, 1H), 7.59 – 7.55 (m, 2H), 7.53 – 7.49 (m, 4H), 7.41 – 7.31 (m, 4H), 6.98 (dd, J = 8.6, 2.1 Hz, 1H), 6.71 (d, J = 4.0 Hz, 1H), 6.59 (d, J = 4.0 Hz, 1H), 2.43 (s, 3H), 2.37 (s, 3H).

<sup>13</sup>C{<sup>1</sup>H} NMR (125 MHz, CDCl<sub>3</sub>) δ 156.0, 137.8, 137.5, 136.7, 134.9, 134.4, 132.5, 130.8, 130.1, 129.9, 129.1, 129.0, 128.8, 128.7, 127.7, 127.5, 126.3, 125.7, 116.6, 116.4, 108.8, 21.0, 19.9.

**HRMS** (ESI) m/z: [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>21</sub>N<sub>2</sub><sup>+</sup>: 349.1699; found: 349.1698.

# X-ray Crystallography of 3ac

Good quality single crystals of **3ac** suitable for X-ray structure analysis were obtained by slow evaporation of hexanes into a saturated solution of **3ac** in dichloromethane. The intensities for the X-ray determinations of **3ac** were collected on a Bruker D8-VENTURE instrument with CuK $\alpha$  radiation ( $\lambda = 1.54178$  Å) at 298 K. Standard procedures were applied for data reduction and absorption correction. Using OLEX,<sup>3</sup> the structure solution and refinement were performed with SHELXT and SHELXL.<sup>4,5</sup> Hydrogen atom positions were calculated for idealized positions and treated with the 'riding mode' option of SHELXL. More details on data collections and structure calculations are contained in Table 1. Additional information on the structure determinations has been deposited with the Cambridge Crystallographic Data Centre with CCDC code: 2414626.



Table 1 Crystal data and structure refinement for **3ac**.

Empirical formula	$C_{23}H_{15}N_{3}O_{2}$
Formula weight	365.38
Crystal system	monoclinic
Space group	$P2_1/n$
a/Å	12.3695(2)
b/Å	7.37850(10)
c/Å	18.8523(4)

$\alpha/\circ$	90
β/°	92.7020(10)
$\gamma/^{\circ}$	90
Volume/Å <sup>3</sup>	1718.71(5)
Z	4
$ ho_{calc}g/cm^3$	1.412
µ/mm <sup>-1</sup>	0.746
F(000)	760.0
Crystal size/mm <sup>3</sup>	0.15  imes 0.12  imes 0.1
Index ranges	$-14 \le h \le 14, -8 \le k \le 8, -22 \le l \le 22$
Reflections collected	22227
Independent reflections	2932 [ $R_{int} = 0.0595, R_{sigma} = 0.0280$ ]
Data/restraints/parameters	2932/0/254
Goodness-of-fit on F <sup>2</sup>	1.122
Final R indexes $[I \ge 2\sigma (I)]$	$]R_1 = 0.0598, wR_2 = 0.1201$
Final R indexes [all data]	$R_1 = 0.0940, wR_2 = 0.1506$

# References

1 (a) C. Xie, L. Feng, W. Li, X. Ma, X. Ma, Y. Liu and C. Ma, *Org. Biomol. Chem.*, 2016, **14**, 8529; (b) B. Budke, W. Tueckmantel, K. Miles, A. P. Kozikowski and P. P. Connell, *ChemMedChem*, 2019, **14**, 1031.

2 D. Hao, Z. Yang, Y. Liu, Y. Li, C. Li, Y. Gu, L. Vaccaro, J. Liu and P. Liu, *Org. Biomol. Chem.*, 2022, **20**, 847.

3 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, *J. Appl. Cryst.*, 2009, **42**, 339.

4 S. M. Sheldrick, Acta Cryst. A71, 2015, 3.

5 S. M. Sheldrick, *Acta Cryst. C71*, 2015, 3.







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S38







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