Supporting Information for

# pH-Activated NIR Fluorescent Probe for Sensitive Mitochondrial Viscosity Detection

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## **Experimental section**

#### Quantum yield calculation

The fluorescence quantum yield  $\Phi_s$  was estimated from the absorption and fluorescence spectra of probe according to equation, where the subscript s and r stand for the sample and reference (fluorescein as standard), respectively.  $\Phi$  is the quantum yields, A represents the absorbance at the excitation wavelength, S refers to the integrated emission band areas and  $n_D$  is the solvent refractive index. The fluorescence quantum yields ( $\Phi_F$ ) were estimated with equation as follows:

$$arphi_{s}=arphi_{r}rac{S_{s}A_{r}n_{DS}^{2}}{S_{R}A_{S}n_{Dr}^{2}}$$

#### **Computational methodology**

Density functional theory (DFT) calculations were employed using the B3LYP functional and  $6-31+G(d, p)^*$  basis set for all calculations. In the calculation, except for the specific instructions, other atom free optimization, the dihedral angle of bond length and bond angle are not fixed, and all optimization is carried out in vacuum environment by default.

### Cytotoxicity of probe SSN

The cytotoxicity of probe **SSN** was tested by MTT method. HepG2 cells were seeded at a density at  $1 \times 10^4$  cells per well into 96-well plate, and incubated in 37 °C cell incubator (containing 5% CO<sub>2</sub>) for 12 hours. The culture medium was high glucose DMEM with fetal bovine serum and appropriate antibodies (penicillin and streptomycin). Then the probe **SSN** (0, 1, 2, 5, 10, 20  $\mu$ M) was added and incubated for 24 h. 50  $\mu$ L MTT was added to each pore and the cells were incubated at 37 °C and 5% CO<sub>2</sub> for 4 h. Then, removed the medium and replaced it with DMSO (150 $\mu$ L), and detected the absorption values at 540 nm.

## Cell culture and fluorescence imaging

DMEM containing 10% fetal bovine serum and 1% penicillin was used for HepG2 cell culture in an incubator supplemented with 95% air and 5% CO<sub>2</sub> at 37 °C. Then, nutrient solution was removed and cells were washed three times with PBS buffer (pH=7.0, 10 mM) before imaging. To compare the difference in viscosity level of

HepG2 cells, they were treated with 5  $\mu$ M SSN for 30 min and then washed three times with PBS buffer.

## **Imaging in zebrafish**

The 3-day-old zebrafish was incubated with SSN (10  $\mu$ M) for 2 h, and then washed with PBS buffer and imaged as control group. The 3-day-old zebrafish firstly incubated with nystatin (50 and 100 mM, respectively) for 8 h, followed by SSN incubation for 2 h. Thereafter, the treated zebrafish was washed with PBS buffer three times and imaged using a confocal microscope ( $\lambda_{ex}/\lambda_{em} = 561/700-760$  nm).



Scheme S1. Synthetic routine for SSN.

## **Synthesis of Compound 1:**

5-Bromo-2-hydroxybenzaldehyde (402 mg, 2 mmol), 2-aminothiophenol (300 mg, 2.4 mmol), and Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> (760 mg, 4 mmol) were added to a 200 mL round-bottom flask. The mixture was dissolved in 20 mL of DMF. Subsequently, the reaction was refluxed at 140°C for 2 hours. After cooling the mixture to room temperature, water was added, and the resulting solution was filtered to obtain a yellow solid compound 5 (367 mg, 60% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  11.71 (s, 1H), 8.39 (d, *J* = 2.4 Hz, 1H), 8.12 (dd, *J* = 27.5, 8.0 Hz, 2H), 7.56 (dd, *J* = 6.2, 2.5 Hz, 2H), 7.47 (t, *J* = 7.5 Hz, 1H), 7.07 (d, *J* = 8.8 Hz, 1H).



Fig.S1 Absorption spectra of SSN (5  $\mu$ M) in different ratios of PBS/glycerol mixtures.



Fig. S2 The fluorescence spectra of probe SSN (5  $\mu$ M) in different solvents.



**Fig. S3** Fluorescence intensity at 760 nm of 5  $\mu$ M **SSN** toward various species (100  $\mu$ M) in PBS/glycerol (v:v = 1:1) mixture. (1) Blank; (2)Ser; (3)Phe; (4)Arg; (5)Cys; (6)GSH; (7)Hcy; (8)BSA; (9)SO<sub>3</sub><sup>2-</sup>; (10)SO<sub>4</sub><sup>2-</sup>; (11)ClO<sup>-</sup>; (12)ClO<sub>4</sub><sup>-</sup>; (13)H<sub>2</sub>O<sub>2</sub>; (14)ONOO<sup>-</sup>; (15)CO<sub>3</sub><sup>2-</sup>; (16)OAc<sup>-</sup>; (17) 50% Glyceral. The excitation wavelength was 520 nm.



Fig.S4 Absorption spectra of SSN (5  $\mu$ M) in different pH buffers.



Fig.S5 Absorption spectra of SSN (5  $\mu$ M) in different ratios of PBS/glycerol mixtures in pH =6.2 and pH =8.6, respectively.



Fig. S6 Photobleaching of SSN (5  $\mu$ M) in 95% glycerol solution under irradiation with660nmlaser(300mW/cm<sup>2</sup>).



**Fig. S7** Calculated electron-hole distributions during photoexcitation of **SSN**. The geometry of **SSN** and **SSN-K** is based on excited-state optimized structures.



Fig. S8 MTT results of HepG2 cells viabilities after incubation with SSN for 24 h. Data are expressed as mean  $\pm$  SD (\*p < 0.05, experiment times n = 3).



Fig. S9 Confocal HepG2 cell images co-stained by SSN (5  $\mu$ M) and MTG (0.2  $\mu$ M). (a) Image of MTG ( $\lambda_{em}$ : 500–550 nm); (b) Image of SSN ( $\lambda_{em}$ : 720–800 nm). (c) merged images with bright-field image. (d) Intensity scatter plot with Pearson's coefficient indicated (PC = 0.9612). Scale bar = 10  $\mu$ m. Data are expressed as mean ± SD (\*\*\* P<0.001, experiment times n = 3).



Fig. S10 Laser Scanning Confocal Microscopy (LSCM) images of HepG2 cells incubated with SSN (5  $\mu$ M), captured at various time intervals. (a-n) Red channel; (o) BF; (p) Merged images. The scale bar = 20  $\mu$ m.









Fig. S12 <sup>1</sup>H NMR spectrum of compound 1 in DMSO- $d_6$ .



Fig. S14 <sup>1</sup>H NMR spectrum of SSN in DMSO-*d*<sub>6</sub>.



Fig. S15 <sup>1</sup>C NMR spectrum of SSN in DMSO-*d*<sub>6</sub>.