

## Supporting Information

### Organocatalytic Asymmetric Synthesis of Axially and Centrally Chiral Heterotriarylmethanes by Friedel-Crafts Reaction

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#### Table of Contents

1. General information .....	S2
2. Methods of synthesizing substrates.....	S3
2.1 Methods of synthesizing 1 <i>H</i> -pyrrol-3-yl carbinol <b>1</b> .....	S3
2.2 Methods of synthesizing 3-(2-methoxynaphthalen-1-yl)-1 <i>H</i> -indole <b>2</b> .....	S7
3. Optimization of reaction conditions.....	S14
4. Copies of <sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>19</sup> F NMR Spectra.....	S19
5. X-Ray single crystal data for product ( <i>S</i> , <i>S</i> )- <b>3b</b> .....	S61
6. Copies of HPLC Spectra .....	S63
7. References.....	S98

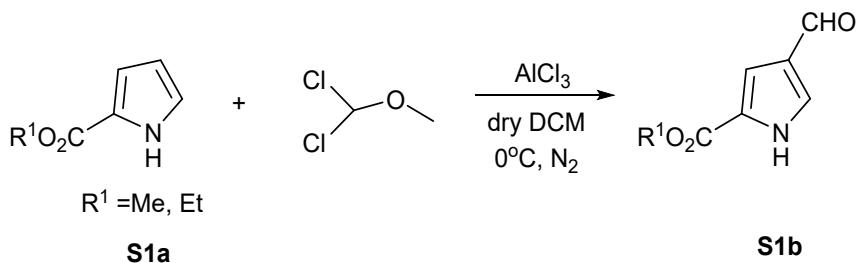
## 1. General information

All solvents and reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Analytical thin layer chromatography (TLC) was performed on precoated silica gel 50 GF254 plates. Flash column chromatography was performed using silica gel (200-300 mesh). Visualization on TLC was achieved by use of UV light (254, 365nm). NMR spectrums were recorded on a Bruker DPX 400 NMR spectrometer at 400 MHz for <sup>1</sup>H NMR, 101 MHz for <sup>13</sup>C NMR. The solvent used for NMR spectroscopy was CDCl<sub>3</sub>. Chemical shifts for <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were reported as  $\delta$  in units of parts per million (ppm) downfield from standard tetramethylsilane (0.0), relative to the signal of chloroform-d. Multiplicities were given as s (singlet), d (doublet), t (triplet), dd (doublets of doublet), or m (multiplets). The number of protons (n) for a given resonance is indicated by nH. Coupling constants were reported as a *J* value in hertz. A high resolution mass spectrum (HRMS) was determined by 1290II-6230 TOF using ESI ionization. Infrared spectra were recorded on an ATR-FTIR spectrometer (NICOLET iS10). Optical rotations were reported as follows:  $[\alpha]_D^{20}$  (c: g/100 mL, in DCM). Enantiomeric excess was determined by chiral high-performance liquid chromatography (chiral HPLC) using DAICEL CHIRALPAK columns such as IA, AD-H, IE-3, IC-3, and IB-3. The melting point of each compound was determined by melting point meter SGW X-4A. The racemic products employed to determine enantiomeric ratios were prepared by using diphenylphosphate as a catalyst. Optical rotation values were measured with instruments operating at  $\lambda = 589$  nm, corresponding to the sodium D line at the temperatures indicated. The X-ray source used for the single crystal X-ray diffraction analysis of compound **3b** was GaK $\alpha$  ( $\lambda = 1.34139$ ). The thermal ellipsoid was drawn at the 50% probability level.

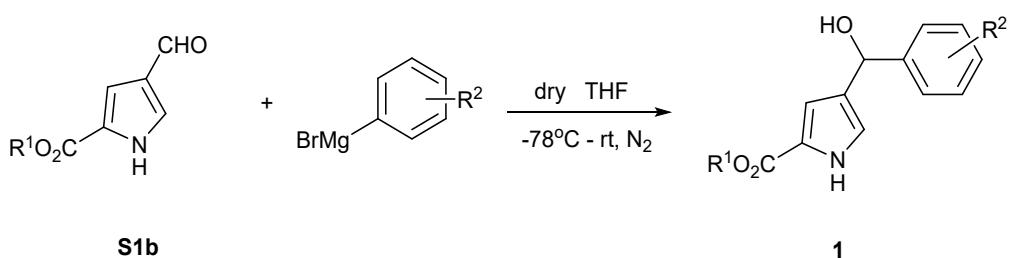
## 2. Methods of synthesizing substrates

### 2.1 Methods of synthesizing 1H-pyrrol-3-yl carbinol 1

4-(Hydroxy(phenyl)methyl)-1H-pyrrole-2-carboxylate derivatives used in the synthesis of substrates 1 were performed by the following method.<sup>1</sup>

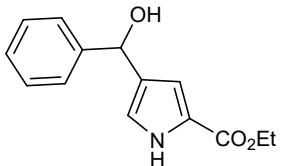


Under a nitrogen atmosphere, 1,1-dichlorodimethyl ether (60 mmol, 1.5 equiv, 6.9 g) was added dropwise to 1H-pyrrole-2-carboxylate derivative (40 mmol, 5.58 g, 1 equiv) and aluminum trichloride (3 equiv, 120 mmol, 16.04 g) dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (80 mL) in a three-neck flask. The reaction mixture was stirred at 0°C for 2 hours. Cold water was slowly added to the mixture in an ice bath until no more gas bubbles evolved. The reaction mixture was then extracted with dichloromethane (3 × 90 mL). The combined organic layer was washed with brine (60 mL) and dried over sodium sulfate. The solution was concentrated under reduced pressure and purified by flash chromatography (PE : EA = 1:8) to obtain **S1b**.



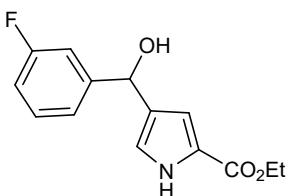
In a three-neck flask, a Grignard reagent (1M in THF, 66 mmol, 3 equiv, 66 mL) was added dropwise to dry THF containing 1H-pyrrole-3-carbaldehyde (20 mL) at -78°C. The mixture was then warmed to room temperature and stirred overnight under a nitrogen atmosphere. After cooling in an ice bath, the resulting mixture was quenched

with a saturated aqueous solution of NH<sub>4</sub>Cl (80 mL). The reaction mixture was then extracted with ethyl acetate (3 × 80 mL). The combined organic layer was washed with brine (60 mL) and dried over Na<sub>2</sub>SO<sub>4</sub>. The solution was concentrated in vacuo and purified by flash chromatography (PE : EA = 1:6) to obtain product **1**.



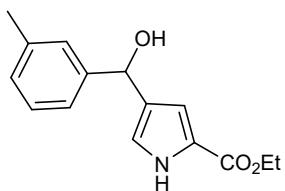
### **Ethyl 4-(hydroxy(phenyl)methyl)-1H-pyrrole-2-carboxylate (1a)**

Yellow solid (2.11g, 86% yield); MP = 102-104 °C; <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 9.87 (s, 1H), 7.33 (d, *J* = 7.0 Hz, 2H), 7.29 - 7.17 (m, 3H), 6.75 (d, *J* = 1.8 Hz, 1H), 6.65 (d, *J* = 2.1 Hz, 1H), 5.68 (s, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 3.55 (s, 1H), 1.23 (t, *J* = 7.1 Hz, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 161.8, 144.2, 129.5, 128.4, 127.4, 126.4, 122.9, 121.6, 113.9, 70.6, 60.5, 14.4 ppm. HRMS (ESI TOF) m/z: [M + Na]<sup>+</sup> calcd for C<sub>14</sub>H<sub>15</sub>NO<sub>3</sub>Na: 268.0945; Found: 268.0952. IR (KBr, cm<sup>-1</sup>): 3426, 3314, 2970, 2914, 2857, 1682, 1605, 1473, 1106, 1023, 828.



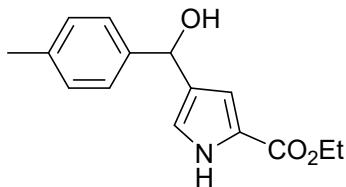
### **Ethyl 4-((3-fluorophenyl)(hydroxy)methyl)-1H-pyrrole-2-carboxylate (1b)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 9.54 (s, 1H), 7.32 – 7.23 (m, 1H), 7.19 – 7.08 (m, 2H), 6.94 (tt, *J* = 8.7, 1.8 Hz, 1H), 6.82 – 6.74 (m, 2H), 5.76 (s, 1H), 4.26 (q, *J* = 7.1 Hz, 2H), 2.87 (s, 1H), 1.31 (t, *J* = 7.1 Hz, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 171.3, 162.92 (d, *J* = 245.7 Hz), 161.3, 146.6, 129.86 (d, *J* = 8.2 Hz), 129.1, 123.3, 121.9, 121.0, 114.24 (d, *J* = 21.2 Hz), 113.5, 113.20 (d, *J* = 22.1 Hz), 60.5, 53.4, 14.4 ppm. <sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -112.92 ppm.



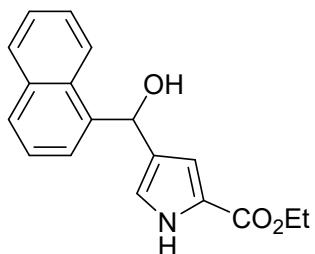
**Ethyl 4-(hydroxy(m-tolyl)methyl)-1H-pyrrole-2-carboxylate (1c)**

$^1\text{H}$  NMR (400 MHz, Chloroform-d)  $\delta$  9.09 (s, 1H), 7.37 – 7.27 (m, 1H), 7.23 – 7.14 (m, 2H), 7.08 (dd,  $J = 12.4, 6.7$  Hz, 2H), 6.83 (dt,  $J = 10.5, 2.2$  Hz, 2H), 5.77 (s, 1H), 4.28 (q, 2H), 2.35 (s, 3H), 1.32 (t,  $J = 7.1$  Hz, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  143.9, 138.1, 128.4, 127.0, 123.4, 120.6, 114.2, 113.5, 70.8, 60.4, 31.5, 30.2, 29.7, 21.5, 14.4 ppm.



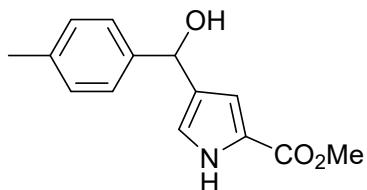
**Ethyl 4-(hydroxy(p-tolyl)methyl)-1H-pyrrole-2-carboxylate (1d)**

Yellow solid (2.33g, 90% yield); MP = 121-123 °C;  $^1\text{H}$  NMR (400 MHz, Chloroform-d)  $\delta$  9.08 (s, 1H), 7.31 – 7.15 (m, 3H), 7.09 (d,  $J = 7.7$  Hz, 2H), 6.74 (dd,  $J = 11.7, 2.5$  Hz, 2H), 5.70 (s, 1H), 4.20 (q, 2H), 2.27 (s, 3H), 1.20 (t,  $J = 11.9$  Hz, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  161.2, 141.0, 137.3, 129.9, 129.2, 126.3, 123.2, 120.7, 113.5, 70.7, 60.4, 31.5, 29.7, 21.1, 14.4 ppm. HRMS (ESI TOF) m/z:  $[\text{M} + \text{Na}]^+$  calcd for  $\text{C}_{15}\text{H}_{17}\text{NO}_3\text{Na}$ : 282.1101; Found: 282.1112. IR (KBr,  $\text{cm}^{-1}$ ): 3406, 3315, 2963, 2910, 2838, 1682, 1605, 1483, 1104, 1006, 847.



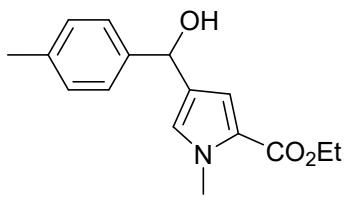
### **Ethyl 4-(hydroxy(p-tolyl)methyl)-1H-pyrrole-2-carboxylate (1e)**

Yellow solid (2.79g , 88% yield); MP = 129-130 °C; <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 9.35 – 9.24 (m, 1H), 7.88 (d, *J* = 1.8 Hz, 1H), 7.84 – 7.77 (m, 3H), 7.51 – 7.43 (m, 3H), 7.25 (m, 1H), 6.80 (q, *J* = 1.6 Hz, 2H), 5.94 (d, *J* = 2.5 Hz, 1H), 4.25 (q, *J* = 7.1 Hz, 2H), 1.29 (t, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 161.3, 141.3, 133.3, 132.9, 129.6, 128.3, 128.1, 127.7, 126.2, 125.9, 124.7, 124.7, 123.3, 121.0, 113.7, 70.8, 60.5, 14.4 ppm. HRMS (ESI TOF) m/z: [M+ Na] <sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>FNO<sub>3</sub>Na: 318.1101; Found: 318.1126. IR (KBr, cm-1): 3429, 3315, 2946, 1689, 1580, 1463, 1379, 1209, 1106, 1024, 825.



### **Methyl 4-(hydroxy(p-tolyl)methyl)-1H-pyrrole-2-carboxylate (1f)**

Yellow solid (1.98g, 81% yield); MP = 115-116 °C; <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 9.24 (s, 1H), 7.50 – 7.24 (m, 2H), 7.15 (d, *J* = 7.8 Hz, 3H), 6.85 – 6.73 (m, 2H), 5.76 (s, 1H), 3.80 (s, 3H), 2.34 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 161.7, 141.0, 137.3, 129.9, 129.2, 126.3, 120.9, 113.7, 70.6, 51.5, 31.5, 30.2, 29.7, 21.1 ppm. HRMS (ESI TOF) m/z: [M+ Na] <sup>+</sup> calcd for C<sub>14</sub>H<sub>15</sub>FNO<sub>3</sub>Na: 268.0944; Found: 268.0932. IR (KBr, cm-1): 3446, 3305, 2981, 1672, 1608, 1565, 1430, 1192, 1017, 792.



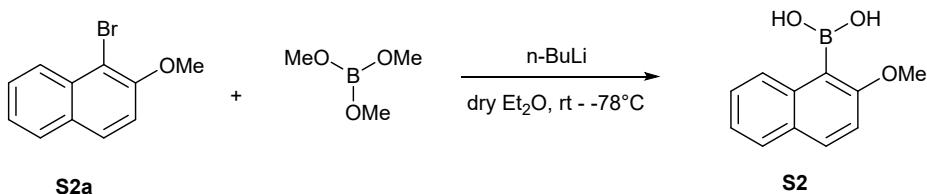
### **Ethyl 4-(hydroxy(p-tolyl)methyl)-1H-pyrrole-2-carboxylate (1g)**

Yellow solid (2.51g, 92% yield); MP = 128-129°C; <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.42 – 7.19 (m, 3H), 7.16 (d, *J* = 7.8 Hz, 2H), 6.84 (d, *J* = 2.0 Hz, 1H), 6.64 (d, *J* = 2.0 Hz, 1H), 5.71 (s, 1H), 4.23 (q, *J* = 7.1 Hz, 2H), 3.84 (s, 3H), 2.34 (s, 3H), 1.32 (t, *J* = 7.2 Hz, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 161.3, 141.1, 137.2, 129.1, 127.3, 126.2, 115.9, 70.6, 67.1, 59.9, 36.8, 31.5, 30.1, 29.7, 21.2, 14.4 ppm. HRMS (ESI TOF) m/z: [M+ Na] <sup>+</sup> calcd for C<sub>15</sub>H<sub>17</sub>FNO<sub>3</sub>Na: 282.1101; Found: 282.1152. IR (KBr, cm<sup>-1</sup>): 3426, 3312, 2952, 1678, 1604, 1506, 1461, 1217, 1014, 832, 736.

## 2.2 Methods of synthesizing naphthalyl-indole substrates 2

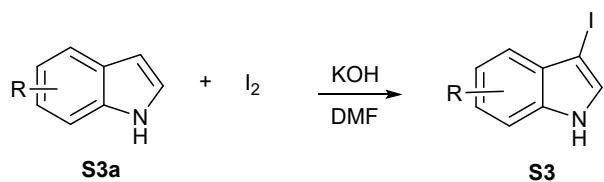
### 2.2.1 Methods of synthesizing boronic acid substrates

Boronic acid substrates used in the construction of substrates **2** were synthesized by the methods described in the literature.<sup>2</sup>



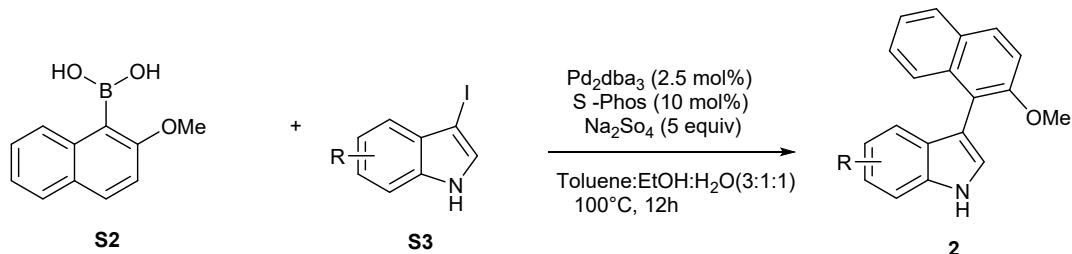
*n*-Butyllithium (7.0 mL, 2.5 M in hexanes, 18 mmol, 1.13 equiv.) was added dropwise to a stirred solution of 1-bromo-2-methoxynaphthalene derivatives (15.9 mmol, 1 equiv.) in dry  $\text{Et}_2\text{O}$  (60 mL) at  $-78^\circ\text{C}$  under nitrogen atmosphere. The solution was warmed to room temperature and stirred for 1 h. After this time, the flask was re-cooled to  $-78^\circ\text{C}$  and trimethylborate (2.3 mL, 21 mmol, 1.32 equiv.) was added drop-wise. The mixture was allowed warm to room temperature overnight (18 h) under nitrogen. 1M HCl (50 mL) was added dropwise and the solution was stirred for an additional 1h. The reaction mixture was evaporated in vacuo and extracted with dichloromethane ( $3 \times 30$  mL). The combined organic layer was washed with brine (30 mL) and dried over  $\text{Na}_2\text{SO}_4$ . The solution was concentrated in vacuo and purified by flash chromatography (PE : EA = 1:6) to get **S2**.

### 2.2.2 Methods of synthesizing 3-Iodoindole derivatives



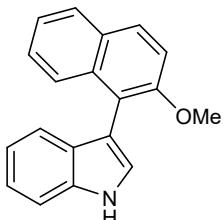
All 3-iodoindole substrates were synthesized according to the reported literature.<sup>3</sup> A solution of indole **S3a** (17.1 mmol, 1 equiv.) in 30 mL of DMF was treated with KOH (2.39 g, 42.7 mmol, 2.5 equiv.) and allowed to stir at room temperature for 20 min. I<sub>2</sub> (4.38 g, 17.1 mmol, 1 equiv.) was dissolved in 5 mL of DMF and added to the reaction. The resulting solution was stirred for an additional 45 min. The reaction mixture was poured into 400 mL of ice water and the precipitate was collected by filtration, washed with water and dried by azeotropic distillation with toluene to yield 3-iodoindole derivatives **S3**.

### 2.2.3 Methods of synthesizing naphthyl-indole substrates



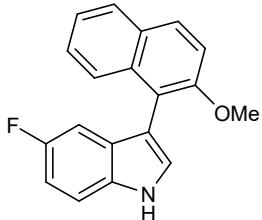
Substrates **2** were prepared according to the general procedure reported by Shi and coworkers.<sup>4</sup> Under nitrogen atmosphere, to the mixture of 3-iodoindole derivatives (10.4 mmol, 1 equiv.), naphthyl boronic acid derivatives (11.4 mmol, 1.1 equiv.), Pd<sub>2</sub>dba<sub>3</sub> (238 mg, 2.5 mol%), S-Phos (436 mg, 10 mol%) and Na<sub>2</sub>CO<sub>3</sub> (5.6 g, 52 mmol, 5 equiv.) was added the mixed solvent of toluene (60 mL), ethanol (20 mL) and H<sub>2</sub>O (20 mL) (3:1:1). Then, the reaction mixture was stirred at 100 °C in an oil bath for 12 h. After the completion of the reaction which was indicated by TLC, the reaction mixture was cooled to room temperature, and the reaction mixture was quenched with water and extracted with ethyl acetate. The combined organic layer was washed with brine and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Then, the organic layer was concentrated in

vacuo to give a residue, which was purified by flash column chromatography (petroleum ether/ethyl acetate = 10/1) to afford the desired compounds **2a-2i**. Compounds **2a-2i** were similar with the previously reported work<sup>4</sup>.



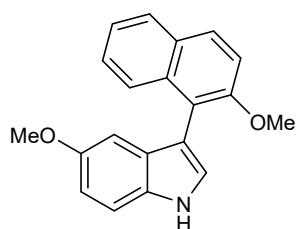
### **3-(2-methoxynaphthalen-1-yl)-1H-indole (2a)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.15 (s, 1H), 7.87 (d, *J* = 9.0 Hz, 2H), 7.82 (dd, *J* = 7.5, 1.5 Hz, 1H), 7.73 – 7.69 (m, 3H), 7.40 – 7.24 (m, 2H), 7.23 – 7.14 (m, 2H), 7.08 – 7.01 (m, 1H), 3.77 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 155.2, 136.1, 134.6, 129.4, 128.9, 128.2, 128.0, 126.2, 126.1, 124.8, 123.6, 122.0, 120.6, 119.7, 118.1, 114.1, 111.3, 110.8, 56.8 ppm.



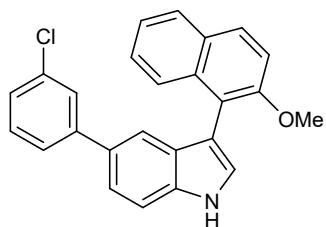
### **5-fluoro-3-(2-methoxynaphthalen-1-yl)-1H-indole (2b)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.18 (s, 1H), 7.87 (d, *J* = 9.1 Hz, 1H), 7.84 – 7.79 (m, 1H), 7.68 (dd, *J* = 7.5, 2.0 Hz, 1H), 7.37 (d, *J* = 9.0 Hz, 3H), 7.30 (tt, *J* = 6.8, 5.1 Hz, 2H), 7.20 (dd, *J* = 8.6, 3.7 Hz, 1H), 6.97 – 6.87 (m, 1H), 3.78 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 158.10 (d, *J* = 234.4 Hz), 155.1, 134.4, 132.6, 129.4, 129.2, 128.64 (d, *J* = 9.9 Hz), 128.1, 126.6, 126.4, 125.7, 123.7, 117.3, 113.8, 111.93 (d, *J* = 9.7 Hz), 111.05 (d, *J* = 4.9 Hz), 110.50 (d, *J* = 26.5 Hz), 105.38 (d, *J* = 23.4 Hz), 56.7 ppm. <sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -124.37 ppm.



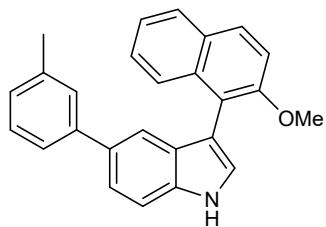
**5-methoxy-3-(2-methoxynaphthalen-1-yl)-1H-indole (2c)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.15 (s, 1H), 7.86 (d, *J* = 9.1 Hz, 1H), 7.83 – 7.79 (m, 1H), 7.72 (dd, *J* = 7.9, 1.8 Hz, 1H), 7.37 (d, *J* = 9.1 Hz, 1H), 7.33 – 7.25 (m, 2H), 7.21 – 7.09 (m, 2H), 6.85 (dd, *J* = 8.8, 2.4 Hz, 1H), 6.67 (d, *J* = 2.4 Hz, 1H), 3.77 (s, 3H), 3.63 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 155.2, 154.3, 134.5, 131.3, 129.4, 128.9, 128.6, 128.0, 126.2, 126.1, 125.6, 123.7, 118.2, 114.1, 112.5, 112.1, 110.5, 102.1, 56.8, 55.9 ppm.



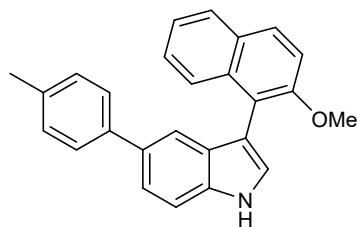
**5-(3-chlorophenyl)-3-(2-methoxynaphthalen-1-yl)-1H-indole (2d)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.31 (s, 1H), 7.90 (d, *J* = 9.0 Hz, 1H), 7.84 (dd, *J* = 7.5, 1.7 Hz, 1H), 7.72 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.50 (d, *J* = 1.9 Hz, 1H), 7.42 (d, *J* = 2.6 Hz, 4H), 7.40 – 7.26 (m, 4H), 7.25 – 7.14 (m, 2H), 3.81 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 155.3, 144.5, 135.9, 134.6, 134.4, 131.9, 129.8, 129.3, 129.1, 128.7, 128.0, 127.4, 126.3, 126.2, 125.9, 125.6, 123.7, 121.7, 119.1, 117.6, 113.9, 111.7, 111.4, 60.5, 56.8 ppm.



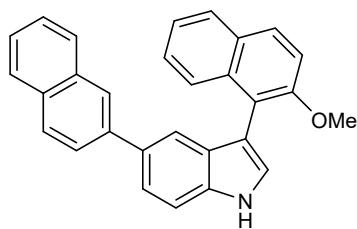
**3-(2-methoxynaphthalen-1-yl)-5-(m-tolyl)-1H-indole (2e)**

<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.16 (s, 1H), 7.87 (d, *J* = 9.0 Hz, 1H), 7.84 – 7.79 (m, 1H), 7.74 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.44 (d, *J* = 8.4 Hz, 2H), 7.41 – 7.23 (m, 6H), 7.22 – 7.13 (m, 2H), 7.06 – 7.01 (m, 1H), 3.77 (s, 3H), 2.31 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 155.3, 142.7, 138.1, 135.7, 134.7, 133.5, 129.4, 129.0, 128.7, 128.5, 128.3, 128.0, 127.1, 126.3, 126.1, 125.4, 124.6, 123.7, 122.0, 119.0, 118.0, 114.0, 111.6, 111.2, 56.8, 21.6 ppm.



**3-(2-methoxynaphthalen-1-yl)-5-(p-tolyl)-1H-indole (2f)**

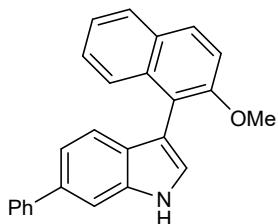
<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.28 (s, 1H), 7.89 (d, *J* = 9.0 Hz, 1H), 7.83 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.79 – 7.73 (m, 1H), 7.47 – 7.38 (m, 6H), 7.31 (qd, *J* = 8.4, 7.6, 1.4 Hz, 3H), 7.21 (s, 1H), 7.13 (d, *J* = 7.8 Hz, 1H), 3.81 (s, 3H), 2.32 (s, 3H) ppm. <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 155.3, 139.7, 135.8, 135.5, 134.6, 133.3, 129.3, 128.9, 128.7, 127.9, 127.3, 126.2, 126.0, 125.3, 123.6, 121.9, 118.8, 117.9, 114.0, 111.4, 111.3, 56.8, 21.0 ppm.



**3-(2-methoxynaphthalen-1-yl)-5-(naphthalen-2-yl)-1H-indole (2g)**

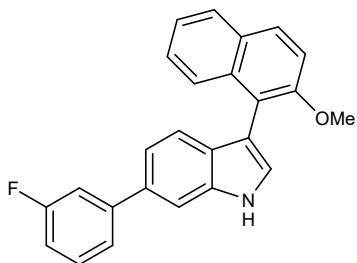
<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.22 (s, 1H), 7.94 (d, *J* = 1.8 Hz, 1H), 7.88 (d, *J* = 9.0 Hz, 1H), 7.82 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.79 – 7.73 (m, 4H), 7.67 (dd, *J* = 8.6, 1.8 Hz, 1H), 7.62 – 7.52 (m, 2H), 7.38 (dt, *J* = 9.2, 6.4 Hz, 4H), 7.34 – 7.24 (m, 2H), 7.20

– 7.13 (m, 1H), 3.77 (s, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  155.3, 140.0, 135.8, 134.7, 133.9, 133.2, 132.2, 129.4, 129.1, 128.8, 128.1, 128.0, 127.7, 126.4, 126.1, 126.0, 125.6, 125.5, 123.7, 122.2, 119.4, 117.9, 114.0, 111.7, 111.3, 60.6, 56.8, 21.2, 14.3 ppm.



### **3-(2-methoxynaphthalen-1-yl)-6-phenyl-1H-indole (2h)**

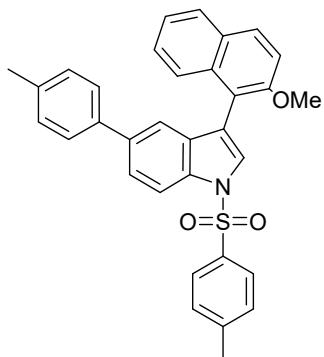
$^1\text{H}$  NMR (400 MHz, Chloroform-d)  $\delta$  8.04 (d,  $J = 2.5$  Hz, 1H), 7.88 – 7.77 (m, 2H), 7.72 (d,  $J = 8.6$  Hz, 1H), 7.56 – 7.51 (m, 2H), 7.45 – 7.39 (m, 1H), 7.36 – 7.20 (m, 8H), 7.07 (d,  $J = 2.4$  Hz, 1H), 3.71 (s, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  155.2, 142.4, 136.7, 135.5, 134.7, 129.4, 129.1, 128.8, 128.1, 127.6, 127.5, 126.7, 126.3, 126.1, 125.7, 123.7, 120.9, 119.7, 118.0, 114.0, 110.7, 109.9, 56.7 ppm.



### **6-(3-fluorophenyl)-3-(2-methoxynaphthalen-1-yl)-1H-indole (2i)**

$^1\text{H}$  NMR (400 MHz, Chloroform-d)  $\delta$  8.22 (s, 1H), 7.90 – 7.79 (m, 1H), 7.71 (dd,  $J = 8.3, 1.4$  Hz, 1H), 7.42 (t,  $J = 1.1$  Hz, 1H), 7.37 (d,  $J = 9.1$  Hz, 2H), 7.34 – 7.22 (m, 6H), 7.15 (d,  $J = 2.4$  Hz, 2H), 6.95 (dd,  $J = 8.8, 7.9, 2.7, 1.5$  Hz, 1H), 3.76 (s, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  163.29 (d,  $J = 244.9$  Hz), 155.2, 144.67 (d,  $J = 7.8$  Hz), 136.6, 134.6, 134.2, 129.4, 129.1, 128.1, 128.0, 126.3, 125.9, 123.7, 123.0,

120.22 (d,  $J = 146.8$  Hz), 117.7, 114.2, 114.0, 113.9, 110.8, 109.9, 60.6, 56.7, 21.2, 14.3 ppm.  $^{19}\text{F}$  NMR (376 MHz, Chloroform-d)  $\delta$  -113.41 ppm.

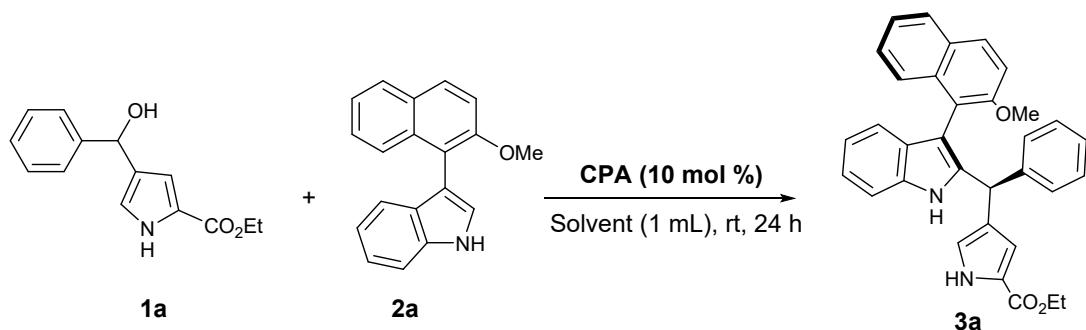


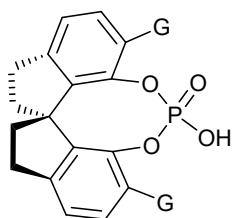
### 3-(2-methoxynaphthalen-1-yl)-5-(p-tolyl)-1-tosyl-1H-indole (2j)

White solid (2.51g, 85% yield); MP = 152-154 °C;  $^1\text{H}$  NMR (400 MHz, Chloroform-d)  $\delta$  8.12 (d,  $J = 8.6$  Hz, 1H), 7.91 (d,  $J = 9.0$  Hz, 1H), 7.88 – 7.78 (m, 3H), 7.67 (s, 1H), 7.54 (dd,  $J = 8.6, 1.8$  Hz, 1H), 7.45 (d,  $J = 8.4$  Hz, 2H), 7.39 – 7.29 (m, 4H), 7.28 – 7.20 (m, 4H), 7.10 (d,  $J = 7.8$  Hz, 2H), 3.77 (s, 3H), 2.35 (s, 3H), 2.29 (s, 3H) ppm.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-d)  $\delta$  155.3, 145.0, 138.4, 136.8, 136.7, 135.5, 134.5, 133.9, 132.3, 130.0, 129.9, 129.4, 129.1, 128.1, 127.2, 127.0, 126.9, 126.6, 125.2, 124.1, 123.8, 119.3, 118.1, 115.1, 114.0, 113.7, 56.7, 21.7, 21.0 ppm; HRMS (ESI TOF) m/z: [M+ H]<sup>+</sup> calcd for  $\text{C}_{33}\text{H}_{28}\text{NO}_3\text{S}$ : 518.1784; Found: 518.1796. IR (KBr, cm<sup>-1</sup>): 3462, 3263, 2942, 1689, 1609, 1528, 1438, 1205, 1075, 831, 716.

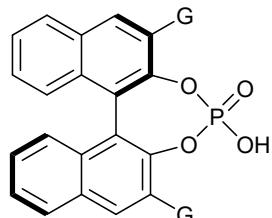
## 3. Optimization of reaction conditions

**Table S1.** Optimization of the reaction catalyst <sup>a</sup>

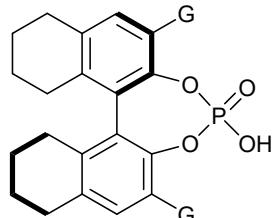




(S) -4.1



(R) -4.2



(R) -4.3

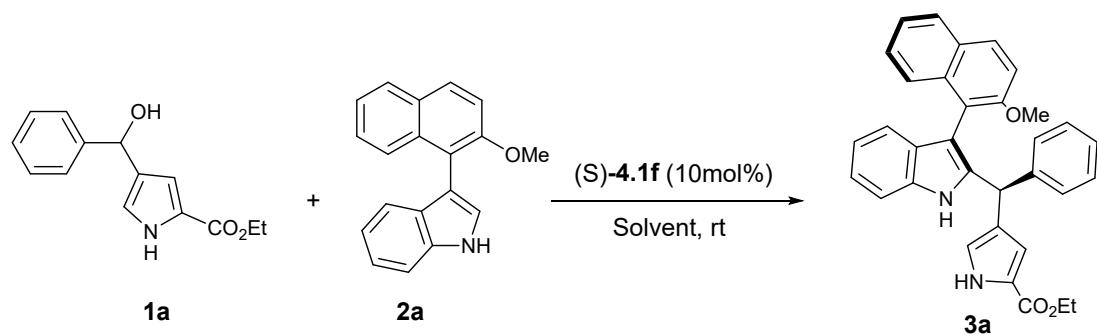
**4.1a:** G = 3,5-(tBu)<sub>2,4</sub>-MeOC<sub>2</sub>H<sub>6</sub>**4.1b:** G = 2,4,6-Me<sub>3</sub>C<sub>6</sub>H<sub>2</sub>**4.1c:** G = H**4.1d:** G = 9-anthracenyl**4.1e:** G = 3,5-(CH<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>**4.1f:** G = 3,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>**4.1g:** G = 1-naphthalene**4.1h:** G = Si-Ph<sub>3</sub>**4.1i:** G = Ph**4.1j:** G = 4-NO<sub>2</sub>-C<sub>6</sub>H<sub>4</sub>**4.1k:** G = 9-phenanthrenyl**4.2a:** G = 3,5-(tBu)<sub>2,4</sub>-MeOC<sub>2</sub>H<sub>6</sub>**4.2b:** G = 2,4,6-Me<sub>3</sub>C<sub>6</sub>H<sub>2</sub>**4.2c:** G = 1-pyrenyl**4.2d:** G = 9-anthracenyl**4.2e:** G = 3,5-(CH<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>**4.2f:** G = 3,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>**4.3a:** G = 2,4,6-i-Pr<sub>3</sub>C<sub>6</sub>H<sub>2</sub>**4.3b:** G = 3,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>**4.3c:** G = 3,5-(tBu)<sub>2,4</sub>-MeOC<sub>2</sub>H<sub>6</sub>**4.3d:** G = 9-anthracenyl

Entry	Cat.	Solvent	Temp( °C)	Yield(%) <sup>b</sup>	dr <sup>c</sup>	ee(%) <sup>d</sup>
1	(S)-4.1a	DCE	rt	64	82:18	78
2	(S)-4.1b	DCE	rt	43	83:17	55
3	(S)-4.1c	DCE	rt	62	86:14	70
4	(S)-4.1d	DCE	rt	78	80:20	76
5	(S)-4.1e	DCE	rt	83	75:25	64
6	(S)-4.1f	DCE	rt	82	86:14	87
7	(S)-4.1g	DCE	rt	59	83:17	48
8	(S)-4.1h	DCE	rt	57	85:15	43
9	(S)-4.1i	DCE	rt	50	68:32	36
10	(S)-4.1j	DCE	rt	79	78:22	68
11	(S)-4.1k	DCE	rt	63	85:15	27

12	(R)-4.2a	DCE	rt	52	58:42	67
13	(R)-4.2b	DCE	rt	84	88:12	47
14	(R)-4.2c	DCE	rt	70	85:15	55
15	(R)-4.2d	DCE	rt	73	82:18	26
16	(R)-4.2e	DCE	rt	41	75:25	59
17	(R)-4.2f	DCE	rt	31	60:40	55
18	(R)-4.3a	DCE	rt	64	87:13	29
19	(R)-4.3b	DCE	rt	55	76:24	71
20	(R)-4.3c	DCE	rt	60	79:21	37
21	(R)-4.3d	DCE	rt	47	84:16	62

<sup>a</sup>Reaction conditions: 1a (0.06 mmol), 2a (0.05 mmol) and catalyst (10 mol%) in DCE (0.5 mL) at room temperature for 12 h. <sup>b</sup> Isolated yields. <sup>c</sup> Determined by HPLC with C18 column. <sup>d</sup> Determined by chiral HPLC analysis.

**Table S2.** Optimization of the reaction solvent <sup>a</sup>

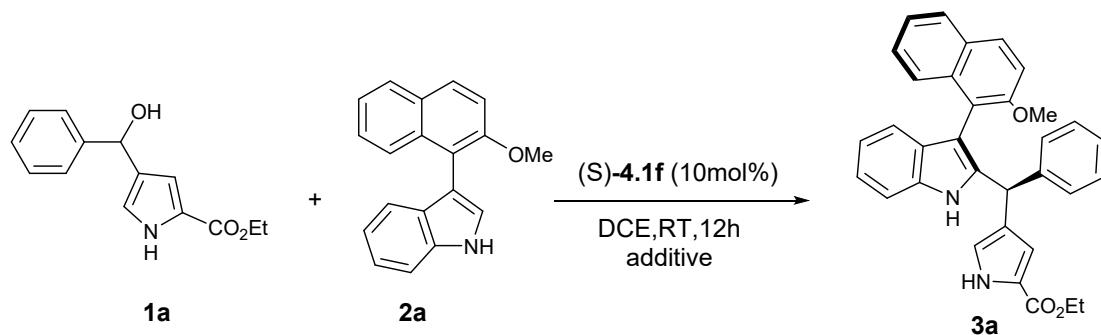


Entry	Solvent	Yield(%) <sup>b</sup>	dr <sup>c</sup>	ee(%) <sup>d</sup>
1	DCM	88	84:16	85
2	DMF	N.R.	--	--

3	Methano	43	73:27	62
4	THF	N.R.	--	--
5	CHCl <sub>3</sub>	86	83:17	72
6	Et <sub>2</sub> O	70	55:45	64
7	EA	63	76:24	78
8	Toluene	77	80:20	60
9	DME	N.R.	--	--
10 <sup>e</sup>	DCE	80	86:14	82
11 <sup>f</sup>	DCE	73	85:15	78

<sup>a</sup> Reaction conditions: 1a (0.05 mmol), 2a (0.06 mmol) and (S)-4.1i (10 mol%) in 0.5 mL solvent at room temperature for 12 h. <sup>b</sup> Isolated yields. <sup>c</sup> Determined by HPLC with C18 column. <sup>d</sup> Determined by chiral HPLC analysis. <sup>e</sup> 1.0 mL DCE. <sup>f</sup> 0.25 mL DCE.

**Table S3.** Optimization of the reaction additive <sup>a</sup>

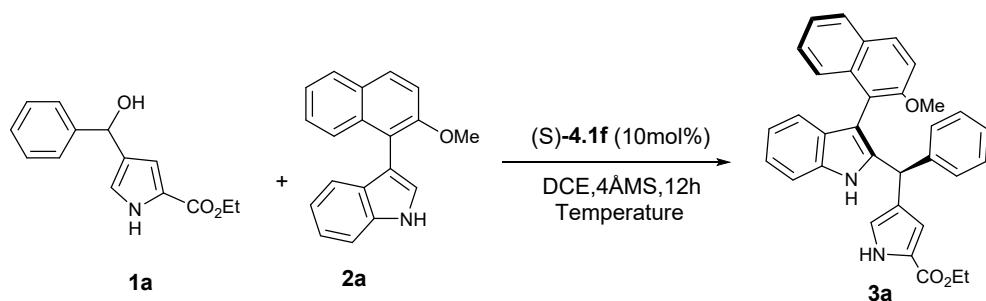


Entry	Additive	Yield(%) <sup>b</sup>	dr <sup>d</sup>	ee(%) <sup>c</sup>
1	3 Å MS	83	88:12	88
2	4 Å MS	85	87:13	89

3	5 Å MS	80	85:15	86
4	Na <sub>2</sub> SO <sub>4</sub>	67	86:14	82
5	MgSO <sub>4</sub>	72	85:15	80
6	3 Å AMS <sup>e</sup>	87	88:12	84
7	4 Å MS <sup>e</sup>	78	88:12	86
8	5 Å MS <sup>e</sup>	80	86:14	85

<sup>a</sup> Reaction conditions: 1a (0.05 mmol), 2a (0.06 mmol), (S)-4.1f (10 mol%) and additive(80 mg) in 0.5 mL solvent for 12 h. <sup>b</sup> Isolated yields. <sup>c</sup> Determined by chiral HPLC analysis. <sup>d</sup> Determined by HPLC with C18 column. <sup>e</sup>120mg

**Table S4.** Optimization of the reaction temperature and catalyst loading <sup>a</sup>



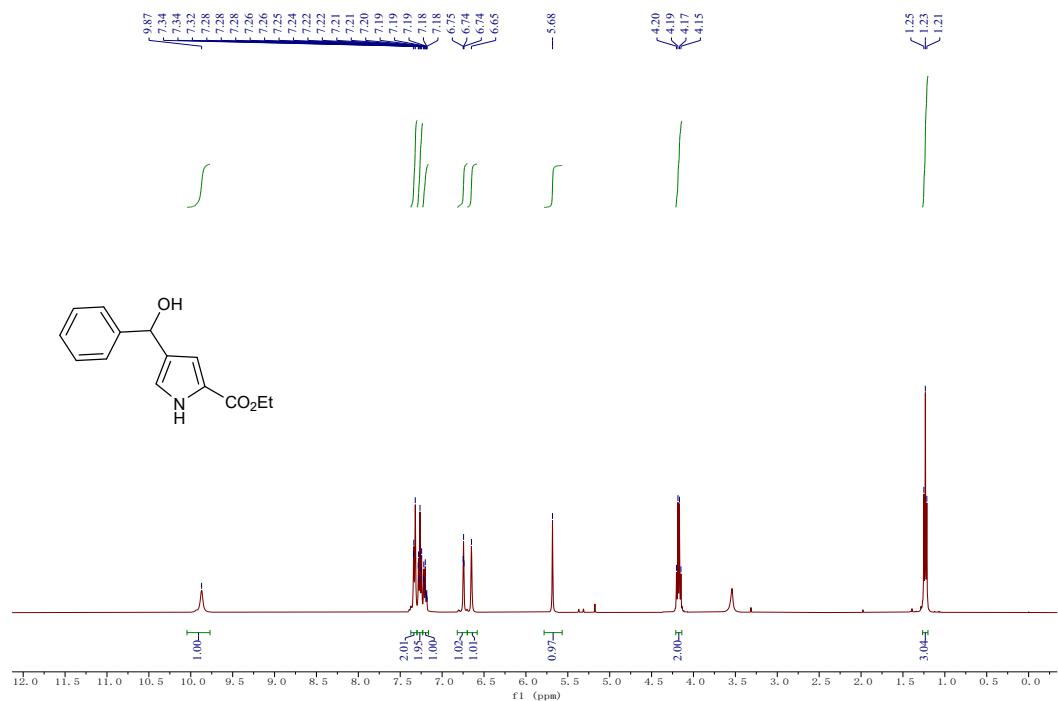
Entry	Temp.( °C)	Yield(%) <sup>b</sup>	dr <sup>c</sup>	ee(%) <sup>d</sup>
1 <sup>e</sup>	-20	88	85:15	92
2 <sup>e</sup>	-10	85	87:13	90
3	0	85	90:10	89
4	10	89	82:18	89
5	rt	84	85:15	86
6	40	55	72:28	63

<sup>a</sup> Reaction conditions: **1a** (0.05 mmol), **2a** (0.06 mmol), (*S*)-**4.1f** (10 mol%) and 4 Å MS (80 mg) in 0.5 mL solvent for 12 h. <sup>b</sup> Isolated yields. <sup>c</sup> Determined by HPLC with C18 column. <sup>d</sup> Determined by chiral HPLC analysis. <sup>e</sup> 24 h

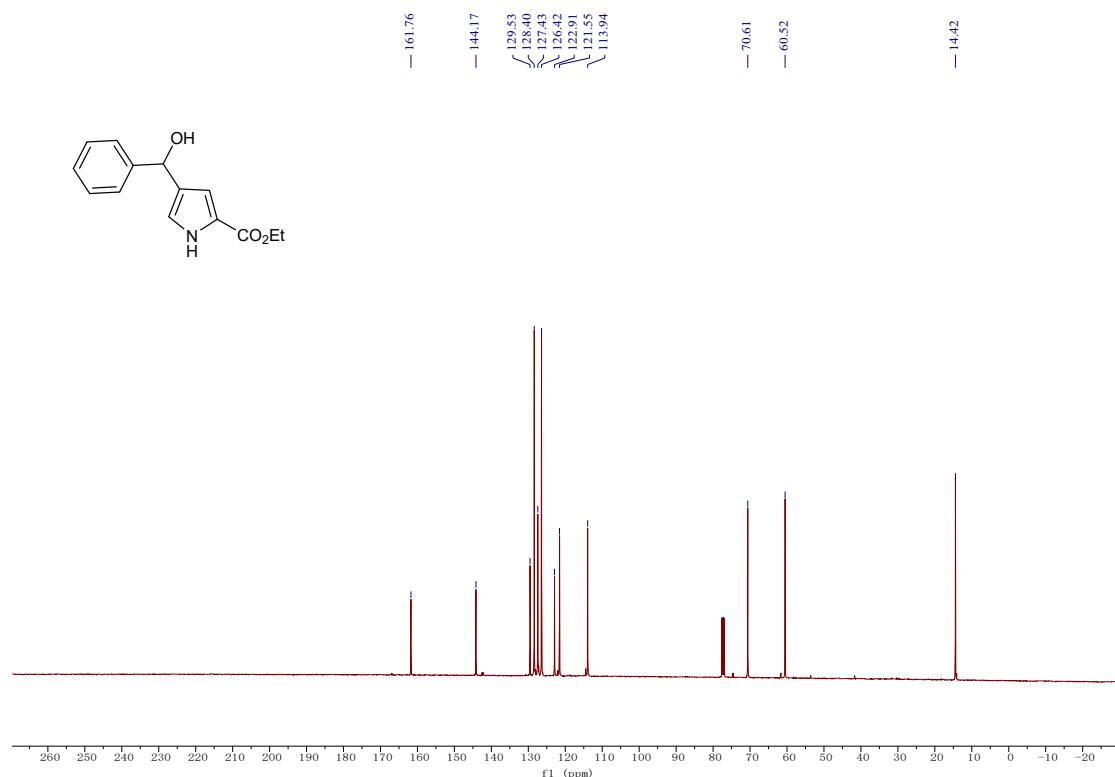
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#### 4. Copies of $^1\text{H}$ NMR, $^{13}\text{C}$ NMR and $^{19}\text{F}$ NMR Spectra

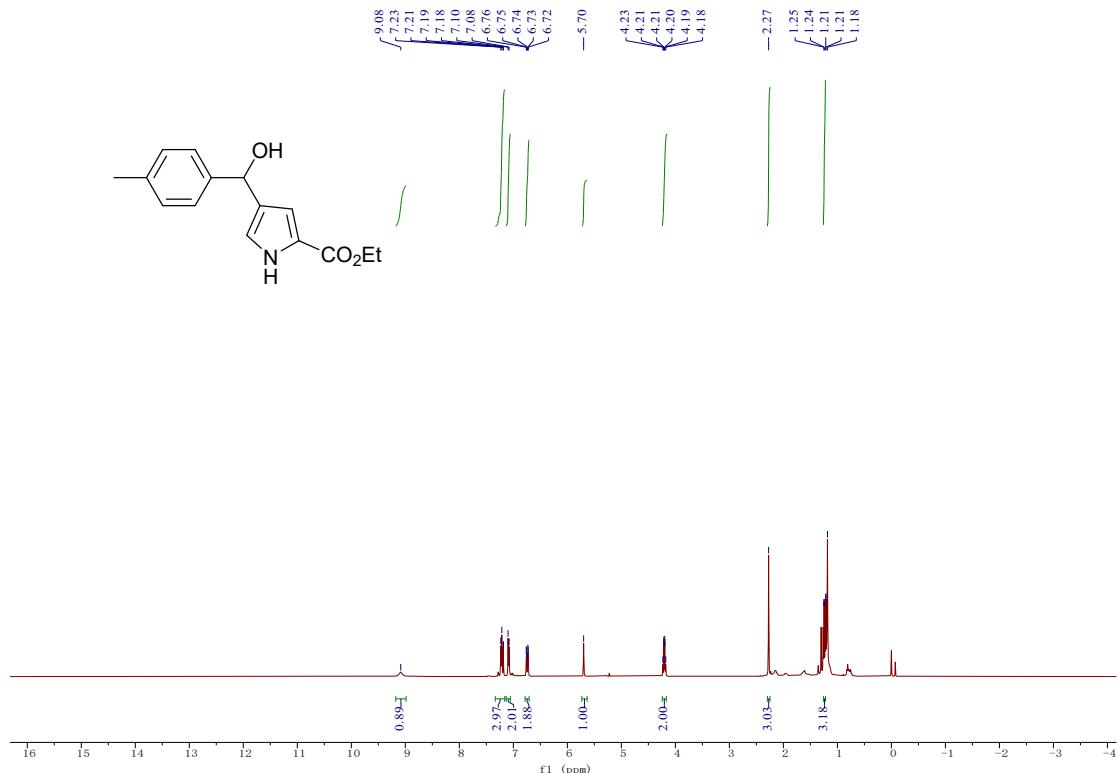
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) of **1a**



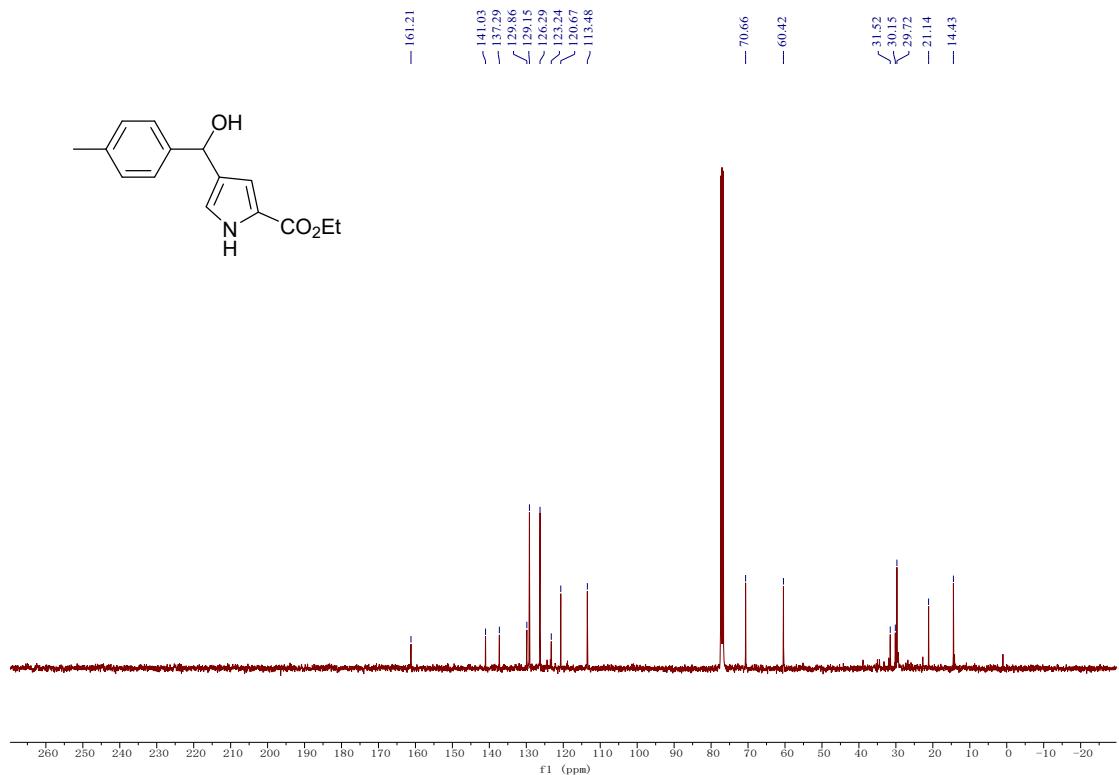
$^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ ) of **1a**



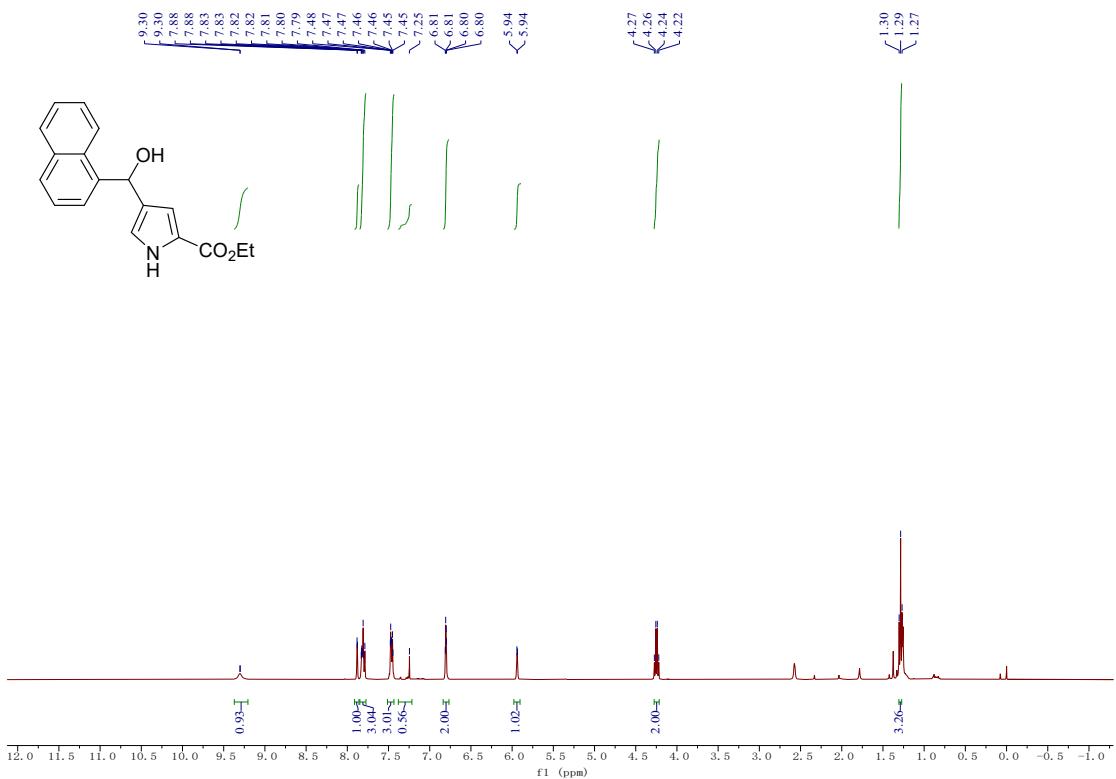
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **1d**



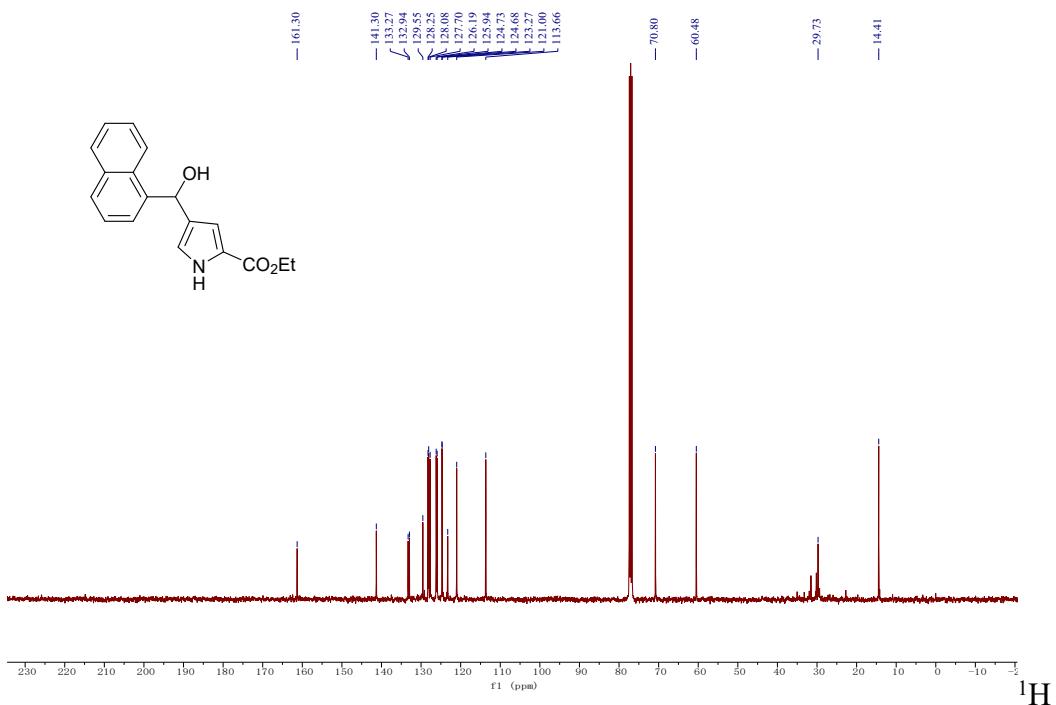
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **1d**



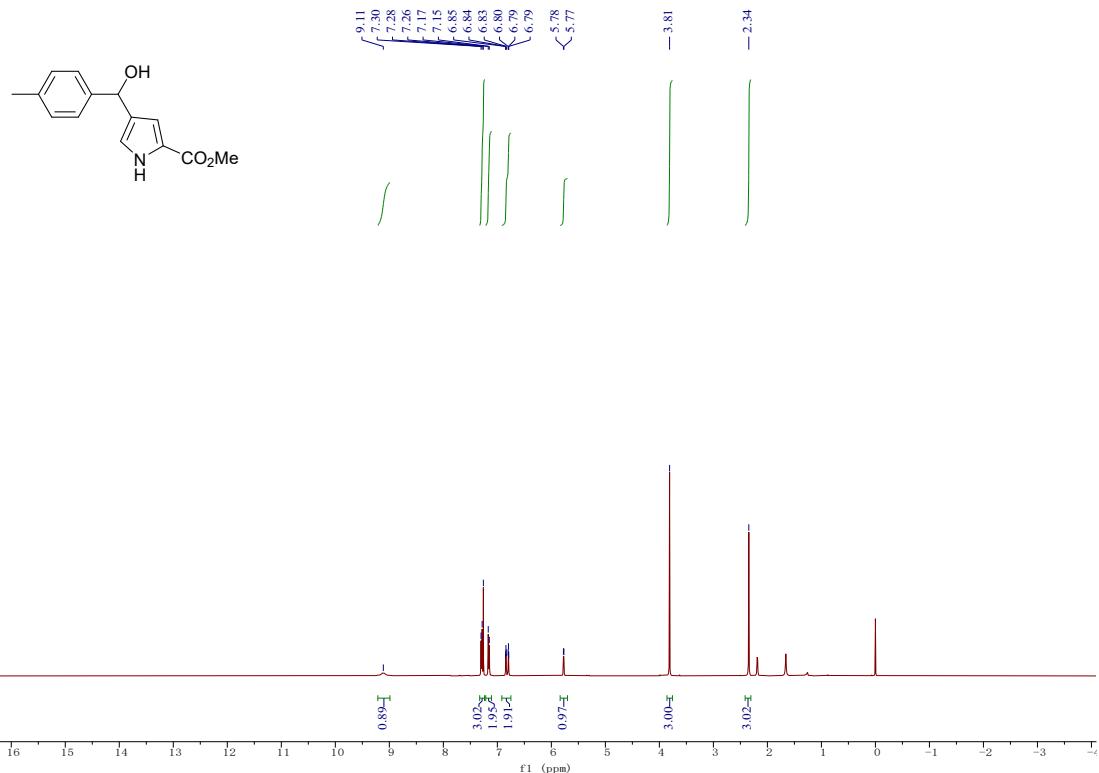
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **1e**



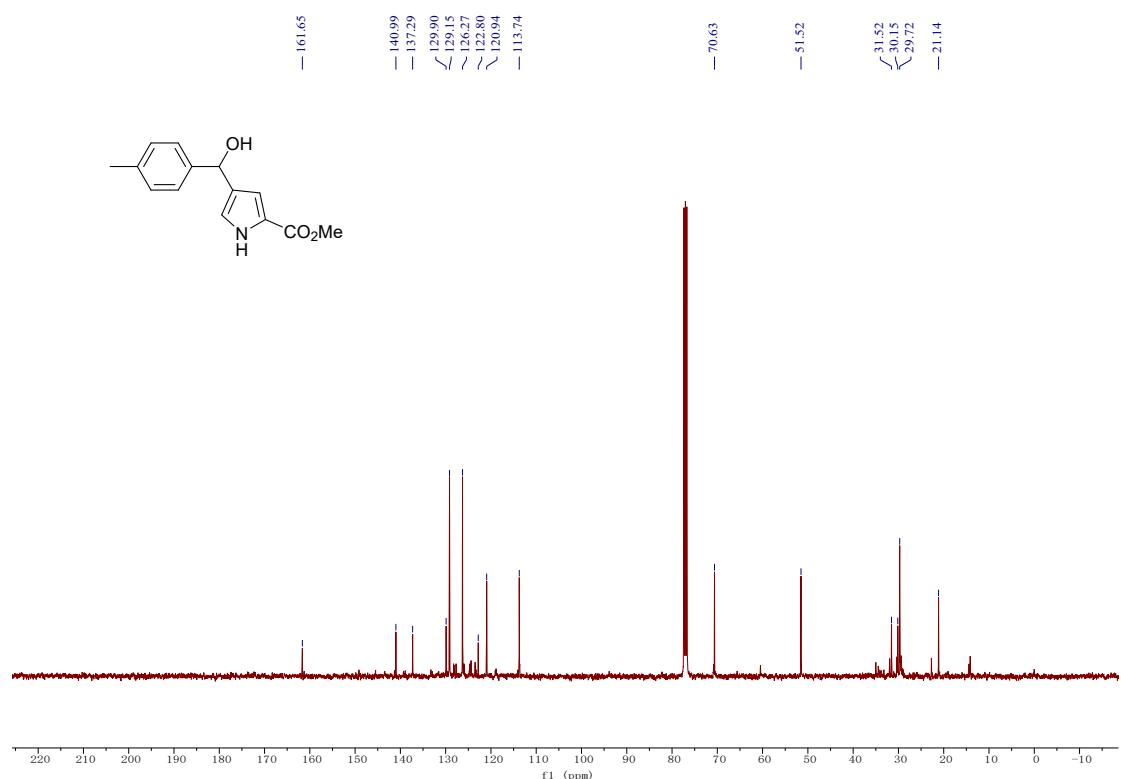
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **1e**



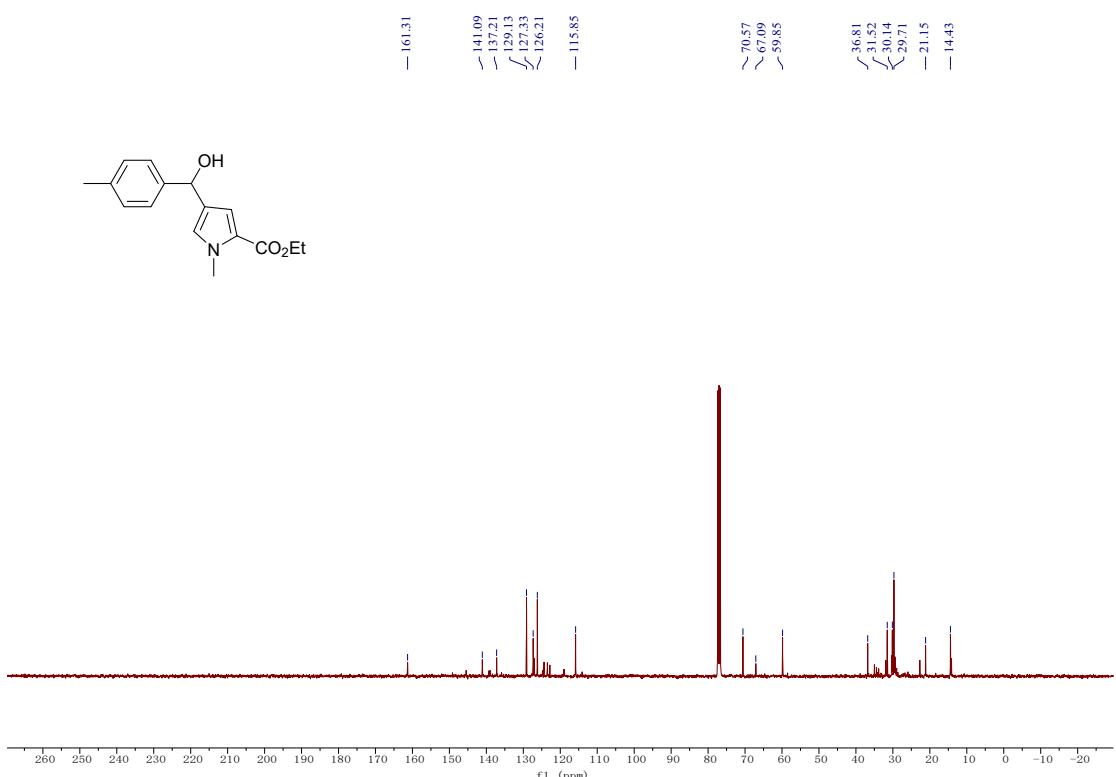
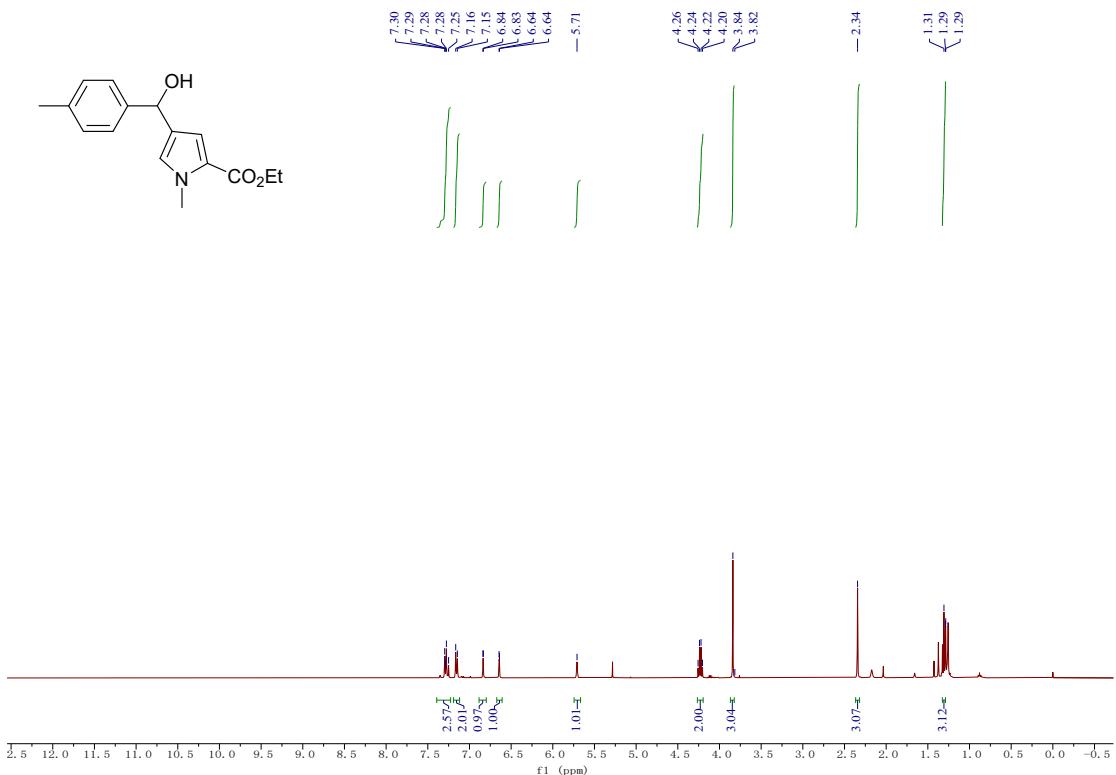
NMR (400 MHz, CDCl<sub>3</sub>) of **1f**



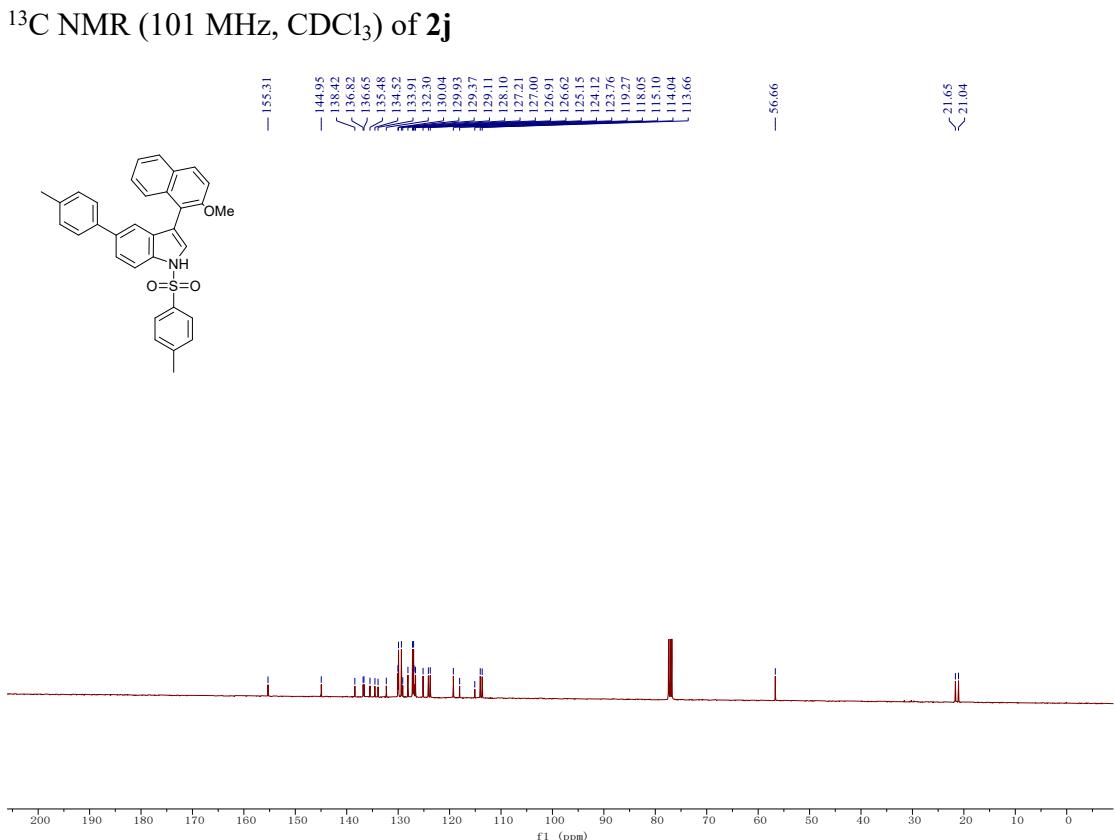
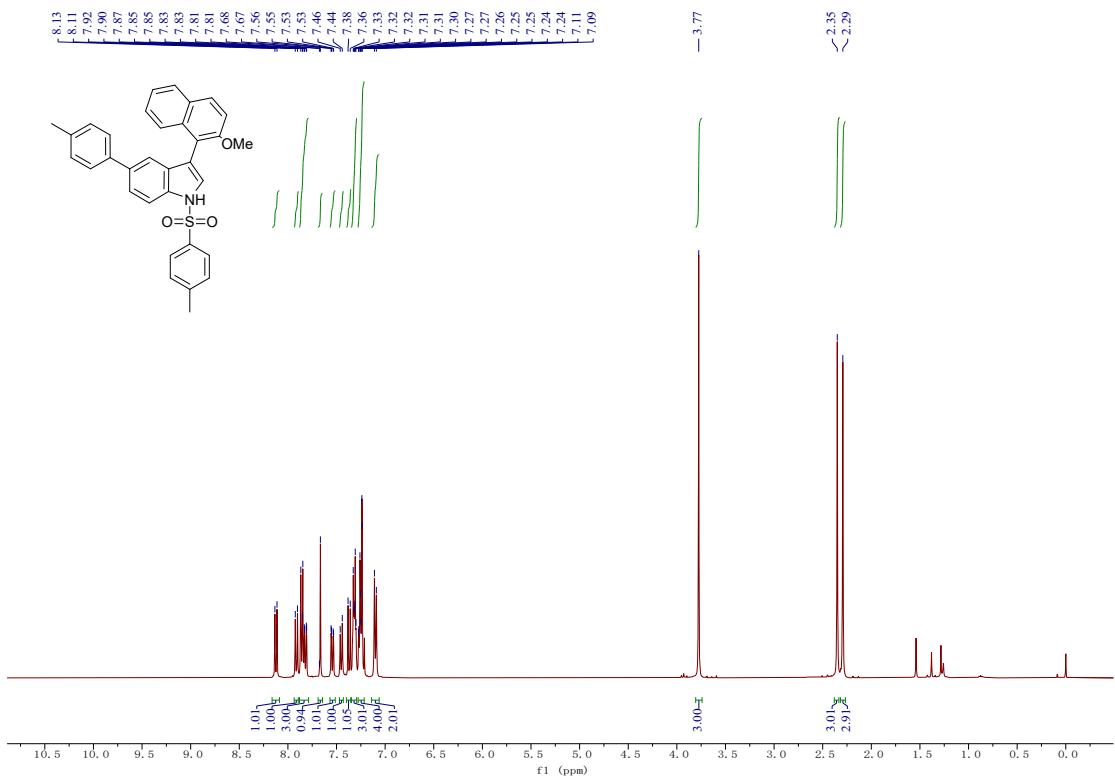
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **1f**



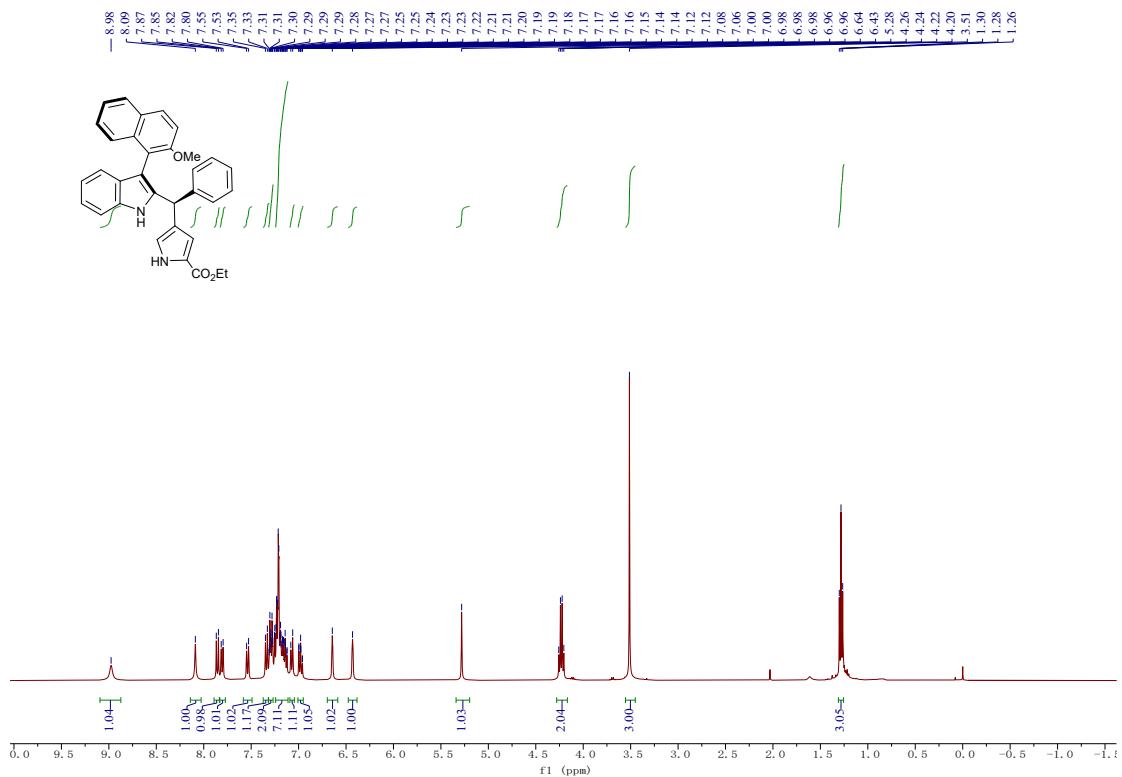
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **1g**



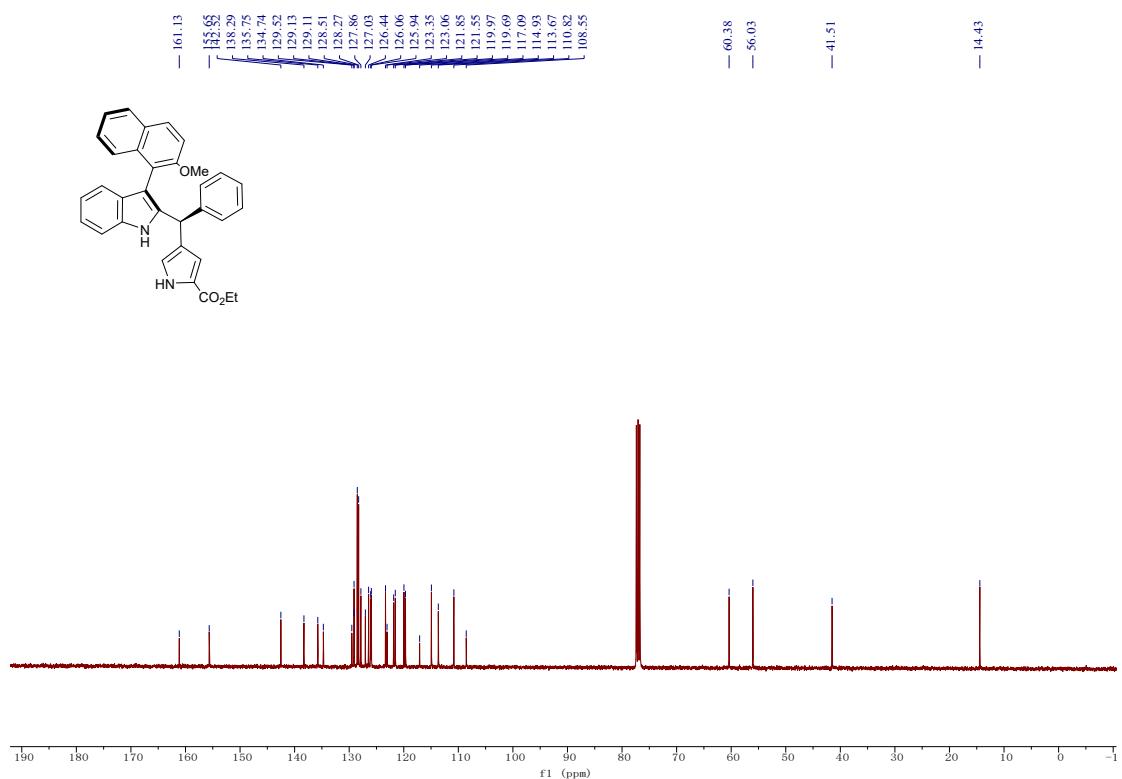
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 2j



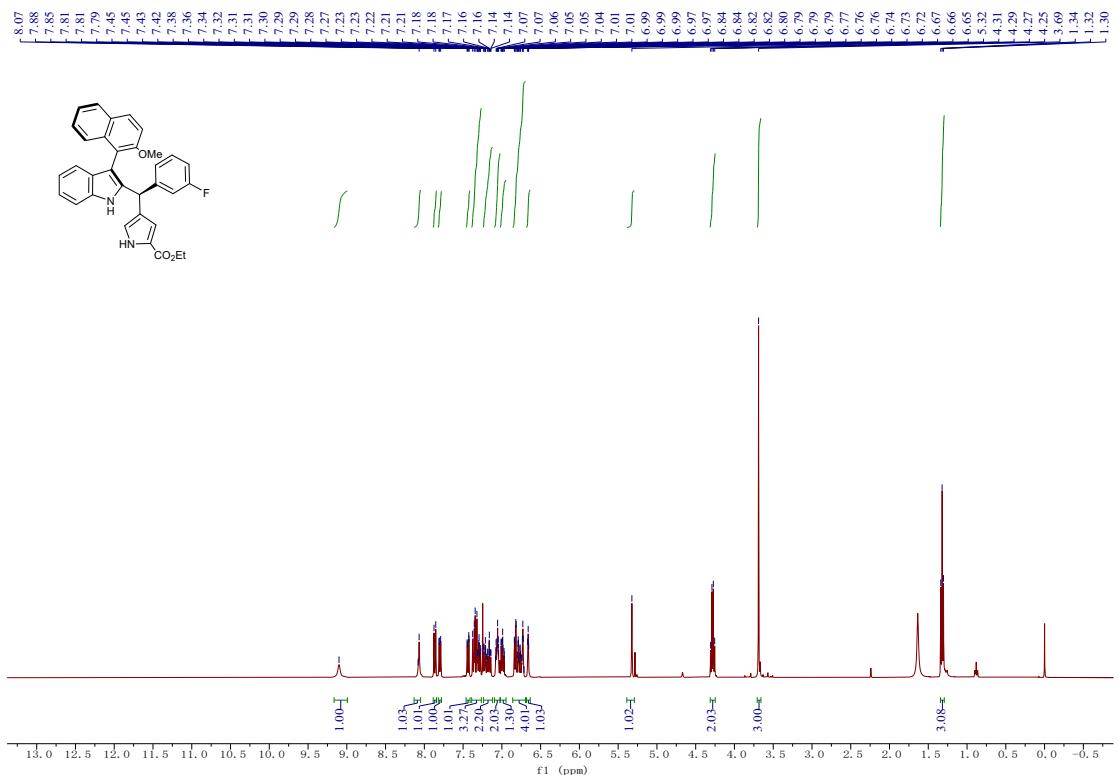
<sup>1</sup>H NMR ( $400\text{ }^1\text{H}$  MHz,  $\text{CDCl}_3$ ) of **3a**



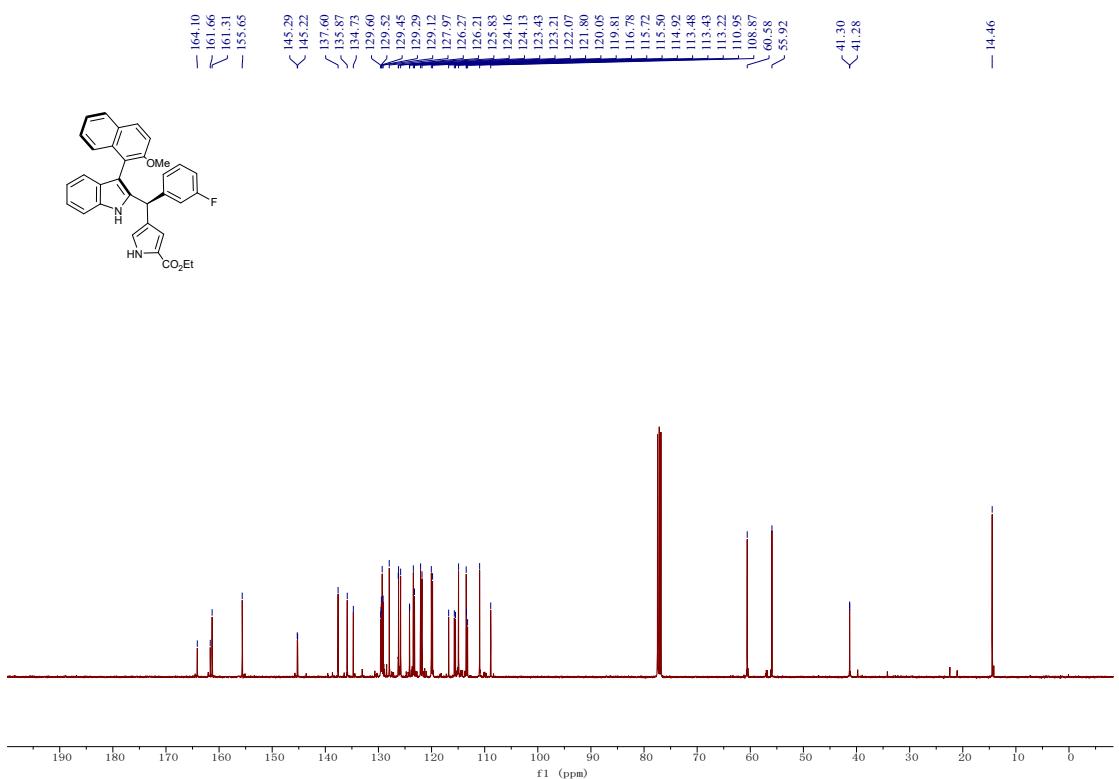
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3a**



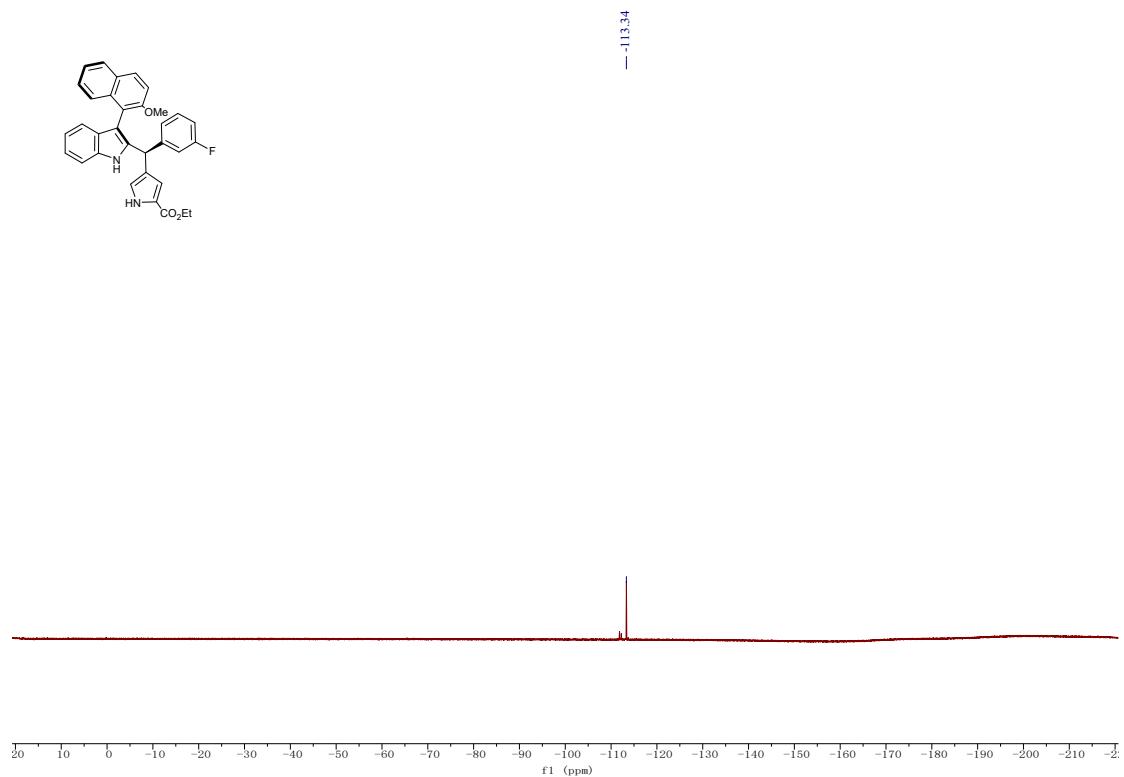
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3b**



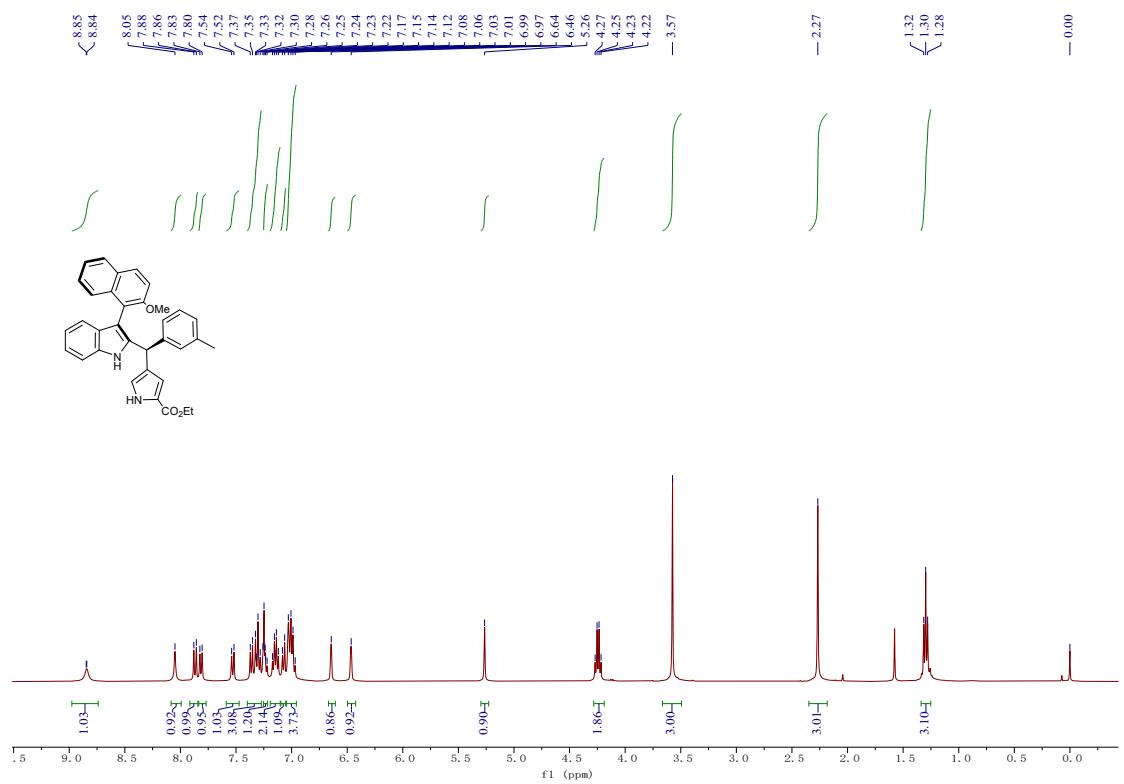
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3b**



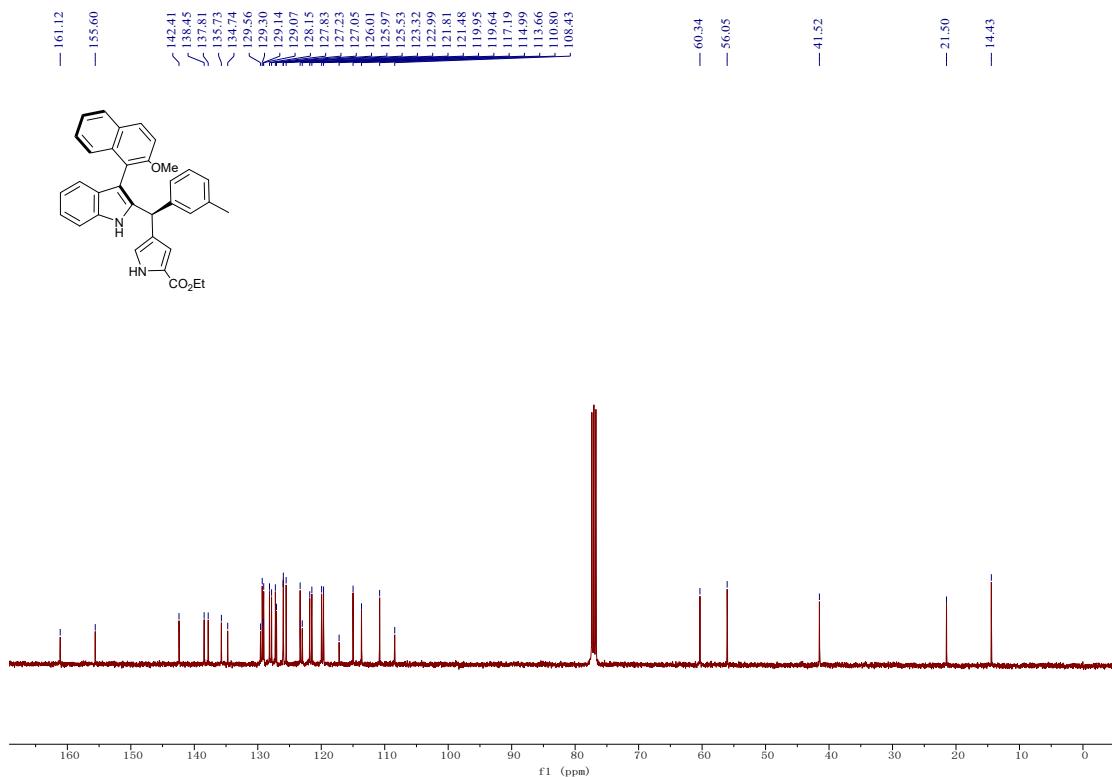
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3b**



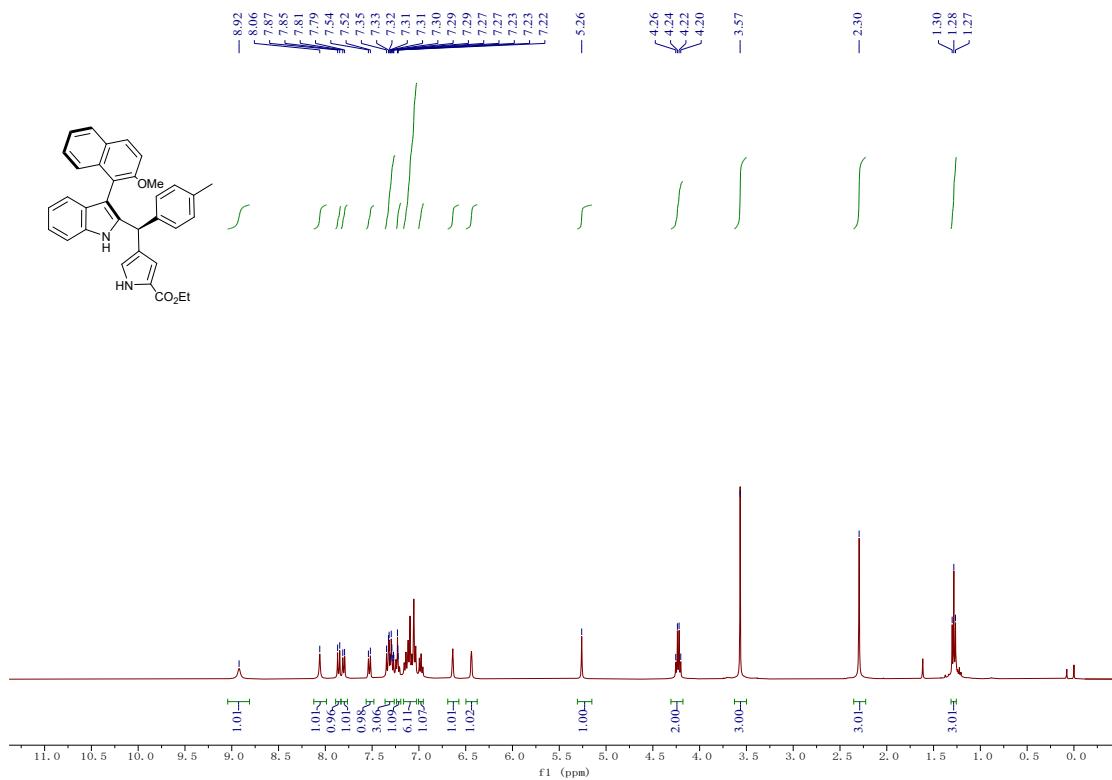
### $^1\text{H}$ NMR (400 MHz, $\text{CDCl}_3$ ) of 3c



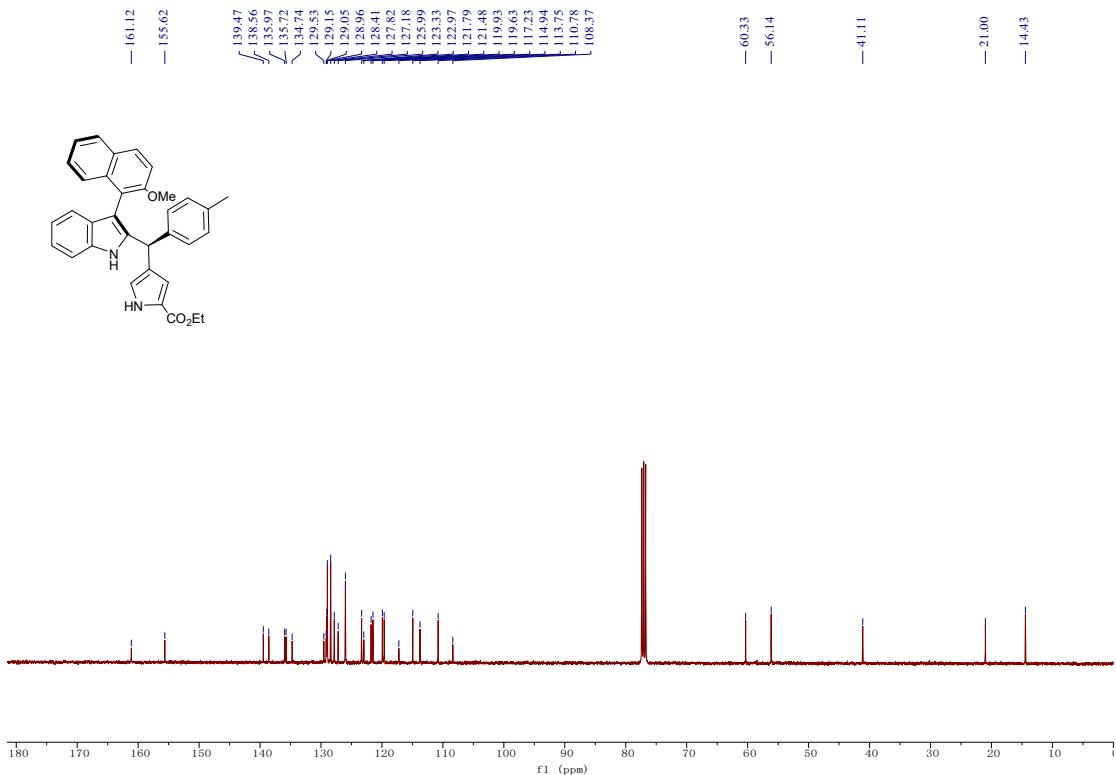
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3c**



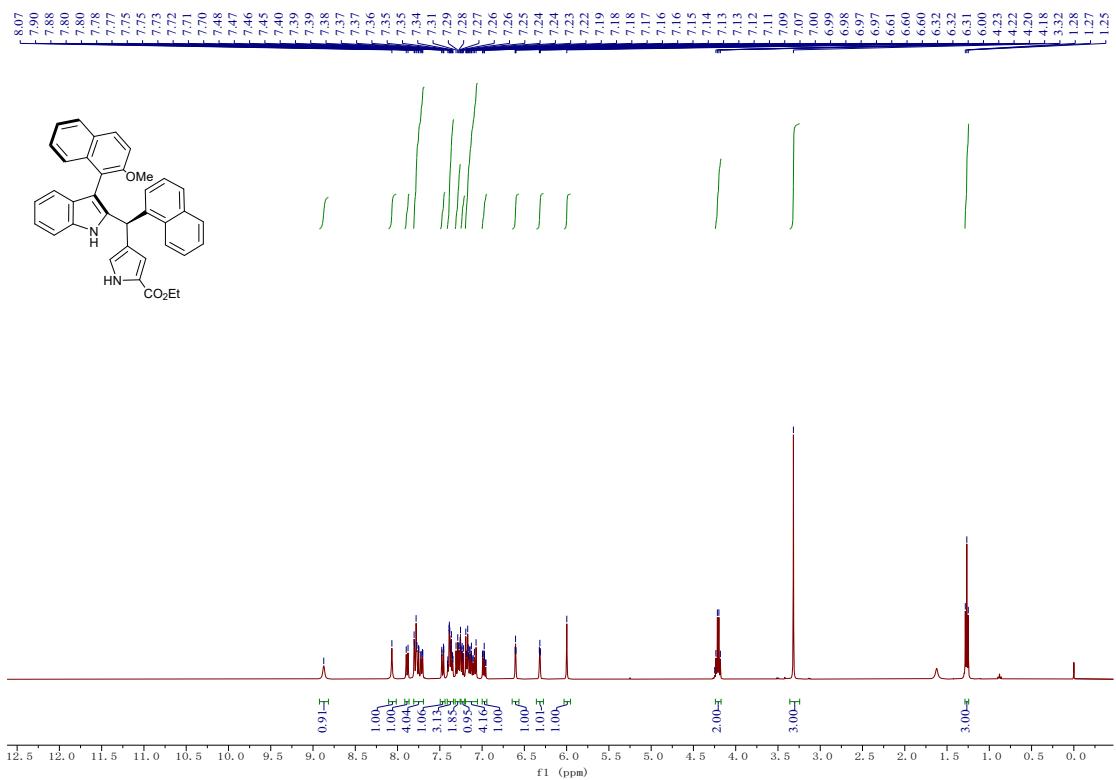
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3d**



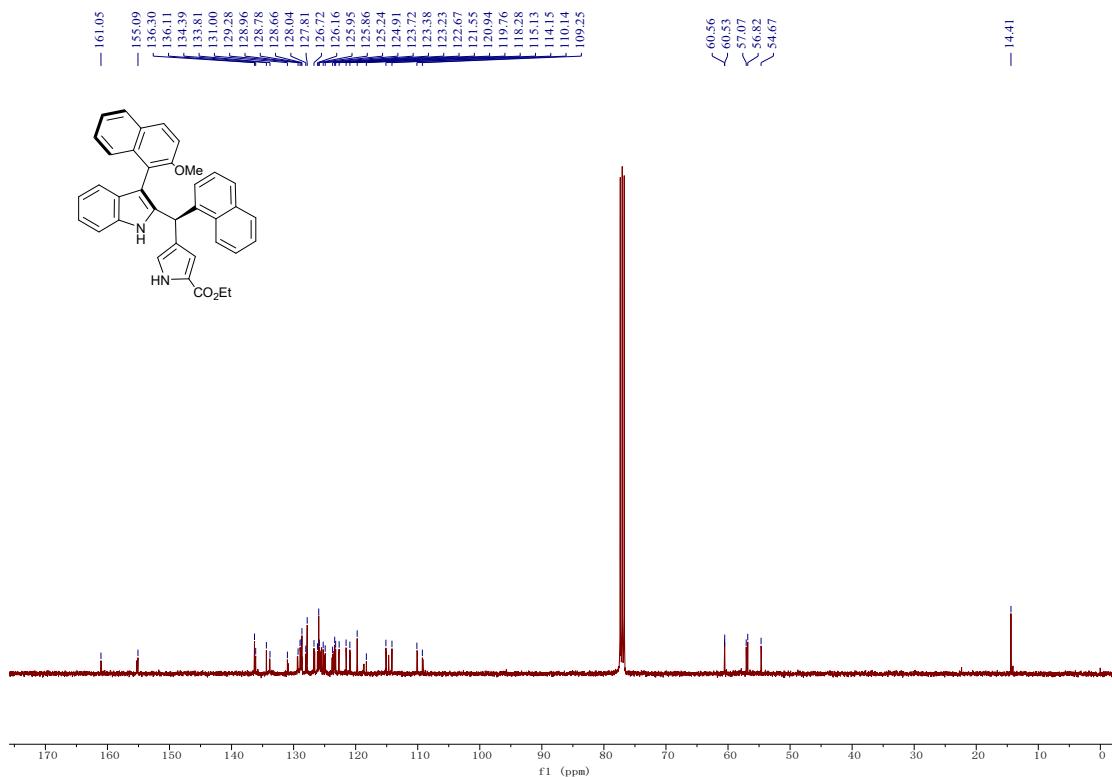
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3d**



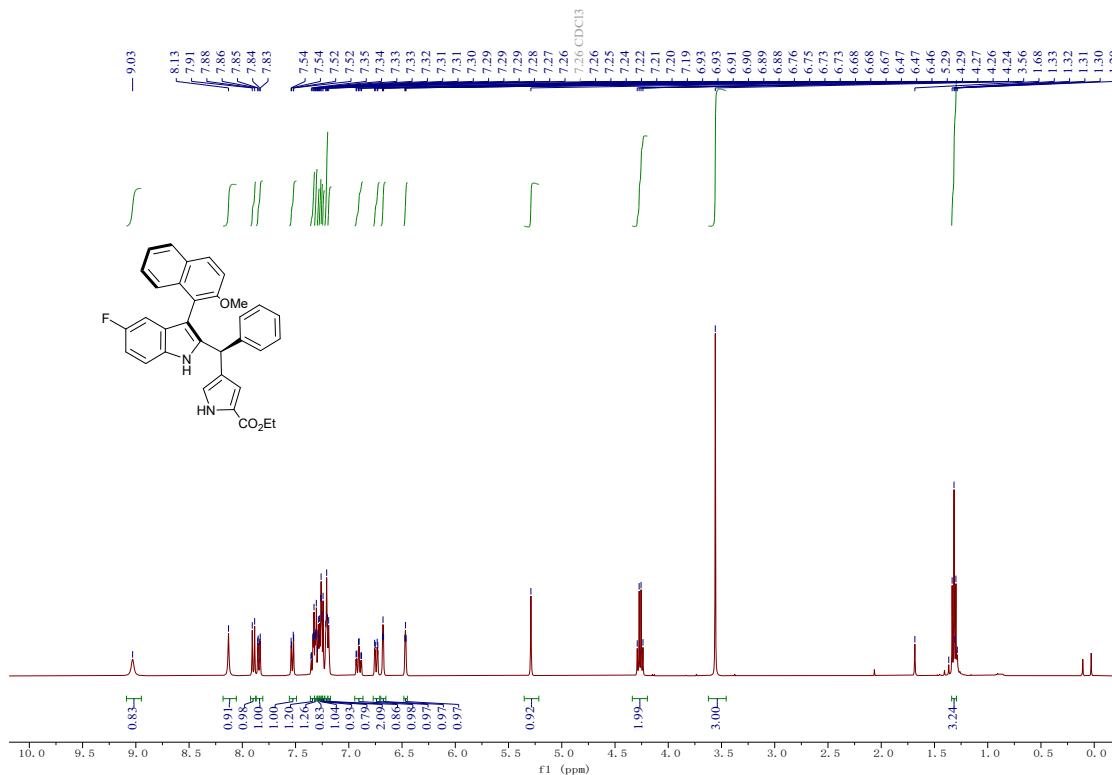
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3e**



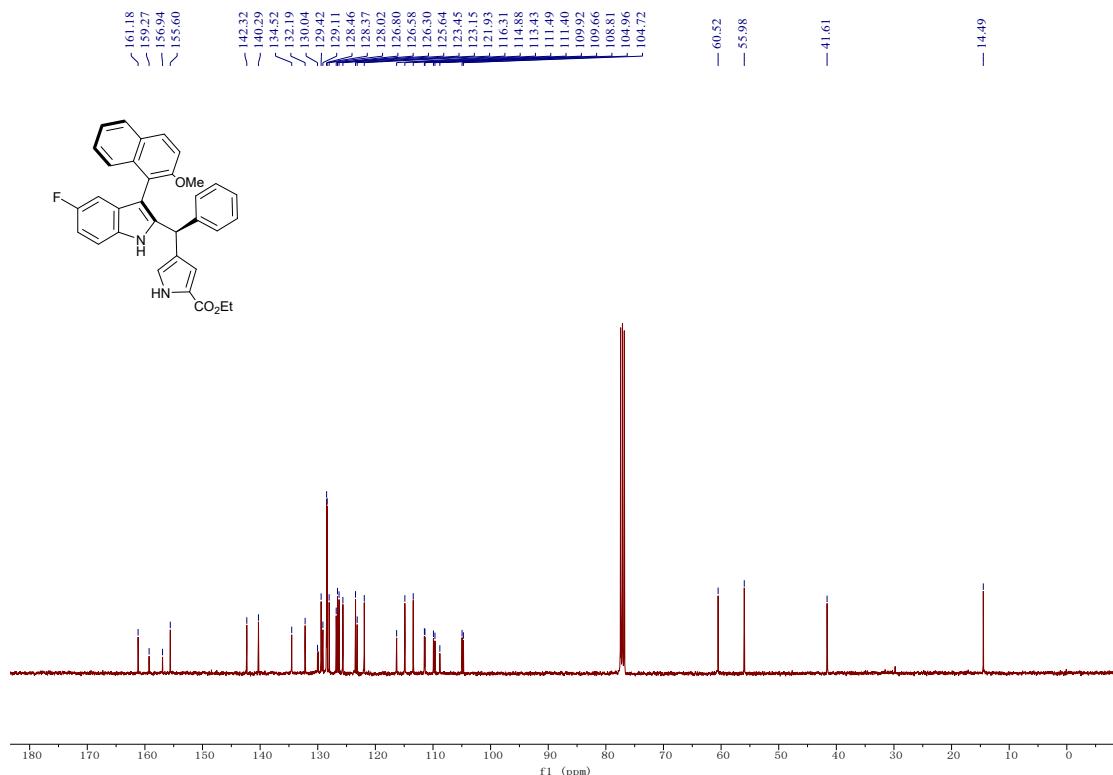
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3e**



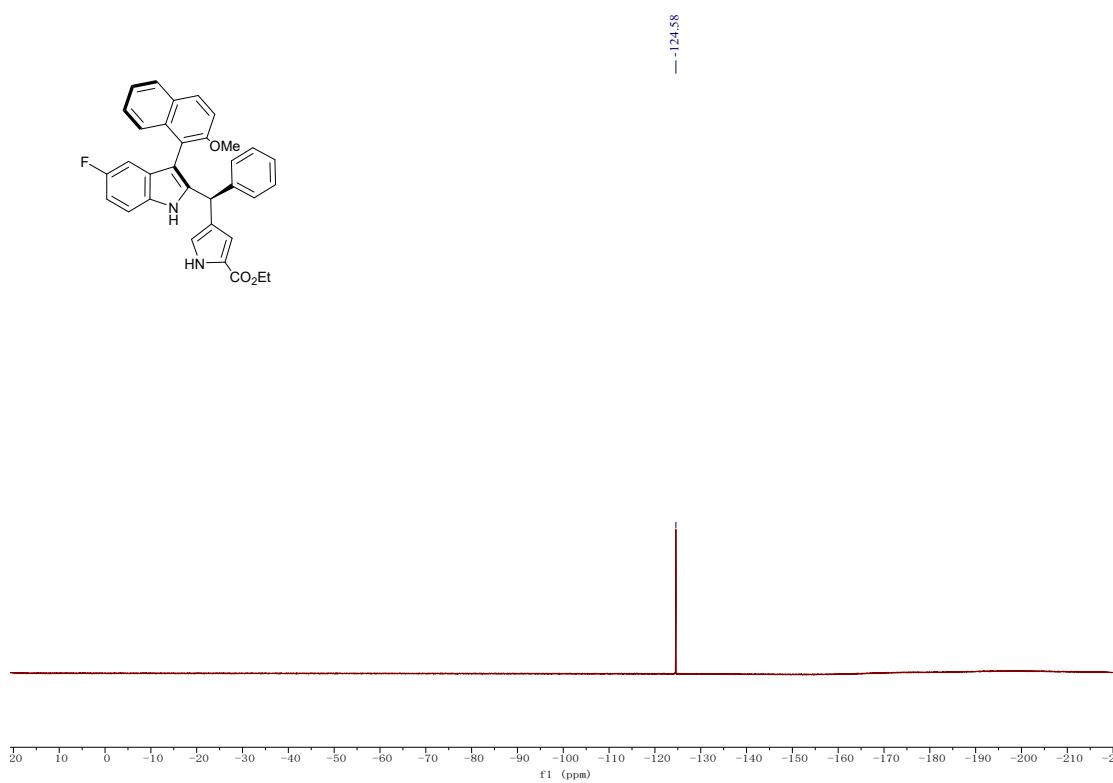
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3f**



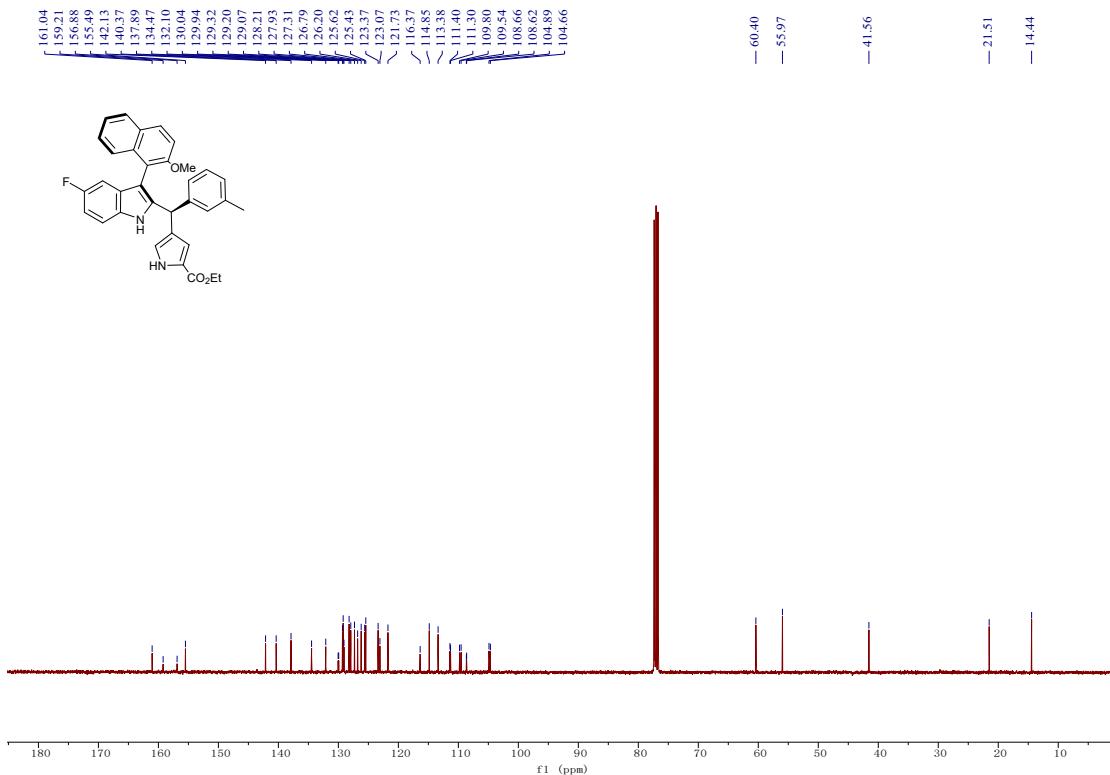
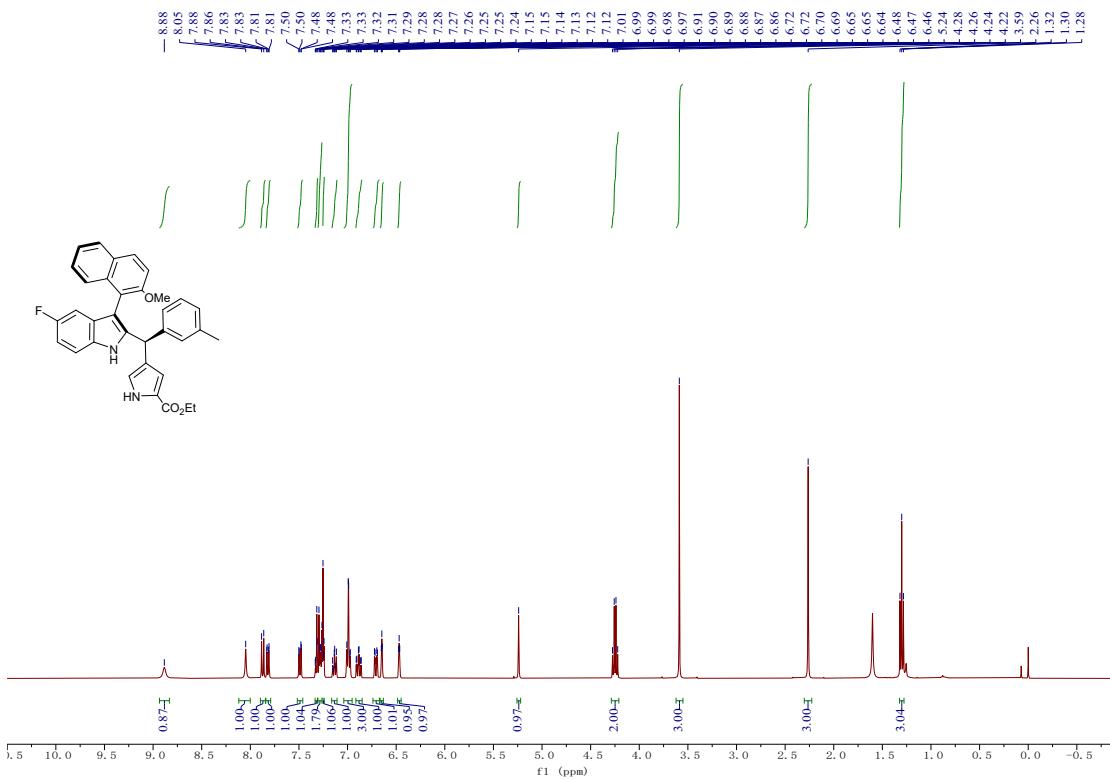
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3f**



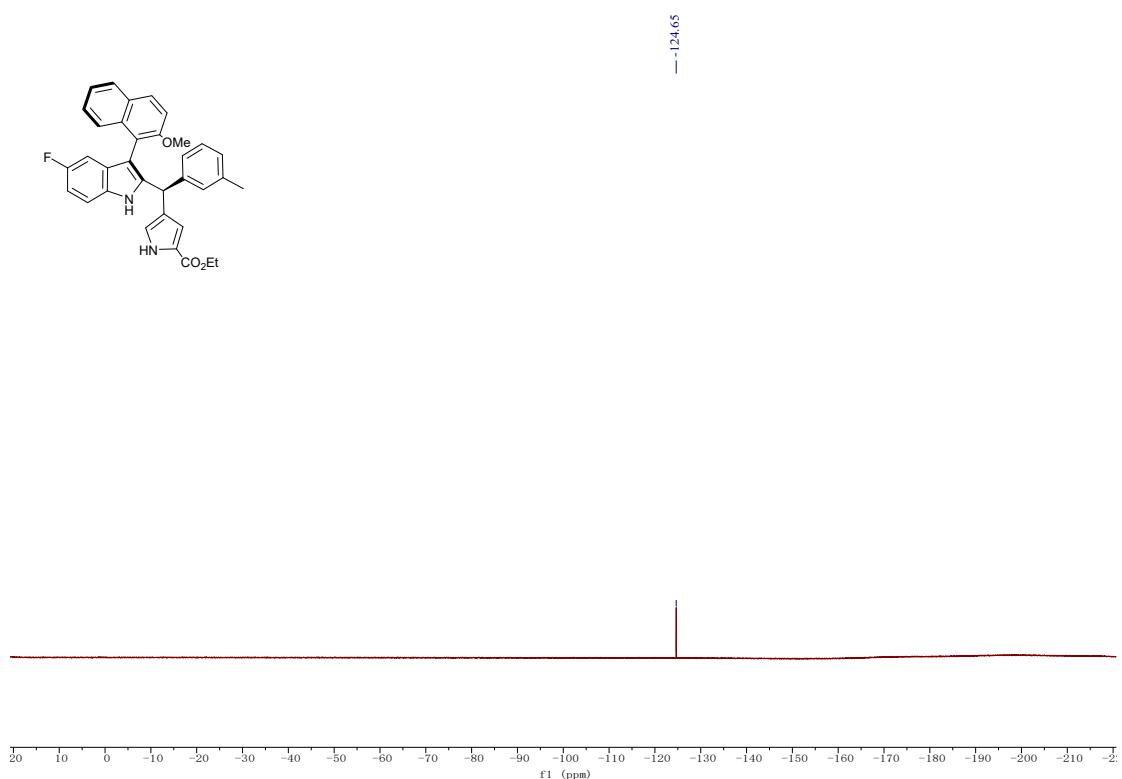
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3f**



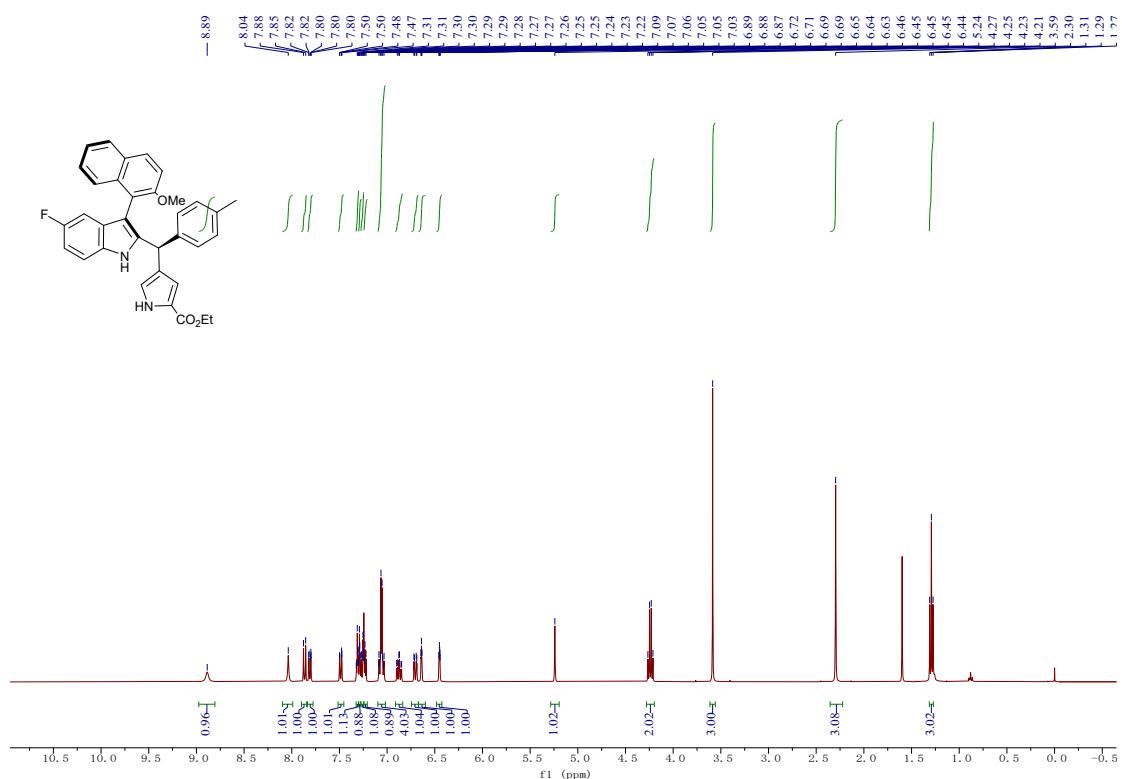
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3g**



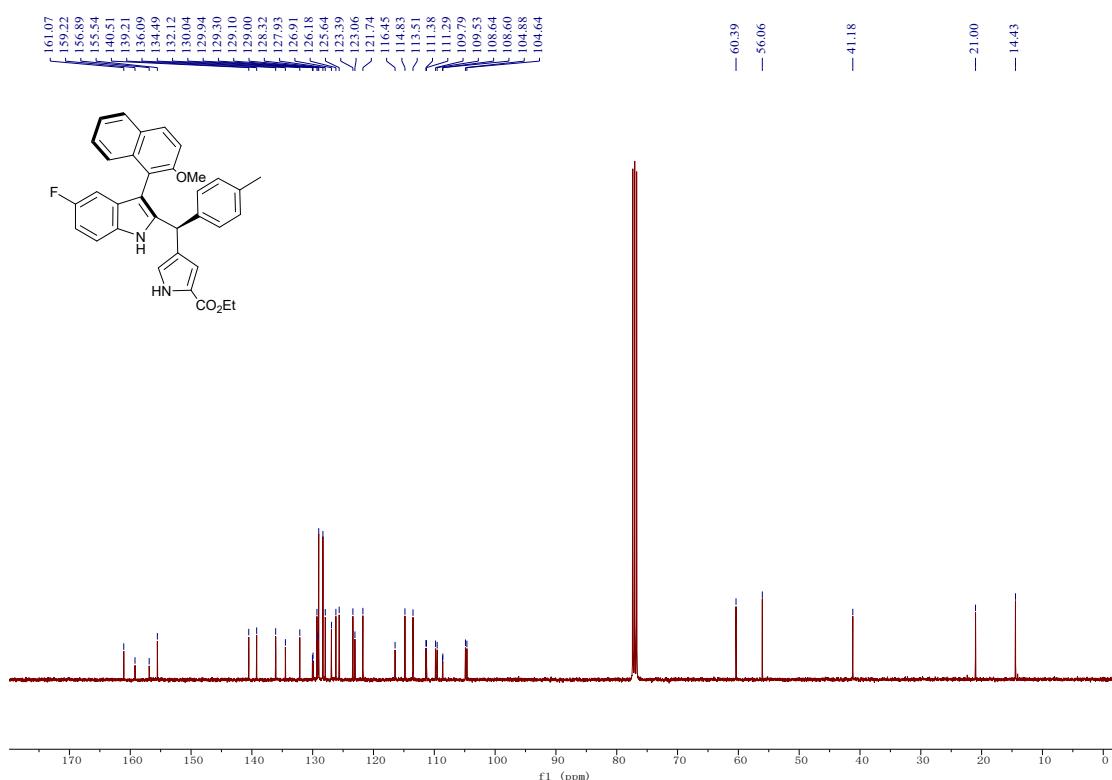
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3g**



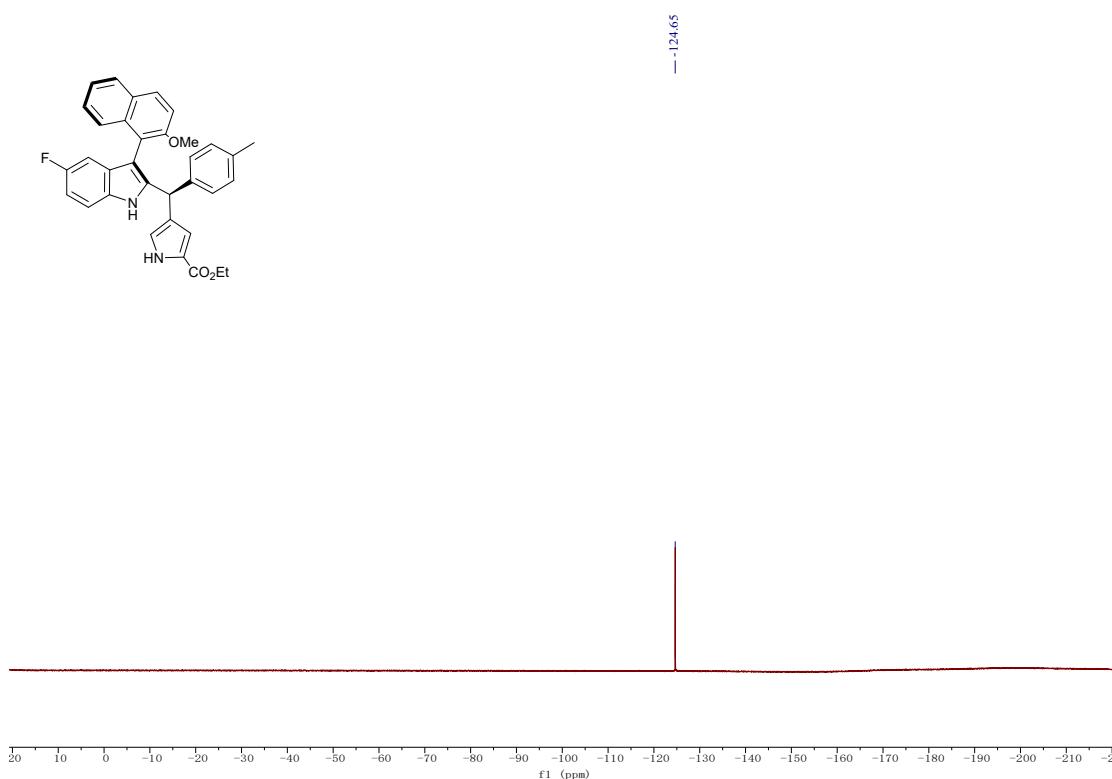
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3h**



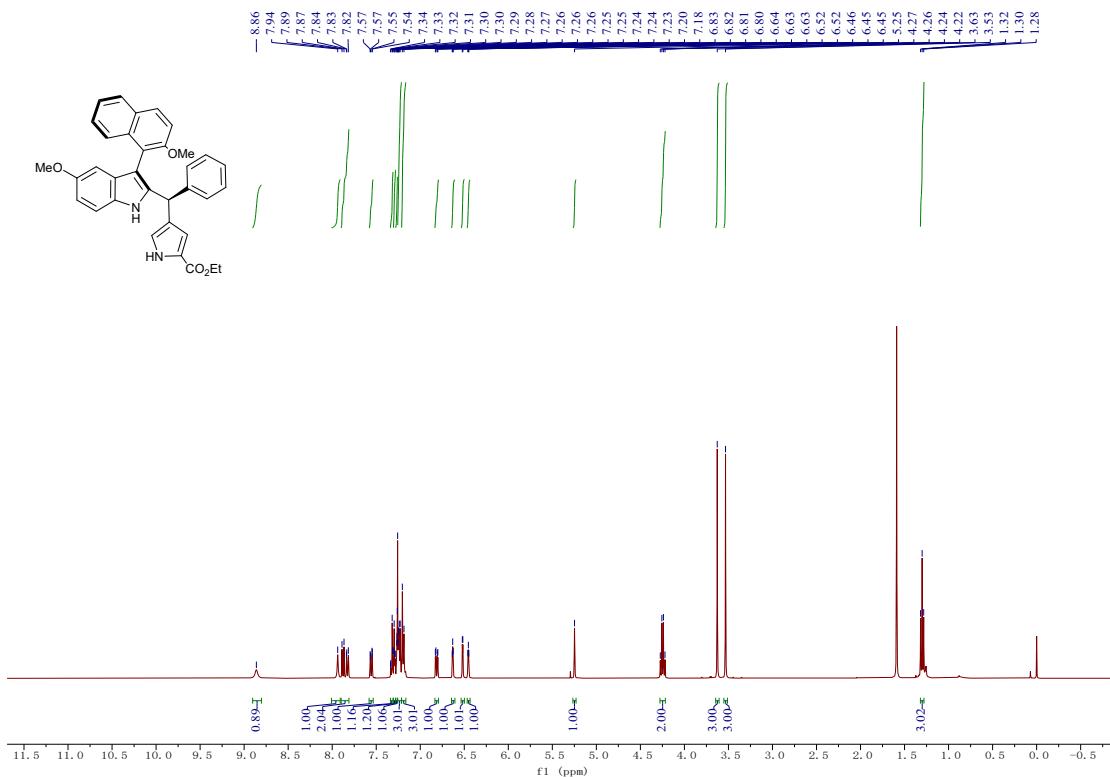
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3h**



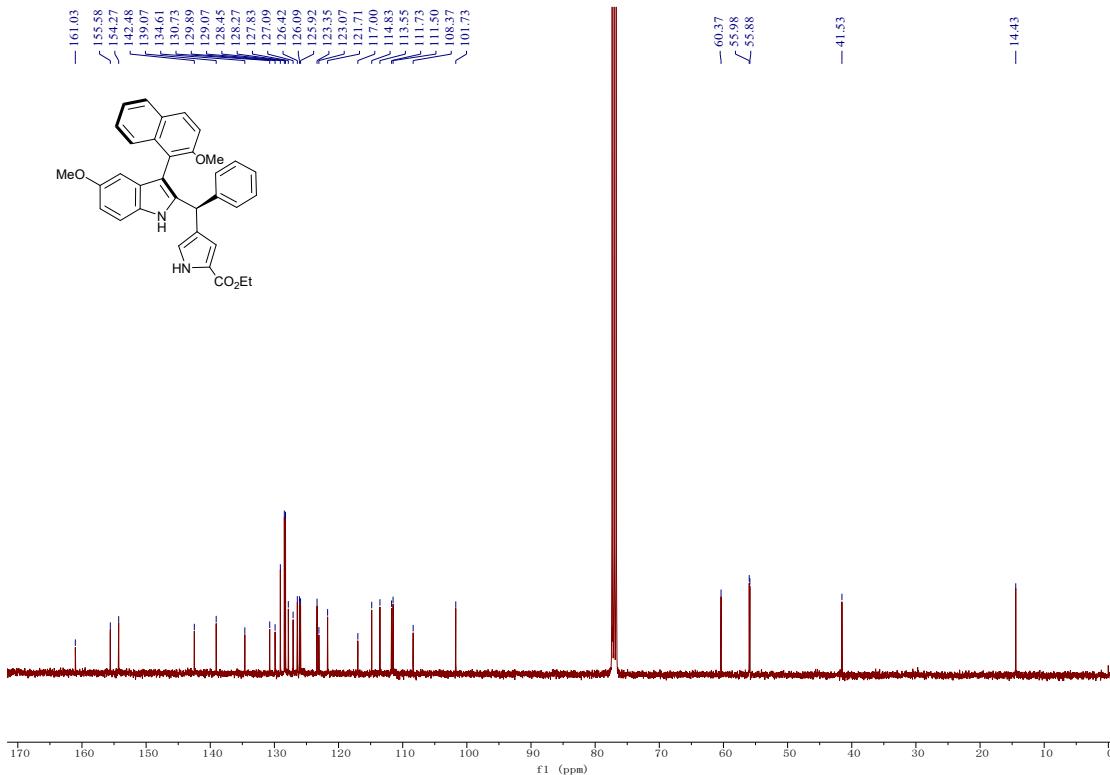
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3h**



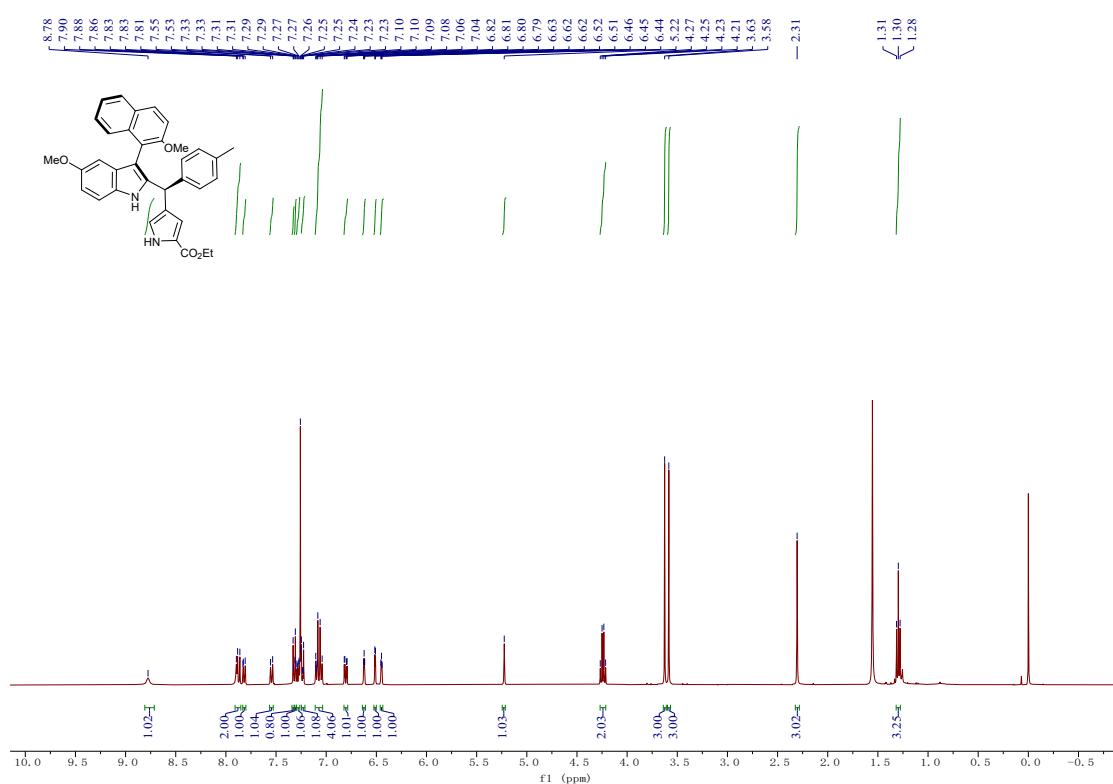
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3i



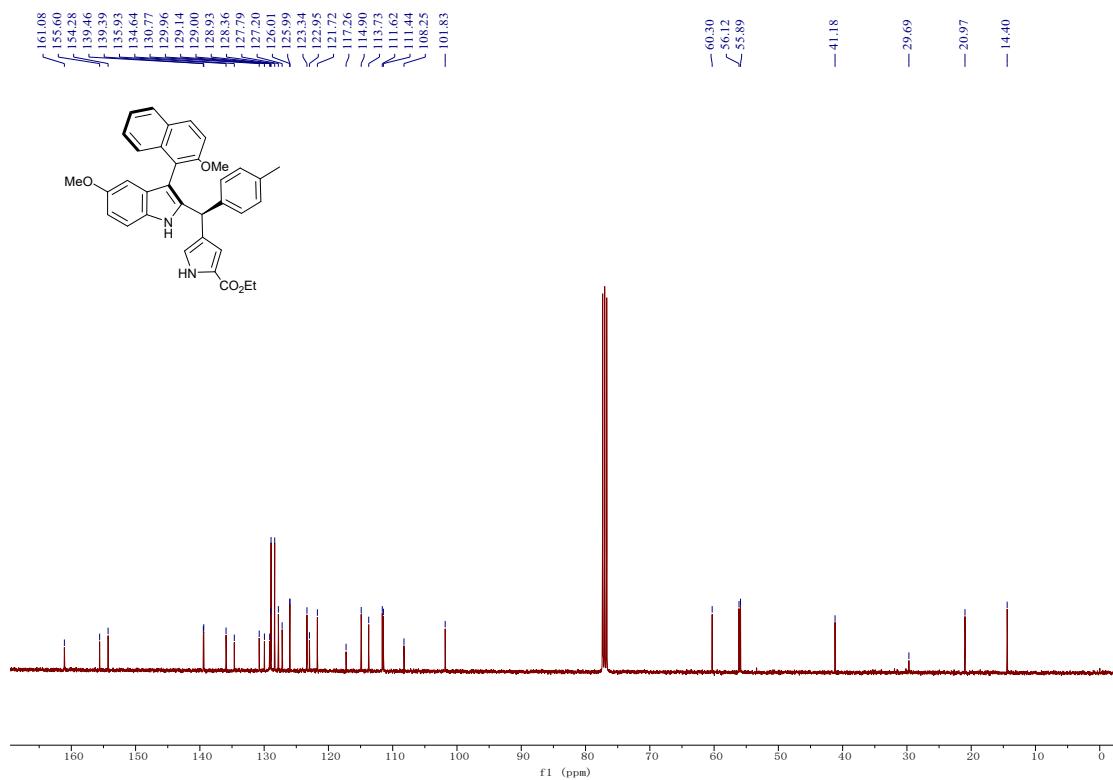
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3i**



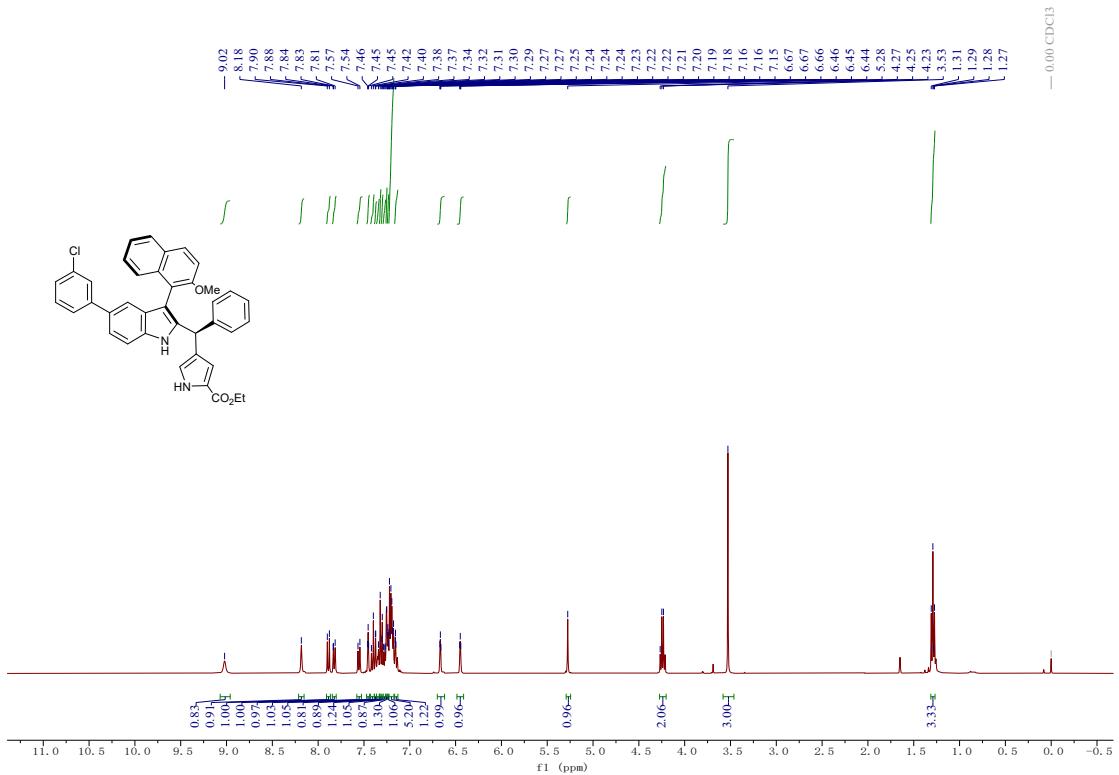
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3j



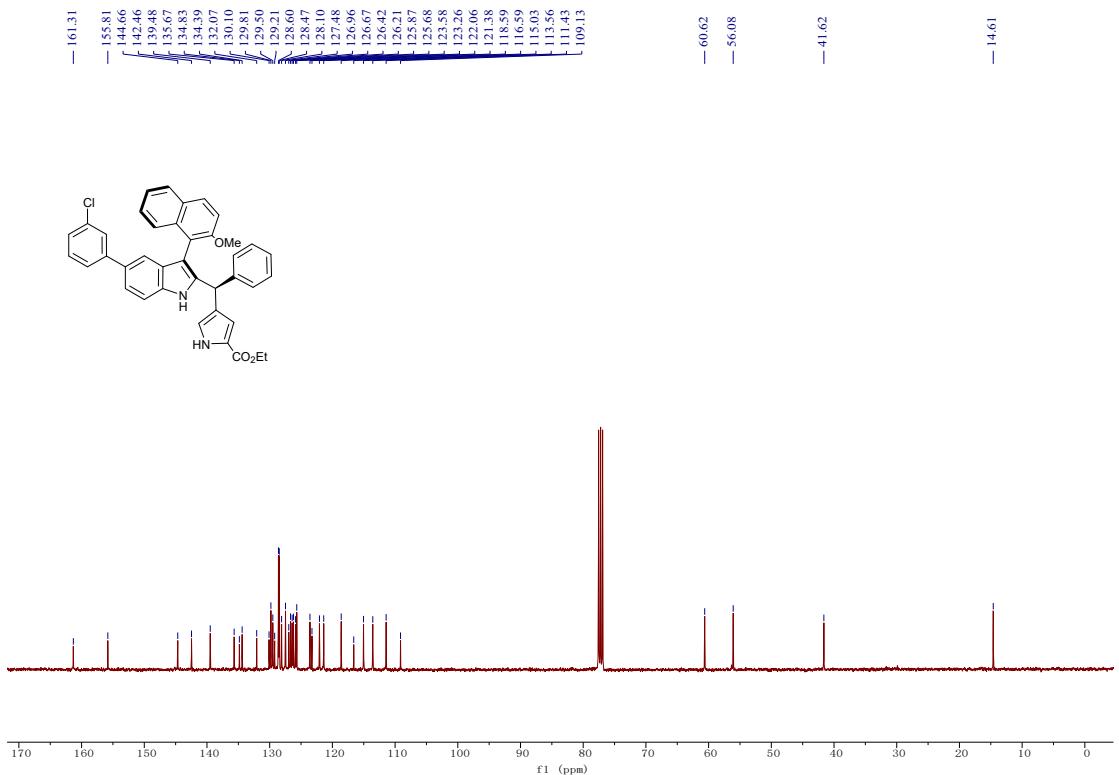
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3j**



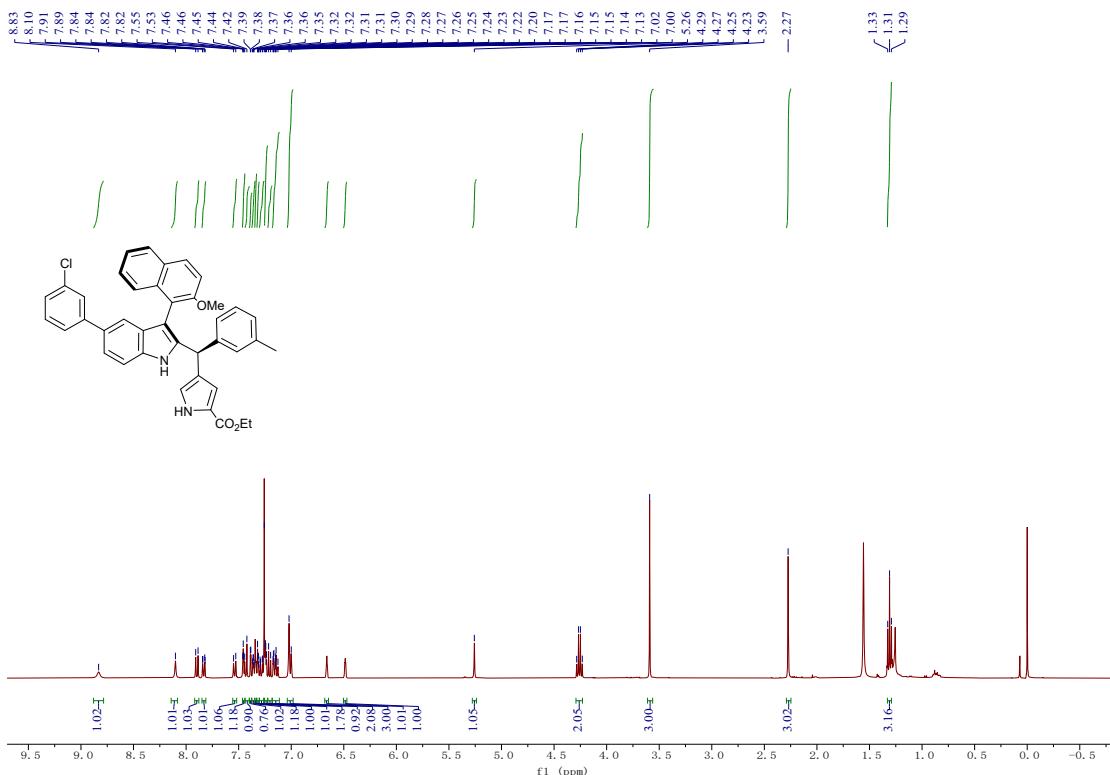
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3k**



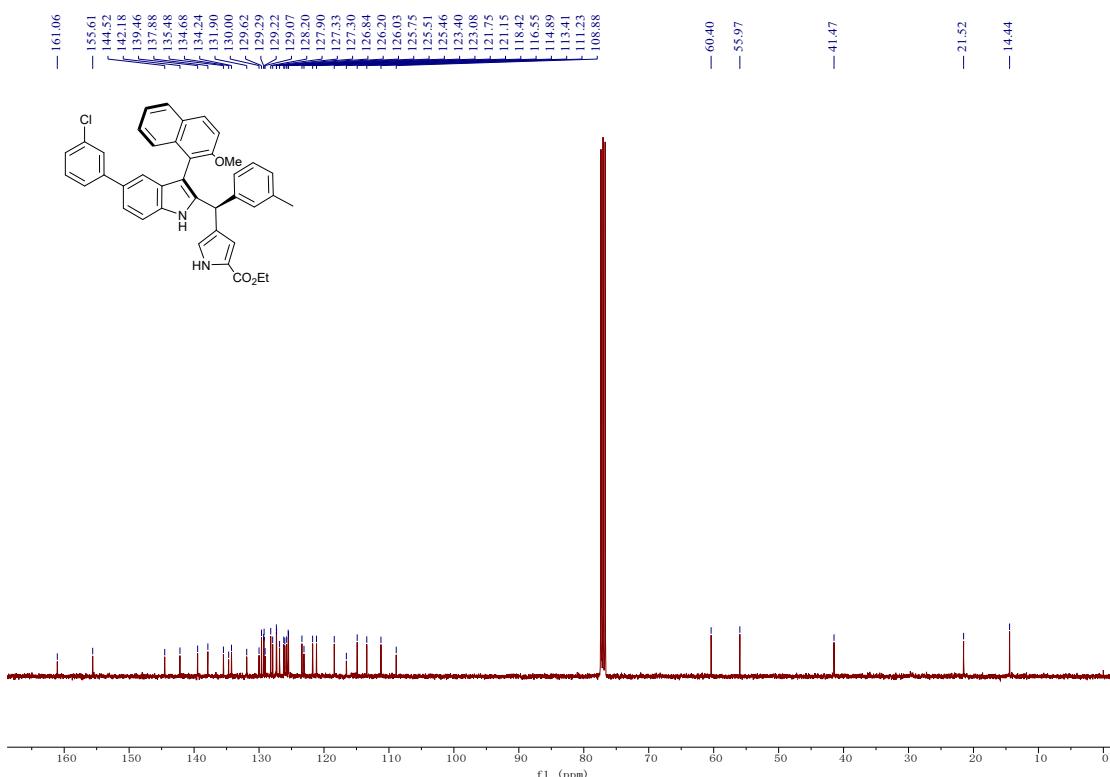
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3k**



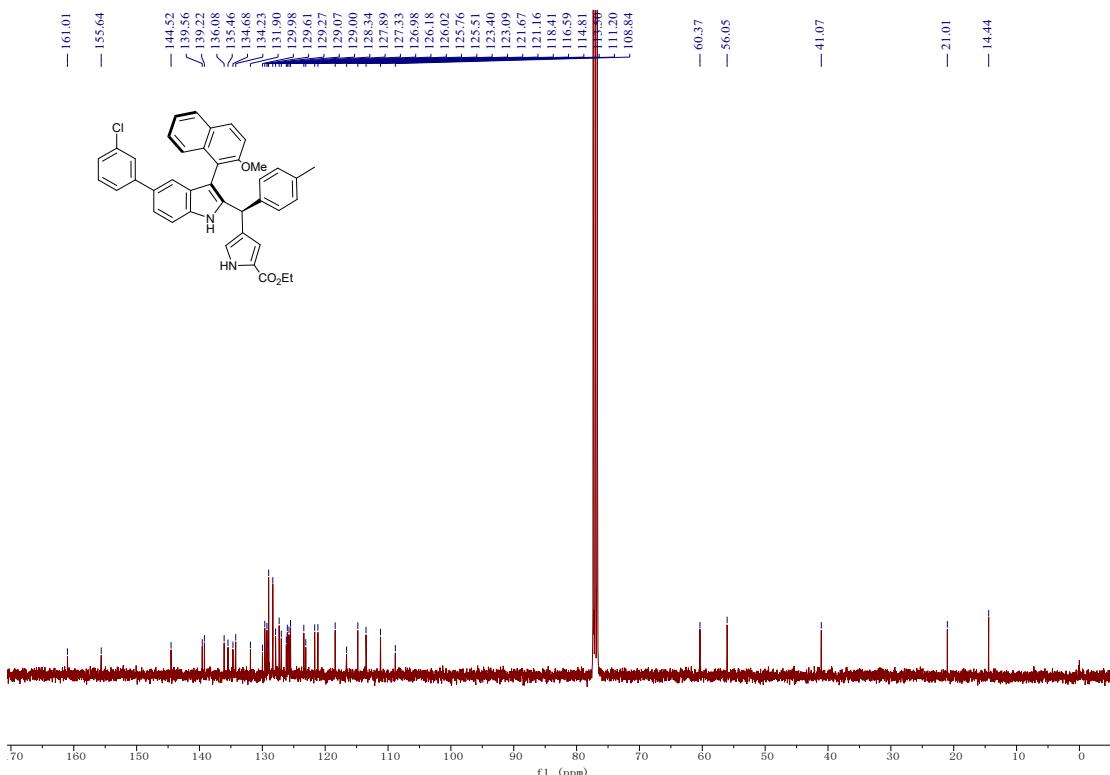
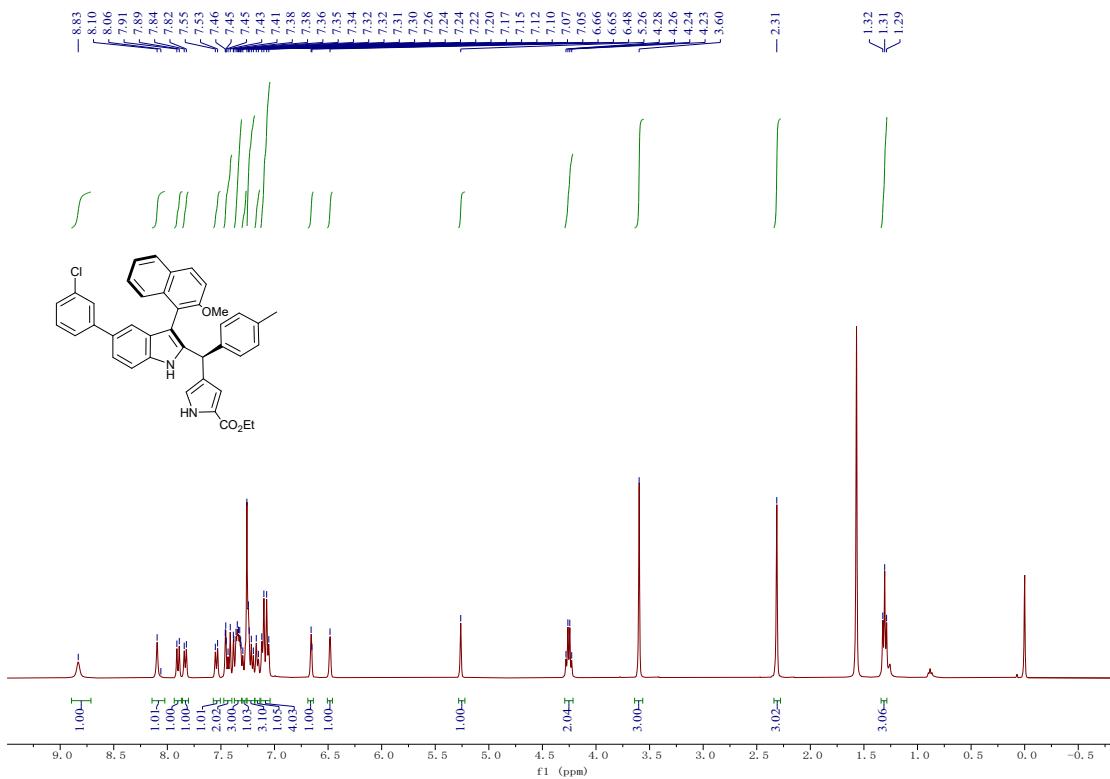
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3l



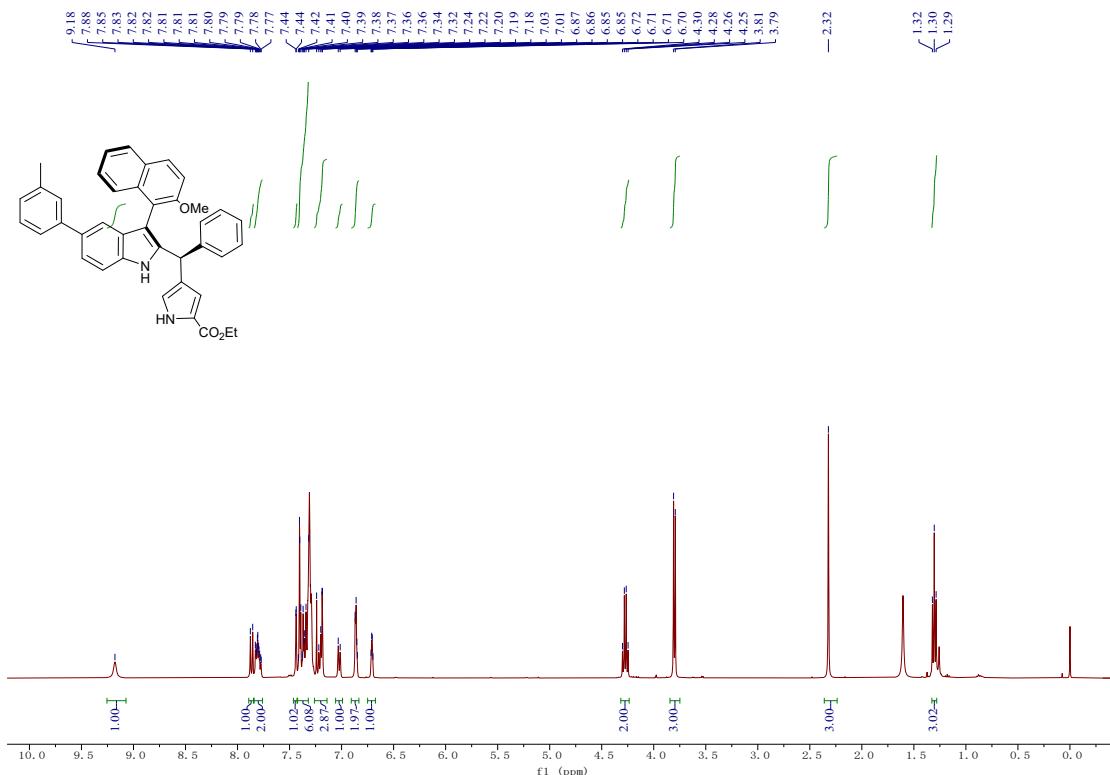
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3l**



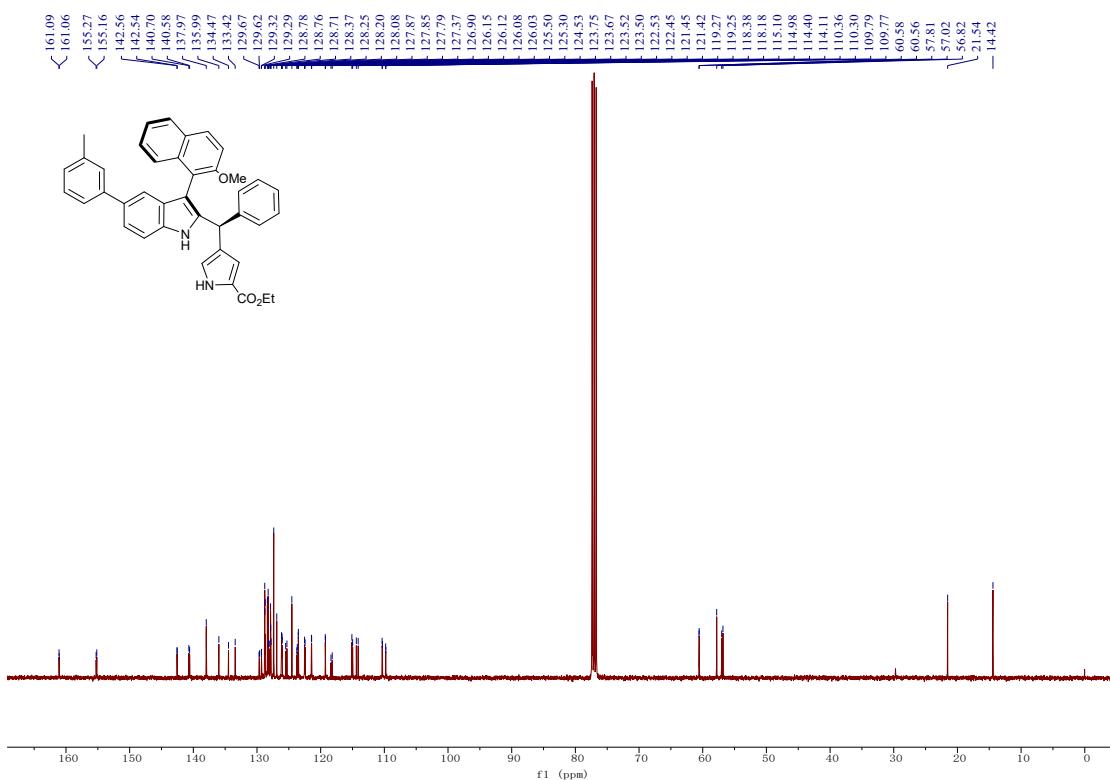
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3m**



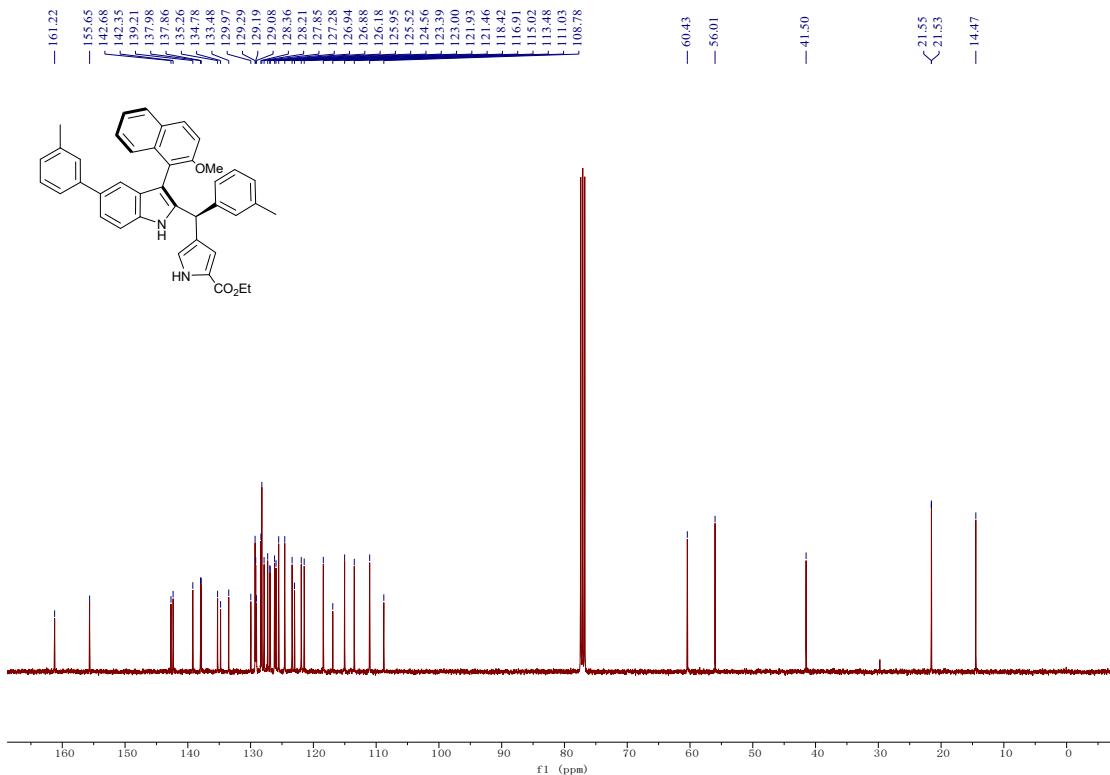
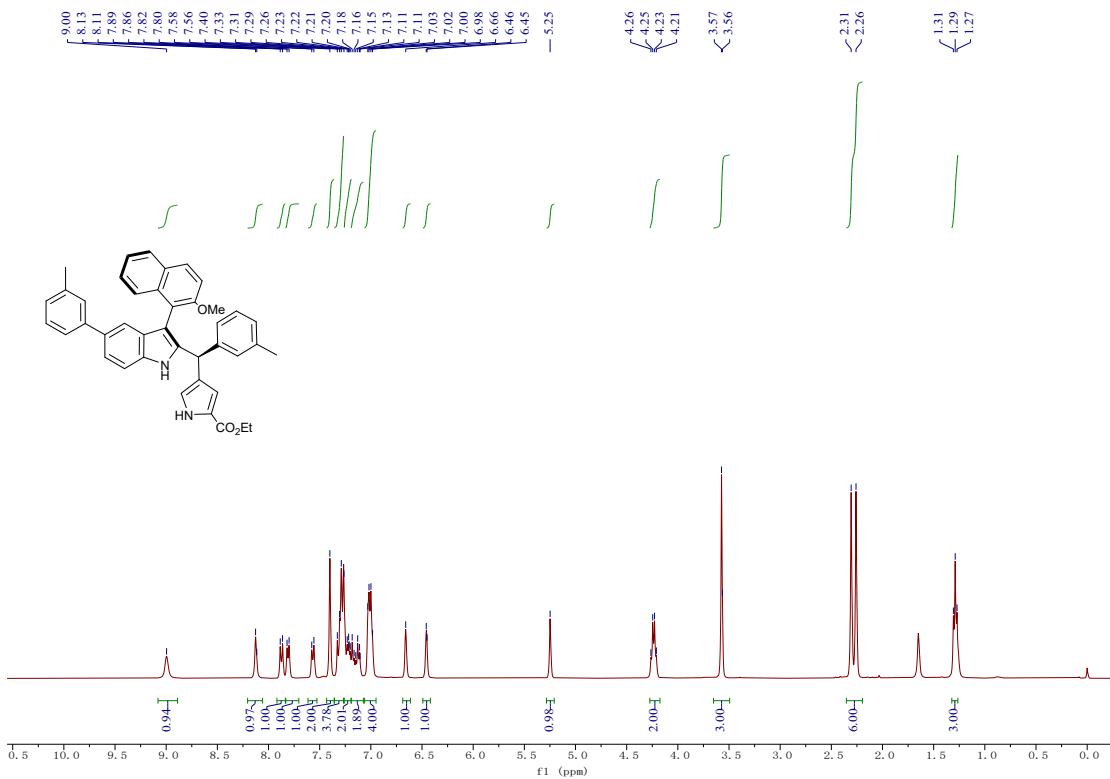
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3n**



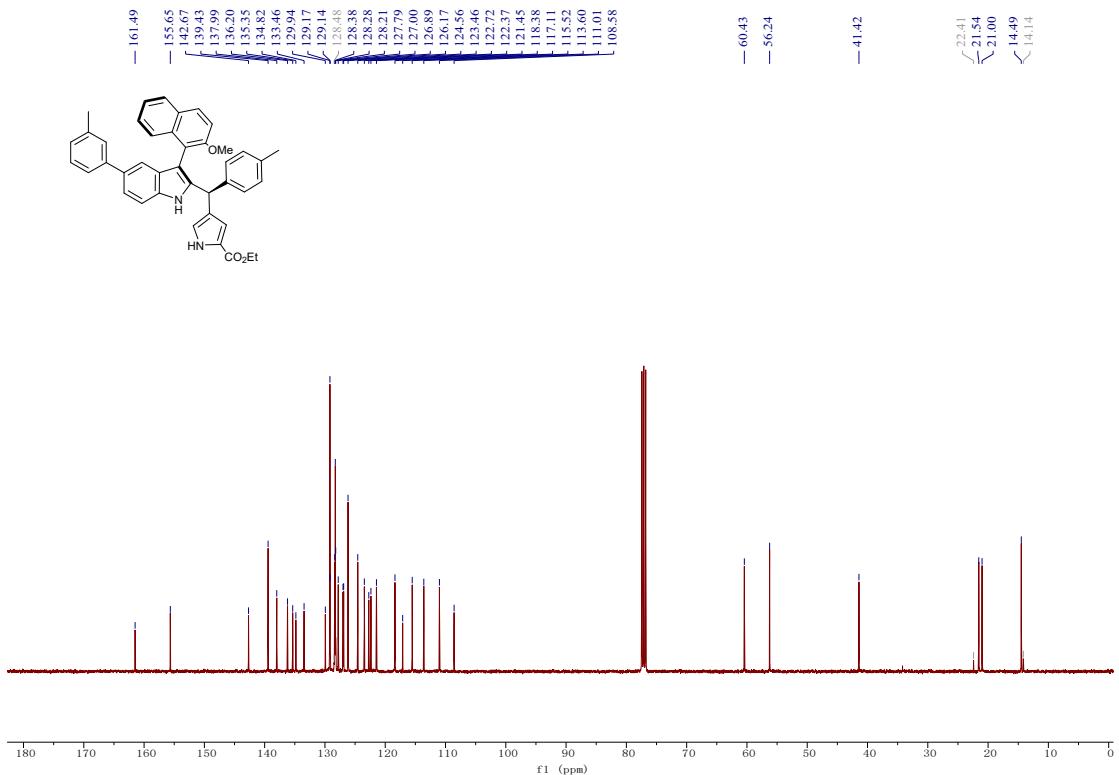
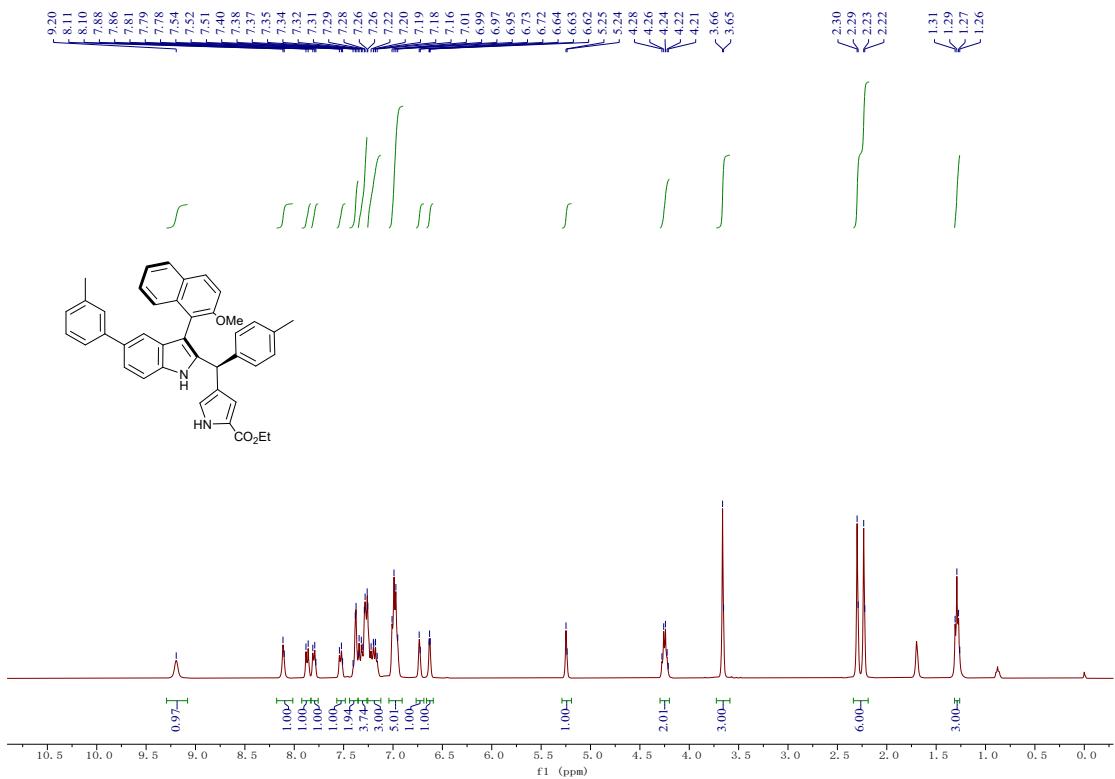
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3n**



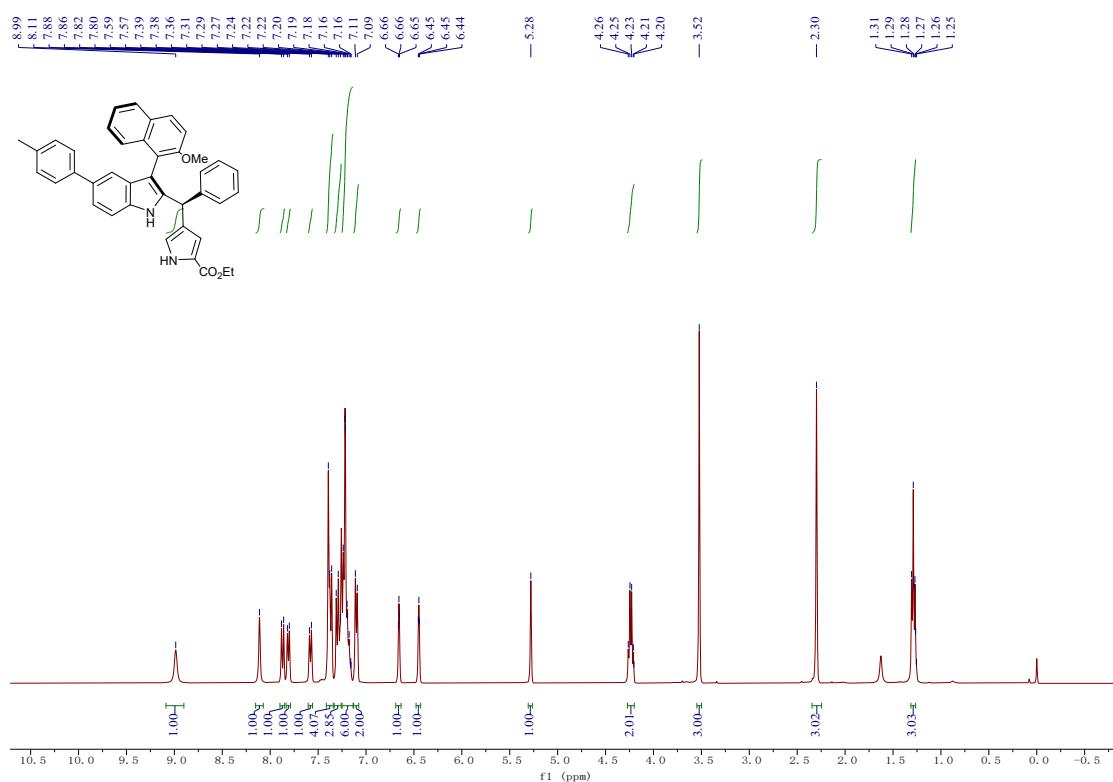
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3o**



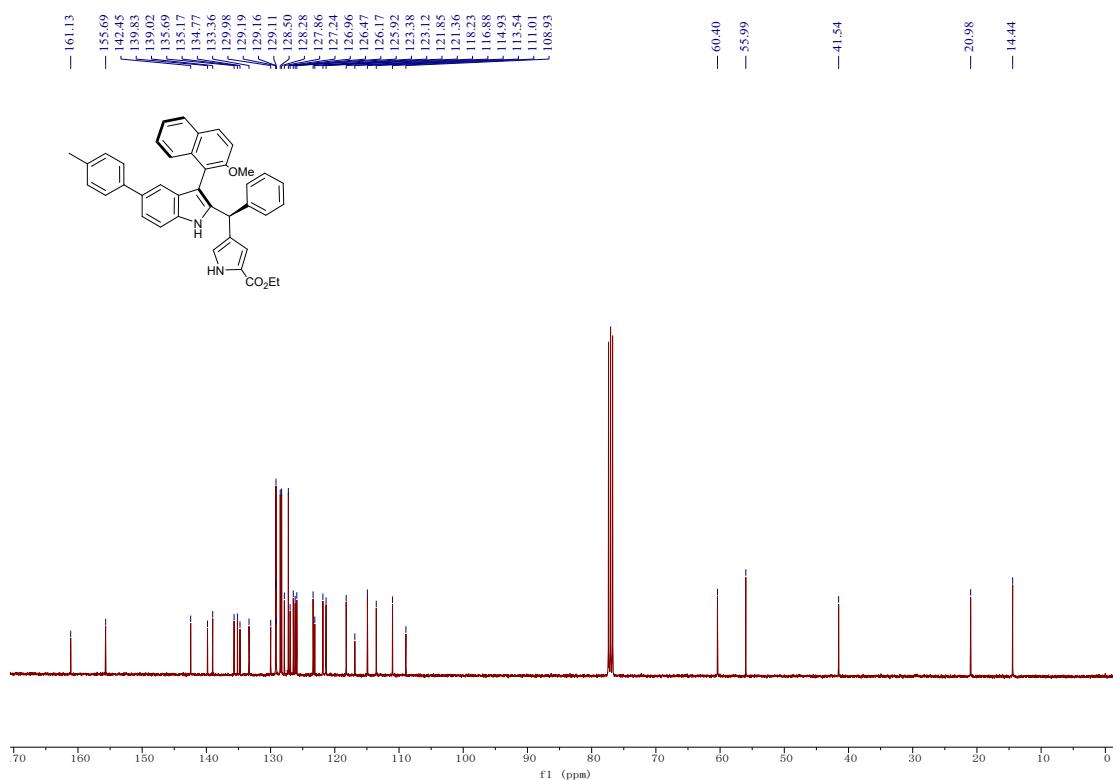
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3p**



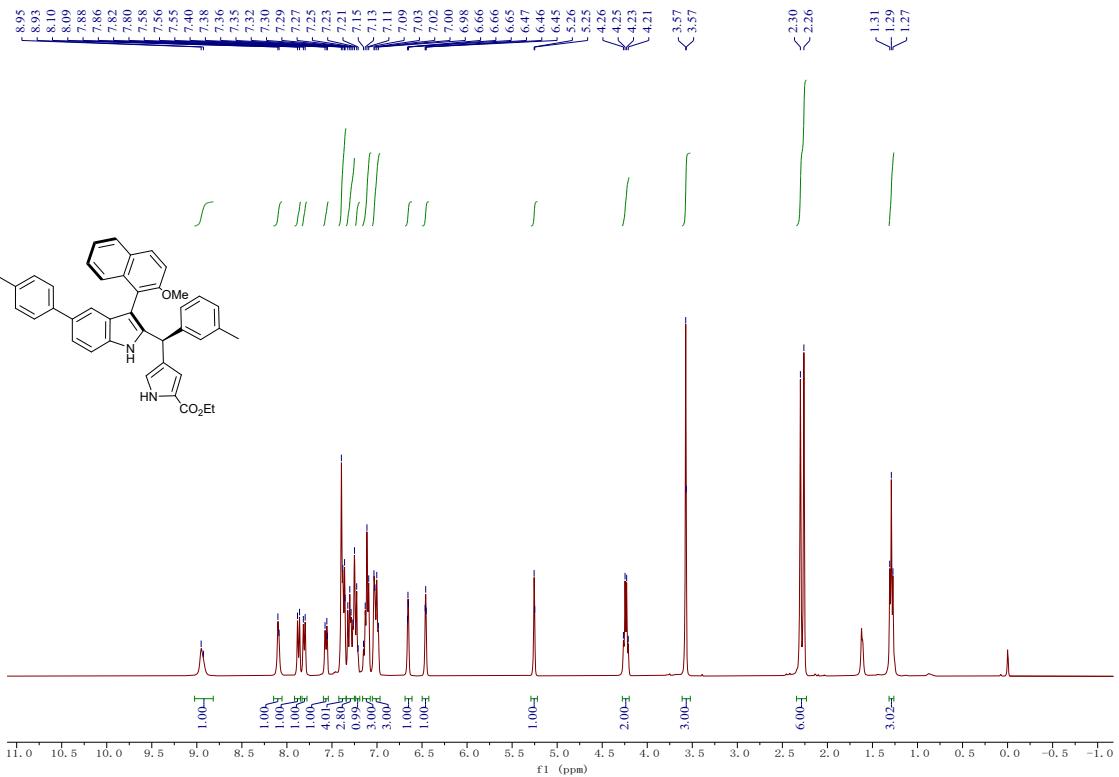
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3q**



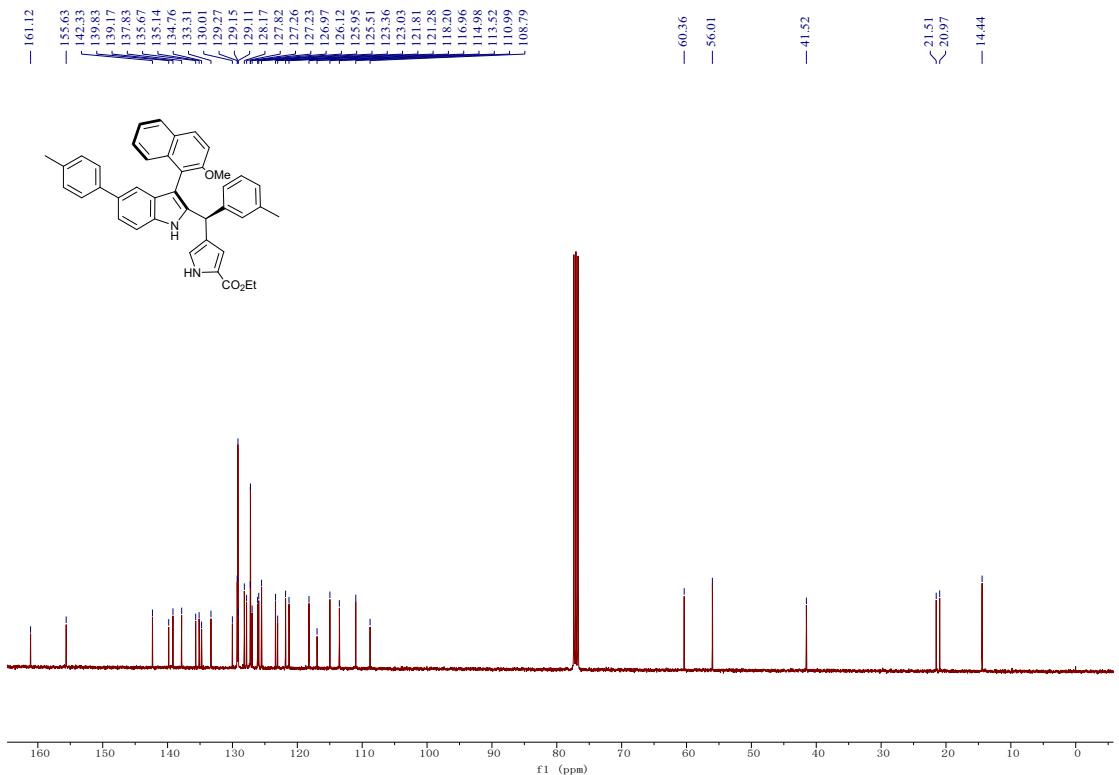
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3q



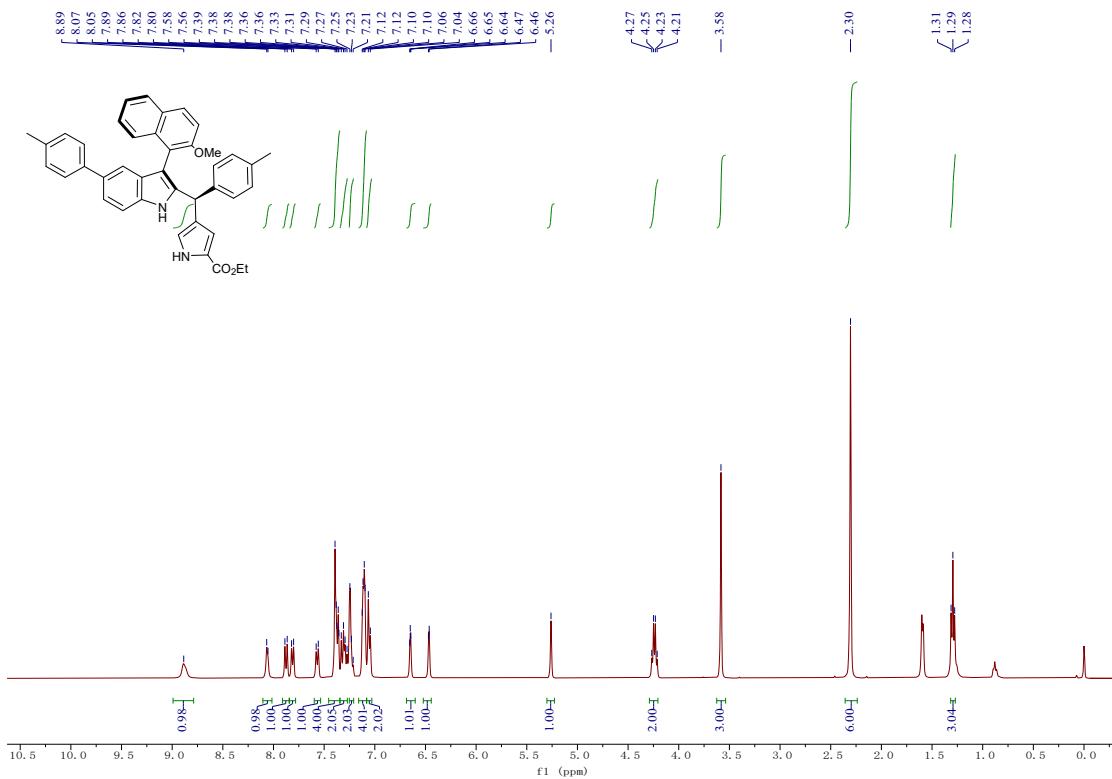
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3r**



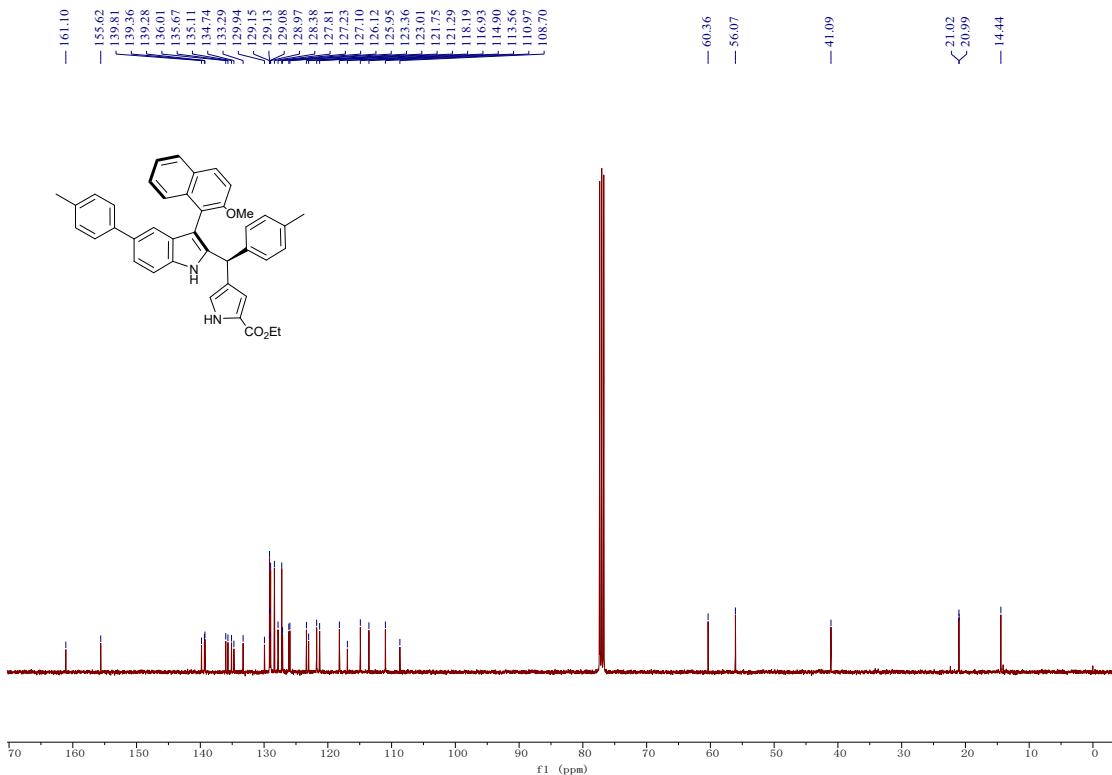
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3r**



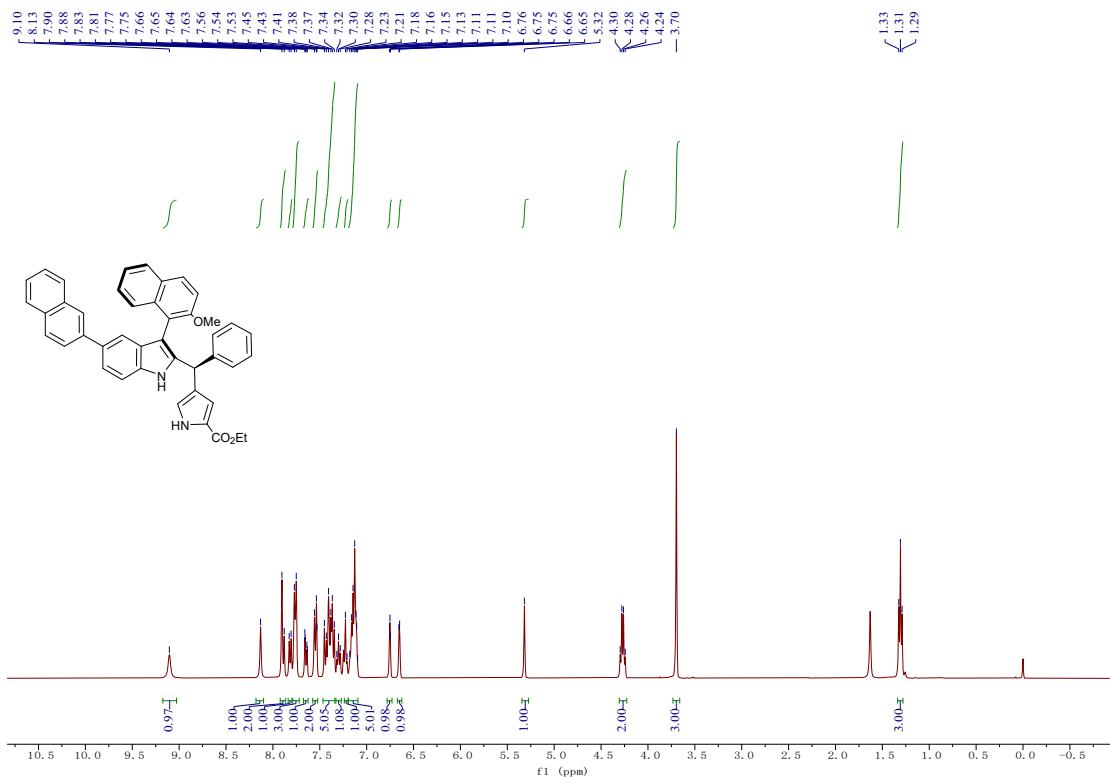
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3s**



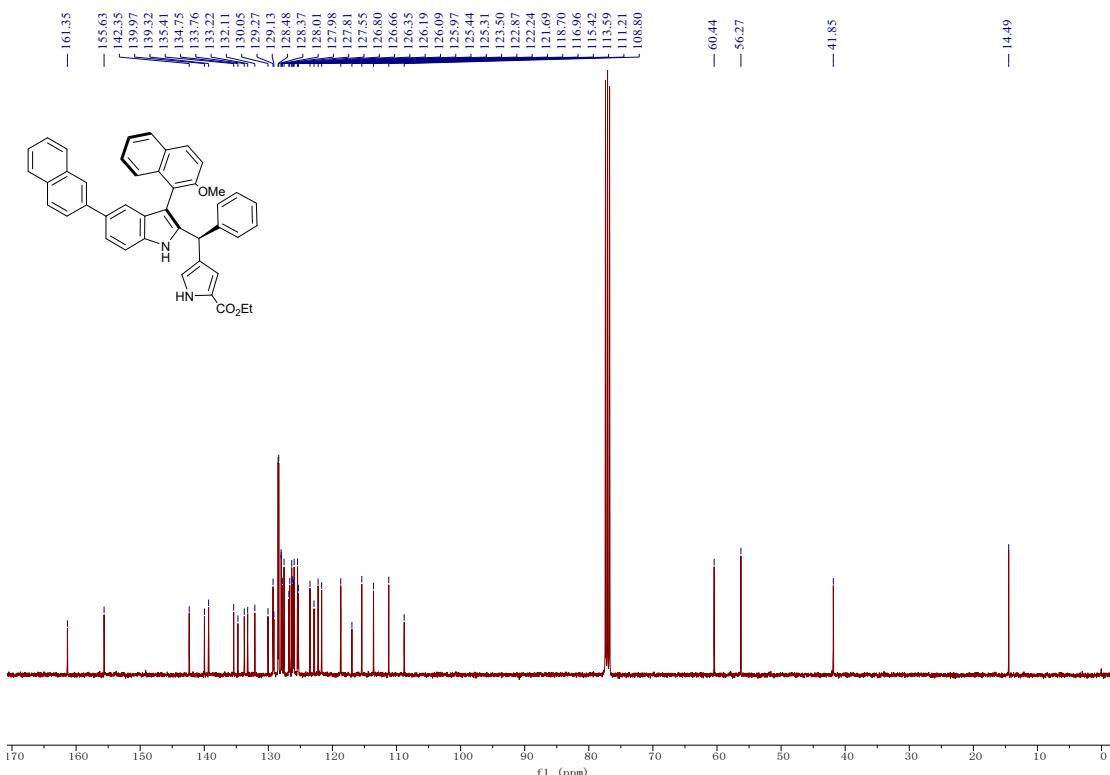
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3s**



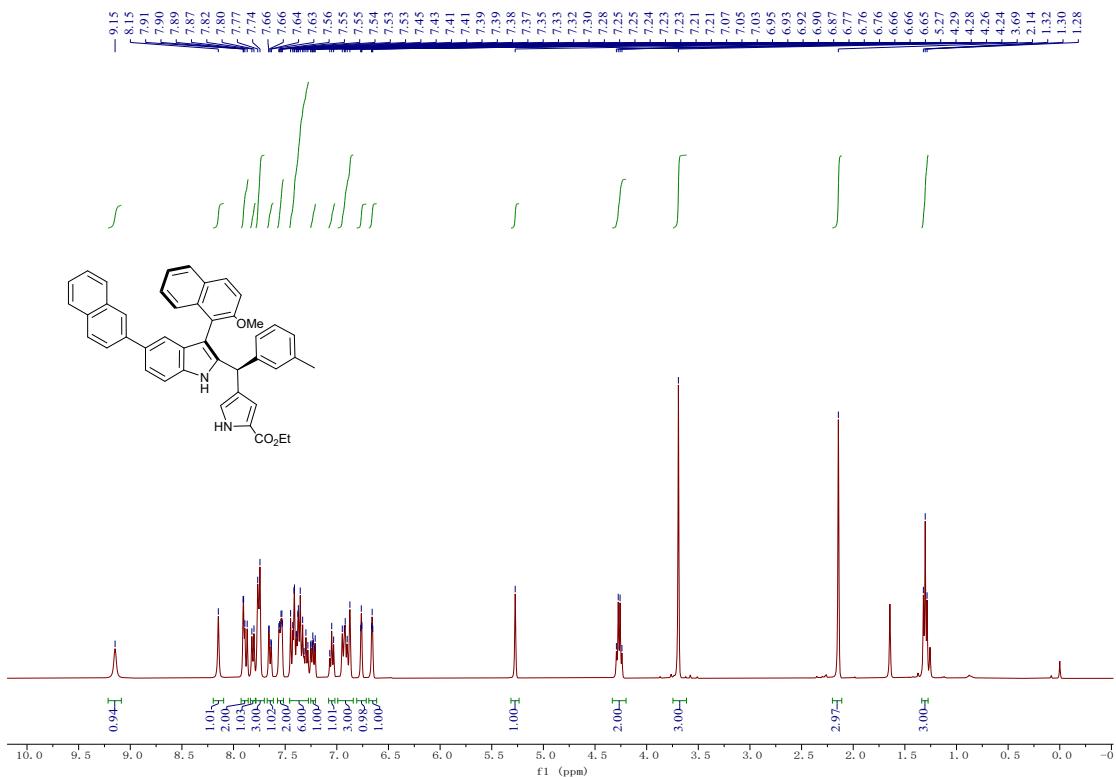
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3t



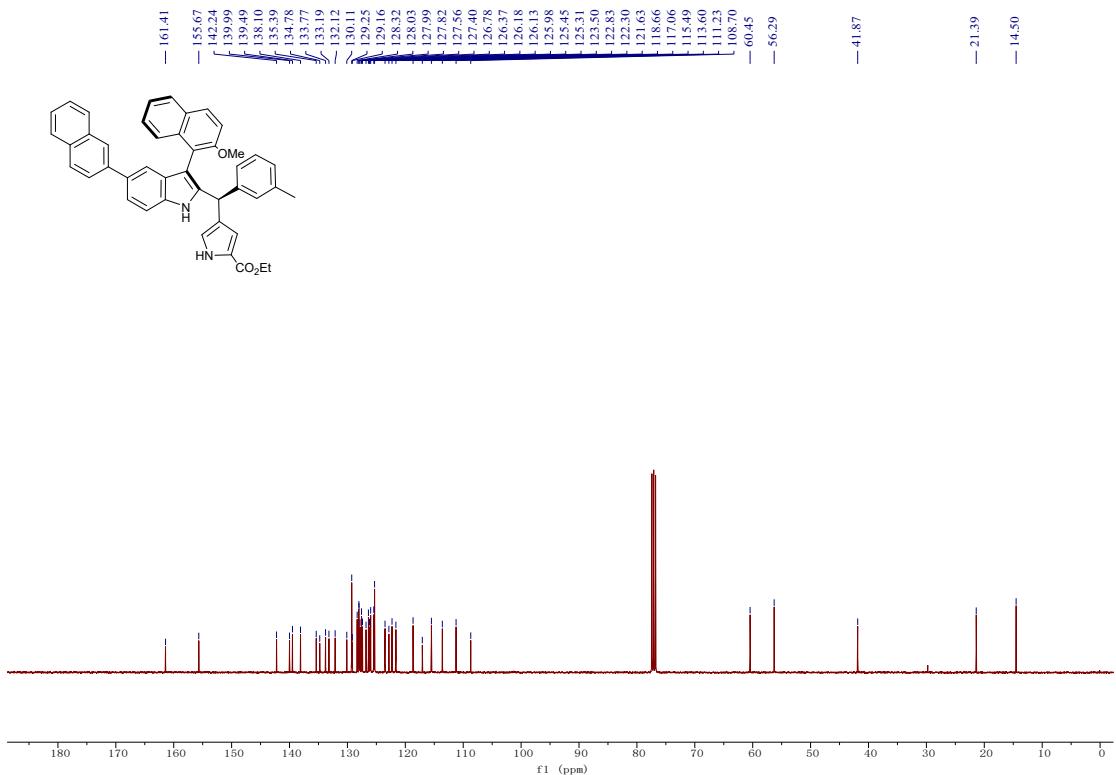
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3t



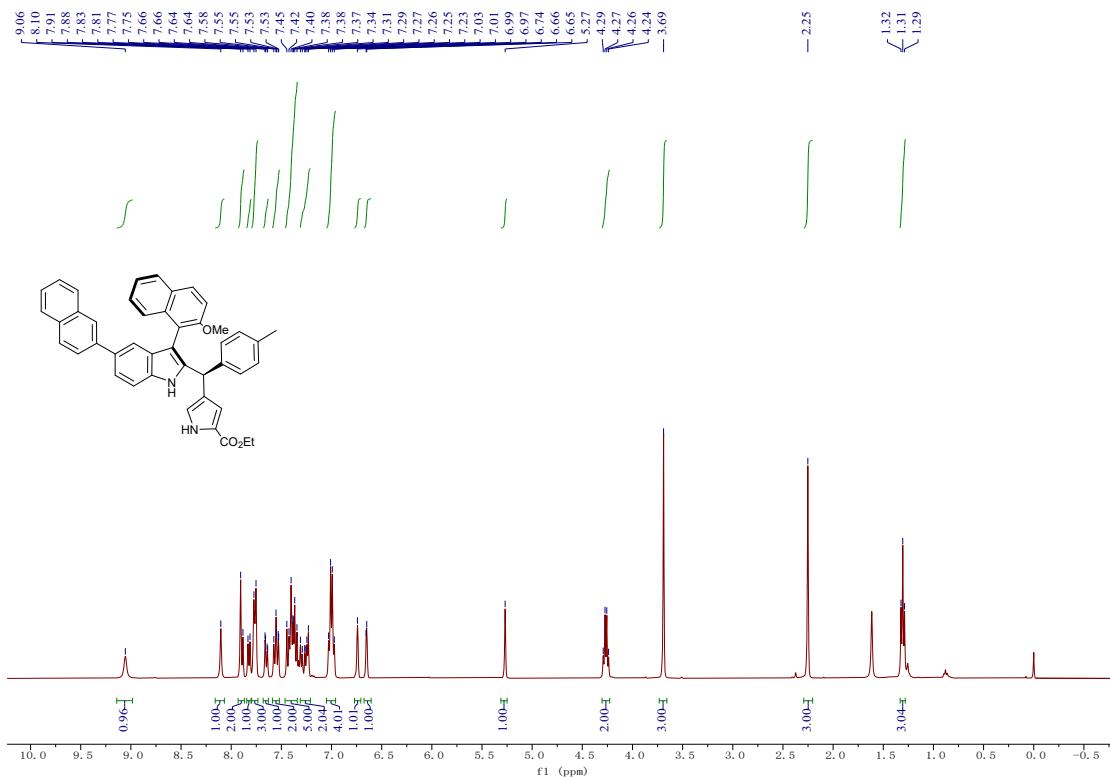
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3u**



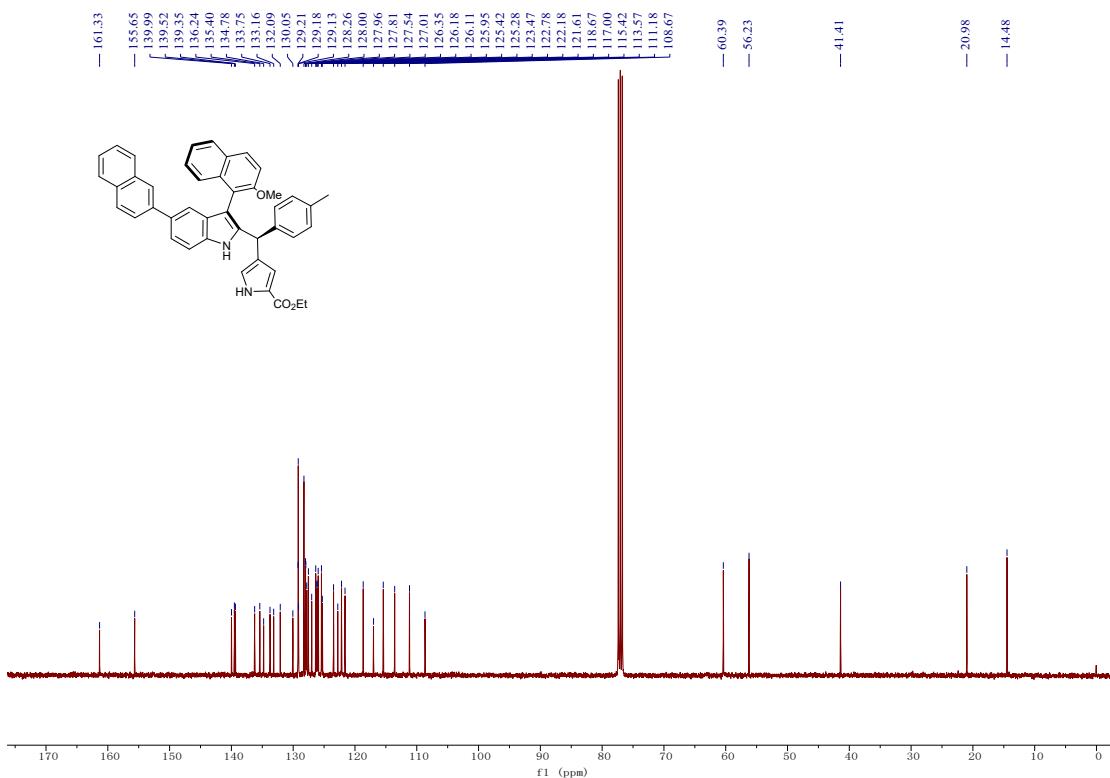
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3u**



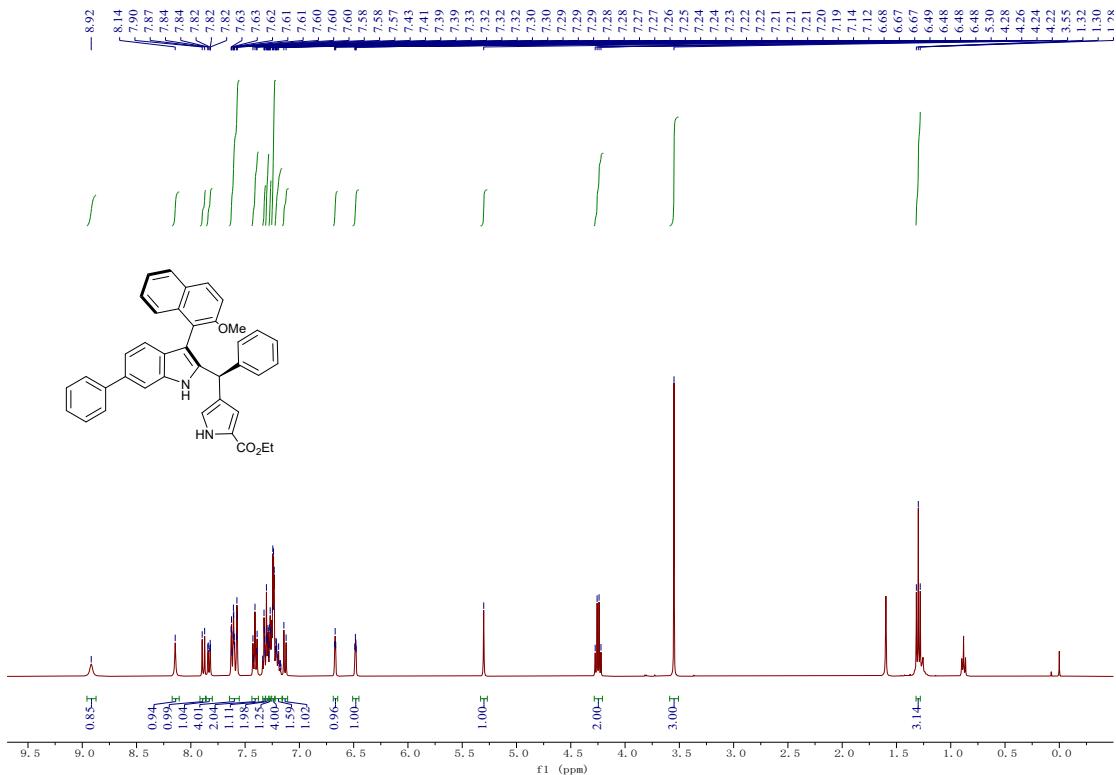
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3v



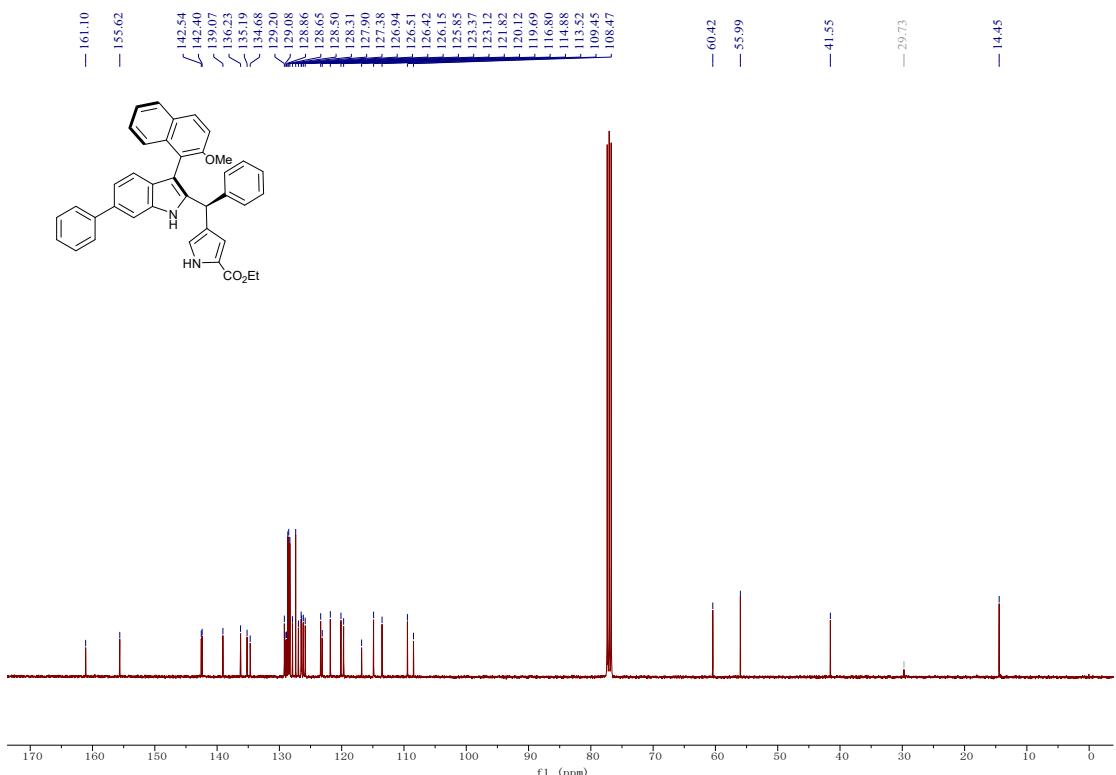
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3v



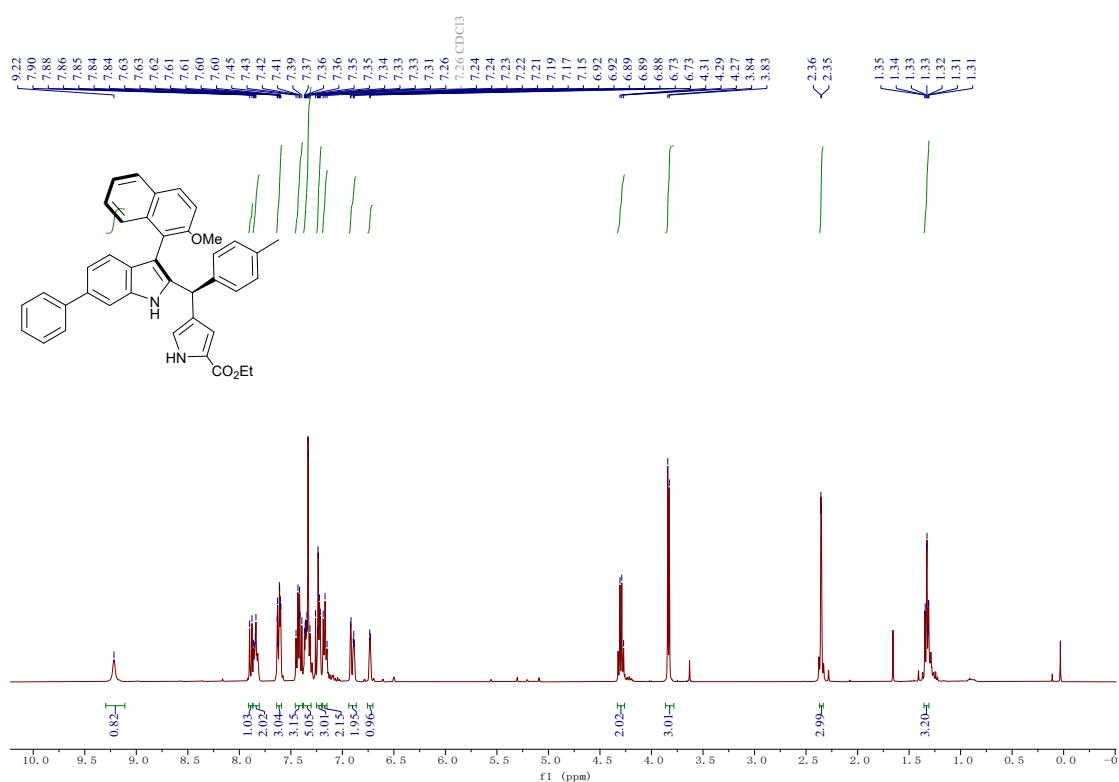
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3w**



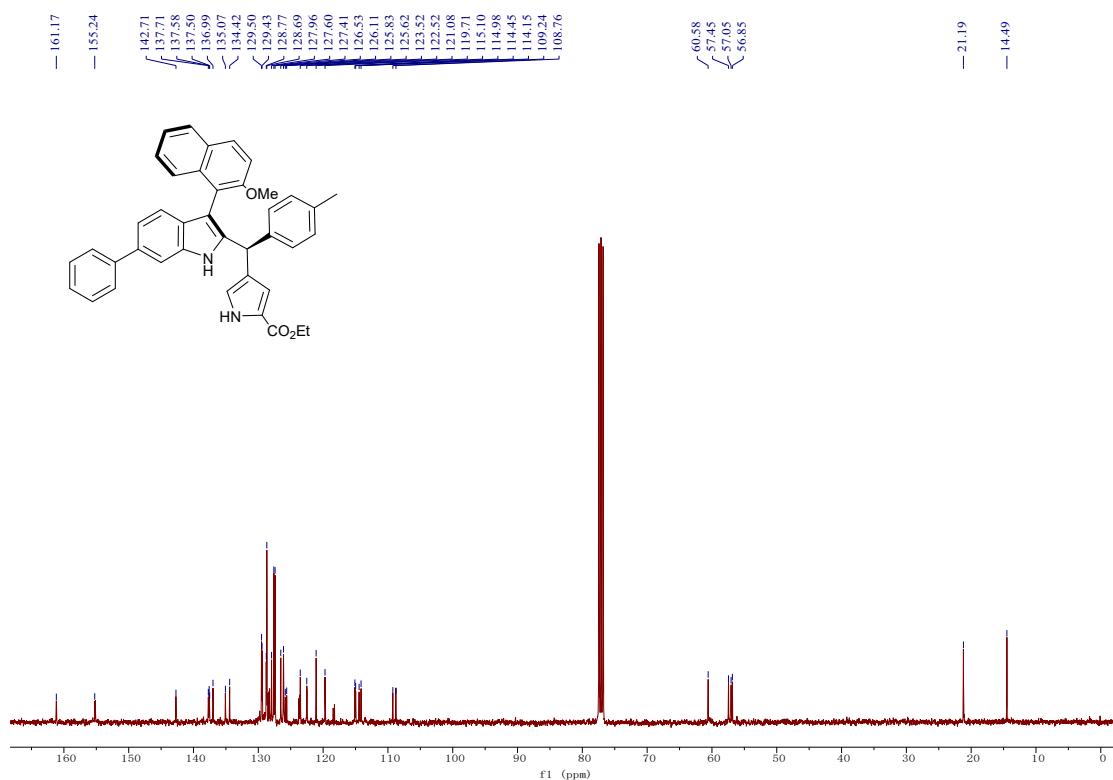
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3w**



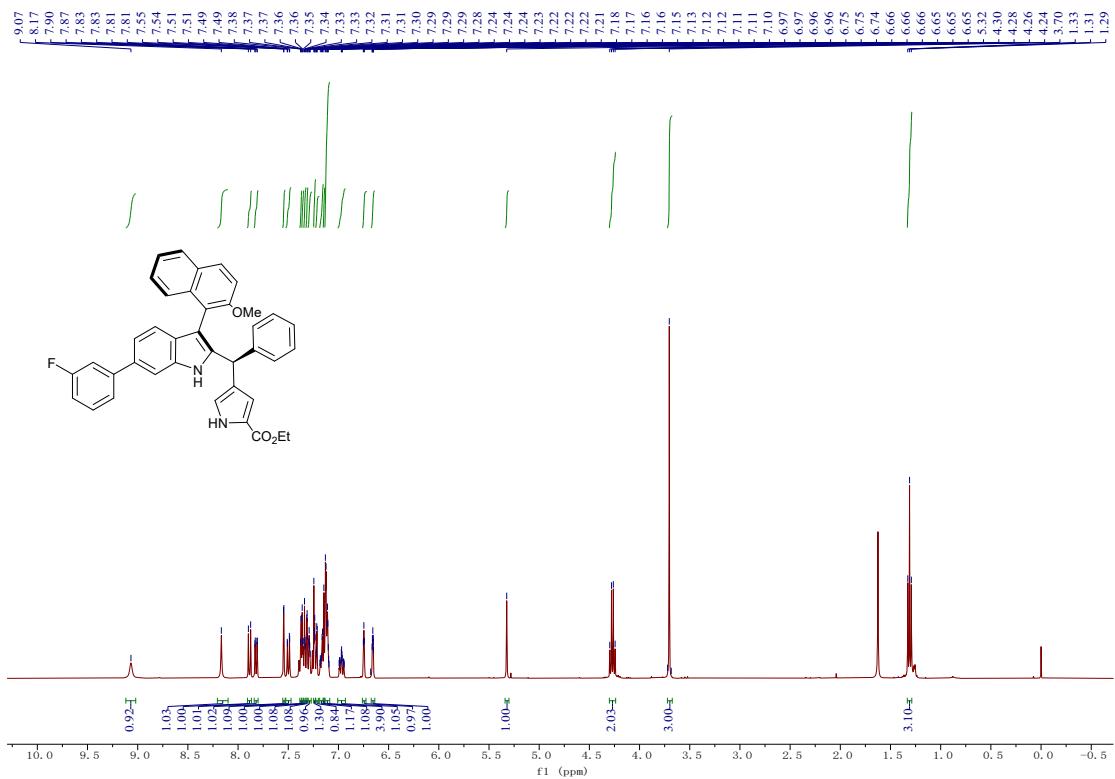
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3x**



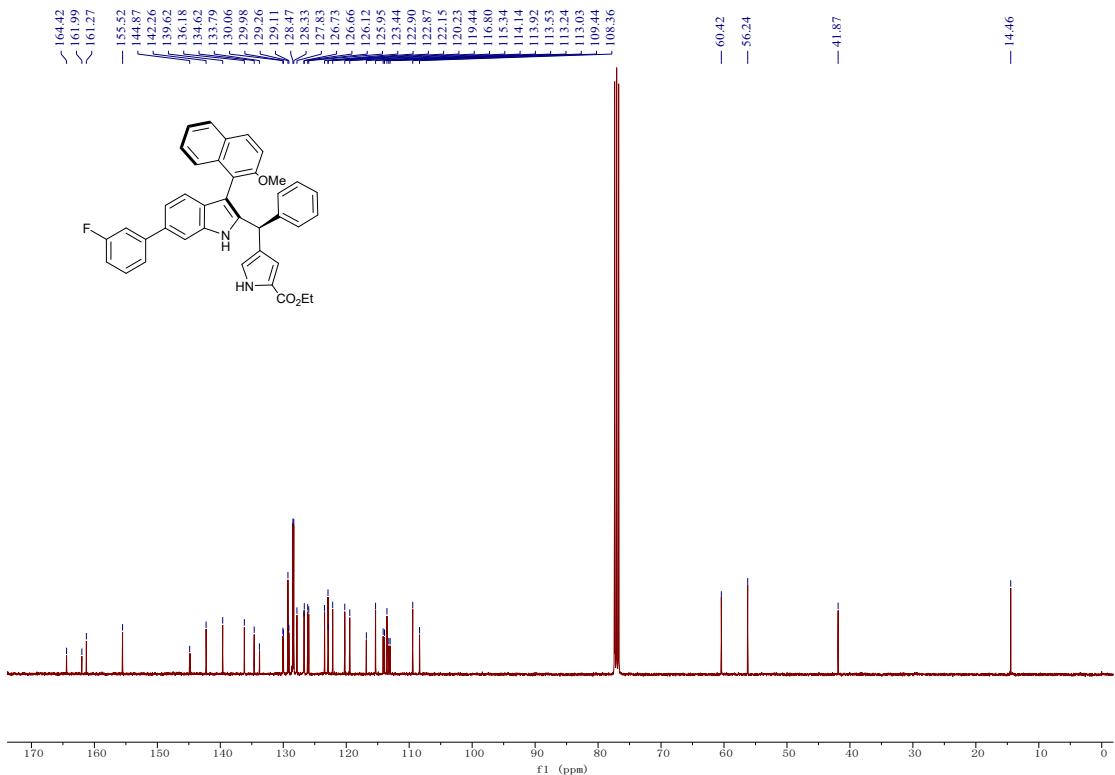
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3x**



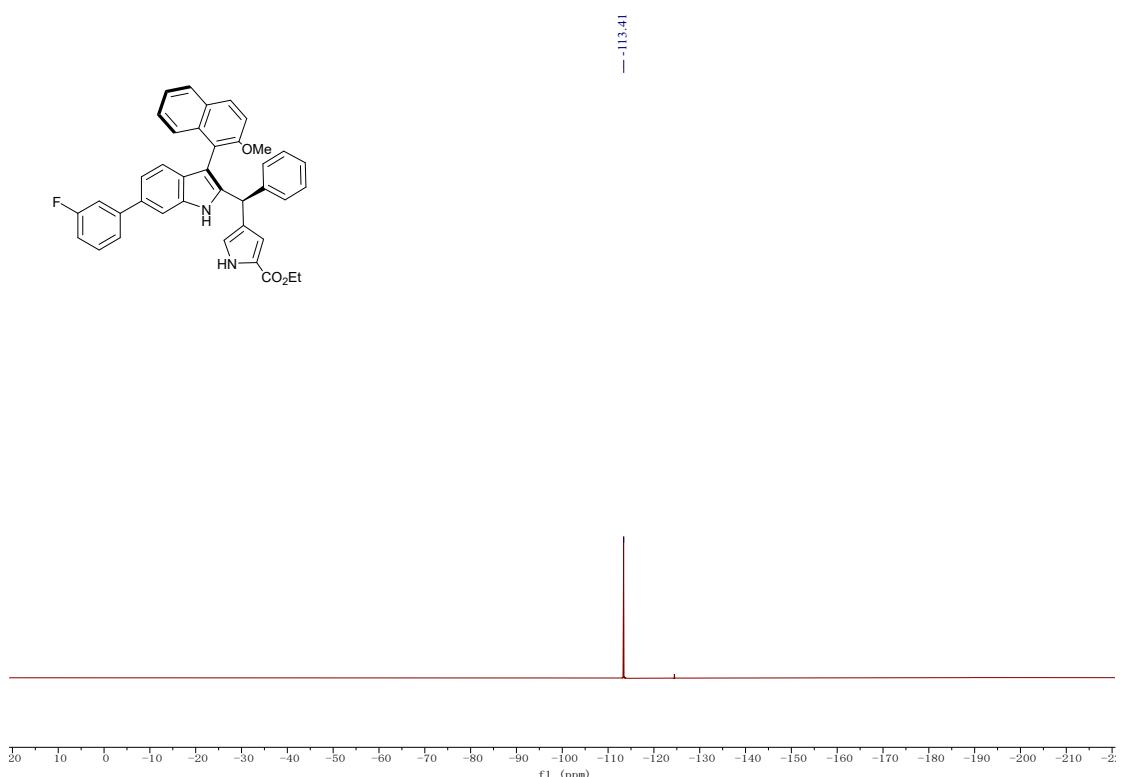
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3y



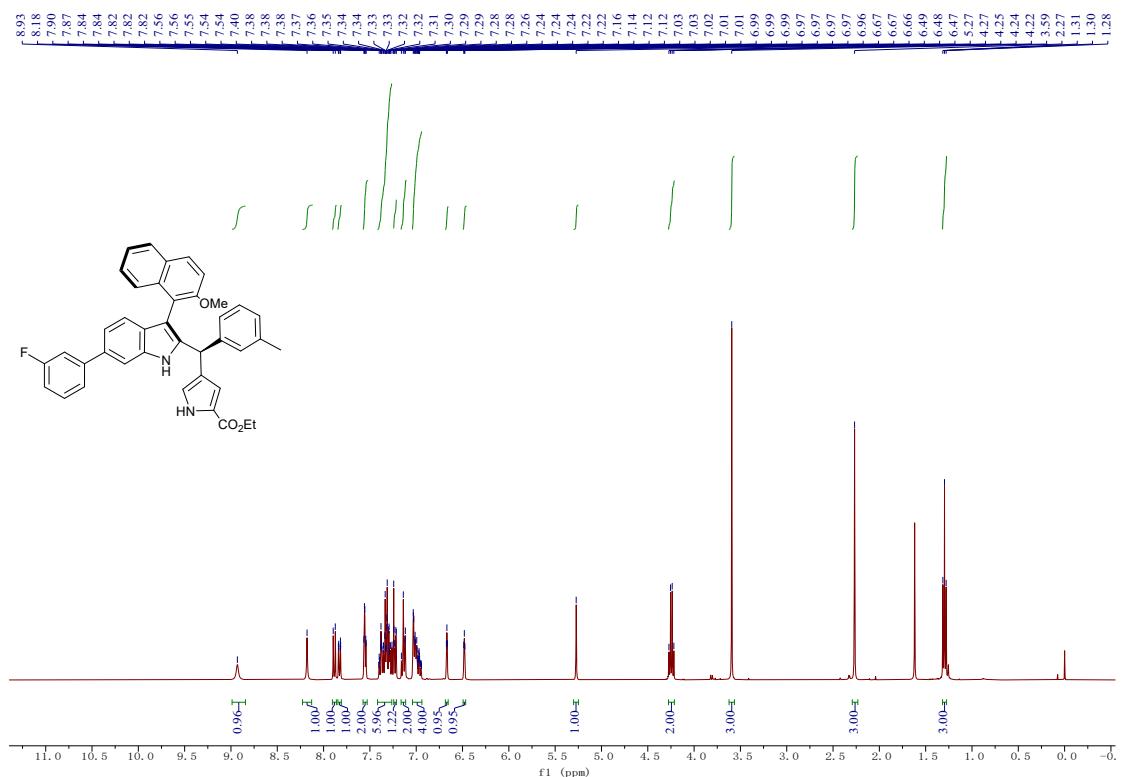
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3y



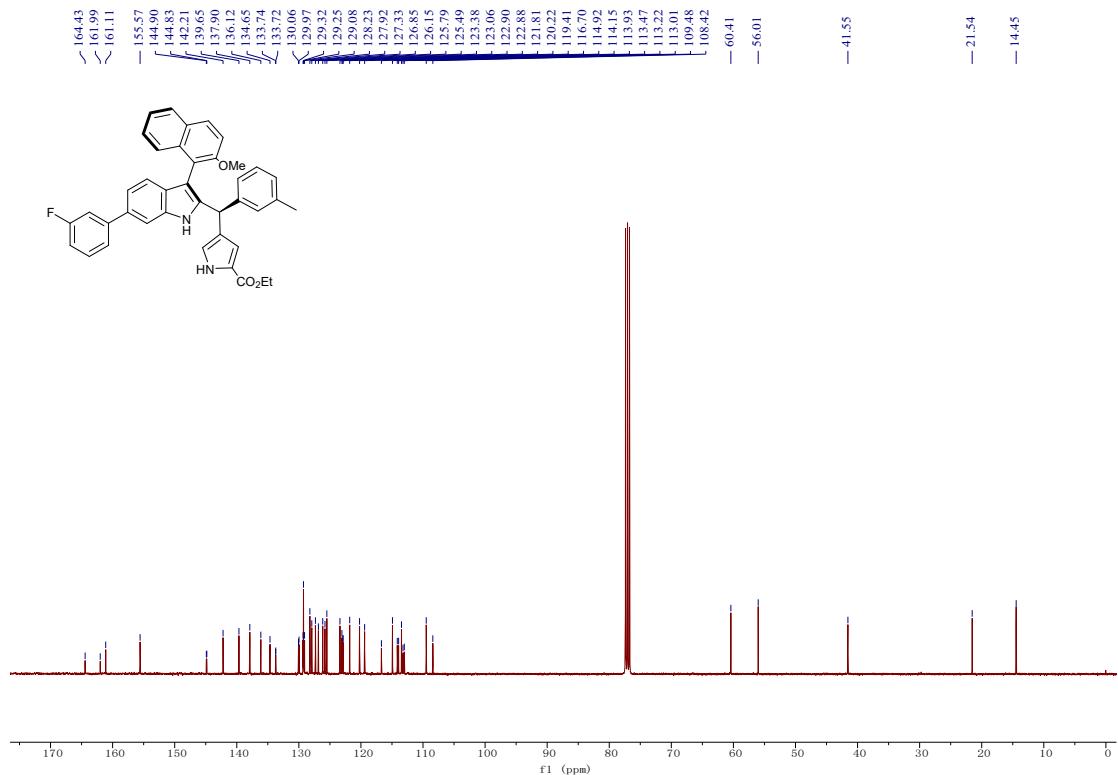
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3y**



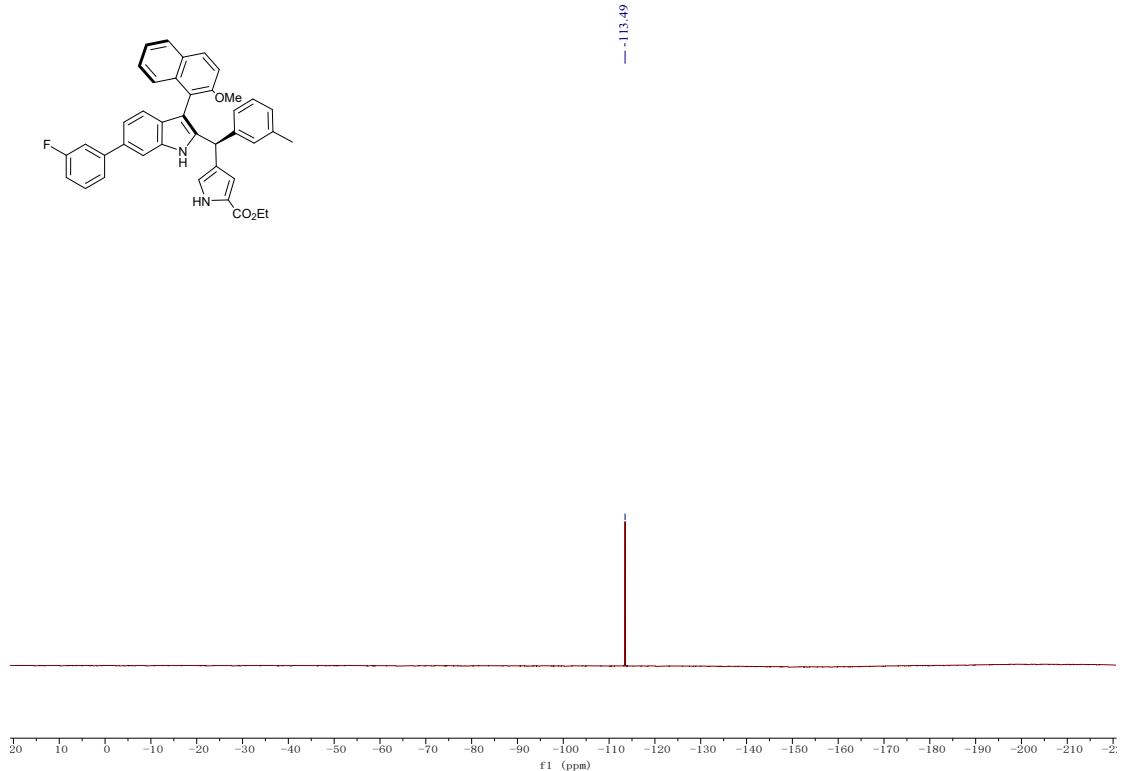
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3z**



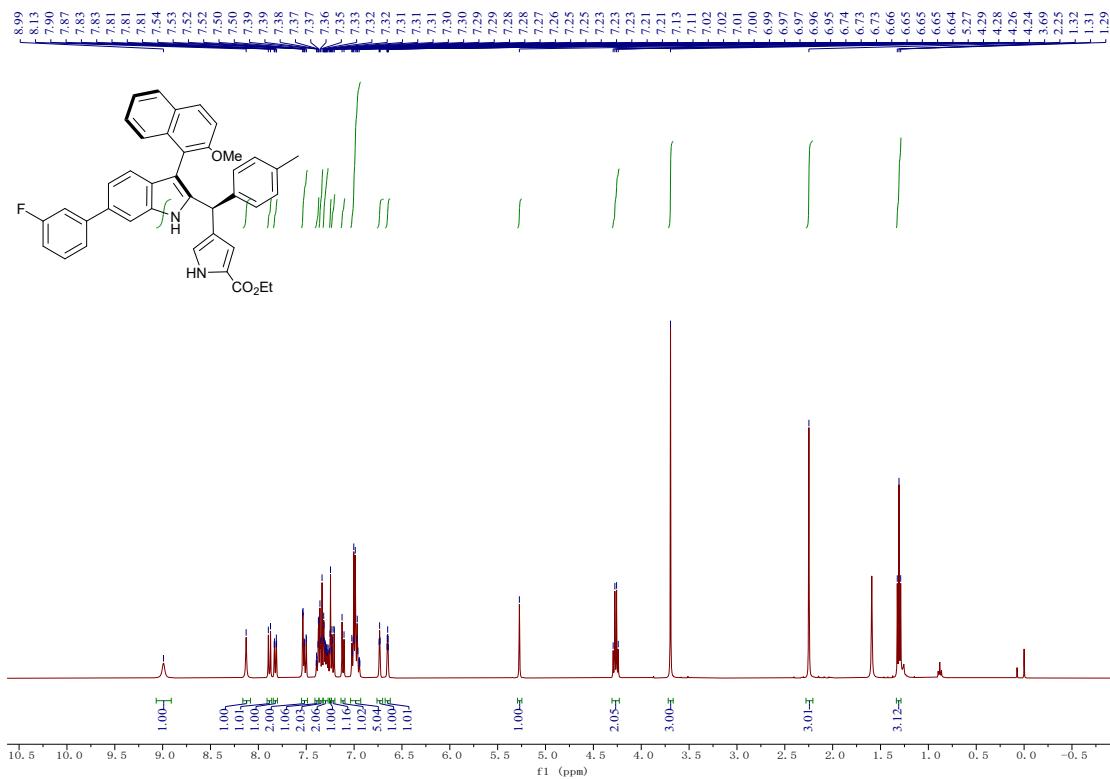
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3z**



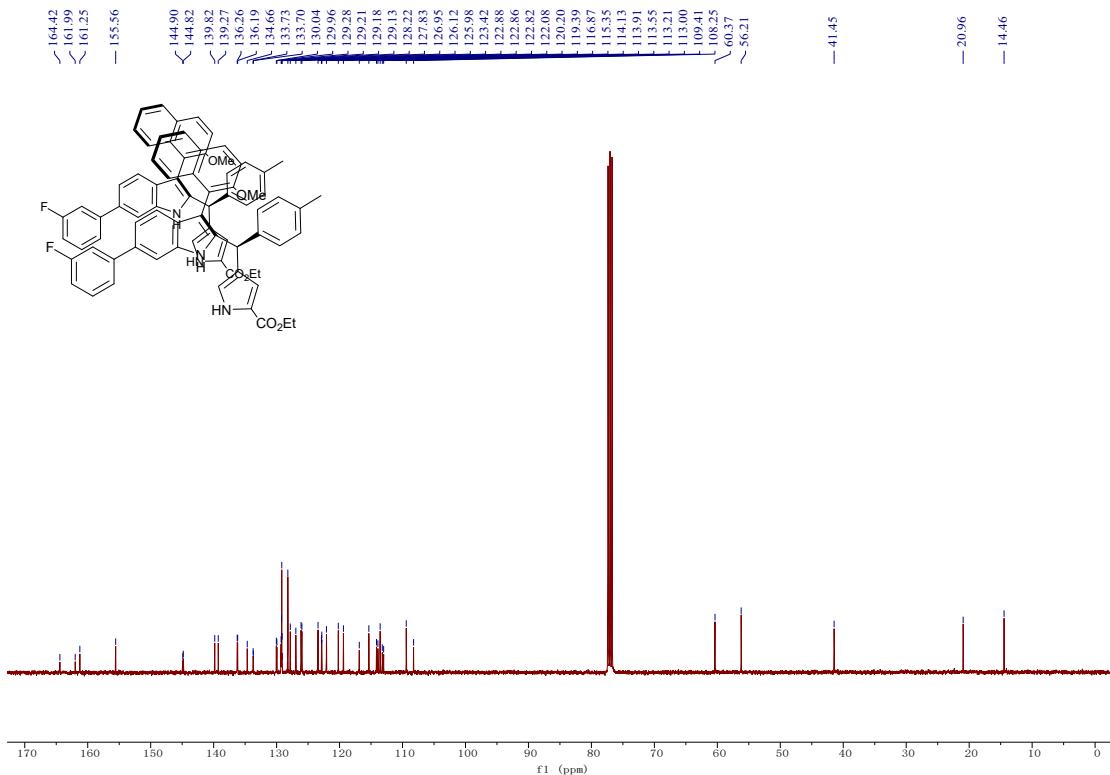
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3z**



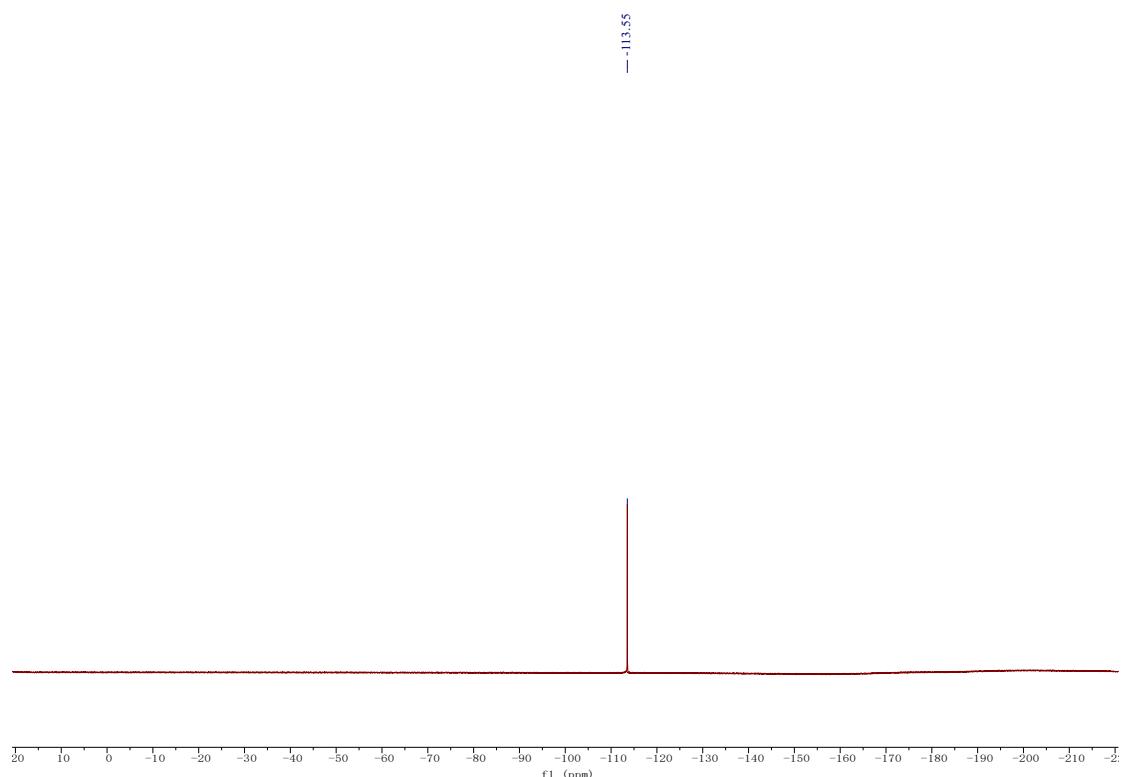
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3za**



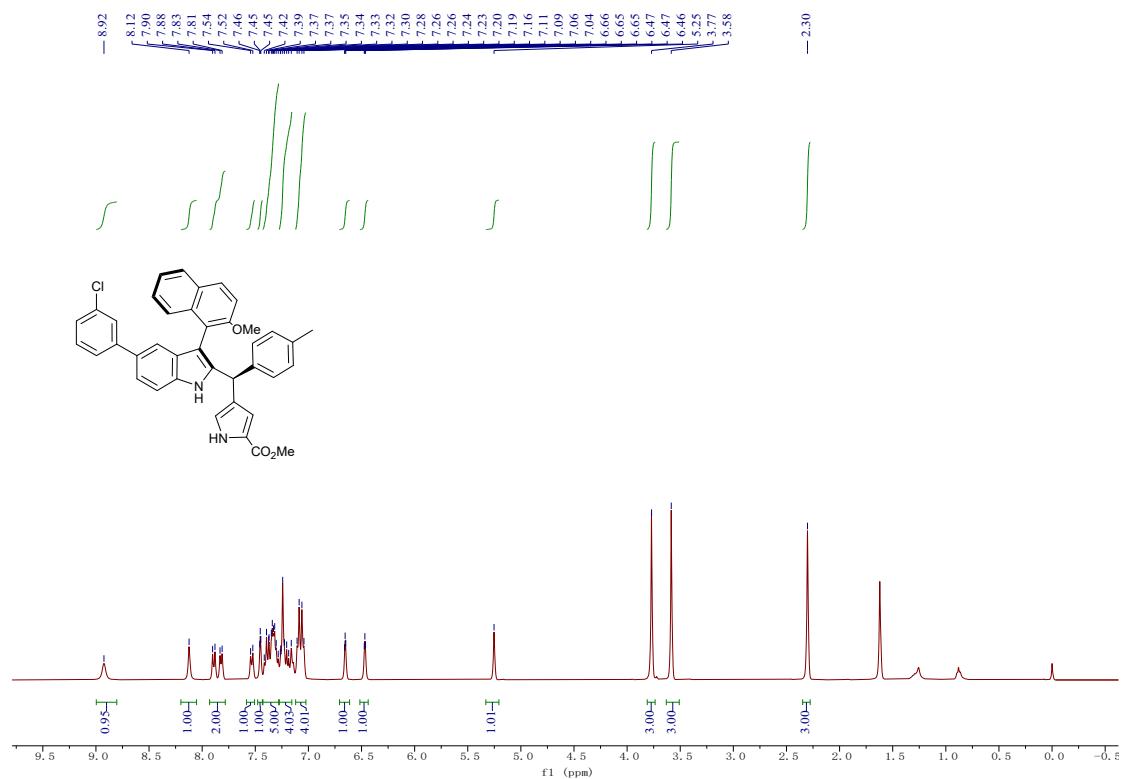
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3za**



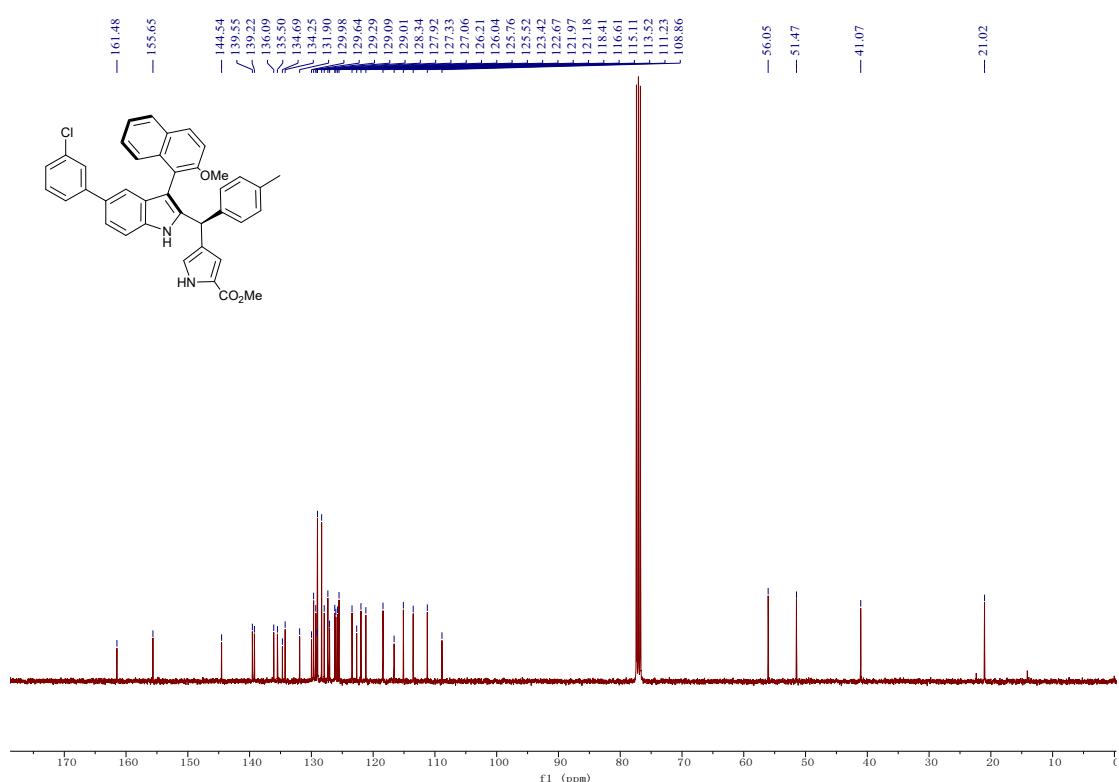
<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) of **3za**



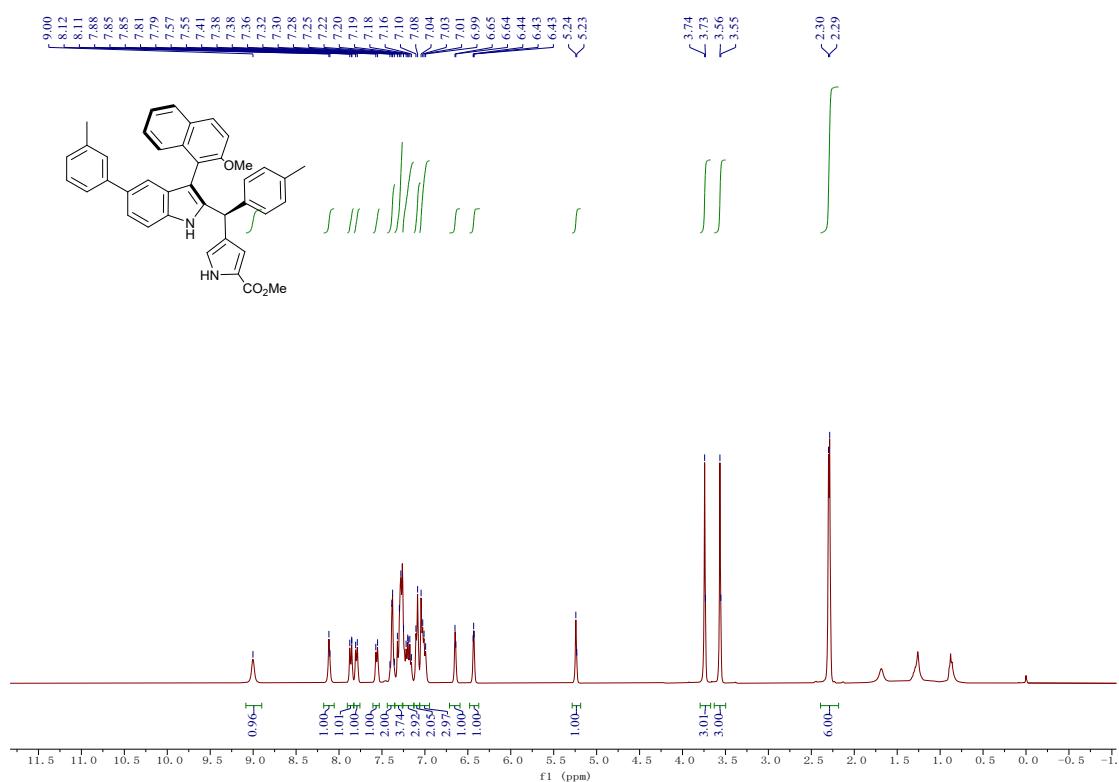
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3zb**



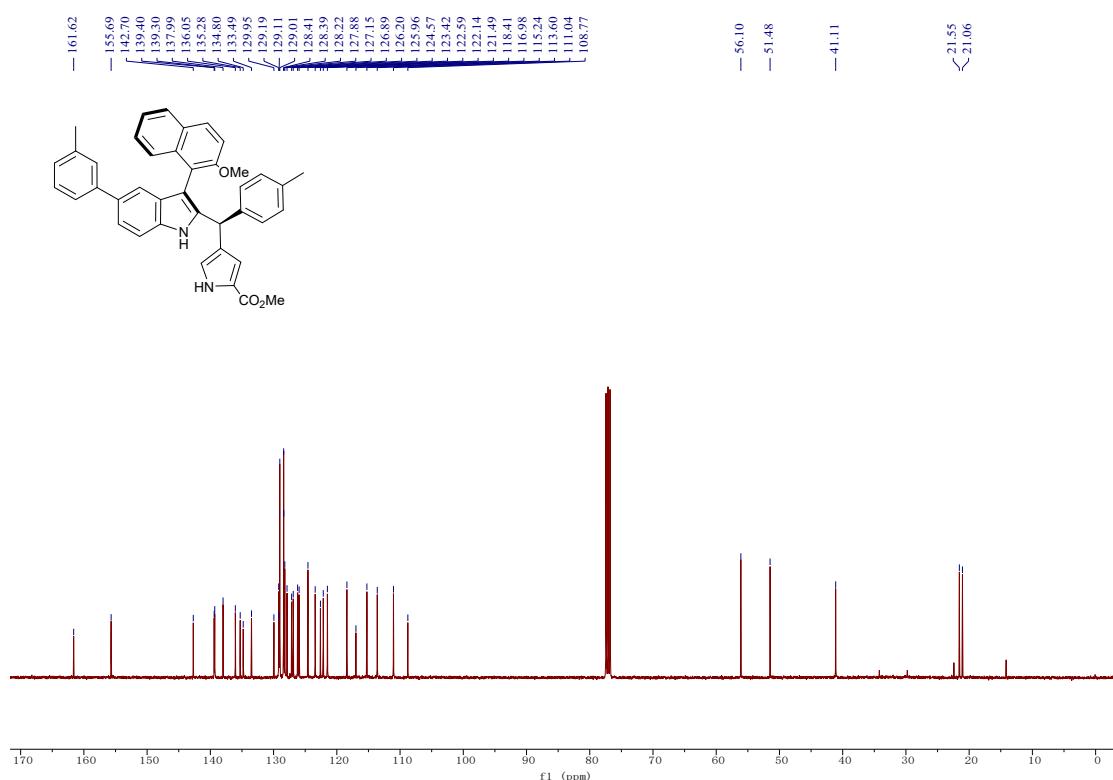
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3zb



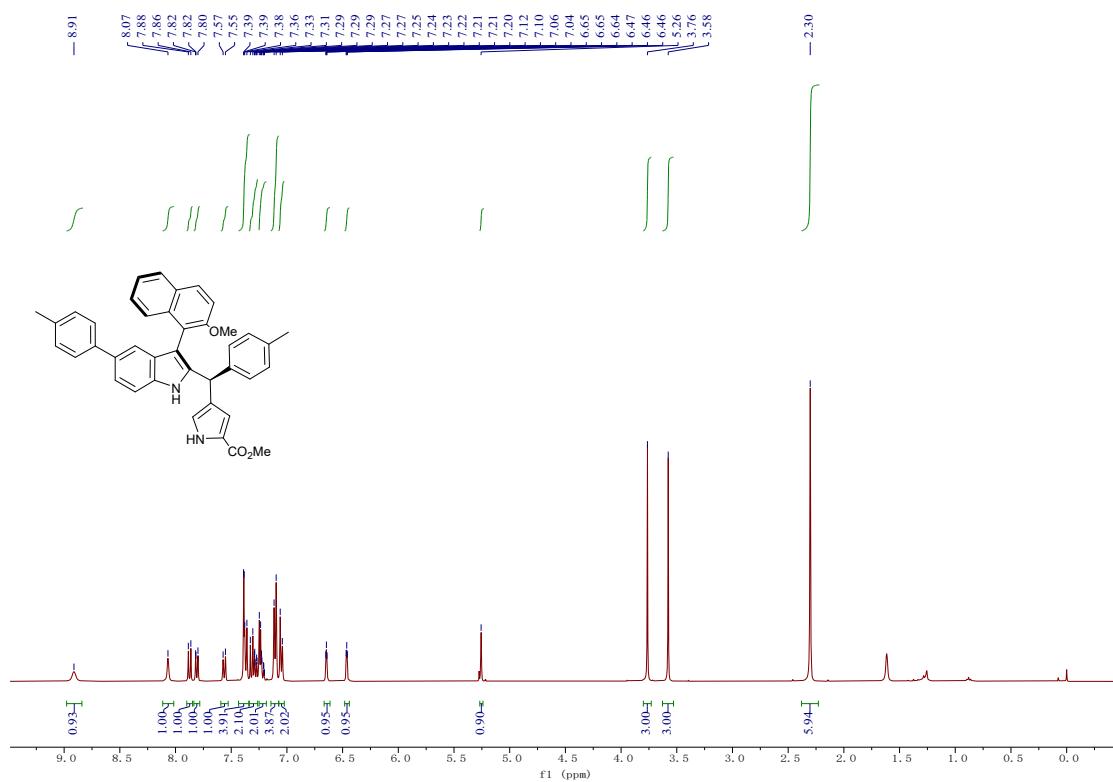
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3zc**



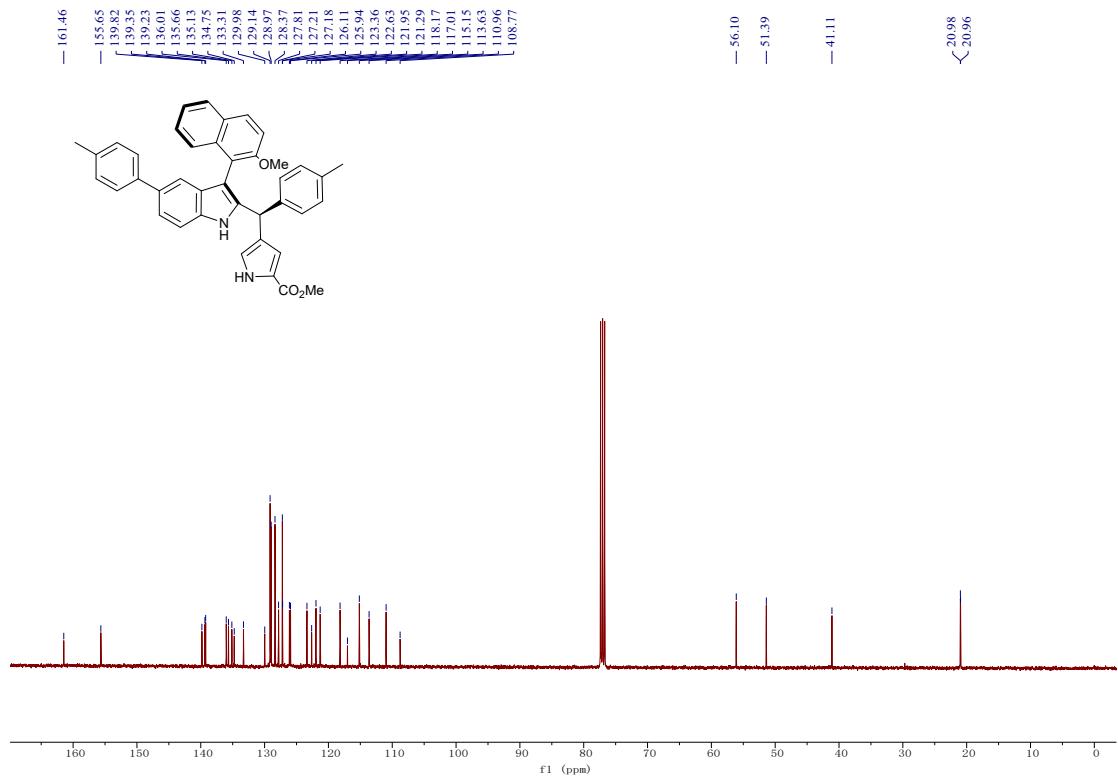
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3zc



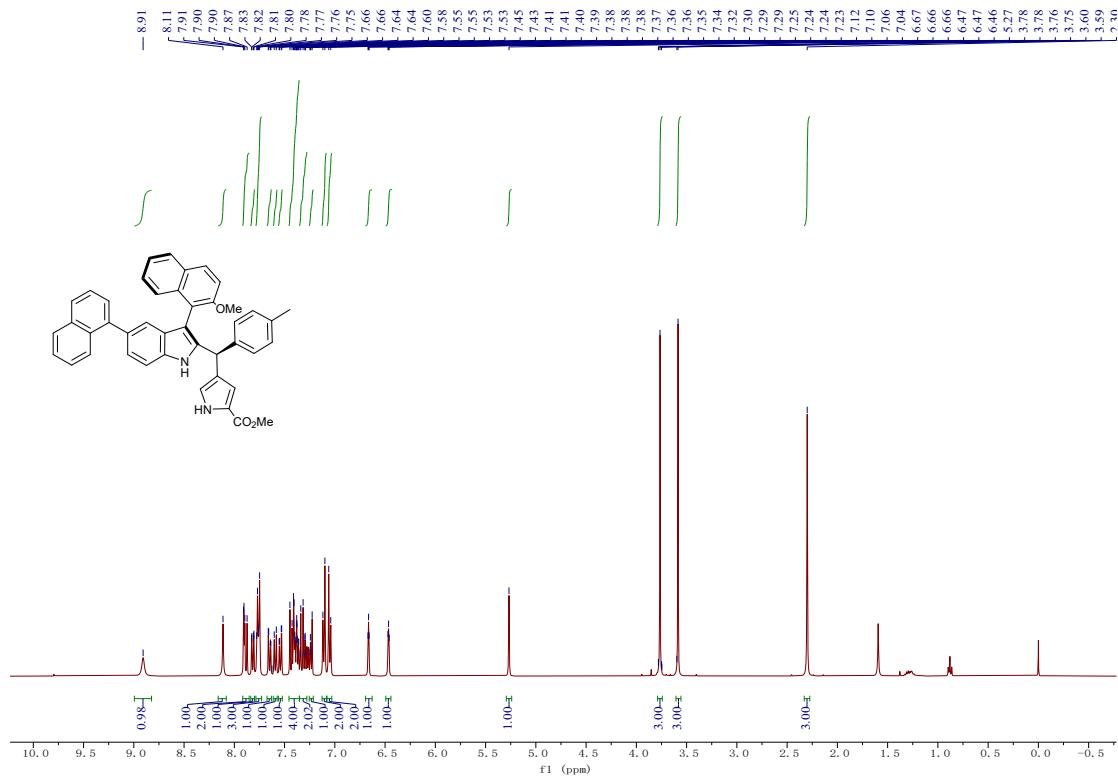
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of 3zd



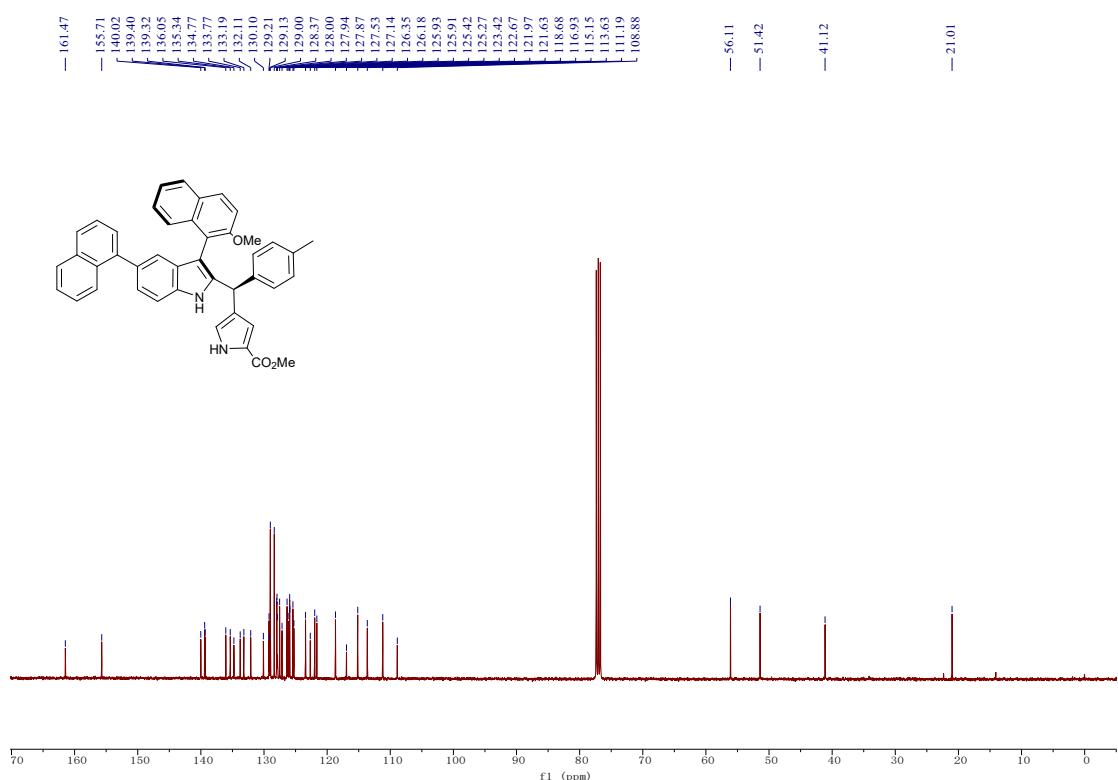
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3zd**



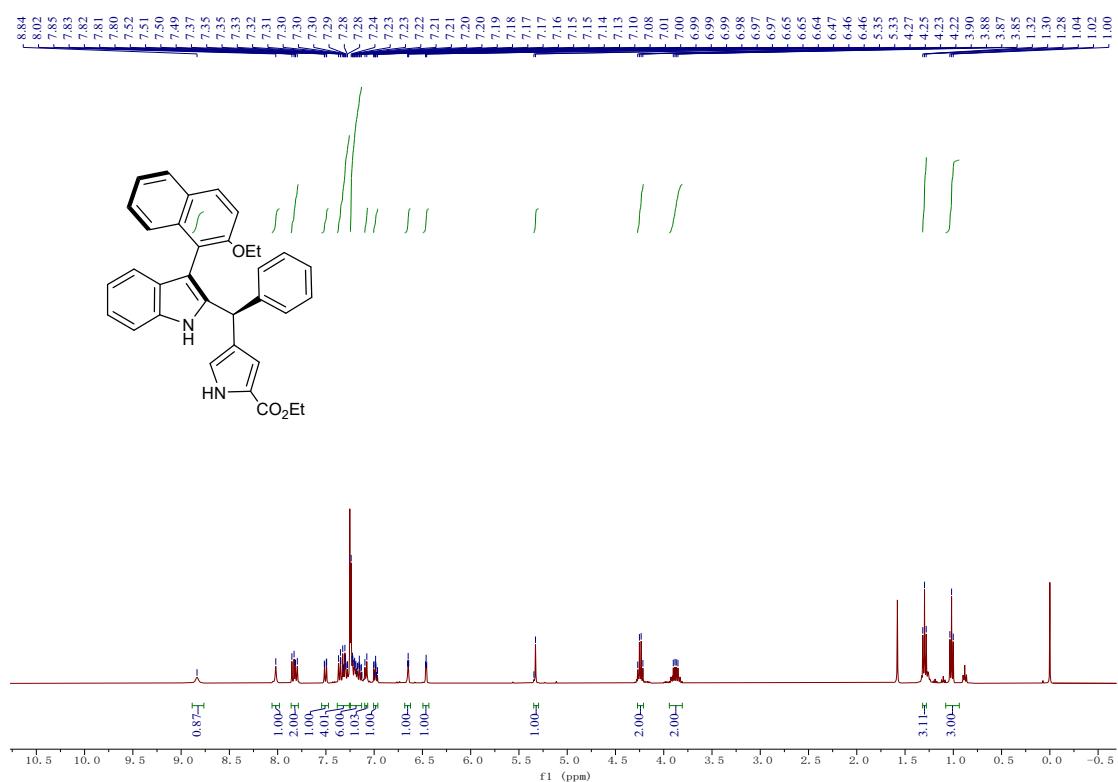
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3ze**



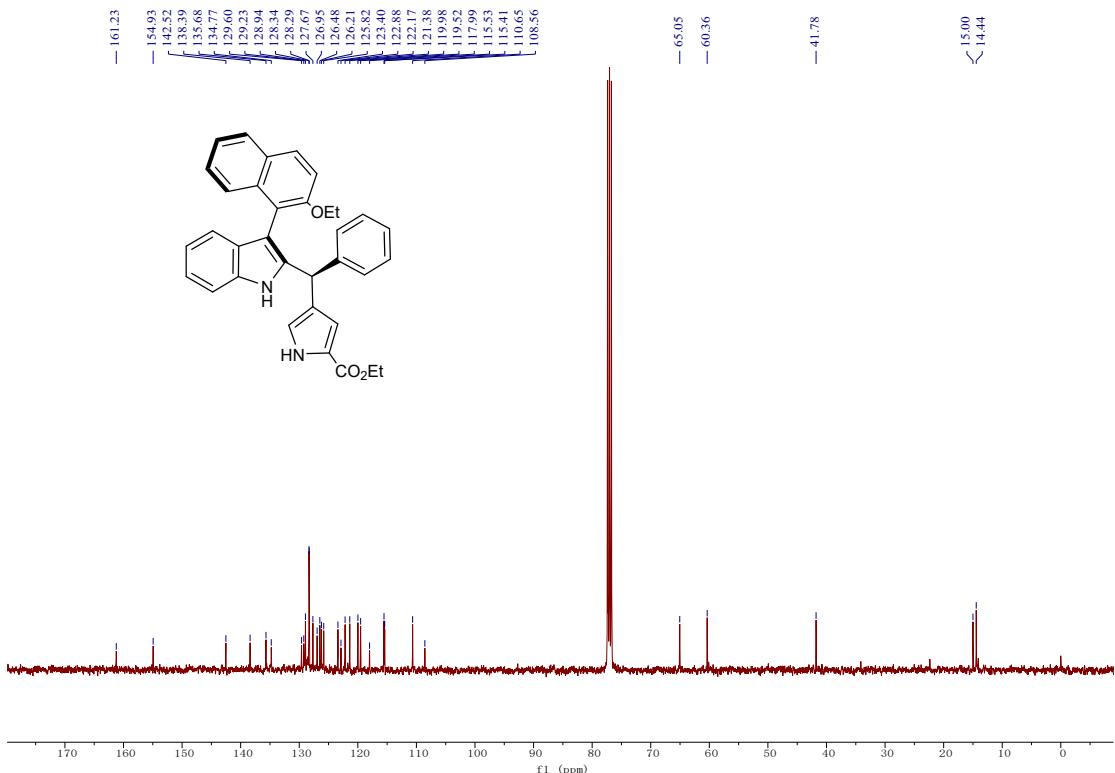
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of 3ze



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of **3zf**

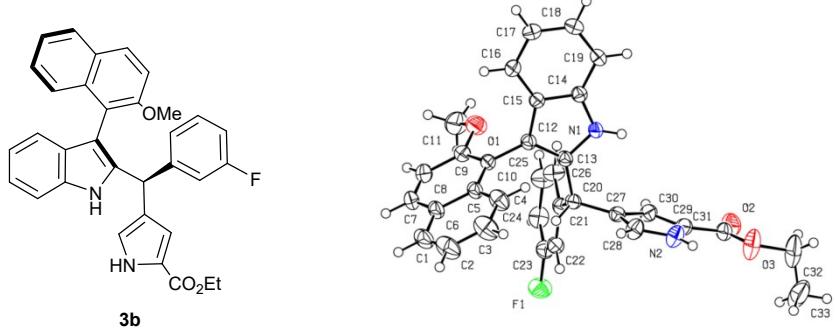


<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of **3zf**



## 5. X-Ray Single Crystal Data for Product (*S*)-**3b**

Single crystals of **3b** were obtained by placing **3b** in a mixed solution of isopropyl alcohol and diethyl ether (volume ratio 1:10) and allowing it to stand at room temperature for a week. The absolute configuration of product **3b** was determined by X-ray diffraction analysis of a single crystal (using a Bruker APEX-II CCD diffractometer). The X-ray data have been deposited at the Cambridge Crystallographic Data Center (CCDC 2431878). The stereochemistry of other products was assumed by analogy.

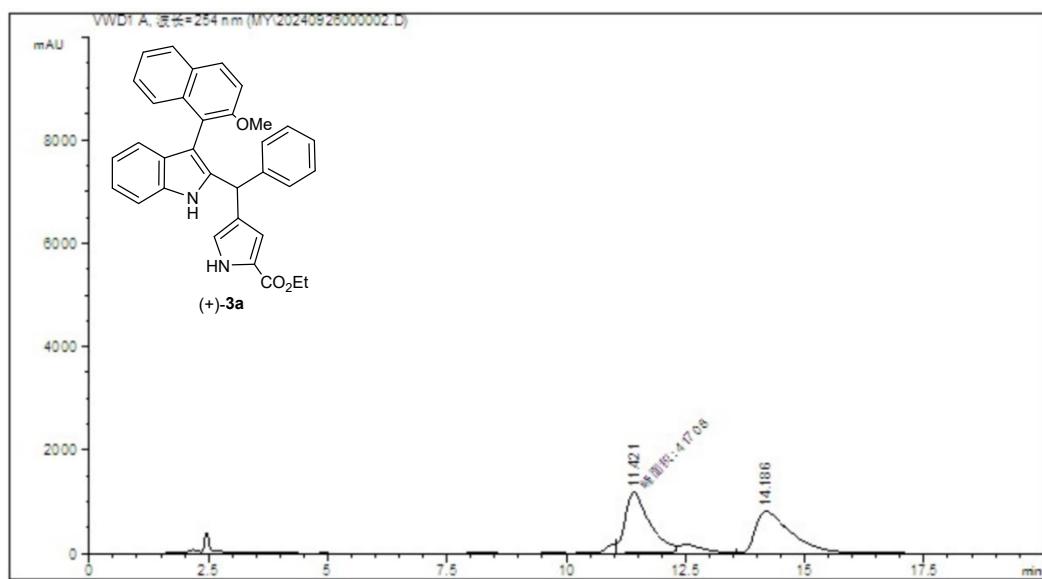


**Table 1 Crystal data and structure refinement for 240918\_my0918\_F1\_TSCl (3b).**

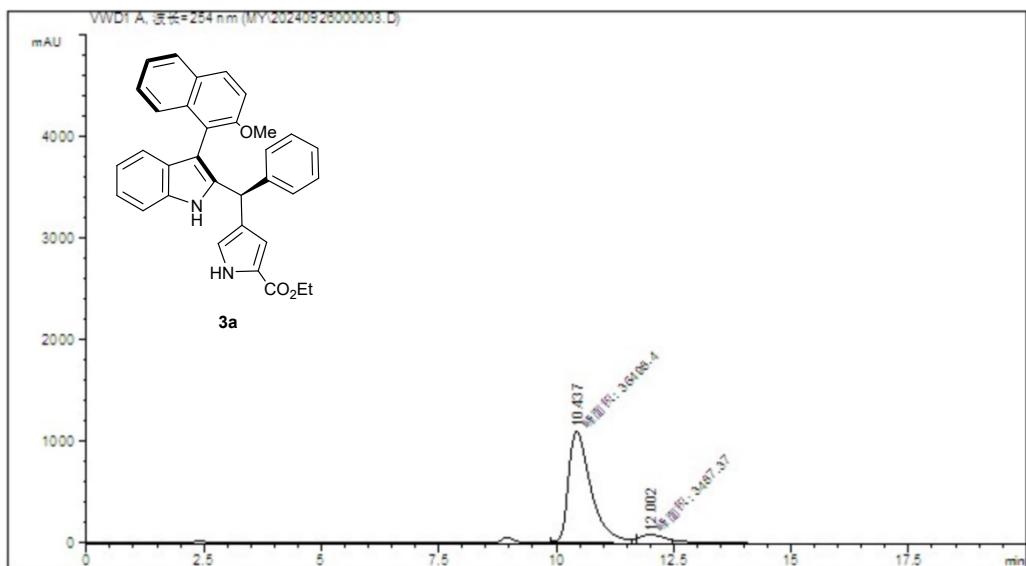
Identification code	240918_my0918_F1_TSCl
Empirical formula	C <sub>33.5</sub> H <sub>27.5</sub> Cl <sub>1.5</sub> FN <sub>2</sub> O <sub>3</sub>
Formula weight	578.25
Temperature/K	170.00
Crystal system	monoclinic
Space group	C2
a/Å	11.5056(7)
b/Å	13.8298(8)
c/Å	18.2204(11)
α/°	90
β/°	98.022(2)
γ/°	90
Volume/Å <sup>3</sup>	2870.9(3)
Z	4
ρ <sub>calc</sub> g/cm <sup>3</sup>	1.338

$\mu/\text{mm}^{-1}$	1.295
F(000)	1204.0
Crystal size/ $\text{mm}^3$	$0.16 \times 0.08 \times 0.06$
Radiation	GaK $\alpha$ ( $\lambda = 1.34139$ )
$2\Theta$ range for data collection/ $^\circ$	8.528 to 121.602
Index ranges	$-14 \leq h \leq 14, -18 \leq k \leq 17, -23 \leq l \leq 23$
Reflections collected	47692
Independent reflections	6564 [ $R_{\text{int}} = 0.0378, R_{\text{sigma}} = 0.0276$ ]
Data/restraints/parameters	6564/47/401
Goodness-of-fit on $F^2$	1.029
Final R indexes [ $I >= 2\sigma(I)$ ]	$R_1 = 0.0470, wR_2 = 0.1264$
Final R indexes [all data]	$R_1 = 0.0494, wR_2 = 0.1294$
Largest diff. peak/hole / e $\text{\AA}^{-3}$	0.38/-0.46
Flack parameter	0.097(8)

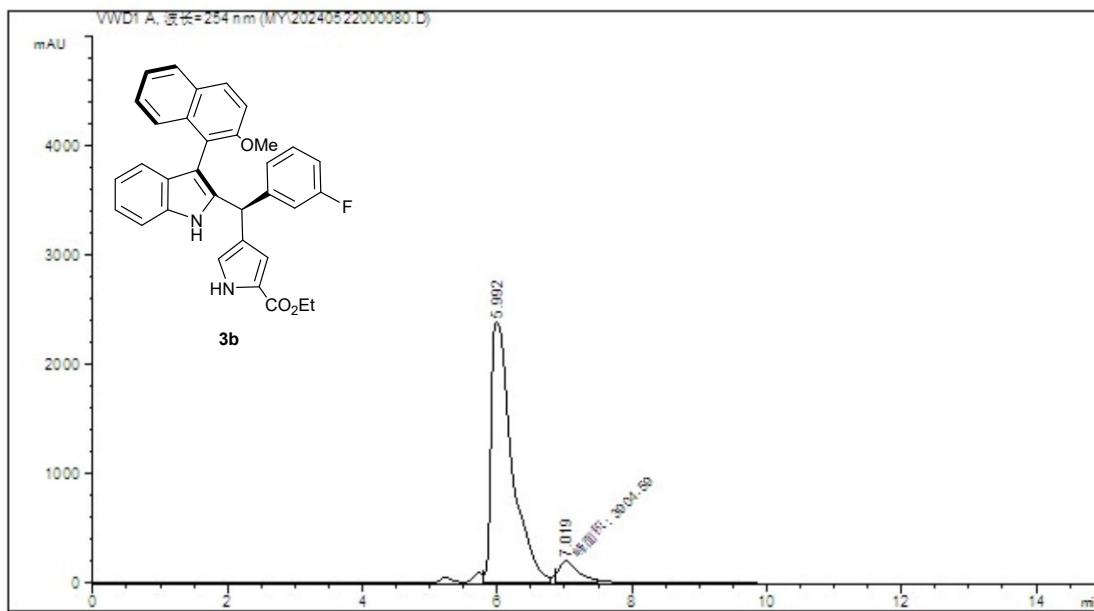
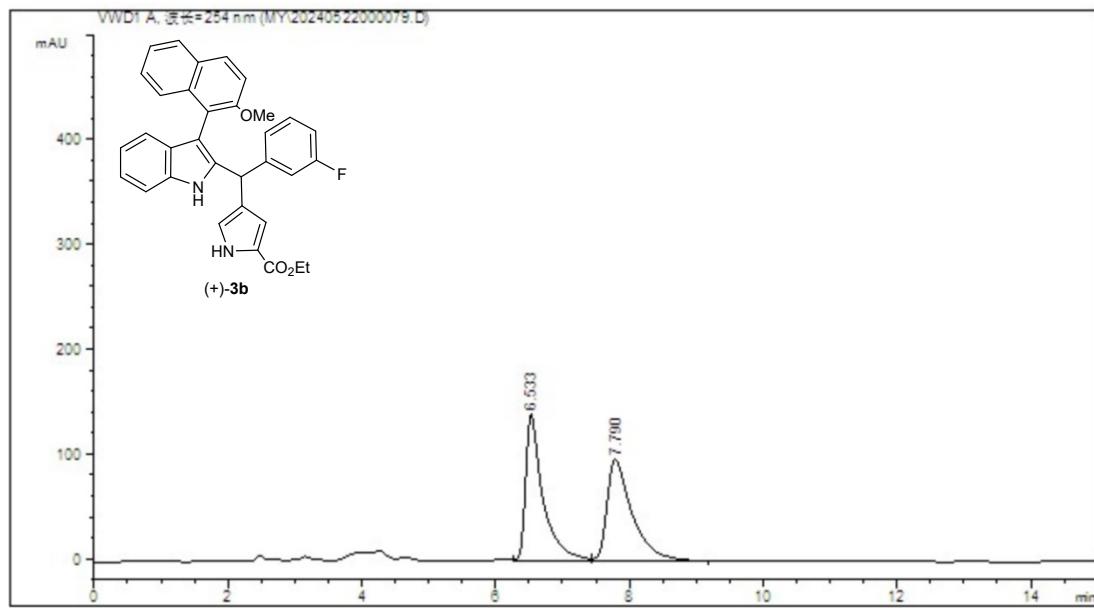
## 6. Copies of HPLC Spectra

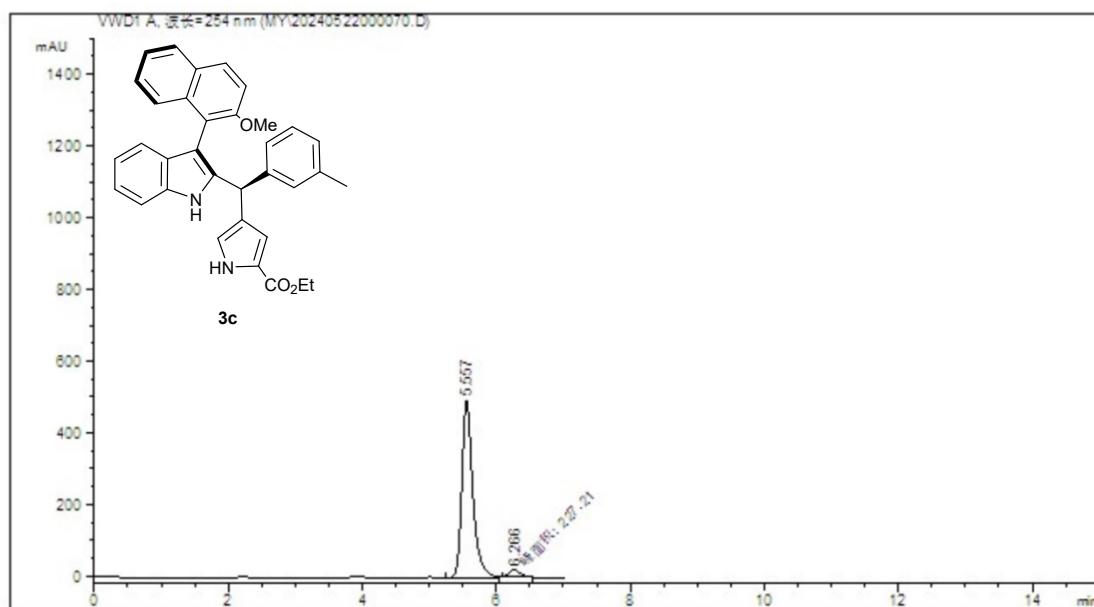
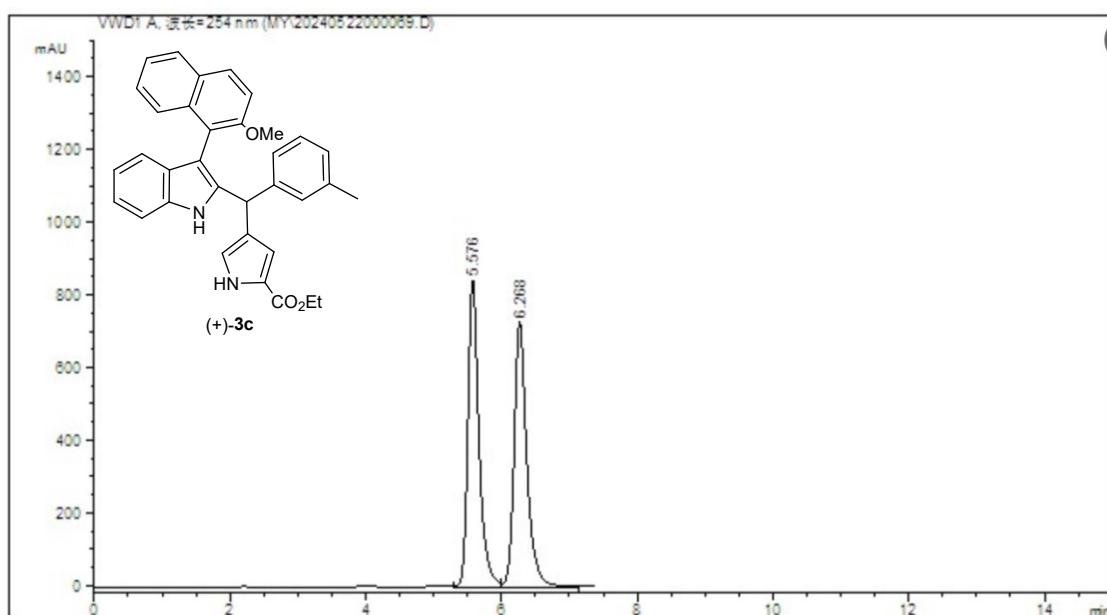


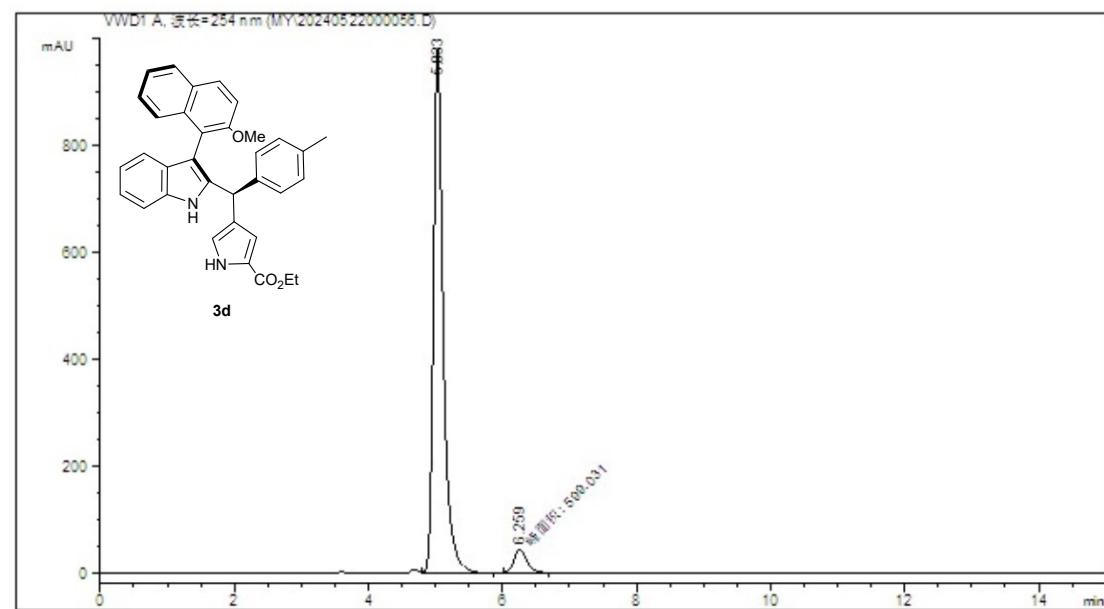
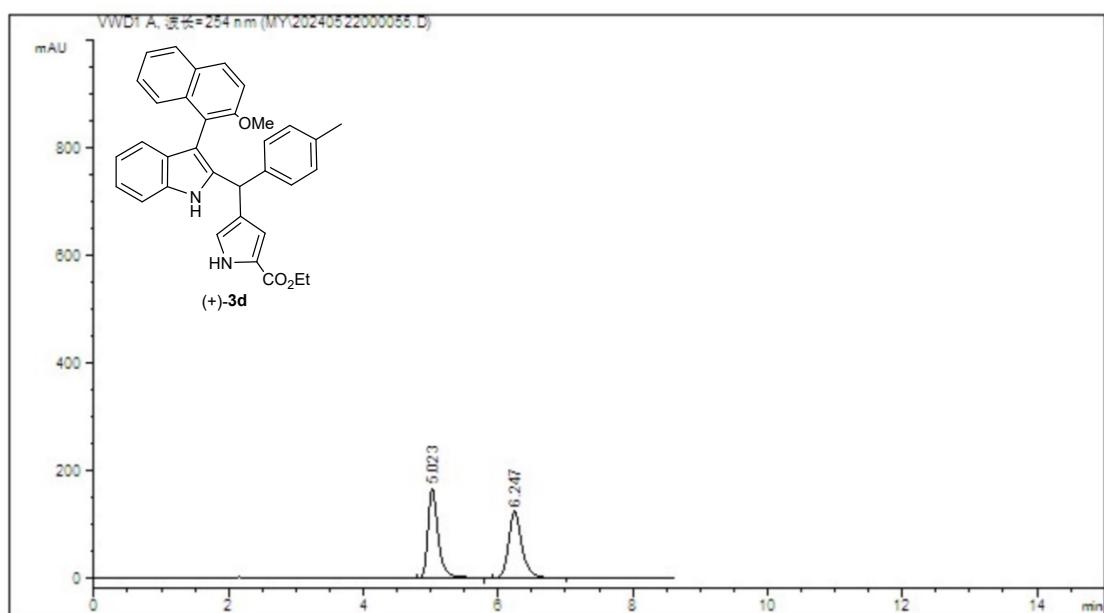
#	[min]		[min]	mAU	*s	[mAU]		#
1	11.421	MM	0.5896	4.17080e4		1179.05383		49.7632
2	14.186	VV	0.7481	4.21049e4		817.23376		50.2368

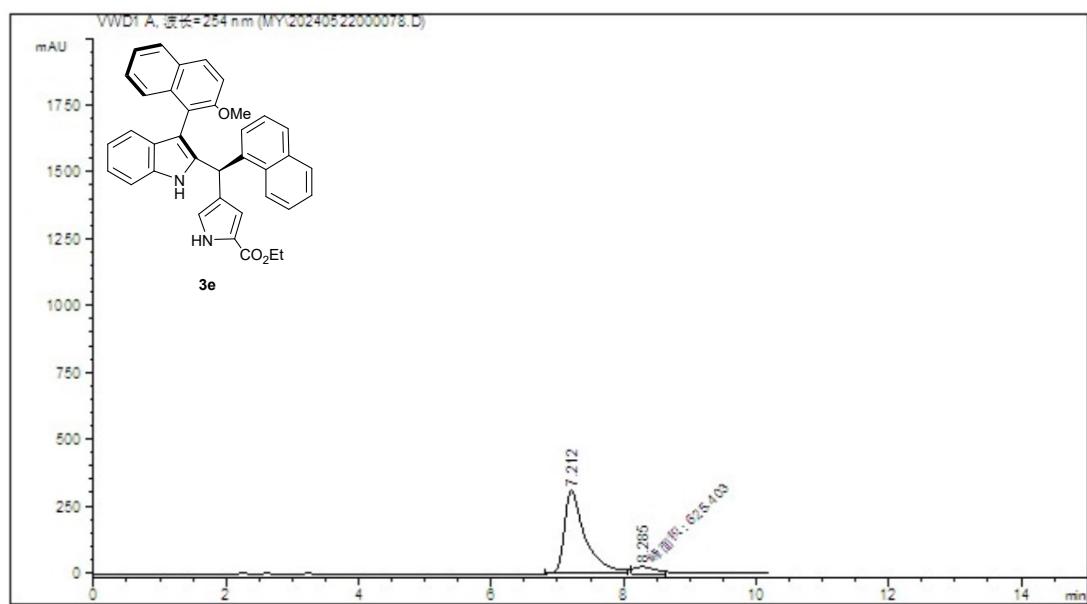
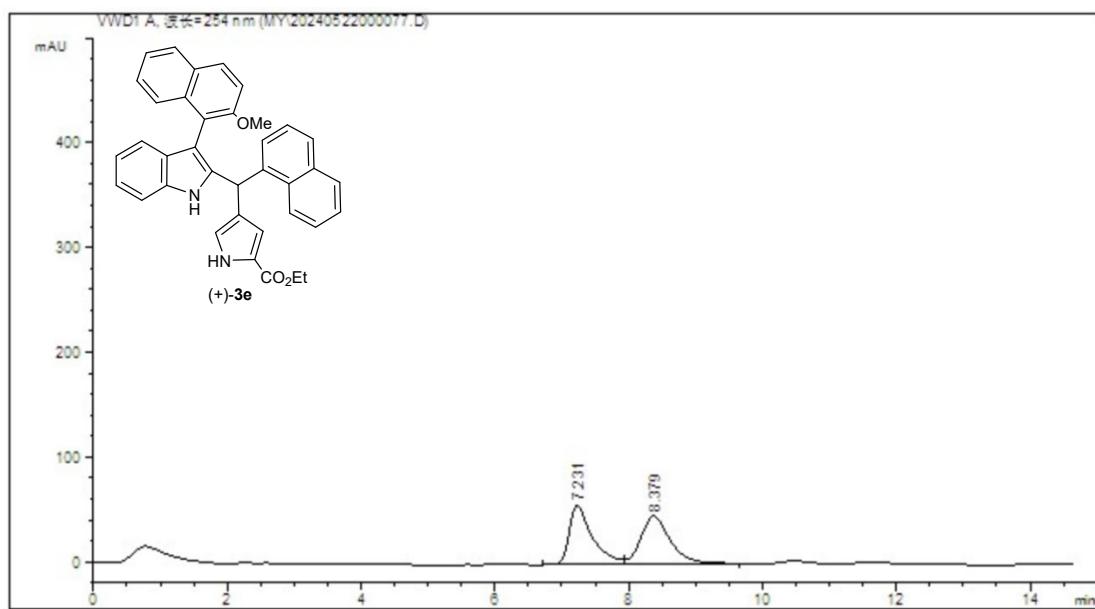


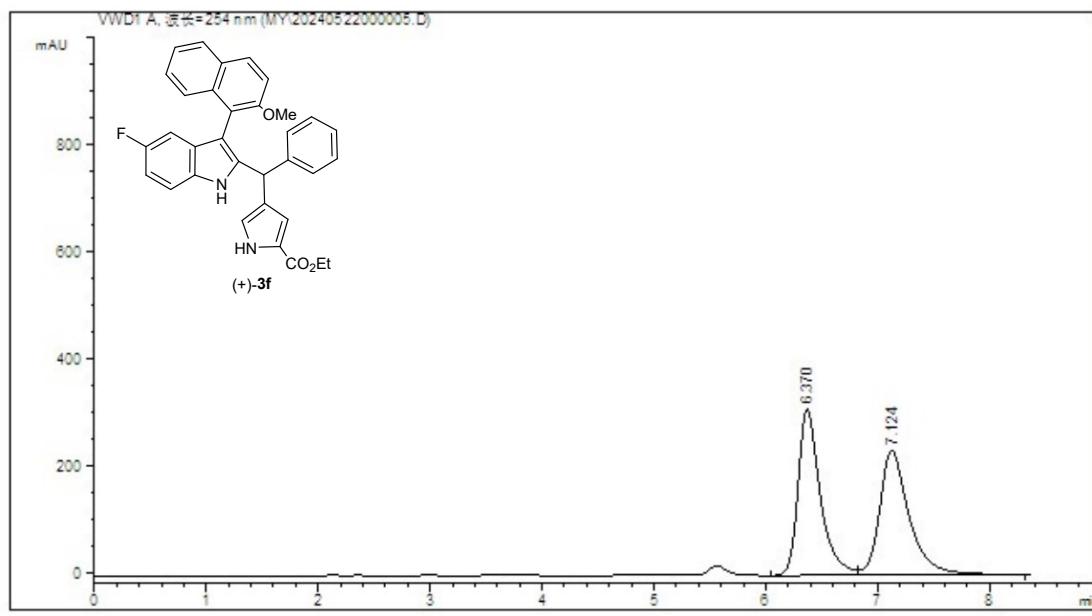
#	[min]		[min]	mAU	*s	[mAU]		#
1	10.437	MM	0.5575	3.64984e4		1091.19482		91.2785
2	12.002	MM	0.6306	3487.36743		92.16611		8.7215



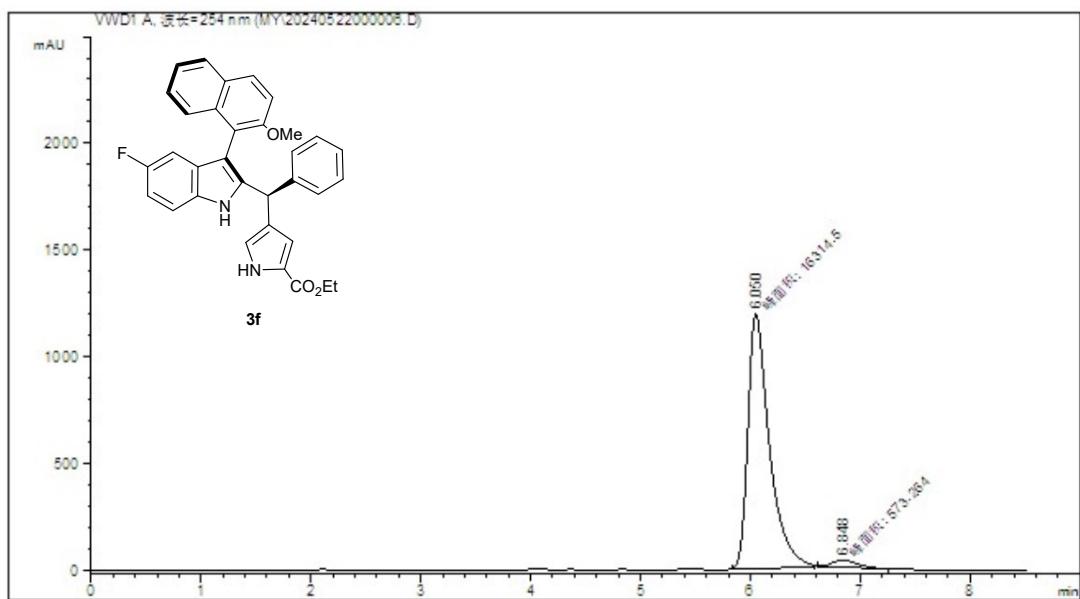




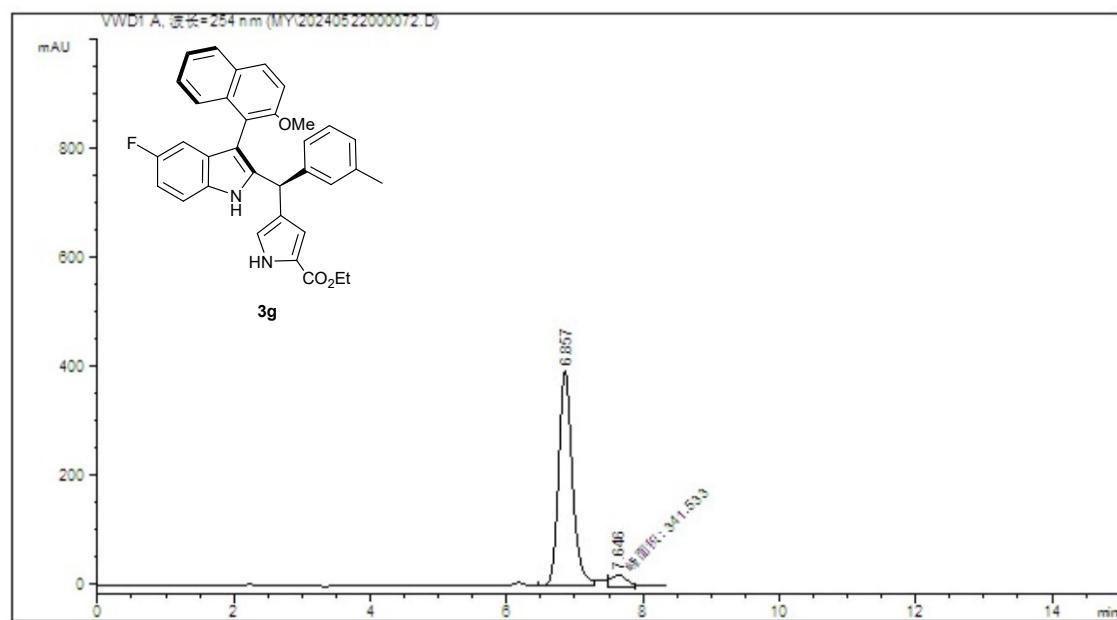
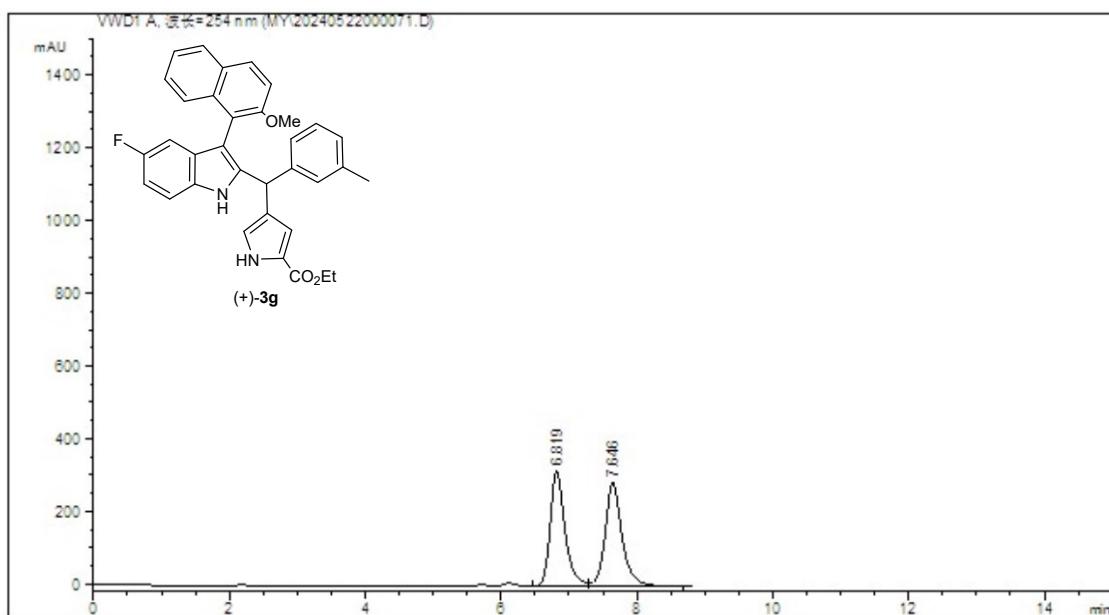


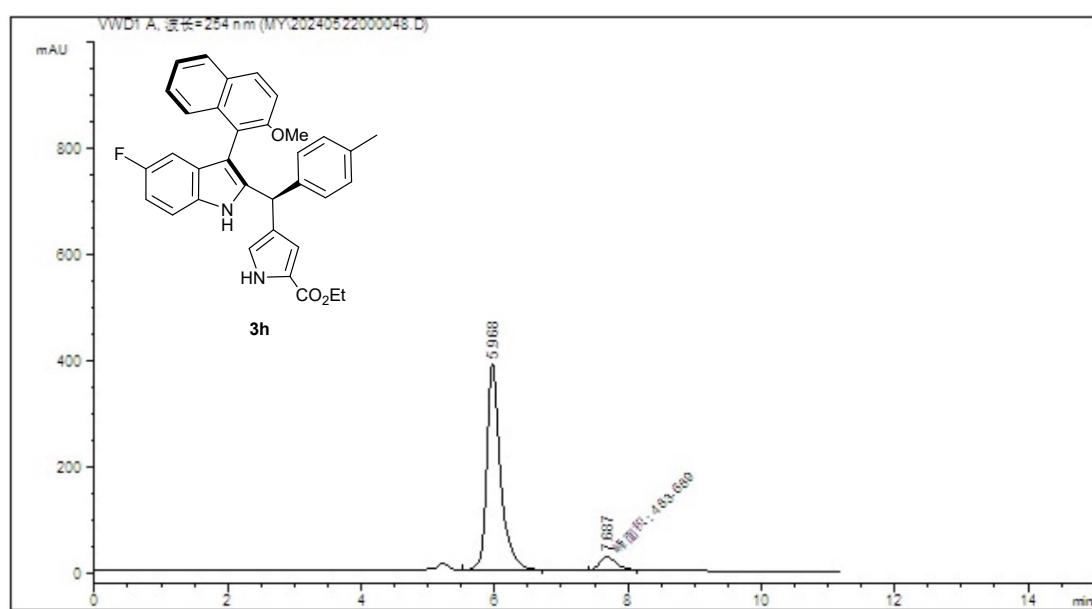
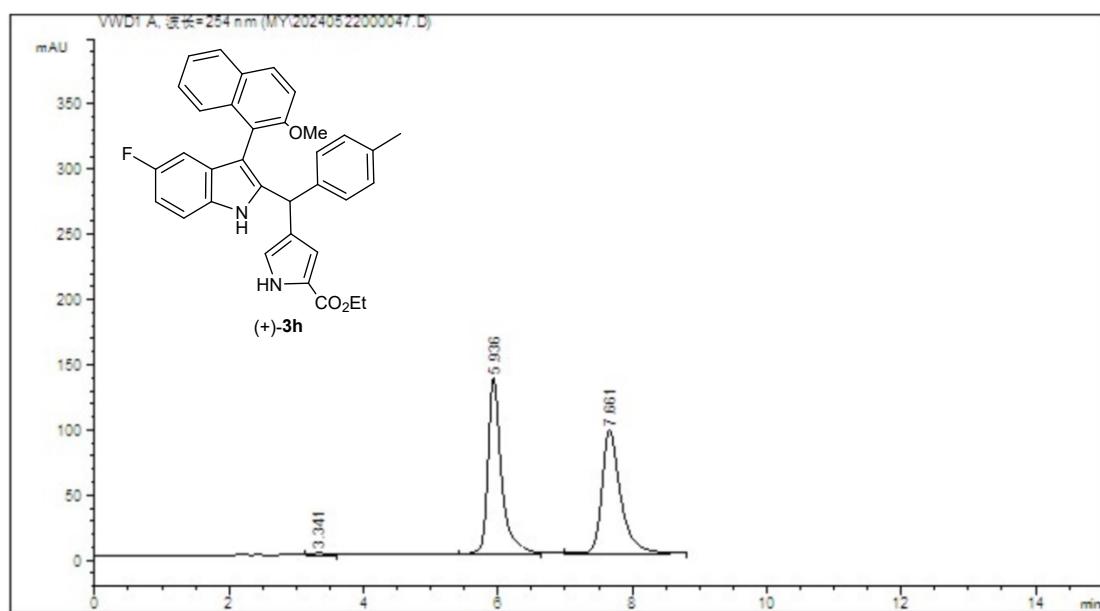


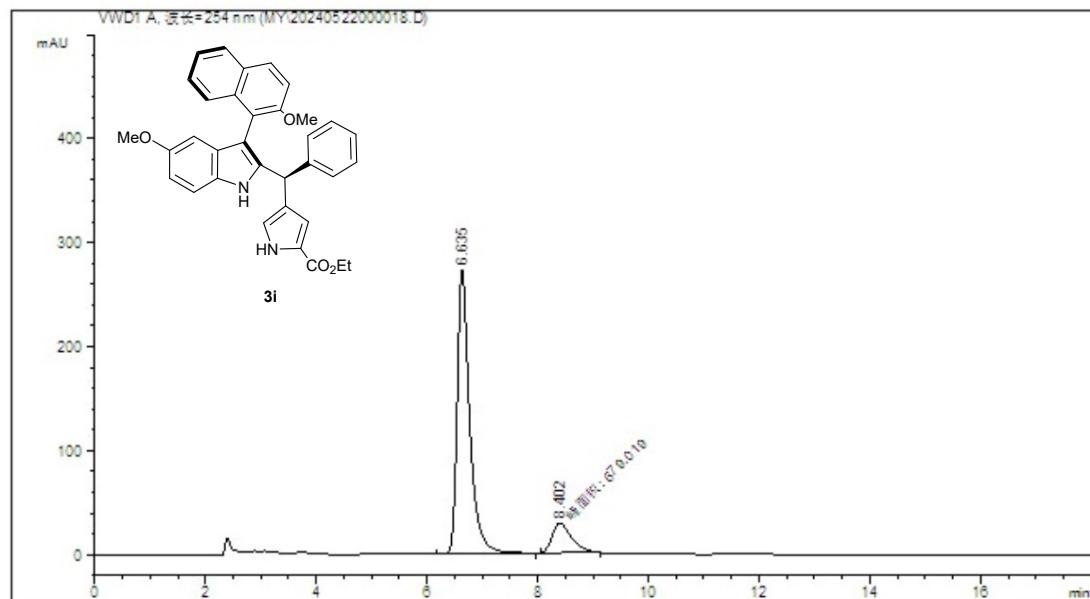
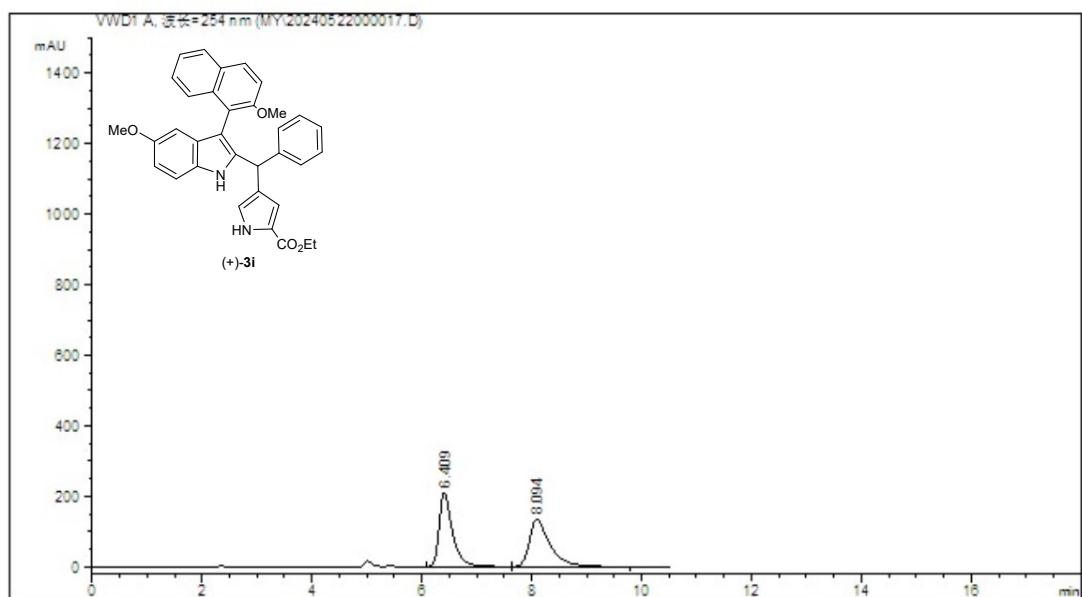
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1	6.370	BV	0.2047	4289.09912		310.71088		50.0645
2	7.124	VB	0.2694	4278.04443		233.15057		49.9355

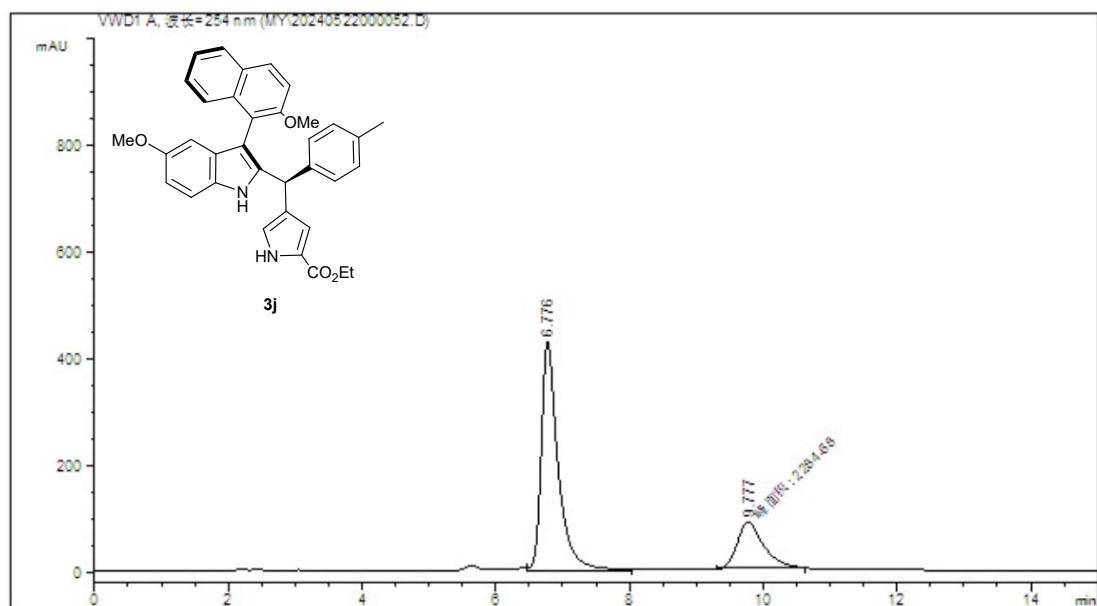
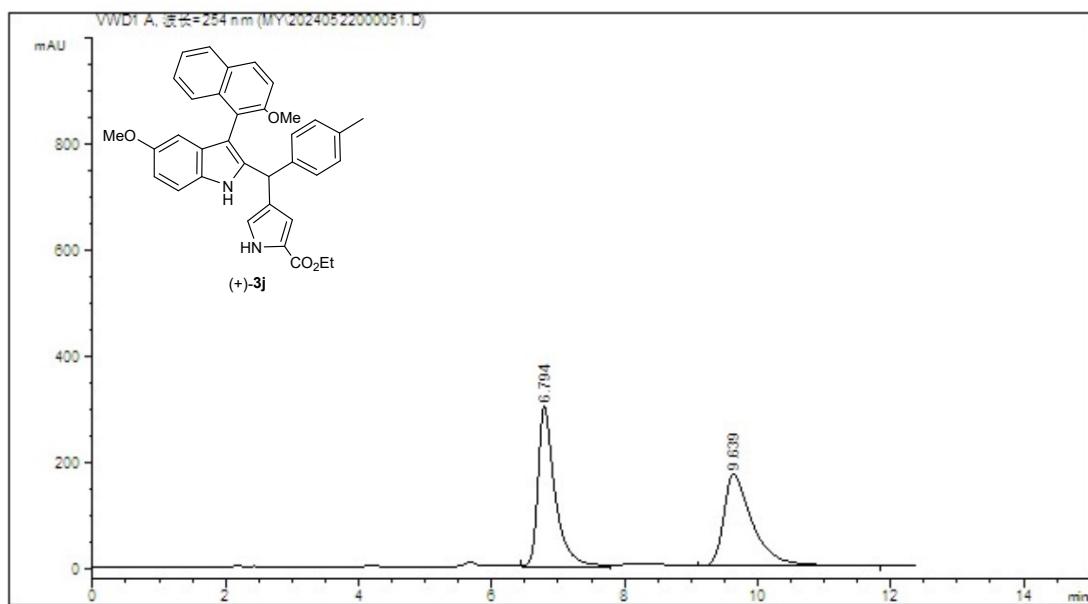


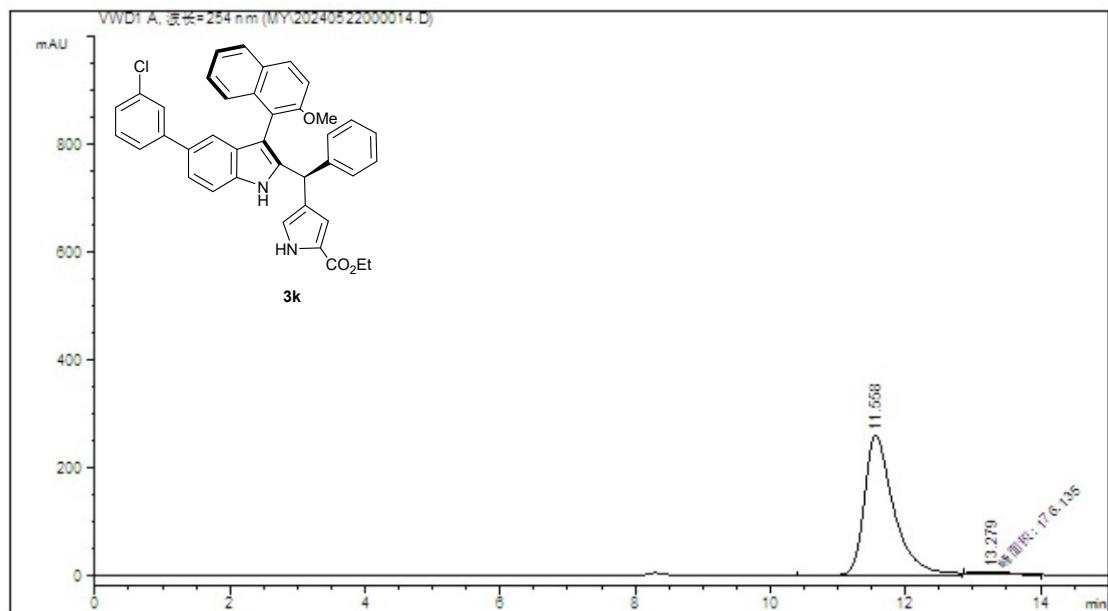
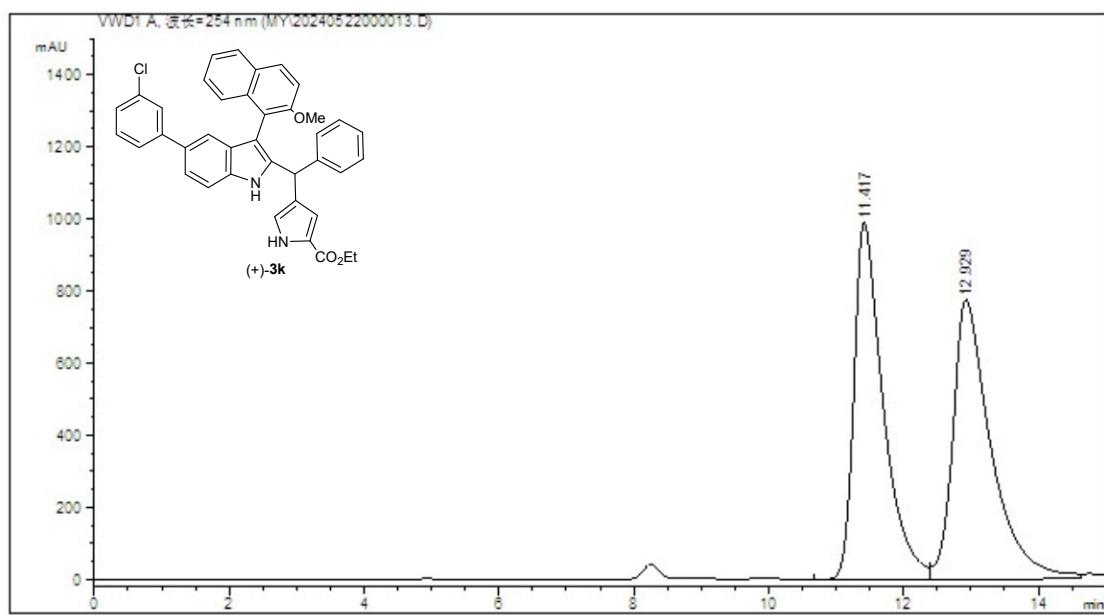
#	[min]		[min]	mAU	*s	[mAU]		%
1	6.050	MM	0.2274	1.63145e4		1195.80151		96.6054
2	6.848	MM	0.2676	573.26428		35.70073		3.3946

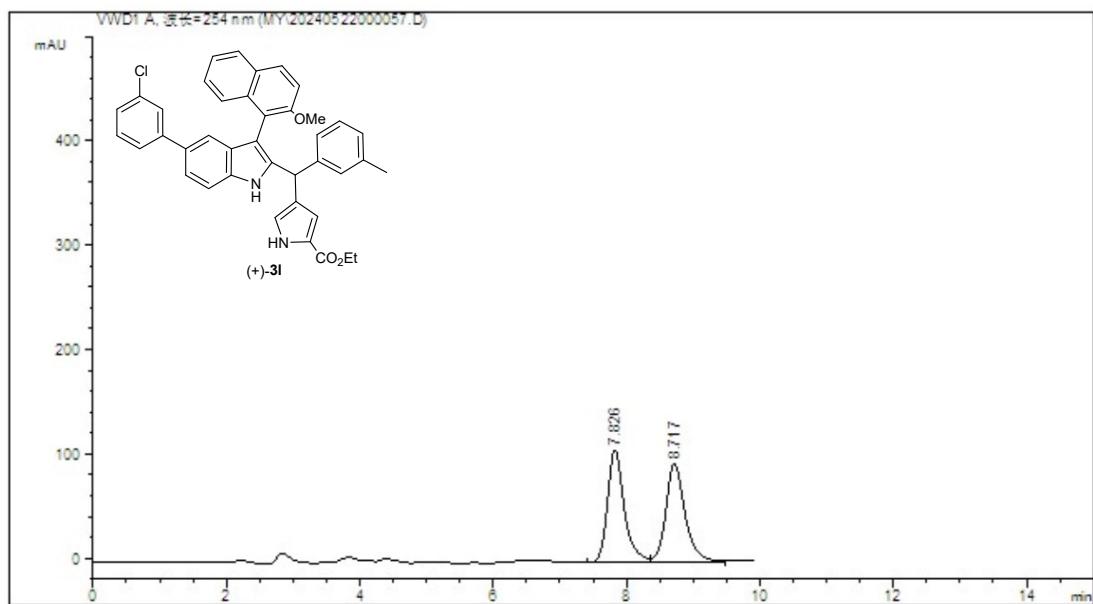




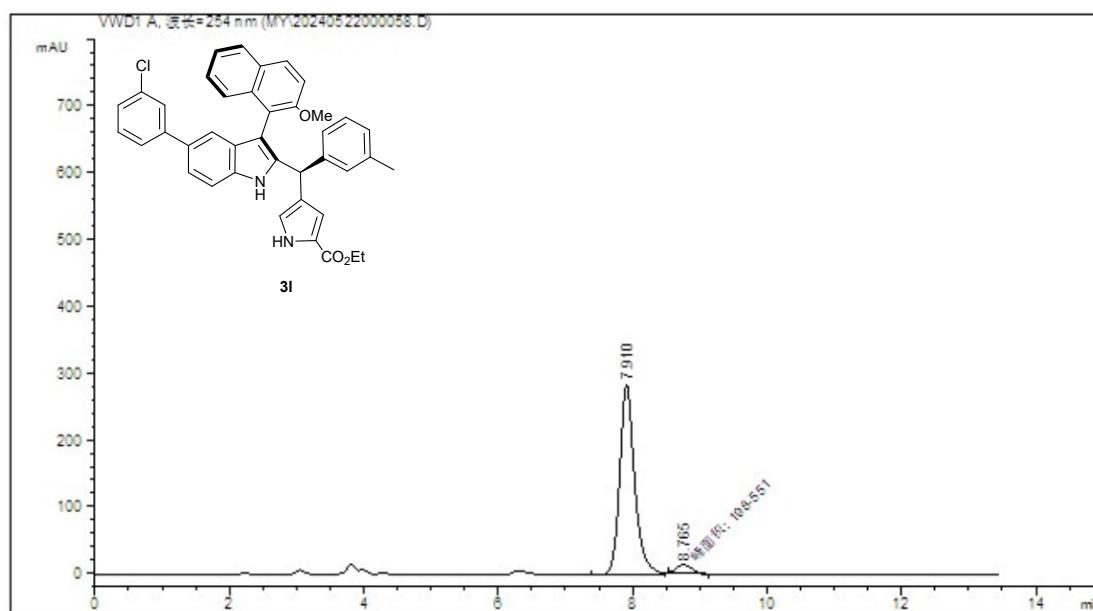




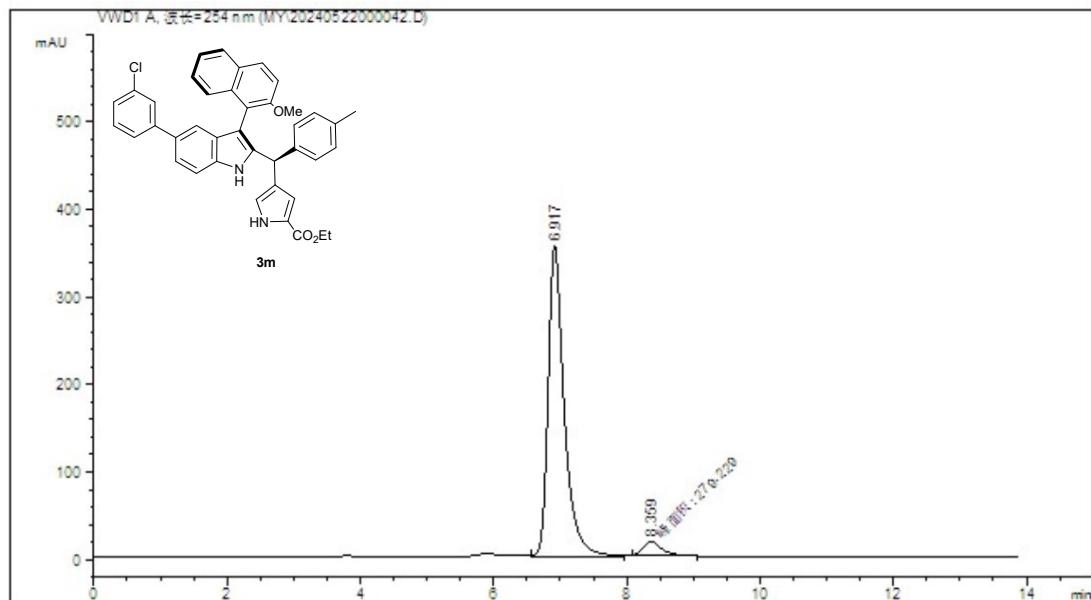
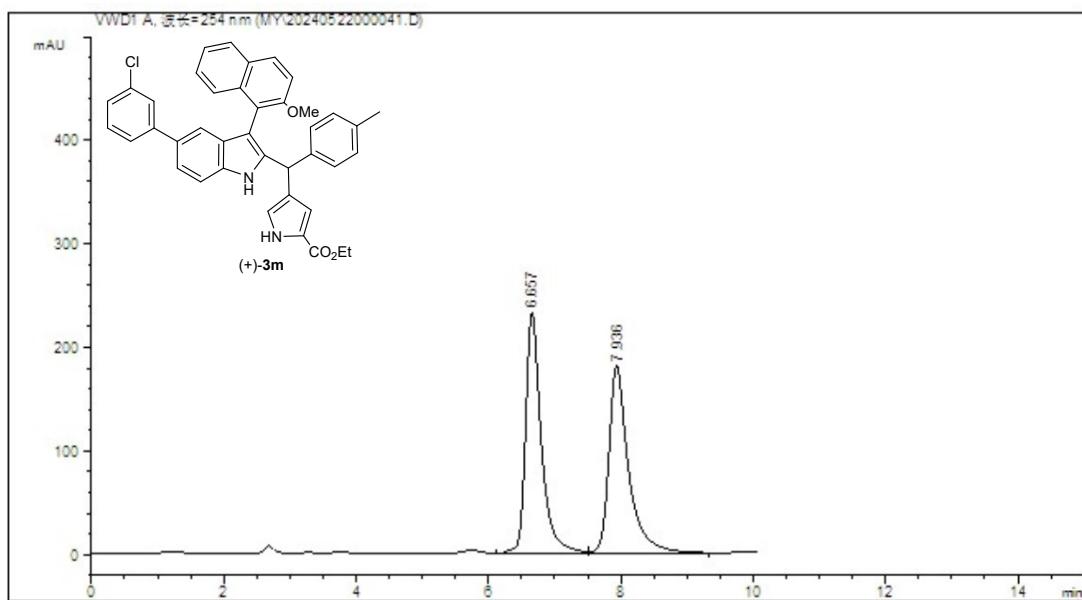


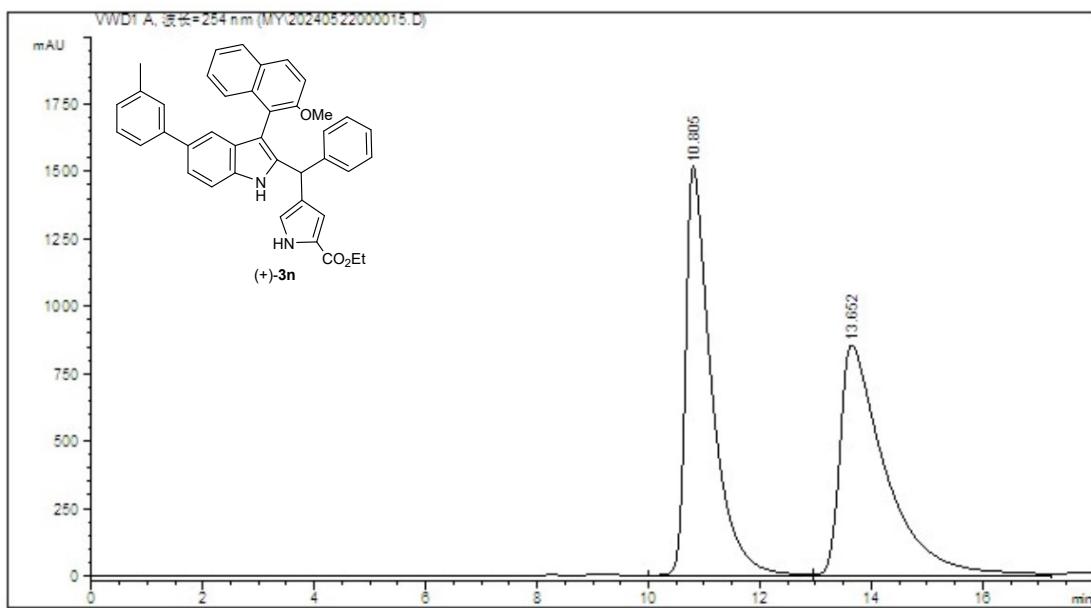


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1	7.826	VV	0.2634	1907.00562	108.41856	49.9849
2	8.717	VB	0.3037	1908.15942	94.62911	50.0151

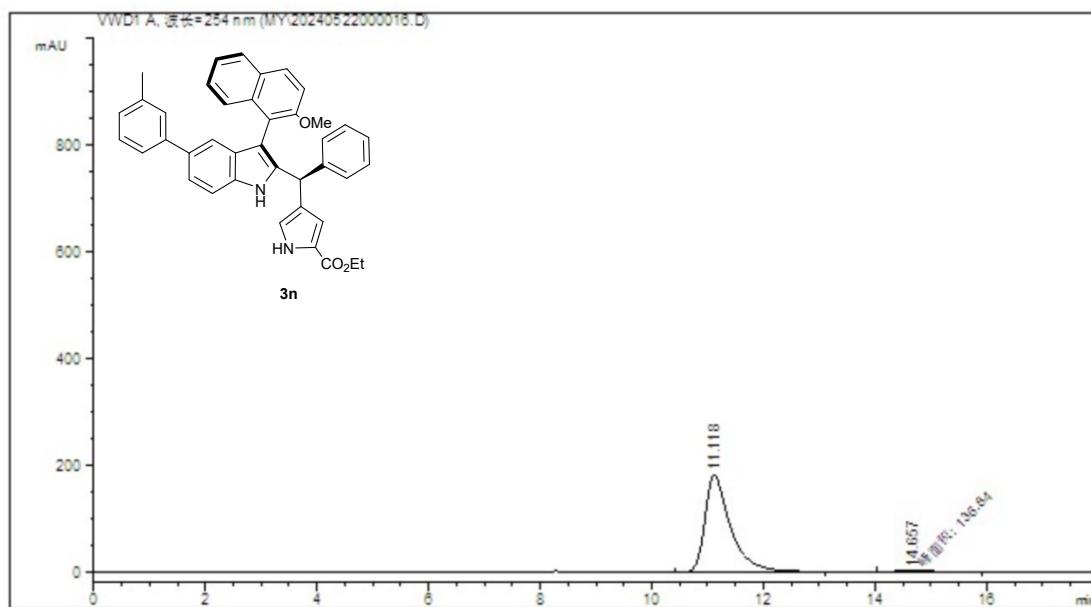


#	[min]	[min]	mAU	*s	[mAU]	%
1	7.910	VV	0.2319	4409.01660	284.31638	95.6908
2	8.765	MM	0.2667	198.55054	12.40960	4.3092

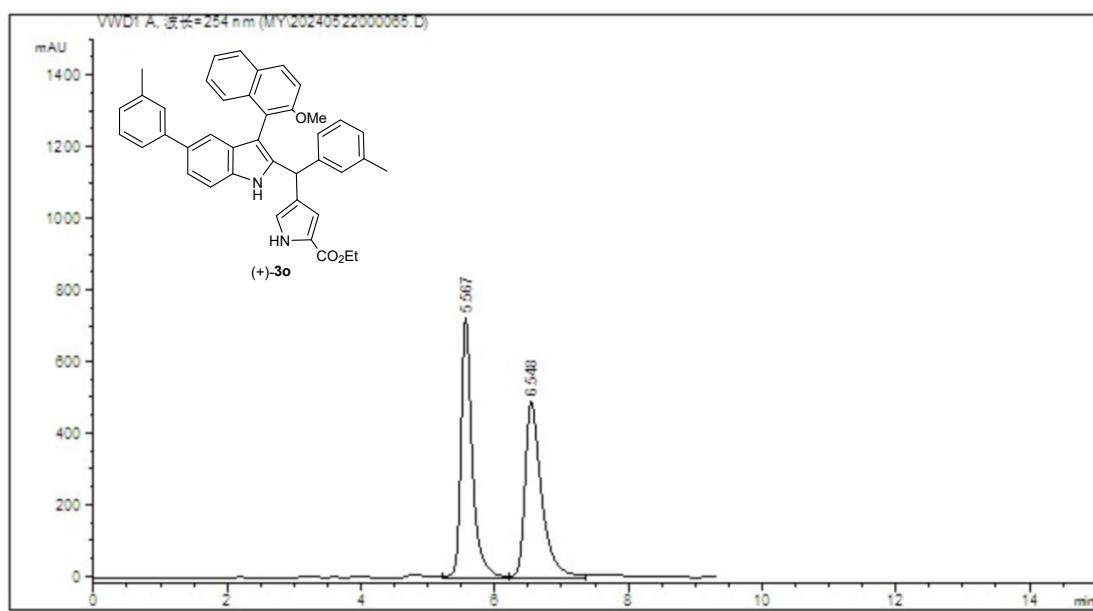




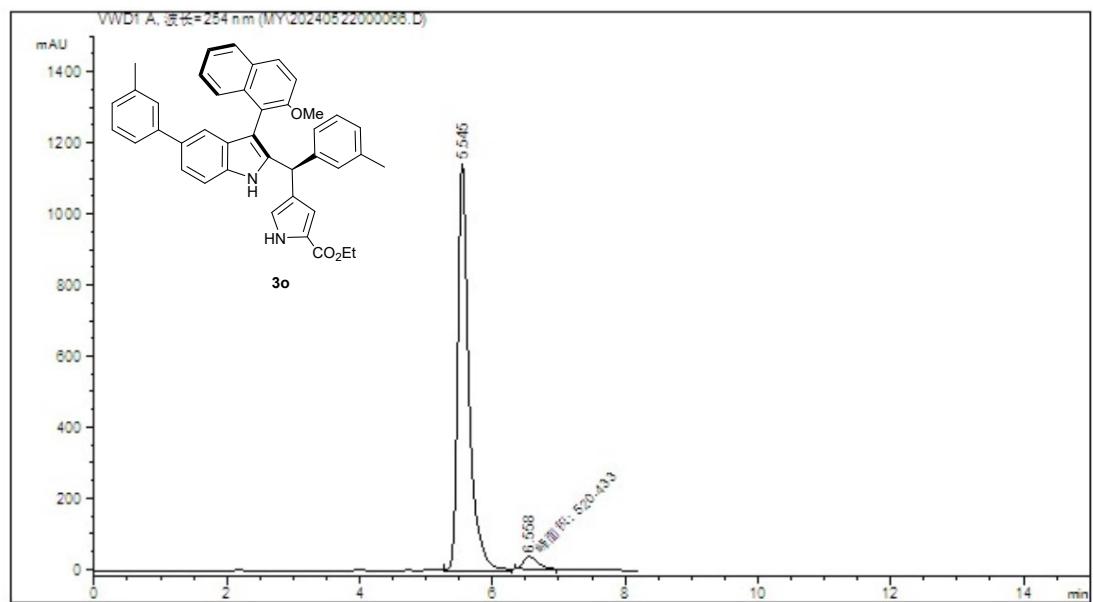
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1	10.805	VV	0.4655	4.90694e4		1519.69031	50.1559
2	13.652	VB	0.8069	4.87643e4		856.55963	49.8441



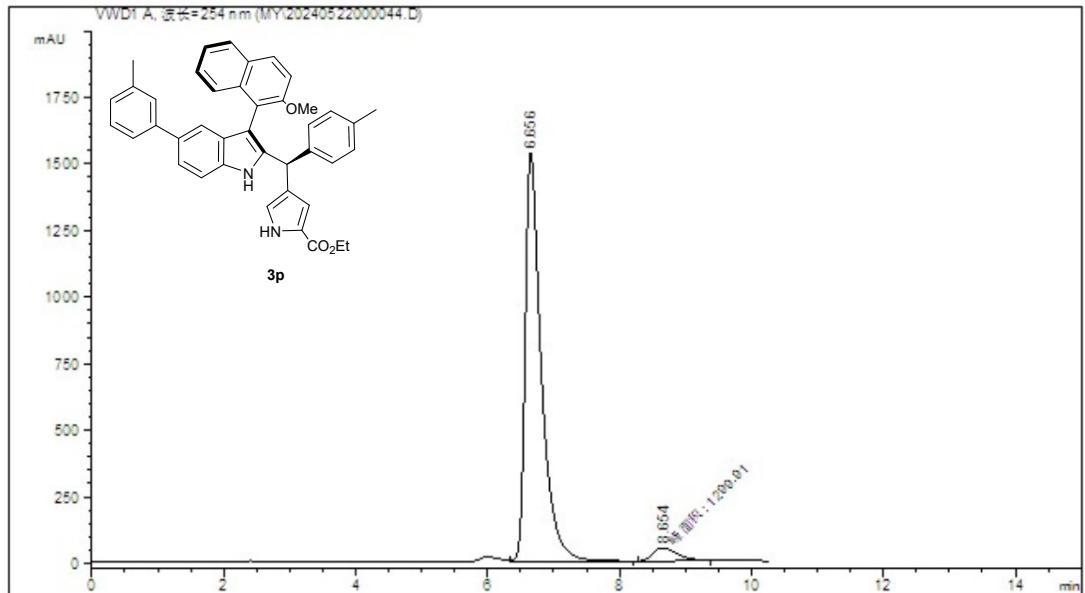
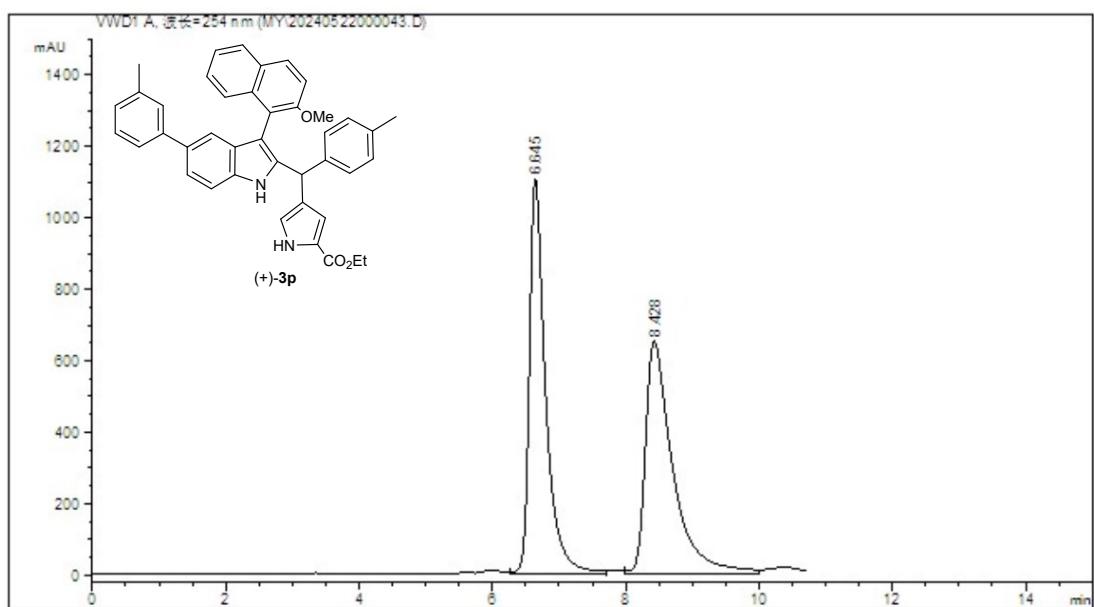
#	[min]		[min]	mAU	*s	[mAU]	%
1	11.118	BB	0.4652	5789.84570		181.58888	97.6911
2	14.657	MM	0.9858	136.84032		2.31345	2.3089

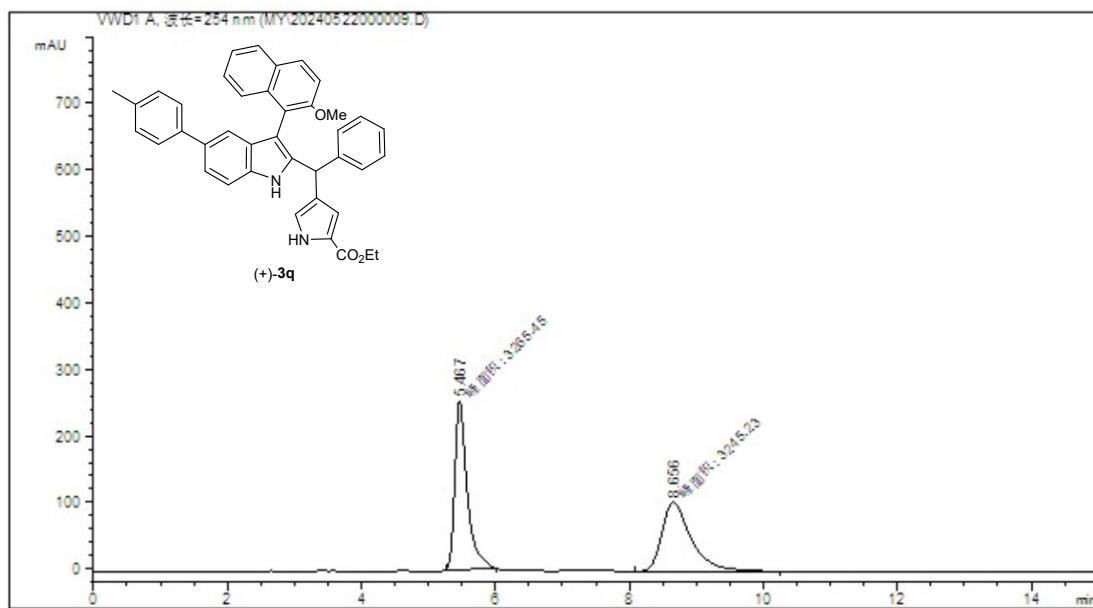


#	[min]		[min]	mAU	*s	[mAU ]	%
1	5.567	VV	0.1740	8552.01172	725.05981	50.1182	
2	6.548	VV	0.2526	8511.66797	492.51593	49.8818	

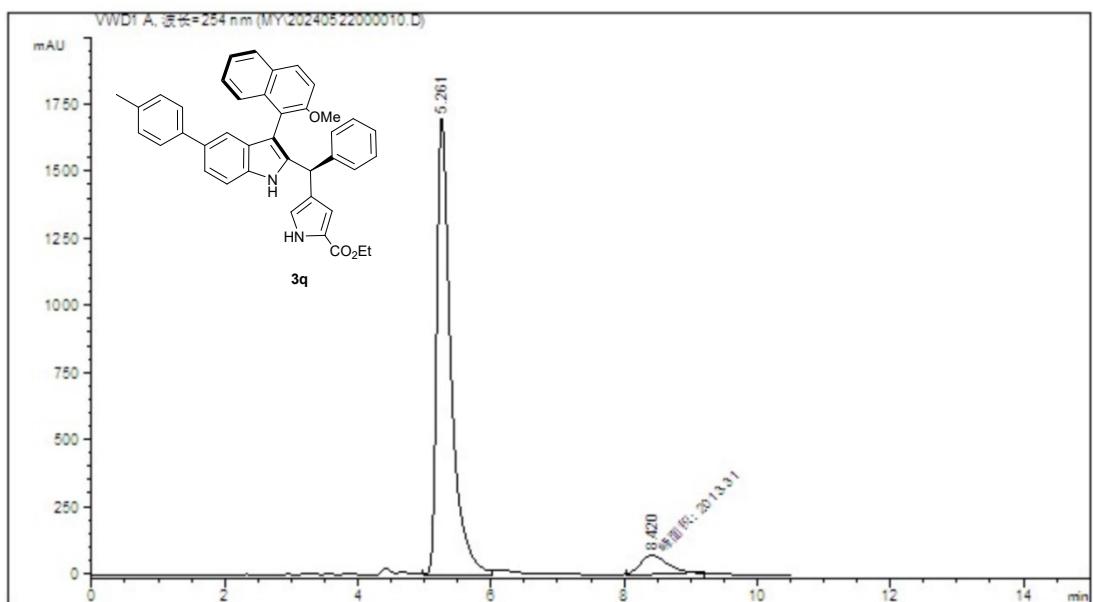


#	[min]		[min]	mAU	*s	[mAU ]	%
1	5.545	VV	0.1752	1.34676e4	1143.96814	96.2794	
2	6.558	MM	0.2515	520.43311	34.48179	3.7206	

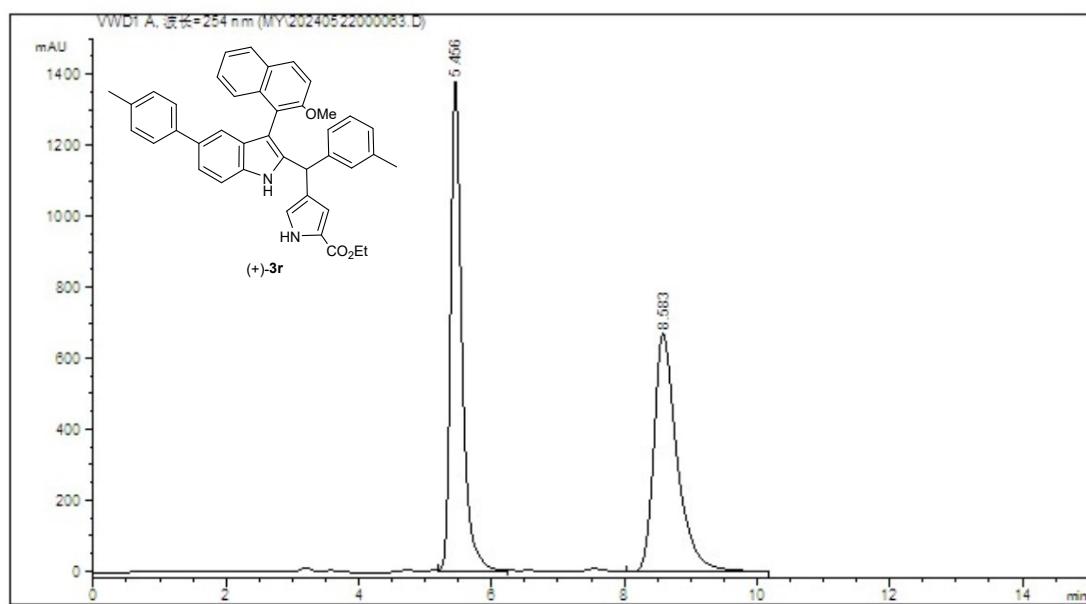




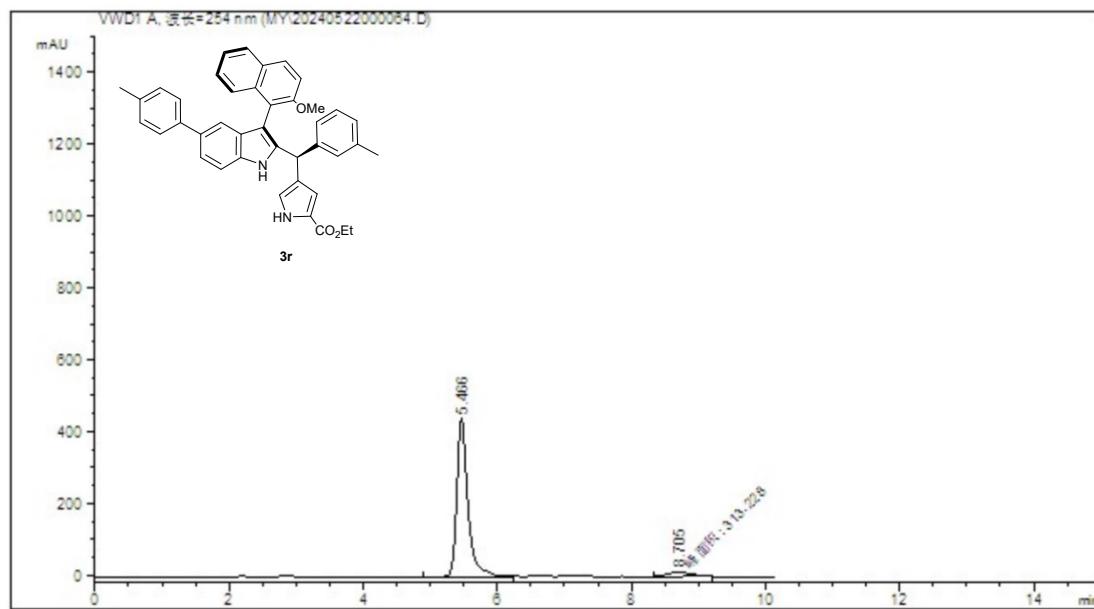
#	[min]	[min]	mAU	*s	[mAU ]	%
1	5.467	MM	0.2148	3265.44946	253.37117	50.1553
2	8.656	MM	0.5158	3245.23022	104.86482	49.8447



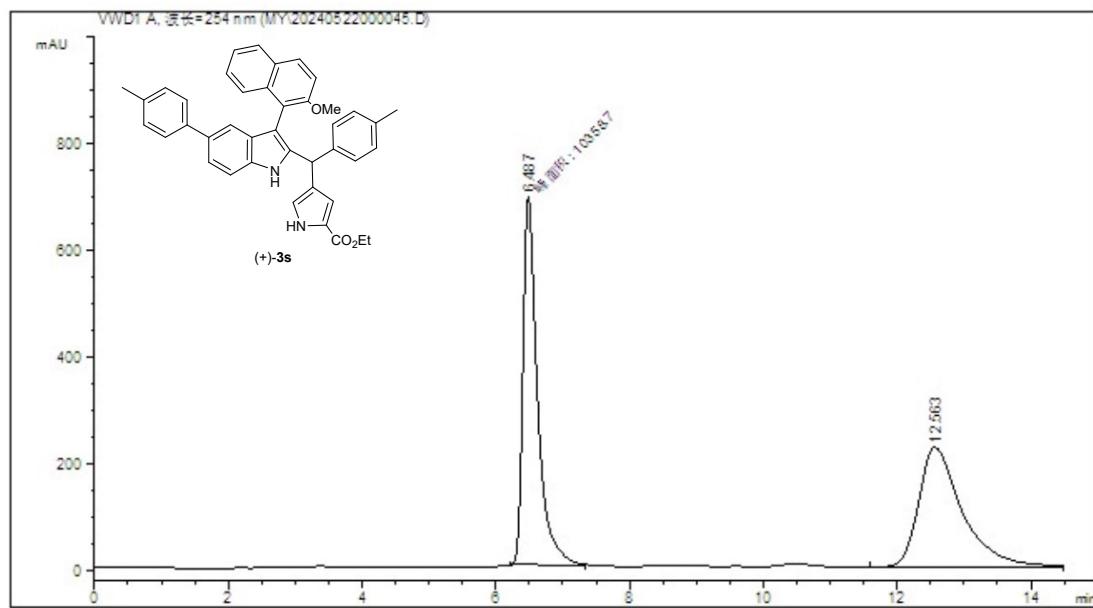
#	[min]	[min]	mAU	*s	[mAU ]	%
1	5.261	VV	0.2189	2.51450e4	1702.90662	92.5868
2	8.420	MM	0.4718	2013.30591	71.11877	7.4132



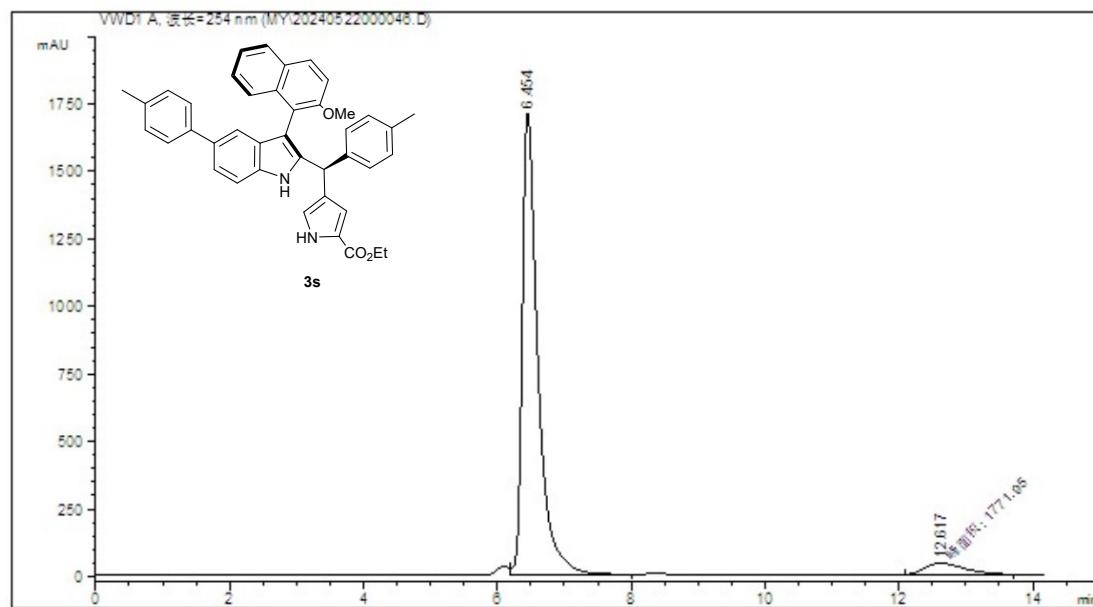
#	[min]		[min]	mAU	*s	[mAU]		%
1	5.456	VB	0.1786	1.65008e4	1381.70605	50.3402		
2	8.583	VBA	0.3608	1.62778e4	669.64618	49.6598		



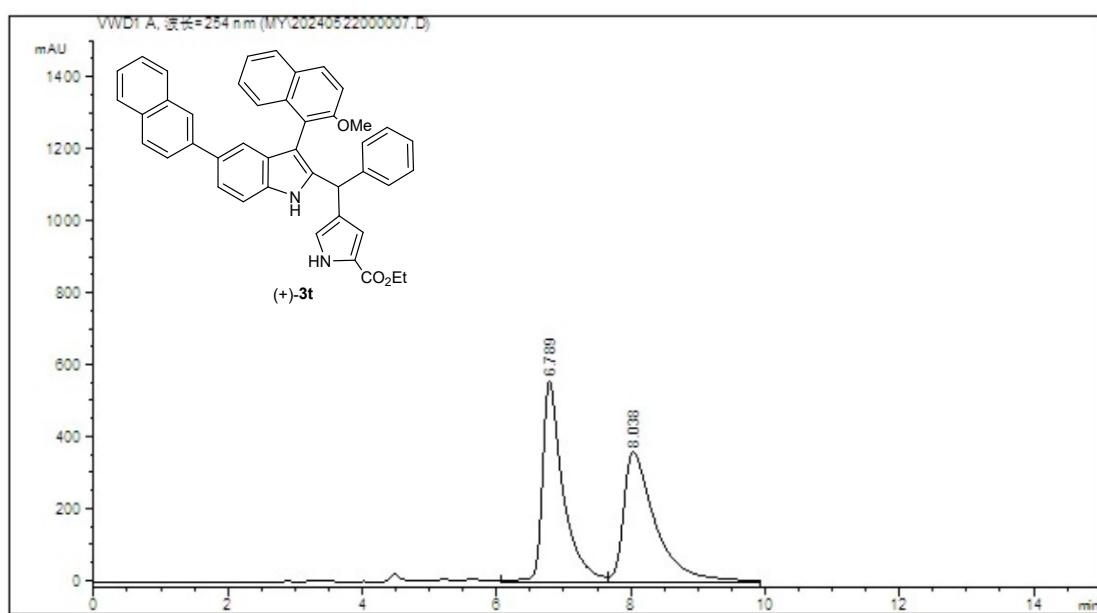
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1	5.466	VB	0.1699	5025.78906	439.15656	94.1332	
2	8.705	MM	0.3835	313.22781	13.61135	5.8668	



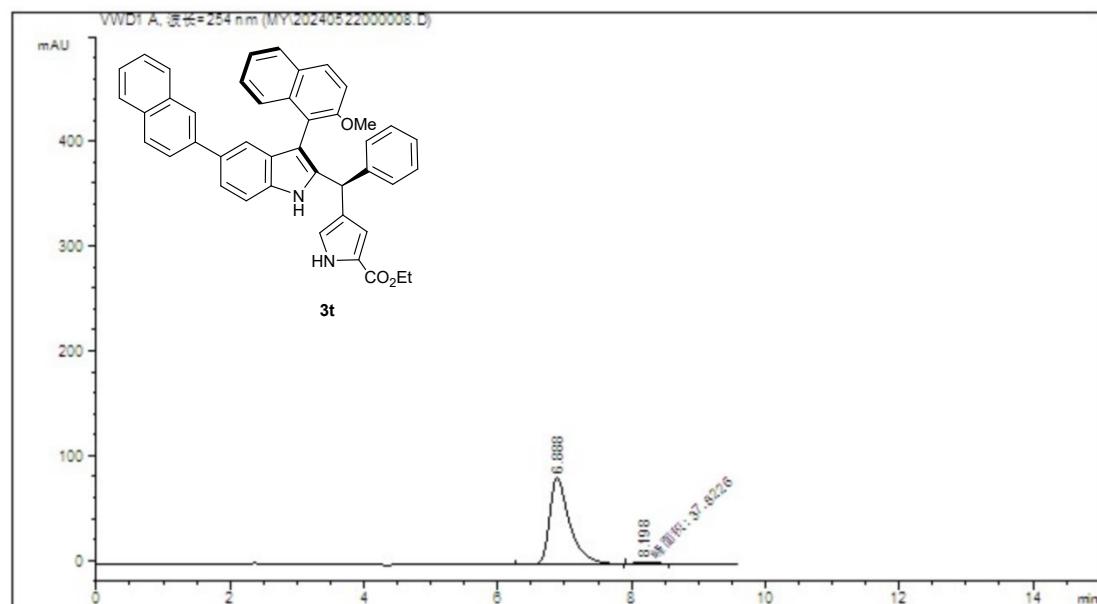
#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.487	MM	0.2507	1.03587e4		688.71466	49.7775
2	12.563	BBA	0.6869	1.04513e4		225.87254	50.2225



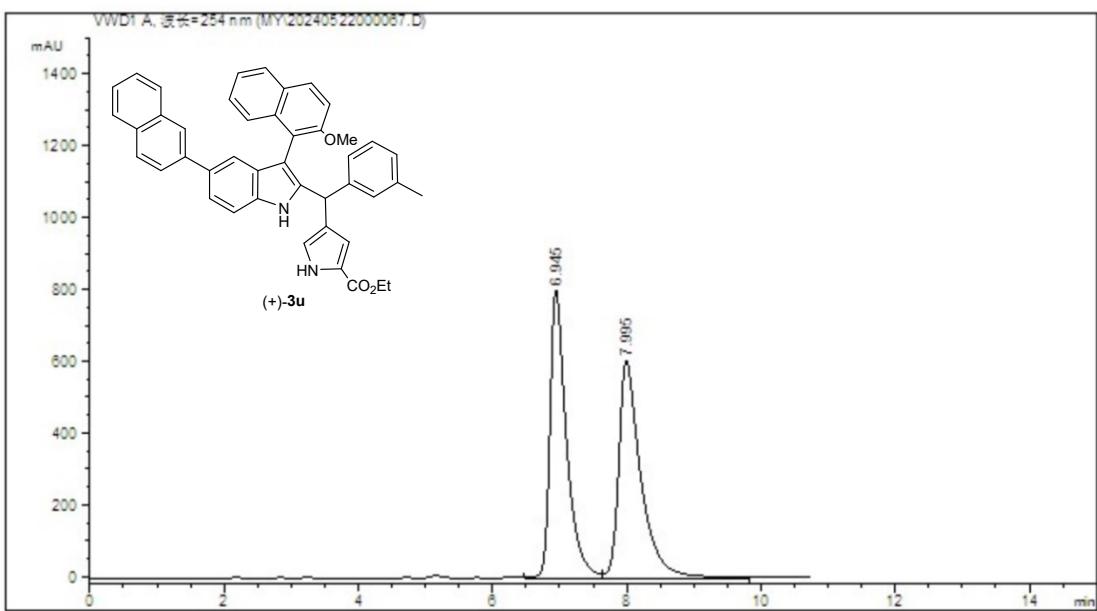
#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.454	VV	0.2412	2.79543e4		1713.93213	94.0391
2	12.617	MM	0.6895	1771.95166		42.83247	5.9609



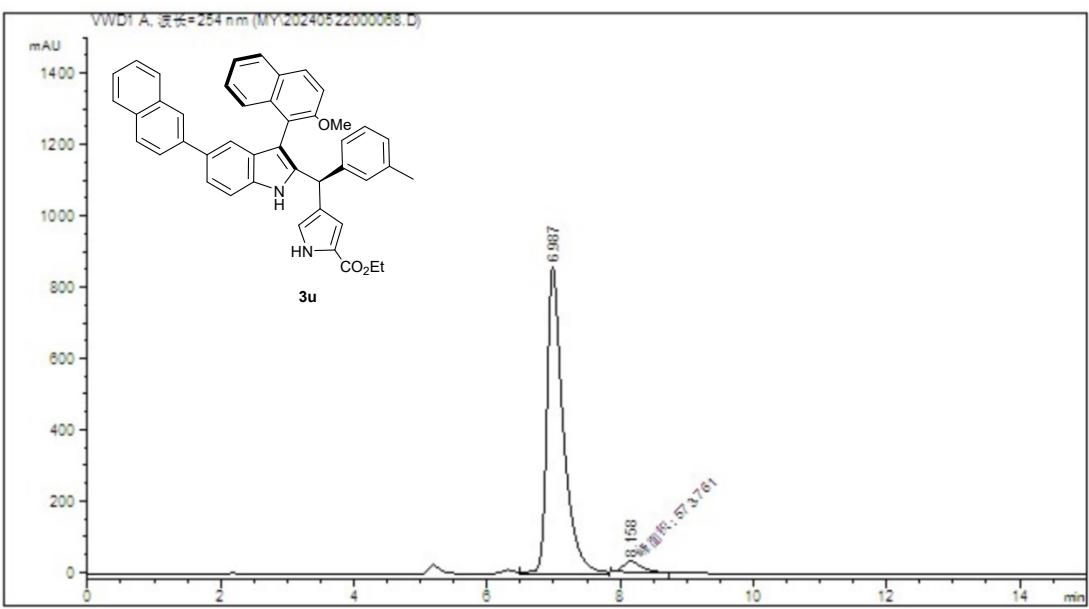
#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.789	VV	0.3065	1.19717e4		559.21002	50.0431
2	8.038	VBA	0.4805	1.19510e4		361.30957	49.9569



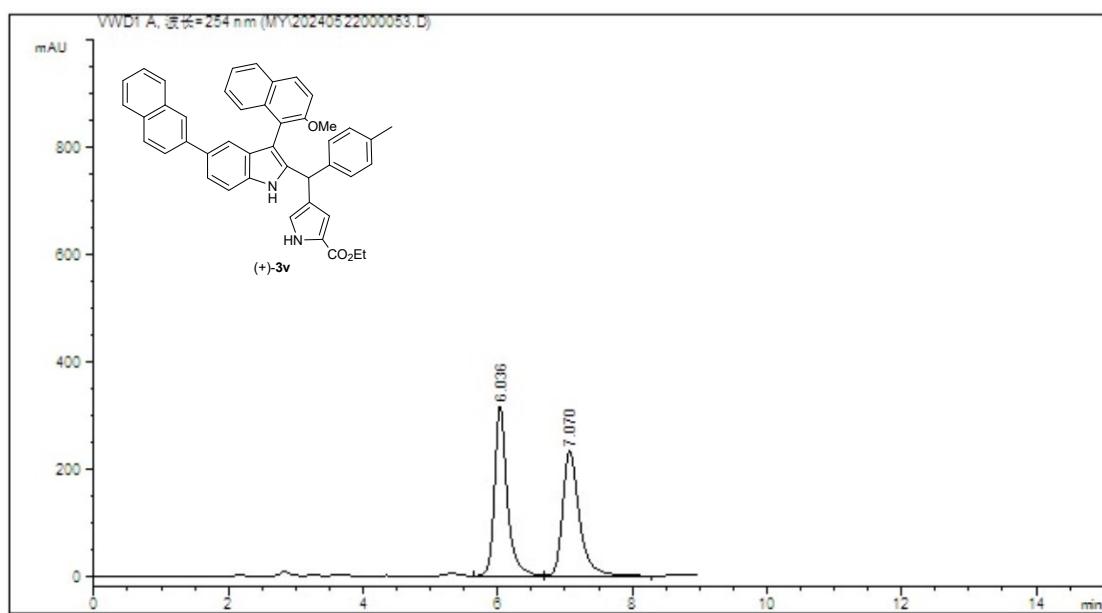
#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.888	BB	0.3142	1773.25452		83.19477	97.9116
2	8.198	MM	0.3554	37.82264		1.77376	2.0884



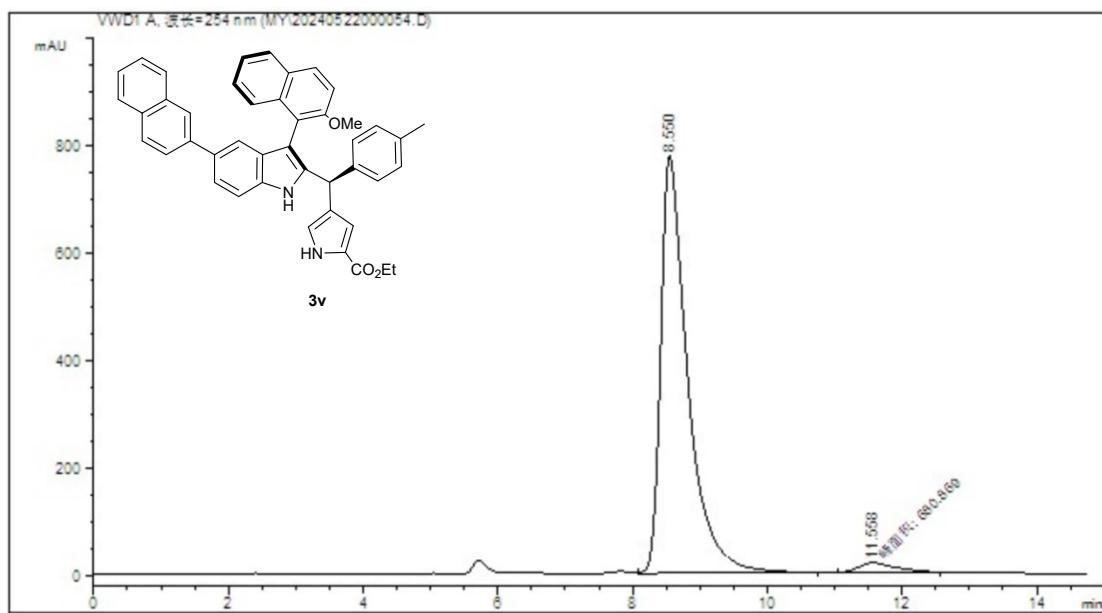
#	[min]		[min]	mAU	*s	[mAU]	%
1	6.945	VV	0.2491	1.37154e4		801.78009	49.6706
2	7.995	VV	0.3303	1.38973e4		605.75421	50.3294



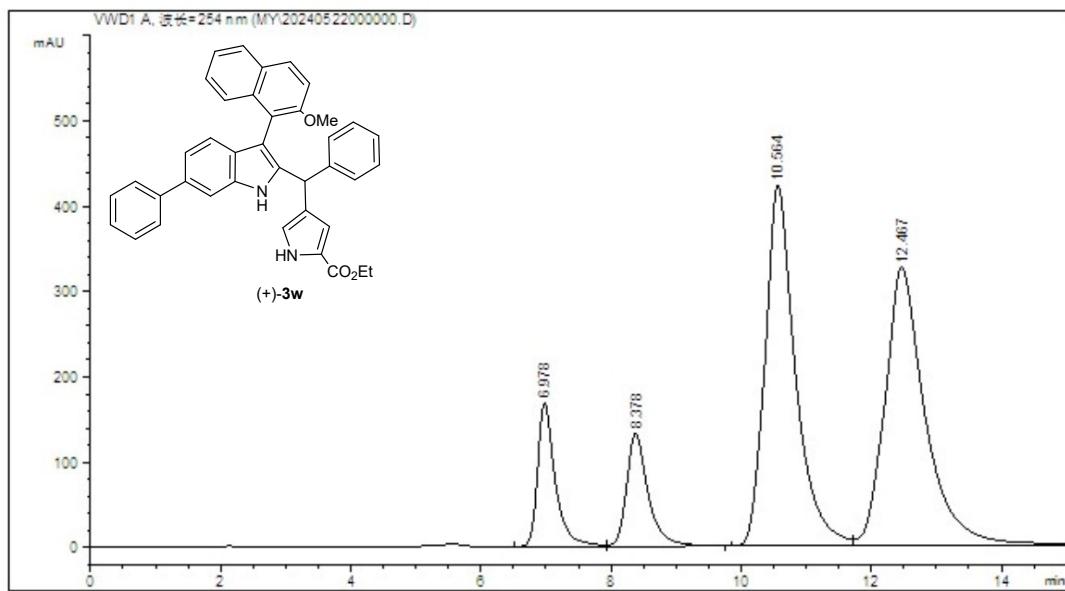
#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.987	VV	0.2470	1.46677e4	859.93134	96.2355	
2	8.158	MM	0.3225	573.76099	29.65424	3.7645	



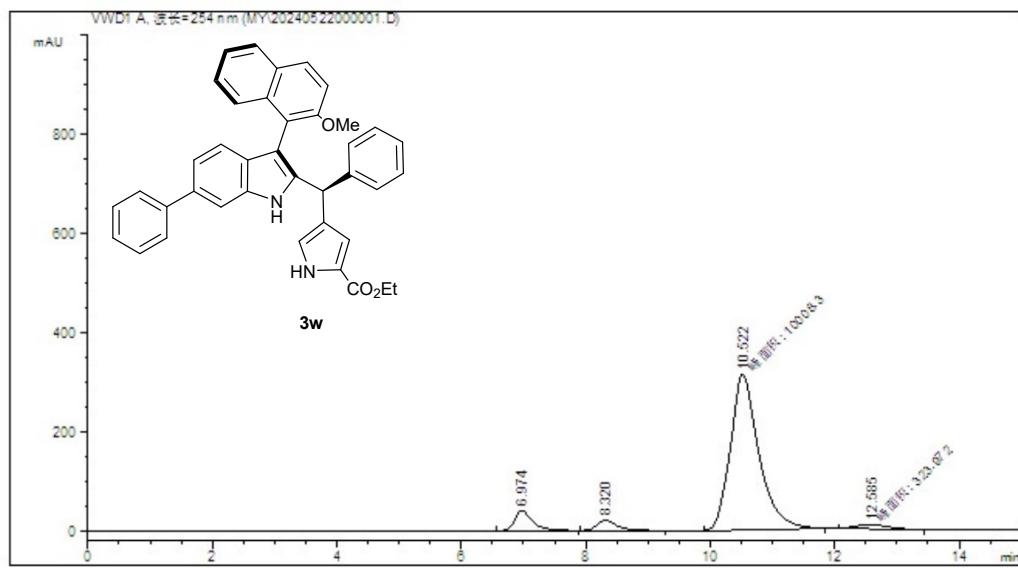
#	[min]		[min]	mAU	*s	[mAU]	*
1	6.036	VV	0.1964	4183.91553		316.59283	50.3790
2	7.070	VV	0.2657	4120.96533		233.37425	49.6210



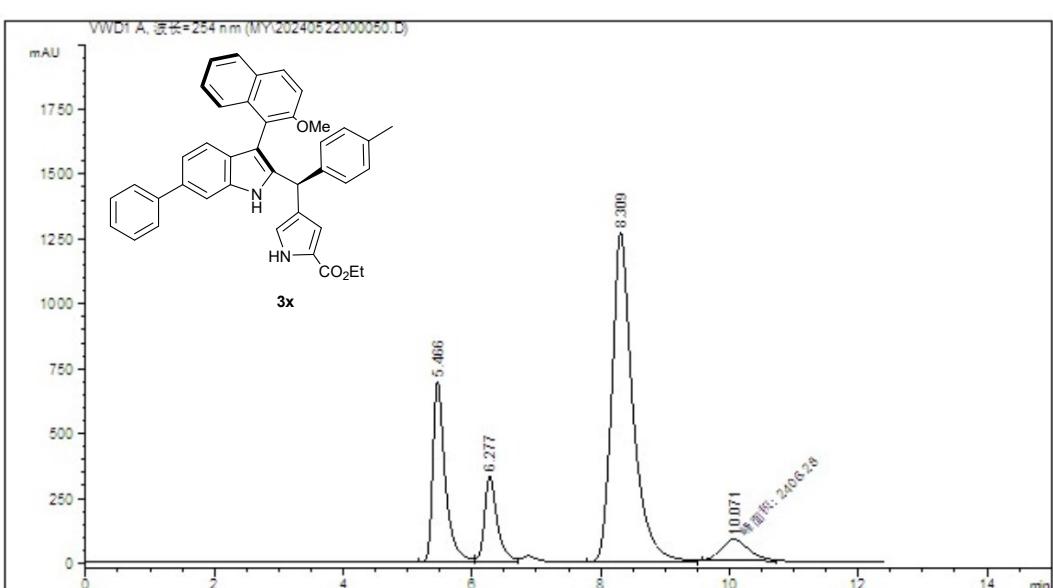
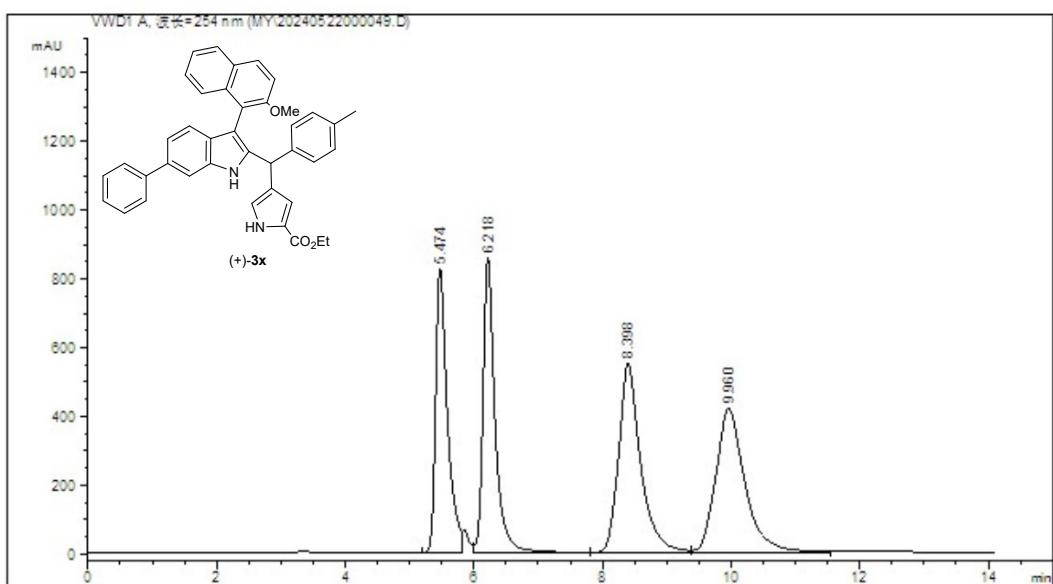
#	[min]		[min]	mAU	*s	[mAU]	*
1	8.550	VB	0.3974	2.14629e4	774.88409	96.9252	
2	11.558	MM	0.6372	680.86920	17.80779	3.0748	

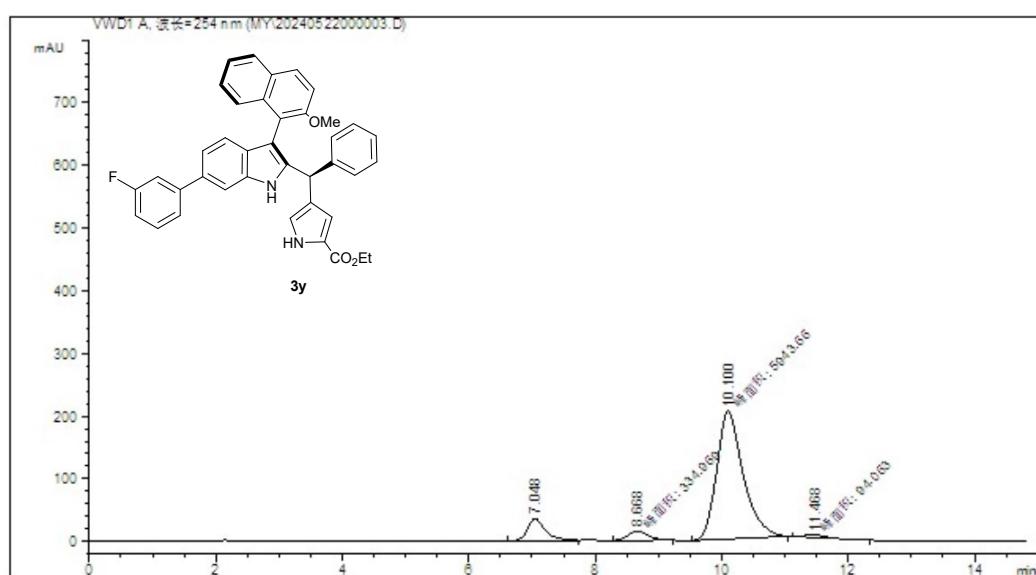
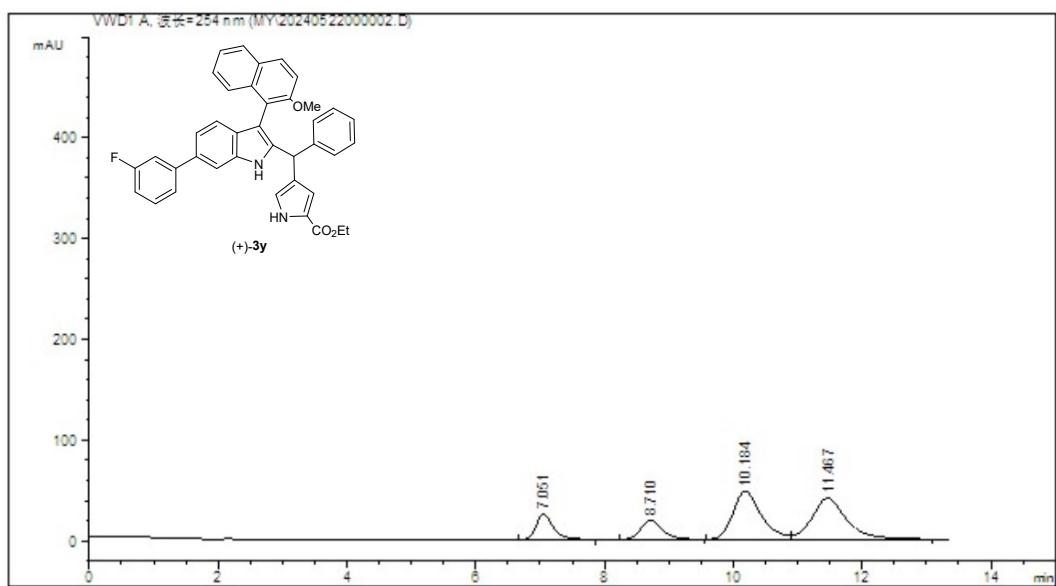


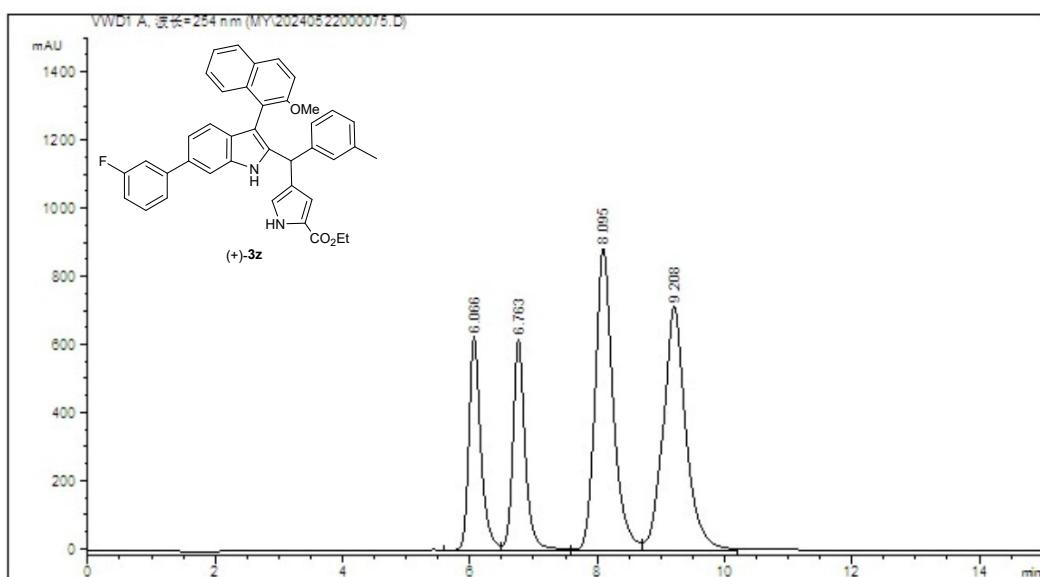
#	[min]		[min]	mAU	*s	[mAU]	%
1	6.978	VV	0.2771	3158.61450	168.40199	9.3382	
2	8.378	VB	0.3391	3035.80640	132.25969	8.9751	
3	10.564	BV	0.4633	1.36798e4	422.86896	40.4432	
4	12.467	VB	0.6067	1.39504e4	326.18692	41.2434	



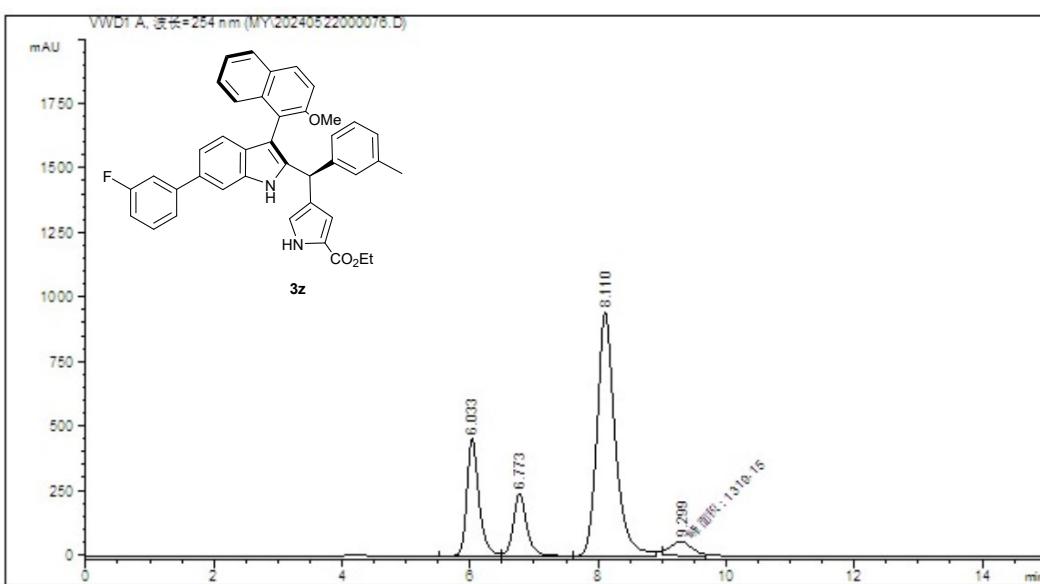
#	[min]		[min]	mAU	*s	[mAU]	%
1	6.974	BB	0.2956	824.44287		41.02570	7.0802
2	8.320	BB	0.3347	487.57346		21.48322	4.1872
3	10.522	MM	0.5307	1.00083e4		314.29877	85.9503
4	12.585	MM	0.6037	323.97229		8.94362	2.7822



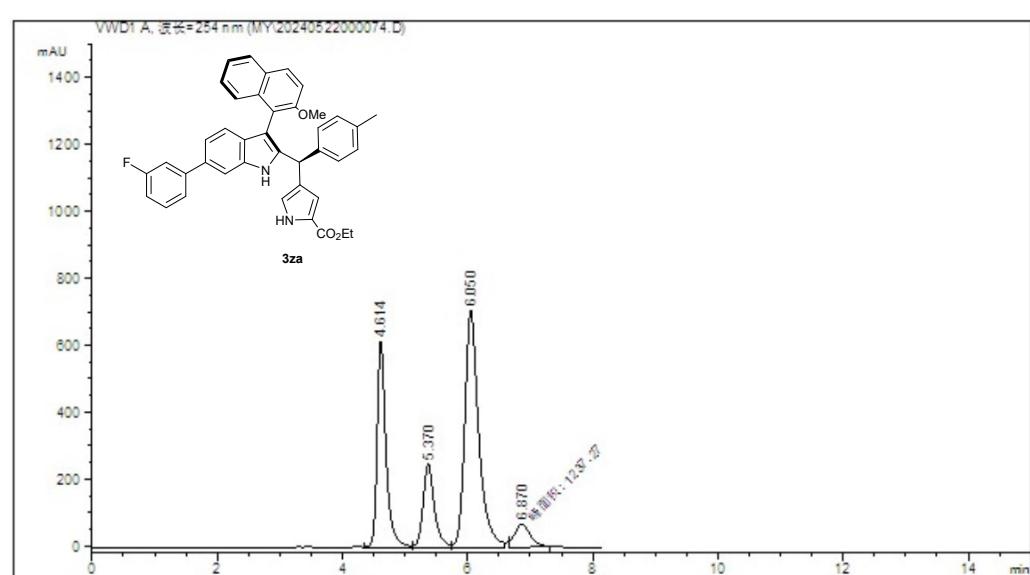
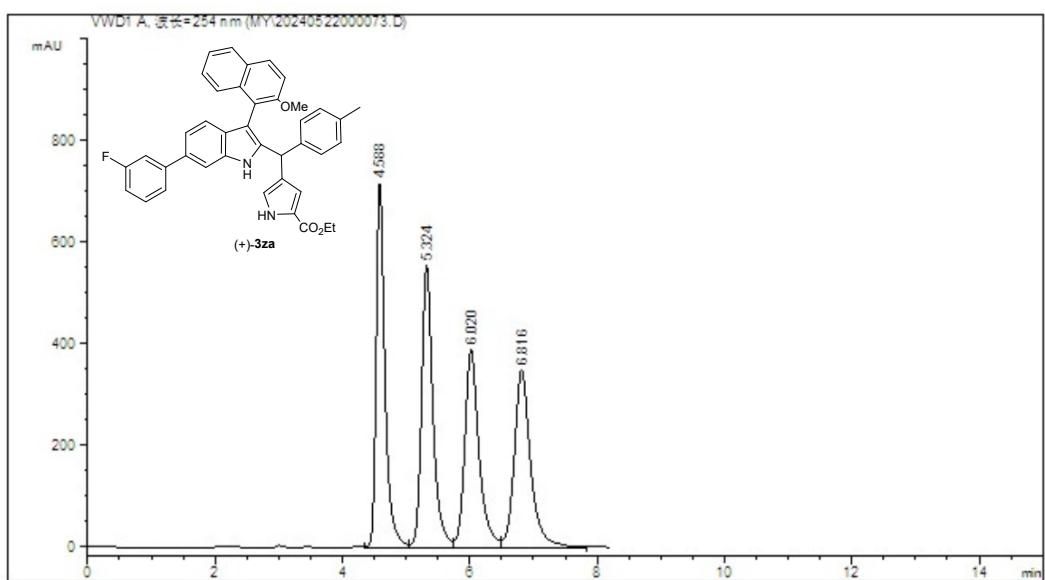


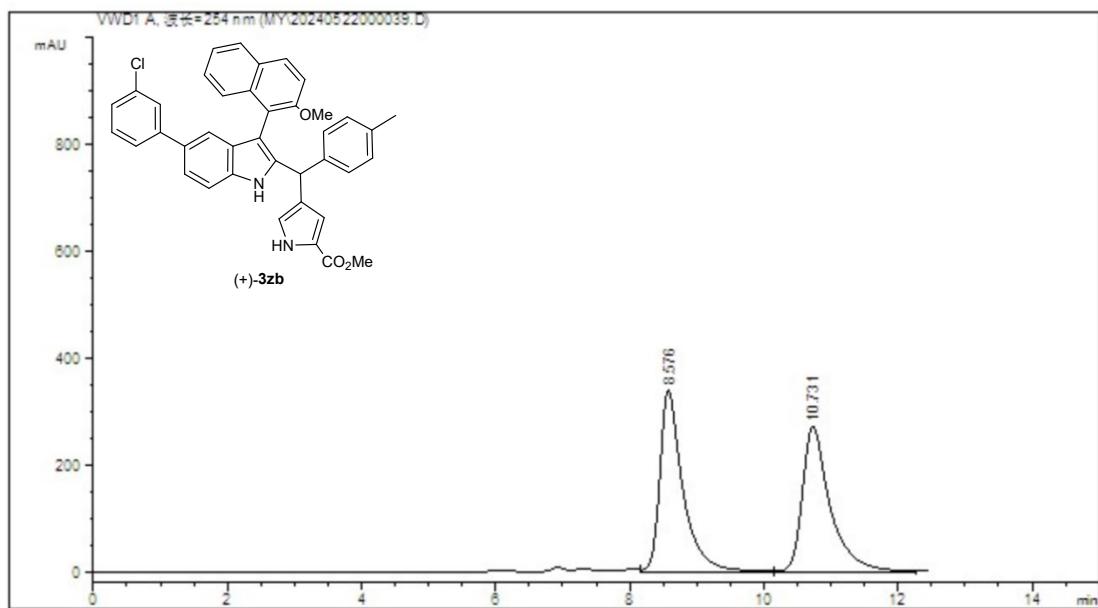


#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.066	VV	0.1975	8289.38086	628.57996	15.9371	
2	6.763	VV	0.2010	8397.34766	622.83313	16.1446	
3	8.095	VV	0.2872	1.74520e4	889.43494	33.5530	
4	9.208	VV	0.3549	1.78745e4	717.08948	34.3653	

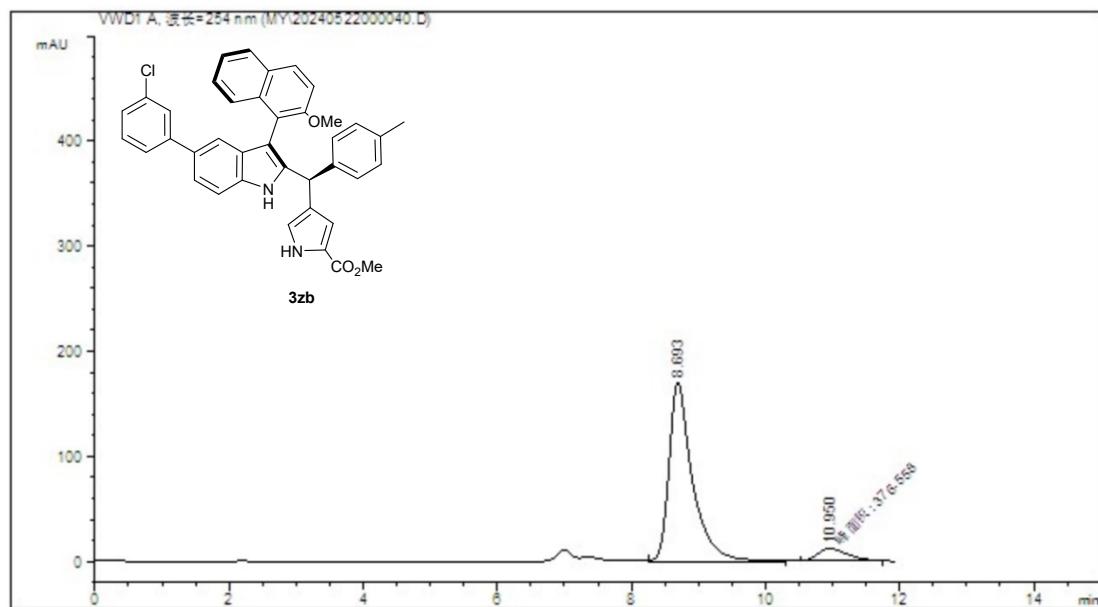


#	[min]		[min]	mAU	*s	[mAU ]	%
1	6.033	VV	0.1996	6035.75391	456.00613	20.4259	
2	6.773	VV	0.2170	3569.91455	244.40172	12.0811	
3	8.110	VV	0.2878	1.86247e4	946.43854	63.0288	
4	9.299	MM	0.3928	1319.15405	55.97678	4.4642	

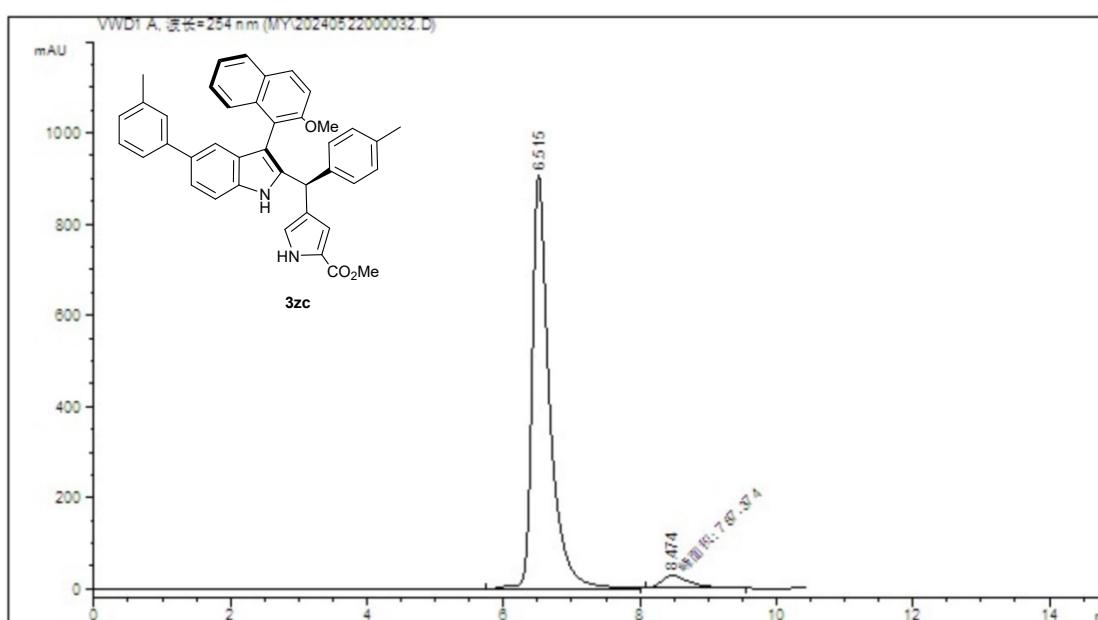
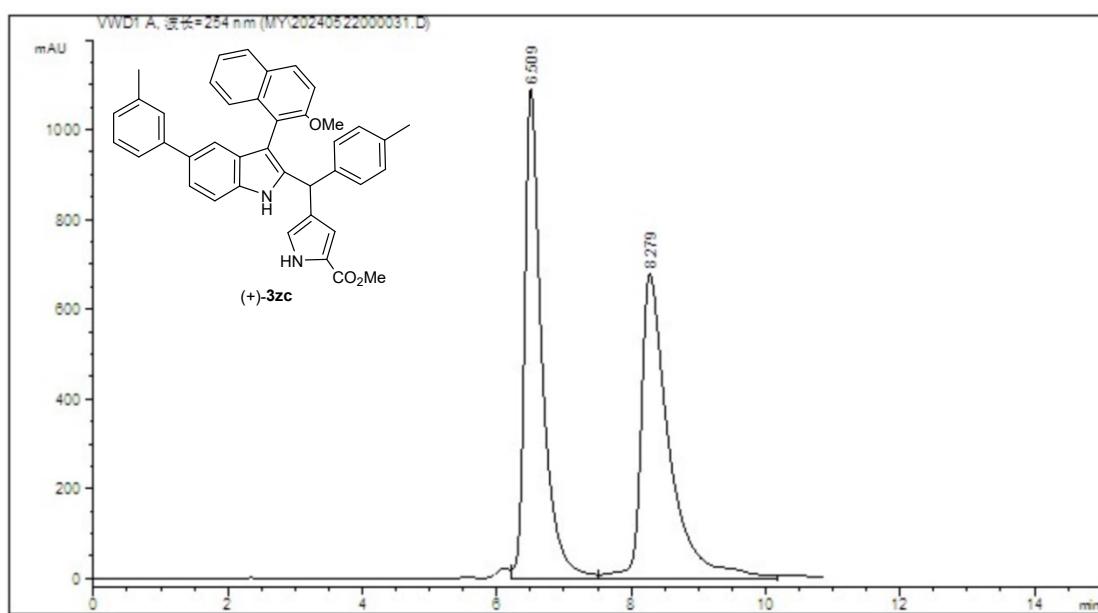


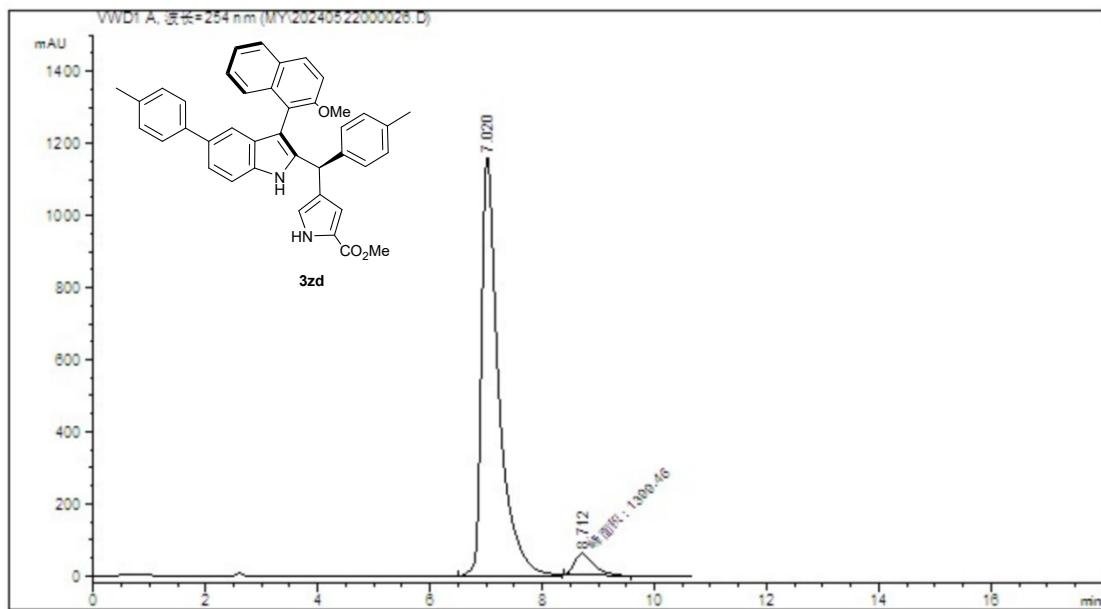
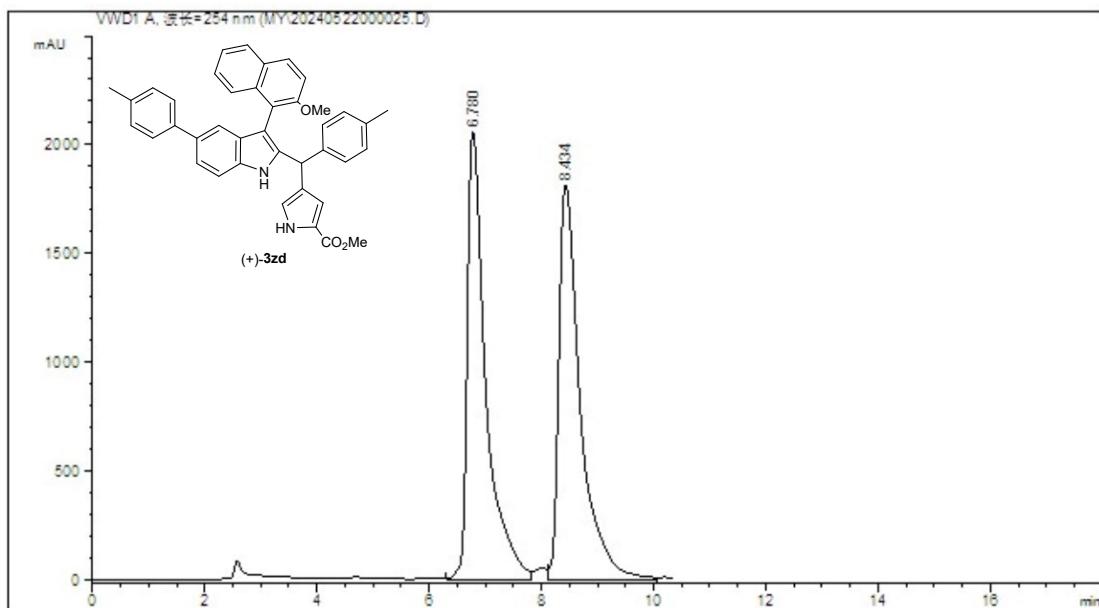


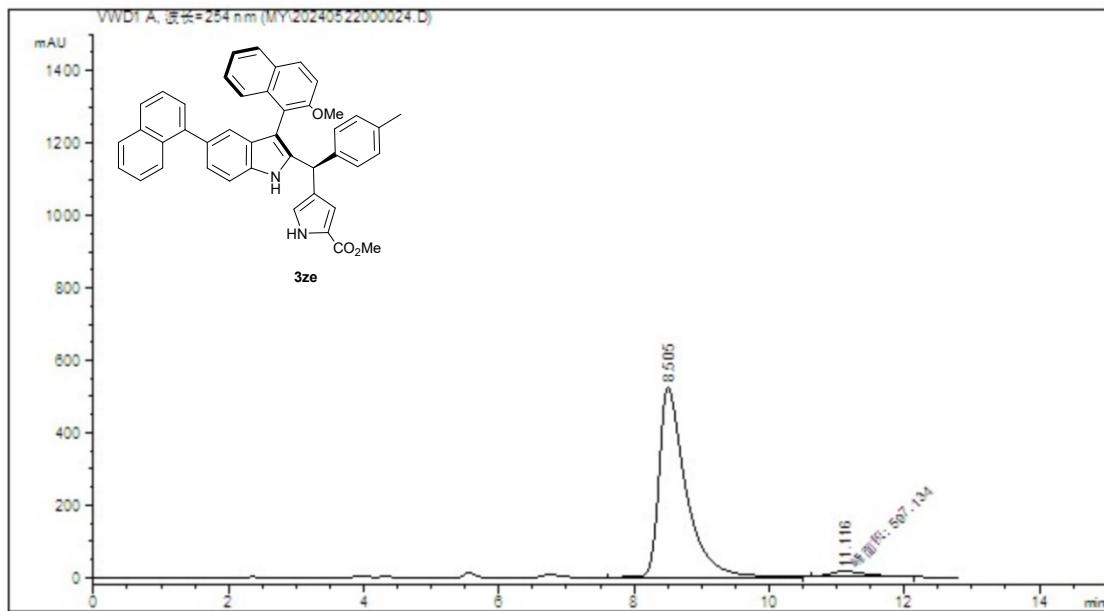
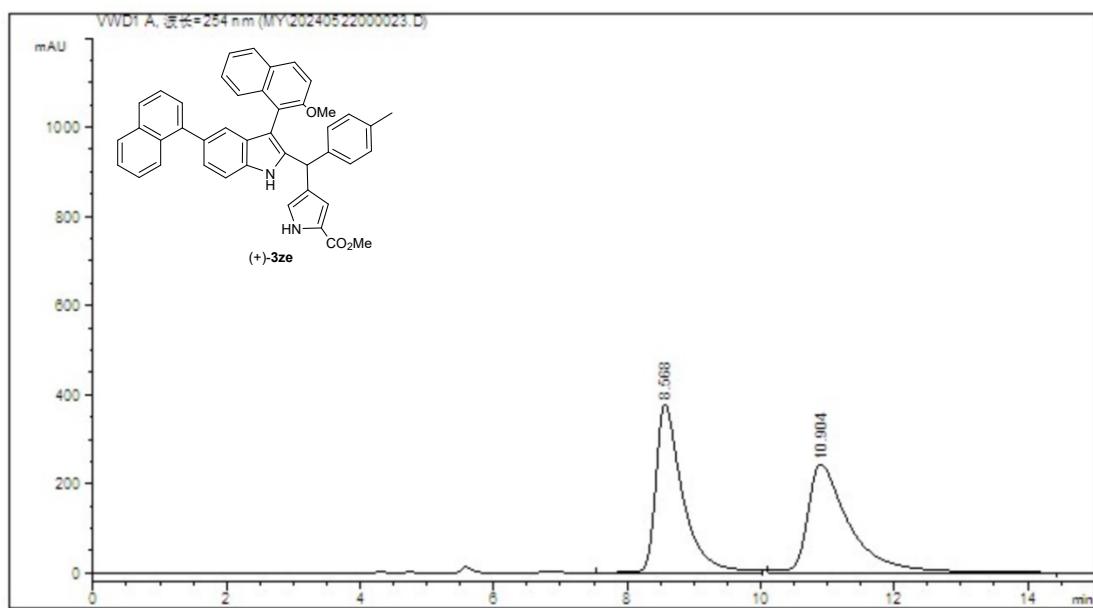
#	[min]		[min]	mAU	*s	[mAU]		%
1	8.576	VV	0.3481	8215.85840	340.90417	340.90417	50.7466	
2	10.731	VB	0.4259	7974.10938	272.45724	272.45724	49.2534	

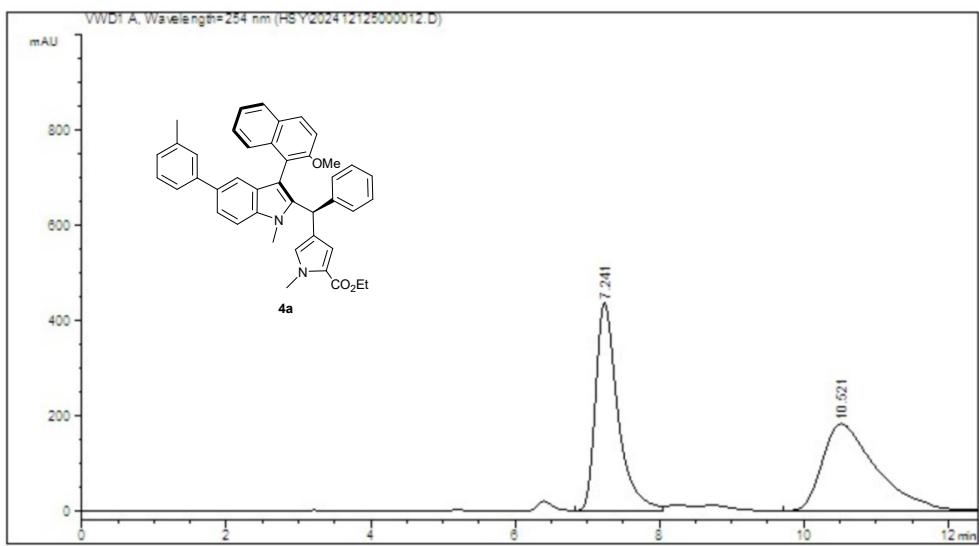


#	[min]		[min]	mAU	*s	[mAU]		%
1	8.693	VB	0.3516	4138.62646	170.53734	170.53734	91.6602	
2	10.950	MM	0.5120	376.55795	12.25847	12.25847	8.3398	







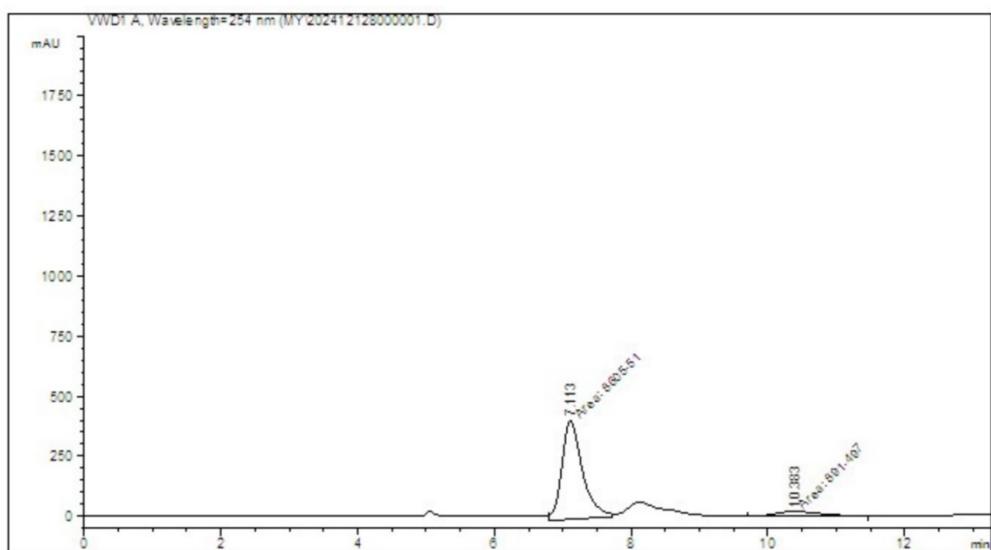


Area Percent Report

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	Height *s	Area [mAU ]	Area %
1	7.241	VV	0.3244	9416.25000	436.54391	49.5612	
2	10.521	BBA	0.7591	9582.97949	183.09856	50.4388	

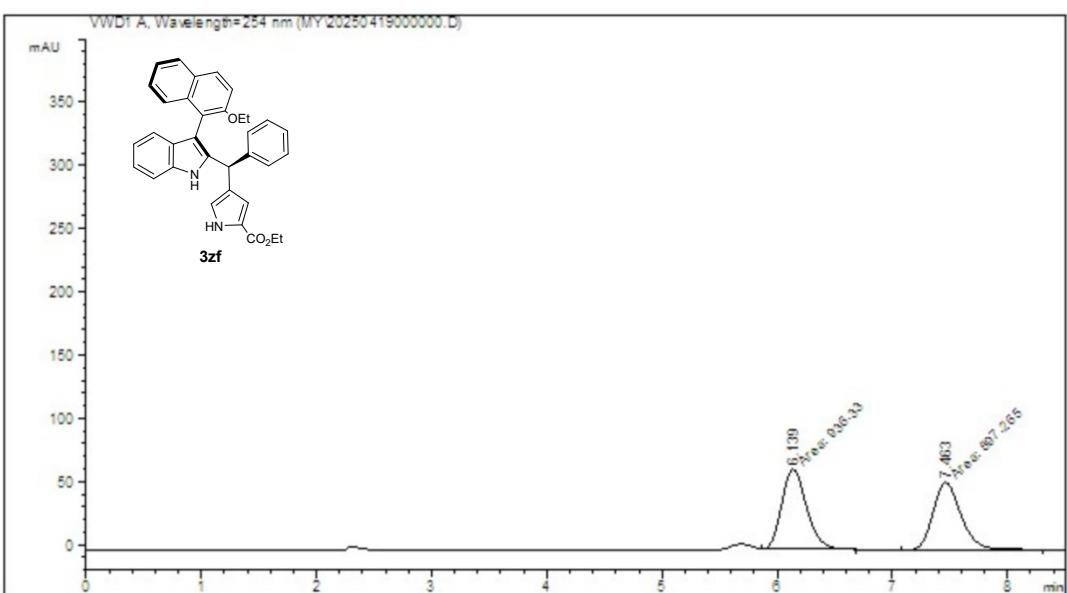


=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	Height *s	[mAU]	Area %
1	7.113	MM	0.3505	8605.50879	409.15839	90.6129	
2	10.383	MM	0.7615	891.49750	19.51092	9.3871	

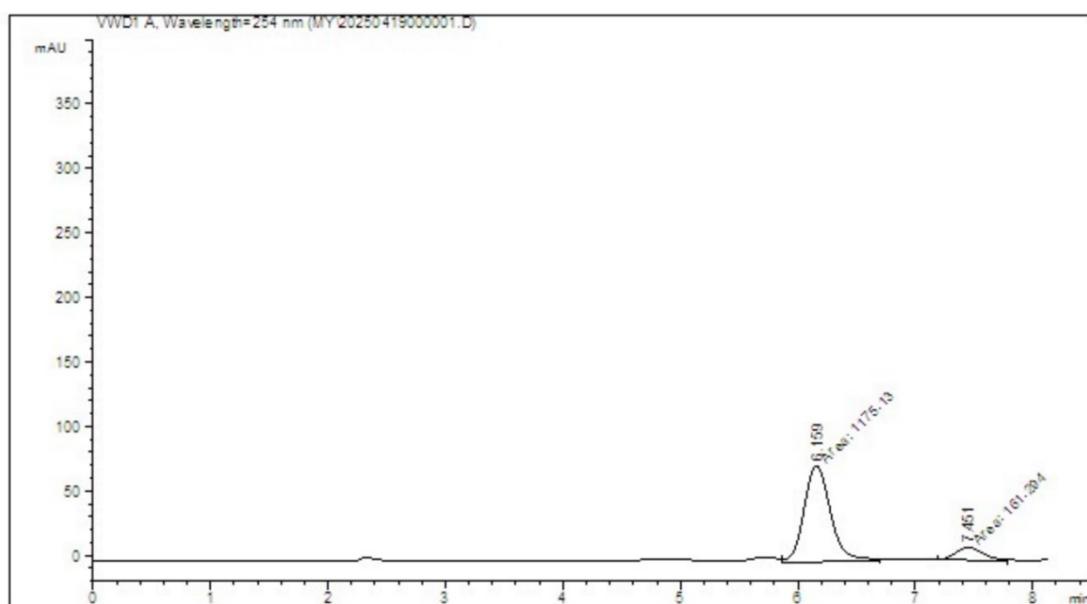


=====  
Area Percent Report  
=====

Sorted By : Signal  
 Multiplier : 1.0000  
 Dilution : 1.0000  
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	Height *s	Area [mAU]	Area %
1	6.139	MM	0.2484	936.33014	62.82299	51.0653	
2	7.463	MM	0.2801	897.26483	53.39255	48.9347	



=====  
Area Percent Report  
=====

Sorted By : Signal  
Multiplier : 1.0000  
Dilution : 1.0000  
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	*s	Height [mAU]	Area %
1	6.159	MM	0.2627	1175.13269		74.54086	87.9309
2	7.451	MM	0.2746	161.29445		9.79115	12.0691

## 7. References

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