Supplementary Information

Superacid-Catalysed α -Deuteration of Ketones with D₂O

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1. General Information

1.1 Reagents and materials.

All chemicals were purchased from commercial suppliers and used directly as received. Deuterated solvents were purchased from Konoscince and directly used. Unless otherwise noted, reagents were commercially available and used without further purification. All reactions were conducted under indicated atmosphere by using standard Schlenk techniques. All glassware and vials were oven-dried prior to use. All work-up and purification procedures were carried out with reagent-grade solvents.

1.2 Analytical methods.

Analytical thin-layer chromatography (TLC) was performed using glass plates precoated with 0.25 mm Yantai silica plates (GF-254). The developed chromatography was analyzed by UV lamp (254 nm and 365 nm). Flash chromatography was performed on 200-300 mesh Yantai silica gel with the indicated eluents. Nuclear magnetic resonance (¹H NMR, ¹³C NMR, ¹⁹F NMR, ¹¹B NMR and spectra were recorded respectively at 500 MHz or 400 MHz and 126 MHz or 101 MHz in indicated deuterated solvents (CDCl₃, or DMSO-*d*₆). Fluorine nuclear magnetic resonance spectra (¹⁹F NMR) were recorded at 376 MHz, and the chemical shifts were accurate to one decimal place or two decimal places to help distinguish overlapping peaks. Chemical shifts were expressed in parts per million (ppm) downfield from tetramethylsilane and refer to the solvent signals (CDCl₃: δ H 7.26 and δ C 77.16 ppm; DMSO-*d*₆: δ H 2.50 and δ C 39.50 ppm). The signals of water were observed at about 1.58 ppm in CDCl₃ and 3.33 ppm in DMSO- d_6 , respectively. Coupling constants (J) were reported in Hertz (Hz). Splitting patterns were designated as s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad; dd, doublet of doublets, etc. High resolution mass spectrometry (HRMS) data were obtained on a Q-Tof MS/MS system (Agilent 1290-6546) using electrospray ionization (ESI) in positive mode.

2. General Procedure

General procedure for the deuteration: To a well-dried Schlenk tube (10 mL) equipped with a magnetic stir bar, $[Ph_3C]^+[B(C_6F_5)_4]^-$ (0.01 mmol, 10 mol%) and substrates (0.1 mmol, 1.0 equiv.) were added. Then, the tube was evacuated and backfilled with argon for three times before 0.2 mL deuterated water and 1.0 mL solvent were charged via syringe. Subsequently, the tube was sealed with screw cap. The resulting solution was heated in an oil bath at 100 °C and stirred vigorously for 10 hours. Once the reaction was complete, the mixture was diluted with ethyl acetate (2 mL), extracted with ethyl acetate (3*5 mL), combined organic solvent, washed with saturated sodium chloride solution, dried with anhydrous sodium sulfate, filtered by a sand core funnel. The filtrate was concentrated under reduced pressure and the residue was purified by preparative thin layer chromatography or flash column chromatography eluted with hexane/ethyl acetate to obtain the products.

3. Optimization of the Reaction Conditions

Table S1. Screening the optical reaction conditions

H ₃ C N N N CH 0.1 from Pe	$ \begin{array}{c} & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ $	H ₃ 4] ⁻ H ₃ 4 →	C N N C H_3 d_5 -Pento	y% y% CD ₃ 96% 95% x%
Entry	Change from the "standard conditions"	x (%ª)	y (%ª)	Yield(% ^b)
1	none	95	96	93(89 ^{<i>d</i>})
2	without [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] ⁻	n.d.	n.d.	99
3	[Ph ₃ C] ⁺ [BF ₄] ⁻ instead of [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] ⁻	n.d.	n.d.	90
4	K ⁺ [B(C ₆ F ₅) ₄] ⁻ instead of [Ph ₃ C] ⁺ [B(C ₆ F ₅) ₄] ⁻	40	34	86
5	DCM instead of DCE	91	96	86
6	90 °C instead of 100 °C	90	93	94
7	8 h instead of 10 h	89	93	86
8	oxygen instead of argon	50	50	82

^{*a*}Reaction conditions: Pentoxifylline (0.1 mmol, 27.8 mg), $[Ph_3C]^+[B(C_6F_5)_4]^-$ (5 mol%, 0.005 mmol), D₂O (55 equiv., 0.2 mL), DCE (1 mL), 100 °C, Argon, 10 hours. ^{*b*}Yield of deuteration was based on hydrogen highlighted in red. ^{*c*}All yields are NMR yields by using CH₃NO₂ as an internal standard. ^{*d*}Isolated yields. n.d.: no deuteration.

4. Gram-scale Reactions



Figure S1. Gram-scale synthesis of *d*₅-Pentoxifylline

10 mmol-scale synthesis of d_5 -Pentoxifylline (1): To a well-dried round bottom flask (250 mL) equipped with a magnetic stir bar, [Ph₃C]⁺[B(C₆F₅)₄]⁻ (461 mg, 0.5 mmol, 0.05 equiv.) and Pentoxifylline (2.78 g, 10 mmol, 1.0 equiv.) were added. Then, the flask was sealed, evacuated and backfilled with Ar for three times before 10 mL deuterated water and 45 mL DCE were charged via syringe. The resulting yellow solution was heated in an oil bath at 100 °C and stirred vigorously for 10 hours. When the reaction was complete, the mixture was diluted with DCE (20 mL), extracted with DCE (3*20 mL), combined organic solvent, washed with saturated sodium chloride solution, dried with anhydrous sodium sulfate, filtered by a funnel. The filtrate was concentrated under reduced pressure and the residue was stirred with the mixture of PE/EA (100:1) for 12 hours and filtered to obtain the desired compound d_5 -Pentoxifylline (2.69 g, 95%) as a white solid. TLC: Rf = 0.35 (silica gel, PE/EA, 5:1).

5. Preliminary Mechanistic Studies



5.1 Reverse D/H exchange.

Figure S2. Reverse D/H exchange using *d*₅-Pentoxifylline

Reverse D/H exchange using d_5 -Pentoxifylline: To a well-dried Schlenk tube (10 mL) equipped with a magnetic stir bar, $[Ph_3C]^+[B(C_6F_5)_4]^-$ (4.6mg, 0.005 mmol, 5 mol%) and d_5 -Pentoxifylline (28.3mg, 0.1 mmol, 1.0 equiv.) were added. Then, the tube was evacuated and backfilled with argon for three times before 0.2 mL water and 1.0 mL DCE were charged via syringe. Subsequently, the tube was sealed with screw cap. The resulting solution was heated in an oil bath at 100 °C and stirred vigorously for 10 hours. Once the reaction was complete, the mixture was diluted with ethyl acetate (2 mL), extracted with ethyl acetate (3*5 mL), combined organic solvent, washed with saturated sodium chloride solution, dried with anhydrous sodium sulfate, filtered by a sand core funnel. The filtrate was concentrated under reduced pressure and the residue was purified by preparative thin layer chromatography or flash column chromatography eluted with hexane/ethyl acetate to obtain the products.

5.2 Deuteration with 1 equiv. Brønsted acids vs. 10 mol% lon pair.



Figure S3. Deuteration with 1 equiv. Brønsted acids vs. 10 mol% lon pair.

Reactions were performed with on a scale of 0.1 mmol Pentoxifylline in presence of 1.0 eq. Brønsted acids in 0.2 mL D₂O (55 equiv.) and 1.0 mL DCE under Ar atmosphere at 100 °C for 10 h. Yield of deuteration was based on hydrogen highlighted in red. All of yields were isolated yields.



Figure S4. H/D exchange using 1.0 eq. Brønsted acids vs. 5 mol% $[Ph_3C]^+[B(C_6F_5)_4]^-$

5.3 Deuteration with 1.0 or 0.1 equiv. K₂CO₃ as catalyst.



Figure S5. Deuteration with 0.1 equiv. K₂CO₃



Figure S6. Deuteration with 1.0 equiv. K₂CO₃

Reactions were performed with on a scale of 0.1 mmol Pentoxifylline in presence of 1.0 eq. or 0.1eq. K_2CO_3 in 0.2 mL D₂O (55 equiv.) and 1.0 mL DCE under Ar atmosphere at 100 °C for 10 h. Yield of deuteration was based on hydrogen highlighted in red. All of yields were isolated yields.

6. Characterization Data of Products

3,7-dimethyl-1-(5-oxohexyl-4,4,6,6,6-*d*₅)-3,7-dihydro-1H-purine-2,6-dione



TLC: R*f* = 0.30 (silica gel, PE/EA, 2:1), (25mg, 89% yield), white solid. ¹H NMR (400 MHz, CDCl₃) δ 7.48 (s, 1H), 3.95 (d, *J* = 7.6 Hz, 5H), 3.52 (d, *J* = 0.7 Hz, 3H), 2.06 (p, *J* = 2.1 Hz, 0.10H), 1.99 (s, 0.12H), 1.68 – 1.57 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 209.17, 155.31, 151.51, 148.80, 141.54, 107.69, 40.86, 33.67, 29.75, 27.44, 20.89, 14.28, 5.95.

1-(3,5,5,6,8,8-hexamethyl-5,6,7,8-tetrahydronaphthalen-2-yl)ethan-1-one-2,2,2- d_3 (3)



TLC: R*f* = 0.35 (silica gel, PE/EA, 2:1), (25.3mg, 97% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 7.68 (s, 1H), 7.22 (s, 1H), 2.56 (dt, *J* = 3.7, 1.8 Hz, 0.15H), 2.54 (s, 3H), 1.91 (dqd, *J* = 13.4, 6.8, 2.6 Hz, 1H), 1.72 – 1.61 (m, 1H), 1.43 (dd, *J* = 13.5, 2.7 Hz, 1H), 1.35 (s, 6H), 1.30 (s, 3H), 1.10 (s, 3H), 1.02 (d, *J* = 6.8 Hz, 3H).

¹³**C NMR** (101 MHz, CDCl₃) δ 201.42, 150.17, 142.21, 135.34, 135.04, 130.62, 128.22, 55.46, 43.44, 37.90, 34.43, 34.03, 32.45, 31.96, 28.33, 24.70, 21.57, 16.79.

(4*R*,4a*S*,6*R*)-4,4a-dimethyl-6-(prop-1-en-2-yl)-4,4a,5,6,7,8hexahydronaphthalen-2(3*H*)-one-1,3,3,8,8-*d*₅ (4)



TLC: R*f* = 0.50 (silica gel, PE/EA, 2:1), (18.5mg, 83% yield), white solid.

¹H NMR (400 MHz, CDCl₃) δ 5.76 (s, 0.03H), 4.76 – 4.68 (m, 2H), 2.38 – 2.22 (m, 1H), 2.22 – 2.19 (m, 0.05H), 2.01 – 1.84 (m, 3H), 1.73 (d, *J* = 1.3 Hz, 3H), 1.38 – 1.28 (m, 1H), 1.11 (d, *J* = 6.0 Hz, 4H), 0.96 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 198.99, 169.63, 148.24, 127.03, 108.36, 43.02, 39.40, 38.34, 30.71, 30.53, 21.77, 19.94, 15.96, 13.98, 13.25.

HRMS (ESI-TOF) Calcd for C₁₅H₁₉D₃O: [M+Na]⁺ 246.1876 Found: m/z 246.1874.

(E)-6,10-dimethylundeca-5,9-dien-2-one-1,1,1,3,3-d₅ (5)



TLC: Rf = 0.30 (silica gel, PE/EA, 10:1), (12.9mg, 65% yield), transparent liquid. ¹H NMR (400 MHz, CDCl₃) δ 5.07 (t, J = 7.4 Hz, 2H), 2.43 (tdd, J = 7.4, 5.1, 2.5 Hz, 0.52H), 2.25 (q, J = 5.6, 4.9 Hz, 2H), 2.19 – 2.12 (m, 0.26H), 2.09 – 1.89 (m, 4H), 1.75 – 1.53 (m, 9H).

¹³**C NMR** (101 MHz, CDCl₃) δ 135.59 (d, *J* = 11.5 Hz), 130.85, 130.59, 123.30 (d, *J* = 2.4 Hz), 121.63, 38.77, 30.99, 25.73, 25.62, 24.85 (d, *J* = 2.9 Hz), 22.50, 16.81 (d, *J* = 4.1 Hz), 15.11.

HRMS (ESI-TOF) Calcd for C₁₃H₁₇D₅O: [M+K]⁺ 238.1616. Found: m/z 238.1630.

4-(2,6,6-trimethylcyclohex-1-en-1-yl)butan-2-one-1,1,1,3,3-*d*₅ (6)



TLC: R*f* = 0.34 (silica gel, PE/EA,10:1), (18.7mg, 94% yield), light liquid.

¹**H NMR** (500 MHz, CDCl₃) δ 2.47 (ddt, *J* = 9.6, 6.6, 2.5 Hz, 0.10H), 2.24 (s, 2H), 2.11 (p, *J* = 2.3 Hz, 0.15H), 1.90 (t, *J* = 6.4 Hz, 2H), 1.61 – 1.52 (m, 5H), 1.44 – 1.38 (m, 2H), 0.98 (s, 6H).

¹³**C NMR** (126 MHz, CDCl₃) δ 209.49, 135.91, 127.81, 89.65, 77.30, 77.05, 76.79, 43.82, 39.72, 35.03, 32.73, 29.09, 28.43, 22.15, 19.73, 19.45.

HRMS (ESI-TOF) Calcd for C₁₃H₁₇D₅O: [M+H]⁺ 200.2057. Found: m/z 200.2058.

(8*R*,9*S*,10*R*,13*S*,14*S*,17*R*)-17-ethynyl-17-hydroxy-10,13-dimethyl-1,2,6,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-3Hcyclopenta[*a*]phenanthren-3-one-2,2,4,6,6-*d*₅ (7)



TLC: R*f* = 0.25 (silica gel, EA), (25.6mg, 81% yield), white solid.

¹**H NMR** (500 MHz, CDCl₃) δ 5.73 (s, 0.07H), 2.58 (s, 1H), 2.30 (ddd, *J* = 13.8, 9.7, 5.5 Hz, 1H), 2.15 – 1.95 (m, 2H), 1.93 (s, 1H), 1.84 (dd, *J* = 12.8, 3.5 Hz, 1H), 1.80 – 1.63 (m, 5H), 1.57 – 1.30 (m, 4H), 1.20 (s, 3H), 1.13 – 0.95 (m, 2H), 0.90 (s, 3H).

¹³**C NMR** (126 MHz, CDCl₃) δ 199.74, 171.07, 87.21, 83.20, 81.20, 79.68, 74.22, 53.40, 49.87, 46.67, 38.85, 38.54, 36.17, 35.53, 32.42, 31.29, 23.06, 20.72, 17.44, 12.71, 1.37.

HRMS (ESI-TOF) Calcd for C₂₁H₂₃D₅O₂: [M+Na]⁺ 340.2295. Found: m/z 340.2294.

1-((3*S*,8*S*,9*S*,10*R*,13*S*,14*S*,17*S*)-3-hydroxy-10,13-dimethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*cyclopenta[*a*]phenanthren-17-yl-17-d)ethan-1-one-2,2,2-*d*₃ (8)



TLC: R*f* = 0.23 (silica gel, EA), (20.8mg, 65% yield), white solid.

¹**H NMR** (500 MHz, CDCl₃) δ 5.34 (dt, *J* = 5.0, 2.0 Hz, 1H), 3.52 (tt, *J* = 11.2, 4.6 Hz, 1H), 2.52 (t, *J* = 8.9 Hz, 0.03H), 2.34 – 2.13 (m, 3H), 2.08 (t, *J* = 2.2 Hz, 0.14H), 2.06 – 1.94 (m, 2H), 1.94 – 1.81 (m, 2H), 1.76 – 1.37 (m, 9H), 1.29 – 1.04 (m, 3H), 0.99 (d, *J* = 7.1 Hz, 4H), 0.62 (s, 3H).

¹³**C NMR** (126 MHz, CDCl₃) δ 209.92, 140.76, 121.44, 77.30, 77.05, 76.79, 71.72, 56.92, 50.55, 49.98, 49.48, 45.43, 43.98, 42.25, 38.79, 37.26, 36.53, 35.08, 32.02, 31.85, 31.78, 31.61, 24.52, 22.67, 21.09, 19.42, 13.26.

HRMS (ESI-TOF) Calcd for $C_{21}H_{28}D_4O_2$: $[M+K]^+$ 359.2285. Found: m/z 359.2285.

(8*S*,9*S*,10*R*,13*S*,14*S*)-17-(acetyl-*d*₃)-10,13-dimethyl-1,2,6,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-3*H*cyclopenta[*a*]phenanthren-3-one-2,2,4,6,6,17-*d*₆ (9)



TLC: Rf = 0.20 (silica ge, EA), (31mg, 96% yield), yellow solid

¹H NMR (400 MHz, CDCl₃) δ 2.51 (t, *J* = 9.0 Hz, 0.03H), 2.44 – 2.10 (m, 1H), 2.10 – 1.93 (m, 2H), 1.82 (dd, *J* = 12.8, 3.7 Hz, 1H), 1.77 – 1.37 (m, 7H), 1.33 – 1.18 (m, 2H), 1.16 (s, 3H), 1.11 – 0.90 (m, 2H), 0.64 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) ¹³C NMR (101 MHz,) δ 209.76, 199.78, 171.05, 56.10, 53.72, 53.13, 49.81, 45.48, 43.97, 38.67, 35.79, 35.58 (d, *J* = 4.5 Hz), 34.96, 31.78, 25.92, 24.28, 22.74, 21.08, 20.73, 17.43, 13.43.

HRMS (ESI-TOF) Calcd for C₂₁H₂₁D₉O₂: [M+H]⁺ 324.2884. Found: m/z 324.2881.

(8*S*,9*S*,10*R*,13*S*,14*S*)-10,13-dimethyl-6,7,8,9,10,12,13,14,15,16-decahydro-1*H*-cyclopenta[*a*]phenanthrene-3,11,17(2*H*)-trione-2,2,4,6,6,16,16-*d*₇ (10)



TLC: R*f* = 0.23 (silica gel, EA), (28.5mg, 94% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 5.71 (s, 0.02H), 2.71 (dd, *J* = 13.6, 1.1 Hz, 1H), 2.44 (d, *J* = 1.7 Hz, 1H), 2.30 (dd, *J* = 12.9, 0.9 Hz, 0.32H), 2.17 – 1.96 (m, 3H), 1.87 (ddd, *J* = 12.2, 10.9, 5.8 Hz, 1H), 1.74 – 1.55 (m, 2H), 1.40 (s, 3H), 1.27 (t, *J* = 12.4 Hz, 1H), 0.85 (d, *J* = 1.2 Hz, 3H).

¹³**C NMR** (101 MHz, CDCl₃) δ 217.10, 207.85, 199.76, 189.72, 167.92, 114.08, 100.56, 63.39, 50.50, 50.45, 49.86, 38.19, 36.25, 34.55, 30.78, 21.44, 17.34, 14.67, 8.59.

HRMS (ESI-TOF) Calcd for C₁₉H₁₇D₇O₃: [M+H]⁺ 308.2238. Found: m/z 308.2237.

(6*S*,8*R*,9*S*,10*R*,13*S*,14*S*,17*R*)-17-(acetyl-*d*₃)-6,10,13-trimethyl-3-oxo-2,3,6,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*cyclopenta[*a*]phenanthren-17-yl-2,2,4,6-*d*₄ acetate (11)



TLC: R*f* = 0.19 (silica gel, EA), (36.6mg, 93% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 5.77 (s, 0.02H), 2.90 (ddd, *J* = 17.2, 11.5, 5.9 Hz, 1H), 2.51 – 2.29 (m, 0.04H), 2.08 (s, 3H), 2.02 (s, 0.45H), 2.00 – 1.89 (m, 2H), 1.82 (dd, *J* = 12.8, 2.6 Hz, 1H), 1.79 – 1.61 (m, 6H), 1.58 – 1.49 (m, 1H), 1.49 – 1.18 (m, 2H), 1.16 (s, 3H), 1.04 (s, 3H), 1.02 – 0.95 (m, 1H), 0.91 – 0.78 (m, 1H), 0.64 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 204.35, 200.06, 174.06, 170.86, 96.77, 53.41, 52.78, 51.60, 51.01, 46.84, 40.84, 38.88, 37.71, 35.82, 35.49, 33.78, 33.41, 31.10, 30.38, 23.87, 21.35, 20.87, 18.34, 14.50.

HRMS (ESI-TOF) Calcd for C₂₄H₂₇D₇O₄: [M+H]⁺ 394.2969. Found: m/z 394.2968.

1-(p-tolyl)ethan-1-one-2,2,2-d₃ (12)



TLC: R*f* = 0.44 (silica gel, PE/EA, 10:1), (12.7mg, 93% yield), yellow liquid.

¹**H NMR** (400 MHz, CDCl₃) δ 7.87 (d, *J* = 8.0 Hz, 2H), 7.28 (d, *J* = 8.0 Hz, 3H), 2.78 – 2.51 (m, 0.33H), 2.43 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 197.96, 143.87, 134.73, 129.23, 128.42, 28.25, 21.63.

1-(4-butylphenyl)ethan-1-one-2,2,2- d_3 (13)



TLC: Rf = 0.30(silica gel, PE/EA, 10:1), (12.5mg, 70% yield), yellow liquid.

¹**H NMR** (500 MHz, CDCl₃) δ 7.88 (d, *J* = 8.3 Hz, 2H), 7.37 – 7.21 (m, 2H), 2.77 – 2.64 (m, 2H), 2.59 (s,0.01H), 1.74 – 1.55 (m, 2H), 1.36 (h, *J* = 7.4 Hz, 2H), 0.93 (t, *J* = 7.4 Hz, 3H).

¹³**C NMR** (126 MHz, CDCl₃) δ 198.12, 148.86, 134.89, 128.64, 128.48, 35.71, 33.29, 25.87, 22.35, 13.94.

HRMS (ESI-TOF) Calcd for C₁₂H₁₃D₃O: [M+H]⁺ 180.1462. Found: m/z 180.1463.

1-(4-methoxyphenyl-3,5-d₂)ethan-1-one-2,2,2-d₃ (14)



TLC: R*f* = 0.40 (silica gel, PE/EA, 2:1), (14.1mg, 91% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 7.98 – 7.90 (m, 2H), 6.98 – 6.90 (m, 2H), 3.87 (s, 3H), 2.53 (p, *J* = 2.2 Hz, 0.13H).

¹³**C NMR** (100 MHz, CDCl₃) δ 200.05, 164.24, 130.69, 130.46, 113.79, 55.57, 32.06.

1-(4-fluorophenyl)ethan-1-one-2,2,2-d₃ (15)



TLC: R*f* = 0.46 (silica gel, PE/EA, 2:1), (13.4mg, 95% yield), yellow liquid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.21 – 7.80 (m, 2H), 7.15 (t, *J* = 8.6 Hz, 2H), 2.58 (p, *J* = 2.2 Hz, 0.21H).

¹³**C NMR** (101 MHz, CDCl₃) δ 196.45, 165.77 (d, *J* = 254.6 Hz), 133.60 (d, *J* = 3.0 Hz), 130.92 (d, *J* = 9.4 Hz), 115.65 (d, *J* = 21.9 Hz), 22.35.

¹⁹**F NMR** (377 MHz, CDCl₃) δ -105.29, -105.31, -105.32, -105.33, -105.34, -105.35, -105.37, -105.39.

1-(4-chlorophenyl)ethan-1-one-2,2,2-d₃ (16)



TLC: Rf = 0.45 (silica gel, PE/EA, 2:1), (15.2mg, 97% yield), yellow solid.

¹**H NMR** (400 MHz, CDCl₃) δ 7.94 – 7.88 (m, 2H), 7.48 – 7.42 (m, 2H), 2.57 (dq, *J* = 4.5, 2.1 Hz, 0.21H).

¹³C NMR (101 MHz, CDCl₃) δ 198.52, 139.56, 136.40, 129.70, 128.21, 28.25.

1-(4-bromophenyl)ethan-1-one-2,2,2- d_3 (17)



TLC: R*f* = 0.41 (silica gel, PE/EA, 2:1), (19mg, 94% yield), yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 7.83 (dd, *J* = 8.4, 1.3 Hz, 2H), 7.65 – 7.59 (m, 2H), 2.62 – 2.54 (m, 0.24H).

¹³C NMR (101 MHz, CDCl₃) δ 197.56, 137.07, 131.89, 129.32, 127.48, 26.48.

1-(4-(trifluoromethyl)phenyl)ethan-1-one-2,2,2-d₃ (18)



TLC: R*f* = 0.39 (silica gel, PE/EA, 2:1), (17.4mg, 91% yield), yellow solid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.04 (dp, *J* = 7.6, 0.8 Hz, 2H), 7.75 – 7.68 (m, 2H), 2.64 – 2.57 (m, 0.27H).

¹³C NMR (100 MHz, CDCl₃) δ 198.21, 139.74, 134.50 (q, *J* = 32.7 Hz), 128.69, 125.75 (q, *J* = 3.9 Hz), 123.67 (q, *J* = 271.5 Hz), 27.39.
¹⁹F NMR (376 MHz, CDCl₃) δ -61.80.

1-(4-nitrophenyl)ethan-1-one-2,2,2- d_3 (19)



TLC: Rf = 0.44 (silica gel, PE/EA, 2:1), (15.6mg, 93% yield), yellow solid.

¹H NMR (400 MHz, CDCl₃) δ 8.36 – 8.29 (m, 2H), 8.16 – 8.09 (m, 2H), 2.71 – 2.64 (m, 0.45H).

¹³C NMR (101 MHz, CDCl₃) δ 196.40, 150.36, 143.41, 130.43, 124.52, 27.59.

4-(acetyl-d₃)benzonitrile (20)



TLC: R*f* = 0.39 (silica gel, PE/EA, 2:1), 13mg, 89% yield), yellow solid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.06 (d, *J* = 8.1 Hz, 1H), 7.88 – 7.68 (m, 1H), 2.63 (dp, *J* = 6.7, 2.2 Hz, 0.39H).

¹³C NMR (101 MHz, CDCl₃) δ 196.63, 139.91, 132.51, 128.68, 117.91, 116.41, 26.48.

methyl 4-(acetyl-d₃)benzoate (21)



TLC: R*f* = 0.33 (silica gel, PE/EA, 2:1), (16.7mg, 92% yield), white wolid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.17 – 8.11 (m, 2H), 8.02 (d, *J* = 8.3 Hz, 2H), 3.97 (d, *J* = 0.9 Hz, 3H), 2.63 (t, *J* = 2.4 Hz, 0.12H).

¹³**C NMR** (101 MHz, CDCl₃) δ 196.87, 166.65, 140.23, 135.00, 129.82, 128.18, 53.34, 27.15.

1-(4-(methylsulfonyl)phenyl)ethan-1-one-2,2,2-d₃ (22)



TLC: R*f* = 0.34 (silica gel, PE/EA, 2:1), (18.9mg, 94% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.15 – 8.07 (m, 2H), 8.07 – 7.99 (m, 2H), 3.06 (d, *J* = 0.5 Hz, 3H), 2.66 – 2.58 (m, 0.34H).

¹³C NMR (100 MHz, CDCl₃) δ 198.21, 144.27, 140.99, 129.22, 127.89, 44.39, 26.50.

1-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)ethan-1-one-2,2,2-*d*₃ (23)



TLC: R*f* = 0.31 (silica gel, PE/EA, 10:1), 18.6mg, 87% yield), white solid.

¹**H NMR** (400 MHz, CDCl₃) δ 7.93 (q, *J* = 8.0 Hz, 4H), 2.61 (dp, *J* = 4.5, 3.2, 2.2 Hz, 0.06H), 1.38 (s, 12H).

¹³**C NMR** (101 MHz, CDCl₃) δ 198.83, 138.99, 134.91, 129.26, 127.79, 127.26, 84.21, 37.33, 24.88.

¹¹**B NMR** (128 MHz, CDCl₃) δ 31.18.

1-phenylbutan-1-one-2,2-d2 (24)



TLC: R*f* = 0.40 (silica gel, PE/EA, 10:1), (14mg, 95% yield), yellow liquid.

¹**H NMR** (400 MHz, CDCl₃) δ 8.01 – 7.95 (m, 2H), 7.61 – 7.53 (m, 1H), 7.52 – 7.44 (m, 2H), 2.94 (t, *J* = 7.4, 2.6 Hz, 0.06H), 1.78 (q, *J* = 7.4 Hz, 2H), 1.03 (td, *J* = 7.4, 0.9 Hz, 3H).

¹³**C NMR** (101 MHz, CDCl₃) δ 200.56, 137.10, 132.86, 128.54, 128.04, 77.35, 77.04, 76.72, 39.82, 17.71, 13.85.

4-phenylbutan-2-one-1,1,1,3,3-*d*₅ (25)



TLC: R*f* = 0.35 (silica gel, PE/EA, 10:1), (14mg, 89% yield), yellow liquid. ¹H NMR (400 MHz, CDCl₃) δ 7.31 – 7.20 (m, 2H), 7.16 (td, *J* = 7.4, 1.2 Hz, 3H), 2.85 (s, 2H), 2.75 – 2.66 (m, 0.2 H), 2.11 – 2.04 (m, 0.21H). ¹³C NMR (101 MHz, CDCl₃) δ 207.99, 141.03, 128.56, 128.34, 126.17, 44.87, 37.49, 29.66.

1-(4-methoxyphenyl)-2-phenylethan-1-one-2,2-d₂ (26)



TLC: R*f* = 0.25(silica gel, PE/EA, 10:1), (21mg, 92% yield), yellow liquid.

¹H NMR (500 MHz, CDCl₃) δ 8.10 – 7.84 (m, 2H), 7.36 – 7.30 (m, 2H), 7.29 – 7.21 (m, 3H), 6.96 – 6.90 (m, 2H), 4.21 (t, *J* = 2.2 Hz, 0.06H), 3.85 (s, 3H).
¹³C NMR (126 MHz, CDCl₃) δ 196.39, 163.54, 134.92, 131.00, 129.61, 129.37, 128.68, 126.82, 113.82, 55.51, 44.83.

HRMS (ESI-TOF) Calcd for C₁₅H₁₂D₂O₂: [M+H]⁺ 229.1192. Found: m/z 229.1192.

2,3-dihydro-1*H*-inden-1-one-2,2-*d*₂ (27)



TLC: R*f* = 0.39 (silica gel, PE/EA, 10:1), (7.5mg, 55% yield), yellow solid.

¹**H NMR** (500 MHz, CDCl₃) δ 7.77 (d, *J* = 7.6 Hz, 1H), 7.59 (td, *J* = 7.5, 1.2 Hz, 1H), 7.49 (dt, *J* = 7.8, 1.0 Hz, 1H), 7.38 (t, *J* = 7.4 Hz, 1H), 3.15 (s, 2H), 2.75 – 2.65 (m, 0.05H).

¹³**C NMR** (126 MHz, CDCl₃) δ 207.33, 155.29, 137.13, 134.66, 127.32, 126.75, 123.76, 89.66, 25.66.

HRMS (ESI-TOF) Calcd for C₉H₆D₂O: [M+K]⁺ 173.0333. Found: m/z 173.0323.

3,4-dihydronaphthalen-1(2H)-one-2,2-d₂ (28)



TLC: R*f* = 0.30 (silica gel, PE/EA, 10:1), (10.3mg, 70% yield), yellow liquid.

¹**H NMR** (500 MHz, CDCl₃) δ 8.04 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.47 (td, *J* = 7.5, 1.5 Hz, 1H), 7.34 – 7.28 (m, 1H), 7.26 (d, *J* = 7.8 Hz, 1H), 2.97 (t, *J* = 6.1 Hz, 2H), 2.64 (ddt, *J* = 9.0, 5.7, 2.5 Hz, 0.04H), 2.13 (t, *J* = 6.1 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 198.63, 144.56, 133.45, 132.62, 128.81, 127.17, 126.66, 38.51, 29.67, 23.12.

HRMS (ESI-TOF) Calcd for C₁₁H₁₀O₂: [M+H]⁺ 149.0930. Found: m/z 149.0930. 6,7,8,9-tetrahydro-5*H*-benzo[7]annulen-5-one-6,6-*d*₂ (29)



TLC: R*f* = 0.29 (silica gel, PE/EA, 10:1), (15mg, 95% yield), yellow liquid.

¹**H NMR** (500 MHz, CDCl₃) δ 7.73 (dd, *J* = 7.7, 1.5 Hz, 1H), 7.42 (td, *J* = 7.5, 1.5 Hz, 1H), 7.30 (td, *J* = 7.5, 1.2 Hz, 1H), 7.20 (d, *J* = 7.5 Hz, 1H), 2.97 – 2.90 (m, 2H), 2.72 (ddt, *J* = 9.8, 5.0, 2.1 Hz, 0.04H), 1.89 (pd, *J* = 6.5, 1.0 Hz, 2H), 1.80 (t, *J* = 6.8 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 206.31, 141.45, 138.82, 132.21, 129.74, 128.58, 126.65, 40.23, 32.57, 25.22, 20.82.

HRMS (ESI-TOF) Calcd for C₁₁H₁₀D₂O: [M+H]⁺ 163.1087. Found: m/z 163.1093.

(Z)-3-hydroxy-1-phenylbut-2-en-1-one-4,4,4- d_3 (30)



TLC: R*f* = 0.30 (silica gel, PE/EA, 10:1), (10mg, 60% yield), white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.95 – 7.84 (m, 2H), 7.59 – 7.51 (m, 1H), 7.49 – 7.42 (m, 2H), 6.19 (s, 1H), 2.21 (s, 0.02H).

¹³**C NMR** (126 MHz, CDCl₃) δ 193.71, 183.55, 134.91, 132.34, 128.65, 127.04, 96.75, 25.47.

HRMS (ESI-TOF) Calcd for C₁₀H₇D₃O₂: [M+H]⁺ 166.0942. Found: m/z 166.0941.

4-phenylbut-3-yn-2-one-1,1,1-d₃ (31)



TLC: R*f* = 0.35 (silica gel, PE/EA, 10:1), (10mg, 66% yield), yellow liquid.

¹**H NMR** (500 MHz, CDCl₃) δ 7.60 – 7.55 (m, 2H), 7.55 – 7.43 (m, 1H), 7.39 (td, *J* = 7.5, 1.6 Hz, 2H), 2.44 (s, 0.06H).

¹³**C NMR** (126 MHz, CDCl₃)δ 184.89, 133.10, 130.79, 128.67, 119.90, 90.41, 88.30, 32.05.

HRMS (ESI-TOF) Calcd for C₁₀H₅D₃O: [M+H]⁺ 148.0836. Found: m/z 146.0830.

7. NMR Spectra of Products



S21



Figure S10. ¹H-NMR spectrum (400 MHz, CDCl₃) of S3







Figure S14. ¹H-NMR spectrum (400 MHz, CDCl₃) of 4



Figure S16. ¹H-NMR spectrum (500 MHz, CDCl₃) of S5



S26







Figure S22. ¹H-NMR spectrum (500 MHz, CDCl₃) of S7



Figure S24. ¹³C-NMR spectrum (126 MHz, CDCl₃) of 7







Figure S26. ¹H-NMR spectrum (500 MHz, CDCl₃) of 8



Figure S28. ¹H-NMR spectrum (400 MHz, CDCl₃) of S9



Figure S30. ¹³C-NMR spectrum (101 MHz, CDCl₃) of 9



Figure S32. ¹H-NMR spectrum (400 MHz, CDCl₃) of 10



Figure S34. ¹H-NMR spectrum (400 MHz, CDCl₃) of S11



Figure S36. ¹³C-NMR spectrum (101 MHz, CDCl₃) of 11









Figure S40. ¹H-NMR spectrum (500 MHz, CDCl₃) of S13



Figure S42. ¹³C-NMR spectrum (126 MHz, CDCl₃) of 13



Figure S44. ¹H-NMR spectrum (400 MHz, CDCl₃) of 14





S41





S43







Figure S58. ¹³C-NMR spectrum (101 MHz, CDCl₃) of **18**





S48



S49





S51



S52



Figure S72. ¹H-NMR spectrum (400 MHz, CDCl₃) of S23



Figure S74. ¹³C-NMR spectrum (101 MHz, CDCl₃) of 23





S56





Figure S82. ¹H-NMR spectrum (500 MHz, CDCl₃) of S26



S59



Figure S86. ¹H-NMR spectrum (500 MHz, CDCI₃) of 27













S65



S66

