Supplementary Information (SI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2025

## Nile Red Derivatives for Dual-Channel Organelle Imaging and LPS Detection Mohini Ghorpade<sup>a</sup>, Sneha Negi<sup>a</sup>, Paramasivam Mahalingam<sup>b</sup> and Sriram Kanvah<sup>a</sup>\*

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## Synthetic procedure and characterization details: N1, N2 and N3

A solution of compound-2 [(9-(diethylamino)-2-hydroxy-5H-benzo[a]phenoxazin-5-one] (0.2 mmol) in anhydrous DMF was cooled to 0 °C. To this, potassium carbonate was added and the mixture was stirred for 10 minutes. Subsequently, 4-nitrobenzyl bromide (5 equiv,1 mmol, N1), diethylene glycol 2-bromoethyl methyl ether (1 mmol, N2), and 1-bromododecane (1 mmol, N3) were added dropwise at ambient temperature under a nitrogen atmosphere. The reaction mixture was then heated to 70 °C and stirred for three hours. The reaction progress was monitored using Thin Layer Chromatography (TLC). Upon completion, the mixture was poured onto crushed ice and extracted with dichloromethane (DCM). The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The crude product was purified by column chromatography using DCM/hexane (15:85) as the eluent, affording the desired compound. The structural characterization details from 1H, 13C, and HRMS are provided in the supporting information.

N1:9-(diethylamino)-2-((4-nitrobenzyl)oxy)-5H-benzo[a]phenoxazin-5-one

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>) δ 8.28 (d, J = 8.7 Hz, 2H), 8.02 (dd, J = 16.4, 5.6 Hz, 1H), 7.79 (d, J = 8.6 Hz, 2H), 7.57 (d, J = 9.1 Hz, 1H), 7.32 (dd, J = 8.7, 2.5 Hz, 1H), 6.78 (dd, J = 9.1, 2.5 Hz, 1H), 6.59 (s, 1H), 6.16 (s, 1H), 5.76 (s, 1H), 5.44 (s, 2H), 3.47 (dd, J = 14.1, 7.0 Hz, 4H), 1.16 (td, J = 7.1, 3.8 Hz, 6H). <sup>13</sup>C NMR (125

**MHz, DMSO**) δ 181.7, 160.9, 152.2 151.3, 147.5, 146.9, 144.9, 138.4, 134.0, 131.3, 128.7, 127.7, 125.7, 124.4, 124.0, 118.4, 118.4, 110.5, 107.3, 104.5, 96.4, 68.8, 55.3, 44.9, 12.9. **Mass** HRMS: Actual mass (M+H)= 470.1716, Observed mass (M+H)= 470.1729, mass error= 2.76 ppm

N2: 9-(diethylamino)-2-(2-(2-methoxyethoxy)ethoxy)-5H-benzo[a]phenoxazin-5-one

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  8.02 (d, J = 8.7 Hz, 1H), 7.93 (d, J = 2.6 Hz, 1H), 7.59 (d, J = 9.1 Hz, 1H), 7.26 (dd, J = 8.7, 2.6 Hz, 1H), 6.80 (dd, J = 9.2, 2.7 Hz, 1H), 6.62 (d, J = 2.7

Hz, 1H), 6.17 (s, 1H), 4.28 (t, J = 7.3, 2H), 3.82 (t, J = 4.5, 2H), 3.63 (dd, J = 6.0, 3.5 Hz, 2H), 3.57 – 3.55 (m, 2H), 3.53 (dt, J = 5.9, 3.2 Hz, 3H), 3.51 – 3.47 (m, 3H), 3.45 – 3.41 (m, 2H), 3.23 (s, 3H), 1.16 (t, J = 7.0 Hz, 7H). <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>)  $\delta$  181.8, 161.5, 152.1, 151.2, 146.9, 138.6, 134.0, 131.4, 127.7, 125.4, 124.4, 118.4, 110.5, 106.7, 104.5, 96.4, 71.7, 70.4, 70.2, 70.0, 69.3, 68.1, 58.5, 44.9, 12.9. Mass HRMS: Actual mass (M+H)= 481.2338, Observed mass (M+H)= 481.2347, mass error= 1.87 ppm.

error=-0.39 ppm.

N3: 9-(diethylamino)-2-(dodecyloxy)-5H-benzo[a]phenoxazin-5-one

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.22 (d, J = 8.7 Hz, 1H), 8.06

(d, J = 2.5 Hz, 1H), 7.62 (d, J = 9.0 Hz, 1H), 7.17 (dd, J = 12.5 Hz, 1H), 6.66 (dd, J = 9.1, 2.7 Hz, 1H), 6.47 (d, J = 1.5 Hz, 1H), 6.31 (s, 1H), 4.17 (t, J = 6.5 Hz, 3H), 3.48 (q, J = 7.1 Hz, 4H), 1.86 (t, J = 7.8, 2H), 1.51 (t, J = 7.7 Hz, 3H), 1.27 (t, J = 7.1 Hz, 18H), 0.88 (t, J = 7.0 Hz, 6H). <sup>13</sup>C NMR (125 MHz, DMSO)  $\delta$  183.3, 161.9, 151.8, 150.9, 146.7, 131.0, 127.7, 125.5, 118.3, 109.4, 106.6, 105.3, 105.3, 96.3, 45.0, 45.0, 31.9, 29.6, 29.6, 29.6, 29.6, 29.5, 29.4, 29.3, 29.2, 26.0, 22.7.Mass HRMS: Actual mass (M+H)= 503.3273 Observed mass (M+H)= 503.3271, mass

Fig S1. Absorption spectra of conmpounds N2 & N3

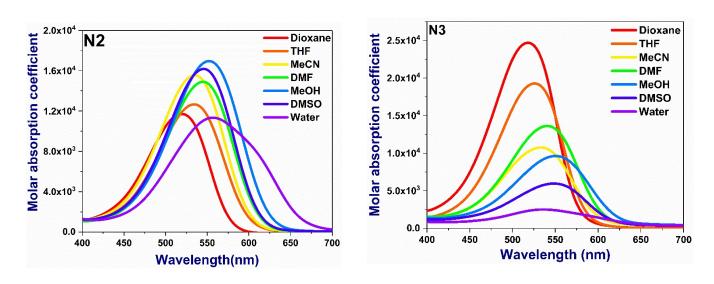


Fig S2: Emission spectrum of compounds N2 & N3 in different solvents

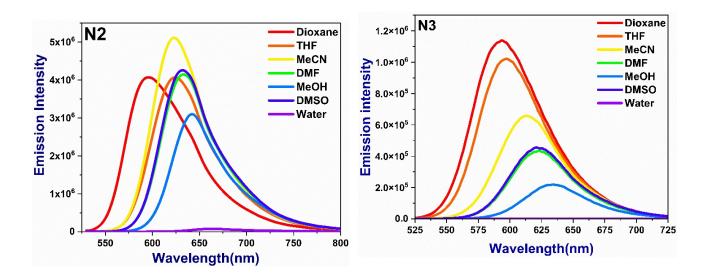


Fig S3: The effect of added LPS on the emission behavior of N2 and N3

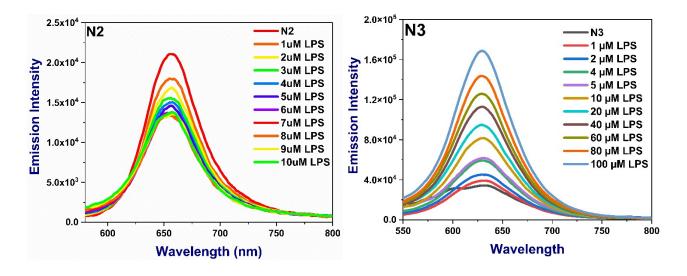


Fig S4: MTT assay of N1, N2 and N3 in COS-7 cells

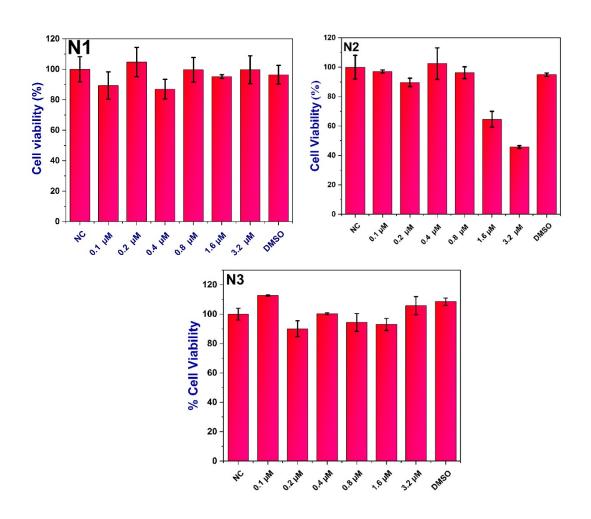


Fig S5: Multiple cellular imaging of N1 and N2 in COS-7 cells. Imaging of N1 and N2 in COS-7 cells (image represents multiple cells). Concentration: (N1= 0.5  $\mu$ M, N2= 0.5  $\mu$ M), \*Live cell imaging condition: 37 °C, 5% CO<sub>2</sub>; Scale bar=10  $\mu$ m.

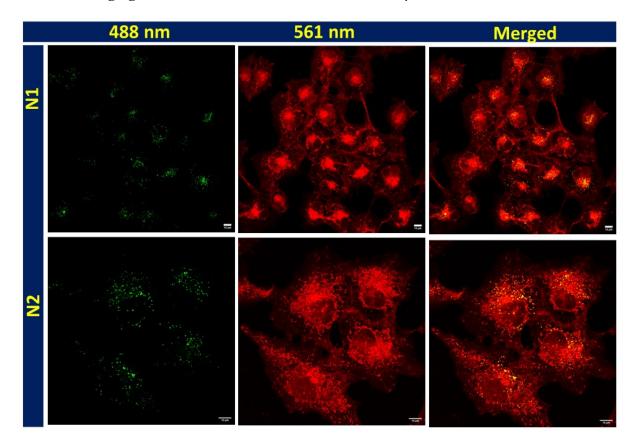


Fig S6: Cellular imaging of **N3** at 488 nm and 561 nm. **N3** unlike others does not show any imaging at 488 nm while we can see lysosomes at 561nm.

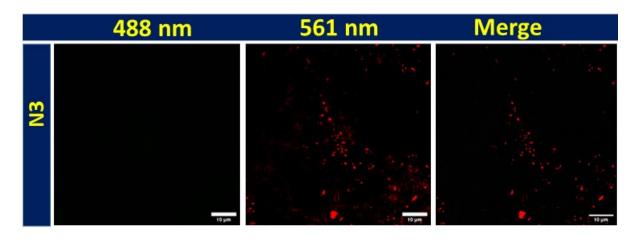
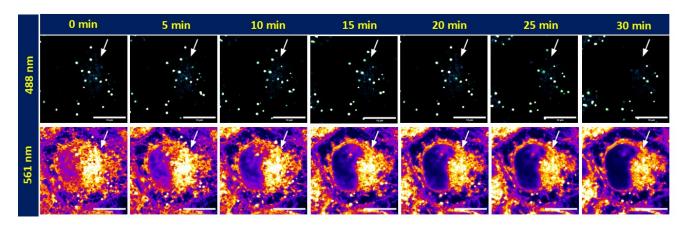


Fig S7: Photostability studies of N2 and N3 in COS-7 cells



Video S1: Video showing continuous photoirradiation studies of N1, COS-7 cells (Video attached as additional information)

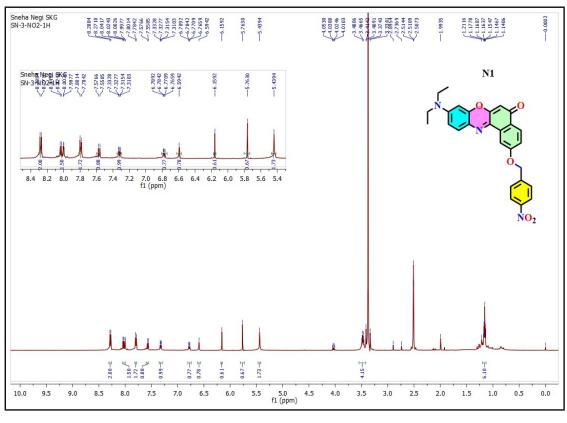
Video S2: Photoirradiation studies of N1 in HeLa cells (Video attached as additional information

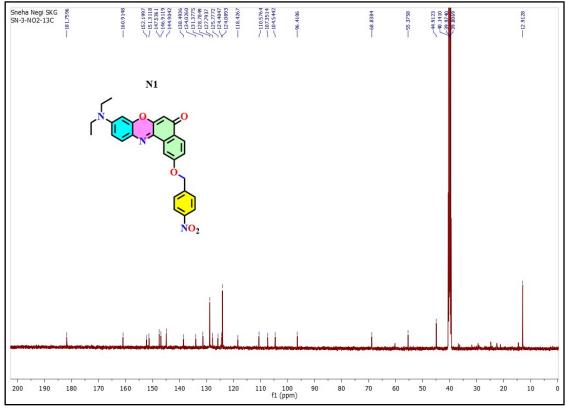
Video S3: Cellular imaging of N2 with continuous photoirradiation for 30 min in COS 7 cells

Video S4 Cellular imaging of N3 with continuous photoirradiation for 30 min in COS 7 cells

## **Spectral characterization**

Fig S8: <sup>1</sup>H and <sup>13</sup>C NMR & Mass of N1





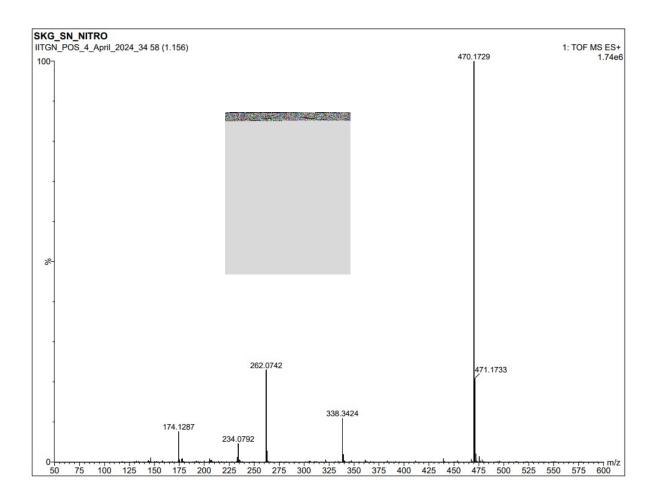
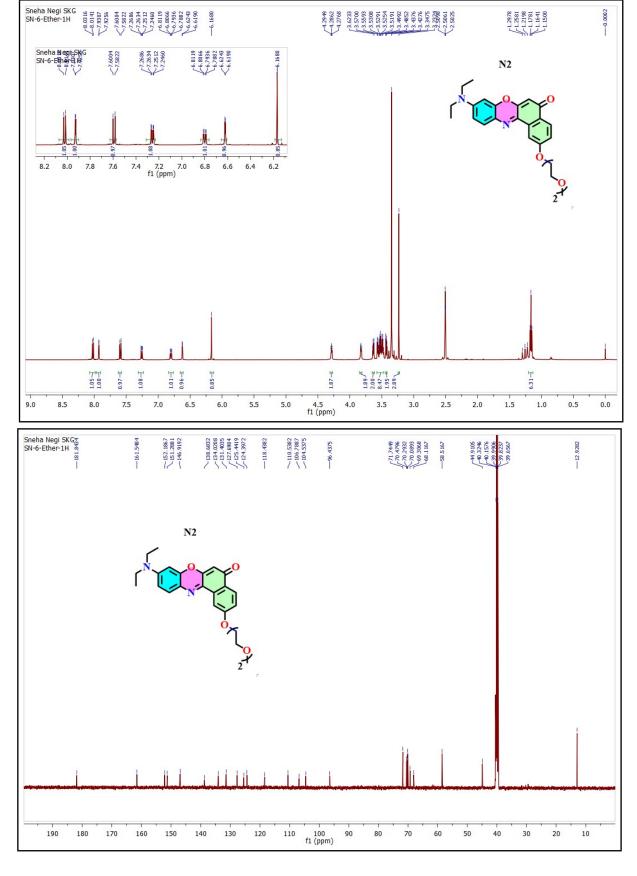


Fig S9: <sup>1</sup>H and <sup>13</sup>C NMR & Mass of N2:



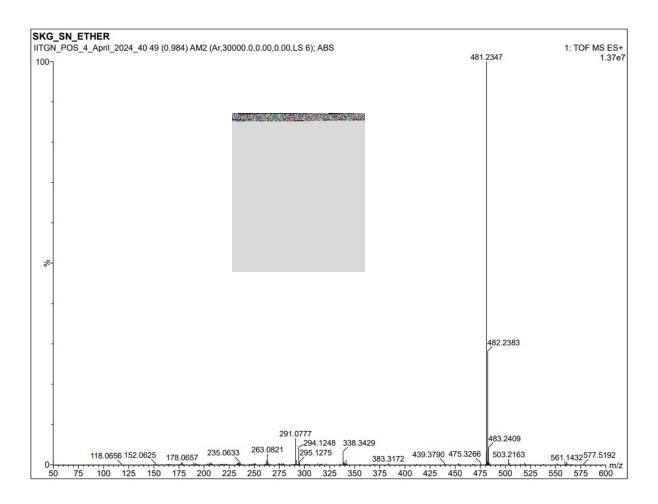


Fig S10: <sup>1</sup>H and <sup>13</sup>C NMR and Mass data of N3:

