

Direct α -halogenation of carboxylic acids with *N*-bromosuccinimide (NBS) or trichloroisocyanuric acid (TCCA) in the presence of catalytic amounts of 4-trifluoromethylbenzoic anhydride and H₂SO₄

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Supplementary Information

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1. General information

Melting points were determined on a Yamato melting point apparatus MP-21 and are uncorrected. IR spectra were recorded using a JEOL FT/IR-4600ST spectrophotometer. NMR spectra were measured on a Varian NMR system 700 spectrometer (700 MHz) using tetramethylsilane as an internal standard. Unless otherwise stated, anhydrous sodium sulfate was employed as the drying agent. Column chromatography was performed on Kanto chemical Silica gel 60 (spherical, 63-210 μm). Anhydride **8** was purchased from TCI. DCE- d_4 , CD_3OD and D_2O was purchased from Merck. Detection of Cl_2 was carried out by the ION SELECTIVE PACK TEST kit, WAK-CIO/DP (Kyoritsu Chemical-Check Lab. Corp.), which DPD (diethyl-*p*-phenylenediamine) was employed as a detection agent, or the test paper, Nissan Aqua Check LC, which syringaldazine (3,5-dimethoxy-4-hydroxybenzaldazine) was applied (purchased from AS ONE Co.). IR and ^1H -NMR spectra of compounds **2**, **9-11**, **15**, **16** and **27** were completely in accordance with those of the commercially available samples purchased from TCI. IR and ^1H -NMR spectra of compounds **12** and **14** were completely in accordance with those of the commercially available samples purchased from Fujifilm-Wako. IR and ^1H -NMR spectra of compounds **13**, **17** and **23** were completely in accordance with those of the commercially available samples purchased from Nakalai Tesque. Compounds **19**,¹ **20**,² **21**,³ **22**,⁴ **24**,⁵ **25**,⁶ and **26**¹ were completely identified by the comparison of spectroscopic behaviours with those of the authentic samples reported in the literature.

2. Experimental procedures

General procedure for the synthesis of α -brominated carboxylic acids

NBS (249 mg, 1.4 mmol, 1.4 equiv.) was added to the mixture of sulfuric acid (9.8 mg, 0.1 mmol), hexanoic acid **1** (116 mg, 1.0 mmol) and anhydride **8** (36 mg, 0.1 mmol) in DCE (0.5 mL). After stirring the mixture at 80 °C for 3 h, the mixture was filtered and the filtrate was evaporated under reduced pressure. In order to remove most of 4-trifluoromethylbenzoic acid (**B**) which was difficult to separate from **2**, DCE (25 μ L) was added and the mixture was filtered. After the evaporation of the filtrate under reduced pressure, the residue was purified by column chromatography on silica gel using heptane-EtOAc (8:2) to give **2** (170 mg, 87%).

General procedure for the synthesis of α -chlorinated carboxylic acids

TCCA (186 mg, 0.8 mmol, 2.4 equiv.) was added to the mixture of sulfuric acid (9.8 mg, 0.1 mmol), isovaleric acid (102 mg, 1.0 mmol) and anhydride **8** (36 mg, 0.1 mmol) in DCE (0.1 mL). After stirring the mixture at 80 °C for 12 h, the mixture was filtered and the filtrate was evaporated under reduced pressure. In order to remove most of 4-trifluoromethylbenzoic acid (**B**), the mixture was filtered and the filtrate was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel using heptane-EtOAc (8:2) to give **24** (127 mg, 93%).

General procedure for the synthesis of α -iodinated carboxylic acids using I₂

I₂ (356 mg, 1.4 mmol, 1.4 equiv.) was added to the mixture of sulfuric acid (9.8 mg, 0.1 mmol), 3-phenylpropanoic acid (**18**) (150 mg, 1.0 mmol) and anhydride **8** (36 mg, 0.1 mmol) in DCE (0.5

mL). The mixture was stirred at 80 °C for 4 h and then filtered. After the evaporation of the filtrate under reduced pressure, 2-iodo-3-phenylpropanoic acid (**28**) was not detected by the ¹H-NMR spectroscopy.

General procedure for the synthesis of α -iodinated carboxylic acids using NIS

NIS (315 mg, 1.4 mmol, 1.4 equiv.) was added to the mixture of sulfuric acid (9.8 mg, 0.1 mmol), 3-phenylpropanoic acid (**18**) (150 mg, 1.0 mmol) and anhydride **8** (36 mg, 0.1 mmol) in DCE (0.5 mL). The mixture was stirred at 80 °C for 4 h and then filtered. After the evaporation of the filtrate under reduced pressure, 2-iodo-3-phenylpropanoic acid (**28**) was not detected by the ¹H-NMR spectroscopy.

Preparative-scale procedure for the synthesis of α -brominated carboxylic acids

NBS (4.98 g, 28 mmol, 1.4 equiv.) was added to the mixture of sulfuric acid (196 mg, 2 mmol), hexanoic acid **1** (2.32 g, 20 mmol) and anhydride **8** (724 mg, 2 mmol) in DCE (10 mL). After stirring the mixture at 80 °C for 4.5 h, the mixture was filtered and the filtrate was evaporated under reduced pressure. In order to remove most of 4-trifluoromethylbenzoic acid (**B**), DCE (0.5 mL) was added and the mixture was filtered. After the evaporation of the filtrate under reduced pressure, the residue was purified by column chromatography on silica gel using heptane-EtOAc (8:2) to give **2** (3.52 g, 90%).

Preparative-scale procedure for the synthesis of α -chlorinated carboxylic acids

TCCA (3.72 g, 16 mmol, 2.4 equiv.) was added to the mixture of sulfuric acid (196 mg, 2 mmol),

isovaleric acid (2.04 g, 20 mmol) and anhydride **8** (724 mg, 2 mmol) in DCE (2 mL). After stirring the mixture at 80 °C for 18 h, the mixture was filtered and the filtrate was evaporated under reduced pressure. In order to remove most of 4-trifluoromethylbenzoic acid (**B**), the mixture was filtered and the filtrate was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel using heptane-EtOAc (8:2) to give **24** (2.56 g, 94%).

Comparative bromination of hexanoic acid (1) by NBS vs. molecular bromine¹

Hexanoic acid **1** (3.25 g, 28 mmol) and sulfuric acid (274 mg, 2.8 mmol) was dissolved in DCE-d₄ (4 mL). The mixture was divided into two equal fractions which were placed in two separate flasks. The latter was cooled in an ice bath. Anhydride **8** (507 mg, 1.4 mmol) and the solution of bromine (3.14 g, 19.6 mmol) in DCE-d₄ (5 mL) were added to one of them. To the other flask, **8** (507 mg, 1.4 mmol), DCE-d₄ (5 mL) and NBS (3.49 g, 19.6 mmol) were added. Both flasks were equipped with condensers fitted with drying tubes and heated simultaneously in the same oil bath at 80 °C. At known intervals of time, aliquots of the reaction mixture were removed from the flasks, which were immediately cooled in an ice bath to quench the reactions. After the filtration, the solution was transferred to NMR tubes. NMR spectra were then measured, and the integration of the triplet of the bromide **2** at δ 4.25 was compared to that of the triplet of **1** at δ 2.35. The resulting ratio increased over time and eventually plateaued, showing completion of the reactions. The results are summarized in Table S2.

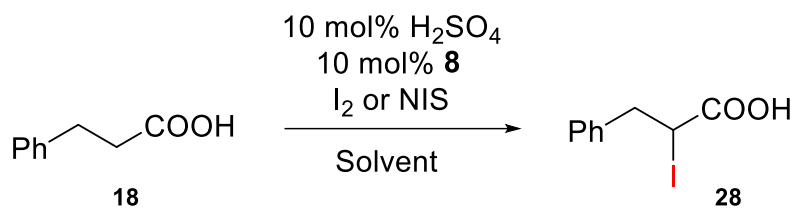
Deuteration of carboxylic acid 1 in the presence of CD₃OD

Anhydride **8** (217 mg, 0.6 mmol) was added to the mixture of sulfuric acid (59 mg, 0.6 mmol), CD₃OD (1.0 mL, 24.8 mmol), DCE (3.0 mL) and hexanoic acid **1** (696 mg, 6.0 mmol). After stirring

the mixture at 80 °C for 6 h, the solution was extracted with ether (70 mL). The extracts were washed with water (20 mL×3), dried and evaporated. In order to remove most of 4-trifluoromethylbenzoic acid (**B**), DCE (0.15 mL) was added and the mixture was filtered. After the evaporation of the filtrate, the residue was purified by column chromatography on silica gel using heptane-EtOAc (20:1) to give CD₃ ester of **1**. Two independent runs afforded 710 mg (89%) and 742 mg (93%), respectively. Deuterium incorporation at the α -position was determined by integration of the ¹H NMR spectra and was 12% in both runs.

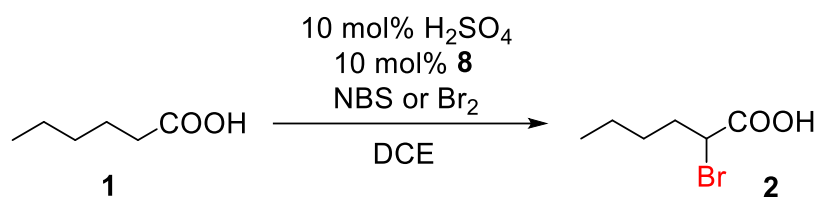
3. Tables

Table S1 α -Iodination of carboxylic acid **18**^a



Entry	Reagent	Solvent	Temp (°C)	Yield (%)
1	I ₂	DCE (0.5 mL)	80	0
2 ^b	I ₂	—	80	0
3	I ₂	DCE (0.1 mL)	80	0
4	I ₂	PhCl (0.5 mL)	130	0
5	I ₂	PhCl (0.1 mL)	130	0
6	NIS	DCE (0.5 mL)	80	0
7	NIS	—	80	0
8	NIS	DCE (0.1 mL)	80	0
9	NIS	PhCl (0.5 mL)	130	0
10 ^b	NIS	—	130	0
11	NIS	PhCl (0.1 mL)	130	0

^a Conditions: carboxylic acid **18** (1.0 mmol), H₂SO₄ (10 mol%), anhydride **8** (10 mol%), I₂ or NIS (1.4 mmol), solvent, 4 h. ^b A large amount of I₂ sublimed.

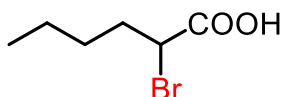
Table S2 Comparative bromination of **1** using NBS and Br₂.^a

Time (h)	Conv. (%) ^b	
	NBS	Br ₂
0	0	0
0.25	35	2
0.5	51	5
0.75	63	6
1.0	75	11
1.5	90	14
2.0	95	20
2.5	100	24
3.0	100	29

^a Conditions: carboxylic acid **1** (14 mmol), H₂SO₄ (10 mol%), anhydride **8** (10 mol%), NBS or Br₂ (19.6 mmol), DCE (7 mL), 80 °C. ^b Determined by ¹H-NMR spectroscopy.

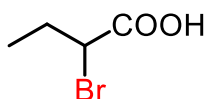
4. Analytical data of compounds 2, 9-11, 15-17 and 21-26

2-Bromohexanoic acid (**2**) (commercially available)



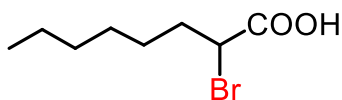
Colourless oil; IR (Neat) 3106, 2959, 2932, 2872, 2667, 1716, 1460, 1424, 1382, 1286, 1245, 1174, 1111, 1065, 929, 733 and 661 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ = 0.95 (3H, t, J = 7.3 Hz), 1.38-1.53 (4H, m), 2.02-2.16 (2H, m) and 4.27 (1H, t, J = 7.1 Hz); $^{13}\text{C-NMR}$ (CDCl_3) δ = 13.8, 22.0, 29.3, 34.4, 45.3 and 175.1.

2-Bromobutyric acid (**9**) (commercially available)



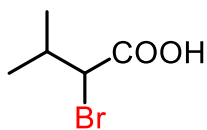
Colourless oil; IR (Neat) 3586, 3438, 3106, 2976, 2940, 2880, 2686, 2596, 1717, 1459, 1421, 1279, 1224, 1172, 1079, 927, 888, 813, 788 and 657 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ = 1.09 (3H, t, J = 7.4 Hz), 2.03-2.19 (2H, m) and 4.22 (1H, dd, J = 7.6 and 6.7 Hz); $^{13}\text{C-NMR}$ (CDCl_3) δ = 11.8, 28.1, 47.0 and 175.5.

2-Bromooctanoic acid (**10**) (commercially available)



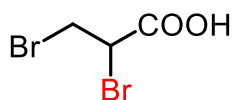
Colourless oil; IR (Neat) 3414, 3102, 2959, 2928, 2858, 2671, 2596, 1717, 1458, 1422, 1379, 1282, 1240, 1186, 1116, 925, 725 and 678 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) δ = 0.91 (3H, t, J = 7.1 Hz), 1.30-1.55 (8H, m), 1.99-2.16 (2H, m) and 4.27 (1H, dd, J = 7.7 and 7.0 Hz); $^{13}\text{C-NMR}$ (CDCl_3) δ = 14.0, 22.5, 27.1, 28.5, 31.5, 34.7, 45.3 and 175.3.

2-Bromo-3-methylbutanoic acid (**11**) (commercially available)



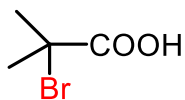
Colourless oil; IR (Neat) 3087, 2979, 2940, 2876, 2686, 2556, 1717, 1469, 1427, 1389, 1369, 1350, 1293, 1226, 1179, 1117, 930, 853, 778, 688, 638 and 489 cm^{-1} ; ^1H -NMR (CDCl_3) δ = 1.08 (3H, d, J = 6.7 Hz), 1.12 (3H, d, J = 6.7 Hz), 2.23-2.28 (1H, m) and 4.09 (1H, d, J = 7.6 Hz); ^{13}C -NMR (CDCl_3) δ = 19.7, 20.1, 32.1, 54.0 and 174.6.

2,3-Dibromopropanoic acid (**15**) (commercially available)



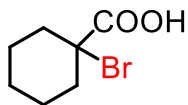
Colourless powder; mp 61-64 $^{\circ}\text{C}$ (EtOAc-heptane); IR (KBr) 3414, 3110, 3045, 2955, 2908, 2755, 2660, 2580, 2496, 1715, 1432, 1336, 1255, 1217, 1154, 951, 909, 828, 790, 661, 633, 553 and 511 cm^{-1} ; ^1H -NMR (CDCl_3) δ = 3.72 (1H, dd, J = 10.1 and 4.3 Hz), 3.94 (1H, dd, J = 11.3 and 10.2 Hz) and 4.51 (1H, dd, J = 11.3 and 4.3 Hz); ^{13}C -NMR (CDCl_3) δ = 29.1, 40.3 and 172.2.

2-Bromo-2-methylpropanoic acid (**16**) (commercially available)



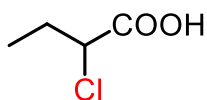
Colourless oil; IR (Neat) 3074, 3011, 2983, 2904, 2825, 2667, 2565, 2473, 1712, 1466, 1449, 1436, 1413, 1375, 1298, 1182, 1111, 1011, 937, 813, 634, 550 and 472 cm^{-1} ; ^1H -NMR (CDCl_3) δ = 1.98 (6H, s); ^{13}C -NMR (CDCl_3) δ = 30.5, 54.9 and 177.3.

1-Bromocyclohexane-1-carboxylic acid (**17**) (commercially available)



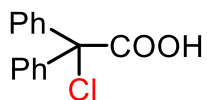
Colourless powder; mp 62-63 °C (EtOAc-heptane); IR (KBr) 3316, 3074, 2952, 2866, 2659, 2540, 1698, 1435, 1411, 1347, 1292, 1266, 1244, 1207, 1142, 1117, 1071, 1059, 937, 907, 874, 800, 741, 689, 529, 510 and 474 cm⁻¹; ¹H-NMR (CDCl₃) δ = 1.45-1.61 (4H, m), 1.76-1.81 (2H, m), and 2.20 (4H, t, *J* = 6.4 Hz); ¹³C-NMR (CDCl₃) δ = 23.7, 24.7, 37.6, 62.3, and 176.4.

2-Chlorobutanoic acid (**21**)³



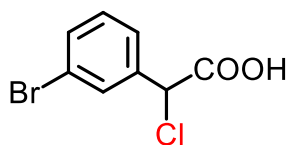
Colourless oil; IR (Neat) 3437, 3117, 2979, 2940, 2884, 2686, 2596, 1729, 1460, 1437, 1420, 1384, 1288, 1204, 1108, 1084, 1035, 930, 881, 827, 795 and 688 cm⁻¹; ¹H-NMR (CDCl₃) δ = 1.10 (3H, t, *J* = 7.4 Hz), 1.99-2.15 (2H, m) and 4.31 (1H, dd, *J* = 7.7 and 5.7 Hz); ¹³C-NMR (CDCl₃) δ = 10.4, 28.3, 58.5 and 175.2.

2-Chloro-2,2-diphenylacetic acid (**22**)⁴



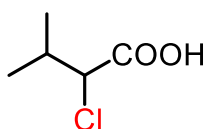
Colourless powder; mp 144-146 °C (toluene-heptane); IR (KBr) 3399, 3066, 3031, 2912, 2631, 2478, 1718, 1496, 1448, 1342, 1248, 1200, 1176, 1091, 1054, 1033, 1006, 982, 925, 773, 762, 741, 720, 699, 662, 627, 593 and 526 cm⁻¹; ¹H-NMR (CDCl₃) δ = 7.35-7.39 (6H, m) and 7.49 (4H, dd, *J* = 8.0 and 2.0 Hz); ¹³C-NMR (CDCl₃) δ = 81.2, 127.4, 128.3, 128.4, 141.2 and 177.7.

2-(3-Bromophenyl)-2-chloroacetic acid (**23**) (commercially available)



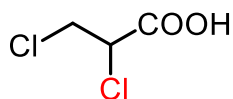
Colourless powder; mp 54-57 °C (toluene-heptane); IR (KBr) 3114, 3057, 2960, 2912, 2676, 2588, 1751, 1726, 1574, 1475, 1432, 1405, 1312, 1303, 1272, 1230, 1201, 1181, 1075, 998, 943, 895, 881, 811, 779, 741, 690, 677, 637, 560, 445 and 429 cm⁻¹; ¹H-NMR (CDCl₃) δ = 5.34 (1H, s), 7.29 (1H, dd, *J* = 7.8 and 7.8 Hz), 7.46 (1H, d, *J* = 7.8 Hz), 7.54 (1H, dt, *J* = 7.8 and 1.7 Hz) and 7.69 (1H, t, *J* = 1.7 Hz); ¹³C-NMR (CDCl₃) δ = 57.9, 122.8, 126.6, 130.4, 131.1, 132.7, 137.2 and 172.1.

2-Chloro-3-methylbutanoic acid (**24**)⁵



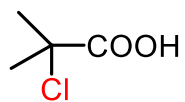
Colourless oil; IR (Neat) 3367, 2972, 2703, 2588, 2500, 1731, 1461, 1424, 1390, 1371, 1336, 1301, 1220, 1119, 1083, 975, 914, 844, 795, 712, 620, 523 and 456 cm⁻¹; ¹H-NMR (CDCl₃) δ = 1.10 (6H, dd, *J* = 10.9 and 6.8 Hz), 2.36-2.41 (1H, m) and 4.22 (1H, d, *J* = 6.0 Hz); ¹³C-NMR (CDCl₃) δ = 17.8, 19.6, 32.5, 64.0 and 175.2.

2,3-Dichloropropanoic acid (**25**)⁶



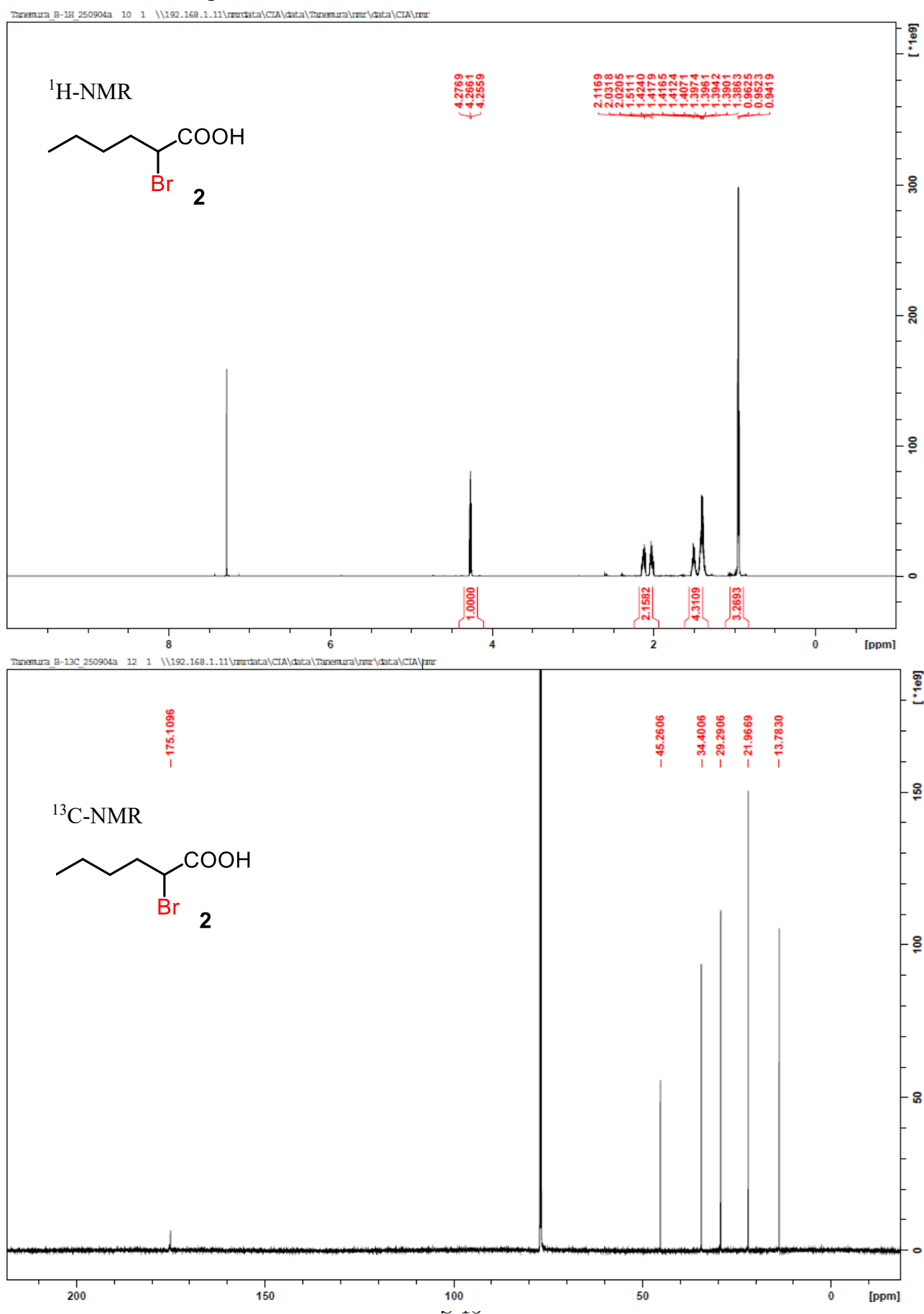
Colourless powder; mp 51-52 °C (EtOAc-heptane); IR (KBr) 3363, 3031, 2943, 2707, 2607, 1731, 1433, 1404, 1302, 1275, 1232, 1205, 1024, 978, 905, 857, 824, 714, 679, 610, 465 and 447 cm⁻¹; ¹H-NMR (CDCl₃) δ = 3.85 (1H, dd, *J* = 11.3 and 5.1 Hz), 3.97 (1H, dd, *J* = 11.3 and 8.4 Hz), 4.50 (1H, dd, *J* = 8.3 and 5.2 Hz) and 7.66 (1H, br s); ¹³C-NMR (CDCl₃) δ = 43.6, 54.9 and 172.1.

2-Chloro-2-methylpropanoic acid (**26**)¹

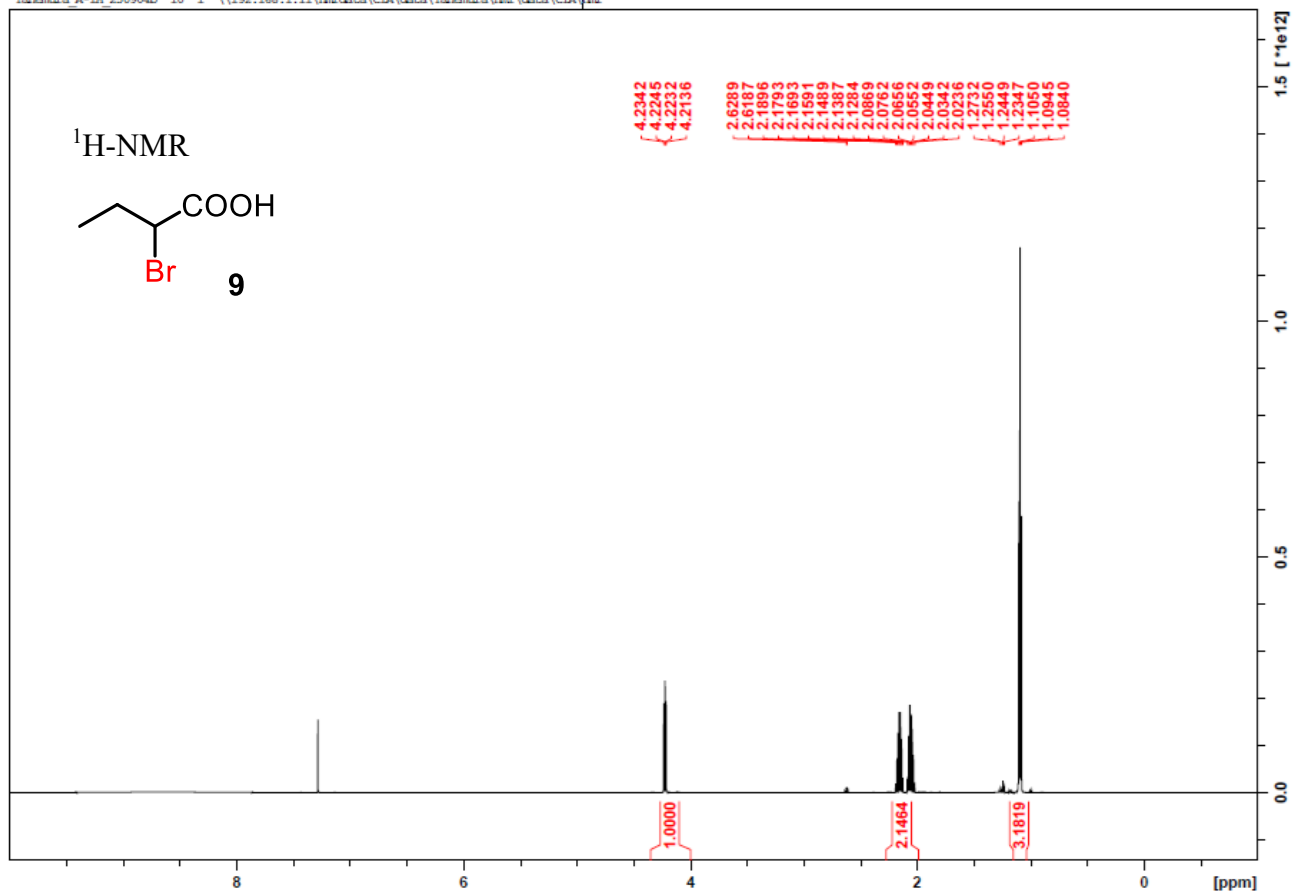


Colourless oil; IR (Neat) 3442, 3098, 2988, 2940, 2667, 2560, 1719, 1465, 1414, 1371, 1288, 1182, 1122, 931, 815, 661 and 544 cm^{-1} ; ^1H -NMR (CDCl_3) δ = 1.82 (6H, s); ^{13}C -NMR (CDCl_3) δ = 29.6, 64.2 and 177.0.

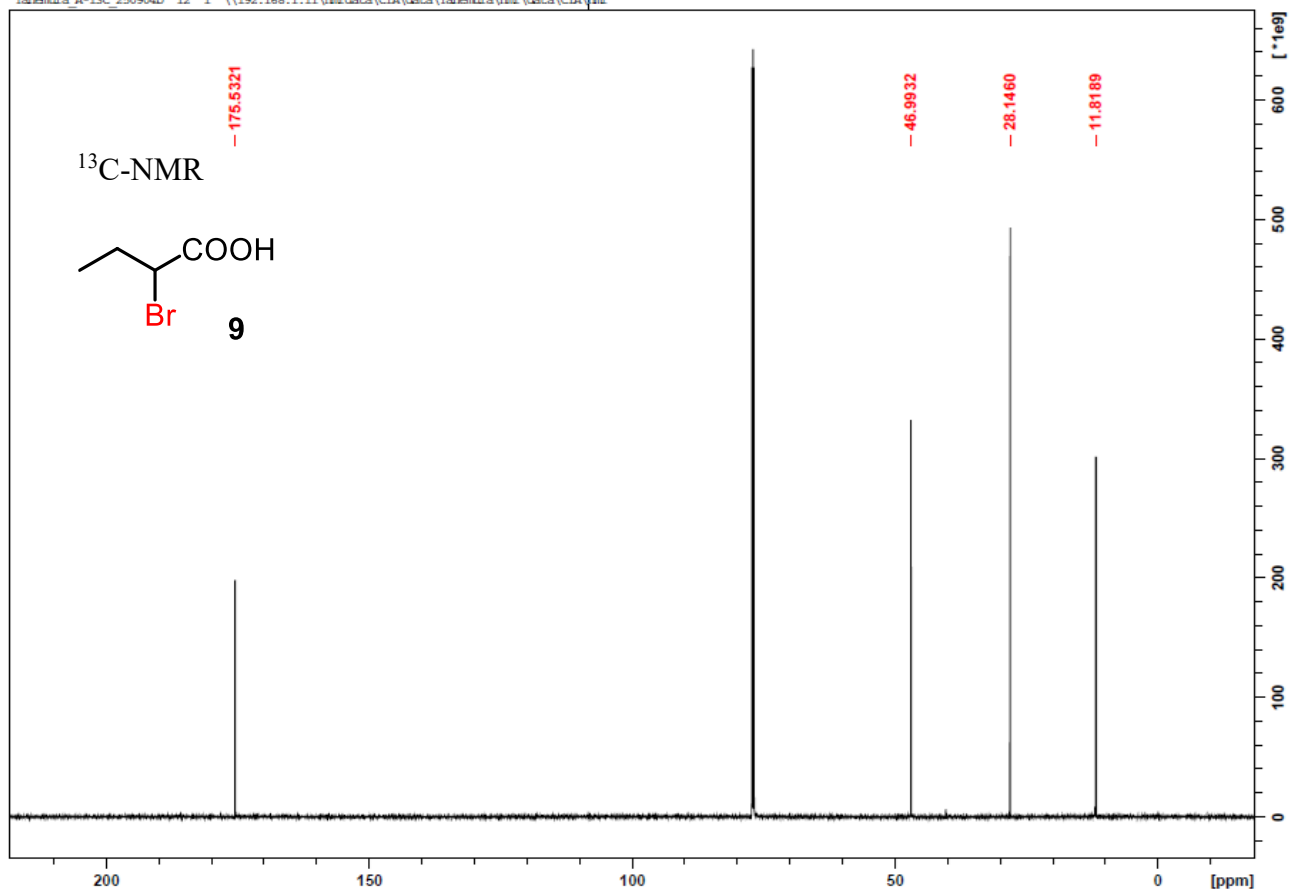
5. ^1H - and ^{13}C -NMR spectra of 2, 9-11, 15-17 and 21-26



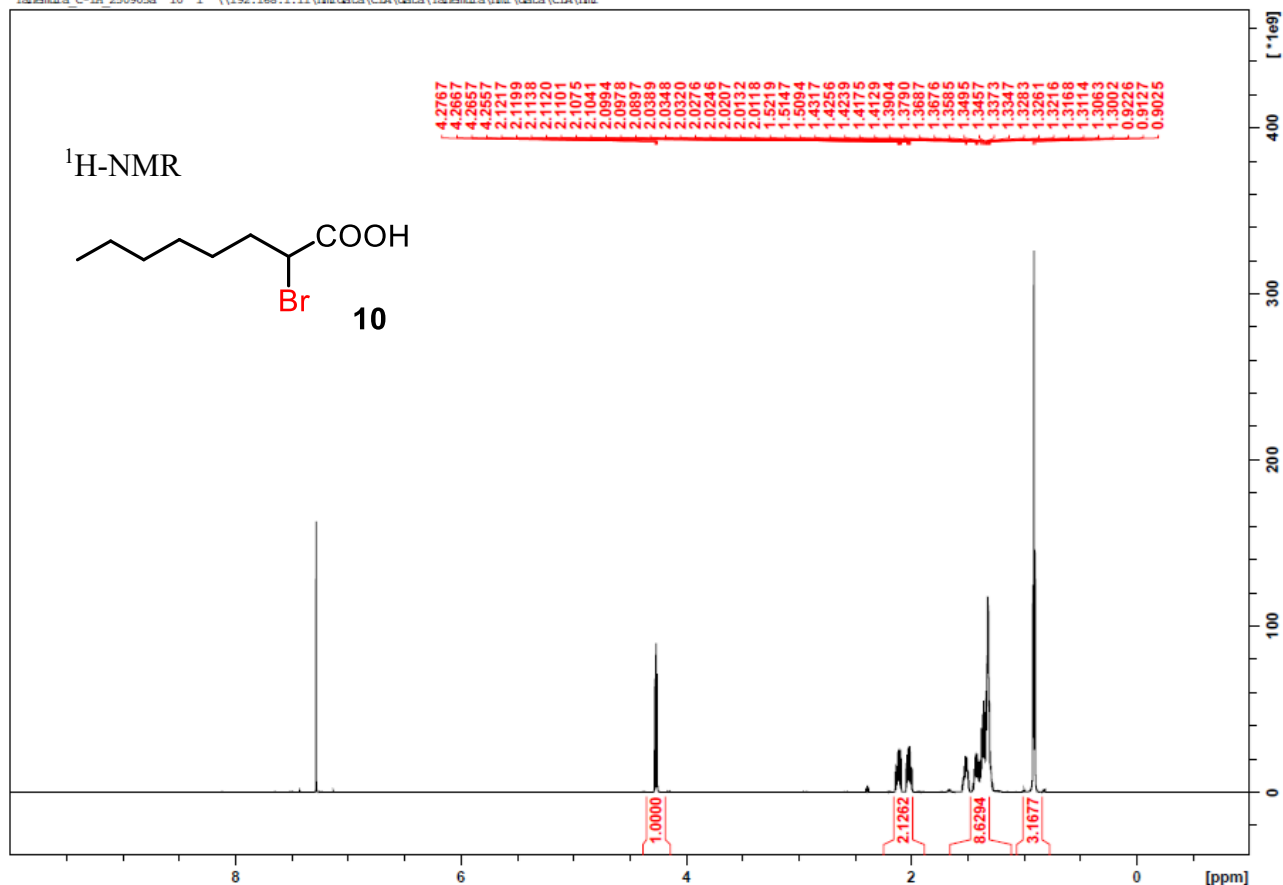
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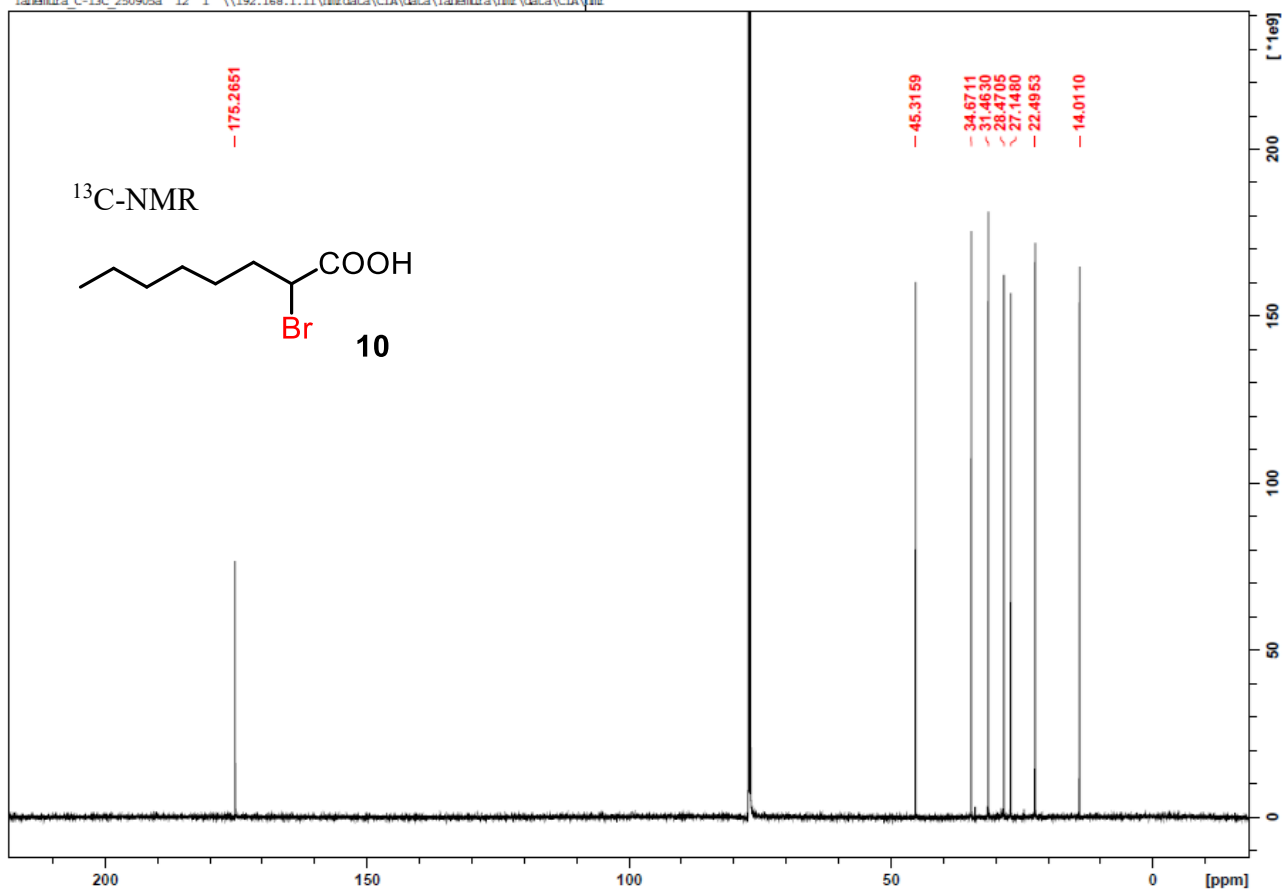
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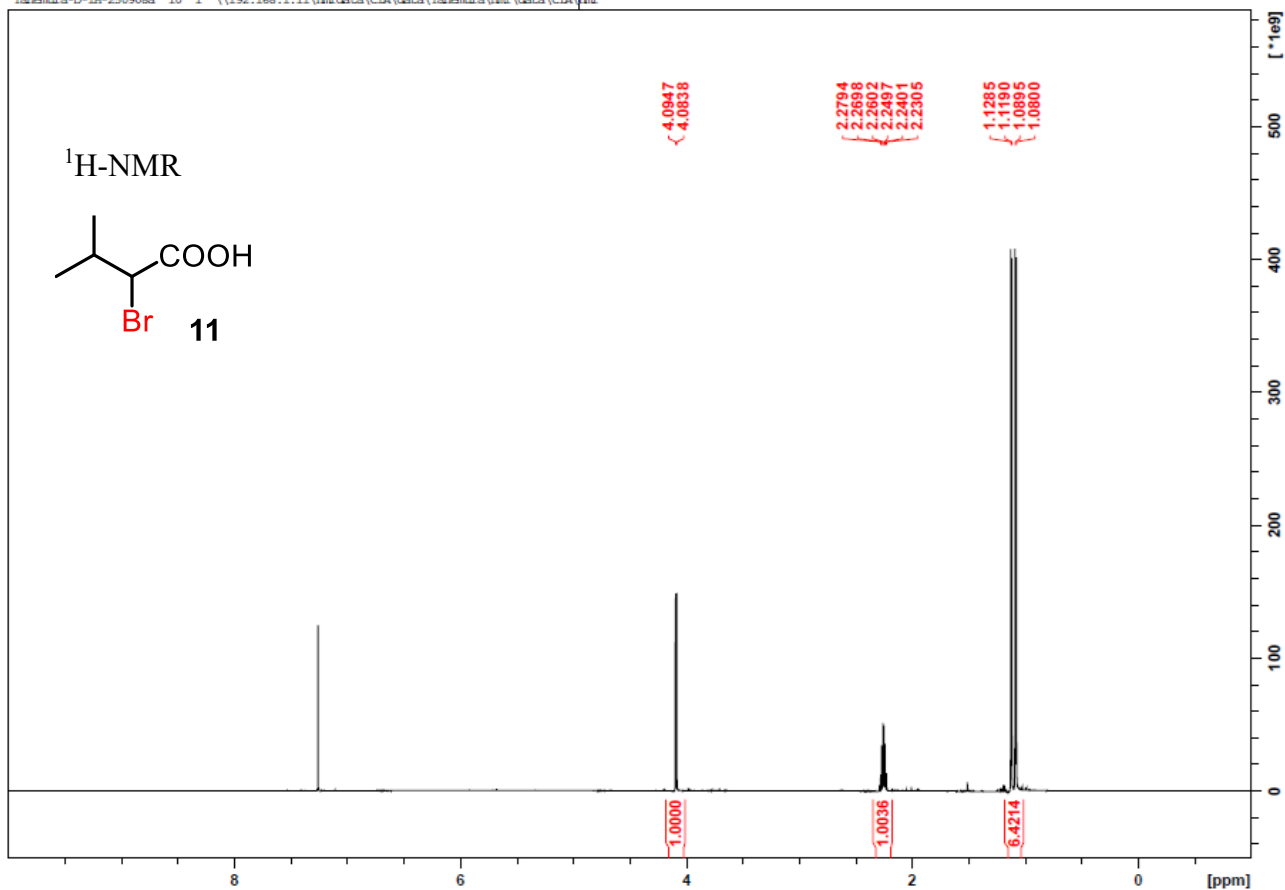
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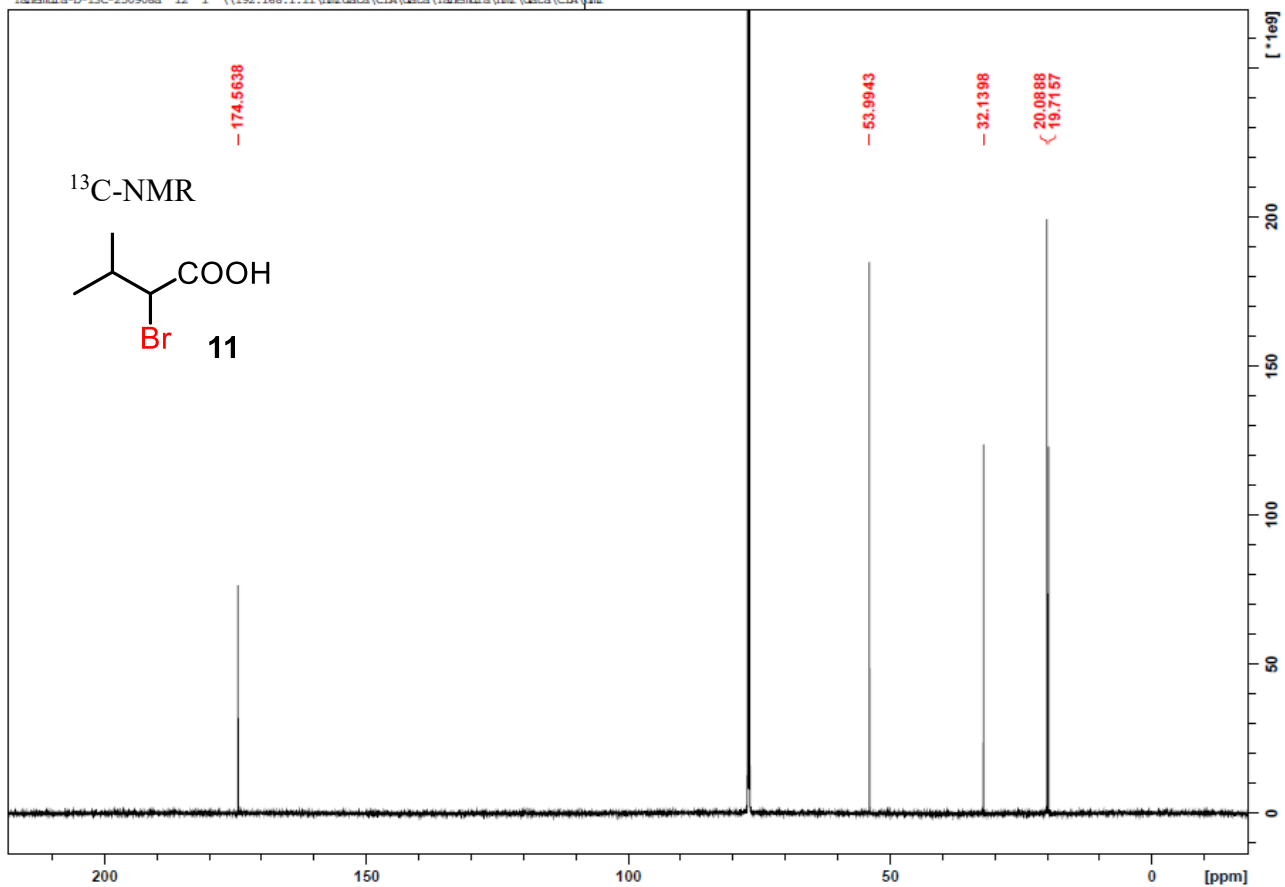
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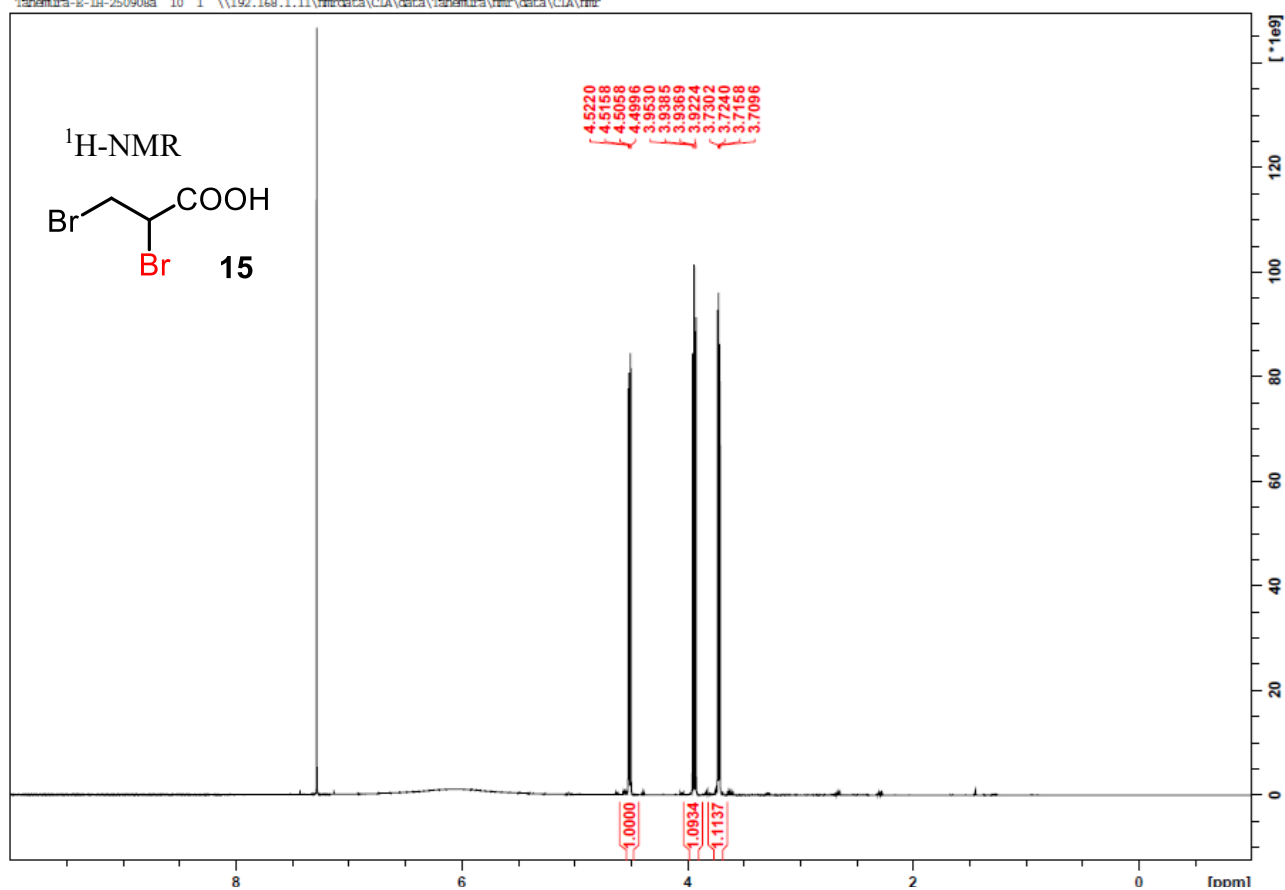
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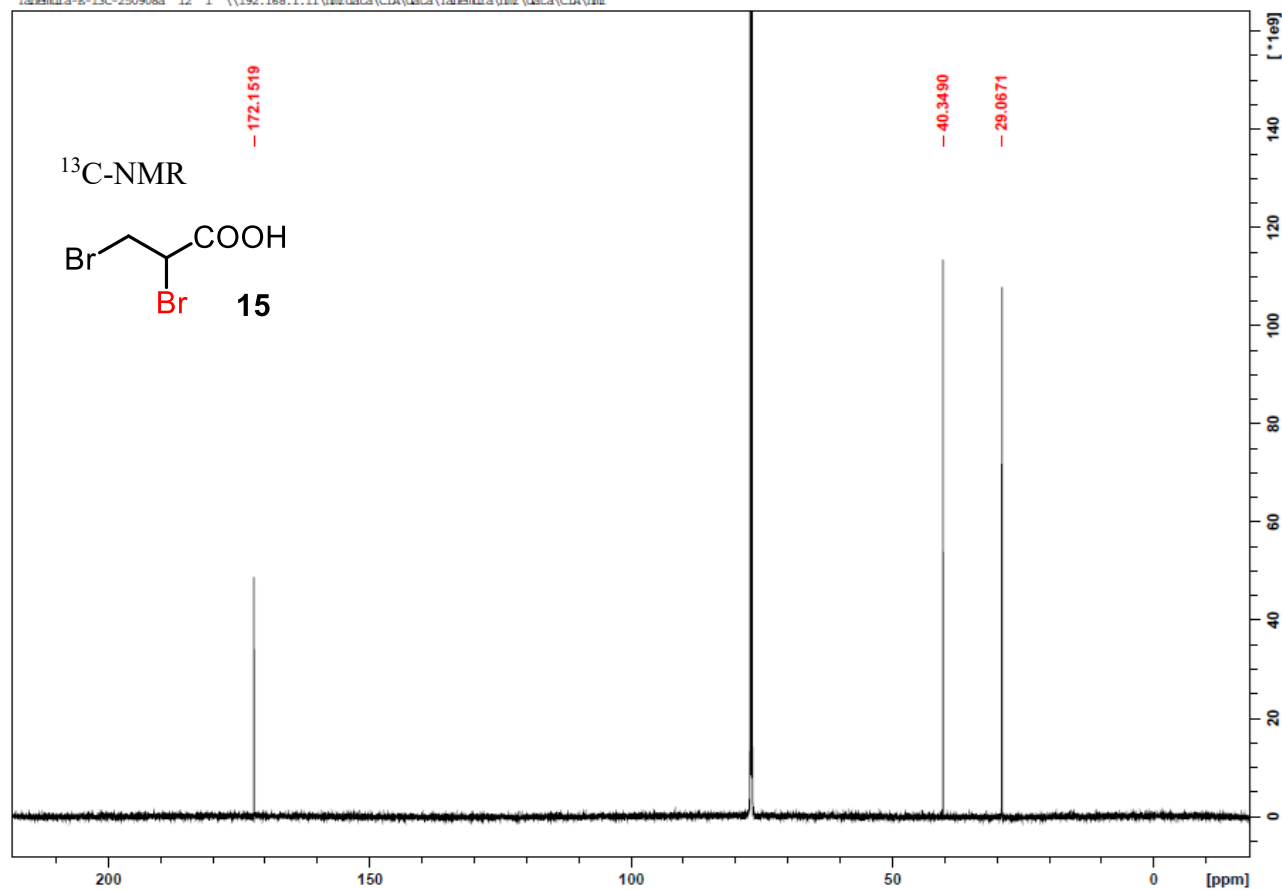
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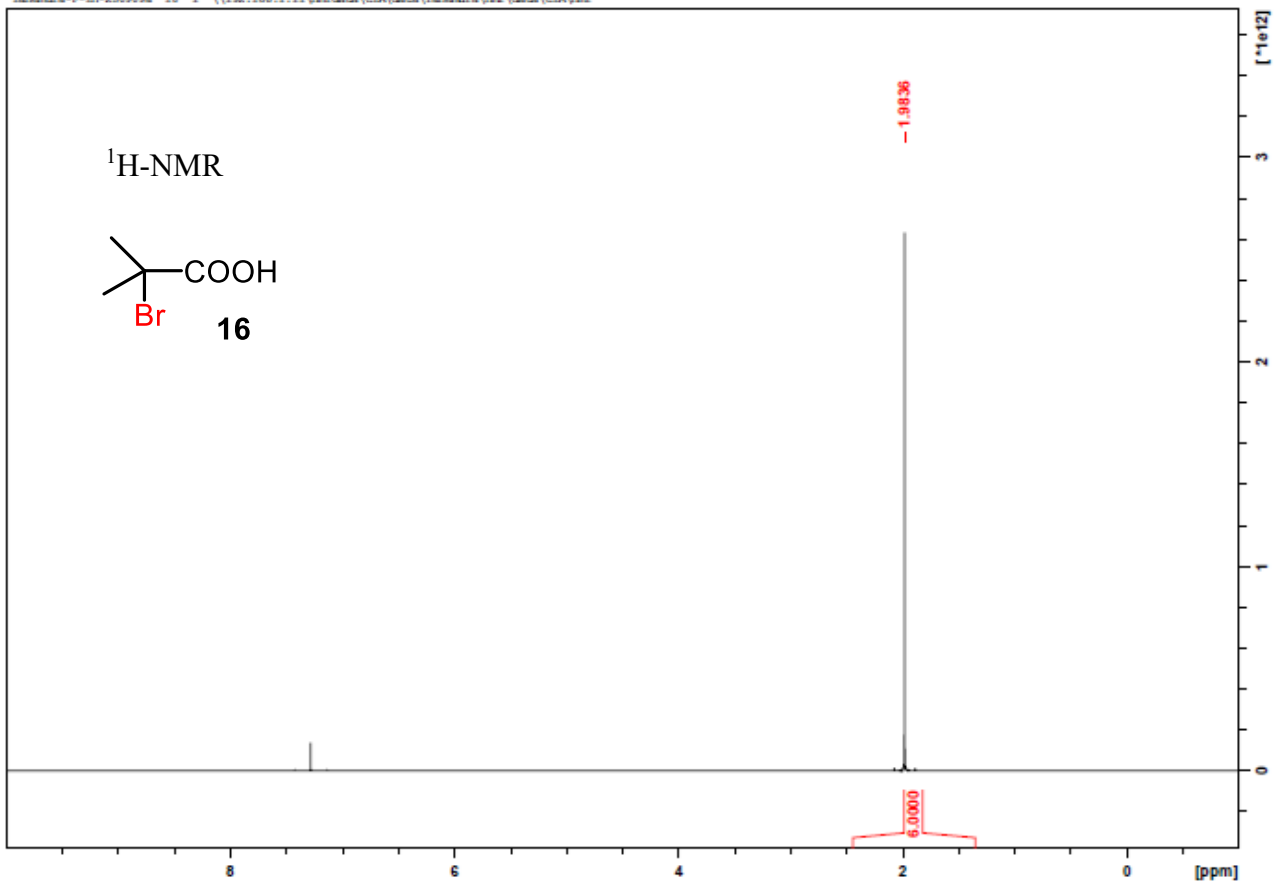
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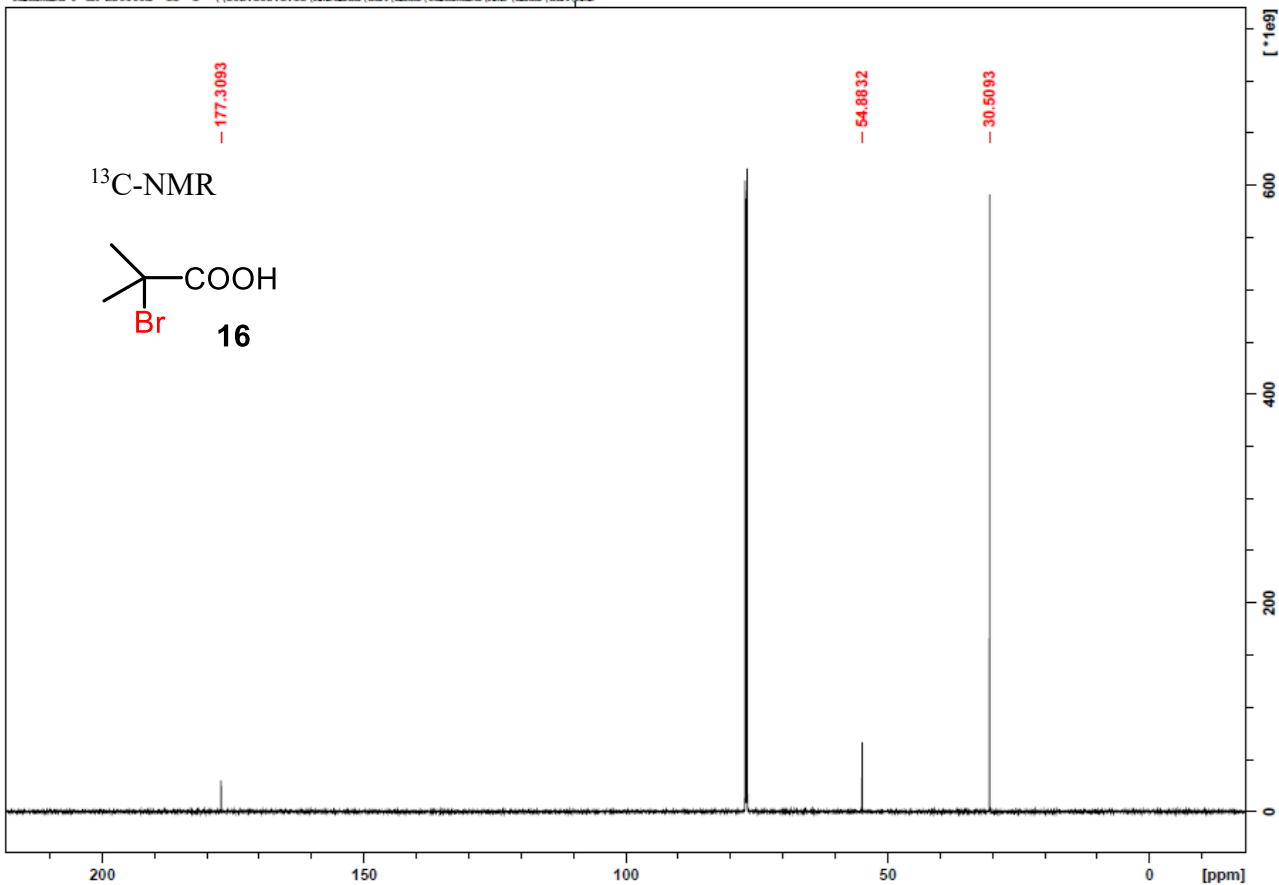
Tanemura-E-13C-250908a 12 1 \\192.168.1.11\rmrdata\CIA\data\tanemura\rmr\data\CIA\rmr



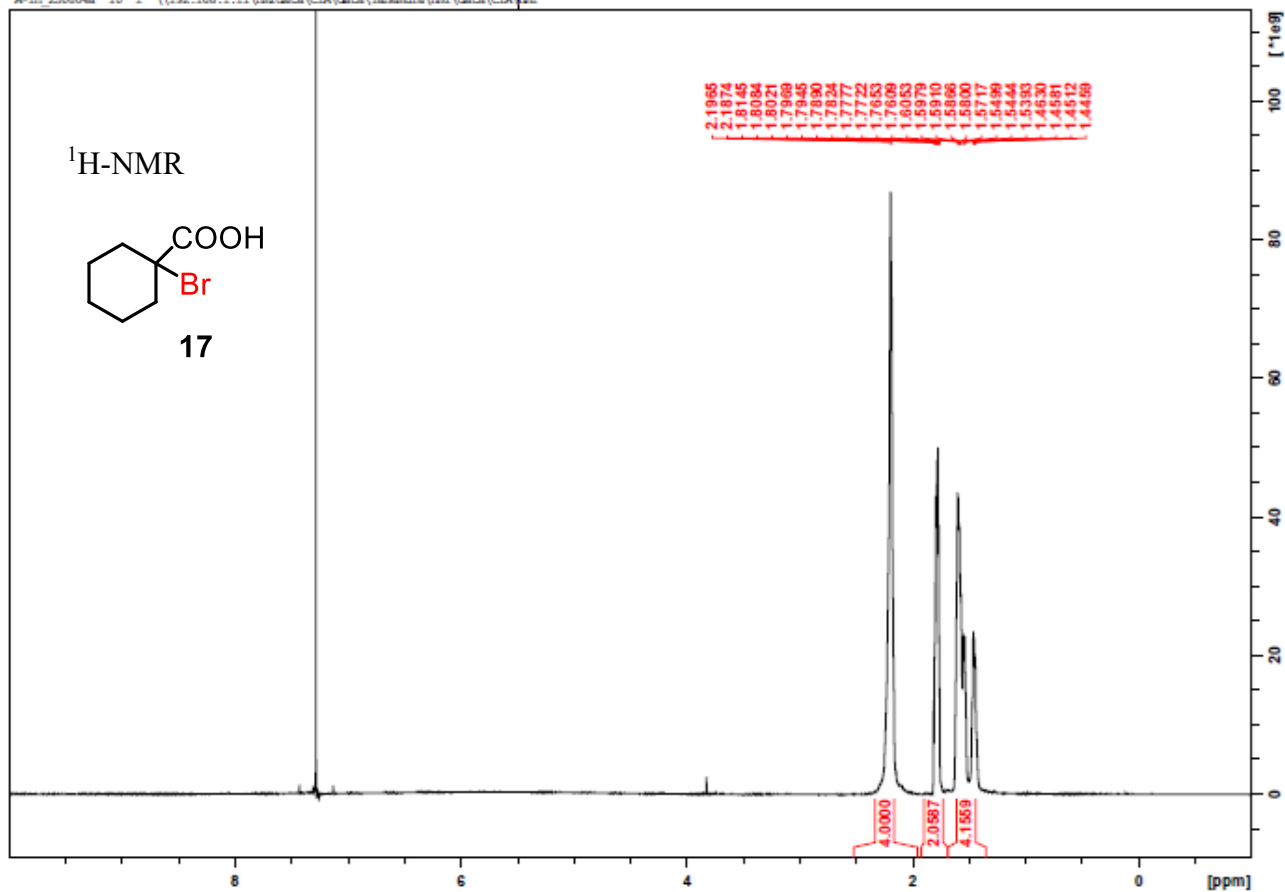
Tanomura-F-1H-250909a 10 1 \\192.168.1.11\rmrdata\CIA\data\tanomura\rmr\data\CIA\rmr



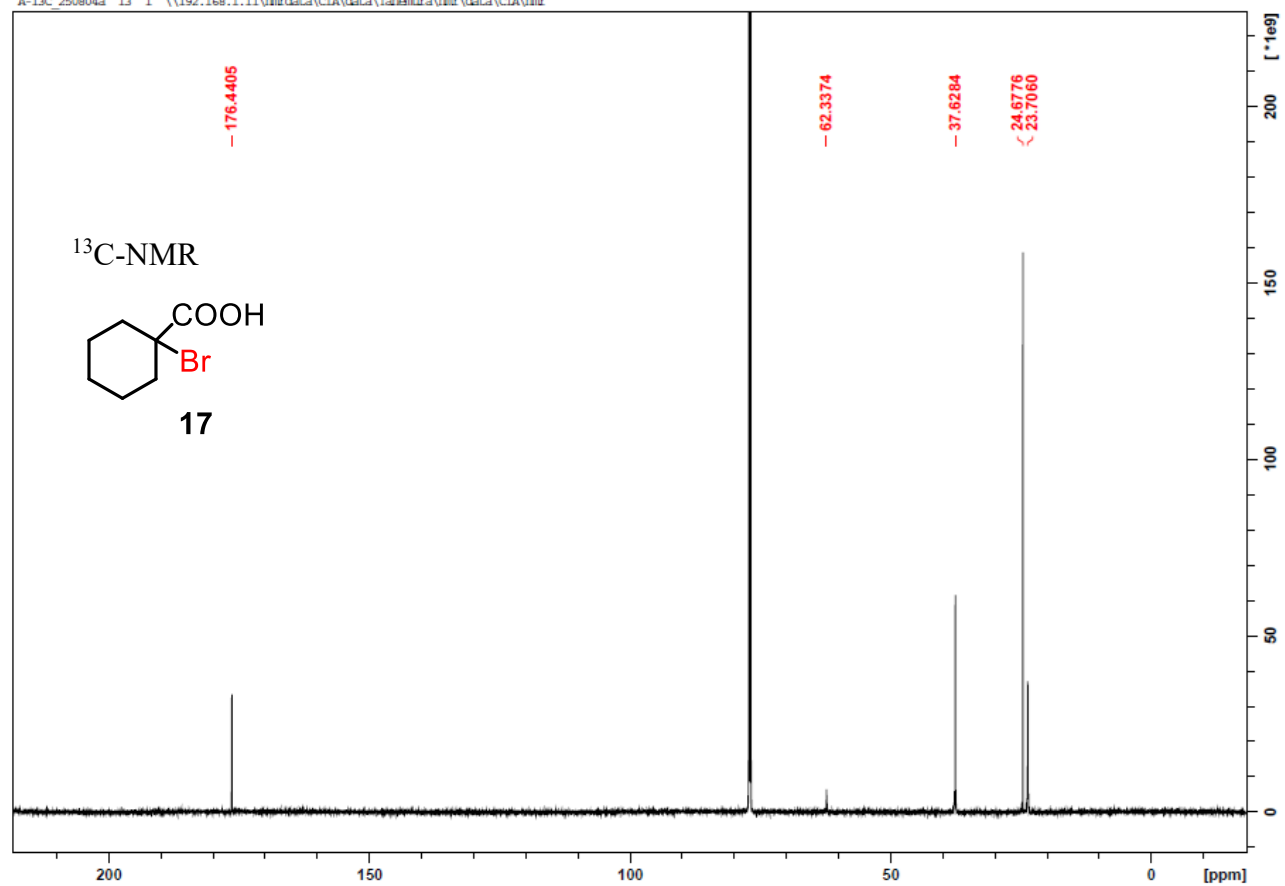
Tanomura-F-1H-250909a 11 1 \\192.168.1.11\rmrdata\CIA\data\tanomura\rmr\data\CIA\rmr



A-1H_250804a 10 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanemura\\nmr\\data\\CIA\\nmr



A-13C_250804a 13 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanemura\\nmr\\data\\CIA\\nmr



¹H-NMR

CCC(Cl)C(=O)O **21**

4.3209
4.3127
4.3099
4.3017
2.1507
2.1469
2.1425
2.1403
2.1320
2.1302
2.1220
2.1198
2.1116
2.1093
2.1011
2.0538
2.0432
2.0325
2.0150
2.0120
2.0014
1.1908
1.1190
1.1085
1.1048
1.1030
1.1009
1.0980
1.0942
1.0923
1.0905
1.0814
1.0786

0.8753
2.0000
2.9930

[ppm]

¹³C-NMR

Chemical structure of compound 21 (2-chloropentan-3-ol) is shown as an inset:

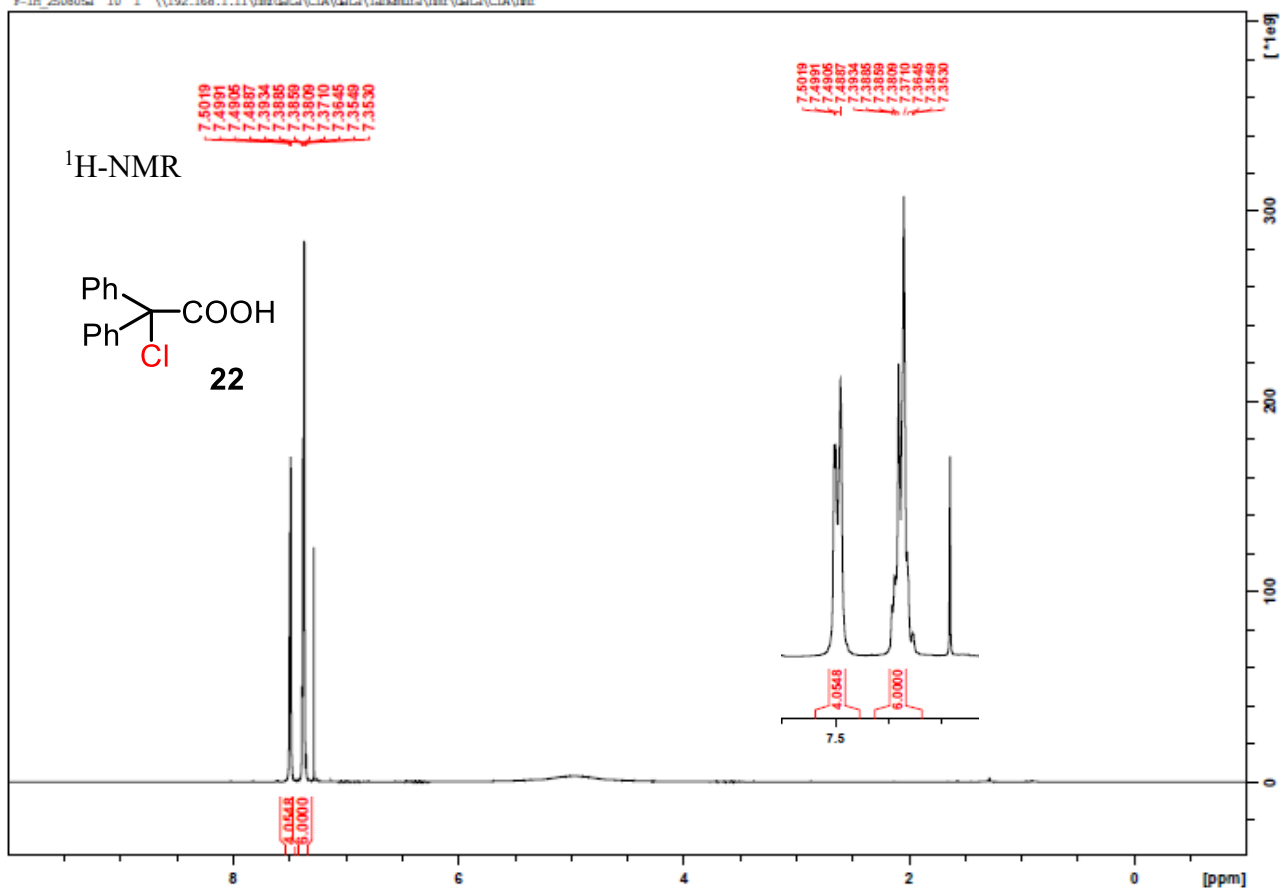
CCCC(Cl)CO

The spectrum displays four main peaks corresponding to the carbon atoms in the molecule:

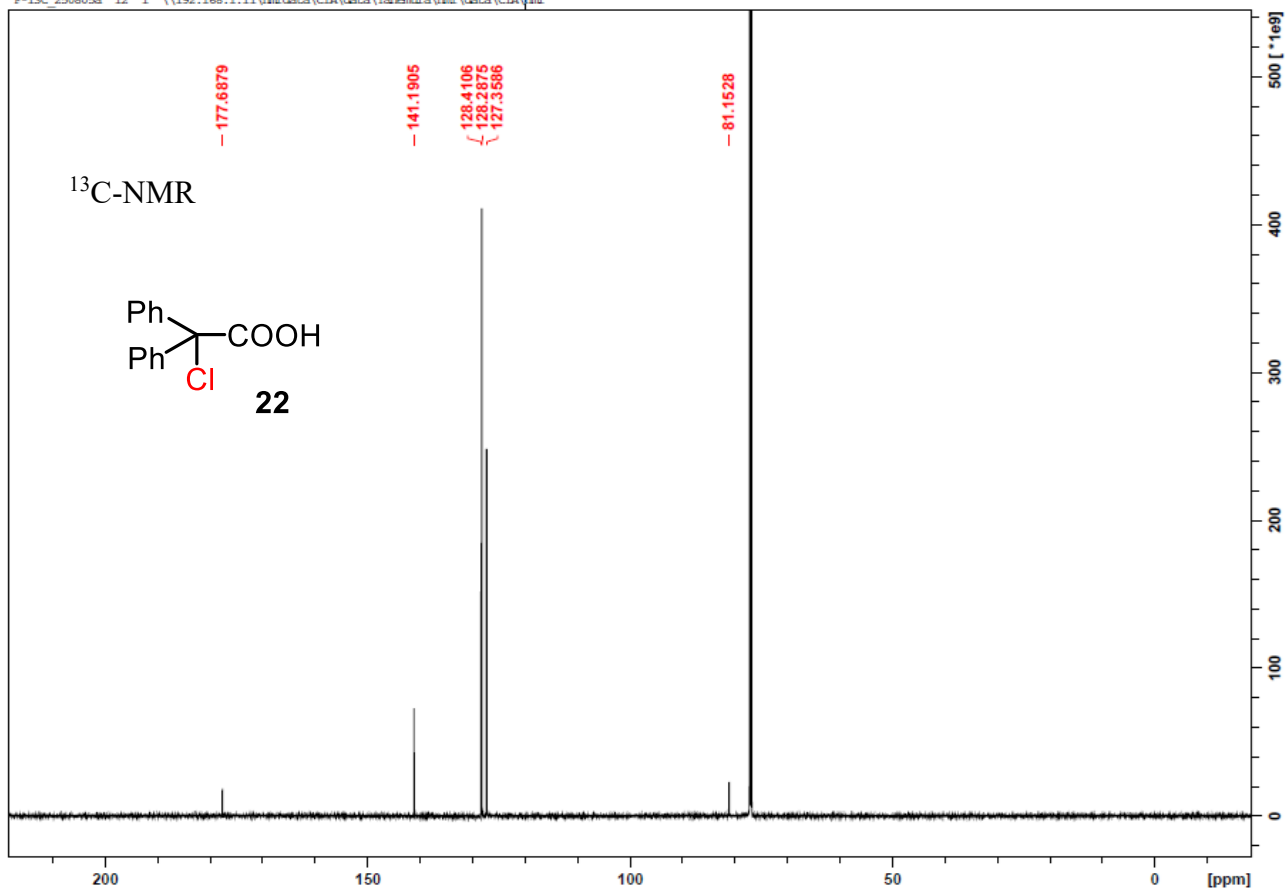
- Peak at 175.2071 ppm (labeled in red) corresponds to the carbonyl carbon (C=O).
- Peak at 58.5304 ppm (labeled in red) corresponds to the carbon atom bonded to the hydroxyl group (C-OH).
- Peak at 28.2544 ppm (labeled in red) corresponds to the carbon atom bonded to the chlorine atom (C-Cl).
- Peak at 10.4255 ppm (labeled in red) corresponds to the methyl carbon (CH₃).

The x-axis is labeled [ppm] and ranges from 0 to 200. The y-axis is labeled [1e9] and ranges from 0 to 500.

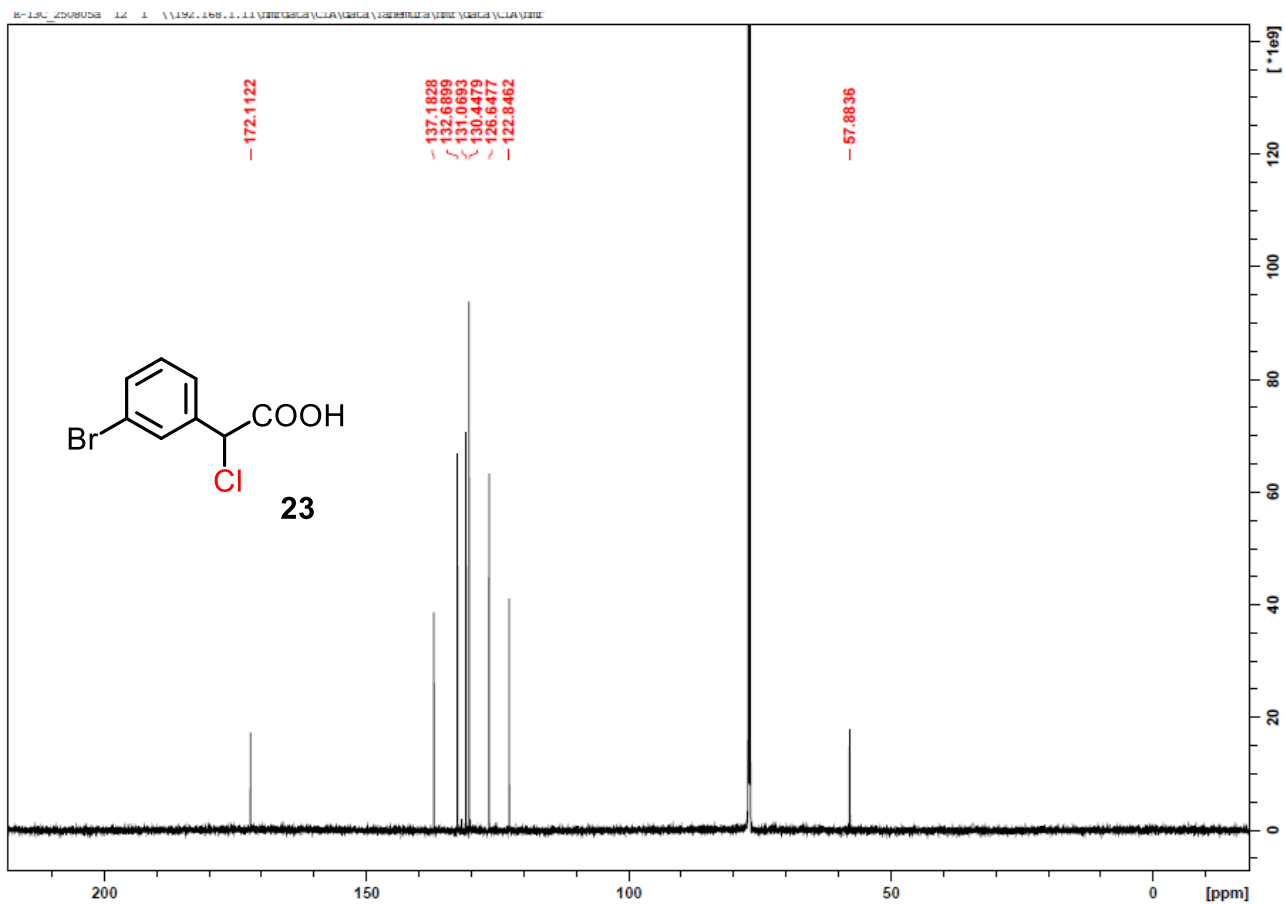
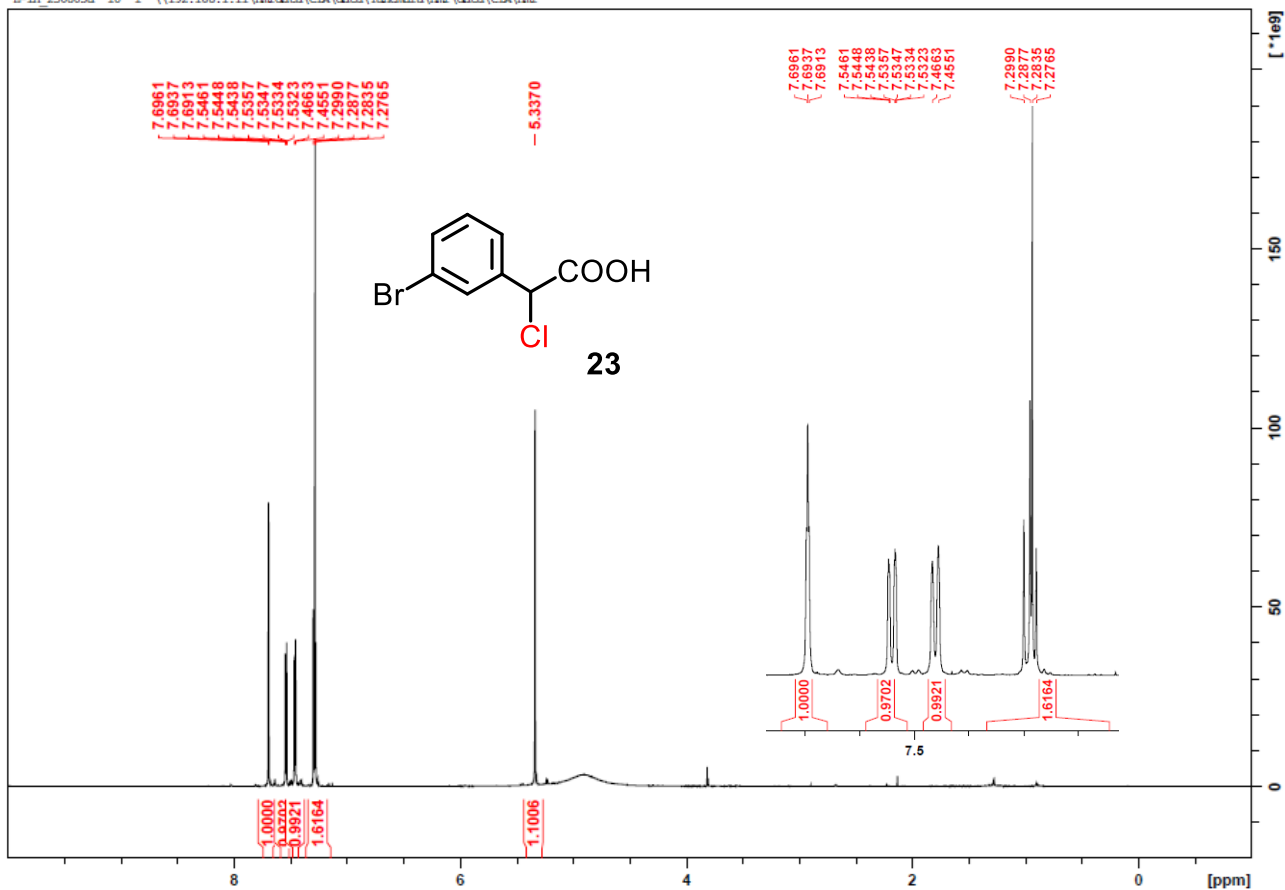
F-1H_250805a 10 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanomura\\nmr\\data\\CIA\\nmr



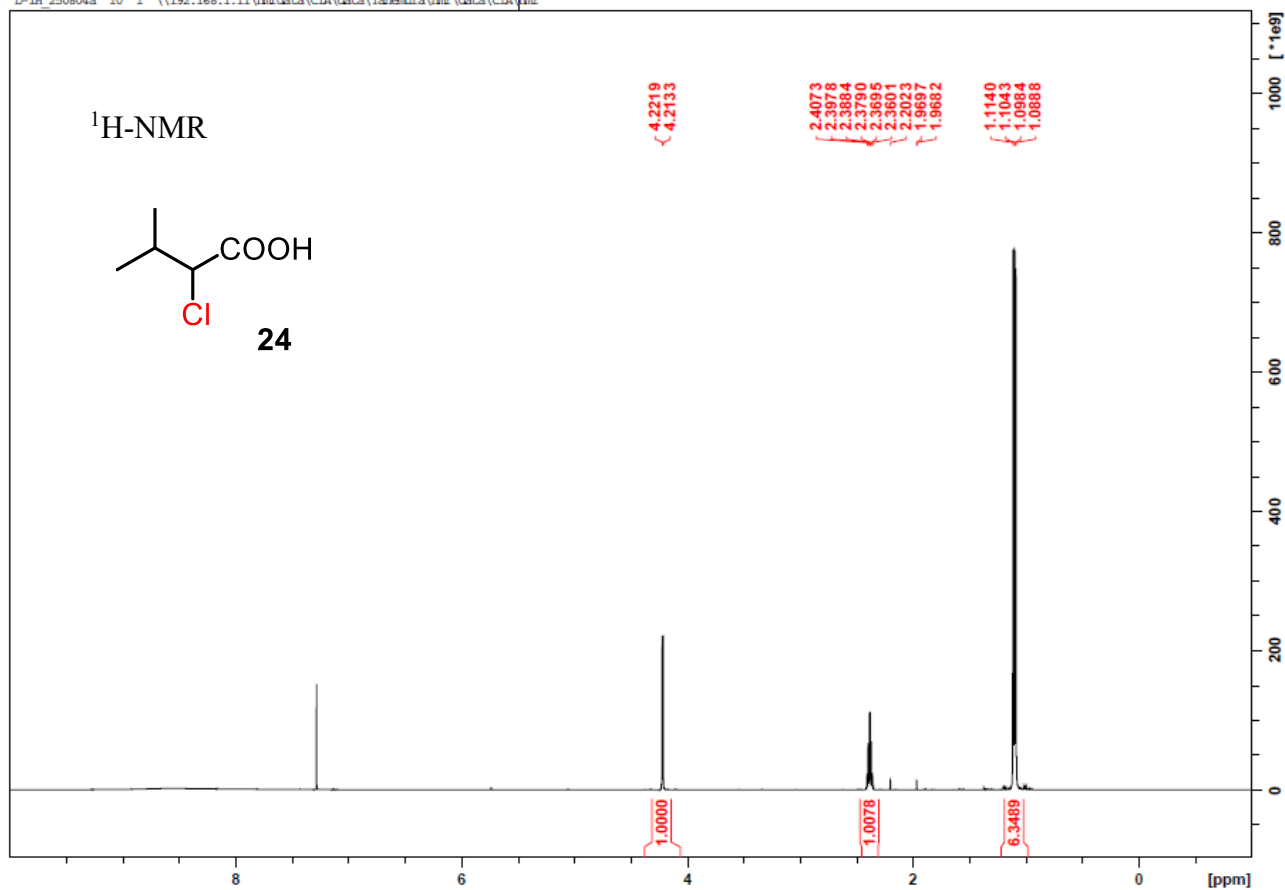
F-13C_250805a 12 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanomura\\nmr\\data\\CIA\\nmr



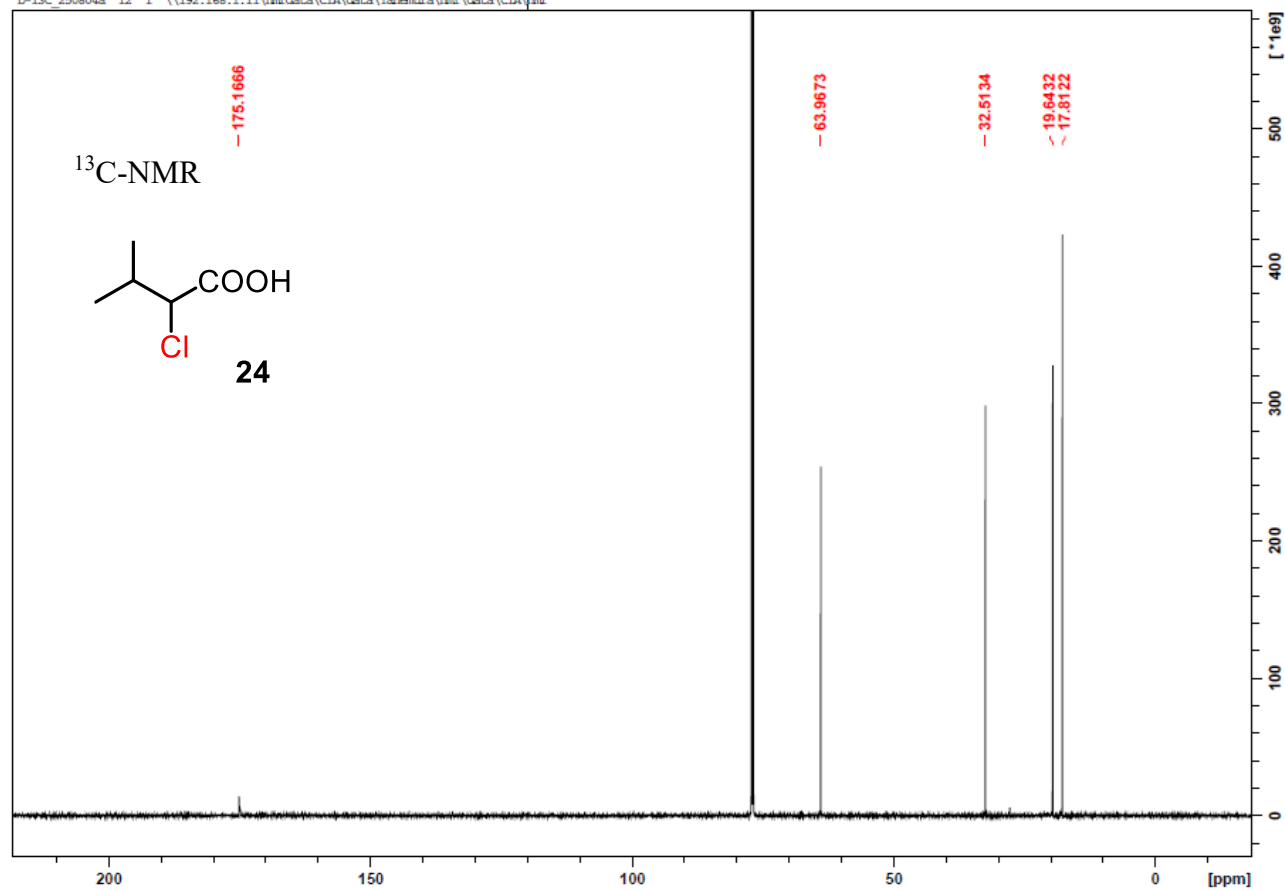
E-1H 250805a 10 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanemura\\nmr\\data\\CIA\\nmr



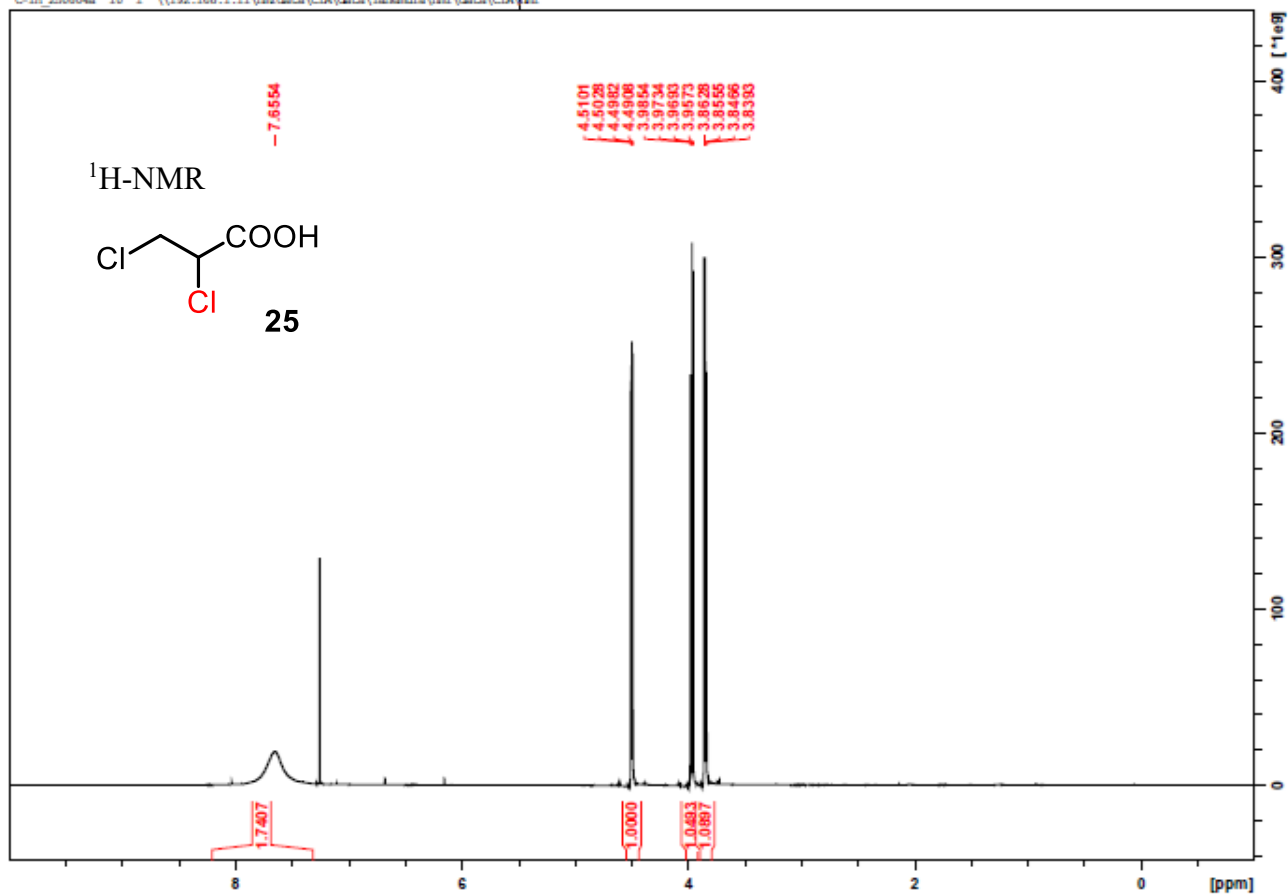
D-1H_250804a 10 1 \\192.168.1.11\vmr\data\CIA\data\Tanemura\vmr\data\CIA\vmr



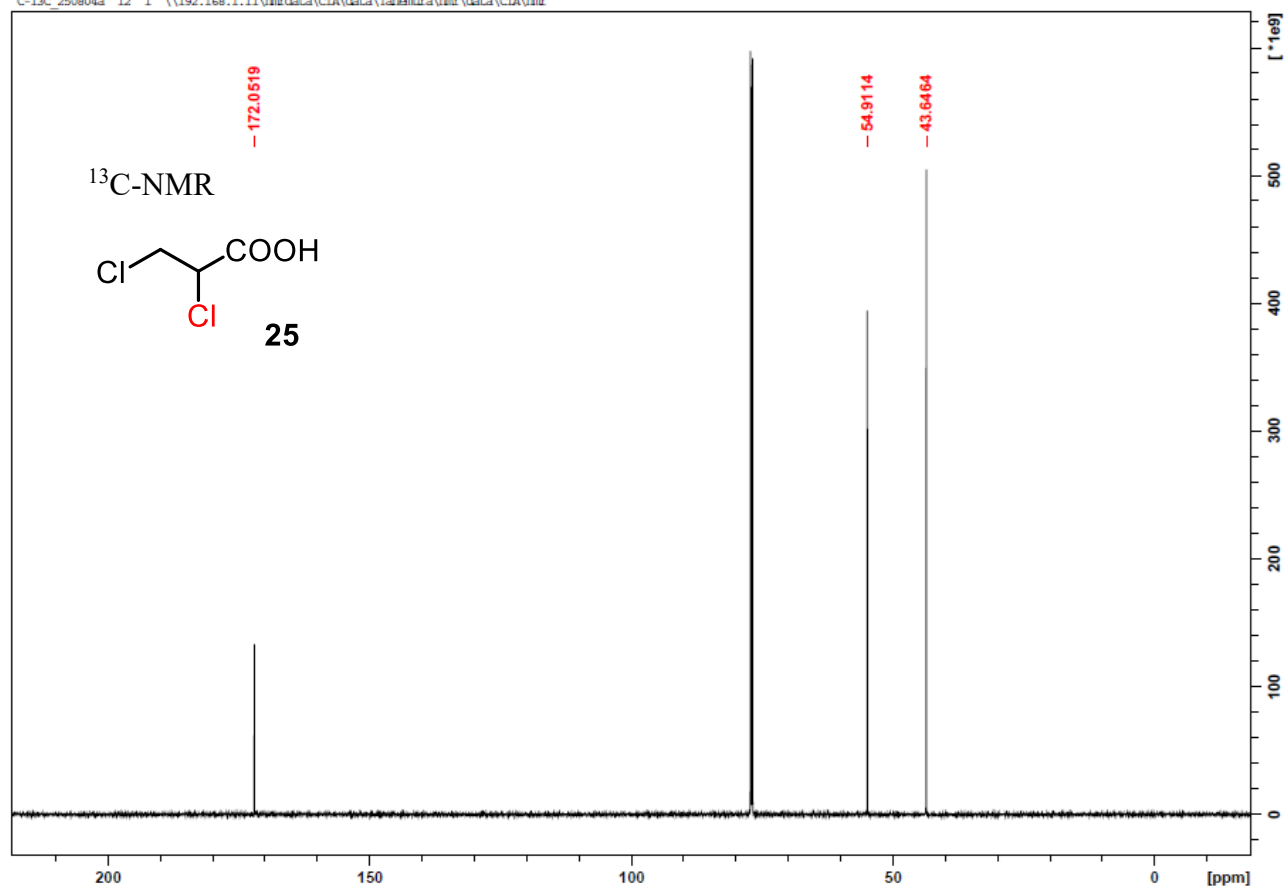
D-13C_250804a 12 1 \\192.168.1.11\vmr\data\CIA\data\Tanemura\vmr\data\CIA\vmr



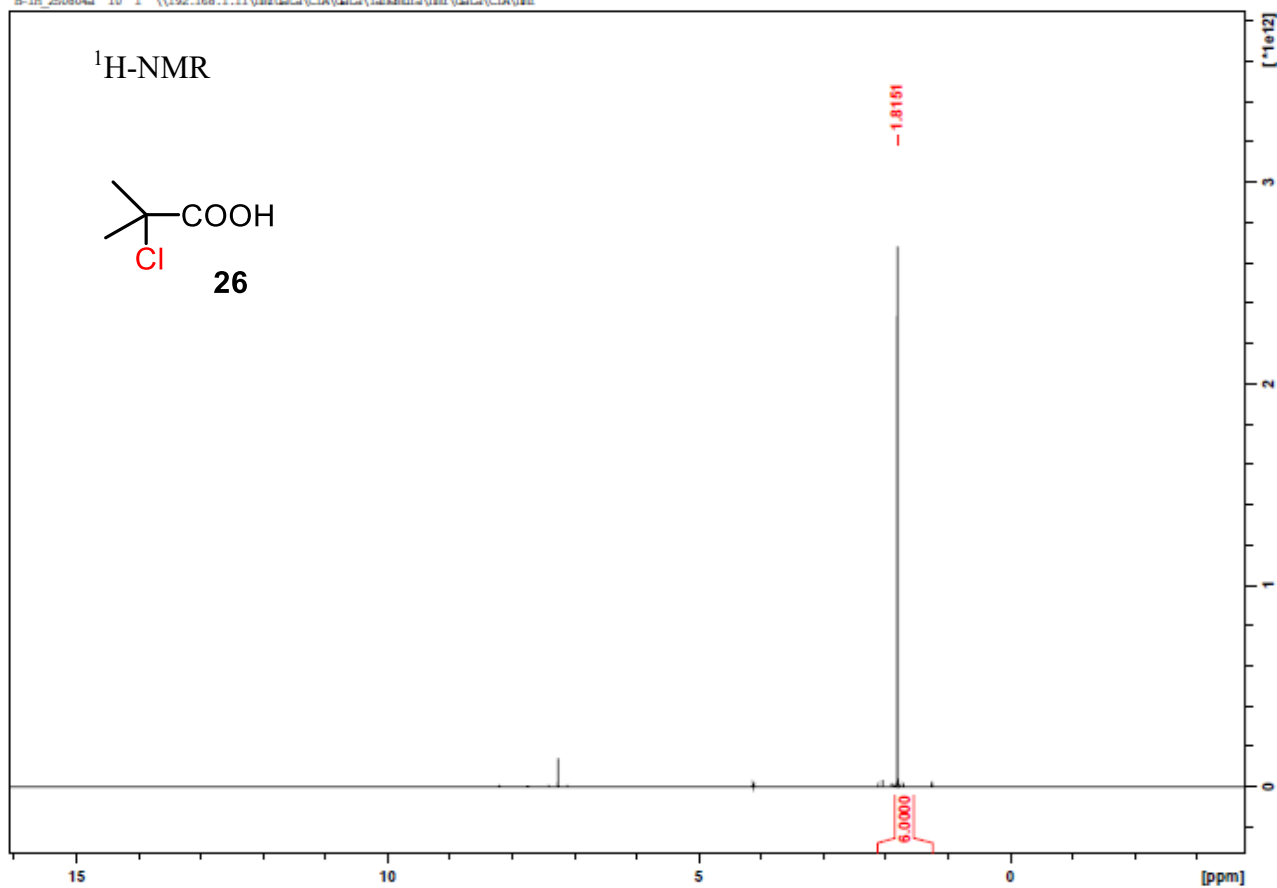
C-1H_250804a 10 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanemura\\nmr\\data\\CIA\\nmr



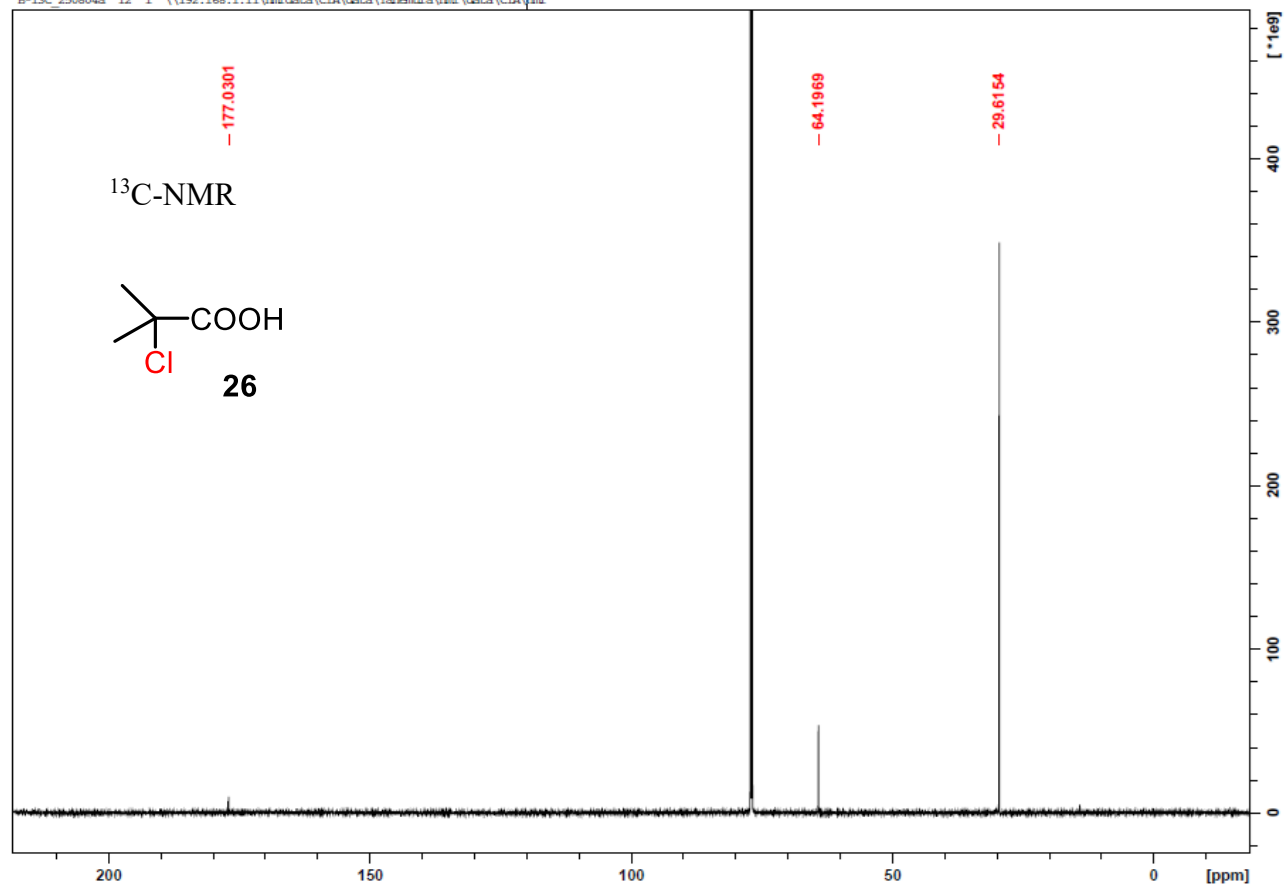
C-13C_250804a 12 1 \\192.168.1.11\\nmrdata\\CIA\\data\\Tanemura\\nmr\\data\\CIA\\nmr



B-1H_250804a 10 1 \\192.168.1.11\\rmrdata\\CIA\\data\\Tanemura\\rmr\\data\\CIA\\rmr



B-13C_250804a 12 1 \\192.168.1.11\\rmrdata\\CIA\\data\\Tanemura\\rmr\\data\\CIA\\rmr



6. References

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