

Electronic Supplementary Information

Mechanochemical synthesis of pyrrolo[1,2-*a*]indoles *via* consecutive C-C and C-N bond formation in presence of ionic liquid: Antimicrobial and photophysical studies

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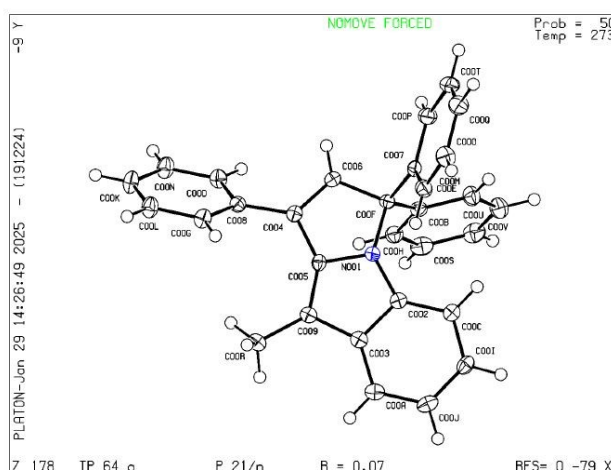
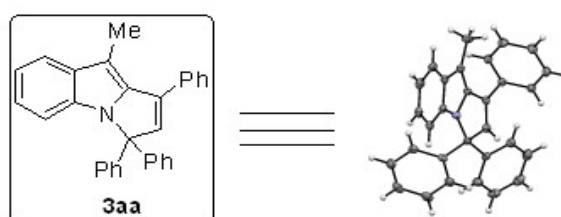
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1. General Information:

No purification of the chemicals has been carried out after purchasing from commercial sources. Solvents, reagents, and chemicals were purchased from Aldrich, Fluka, Merck, SRL, Spectrochem, and Process Chemicals. ^1H NMR spectra were determined as solutions in CDCl_3 on a 400 MHz spectrometer. $^{13}\text{C}\{^1\text{H}\}$ NMR and ^{19}F NMR spectra were recorded at 100 MHz and 376 MHz in CDCl_3 solution, respectively. Chemical shifts are in parts per million (δ) and reported as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (double doublet), and J (coupling constants) in Hz. Slide (made of glass) coated with silica gel has been used for thin-layer chromatography. Silica gel (100–200 mesh) as the stationary phase and ethyl acetate/petroleum ether was used for column chromatography. Melting points were determined on a glass disk with an electric hot plate and are uncorrected. A Borosilicate glass tube was used as a reaction tube.

2. Structure Determination (X-ray crystallographic data of **3aa**):

The colorless crystals of **3aa** were obtained by crystallization from a solution in dichloromethane/ hexane. Chemical Formula: $\text{C}_{30}\text{H}_{23}\text{N}$.

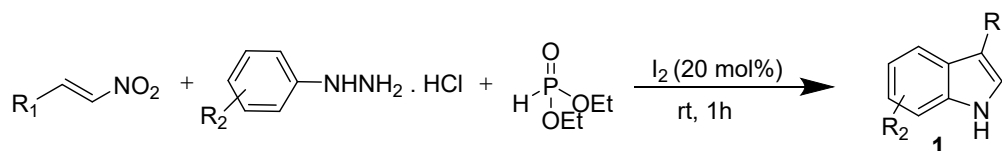


Wavelength	0.71073 Å	
Formula	C ₃₀ H ₂₃ N	
Crystal system	Monoclinic	
Space group	P 21/n	
Unit cell dimensions	a = 11.3687(4) Å	α = 90 °
	b = 17.6290 (7) Å	β = 108.254 °
	c = 11.5663 (5) Å	γ = 90 °
Volume	2201.45 (19) Å ³	
Z	4	
R-factor (%)	7	

The crystallographic data have been deposited with the Cambridge Crystallographic Data Centre as a supplementary publication with a CCDC reference number CCDC 2457288.

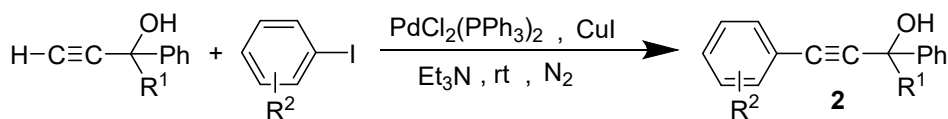
3. Experimental Procedures:

3.1. Typical procedure for the synthesis of 3-substituted indoles 1:



The synthesis of **1** was carried out according to the described method.¹ In a round bottom flask, 10 mmol of nitro styrene, 10 mmol of phenyl hydrazine hydrochloride, 20 mmol of diethyl phosphite and 20 mol% of iodine were stirred at room temperature for 1h. The mixture was then poured into 40 mL of a saturated sodium thiosulfate solution and extracted with dichloromethane. The extract was dried over anhydrous Na₂SO₄. Dichloromethane was removed in a vacuum and evaporated into a crude product. The crude product was passed through a silica gel column using petroleum ether as eluent. The eluent was removed in a vacuum, and the corresponding indole was obtained.

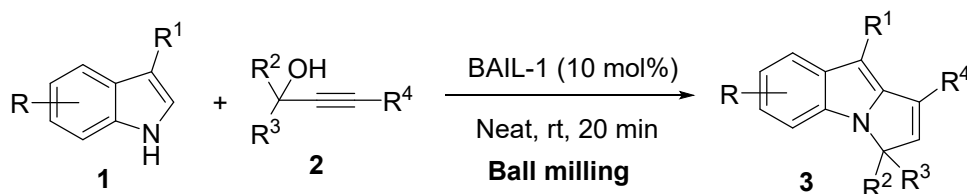
3.2. General procedure for the synthesis of substituted propargyl alcohols (2):



The starting propargyl alcohols were prepared according to the previously reported method.² To a mixture of bis(triphenylphosphine)palladium dichloride (0.1 mmol) and tri-ethylamine solution (60 mL) of iodoarene (10 mmol) under a nitrogen atmosphere in a flask equipped with a magnetic stirrer. Then, add alkyne moiety and the reaction mixture was allowed to stir for 6 h at room temperature. After completion of the reaction (monitored by TLC), the mixture was diluted with saturated saline water (3×15 mL), and extracted with ethyl acetate. The combined organic layer was collected and dried over anhydrous Na_2SO_4 . The residue was purified by column chromatography on silica gel to get the desired products.

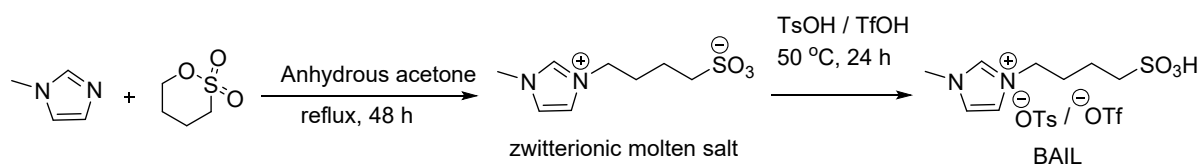
The propargylic alcohol **2a** ($\text{R}^1 = \text{Ph}$, $\text{R}^2 = \text{H}$) was purchased commercially from TCI Chemicals. **2b**,³ **2c**,³ **2d**,⁴ **2e**⁵ are known compounds, and their spectral data showed good agreement with the literature data.

3.3. General procedure for the synthesis of 3:



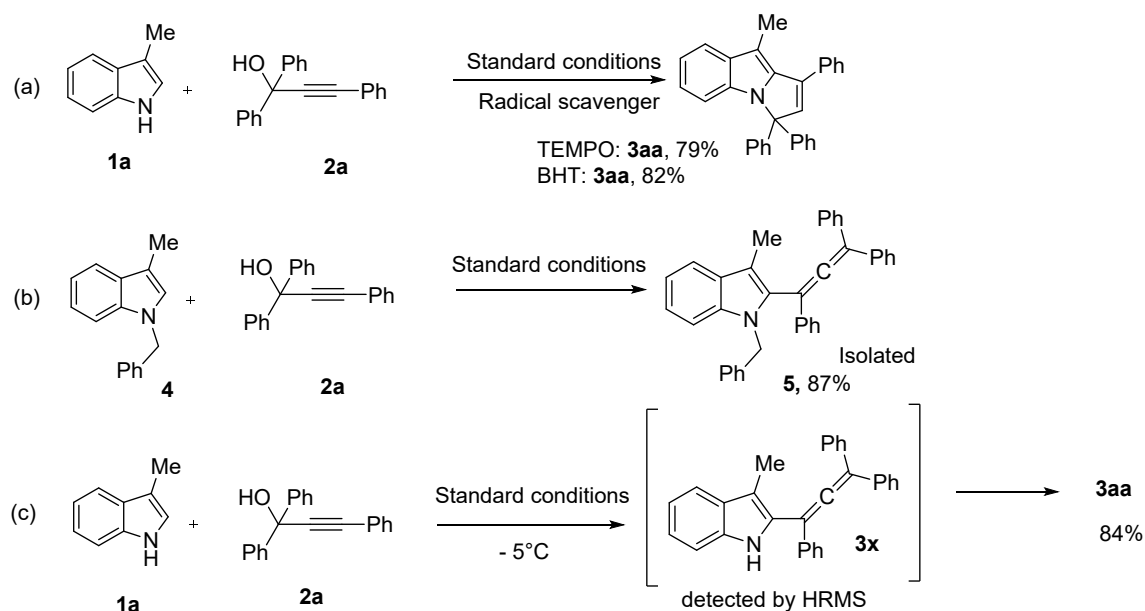
A grinding beaker (50 mL) and milling balls (4×10 mm) were set as a reaction chamber. For each reaction, a mixture of propargyl alcohol (1 mmol), 3-substituted indole (1 mmol), with 10 mol% of Brønsted acidic ionic liquid were milled for 20 min at 500 rpm at room temperature. After completion of reaction (monitored by TLC), the mixture was diluted with saturated saline water (3×15 mL), and extracted with ethyl acetate. The combined organic layer was collected and dried over anhydrous Na_2SO_4 . The residue was purified by column chromatography on silica gel to get the desired products.

3.4. General procedure for the synthesis of Brønsted acidic ionic liquids (BAILs):



Brønsted acidic ionic liquids (BAILS) were prepared according to the previously reported method.⁶ A mixture of an equivalent amount of 1-methylimidazole and corresponding butane sultone was refluxed in anhydrous acetone for 48 h to get the zwitterionic molten salt in about 99% yield. After washing the salt with diethyl ether and toluene to remove any unreacted starting materials, the solid was dried in vacuo. Then, a stoichiometric amount of corresponding sulfonic acid was added and the mixture was stirred for 240 h at 50 °C, during which time the solid zwitterion dissolved/liqefied and resulted in the formation of –SO₃H acidic ionic liquids (BAILS) with yield of ~98%. The IL phase was then washed repeatedly with toluene and ether to remove non-ionic residues and dried in vacuo.

4. Mechanistic investigation:



5. Antimicrobial assay for the compounds **3am** & **3ak**

5.1 Antimicrobial activity using Agar well diffusion method

The agar well diffusion method was used to evaluate the antibacterial activity of test compounds **3am** and **3ak**.⁷ To begin the assay, microbial inocula were evenly spread on sterile nutrient agar plates. Uniform wells were then made using a sterile cork borer. Five bacterial strains were tested: Gram-positive *Bacillus cereus* (ATCC 13061), *Listeria monocytogenes* (MTCC 657), *Staphylococcus aureus* (MTCC 96), Gram-negative *Salmonella typhimurium* (MTCC 98) and *Escherichia coli* (MTCC 1667). All strains were cultured in nutrient broth to the logarithmic phase and standardized to an optical density (OD) of 0.5 at 600 nm. Test compounds, dissolved in DMSO, were added to the wells at concentrations of 50, 25, and 10 mg/ml (25 μ l per well). The plates were then incubated at 37°C for 12–16 hours. Following incubation, the zones of inhibition were measured to assess antimicrobial activity.⁸

5.2 Minimum Inhibitory Concentration (MIC) value determination

The lowest concentration of an antimicrobial agent that inhibits bacterial growth is known as the minimum inhibitory concentration or MIC.⁹ A modified resazurin-based assay determined the MIC value of **3am** and **3ak**.¹⁰ Resazurin is a blue dye that is reduced to pink resorufin by metabolically active cells, serving as an indicator of cell viability. In this assay, test compounds were prepared at concentrations of 1000, 500, 250, and 125 μ g/mL in a 96-well microtiter plate. Each well was filled using a micropipette with the test compound, bacterial suspension, and 0.01% resazurin solution. The negative control consisted of bacterial cultures in DMSO without any test compound, while ciprofloxacin served as the positive control. After 10 hours of incubation at 37°C, the minimum inhibitory concentration (MIC) was determined as the lowest concentration that retained the blue colour of resazurin, indicating the absence of bacterial growth.¹¹

5.3 SEM analysis of bacterial strains

To evaluate the effectiveness of compound **3am**, which was found to be more effective than the others, morphological changes in bacterial cells were examined using scanning electron microscopy (SEM). The analysis provided insights into structural alterations caused by the

treatment. Two bacterial strains, *Bacillus cereus* and *Staphylococcus aureus*, were cultured in fresh nutrient broth and treated with compound **3am** at its minimum inhibitory concentration (MIC). After overnight incubation, the bacterial cells were fixed with 2.5% glutaraldehyde, centrifuged, and dehydrated through a graded ethanol series (10% to 90%). Following critical point drying, the samples were mounted on metal stubs, coated with gold using an ion sputter, and imaged using a high-resolution SEM (Carl Zeiss Gemini 450).¹²

5.4 Results

5.4.1. Antimicrobial activity using the Agar well diffusion method

Significant variations in the antibacterial activity were found for the compounds **3am**, **3ak**, **3ap**, **3ar**, **3ax**, but two of them showed the best activity against five bacterial strains found using the agar well diffusion experiment. The compound **3am** (25 mg/ml) demonstrated greater activity with inhibitory zones that ranged from 8 ± 0.5 mm for *Bacillus cereus* to 6 ± 1.5 mm for *Listeria monocytogenes*. On the other hand, smaller zones were produced by the compound **3ak** (25 mg/ml), with zones measuring 3.5 ± 1.0 mm for *Listeria monocytogenes* and for *Escherichia coli* inhibition zones measuring 5.5 ± 1.0 mm (Table S1). In the antibacterial assay, the compound **3am** exhibited a greater inhibitory zone than **3ak**, as shown in Figure S1.

Table S1 Inhibition zone of antimicrobial activity for the **3ak** and **3am** using the agar well diffusion method

Test materials (25 mg/mL)	Zone of Inhibition (mm) (Mean \pm SD)				
	<i>Escherichia coli</i>	<i>Staphylococcus aureus</i>	<i>Salmonella typhimurium</i>	<i>Bacillus cereus</i>	<i>Listeria monocytogenes</i>
3ak	5.5 ± 1.0	4.5 ± 1.5	4.5 ± 1.5	4 ± 1.0	3.5 ± 1.0
3am	7.0 ± 1.5	7.5 ± 0.5	7 ± 1.5	8 ± 0.5	6 ± 1.5

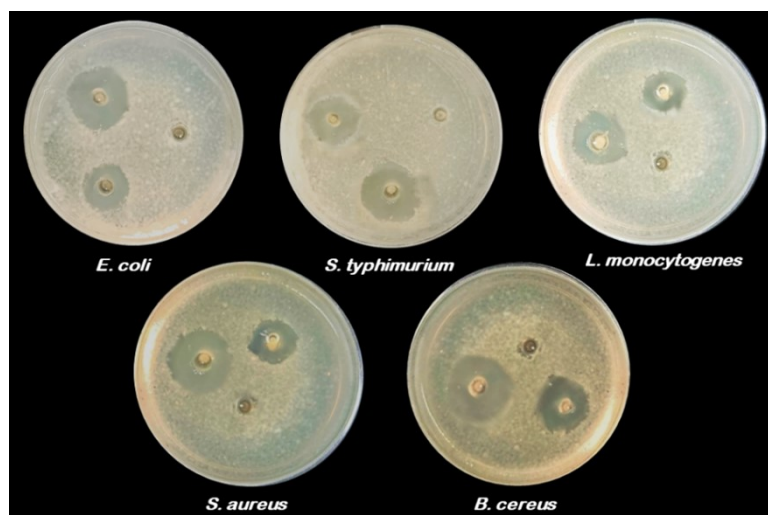


Figure S1 Antibacterial screening of **3am** and **3ak** against five bacterial strains (agar-well diffusion method). A larger inhibition zone indicates for **3am**, whereas a smaller one indicates **3ak**.

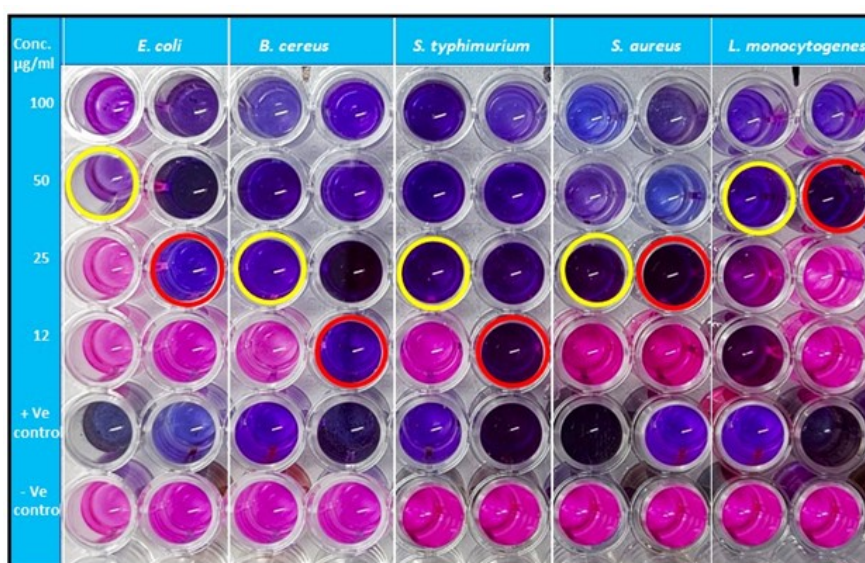


Figure S2 The microtiter plate, after a 14-hour incubation period, showed the antibacterial activity of the test compounds against the five bacterial strains. Blue wells signify the inhibition of bacterial growth, while pink wells show the growth of bacteria. Red circles represent the value of **3am**, while yellow circles represent the value of **3ak**. Positive and negative controls are below

Table S2 MIC values of the compounds **3ak** and **3am** against different bacterial strains

List of Bacterial strains	Gram-positive/Gram-negative	MIC values of 3ak (µg/mL)	MIC values of 3am (µg/mL)	MIC values of ciprofloxacin (µg/mL)
<i>Bacillus cereus</i>	Gm +ve	25	12	03
<i>Salmonella typhimurium</i>	Gm -ve	25	12	03
<i>Escherichia coli</i>	Gm -ve	50	25	02
<i>Staphylococcus aureus</i>	Gm +ve	25	25	01
<i>Listeria monocytogenes</i>	Gm +ve	50	50	02

5.4.2 Evaluation of Minimum Inhibitory Concentration (MIC):

The minimum concentration of an antimicrobial agent needed to inhibit bacterial growth can be determined by the Minimum Inhibitory Concentration (MIC). A redox indicator called resazurin detects the metabolic activities of bacteria. Positive responses were indicated by a continuous blue coloring, whereas responses that were negative were indicated by changing the color to pink or colorless. Although the compounds **3am**, **3ak**, **3ap**, **3ar**, and **3ax** exhibited good antibacterial activity, MIC values were determined only for **3am** and **3ak**, as these two compounds demonstrated the best bioactivity. MIC values for compound **3ak** ranged from 25 to 50 µg/ml against each tested bacterial strain. Meanwhile, MIC values for **3am** ranged from 12 to 25 µg/ml against test bacteria and exhibited stronger antibacterial activity (Table S2). The higher antibacterial potency of compound **3am** is demonstrated by these MIC values over **3ak**, which were also confirmed by triple replicates (Figure S2).

Similarly, scanning electron microscopy (SEM) provides a direct and effective method for evaluating the mechanism of antibacterial action by revealing structural damage in bacterial cells. The images clearly differentiate between live and dead cells, demonstrating the impact of the treatment. A comparison between untreated cells and those treated with compound **3am** allows for a more precise assessment of cell integrity. This visual evidence

underscores the compound's antibacterial potential by highlighting its ability to disrupt and destroy the bacterial cell wall (Figure S3).

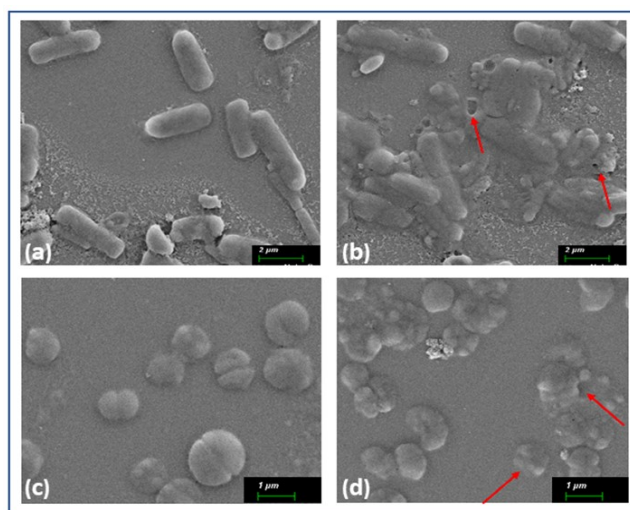


Figure S3 FE-SEM images of a Gram-negative and a Gram-positive bacterium: (a) *E. coli* before treatment; (b) *E. coli* after treatment with **3am**; (c) *S. aureus* before treatment; (d) *S. aureus* after treatment with **3am**. Red arrows indicate dead bacterial cells.

6. Photophysical Studies:

We studied the photophysical behavior of these 3*H*-pyrrolo[1,2-*a*]indole derivatives. The spectroscopic results are shown for **3ae** (Figure S4). The principal absorption band falls between 325 and 346 nm in the experiment. All of the examined compounds were found to be fluorescent in the blue region (λ_{em} = 422-463 nm, Figure S5).¹³ The electron donating group present in **3ad** and **3ae** have relatively high emission wavelengths while the electron withdrawing group present in **3an** showed the low emission wavelength. The fluorescence quantum yield (Φ) of the synthesized compounds are in the range of 0.02-0.43 in acetonitrile. Here tryptophan is used as a reference at λ_{ex} = 280 nm (Φ_R = 0.14 at 298 K).¹⁴ The thiomethyl group on the arene moiety of 3*H*-pyrrolo[1,2-*a*]indole in compound **3ae** shows higher Φ (0.43) value and higher stokes shift value (123 nm). These values indicate the distortion of the π conjugated system in excited state.¹⁵ Compound **3an** showed the lower Stokes shift value (97 nm) and relatively lower quantum yield value (0.16). Remarkable differences between alkyl and aryl substituted 3*H*-pyrrolo[1,2-*a*]indole derivatives were also found in fluorescence quantum yield. Compound **3ae** exhibited ~21-fold higher quantum yield value and longer emission wavelength than compound **3aa**. Additionally, the molar absorptivity (ϵ_{max}) for **3ab** (-OMe) is higher (25000 M⁻¹cm⁻¹) and lower (7000 M⁻¹cm⁻¹) for **3an** (-CF₃),

which indicates the probability of transition from ground state(S_0) to first excited electronic state (S_1) is higher for those derivatives with strong electron donating group comparative to strong electron withdrawing group.¹⁶ These photophysical properties with high quantum yield and Stokes shift value establish the compounds as unique material for potential bioimaging¹⁷ and further utilized them as fluoroprobes.

In addition, we examine the solvatofluorochromic properties of selected five synthesized compounds by changing the solvent polarity from non-polar to polar solvents. Figure S4(c) shows the emission spectra of **3ae** in *n*-hexane, THF, methanol, acetonitrile and DMSO. The results of solvent effect with **3ai**, **3at**, **3an** and **3aw** are shown here. With increase in the polarity of solvents, the compounds show a moderate bathochromic shift. The emission maxima of **3ae** locates at 448 nm in *n*-hexane, while the emission spectra shift to 464 nm in DMSO. In general, the compounds have high dipole moments in excited state as compared to their corresponding ground state due to electronic excitations.¹⁸ The high dipole moment of compounds interacts with the environment of the polarity of solvents and results the slight shifts of emission maxima to longer wavelengths.¹⁹ The observed trend in photoluminescence characteristics is evidently showed that 3*H*-pyrrolo[1,2-*a*]indole derivatives have good tendency to photoluminescence properties.

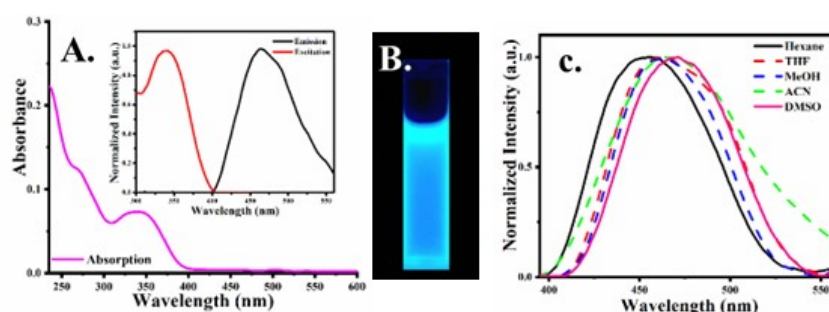


Figure S4 Photophysical behavior of compound **3ae**: (a) The UV-Vis absorption characteristic of compound **3ae** in acetonitrile (ACN) solvent medium, inset indicates the normalized excitation and emission spectra of the same, (b) the high emissive image of compound **3ae** in ACN solution under fluorescence light and (c) the solvent dependent normalized emission spectra of compound **3ae**.

6.1. Steady state UV-Vis absorbance and fluorescence spectroscopic measurements:

UV-Vis absorption spectra of the selected thirteen compounds were recorded in acetonitrile solution with the help of a UV-Vis-NIR spectrophotometer (Shimadzu UV 1800 pc) within

the wavelength range of 200-600 nm. The fluorescence emission and corresponding excitation spectra of the same compounds in acetonitrile solution were obtained using a Perkin Elmer LS55 fluorescence spectrophotometer at the excitation wavelength (λ_{ex}) 290 nm to get the maximum fluorescence intensity. The slit width was fixed at 7/2.5 nm for both the excitation and emission beams. The spectroscopic studies were carried out at 25° C and no change was observed in the spectra of all experimental solutions for a long time period. During experimental studies, this nullifies any possibility of degradation of the sample solutions. Molar extinction coefficient (ϵ) of the thirteen compounds in acetonitrile solution for the maximum absorption band was determined with the help of Beer-Lambert's law ($A = \epsilon cl$) where concentration (c) of the compounds in micro molar range and path length (l) was kept as 1 cm and 'A' represent the absorbance. For calculation of the Stokes shifts, considered the lowest energetic absorption band of compounds.

6.2. Fluorescence Quantum Yield (Φ) calculations of compounds:

The relative fluorescence quantum yields (Φ) of the selected thirteen compounds were calculated in acetonitrile solution by using the following equation:

$$\Phi = \Phi_R \times \frac{I}{I_R} \times \frac{OD_R}{OD} \times \frac{\eta_R^2}{\eta^2} \dots (1)$$

Where Φ_R represents the quantum yield of reference, here tryptophan in a water medium is used as reference; I and I_R denote the area under the fluorescence peak of samples and reference respectively; OD and OD_R indicate the absorbance for samples and reference respectively; η and η_R are the refractive index of respective solvent. Here, refractive indexes of the medium ($\eta = 1.333$, water) and ($\eta = 1.344$, acetonitrile). The absorbance value of sample compounds and reference was kept between 0.01 to 0.1 to avoid self-quenching and self-aggregation. Quantum yield value ($\Phi_R = 0.14$ at 298 K) of tryptophan in water medium at $\lambda_{\text{ex}} = 280$ nm used as reference.¹⁴

UV-Vis absorbance and steady-state fluorescence spectra were measured to understand the photo-physical behaviour of 3*H*-pyrrolo[1,2-*a*]indole derivatives. Here we selected thirteen 3*H*-pyrrolo[1,2-*a*]indole derivatives (**3ae**, **3au**, **3ab**, **3af**, **3at**, **3ai**, **3ah**, **3ad**, **3an**, **3aw**, **3ax**, **3aj** and **3aa**) in acetonitrile medium for UV-Vis absorbance measurement and the concentration of all measured solutions is fixed at 5 μ M. Similarly, the excitation and emission spectra of the same solutions were recorded in the same medium. Selected

compounds manifest captivating photo-physical behaviours; the detailed spectral results are summarized in **Table S3**. **Table S3** contains the values of λ_{abs} (wavelengths related to several absorption bands), the excitation wavelength (λ_{ex}), maxima of emission wavelength (λ_{max}), molar extinction coefficients, ϵ (at the wavelength having the highest absorbance), Stokes shifts and fluorescence quantum yields. **Figure S5** indicates the absorption and normalized excitation and emission spectra of compounds (**3au-3aa**). From all the figures, it is evidently noticed that the excitation and absorption spectra nearly resemble, signifying the purity of the compounds. The UV-Vis spectral patterns show mainly one absorption band (λ_{abs}) at ~325-346 nm. No remarkable changes are observed in the absorption spectra upon the substitution of 3-position of the indole ring. Electron donating groups as substituents show the red shift as compared to the electron withdrawing group as substituents. The fluorescence emission studies show an intense emission band in visible region at ~ 422-463 nm (λ_{em}) (**Table S3**). The emission studies of compound (**3ae** and **3ad**) show a red shift in the emission maximum upon increase the conjugation compared to other compounds.

Table S3 Summarized photo-physical parameters of the selected thirteen indole derivatives

Compounds	Absorption (λ_{abs}) (nm)	Excitation (λ_{ex}) (nm)	Emission λ_{max} (nm)	Molar Extinction Coefficient (ϵ) ($10^4\text{M}^{-1}\text{cm}^{-1}$)	Stokes Shift (nm)	Quantum Yield (Φ)
3ae	340	290	463	1.0	123	0.43
3au	346	290	457	1.5	111	0.41
3ab	335	290	452	2.5	117	0.38
3af	335	290	456	1.8	121	0.37
3at	330	290	450	2.2	120	0.31
3ai	331	290	451	1.2	120	0.29
3ah	330	290	448	1.3	118	0.24
3ad	337	290	463	1.2	126	0.19
3an	325	290	422	0.7	97	0.16
3aw	333	290	451	1.8	118	0.14
3ax	330	290	453	1.0	123	0.11
3aj	333	290	450	1.3	117	0.09
3aa	330	290	430	1.2	100	0.02

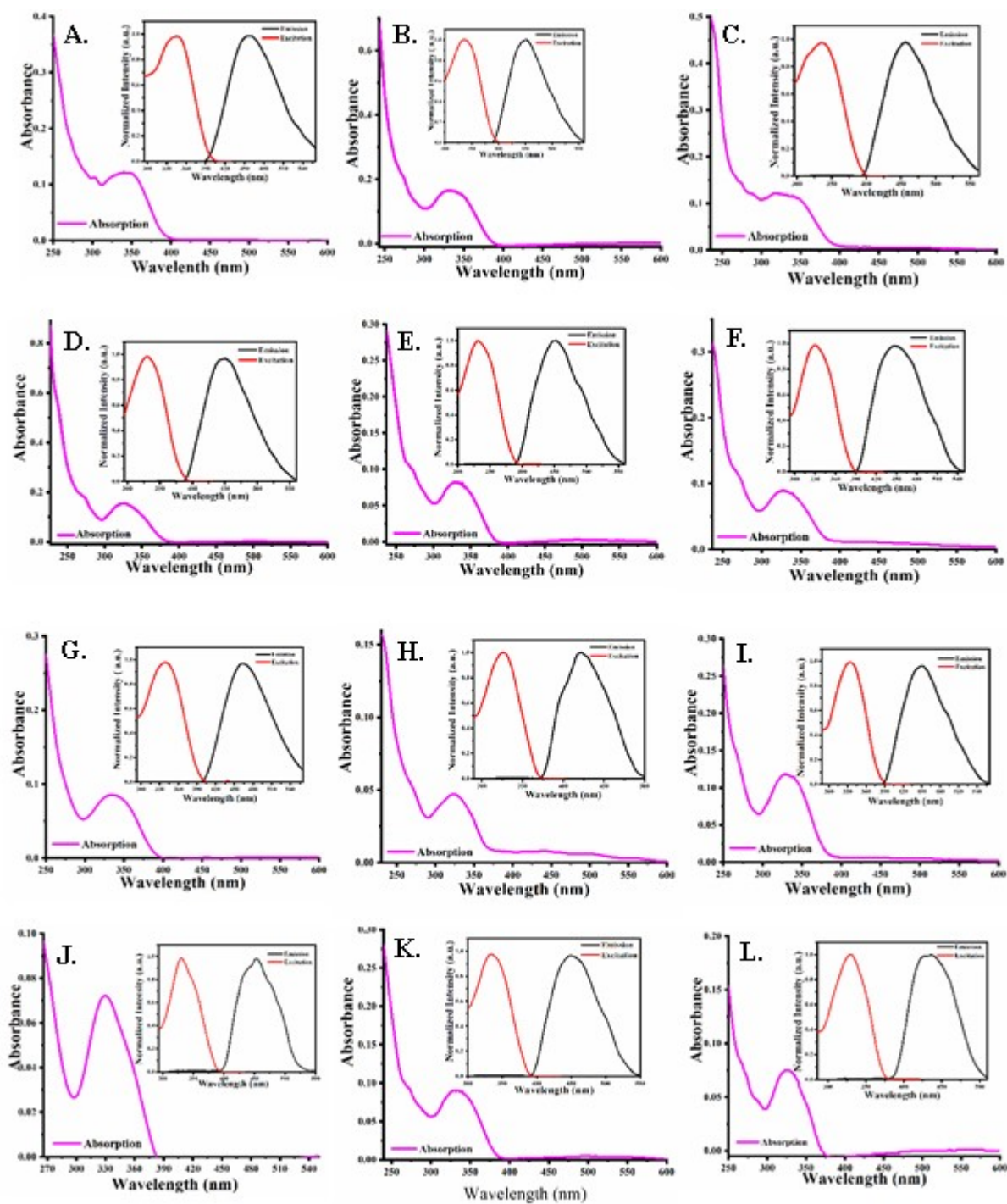


Figure S5 (A-L) UV-Vis absorption spectra of compounds (**3au-3aa**) in acetonitrile medium respectively (compound final concentration: $5\mu\text{M}$), inset shows the normalized excitation and emission spectra of the same.

Furthermore, the solvatofluorochromic properties of selected five compounds are examined in various solvents as the polar substituents of aromatic ring are sensitive to the variation of solvents. **Figure S6** shows the emission spectra of selected compounds are in *n*-hexane, THF, methanol, acetonitrile and DMSO. With increase in the polarity of solvents, the compounds show a moderate bathochromic shift. Here all the five compounds exhibited same observations with the effect of solvents. The emission maxima of **3ai**, **3at** and **3aw** locate at 439 nm, and for **3an** at 410 nm in *n*-hexane, while the emission spectra shift to 453 nm for first three compounds and 425 nm for **3an** in DMSO. In general, the compounds have high dipole moments in excited state as compared to their corresponding ground state due to electronic excitations.²⁰ The high dipole moment of the compounds interacts with the environment of the polarity of solvents and results the slight shifts of emission maxima to longer wavelengths.²¹ The observed trend in photoluminescence characteristics are evidently showed that 3*H*-pyrrolo[1,2-*a*]indole derivatives have good tendency to photoluminescence properties.

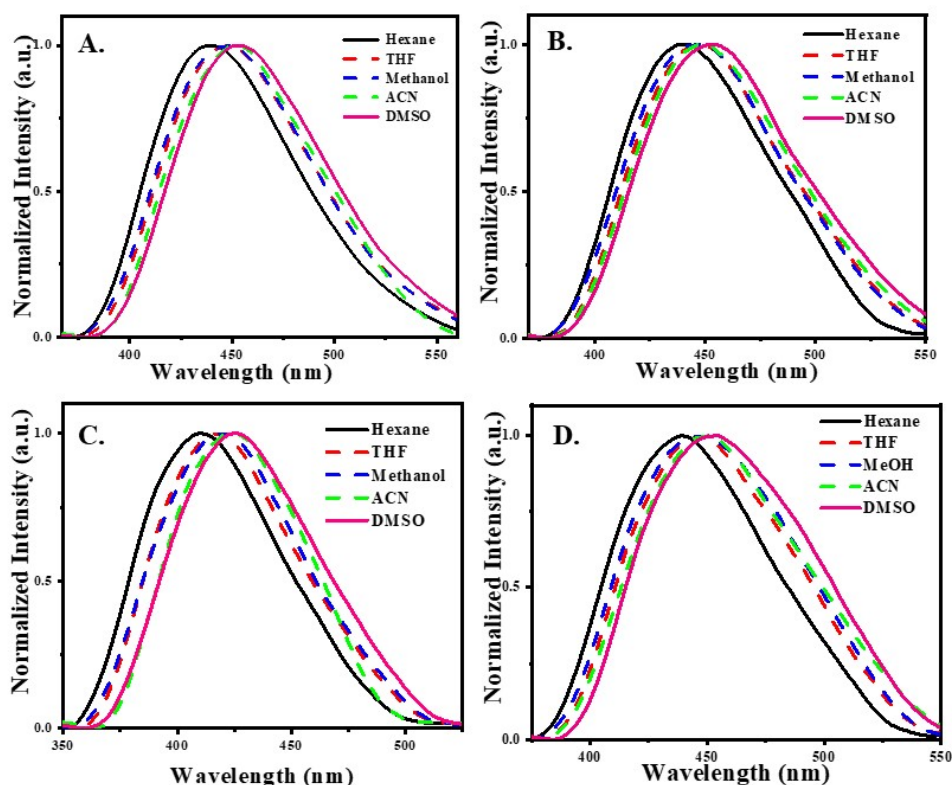
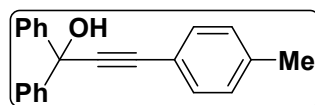
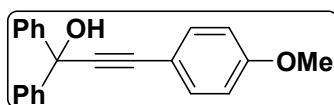


Figure S6 Normalized fluorescence emission spectra of compounds; **A. 3ai**, **B. 3at**, **C. 3an** and **D. 3aw** in different solvent medium.

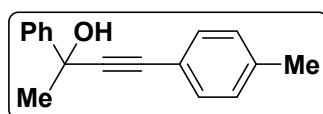
7. Spectroscopic data of synthesized compounds:



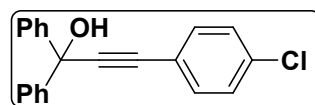
1,1-Diphenyl-3-(p-tolyl)prop-2-yn-1-ol (2b)³: Yield: 72%; ¹H NMR (CDCl₃, 400 MHz): δ 7.72 (t, J = 7.2 Hz, 4H), 7.44 (d, J = 8 Hz, 2H), 7.40-7.36(m, 4H), 7.33-7.29 (m, 2H), 7.18 (d, J = 8.0 Hz, 2H), 2.93 (s, 1H), 2.39 (s, 3H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 145.2, 139.0, 131.8, 129.2, 128.4, 127.8, 126.2, 119.4, 91.0, 87.5, 74.9, 21.7.



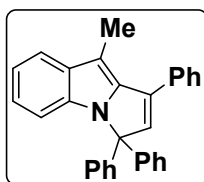
3-(4-Methoxyphenyl)-1,1-diphenylprop-2-yn-1-ol (2c)³: Yield: 74%; ¹H NMR (CDCl₃, 400 MHz): δ 7.60 (d, J = 8.0 Hz, 4H), 7.37 (t, J = 6.8 Hz, 2H), 7.27 (t, J = 7.6 Hz, 4H), 7.21-7.17 (m, 2H), 6.78 (d, J = 8.8 Hz, 2H), 3.73 (s, 3H), 2.85 (s, 1H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 160.0, 145.3, 133.4, 128.4, 127.8, 126.2, 114.5, 114.0, 90.4, 87.3, 75.0, 55.4.



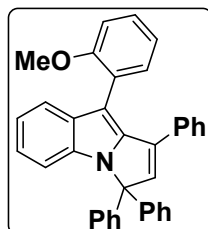
2-Phenyl-4-(p-tolyl)but-3-yn-2-ol (2d)⁴: Yield: 76%; ¹H NMR (CDCl₃, 400 MHz): δ 7.75-7.73 (m, 2H), 7.41-7.37 (m, 4H), 7.32 (t, J = 7.6 Hz, 1H), 7.14 (d, J = 8.0 Hz, 2H), 2.52 (s, 1H), 2.36 (s, 3H), 1.87 (s, 3H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 145.9, 138.8, 131.7, 129.2, 128.4, 127.8, 125.1, 119.5, 91.8, 85.2, 70.5, 33.5, 21.6.



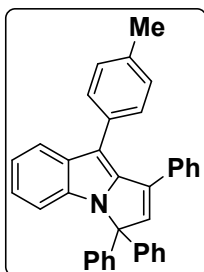
1,1-Diphenyl-3-(p-tolyl)prop-2-yn-1-ol (2e)⁵: Yield: 71%; ¹H NMR (CDCl₃, 400 MHz): δ 7.67 (t, J = 7.2 Hz, 4H), 7.44 (d, J = 8.4 Hz, 2H), 7.38-7.34 (m, 4H), 7.32-7.28 (m, 4H), 2.92 (s, 1H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 144.9, 134.9, 133.1, 128.8, 128.5, 128.0, 126.1, 121.0, 92.7, 86.2, 74.9.



9-Methyl-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3aa)²²: Yield: 84%; 333 mg; white solid; mp 120-122 °C; R_f = 0.60 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.68 (t, J = 6.8 Hz, 2H), 7.61 (d, J = 8 Hz, 1H), 7.48-7.42 (m, 3H), 7.35-7.26 (m, 10H), 7.06-6.94 (m, 3H), 6.67 (s, 1H), 2.37 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 140.5, 140.2, 139.6, 136.4, 134.4, 134.1, 133.8, 128.6, 128.4, 128.3, 127.4, 121.9, 119.7, 118.8, 110.5, 103.0, 75.5, 9.7.

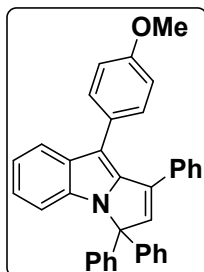


9-(2-Methoxyphenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ab): Yield: 77%; 377 mg; white solid; mp 180-182 °C; R_f = 0.50 (petroleum ether/EtOAc = 90/10); ^1H NMR (CDCl_3 , 400 MHz): δ 7.70-7.68 (m, 1H), 7.55-7.53 (m, 1H), 7.39-7.21 (m, 14H), 7.16-7.01 (m, 6H), 6.87 (s, 1H), 6.67 (d, J = 8 Hz, 1H), 3.16 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 157.1, 141.1, 140.0, 139.9, 139.6, 136.9, 134.4, 133.6, 132.8, 131.9, 128.7, 128.7, 128.3, 128.0, 127.8(3C), 127.4, 123.5, 121.9, 120.8, 120.1, 119.5, 110.6, 110.5, 104.7, 75.7, 54.4; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{36}\text{H}_{28}\text{NO}]^+$: 490.2165; Found : 490.2169.

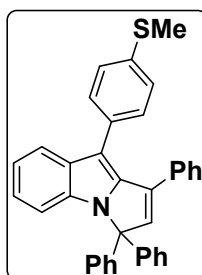


1,3,3-Triphenyl-9-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3ac): Yield: 83%; 393 mg; white solid; mp 134-136 °C; R_f = 0.6 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.96-7.94 (m, 1H), 7.53-7.49 (m, 9H), 7.48-7.44 (m, 3H), 7.41-7.36 (m, 3H), 7.33-7.29 (m, 2H), 7.26-7.20 (m, 5H), 7.00 (s, 1H), 2.54 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ

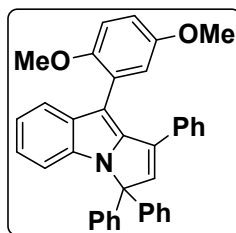
140.7, 139.7, 139.7, 136.2, 135.7, 134.3, 132.7, 132.5, 131.2, 130.7, 128.7, 128.6, 128.4, 128.3, 128.2, 127.9, 122.3, 120.7, 119.6, 110.7, 110.3, 75.6, 21.3; HRMS (ESI-TOF) m/z: $[M+H]^+$ Calculated for $[C_{36}H_{28}N]^+$: 474.2222; Found : 474.2209.



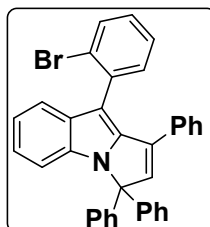
9-(4-Methoxyphenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ad): Yield: 85%; 416 mg; off white solid; mp 182-184 °C; R_f = 0.50 (petroleum ether/EtOAc = 90/10); 1H NMR ($CDCl_3$, 400 MHz): δ 7.65-7.63 (m, 1H), 7.24-7.7.20 (m, 9H), 7.19-7.16 (m, 3H), 7.12-7.10 (m, 3H), 7.03 (t, J = 8 Hz, 2H), 6.97-6.94 (m, 3H), 6.70 (s, 1H), 6.68-6.66 (m, 2H), 3.70 (s, 3H); $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 158.2, 140.6, 139.7, 139.6, 136.1, 134.2, 132.7, 132.5, 131.8, 128.7, 128.4, 128.3, 128.2, 127.9(2C), 126.6, 122.3, 120.6, 119.5, 113.4, 110.6, 109.9, 75.6, 55.4; HRMS (ESI-TOF) m/z: $[M+H]^+$ Calculated for $[C_{36}H_{28}NO]^+$: 490.2165; Found : 490.2160.



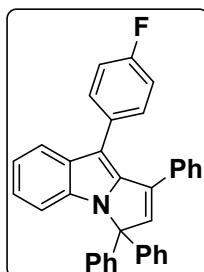
9-(4-(Methylthio)phenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ae): Yield: 80%; 405 mg; off white solid; mp 184-186 °C; R_f = 0.65 (petroleum ether/EtOAc = 90/10); 1H NMR ($CDCl_3$, 400 MHz): δ 7.81-7.79 (m, 1H), 7.40-7.35 (m, 9H), 7.34-7.24 (m, 6H), 7.21-7.15 (m, 4H), 7.13-7.09 (m, 3H), 6.87 (s, 1H), 2.53 (s, 3H); $^{13}C\{^1H\}$ NMR ($CDCl_3$, 100 MHz): δ 141.0, 139.9, 139.5, 136.0, 135.8, 134.3, 132.6, 132.2, 131.2, 131.2, 130.2, 128.7, 128.4, 128.3, 128.0, 127.9, 126.4, 122.4, 120.5, 119.7, 110.7, 109.6, 75.8, 16.38; HRMS (ESI-TOF) m/z: $[M+H]^+$ Calculated for $[C_{36}H_{28}NS]^+$: 506.1973; Found : 506.1967.



9-(2,5-Dimethoxyphenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3af): Yield: 80%; 416 mg; white solid; mp 214-216 °C; R_f = 0.3 (petroleum ether/EtOAc = 90/10); ^1H NMR (CDCl_3 , 400 MHz): δ 7.74-7.72 (m, 1H), 7.68-7.65 (m, 2H), 7.39-7.31 (m, 13H), 7.26-7.22 (m, 1H), 7.18-7.15 (m, 2H), 7.09-7.07 (m, 3H), 6.88 (s, 1H), 6.85-6.82 (m, 1H), 6.63(d, J = 8 Hz, 1H), 3.74 (s, 3H), 3.17 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 153.1, 151.4, 141.1, 140.2, 139.8, 139.6, 136.8, 134.3, 133.5, 132.6, 128.7(2C), 128.3, 127.9, 127.8(2C), 127.5, 124.1, 122.0, 120.9, 119.5, 117.4, 113.2, 111.5, 110.6, 104.8, 75.7, 55.8, 55.0; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{37}\text{H}_{30}\text{NO}_2]^+$: 520.2271; Found : 520.2303.

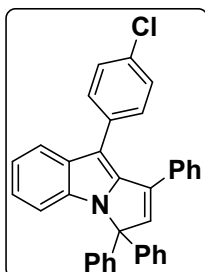


9-(2-Bromophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ag): Yield: 77%; 415 mg; off white solid; mp 150-152 °C; R_f = 0.50 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.49-7.47 (m, 1H), 7.42-7.340 (m, 1H), 7.38-7.36 (m, 1H), 7.33 (d, J = 4.4 Hz, 4H), 7.30-7.260 (m, 6H), 7.21-7.18 (m, 3H), 7.15-7.08 (m, 2H), 7.04-7.00 (m, 5H) 6.83 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.1, 140.4, 139.6, 139.4, 136.1, 135.8, 133.9, 133.1, 132.6(2C), 128.8, 128.7, 128.6, 128.4, 128.2, 128.1, 127.9(2C), 127.8, 127.7, 126.9, 126.4, 122.2, 120.7, 119.6, 110.7, 108.7, 75.9; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{BrN}]^+$: 538.1165; Found : 538.1167.

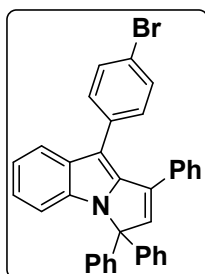


9-(4-Fluorophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ah): Yield: 76%; 363 mg; off white solid; mp 198-200 °C; R_f = 0.4 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.76-7.74 (m, 1H), 7.44-7.34 (m, 10H), 7.31-7.26 (m, 5H), 7.20 (t, J =

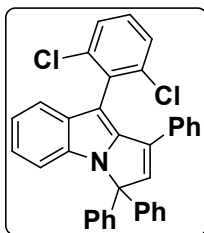
7.6 Hz, 2H), 7.15-7.08 (m, 3H), 6.95 (t, $J = 8.8$ Hz, 2H), 6.87 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 161.6 (d, $J = 243$ Hz), 141.0, 140.0, 139.5, 135.9, 134.2, 132.5, 132.2 (d, $J = 9$ Hz), 130.2, 128.8, 128.3(2C), 128.0(2C), 122.4, 120.0 (d, $J = 56$ Hz), 114.7 (d, $J = 22$ Hz), 109.9 (d, $J = 171$ Hz), 75.8; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{FN}]^+$: 478.1971; Found : 478.1954.



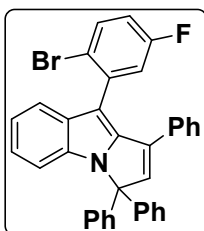
9-(4-Chlorophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ai): Yield: 80%, 395 mg; off white solid; mp 208-210 °C; $R_f = 0.50$ (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.77-7.75 (m, 1H), 7.42-7.35 (m, 10H), 7.30-7.28 (m, 3H), 7.24-7.20 (m, 6H), 7.13 (t, $J = 3.6$ Hz, 1H), 7.12-7.09 (m, 2H), 6.87 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.3, 140.1, 139.4, 135.9, 134.2, 132.7, 132.5, 132.0(2C), 131.9, 128.8, 128.4(2C), 128.3, 128.1, 128.0, 122.5, 120.3, 119.9, 110.8, 108.9, 75.9; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{ClN}]^+$: 494.1670; Found : 494.1638.



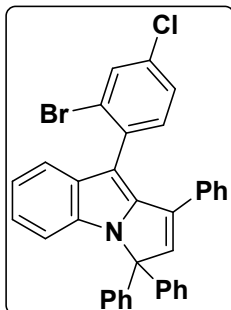
9-(4-Bromophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3aj): Yield: 62%; 334 mg; radish solid; mp 216-218 °C; $R_f = 0.60$ (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.72-7.70 (m, 1H), 7.37-7.32 (m, 6H), 7.31-7.29 (m, 5H), 7.27-7.24 (m, 4H), 7.18-7.12 (m, 4H), 7.09-7.04 (m, 3H), 6.83 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.4, 140.1, 139.4, 135.9, 134.3, 133.2, 132.5, 132.3, 132.0, 131.0, 128.8, 128.5, 128.4, 128.3, 128.1, 128.0, 122.5, 120.3, 120.1, 119.9, 110.9, 108.9, 75.9; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{BrN}]^+$: 338.1165; Found : 338.1156.



9-(2,6-Dichlorophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ak): Yield: 77%; 407 mg; off white solid; mp 128-130 °C; R_f = 0.45 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.33-7.28 (m, 10H), 7.24-7.21 (m, 5H), 7.14-7.10 (m, 2H), 7.06-7.00 (m, 5H), 6.84 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 140.4, 139.4, 137.3, 136.0, 134.3, 133.1, 132.5, 132.0, 129.0, 128.8, 128.3, 128.2, 128.0, 127.8, 127.7, 127.4, 122.1, 120.8, 119.7, 110.9, 103.9, 76.2; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{24}\text{Cl}_2\text{N}]^+$: 528.1280; Found : 528.1254.

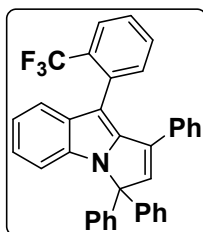


9-(2-Bromo-5-fluorophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3al): Yield: 80%; 445 mg; off white solid; mp 152-154 °C; R_f = 0.35 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.51-7.47 (m, 2H), 7.42-7.32 (m, 10H), 7.29 (d, J = 7.2 Hz, 2 H), 7.24-7.19 (m, 2H), 7.16-7.09 (m, 5H), 6.94-6.90 (m, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 161.6 (d, J = 245 Hz), 141.1 (d, J = 53 Hz), 139.4, 139.1, 137.8 (d, J = 9 Hz), 135.9, 133.9, 133.7 (2C), 132.4 (d, J = 36 Hz), 128.8 (2C), 128.6, 128.3, 128.2, 128.0 (2C), 127.9, 127.6, 122.4, 120.7 (2C), 120.5, 119.9 (2C), 119.7, 115.7 (d, J = 22 Hz), 110.8, 107.6, 76.1; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{24}\text{BrFN}]^+$: 556.1071; Found : 556.1054.

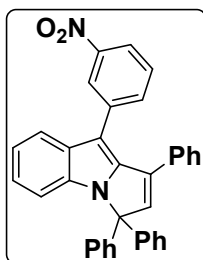


9-(2-Bromo-4-chlorophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3am): Yield: 71%; 406.7 mg; light yellow solid; mp 110-112 °C; R_f = 0.50 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.56 (d, J = 2 Hz, 1H), 7.43-7.41 (m, 1H), 7.39-7.36 (m, 5H),

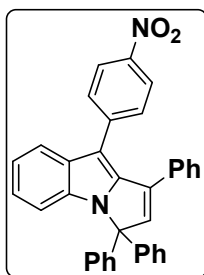
7.35-7.33 (m, 3H), 7.31-7.29 (m, 2H), 7.27 (s, 1H), 7.25-7.21 (m, 4H), 7.14-7.06 (m, 5H), 6.88 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.4, 140.7, 139.5, 139.3, 135.9, 134.6, 134.0, 133.7, 133.4, 132.5, 132.4, 132.3, 128.8(2C), 128.3, 128.2, 128.0(3C), 127.7, 127.2, 126.7, 122.3, 120.5, 119.8, 110.9, 107.4, 76.1; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{24}\text{BrClN}]^+$: 572.0775; Found : 572.0775.



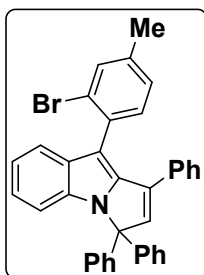
1,3,3-Triphenyl-9-(2-(trifluoromethyl)phenyl)-3H-pyrrolo[1,2-a]indole (3an): Yield: 76%; 401 mg; off white solid; mp 174-176 °C; R_f = 0.20 (petroleum ether/EtOAc = 90/10); ^1H NMR (CDCl_3 , 400 MHz): δ 7.75 (d, J = 8 Hz, 1H), 7.67-7.62 (m, 2H), 7.56-7.38 (m, 12H), 7.31 (t, J = 7.6 Hz, 2H), 7.23 (d, J = 7.6 Hz, 1H), 7.17-7.12 (m, 5H), 7.98 (t, J = 1.6 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 140.3 (q, J = 239 Hz), 135.9, 134.1, 134.0, 133.9, 133.7, 132.5, 131.3, 131.2, 131.0, 130.7, 128.8, 128.7, 128.4, 127.9 (q, J = 14 Hz), 127.6, 127.5, 126.1 (q, J = 15 Hz), 125.4, 123.0, 122.7, 122.2, 120.6, 119.9, 119.6, 110.6, 106.0, 75.8; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{36}\text{H}_{25}\text{F}_3\text{N}]^+$: 528.1934; Found : 528.1938.



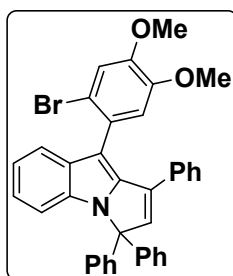
9-(3-Nitrophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ao): Yield: 79%; 399 mg; yellow solid; mp 246-248 °C; R_f = 0.40 (petroleum ether/EtOAc = 85/15); ^1H NMR (CDCl_3 , 400 MHz): δ 8.10 (s, 1H), 8.06 (d, J = 8.4 Hz, 1H), 7.80 (d, J = 8 Hz, 1H), 7.65 (d, J = 7.6 Hz, 1H), 7.42-7.34 (m, 11H), 7.27-7.25 (m, 3H), 7.19-7.13 (m, 5H), 6.91 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 147.8, 142.1, 141.0, 139.0, 136.3, 135.9, 135.5, 134.3, 132.3, 131.4, 128.8, 128.6, 128.4, 128.2(2C), 128.1, 125.4, 122.9, 120.7, 120.4, 119.8, 111.1, 107.5, 76.2; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{N}_2\text{O}_2]^+$: 505.1911; Found : 505.1939.



9-(4-Nitrophenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ap): Yield: 80%; 404 mg; yellow solid; mp 247-249 °C; R_f = 0.40 (petroleum ether/EtOAc = 85/15); ^1H NMR (CDCl_3 , 400 MHz): δ 8.06 (d, J = 8 Hz, 2H), 7.80 (d, J = 8 Hz, 1H), 7.43-7.34 (m, 12H), 7.31-7.26 (m, 3H), 7.21-7.12 (m, 5H), 6.93 (d, J = 1.6 Hz, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 145.7, 142.7, 141.5, 141.3, 138.8, 135.5, 134.3, 132.4, 131.3, 130.9, 128.9, 128.8, 128.3(2C), 128.2(2C), 123.1, 123.0, 120.5, 120.0, 111.2, 108.0, 76.3; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{35}\text{H}_{25}\text{N}_2\text{O}_2]^+$: 505.1911; Found : 505.1933.

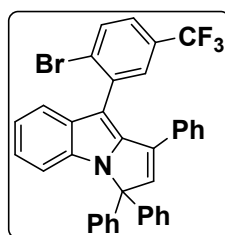


9-(2-Bromo-4-methylphenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3aq): Yield: 72%; 398 mg; off white solid; mp 88-90 °C; R_f = 0.50 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.47-7.45 (m, 1H), 7.40-7.38 (m, 5H), 7.36-7.31 (m, 6H), 7.29-7.26 (m, 3H), 7.22-7.18 (m, 1H), 7.11-7.06 (m, 6H), 6.88 (d, J = 1.2 Hz, 1H), 2.38 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.1, 140.3, 139.8, 139.5, 138.7, 136.2, 134.0, 133.1, 132.8, 132.7, 128.8, 128.7, 128.4, 128.3, 128.1, 127.9, 127.8, 127.7, 126.1, 122.1, 120.8, 119.5, 110.7, 108.6, 75.9, 20.9; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{36}\text{H}_{27}\text{BrN}]^+$: 552.1321; Found : 552.1299.



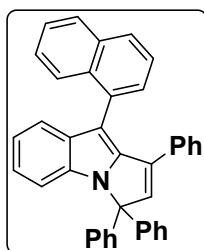
9-(2-Bromo-4,5-dimethoxyphenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3ar): Yield: 78%; 500 mg; brown solid; mp 241-243 °C; R_f = 0.60 (petroleum ether/EtOAc = 85/15); ^1H

NMR (CDCl₃, 400 MHz): δ 7.57-7.53 (m, 1H), 7.40-7.28 (m, 12H), 7.26-7.22 (m, 1H), 7.15-7.07 (m, 6H), 7.92-6.85 (m, 2H), 3.93 (s, 3H), 3.67 (s, 3H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 148.7, 147.9, 141.0, 140.4, 139.6, 139.4, 136.1, 133.9, 132.7, 132.4, 128.8, 128.7, 128.4, 128.2, 127.9(2C), 127.8, 127.7, 122.2, 121.0, 119.6, 116.1, 115.7, 115.4, 110.8, 108.7, 75.9, 56.3, 56.0; HRMS (ESI-TOF) m/z: [M+H]⁺ Calculated for [C₃₇H₂₉BrNO₂]⁺: 598.1376; Found: 598.1386.

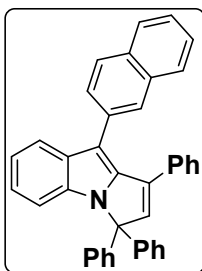


9-(2-Bromo-5-(trifluoromethyl)phenyl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3as):

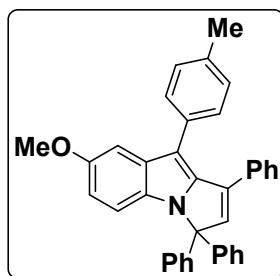
Yield: 80%; 485 mg; grey solid; mp 102-104 °C; *R_f* = 0.60 (petroleum ether/EtOAc = 85/15); ¹H NMR (CDCl₃, 400 MHz): δ 7.68 (d, *J* = 8.4 Hz, 1H), 7.61 (s, 1H), 7.46 (t, *J* = 5.2 Hz, 1H), 7.40-7.34 (m, 11H), 7.20 (d, *J* = 7.6 Hz, 3H), 7.12-7.07 (m, 5H), 6.91 (s, 1H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 141.7, 141.0, 139.3, 139.1, 136.9, 135.8, 132.9 (q, *J* = 202 Hz), 130.0 (q, *J* = 13 Hz), 129.6, 129.2, 128.8, 128.3(2C), 128.3 (q, *J* = 28 Hz), 127.7, 125.2, 125.0, 124.9, 122.5, 120.6, 119.9, 111.0, 107.2, 76.3; HRMS (ESI-TOF) m/z: [M+H]⁺ Calculated for [C₃₆H₂₄BrF₃N]⁺: 606.1039; Found: 606.1021.



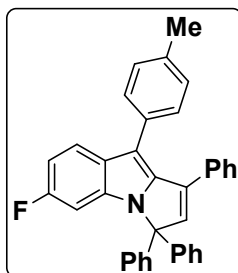
9-(Naphthalen-1-yl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3at): Yield: 73%; 372 mg; radish solid; mp 182-184 °C *R_f* = 0.80 (petroleum ether/EtOAc = 95/05); ¹H NMR (CDCl₃, 400 MHz): δ 7.89 (d, *J* = 8 Hz, 1H), 7.85 (d, *J* = 8 Hz, 1H), 7.80 (d, *J* = 8 Hz, 1H), 7.42-7.40 (m, 5H), 7.38-7.30 (m, 9H), 7.29-7.25 (m, 1H), 7.13-6.97 (m, 6H), 6.87 (s, 1H), 6.82 (t, *J* = 7.6 Hz, 2H); ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 141.4, 140.3, 139.8, 139.7, 136.1, 134.2, 133.8, 133.6, 133.3, 132.2, 129.6, 128.8(2C), 128.4, 128.3, 128.0(2C), 127.9, 127.5, 127.3, 126.9, 125.6(2C), 125.2, 122.2, 121.3, 119.5, 110.8, 107.8, 75.9; HRMS (ESI-TOF) m/z: [M+H]⁺ Calculated for [C₃₉H₂₈N]⁺: 510.2216; Found: 510.2221.



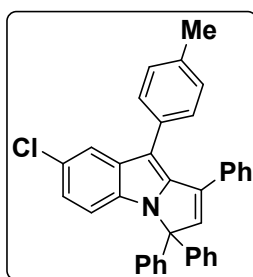
9-(Naphthalen-2-yl)-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3au): Yield: 60%; 306 mg; off white solid; mp 224-226 °C; R_f = 0.80 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.87-7.85 (m, 1H), 7.81-7.79 (m, 1H), 7.73 (s, 1H), 7.65 (d, J = 8 Hz, 1H), 7.59-7.57 (m, 1H), 7.44-7.41 (m, 3H), 7.37-7.33 (m, 9H), 7.27-7.25 (m, 3H), 7.14-7.07 (m, 4H), 7.02-6.98 (m, 2H), 6.85 (s, 1H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.2, 140.2, 139.5, 136.2, 134.4, 133.4, 132.7, 132.3, 132.0, 131.7, 129.5, 129.3, 128.8, 128.4, 128.4, 128.3, 128.1, 127.9, 127.6, 127.1, 125.8, 125.4, 122.5, 120.7, 119.8, 110.8, 110.2, 104.8, 75.8; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{39}\text{H}_{28}\text{N}]^+$: 510.2216; Found: 510.2211.



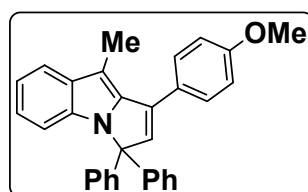
7-Methoxy-1,3,3-triphenyl-9-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3av): Yield: 76%; 383 mg; yellow solid; mp 138-140 °C; R_f = 0.50 (petroleum ether/EtOAc = 90/10); ^1H NMR (CDCl_3 , 400 MHz): δ 7.28-7.19 (m, 10H), 7.17-7.12 (m, 4H), 7.08 (d, J = 8 Hz, 2H), 7.03 (t, J = 7.6, 2H), 6.95 (d, J = 7.6 Hz, 2H), 6.84 (d, J = 8.8 Hz, 1H), 6.71 (s, 1H), 6.65-6.62 (m, 1H), 3.69 (s, 3H), 2.27 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 154.2, 140.6, 140.3, 139.7, 136.2, 135.6, 132.8, 132.7, 131.3, 130.6, 129.5, 128.7, 128.6, 128.4, 128.3, 128.1, 127.9, 112.3, 111.3, 109.9, 102.5, 75.6, 55.9, 21.4; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{37}\text{H}_{30}\text{NO}]^+$: 504.2322; Found: 504.2353.



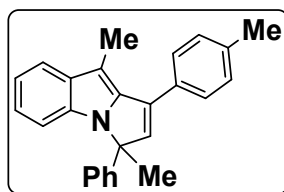
6-Fluoro-1,3,3-triphenyl-9-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3aw): Yield: 79%; 388 mg; off white solid; mp 138-140 °C; R_f = 0.40 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.75-7.72 (m, 1H), 7.44-7.39 (m, 9H), 7.35-7.29 (m, 3H), 7.27-7.18 (m, 5H), 7.11(d, 2H), 6.93-6.91 (m, 1H), 6.87 (s, 1H), 6.84-6.81 (m, 1H), 2.43 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 160.0 (d, J = 237 Hz), 141.2, 140.4, 140.0, 139.3, 139.2, 136.0 (d, J = 13 Hz), 134.2, 132.5, 131.3, 130.8, 130.6, 129.0, 128.8(2C), 128.6, 128.4, 128.2(2C), 128.0, 127.9, 127.8, 121.4 (d, J = 10 Hz), 110.4, 108.1 (d, J = 24 Hz), 97.1 (d, J = 27 Hz), 97.0, 75.7, 21.3;; ^{19}F NMR (CDCl_3 , 376 MHz): δ -119.7; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{36}\text{H}_{27}\text{FN}]^+$: 492.2122; Found : 492.2145.



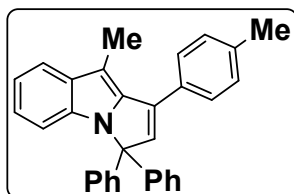
7-Chloro-1,3,3-triphenyl-9-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3ax): Yield: 73%; 371 mg; white solid; mp 220-222 °C; R_f = 0.50 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.69 (t, J = 9.2 Hz, 13H), 7.34-7.22 (m, 13H), 7.11 (d, J = 6 Hz, 4H), 7.02-7.6.96 (m, 3H), 6.94-6.89 (m, 3H), 6.84 (s, 1H), 2.34 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.2, 141.0, 139.4, 136.1, 133.6, 132.6, 132.4, 130.6, 130.5, 128.4, 128.7, 128.4, 128.3, 128.2, 128.1, 128.0, 125.6, 122.6, 120.2, 111.5, 110.0, 75.9, 21.3; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{36}\text{H}_{27}\text{ClN}]^+$: 508.1827; Found : 508.1853.



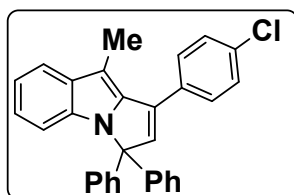
1-(4-Methoxyphenyl)-9-methyl-3,3-diphenyl-3H-pyrrolo[1,2-a]indole (3ay): Yield: 76%; 325 mg; off white solid; mp 184-186 °C; R_f = 0.50 (petroleum ether/EtOAc = 90/10); ^1H NMR (CDCl_3 , 400 MHz): δ 7.60 (d, J = 8.8 Hz, 3H), 7.33-7.24 (m, 10H), 7.05-6.93 (m, 5H), 6.59 (s, 1H), 3.87 (s, 3H), 2.37 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 159.9, 140.8, 140.3, 138.6, 135.9, 134.4, 134.1, 129.6, 128.6, 128.2, 127.7, 126.1, 121.8, 119.6, 118.7, 113.9, 110.5, 102.8, 75.4, 55.5, 9.8; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{31}\text{H}_{26}\text{NO}]^+$: 428.2009; Found : 428.2036.



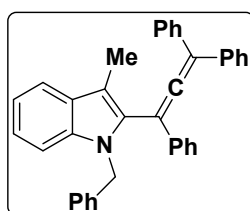
3,9-Dimethyl-3-phenyl-1-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3az): Yield: 74%; 259 mg; off white solid; mp 122-124 °C; R_f = 0.70 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.68-7.66 (m, 1H), 7.53 (d, J = 8 Hz, 1H), 7.34-7.26 (m, 5H), 7.20 (d, J = 7.6 Hz, 2H), 7.13-7.08 (m, 2H), 7.02-7.00 (m, 1H), 6.38 (s, 1H), 2.44 (s, 3H), 2.39 (s, 3H), 2.01 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 141.7, 141.1, 140.1, 138.2, 135.6, 134.0, 133.4, 131.0, 129.2, 128.8, 128.2, 127.4, 126.0, 121.6, 119.7, 118.6, 109.9, 101.9, 68.5, 24.0, 21.4, 9.7; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{26}\text{H}_{24}\text{N}]^+$: 350.1903; Found : 350.1906.



9-Methyl-3,3-diphenyl-1-(p-tolyl)-3H-pyrrolo[1,2-a]indole (3ba): Yield: 68%; 280 mg; off white solid; mp 141-143 °C; R_f = 0.55 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.63-7.57 (m, 3H), 7.36-7.27 (m, 12H), 7.09-7.00 (m, 2H), 6.96 (d, J = 7.6 Hz, 1H), 6.65 (s, 1H), 2.43 (s, 3H), 2.39 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 140.6, 140.2, 139.1, 138.4, 136.2, 134.3, 134.1, 130.8, 129.2, 128.6, 128.2, 127.7, 121.8, 119.6, 118.7, 110.4, 102.9, 75.4, 21.5, 9.8; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{31}\text{H}_{26}\text{N}]^+$: 412.2060; Found : 412.2073.



1-(4-Chlorophenyl)-9-methyl-3,3-diphenyl-3H-pyrrolo[1,2-a]indole (3bb): Yield: 76%; 328 mg; white solid; mp 177-179 °C; R_f = 0.6 (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.58 (d, J = 8.4 Hz, 3H), 7.41 (d, J = 8.4 Hz, 2H), 7.32-7.27 (m, 6H), 7.23 (t, J = 6.4 Hz, 4H), 7.04-6.98 (m, 2H), 6.92 (d, J = 7.6 Hz, 1H), 6.64 (s, 1H), 2.33 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 140.0, 139.9, 139.8, 135.2, 134.4, 134.0, 132.2, 129.7, 128.8, 128.7, 128.2, 127.8, 122.1, 119.8, 118.9, 110.5, 103.1, 75.5, 9.73; HRMS (ESI-TOF) m/z : $[\text{M}+\text{H}]^+$ Calculated for $[\text{C}_{30}\text{H}_{23}\text{ClN}]^+$: 432.1514; Found: 432.1505.



1-Benzyl-3-methyl-2-(1,3,3-triphenylpropa-1,2-dien-1-yl)-1H-indole (5): Yield: 87%; 424 mg; off white gummy; $R_f = 0.70$ (petroleum ether/EtOAc = 95/05); ^1H NMR (CDCl_3 , 400 MHz): δ 7.60 (t, $J = 5.2$ Hz, 1H), 7.27-7.25 (m, 4H), 7.22-7.18 (m, 11H), 7.14-7.10 (m, 3H), 7.07-7.02 (m, 3H), 6.73 (d, $J = 6.4$ Hz, 1H), 5.10 (s, 2H), 2.13 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (CDCl_3 , 100 MHz): δ 209.6, 138.1, 137.2, 135.8, 135.7, 130.9, 128.9, 128.8, 128.7, 128.5, 128.4, 127.8, 127.7, 126.9, 126.7, 126.1, 122.1, 119.3, 119.0, 112.8, 111.2, 110.3, 102.7, 47.6, 9.7.

7. Calculation of Green Chemistry Metrics (EcoScale and E-factor):

(a) Calculation of EcoScale indexes under ball milling

The penalty points for synthesis of coumarin derivatives for the synthesis of pyrrolo[1,2-*a*]indoles (**3**) under ball milling

Parameter	Penalty
1. Yields 60-85%	20-7.5
2. Price of reaction components	
Indoles	0
Propargyl alcohols	0
BAIL-1	0
3. Safety	
non-dangerous for environment, non-toxic, non-flammable	0
4. Technical setup	
Unconventional activation technique	2
5. Temperature/time	
Room temperature < 1 h	0
6. Workup and purification	
Classical chromatography	10
Penalty points total:	32-19.5

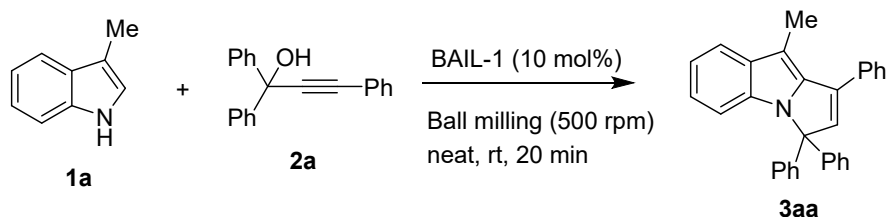
$$\text{EcoScale Score} = 100 - \text{Total penalty points} = 68-80.5$$

EcoScale Score for the synthesis of 9-methyl-1,3,3-triphenyl-3*H*-pyrrolo[1,2-*a*]indole (3aa) under the ball-milling conditions:

Parameter	Penalty
1. Yields 84%	8
2. Price of reaction components	
3-Methyl-1 <i>H</i> -indole (1a)	0
Propargyl alcohol (2a)	0
BAIL-1	0
3. Safety	
non-dangerous for environment, non-toxic, non-flammable	0
4. Technical setup	
Unconventional activation technique	2
5. Temperature/time	
Room temperature < 1 h	0
6. Workup and purification	
Classical chromatography	10
Penalty points total:	20

$$\begin{aligned}\text{EcoScale Score} &= 100 - \text{Total penalty points} \\ &= 80\end{aligned}$$

(c) Calculation of E-factor for the synthesis of 9-methyl-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole (3aa) under the ball-milling conditions:



E-factor calculation for the synthesis of 3aa under the ball-milling conditions:

Reactant 1 (1a):	3-Methyl-1H-indole	0.131 g	1 mmol	FW 131.18
Reactant 2 (2a):	1,1,3-Triphenylprop-2-yn-1-ol	0.284 g	1 mmol	FW 284.36
Reagent:	BAIL-1	0.039 g	0.1 mmol	FW 390
Solvent:	—	—	—	—
Auxiliary (grinding):	—	—	—	—
Product (3aa):	9-methyl-1,3,3-triphenyl-3H-pyrrolo[1,2-a]indole	0.333 g	0.84 mmol	FW 397.52

Product yield = 84%

$$\text{E-factor} = \frac{0.131 + 0.284 + (0.039 - 0.036) - (0.333)}{0.333} = 0.25 \text{ Kg waste/1 Kg product}$$

Note: (i) Calculations were done on 1 mmol scale. (ii) When the authors have not reported the amount of solvent used in the work-up and purification procedure, we have not accounted for solvent and considered that solvent can be recovered. The catalyst was recovered in 94% amount.

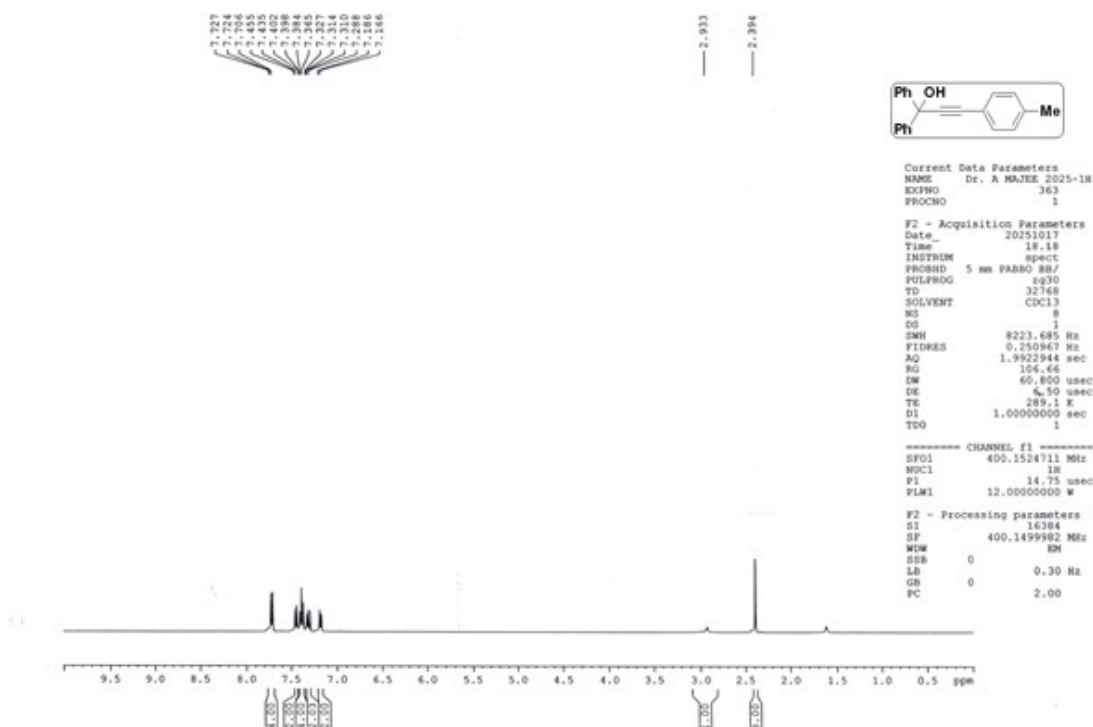
8. References:

1. M. Singsardar, A. Chakraborty, S. Jana and A. Hajra, *ChemistrySelect*, 2017, **2**, 8893–8897.
2. K. Sonogashira, Y. Tohda and N. Hagihara, *Tetrahedron Lett.*, 1975, **16**, 4467–4470.
3. X. Zhang, W. T. Teo and P. W. H. Chan, *Org. Lett.*, 2009, **11**, 4990–4993.
4. A. O. Shchukin, A. V. Vasil'ev, and E. V. Grinenko, *Russ. J. Org. Chem.*, 2010, **46**, 82–97.
5. A. Horita, H. Tsurugi, A. Funayama, T. Satoh and M. Miura, *Org. Lett.*, 2007, **9**, 2231–2233.
6. Z. Du., Z. Li. and Y. Deng, *Synth. Commun.*, 2005, **35**, 1343–1349.
7. I. A. Holder and S. T. Boyce, *Burns*, 1994, **20**, 426–429.

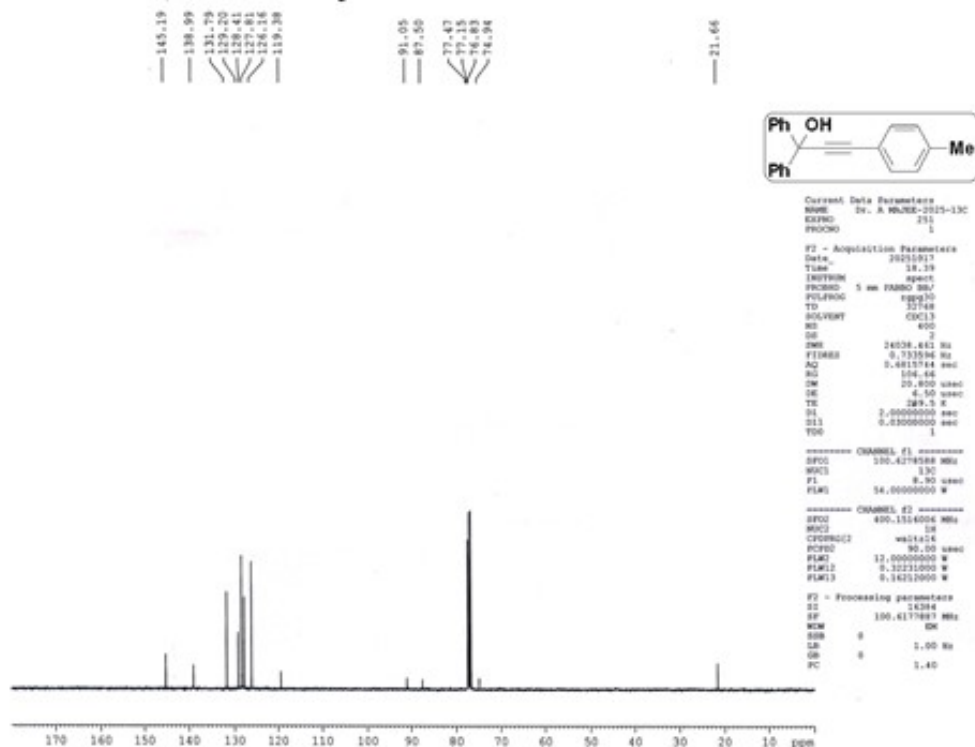
8. (a) I. Pal, S. Ghosh, S. Sadhu, A. Kundu, S. Majumdar, S. Kumar Yatirajula, J. Rath, P. Pratim Ray and B. Dey, *Inorganica Chim. Acta*, 2025, **574**, 122413; (b) S. Sarkar, R. Chatterjee, A. Mukherjee, D. Mukherjee, N. Chandra Mandal, S. Mahato, S. Santra, G. V. Zyryanov and A. Majee, *ACS Sustain. Chem. Eng.*, 2021, **9**, 5557–5569.
9. (a) P. C. Hannan, *Vet. Res.*, 2000, **31**, 373–395; (b) Ruangpan, L. Laboratory manual of standardized methods for antimicrobial sensitivity tests for bacteria isolated from aquatic animals and environment 2004, 31-55.
10. S. D. Sarker, L. Nahar and Y. Kumarasamy, *Methods*, 2007, **42**, 321–324.
11. S. Pal, S. Sarkar, A. Mukherjee, A. Kundu, A. Sen, J. Rath, S. Santra, G. V. Zyryanov and A. Majee, *Green Chem.*, 2023, **25**, 9847–9856.
12. M. Hartmann, M. Berditsch, J. Hawecker, M. F. Ardakani, D. Gerthsen and A. S. Ulrich, *Antimicrob. Agents Chemother.*, 2010, **54**, 3132–3142.
13. D. Carta, A. Balasso, P. Caliceti and M. G. Ferlin, *ChemMedChem*, 2015, **10**, 1846–1862.
14. E. P. Kirby and R. F. Steiner, *J. Phys. Chem.*, 1970, **74**, 4480–4490.
15. B. Ojha, K. Laxman and M. Ravikanth, *Asian J. Org. Chem.*, 2021, **10**, 857–867.
16. Z. Gao, Y. Hao, M. Zheng and Y. Chen, *RSC Adv.*, 2017, **7**, 7604–7609.
17. R. Hernández-Ruiz, R. Rubio-Presa, S. Suárez-Pantiga, M. R. Pedrosa, M. A. Fernández-Rodríguez, M. J. Tapia and R. Sanz, *Chemistry*, 2021, **27**, 13613–13623.
18. N. Sharma, S. K. Jain and R. C. Rastogi, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 2007, **66**, 171–176.
19. K. Pal, T. Dutta and A. L. Koner, *ACS Omega*, 2021, **6**, 28–37.
20. N. Sharma, S. K. Jain and R. C. Rastogi, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 2007, **66**, 171–176.
21. K. Pal, T. Dutta and A. L. Koner, *ACS Omega*, 2021, **6**, 28–37.
22. L. Hao, Y. Pan, T. Wang, M. Lin, L. Chen and Z.-P. Zhan, *Adv. Synth. Catal.*, 2010, **352**, 3215–3222.

9. ^1H , and ^{13}C NMR spectra of synthesized compounds:

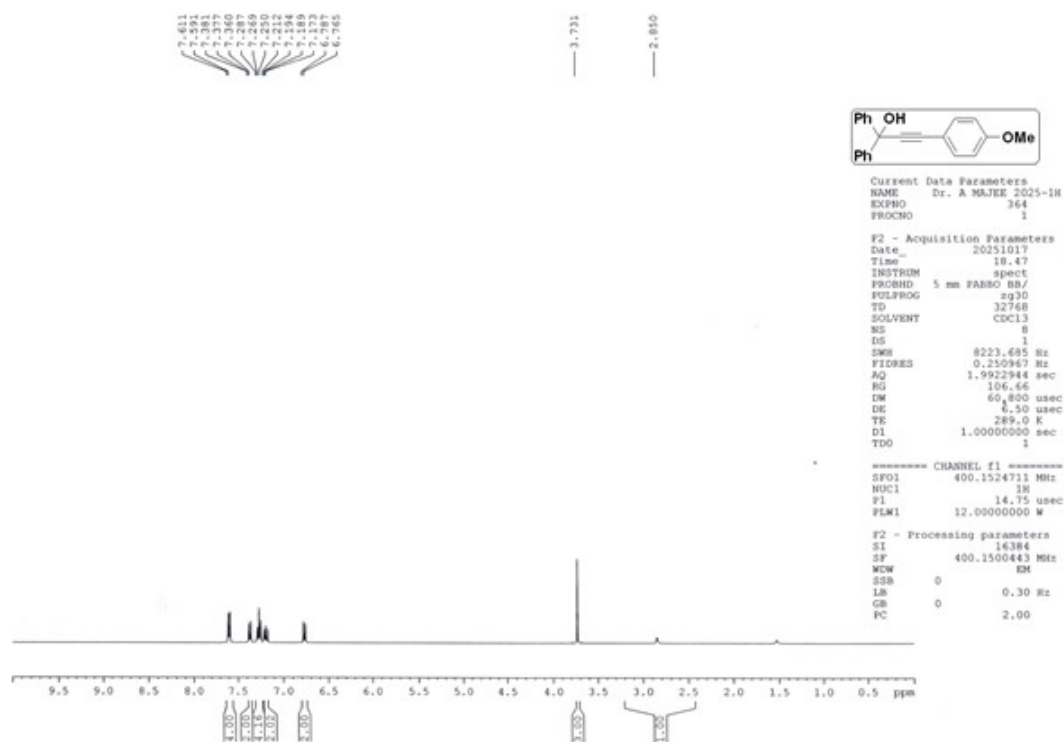
^1H NMR : 400 MHz, Solvent : CDCl_3



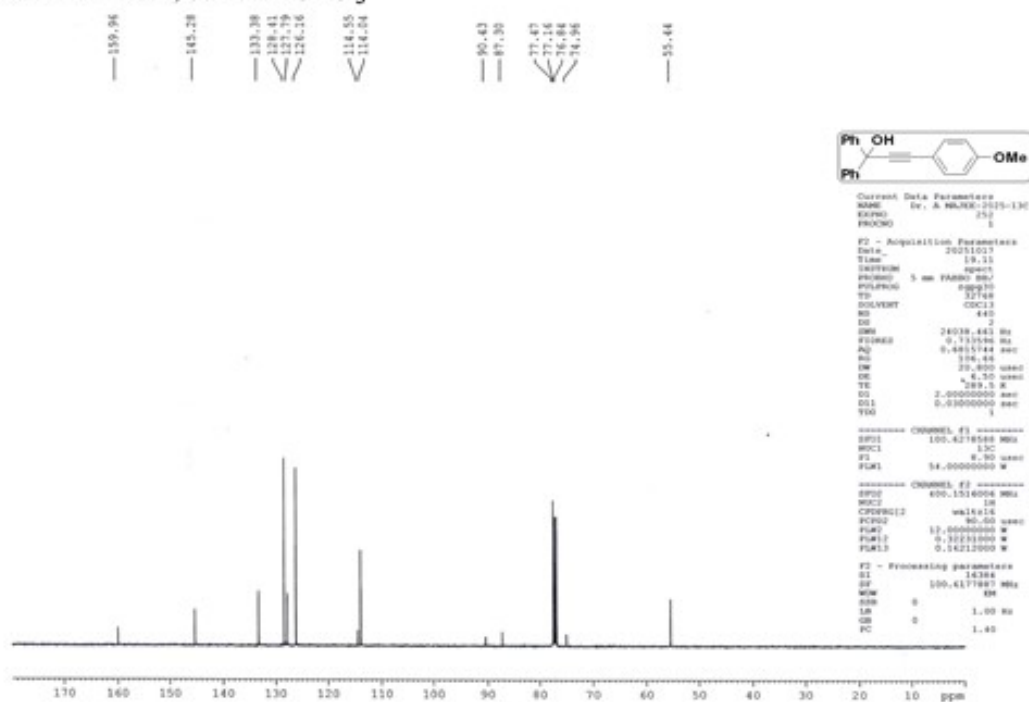
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



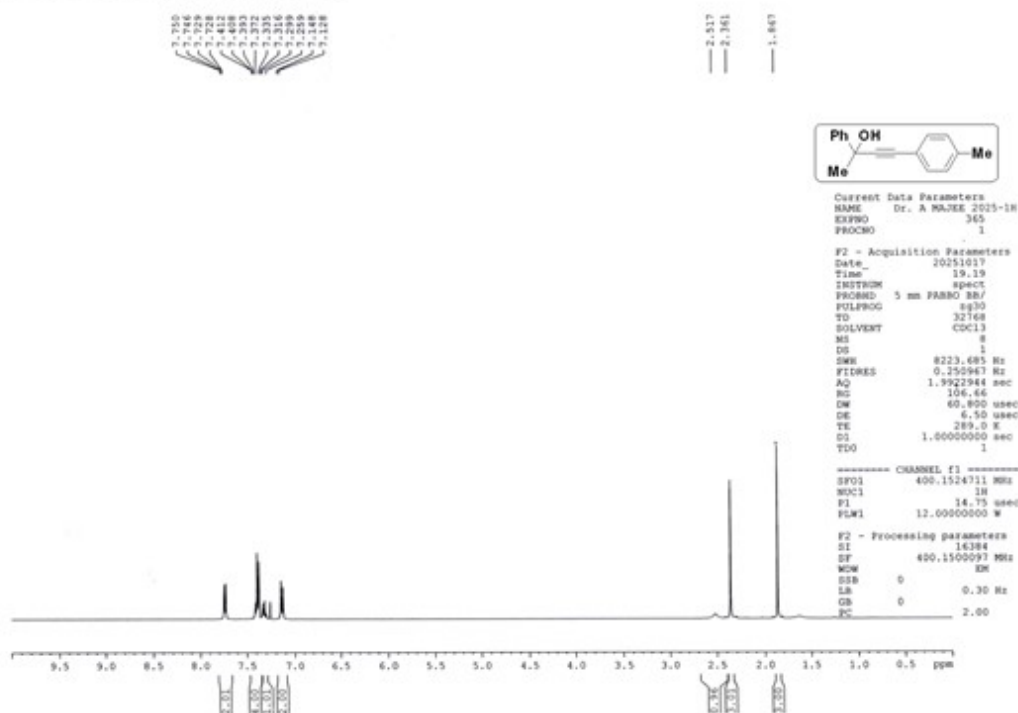
¹H NMR : 400 MHz, Solvent : CDCl₃



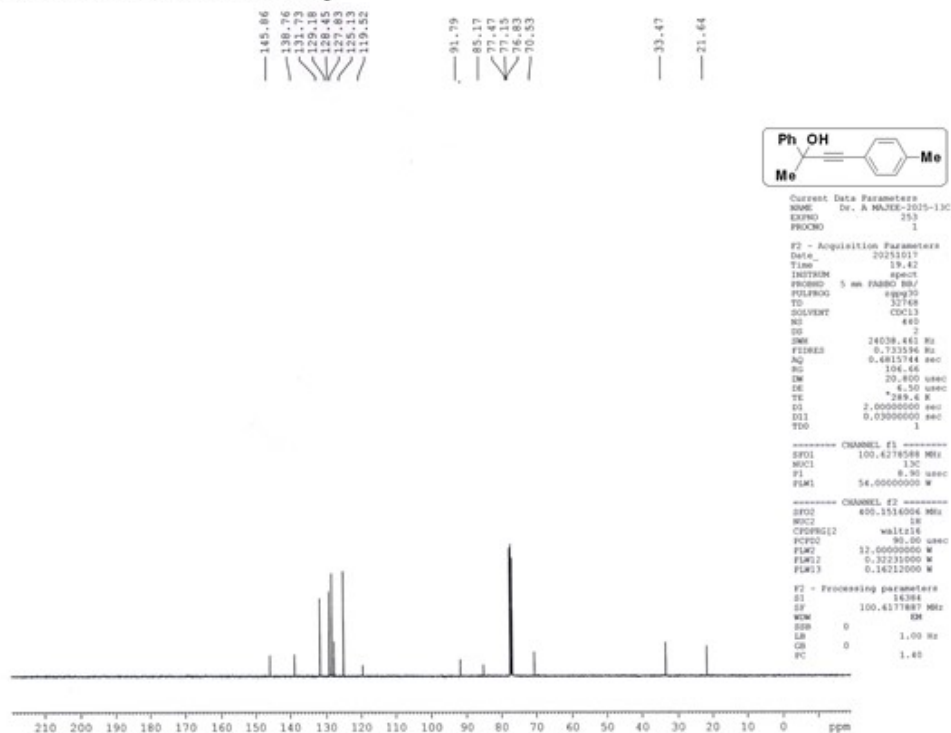
¹³C{¹H} NMR : 100 MHz, Solvent : CDCl₃

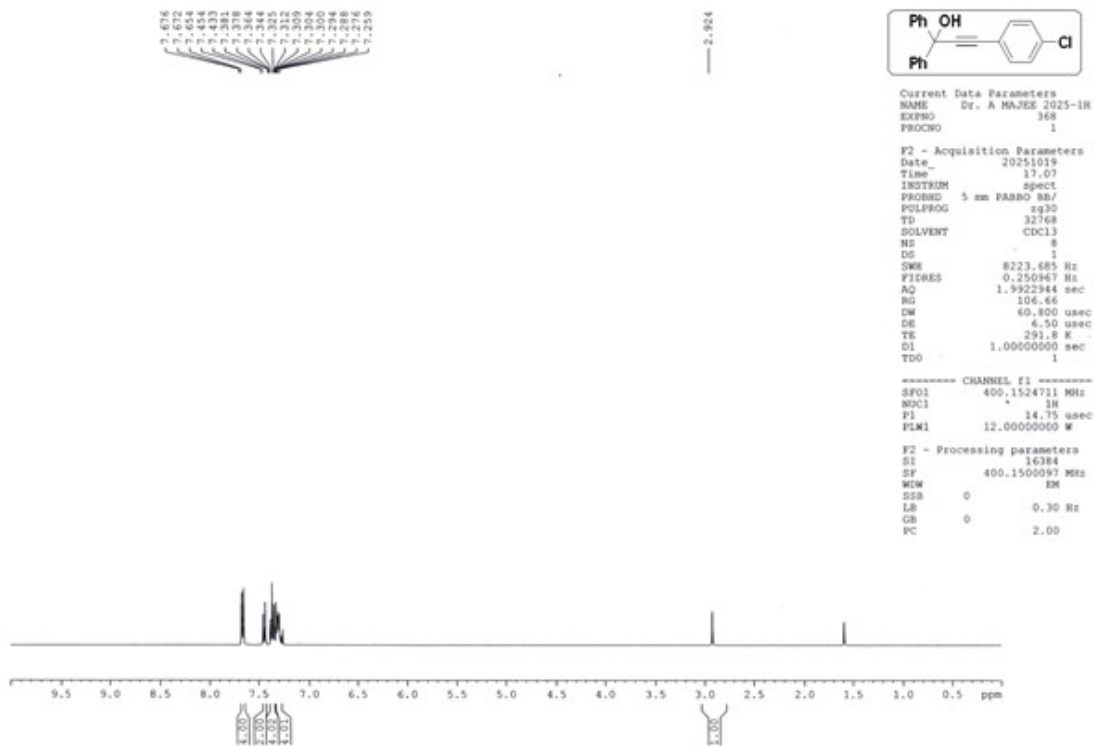
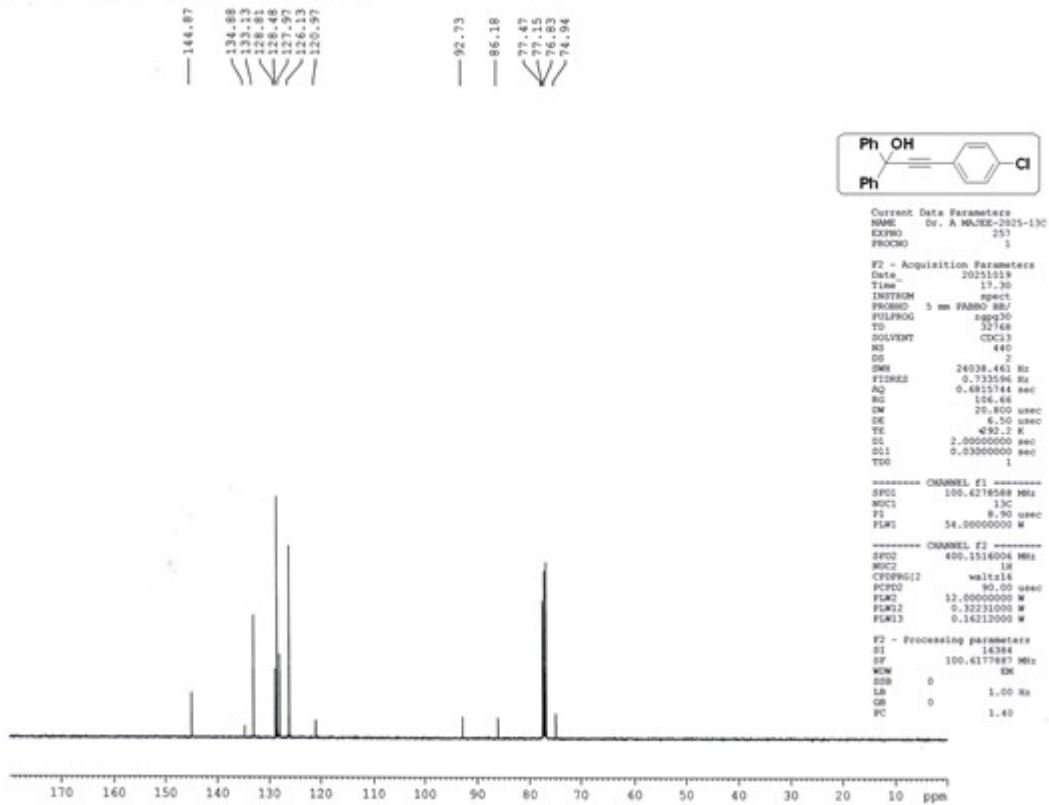


^1H NMR : 400 MHz, Solvent : CDCl_3

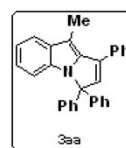
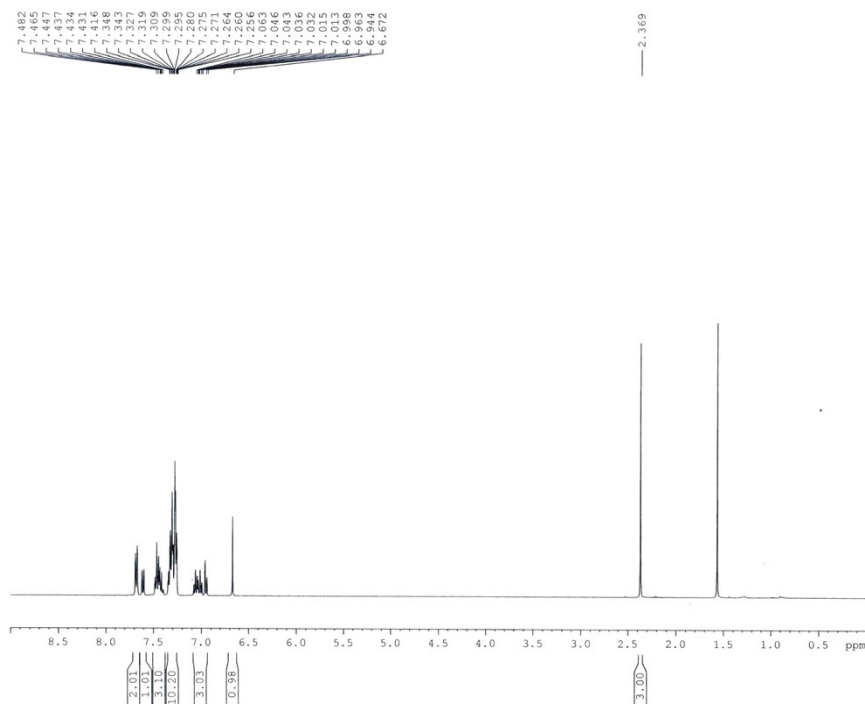


$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



¹H NMR : 400 MHz, Solvent : CDCl₃ $^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3 

^1H NMR : 400 MHz, Solvent : CDCl_3



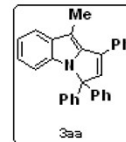
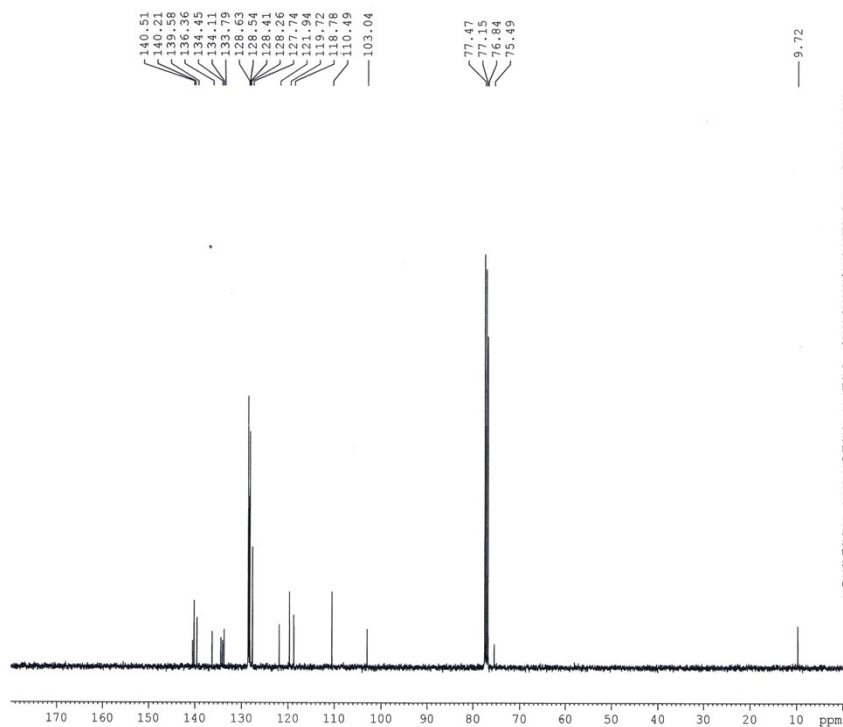
Current Data Parameters
NAME Dr. A MAJEE 2020
EXPNO 417
PROCNO 1

F2 - Acquisition Parameters
Date_ 20201004
Time_ 19.17
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 32
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 186.42
DW 60.800 usec
DE 6.50 usec
TE 295.7 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500046 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2020
EXPNO 448
PROCNO 1

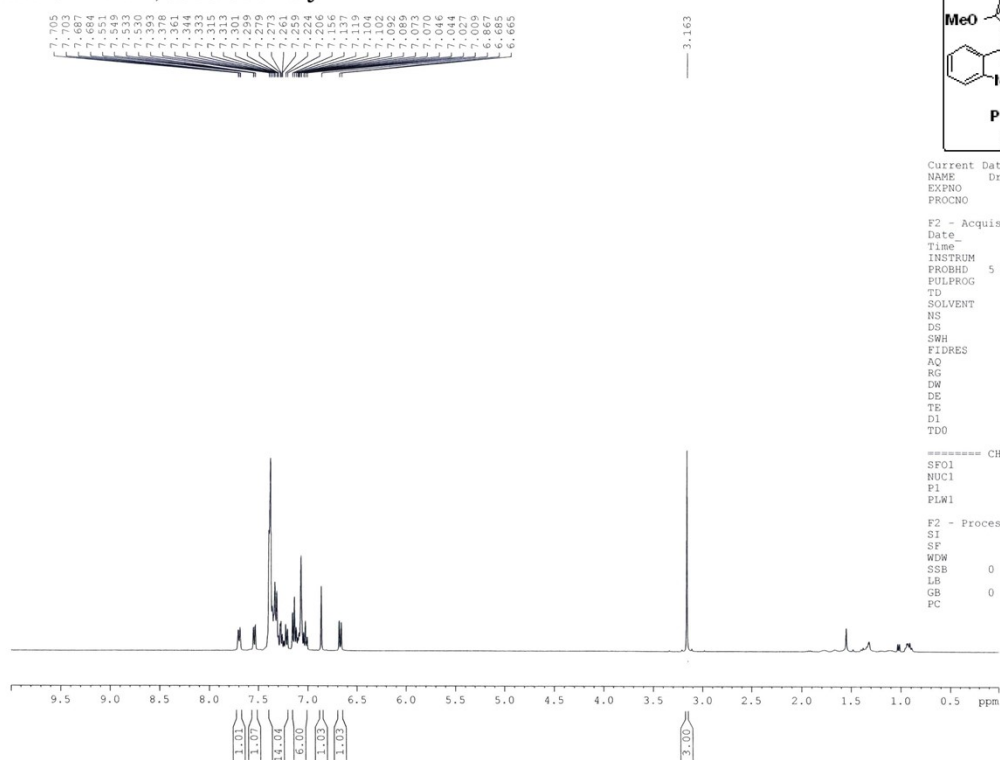
F2 - Acquisition Parameters
Date_ 20201014
Time_ 13.03
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 512
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 186.42
DW 20.800 usec
DE 6.50 usec
TE 300.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

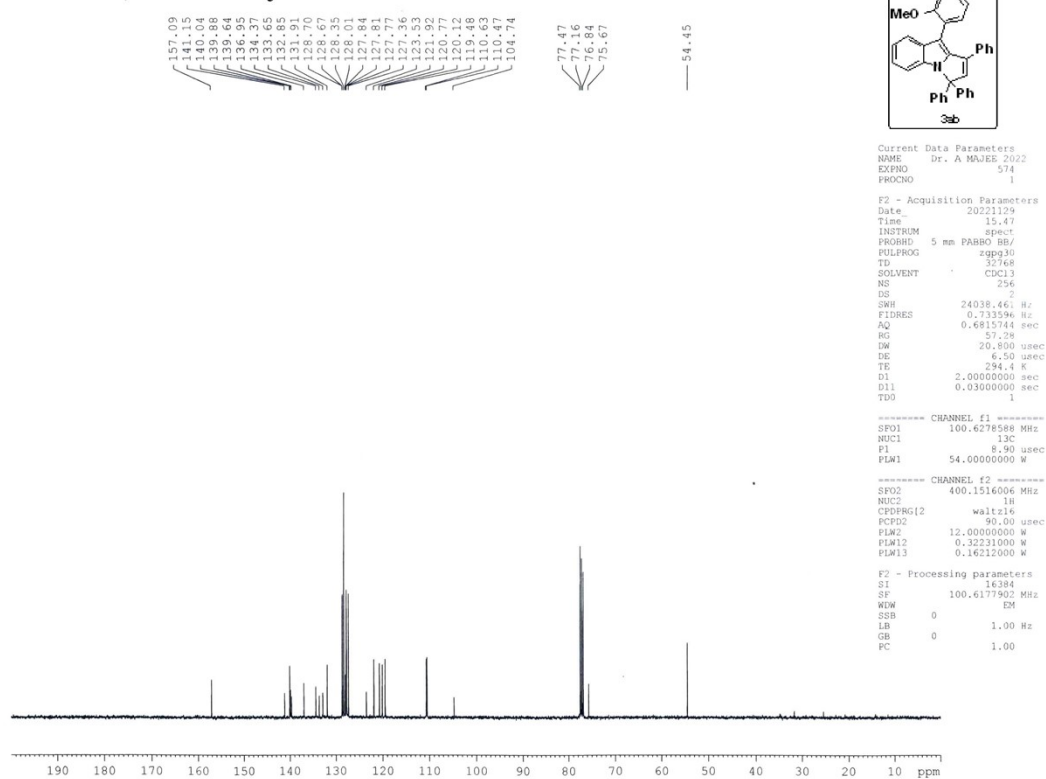
===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 ^1H
CPDPRG12 waltz16
PCPD 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

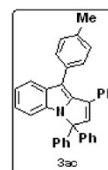
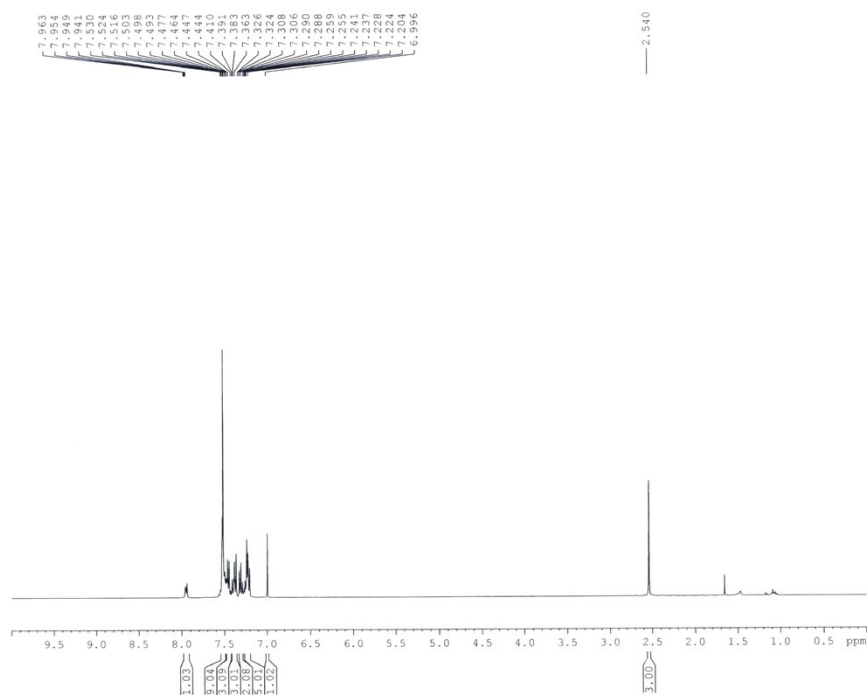
F2 - Processing parameters
SI 16384
SF 100.6177844 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



$^1\text{H NMR}$: 400 MHz, Solvent : CDCl_3 

```

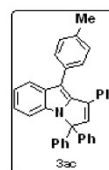
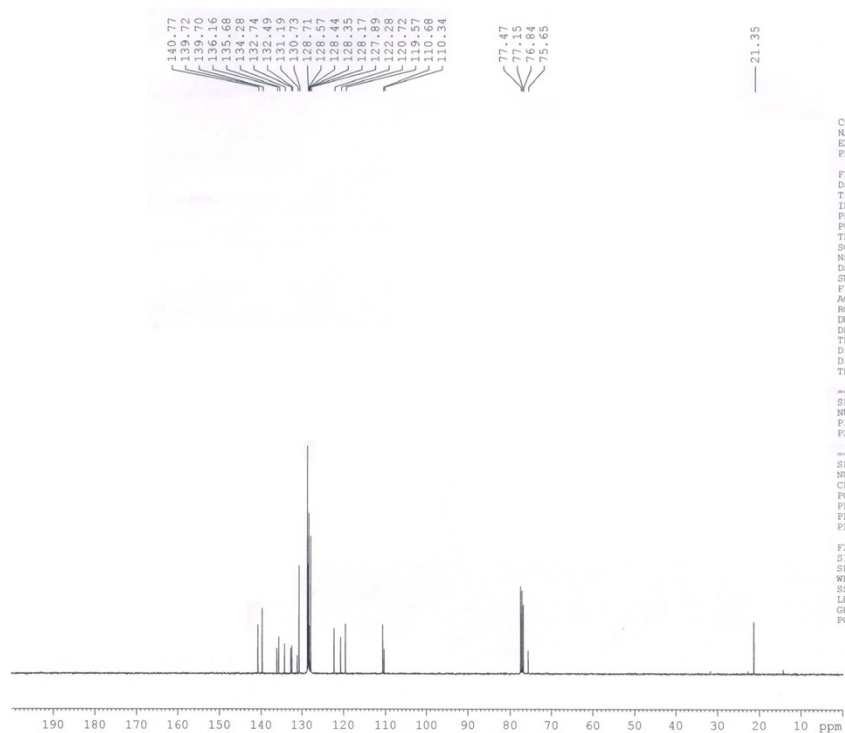
Current Data Parameters
NAME      Dr. A MAJEE 2020
EXPNO     444
PROCNO    1

F2 - Processing Parameters
Date_     20201014
Time      12.02
INSTRUM    spect
PROBHD     5 mm PABBO Bb
PULPROG    zgpg30
TD          32768
SOLVENT     CDCl3
NS          8
DS          1
SWH         8223.685 Hz
FIDRES     0.250967 Hz
AQ          1.952294 sec
RG          40.87
LW          60.800 usec
DE          6.50 usec
TE          298.6 K
D1          1.0000000 sec
TDO         1

===== CHANNEL f1 =====
SF01       400.1524711 MHz
NUC1        1H
P1          14.75 usec
PLW1       12.0000000 W

F2 - Processing parameters
SI          16384
SF          400.1499555 MHz
WDW         EM
SSB         0
GB          0.30 Hz
LB          0
PC          0

```

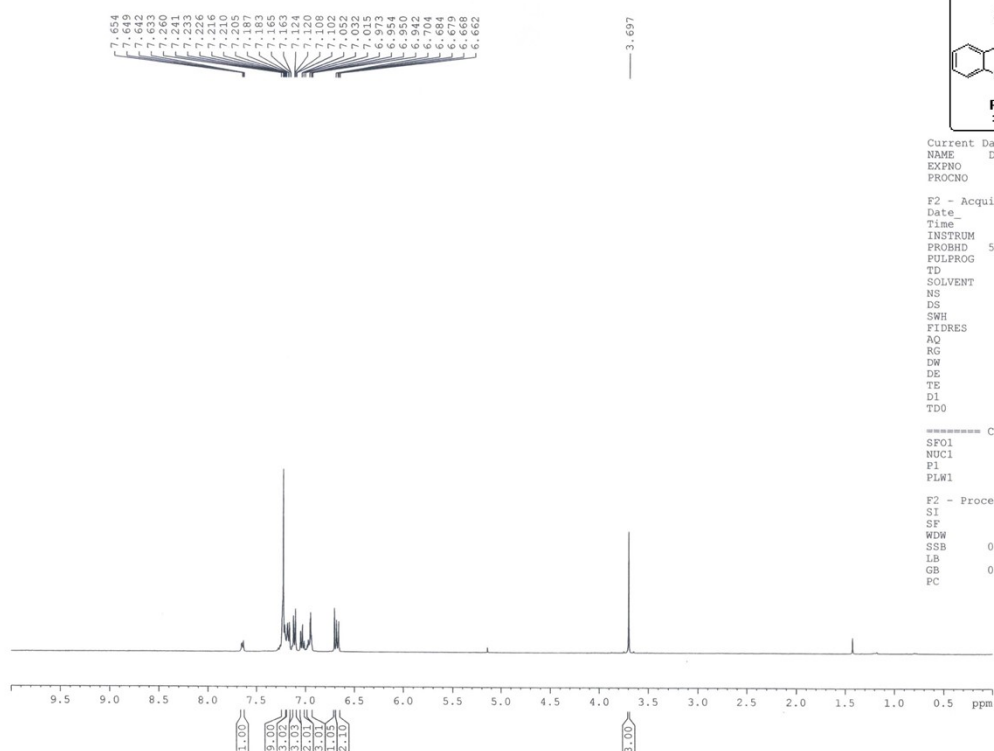
 $^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3 

```

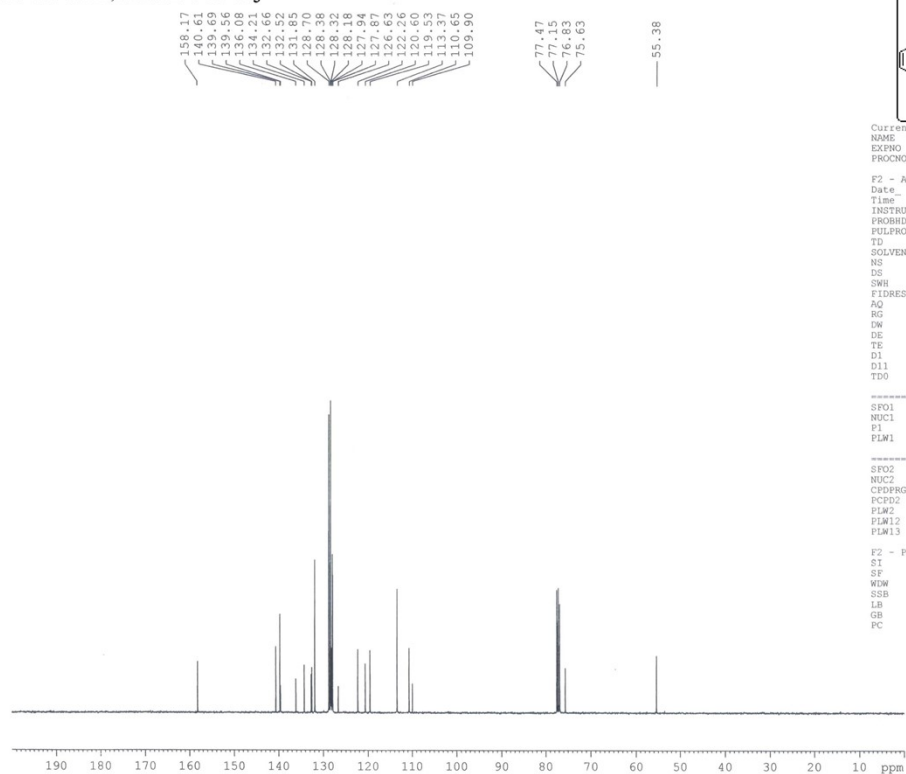
Current Data Parameters
Name      Dr. A MAJEE 2020
EXPNO     1          445
PROCNO    1
=====
F2 - Acquisition Parameters
Date_      20200104
Time       12.15
INSTRUM    spect
PROBHD     5 mm PABBO-1H/13C
PULPROG    zgpg30
TD         65536
SOLVENT    CDCl3
NS          256
DS          2
SWH         24038.461 Hz
FIDRES     0.733556 Hz
AQ          0.681744 sec
RG          40.87
LW          20.800 usec
DE          50 usec
TE          299.3 K
D1          2.00000000 sec
D11         0.03000000 sec
TDO         1
=====
CHANNEL f1
SF01       100.6278568 MHz
NUC1        13C
P1          8.90 usec
PL1         54.00000000 W
=====
CHANNEL f2
SF02       400.1556066 MHz
NUC2        1H
P2          wait16
PL2         12.00000000 W
PL12        0.32231000 W
PL13        0.16212000 W
=====
F2 - Processing parameters
SF         100.6277971 MHz
RG         4096
SI          32768
GB          0
LB          0.100 Hz
GB          0
GB          0
GB          1.40

```

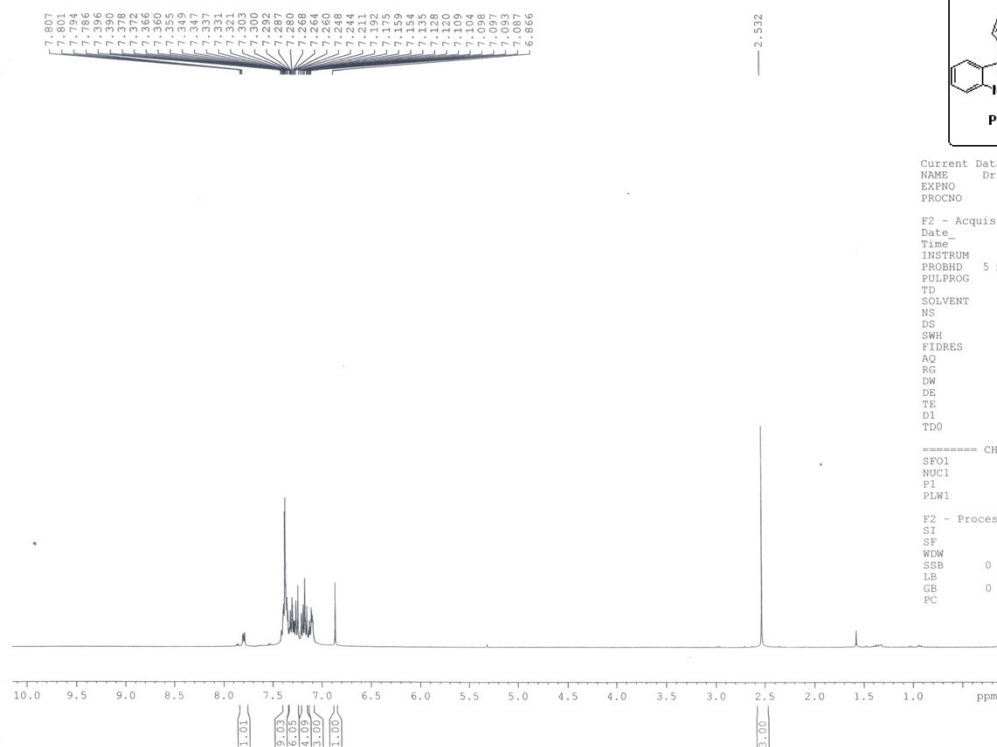

¹H NMR : 400 MHz, Solvent : CDCl₃



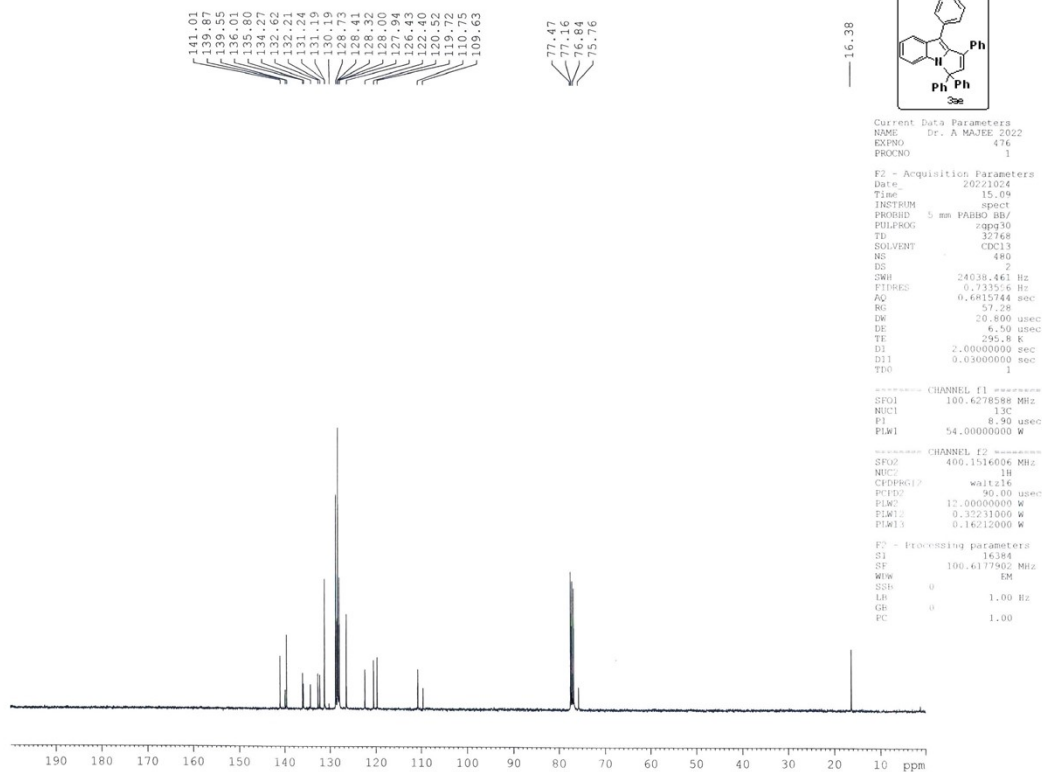
IR : 100 MHz, Solvent : CDCl₃



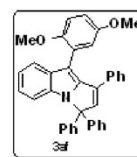
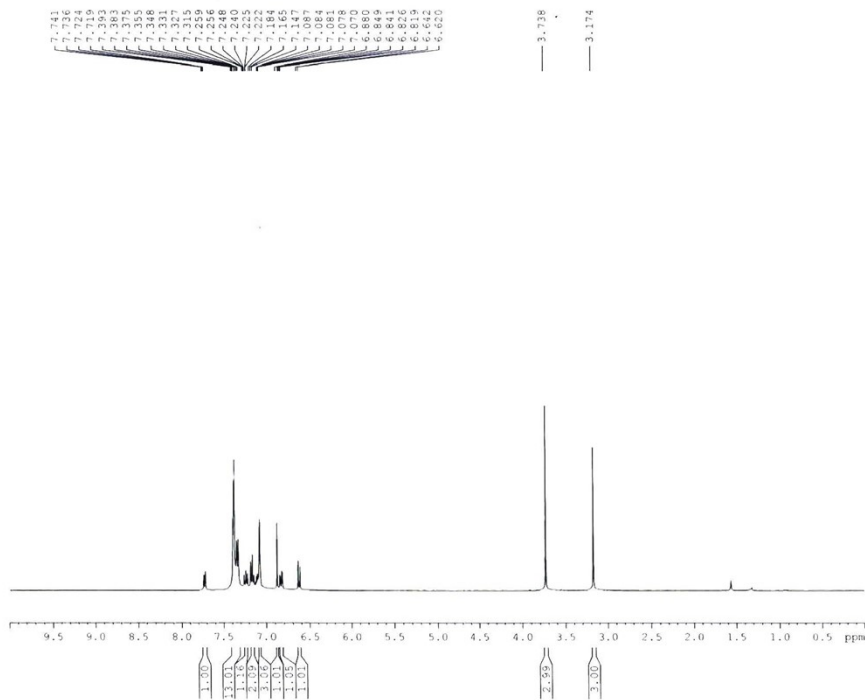
^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



¹H NMR : 400 MHz, Solvent : CDCl₃



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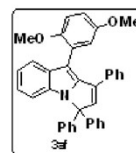
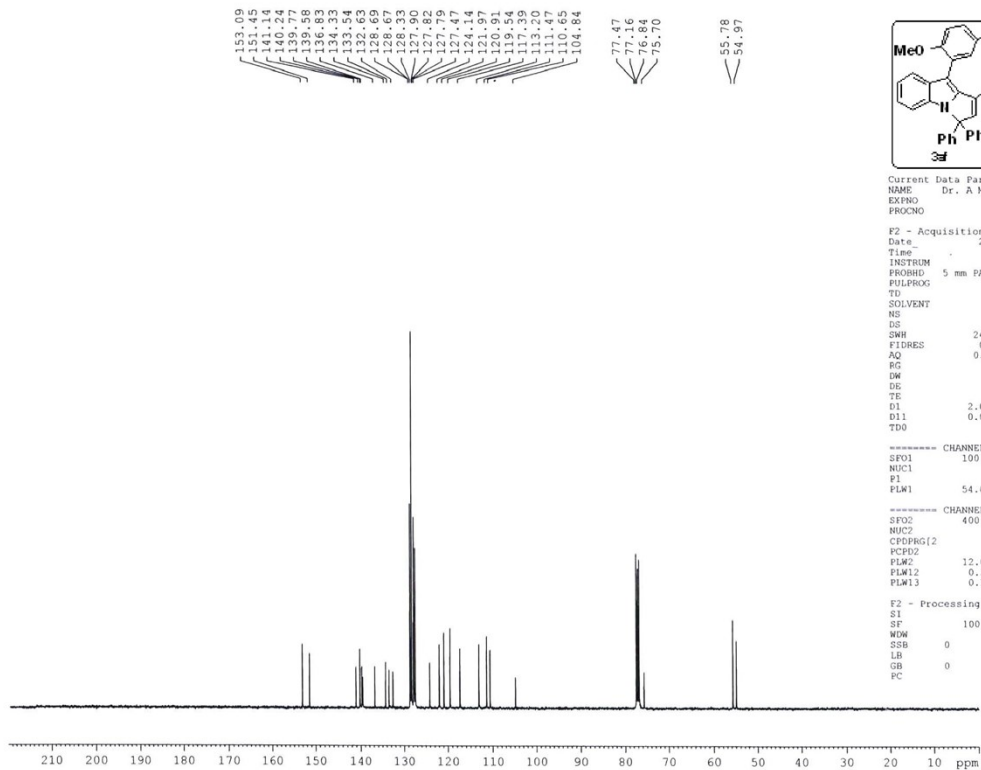
Current Data Parameters
NAME      Dr. A MAJEE 2022
EXPNO     506
PROCNO    1

P2 - Acquisition Parameters
Date_     202211107
Time      13.09
INSTRUM    spect
PROBHD     5 mm F4BBO BB/
PULPROG    zgpg30
TD          69300
SOLVENT    CDCl3
NS          8
DS          1
SWH         8223.685 Hz
FIDRES     0.25097 Hz
AQ          1.950284 sec
RG          62.69
DE          60.800 usec
TE          300.2
DW          294.0 K
D1          1.00000000 sec
TD0         1

===== CHANNEL f1 =====
SFO1       400.1524711 MHz
NUC1        131
P1          14.75 usec
PLM1       12.00000000 W

F2 - Processing parameters
S1          131
F1          16.84
SF           400.1500000 MHz
WDW          EM
SS           0
LB           0.30 Hz
GB           0
PC           2.00

```

 $^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3 

```

Current Data Parameters
NAME      Dr. A. MAJEE 2022
EXPNO     507
PROCNO    1

F2 - Acquisition Parameters
Date_      20221107
Time       13.35
INSTRUM    HMNMR
PROBHD     5 mm PABBO / 1H-13
PULPROG    zgpg30
TD          65536
SOLVENT    CDCl3
NS          512
DS          2
SWH         24039.461 Hz
FIDRES      0.733596 Hz
AQ          0.683754 sec
RG          62.65
WDW          20.950 usec
SSB          0
LB           1.00 Hz
GB          0
DE          294.9 K
TE          300.2 sec
D1           0.30000000 sec
T2           1
T20          1

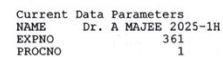
===== CHANNEL f1 =====
SPF1        100.627585000 W
NUC1         13C
P1           1.80 usec
PLW1         54.00000000 W

===== CHANNEL f2 =====
MUC2         400.15160000 W
NUC2         1H
PCPGPRG2(C) walz16b
PCPDPRG2(C)
P2           12.00000000 W
PLW2         0.32231000 W
PLW12        0.16212000 W

F2 - Processing parameters
SF           400.1517932 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1
LB           0.100 Hz

```

7.492
7.489
7.471
7.469
7.420
7.413
7.401
7.399
7.380
7.376
7.361
7.357
7.339
7.328
7.303
7.289
7.277
7.259
7.228
7.209
7.204
7.183
7.181
7.148
7.129
7.114
7.112
7.109
7.096
7.081
7.077
7.036
7.026
7.016
7.010
7.002
6.827



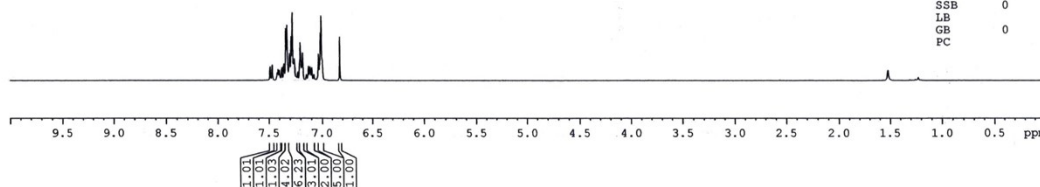
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F2 - Acquisition Parameters
Date_      20251017
Time       11:34
INSTRUM    spect
PROBHD     5 mm PABBO BB/
PULPROG    zg30
TD          32768
SOLVENT    CDCl3
NS          8
DS          1
SWH         8223.685 Hz
FIDRES     0.250967 Hz
AQ          1.9922944 sec
RG          77.59
BW          60.800 usec
DE          6.50 usec
TE          290.6 K
D1          1.00000000 sec
TDO        1

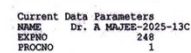
```

```
===== CHANNEL f1 =====
SFO1      400.1524711 MHz
NUC1              1H
P1              14.75 usec
PLW1      12.00000000 W
```

F2 - Processing parameters	
SI	16384
SF	400.1500318 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	2.00



141.14
140.41
139.65
139.38
136.08
135.85
133.92
133.13
132.64
132.60
128.77
128.74
128.57
128.36
128.25
128.25
128.12
127.94
127.92
127.82
127.69
126.93
126.40
122.18
120.74
119.63
110.75
108.66



```

F2 - Acquisition Parameters
Date_      20251017
Time       12.03
INSTRUM    spect
PROBHD     5 mm PABBO BB/
PULPROG    zgpg30
TD         32768
SOLVENT    CDC13
NS         600
DS         2
SWH        240398.461 Hz
FIDRES     0.733596 Hz
AQ         0.6815744 sec
RG         77.59
DW         20.800 usec
DE         6.50 usec
TE         290.8 K
D1         2.00000000 sec
D11        0.030000000 sec
TD0        1

```

```

*** CHANNEL f1 ***
SFO1      100.6278588 MHz
NUC1      13C
P1         8.90 usec
PLW1      54.00000000 W

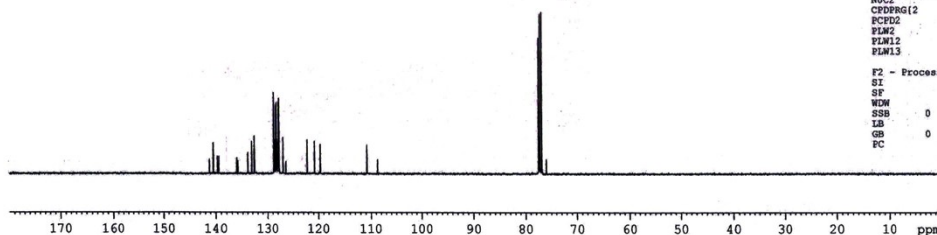
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```

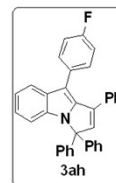
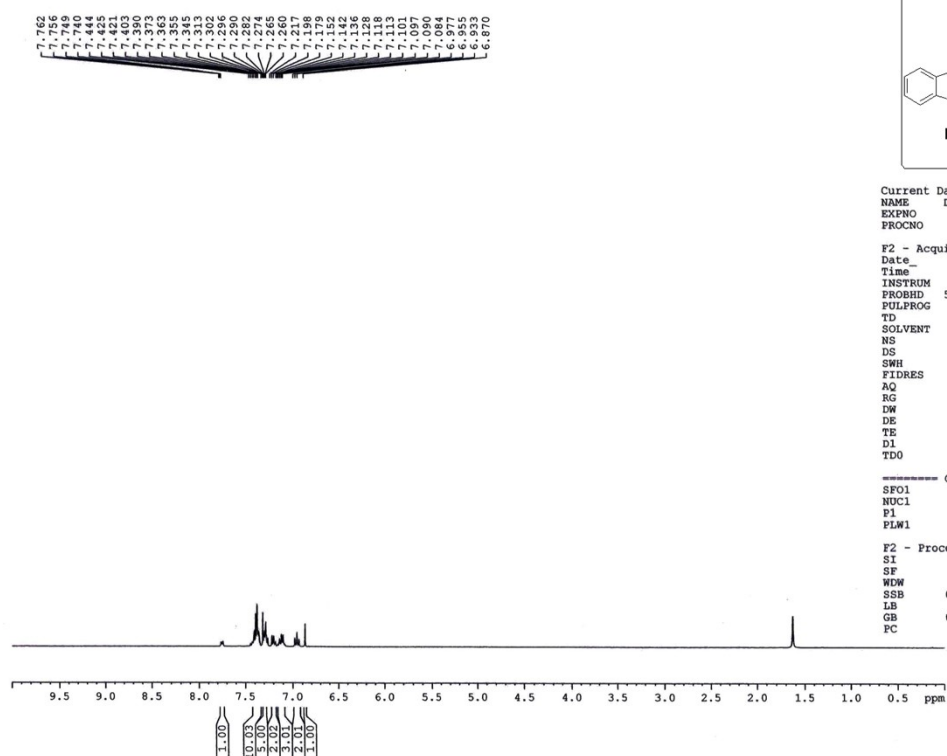
CHANNEL F2
SFO2      400.1516006 MHz
NUC2      1H
CPDRG(2   waltz16
PCPD2     90.00 usec
PLW2      12.000000000 W
PLW12     0.32231000 W
PLW13     0.16212000 W

```

```
F2 - Processing parameters
SI                      16384
SF                      100.6177902 MHz
WDW                      EM
SSB                      0
LB                      1.00 Hz
GB                      0
PC                      1.40
```



^1H NMR : 400 MHz, Solvent : CDCl_3



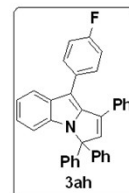
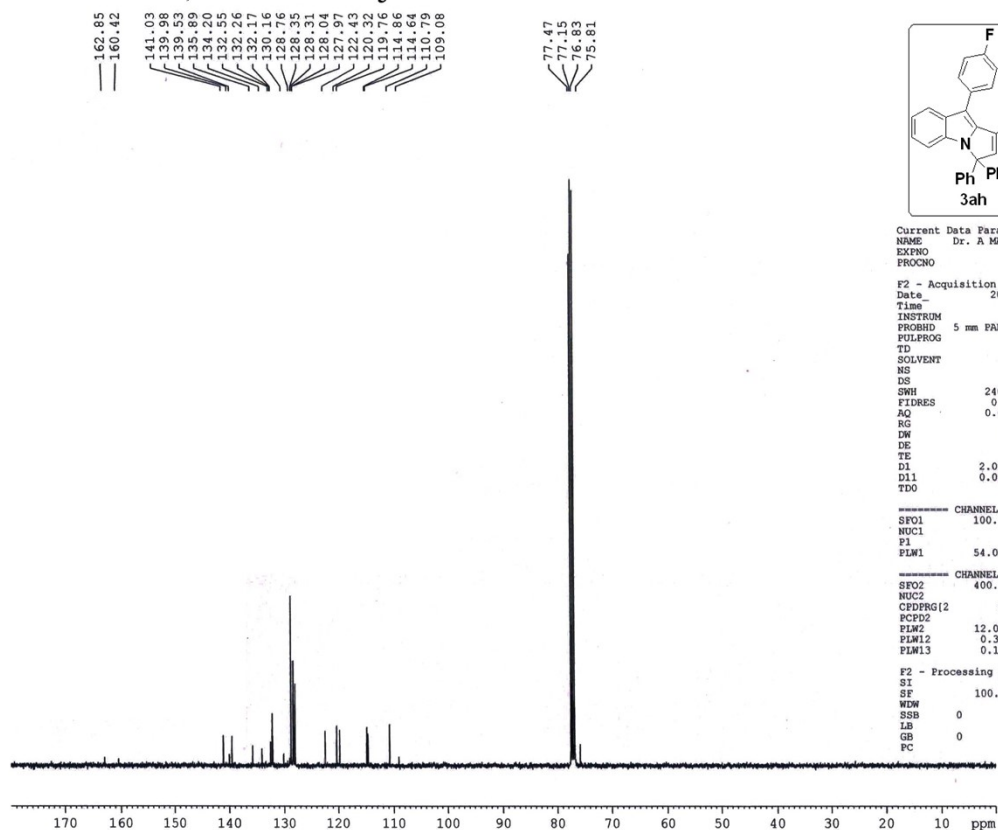
Current Data Parameters
NAME Dr. A MAJEE 2025-1H
EXPNO 372
PROCNO 1

F2 - Acquisition Parameters
Date_ 20251022
Time 17.04
INSTRUM spect
PROBHD 5 mm PABBO BB/
FULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 186.42
DW 60.800 usec
DE 6.50 usec
TE 293.8 K
D1 1.00000000 sec
TDO 1

CHANNEL f1
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1499881 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

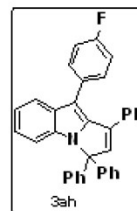
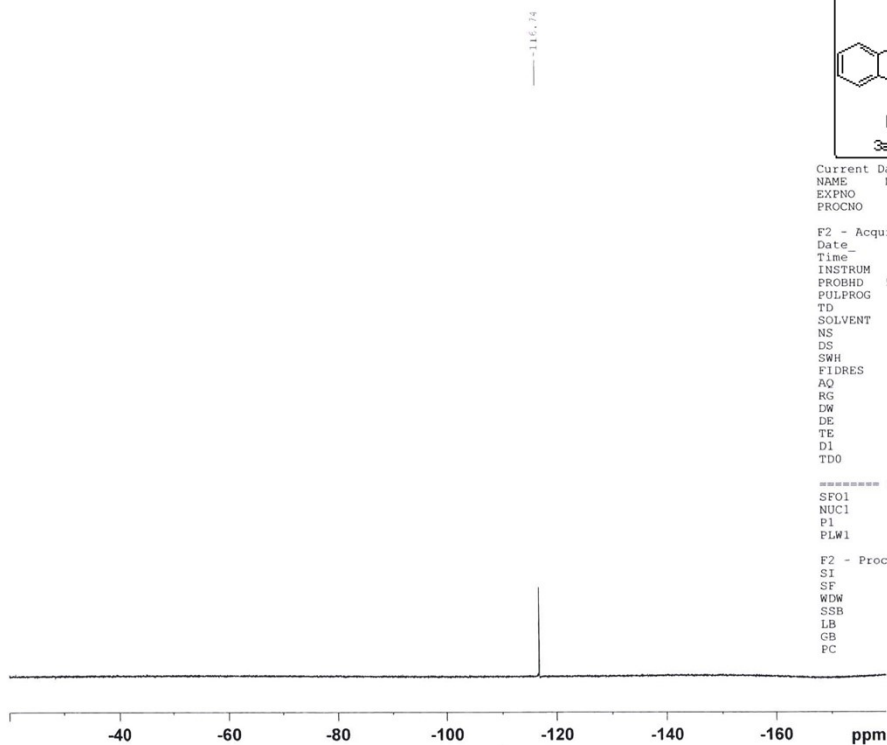
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2025-13C
EXPNO 260
PROCNO 1

F2 - Acquisition Parameters
Date_ 2

^{19}F NMR : 376 MHz, Solvent : CDCl_3

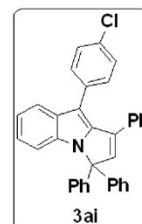
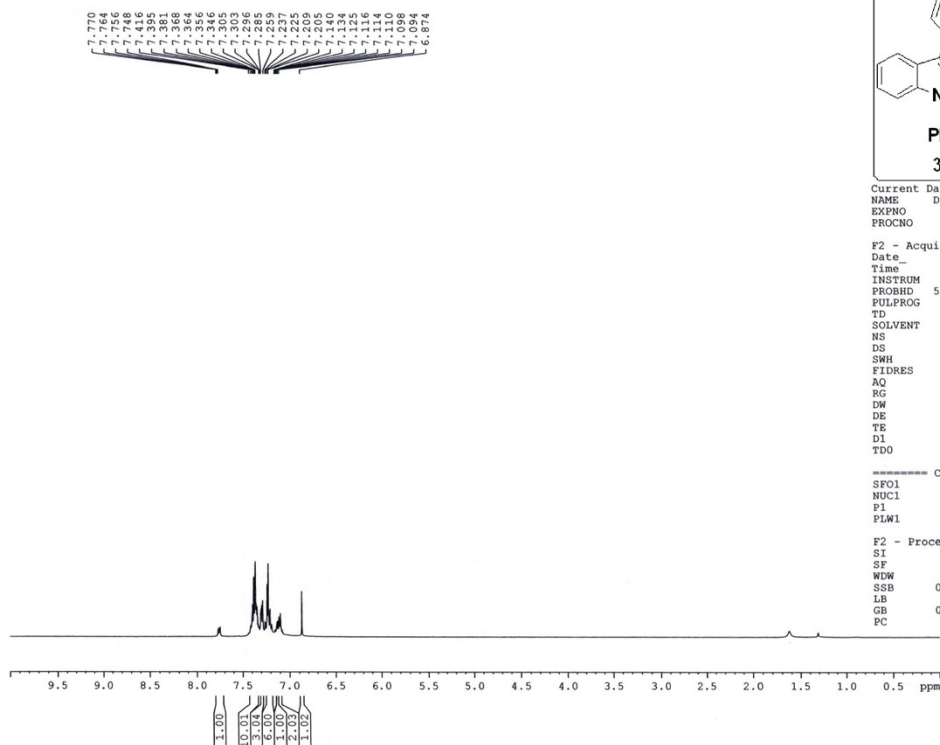


Current Data Parameters
NAME Dr. A MAJEE 2024
EXPNO 488
PROCNO 1

F2 - Acquisition Parameters
Date 20241217
Time 17.50
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 16
DS 2
SWH 89285.711 Hz
FIDRES 2.724784 Hz
AQ 0.1835008 sec
RG 186.42
DW 5.600 usec
DE 6.50 usec
TE 289.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SF01 376.4795333 MHz
NUC1 ^{19}F
P1 12.50 usec
PLW1 20.00000000 W

F2 - Processing parameters
SI 16384
SF 376.5171850 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

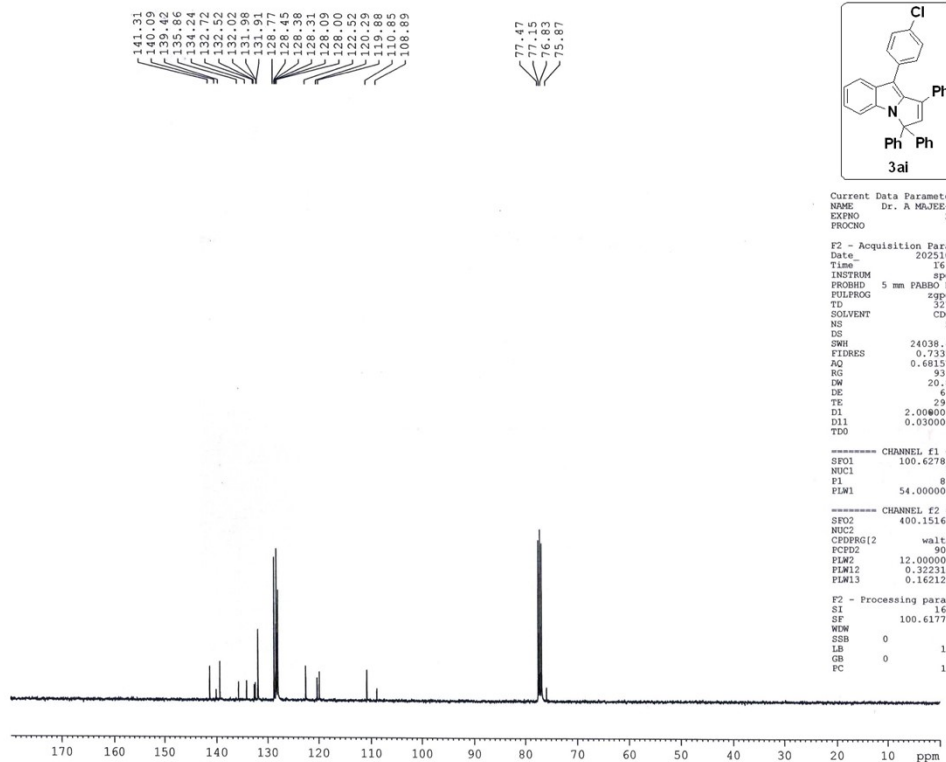


Current Data Parameters
NAME Dr. A MAJEE 2025-1H
EXPNO 371
PROCNO 1

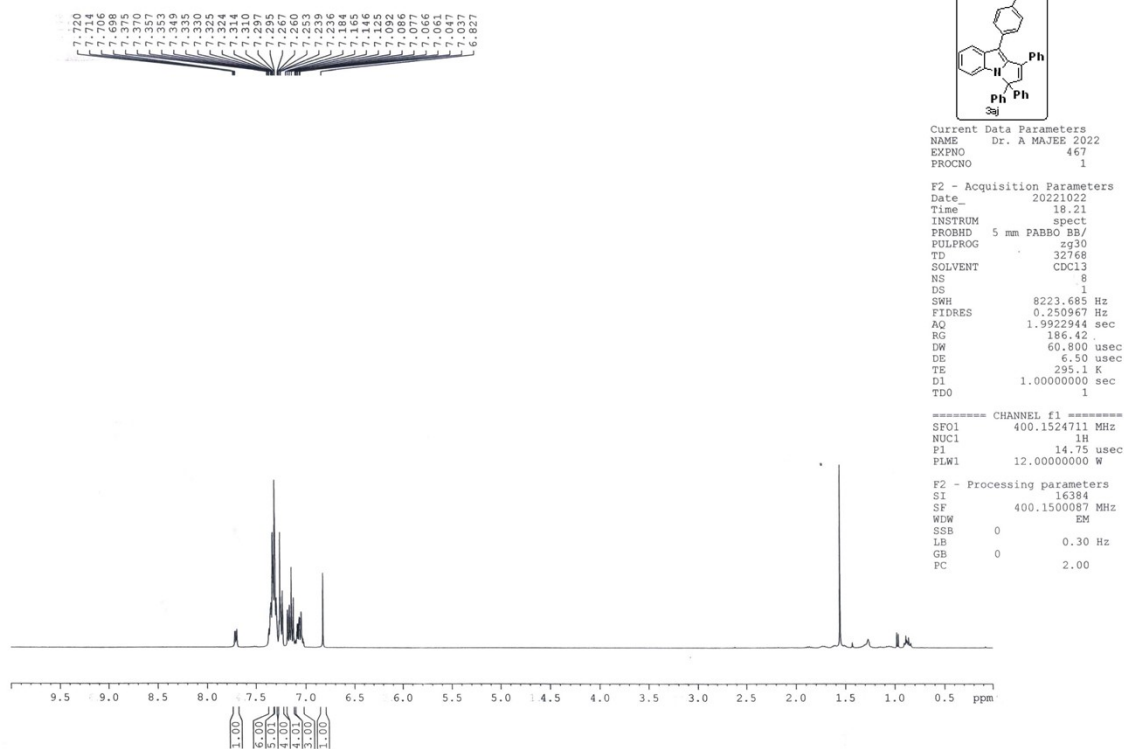
F2 - Acquisition Parameters
Date 20251022
Time 16.26
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.230967 Hz
AQ 1.9922944 sec
RG 93.46
DW 60.800 usec
DE 6.50 usec
TE 294.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SF01 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

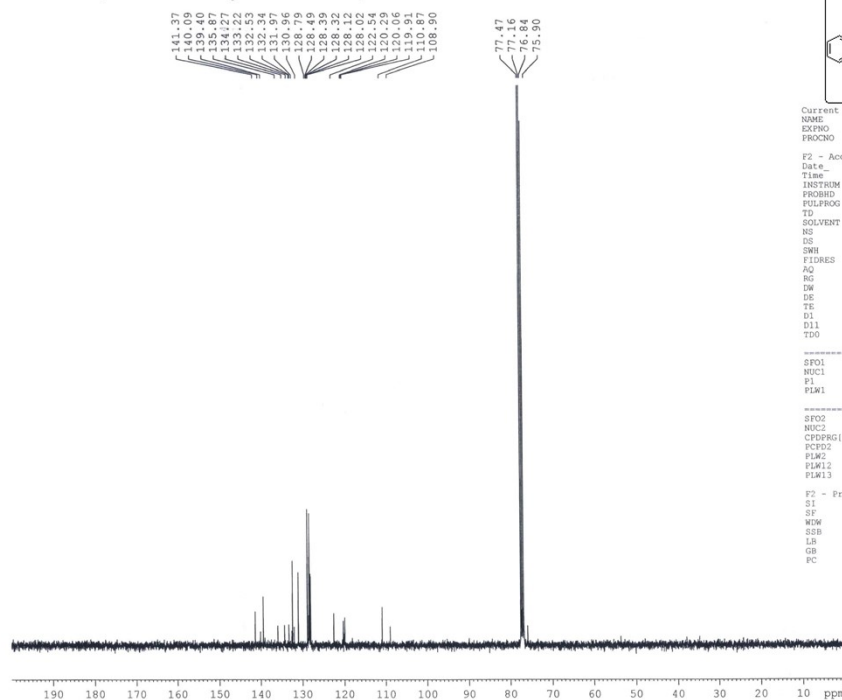
F2 - Processing parameters
SI 16384
SF 400.1499952 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00



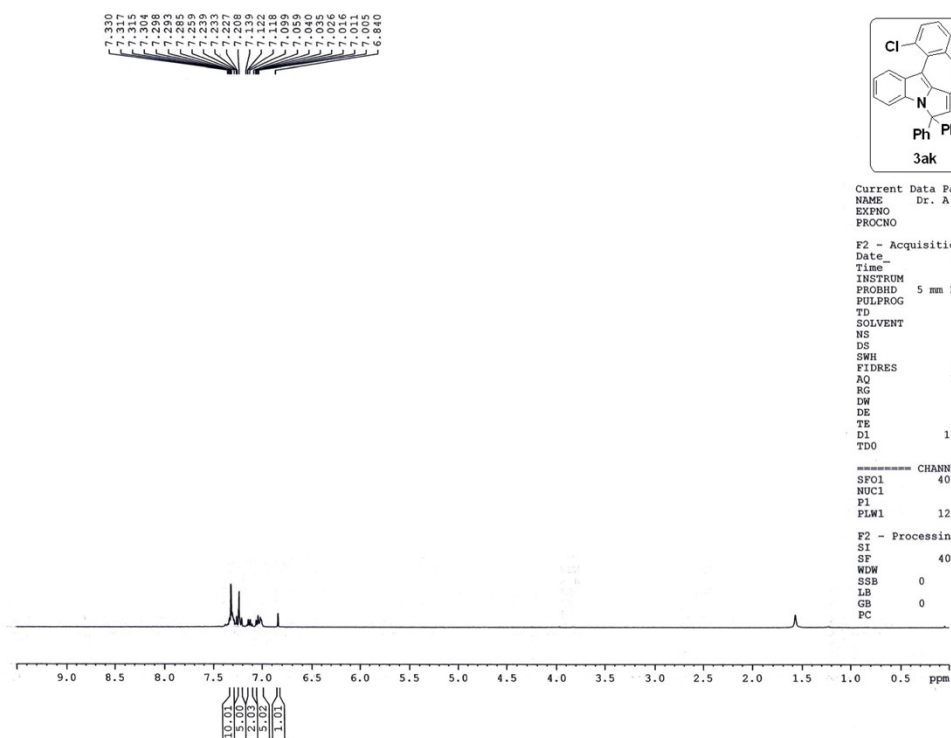
¹H NMR : 400 MHz, Solvent : CDCl₃



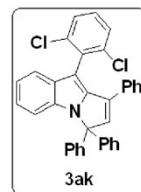
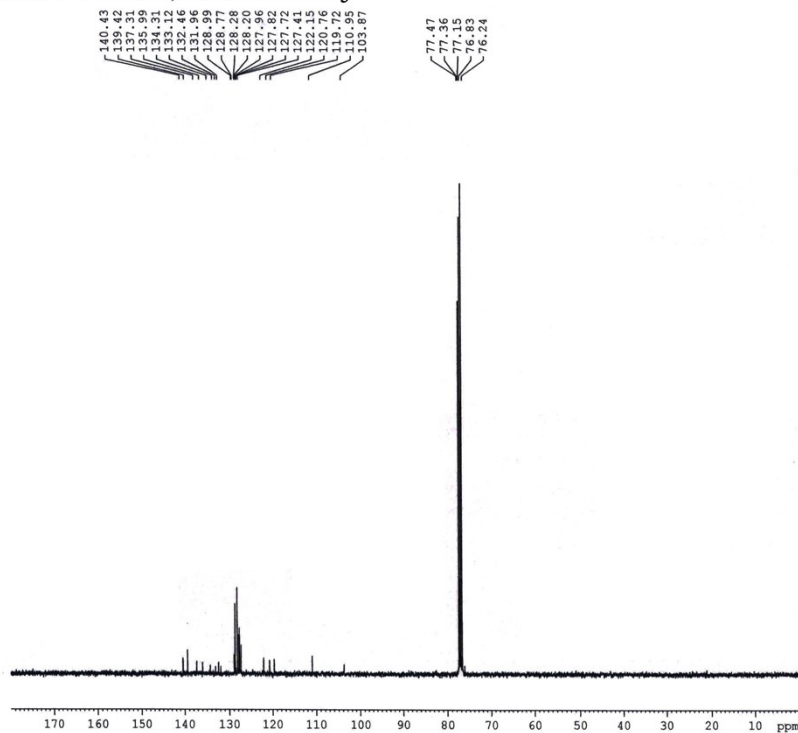
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE-2025-13c
EXPNO 247
PROCNO 1

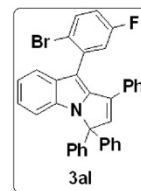
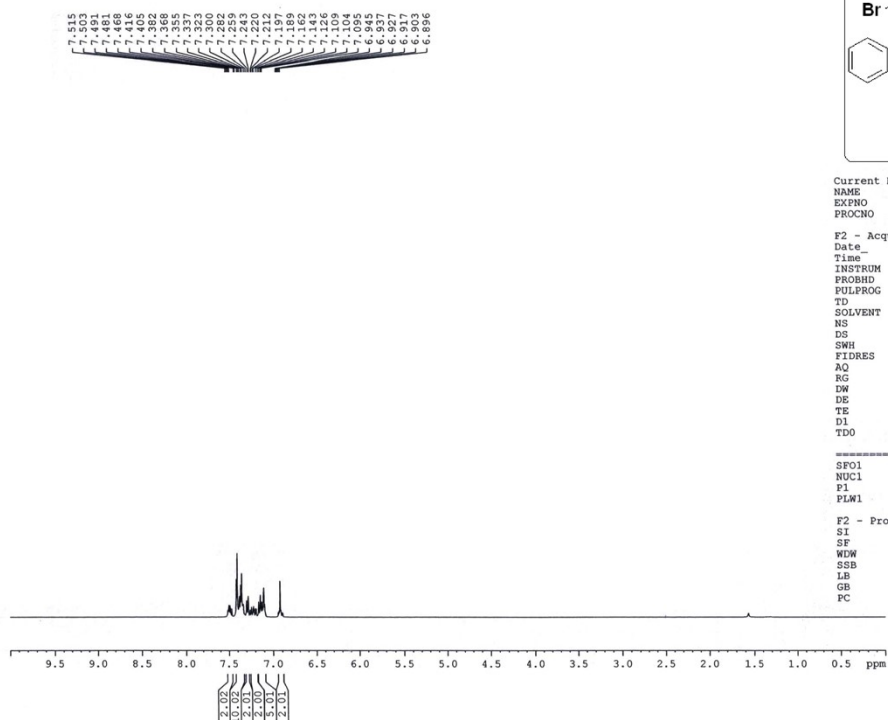
F2 - Acquisition Parameters
Date 20251016
Time 17.17
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 720
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6915744 sec
RG 186.42
DW 20.800 usec
DE 6.20 usec
TE 290.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 100.627869 MHz
NUC1 13C
P1 8.30 usec
PLW1 54.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177873 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

^1H NMR : 400 MHz, Solvent : CDCl_3



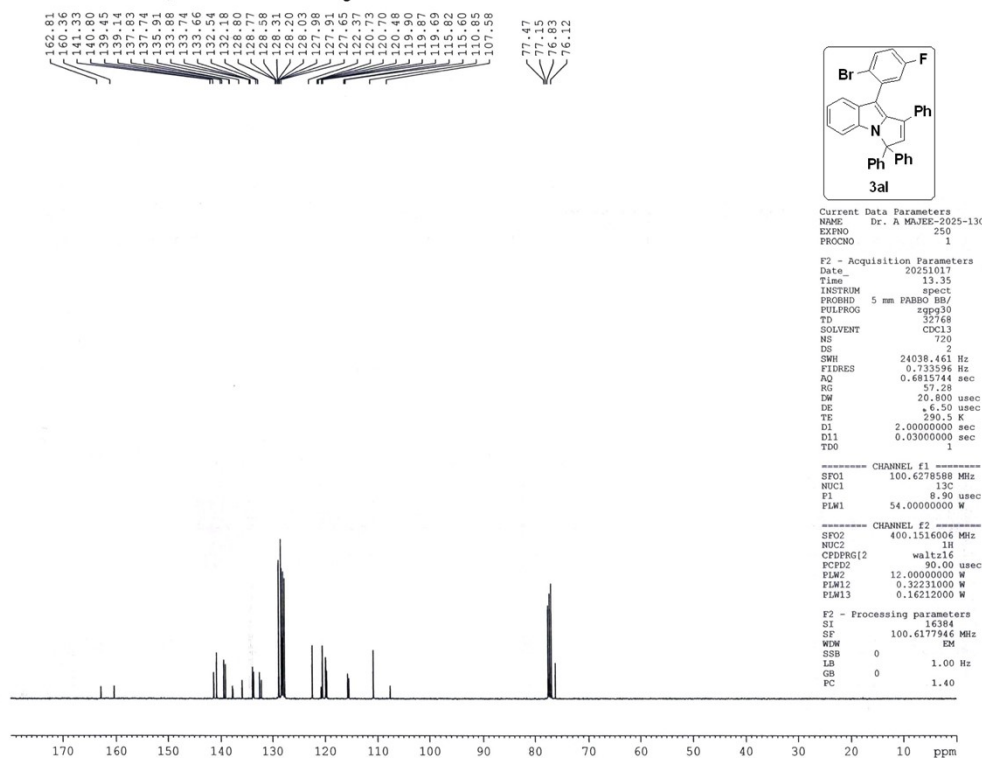
Current Data Parameters
NAME Dr. A MAJEE 2025-1H
EXPNO 362
PROCNO 1

F2 - Acquisition Parameters
Date 20251017
Time 12.57
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 52.28
DW 60.800 usec
DE 6.50 usec
TE 290.0 K
D1 1.00000000 sec
TDO 1

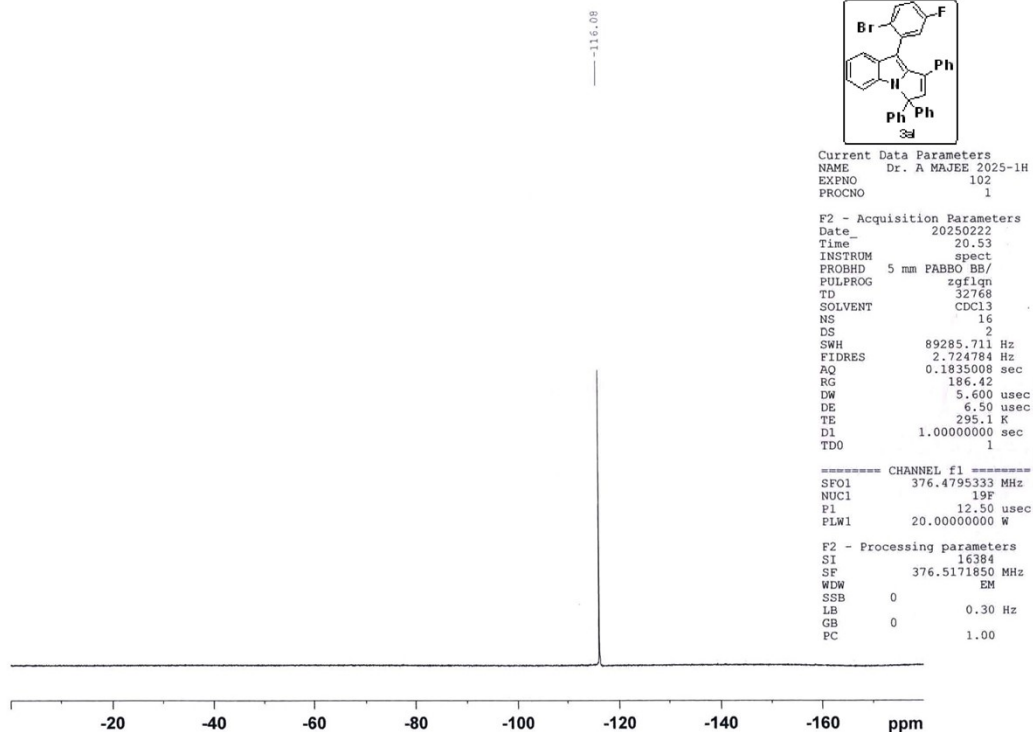
===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



^{19}F NMR : 376 MHz, Solvent : CDCl_3



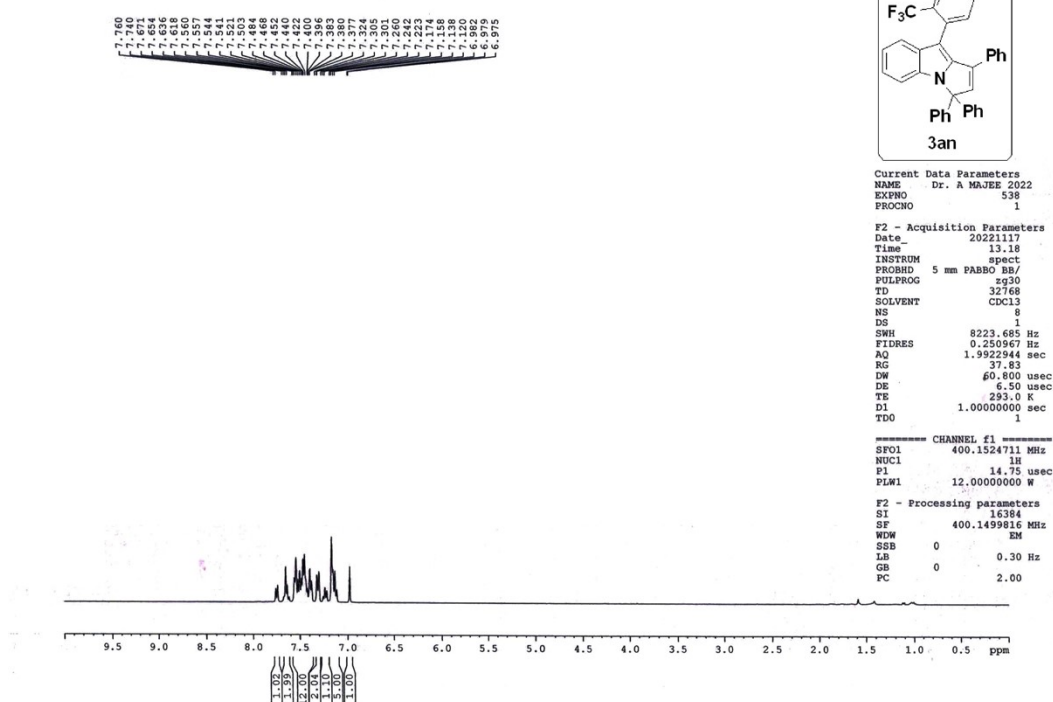
1.03
 1.05
 1.05
 1.07
 1.04
 1.02
 1.02
 1.00
 1.00
 1.00
 1.00

9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 ppm

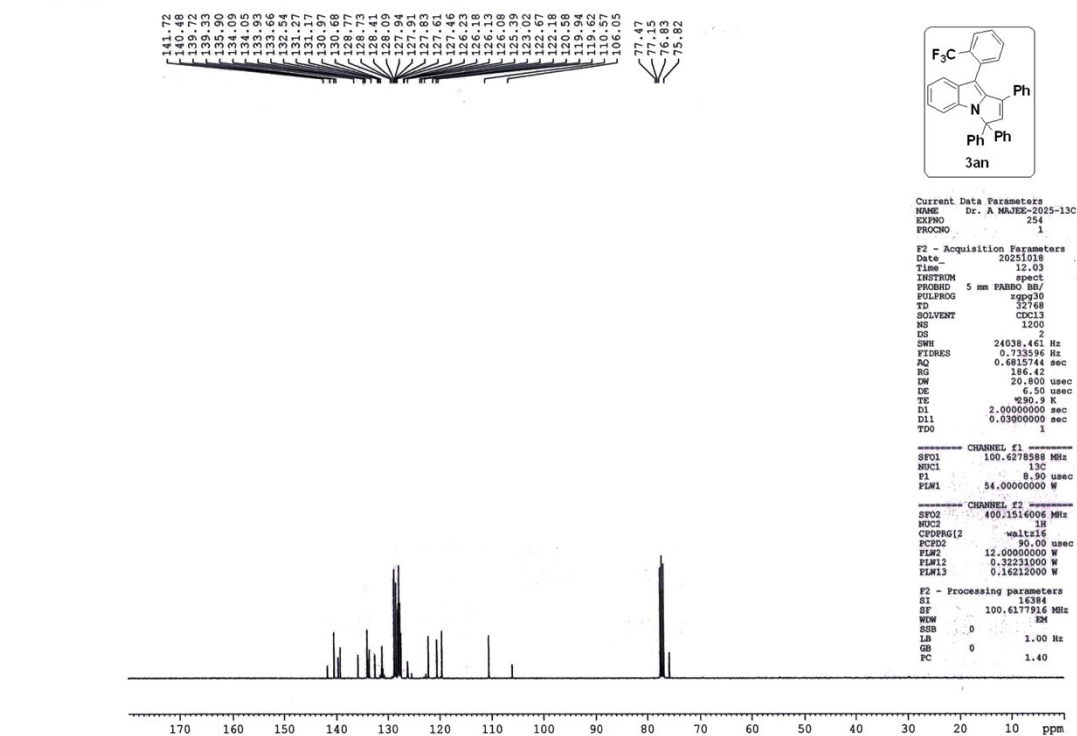
1.572
 7.608
 7.604
 7.600
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 7.592
 7.588
 7.584
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 7.564
 7.560
 7.556
 7.552
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 7.476
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 7.456
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 7.440
 7.436
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 7.424
 7.420
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 6.780
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 6.740
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 6.200
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 6.192
 6.188
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 6.180
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 6.124
 6.1

[illegible]

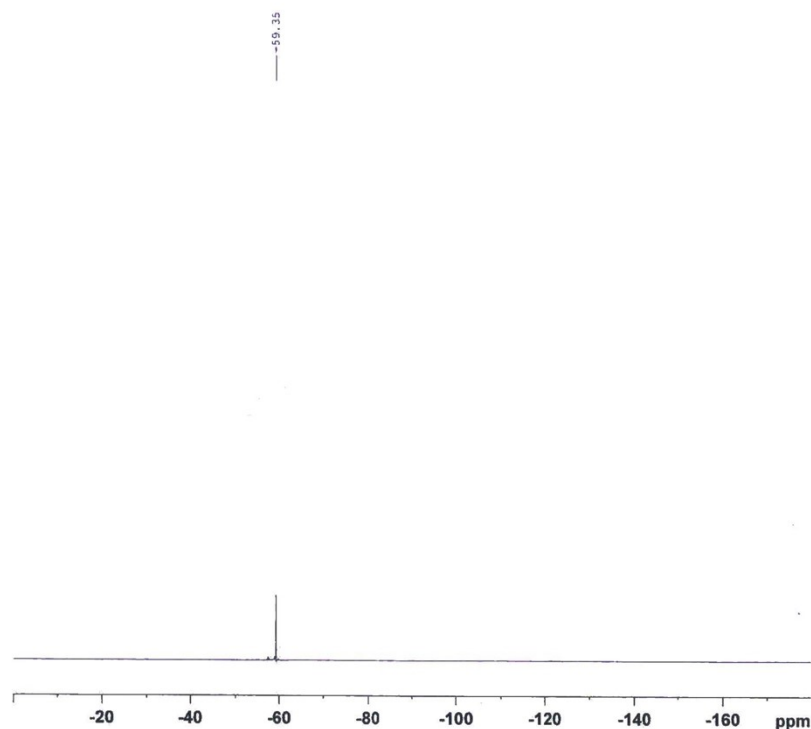
^1H NMR : 400 MHz, Solvent : CDCl_3



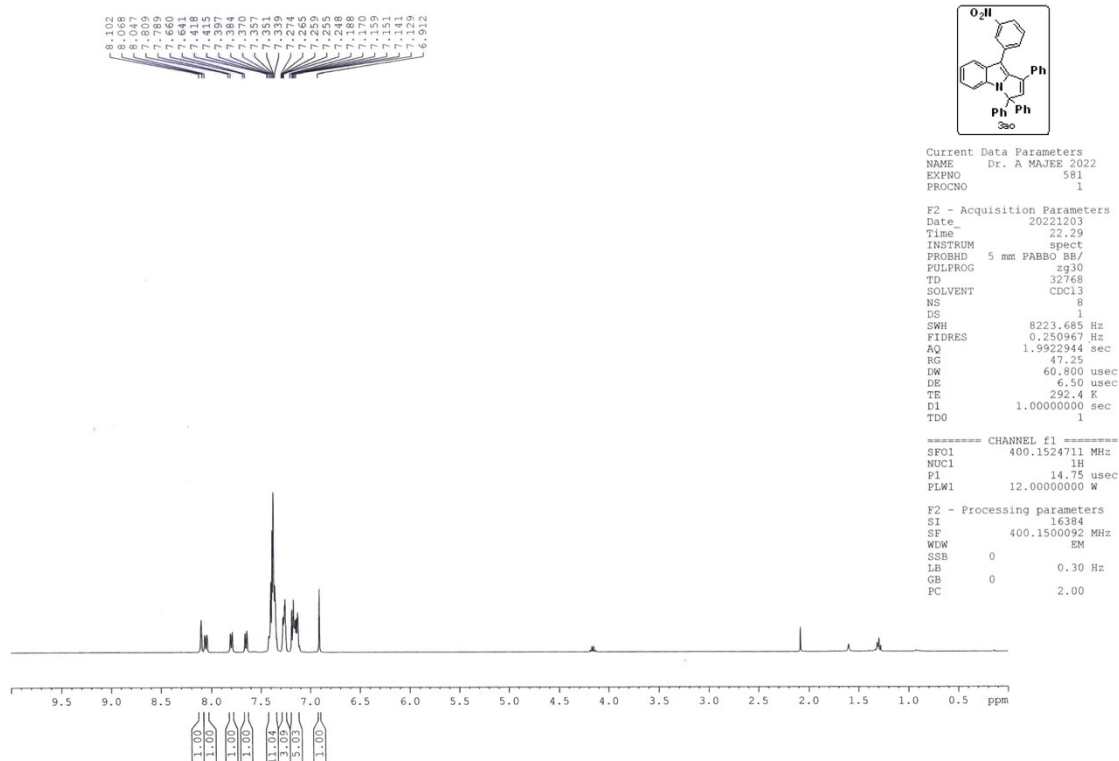
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



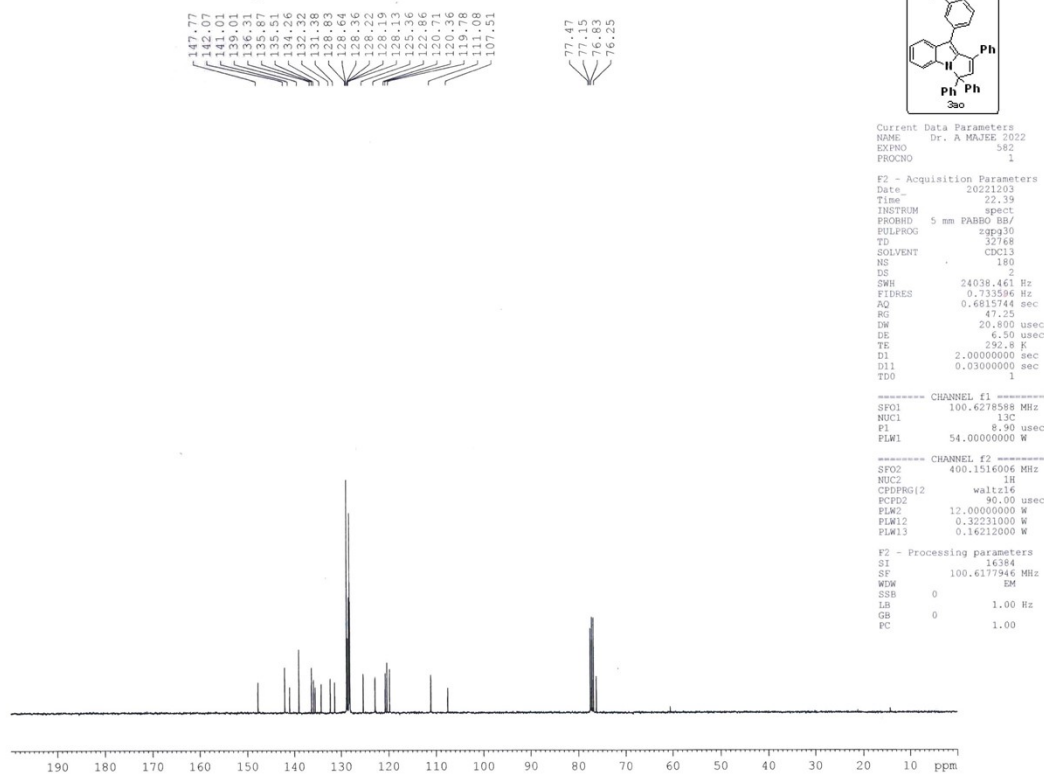
^{19}F NMR : 376 MHz, Solvent : CDCl_3



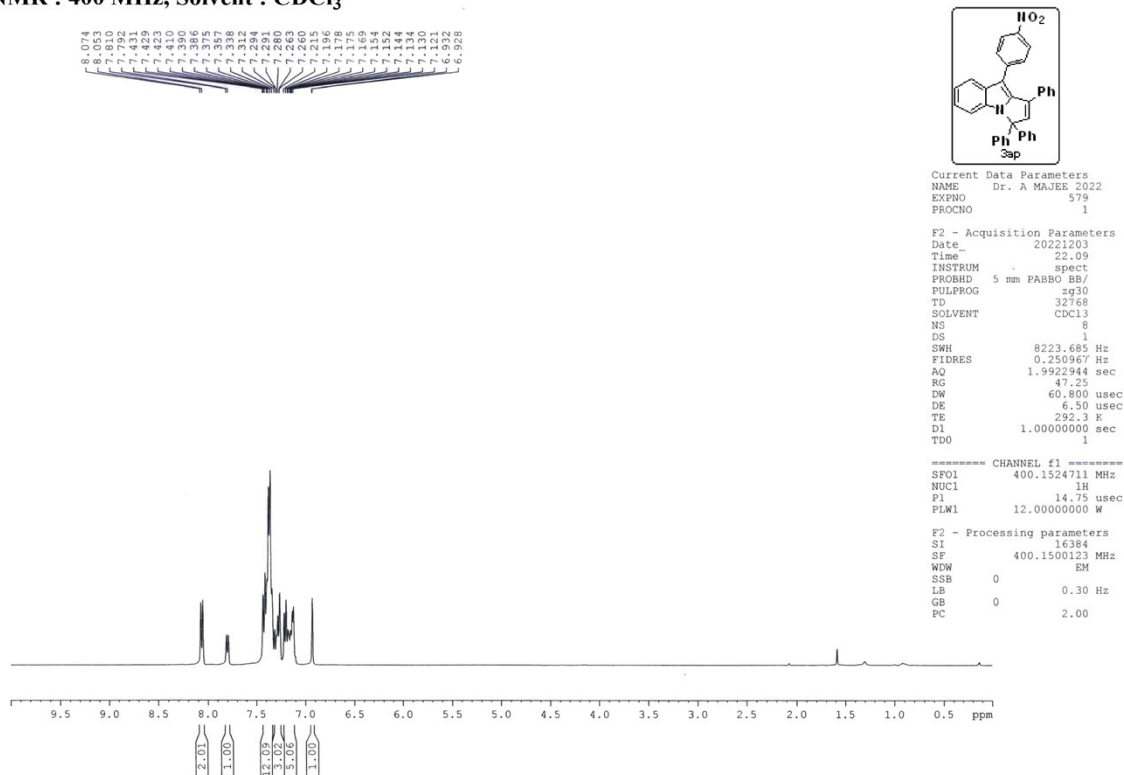
^1H NMR : 400 MHz, Solvent : CDCl_3



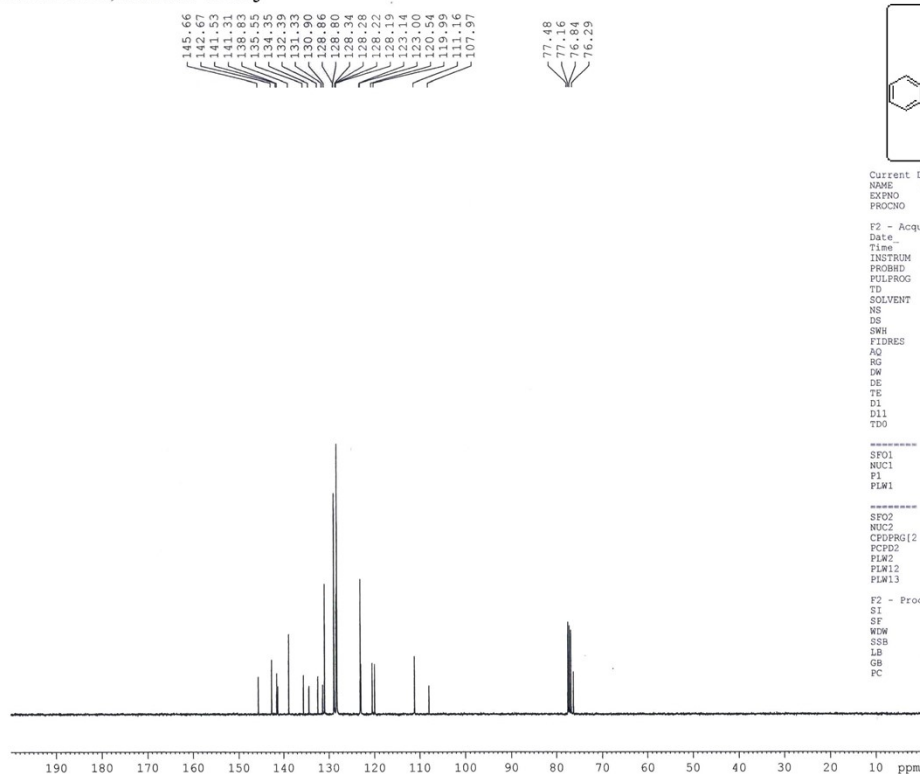
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



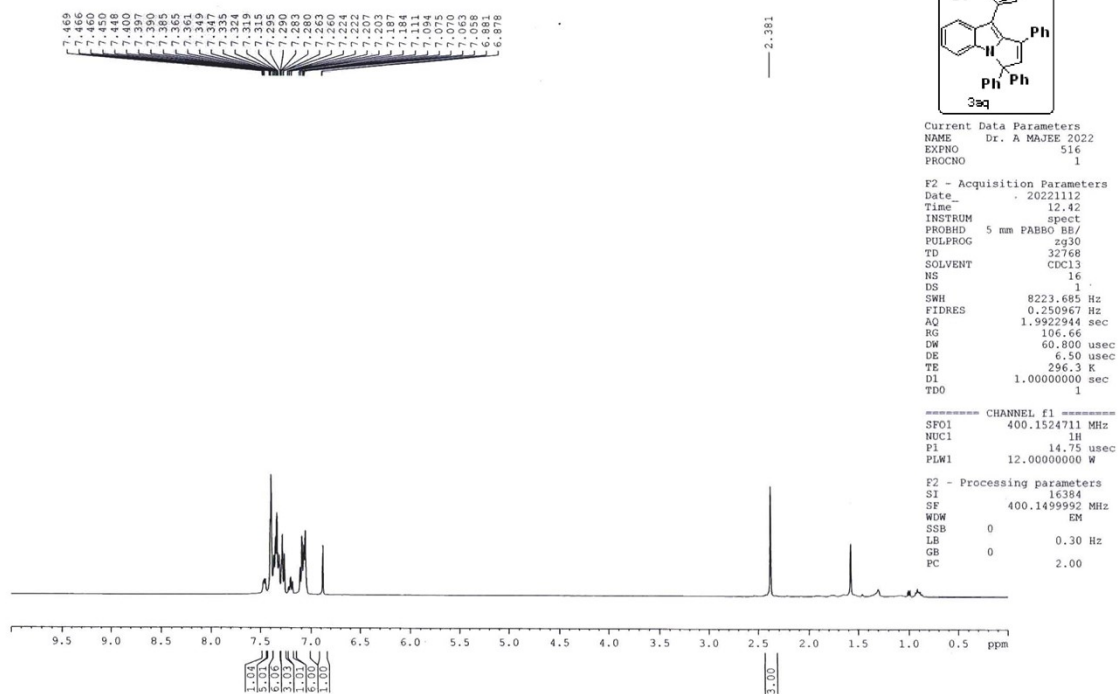
^1H NMR : 400 MHz, Solvent : CDCl_3



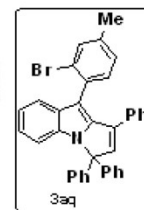
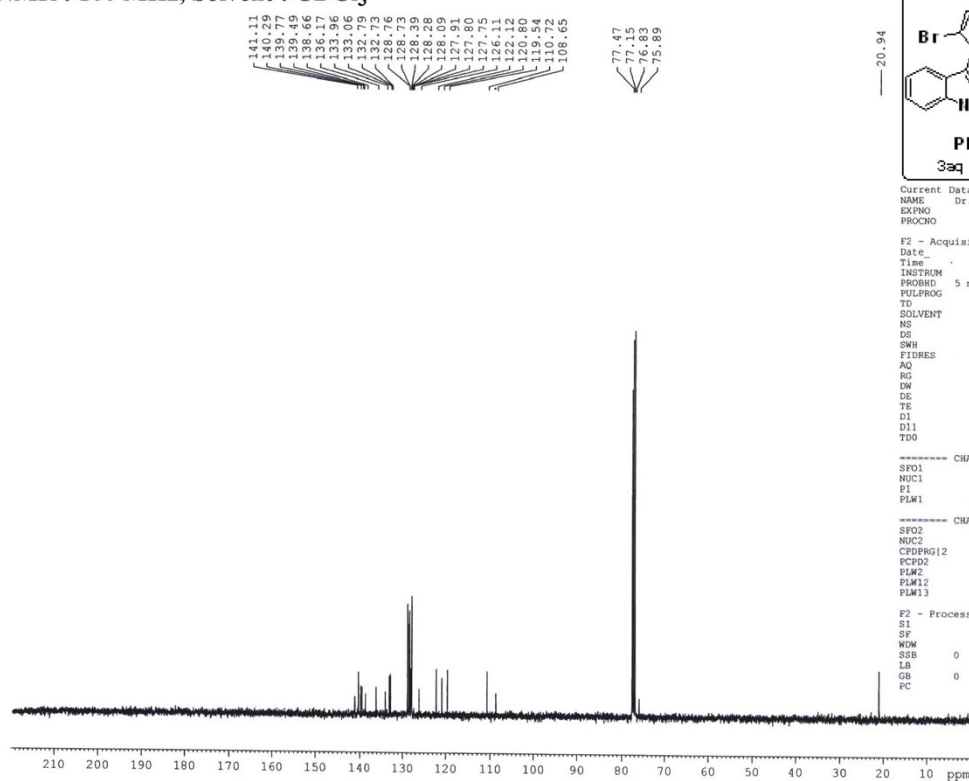
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2022
EXPNO 517
PROCNO 1

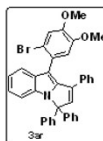
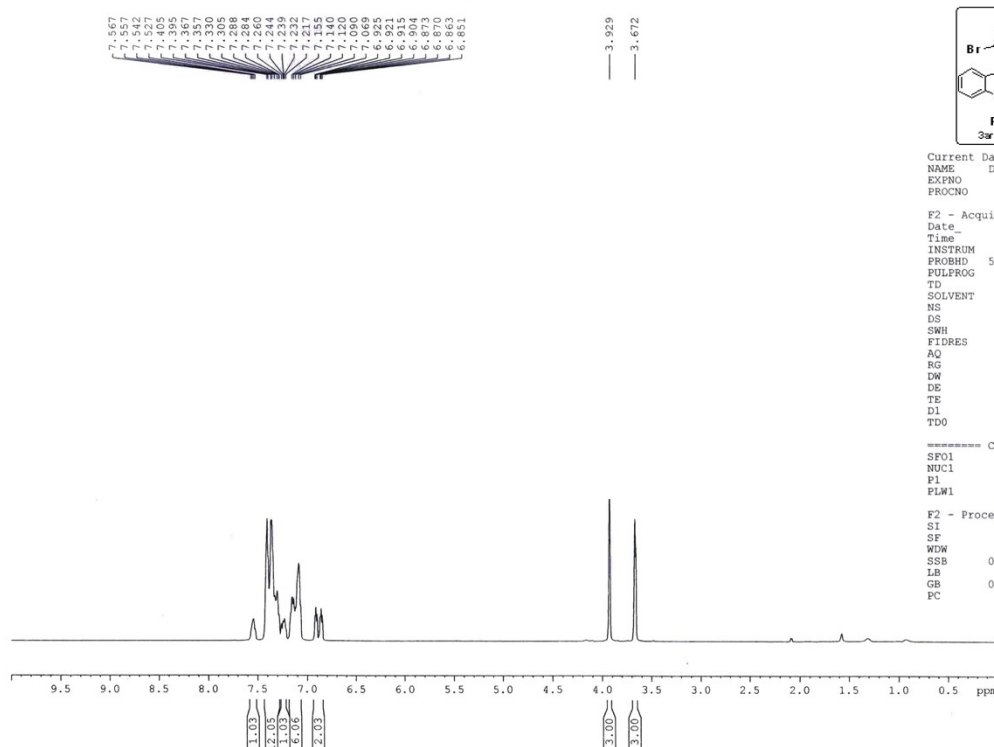
F2 - Acquisition Parameters
Date 20221112
Time 12.55
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 256
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 106.66
DW 20.800 usec
DE 6.50 usec
TE 296.7 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

----- CHANNEL f2 -----
SFO2 400.1516006 MHz
NUC2 ^1H
PCPDPRG2 waltz16
CPDPRG2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6171858 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

^1H NMR : 400 MHz, Solvent : CDCl_3



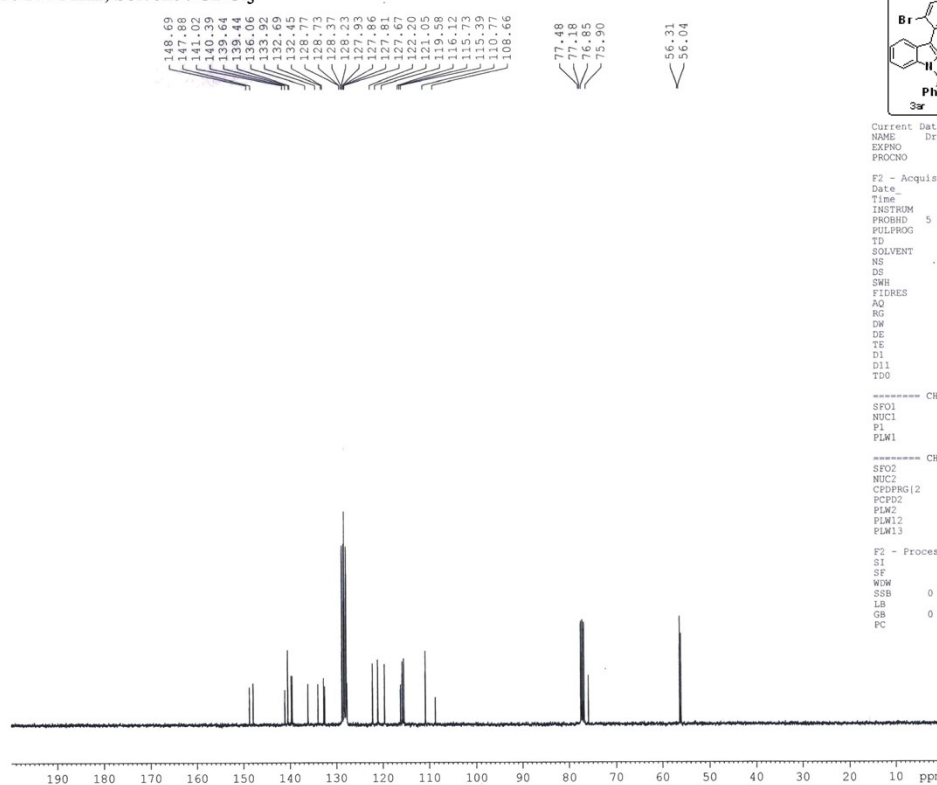
Current Data Parameters
NAME Dr. A MAJEE 2022
EXPNO 597
PROCNO 1

F2 - Acquisition Parameters
Date 20221211
Time 11.13
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 47.25
DW 60.800 usec
DE 6.50 usec
TE 293.3 K
D1 1.00000000 sec
TD0 1

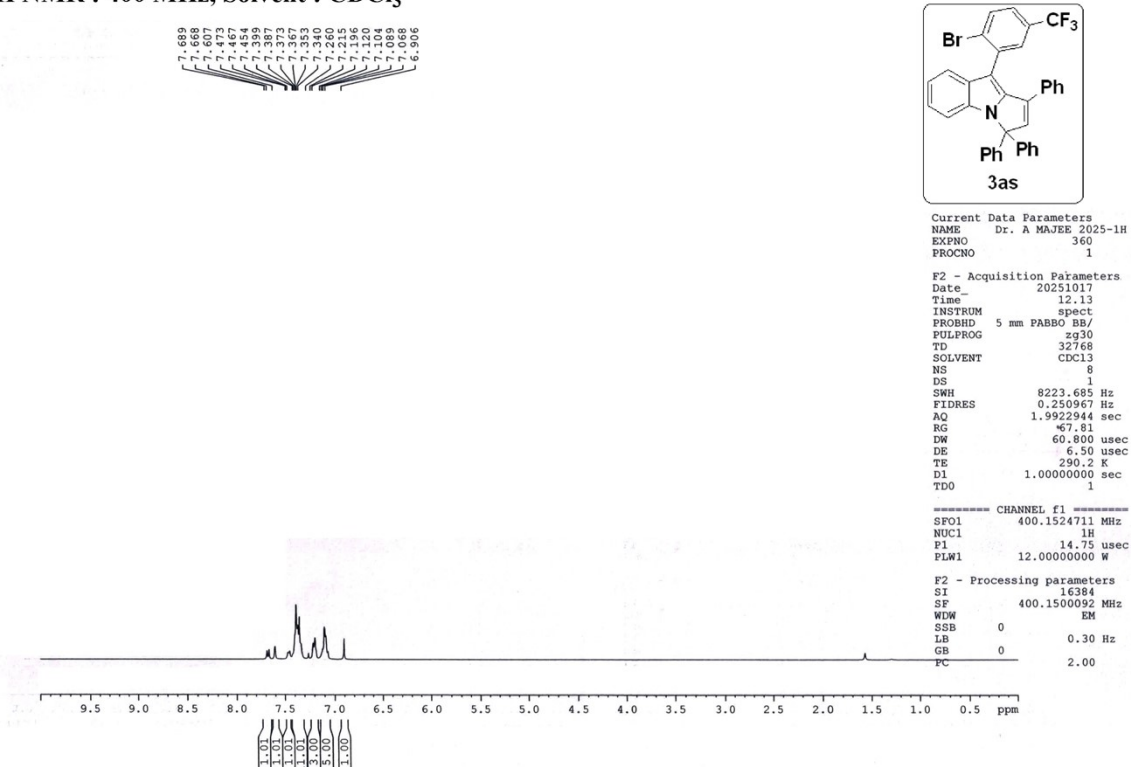
----- CHANNEL f1 -----
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500072 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

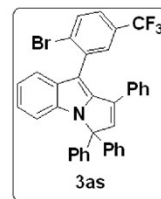
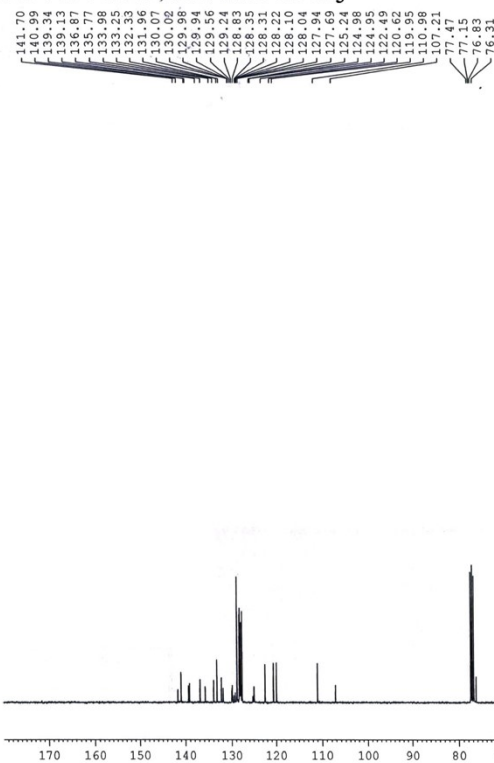
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
 NAME Dr. A MAJEE-2025-13C
 EXPNO 249
 PROCNO 1

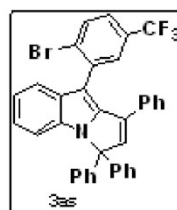
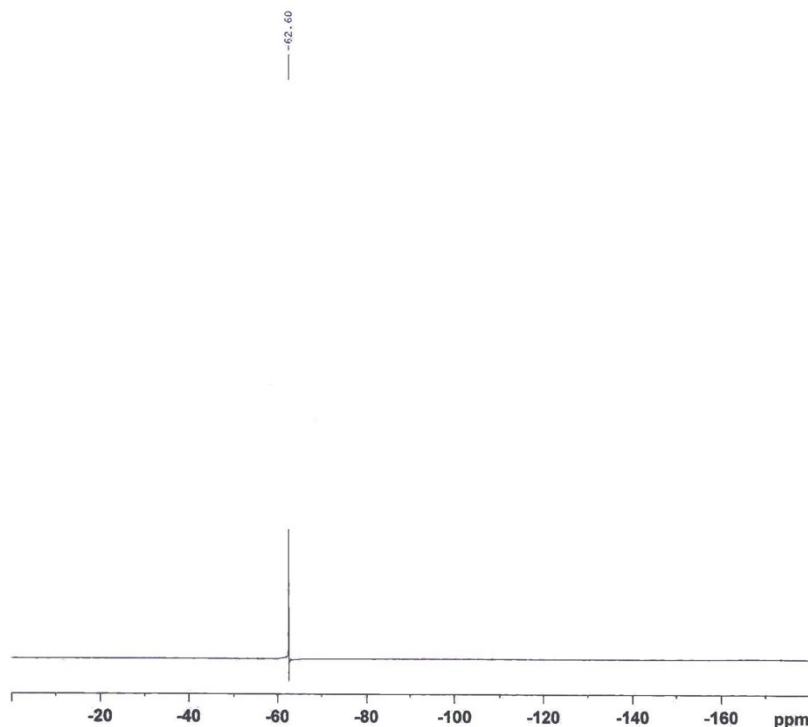
F2 - Acquisition Parameters
 Date_ 20251017
 Time 12.46
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 600
 DS 2
 SWH 24038.461 Hz
 FIDRES 0.733594 Hz
 AQ 0.6815744 sec
 RG 62.69
 LW 20.800 usec
 DE 6.50 usec
 TE 290.6 K
 D1 2.00600000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 100.627858 MHz
 NUC1 13C
 P1 8.90 usec
 PLW1 54.00000000 W

===== CHANNEL f2 =====
 SF02 400.1516006 MHz
 NUC2 1H
 CYPRG2 waltz16
 PCPD2 90.00 usec
 PLW2 12.00000000 W
 PLM12 0.32231000 W
 PLW13 0.16212000 W

F2 - Processing parameters
 SI 16384
 SF 100.6177902 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

^{19}F NMR : 376 MHz, Solvent : CDCl_3

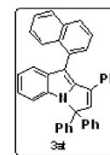
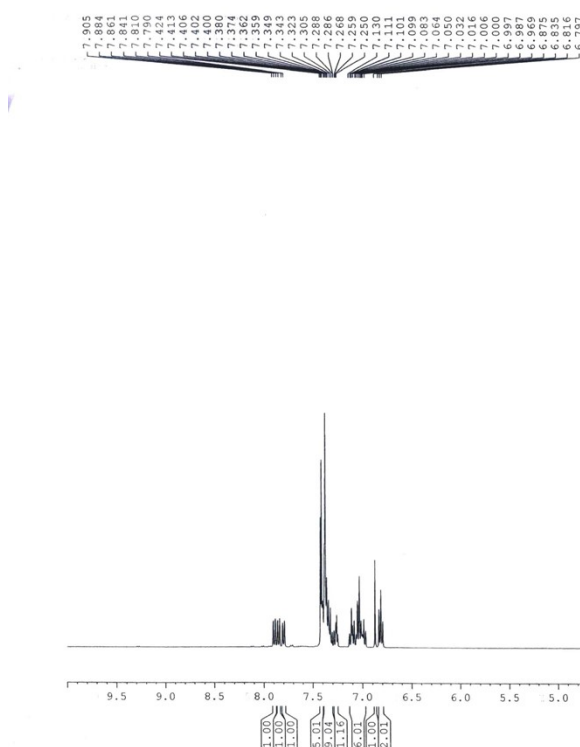


Current Data Parameters
 NAME Dr. A MAJEE 2024
 EXPNO 491
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20241217
 Time 18.13
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 89285.711 Hz
 FIDRES 2.724784 Hz
 AQ 0.1835008 sec
 RG 186.42
 LW 5.600 usec
 DE 6.50 usec
 TE 289.1 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 376.4795333 MHz
 NUC1 19F
 P1 12.50 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 16384
 SF 376.5171850 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

^1H NMR : 400 MHz, Solvent : CDCl_3 

```

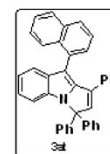
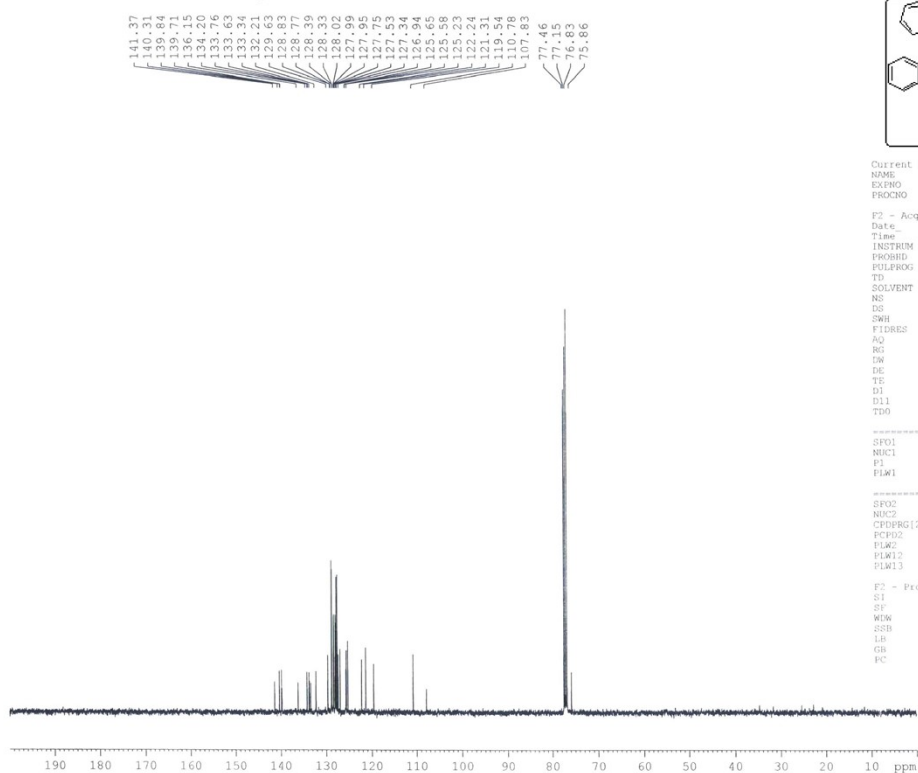
Current Data Parameters
NAME      Dr. A MAJEE 2022
EXPNO     465
PROCNO    1

F2 - Processing Parameters
Date_     20221022
Time      17:55
INSTRUM    spect
PROBHD     5 mm PABBO B1
PULPROG    zgpg30
TD          32768
NS          8
DS          1
SWH         8223.685 Hz
FIDRES     0.250967 Hz
AQ          1.952294 sec
RG          93.46
DW          60.800 usec
DE          6.50 usec
TE          295.3 K
D1          1.0000000 sec
TDO         1

===== CHANNEL f1 =====
SF01       400.1524711 MHz
NUC1        1H
P1          14.75 usec
PLW1       12.0000000 W

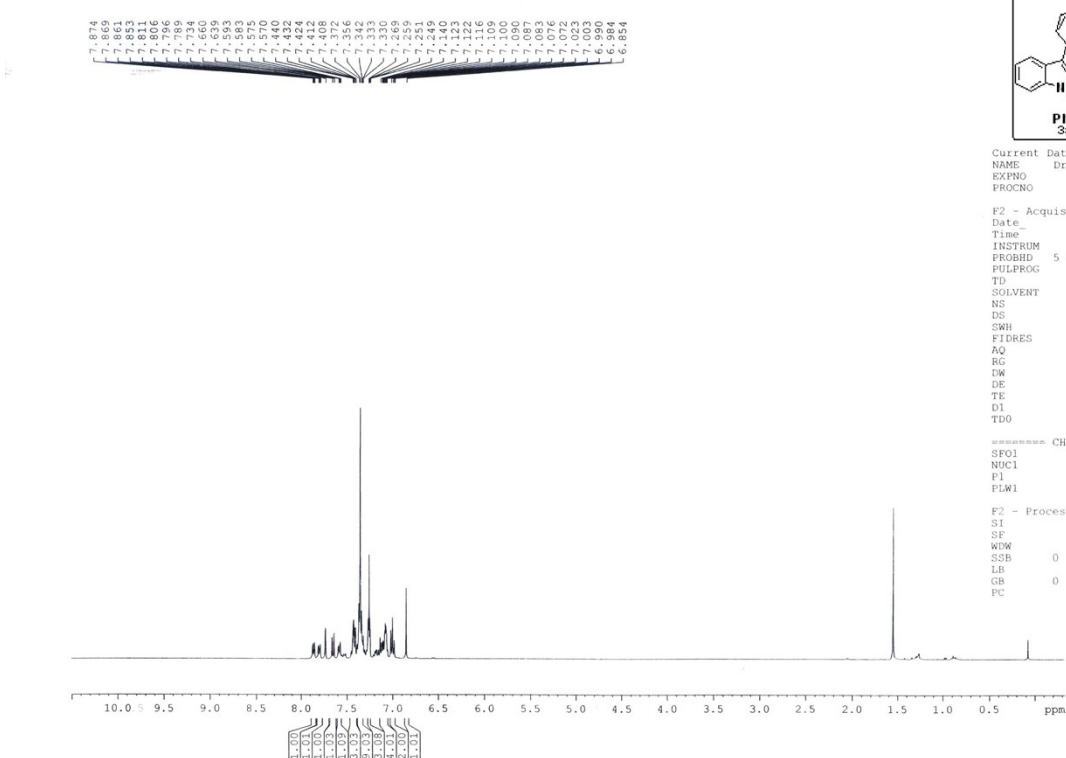
F2 - Processing parameters
SI          16384
SF          400.1500087 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          2.00

```

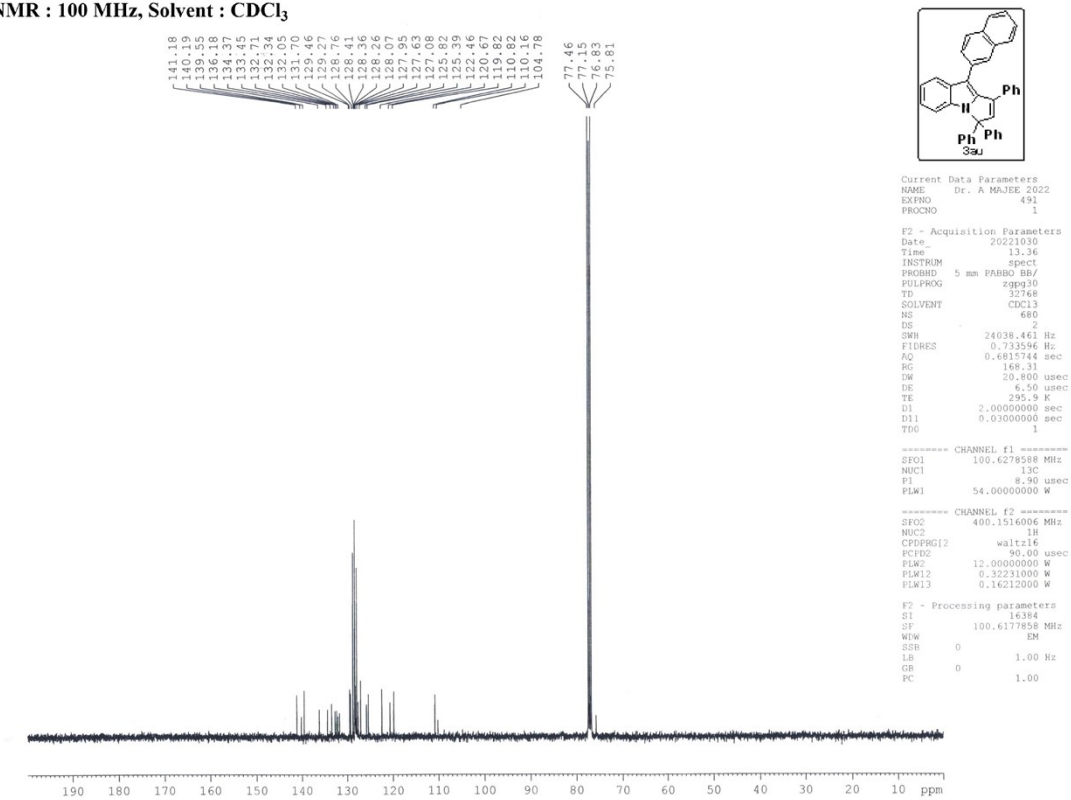
 $^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3 

```
Current Data Parameters
NAME      Dr. A MAJEE 2022
EXPNO     1          466
PROCNO    1
F2 - Acquisition Parameters
=====
NAME      202201122
INSTRUM    18.15
PROBHD     5 mm PABBO/HB
PULPROG    zgpg30
TD          65536
SOLVENT    CDCl3
NS          2
DS          2
SWH         24038.941 Hz
FIDRES     0.733596 sec
AQ          0.6815714 sec
RG          93.46
DE         20.800 usec
RG          6.50 usec
TE          295.7 K
D1          2.00000000 sec
D11         0.03000000 sec
TDO         1
===== CHANNEL f1 =====
SPFO1      100.627858 MHz
NUC1        13C
P1          1
PL1         54.00000000 W
===== CHANNEL f2 =====
SPFO2      400.1516264 MHz
NUC2        1H
P2          12.00000000 usec
PCPD2       wait16
PL2         12.00000000 usec
PL1P2       0.32310000 W
PL1P1       0.16212050 W
F2 - Processing parameters
=====
SI          32
SF          100.617773 MHz
GB          0
WDW         HANN
GB          0
PC          1.00 Hz
RG          0
SC          1.00
```

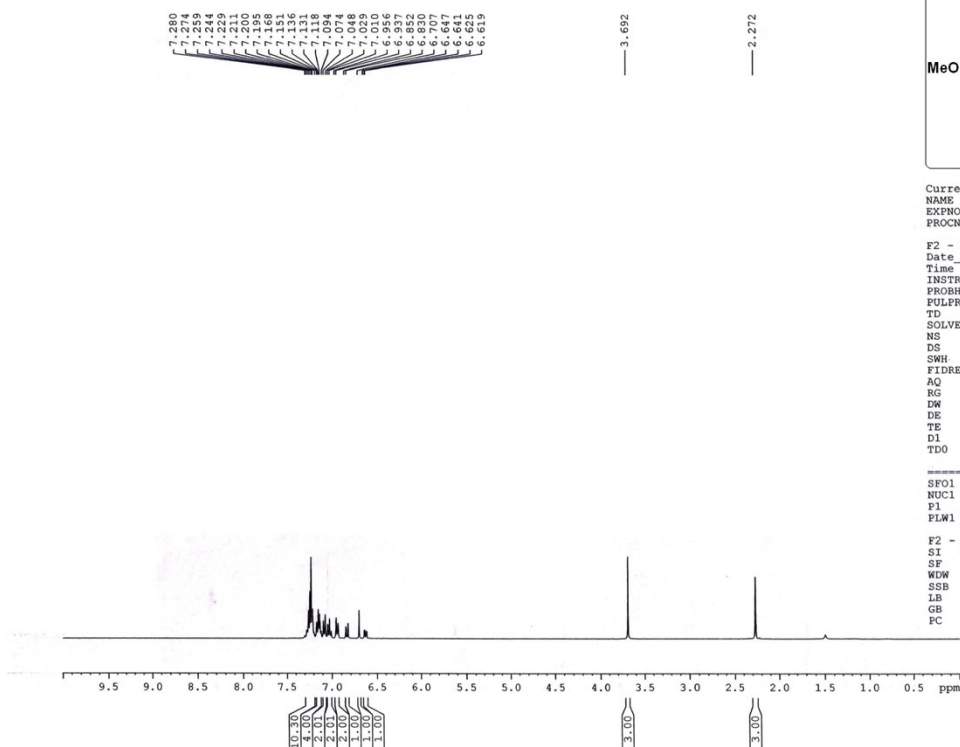
¹H NMR : 400 MHz, Solvent : CDCl₃



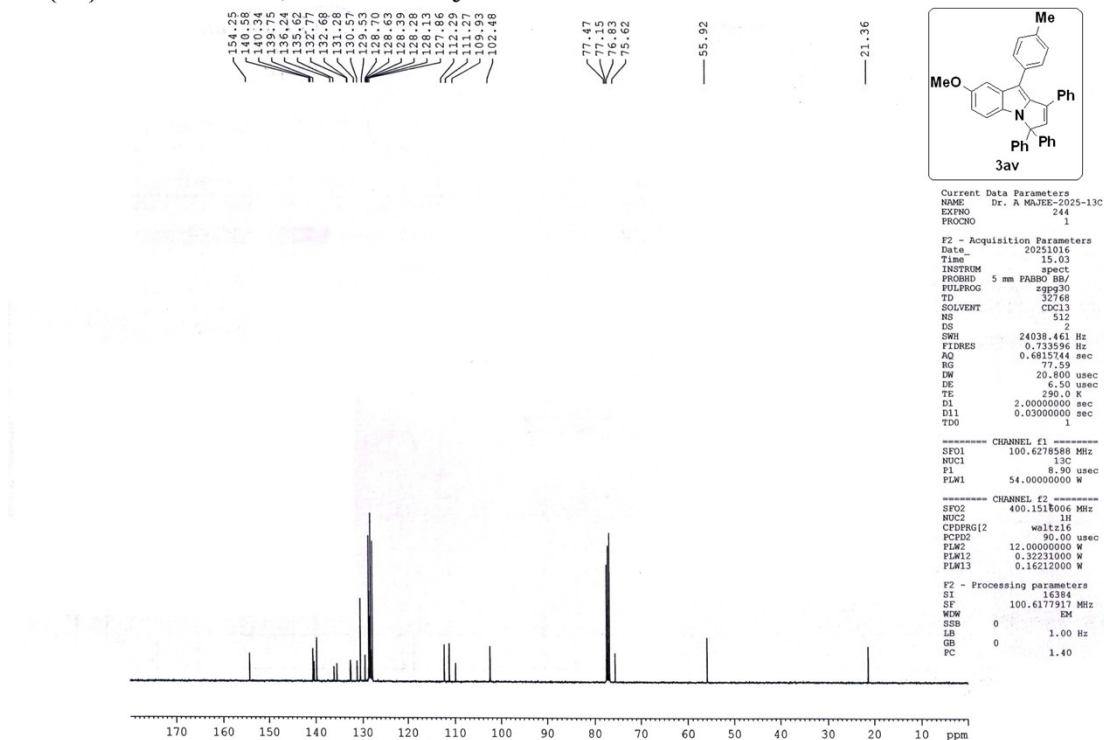
¹³C{¹H} NMR : 100 MHz, Solvent : CDCl₃



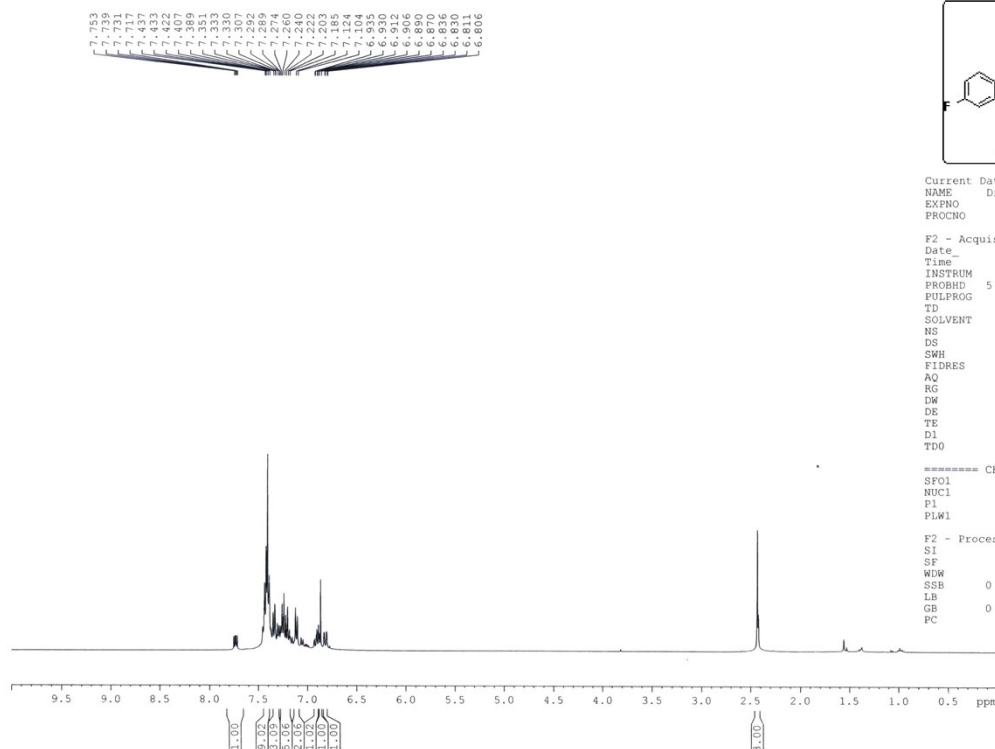
^1H NMR : 400 MHz, Solvent : CDCl_3



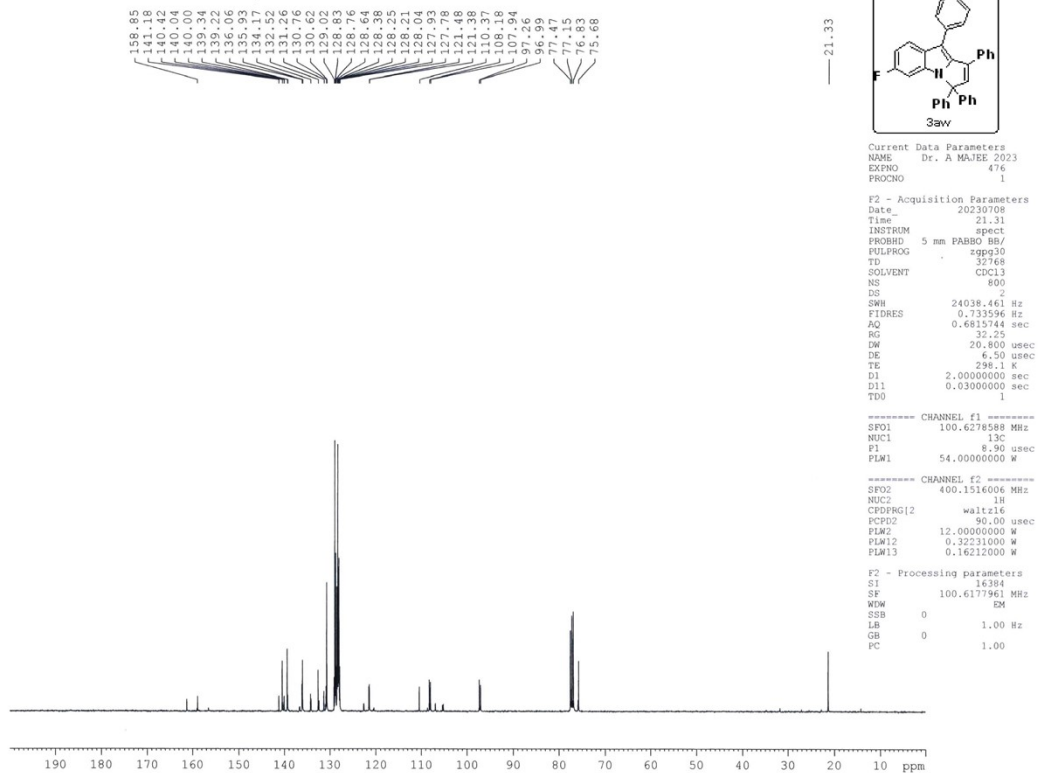
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



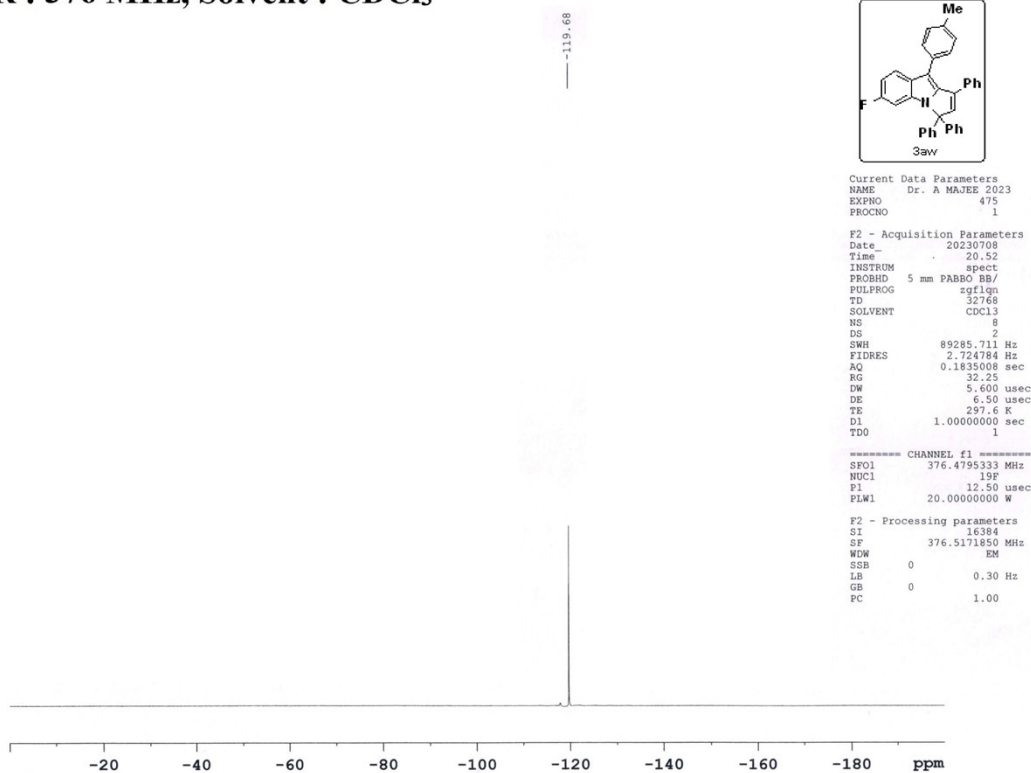
^1H NMR : 400 MHz, Solvent : CDCl_3



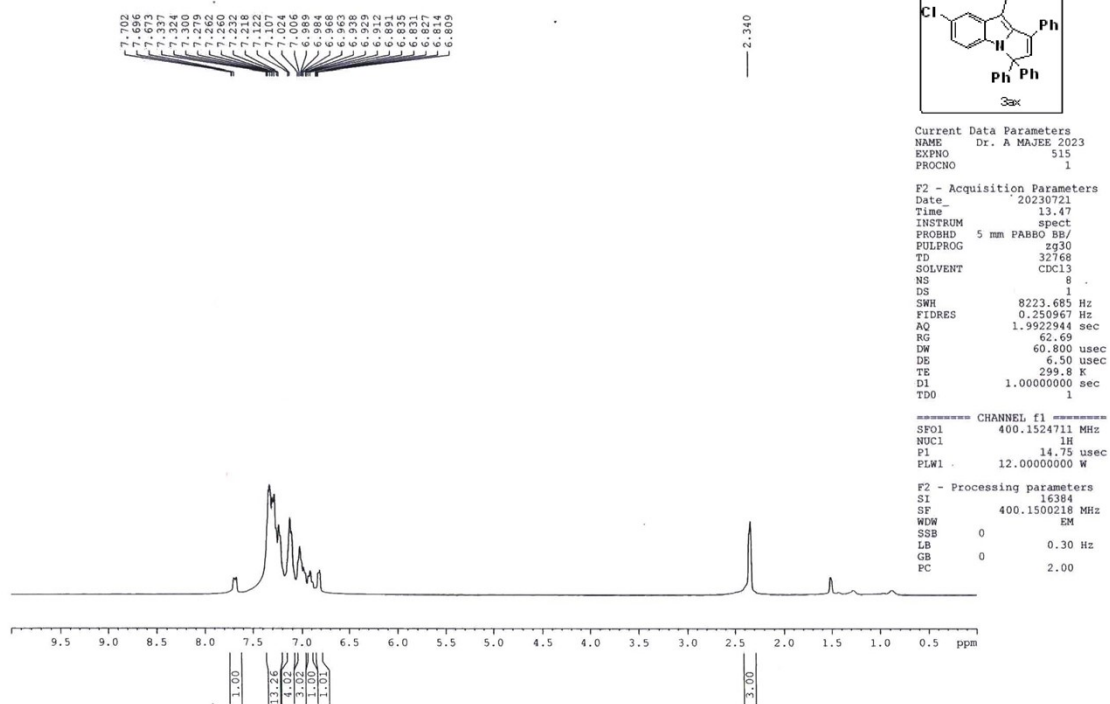
$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



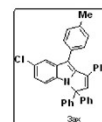
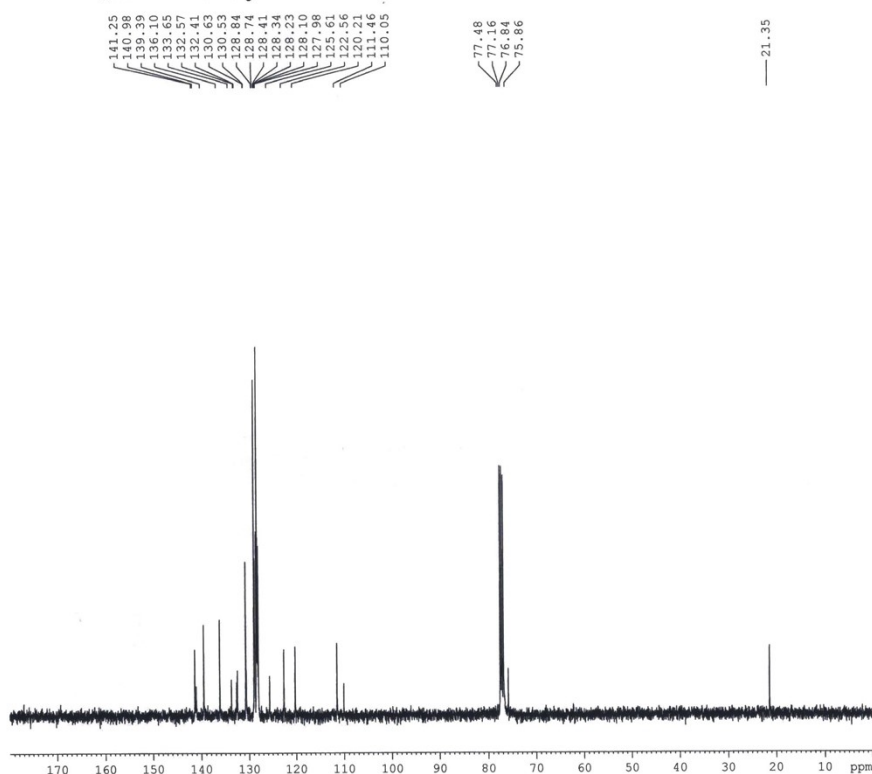
^{19}F NMR : 376 MHz, Solvent : CDCl_3



^1H NMR : 400 MHz, Solvent : CDCl_3



$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



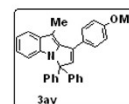
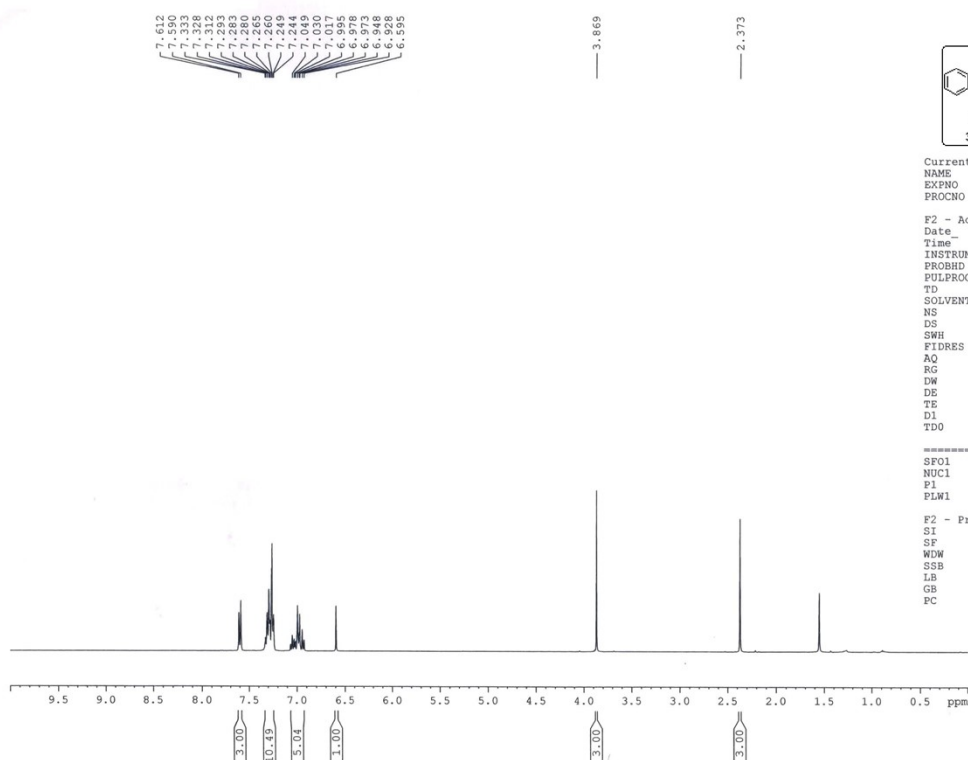
Current Data Parameters
NAME Dr. A MAJEE 2023
EXPNO 516
PROCNO 1

F2 - Acquisition Parameters
Date 20230721
Time 14.08
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 256
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 62.69
DW 20.800 usec
DE 6.50 usec
TE 300.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177858 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



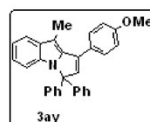
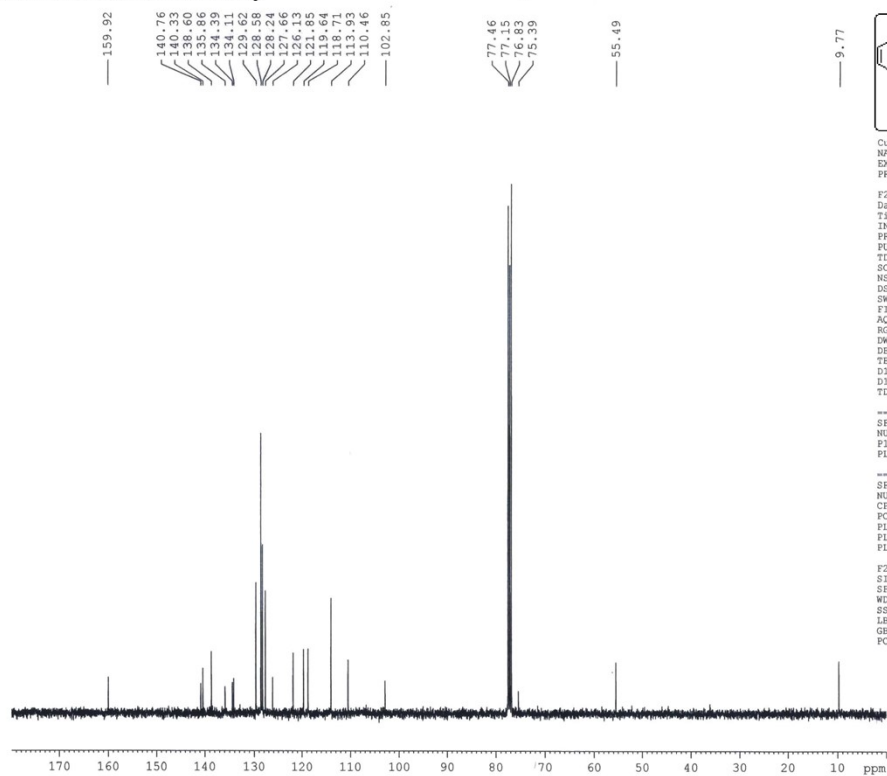
Current Data Parameters
NAME Dr. A MAJEE 2023
EXPNO 730
PROCNO 1

F2 - Acquisition Parameters
Date 20231014
Time 12.33
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 135.7
DW 60.800 usec
DE 6.50 usec
TE 295.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2023
EXPNO 731
PROCNO 1

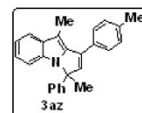
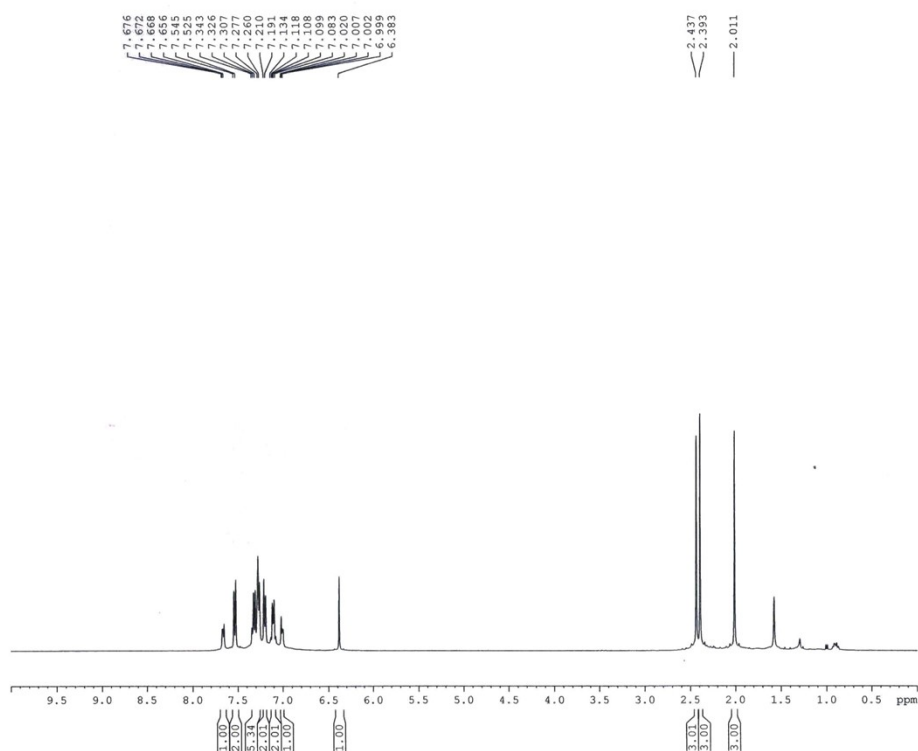
F2 - Acquisition Parameters
Date 20231014
Time 12.58
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 400
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 186.42
DW 20.800 usec
DE 6.50 usec
TE 296.2 K
D1 2.00000000 sec
D11 0.83000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177857 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

^1H NMR : 400 MHz, Solvent : CDCl_3



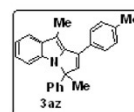
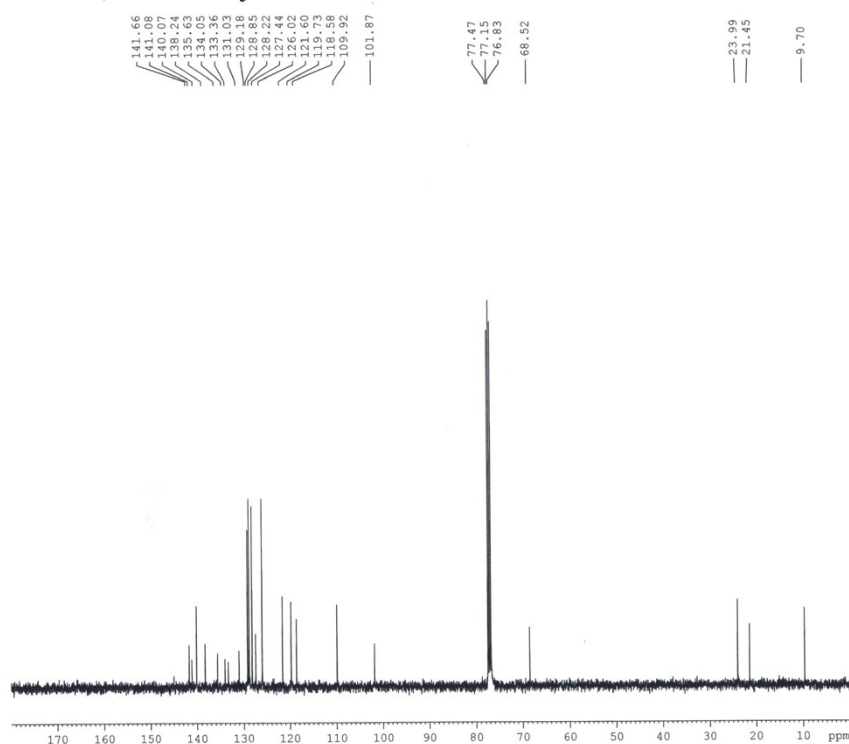
Current Data Parameters
NAME Dr. A MAJEE 2023
EXPNO 635
PROCNO 1

F2 - Acquisition Parameters
Date 20230914
Time 14.09
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 87.66
DW 60.800 usec
DE 6.50 usec
TE 296.4 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500037 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2023
EXPNO 636
PROCNO 1

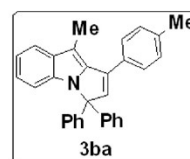
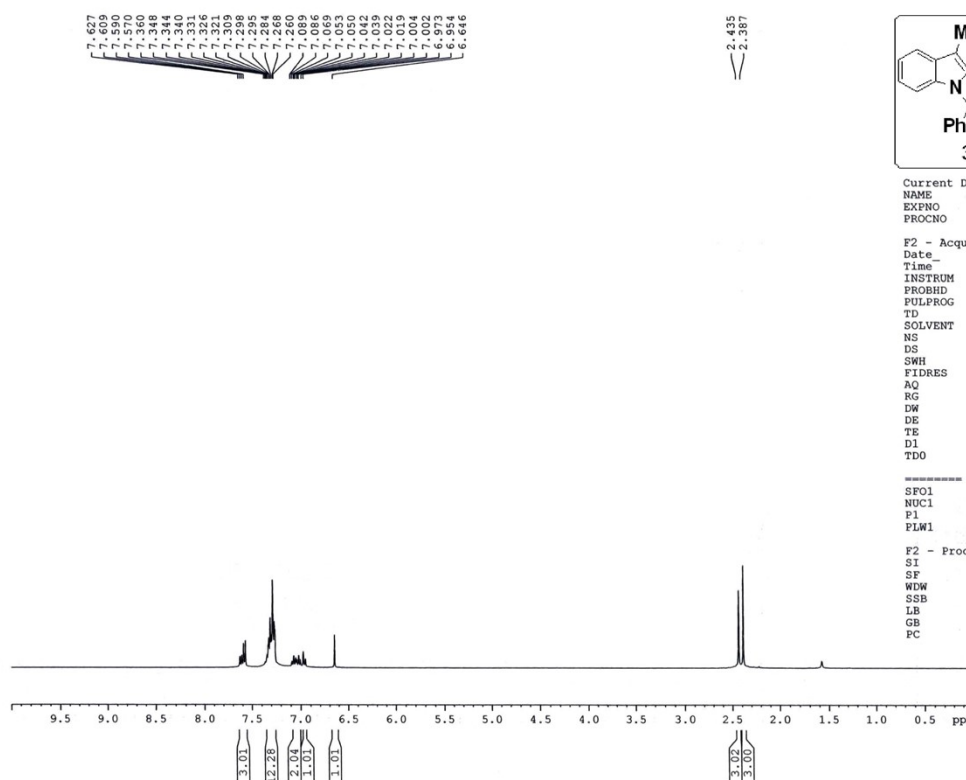
F2 - Acquisition Parameters
Date_ 20230914
Time 14.23
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 256
DS 2
SWH 24039.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 87.46
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177858 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

^1H NMR : 400 MHz, Solvent : CDCl_3

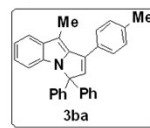
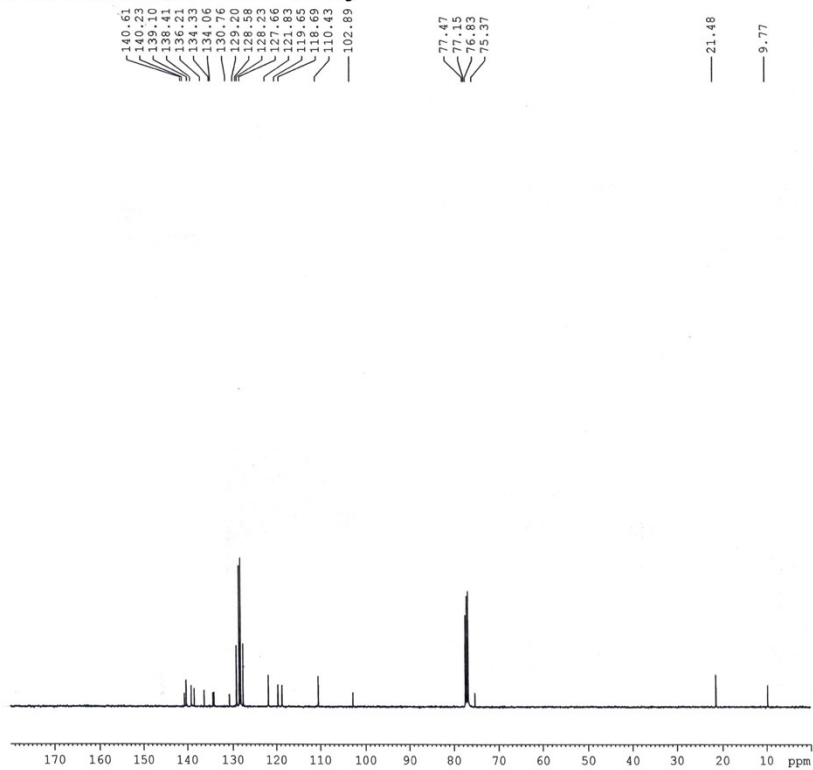


Current Data Parameters
NAME Dr. A MAJEE 2025-1H
EXPNO 356
PROCNO 1

F2 - Acquisition Parameters
Date_ 20251016
Time 15.09
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 77.59
DW 60.800 usec
DE 6.50 usec
TE 289.5 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3 

```
Current Data Parameters
NAME      Dr. A MAJEE-2025-13C
EXPNO      245
PROCNO     1
```

```

F2 - Acquisition Parameters
Date_      20251016
Time       15.39
INSTRUM    spect
PROBHD     5 mm PABBO BB/
FULPROG    zgpg30
TD          32768
SOLVENT     CDCl3
NS          600
DS          2
SWH          24038.46 Hz
FIDRES      0.733596 Hz
AQ          0.6815744 sec
RG          77.59
DW          20.800 usec
DE          6.50 usec
TE          290.4 K
D1          2.00000000 sec
D11         0.03000000 sec
TDO         * 1

```

```

----- CHANNEL f1 -----
SFO1      100.6278588 MHz
NUC1      13C
P1         8.90 usec
PLW1      54.00000000 W

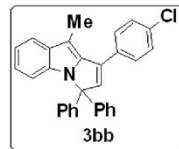
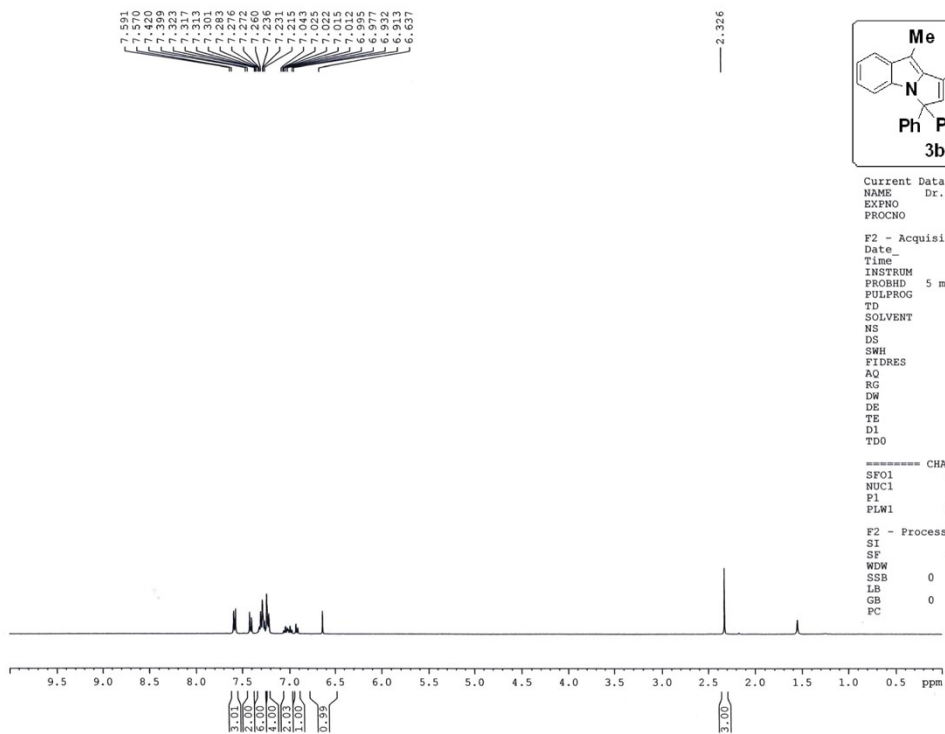
```

```

                                CHANNEL f2
SFO2                400.1516006 MHz
NUC2                  1H
CPDPRG2              waltz16
PCPD2                 90.00 usec
PLW2                 12.00000000 W
PLW12                0.32231000 W
PLW13                0.16212000 W

```

```
F2 - Processing parameters
SI                      16384
SF                      100.6177902 MHz
WDW                      EM
SSB                      0
LB                      1.00 Hz
GB                      0
PC                      1.40
```

 $^1\text{H NMR}$: 400 MHz, Solvent : CDCl_3 

```
Current Data Parameters
NAME      Dr. A MAJEE 2025-1H
EXPNO      370
PROCNO     1
```

```

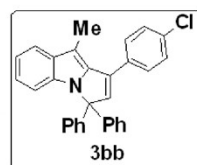
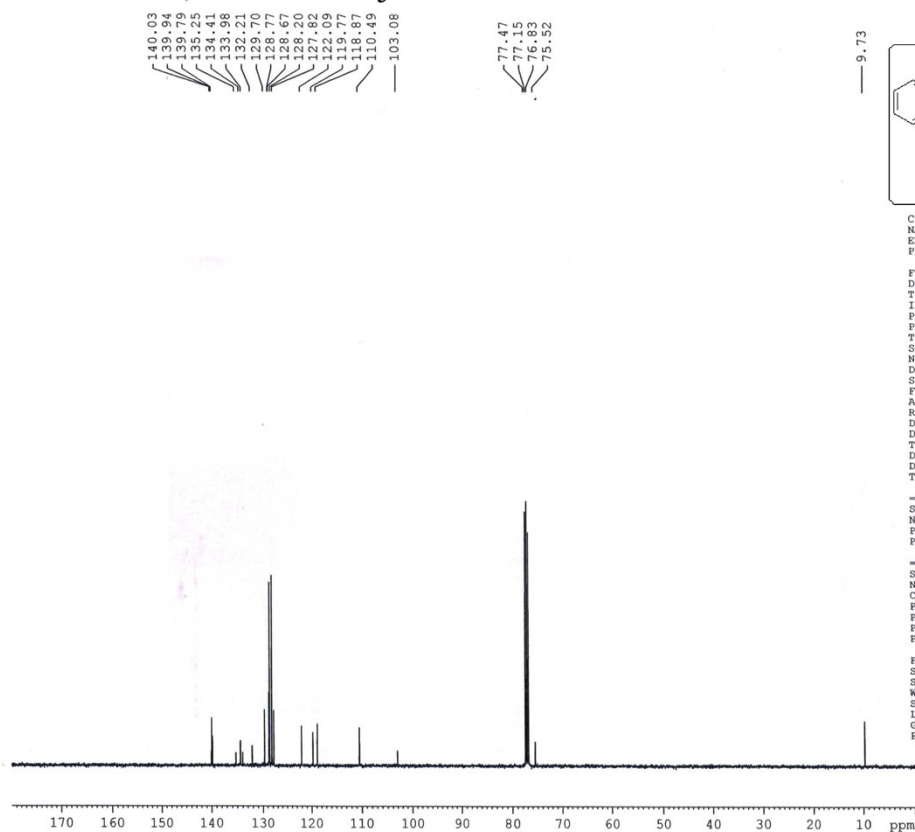
F2 - Acquisition Parameters
-----
Date_      20151021
Time       17.01
INSTRUM    spect
PROBHD     5 mm PABBO BB/
PULPROG    zg30
TD          32768
SOLVENT     CDCl3
NS          8
DS          1
SWH         8223.685 Hz
FID         0.250967 Hz
AQ          1.992294 sec
RG          186.42
DW          68.800 usec
DE          6.50 usec
TE          292.5 K
D1          1.00000000 sec
TD0         1

```

```
===== CHANNEL f1 =====
SFO1      400.1524711 MHz
NUC1      1H
P1         14.75 usec
PLW1      12.00000000 W
```

```
F2 - Processing parameters
SI                      16384
SF                      400.1500187 MHz
WDW                      EM
SSB                      0
LB                      0.30 Hz
GB                      0
PC                      2.00
```

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE-2025-13C
EXPNO 258
PROCNO 1

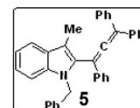
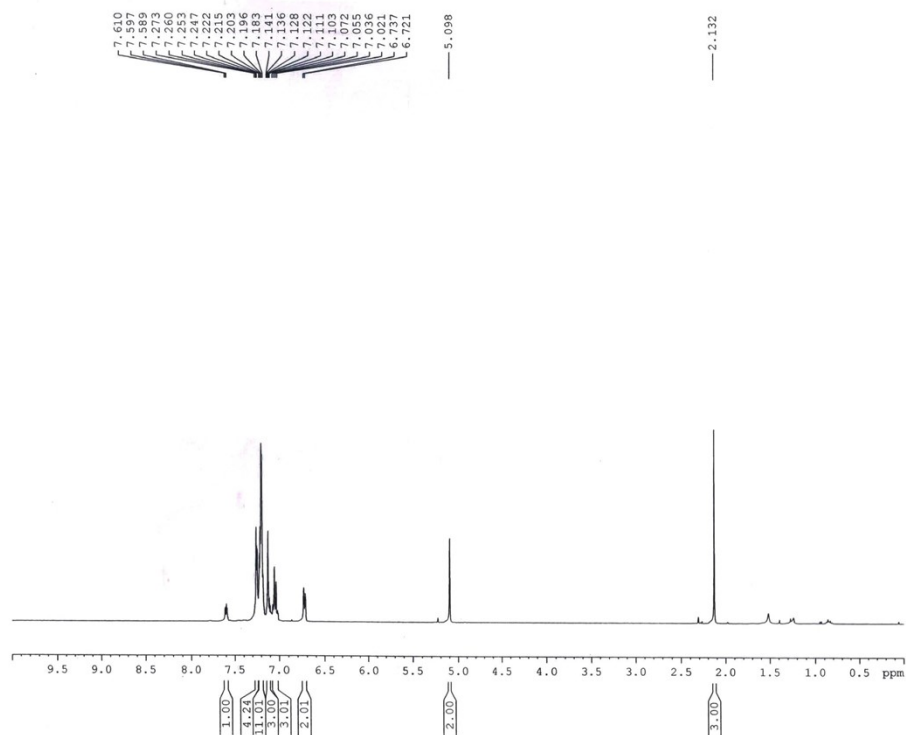
F2 - Acquisition Parameters
Date_ 20251021
Time 16.55
INSTRUM spect
PROBHD 5 mm PABBO BB/
FULPROG zgpg30
TD 32768
SOLVENT CDCl_3
NS 512
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 186.42
DW 20.800 usec
DE 6.50 usec
TE 293.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SF01 100.6278588 MHz
NUC1 ^{13}C
P1 8.90 usec
PLW1 54.00000000 W

===== CHANNEL f2 =====
SF02 400.1516006 MHz
NUC2 ^1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177873 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

^1H NMR : 400 MHz, Solvent : CDCl_3



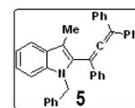
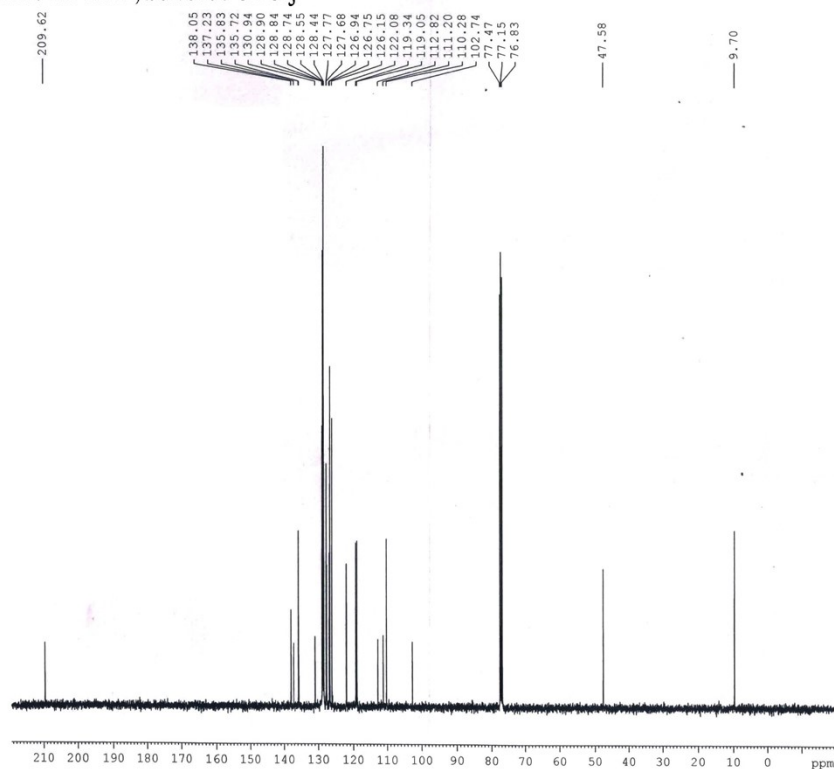
Current Data Parameters
NAME Dr. A MAJEE 2024
EXPNO 407
PROCNO 1

F2 - Acquisition Parameters
Date_ 20241119
Time 19.00
INSTRUM spect
PROBHD 5 mm PABBO BB/
FULPROG zg30
TD 32768
SOLVENT CDCl_3
NS 8
DS 1
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9922944 sec
RG 77.55
DW 60.800 usec
DE 6.50 usec
TE 293.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SF01 400.1524711 MHz
NUC1 ^1H
P1 14.75 usec
PLW1 12.00000000 W

F2 - Processing parameters
SI 16384
SF 400.1500313 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

$^{13}\text{C}\{^1\text{H}\}$ NMR : 100 MHz, Solvent : CDCl_3



Current Data Parameters
NAME Dr. A MAJEE 2024
EXPNO 408
PROCNO 1

F2 - Acquisition Parameters
Date_ 20241119
Time 19:31
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 512
DS 2
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 77.59
DW 20.800 usec
DE 6.50 usec
TE 293.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

----- CHANNEL f1 -----
SFO1 100.6278588 MHz
NUC1 13C
P1 8.90 usec
PLW1 54.00000000 W

----- CHANNEL f2 -----
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLW2 12.00000000 W
PLW12 0.32231000 W
PLW13 0.16212000 W

F2 - Processing parameters
SI 16384
SF 100.6177887 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

