

# Nickel-Catalyzed Isocyanide Insertion Reaction with Aromatic Amines: Direct Access to Open-Chain Guanidines

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## 1. General Information

All reactions were carried out in flame-dried reaction vessels with Teflon screw caps under air. Solvents were purified and dried according to standard methods prior to use. Flash column chromatography was performed on silica gel (200-300 mesh) with the indicated solvent mixtures. TLC analysis was performed on pre-coated, glass-backed silica gel plates and visualized with UV light. The heating reaction used oil bath as heat source.

The  $^1\text{H}$  and  $^{13}\text{C}$  spectra were recorded on a Bruker 400 or 500 AV spectrometer. Chemical shifts ( $\delta$ ) were reported as parts per million (ppm) downfield from tetramethylsilane. Coupling constant (J) was reported in hertz unit (Hz). The high-resolution mass spectra (HRMS) were recorded on an Agilent 6210 LC/TOF spectrometer. The crystal measurement was recorded on an Empyrean X-ray diffractometer.

All the amines and *tert*-butyl isocyanide are commercially available and used as received.

## 2. Nickel-Catalyzed Isocyanide Insertion Reaction with Aromatic Amines

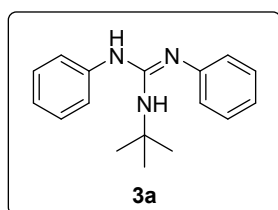
### 2.1 General procedure and characterization of products

To a 25-mL flame-dried Schlenk tube containing a stir bar was added aromatic amine **1** (0.6 mmol), *tert*-butyl isocyanide **2a** (0.2 mmol, 16.7 mg), NiBr<sub>2</sub> (0.01 mmol, 5 mol%, 2.18 mg), anisole (0.5 mL) sequentially. The tube was sealed and stirred at 90 °C for 3 h. After the completion, the reaction mixture was concentrated and purified by silica gel column chromatography to give the product **3**.

#### Characterization of products

*Note: The <sup>13</sup>C NMR spectra of guanidines seem to be abnormal presumably due to the intrinsic nature of these compounds.*

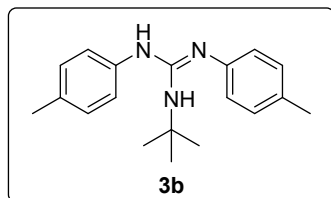
#### *N*-(1,1-Dimethylethyl)-*N'*,*N''*-diphenylguanidine (**3a**)<sup>1</sup>



Purified by silica gel column chromatography (silica gel, petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as dark brown solid (27.8 mg, 75% yield).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.30 (dd, *J* = 8.0, 7.2 Hz, 4H), 7.03 (d, *J* = 7.2 Hz, 6H), 5.58 (s, 1H), 4.00 (s, 1H), 1.46 (s, 9H).

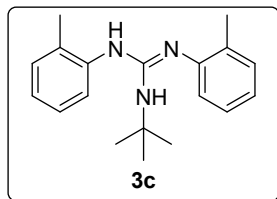
#### *N*-(1,1-dimethylethyl)-*N'*,*N''*-di-*p*-toluenylguanidine (**3b**)<sup>1</sup>



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (38.4 mg, 65% yield).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.13 (d, *J* = 6.4 Hz, 4H), 6.95 (s, 4H), 5.55 (s, 1H), 3.95 (s, 1H), 2.34 (s, 6H), 1.48 (s, 9H).

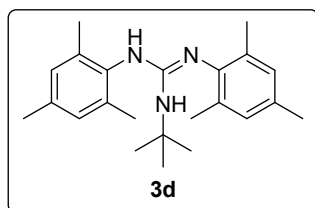
#### *N*-(1,1-dimethylethyl)-*N'*,*N''*-di-*o*-methylphenyl guanidine (**3c**)



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as red liquid (27.1 mg, 46% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.28–6.94 (m, 8H), 5.28 (d, *J* = 15.2 Hz, 1H), 3.77 (s, 1H), 2.30 (d, *J* = 44.0 Hz, 6H), 1.53 (d, *J* = 6 Hz, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 145.6, 130.8, 126.8, 125.2, 122.6, 121.9, 50.9, 29.4, 18.1. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>26</sub>N<sub>3</sub> 296.21212; found 296.21154.

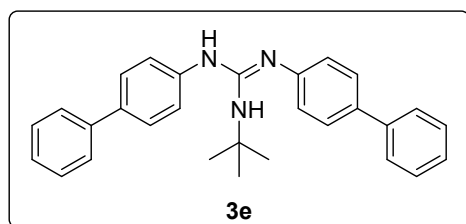
***N*-(1,1-dimethylethyl)-*N'*,*N''*-bis-2,4,6-trimethylphenyl guanidine (3d)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (38.6 mg, 55% yield). mp: 125-127 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 6.96 (s, 2H), 6.86 (s, 2H), 4.67 (s, 1H), 3.42 (s, 1H), 2.35 (d, *J* = 7.2 Hz, 9H), 2.30 (s, 3H), 2.24 (s, 6H), 1.49 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 145.1, 136.8, 132.3, 131.0, 129.3, 128.8, 29.4, 20.8, 18.5. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>34</sub>N<sub>3</sub> 352.27472; found 352.27381.

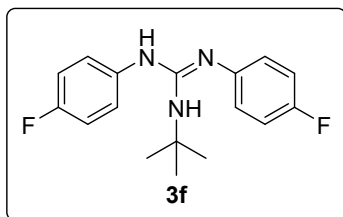
***N*-(1,1-dimethylethyl)-*N'*,*N''*-biphenyl guanidine (3e)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as dark brown solid (52.8 mg, 63% yield). mp: 64-66 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.56 (dd, *J* = 15.6, 7.2 Hz, 8H), 7.44 (t, *J* = 7.4 Hz, 4H), 7.33 (t, *J* = 7.3 Hz, 2H), 7.12 (d, *J* = 8.0 Hz, 4H), 5.80 (s, 1H), 4.31 (s, 1H), 1.51 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 146.4, 140.9, 135.4, 128.8, 128.7, 128.1, 126.8, 126.7, 122.6, 51.4, 29.6. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>29</sub>H<sub>39</sub>N<sub>3</sub> 420.24342; found 420.24296.

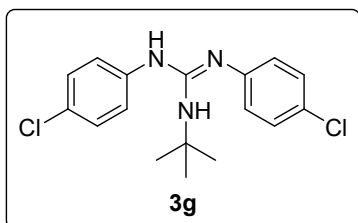
***N*-(1,1-dimethylethyl)-*N'*,*N''*-di-*p*-fluorophenylguanidine (3f)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (24 mg, 55% yield). mp: 129-131 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.26 (d, *J* = 7.6 Hz, 4H), 6.96 (s, 4H), 5.51 (s, 1H), 3.91 (s, 1H), 1.44 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 146.4, 129.4, 123.4, 51.4, 29.5. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>20</sub>F<sub>2</sub>N<sub>3</sub> 304.16198; found 304.16146.

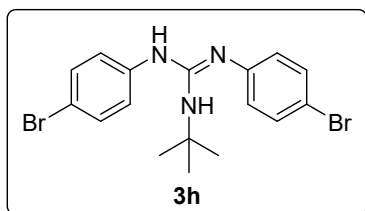
***N*-(1,1-dimethylethyl)-*N'*,*N''*-di-*p*-chlorophenylguanidine (3g)<sup>2</sup>**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (34.2 mg, 51% yield). mp: 133-135 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.23 (d, *J* = 8.8 Hz, 4H), 6.93 (s, 4H), 5.48 (s, 1H), 3.88 (s, 1H), 1.41 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 146.4, 129.4, 128.0, 123.5, 51.4, 29.5. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>20</sub>N<sub>3</sub>Cl<sub>2</sub> 336.10288; found 336.10254..

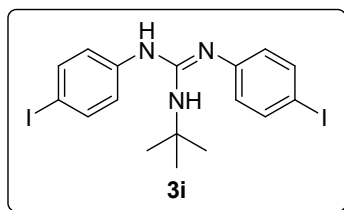
***N*-(1,1-dimethylethyl)-*N'*,*N''*-di-*p*-bromophenylguanidine (3h)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (42.5 mg, 50% yield). mp: 133-135 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.38–7.34 (m, 4H), 6.87 (d, *J* = 9.6 Hz, 4H), 5.46 (s, 1H), 3.90 (s, 1H), 1.41 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 146.2, 132.3, 123.8, 51.4, 29.6. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>20</sub>N<sub>3</sub>Br<sub>2</sub> 424.00185; found 424.00186.

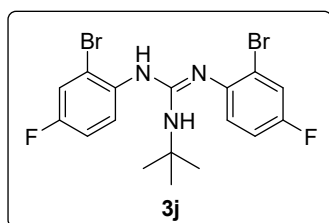
***N*-(1,1-dimethylethyl)-*N'*,*N''*-bis-*p*-iodophenylguanidine (3i)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as white solid (48.8 mg, 47% yield). mp: 134-136 °C.

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.55 (d, *J* = 6.8 Hz, 4H), 6.75 (s, 4H), 5.47 (s, 1H), 3.92 (s, 1H), 1.41 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 146.0, 138.3, 124.1, 51.5, 29.5. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>20</sub>I<sub>2</sub>N<sub>3</sub> 519.97411; found 519.97408.

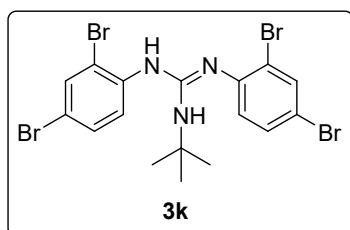
***N*-(1,1-dimethylethyl)- *N'*,*N''*-di-2-bromo-4-fluorophenylguanidine (3j)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as dark brown liquid (59.9 mg, 65% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.30 (d, *J* = 7.6 Hz, 3H), 6.97 (s, 3H), 5.75 (s, 1H), 3.85 (s, 1H), 1.47 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 144.0, 133.9, 124.3, 119.8, 118.2, 115.1, 51.5, 29.8. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>N<sub>3</sub>Br<sub>2</sub>F<sub>2</sub> 459.98301; found 459.98297.

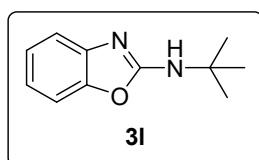
***N*-(1,1-dimethylethyl)- *N'*,*N''*-bis-2,4-dibromophenylguanidine (3k)**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine = 15:1:0.1) as dark brown liquid (53.5 mg, 46% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.67 (dd, *J* = 15.2, 6 Hz, 3H), 7.40–7.28 (m, 2H), 6.76 (dd, *J* = 29.6, 8 Hz, 1H), 5.96 (s, 1H), 3.89 (s, 1H), 1.44 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 148.0, 147.7, 135.1, 134.4, 131.2, 131.0, 52.4, 51.7, 29.9, 28.0. **HRMS (ESI) m/z:** [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>Br<sub>4</sub>N<sub>3</sub> 579.82287; found 579.82282.

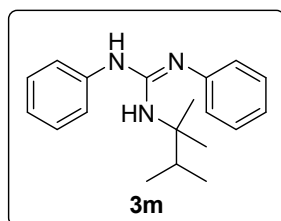
***N*-(1,1-Dimethylethyl)-2-benzoxazamine (3l)<sup>3</sup>**



Purified by silica gel column chromatography (petroleum ether/ethyl acetate = 50:1) as dark brown liquid (30.6 mg, 60% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.41 (d, *J* = 7.8 Hz, 1H), 7.29 (d, *J* = 7.8 Hz, 1H), 7.19 (t, *J* = 7.7 Hz, 1H), 7.05 (t, *J* = 7.7 Hz, 1H), 6.08 (br, 1H), 1.54 (s, 9H).

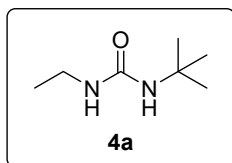
### 1,2-diphenyl-3-(2,4,4-trimethylpentan-2-yl)guanidine (3m)



Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine =10:1:0.1) as white solid (23.3 mg, 36% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 7.19 (t, *J* = 7.8 Hz, 4H), 6.92 (s, 6H), 5.45 (s, 1H), 3.86 (s, 1H), 1.77 (s, 2H), 1.38 (s, 6H), 0.92 (s, 9H). **<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 145.9, 129.4, 122.9, 55.0, 51.8, 31.7, 31.62, 29.9. **HRMS (ESI)** *m/z*: [M+H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>30</sub>N<sub>3</sub> 324.24342; found 324.24337.

### 1-(tert-butyl)-3-ethylurea (4a)<sup>4</sup>

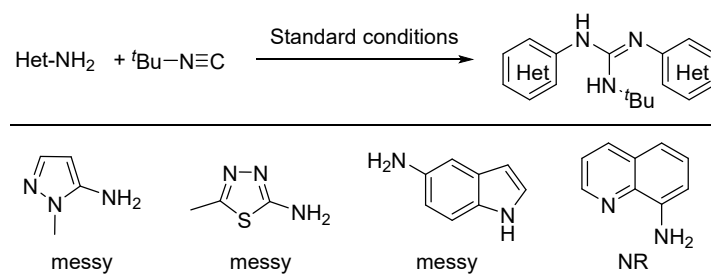


Purified by silica gel column chromatography (petroleum ether/ethyl acetate/triethylamine =10:1:0.1) as white solid (10.9 mg, 38% yield).

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)** δ 4.17 (s, 1H), 4.09 (s, 1H), 3.15 (dd, *J* = 7.2, 5.6 Hz, 2H), 1.33 (s, 9H), 1.11 (t, *J* = 7.2 Hz, 3H).

## 2.2 Test on the reactivity of heteroaromatic amines

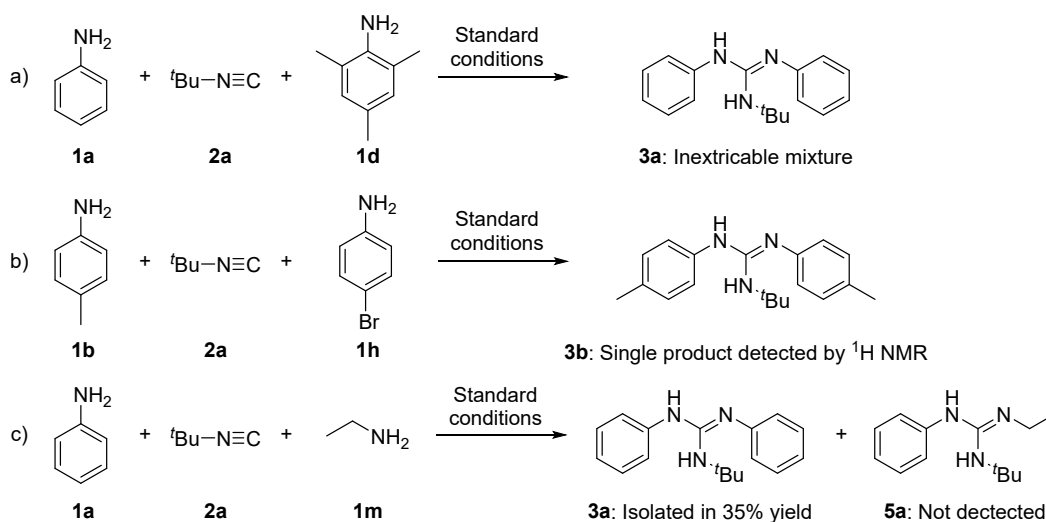
The substrate scope of heterocyclic amines has been evaluated under the standard conditions. As exhibited in Scheme S1, five-membered heterocyclic amines based on pyrazole, thiadiazole and indole frameworks resulted in messy mixtures. This is probably ascribed to the electron-rich property for decomposition under the reaction conditions. In contrast, quinolin-8-amine was totally inert in the reaction. We speculated that the coordination effect of the N-atom to the nickel catalyst might inhibit the expected reaction.



Scheme S1. The reactivity survey of heteroaromatic amines

## 2.3 Cross-over reactions of two different amines with isocyanide

The isocyanide insertion reactions with two different aromatic amines were examined. First, in the cross-over reaction of aniline **1a** and 2,4,6-trimethylaniline **1d**, an inextricable mixture was detected (Scheme S2a). After meticulous analysis of its  $^1\text{H}$  NMR spectrum, guanidine **3a** was determined as major product. Next, the cross-over reaction with *p*-toluidine **1b** and 4-bromoaniline **1h** was explored (Scheme S2b). Peculiarly, guanidine **3b** was exclusively detected by  $^1\text{H}$  NMR analysis, which indicated that the electron-rich aromatic amines might be preferable than electron-deficient ones in this isocyanide insertion protocol. In addition, the reaction of aniline **1a** and ethanamine **1m** was also tested under the standard reaction conditions (Scheme S2c). As shown, only guanidine **3a** was isolated in 35% yield and the unsymmetric guanidine **5a** could not be detected.



Scheme S2. Cross-over reactions of two different amines with isocyanide



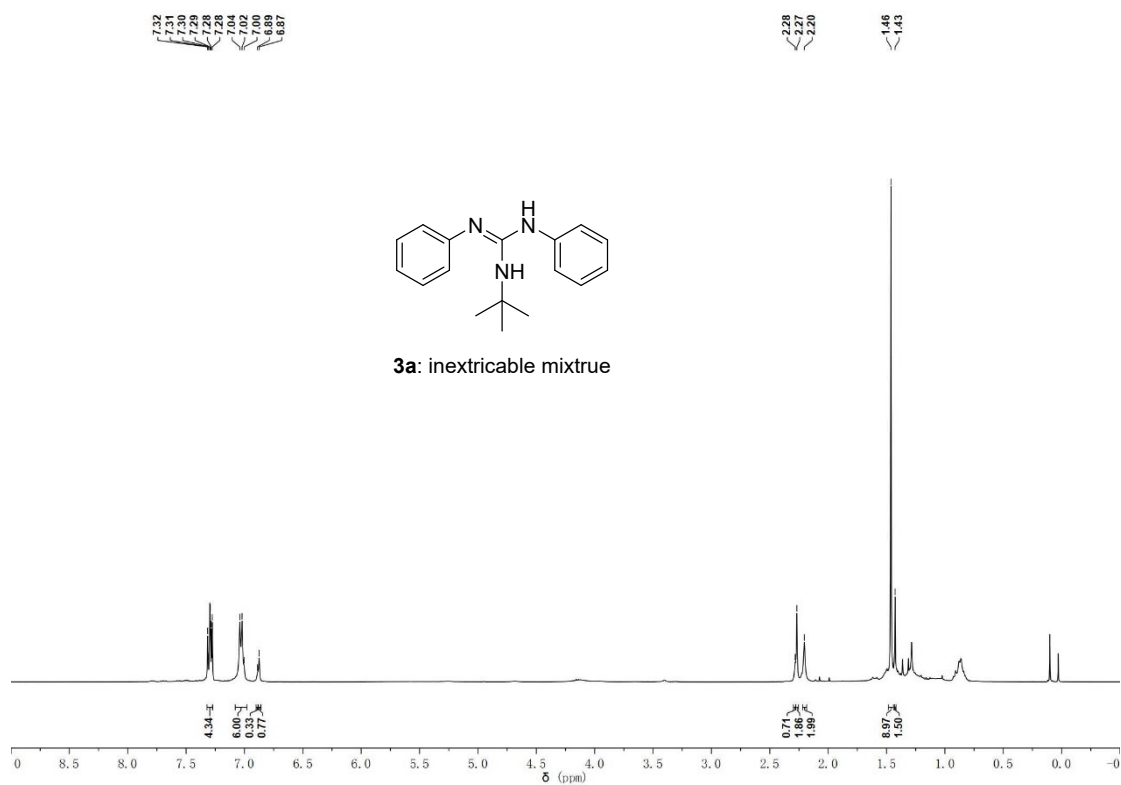


Figure S1. <sup>1</sup>H NMR spectrum of inextricable mixture in Scheme S2a

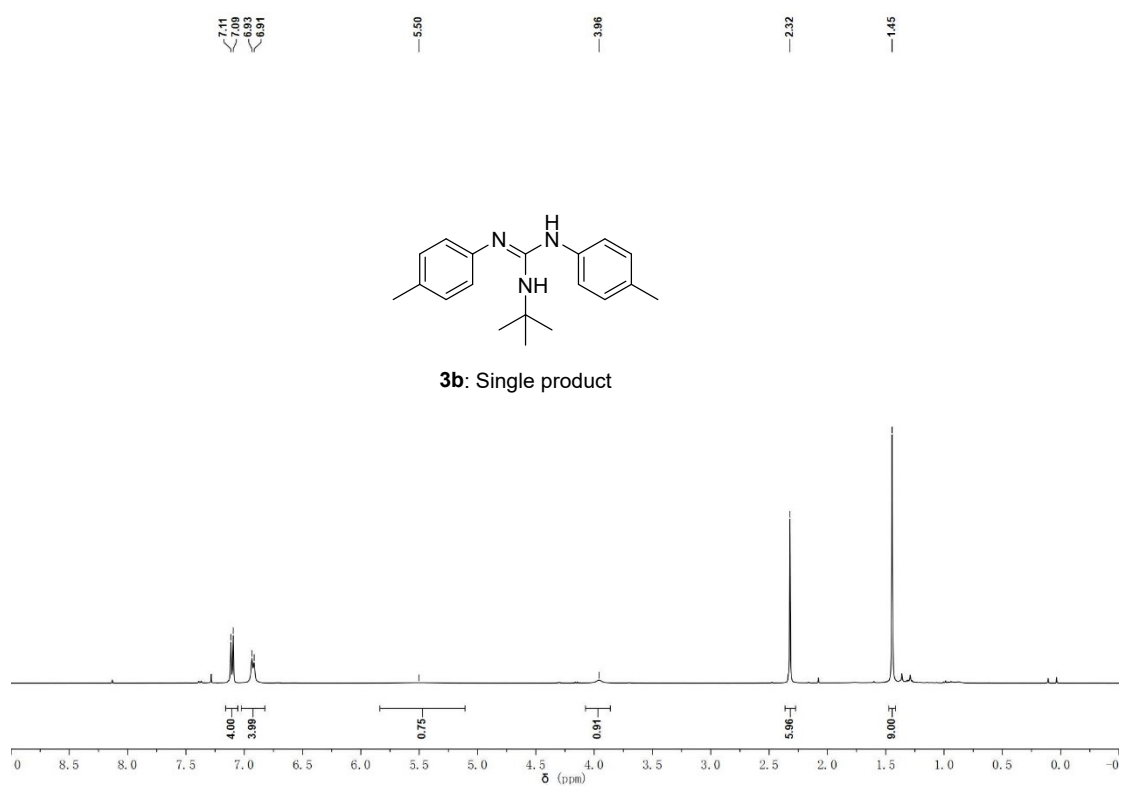


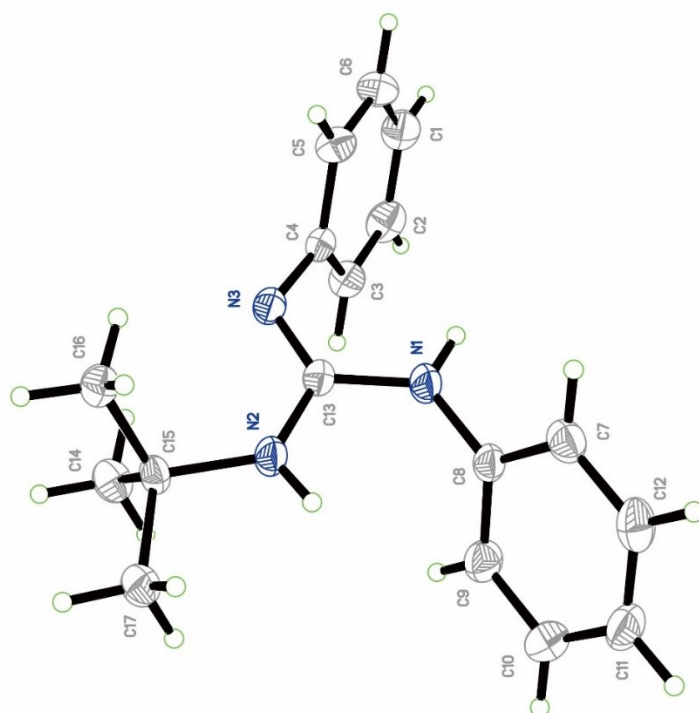
Figure S2. <sup>1</sup>H NMR spectrum of product **3b** in Scheme S2b

### 3. References

1. T.-H. Zhu, S.-Y. Wang, T.-Q. Wei and S.-J. Ji, *Adv. Synth. Catal.*, 2015, **357**, 823.
2. H. Yu, M. Zhang, W. Sun, Y. Li and R. Gao, *Lett. Org. Chem.*, 2010, **7**, 566.
3. B. Liu, M. Yin, H. Gao, W. Wu and H. Jiang, *J. Org. Chem.*, 2013, **78**, 3009.
4. J. Pawlas, J.-H. Choi, C. von Bargaen, S. Maibom-Thomsen, J. H. Rasmussen and O. Ludemann-Hombourger, *Org. Process Res. Dev.* 2023, **27**, 1348.

#### 4. X-Ray Crystallographic Data of **3a**

The crystal of **3a** was crystallized by slow vapor diffusion of acetate and petroleum ether.



**Figure S1.** Ortep drawing of compound **3a** with 50% ellipsoids

## Datablock: mo\_JS\_10045\_0m\_tw

Bond precision: C-C = 0.0065 Å

Wavelength=0.71073

Cell: a=5.9783(10) b=10.038(2) c=25.696(5)  
alpha=90.518(8) beta=91.336(7) gamma=93.881(7)  
Temperature: 223 K

	Calculated	Reported
Volume	1538.0(5)	1537.9(5)
Space group	P -1	P -1
Hall group	-P 1	-P 1
Moiety formula	C17 H21 N3	C17 H21 N3
Sum formula	C17 H21 N3	C17 H21 N3
Mr	267.37	267.38
Dx, g cm <sup>-3</sup>	1.155	1.155
Z	4	4
Mu (mm <sup>-1</sup> )	0.070	0.070
F000	576.0	576.3
F000'	576.17	
h,k,lmax	7,12,30	7,12,30
Nref	5634	5512
Tmin,Tmax	0.990,0.993	0.024,0.047
Tmin'	0.990	

Correction method= # Reported T Limits: Tmin=0.024 Tmax=0.047  
AbsCorr = PSI-SCAN

Data completeness= 0.978

Theta(max)= 25.350

R(reflections)= 0.1013( 4038)

wR2(reflections)=  
0.2901( 5512)

S = 1.056

Npar= 368

The following ALERTS were generated. Each ALERT has the format

**test-name\_ALERT\_alert-type\_alert-level.**

Click on the hyperlinks for more details of the test.



### Alert level C

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PLAT340\_ALERT\_3\_C Low Bond Precision on C-C Bonds ..... 0.0065 Ang.  
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PLAT420\_ALERT\_2\_C D-H Bond Without Acceptor N2 --H2 . Please Check  
PLAT420\_ALERT\_2\_C D-H Bond Without Acceptor N4 --H4 . Please Check  
PLAT420\_ALERT\_2\_C D-H Bond Without Acceptor N5 --H5 . Please Check  
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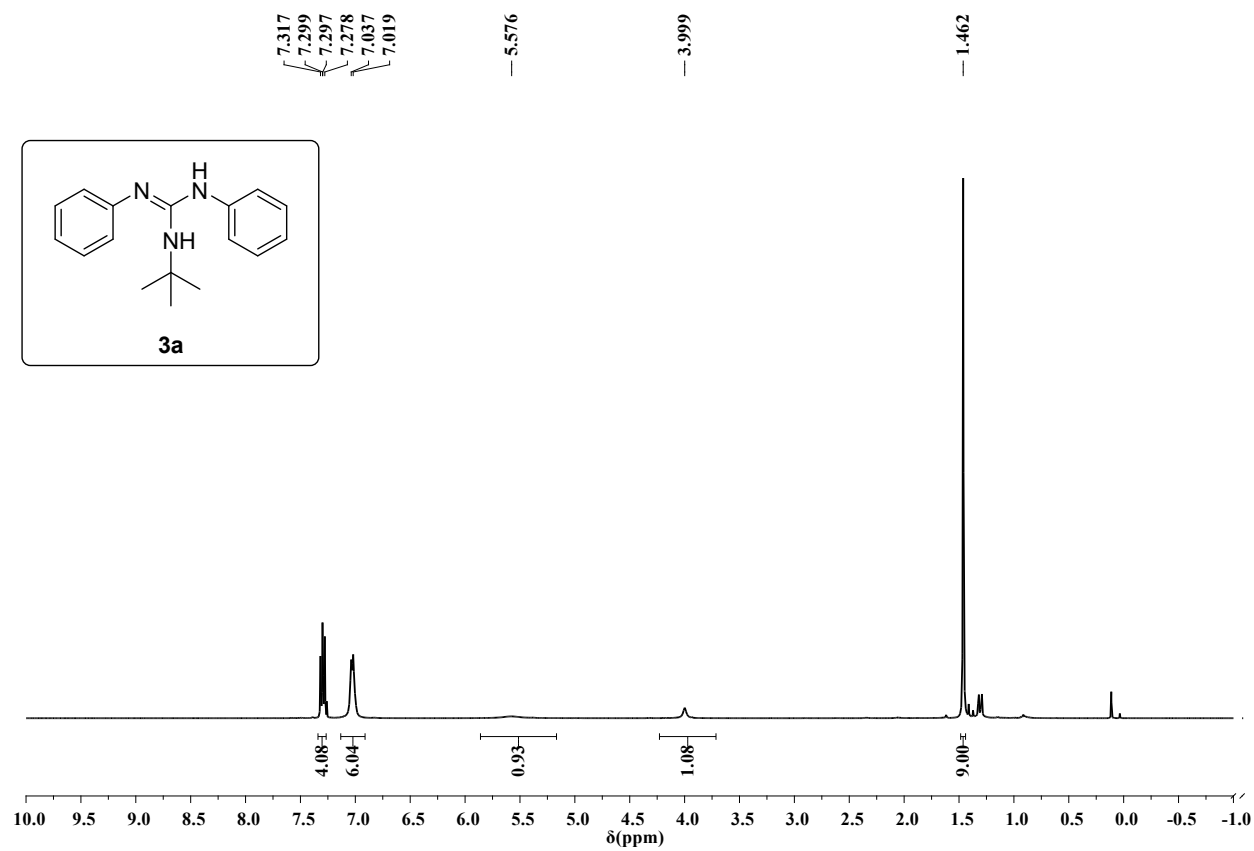
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PLAT073_ALERT_1_G	H-atoms ref., but hydrogen treatment Reported as	constr Check
PLAT769_ALERT_4_G	CIF Embedded Explicitly Supplied Scattering Data	3 Note
PLAT870_ALERT_4_G	ALERTS Related to Twinning Effects Suppressed ..	! Info
PLAT910_ALERT_3_G	Missing FCF Reflection(s) Below Theta(Min) [Deg]= 0 0 1, 0 0 2,	2.03 Note
PLAT930_ALERT_2_G	FCF-based Twin Law ( 0 1 0) Est.d BASF	0.39 Check
PLAT931_ALERT_5_G	CIFcalcFCF Twin Law ( 0 1 0) Est.d BASF	0.39 Check
PLAT933_ALERT_2_G	Number of HKL-OMIT Records in Embedded .res File -5 -2 13, -5 0 12, -4 -7 5, -4 -4 2, -4 -3 1, -4 -2 1, -4 -2 3, -4 5 0, -1 -5 19, -1 -3 23, -1 1 18, -1 4 21, -1 5 23, 0 -9 3, 0 -7 2, 0 -7 10, 0 -6 1, 0 -6 4, 0 -5 1, 0 -5 2, 0 -5 3, 0 -5 5, 0 -5 8, 0 -4 2, 0 -4 5, 0 -4 6, 0 -3 3, 0 -3 4, 0 -3 5, 0 -3 6, 0 -3 17, 0 -2 1, 0 -2 3, 0 -2 4, 0 -2 5, 0 -2 6, 0 -2 10, 0 -2 11, 0 -1 1, 0 -1 2, 0 0 6, 0 1 6, 0 2 1, 0 2 2, 0 2 3, 0 2 4, 0 2 5, 0 3 2, 0 3 7, 0 4 0,	104 Note
PLAT941_ALERT_3_G	Average HKL Measurement Multiplicity .....	1.0 Low
PLAT948_ALERT_5_G	Externally Supplied Scattering Factors CIF	3 Note
PLAT969_ALERT_5_G	The 'Henn et al.' R-Factor-gap value ..... Predicted wR2: Based on SigI**2 8.52 or SHELX Weight 27.47	3.404 Note

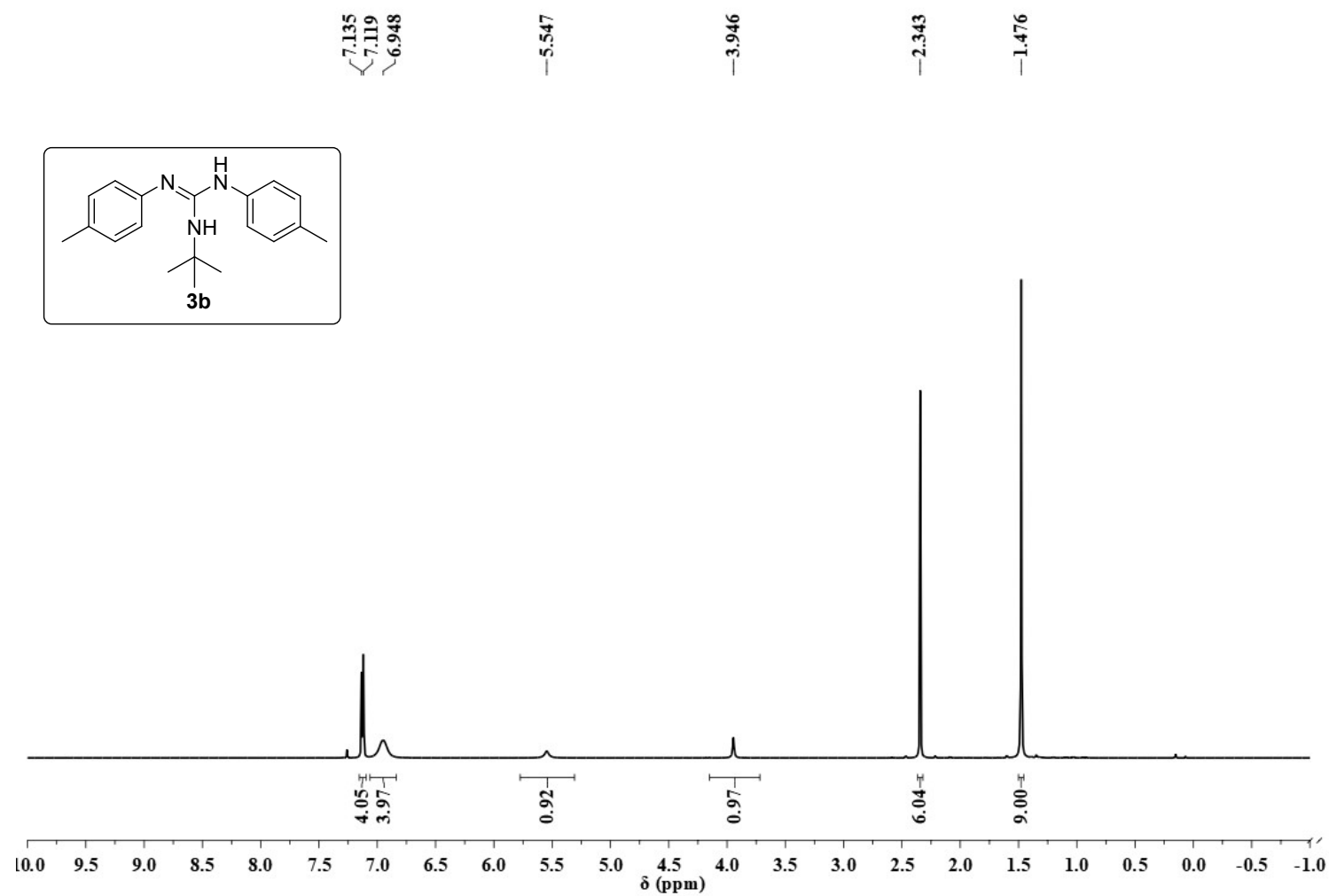
- 
- 0 **ALERT level A** = Most likely a serious problem - resolve or explain  
0 **ALERT level B** = A potentially serious problem, consider carefully  
9 **ALERT level C** = Check. Ensure it is not caused by an omission or oversight  
12 **ALERT level G** = General information/check it is not something unexpected
- 2 ALERT type 1 CIF construction/syntax error, inconsistent or missing data  
7 ALERT type 2 Indicator that the structure model may be wrong or deficient  
7 ALERT type 3 Indicator that the structure quality may be low  
2 ALERT type 4 Improvement, methodology, query or suggestion  
3 ALERT type 5 Informative message, check
-

## 5. $^1\text{H}$ and $^{13}\text{C}$ NMR Spectra

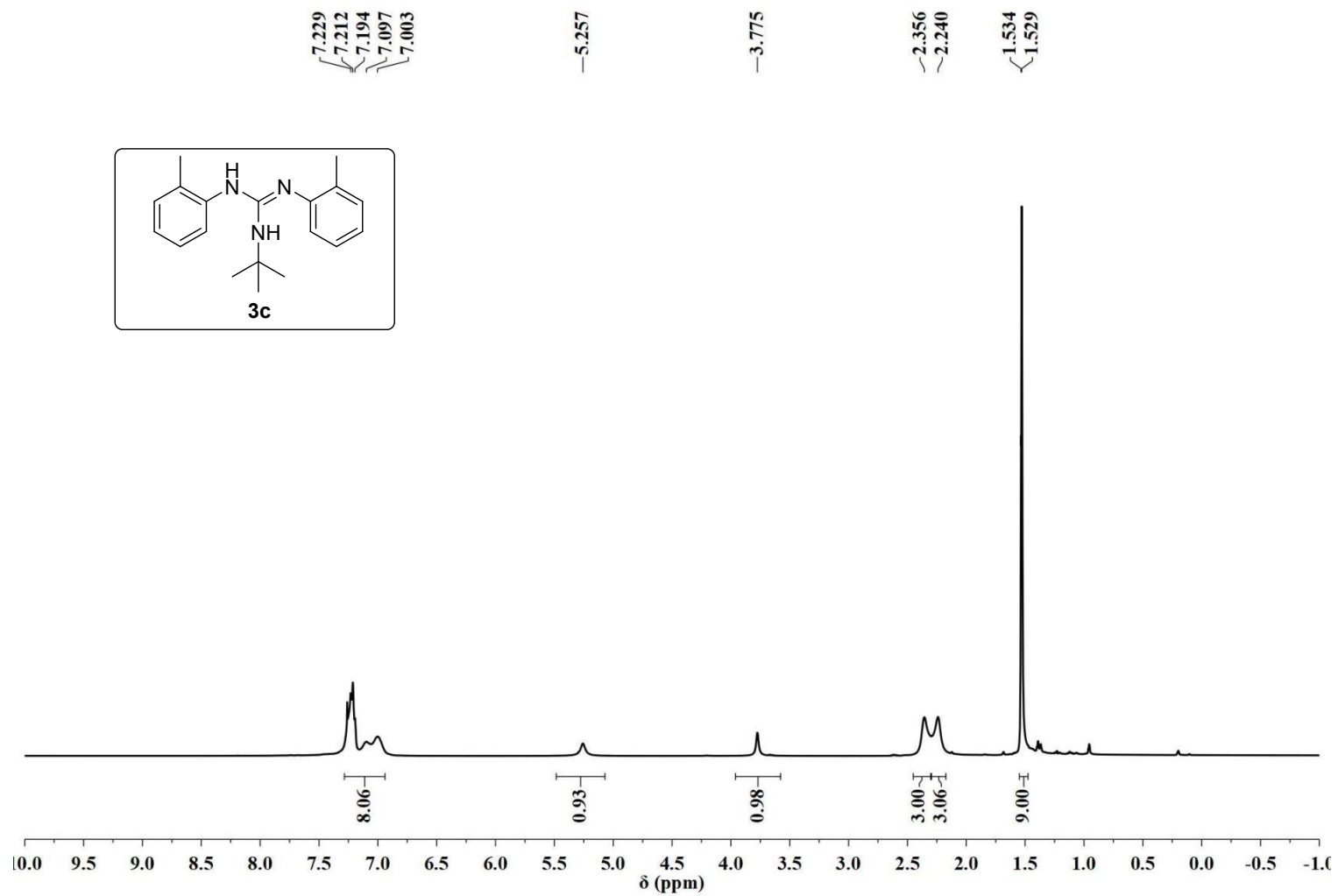
$^1\text{H}$  NMR of **3a** (400 MHz,  $\text{CDCl}_3$ )



**<sup>1</sup>H NMR of 3b (400 MHz, CDCl<sub>3</sub>)**

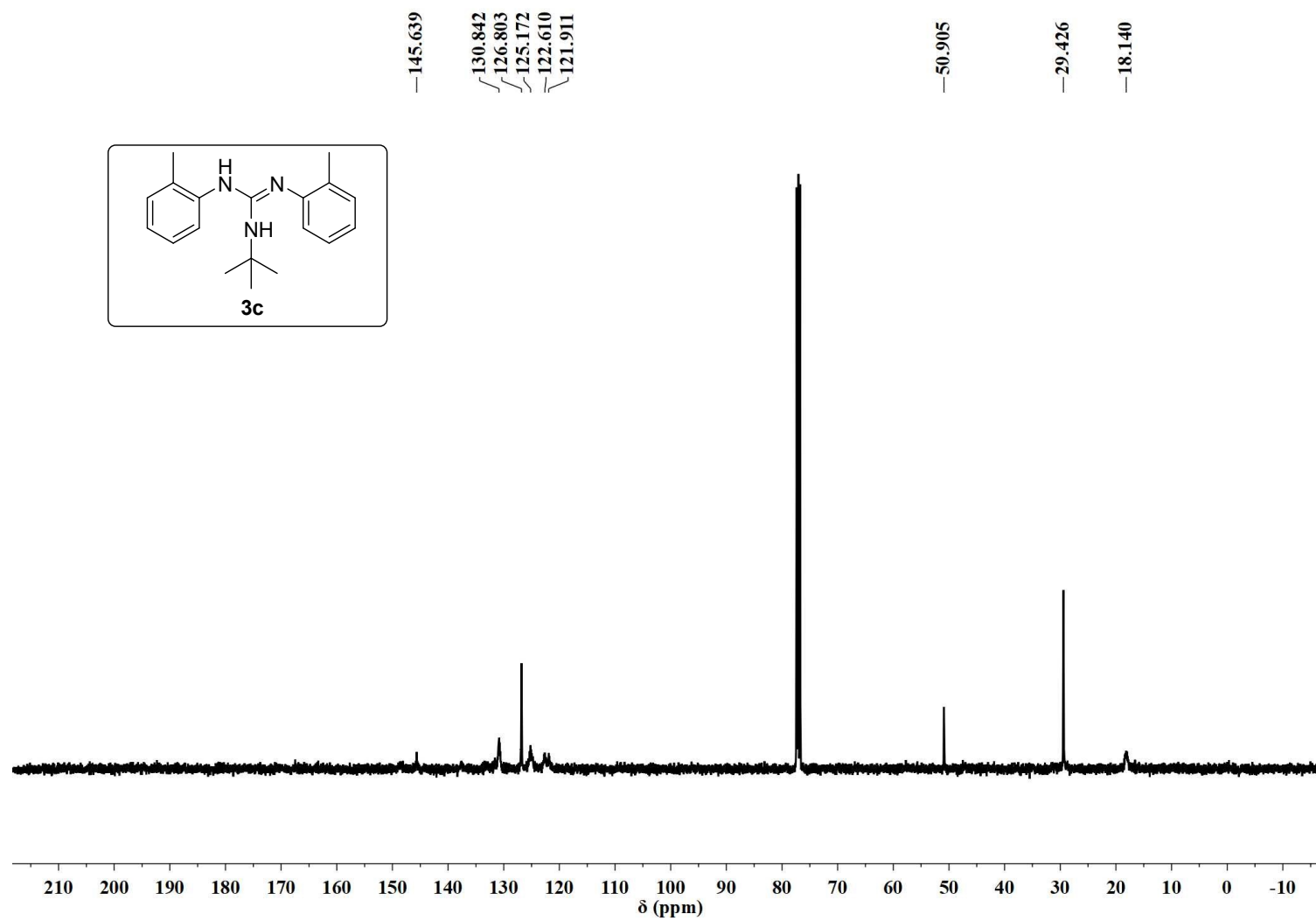


**<sup>1</sup>H NMR of 3c (400 MHz, CDCl<sub>3</sub>)**

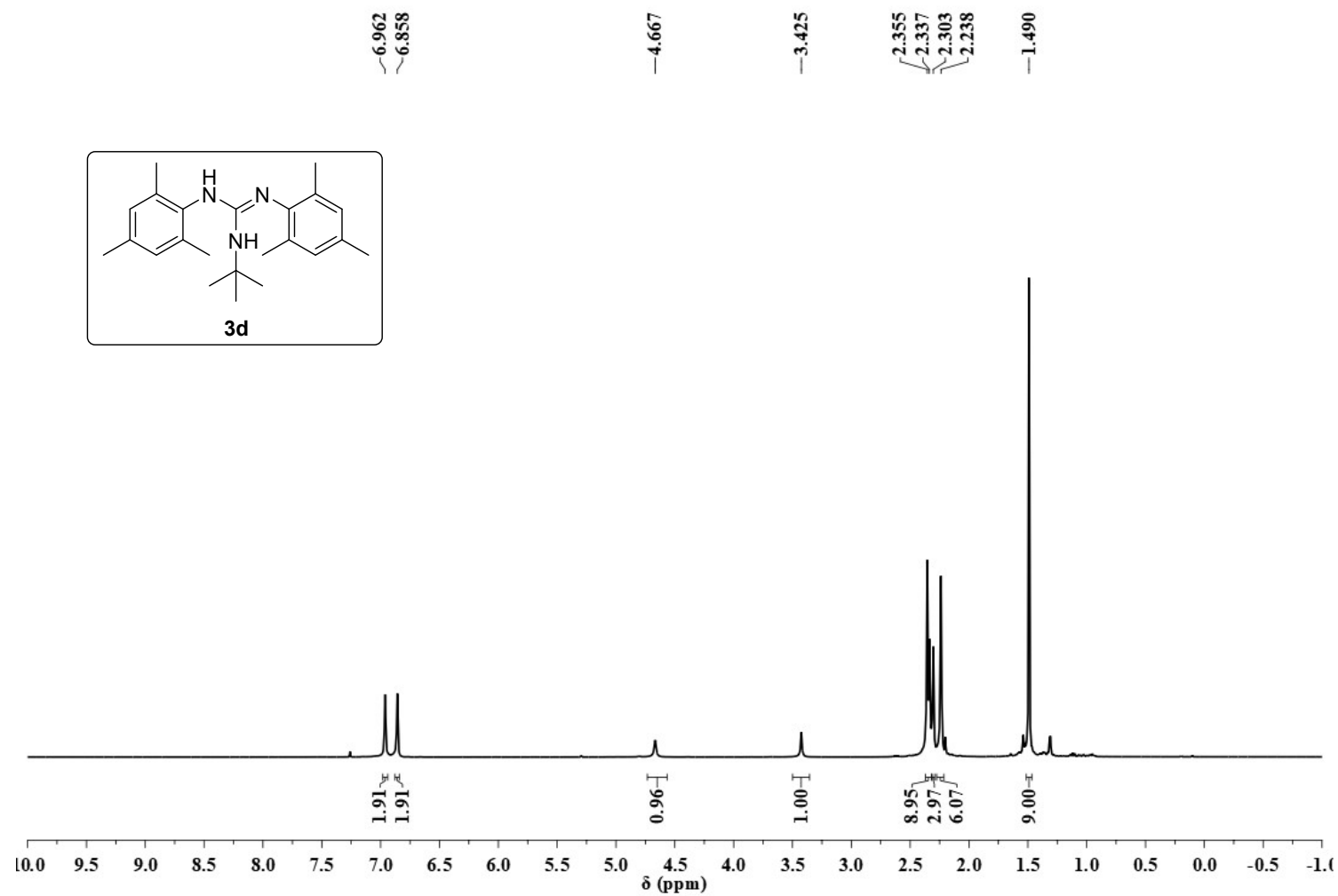




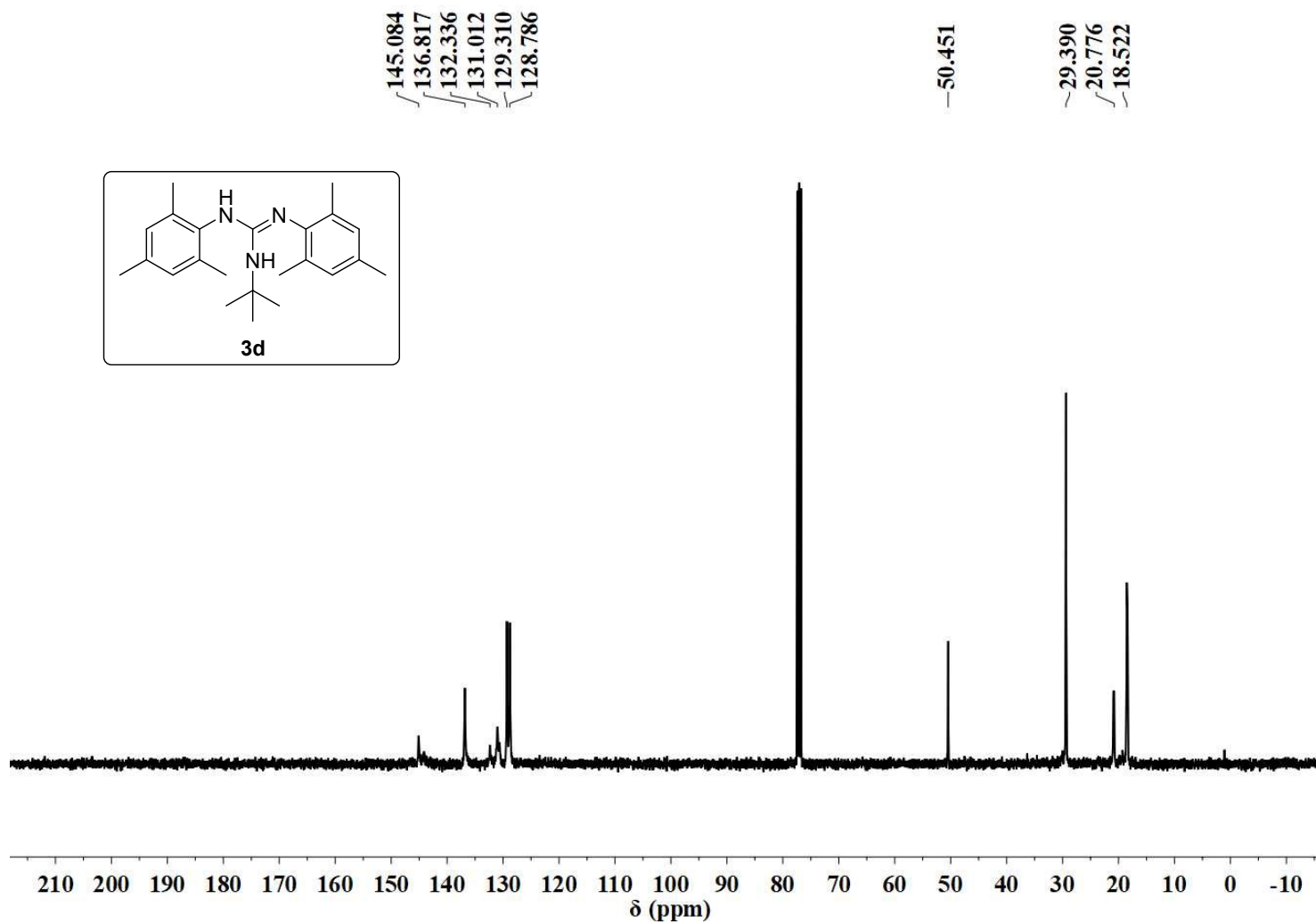
**$^{13}\text{C}$  NMR of 3c (101 MHz,  $\text{CDCl}_3$ )**



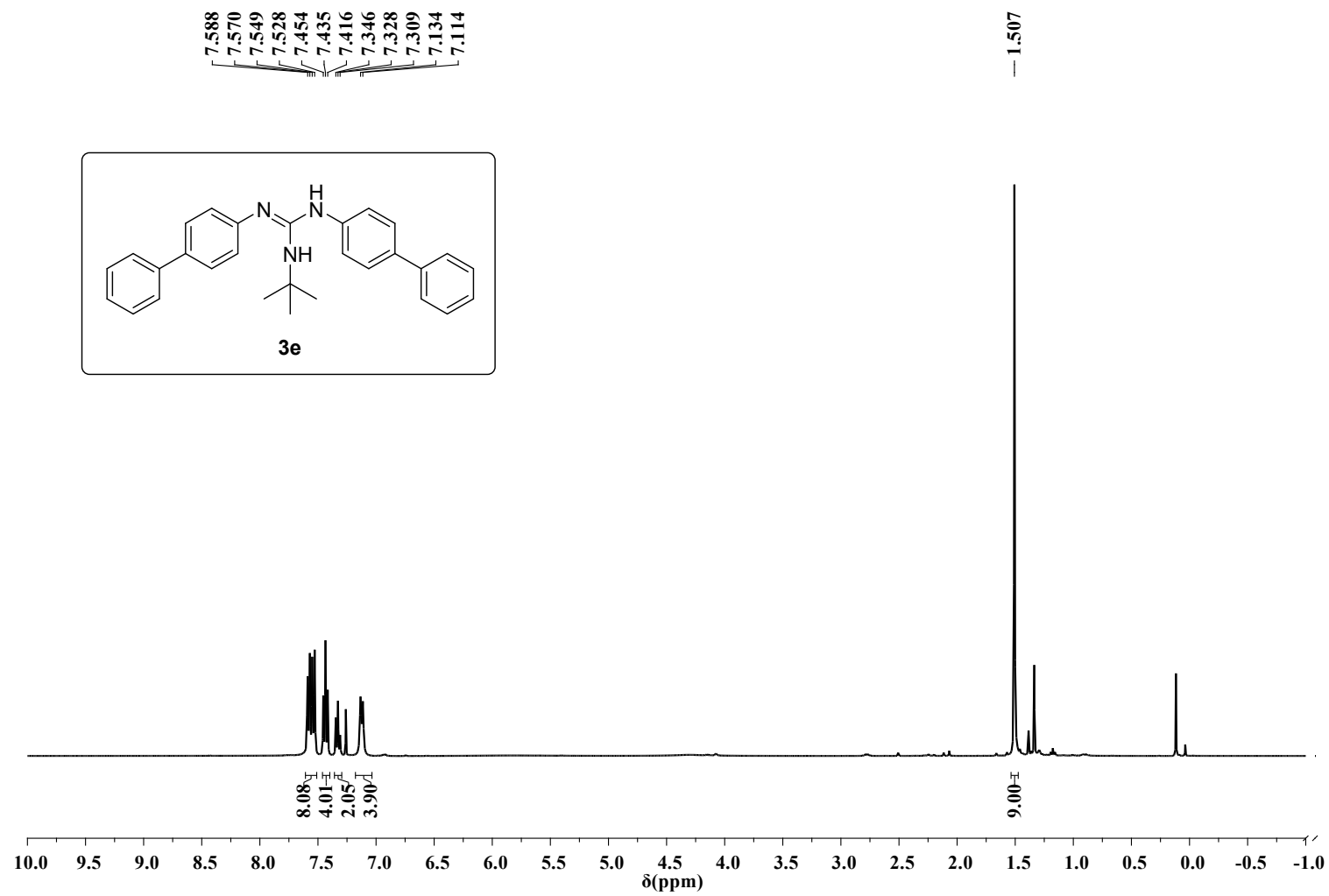
**<sup>1</sup>H NMR of 3d (400 MHz, CDCl<sub>3</sub>)**



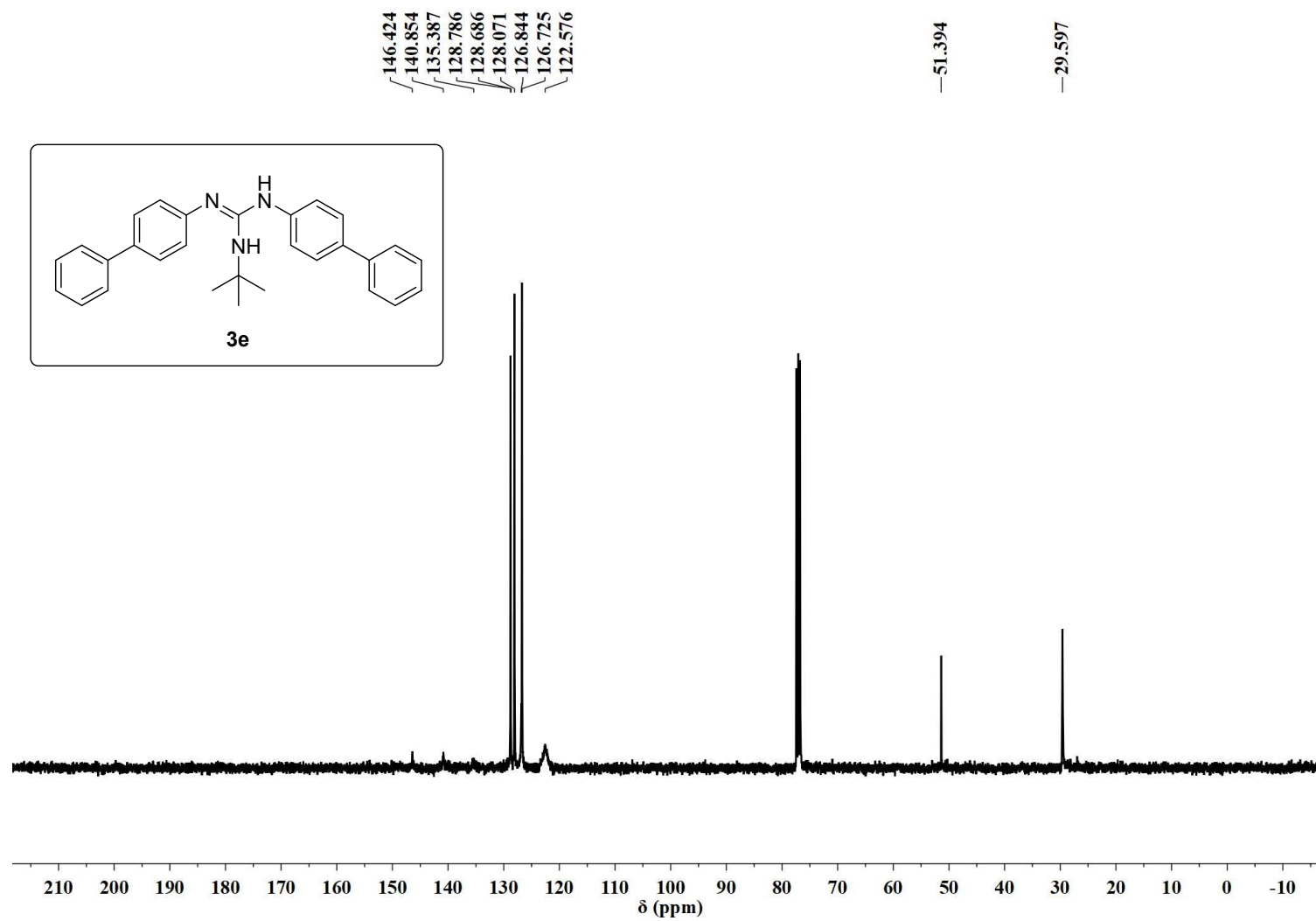
<sup>13</sup>C NMR of 3d (101 MHz, CDCl<sub>3</sub>)



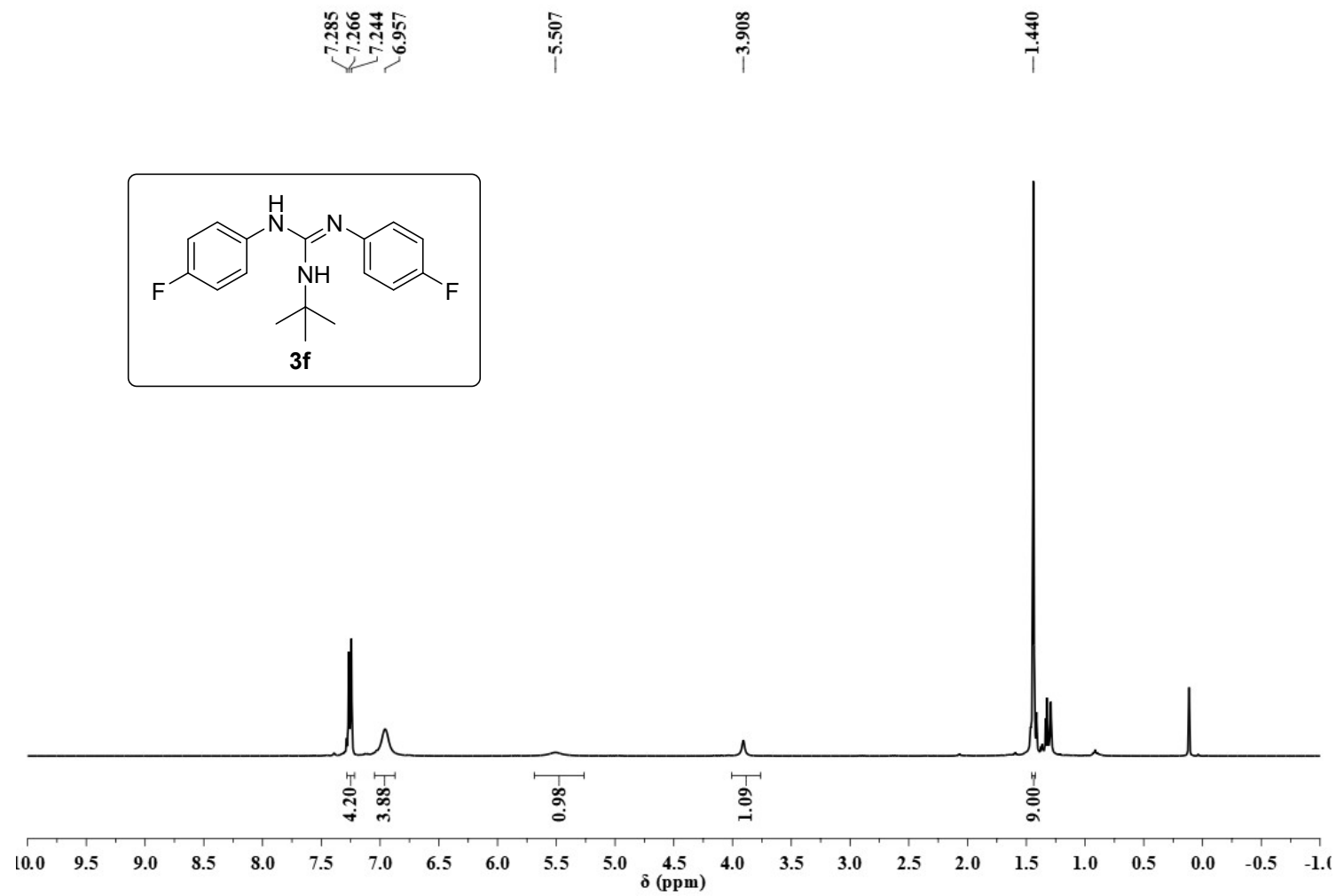
**<sup>1</sup>H NMR of 3e (400 MHz, CDCl<sub>3</sub>)**



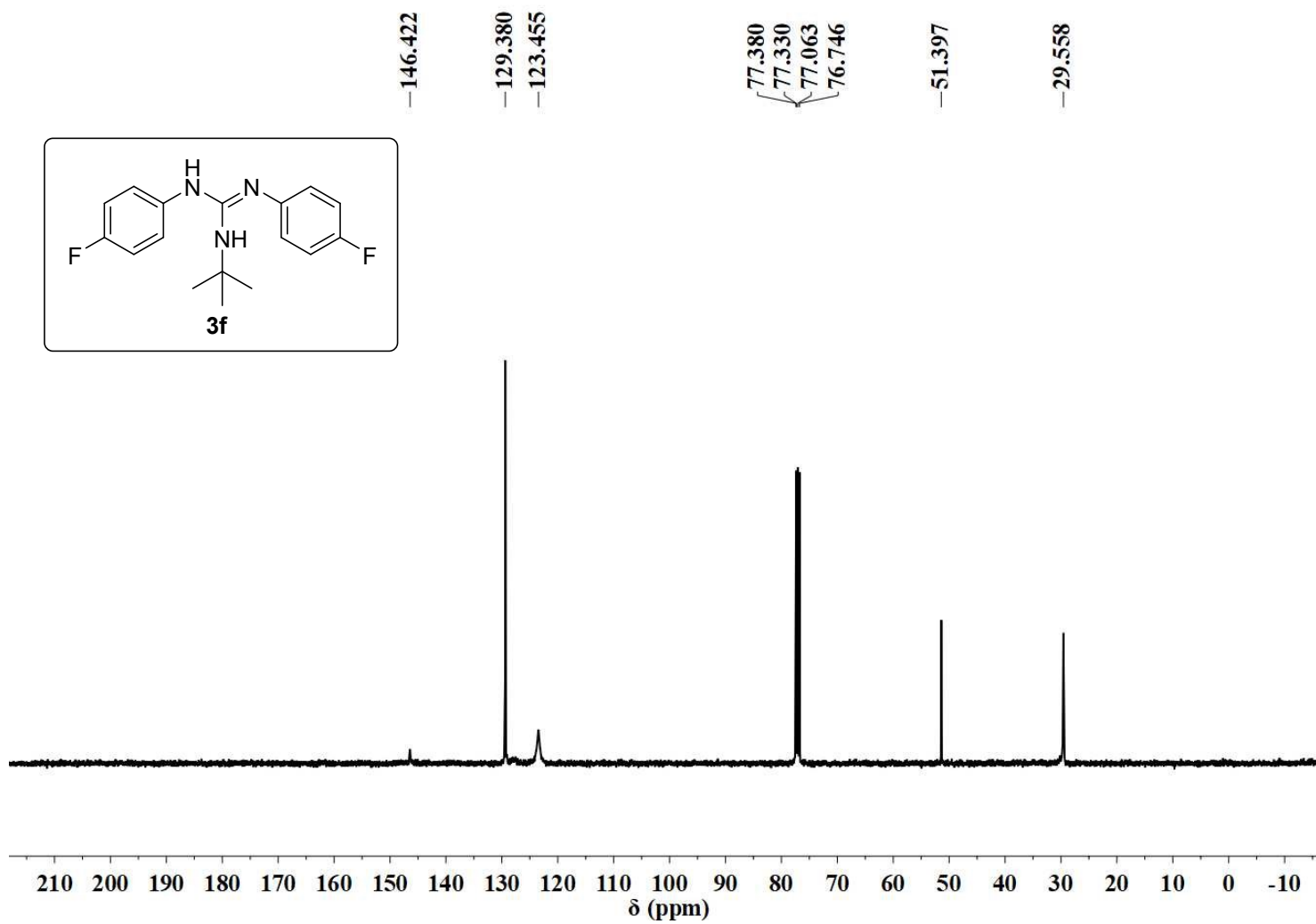
**$^{13}\text{C}$  NMR of 3e (101 MHz,  $\text{CDCl}_3$ )**



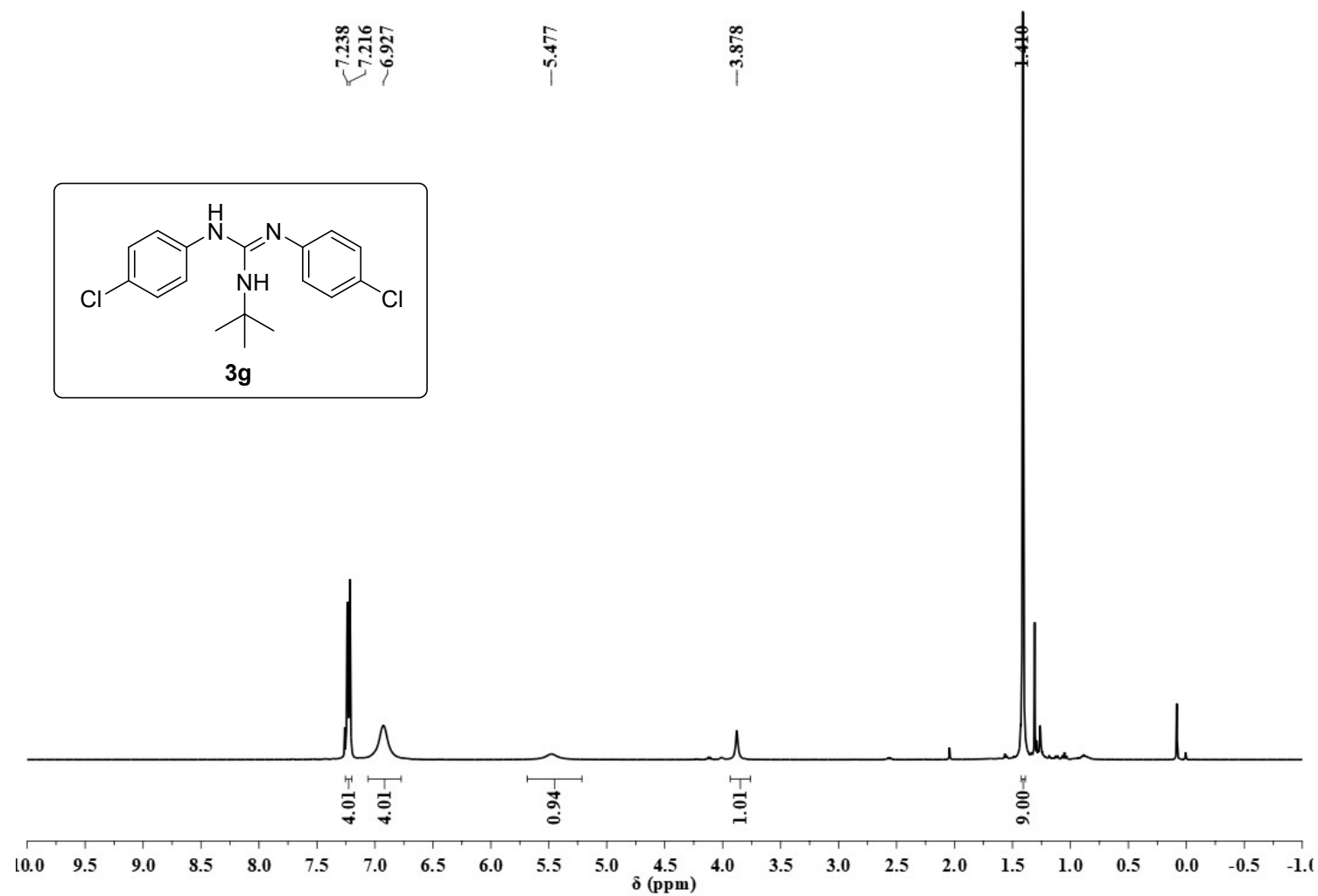
**<sup>1</sup>H NMR of 3f (400 MHz, CDCl<sub>3</sub>)**



$^{13}\text{C}$  NMR of **3f** (101 MHz,  $\text{CDCl}_3$ )

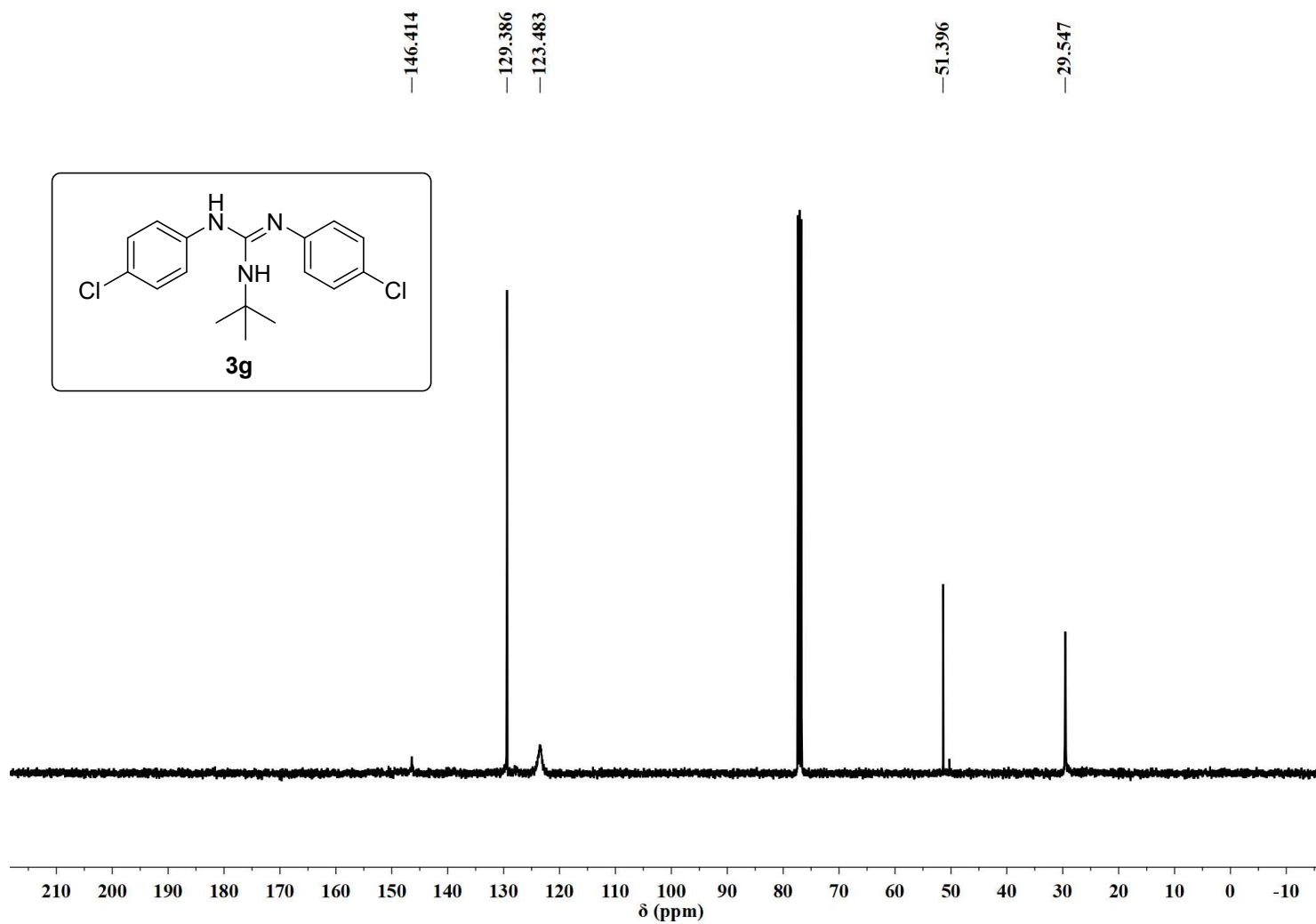


**<sup>1</sup>H NMR of 3g (400 MHz, CDCl<sub>3</sub>)**

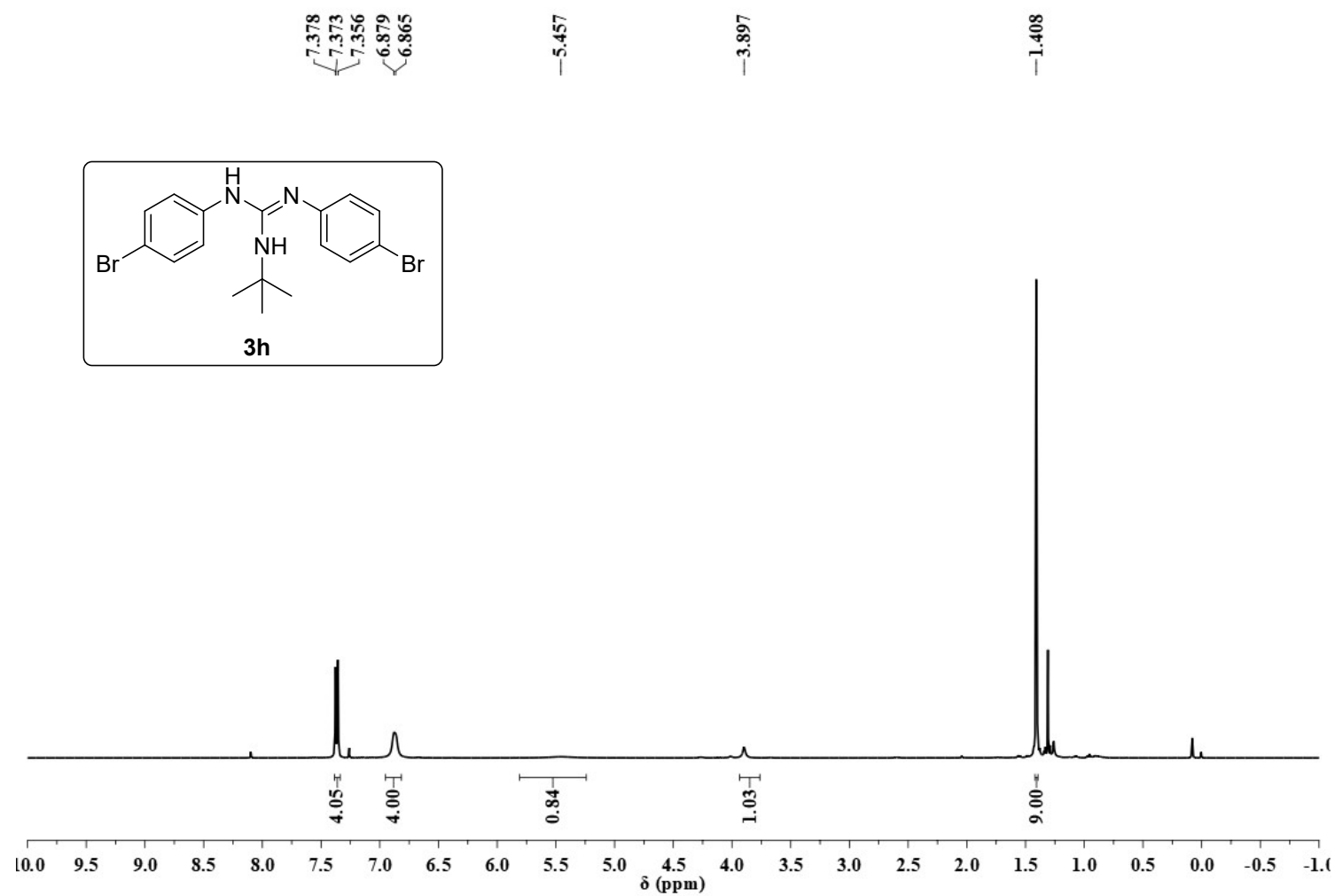




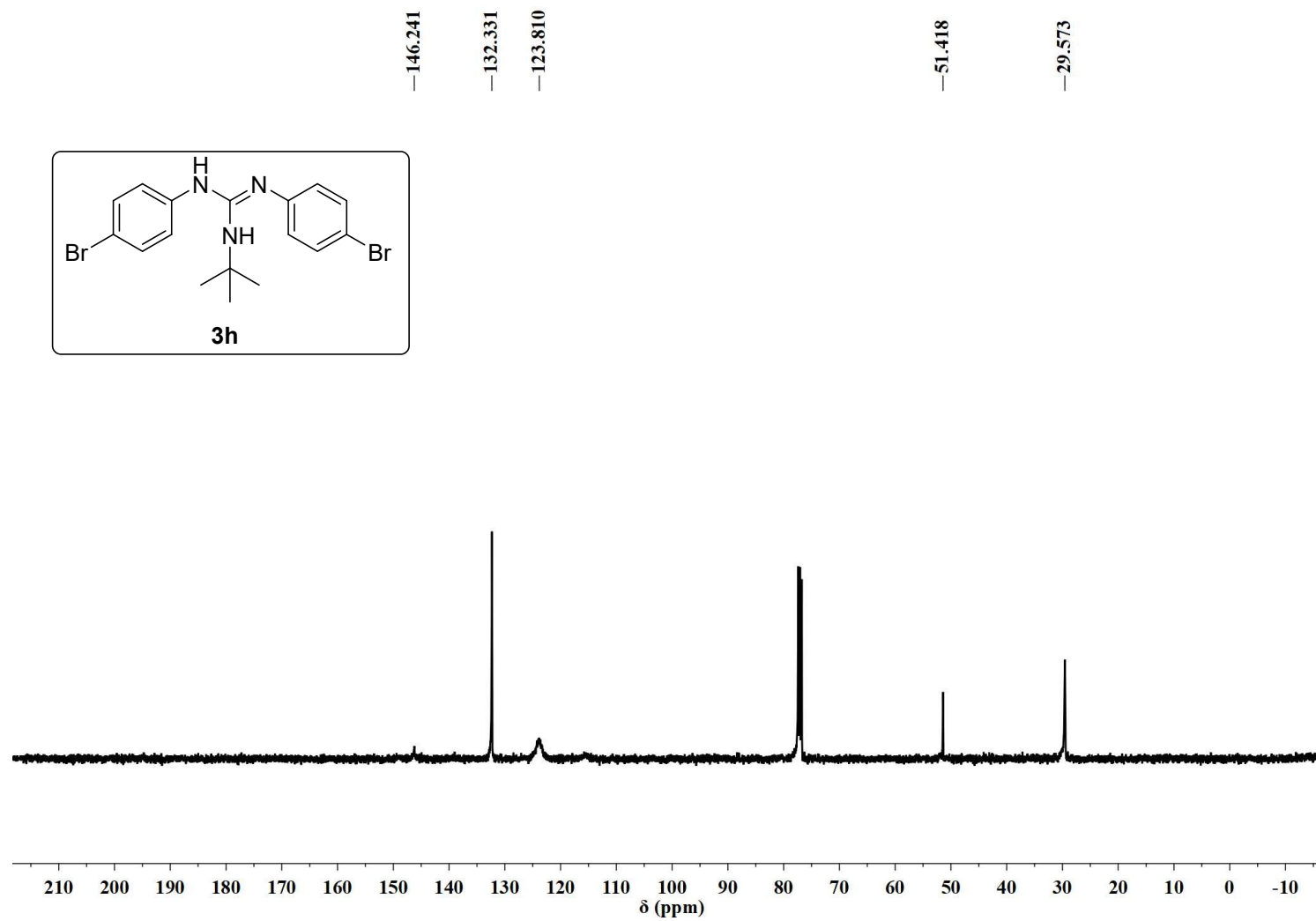
**$^{13}\text{C}$  NMR of 3g (101 MHz,  $\text{CDCl}_3$ )**



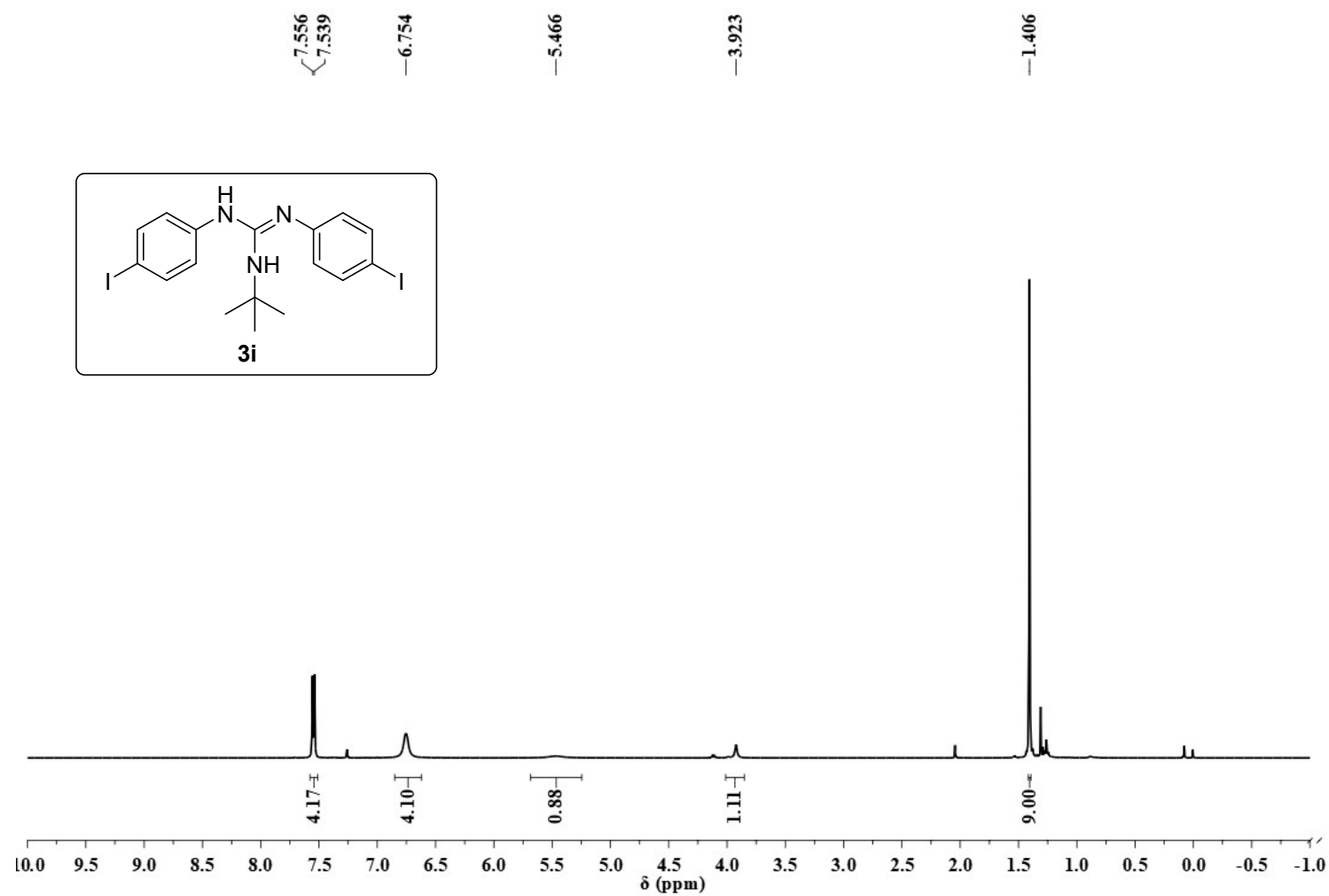
**<sup>1</sup>H NMR of 3h (400 MHz, CDCl<sub>3</sub>)**



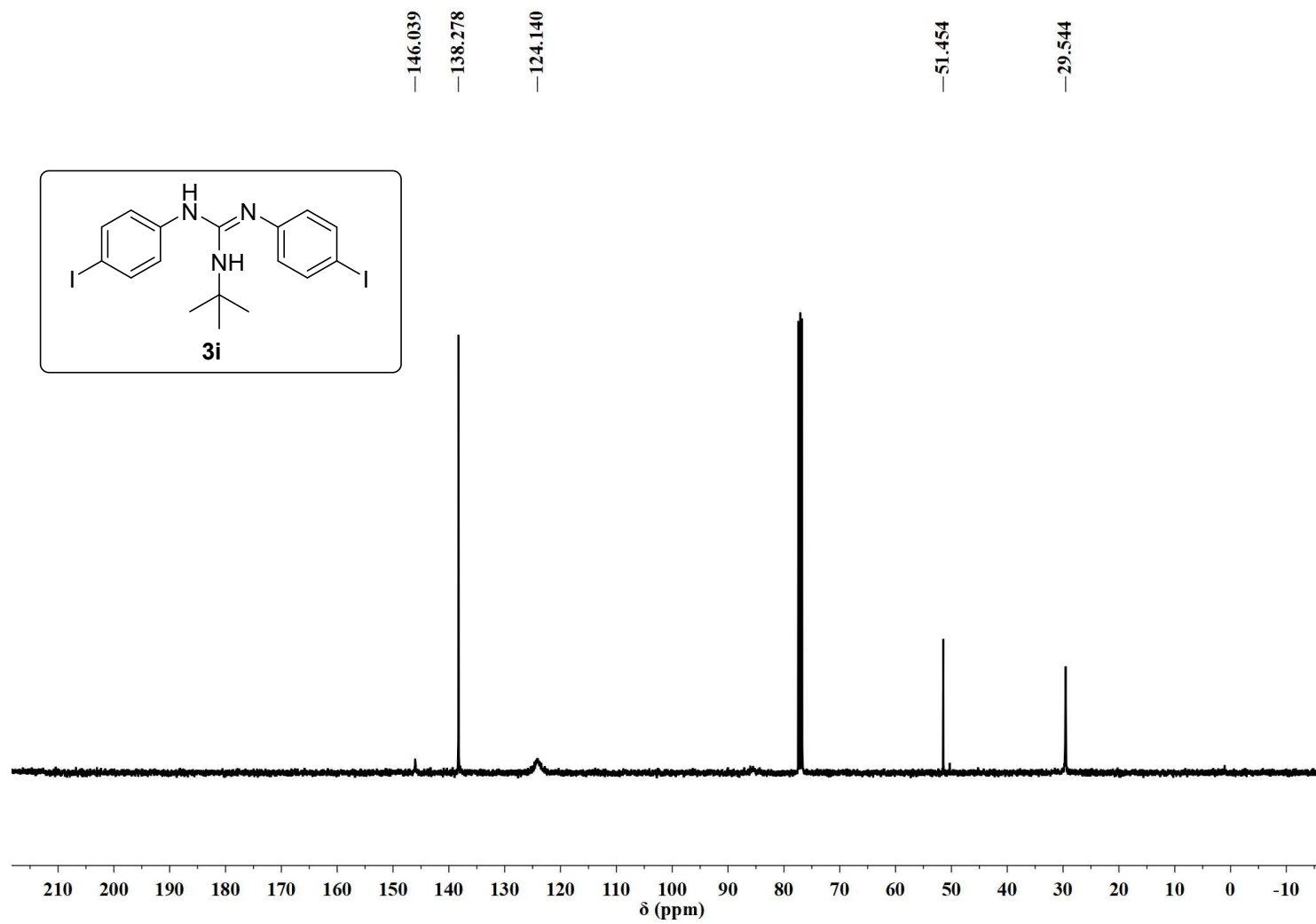
**$^{13}\text{C}$  NMR of 3h (101 MHz,  $\text{CDCl}_3$ )**



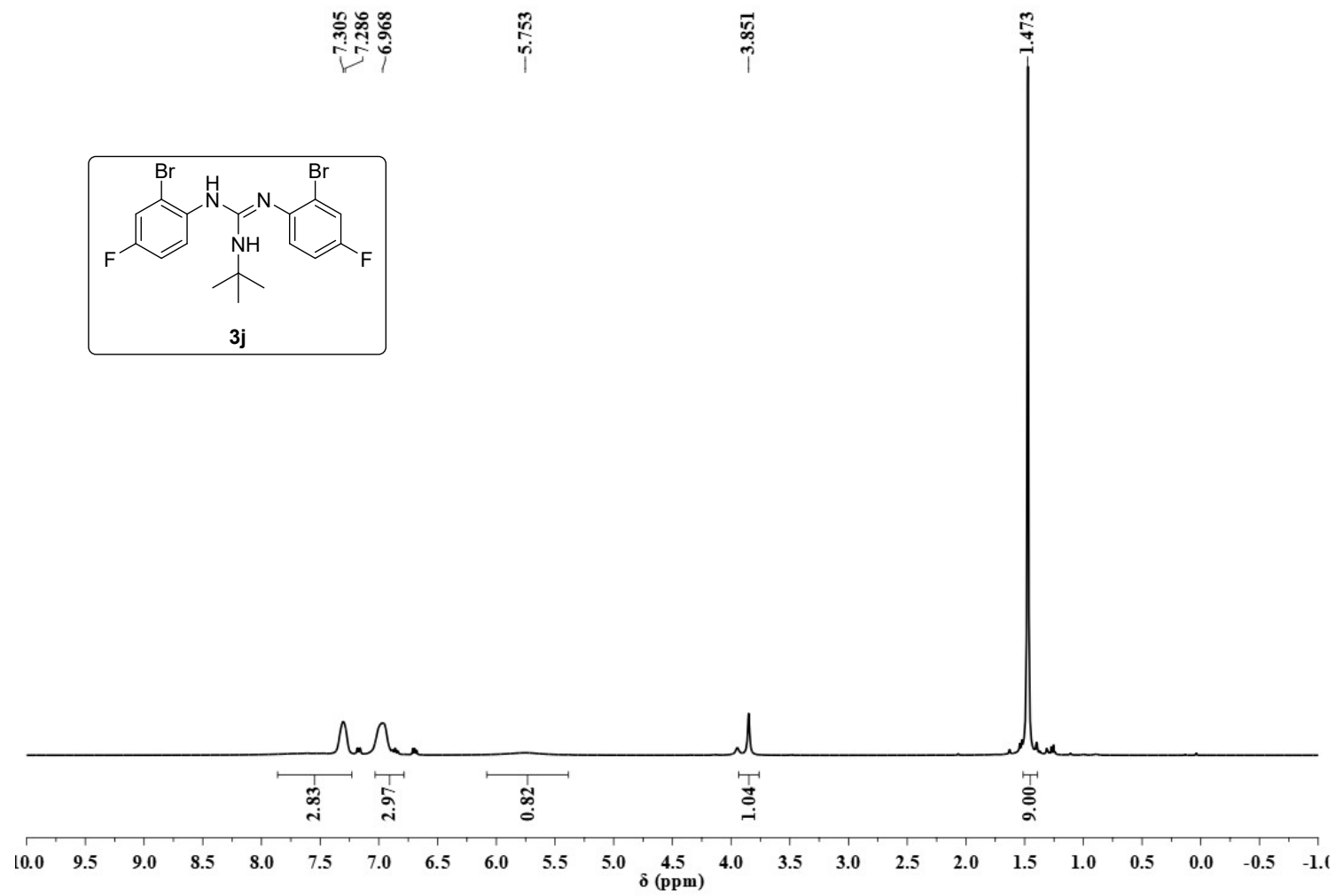
**<sup>1</sup>H NMR of 3i (400 MHz, CDCl<sub>3</sub>)**



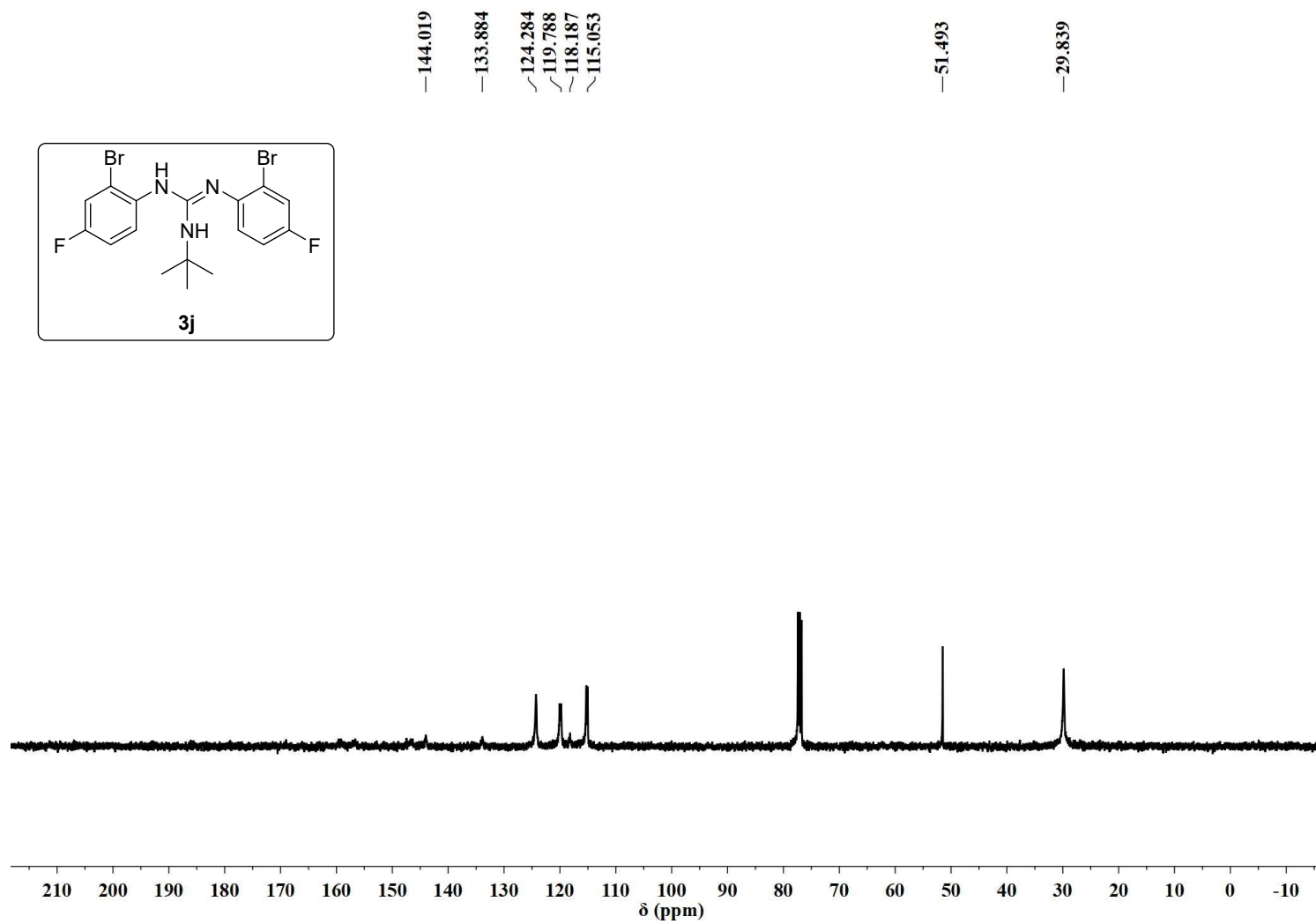
**$^{13}\text{C}$  NMR of 3i (400 MHz,  $\text{CDCl}_3$ )**



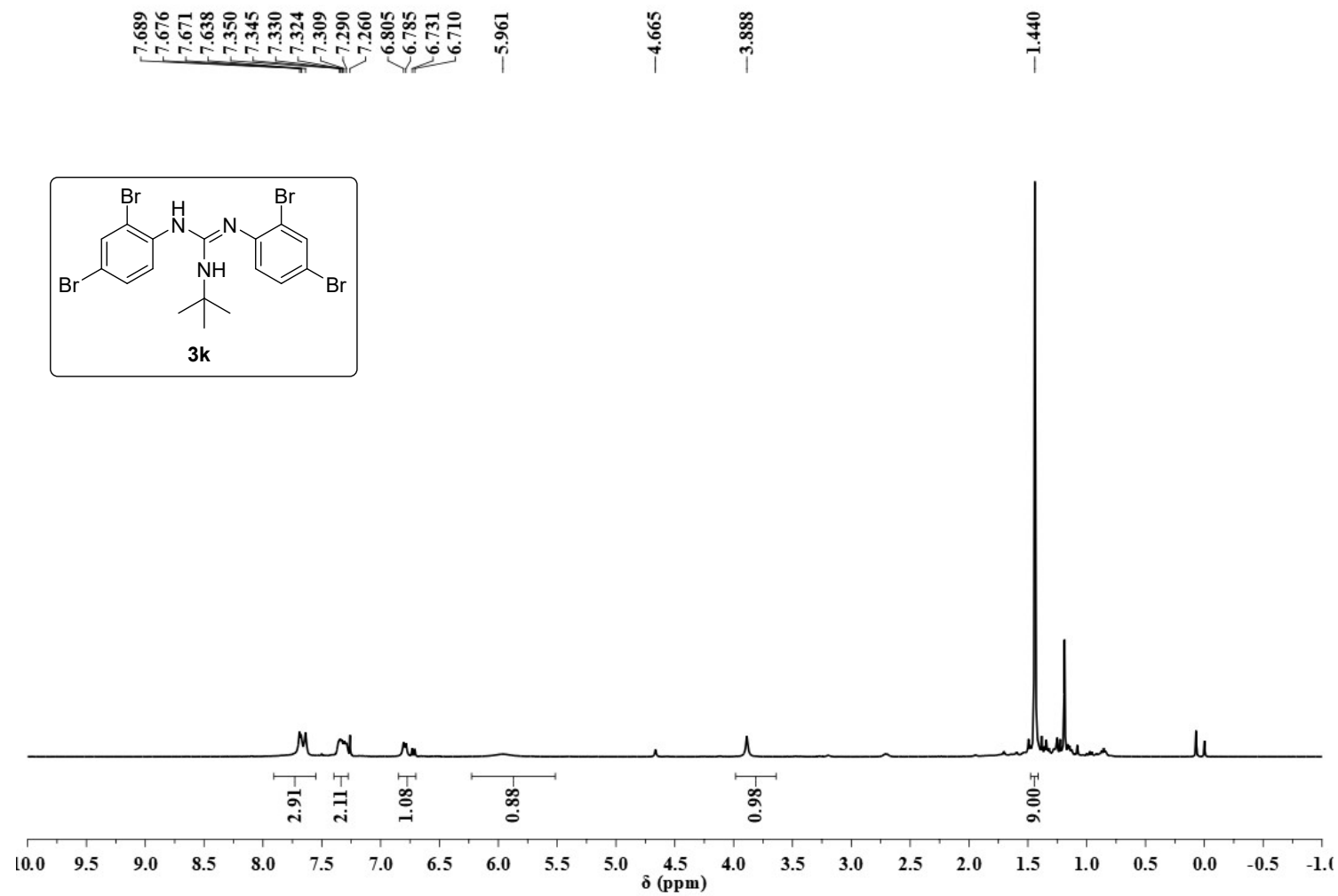
**<sup>1</sup>H NMR of 3j (400 MHz, CDCl<sub>3</sub>)**



**<sup>13</sup>C NMR of 3j (101 MHz, CDCl<sub>3</sub>)**

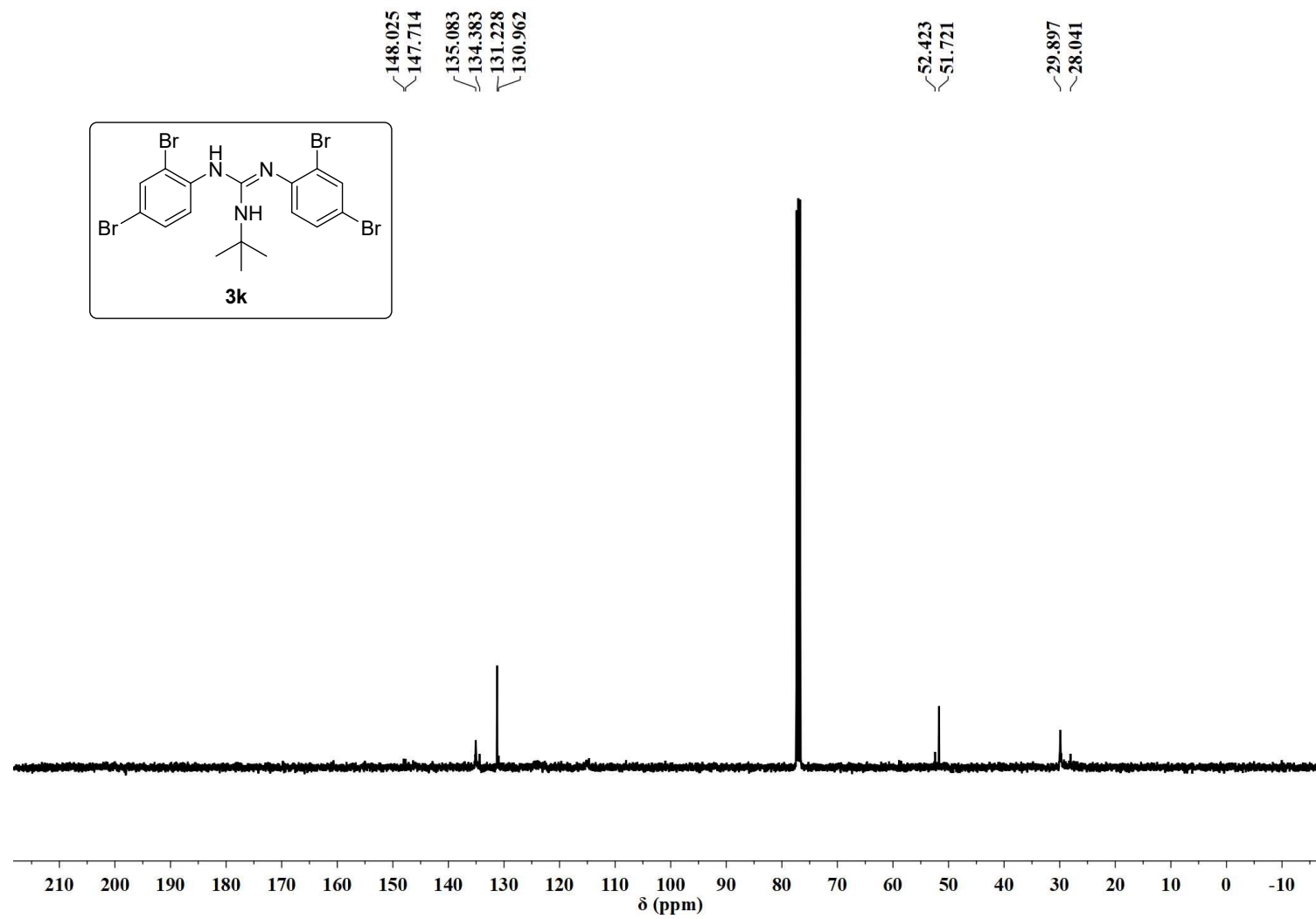


**<sup>1</sup>H NMR of 3k (400 MHz, CDCl<sub>3</sub>)**

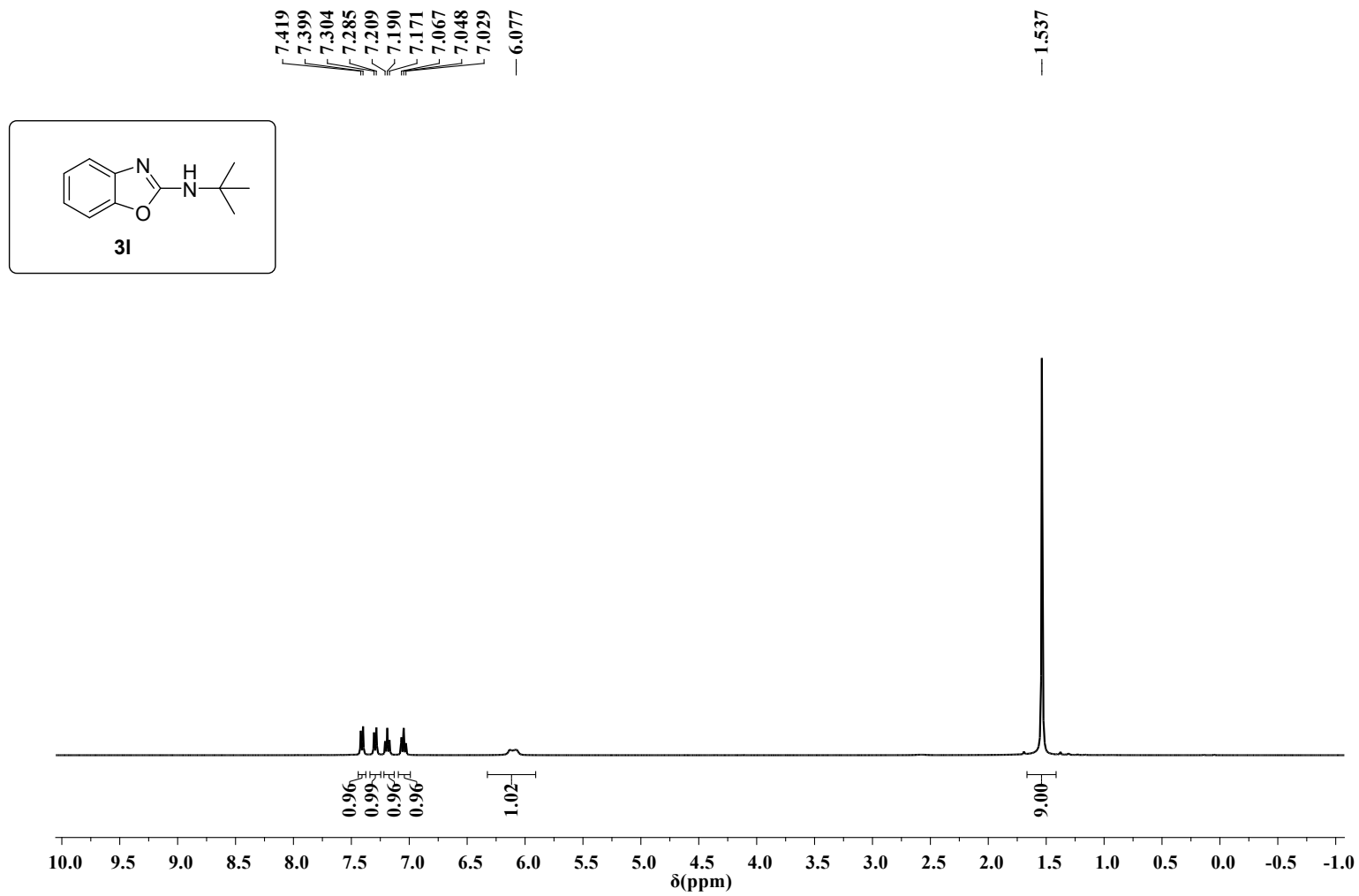




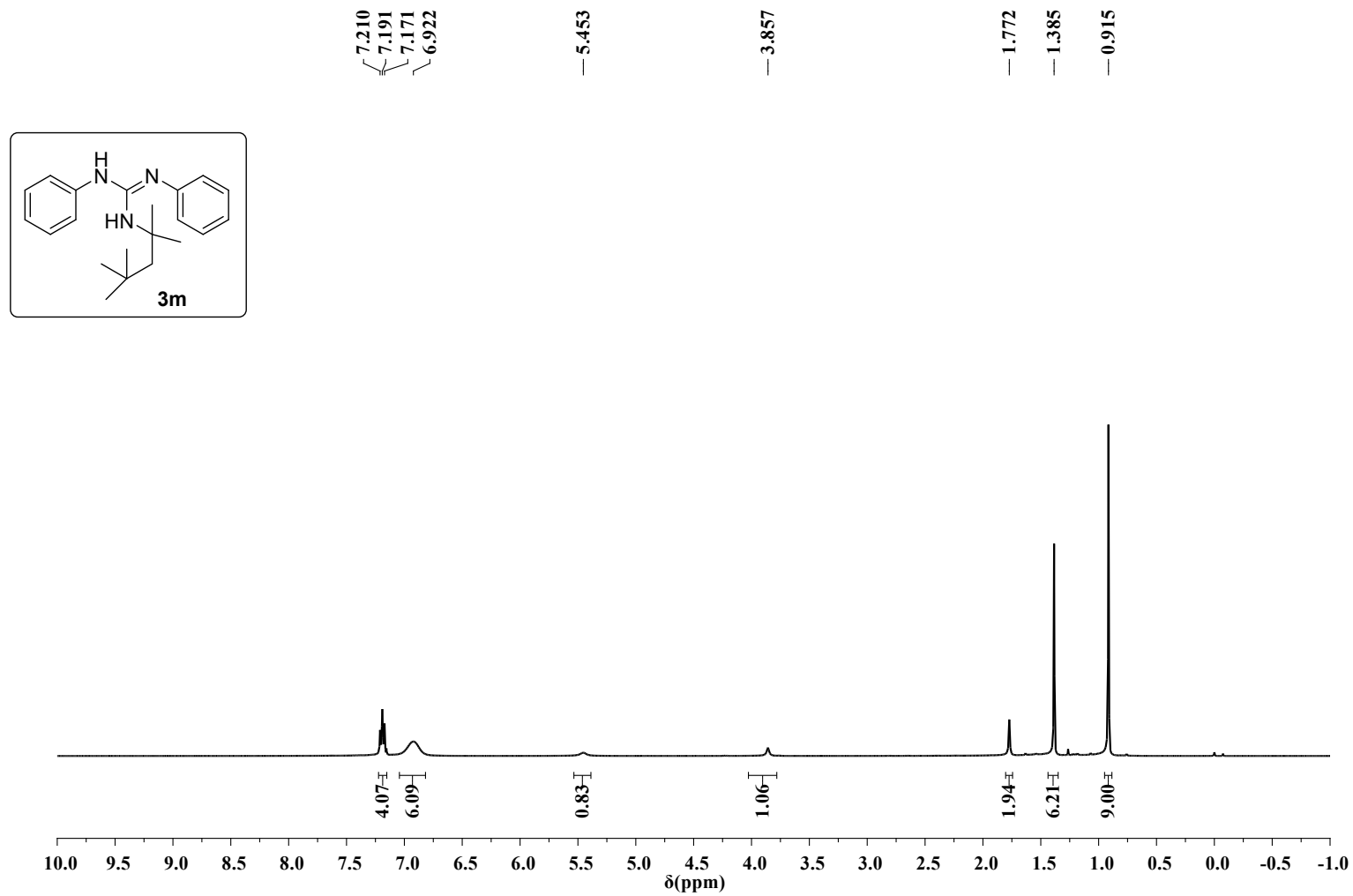
**$^{13}\text{C}$  NMR of 3k (101 MHz,  $\text{CDCl}_3$ )**



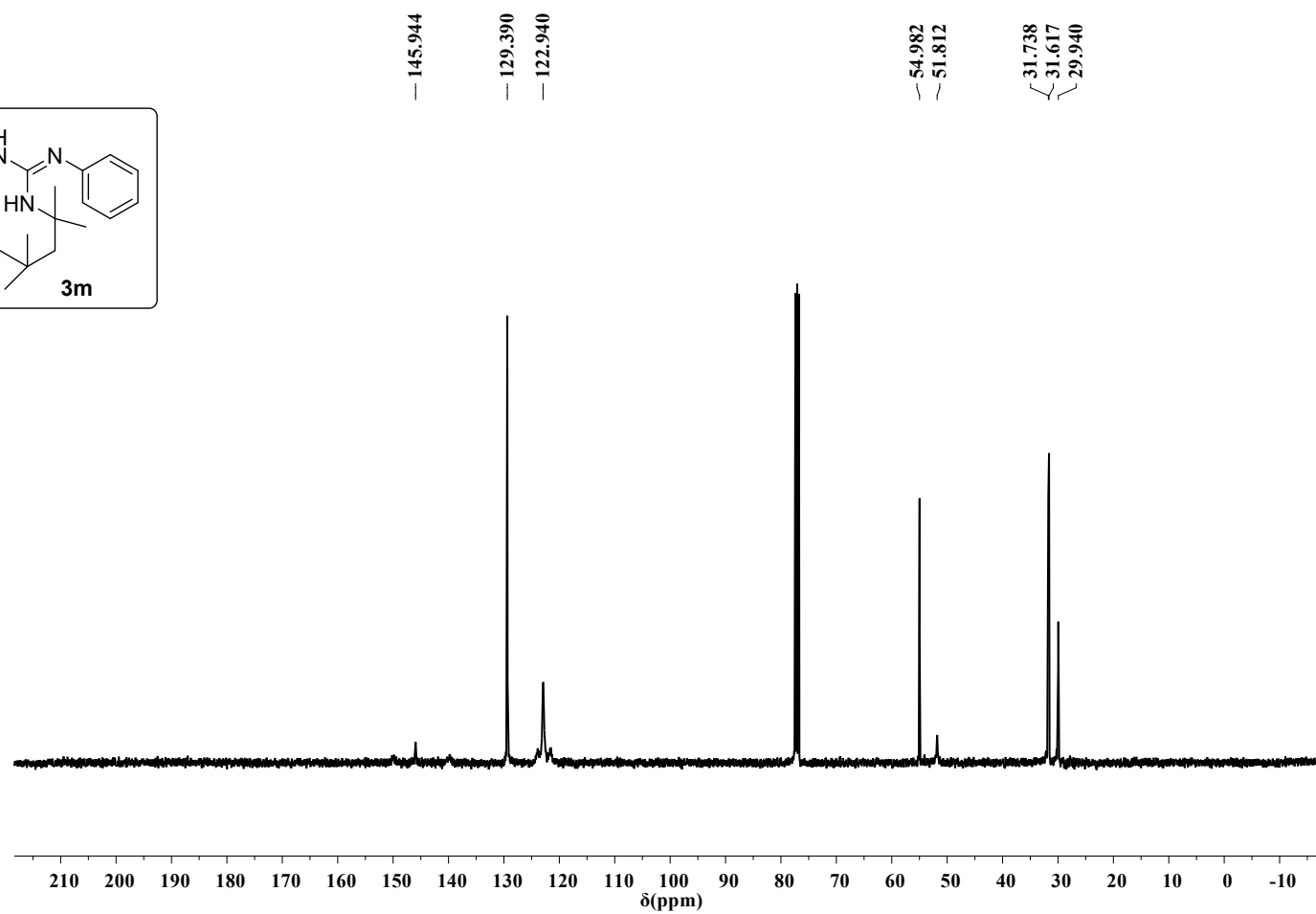
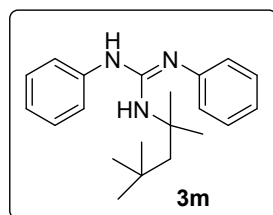
**<sup>1</sup>H NMR of 3l (400 MHz, CDCl<sub>3</sub>)**



**$^1\text{H}$  NMR of 3m (400 MHz,  $\text{CDCl}_3$ )**



**$^{13}\text{C}$  NMR of 3m (101 MHz,  $\text{CDCl}_3$ )**



**<sup>1</sup>H NMR of 4a (400 MHz, CDCl<sub>3</sub>)**

