

Development and Applications of Heterogeneous Chiral Bifunctional Alkaloid-Thioureas as Recyclable Organocatalysts

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General Experimental Information

Solvents and chemicals were obtained from typical commercial vendors and were used as received, without any further purification. Starting materials were synthesized according to literature protocols.

Solvents for extraction, chromatography and crystallization were technical grade. All solvents used in reactions were treated with dry molecular sieves as specified in the literature.¹

When required, chromatographic purification was performed by flash chromatography using commercial grades of silica gel finer than 230 mesh with pressure. Analytical thin-layer chromatography (TLC) was carried out using 60 F₂₅₄ TLC aluminum plates. Compounds were visualized by means of UV or by using KMnO₄.

¹H, and ¹³C-NMR spectra were recorded on a Bruker Avance 400 spectrometer instrument at room temperature, in CDCl₃ or acetone-d₆ as a solvent, at 300 MHz and 75 MHz, respectively. Chemical shifts (δ) are reported in ppm relative to the residual solvent peak (CDCl₃, ¹H: 7.26 ppm; ¹³C: 77.16 ppm. Acetone-d₆ ¹H: 2.09 ppm; ¹³C: 205.87, 30.60 ppm). External reference CFCI₃ (δ F = 0.0 ppm) is used for ¹⁹F NMR spectra. Coupling constants (*J*) are reported in Hertz. ¹³C NMR spectra were recorded in a broad band decoupled mode, and peak assignments were supported by Distortionless Enhanced Polarization Transfer (DEPT). Multiplicity is reported with the usual abbreviations.

High resolution mass spectra (HRMS) spectra were obtained by positive-ion electrospray ionization (ESI) using a LC-QTOF method. Data are reported in the form *m/z* (intensity relative to base = 100).

Chiral HPLC analysis was performed on a HPLC Agilent 1100/1200 Series system, with Chiracel® IA, Chiracel® IC and Chiracel® OD-H columns as stationary phases.

The absolute configuration was determined by comparison of chiral HPLC data with literature reports for compound **4a** and **6c**, and the absolute configurations of other compounds were assigned by analogy.^{2,3}

The infrared spectra (FTIR) were registered using a Nicolet iS10 spectrophotometer (Thermo Scientific), using Smart iTR accessory (ATR) and reporting the value of the peaks in cm⁻¹. IR spectra were taken as neat solids.

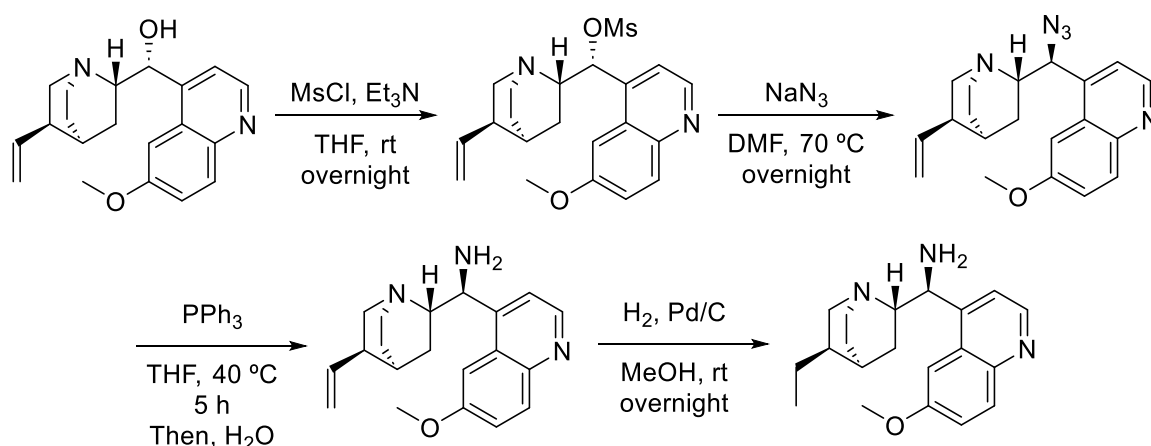
Elemental analysis was carried out according to the protocol described below. For calibration, five tin capsules were packed with acetanilide test samples of varying mass. The mass of samples was chosen so as to make the absolute content of the detected elements cover all their expected concentration range in the analyzed samples. Samples to be analyzed were placed in tin capsules, weighed and packed carefully. The prepared calibration and analysis samples were placed in the auto-sampler wherefrom they were periodically tipped into a vertical quartz reactor heated at a temperature of 980 °C with a constant flow of helium stream. A few seconds before introduction, the helium stream was enriched with high purity oxygen. The combustion gas mixture is driven through a chromium oxide zone to achieve a complete quantitative oxidation followed by a reduction step in a copper zone to reduce nitrogen oxides to nitrogen. The resulting components N₂, CO₂, H₂O are separated in a chromatographic column and detected by a thermo conductivity detector. The resulting signals, proportional to the amount of eluted gasses, are analyzed by Callidus® software which automatically provides the sample elemental composition report.

Orbital shakers IKA® KS 130 basic were used for all reactions involving the Merrifield resin derived polymer-supported catalysts.

Scanning electron microscopy (SEM) was used to study the morphology. A Hitachi TM3030Plus microscope (Tokyo, Japan), equipped with a secondary electron detector and an accelerating voltage of 15 kV, was employed. The samples were gold-coated prior to analysis. Morphological observations were also carried out using a Nikon Eclipse TS100 optical microscope equipped with a biocular head, Nikon C-W10 B/12 eyepieces, and an ELWD 0.3/OD75 objective lens. The internal calibration of the optical microscope was done using a Neubauer cell.

Synthesis and characterization of Merrifield-supported thiourea catalysts

Synthesis of cinchona alkaloid derived chiral amines:



General procedure is described for Quinine. The same procedures were used for other Quinidine, Cinchonine and Cinchonidine.

Step 1, alcohol mesylation: Quinine (1 equiv., 8.11 g, 25 mmol) was dissolved in a mixture of dry THF (50 mL) and Et_3N (3.6 equiv., 12.5 mL, 90 mmol). After cooling the resulting solution to 0°C , methanesulfonyl chloride (1.8 equiv., 3.48 mL, 45 mmol) was added dropwise. The mixture was stirred at room temperature overnight. Afterwards, the reaction was quenched with water (40 mL) and extracted with DCM (3 x 60 mL). The combined organic extracts were dried over MgSO_4 , filtered and the solvent was evaporated. The product was used directly in the next step without further purification.

Step 2, azide SN_2 : The crude mesylated alkaloid (1 equiv., 10.0 g, 25 mmol) was dissolved in dry DMF (50 mL). The solution was cooled to 0°C and NaN_3 (2 equiv., 3.25 g, 50 mmol) was added gradually in portions. The mixture was stirred at 70°C overnight and subsequently the reaction was quenched with water (40 mL) and extracted with ethyl acetate (3 x 60 mL). The organic layers were combined and washed with brine several times (15 x 50 mL) to remove DMF traces, dried over MgSO_4 and concentrated under vacuum to afford the crude product, which was used in the next step without additional purification.

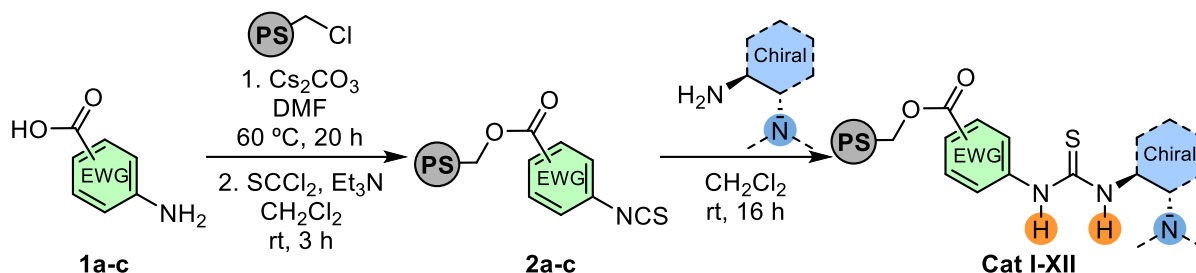
Step 3, Staudinger reaction: The crude azide (1 equiv., 8.74 g, 25 mmol) was dissolved in anhydrous THF (50 mL) under N_2 atmosphere and PPh_3 (1.05 equiv., 6.89 g, 26.25 mmol) was added in portions. The reaction mixture was stirred at 40°C for 5 h. Then, water (5 mL) was added and the mixture was stirred overnight at 40°C . After solvent removal, the residue was dissolved in aqueous HCl (2N, 80 mL) and wash with DCM (3 x 60 mL). The aqueous layer was basified with NaOH 40 % and extracted with Et_2O (3 x 60 mL). The combined Et_2O phases were dried over MgSO_4 , filtered and the solvent was evaporated to afford 5.73 g (71% from Quinine) of 9-amino-(9- deoxy)epiquinine as a yellow viscous oil.

Step 4, alkene reduction: 9-amino-(9- deoxy)epiquinine (1 equiv., 0.647 g, 2 mmol) was dissolved in MeOH (10 mL) and added to a solution of Pd on activated charcoal 10% (0.1 equiv., 0.21 g, 0.2 mmol) in MeOH (20 mL). The reaction mixture then purged with H_2 and shaken overnight at 5 bar of H_2 . The reaction mixture was filtered through a short pad of celite and washed with MeOH (3x30 mL) and

concentrated under reduced pressure to obtain 0.5 g (77%) (S)-((1S,2S,4S,5R)-5-ethylquinuclidin-2-yl)(6-methoxyquinolin-4-yl)methanamine as a dark viscous oil.

The reported data match the literature.⁴

General procedure for the synthesis of Merrifield-supported chiral catalysts I-XII:



The synthesis of catalysts **V** is used as example. Other thiourea catalysts were synthesized using analogous procedures. Squaramide catalyst **III** was synthesized following the procedure described in the literature.^{5,6}

Step 1, PS-NH₂: Merrifield resin (1 equiv., 1 g, *f* = 1.6 mmol/g, 1.6 mmol) was swollen in dry DMF (6 mL) under N₂ atmosphere, stirring in an orbital shaker. Then, Cs₂CO₃ (1.5 equiv., 0.78 g, 2.4 mmol) and 4-amino-2-(trifluoromethyl)benzoic acid (1.5 equiv., 0.49 g, 2.4 mmol) were added to the previous suspension and the reaction mixture was shaken at room temperature for 1h and afterwards, at 65 °C overnight. The resulting resin was allowed to cool down to room temperature, filtered and washed successively with water (60 mL), water/MeOH 1:1 (60 mL), MeOH/CH₂Cl₂ 1:1 (60 mL) and (60 mL). The resin was dried overnight at 50 °C.

Step 2, PS-NCS: PS-NH₂ (1 equiv., 0.9 g, 1.48 mmol) was swollen for 30 min in dry DCM. Then, Et₃N (4 equiv., 0.82 mL, 5.92 mmol) and thiophosgene (1.2 equiv., 0.14 mL, 1.77 mmol) were added dropwise to the suspension at room temperature under N₂ atmosphere. After 3h, the dark brown mixture was filtered and the resin was washed with DCM (60 mL), THF (60 mL) and DCM (60 mL). The resin was dried under vacuum for 30 min and used directly in the next step.

Step 3, Cat. V: PS-NCS (1 equiv., 0.7 g, 1.08 mmol) was pre-swollen in dry DCM for 30 min, and 9-amino-9-deoxyepiquinine (1.5 equiv., 0.52 g, 1.6 mmol) was subsequently added at room temperature. After shaking 16h, the dark brown reaction mixture was filtered and the resin was washed with DCM (60 mL), THF (60 mL) and DCM (60 mL). The resin was dried under vacuum for 12h.

The reported data match the literature.⁷

The catalyst loading of the resin was calculated based on the N elemental analysis by using the following formula:

$$f \left(\frac{\text{mmol}}{\text{g}} \right) = \frac{\%N \times 1000}{\text{number of N atoms} \times MW(N) \times 100}$$

Elem. Anal. N: 5.04%.

f = 0.90 mmol/g

As-prepared catalyst

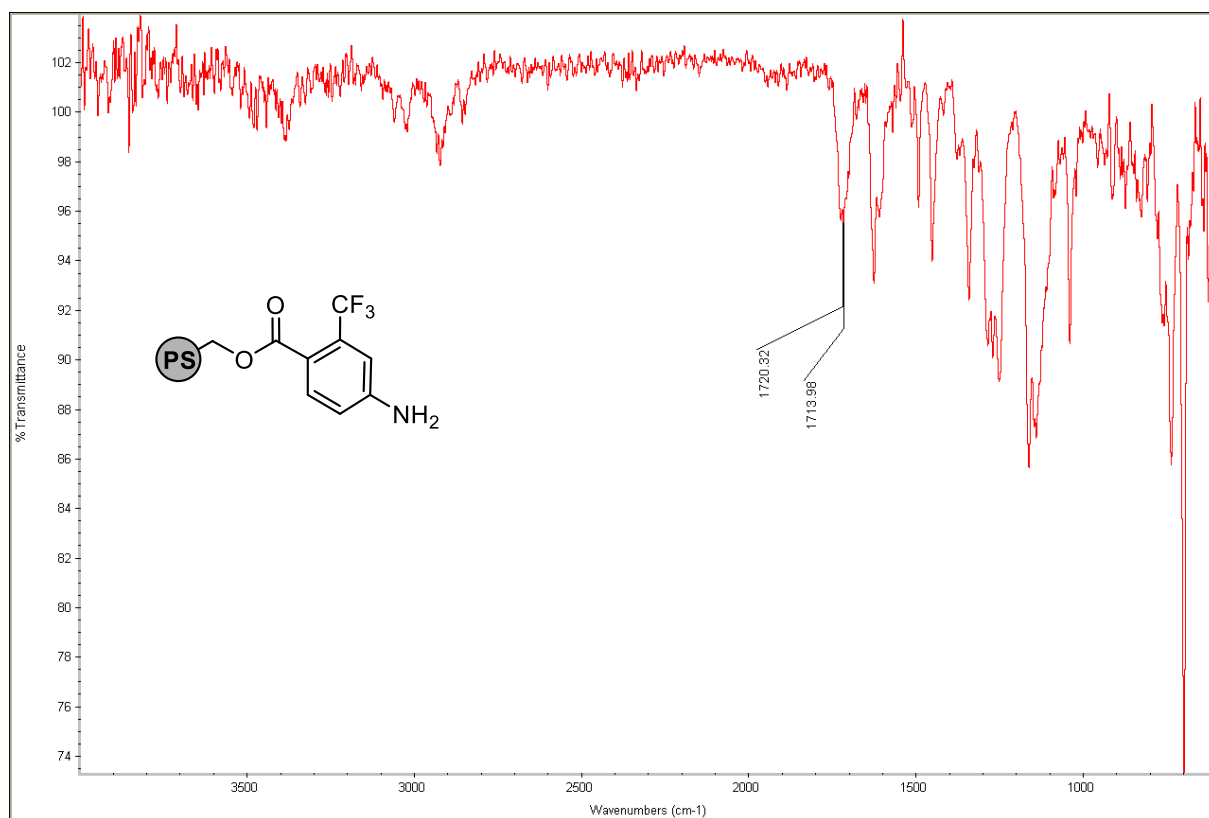


Figure S1. FTIR of the as-prepared PS-NH₂.

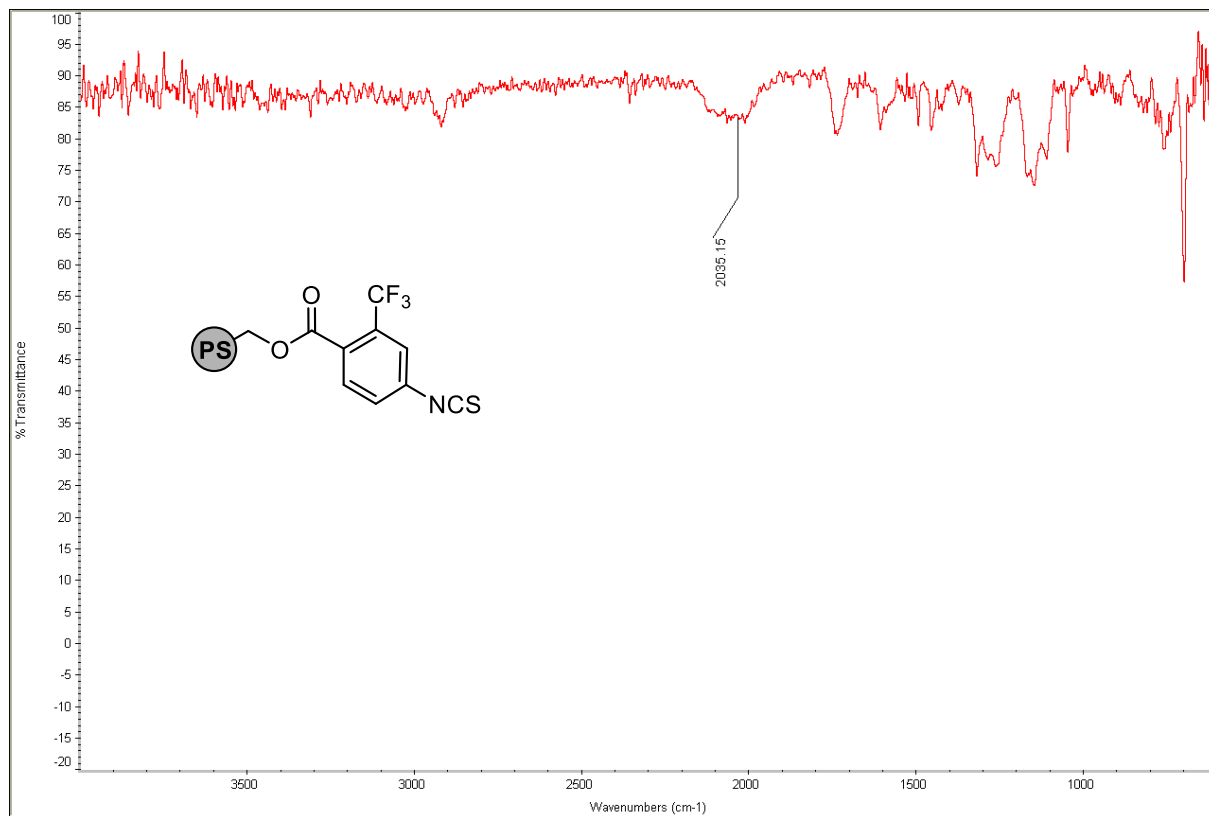


Figure S2. FTIR of the as-prepared PS-NCS.

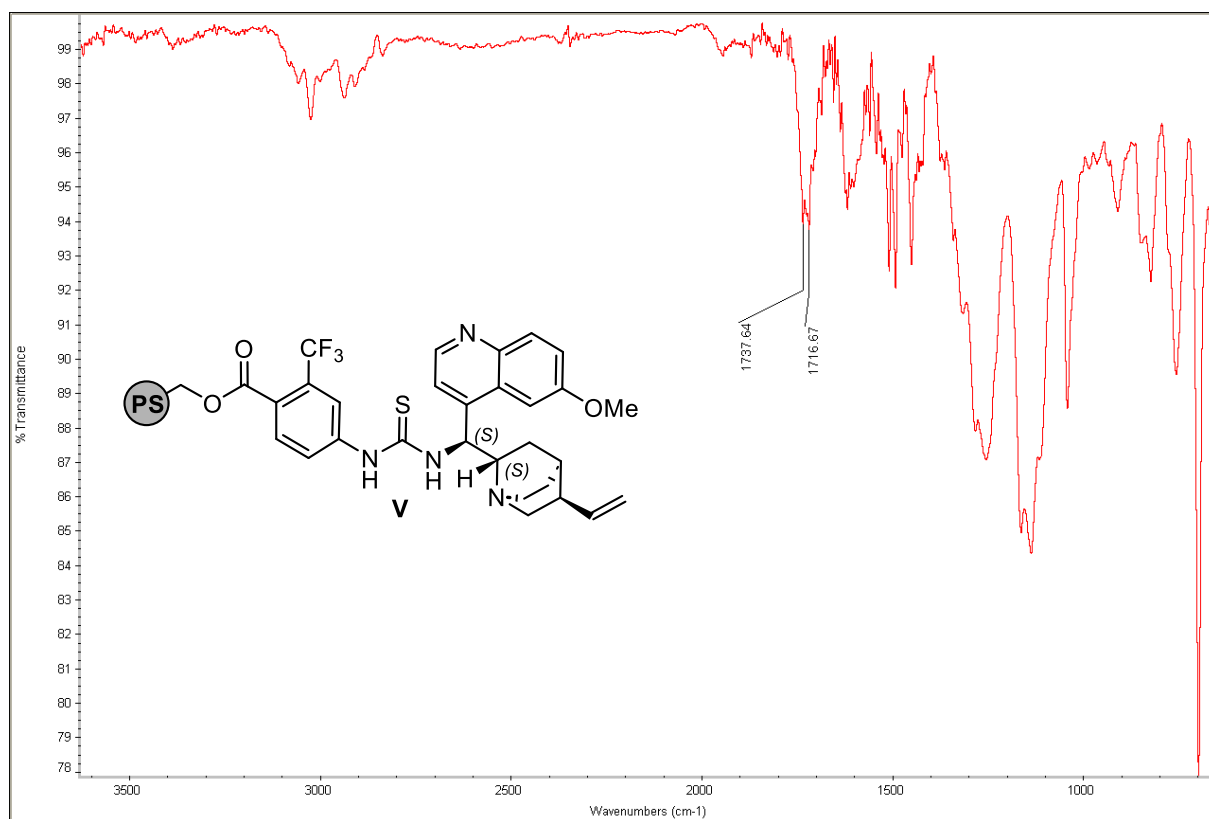


Figure S3. FTIR of the as-prepared catalyst **V**.

Used catalyst (aza-Henry reaction)

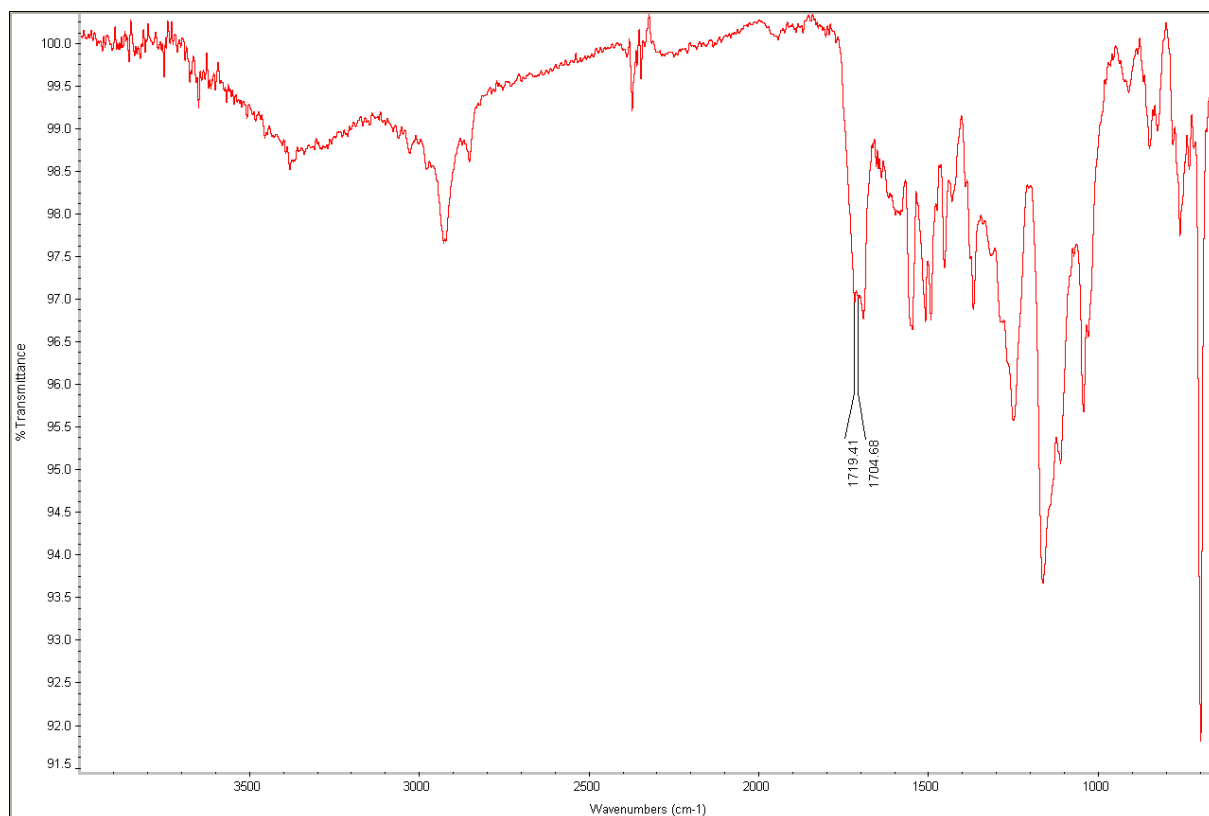


Figure S4. FTIR of catalyst **V** after 15 reaction cycles (aza-Henry reaction).

Used catalyst (Michael reaction)

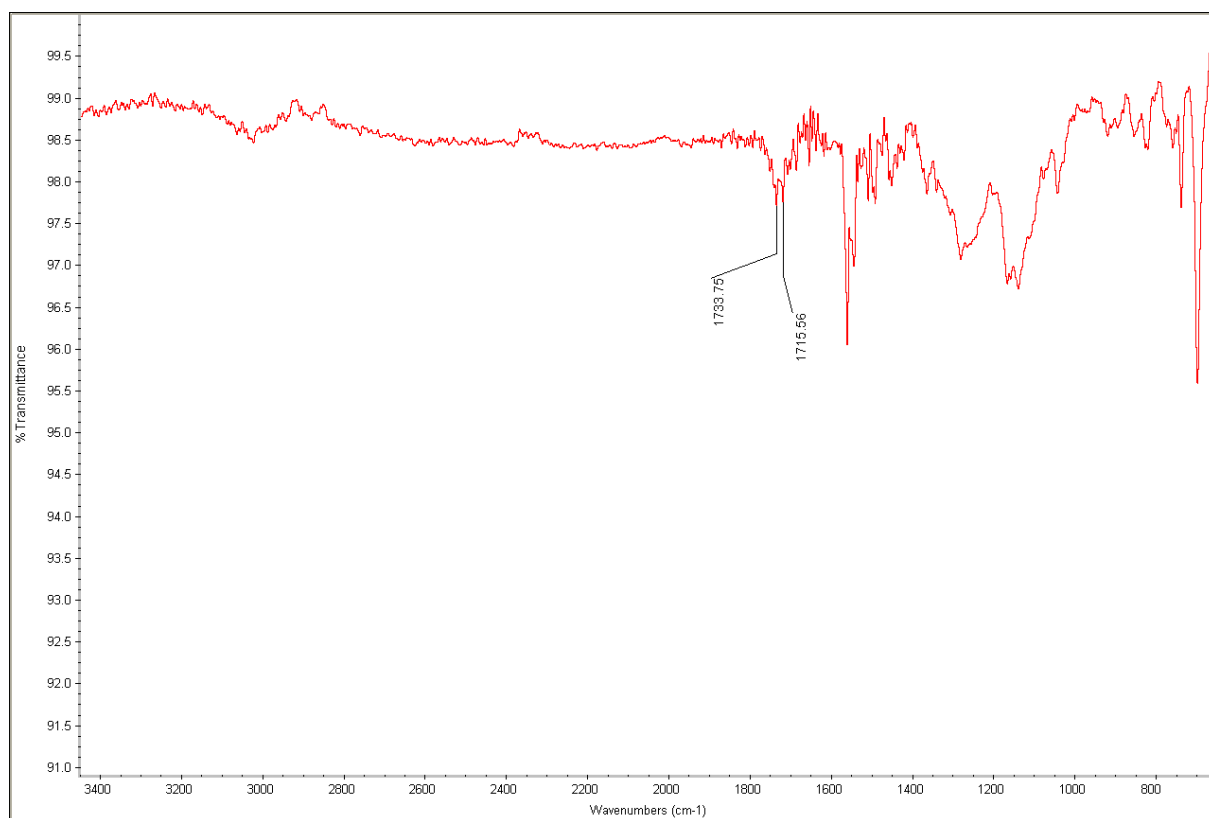


Figure S5. FTIR of catalyst **V** after 10 reaction cycles (Michael reaction).

As-prepared catalyst

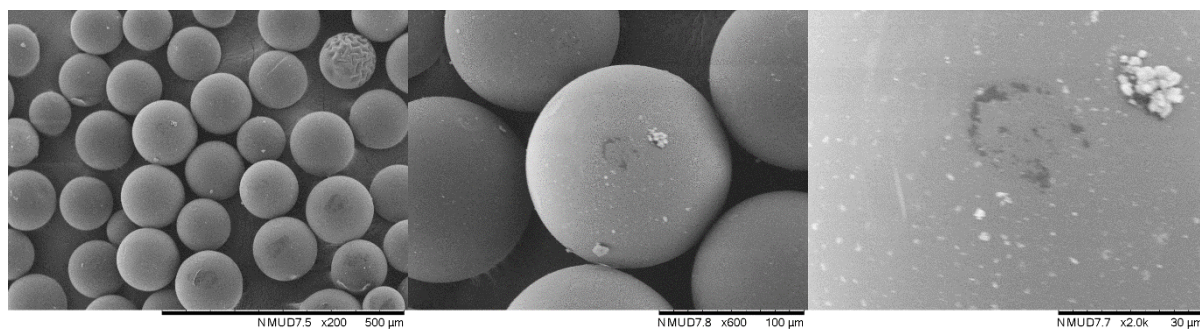


Figure S6. SEM images of the as-prepared catalyst **V**.

Used catalyst (aza-Henry reaction)

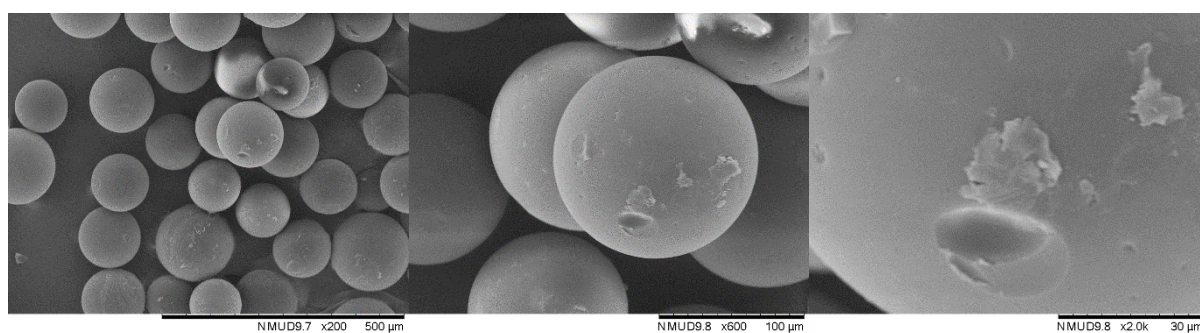


Figure S7. SEM images of catalyst **V** after 15 reaction cycles (aza-Henry reaction).

Used catalyst (Michael reaction)

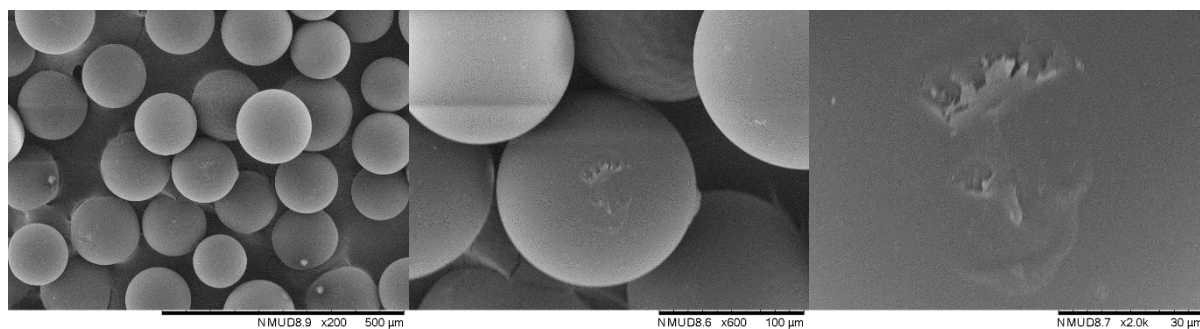


Figure S8. SEM images of catalyst **V** after 10 reaction cycles (Michael reaction).

As-prepared catalyst

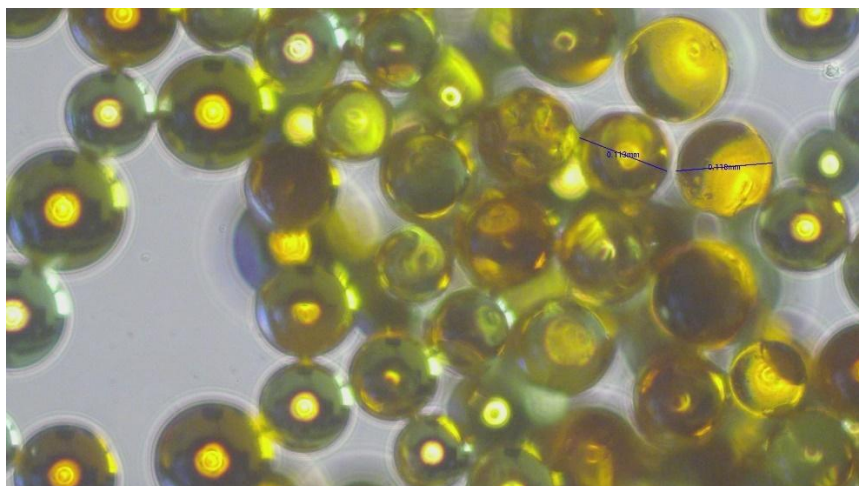


Figure S9. Optical microscope image of the as-prepared catalyst **V**.

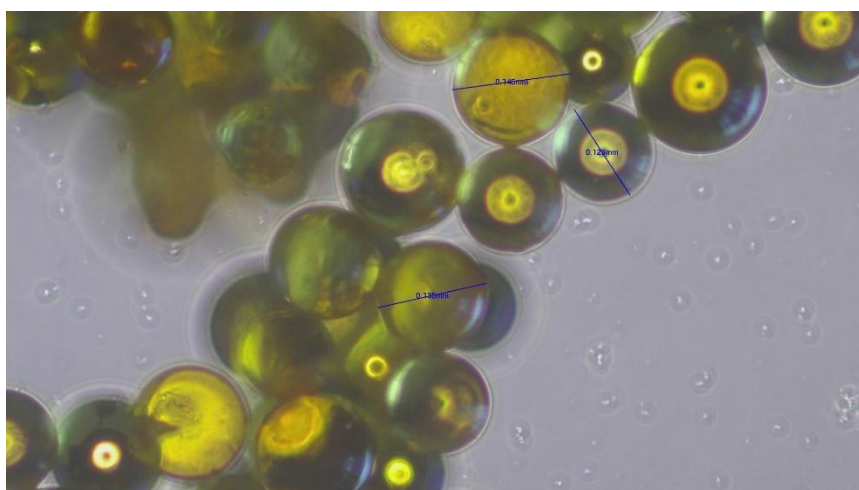


Figure S10. Optical microscope image of catalyst **V** after 15 reaction cycles (aza-Henry reaction).

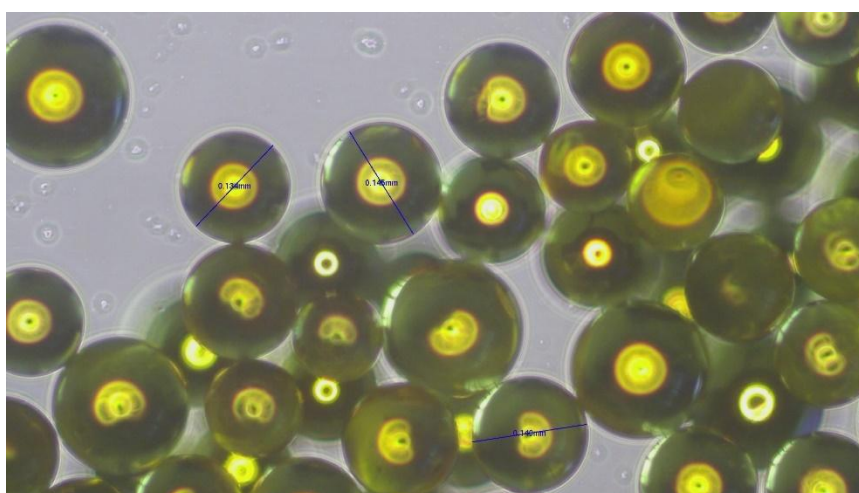


Figure S11. Optical microscope image of catalyst **V** after 10 reaction cycles (Michael reaction).

Reactor setup

All experiments involving the supported catalyst were conducted in filters so that, once the reaction was complete, the product and solvent could be easily separated from the catalyst (Figure S12). Additionally, an orbital shaker was used to stir the reactions, as magnetic stirrers were avoided to prevent mechanical degradation of the polymer-supported catalysts (Figure S13). High and low temperature experiments were carried out as shown (Figure S14).

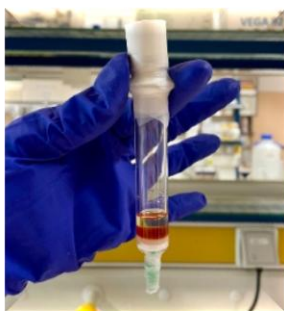


Figure S12. The filter where all the tests have been performed



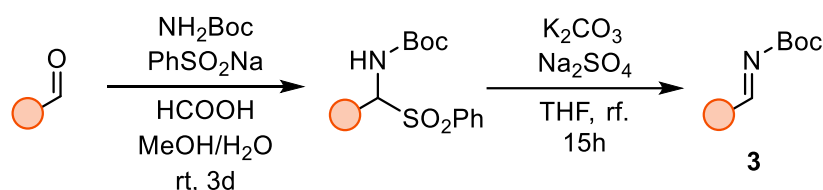
Figure S13. Orbital shaker used to stir the reaction at room temperature



Figure S14. The setup for temperature optimization

Synthesis and characterization of imines **3**

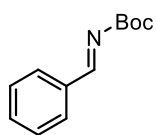
General procedure for the preparation of *N*-Boc imine:



Step 1, α -sulfonyl amine: A reaction mixture was prepared by combining benzaldehyde (1.5 equiv., 3.83 mL, 37.5 mmol), tert-butyl carbamate (1 equiv., 2.93 g, 25 mmol), sodium benzenesulfinate (2 equiv., 8.2 g, 50 mmol) and formic acid (4 equiv., 3.75 mL, 100 mmol) in a solvent mixture of methanol (50 mL) and water (100 mL). The reaction was allowed to stir 3 days at room temperature, during which time the product precipitated as a white solid. It was isolated via Büchner funnel filtration and washed with water and diethyl ether. After drying in vacuum, the product was obtained as a white solid (6.6 g, 76%).

Step 2, *N*-Boc imine: A 100 mL round-bottom flask containing potassium carbonate (6.00 equiv., 4.14 g, 30.0 mmol) and sodium sulfate (5.00 equiv., 5 g, 5 mmol) was flame-dried under vacuum and cooled under an inert atmosphere. Subsequently, the corresponding sulfonyl amine (1 equiv., 1.19 g, 5 mmol) was added with dry THF (40 mL). The mixture was refluxed under N_2 for 15h. After cooling to room temperature, it was filtered and the solvent was evaporated to afford imines **3a-l** as an oil. The appearance of the oil varied in color depending on the nature of the substituent in the starting material. The crude imine was used directly in the step without further purification.⁸

Tert-butyl (*E*)-benzylidenecarbamate (3a)

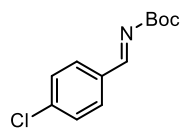


The general procedure was followed to afford 0.46 g (74 %) of the product as a white-yellow liquid. The reported data match the literature.⁸

¹H NMR (400 MHz, CDCl₃) δ 8.86 (s, 1H), 7.90 (d, ³J_{HH} = 9.6 Hz, 2H), 7.55 (t, 1H), 7.45 (t, ³J_{HH} = 6.7 Hz, 2H), 1.58 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 169.7, 162.7, 134.2, 133.6, 130.3, 129.0, 82.4, 28.0.

Tert-butyl (*E*)-(4-chlorobenzylidene)carbamate (3b)

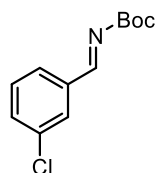


The general procedure was followed to afford 1.16 g (97 %) of the product as a white-yellow solid. The reported data match the literature.⁸

¹H NMR (400 MHz, CDCl₃) δ 8.64 (s, 1H), 7.66 (d, ³J_{HH} = 8.6 Hz, 2H), 7.25 (d, ³J_{HH} = 8.5 Hz, 2H), 1.43 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.2, 161.6, 139.0, 132.0, 130.6, 128.6, 81.5, 27.3.

Tert-butyl (*E*)-(3-chlorobenzylidene)carbamate (3c)

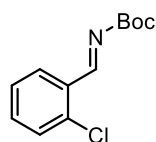


The general procedure was followed to afford 0.50 g (49 %) of the product as a white liquid. The reported data match the literature.⁹

¹H NMR (400 MHz, CDCl₃) δ 8.79 (s, 1H), 7.94 (s, 1H), 7.74 (d, ³J_{HH} = 7.8 Hz, 1H), 7.52 (d, ³J_{HH} = 6.9 Hz, 1H), 7.40 (t, ³J_{HH} = 7.8 Hz, 1H), 1.58 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.4, 161.7, 135.4, 134.7, 132.9, 129.8, 128.9, 128.2, 82.2, 27.5.

Tert-butyl (*E*)-(2-chlorobenzylidene)carbamate (3d)

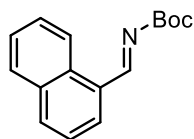


The general procedure was followed to afford 0.75 g (87 %) of the product as a white liquid. The reported data match the literature.⁹

¹H NMR (400 MHz, CDCl₃) δ 9.25 (s, 1H), 8.16 (t, ³J_{HH} = 8.0 Hz, 1H), 7.41 (t, 2H), 7.30 (t, ³J_{HH} = 7.8 Hz, 1H), 1.57 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 165.4, 162.1, 137.6, 134.0, 131.1, 129.9, 128.9, 126.9, 82.4, 27.7.

Tert-butyl (*E*)-(naphthalen-1-ylmethylene)carbamate (3e)

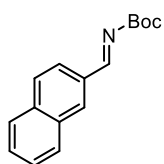


The general procedure was followed to afford 1.26 g (99 %) of the product as a yellow liquid. The reported data match the literature.⁹

¹H NMR (400 MHz, CDCl₃) δ 9.53 (s, 1H), 8.93 (d, ³J_{HH} = 8.6 Hz, 1H), 8.16 (d, ³J_{HH} = 7.3 Hz, 1H), 8.02 (d, ³J_{HH} = 8.3 Hz, 1H), 7.89 (d, ³J_{HH} = 8.1 Hz, 1H), 7.63 (t, ³J_{HH} = 7.8 Hz, 1H), 7.54 (d, ³J_{HH} = 8.4 Hz, 2H), 1.64 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 169.0, 162.9, 134.3, 133.8, 132.0, 131.9, 129.3, 128.9, 128.2, 126.6, 125.2, 124.0, 82.3, 28.1.

Tert-butyl (*E*)-(naphthalen-2-ylmethylene)carbamate (3f)

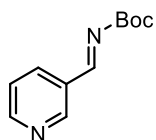


The general procedure was followed to afford 1.16 g (91 %) of the product as a white solid. The reported data match the literature.¹⁰

¹H NMR (400 MHz, CDCl₃) δ (s, 1H), 8.21 (d, 1H), 8.03 (d, ³J_{HH} = 8.6 Hz, 1H), 7.83 (t, ³J_{HH} = 13.1 Hz, 3H), 7.50 (d, ³J_{HH} = 14.6 Hz, 2H), 1.60 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 170.2, 163.1, 136.4, 134.5, 133.1, 132.2, 129.6, 129.2, 129.0, 128.3, 127.6, 124.3, 82.6, 28.4.

Tert-butyl (E)-(pyridin-3-ylmethylene)carbamate (3g)

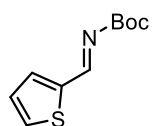


The general procedure was followed to afford 0.98 g (95 %) of the product as an orange liquid. The reported data match the literature.⁹

¹H NMR (400 MHz, CDCl₃) δ 8.99 (s, 1H), 8.85 (d, ³J_{HH} = 7.8 Hz, 1H), 8.75 (d, ⁴J_{HH} = 4.8 Hz, 1H), 8.26 (dd, ³J_{HH} = 8.0 Hz, ⁴J_{HH} = 2.0 Hz, 1H), 7.39 (d, ³J_{HH} = 7.1 Hz, 1H), 1.57 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 166.8, 162.1, 154.0, 152.3, 136.0, 130.0, 124.0, 83.0, 28.0.

Tert-butyl (E)-(thiophen-2-ylmethylene)carbamate (3h)

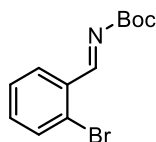


The general procedure was followed to afford 0.92 g (97 %) of the product as a brown liquid. The reported data match the literature.¹⁰

¹H NMR (400 MHz, CDCl₃) δ 9.02 (s, 1H), 7.66 (d, ⁴J_{HH} = 3.9 Hz, 1H), 7.62 (s, 1H), 7.15 – 7.10 (m, 1H), 1.52 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 163.5, 162.2, 140.3, 137.1, 134.2, 128.4, 82.1, 28.0.

Tert-butyl (E)-(2-bromobenzylidene)carbamate (3i)

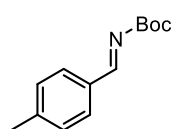


The general procedure was followed to afford 1.29 g (91 %) of the product as a white liquid. The reported data match the literature.¹¹

¹H NMR (400 MHz, CDCl₃) δ 9.15 (s, 1H), 8.12 (t, ³J_{HH} = 9.6 Hz, 1H), 7.56 (t, ³J_{HH} = 9.3 Hz, 1H), 7.32 (d, ³J_{HH} = 9.5 Hz, 2H), 1.55 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.2, 162.5, 135.6, 134.7, 133.8, 133.0, 129.8, 128.1, 82.9, 28.3.

Tert-butyl (E)-(4-methylbenzylidene)carbamate (3j)

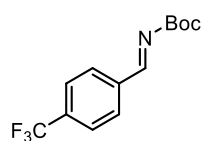


The general procedure was followed to afford 0.42 g (53 %) of the product as a yellow solid. The reported data match the literature.⁸

¹H NMR (400 MHz, CDCl₃) δ 8.87 (s, 1H), 7.81 (d, ³J_{HH} = 8.3 Hz, 2H), 7.27 (d, ³J_{HH} = 8.1 Hz, 2H), 2.42 (s, 3H), 1.59 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 170.4, 163.2, 145.0, 132.0, 130.8, 130.1, 82.5, 28.4, 22.3.

Tert-butyl (E)-(4-(trifluoromethyl)benzylidene)carbamate (3k)



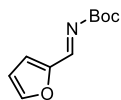
The general procedure was followed to afford 1.29 g (94 %) of the product as a white solid. The reported data match the literature.⁸

¹H NMR (400 MHz, CDCl₃) δ 8.81 (s, 1H), 7.97 (d, ³J_{HH} = 8.1 Hz, 2H), 7.68 (d, ³J_{HH} = 8.1 Hz, 2H), 1.55 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.2 (s), 162.1 (s), 137.2 (s), 134.6 (q), 130.1 (s), 125.8 (s), 82.7 (s), 27.8 (s).

¹⁹F {¹H} NMR (376 MHz, CDCl₃) δ -63.19.

Tert-butyl (*E*)-(furan-2-ylmethylene)carbamate (**3l**)



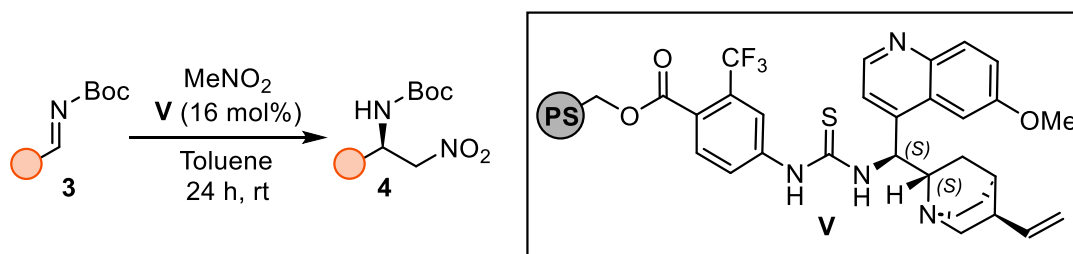
The general procedure was followed to afford 0.77 g (79 %) of the product as a brown liquid. The reported data match the literature.¹⁰

¹H NMR (400 MHz, CDCl₃) δ 8.75 (s, 1H), 7.67 (s, 1H), 7.21 (d, ⁴J_{HH} = 3.6 Hz, 2H), 6.58 (d, ⁴J_{HH} = 1.9 Hz, 1H), 1.55 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 162.3, 157.6, 150.9, 148.2, 121.4, 113.1, 82.2, 28.0.

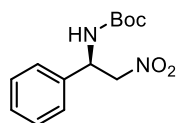
Synthesis and characterization of β-nitro amines **4**

General procedure for the enantioselective aza-Henry reaction:



In a filtered tube were added a solution of the imine in toluene (1 equiv., 1 mL, 0.2 mmol, 0.2 M), the supported-catalyst (0.16 equiv., *f* = 0.7 mmol/g, 44.0 mg, 0.032 mmol) and nitromethane (5 equiv., 53 μL, 1 mmol). The tube was shaken at room temperature overnight. Afterward, the reaction mixture was filtered off and the supported catalyst was washed three times with DCM (2 mL). After evaporating the solvent, the reaction crude was purified by chromatography in hexane/EtOAc to afford aza-Henry adducts **4a-l**. Racemic products **4** were synthesized using trimethylamine (10 mol%) as the catalyst.¹²

Tert-butyl (*R*)-(2-nitro-1-phenylethyl)carbamate (**4a**)



The general procedure was followed to afford 50.0 mg (94%) of the product as a white solid. The reported data match the literature.²

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.29 (m, 5H), 5.42 – 5.19 (m, 2H), 4.85 (s, 1H), 4.71 (dd, ³J_{HH} = 12.6 Hz, ³J_{HH} = 5.6 Hz, 1H), 1.44 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.9, 137.0, 129.4, 128.9, 126.5, 80.9, 79.0, 53.1, 28.4.

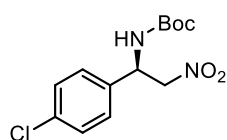
The absolute configuration of **4a** was determined by comparing the HPLC data with the literature:

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 98:2, 1.0 mL/min, 40 °C, detection at 230 nm. Residence time: 18.8 min (major), 20.7 (minor). 91:9 er.

Lit. for (*R*) isomer HPLC (chiral): Chiralpak IA, Hexanes/*i*-PrOH 90:10, 1.0 mL/min, temperature not specified, detection at 210 nm. Residence time: 12.0 min (major), 13.0 (minor).

The accumulated turnover number (TON) was calculated using the average yield of all the recycling tests for compound **4a**.

$$\text{TON} = \frac{\text{mmoles limiting reactant}}{\text{mmoles catalyst}} \times \text{yield} = \frac{3.0}{0.032} \times 0.89 = 83.4$$

Tert-butyl (R)-(1-(4-chlorophenyl)-2-nitroethyl)carbamate (4b)

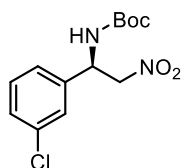
The general procedure was followed to afford 55.0 mg (93%) of the product as a white solid. The reported data match the literature.²

¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, ³J_{HH} = 8.5 Hz, 2H), 7.27 (, ³J_{HH} = 9.0 Hz, 2H), 5.37 (s, 2H), 4.84 (s, 1H), 4.71 (d, ³J_{HH} = 7.6 Hz, 1H), 1.46 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.8, 135.6, 134.8, 131.1, 129.5, 127.9, 78.8, 52.4, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 95:5, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 10.65 min (major), 12.52 (minor). 86:14 er.

Tert-butyl (R)-(1-(3-chlorophenyl)-2-nitroethyl)carbamate (4c)

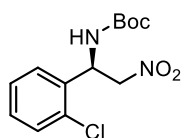
The general procedure was followed to afford 56.0 mg (93%) of the product as a white solid. The reported data match the literature.¹³

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, ²J_{HH} = 4.4 Hz, 1H), 7.36 (d, ²J_{HH} = 4.4 Hz, 1H), 7.30 (d, ³J_{HH} = 5.0 Hz, 2H), 5.73 (s, 1H), 4.88 - 4.80 (m, 1H), 1.43 (s, 9H), 1.25 (s, 2H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 132.8, 130.5, 130.2, 130.1, 128.2, 127.7, 83.6, 50.8, 29.9, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 230 nm.

Residence time: 15.12 min (major), 16.77 (minor). 92:8 er.

Tert-butyl (R)-(1-(2-chlorophenyl)-2-nitroethyl)carbamate (4d)

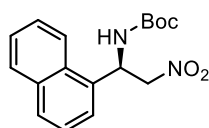
The general procedure was followed to afford 56.0 mg (93%) of the product as a white solid. The reported data match the literature.¹⁴

¹H NMR (400 MHz, CDCl₃) δ 7.40 (dd, ³J_{HH} = 5.9, ⁴J_{HH} = 3.4 Hz, 1H), 7.37 – 7.33 (m, 1H), 7.29 (dd, ³J_{HH} = 5.9, ⁴J_{HH} = 3.6 Hz, 2H), 5.73 (s, 2H), 4.94 – 4.68 (m, 2H), 1.43 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.7, 134.4, 132.7, 130.4, 130.0, 129.8, 128.2, 127.7, 80.9, 51.7, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 98:2, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 13.94 min (major), 14.60 (minor). 86:14 er.

Tert-butyl (R)-(1-(naphthalen-1-yl)-2-nitroethyl)carbamate (4e)

The general procedure was followed to afford 57.0 mg (90%) of the product as a white solid. The reported data match the literature.²

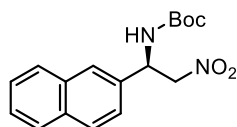
¹H NMR (400 MHz, CDCl₃) δ 8.12 (d, ³J_{HH} = 8.6 Hz, 1H), 7.91 (d, ³J_{HH} = 9.9 Hz, 1H), 7.86 (dd, ³J_{HH} = 6.8, ⁴J_{HH} = 2.8 Hz, 1H), 7.62 (t, 1H), 7.55 (t, 1H), 7.46 (t, 2H), 6.28 (d,

⁴J_{HH} = 6.6 Hz, 1H), 5.30 (s, 1H), 4.91 (s, 2H), 1.44 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.8, 134.2, 132.7, 130.4, 129.7, 129.4, 127.4, 126.4, 125.4, 123.4, 122.3, 80.9, 78.4, 49.4, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 15.45 min (major), 19.64 (minor). 88:12 er.

Tert-butyl (R)-(1-(naphthalen-2-yl)-2-nitroethyl)carbamate (4f)

The general procedure was followed to afford 60.0 mg (95%) of the product as a white solid. The reported data match the literature.¹³

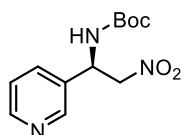
¹H NMR (400 MHz, CDCl₃) δ 7.85 (t, 3H), 7.76 (s, 1H), 7.51 (d, ³J_{HH} = 9.5 Hz, 2H), 7.40 (dd, ³J_{HH} = 8.5 Hz, ⁴J_{HH} = 2.0 Hz, 1H), 5.48 (d, 2H), 4.93 (s, 1H), 4.80 (dd, ³J_{HH} =

12.7 Hz, ⁴J_{HH} = 5.7 Hz, 1H), 1.45 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.8, 134.2, 133.3, 133.2, 129.3, 128.1, 127.7, 126.8, 126.6, 125.6, 123.8, 80.8, 78.9, 53.1, 28.3.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 20.31 min (major), 24.57 (minor). 89:11 er.

Tert-butyl (R)-(2-nitro-1-(pyridin-3-yl)ethyl)carbamate (4g)

The general procedure was followed to afford 40.0 mg (75%) of the product as a white solid. The reported data match the literature.¹⁴

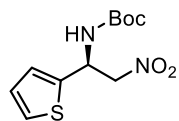
¹H NMR (400 MHz, CDCl₃) δ 8.60 (d, ³J_{HH} = 8.0 Hz, 2H), 7.65 (d, ³J_{HH} = 8.0 Hz, 1H), 7.32 (dd, ³J_{HH} = 8.0 Hz, ⁴J_{HH} = 4.9 Hz, 1H), 5.42 (d, ⁴J_{HH} = 5.7 Hz, 2H), 4.89 (s, 1H), 4.75

(dd, ³J_{HH} = 13.3 Hz, ⁴J_{HH} = 4.8 Hz, 1H), 1.44 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.8, 150.2, 148.3, 134.2, 132.9, 124.0, 81.3, 78.4, 51.0, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 90:10, 1.0 mL/min, 40 °C, detection at 210 nm.

Residence time: 10.90 min (major), 13.41 (minor). 94:6 er.

Tert-butyl (R)-(2-nitro-1-(thiophen-2-yl)ethyl)carbamate (4h)

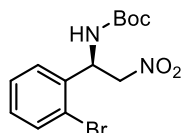
The general procedure was followed to afford 48.0 mg (90%) of the product as a white solid. The reported data match the literature.¹⁵

¹H NMR (400 MHz, CDCl₃) δ 7.27 (dd, ⁴J_{HH} = 5.0, ⁴J_{HH} = 1.4 Hz, 1H), 6.98 (dd, ³J_{HH} = 9.1 Hz, ⁴J_{HH} = 4.1 Hz, 2H), 5.66 – 5.58 (m, 1H), 5.33 (s, 1H), 4.89 (s, 1H), 4.75 (dd, ³J_{HH} = 12.9 Hz, ⁴J_{HH} = 5.6 Hz, 1H), 1.45 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 155.0, 140.5, 127.8, 126.1, 125.7, 81.4, 79.1, 49.3, 28.7.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 20.43 min (major), 22.73 (minor). 88:12 er.

Tert-butyl (R)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4i)

The general procedure was followed to afford 54.0 mg (80%) of the product as a white solid. The reported data match the literature.¹⁶

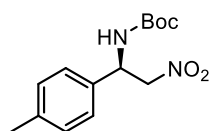
¹H NMR (400 MHz, CDCl₃) δ 7.59 (d, ³J_{HH} = 7.6 Hz, 1H), 7.36 – 7.32 (m, 2H), 7.23 – 7.18 (m, 1H), 5.73 (s, 2H), 4.91 – 4.67 (m, 2H), 1.43 (s, 9H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.64, 136.06, 133.72, 130.56, 130.24, 128.24, 128.04, 122.82, 80.92, 52.61, 28.36.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 98:2, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 15.78 min (minor), 25.48 (major). 10:90 er.

Tert-butyl (R)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4j)



The general procedure was followed to afford 30.0 mg (54%) of the product as a white solid. The reported data match the literature.¹⁴

¹H NMR (400 MHz, Acetone-d₆) δ 7.36 (d, ³J_{HH} = 8.1 Hz, 2H), 7.20 (d, ³J_{HH} = 7.9 Hz, 2H), 6.80 (s, 1H), 5.44 (d, ³J_{HH} = 7.4 Hz, 1H), 4.87 (d, ³J_{HH} = 8.1 Hz, 2H), 2.31 (s, 3H),

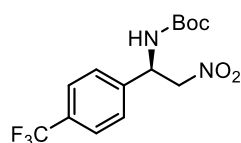
1.38 (s, 9H).

¹³C {¹H} NMR (101 MHz, Acetone-d₆) δ 192.6, 155.9, 138.7, 136.7, 130.3, 127.7, 79.8, 53.8, 28.6, 21.2.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 97:3, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 11.33 min (major), 12.76 (minor). 91:9 er.

Tert-butyl (R)-(2-nitro-1-(4-(trifluoromethyl)phenyl)ethyl)carbamate (4k)



The general procedure was followed to afford 44.0 mg (66%) of the product as a white solid. The reported data match the literature.²

¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, ³J_{HH} = 8.4 Hz, 2H), 7.45 (d, ³J_{HH} = 8.4 Hz, 2H), 5.44 (s, 2H), 4.85 (s, 1H), 4.74 (s, 1H), 1.44 (s, 9H).

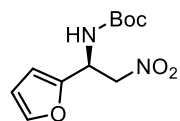
¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.7(s), 141.1 (s), 131.1 (s), 126.9 (s), 126.3 (q, ²J_{HH} = 4.0 Hz), 126.3 (s), 81.2 (s), 78.7 (s), 52.5 (s), 28.3 (s).

¹⁹F {¹H} NMR (376 MHz, CDCl₃) δ -62.78.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 97:3, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 13.45 min (major), 15.29 (minor). 62:38 er.

Tert-butyl (R)-(1-(furan-2-yl)-2-nitroethyl)carbamate (4l)



The general procedure was followed to afford 38.0 mg (75%) of the product as a white solid. The reported data match the literature.²

¹H NMR (400 MHz, CDCl₃) δ 7.38 (s, 1H), 6.32 (dd, ³J_{HH} = 13.0 Hz, ⁴J_{HH} = 2.6 Hz, 2H), 5.51-5.22 (m, 2H), 4.83 (s, 1H), 4.72 (dd, ³J_{HH} = 12.9 Hz, ⁴J_{HH} = 5.8 Hz, 1H), 1.45 (s, 9H).

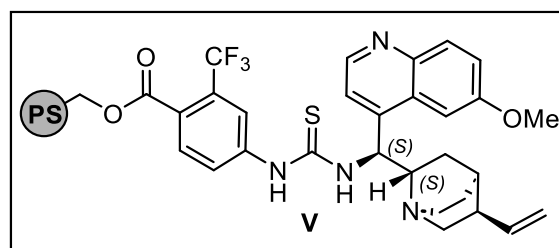
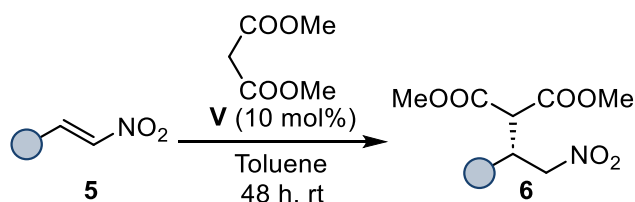
¹³C {¹H} NMR (101 MHz, CDCl₃) δ 154.8, 149.6, 143.0, 110.8, 107.9, 81.0, 76.7, 47.3, 28.4.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 98:2, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 15.03 min (minor), 20.16 (major). 37:63 er.

Synthesis and characterization of Michael adducts 6

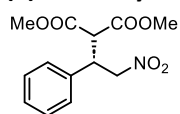
General procedure for the enantioselective Michael addition:



A solution of β-Nitrostyrene **5a** (1 equiv., 74.5 mg, 0.5 mmol) and dimethyl malonate (2 equiv., 114 μL, 1.0 mmol) in toluene (1.0 mL, 1 M) was added to a fritted tube containing the catalyst **V** (0.1 equiv., 74 mg, 0.05 mmol). The tube was then closed with a rubber septum and shaken at room temperature for 2 days. The reaction mixture was filtered off and the supported catalyst was washed three times with

DCM (2 mL). After evaporating the solvent, the reaction crude was purified by chromatography in Hexanes/EtOAc to afford Michael adduct **6a**. Racemic products **6** were synthesized using trimethylamine (10 mol%) as the catalyst.

(S)-dimethyl 2-(2-nitro-1-phenylethyl)malonate (6a)



The general procedure was followed to afford 116.0 mg (83%) of the product as a yellow solid. The reported data match the literature.¹⁷

¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.19 (m, 5H), 4.99 – 4.79 (m, 2H), 4.29 – 4.19 (m, 1H), 3.86 (d, ³J_{HH} = 9.0 Hz, 1H), 3.76 (s, 3H), 3.56 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.0, 167.4, 136.3, 129.2, 128.6, 128.0, 77.5, 54.9, 53.2, 53.0, 43.1.

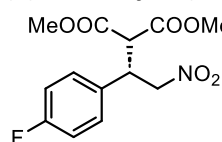
HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 18.5 min (major), 26.8 (minor). 90:10 er.

The accumulated turnover number (TON) was calculated using the average yield of all the recycling tests for compound **6a**.

$$\text{TON} = \frac{\text{mmoles limiting reactant}}{\text{mmoles catalyst}} \times \text{yield} = \frac{5.0}{0.05} \times 0.61 = 61.0$$

(S)-dimethyl 2-(1-(4-fluorophenyl)-2-nitroethyl)malonate (6b)



The general procedure was followed to afford 135.0 mg (90 %) of the product as a yellow solid. The reported data match the literature.¹⁸

¹H NMR (400 MHz, CDCl₃) δ 7.24 – 7.19 (m, 2H), 7.01 (t, ³J_{HH} = 8.6 Hz, 2H), 4.93 – 4.80 (m, 2H), 4.26 – 4.20 (m, 1H), 3.82 (d, ³J_{HH} = 9.1 Hz, 1H), 3.76 (s, 3H), 3.57 (s, 3H).

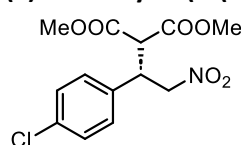
¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.1, 167.6, 164.2, 161.7, 130.2, 130.1 (d, ³J_{HH} = 8.4 Hz), 116.6 (d, ³J_{HH} = 21.4 Hz), 116.4, 55.1, 53.5, 53.3, 42.7.

¹⁹F NMR (376 MHz, CDCl₃) δ -113.2.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 26.6 min (major), 34.4 (minor). 92:8 er.

(S)-dimethyl 2-(1-(4-chlorophenyl)-2-nitroethyl)malonate (6c)



The general procedure was followed to afford 0.140 g (89%) of the product as a yellow solid. The reported data match the literature.³

¹H NMR (400 MHz, CDCl₃) δ 7.25 (d, ³J_{HH} = 8.6 Hz, 2H), 7.16 (d, ³J_{HH} = 8.8 Hz, 2H), 4.91 – 4.77 (m, 2H), 4.23 – 4.15 (m, 1H), 3.81 (d, ³J_{HH} = 9.1 Hz, 1H), 3.69 (s, 3H), 3.51 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.6, 167.0, 134.7, 134.1, 129.3, 129.1, 77.1, 54.4, 52.9, 52.8, 42.3.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 30.4 min (major), 36.9 (minor). 90:10 er.

The absolute configuration of **6c** was determined by comparing the HPLC data with the literature:

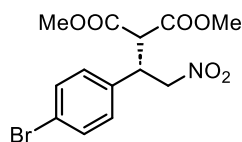
HPLC (chiral): Chiralpak-OD-H, n-heptane/EtOH 95:5, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 17.1 min (major), 20.3 (minor). 91:9 er.

Lit. for (S) isomer HPLC (chiral): Chiralcel-OD-H, Hexanes/i-PrOH 70:30, 1.0 mL/min, temperature not specified, detection at 220 nm.

Residence time: 10.2 min (major), 13.6 (minor).

(S)-dimethyl 2-(1-(4-bromophenyl)-2-nitroethyl)malonate (6d)



The general procedure was followed to afford 0.142 g (80%) of the product as a yellow solid. The reported data match the literature.¹⁸

¹H NMR (400 MHz, CDCl₃) δ 7.45 (d, ³J_{HH} = 8.0 Hz, 2H), 7.12 (d, ³J_{HH} = 8.0 Hz, 2H), 4.97 – 4.77 (m, 2H), 4.28 – 4.13 (m, 1H), 3.82 (d, ³J_{HH} = 9.0 Hz, 1H), 3.76 (s, 3H),

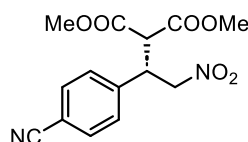
3.59 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.7, 167.2, 135.3, 132.3, 129.7, 122.7, 77.2, 54.6, 53.3, 53.1, 42.5.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 32.9 min (major), 39.9 (minor). 92:8 er.

(S)-dimethyl 2-(1-(4-cyanophenyl)-2-nitroethyl)malonate (6e)



The general procedure was followed to afford 0.097 g (64%) of the product as a yellow solid. The reported data match the literature.¹⁹

¹H NMR (400 MHz, CDCl₃) δ 7.64 (d, ³J_{HH} = 8.1 Hz, 2H), 7.38 (d, ³J_{HH} = 8.1 Hz, 2H), 5.01 – 4.82 (m, 2H), 4.37 – 4.22 (m, 1H), 3.85 (d, ³J_{HH} = 8.8 Hz, 1H), 3.77 (s,

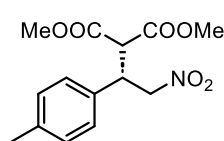
3H), 3.60 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 167.5, 166.9, 141.7, 132.9, 129.0, 118.2, 112.8, 76.8, 54.3, 53.4, 53.2, 42.9.

HPLC (chiral): Chiralpak-IC, n-heptane/EtOH 97:3, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 61.6 min (major), 73.4 (minor). 88:12 er.

(S)-dimethyl 2-(2-nitro-1-(p-tolyl)ethyl)malonate (6f)



The general procedure was followed to afford 0.095 g (64%) of the product as a yellow solid. The reported data match the literature.¹⁷

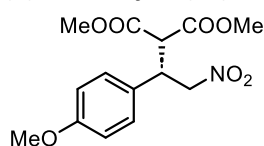
¹H NMR (400 MHz, CDCl₃) δ 7.18 – 7.01 (m, 4H), 4.99 – 4.76 (m, 2H), 4.29 – 4.10 (m, 1H), 3.84 (d, ³J_{HH} = 9.0 Hz, 1H), 3.75 (s, 3H), 3.57 (s, 3H), 2.30 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.0, 167.4, 138.3, 133.1, 129.8, 127.8, 77.7, 54.9, 53.2, 52.9, 42.7, 21.2.

HPLC (chiral): Chiralpak-IA, n-heptane/EtOH 99:1, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 16.1 min (major), 19.8 (minor). 84:16 er.

(S)-dimethyl 2-(1-(4-methoxyphenyl)-2-nitroethyl)malonate (6g)



The general procedure was followed to afford 0.097 g (62%) of the product as a yellow liquid. The reported data match the literature.¹⁷

¹H NMR (400 MHz, CDCl₃) δ 7.14 (d, ³J_{HH} = 8.1 Hz, 2H), 6.83 (d, ³J_{HH} = 8.1 Hz, 2H), 4.96 – 4.71 (m, 2H), 4.27 – 4.06 (m, 1H), 3.82 (d, ³J_{HH} = 9.0 Hz, 1H), 3.80 –

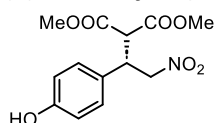
3.68 (m, 6H), 3.57 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.0, 167.4, 159.6, 129.1, 128.0, 114.5, 77.8, 55.3, 55.0, 53.1, 53.0, 42.4.

HPLC (chiral): Chiralpak-IC, n-heptane/EtOH 97:3, 1.0 mL/min, 40 °C, detection at 254 nm.

Residence time: 21.3 min (major), 25.0 (minor). 91:9 er.

(S)-dimethyl 2-(1-(4-hydroxyphenyl)-2-nitroethyl)malonate (6h)



The general procedure was followed to afford 0.112 g (76%) of the product as a yellow solid. The reported data match the literature.²⁰

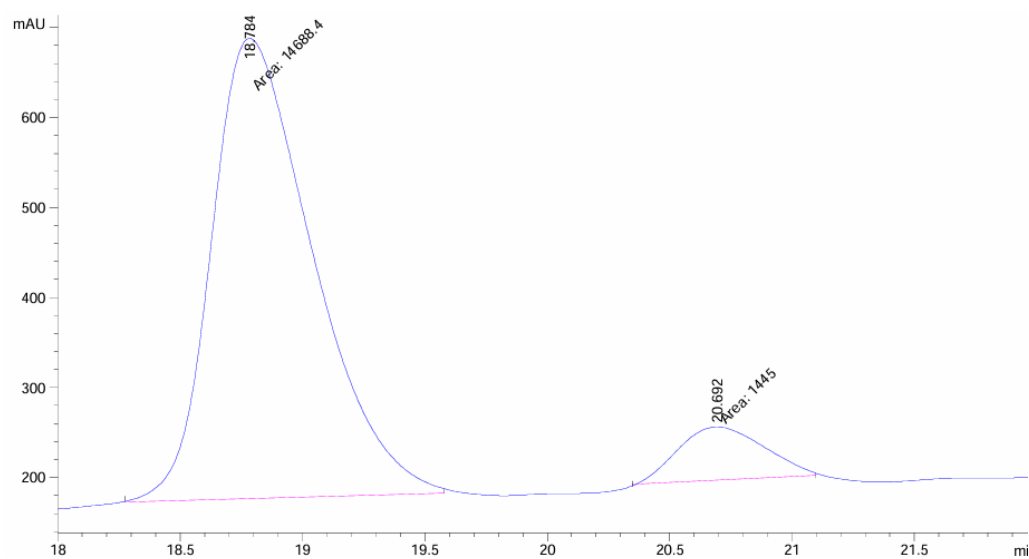
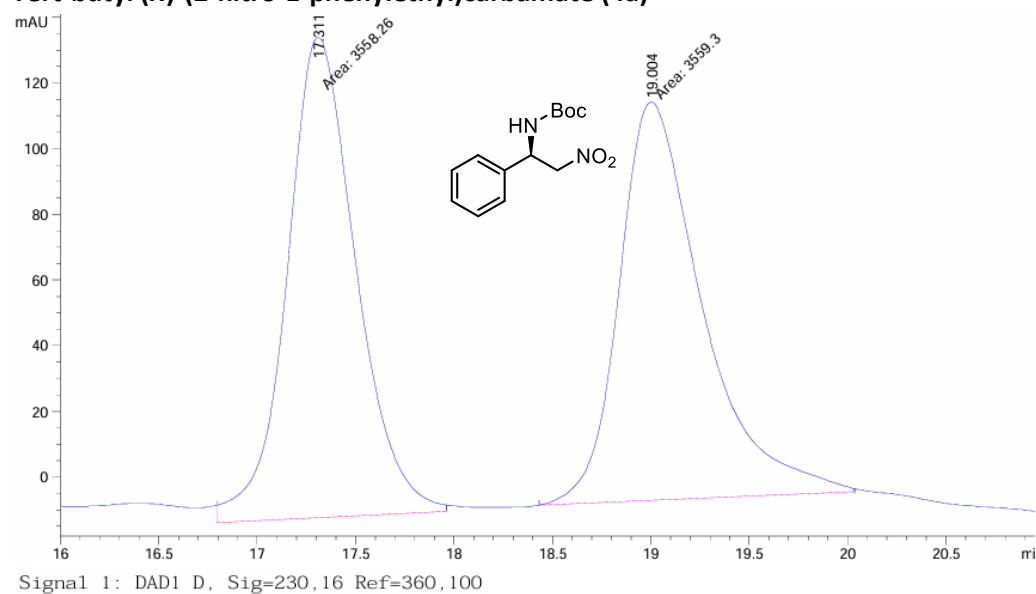
¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, ³J_{HH} = 8.2 Hz, 2H), 6.73 (d, ³J_{HH} = 8.0 Hz, 2H), 5.19 (s, 1H), 4.98 – 4.71 (m, 2H), 4.25 – 4.11 (m, 1H), 3.82 (d, ³J_{HH} = 9.2 Hz, 1H), 3.76 (s, 3H), 3.57 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 168.0, 167.6, 155.8, 129.4, 128.0, 116.1, 77.8, 55.0, 53.2, 53.1, 42.45.

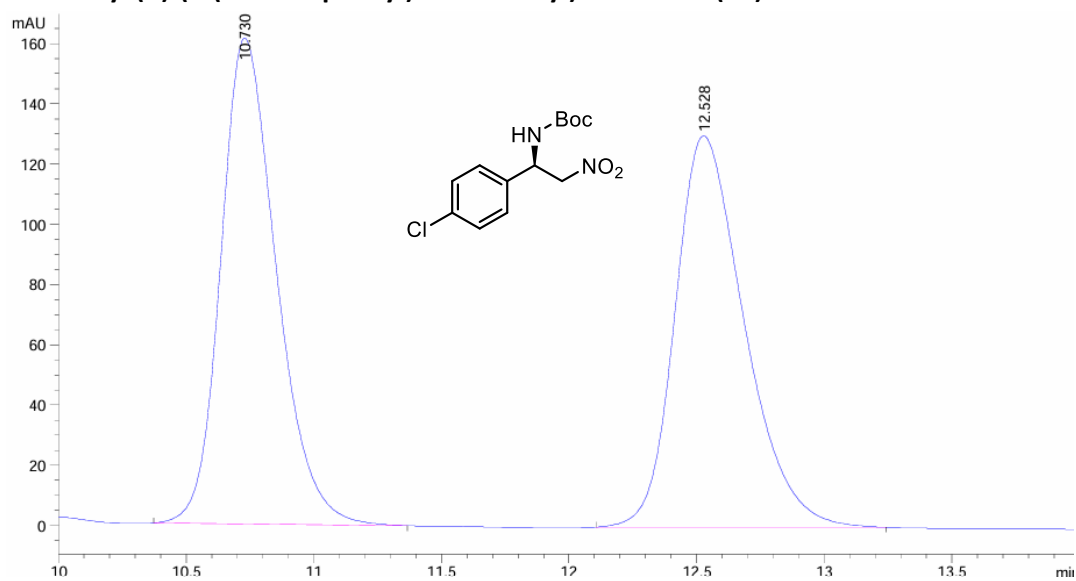
HPLC (chiral): Chiralpak-IC, n-heptane/EtOH 97:3, 1.0 mL/min, 40 °C, detection at 254 nm.
Residence time: 48.1 min (major), 51.5 (minor). 88:12 er.

Chiral HPLC chromatograms of 4

Tert-butyl (R)-(2-nitro-1-phenylethyl)carbamate (4a)



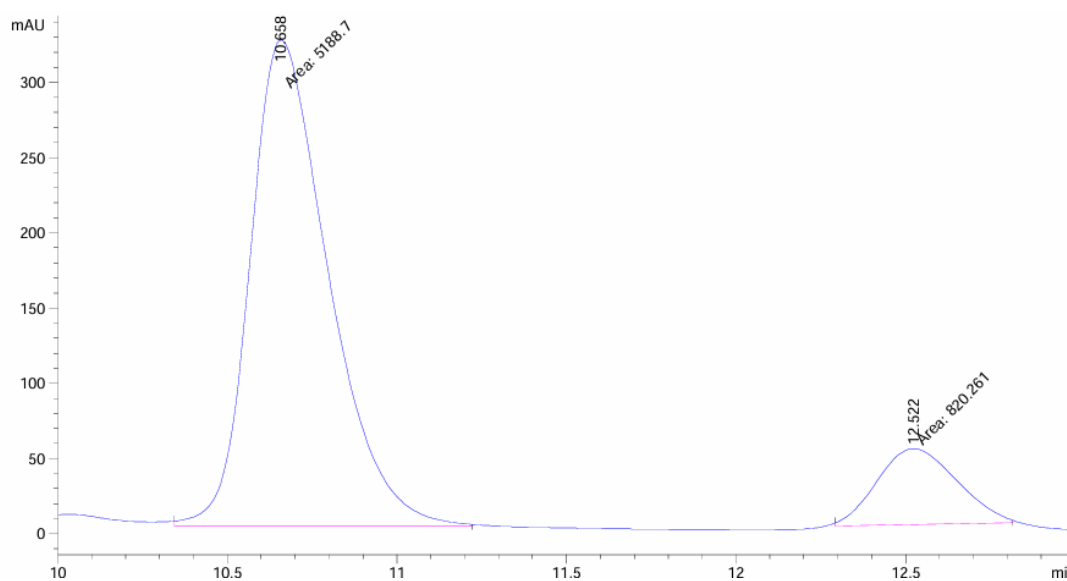
Tert-butyl (*R*)-(1-(4-chlorophenyl)-2-nitroethyl)carbamate (4b)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.730	BB	0.2308	2443.55518	161.28192	49.7940
2	12.528	BB	0.2888	2463.77222	130.16356	50.2060

Totals : 4907.32739 291.44548

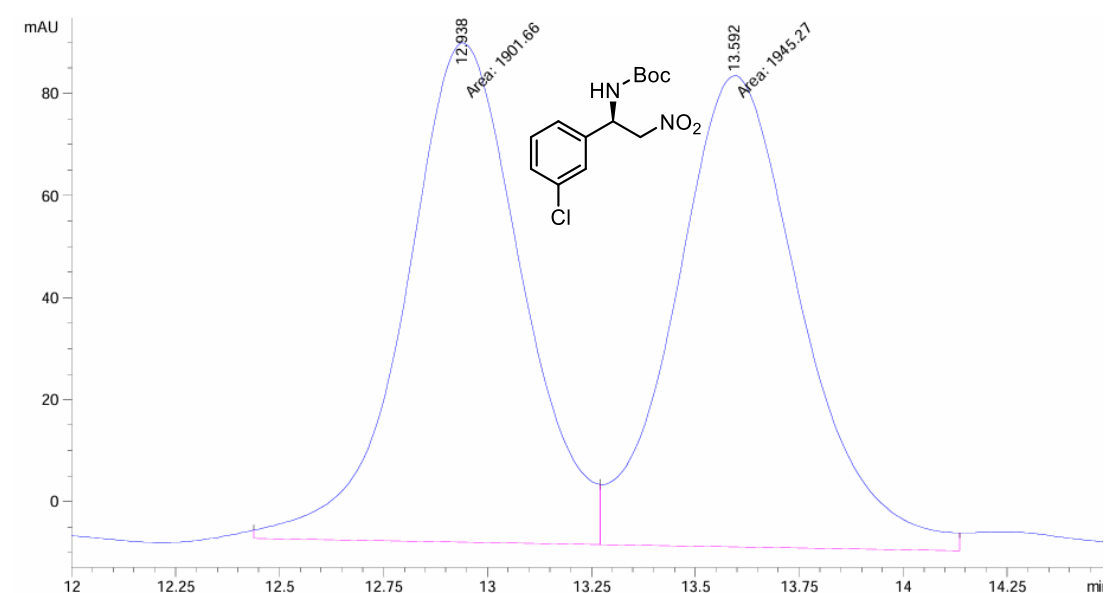


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.658	MM	0.2677	5188.70068	323.03860	86.3494
2	12.522	MM	0.2713	820.26141	50.38549	13.6506

Totals : 6008.96210 373.42409

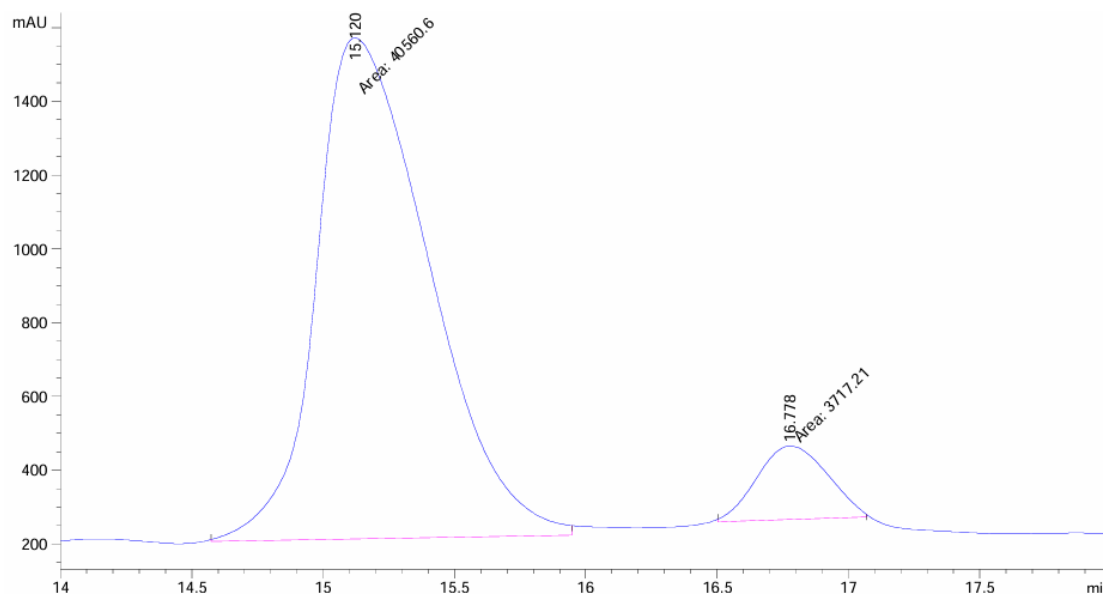
Tert-butyl (R)-(1-(3-chlorophenyl)-2-nitroethyl)carbamate (4c)



Signal 1: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.938	MF	0.3241	1901.65735	97.79926	49.4332
2	13.592	FM	0.3513	1945.26550	92.28041	50.5668

Totals : 3846.92285 190.07967

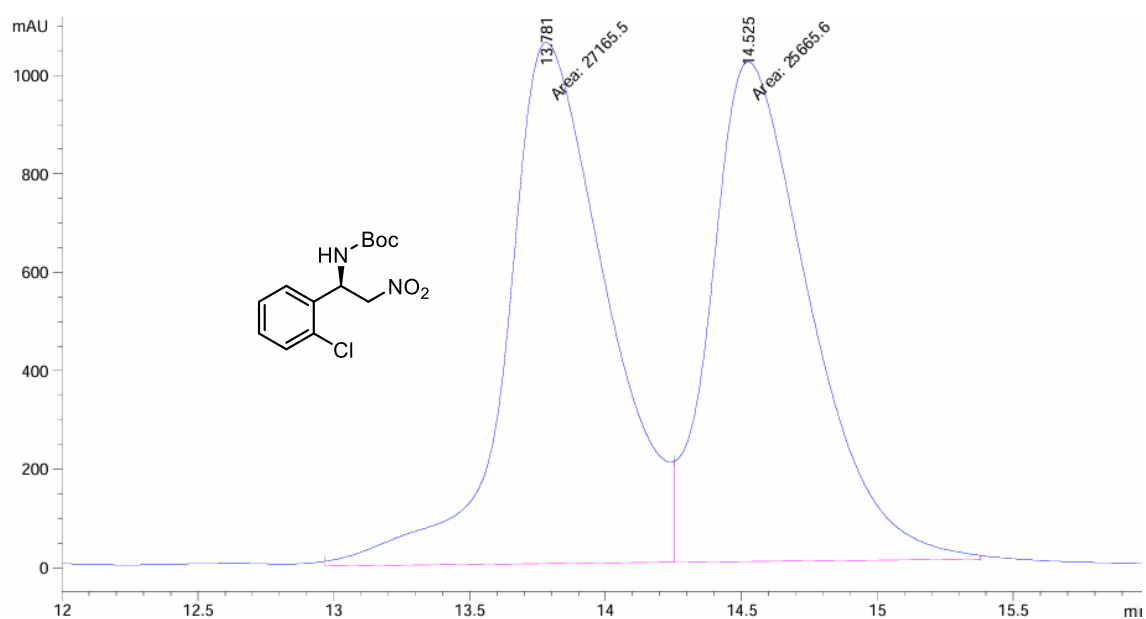


Signal 1: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.120	MM	0.4976	4.05606e4	1358.56177	91.6048
2	16.778	MM	0.3107	3717.20703	199.42900	8.3952

Totals : 4.42778e4 1557.99077

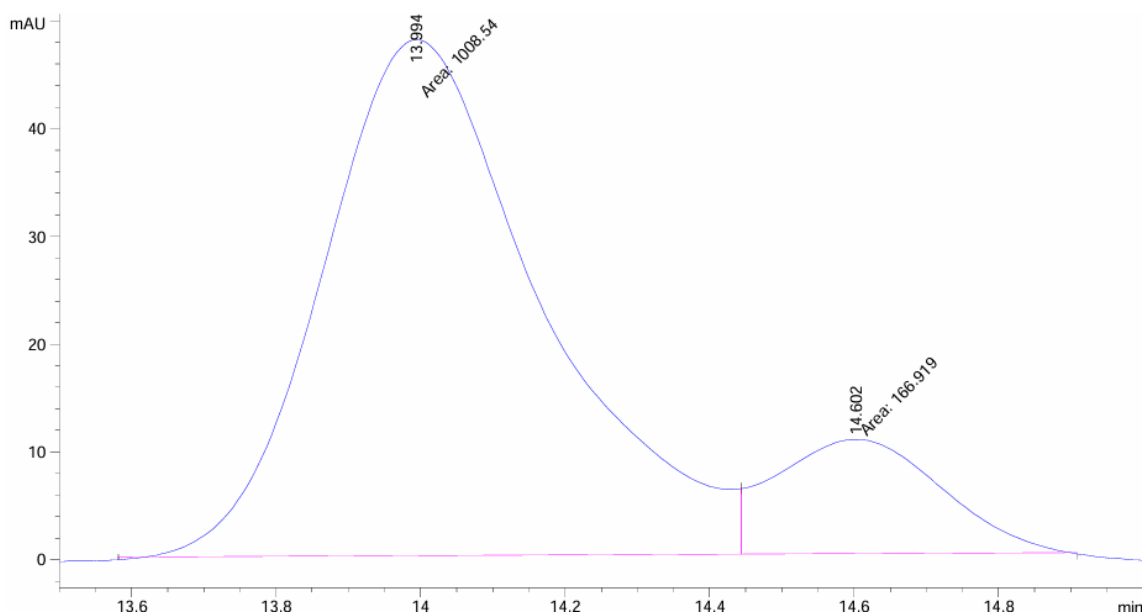
Tert-butyl (*R*)-(1-(2-chlorophenyl)-2-nitroethyl)carbamate (4d)



Signal 1: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.781	MF	0.4276	2.71655e4	1058.89563	51.4195
2	14.525	FM	0.4213	2.56656e4	1015.36279	48.5805

Totals : 5.28311e4 2074.25842

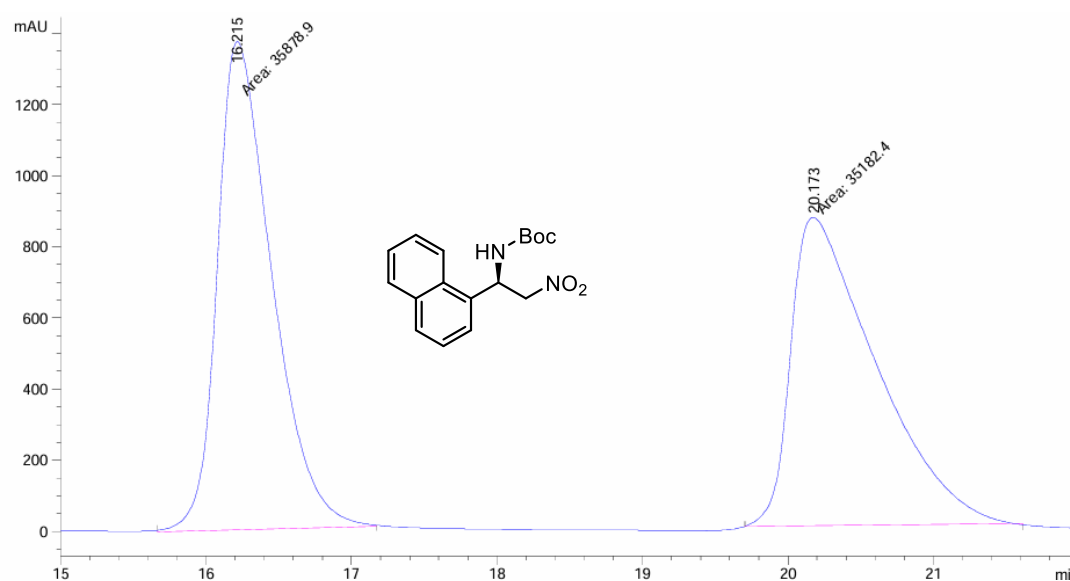


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.994	MF	0.3509	1008.54425	47.90654	85.7997
2	14.602	FM	0.2620	166.91896	10.61637	14.2003

Totals : 1175.46321 58.52291

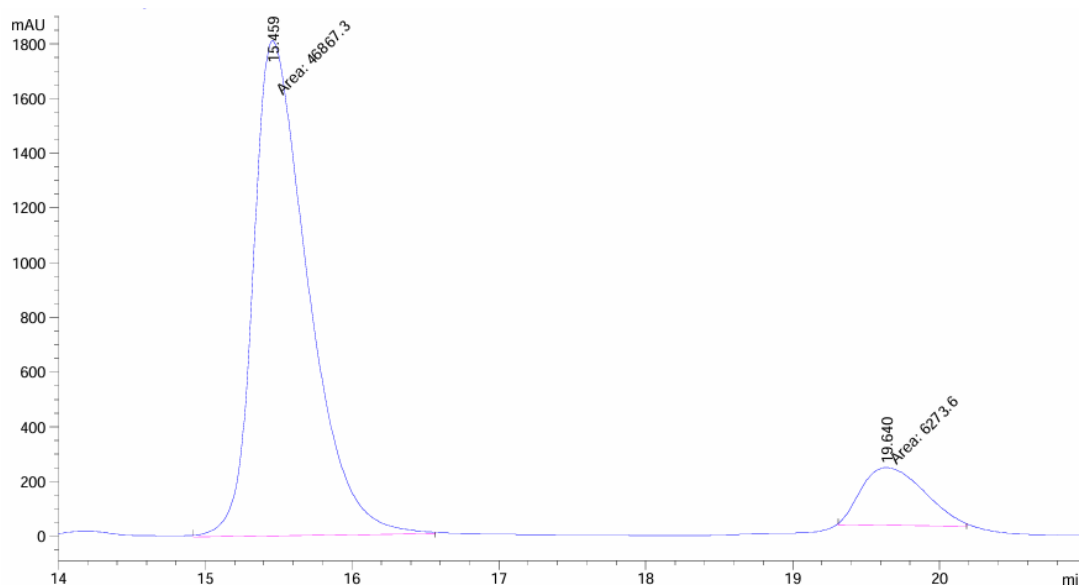
Tert-butyl (R)-(1-(naphthalen-1-yl)-2-nitroethyl)carbamate (4e)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.215	MM	0.4353	3.58789e4	1373.73767	50.4901
2	20.173	MM	0.6761	3.51824e4	867.30377	49.5099

Totals : 7.10613e4 2241.04144

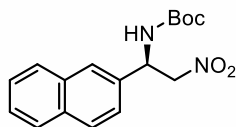
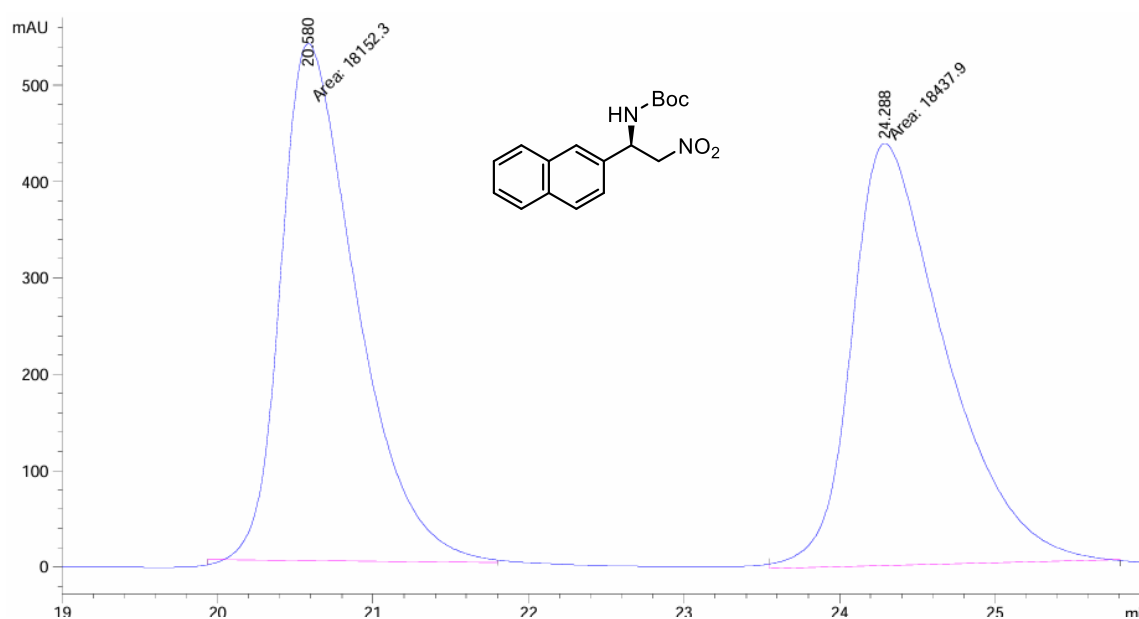


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.459	MM	0.4314	4.68673e4	1810.73157	88.1944
2	19.640	MM	0.4953	6273.59863	211.09323	11.8056

Totals : 5.31409e4 2021.82480

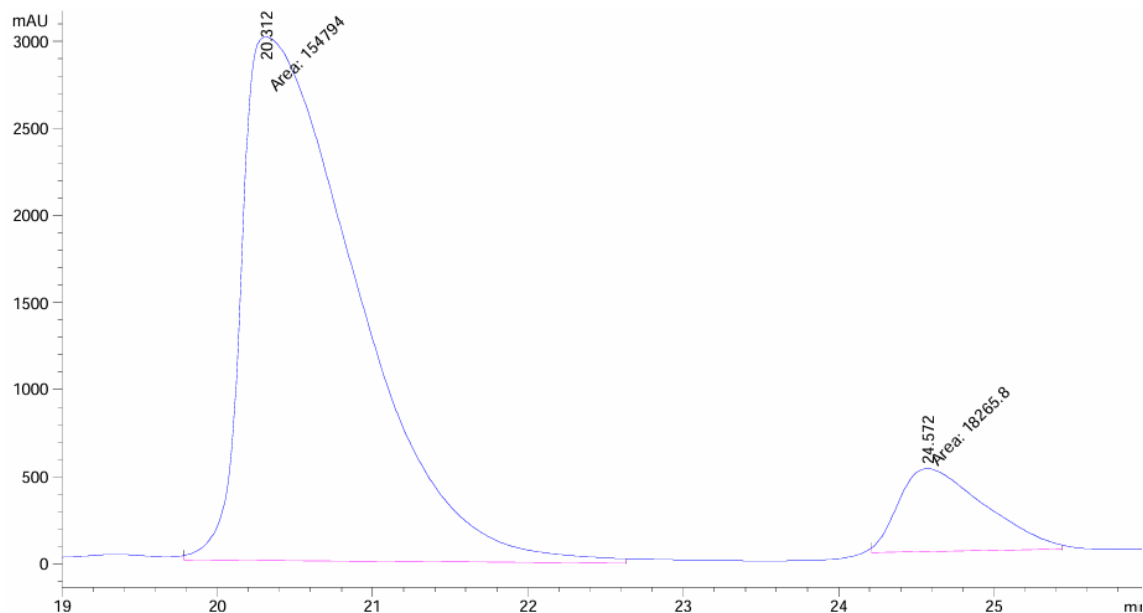
Tert-butyl (*R*)-(1-(naphthalen-2-yl)-2-nitroethyl)carbamate (4f)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.580	MM	0.5635	1.81523e4	536.86047	49.6097
2	24.288	MM	0.7011	1.84379e4	438.31787	50.3903

Totals : 3.65903e4 975.17834

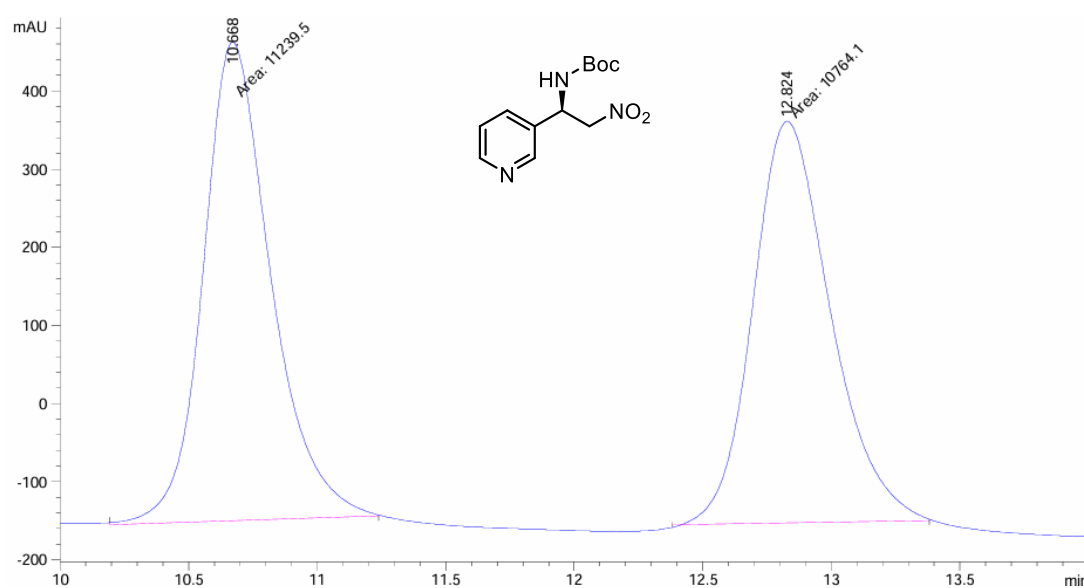


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.312	MM	0.8579	1.54794e5	3007.25537	89.4454
2	24.572	MM	0.6396	1.82658e4	475.93234	10.5546

Totals : 1.73060e5 3483.18771

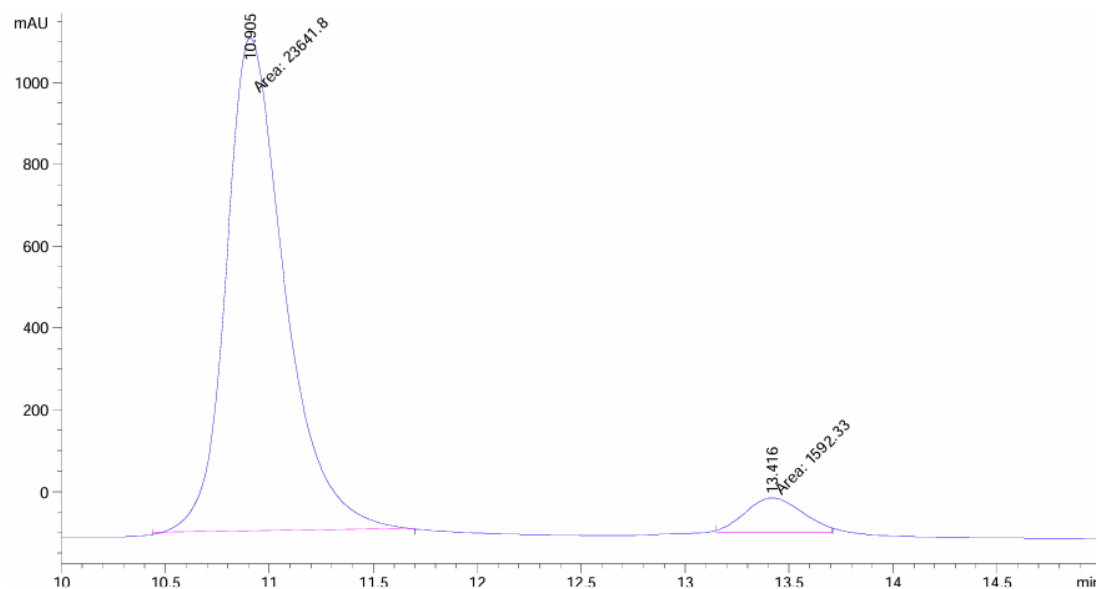
Tert-butyl (R)-(2-nitro-1-(pyridin-3-yl)ethyl)carbamate (4g)



Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.668	MM	0.3058	1.12395e4	612.61719	51.0805
2	12.824	MM	0.3492	1.07641e4	513.78003	48.9195

Totals : 2.20036e4 1126.39722

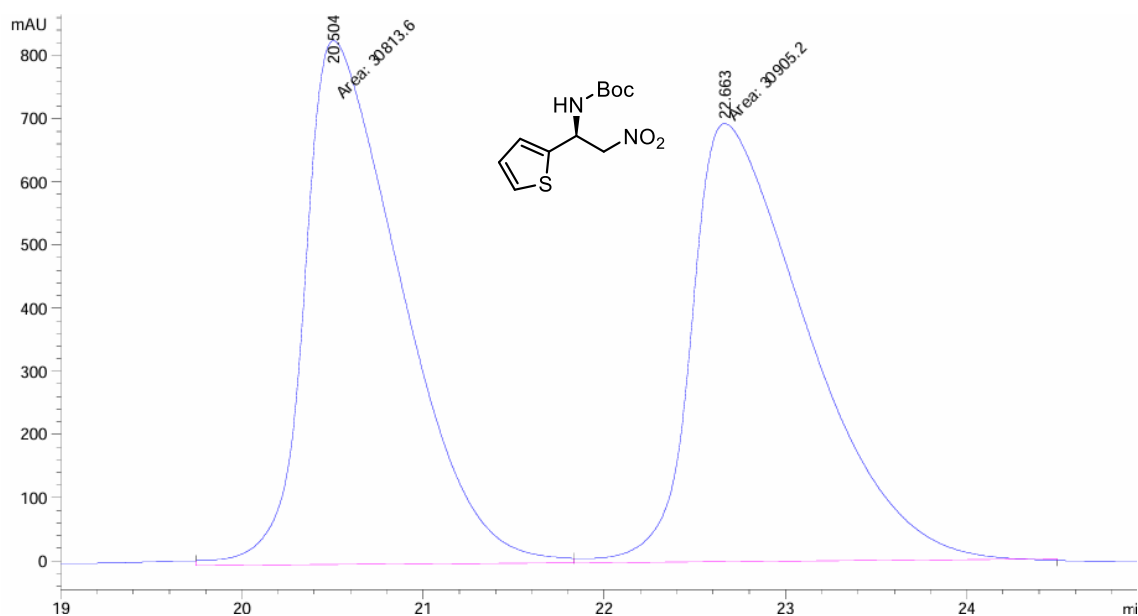


Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.905	MM	0.3271	2.36418e4	1204.67261	93.6898
2	13.416	MM	0.3169	1592.33142	83.73948	6.3102

Totals : 2.52342e4 1288.41209

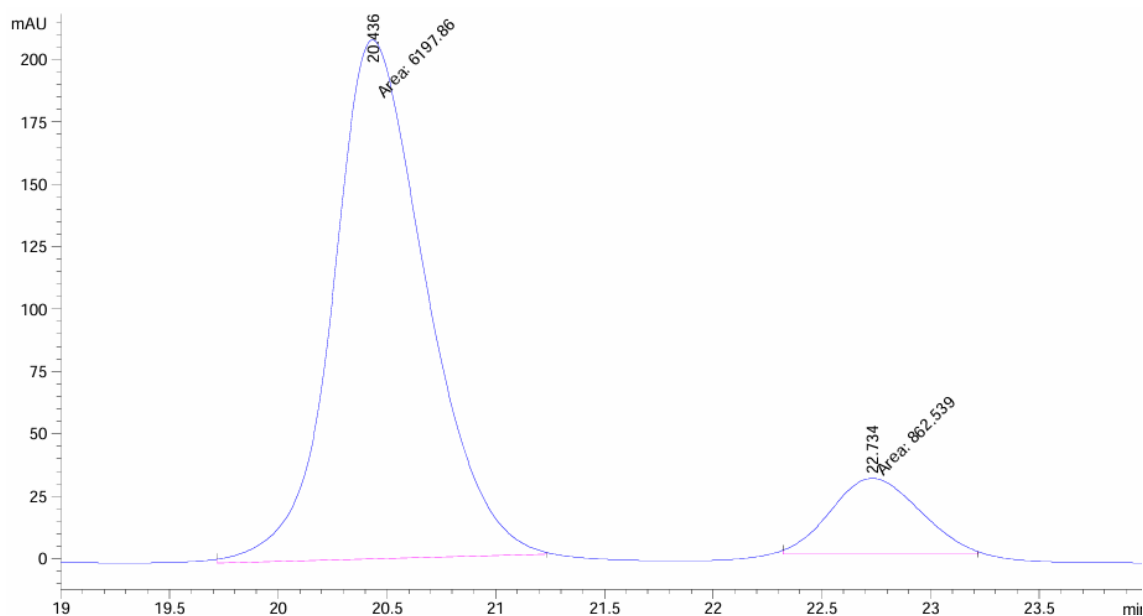
Tert-butyl (R)-(2-nitro-1-(thiophen-2-yl)ethyl)carbamate (4h)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.504	MF	0.6191	3.08136e4	829.49475	49.9258
2	22.663	FM	0.7427	3.09052e4	693.54529	50.0742

Totals : 6.17188e4 1523.04004

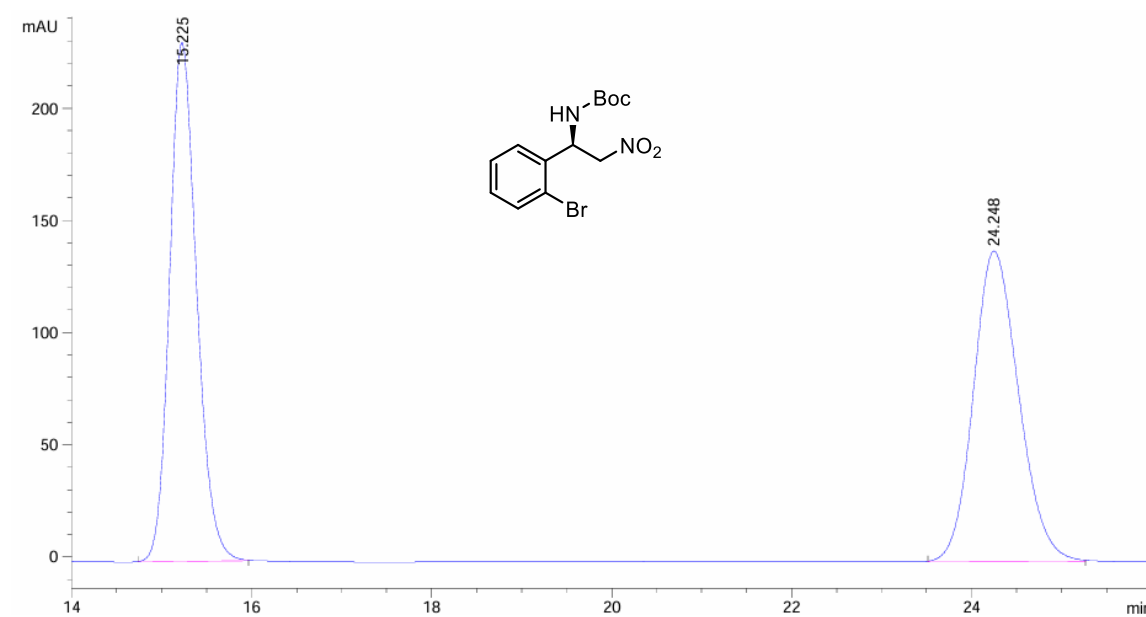


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.436	MM	0.4966	6197.85547	208.00462	87.7834
2	22.734	MM	0.4725	862.53937	30.42364	12.2166

Totals : 7060.39484 238.42826

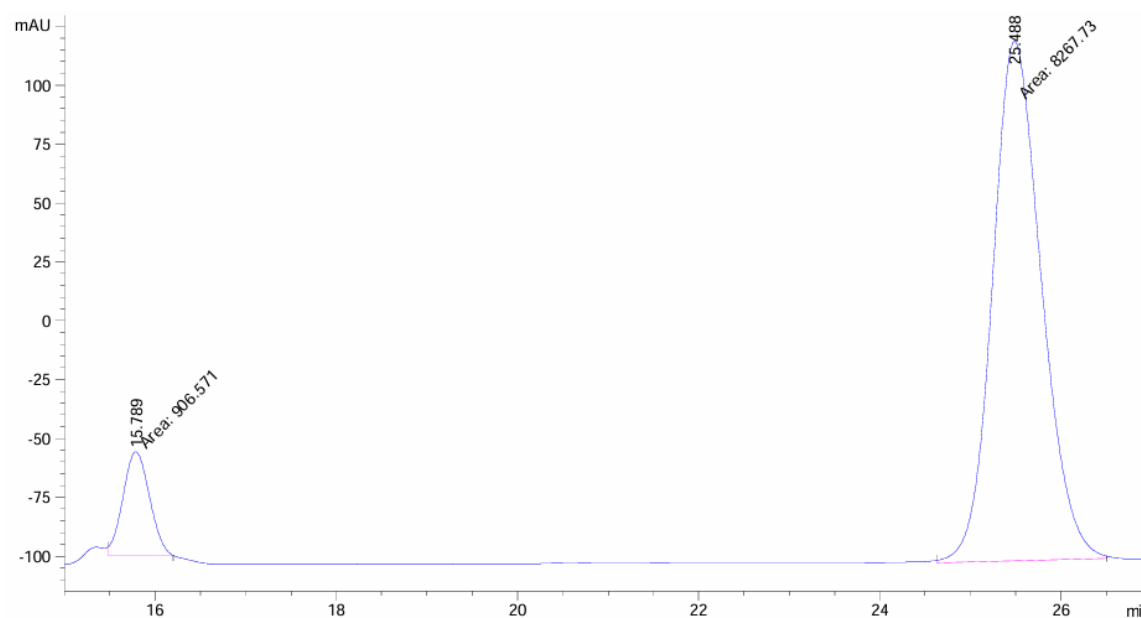
Tert-butyl (R)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4i)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.225	BB	0.3193	4821.98438	231.21657	49.9596
2	24.248	BB	0.5393	4829.78369	138.33565	50.0404

Totals : 9651.76807 369.55222

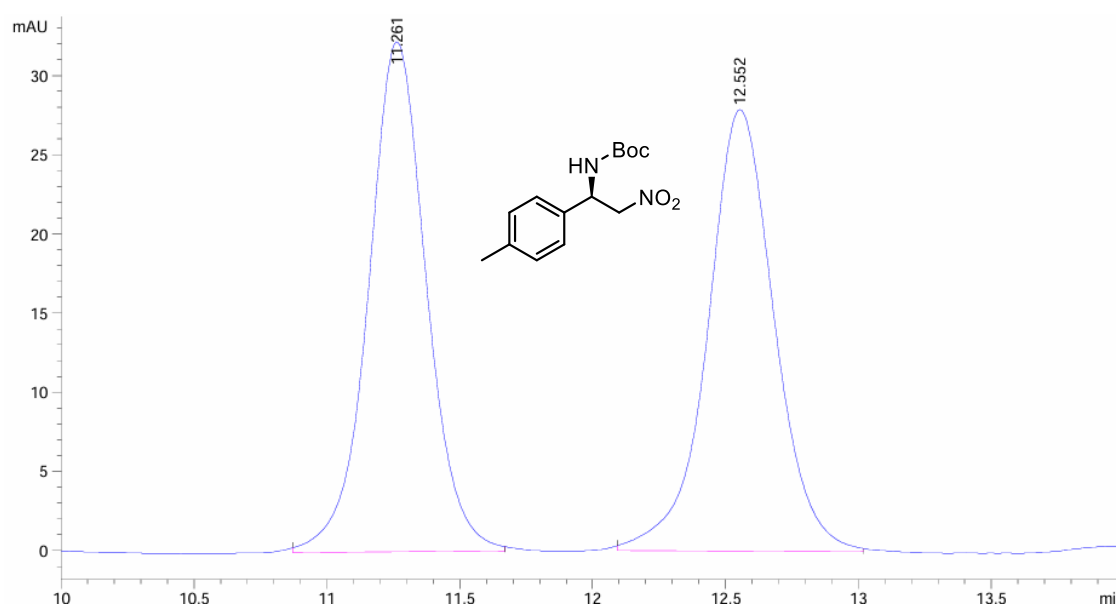


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.789	MM	0.3414	906.57147	44.26062	9.8816
2	25.488	MM	0.6248	8267.72656	220.55910	90.1184

Totals : 9174.29803 264.81972

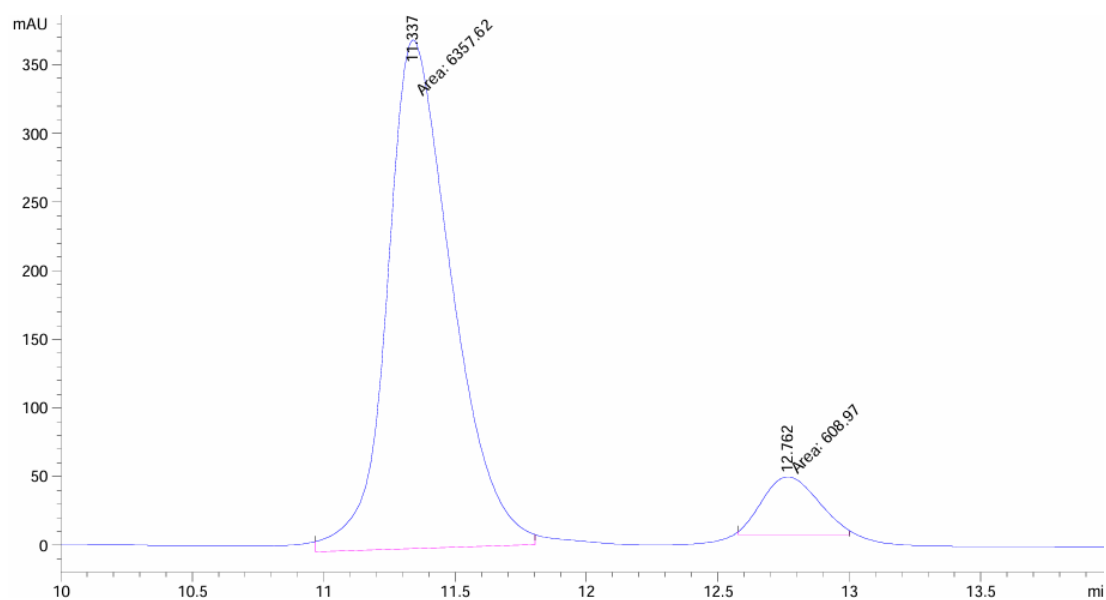
Tert-butyl (R)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4j)



Signal 1: DAD1 B, Sig=254,16 Ref=550,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.261	BB	0.2309	488.31039	32.21157	50.1146
2	12.552	BB	0.2633	486.07770	27.89753	49.8854

Totals : 974.38809 60.10910

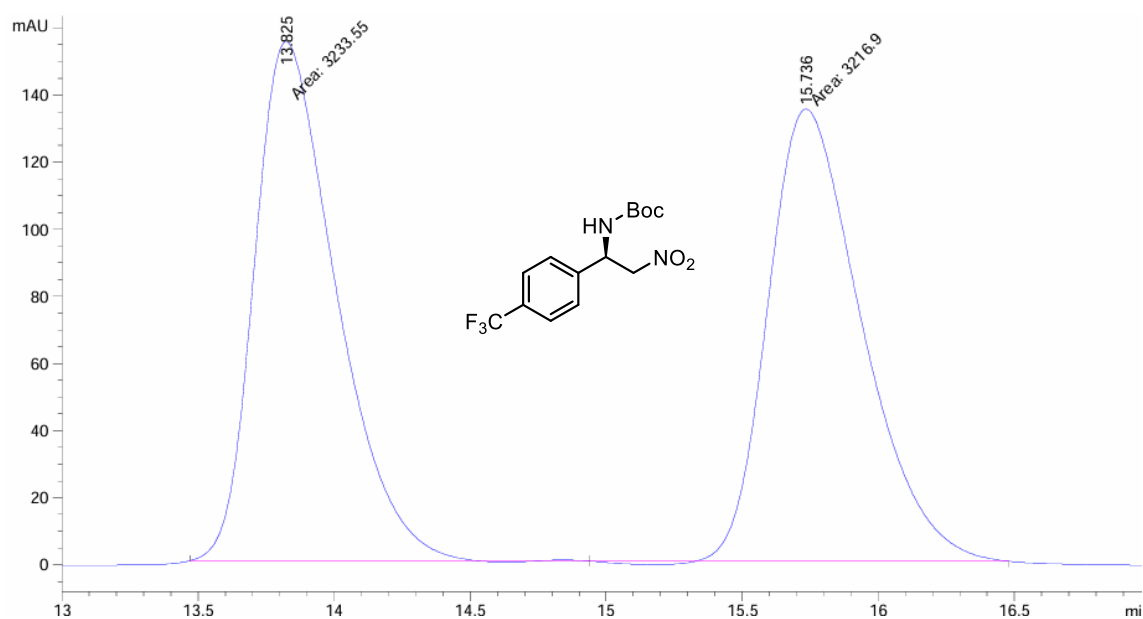


Signal 1: DAD1 A, Sig=254,4 Ref=550,99.7

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.337	MM	0.2860	6357.62061	370.54602	91.2587
2	12.762	MM	0.2421	608.96997	41.92290	8.7413

Totals : 6966.59058 412.46892

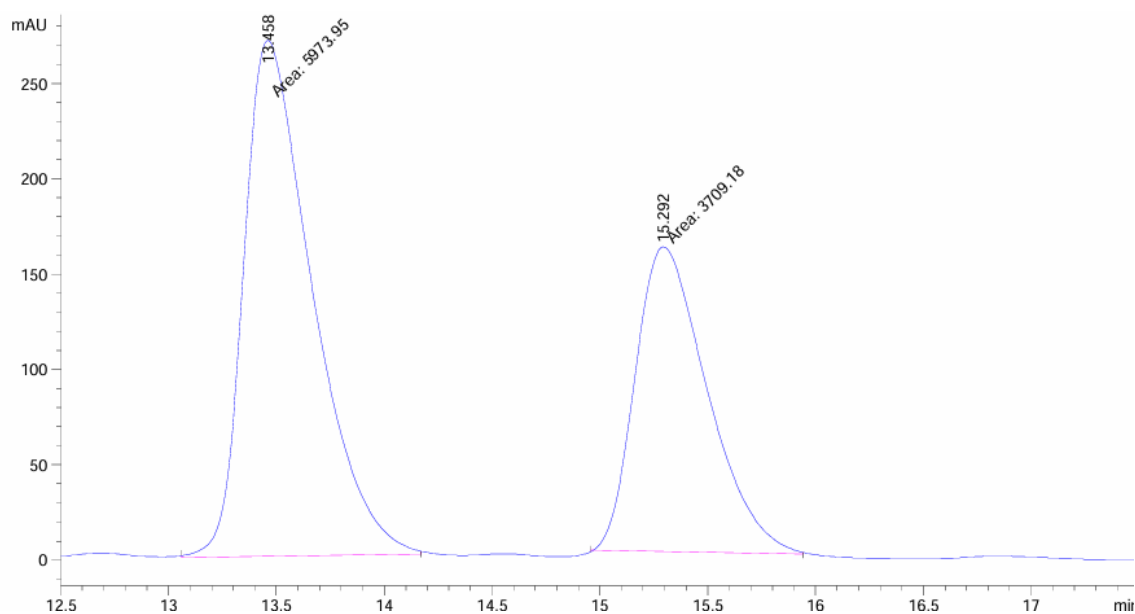
Tert-butyl (*R*)-(2-nitro-1-(4-(trifluoromethyl)phenyl)ethyl)carbamate (4k)



Signal 1: DAD1 B, Sig=254,16 Ref=550,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.825	MF	0.3483	3233.54590	154.74586	50.1290
2	15.736	FM	0.3982	3216.89966	134.63853	49.8710

Totals : 6450.44556 289.38440

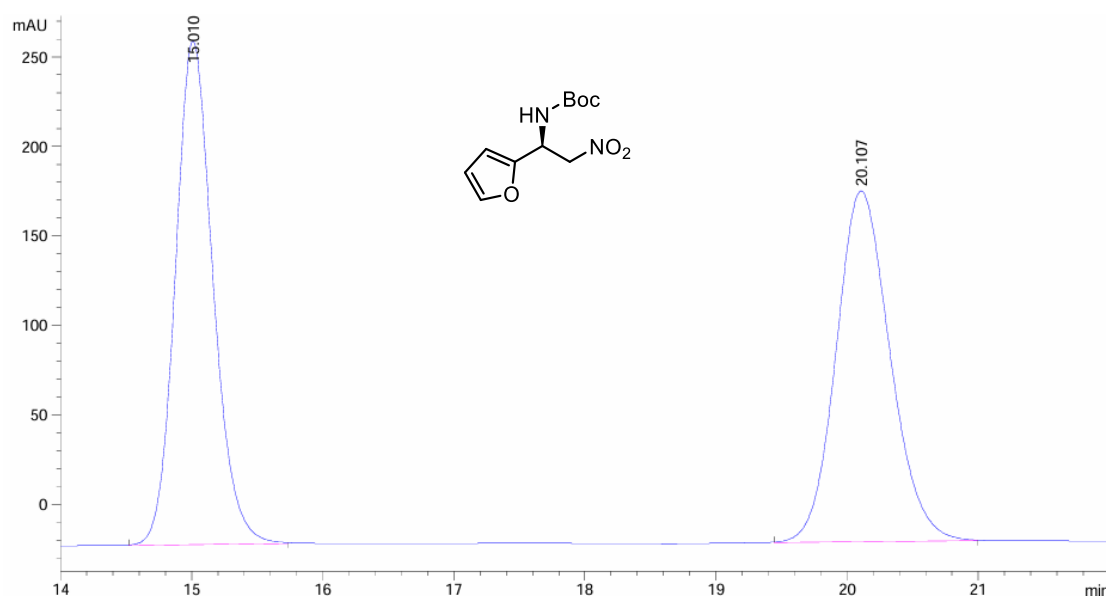


Signal 1: DAD1 B, Sig=254,16 Ref=550,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.458	MM	0.3682	5973.95068	270.43240	61.6944
2	15.292	MM	0.3871	3709.17822	159.69753	38.3056

Totals : 9683.12891 430.12993

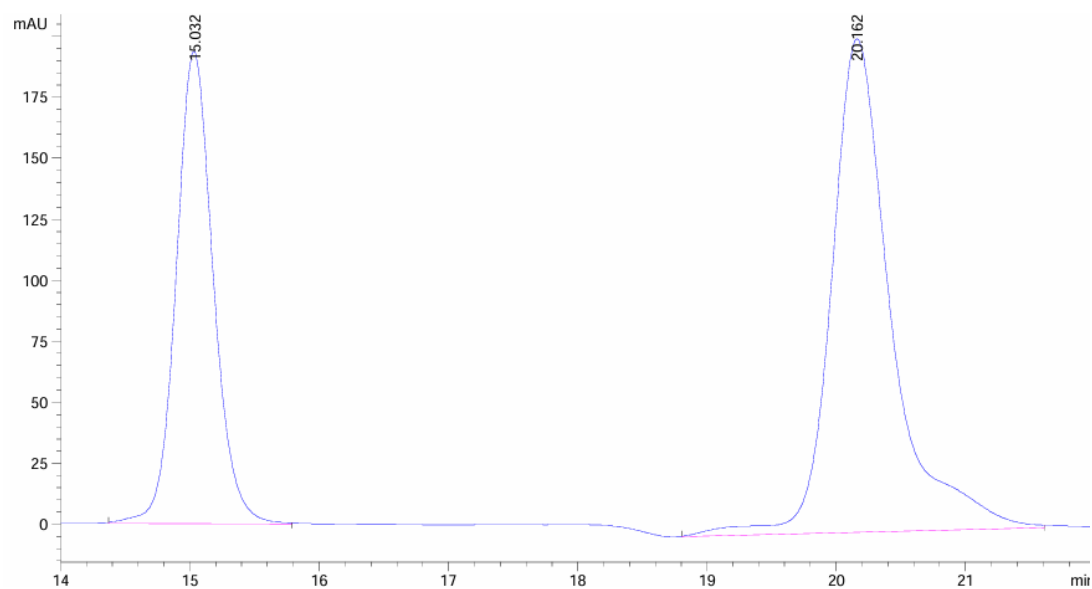
Tert-butyl (R)-(1-(furan-2-yl)-2-nitroethyl)carbamate (4I)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.010	BB	0.3047	5614.80566	281.54291	50.0372
2	20.107	BB	0.4415	5606.45557	196.10039	49.9628

Totals : 1.12213e4 477.64330



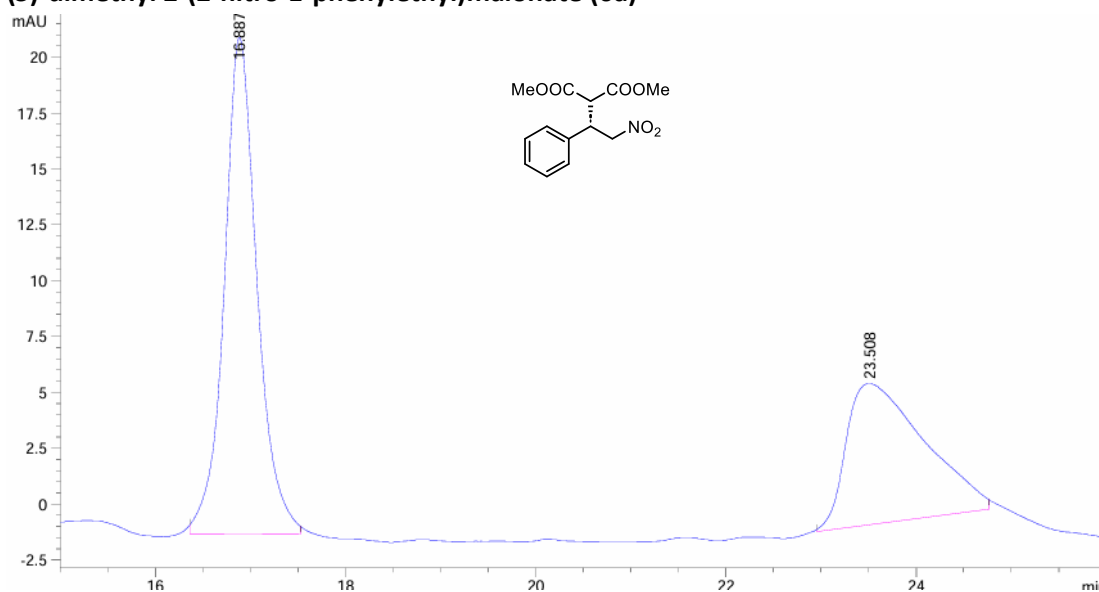
Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.032	BB	0.3055	3875.34839	193.62755	37.3029
2	20.162	BB	0.4778	6513.52832	202.19289	62.6971

Totals : 1.03889e4 395.82043

Chiral HPLC chromatograms of 6

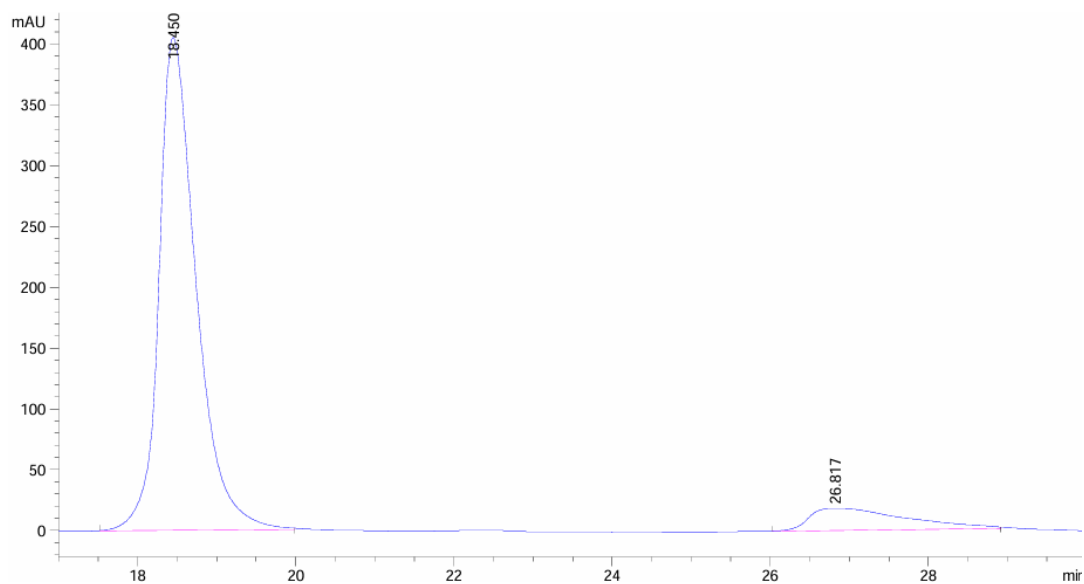
(S)-dimethyl 2-(2-nitro-1-phenylethyl)malonate (6a)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.887	BB	0.3607	535.05011	22.23783	49.5488
2	23.503	MM	1.2590	544.79376	7.21205	50.4512

Totals : 1079.84387 29.44987

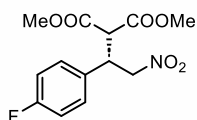
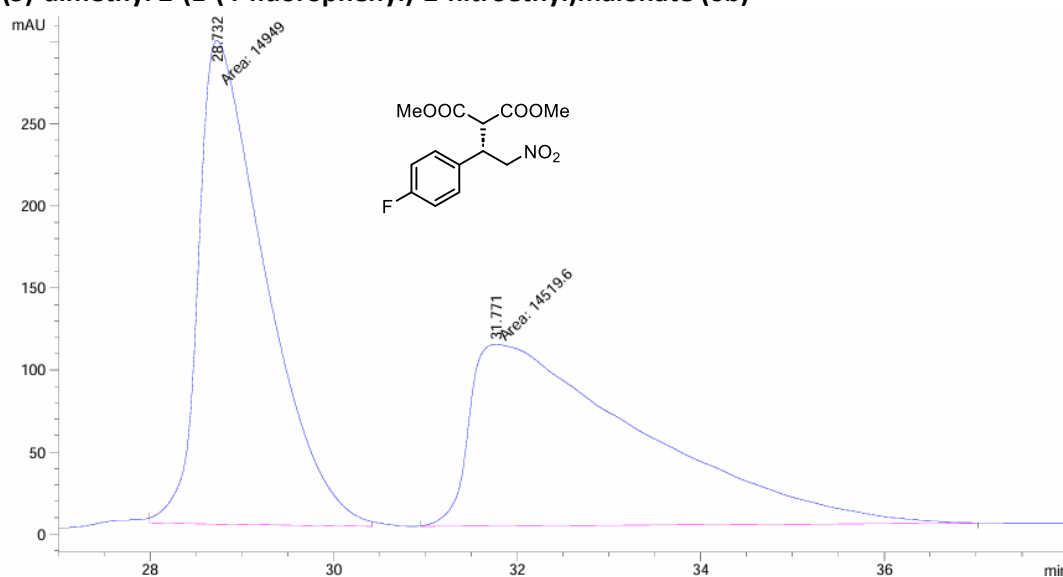


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.450	BB	0.4942	1.32816e4	405.38852	89.6975
2	26.817	BB	1.2262	1525.50781	18.23069	10.3025

Totals : 1.48071e4 423.61921

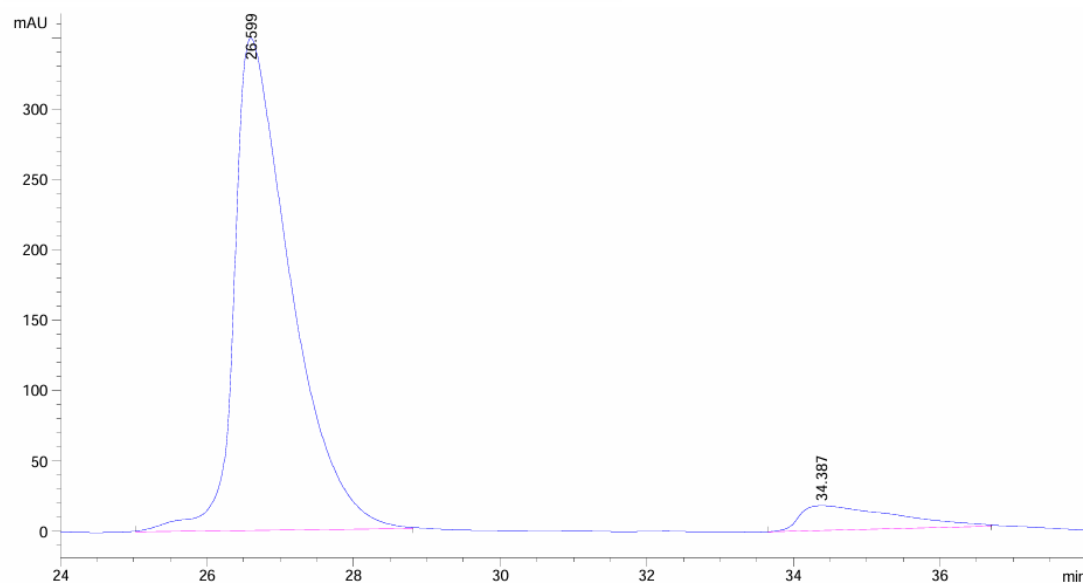
(S)-dimethyl 2-(1-(4-fluorophenyl)-2-nitroethyl)malonate (6b)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	28.732	MM	0.8452	1.49490e4	294.78055	50.7286
2	31.771	MM	2.1865	1.45196e4	110.67889	49.2714

Totals : 2.94687e4 405.45944

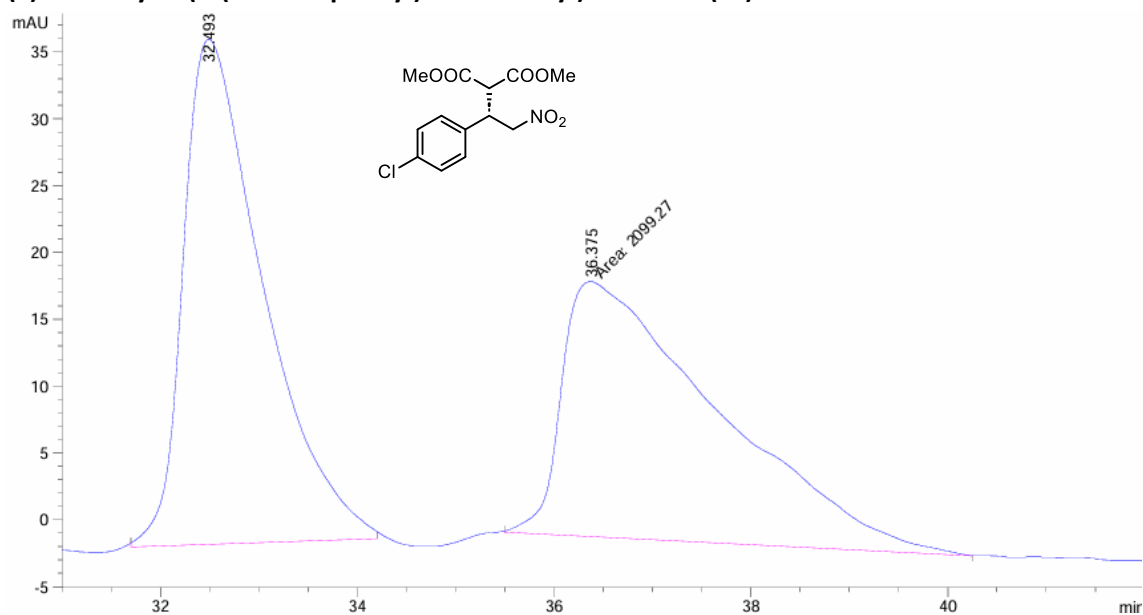


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	26.599	BB	0.8085	1.87842e4	349.78241	92.2168
2	34.387	BB	1.2584	1585.40247	17.60824	7.7832

Totals : 2.03696e4 367.39065

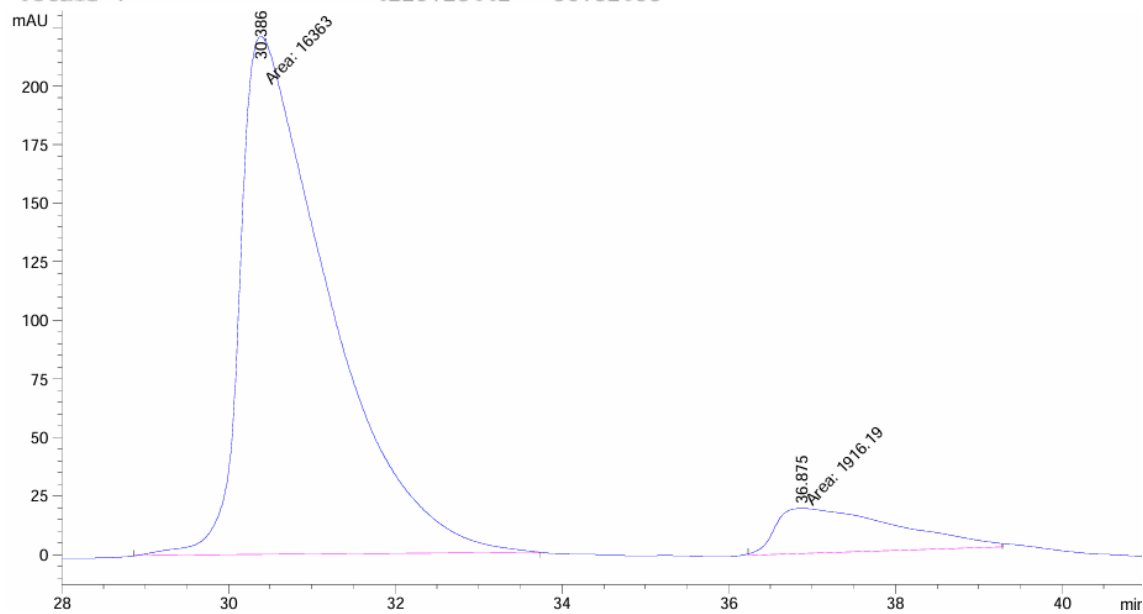
(S)-dimethyl 2-(1-(4-chlorophenyl)-2-nitroethyl)malonate (6c)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	32.493	BB	0.8260	2130.01782	37.76342	50.3636
2	36.375	MM	1.8358	2099.26660	19.05853	49.6364

Totals : 4229.28442 56.82195

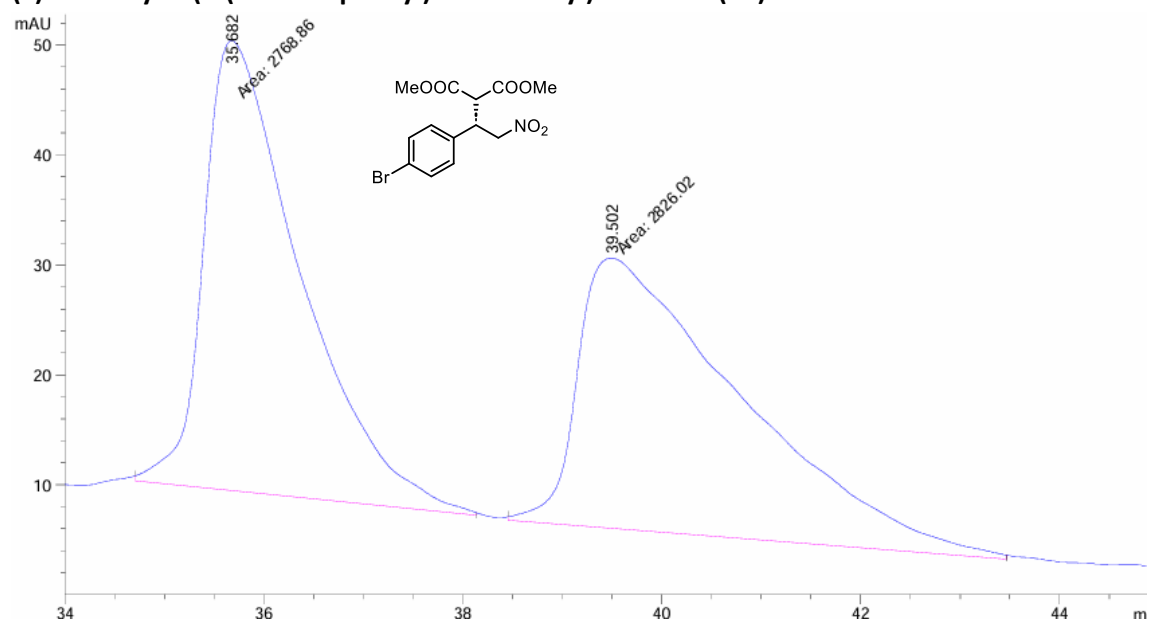


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	30.386	MM	1.2335	1.63630e4	221.08369	89.5171
2	36.875	MM	1.6546	1916.18994	19.30120	10.4829

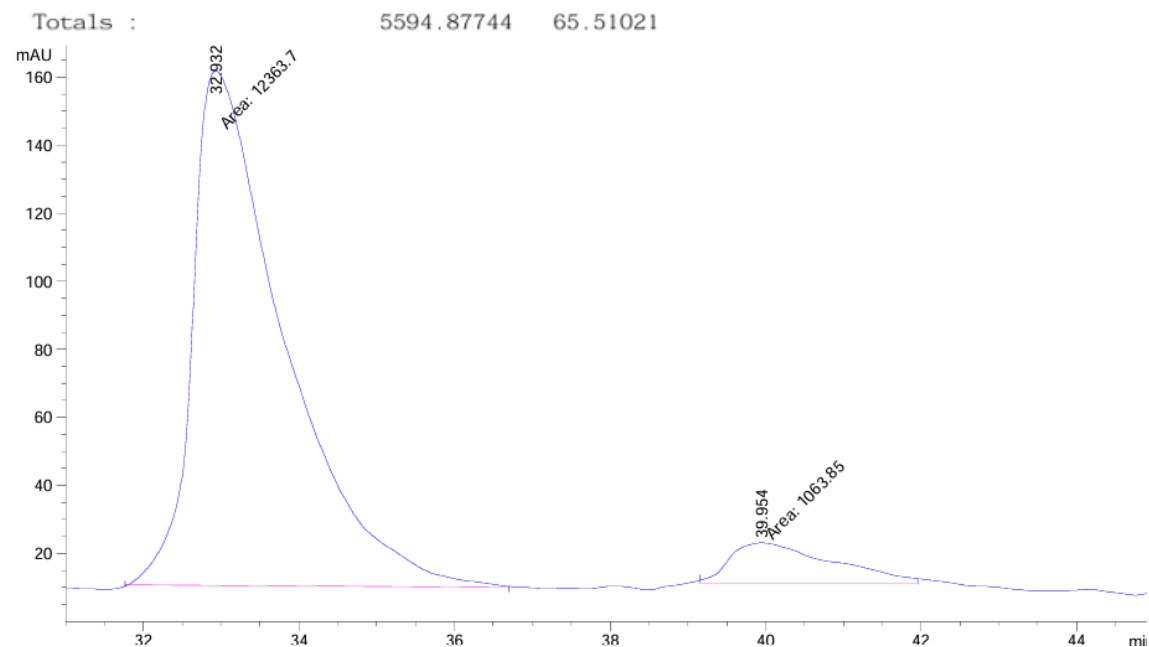
Totals : 1.82792e4 240.38489

(S)-dimethyl 2-(1-(4-bromophenyl)-2-nitroethyl)malonate (6d)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	35.682	MM	1.1275	2768.85547	40.93092	49.4891
2	39.502	MM	1.9163	2826.02197	24.57929	50.5109

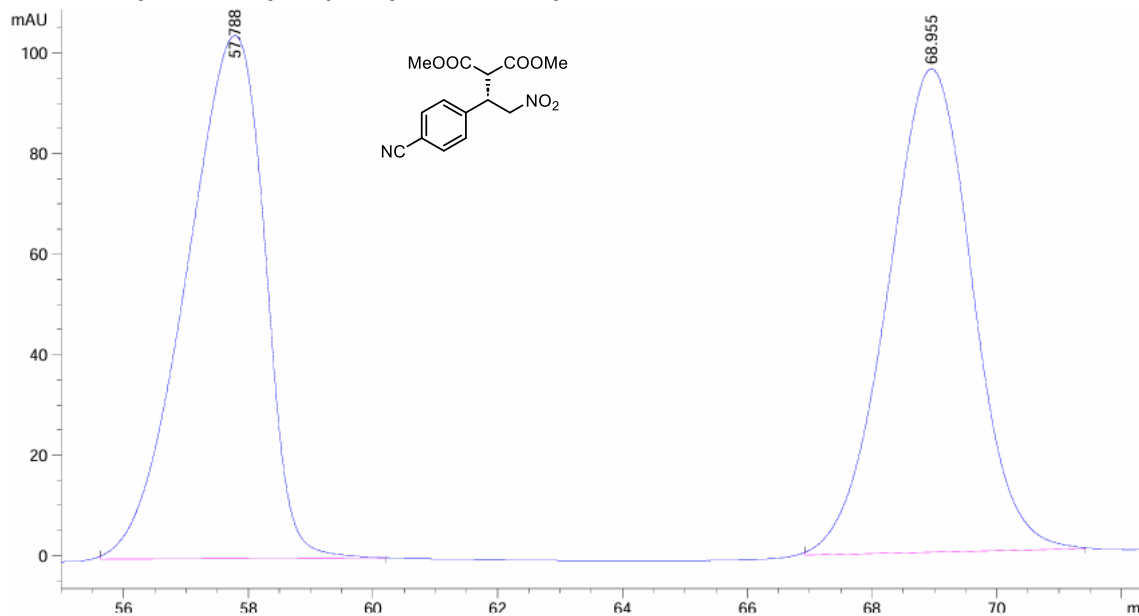


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	32.932	MM	1.3638	1.23637e4	151.09863	92.0771
2	39.954	MM	1.5060	1063.84729	11.77319	7.9229

Totals : 1.34276e4 162.87182

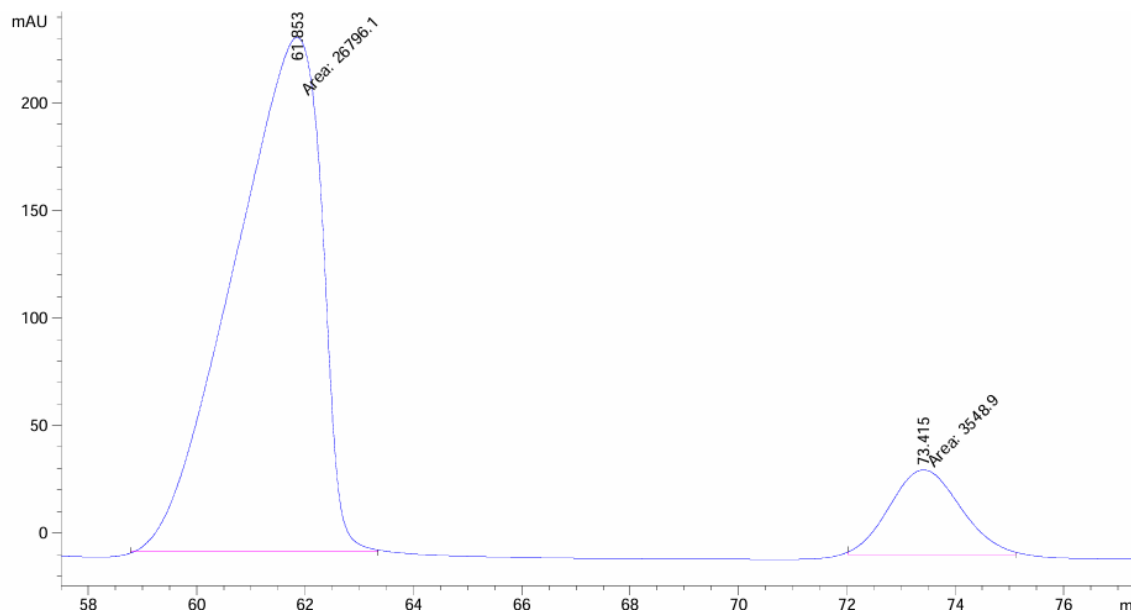
(S)-dimethyl 2-(1-(4-cyanophenyl)-2-nitroethyl)malonate (6e)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	57.788	BB	1.3769	9072.02930	104.07824	49.9349
2	68.955	BB	1.4512	9095.68652	96.18860	50.0651

Totals : 1.81677e4 200.26684

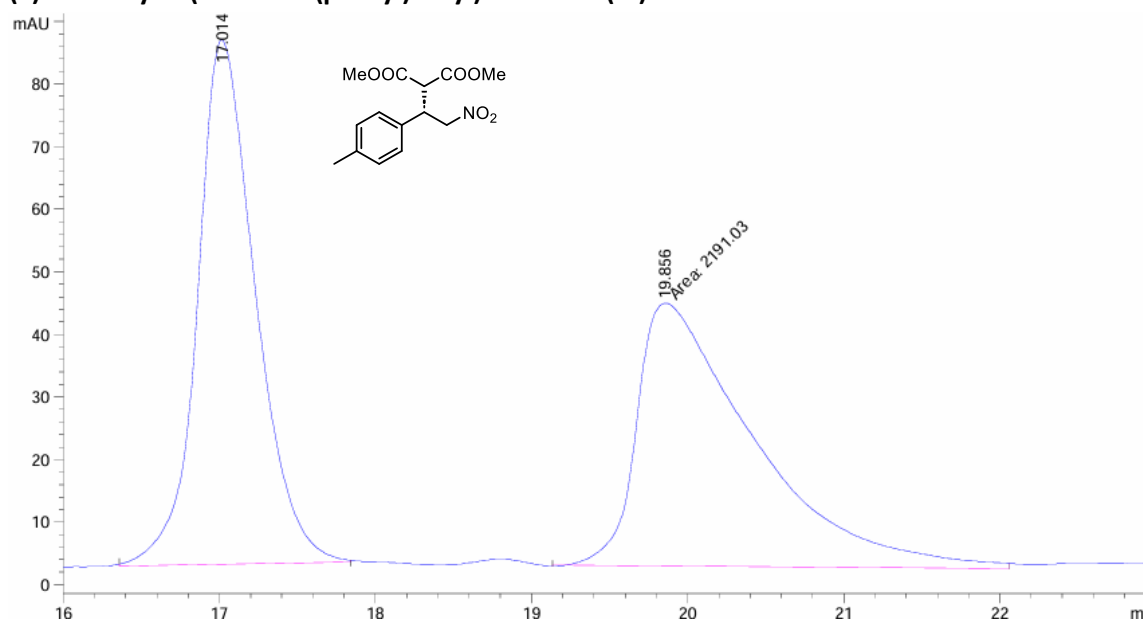


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	61.853	MM	1.8680	2.67961e4	239.08089	88.3048
2	73.415	MM	1.5085	3548.89819	39.20918	11.6952

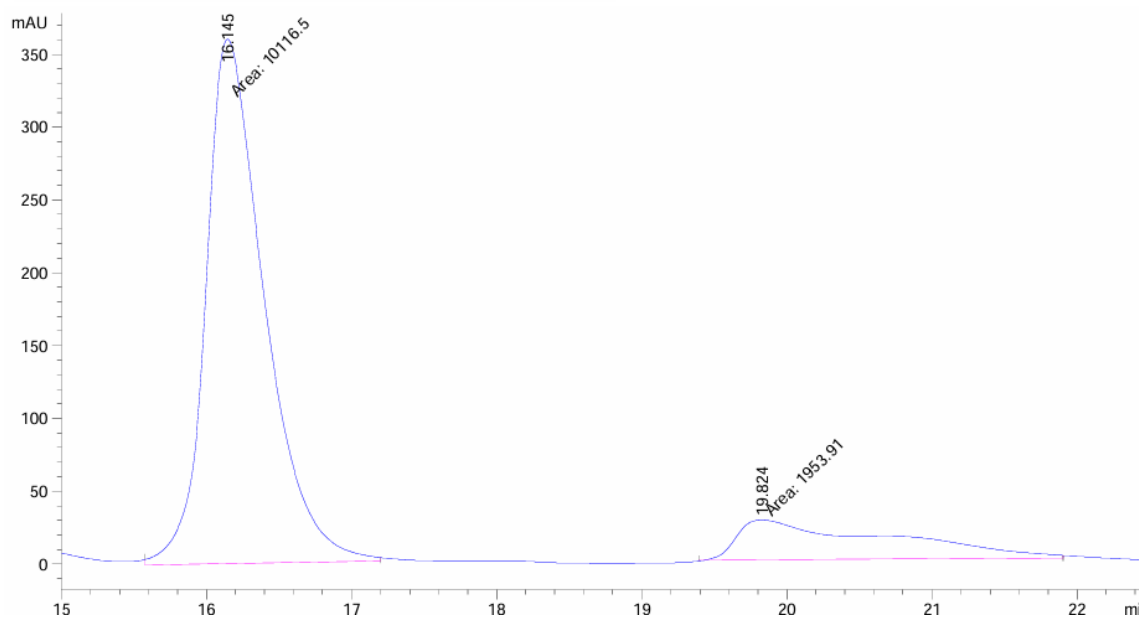
Totals : 3.03450e4 278.29007

(S)-dimethyl 2-(2-nitro-1-(p-tolyl)ethyl)malonate (6f)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.014	BB	0.3871	2163.00879	83.81332	49.6782
2	19.856	MM	0.8702	2191.03101	41.96473	50.3218

Totals : 4354.03979 125.77804

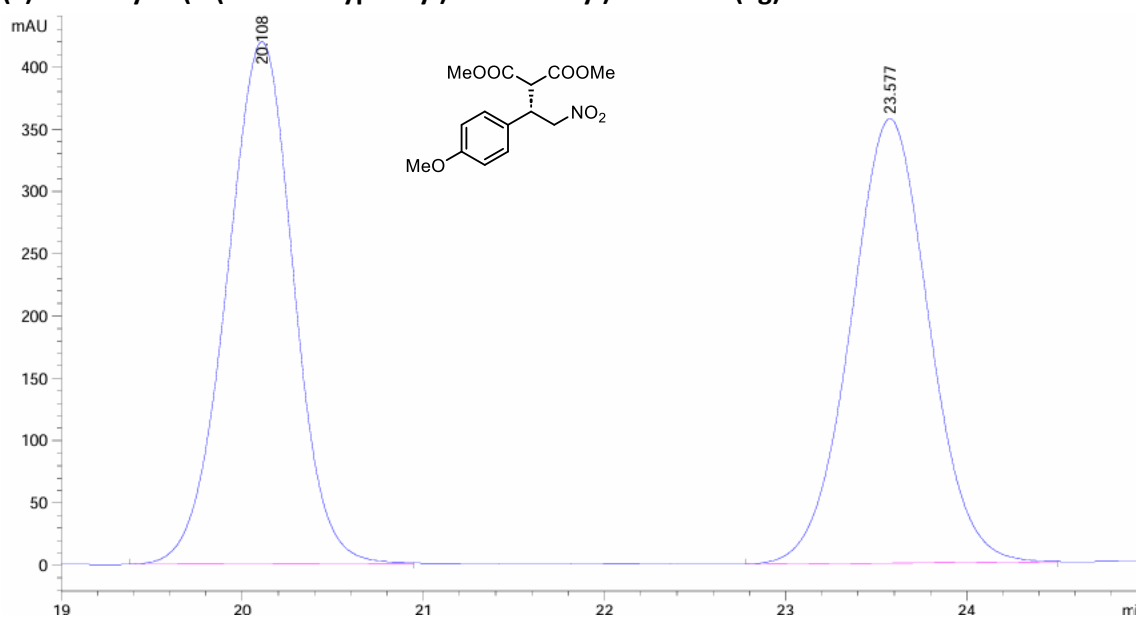


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.145	MM	0.4683	1.01165e4	360.00928	83.8124
2	19.824	MM	1.1887	1953.91248	27.39496	16.1876

Totals : 1.20705e4 387.40424

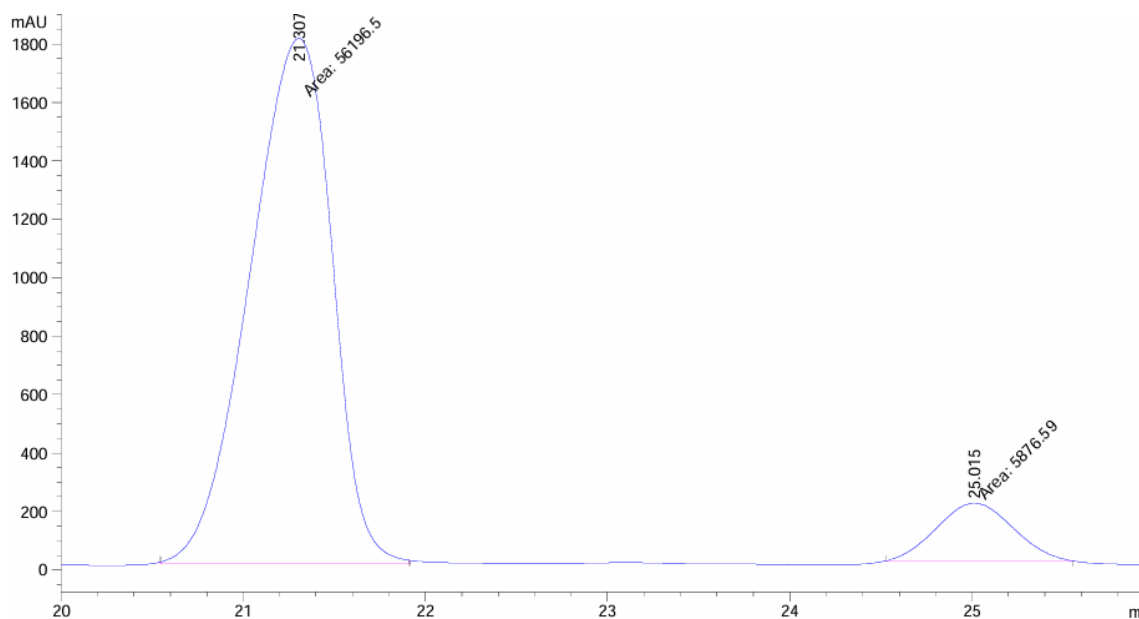
(S)-dimethyl 2-(1-(4-methoxyphenyl)-2-nitroethyl)malonate (6g)



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.108	BB	0.3997	1.07658e4	419.02090	50.1148
2	23.577	BB	0.4646	1.07165e4	356.56686	49.8852

Totals : 2.14822e4 775.58777

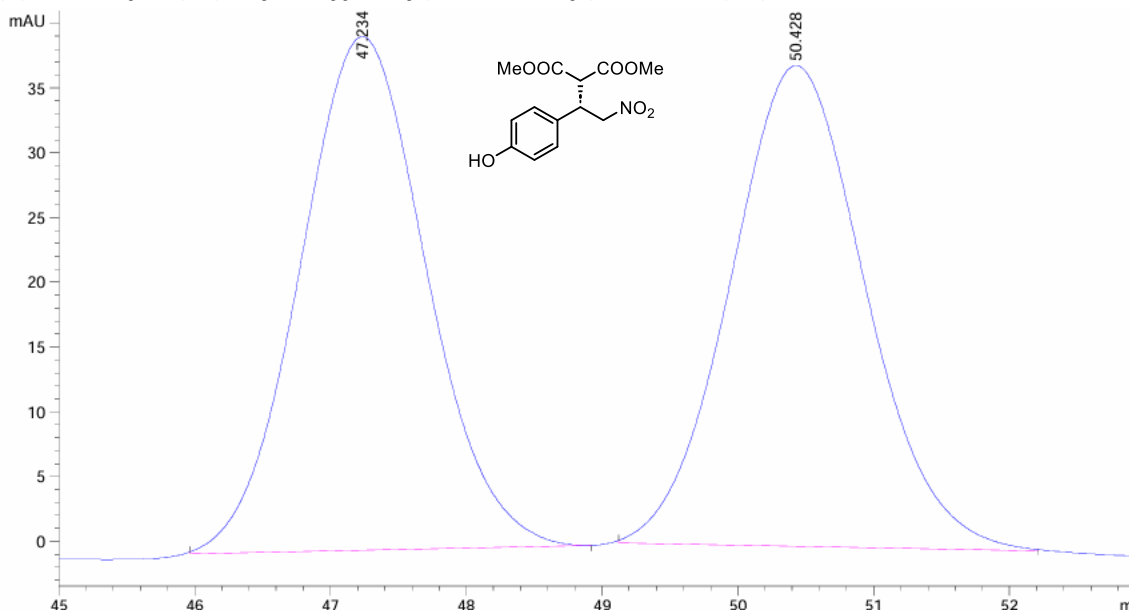


Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.307	MM	0.5208	5.61965e4	1798.35657	90.5328
2	25.015	MM	0.4904	5876.58643	199.72989	9.4672

Totals : 6.20731e4 1998.08646

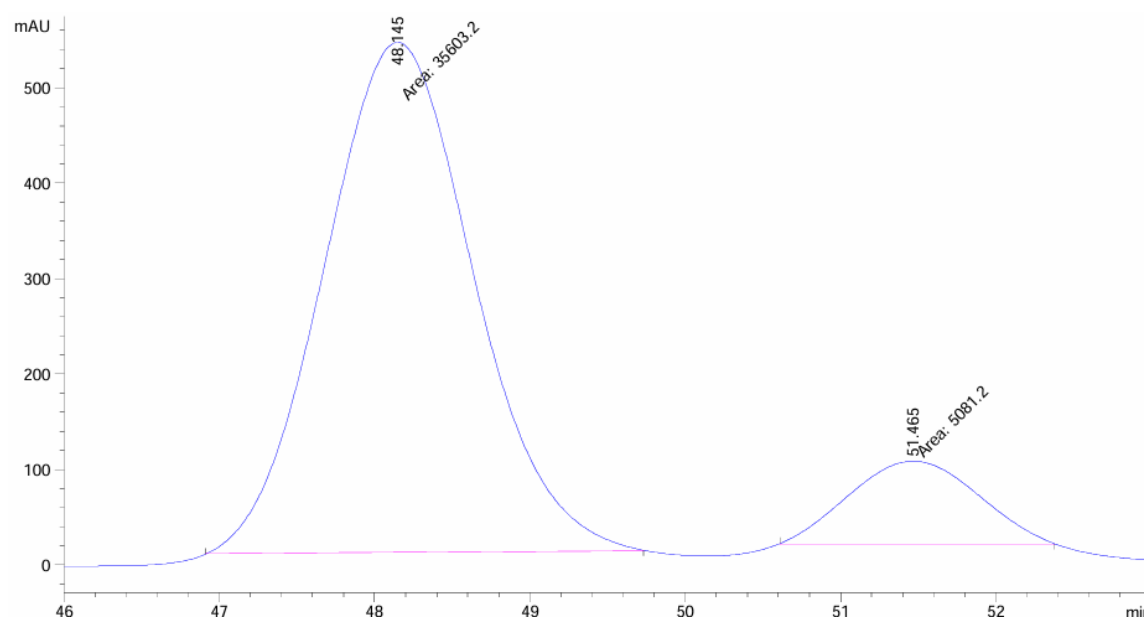
(S)-dimethyl 2-(1-(4-hydroxyphenyl)-2-nitroethyl)malonate (6h)



Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	47.234	BB	1.0009	2560.21094	39.66983	50.0256
2	50.428	BB	1.0638	2557.59497	37.12057	49.9744

Totals : 5117.80591 76.79041



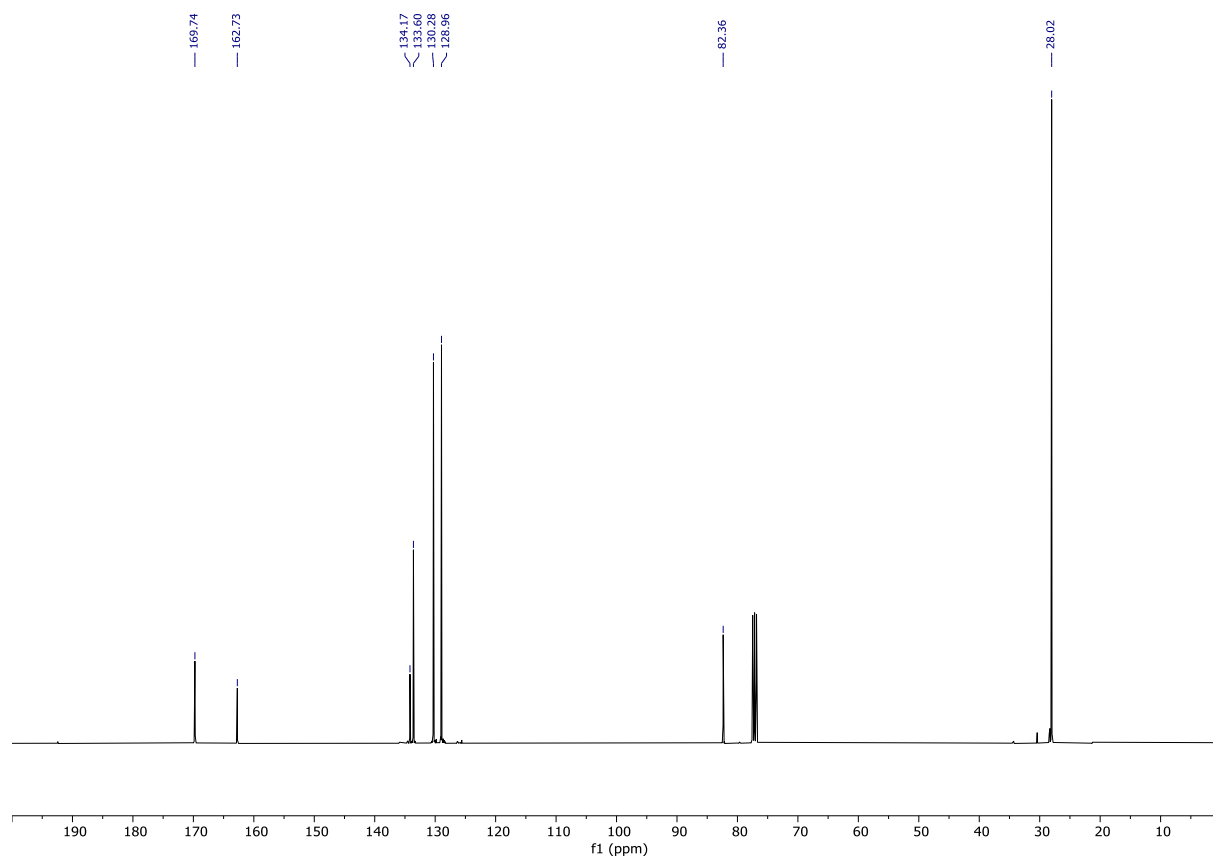
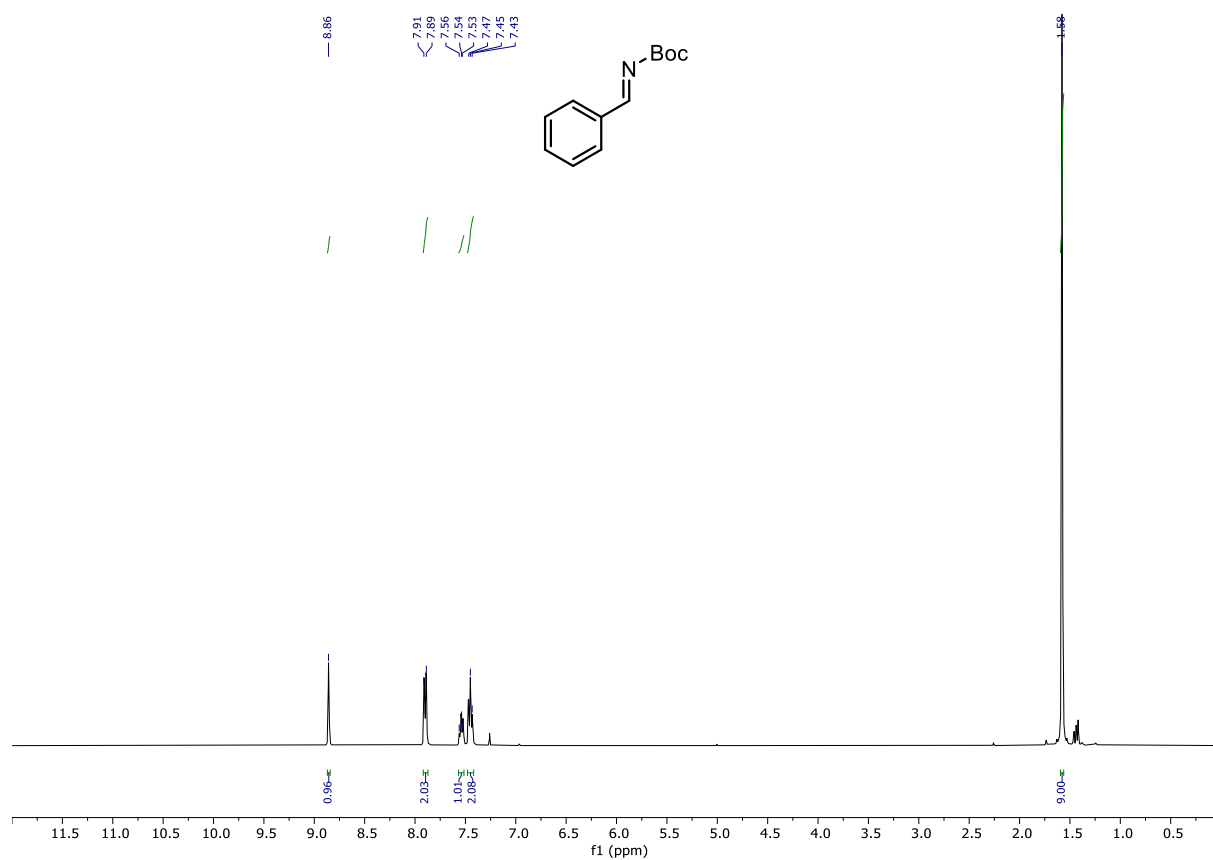
Signal 1: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	48.145	MM	1.1110	3.56032e4	534.12360	87.5107
2	51.465	MM	0.9644	5081.19775	87.80897	12.4893

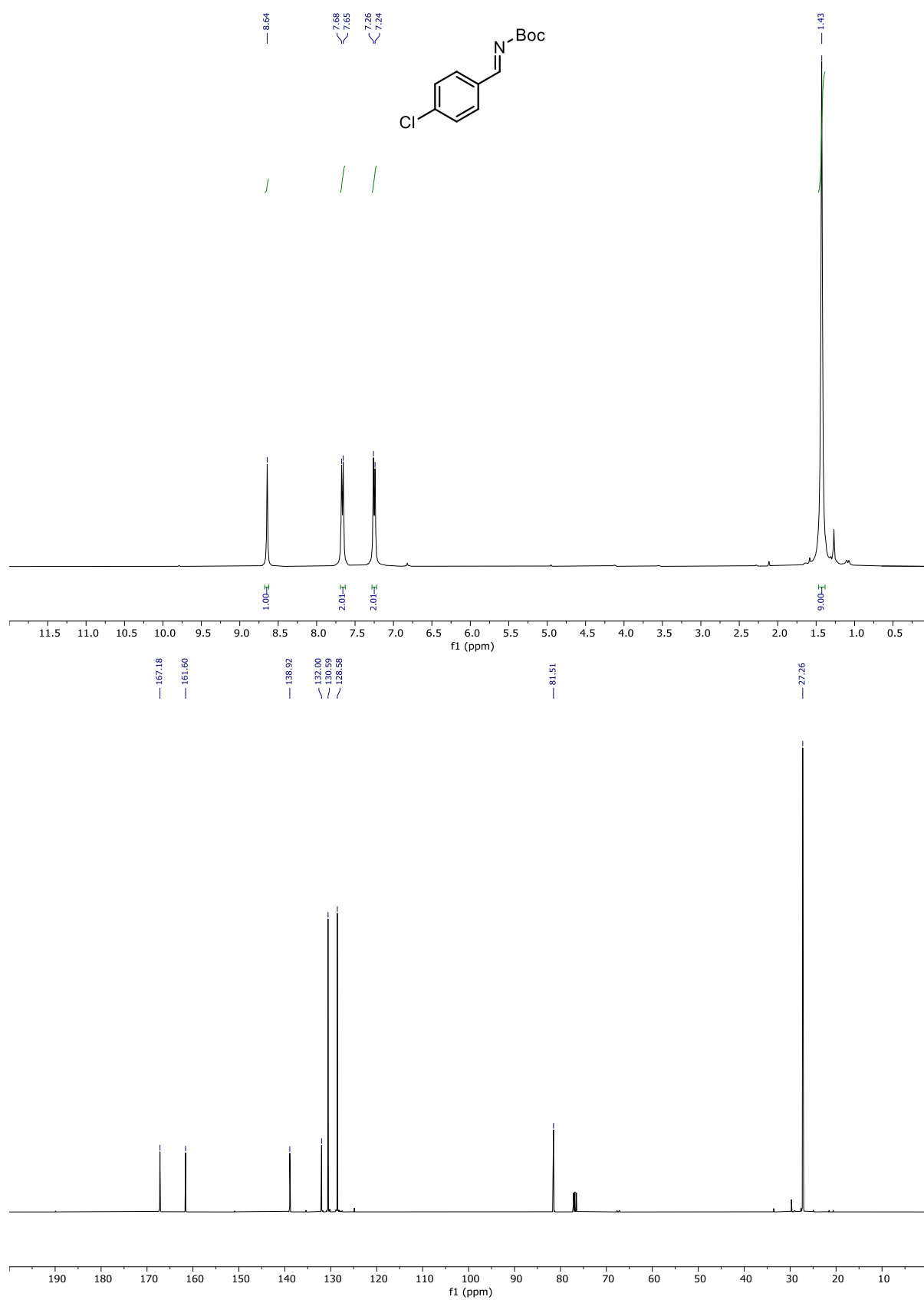
Totals : 4.06844e4 621.93256

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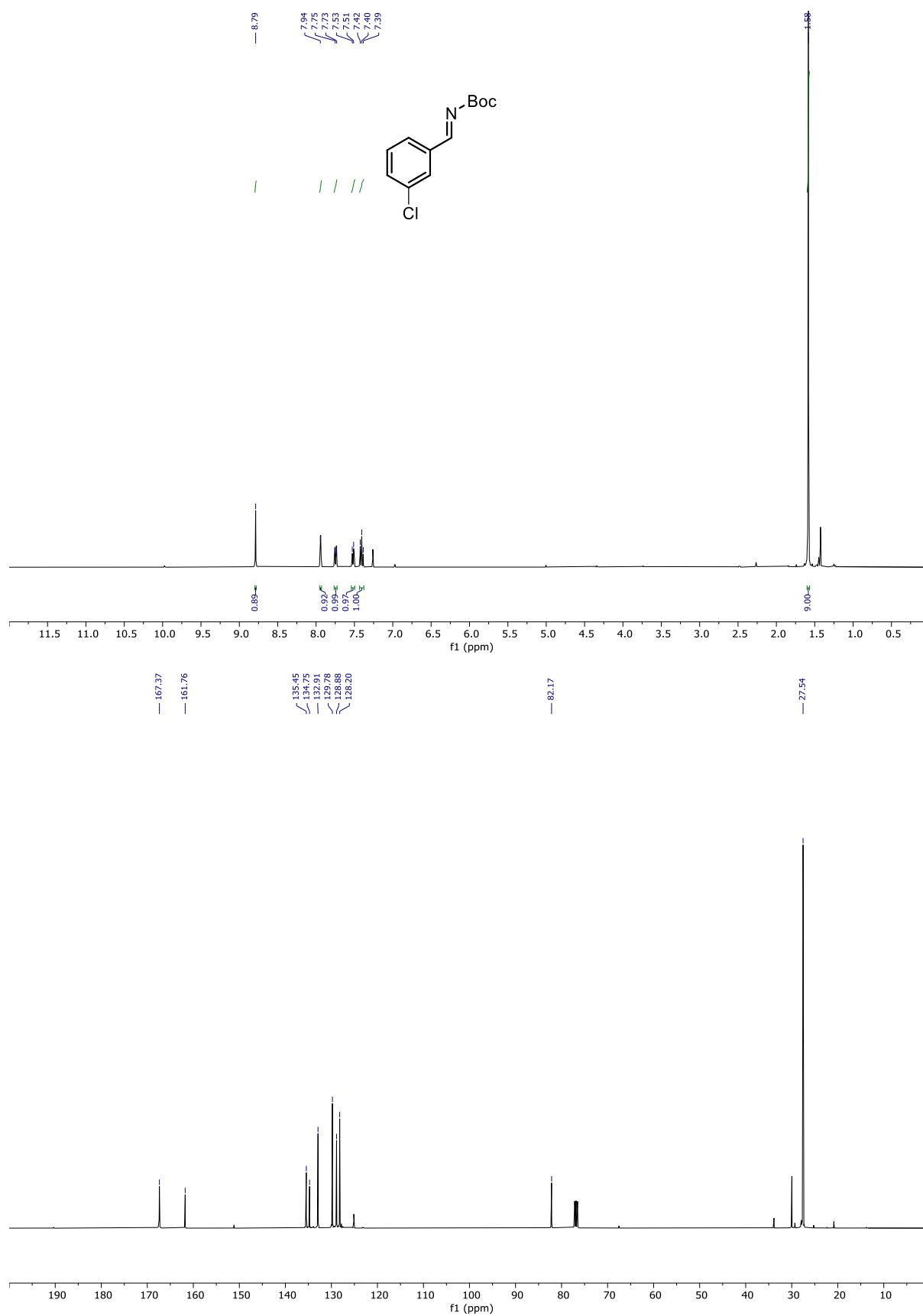
Tert-butyl (*E*)-benzylidenecarbamate (3a) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



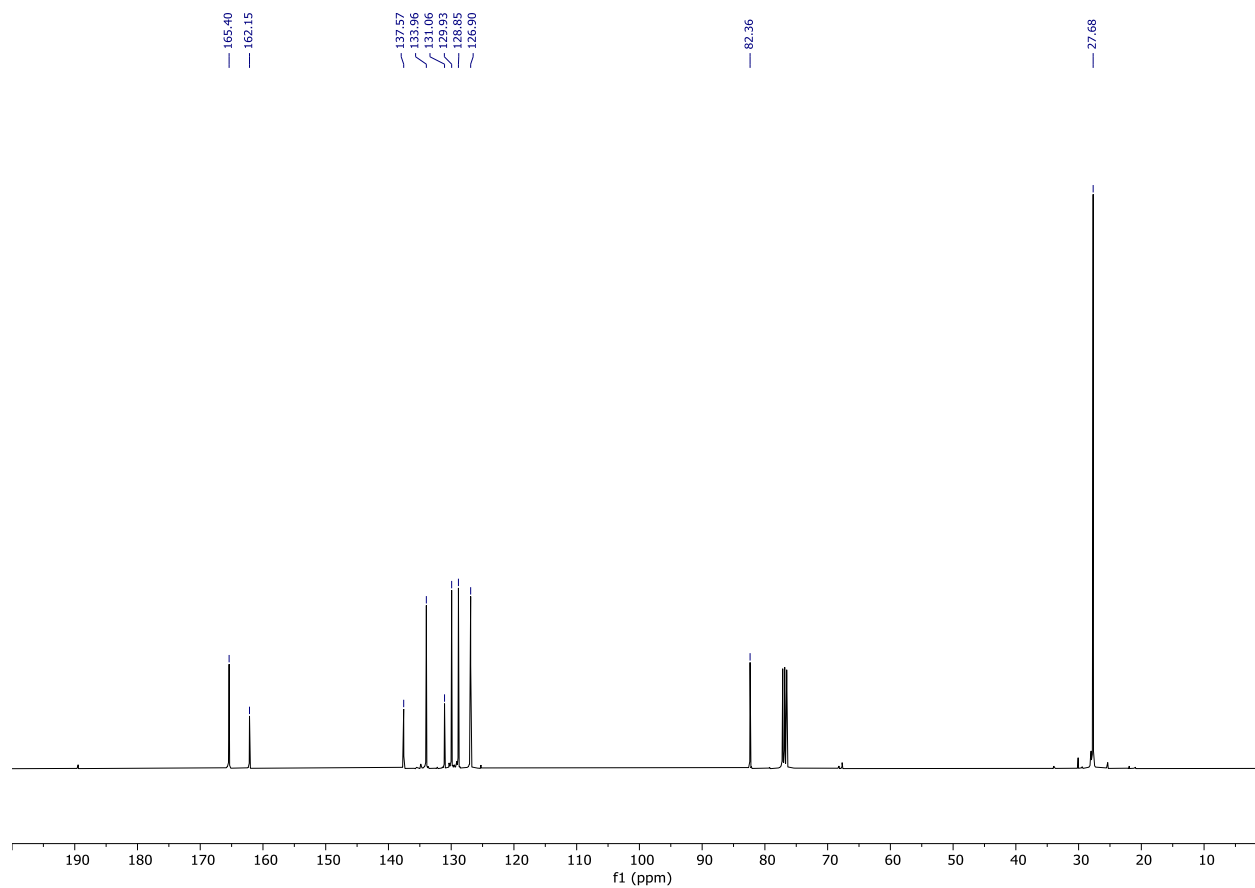
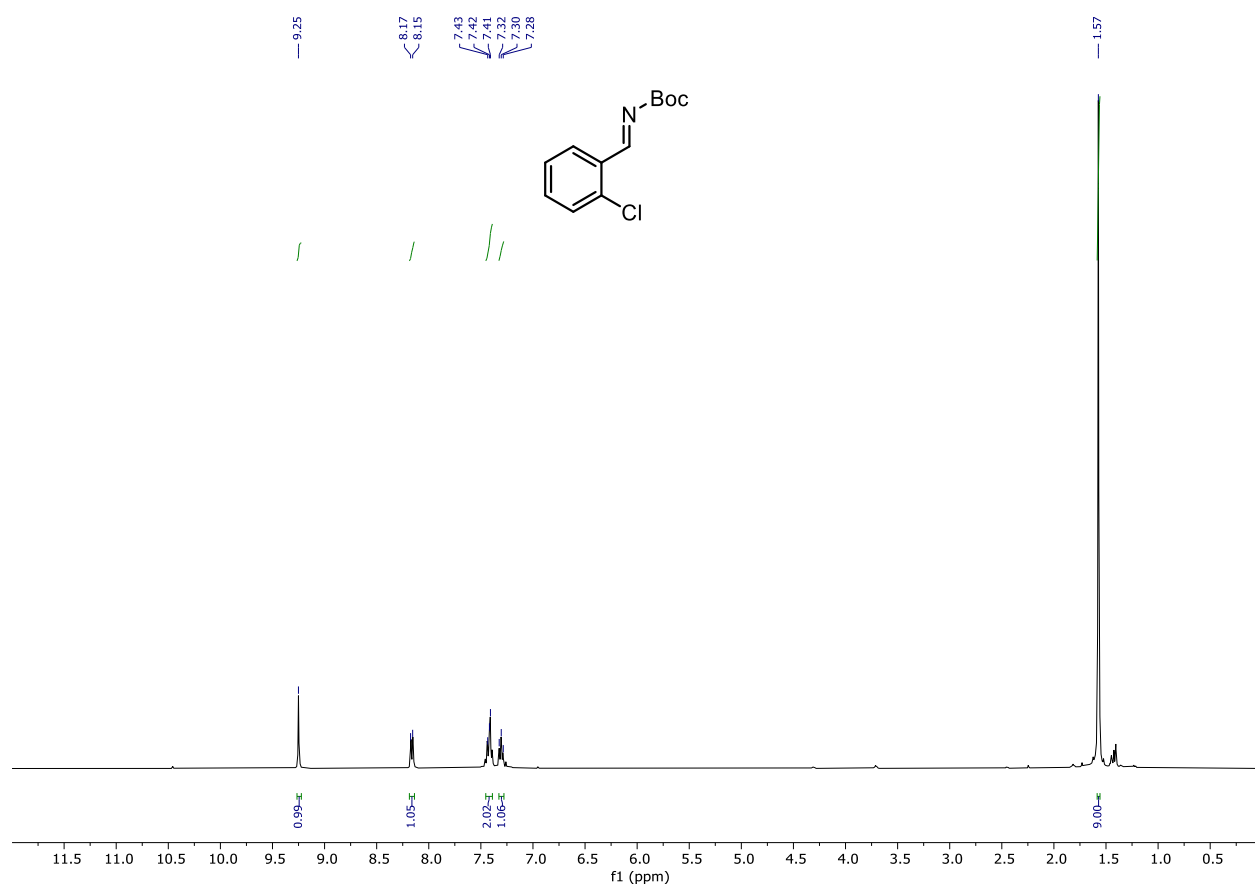
Tert-butyl (*E*)-(4-chlorobenzylidene)carbamate (3b) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



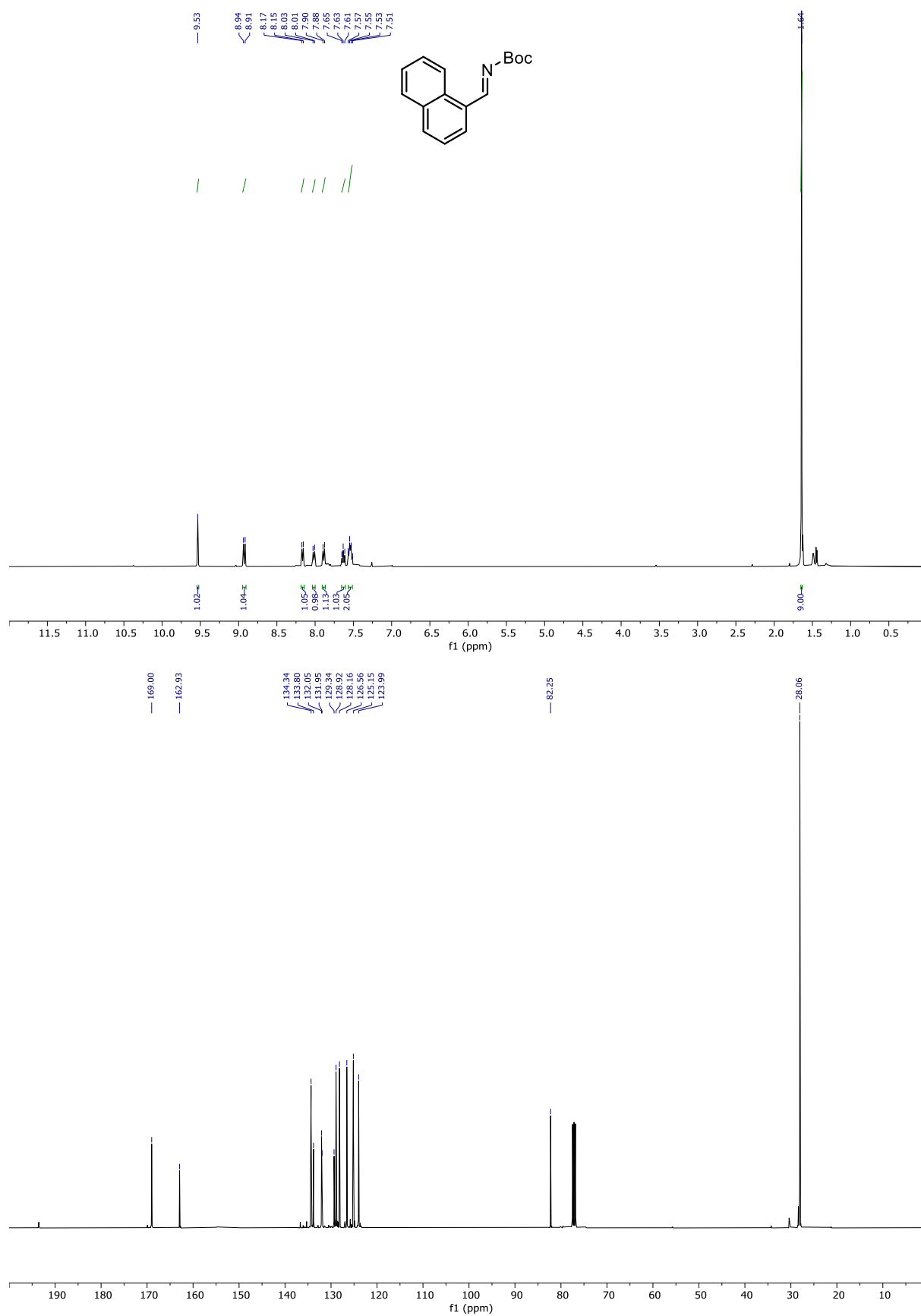
Tert-butyl (*E*)-(3-chlorobenzylidene)carbamate (**3c**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



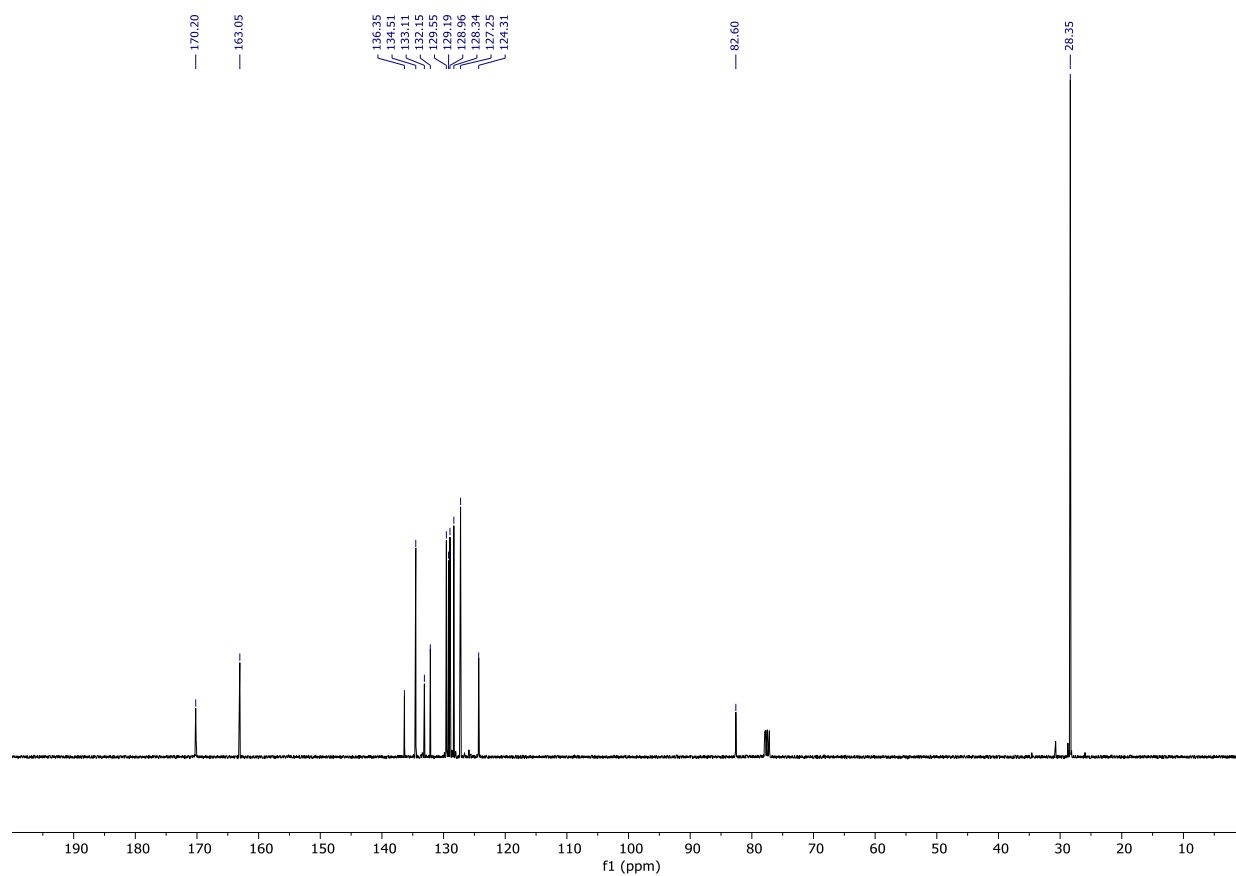
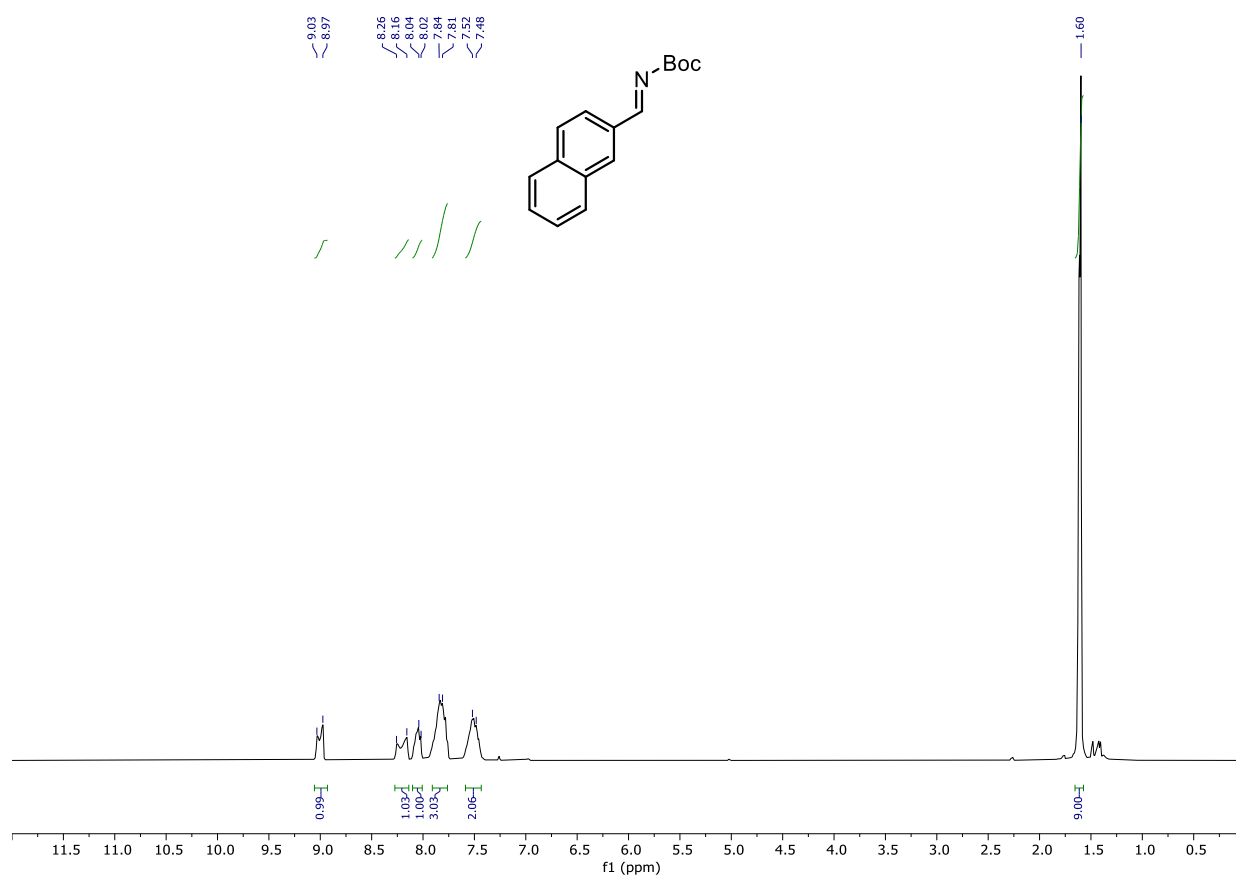
Tert-butyl (*E*)-(2-chlorobenzylidene)carbamate (3d) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



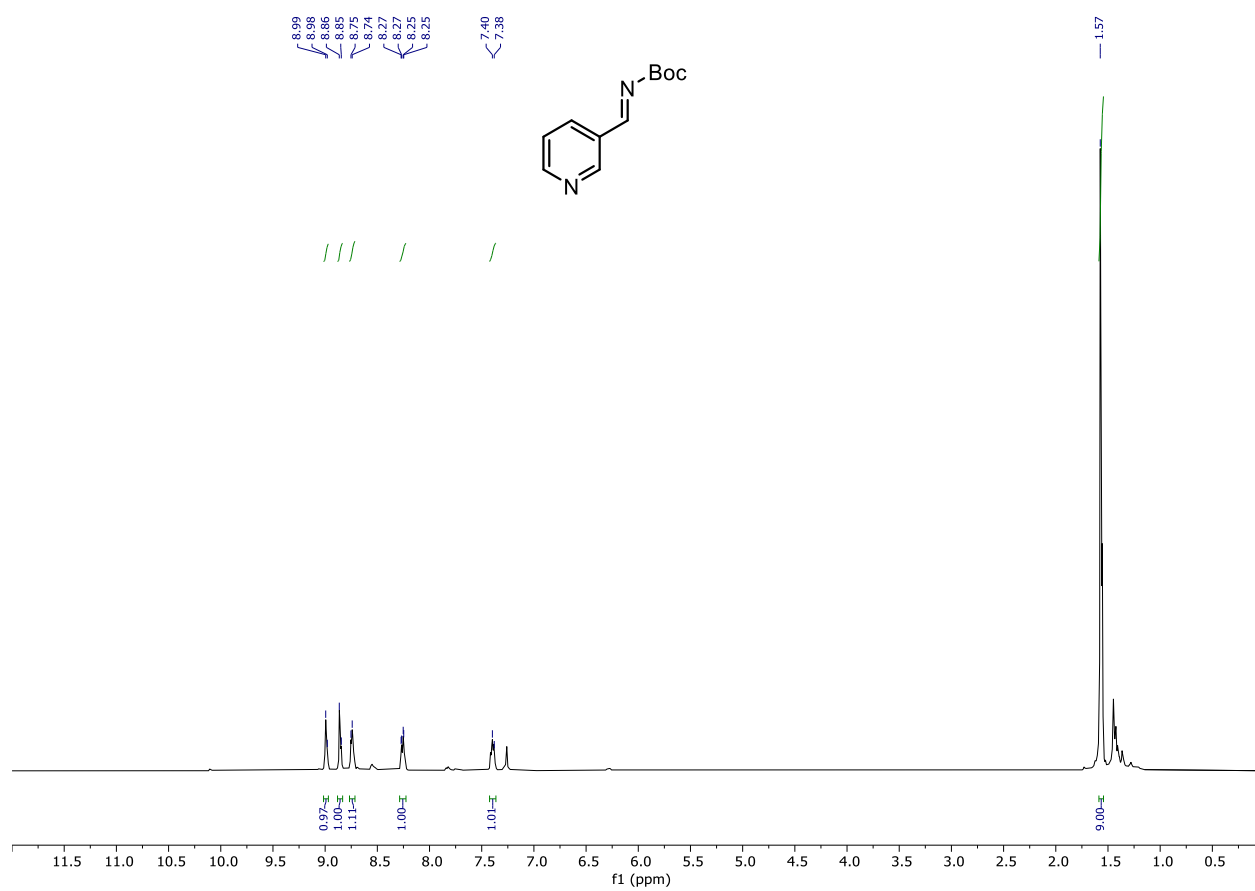
Tert-butyl (*E*)-(naphthalen-1-ylmethylene)carbamate (3e) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

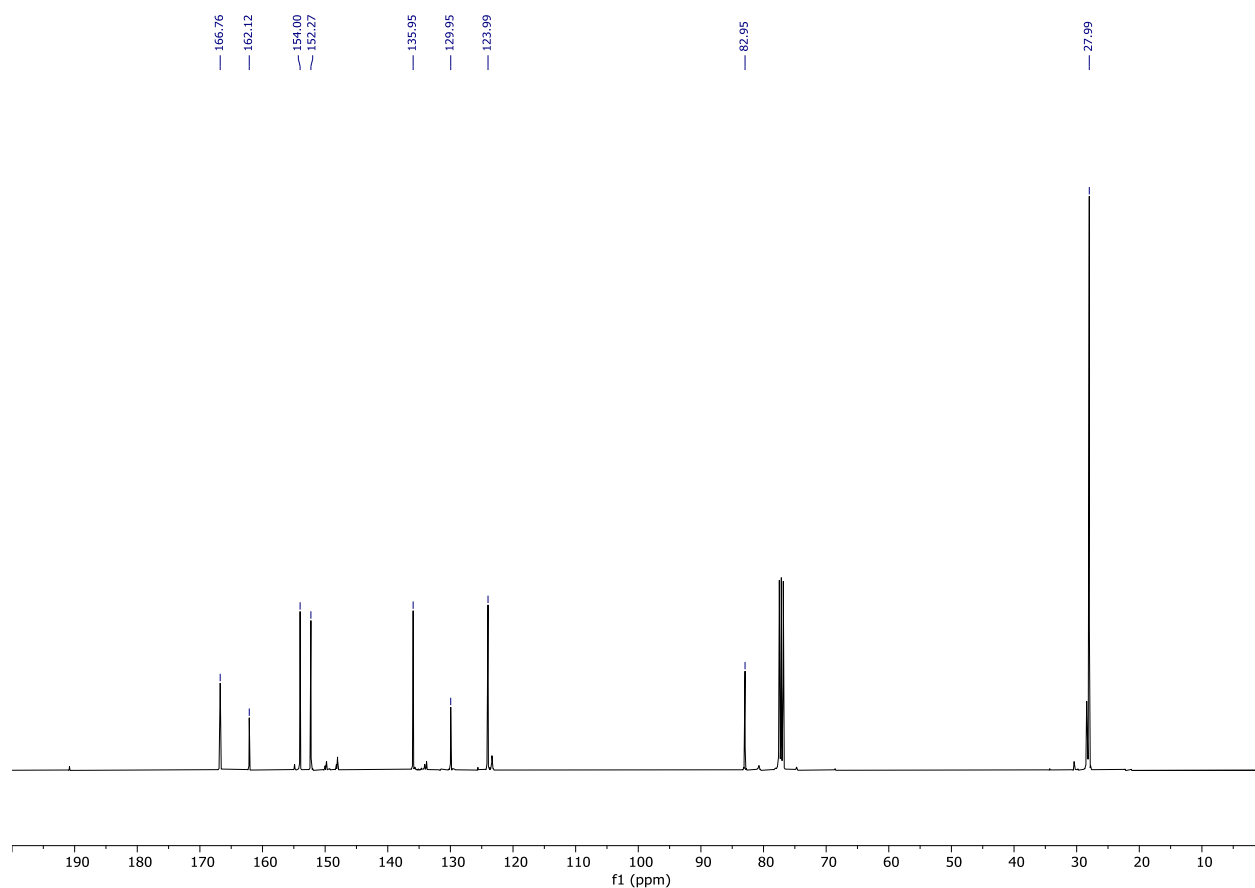


Tert-butyl (*E*)-(naphthalen-2-ylmethylene)carbamate (**3f**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

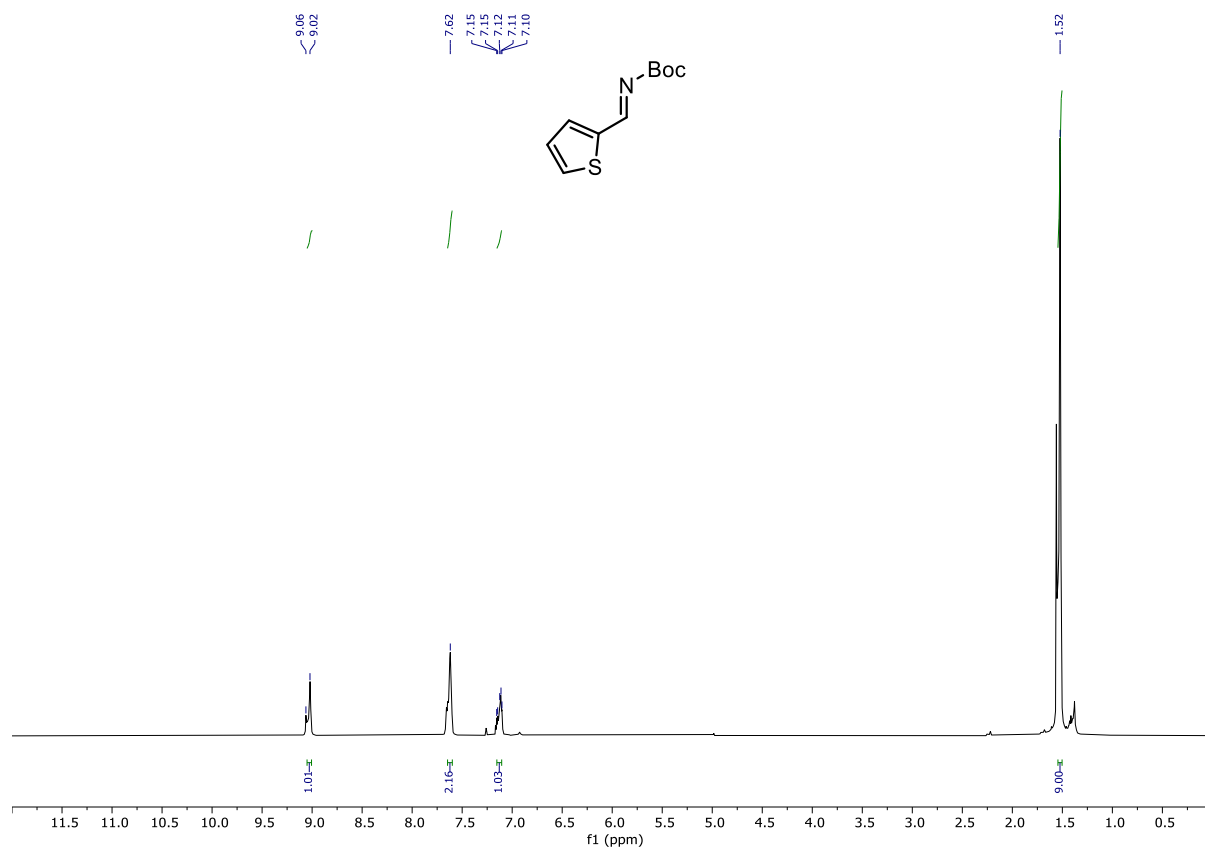


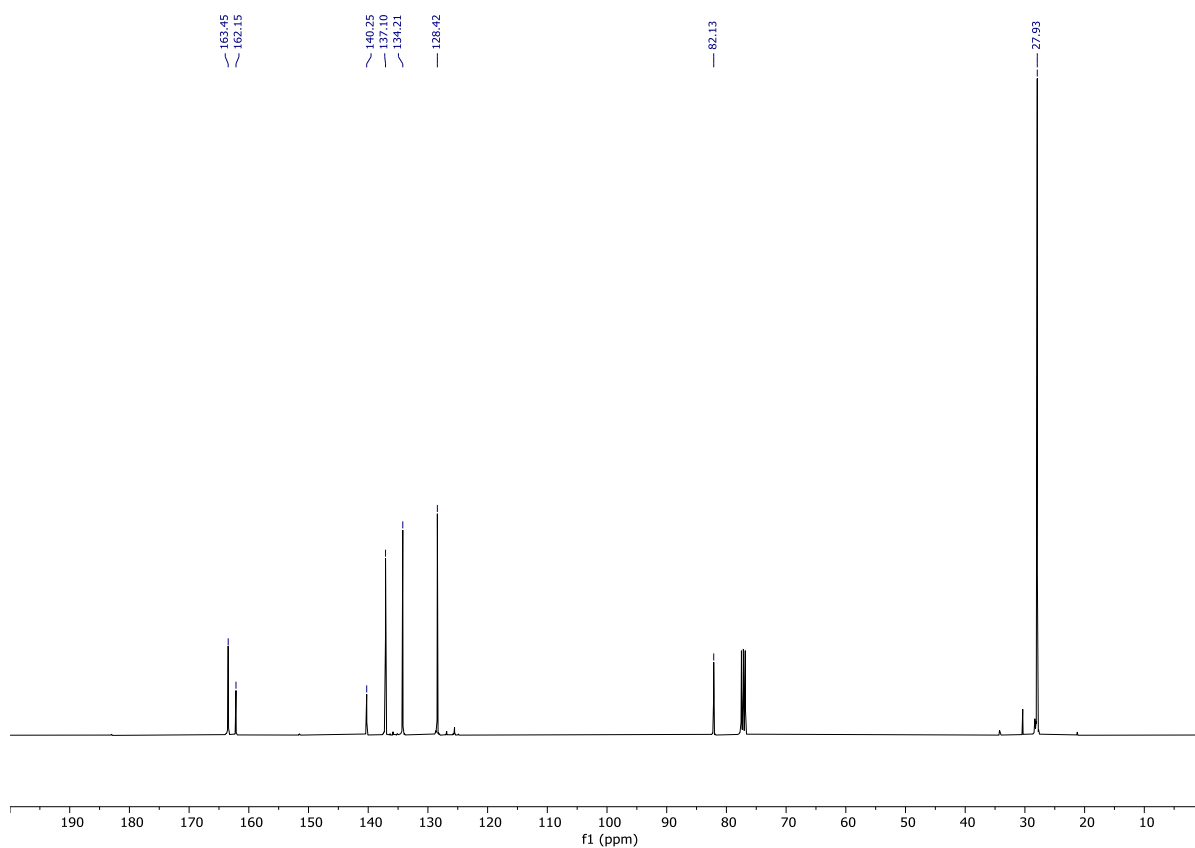
Tert-butyl (*E*)-(pyridin-3-ylmethylene)carbamate (3g) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



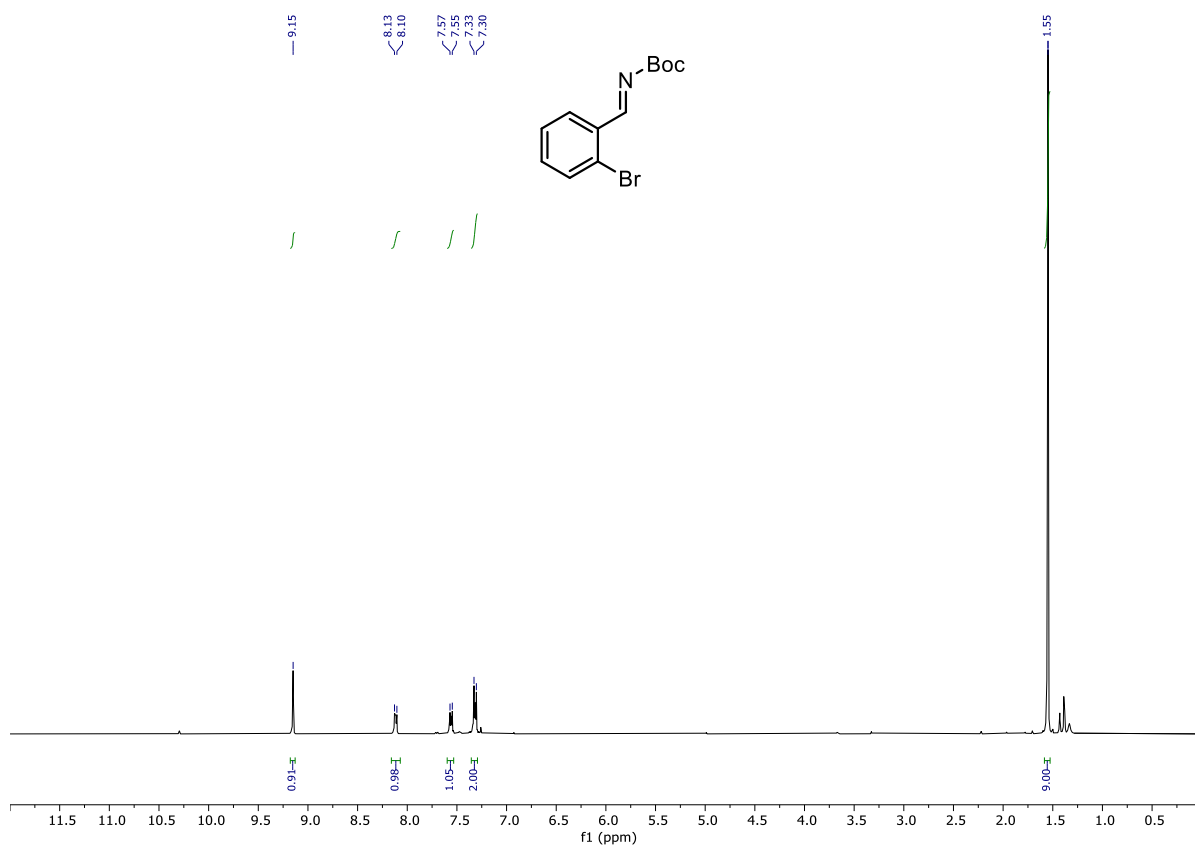


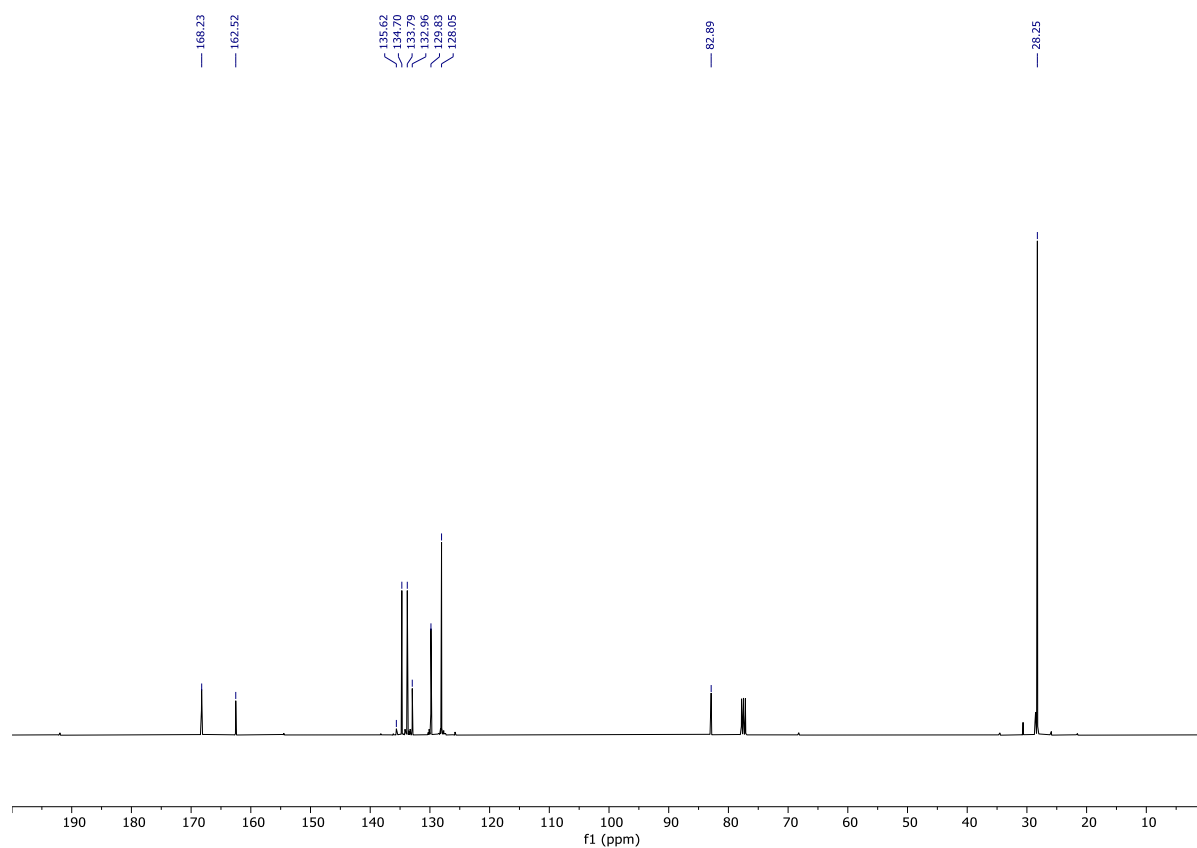
Tert-butyl (*E*)-(thiophen-2-ylmethylene)carbamate (3h) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



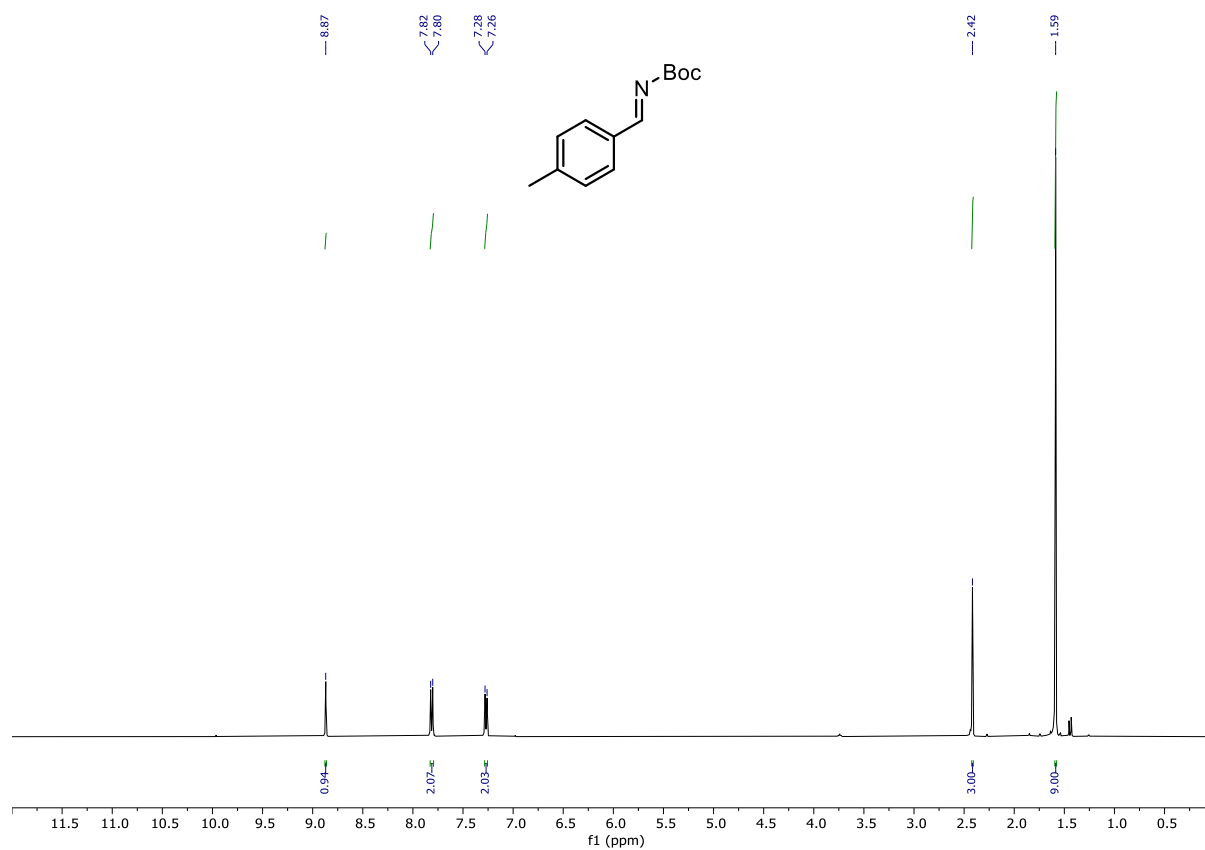


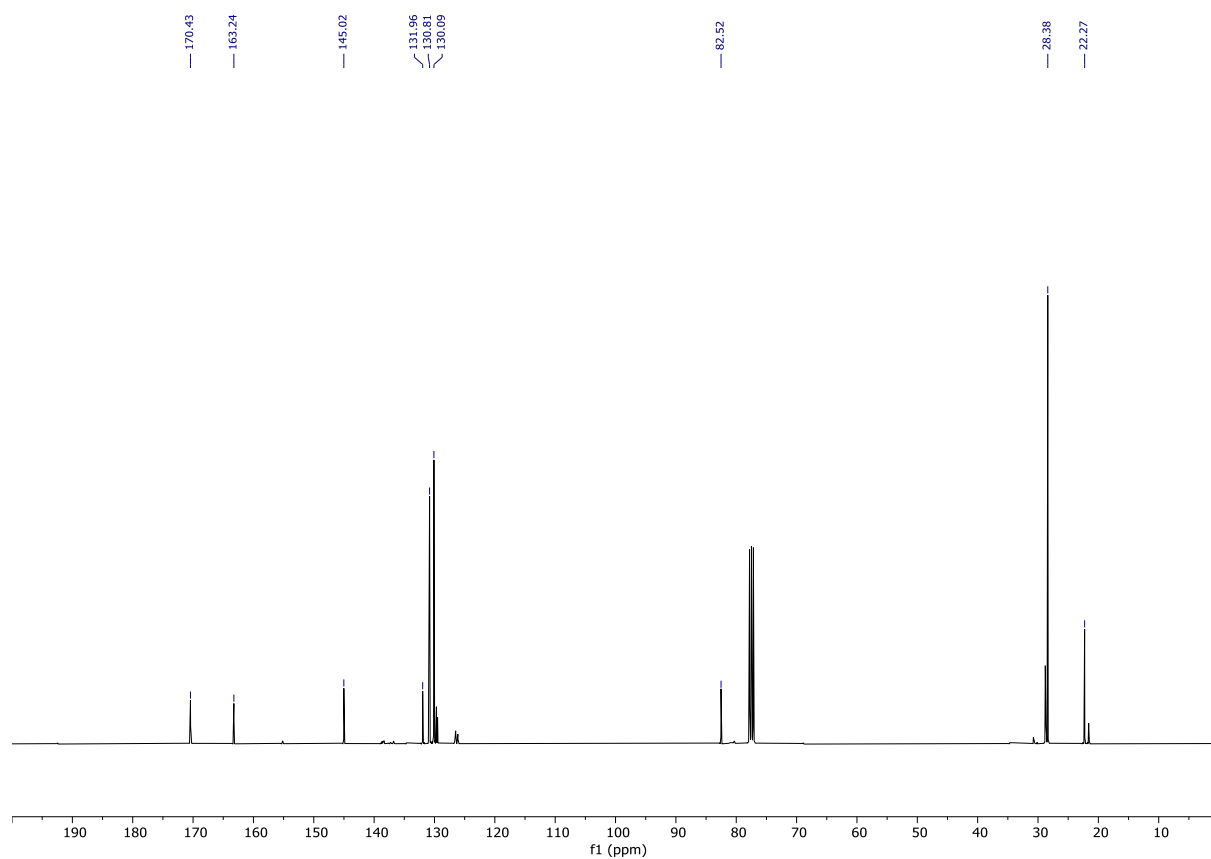
Tert-butyl (E)-(2-bromobenzylidene)carbamate (3i) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



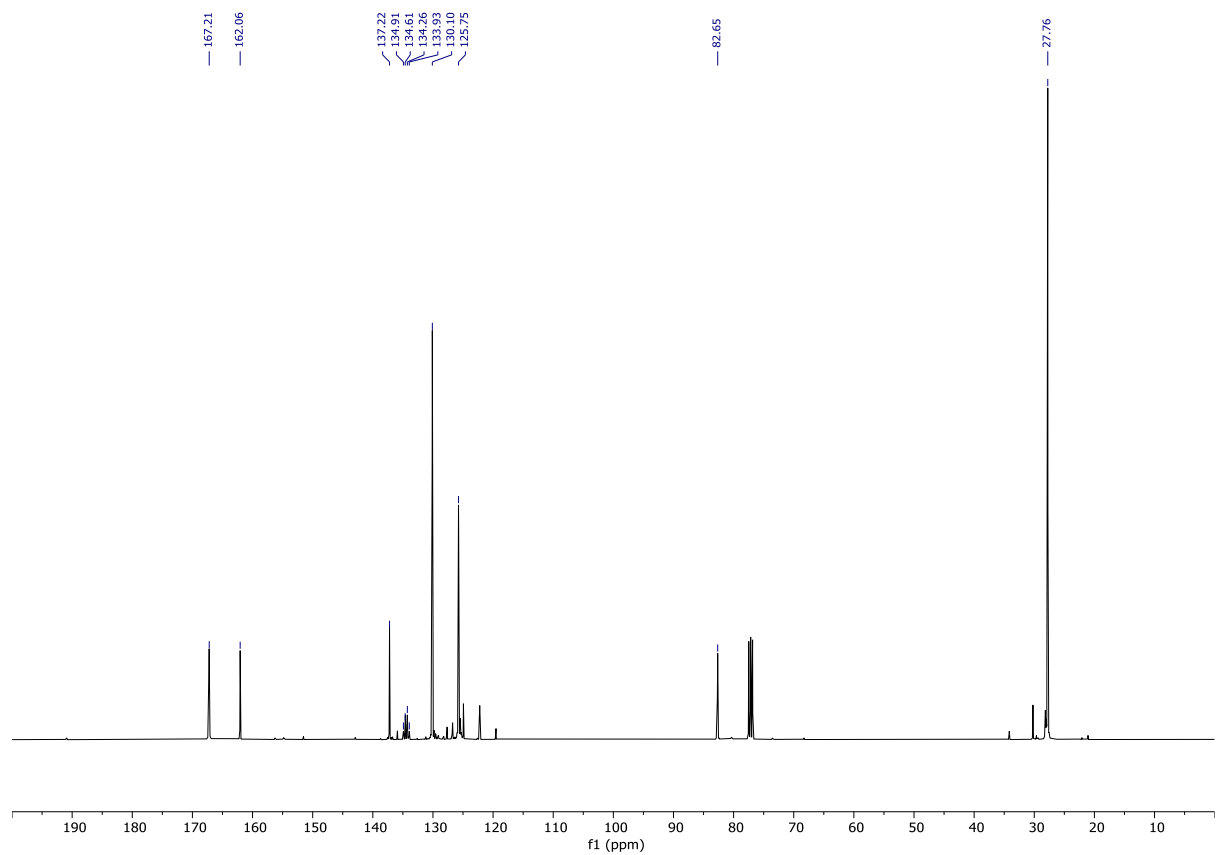
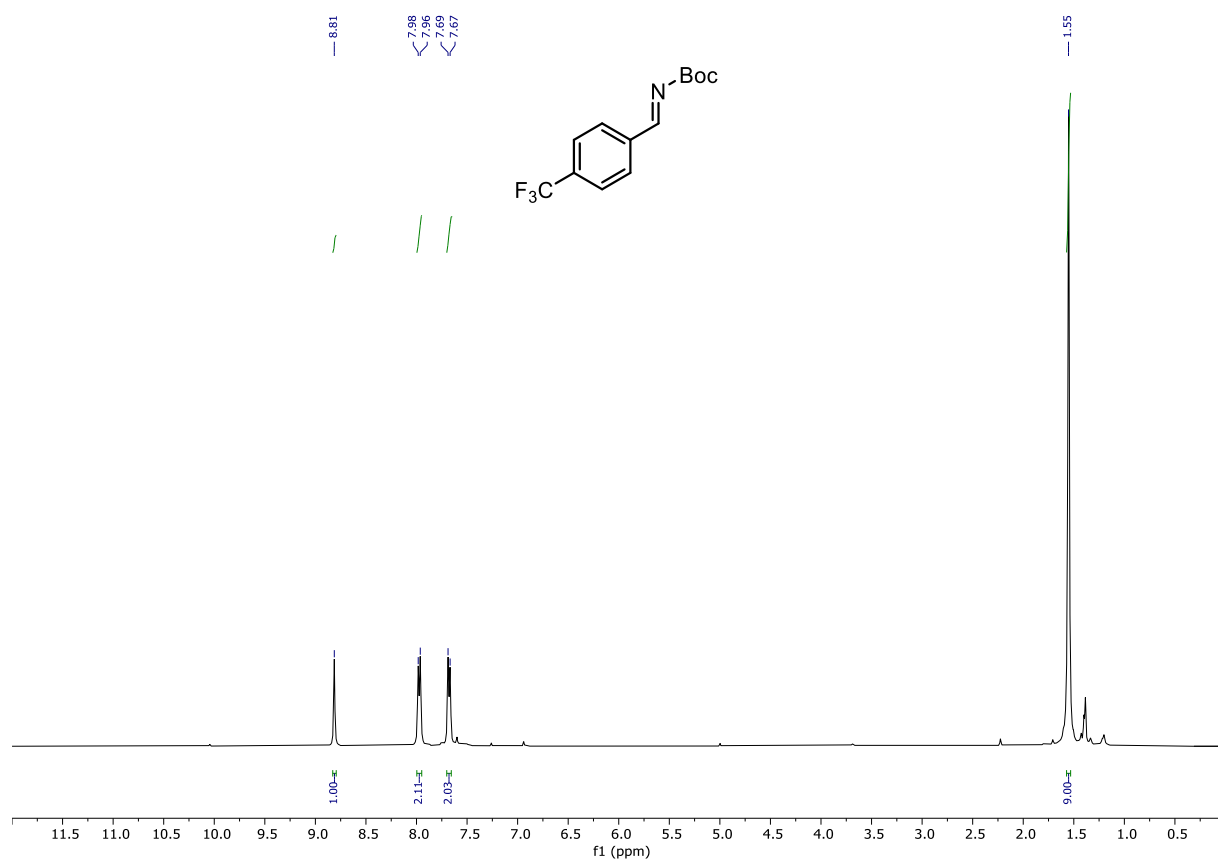


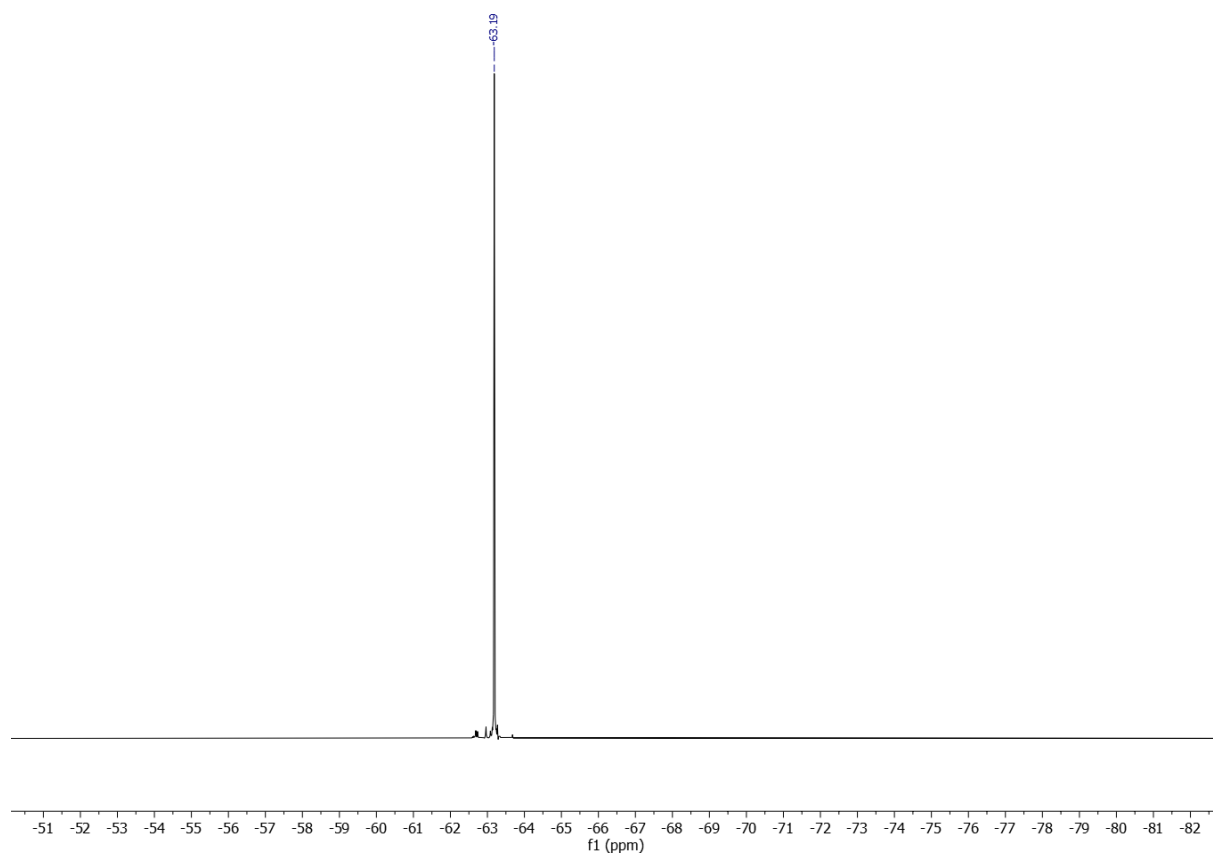
Tert-butyl (*E*)-(4-methylbenzylidene)carbamate (3j) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



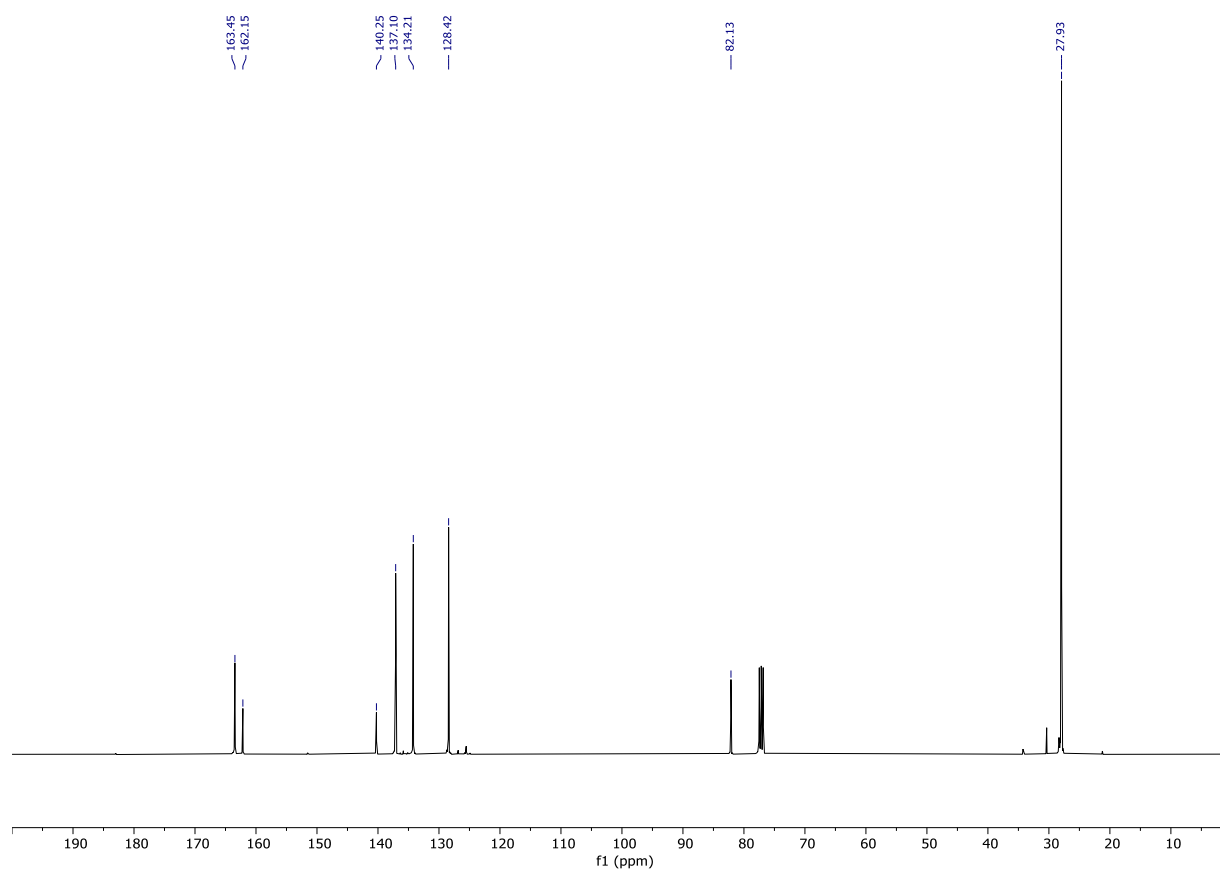
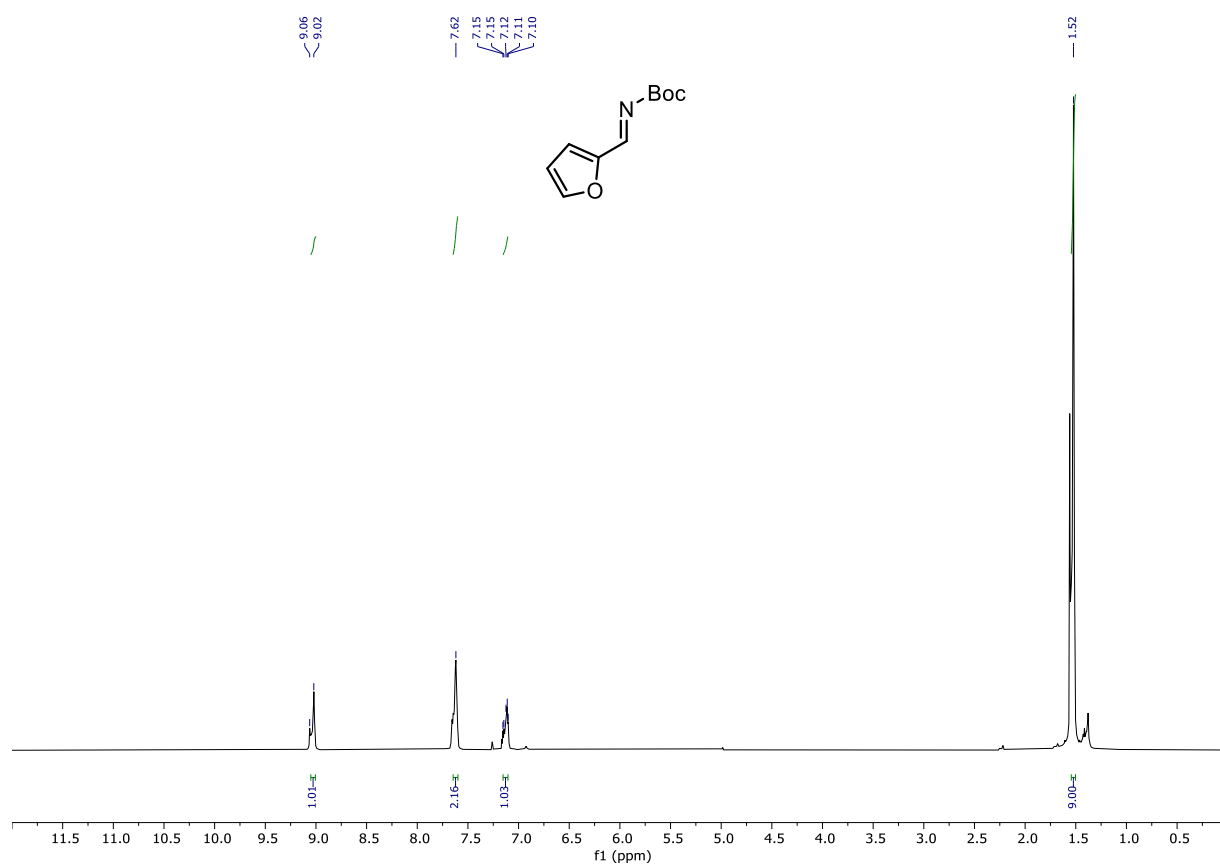


Tert-butyl (*E*)-(4-(trifluoromethyl)benzylidene)carbamate (3k) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz, ¹⁹F 376 MHz)



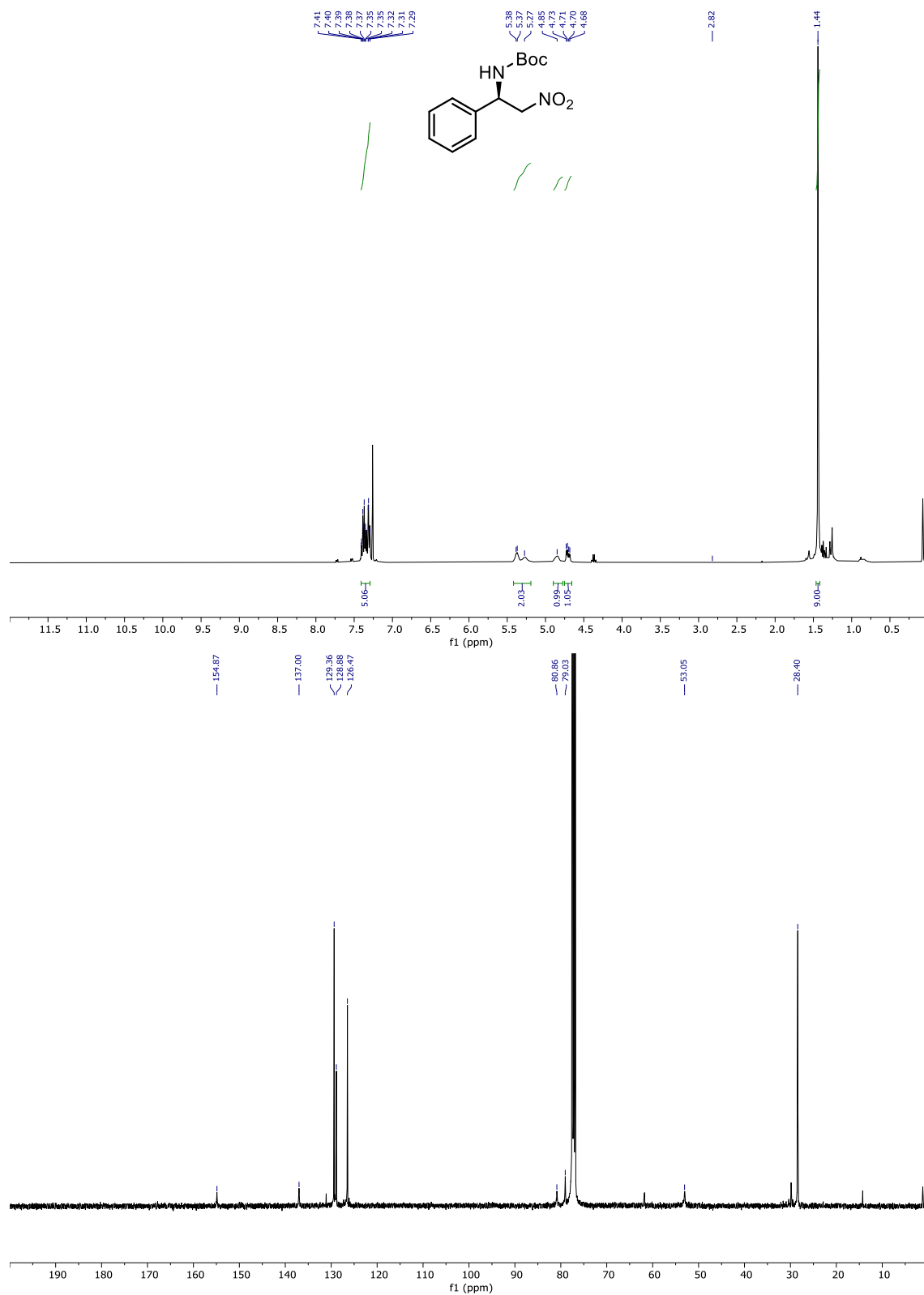


Tert-butyl (*E*)-(furan-2-ylmethylene)carbamate (3l) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

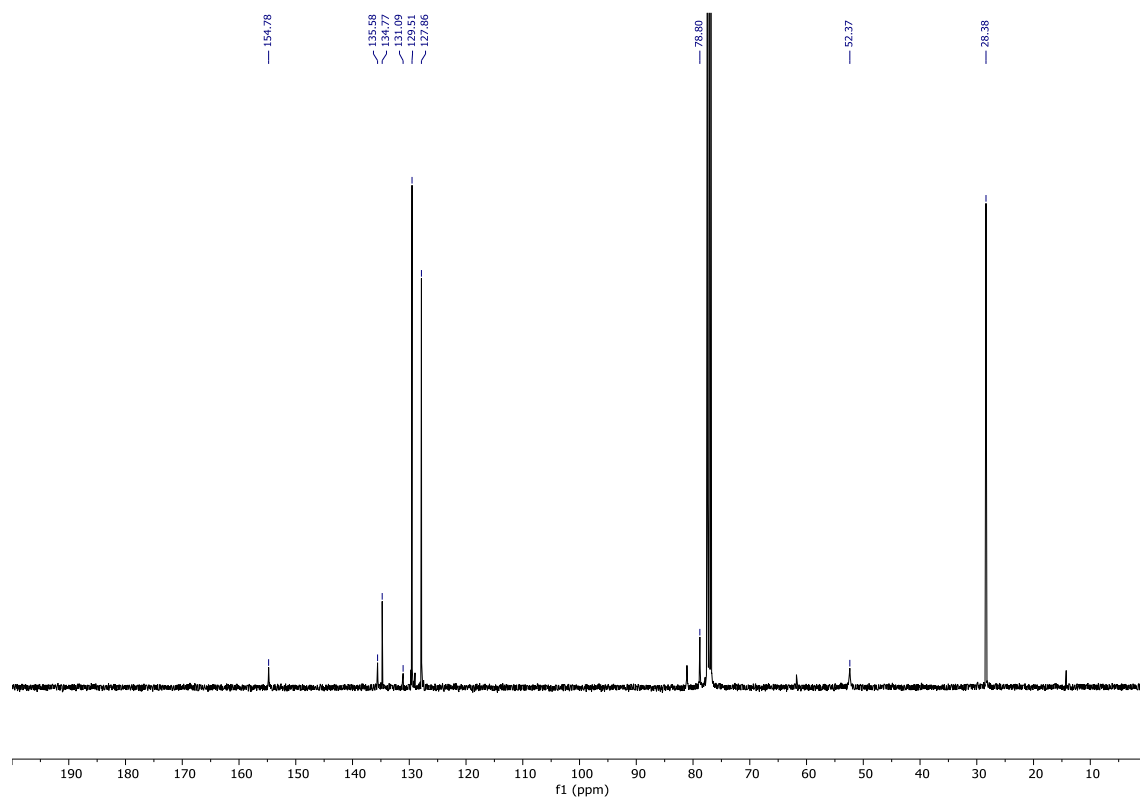
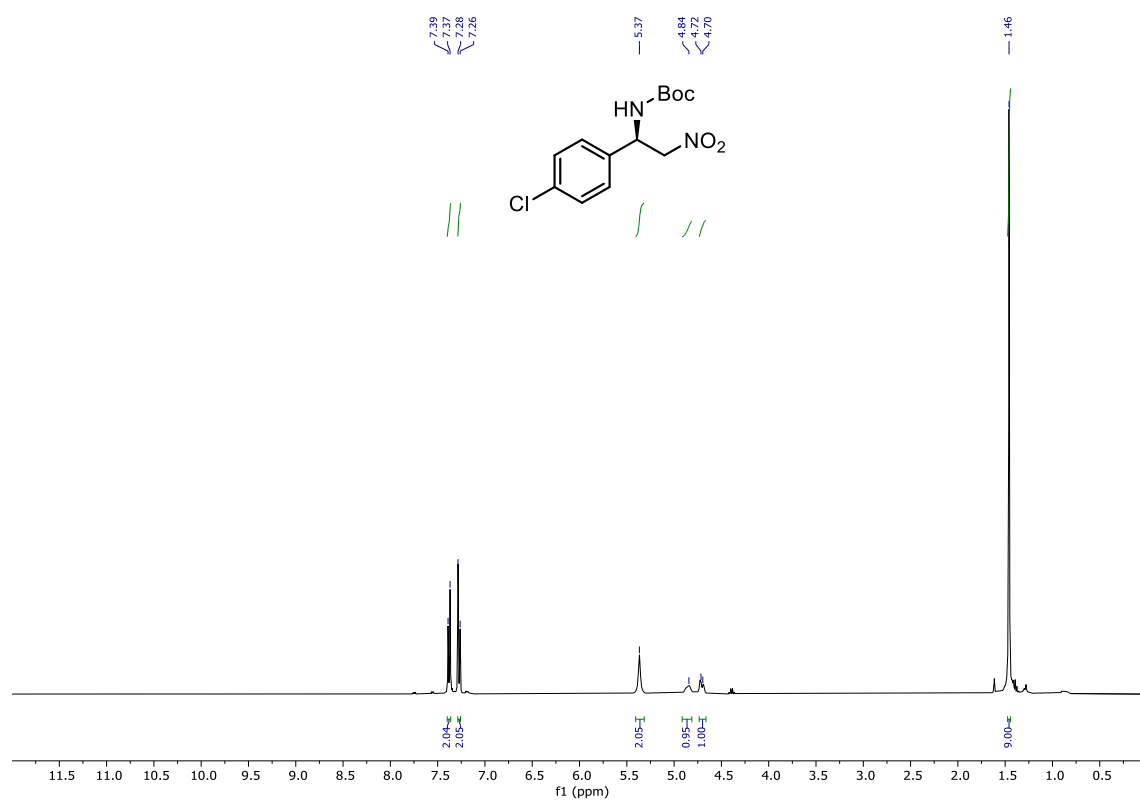


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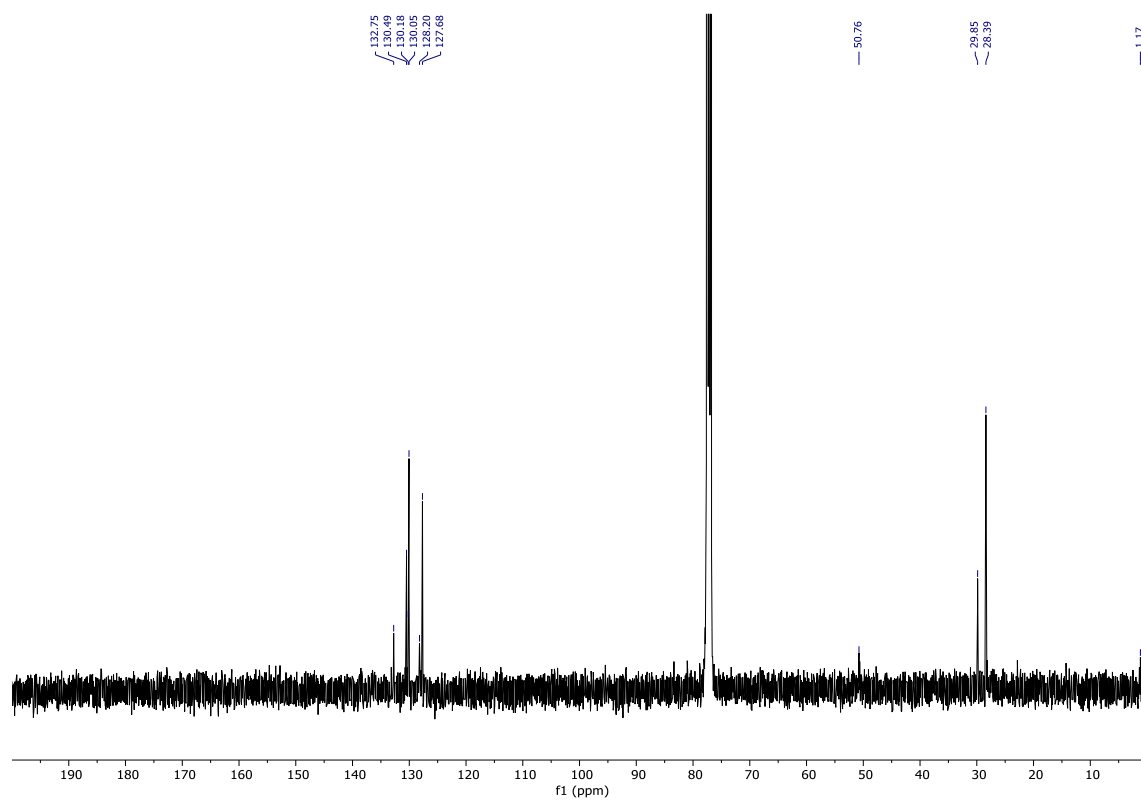
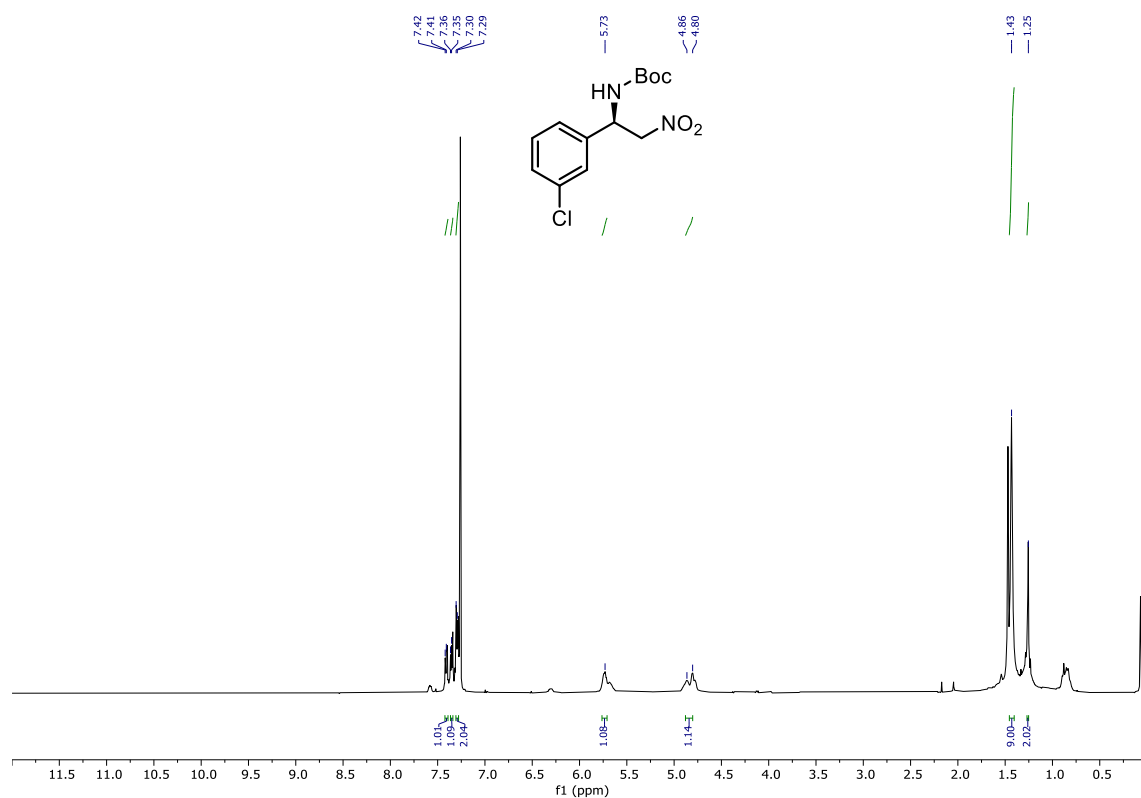
Tert-butyl (*R*)-(2-nitro-1-phenylethyl)carbamate (**4a**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



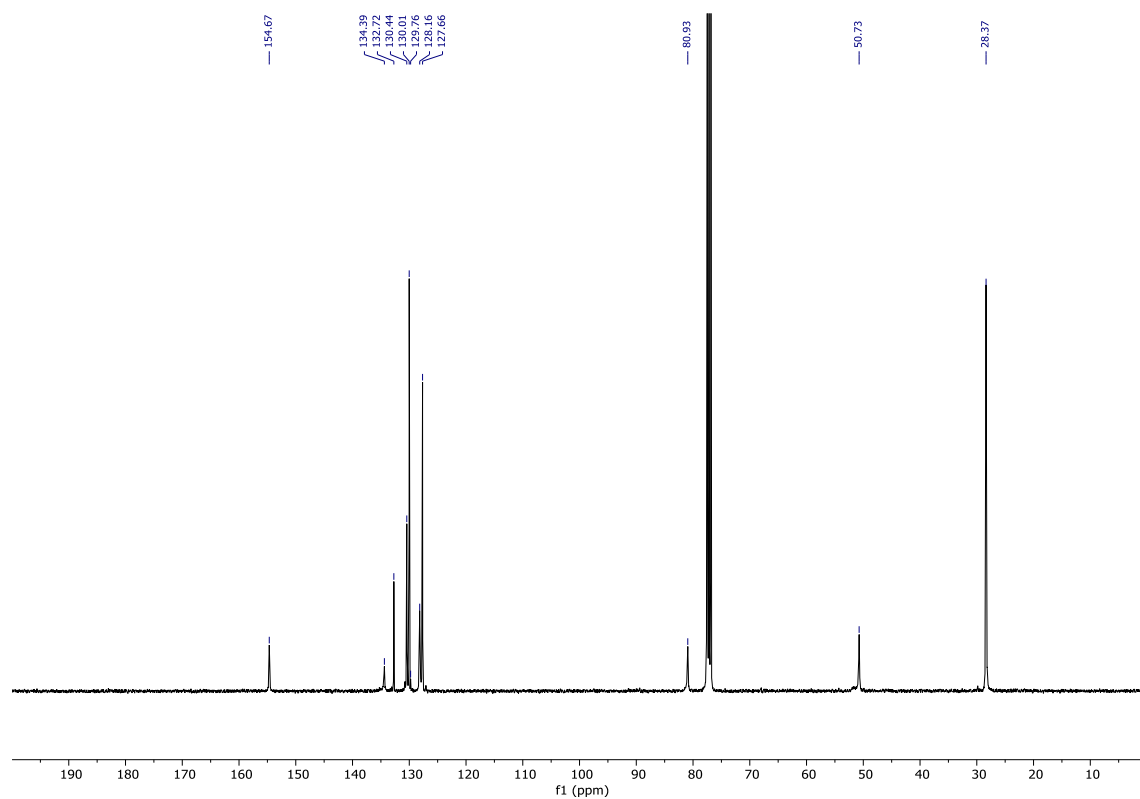
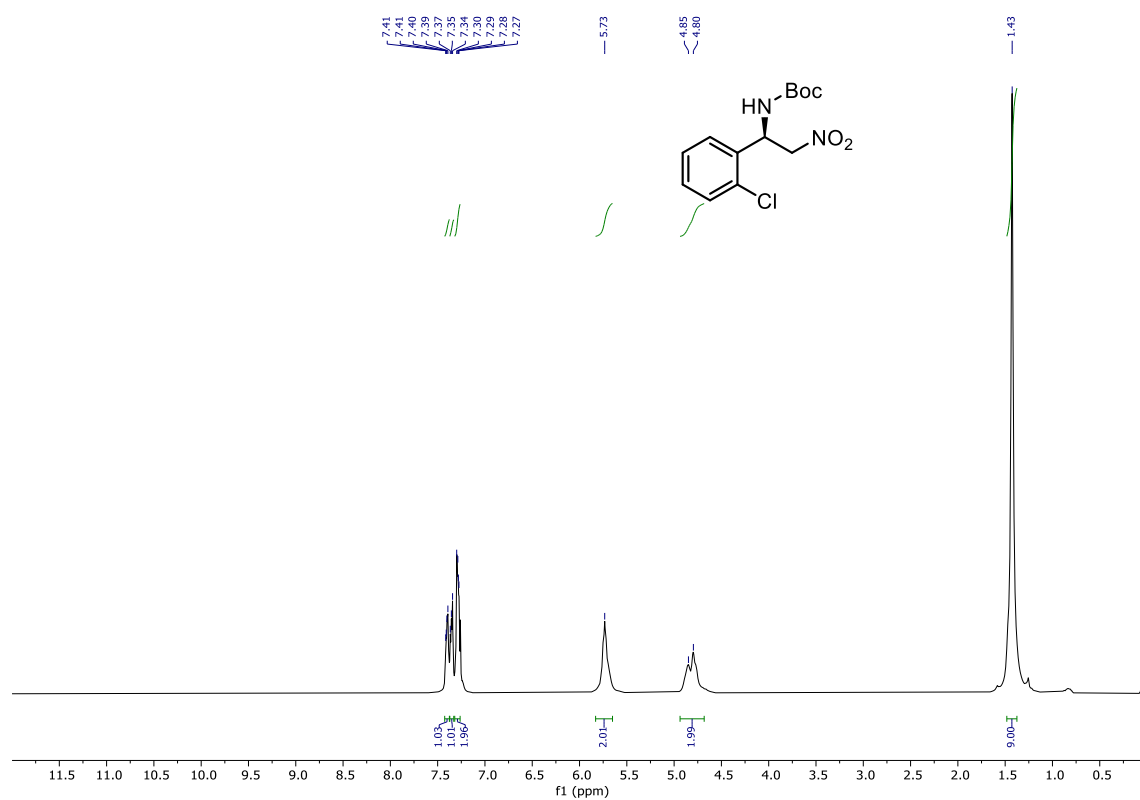
Tert-butyl (*R*)-(1-(4-chlorophenyl)-2-nitroethyl)carbamate (**4b**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



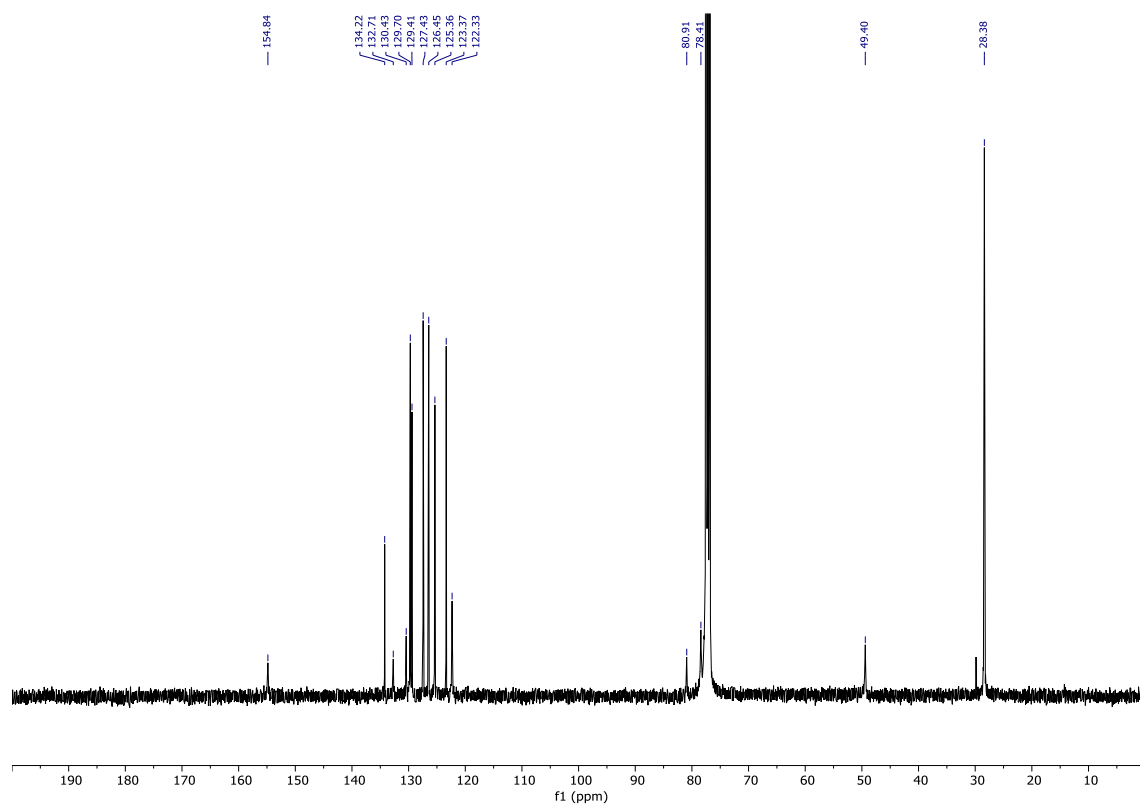
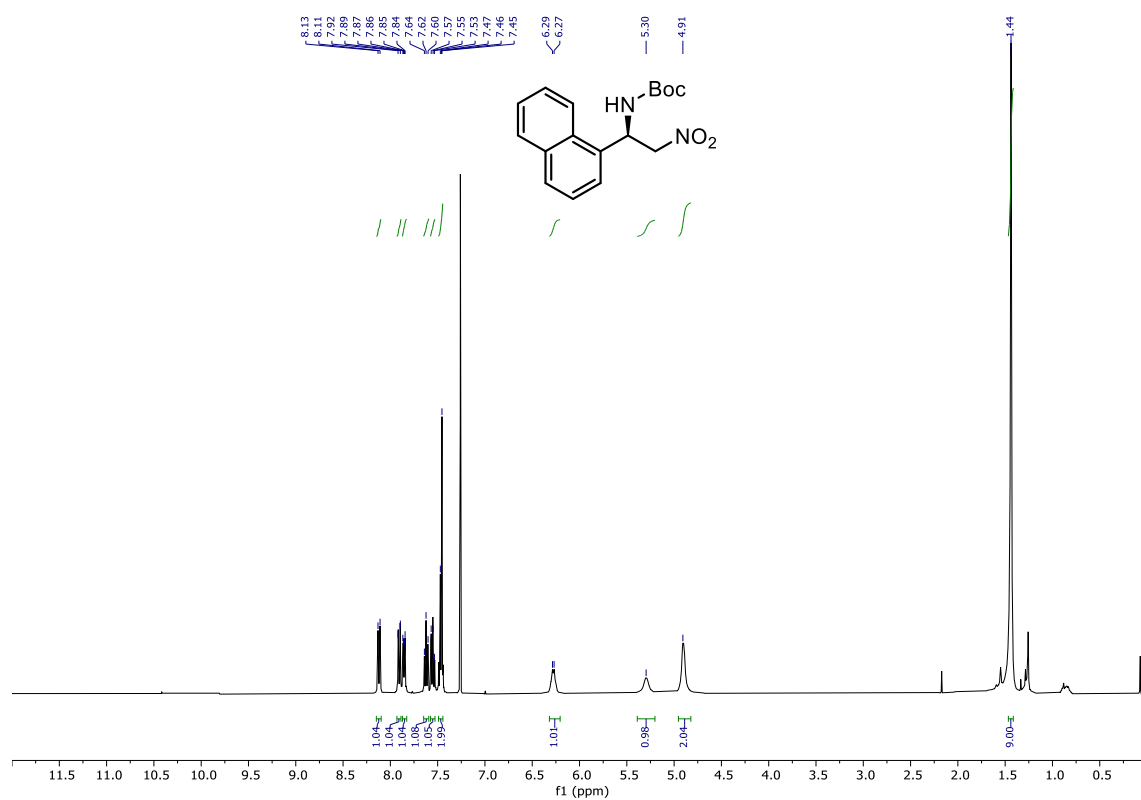
Tert-butyl (*R*)-(1-(3-chlorophenyl)-2-nitroethyl)carbamate (**4c**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



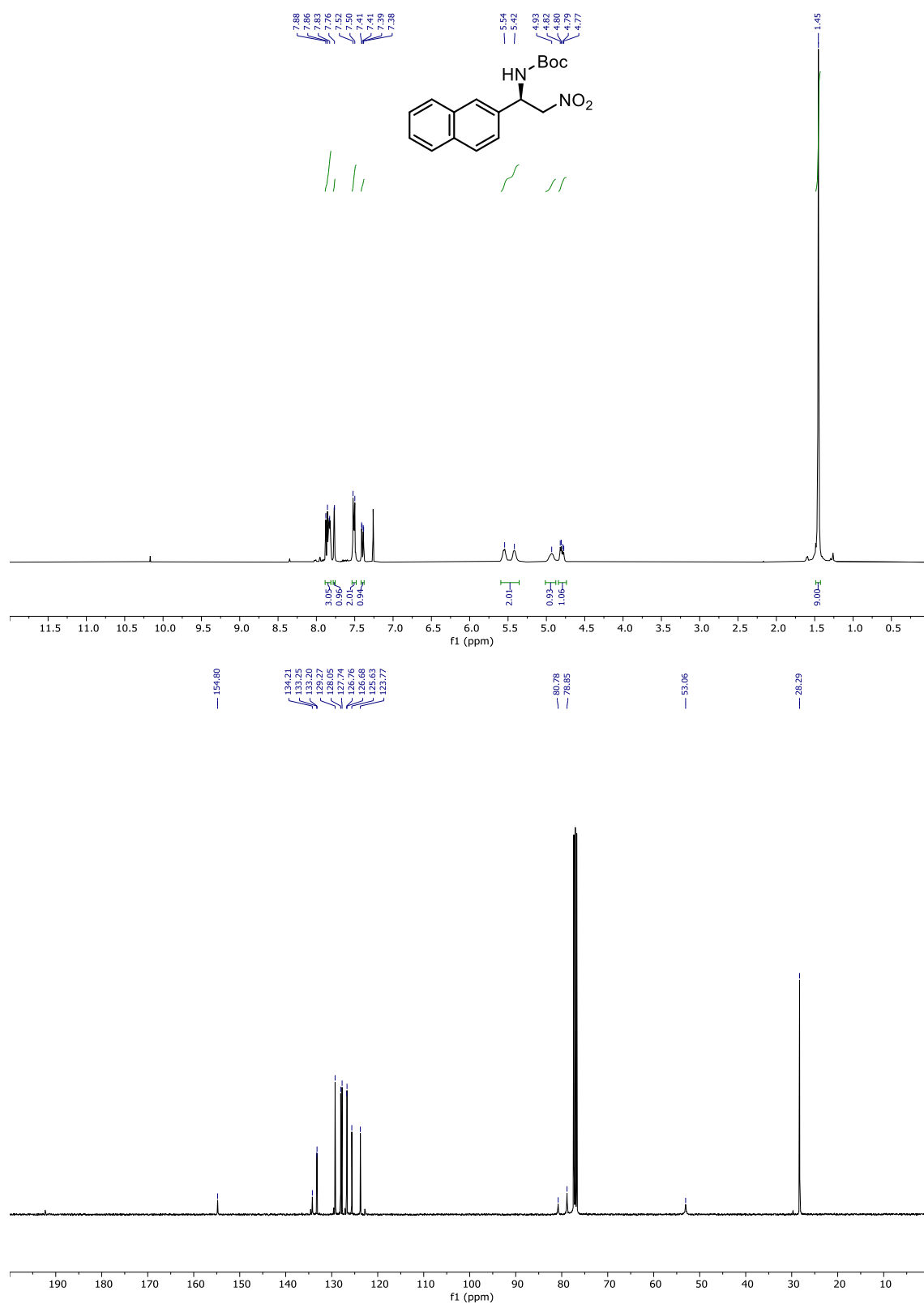
Tert-butyl (*R*)-(1-(2-chlorophenyl)-2-nitroethyl)carbamate (4d) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



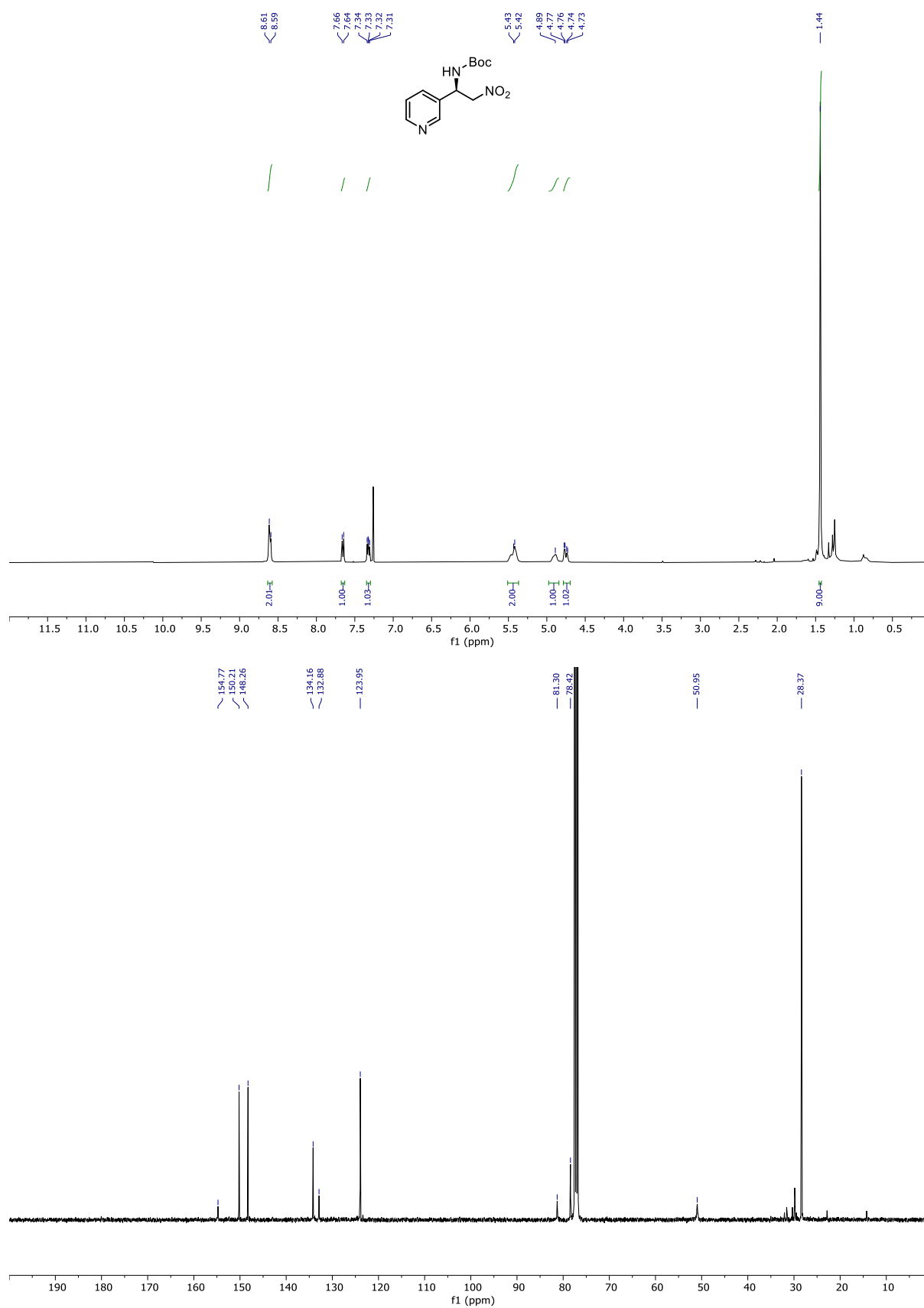
Tert-butyl (*R*)-(1-(naphthalen-1-yl)-2-nitroethyl)carbamate (4e) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



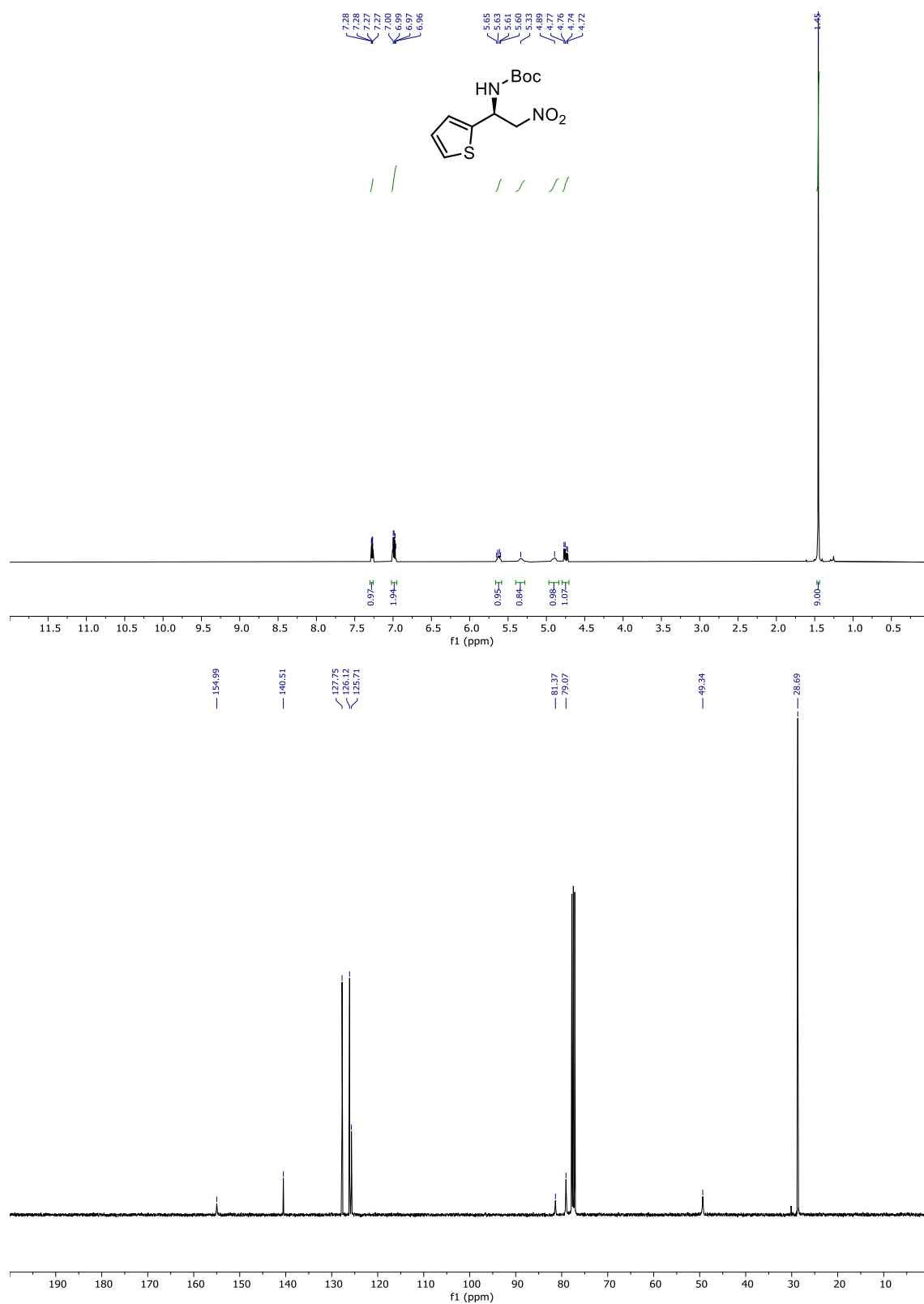
Tert-butyl (*R*)-(1-(naphthalen-2-yl)-2-nitroethyl)carbamate (4f) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



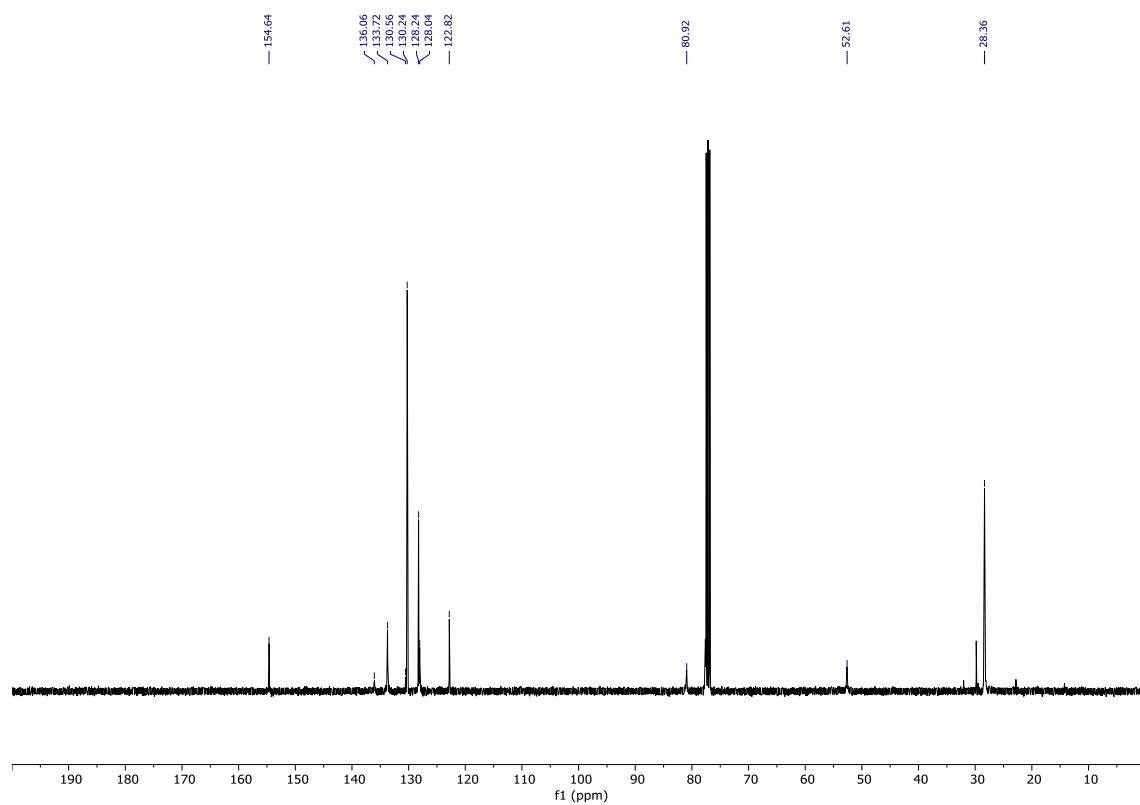
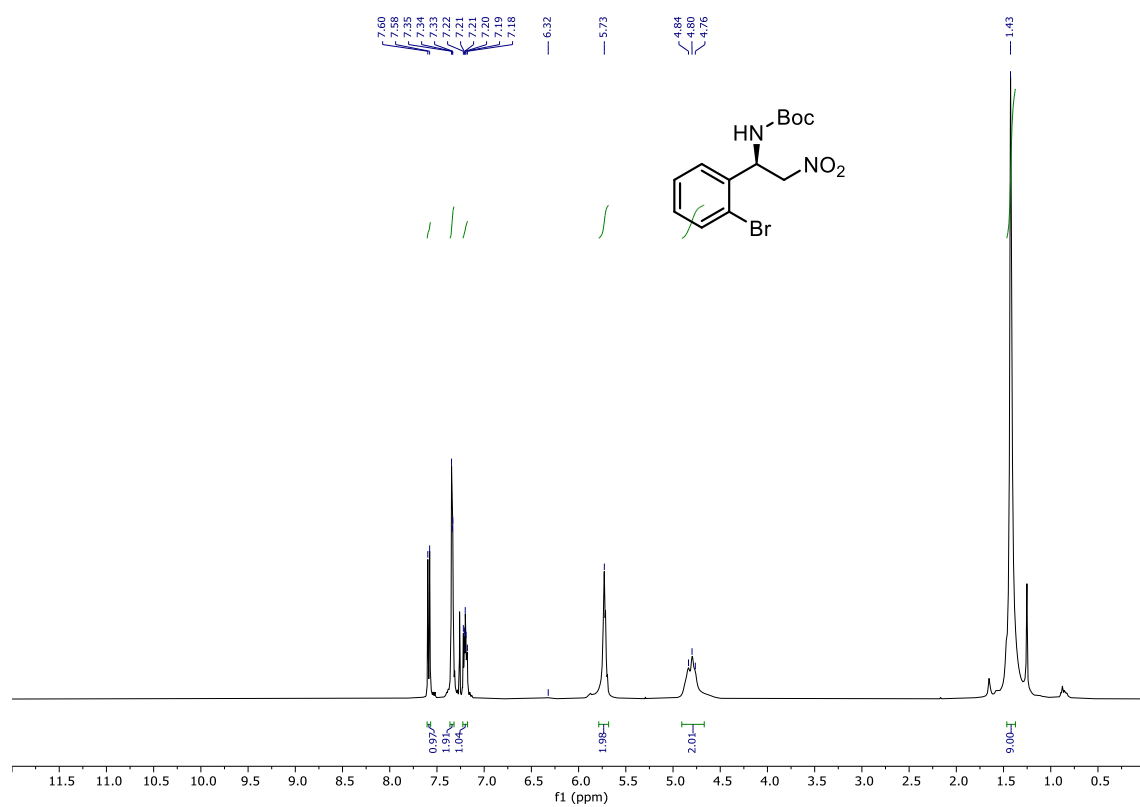
Tert-butyl (*R*)-(2-nitro-1-(pyridin-3-yl)ethyl)carbamate (4g) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



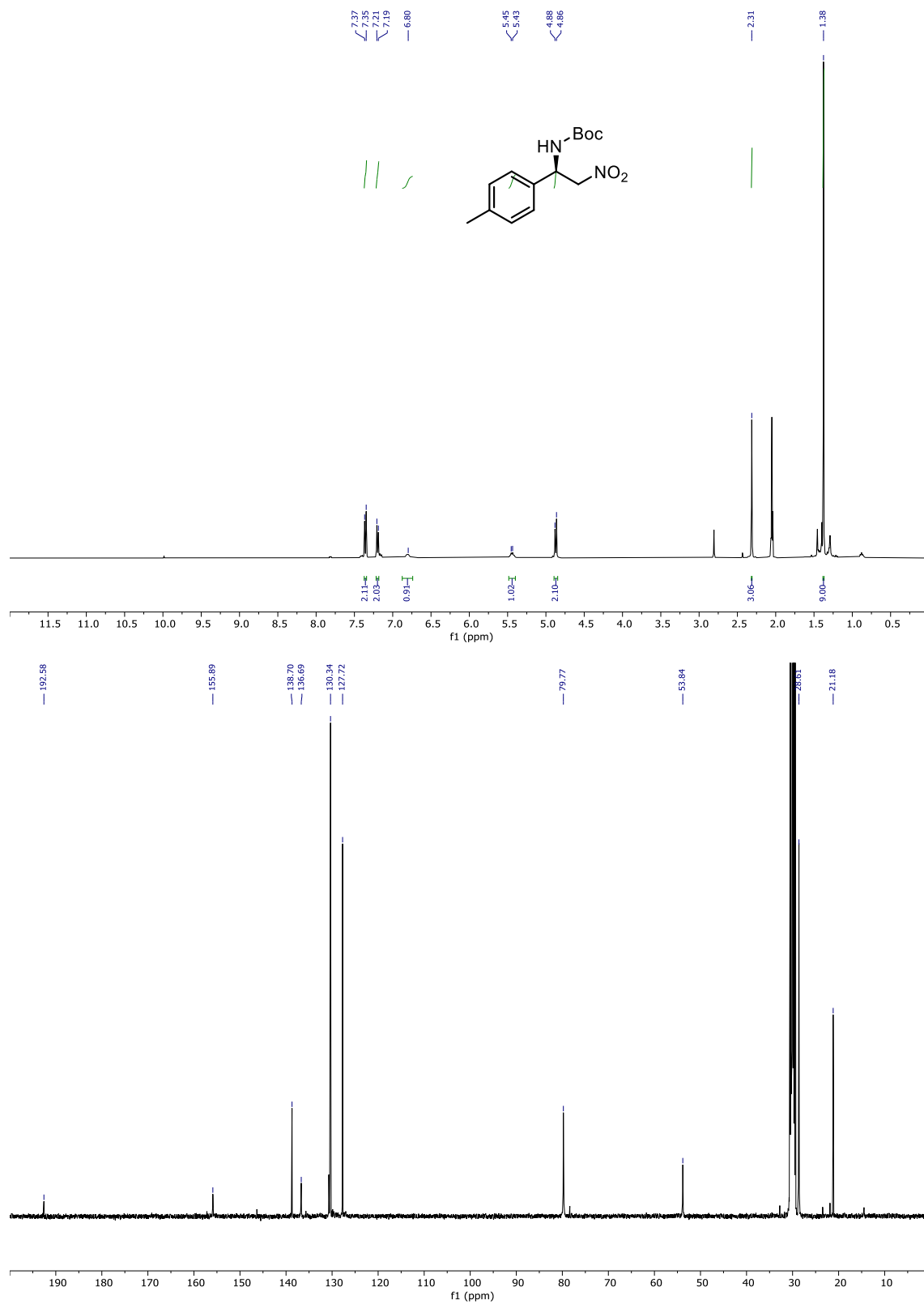
Tert-butyl (*R*)-(2-nitro-1-(thiophen-2-yl)ethyl)carbamate (**4h**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



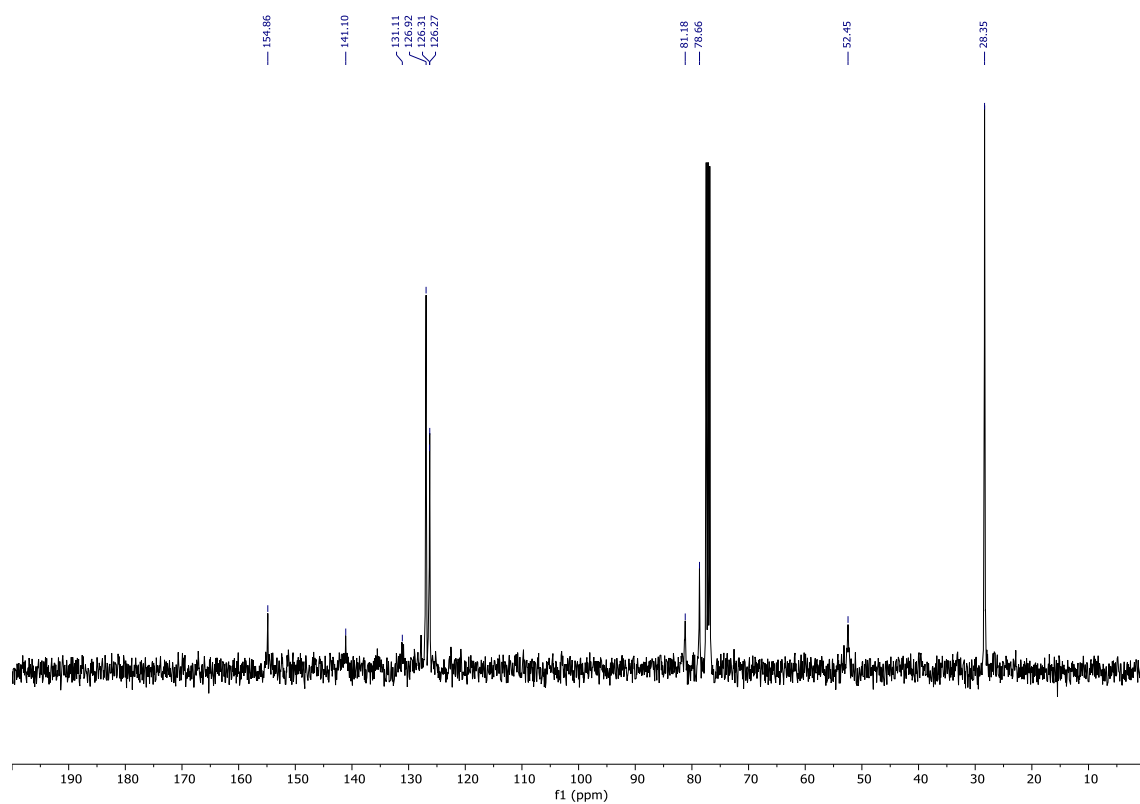
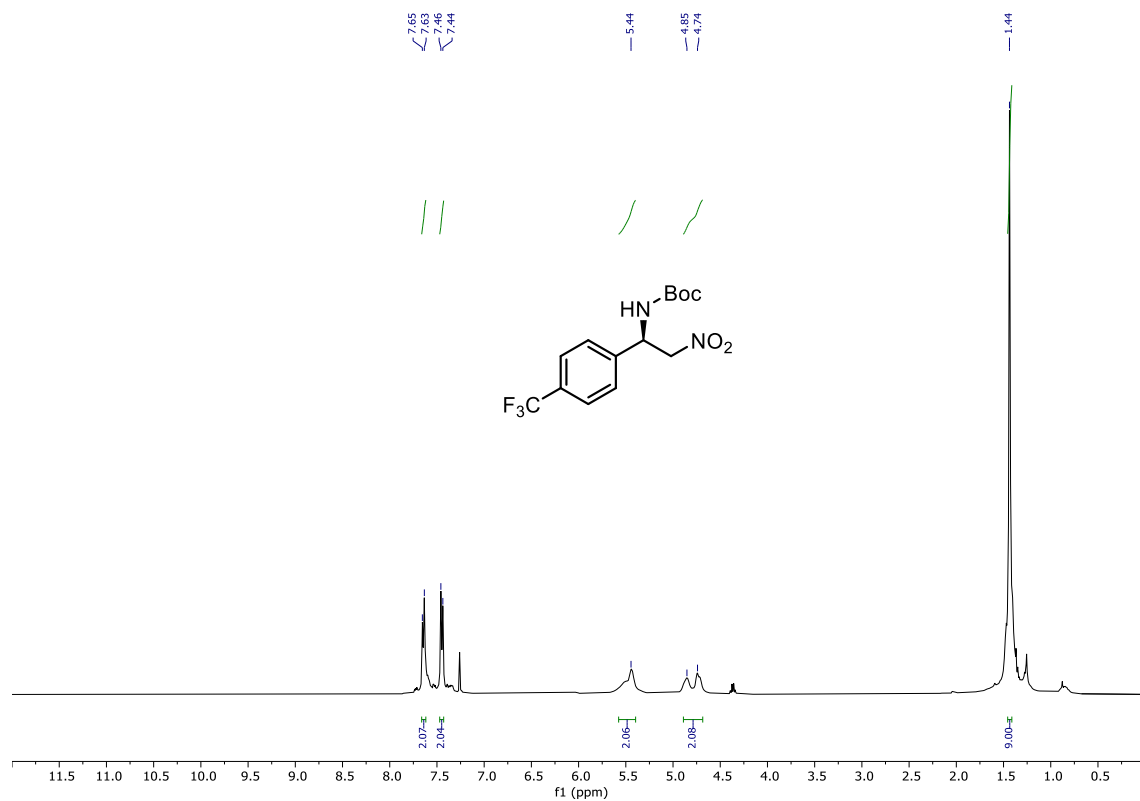
Tert-butyl (*R*)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4i) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

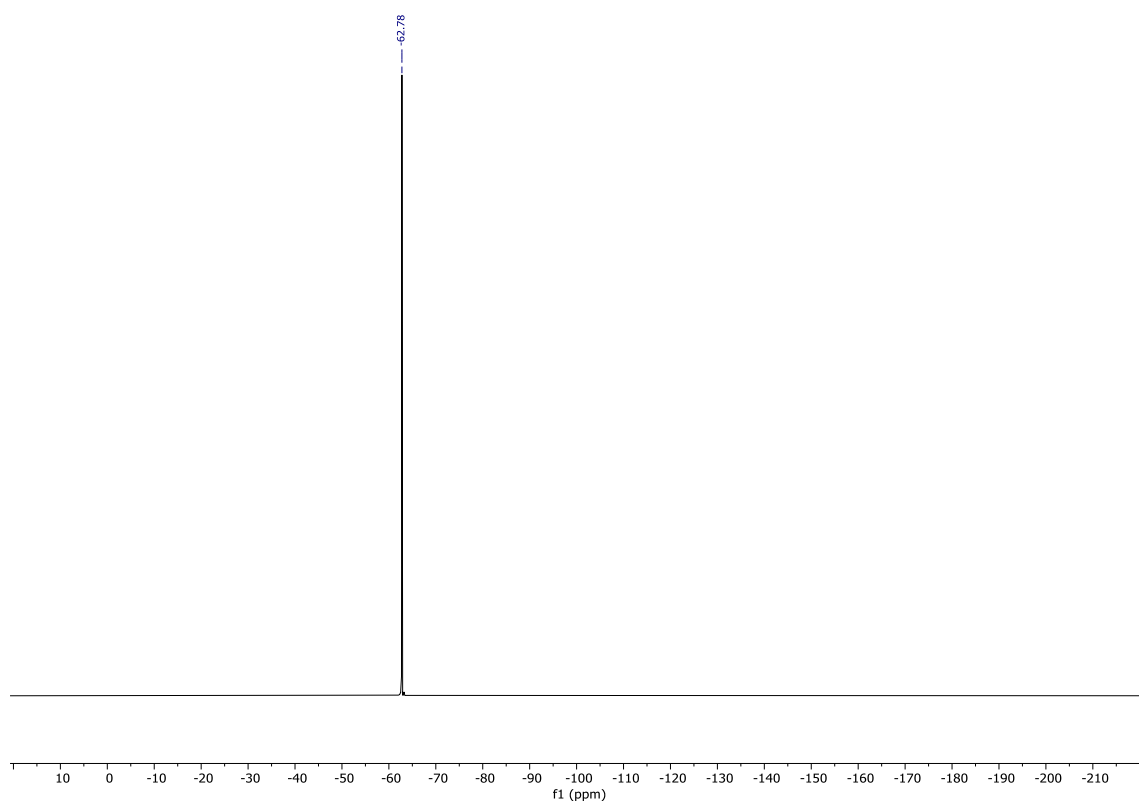


Tert-butyl (*R*)-(1-(2-bromophenyl)-2-nitroethyl)carbamate (4j) (Acetone-d₆, ¹H 400 MHz, ¹³C 101 MHz)

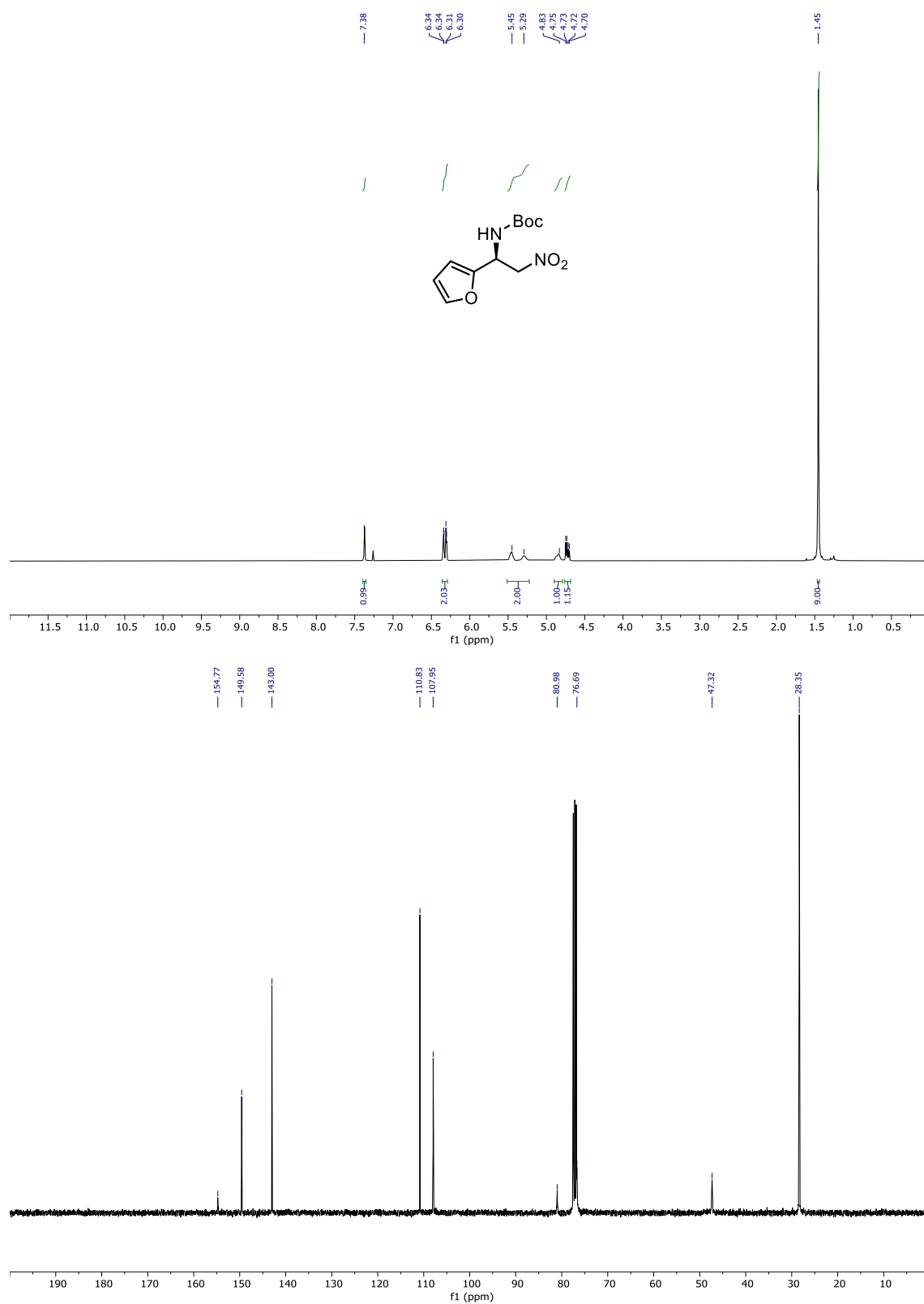


Tert-butyl (*R*)-(2-nitro-1-(4-(trifluoromethyl)phenyl)ethyl)carbamate (**4k**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz, ¹⁹F 376 MHz)



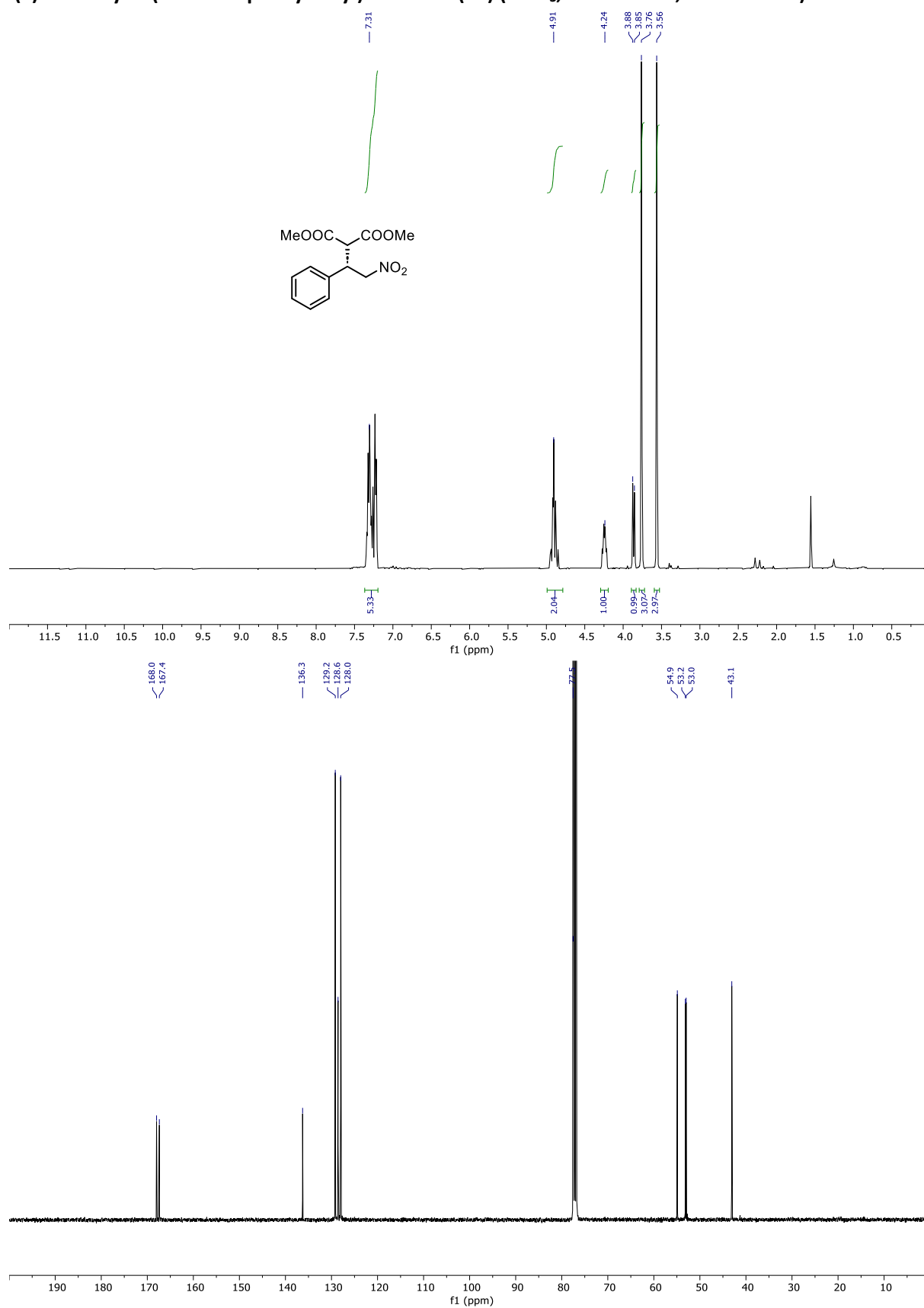


Tert-butyl (*R*)-(1-(furan-2-yl)-2-nitroethyl)carbamate (**4l**) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

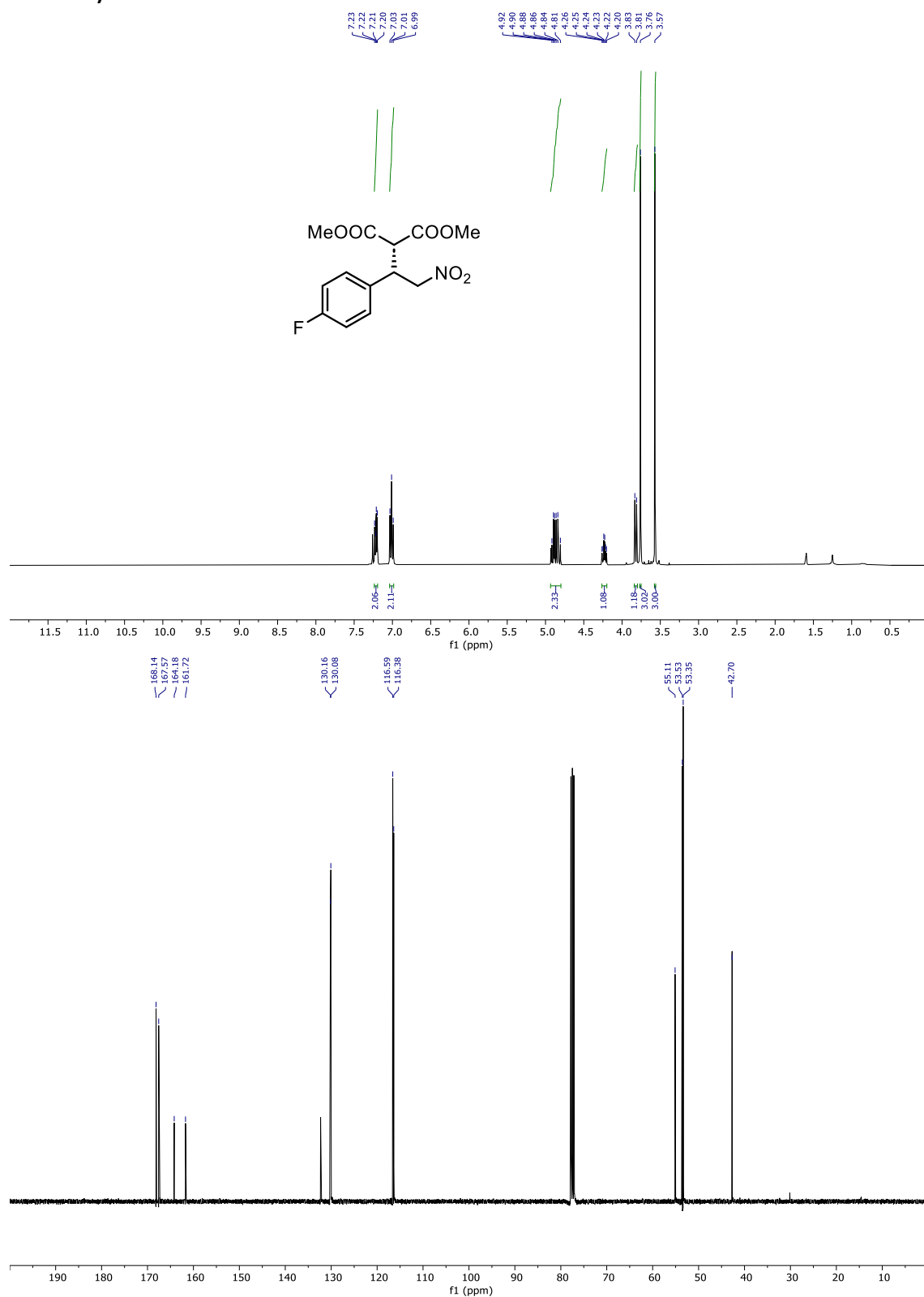


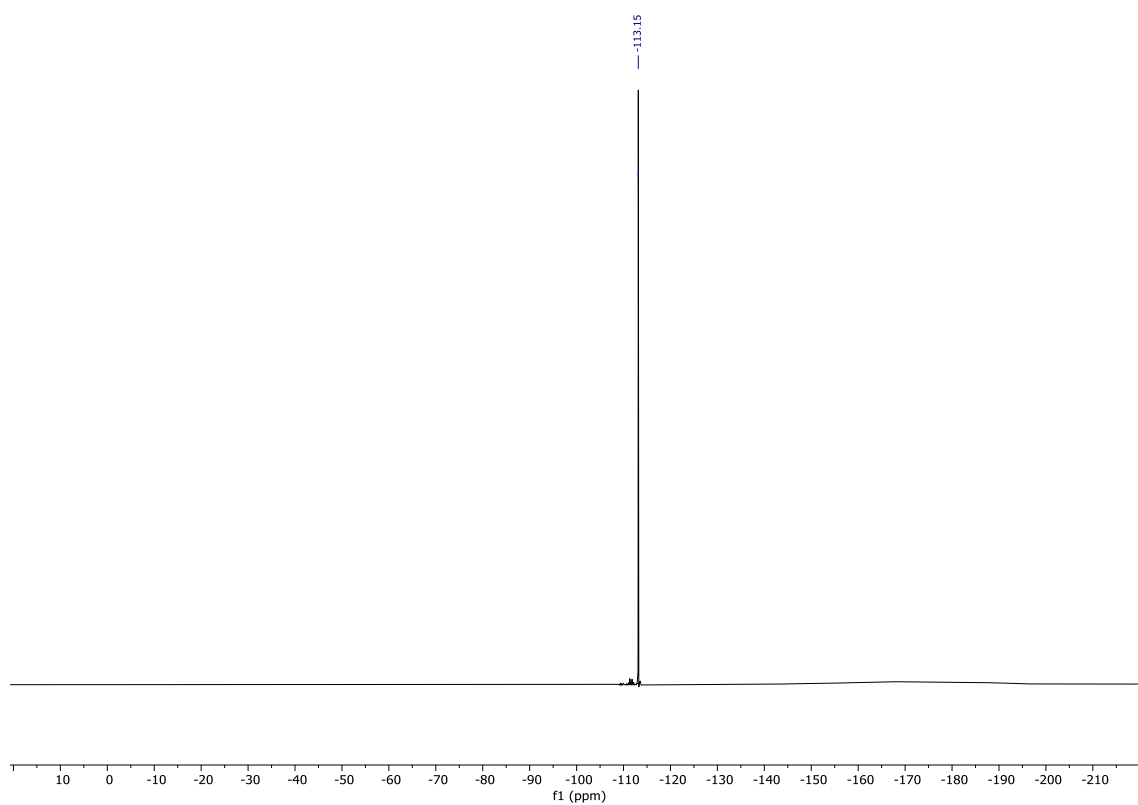
NMR spectra of 6

(S)-dimethyl 2-(2-nitro-1-phenylethyl)malonate (6a) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)

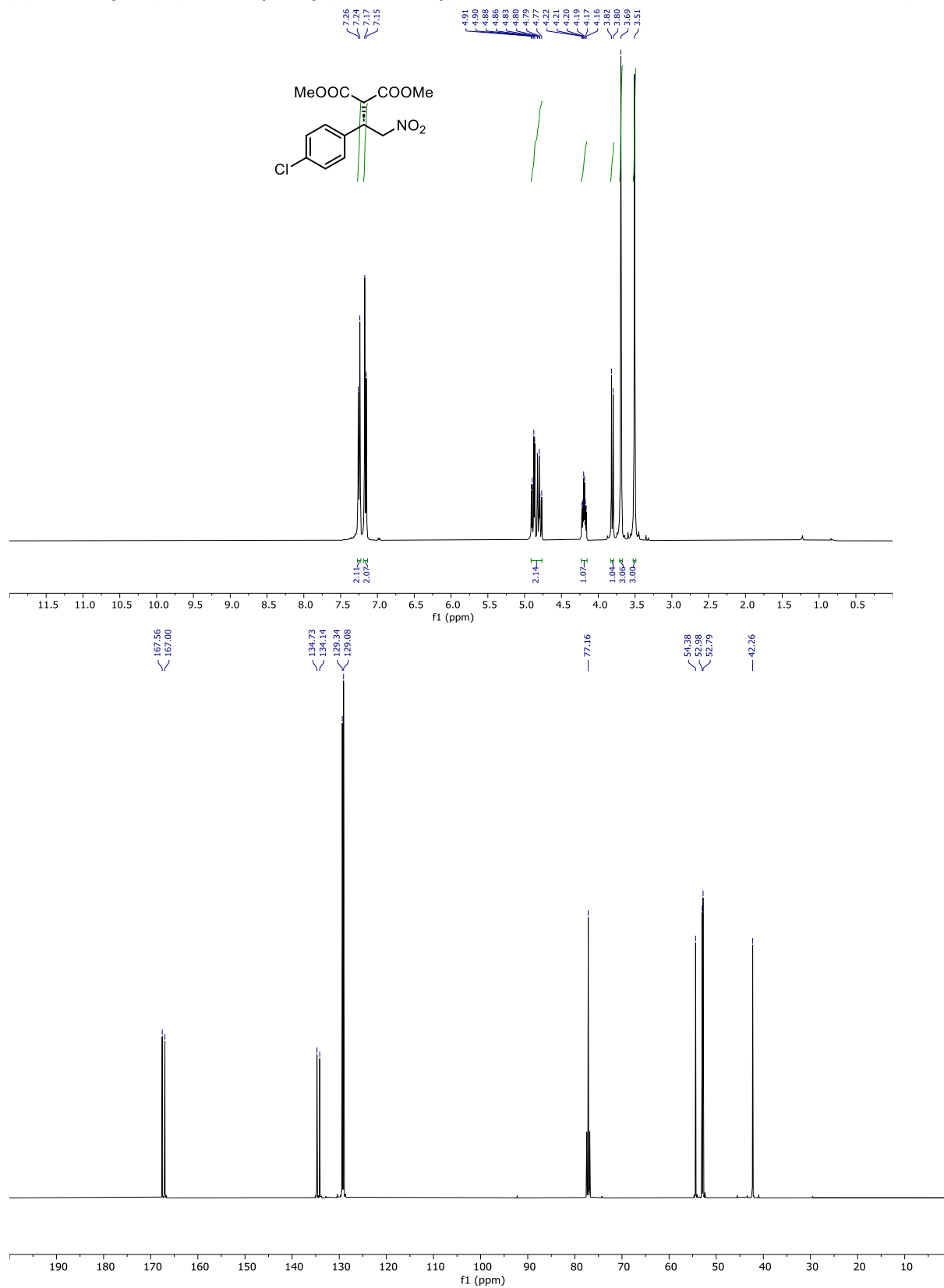


(S)-dimethyl 2-(1-(4-fluorophenyl)-2-nitroethyl)malonate (6b) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz, ¹⁹F 376 MHz)

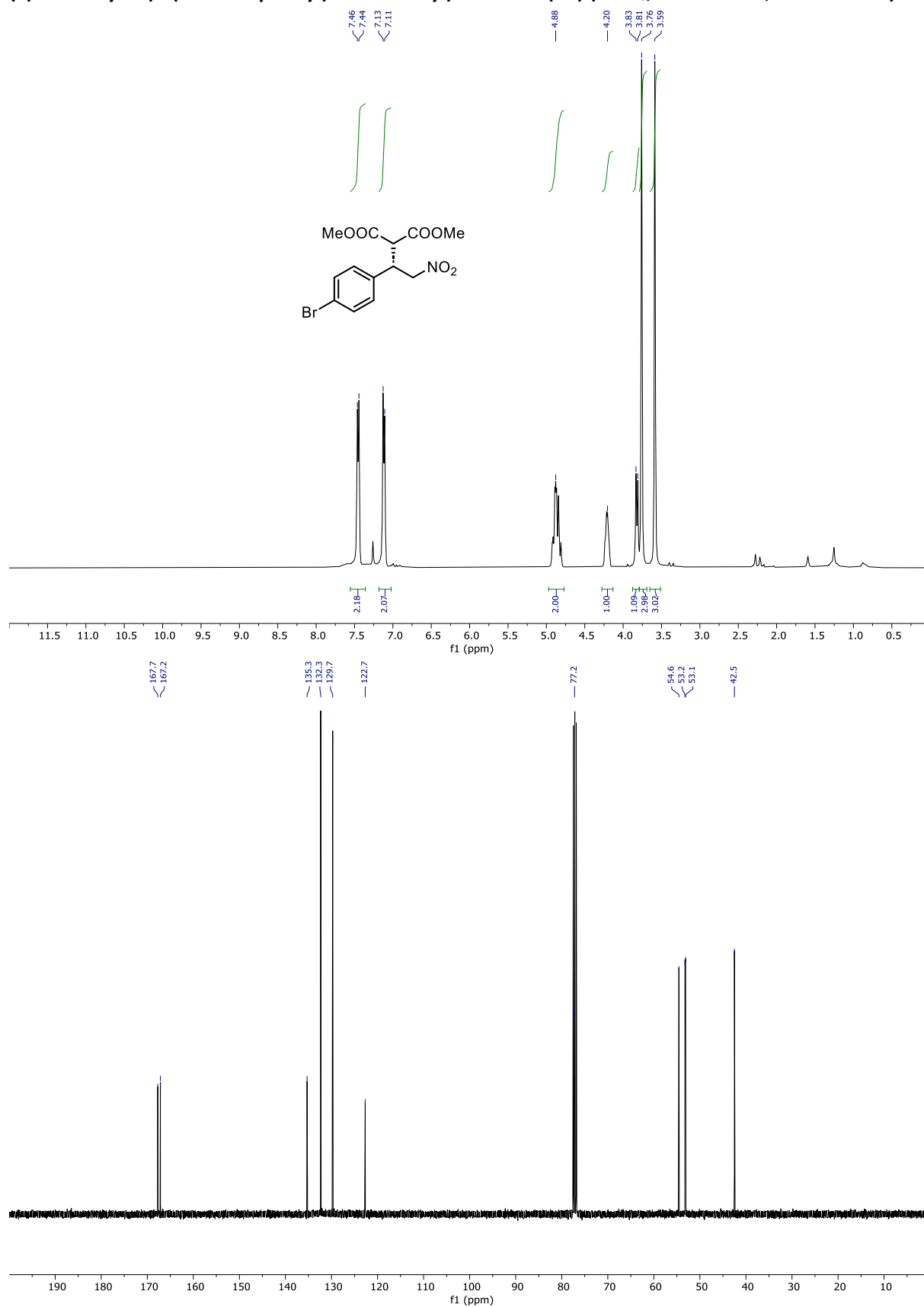




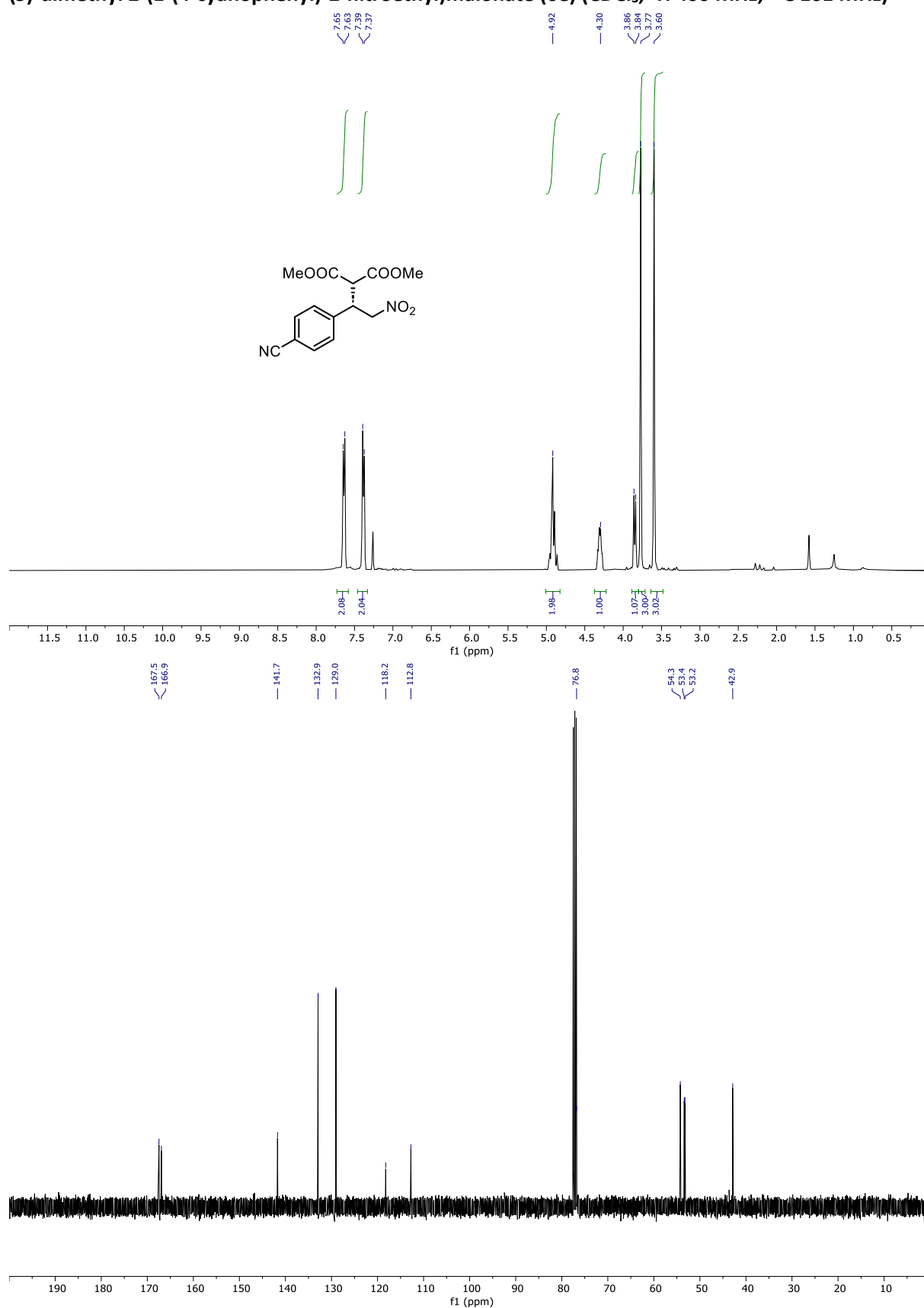
(S)-dimethyl 2-(1-(4-chlorophenyl)-2-nitroethyl)malonate (6c) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



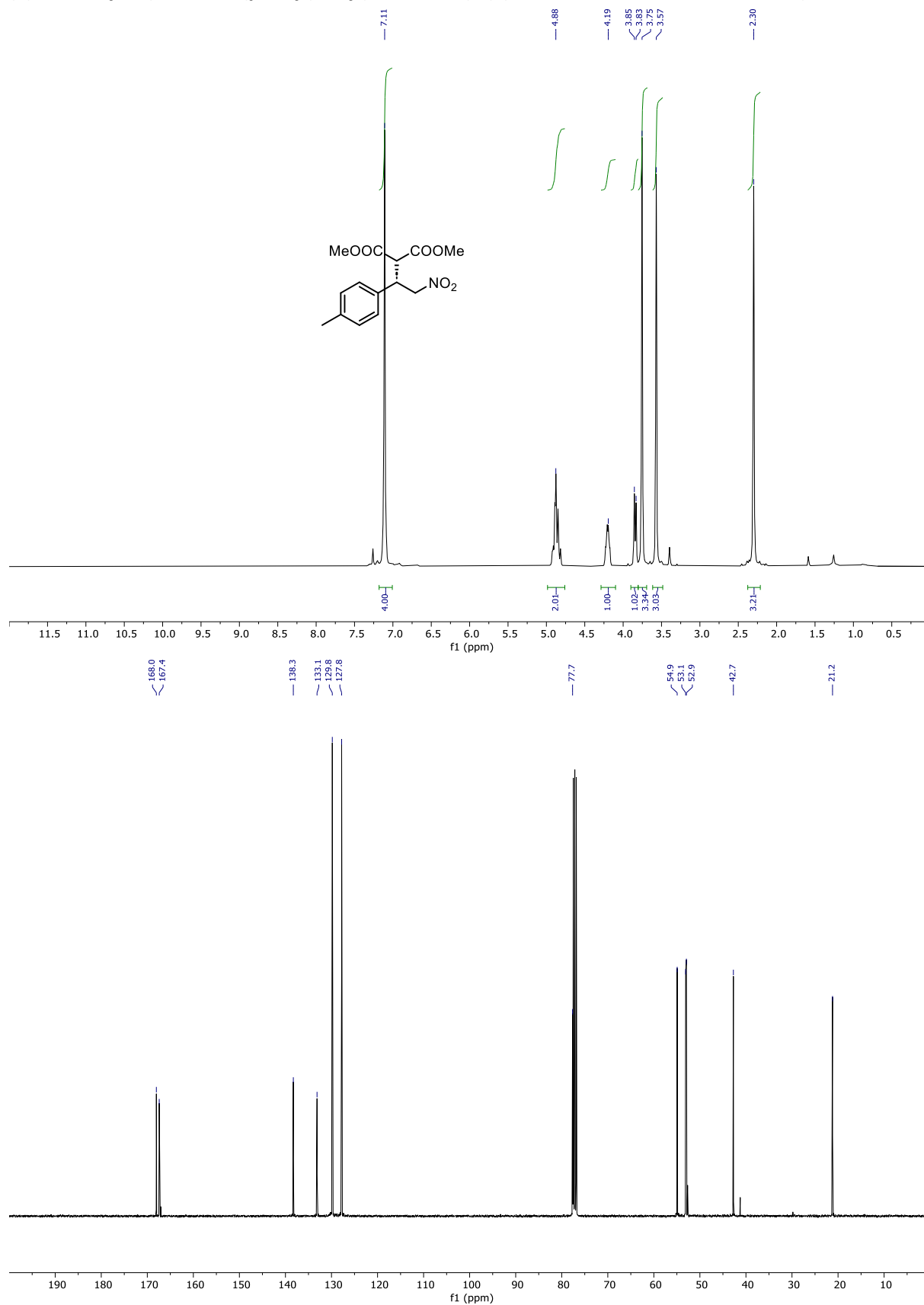
(S)-dimethyl 2-(1-(4-bromophenyl)-2-nitroethyl)malonate (6d) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



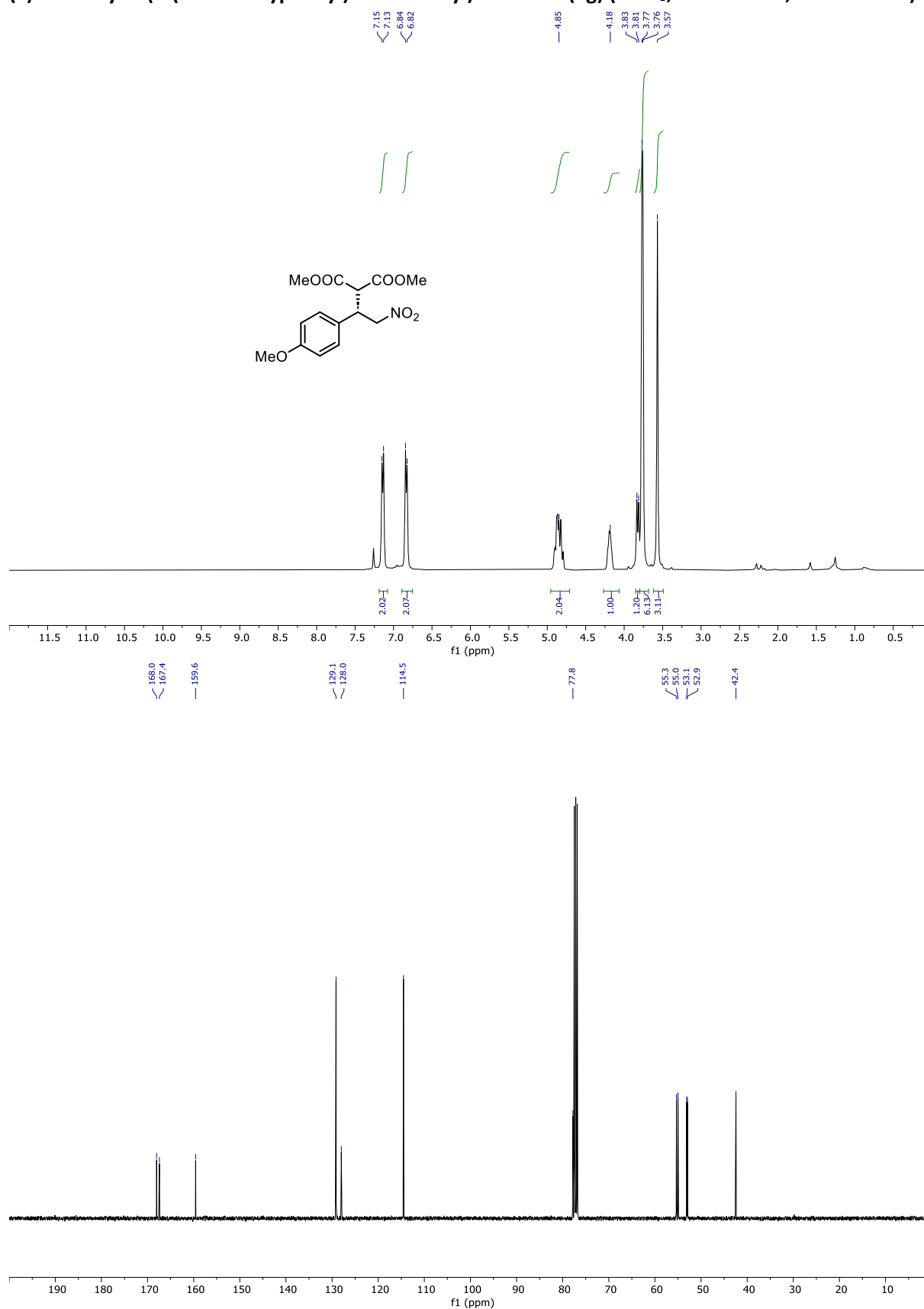
(S)-dimethyl 2-(1-(4-cyanophenyl)-2-nitroethyl)malonate (6e) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



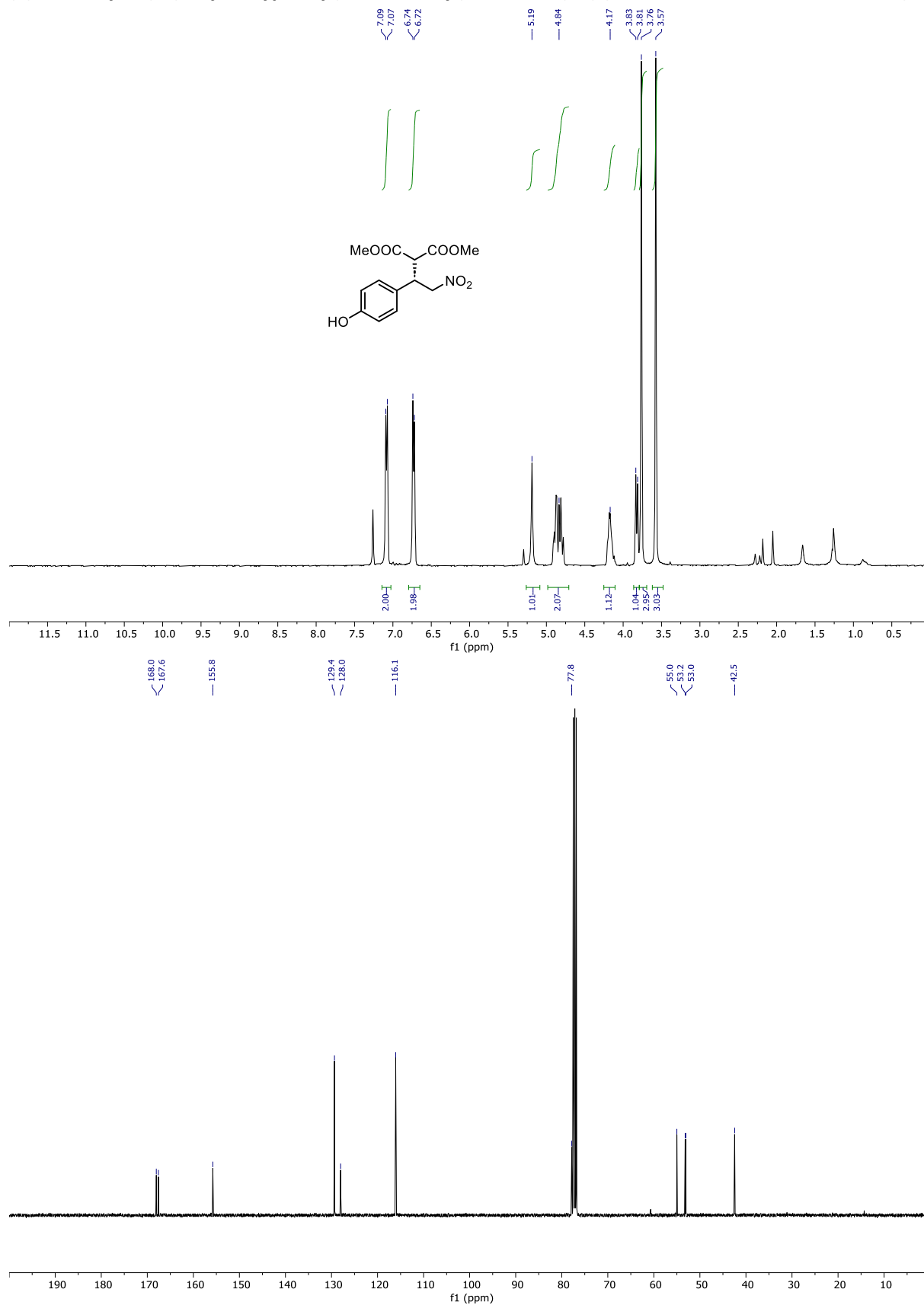
(S)-dimethyl 2-(2-nitro-1-(p-tolyl)ethyl)malonate (6f) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



(S)-dimethyl 2-(1-(4-methoxyphenyl)-2-nitroethyl)malonate (6g) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



(S)-dimethyl 2-(1-(4-hydroxyphenyl)-2-nitroethyl)malonate (6h) (CDCl₃, ¹H 400 MHz, ¹³C 101 MHz)



References

- S1 D. B. G. Williams and M. Lawton, Drying of organic solvents: Quantitative evaluation of the efficiency of several desiccants, *J. Org. Chem.*, 2010, **75**, 8351–8354.
- S2 C. Palomo, M. Oiarbide, A. Laso and R. López, Catalytic enantioselective aza-Henry reaction with broad substrate scope, *J. Am. Chem. Soc.*, 2005, **127**, 17622–17623.
- S3 H. Ishitani, Y. Furiya and S. Kobayashi, Enantioselective Sequential-Flow Synthesis of Baclofen Precursor via Asymmetric 1,4-Addition and Chemoselective Hydrogenation on Platinum/Carbon/Calcium Phosphate Composites, *Chem. Asian J.*, 2020, **15**, 1688–1691.
- S4 S. Del Pozo, S. Vera, M. Oiarbide and C. Palomo, Catalytic Asymmetric Synthesis of Quaternary Barbituric Acids, *J. Am. Chem. Soc.*, 2017, **139**, 15308–15311.
- S5 P. Kasaplar, P. Riente, C. Hartmann and M. A. Pericàs, A polystyrene-supported, highly recyclable squaramide organocatalyst for the enantioselective Michael addition of 1,3-dicarbonyl compounds to β -nitrostyrenes, *Adv. Synth. Catal.*, 2012, **354**, 2905–2910.
- S6 L. Osorio-Planes, C. Rodríguez-Esrich and M. A. Pericàs, Removing the superfluous: A supported squaramide catalyst with a minimalistic linker applied to the enantioselective flow synthesis of pyranonaphthoquinones, *Catal. Sci. Technol.*, 2016, **6**, 4686–4689.
- S7 P. Kasaplar, E. Ozkal, C. Rodríguez-Esrich and M. A. Pericàs, Enantioselective α -amination of 1,3-dicarbonyl compounds in batch and flow with immobilized thiourea organocatalysts, *Green Chem.*, 2015, **17**, 3122–3129.
- S8 L. Huang and W. D. Wulff, Catalytic asymmetric synthesis of trisubstituted aziridines, *J. Am. Chem. Soc.*, 2011, **133**, 8892–8895.
- S9 C. Rampalakos and W. D. Wulff, A novel bis-thiourea organocatalyst for the asymmetric aza-Henry reaction, *Adv. Synth. Catal.*, 2008, **350**, 1785–1790.
- S10 A. G. Wenzel and E. N. Jacobsen, Asymmetric catalytic Mannich reactions catalyzed by urea derivatives: Enantioselective synthesis of β -aryl- β -amino acids, *J. Am. Chem. Soc.*, 2002, **124**, 12964–12965.
- S11 B. Karimi, E. Jafari and D. Enders, Highly efficient catalytic enantioselective mannich reaction of malonates with N-tert-butoxycarbonyl imines by using Yb(OTf)₃/pybox catalysts at room temperature, *Chem. Eur. J.*, 2013, **19**, 10142–10145.
- S12 L. Bernardi, F. Fini, R. P. Herrera, A. Ricci and V. Sgarzani, Enantioselective aza-Henry reaction using cinchona organocatalysts, *Tetrahedron*, 2006, **62**, 375–380.
- S13 X. Jiang, Y. Zhang, L. Wu, G. Zhang, X. Liu, H. Zhang, D. Fu and R. Wang, Doubly stereocontrolled asymmetric aza-henry reaction with in situ generation of n-boc-imines catalyzed by novel rosin-derived amine thiourea catalysts, *Adv. Synth. Catal.*, 2009, **351**, 2096–2100.
- S14 C. J. Wang, X. Q. Dong, Z. H. Zhang, Z. Y. Xue and H. L. Teng, Highly anti-selective asymmetric nitro-mannich reactions catalyzed by bifunctional amine-thiourea-bearing multiple hydrogen-bonding donors, *J. Am. Chem. Soc.*, 2008, **130**, 8606–8607.
- S15 D. Cao, Z. Chai, J. Zhang, Z. Ye, H. Xiao, H. Wang, J. Chen, X. Wu and G. Zhao, Thiourea-phosphonium salts from amino acids: Cooperative phase-transfer catalysts in the enantioselective aza-Henry reaction, *Chem. Commun.*, 2013, **49**, 5972–5974.
- S16 F. Fini, V. Sgarzani, D. Pettersen, R. P. Herrera, L. Bernardi and A. Ricci, Phase-transfer-catalyzed asymmetric aza-Henry reaction using N-carbamoyl imines generated in situ from α -amido

- sulfones, *Angew. Chem. Int. Ed.*, 2005, **44**, 7975–7978.
- S17 H. Ishitani, K. Kanai and S. Kobayashi, Continuous-Flow Enantioselective 1,4-Addition Reactions of Malonates with Nitroolefins on Ni-Supported Mesoporous Silica Materials with Co-feeding of a Chiral Ligand, *Adv. Synth. Catal.*, 2023, **365**, 1526–1530.
- S18 Y. Ren, M. Wang, Q. Yang and J. Zhu, Merging Chiral Diamine and Ni/SiO₂ for Heterogeneous Asymmetric 1,4-Addition Reactions, *ACS Catal.*, 2023, **13**, 1974–1982.
- S19 D. Bécart, V. Diemer, A. Salaün, M. Oiarbide, Y. R. Nelli, B. Kauffmann, L. Fischer, C. Palomo and G. Guichard, Helical Oligoureia Foldamers as Powerful Hydrogen Bonding Catalysts for Enantioselective C-C Bond-Forming Reactions, *J. Am. Chem. Soc.*, 2017, **139**, 12524–12532.
- S20 F. X. Chen, C. Shao, Q. Wang, P. Gong, D. Y. Zhang, B. Z. Zhang and R. Wang, An enantioselective Michael addition of malonate to nitroalkenes catalyzed by low loading demethylquinine salts in water, *Tetrahedron Lett.*, 2007, **48**, 8456–8459.