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Supporting Information

Deoxygenative Functionalization of Trifluoromethyl Ketones

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Table of Contents

1.	General	S2-S3
2.	Optimization of Reaction Conditions	S4-S5
3.	Synthesis of Trifluoromethyl Phosphinate 2	S6-S7
4.	Deoxygenative Functionalization of Trifluoromethylketones	S8-S17
5.	Deoxygenative Arylation of Trifluoromethylketones	S18-S20
6.	Application	S21-S24
7.	References	S25-S26
8.	¹ H, ¹³ C, ¹⁹ F, and ³¹ P NMR Spectra	S27-S132

1. General

Unless otherwise noted, all reactants or reagents including drying solvents were obtained from commercial suppliers and used as received. Diphenylphosphine oxide was purchased from BLD pharm. 1,8-Diazabicyclo[5.4.0]-7-undecene (DBU), magnesium turnings, mesitylene, tetrakis(triphenylphosphine)palladium(0), and thiophenol were purchased from KANTO Chemical. Lithium bis(trimethylsilyl)amide(1 M in toluene solution) and trimethylsilyl trifluoromethanesulfonate (TMSOTf), were purchased from Sigma-Aldrich (Note: TMSOTf was found to lose activity upon prolonged storage; therefore, freshly opened reagent was used in all experiments.). Allyltrimethylsilane, benzoic acid, diethyl phosphite, 4,4'-di-tert-butyl-2,2'-bipyridyl, methallyltrimethylsilane, N-methyl-ptoluenesulfonamide, nickel(II) bromide ethylene glycol dimethyl ether complex, phenyl triflimide, sodium formate, p-toluenethiol, thiobenzoic acid, thiophene, trifluoromethanesulfonic acid, 1-(trimethylsilyloxy)cyclohexene, 2,2,2-trifluoro-1-(4-methoxyphenyl)ethan-1-one (1A), 2,2,2-trifluoro-1-phenylethan-1-one (1B), 2,2,2-trifluoro-1-(p-tolyl)ethan-1-one (1C), 1-(4-chlorophenyl)-2,2,2trifluoroethan-1-one (1G) were purchased from Tokyo Chemical Industry (TCI). Zinc cyanide was purchased from FUJIFILM Wako Pure Chemical Corporation. 1-(4-(tert-butyl)phenyl)-2,2,2-(**1D**),^[1] 1-(3,5-dimethylphenyl)-2,2,2-trifluoroethan-1-one (**1E**),^[2] 1-(4trifluoroethan-1-one bromophenyl)-2,2,2-trifluoroethan-1-one (1F),[3] 2,2,2-trifluoro-1-(4-morpholinophenyl)ethan-1-one (1H), [1] 1-(benzo[d][1,3]dioxol-5-yl)-2,2,2-trifluoroethan-1-one (1I), [1] 1-([1,1'-biphenyl]-4-yl)-2,2,2-(1J),^[1] 2,2,2-trifluoro-1-(naphthalen-2-yl)ethan-1-one trifluoroethan-1-one $(benzo[b]thiophen-2-yl)-2,2,2-trifluoroethan-1-one (1L),^{[1]} 2,2,3,3,3-pentafluoro-1-phenylpropan-1$ one (1U), [4] 2,2-difluoro-1-phenylethan-1-one (1V), [1] 2,2,2-trichloro-1-phenylethan-1-one (1W), [5] and 2,2,2-tribromo-1-phenylethan-1-one $(1X)^{[6]}$ were synthesized according to procedures and the spectra matched with those of compounds reported in the literature. Unless otherwise noted, all reactions were performed with drying solvents under an atmosphere of N₂ in dried glassware using standard vacuumline techniques. All reactions were performed in 8-mL glass vessel tubes equipped with a screw cap and heated (IKA Plate RCT digital) in a 16-well aluminum reaction block (IKA DB4.3 Block) unless otherwise noted. All work-up and purification procedures were carried out with reagent-grade solvents under air unless otherwise noted.

Analytical thin-layer chromatography (TLC) was performed using Silica-gel 70 TLC Plate-Wako (0.25 mm). The developed chromatogram was analyzed by UV lamp (254 nm). Flash column chromatography was performed with Biotage Isolera® equipped with Biotage Sfär Cartridge Silica D columns. Preparative thin-layer chromatography (PTLC) was performed using Wakogel B5-F silica coated plates (0.75 mm) prepared in our laboratory. High-resolution mass spectra (HRMS) were conducted on Thermo Fisher Scientific ExactivePlus Orbitrap (ESI) and Bruker Compact QTOF (APCI). Nuclear magnetic resonance (NMR) spectra were recorded on a JEOL JNM-ECS-400 (¹H 400 MHz, ¹³C 101 MHz, ³¹P 162 MHz, ¹³F 376 MHz) and JEOL JNM-ECZ-400 (¹H 400 MHz, ¹³C 101 MHz, ³¹P 162 MHz, ¹°F 376 MHz). Chemical shifts for ¹H NMR are expressed in parts per million (ppm) relative

to tetramethylsilane (δ 0.00 ppm). Chemical shifts for $^{13}C\{^1H\}$ NMR are expressed in ppm relative to CDCl₃ (δ 77.0 ppm). Chemical shifts for ^{31}P NMR are expressed in ppm relative to H₃PO₄ (δ 0.00 ppm) as an external standard. Chemical shifts for ^{19}F NMR are expressed in ppm relative to fluorobenzene (δ –113.15 ppm) as an internal standard. Preparative recycling gel permeation chromatography (GPC) was performed with a JAI LaboACE LC-5060 instrument equipped with JAIGEL-2HR columns using CHCl₃ as an eluent.

Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, dt = doublet of triplets, tt = triplet of triplets, tq = triplet of quartets, tq = triplet of quartets, tq = triplet of doublets, tq = triplet of doublets of triplets of doublets, tq = triplet of doublets of doublets of doublets of doublets of doublets, tq = triplet of doublets of doublets of doublets, tq = triplet of dou

2. Optimization of Reaction Conditions

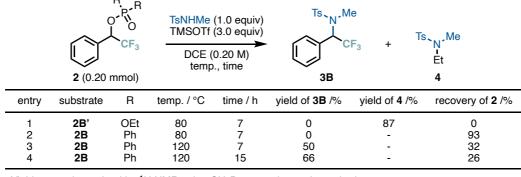
2.1. Pudovik Addition/Phospha-Brook Rearrangement of 1A

Yields were determined by ¹H NMR using CH₂Br₂ as an internal standard.

2.2. Benzylic Substitution of 2

Yields were determined by ¹H NMR using CH₂Br₂ as an internal standard.

2.3. Re-examination with 3B



Yields were determined by ¹H NMR using CH₂Br₂ as an internal standard.

2.4. Electron-deficient Trifluoromethyl Ketones

2.5. Other Fluorinated or Halogenated Ketones

3. Synthesis of Trifluoromethyl Phosphinate 2

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N₂ gas after cooling to room temperature. To this tube was added phosphine oxide (1.0 equiv). The tube was placed under vacuum and refilled three times with N₂ gas, and to this tube were added trifluoromethylketone 1 (1.0 equiv), DBU (10 mol%), and 1,2-dichloroethane (0.20 M). The vessel was sealed with a screw cap and then heated at 80 °C for 2 h while its contents were being stirred. After the reaction mixture had been cooled to room temperature, the reaction mixture was concentrated *in vacuo*. The residue was purified by Isolera® to afford the corresponding product 2.

2,2,2-Trifluoro-1-(4-methoxyphenyl)ethyl Diphenylphosphinate (2A)

Reaction was conducted in 4 h. Purification by Isolera® (9:1 to 3:2 hexane/EtOAc) afforded **2A** as a colorless oil (0.50 mmol scale: 189.5 mg, 93% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.86–7.80 (m, 2H), 7.63–7.55 (m, 3H), 7.52–7.41 (m, 3H), 7.34–7.28 (m, 4H), 6.81 (d, J = 8.8 Hz, 2H), 5.65 (dq, J_{H-P} = 10.8 Hz, J_{H-F} = 6.4 Hz, 1H), 3.79 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 160.6, 132.5 (d, J_{C-P} = 2.9 Hz), 132.3 (d, J_{C-P} = 2.9 Hz), 131.5 (d, J_{C-P} = 10.7 Hz), 130.5 (d, J_{C-P} = 136.2 Hz), 130.4 (d, J_{C-P} = 138.1 Hz), 129.5, 128.5 (d, J_{C-P} = 13.6 Hz), 128.2 (d, J_{C-P} = 13.5 Hz), 123.5, 123.2 (qd, J_{C-F} = 281.2 Hz, J_{C-P} = 8.3 Hz), 113.8, 73.3 (qd, J_{C-F} = 33.8 Hz, J_{C-P} = 5.0 Hz), 55.1 (one peak is missing due to overlapping); ³¹P NMR (162 MHz, CDCl₃) δ 34.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –76.7 (d, J_{F-H} = 6.4 Hz); HRMS (ESI) m/z [M+H]⁺ Calcd for C₂₁H₁₉F₃O₃P 407.1018; Found 407.1008.

Diethyl (2,2,2-trifluoro-1-(4-methoxyphenyl)ethyl) Phosphate (2A')

Purification by Isolera[®] (9:1 to 3:2 hexane/EtOAc) afforded **2A'** as a colorless oil (0.50 mmol scale: 168.9 mg, 98% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, J = 8.8 Hz, 2H), 6.93 (d, J = 8.8 Hz, 2H), 5.54 (dq, J_{H-P} = 10.0 Hz, J_{H-F} = 6.4 Hz, 1H), 4.22–4.04 (m, 2H), 3.98–3.84 (m, 2H), 3.83 (s, 3H), 1.31

(td, J = 7.2 Hz, $J_{H-P} = 1.2$ Hz, 3H), 1.15 (td, J = 7.2 Hz, $J_{H-P} = 1.2$ Hz, 3H); $^{13}C\{^{1}H\}$ NMR (101 MHz, CDCl₃) δ 160.8, 129.4, 123.3, 123.0 (qd, $J_{C-F} = 280.8$ Hz, $J_{C-P} = 10.8$ Hz), 113.9, 75.8 (qd, $J_{C-F} = 34.1$ Hz, $J_{C-P} = 4.5$ Hz), 64.3 (d, $J_{C-P} = 6.0$ Hz), 64.1 (d, $J_{C-P} = 5.7$ Hz), 55.1, 15.7 (d, $J_{C-P} = 7.5$ Hz), 15.6 (d, $J_{C-P} = 7.5$ Hz); ^{31}P NMR (162 MHz, CDCl₃) δ –2.5; ^{19}F NMR (376 MHz, CDCl₃) δ –77.4 (d, $J_{F-H} = 6.4$ Hz); HRMS (ESI) m/z [M+H]⁺ Calcd for $C_{13}H_{19}F_{3}O_{5}P$ 343.0917; Found 343.0919.

2,2,2-Trifluoro-1-phenylethyl Diphenylphosphinate (2B)^[7]

Purification by Isolera® (9:1 to 3:2 hexane/EtOAc) afforded **2B** as a colorless oil (1.0 mmol scale: 350.4 mg, 93% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.88–7.81 (m, 2H), 7.63–7.58 (m, 3H), 7.53–7.48 (m, 2H), 7.44–7.38 (m, 3H), 7.35–7.27 (m, 5H), 5.71 (dq, $J_{\text{H-P}}$ = 10.8 Hz, $J_{\text{H-F}}$ = 6.4 Hz, 1H); 13 C { 1 H} NMR (101 MHz, CDCl₃) δ 132.7 (d, $J_{\text{C-P}}$ = 2.9 Hz), 132.4 (d, $J_{\text{C-P}}$ = 2.9 Hz), 131.6 (d, $J_{\text{C-P}}$ = 10.6 Hz), 131.5 (d, $J_{\text{C-P}}$ = 10.6 Hz), 130.4 (d, $J_{\text{C-P}}$ = 136.6 Hz), 130.3 (d, $J_{\text{C-P}}$ = 138.2 Hz), 129.8, 128.6 (d, $J_{\text{C-P}}$ = 13.6 Hz), 128.4, 128.3 (d, $J_{\text{C-P}}$ = 13.6 Hz), 128.1, 123.1 (qd, $J_{\text{C-F}}$ = 281.5 Hz, $J_{\text{C-P}}$ = 8.0 Hz), 73.5 (qd, $J_{\text{C-F}}$ = 33.7 Hz, $J_{\text{C-P}}$ = 4.8 Hz) (one peak is missing due to overlapping); 31 P NMR (162 MHz, CDCl₃) δ 34.8; 19 F NMR (376 MHz, CDCl₃) δ –76.6 (d, $J_{\text{F-H}}$ = 6.4 Hz); HRMS (ESI) m/z [M+H]⁺ Calcd for C₂₀H₁₇F₃O₂P 377.0913; Found 377.0917. The spectra matched with those of this compound reported in the literature.[7]

Diethyl (2,2,2-trifluoro-1-phenylethyl) Phosphate (2B')[8]

Purification by Isolera® (9:1 to 3:2 hexane/EtOAc) afforded **2B'** as a colorless oil (0.50 mmol scale: 120.3 mg, 77% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.52–7.47 (m, 2H), 7.45–7.39 (m, 3H), 5.59 (dq, $J_{\text{H-P}} = 10.4$ Hz, $J_{\text{H-F}} = 6.4$ Hz, 1H), 4.22–4.04 (m, 2H), 3.99–3.82 (m, 2H), 1.31 (td, J = 7.2 Hz, $J_{\text{H-P}} = 1.2$ Hz, 3H), 1.15 (td, J = 7.2 Hz, $J_{\text{H-P}} = 1.2$ Hz, 3H); 13 C{ 1 H} NMR (101 MHz, CDCl₃) δ 131.4, 130.1, 128.6, 127.9, 122.9 (qd, $J_{\text{C-F}} = 280.9$ Hz, $J_{\text{C-P}} = 9.8$ Hz), 76.1 (qd, $J_{\text{C-F}} = 34.0$ Hz, $J_{\text{C-P}} = 4.4$ Hz), 64.4 (d, $J_{\text{H-P}} = 5.9$ Hz), 64.2 (d, $J_{\text{H-P}} = 5.8$ Hz), 15.8 (d, $J_{\text{H-P}} = 7.4$ Hz), 15.7 (d, $J_{\text{H-P}} = 7.1$ Hz); 31 P NMR (162 MHz, CDCl₃) δ –2.5; 19 F NMR (376 MHz, CDCl₃) δ –77.3 (d, $J_{\text{F-H}} = 6.4$ Hz); HRMS (ESI) m/z [M+H] $^+$ Calcd for C₁₂H₁₇F₃O₄P 313.0811; Found 313.0804. The spectra matched with those of this compound reported in the literature. $^{[8]}$

4. Deoxygenative Functionalization of Trifluoromethylketones

General Procedure A

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N₂ gas after cooling to room temperature. To this tube was added diphenylphosphine oxide (40.4 mg, 0.20 mmol, 1.0 equiv). The tube was placed under vacuum and refilled three times with N₂ gas, and to this tube were added trifluoromethylketone 1 (0.20 mmol, 1.0 equiv), DBU (3 μL, 20 μmol, 10 mol%), and 1,2-dichloroethane (DCE: 1.0 mL). The vessel was sealed with a screw cap and then heated at 80 °C for 2 h while its contents were being stirred. After cooling to room temperature, to this tube was added nucleophile (0.20 mmol, 1.0 equiv). The mixture was stirred at 0 °C, and to this tube was added TMSOTf (108 μL, 0.60 mmol, 3.0 equiv) slowly. The vessel was sealed with a screw cap and then heated at 120 °C for 15 h while its contents were being stirred. After the reaction mixture had been cooled to room temperature, the reaction was quenched by saturated NaHCO₃ aq. The mixture was extracted three times with CH₂Cl₂. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by PTLC to afford the corresponding product 3.

General Procedure B

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N_2 gas after cooling to room temperature. The tube was placed under vacuum and refilled three times with N_2 gas, and to this tube were added trifluoromethylketone 1 (0.20 mmol, 1.0 equiv), diethyl phosphite (26 μ L, 0.20 mmol, 1.0 equiv), DBU (3 μ L, 20 μ mol, 10 mol%), and chloroform (1.0 mL). The vessel was sealed with a screw cap and then heated at 80 °C for 2 h while its contents were being stirred. After cooling to room temperature, to this tube was added nucleophile (0.20 mmol, 1.0 equiv). The mixture was stirred at 0 °C, and to this tube was added TMSOTf (108 μ L, 0.60 mmol, 3.0 equiv) slowly. The vessel was sealed with a screw cap and then heated at 80 °C for 7 h while its contents were being stirred. After the reaction mixture had been cooled to room temperature, the reaction was quenched by saturated NaHCO₃ aq. The mixture was extracted three times with CH₂Cl₂. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by PTLC to afford the corresponding product 3.

N,4-Dimethyl-N-(2,2,2-trifluoro-1-(4-methoxyphenyl)ethyl)benzenesulfonamide (3A)

The reaction was conducted according to **General Procedure B**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3A** as a colorless oil (58.4 mg, 78% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.73 (d, J = 8.4 Hz, 2H), 7.34–7.29 (m, 4H), 6.89 (d, J = 8.4 Hz, 2H), 5.81 (q, J_{H-F} = 8.4 Hz, 1H), 3.81 (s, 3H), 2.66 (s, 3H), 2.44 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 160.0, 143.8, 136.1, 130.1, 129.6, 127.5, 125.0 (q, J_{C-F} = 284.3 Hz), 122.3, 114.2, 59.9 (q, J_{C-F} = 31.2 Hz), 55.3, 30.5, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ – 68.4 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₇H₁₈F₃NNaO₂S 396.0852; Found 396.0850.

N,4-Dimethyl-N-(2,2,2-trifluoro-1-phenylethyl)benzenesulfonamide (3B)

N,4-Dimethyl-N-(2,2,2-trifluoro-1-(p-tolyl)ethyl)benzenesulfonamide (3C)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3C** as a colorless oil (38.3 mg, 53% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.74 (d, J = 8.0 Hz, 2H), 7.32–7.26 (m, 4H), 7.18 (d, J = 8.0 Hz, 2H), 5.82 (q, J_{H-F} = 8.4 Hz, 1H), 2.65 (s, 3H), 2.44 (s, 3H), 2.35 (s, 3H); 13 C{ 1 H} NMR (101 MHz, CDCl₃) δ 143.8, 139.1, 136.1, 129.61, 129.55, 128.6, 127.5, 127.4, 125.0 (q, J_{C-F} = 284.5 Hz), 60.2 (q, J_{C-F} = 30.8 Hz), 30.5, 21.5, 21.0; 19 F NMR (376 MHz, CDCl₃)

 δ –68.2 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for $C_{17}H_{18}F_3NNaO_2S$ 380.0903; Found 380.0906.

N-(1-(4-(tert-Butyl)phenyl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3D)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3D** as a white solid (71.1 mg, 88% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.74 (d, J = 8.4 Hz, 2H), 7.38 (d, J = 8.4 Hz, 2H), 7.34–7.29 (m, 4H), 5.84 (q, J_{H-F} = 8.4 Hz, 1H), 2.67 (s, 3H), 2.44 (s, 3H), 1.31 (s, 9H); ¹³C { ¹H } NMR (101 MHz, CDCl₃) δ 152.1, 143.8, 136.1, 129.6, 128.4, 127.6, 127.4, 125.8, 125.0 (q, J_{C-F} = 284.4 Hz), 60.2 (q, J_{C-F} = 31.1 Hz), 34.6, 31.1, 30.6, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –68.0 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₂₀H₂₄F₃NNaO₂S 422.1372; Found 422.1372.

N-(1-(3,5-Dimethylphenyl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3E)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3E** as a colorless oil (32.7 mg, 44% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.74 (d, J = 8.4 Hz, 2H), 7.31 (d, J = 8.4 Hz, 2H), 6.98 (s, 1H), 6.91 (s, 2H), 5.73 (q, J_{H-F} = 8.4 Hz, 1H), 2.68 (s, 3H), 2.44 (s, 3H), 2.28 (s, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 143.8, 138.4, 136.2, 130.7, 130.1, 129.6, 127.6, 126.4, 125.0 (q, J_{C-F} = 284.5 Hz), 60.5 (q, J_{C-F} = 31.0 Hz), 30.8, 21.5, 21.3; ¹⁹F NMR (376 MHz, CDCl₃) δ -68.0 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₈H₂₀F₃NNaO₂S 394.1059; Found 394.1048.

N-(1-(4-Bromophenyl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3F)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3E** as a white solid (62.3 mg, 73% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.73 (d, J = 8.4 Hz,

2H), 7.52 (d, J = 8.4 Hz, 2H), 7.32 (d, J = 8.4 Hz, 2H), 7.29 (d, J = 8.4 Hz, 2H), 5.83 (q, $J_{\text{H-F}}$ = 8.4 Hz, 1H), 2.66 (s, 3H), 2.45 (s, 3H); $^{13}\text{C}\{^{1}\text{H}\}$ NMR (101 MHz, CDCl₃) δ 144.1, 135.8, 132.2, 130.2, 129.7, 129.6, 127.5, 124.6 (q, $J_{\text{C-F}}$ = 284.5 Hz), 123.5, 59.9 (q, $J_{\text{C-F}}$ = 31.2 Hz), 30.5, 21.5; ^{19}F NMR (376 MHz, CDCl₃) δ -68.0 (d, $J_{\text{F-H}}$ = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₆H₁₅BrF₃NNaO₂S 443.9851; Found 443.9847.

N-(1-(4-Chlorophenyl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3G)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3G** as a colorless oil (46.3 mg, 61% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.73 (d, J = 8.4 Hz, 2H), 7.38–7.35 (m, 4H), 7.32 (d, J = 8.4 Hz, 2H), 5.84 (q, J_{H-F} = 8.4 Hz, 1H), 2.66 (s, 3H), 2.44 (s, 3H); 13 C{ 1 H} NMR (101 MHz, CDCl₃) δ 144.0, 135.8, 135.3, 130.0, 129.7, 129.2, 129.1, 127.5, 124.7 (q, J_{C-F} = 284.4 Hz), 59.8 (q, J_{C-F} = 31.4 Hz), 30.5, 21.5; 19 F NMR (376 MHz, CDCl₃) δ –68.0; HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₆H₁₅ClF₃NNaO₂S 400.0356; Found 400.0354.

N,4-Dimethyl-N-(2,2,2-trifluoro-1-(4-morpholinophenyl)ethyl)benzenesulfonamide (3H)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) and GPC afforded **3H** as a white solid (41.4 mg, 48% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.74 (d, J = 8.4 Hz, 2H), 7.32 (d, J = 8.0 Hz, 2H), 7.28 (d, J = 8.0 Hz, 2H), 6.87 (d, J = 8.4 Hz, 2H), 5.79 (q, $J_{\text{H-F}}$ = 8.4 Hz, 1H), 3.88–3.83 (m, 4H), 3.20–3.16 (m, 4H), 2.65 (s, 3H), 2.43 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 151.3, 143.7, 136.1, 129.8, 129.6, 127.5, 125.0 (q, $J_{\text{C-F}}$ = 284.3 Hz), 120.8, 115.1, 66.7, 59.9 (q, $J_{\text{C-F}}$ = 31.1 Hz), 48.4, 30.5, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –68.4 (d, $J_{\text{F-H}}$ = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₂₀H₂₃F₃N₂NaO₃S 451.1274; Found 451.1270.

N-(1-(Benzo[d][1,3]dioxol-5-yl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3I)

The reaction was conducted according to **General Procedure B**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3I** as a white solid (77.4 mg, 99% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.73 (d, J = 8.4 Hz, 2H), 7.31 (d, J = 8.4 Hz, 2H), 6.88 (d, J = 8.8 Hz, 1H), 6.87 (s, 1H), 6.79 (d, J = 8.8 Hz, 1H), 5.99 (s, 2H), 5.77 (q, J_{H-F} = 8.4 Hz, 1H), 2.68 (s, 3H), 2.44 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 148.2, 148.1, 143.9, 136.0, 129.6, 127.5, 124.8 (q, J_{C-F} = 284.7 Hz), 123.9, 122.8, 109.0, 108.5, 101.5, 60.1 (q, J_{C-F} = 31.1 Hz), 30.5, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ -68.3 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₇H₁₆F₃NNaO₄S 410.644; Found 410.0650.

N-(1-([1,1'-Biphenyl]-4-yl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3J)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3J** as a white solid (41.0 mg, 49% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, J = 8.4 Hz, 2H), 7.60 (d, J = 8.0 Hz, 2H), 7.58 (d, J = 8.4 Hz, 2H), 7.49–7.43 (m, 4H), 7.38 (t, J = 7.2 Hz, 1H), 7.32 (d, J = 8.4 Hz, 2H), 5.91 (q, J_{H-F} = 8.4 Hz, 1H), 2.71 (s, 3H), 2.44 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 143.9, 141.9, 139.9, 136.0, 129.7, 129.4, 129.1, 128.9, 127.8, 127.6, 127.5, 127.1, 124.9 (q, J_{C-F} = 284.5 Hz), 60.2 (q, J_{C-F} = 31.2 Hz), 30.6, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –68.0 (d, J_{F-H} = 8.4 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₂₂H₂₀F₃NNaO₂S 442.1059; Found 442.1055.

N,4-Dimethyl-N-(2,2,2-trifluoro-1-(naphthalen-2-yl)ethyl)benzenesulfonamide (3K)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3K** as a white solid (31.4 mg, 39% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.88–7.80 (m, 4H), 7.77 (d, J = 8.4 Hz, 2H), 7.55–7.51 (m, 2H), 7.50–7.46 (m, 1H), 7.32 (d, J = 8.0 Hz, 2H), 6.02 (q, $J_{\text{H-F}} = 8.4$ Hz, 1H), 2.69 (s, 3H), 2.44 (s, 3H); ¹³C { ¹H} NMR (101 MHz, CDCl₃) δ 143.9, 136.1, 133.1, 132.9, 129.7, 128.9, 128.3, 128.2, 127.7, 127.6, 127.1, 126.7, 125.7, 125.0 (q, $J_{\text{C-F}} = 284.7$ Hz), 60.6 (q, $J_{\text{C-F}} = 31.2$ Hz), 30.7, 21.5 (one peak is missing due to overlapping); ¹⁹F NMR (376 MHz, CDCl₃) δ –67.8 (d, $J_{\text{F-H}} = 8.4$ Hz).; HRMS (ESI) m/z [M+Na]⁺ Calcd for C₂₀H₁₈F₃NNaO₂S 416.0903; Found 416.0910.

N-(1-(Benzo[b]thiophen-2-yl)-2,2,2-trifluoroethyl)-N,4-dimethylbenzenesulfonamide (3L)

The reaction was conducted according to **General Procedure A**, *N*-methyl-*p*-toluenesulfonamide (37.1 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (17:3 hexane/EtOAc) afforded **3L** as a yellow oil (37.1 mg, 46% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.80–7.74 (m, 4H), 7.39–7.35 (m, 3H), 7.33 (d, J = 8.8 Hz, 2H), 6.10 (q, $J_{H-F} = 7.6$ Hz, 1H), 2.80 (s, 3H), 2.45 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 144.1, 139.7, 138.9, 135.6, 133.1, 129.7, 127.7, 125.6, 125.4, 124.8, 124.1, 122.1, 57.7 (q, $J_{C-F} = 32.6$ Hz), 30.6, 21.6 (one peak is missing due to overlapping); ¹⁹F NMR (376 MHz, CDCl₃) δ –68.3 (d, $J_{F-H} = 7.6$ Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₈H₁₆F₃NNaO₂S₂ 422.0467; Found 422.0467.

2,2,2-Trifluoro-1-phenylethyl Benzoate (3M)^[9]

The reaction was conducted according to **General Procedure A**, benzoic acid (24.4 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (4:1 hexane/EtOAc) afforded **3M** as a colorless oil (28.6 mg, 51% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.15–8.12 (m, 2H), 7.63 (t, J = 7.6 Hz, 1H), 7.58–7.54 (m, 2H), 7.40 (t, J = 7.6 Hz, 2H), 7.44–7.41 (m, 3H), 6.37 (q, $J_{\text{H-F}}$ = 6.8 Hz, 1H); ${}^{13}\text{C}\{{}^{1}\text{H}\}$ NMR (101 MHz, CDCl₃) δ 164.3, 133.9, 131.3, 130.0, 129.9, 128.73, 128.69, 128.62, 128.0, 123.3 (q, $J_{\text{C-F}}$ = 280.7 Hz), 72.4 (q, $J_{\text{C-F}}$ = 33.2 Hz); ${}^{19}\text{F}$ NMR (376 MHz, CDCl₃) δ –75.9; HRMS (ESI) m/z [M+Na]⁺ Calcd for $C_{15}H_{11}F_{3}\text{NaO}_{2}$ 303.0603; Found 303.0609. The spectra matched with those of this compound reported in the literature. [9]

Phenyl(2,2,2-trifluoro-1-phenylethyl)sulfane (3N)^[10]

The reaction was conducted according to **General Procedure A**, thiophenol (21 μ L, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (19:1 hexane/Chloroform) afforded **3N** as a colorless oil (15.2 mg, 28% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.45–7.41 (m, 2H), 7.37–7.33 (m, 5H), 7.31–7.27 (m, 3H), 4.52 (q, J_{H-F} = 8.4 Hz, 1H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 133.8, 133.6, 132.7, 129.1, 128.9, 128.8, 128.72, 128.67, 125.5 (q, J_{C-F} = 279.9 Hz), 56.8 (q, J_{C-F} = 29.5 Hz); ¹⁹F

NMR (376 MHz, CDCl₃) δ –67.8 (d, J_{F-H} = 8.4 Hz); HRMS (APCI) m/z [M+H]⁺ Calcd for C₁₄H₁₂F₃S 269.0606; Found 269.0600. The spectra matched with those of this compound reported in the literature.^[10]

p-Tolyl(2,2,2-trifluoro-1-phenylethyl)sulfane (3O)^[11]

The reaction was conducted according to **General Procedure A**, *p*-toluenethiol (24.8 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile. Purification by PTLC (9:1 hexane/EtOAc) afforded **3O** as a colorless oil (29.7 mg, 52% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.33 (m, 5H), 7.32 (d, J = 8.4 Hz, 2H), 7.09 (d, J = 8.4 Hz, 2H), 4.45 (q, J_{H-F} = 8.4 Hz, 1H), 2.33 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 139.1, 134.2, 133.8, 129.9, 129.1, 128.9, 128.73, 128.67, 125.6 (q, J_{C-F} = 279.6 Hz), 57.2 (q, J_{C-F} = 29.3 Hz), 21.2; ¹⁹F NMR (376 MHz, CDCl₃) δ –67.8 (d, J_{F-H} = 8.4 Hz); HRMS (APCI) m/z [M+H]⁺ Calcd for C₁₅H₁₄F₃S 283.0763; Found 283.0755. The spectra matched with those of this compound reported in the literature.^[11]

1-Methoxy-4-(1,1,1-trifluoropent-4-en-2-yl)benzene (3P)^[12]

The reaction was conducted according to **General Procedure B**. Allyltrimethylsilane (95 µL, 0.60 mmol, 3.0 equiv) was used as a nucleophile, and TMSOTf (1.0 equiv; reduced from 3.0 equiv in the standard conditions) was employed. Purification by Isolera® (99:1 to 9:1 hexane/EtOAc) afforded **3P** as a colorless oil (34.2 mg, 74% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.20 (d, J = 8.4 Hz, 2H), 6.88 (d, J = 8.4 Hz, 2H), 5.57 (ddt, J = 17.2, 10.0, 6.8 Hz, 1H), 5.06–5.00 (m, 1H), 4.97 (dd, J = 10.0, 0.8 Hz, 1H), 3.80 (s, 3H), 3.27 (dtd, J = 19.6, 9.2, 4.4 Hz, 1H), 2.79–2.71 (m, 1H), 2.65–2.56 (m, 1H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 159.4, 134.0, 130.1, 126.8 (q, J_{C-F} = 280.0 Hz), 126.2, 117.6, 114.0, 55.2, 49.2 (q, J_{C-F} = 26.4 Hz), 33.2; ¹⁹F NMR (376 MHz, CDCl₃) δ –70.2 (d, J_{F-H} = 8.8 Hz); HRMS (APCI) m/z [M+H]⁺ Calcd for C₁₂H₁₄F₃O 231.0991; Found 231.1000. The spectra matched with those of this compound reported in the literature. ^[12]

1-Methoxy-4-(1,1,1-trifluoro-4-methylpent-4-en-2-yl)benzene (3Q)

The reaction was conducted according to **General Procedure B**. Methallyltrimethylsilane (104 μ L, 0.60 mmol, 3.0 equiv) was used as a nucleophile, and TMSOTf (1.0 equiv; reduced from 3.0 equiv in the standard conditions) was employed. Purification by Isolera® (99:1 to 9:1 hexane/EtOAc) afforded **3Q** as a colorless oil (43.5 mg, 89% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.20 (d, J = 8.8 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 4.71–4.68 (m, 1H), 4.61–4.59 (m, 1H), 3.80 (s, 3H), 3.42 (dqd, J = 11.2, 9.2, 4.0 Hz, 1H), 2.70 (dd, J = 14.4, 4.4 Hz, 1H), 2.58 (ddd, J = 14.4, 11.2, 0.8 Hz, 1H), 1.63–1.62 (m, 3H); 13 C{ 1 H} NMR (101 MHz, CDCl₃) δ 159.3, 140.8, 130.1, 126.9 (q, J_{C-F} = 280.0 Hz), 126.3, 113.9, 55.1, 47.6 (q, J_{C-F} = 26.4 Hz), 36.7, 22.0; 19 F NMR (376 MHz, CDCl₃) δ –70.3 (d, J_{F-H} = 8.8 Hz); HRMS (APCI) m/z [M+H] $^{+}$ Calcd for C₁₃H₁₆F₃O 245.1148; Found 245.1145.

2-(2,2,2-Trifluoro-1-(4-methoxyphenyl)ethyl)cyclohexan-1-one (3R)

The reaction was conducted according to **General Procedure B**. (Cyclohex-1-en-1-yloxy)trimethylsilane (115 μ L, 0.60 mmol, 3.0 equiv) was used as a nucleophile, and TMSOTf (1.0 equiv; reduced from 3.0 equiv in the standard conditions) was employed. Crude ¹H NMR showed *major*-3**R**:*minor*-3**R** = 63:37; ¹H NMR peaks at 4.31 (qd, J_{H-F} = 10.8 Hz, J = 5.6 Hz, 1H) and 3.96–3.84 (m, 0.59H) were used. Purification by PTLC (9:1 hexane/EtOAc) afforded *major*-3**R** as a colorless oil (20.3 mg, 35% yield) and *minor*-3**R** as a white solid (9.7 mg, 17% yield)

For *major*-3R: ¹H NMR (400 MHz, CDCl₃) δ 7.25 (d, J = 8.8 Hz, 2H), 6.86 (d, J = 8.8 Hz, 2H), 4.31 (qd, $J_{\text{H-F}}$ = 10.8 Hz, J = 5.6 Hz, 1H), 3.80 (s, 3H), 2.97 (dt, J = 11.2, 5.6 Hz, 1H), 2.51 (dt, J = 14.8, 4.4 Hz, 1H), 2.43–2.34 (m, 1H), 2.04–1.96 (m, 2H), 1.84–1.78 (m, 1H), 1.72–1.60 (m, 2H), 1.18–1.08 (m, 1H); ¹³C{¹H} NMR (101 MHz, CDCl₃, 323 K) δ 208.9, 159.4, 131.2, 127.4 (q, $J_{\text{C-F}}$ = 280.2 Hz), 124.8, 113.9, 55.2, 50.1, 46.2 (q, $J_{\text{C-F}}$ = 26.5 Hz), 41.7, 31.2, 27.6, 24.6; ¹⁹F NMR (376 MHz, CDCl₃) δ –67.1 (d, $J_{\text{F-H}}$ = 8.8 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₅H₁₇F₃NaO₂ 309.1073; Found 309.1074.

For *minor*-3R: ¹H NMR (400 MHz, CDCl₃) δ 7.25 (d, J = 8.8 Hz, 2H), 6.84 (d, J = 8.8 Hz, 2H), 3.96–3.84 (m, 1H), 3.78 (s, 3H), 3.11–3.03 (m, 1H), 2.54–2.44 (m, 1H), 2.36–2.30 (m, 1H), 2.28–2.23 (m, 1H), 2.12–2.05 (m, 1H), 2.01–1.95 (m, 1H), 1.80–1.68 (m, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 209.2, 159.1, 130.2, 127.9, 126.9 (q, J_{C-F} = 280.5 Hz), 113.9, 55.2, 53.7, 47.1 (q, J_{C-F} = 26.5 Hz), 42.6, 32.3, 28.6, 25.2; ¹⁹F NMR (376 MHz, CDCl₃) δ –63.4 (d, J_{F-H} = 11.6 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₅H₁₇F₃NaO₂ 309.1073; Found 309.1084.

2,2,4,4,4-Pentafluoro-1,3-bis(4-methoxyphenyl)butan-1-one (3S)

The reaction was conducted according to **General Procedure B**. 2,2-Difluoro-1-(4-methoxyphenyl)vinyl diethyl phosphate (64.4 mg, 0.20 mmol, 1.0 equiv) was used as a nucleophile, and TMSOTf (1.0 equiv; reduced from 3.0 equiv in the standard conditions) was employed. Purification by PTLC (4:1 hexane/EtOAc) and GPC afforded **3S** as a colorless oil (32.4 mg, 43% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 8.8 Hz, 2H), 7.36 (d, J = 8.8 Hz, 2H), 6.93 (d, J = 8.8 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 4.57 (ddq, $J_{\text{H-F}}$ = 17.6, 13.2, 8.8 Hz, 1H), 3.89 (s, 3H), 3.79 (s, 3H); ¹³C { ¹H } NMR (101 MHz, CDCl₃) δ 186.1 (t, $J_{\text{C-F}}$ = 29.1 Hz), 164.6, 160.2, 132.6 (t, $J_{\text{C-F}}$ = 3.8 Hz), 131.9, 124.5, 124.3 (qd, $J_{\text{C-F}}$ = 281.2, 3.6 Hz), 119.4, 116.8 (t, $J_{\text{C-F}}$ = 254.3 Hz), 114.1, 114.0, 55.6, 55.2, 51.7 (qdd, $J_{\text{C-F}}$ = 27.8, 23.2, 20.8 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -63.5 (dt, $J_{\text{F-F}}$ = 14.6 Hz, $J_{\text{F-H}}$ = 8.8 Hz), -101.0 (dqd, $J_{\text{F-F}}$ = 289.5, 13.2 Hz, $J_{\text{F-H}}$ = 13.2 Hz), -103.4 (dq, $J_{\text{F-F}}$ = 289.5, 8.6 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₈H₁₅F₅NaO₃ 397.0834; Found 397.0827.

S-(2,2,2-Trifluoro-1-(4-methoxyphenyl)ethyl) benzothioate (3T)

The reaction was conducted according to **General Procedure B**, thiobenzoic acid (82.9 mg, 0.60 mmol, 3.0 equiv) was used as a nucleophile, and TMSOTf (1.0 equiv; reduced from 3.0 equiv in the standard conditions) was employed. Purification by PTLC (4:1 hexane/EtOAc) afforded **3T** as a pink oil (22.0 mg, 34% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.97 (dd, J = 8.4, 1.2 Hz, 2H), 7.61 (tt, J = 7.2, 1.2 Hz, 1H), 7.50–7.44 (m, 2H), 7.39 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 8.8 Hz, 2H), 5.44 (q, J_{H-F} = 9.2 Hz, 1H), 3.81 (s, 3H); 13 C{ 1 H} NMR (101 MHz, CDCl₃) δ 187.7, 160.0, 135.7, 134.2, 130.5, 128.8, 127.6, 125.3 (q, J_{C-F} = 278.6 Hz), 125.1, 114.2, 55.3, 48.7 (q, J_{C-F} = 31.1 Hz); 19 F NMR (376 MHz, CDCl₃) δ –68.4 (d, J_{F-H} = 9.2 Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₆H₁₃F₃NaO₂S 349.0481; Found 349.0486.

N,4-Dimethyl-N-(2,2,3,3,3-pentafluoro-1-phenylpropyl)benzenesulfonamide (3U)

The reaction was conducted according to **General Procedure A**, imidazole (20 mol%) was used instead of DBU. Purification by PTLC (9:1 hexane/EtOAc) afforded **3U** as a colorless oil (29.8 mg, 38% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.64 (d, J = 8.4 Hz, 2H), 7.45–7.42 (m, 2H), 7.39–7.35 (m,

3H), 7.24 (d, J = 8.4 Hz, 2H), 5.89 (t, $J_{H-F} = 16.0$ Hz, 1H), 2.79 (s, 3H), 2.40 (s, 3H); $^{13}C\{^{1}H\}$ NMR (101 MHz, CDCl₃) δ 143.9, 135.5, 129.9, 129.5, 128.8, 127.7, 118.6 (qt, $J_{C-F} = 287.1$, 35.3 Hz), 115.0 (tq, $J_{C-F} = 260.9$, 36.0 Hz), 58.7 (t, $J_{C-F} = 22.6$ Hz), 31.0, 21.5 (two peaks are missing due to overlapping); ^{19}F NMR (376 MHz, CDCl₃) δ -81.7, -116.5 (t, $J_{F-H} = 16.0$ Hz); HRMS (ESI) m/z [M+Na]⁺ Calcd for $C_{17}H_{16}F_{5}NNaO_{2}S$ 416.0714; Found 416.0710.

5. Deoxygenative Arylation of Trifluoromethylketones

General Procedure

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N_2 gas after cooling to room temperature. The tube was placed under vacuum and refilled three times with N_2 gas, and to this tube were added trifluoromethylketone 1 (0.20 mmol, 1.0 equiv), diethyl phosphite (26 μ L, 0.20 mmol, 1.0 equiv), DBU (3 μ L, 20 μ mol, 10 mol%), and chloroform (1.0 mL). The vessel was sealed with a screw cap and then heated at 80 °C for 2 h while its contents were being stirred. After cooling to room temperature, the mixture was concentrated *in vacuo*. To the residue were added arene (1.0 mL, 0.20 M) and TfOH (35 μ L, 0.40 mmol, 2.0 equiv) at 0 °C. After being stirred the mixture for 15 minutes while the reaction progress was being monitored by TLC, the reaction was quenched by saturated NaHCO₃ aq. The mixture was extracted three times with CH₂Cl₂. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by PTLC to afford the corresponding product 5.

1-Methoxy-4-(2,2,2-trifluoro-1-(p-tolyl)ethyl)benzene (5A)^[13]

The reaction was conducted with toluene as solvent. Crude ¹H NMR showed p-5A:o-5A = 95:5; ¹H NMR peaks at 4.59 (q, $J_{H-F} = 10.0$ Hz, 1H) and 4.83 (q, $J_{H-F} = 10.0$ Hz, 0.05H) were used. Purification by PTLC (9:1 hexane/EtOAc) afforded **5A** as a colorless oil (52.6 mg, 93% yield, as a mixture of structural isomer; p-5A:o-5A = 95:5).

For p-5A: ¹H NMR (400 MHz, CDCl₃) δ 7.27 (d, J = 8.8 Hz, 2H), 7.25–7.22 (m, 2H), 7.15 (d, J = 7.6 Hz, 2H), 6.86 (d, J = 8.8 Hz, 2H), 4.59 (q, $J_{\text{H-F}}$ = 10.0 Hz, 1H), 3.78 (s, 3H), 2.33 (s, 3H); ¹³C { ¹H } NMR (101 MHz, CDCl₃) δ 159.1, 137.5, 132.7, 130.1, 129.4, 128.8, 127.7, 126.3 (q, $J_{\text{C-F}}$ = 280.3 Hz), 114.0, 55.2, 54.4 (q, $J_{\text{C-F}}$ = 27.6 Hz), 21.0; ¹⁹F NMR (376 MHz, CDCl₃) δ –66.3 (d, $J_{\text{F-H}}$ = 10.0 Hz). HRMS (APCI) m/z [M+H]⁺ Calcd for C₁₆H₁₆F₃O 281.1148; Found 281.1138. The spectra matched with those of this compound reported in the literature. ^[13]

4,4'-(2,2,2-Trifluoroethane-1,1-diyl)bis(methoxybenzene) (5B)[14]

The reaction was conducted with anisole as solvent. Purification by PTLC (9:1 hexane/EtOAc) afforded **5B** as a colorless oil (60.0 mg, 98% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.27 (d, J = 8.4 Hz, 4H), 6.87 (d, J = 8.4 Hz, 4H), 4.58 (q, $J_{\text{H-F}}$ = 10.0 Hz, 1H), 3.79 (s, 6H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 159.1, 130.1, 127.8, 126.4 (q, $J_{\text{C-F}}$ = 280.8 Hz), 114.0, 55.2, 53.9 (q, $J_{\text{C-F}}$ = 27.5 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ –66.5 (d, $J_{\text{F-H}}$ = 10.0 Hz); HRMS (APCI) m/z [M]⁺ Calcd for C₁₆H₁₅F₃O₂ 296.1019; Found 296.1011. The spectra matched with those of this compound reported in the literature. ^[14]

1,3,5-Trimethyl-2-(2,2,2-trifluoro-1-(4-methoxyphenyl)ethyl)benzene (5C)^[14]

The reaction was conducted with mesitylene as solvent. Purification by PTLC (9:1 hexane/EtOAc) afforded **5C** as a colorless oil (59.9 mg, 97% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.15 (d, J = 8.8 Hz, 2H), 6.94 (s, 1H), 6.83 (d, J = 8.8 Hz, 2H), 6.80 (s, 1H), 5.30 (q, J_{H-F} = 10.4 Hz, 1H), 3.79 (s, 3H), 2.46 (s, 3H), 2.27 (s, 3H), 1.85 (s, 3H); 13 C { 1 H} NMR (101 MHz, CDCl₃) δ 158.2, 138.5, 137.6, 131.6, 130.0, 129.3, 128.6, 127.9, 126.9 (q, J_{C-F} = 281.3 Hz), 113.8, 55.2, 48.4 (q, J_{C-F} = 27.5 Hz), 21.6, 21.0, 20.7 (one peak is missing due to overlapping); 19 F NMR (376 MHz, CDCl₃) δ –62.7 (d, J_{F-H} = 10.4 Hz); HRMS (APCI) m/z [M+H] $^{+}$ Calcd for C₁₈H₂₀F₃O 309.1461; Found 309.1458. The spectra matched with those of this compound reported in the literature. $^{[14]}$

2-(2,2,2-Trifluoro-1-(4-methoxyphenyl)ethyl)thiophene (5D)^[13.15]

The reaction was conducted with thiophene as solvent. Crude 1 H NMR showed C_{2} - $5D:C_{3}$ -5D = 88:12; 1 H NMR peaks at 4.84 (q, $J_{H-F} = 9.6$ Hz, 1H) and 4.70 (q, $J_{H-F} = 9.6$ Hz, 0.14H) were used. Purification by PTLC (9:1 hexane/EtOAc) afforded 5D as a colorless oil (24.6 mg, 45% yield, as a mixture of structural isomer; C_{2} - $5D:C_{3}$ -5D = 88:12).

For C₂-**5D**: ¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, J = 8.8 Hz, 2H), 7.27 (dd, J = 5.2, 1.2 Hz 1H), 7.09–7.07 (m, 1H), 6.99 (dd, J = 5.2, 3.6 Hz, 1H), 6.90 (d, J = 8.8 Hz, 2H), 4.84 (q, $J_{\text{H-F}}$ = 9.6 Hz, 1H), 3.81 (s, 3H); ¹³C { ¹H } NMR (101 MHz, CDCl₃) δ 159.6, 137.6, 130.2, 127.2, 126.9, 125.7, 125.6 (q, $J_{\text{C-F}}$ = 280.5 Hz), 114.1, 55.2, 50.4 (q, $J_{\text{C-F}}$ = 29.2 Hz) (one peak is missing due to overlapping); ¹⁹F NMR (376 MHz, CDCl₃) δ –68.1 (d, $J_{\text{F-H}}$ = 8.8 Hz); HRMS (APCI) m/z [M+H]⁺ Calcd for C₁₃H₁₂F₃OS 273.0555; Found 273.0551. The spectra matched with those of this compound reported in the literature. ^[13.15]

6. Application

6.1. Detosylation of 3J

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar and magnesium turnings (48.6 mg, 2.0 mmol, 10 equiv) was dried with a heat gun *in vacuo*. The tube was filled with N_2 gas after cooling to room temperature. To this tube was added **3J** (83.9 mg, 0.20 mmol, 1.0 equiv). The tube was placed under vacuum and refilled three times with N_2 gas, and to this tube was added methanol (1.0 mL). The vessel was sealed with a screw cap, and the mixture was stirred at room temperature for 2 h while the reaction progress was being monitored by TLC. The reaction was quenched with NH₄Cl aq. The mixture was extracted three times with Et₂O. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by PTLC (4:1 hexane/EtOAc) to afford 1-([1,1'-biphenyl]-4-yl)-2,2,2-trifluoro-*N*-methylethan-1-amine (**6**) as white solid (45.1 mg, 85% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.61 (d, J = 8.0 Hz, 4H), 7.49–7.43 (m, 4H), 7.36 (t, J = 7.2 Hz, 1H), 4.07 (q, J_{H-F} = 7.6 Hz, 1H), 2.45 (s, 3H), 1.70 (brs, 1H); 13 C (1 H) NMR (101 MHz, CDCl₃) δ 141.9, 140.4, 133.0, 128.9, 128.8, 127.6, 127.4, 127.1, 125.4 (q, J_{C-F} = 281.6 Hz), 66.2 (q, J_{C-F} = 28.5 Hz), 34.6; 19 F NMR (376 MHz, CDCl₃) δ -74.1 (d, J_{F-H} = 7.6 Hz). HRMS (ESI) m/z [M+H]⁺ Calcd for C₁₅H₁₅F₃N 266.1151; Found 266.1158.

6.2. Defluorination of 3F

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N_2 gas after cooling to room temperature. To this tube was added **3F** (84.5 mg, 0.20 mmol, 1.0 equiv). The tube was placed under vacuum and refilled three times with N_2 gas, and to this tube was added THF (1.0 mL). To this tube was added lithium bis(trimethylsilyl)amide (LiHMDS: 0.24 mL, 1.0 M in toluene 0.24 mmol, 1.2 equiv) at -30 °C. The mixture was stirred for 1 h while the reaction progress was being monitored by TLC. The reaction was quenched with water. The mixture was extracted three times with Et₂O. The combined organic layer was dried over Na_2SO_4 , filtered, and concentrated *in vacuo*. The residue was purified by PTLC (9:1 hexane/EtOAc) to afford N-(1-(4-bromophenyl)-2,2-difluorovinyl)-N,4-dimethylbenzenesulfonamide (7) as colorless oil (49.1 mg, 61% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.71 (d, J = 8.4 Hz, 2H), 7.49 (d, J = 8.4 Hz, 2H), 7.35–7.30 (m, 4H), 3.06 (s, 3H), 2.45 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ

156.9 (dd, $J_{C-F} = 303.3$, 298.8 Hz), 144.0, 135.4, 131.8, 129.9 (dd, $J_{C-F} = 6.9$, 1.8 Hz), 129.6, 128.7 (dd, $J_{C-F} = 6.3$, 3.4 Hz), 127.5, 122.3, 100.6 (dd, $J_{C-F} = 30.7$, 17.7 Hz), 36.8, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –82.4 (d, $J_{F-F} = 20.1$ Hz), –88.2 (dd, $J_{F-F} = 20.1$, 6.5 Hz). HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₆H₁₄BrF₂NNaO₂S 423.9789; Found 423.9789.

6.3. Carboxylation of 3F

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun in vacuo. The tube was filled with N2 gas after cooling to room temperature. To this tube were added 3F (84.5 mg, 0.20 mmol, 1.0 equiv), sodium formate (20.4 mg, 0.30 mmol, 1.5 equiv), 4CzIPN (3.2 mg, 4.0 μmol, 2.0 mol%), N-phenylbis(trifluoromethanesulfonimide) (10.7 mg, 30 μmol, 15 mol%), NiBr₂·DME (6.2 mg, 20 μmol, 10 mol%), and 4,4'-di-*tert*-butyl-2,2'-bipyridyl (dtbbpy: 5.4 mg, 20 μmol, 10 mol%). The tube was placed under vacuum and refilled three times with N_2 gas, and to this tube were added DMSO (0.50 mL) and 1,4-dioxane (0.50 mL). The reaction mixture was purged with N₂ gas for 5 min. The vessel was sealed with a screw cap, and the mixture was irradiated with blue LEDs (Kessil®, 456 nm) for 16 h. The reaction was quenched with HCl aq. The mixture was extracted three times with EtOAc. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue purified chloroform/methanol) was by **PTLC** (9:1)to afford 4-(1-((*N*,4dimethylphenyl)sulfonamido)-2,2,2-trifluoroethyl)benzoic acid (8) as colorless oil (62.4 mg, 81% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.13 (d, J = 8.0 Hz, 2H), 7.76 (d, J = 8.0 Hz, 2H), 7.55 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 5.95 (q, $J_{H-F} = 8.4$ Hz, 1H), 2.68 (s, 3H), 2.46 (s, 3H); ${}^{13}C\{{}^{1}H\}$ NMR (101) MHz, CDCl₃) δ 171.2, 144.2, 136.4, 135.7, 130.7, 129.8, 128.7, 127.6, 124.6 (q, J_{C-F} = 284.4 Hz), 123.4, 60.2 (q, $J_{C-F} = 31.5$ Hz), 30.6, 21.5; ¹⁹F NMR (376 MHz, CDCl₃) δ –67.7 (d, $J_{F-H} = 8.4$ Hz). HRMS (ESI) m/z [M+Na]⁺ Calcd for C₁₇H₁₆F₃NNaO₄S 410.0644; Found 410.0648.

Photochemical Reaction Setup

The blue LED lamps (PR160L-456 nm Kessil[®] LED lamp, $\lambda_{max} = 456$ nm) were used with the intensity dial set to 100. The reaction tubes were placed 4.0 cm away from the LED lamps (**Figure S1**). During the photochemical reaction, fan cooling was used to maintain the temperature at approximately 35 °C. All reactions were performed with 1 or 2 tubes per 1 Kessil LED lamp.

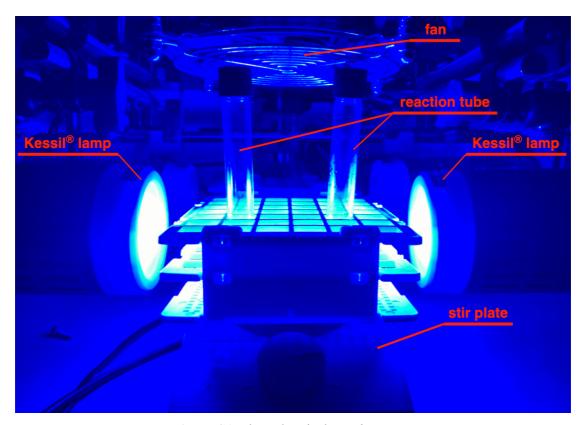


Figure S1. Photochemical reaction setup

6.4. Cyanation of 3F

An 8-mL glass tube equipped with a screw cap containing a magnetic stirring bar was dried with a heat gun *in vacuo*. The tube was filled with N₂ gas after cooling to room temperature. To this tube were added **3F** (84.5 mg, 0.20 mmol, 1.0 equiv), Pd(PPh₃)₄ (13.9 mg, 12 μ mol, 6.0 mol%), zinc cyanide (14.1 mg, 0.12 mmol, 0.60 equiv). The tube was placed under vacuum and refilled three times with N₂ gas, and to this tube were added DMF (1.0 mL). The vessel was sealed with a screw cap and then heated at 90 °C for 10 h while its contents were being stirred. After the reaction mixture had been cooled to room temperature, the reaction was quenched by water. The mixture was extracted three times with EtOAc. The combined organic layer was dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The residue was purified by PTLC (3:2 hexane/EtOAc) to afford *N*-(1-(4-cyanophenyl)-2,2,2-trifluoroethyl)-*N*,4-dimethylbenzenesulfonamide (**9**) as yellow solid (63.1 mg, 86% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, J = 8.0 Hz, 2H), 7.71 (d, J = 8.0 Hz, 2H), 7.58 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 5.93 (q, J_{H-F} = 8.0 Hz, 1H), 2.66 (s, 3H), 2.46 (s, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 144.3, 135.8, 135.4, 132.6, 129.7, 129.2, 127.5, 124.3 (q, J_{C-F} = 284.8 Hz), 117.8, 113.2, 60.0 (q, J_{C-F} = 31.7 Hz),

30.5, 21.5; 19 F NMR (376 MHz, CDCl₃) δ –67.5 (t, J_{F-H} = 8.0 Hz). HRMS (ESI) m/z [M+Na]⁺ Calcd for $C_{17}H_{15}F_3N_2NaO_2S$ 391.0699; Found 391.0690.

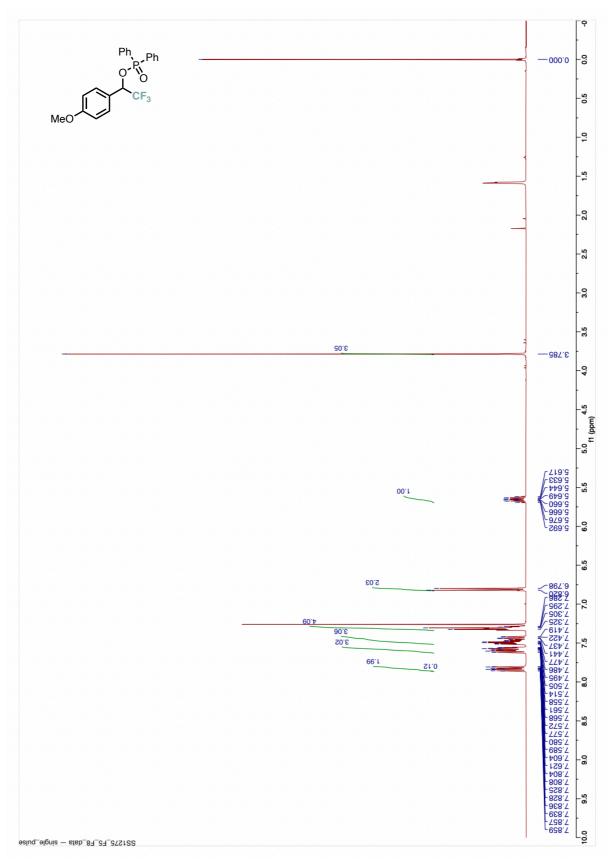
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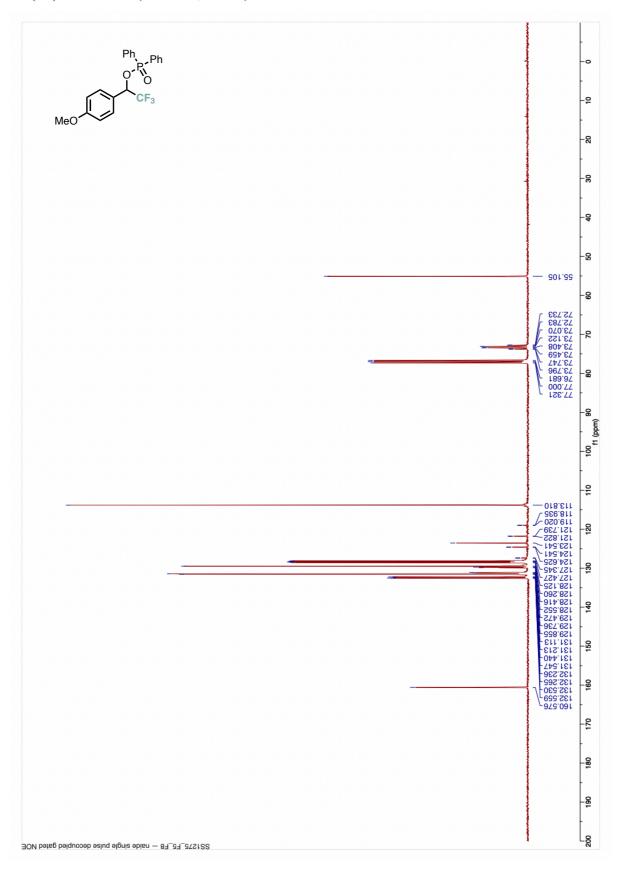
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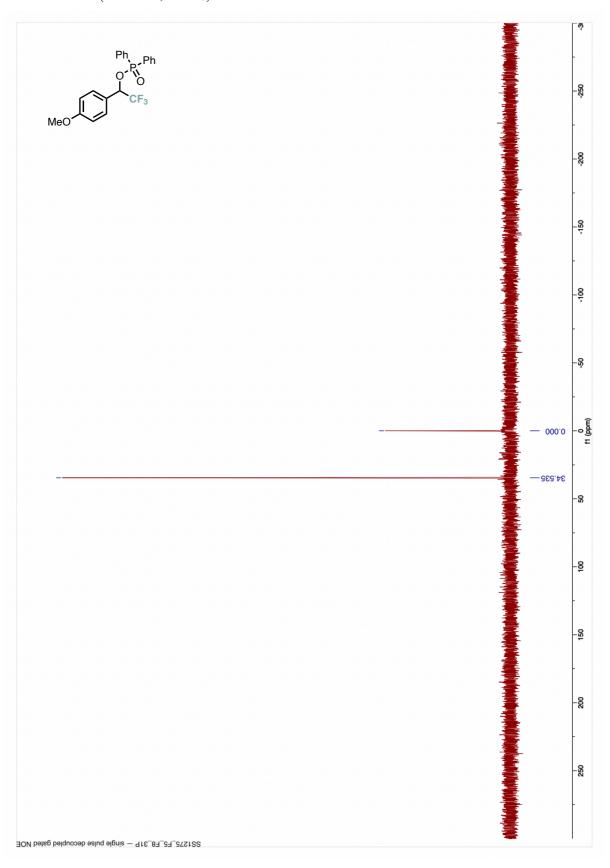
8. ¹H , ¹³C, ¹⁹F, and ³¹P NMR Spectra

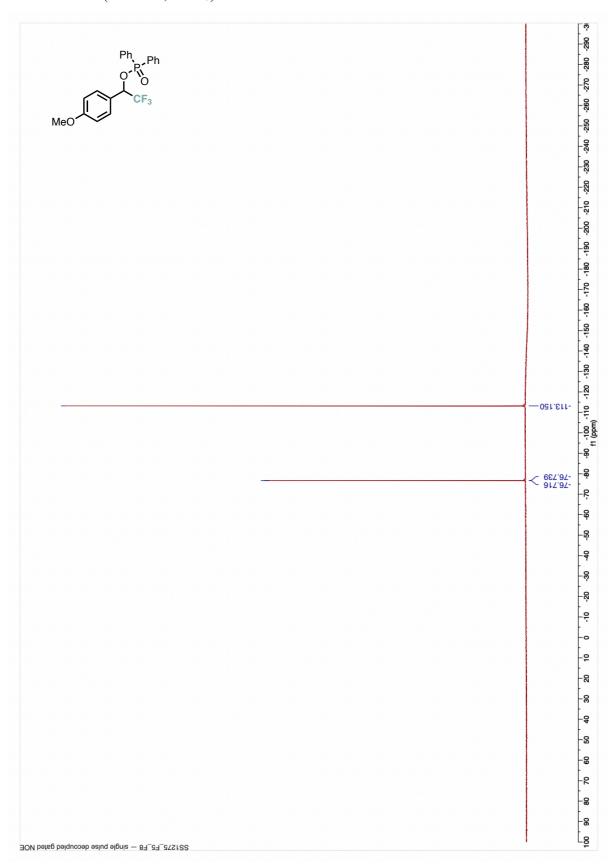
¹H NMR of **2A** (400 MHz, CDCl₃)

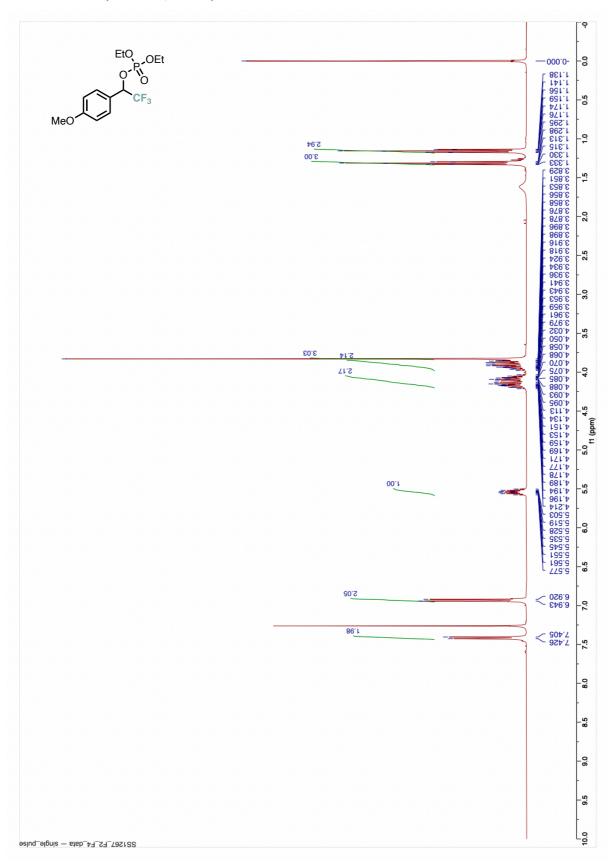


¹³C{¹H} NMR of **2A** (101 MHz, CDCl₃)

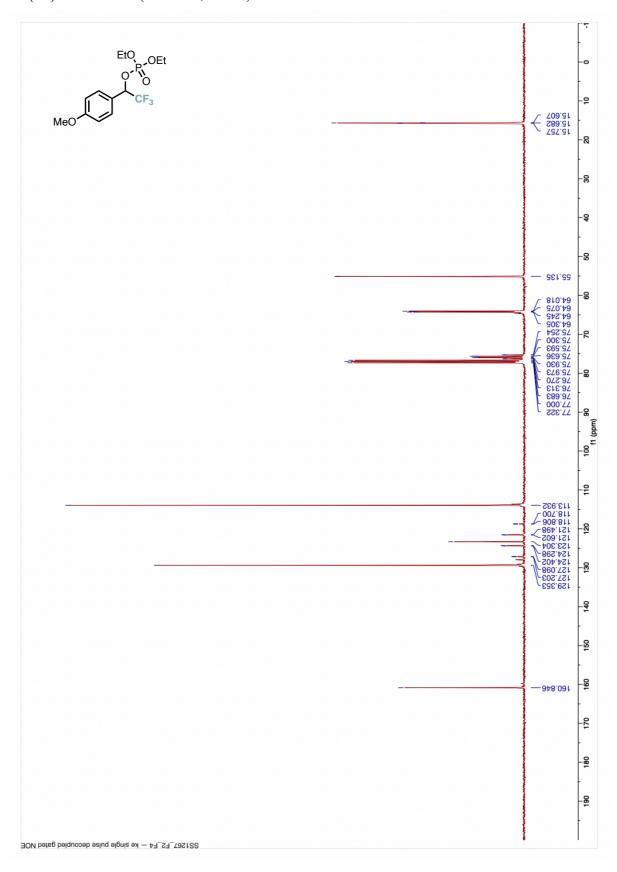


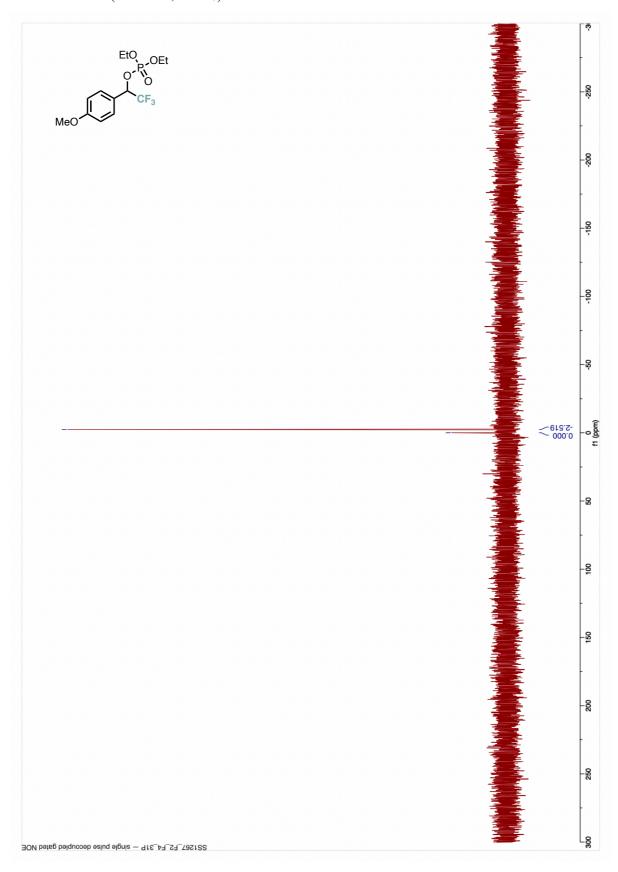


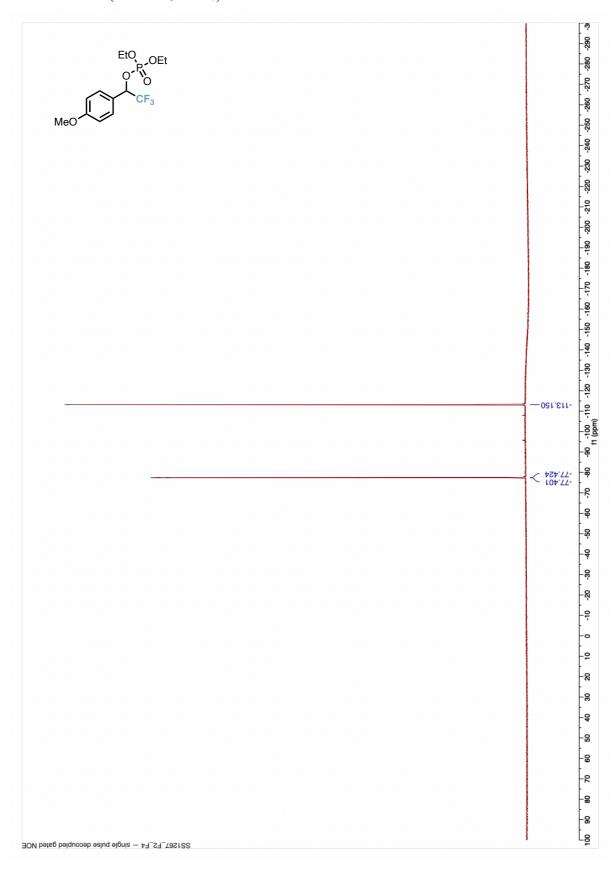


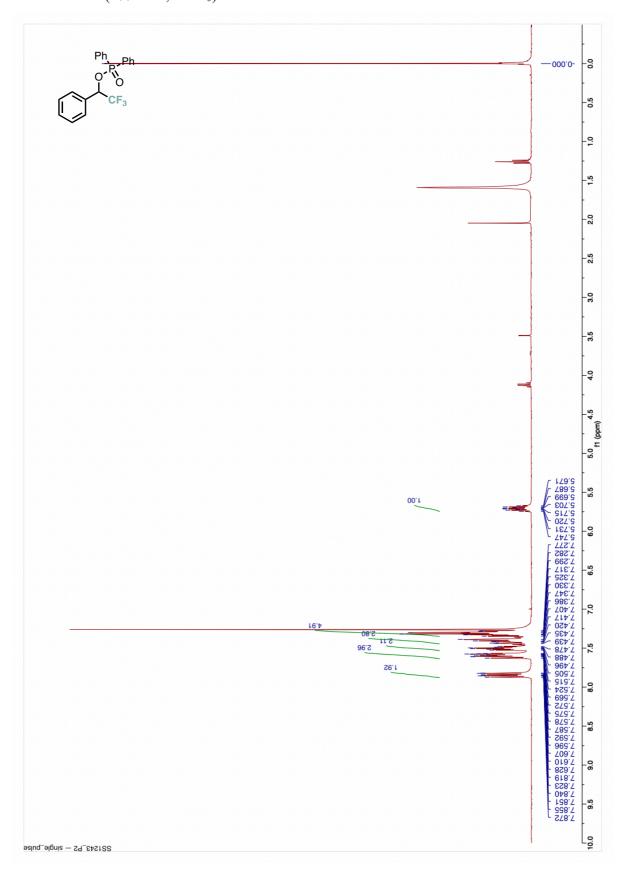


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{2A'}$ (101 MHz, CDCl₃)

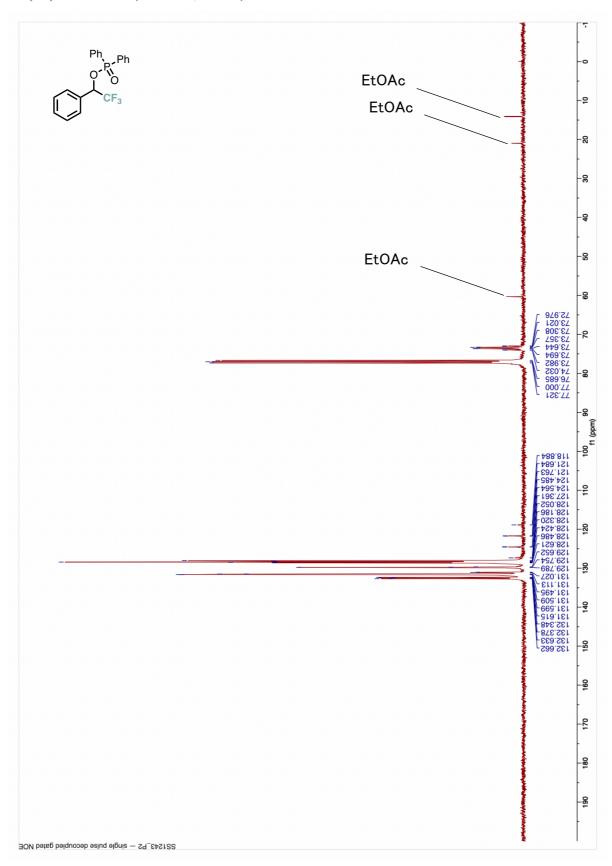




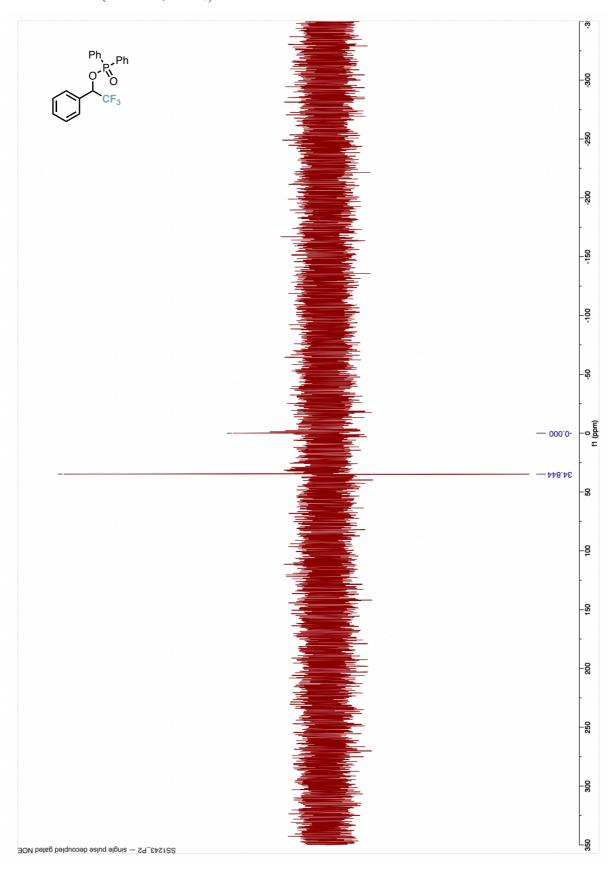


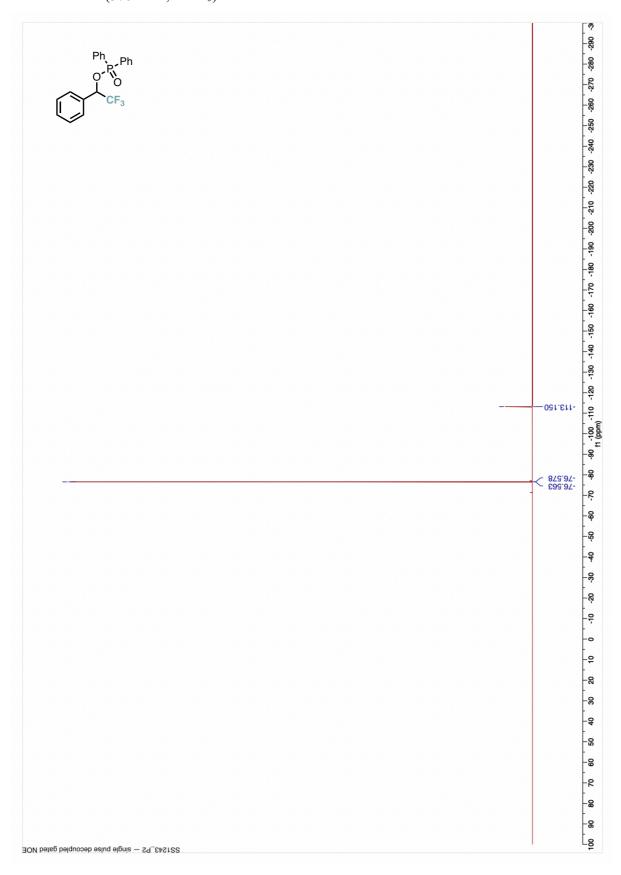


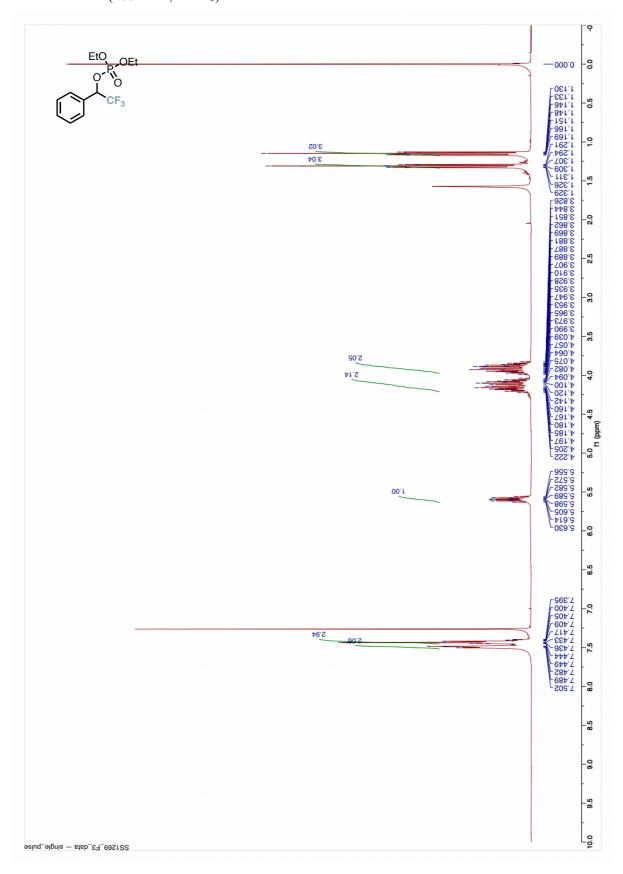
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{2B}$ (101 MHz, CDCl₃)



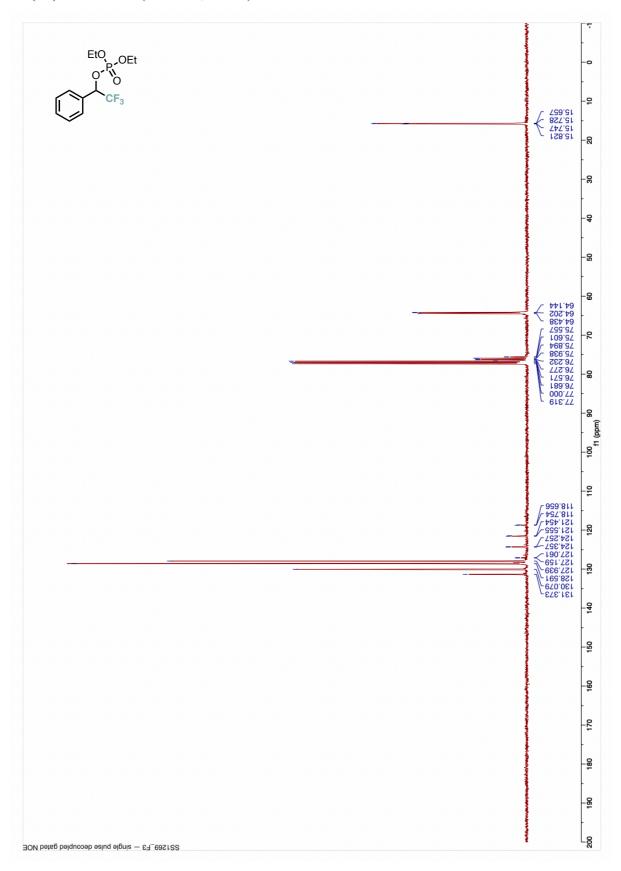
³¹P NMR of **2B** (162 MHz, CDCl₃)

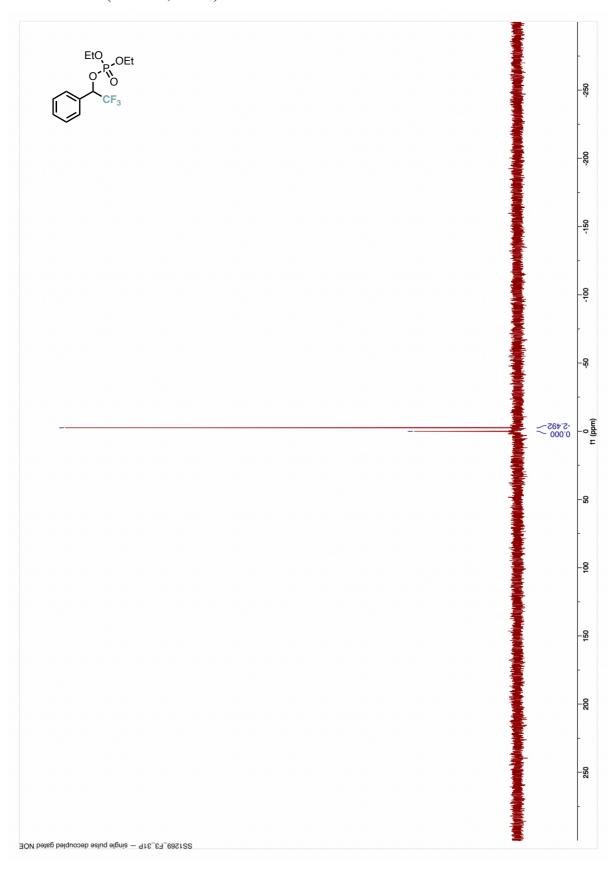


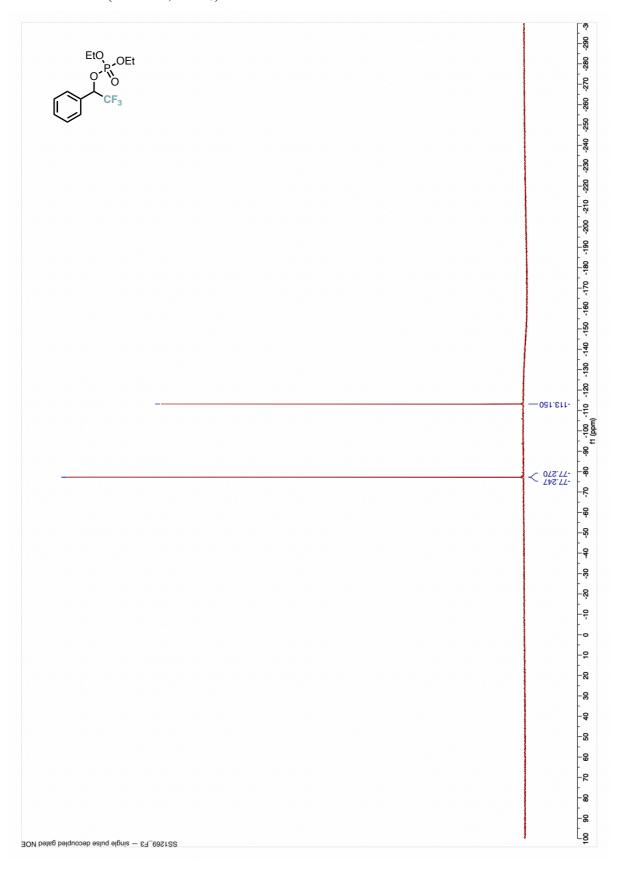


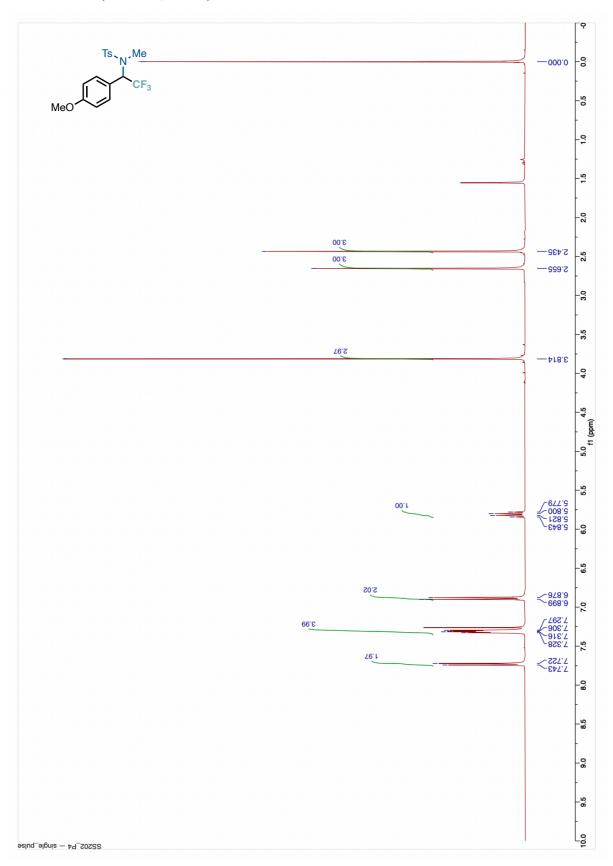


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{2B'}$ (101 MHz, CDCl₃)

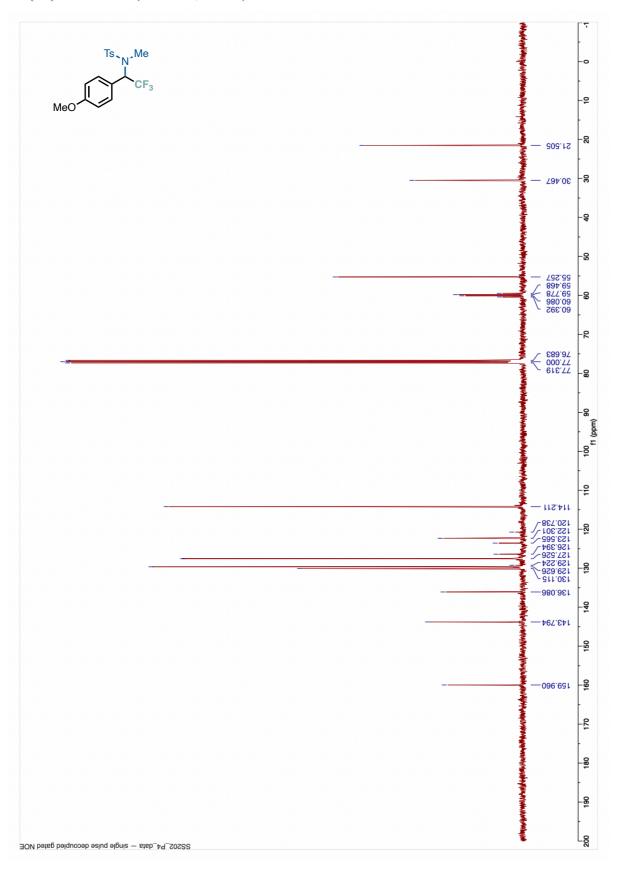


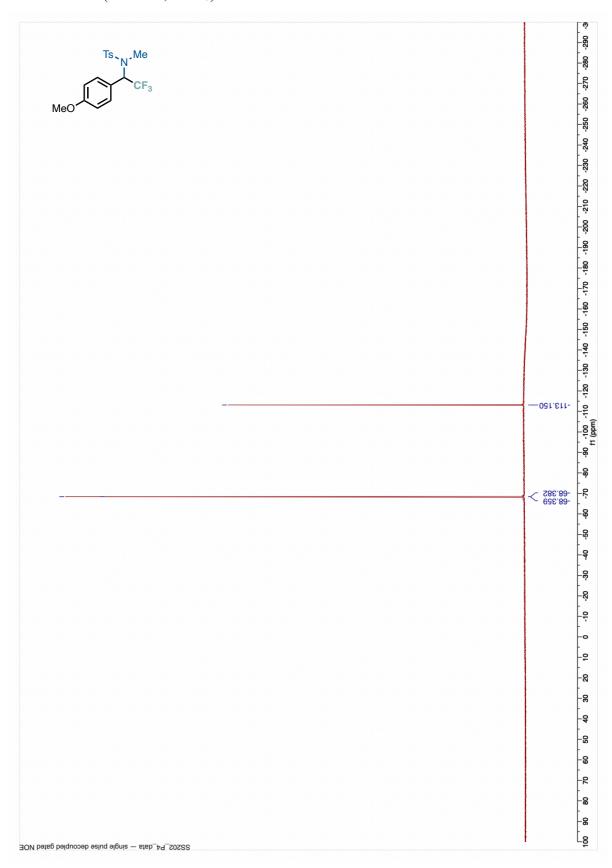




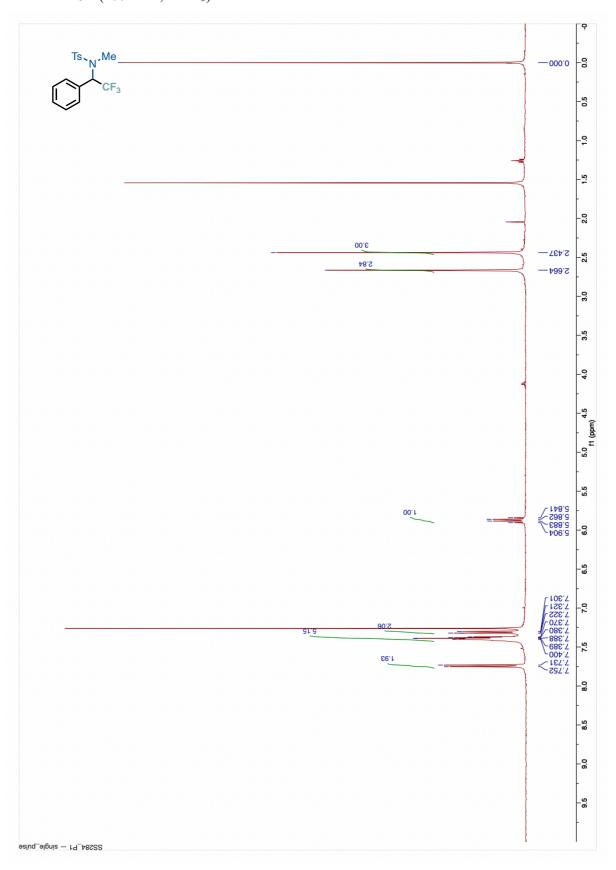


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3A}$ (101 MHz, CDCl₃)

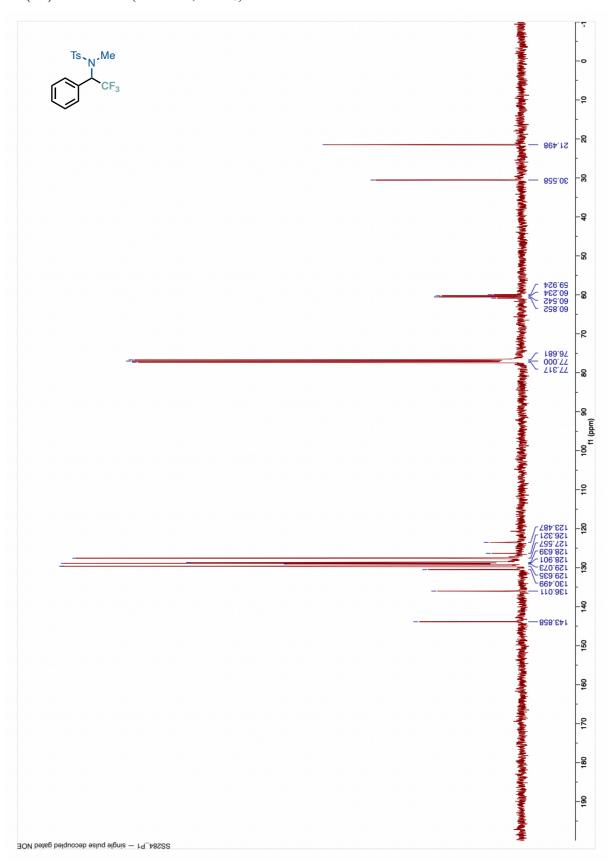


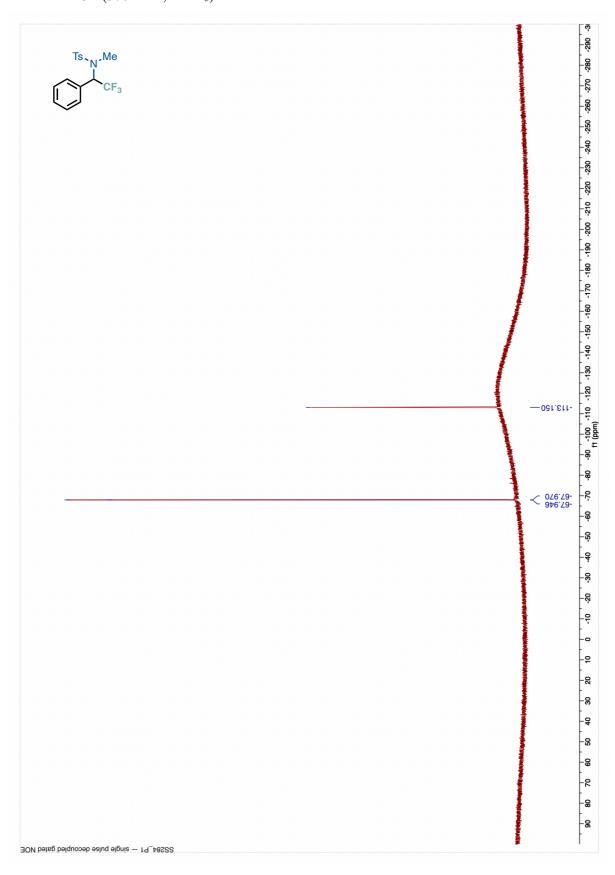


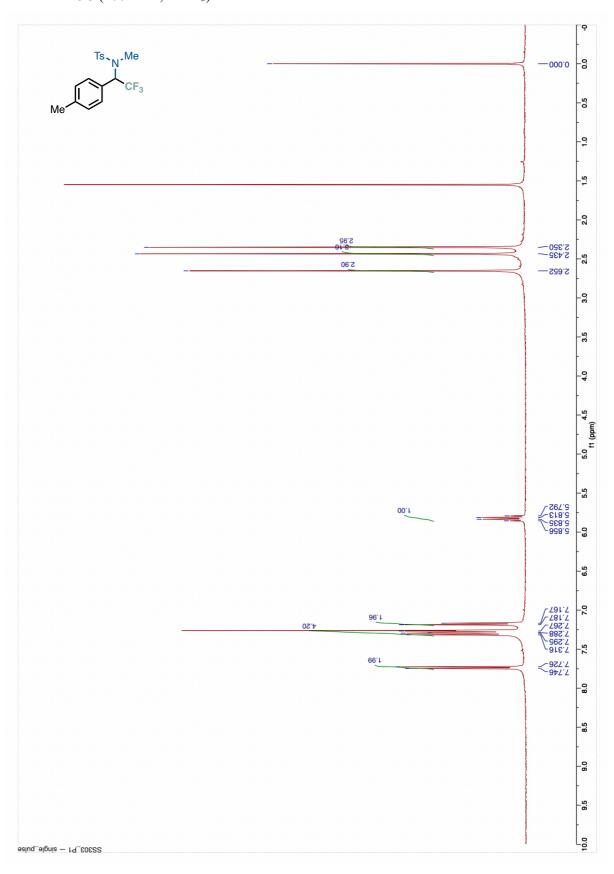
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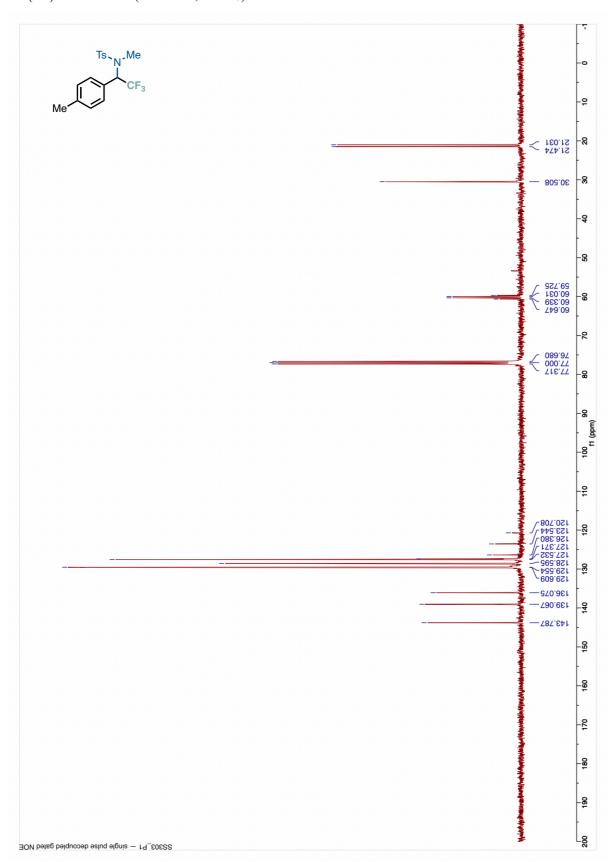
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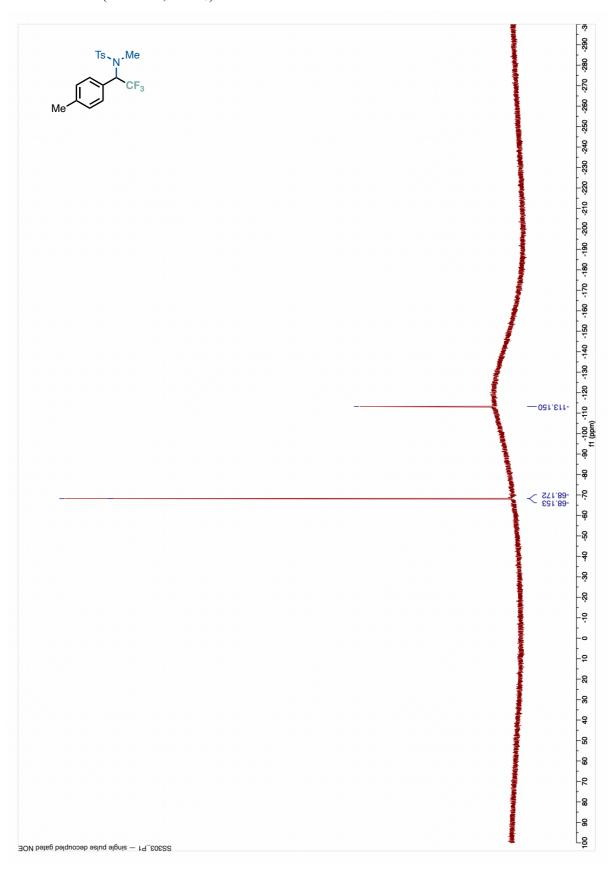


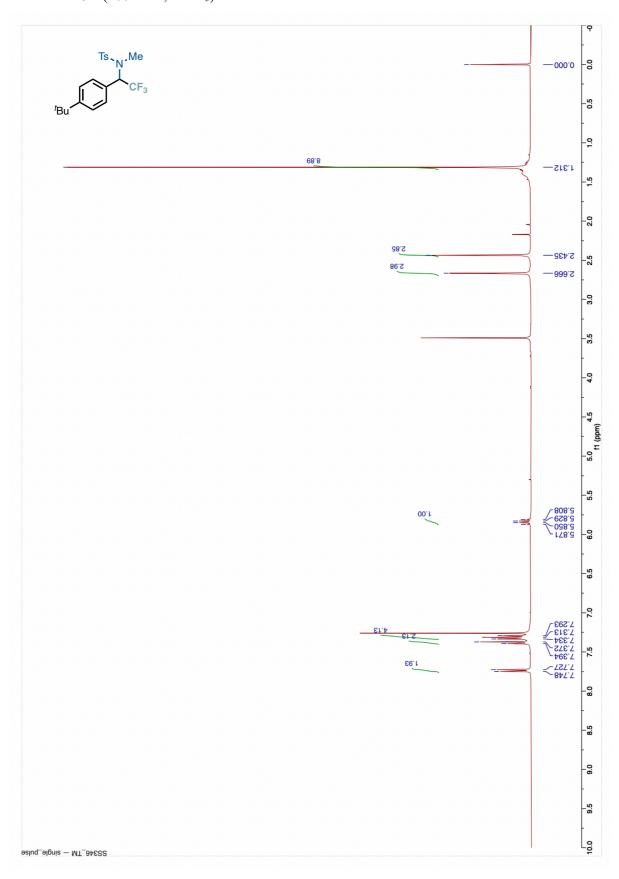




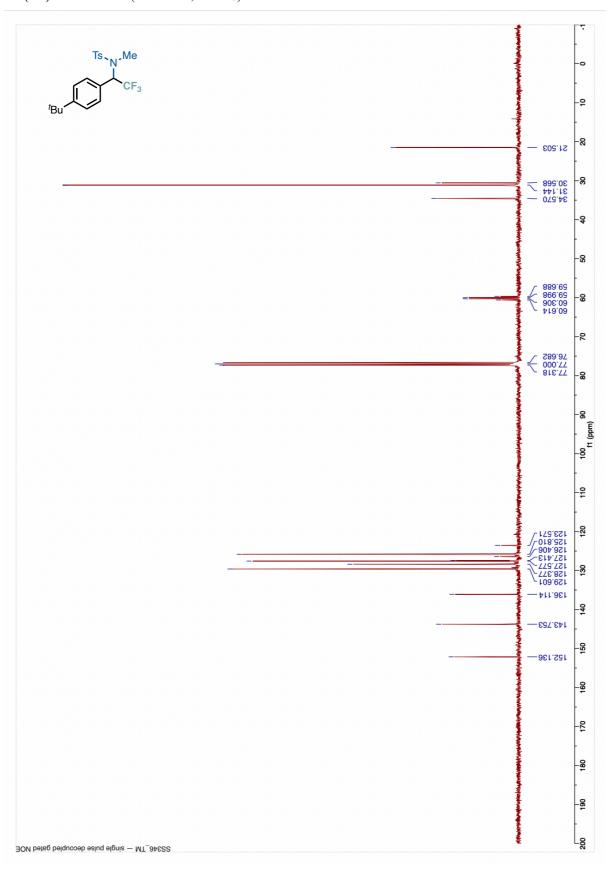
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3C}$ (101 MHz, CDCl₃)

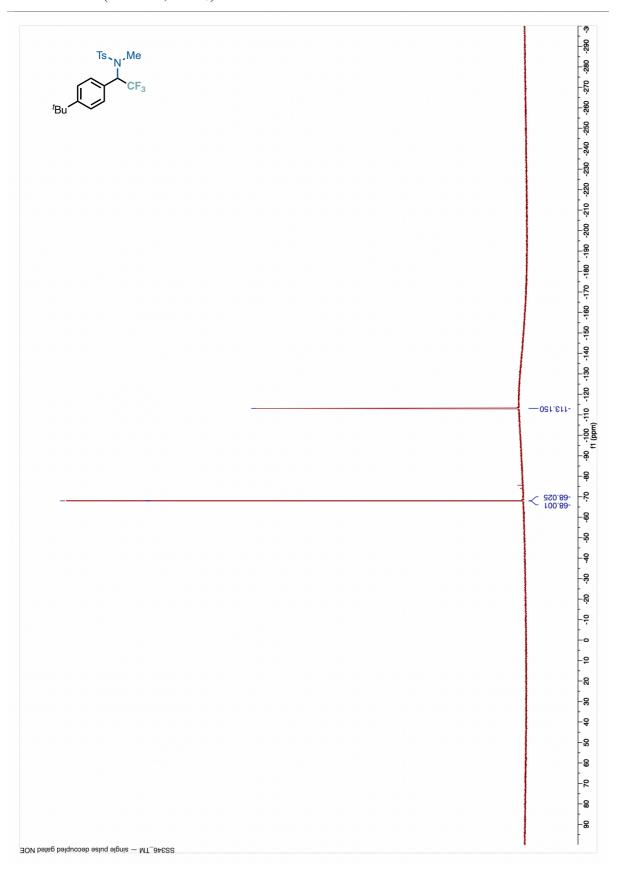




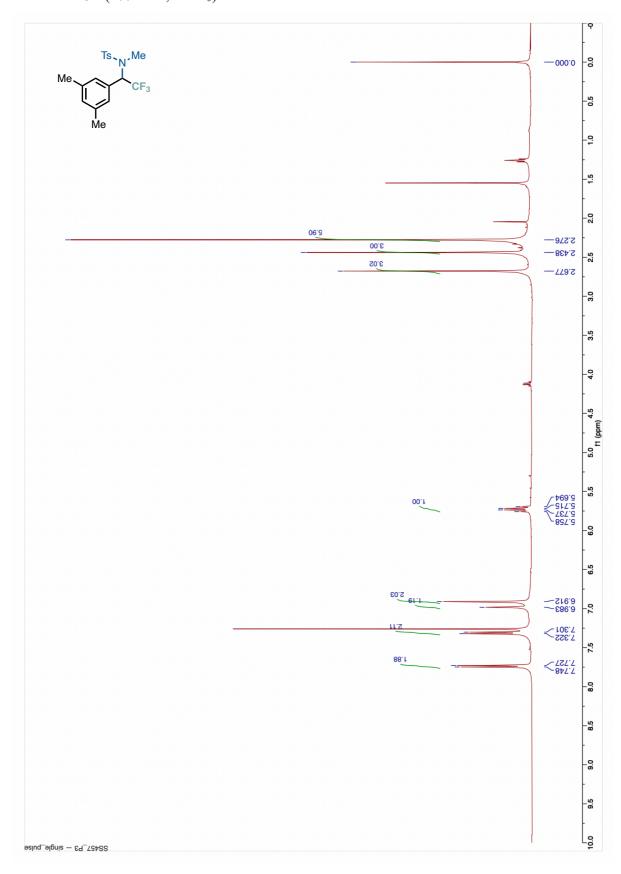


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3D}$ (101 MHz, CDCl₃)

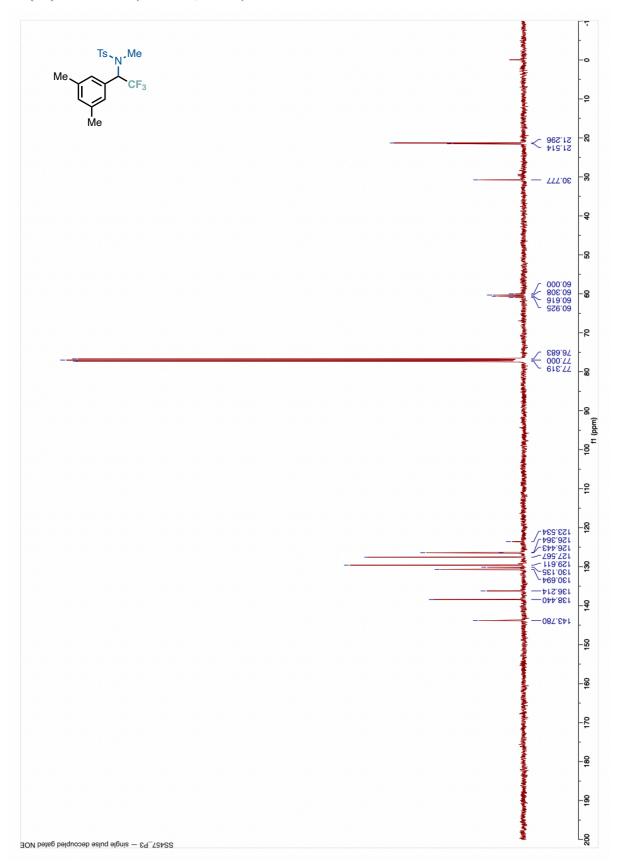


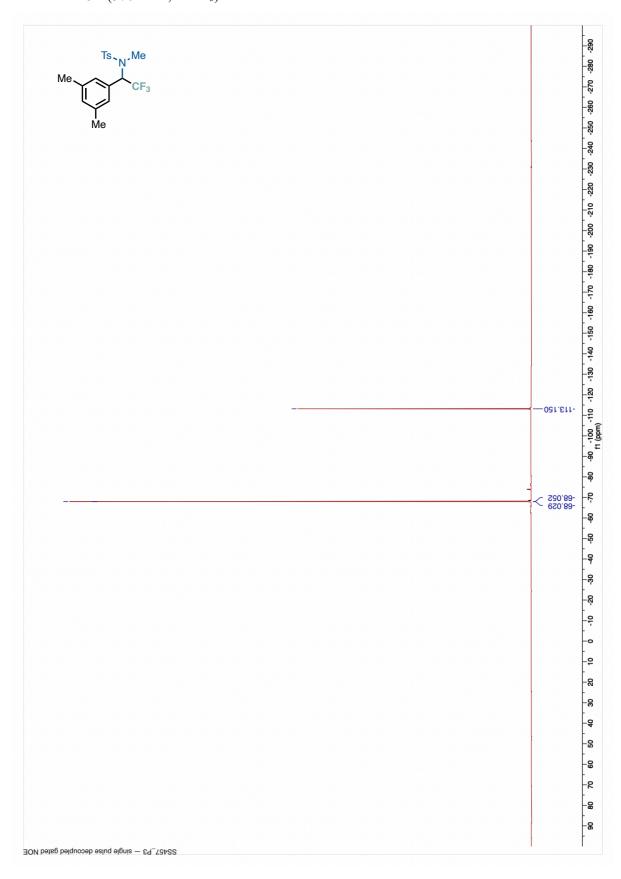


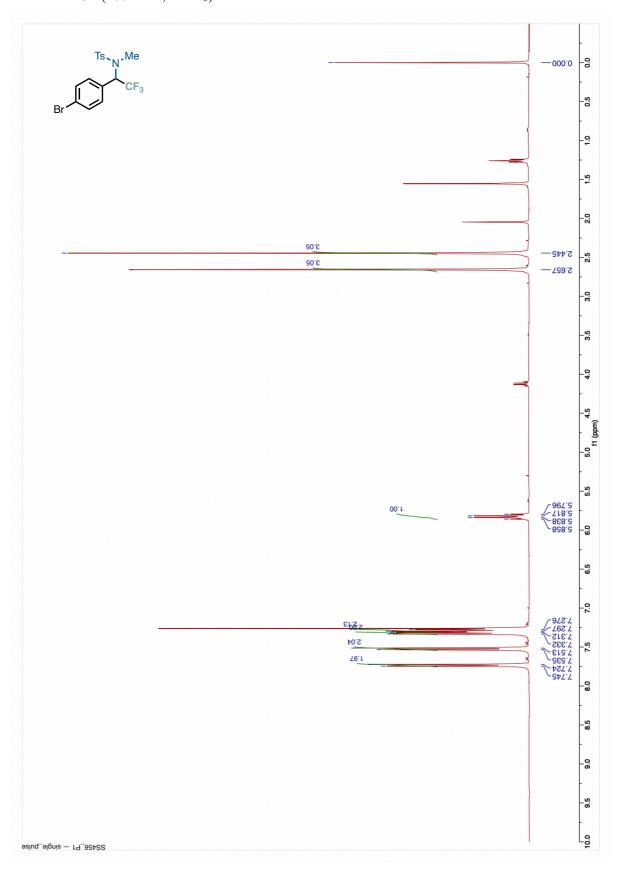
¹H NMR of **3E** (400 MHz, CDCl₃)



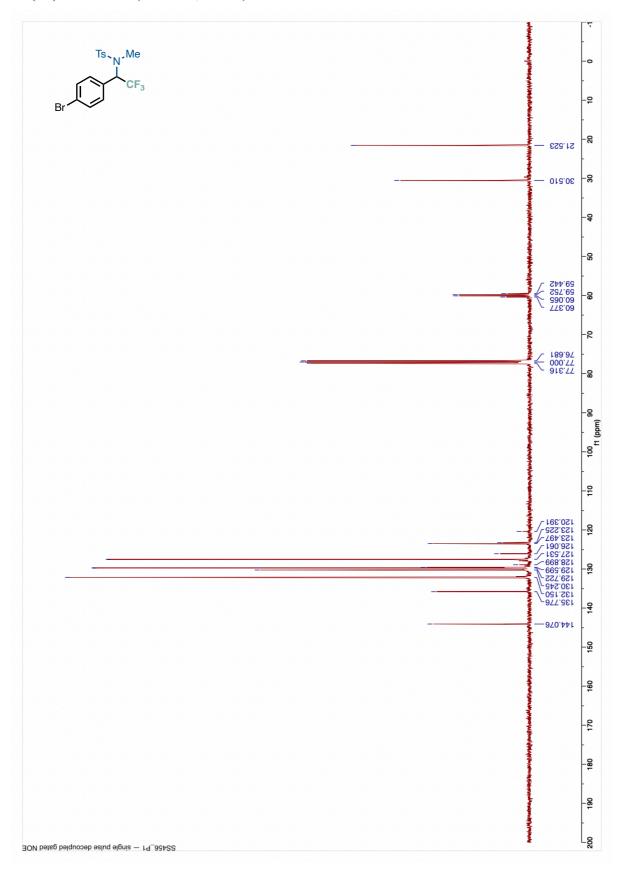
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3E}$ (101 MHz, CDCl₃)

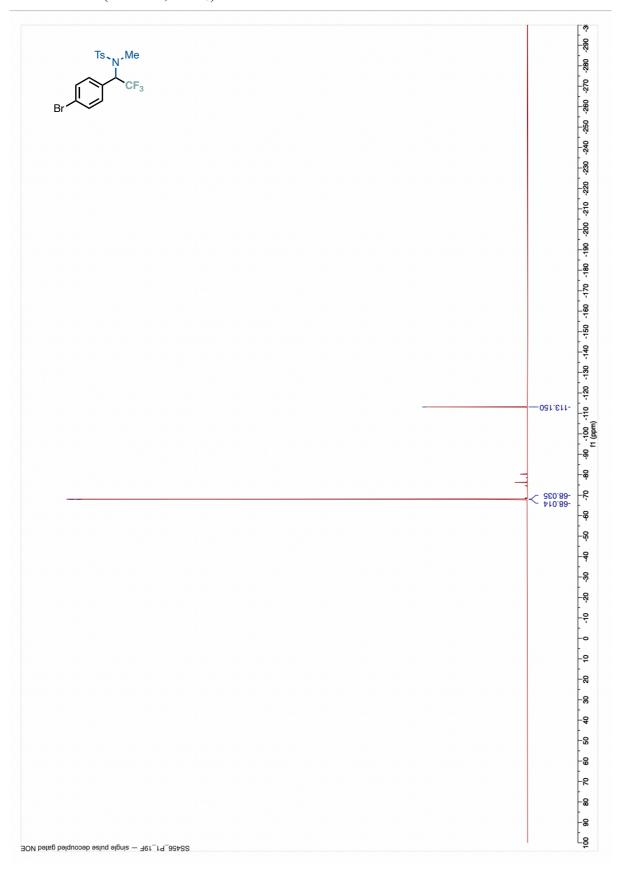


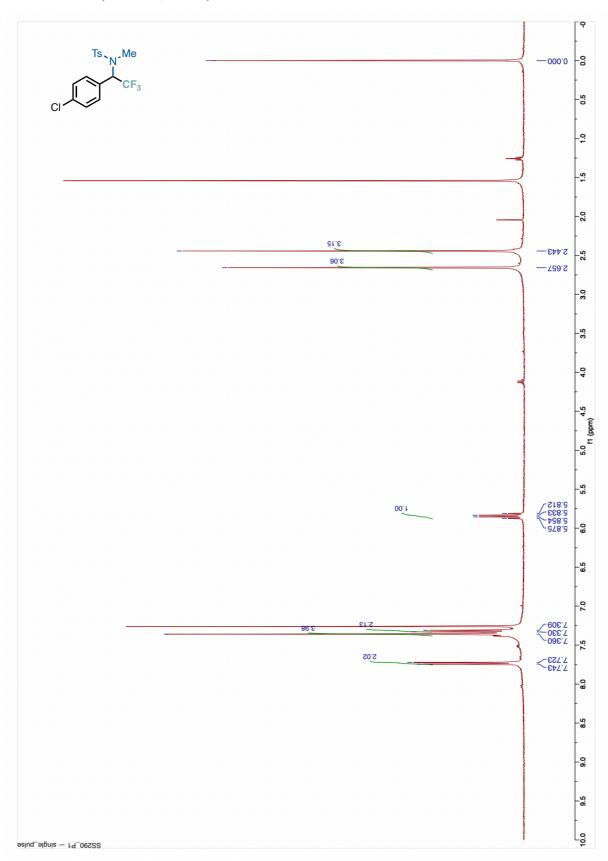




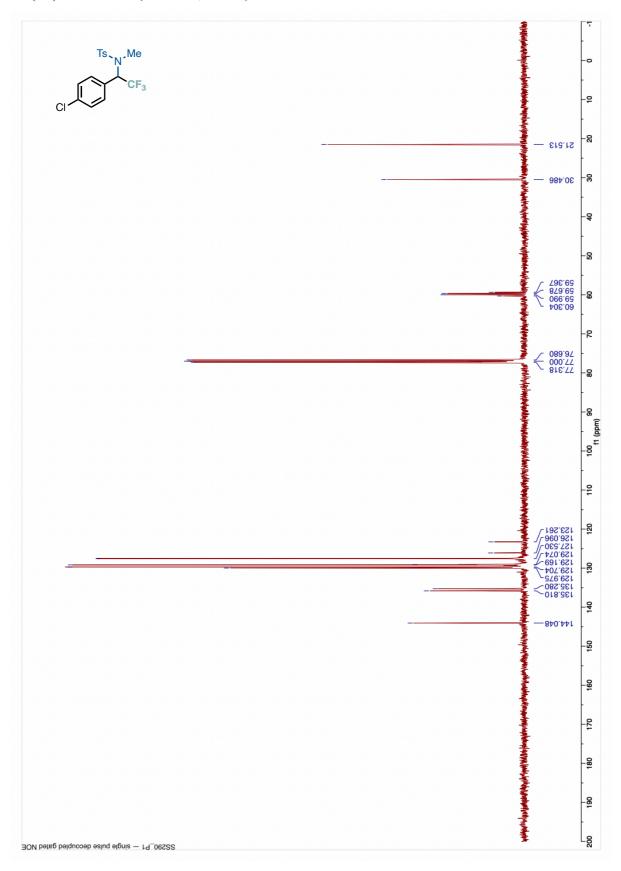
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3F}$ (101 MHz, CDCl₃)

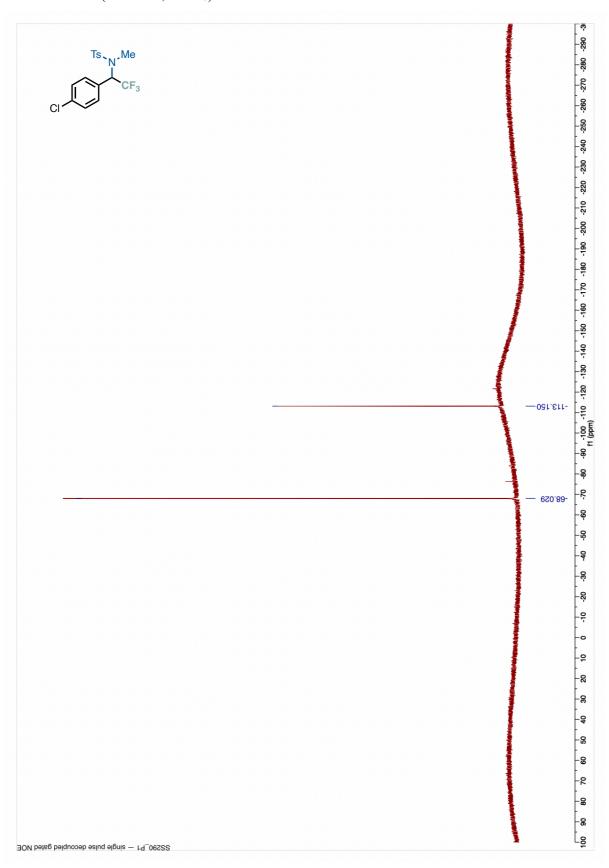


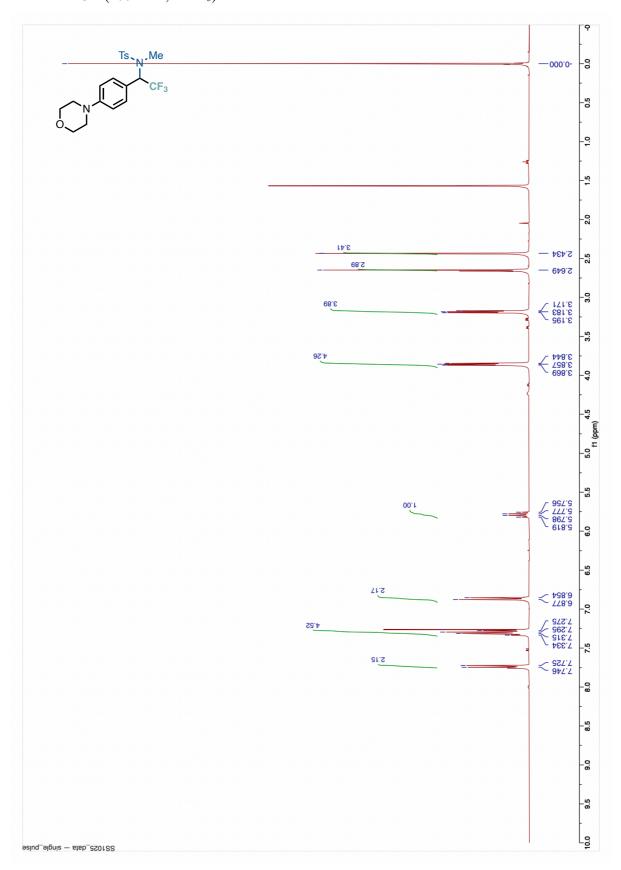




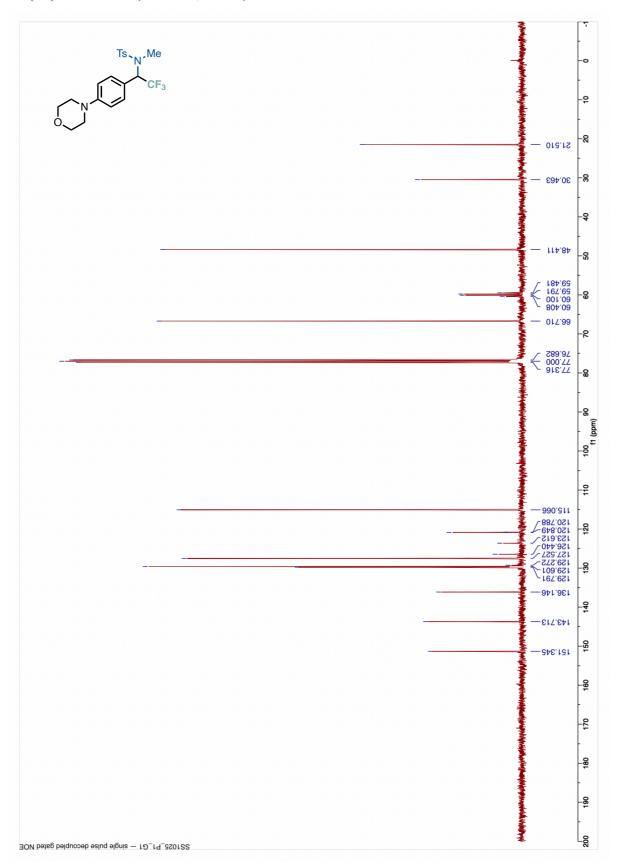
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3}\pmb{G}$ (101 MHz, CDCl₃)

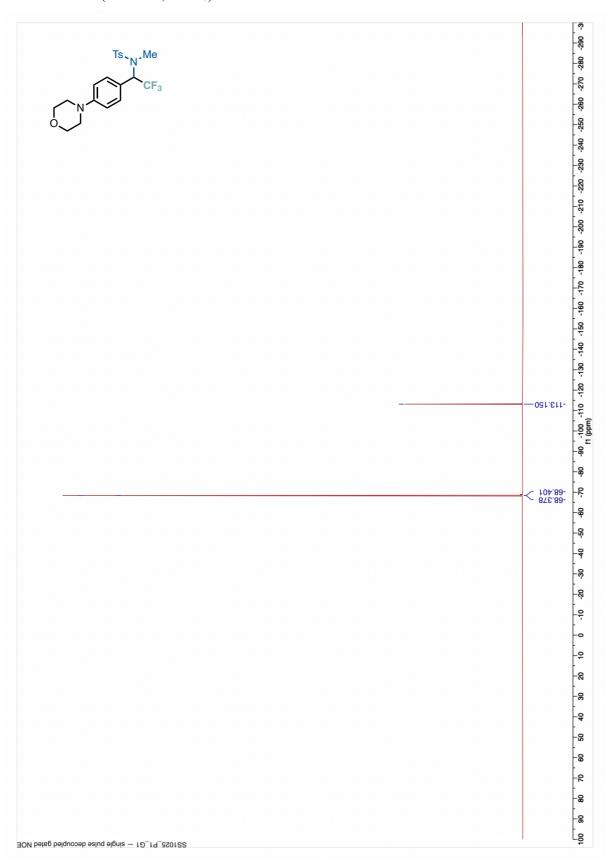


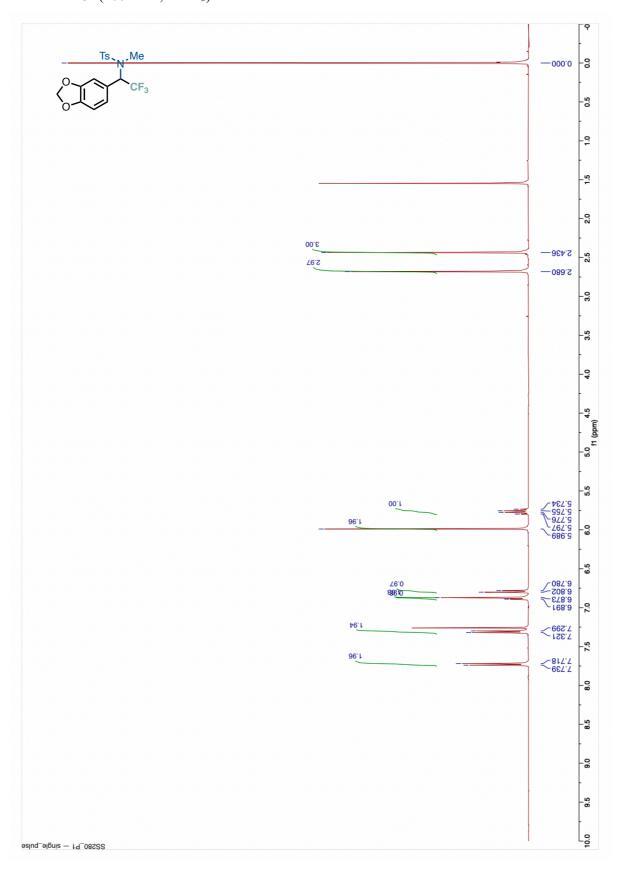




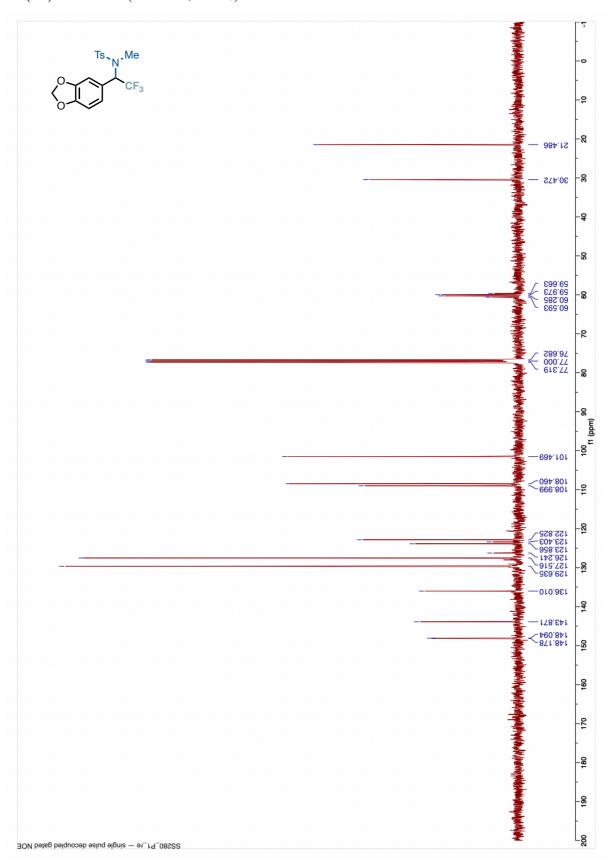
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3H}$ (101 MHz, CDCl₃)

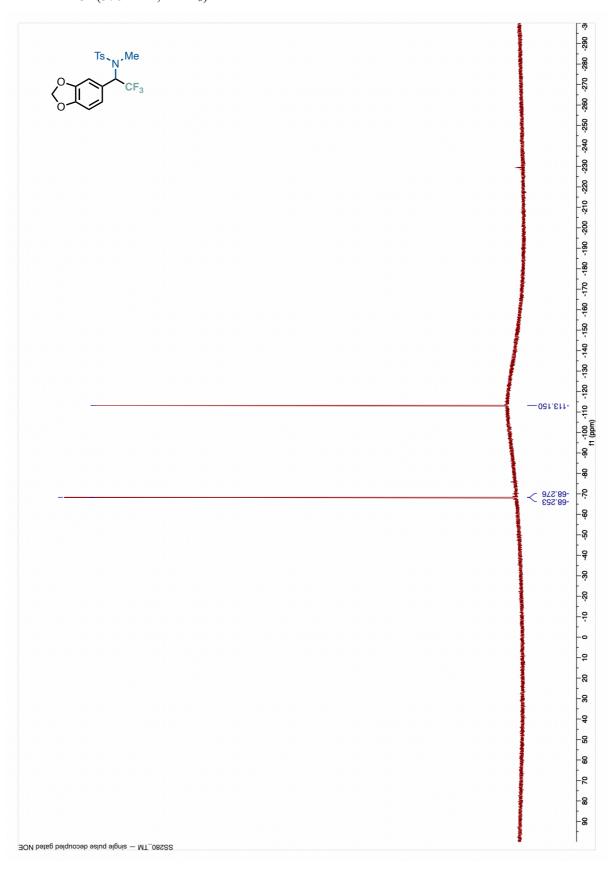


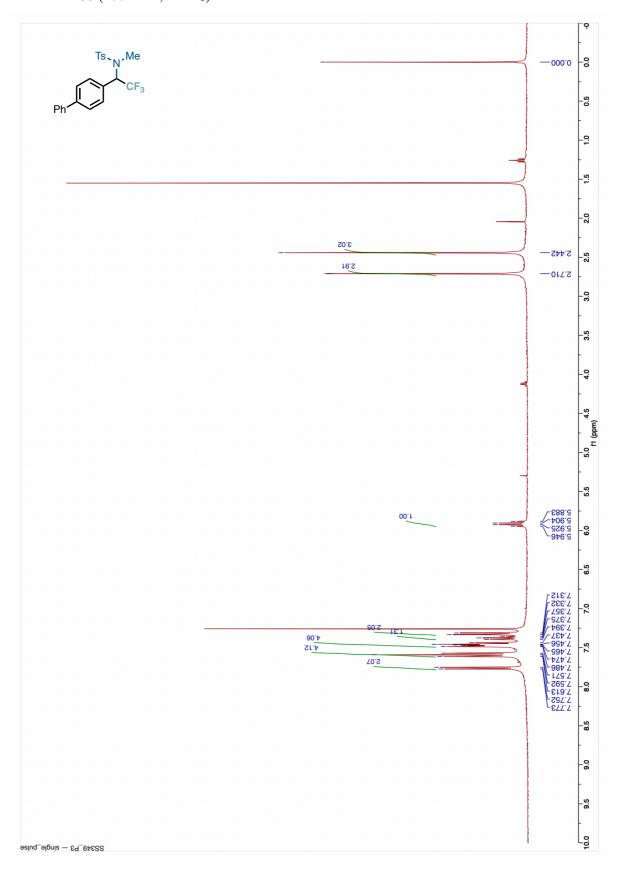




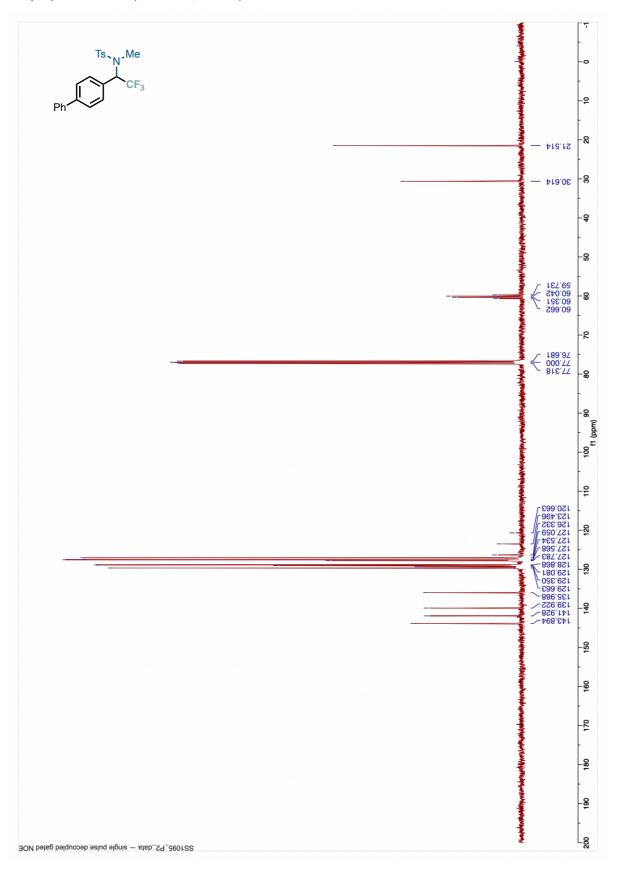
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3I}$ (101 MHz, CDCl₃)

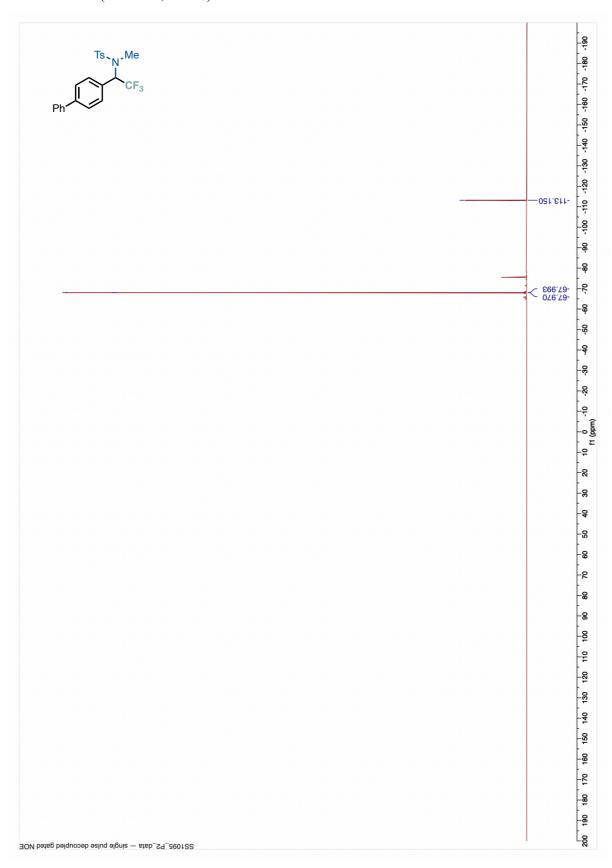


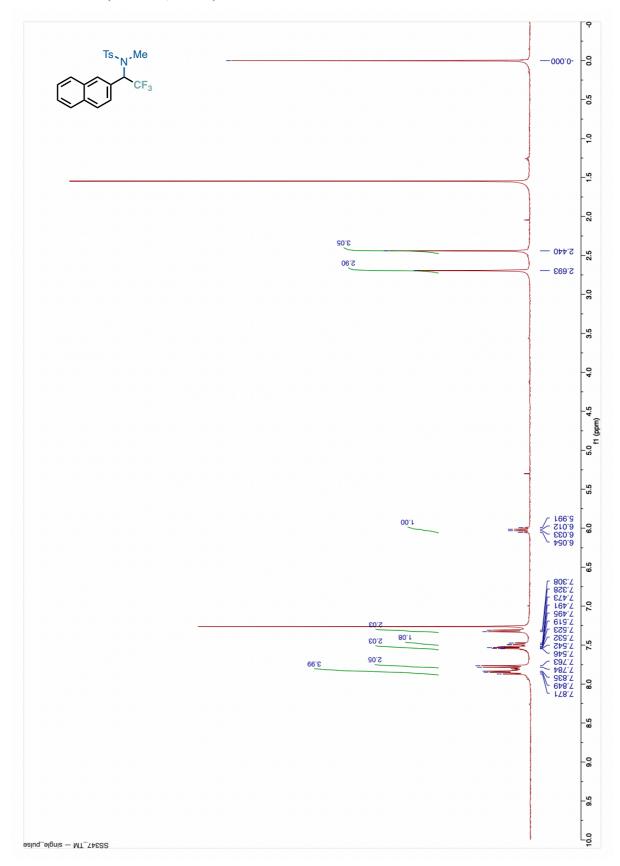




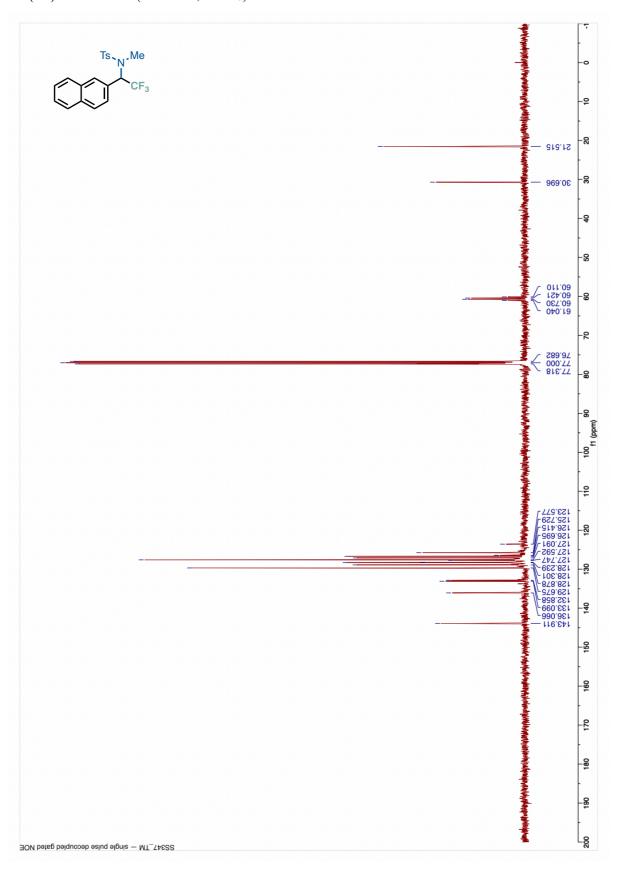
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3J}$ (101 MHz, CDCl₃)

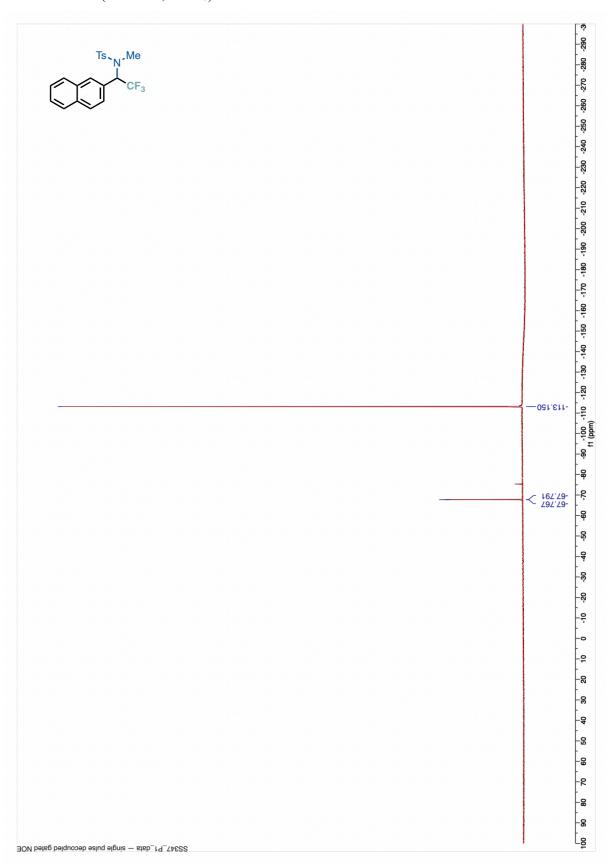


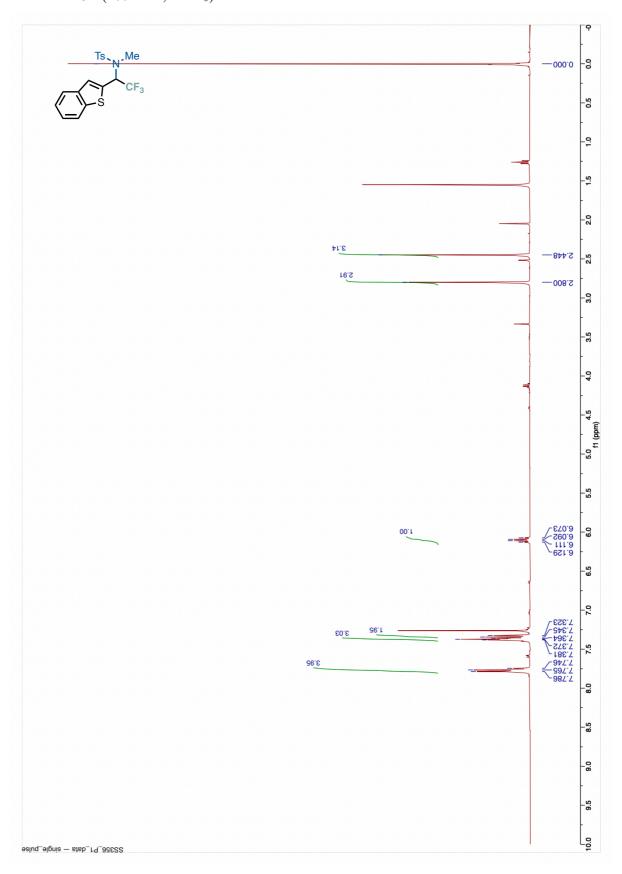




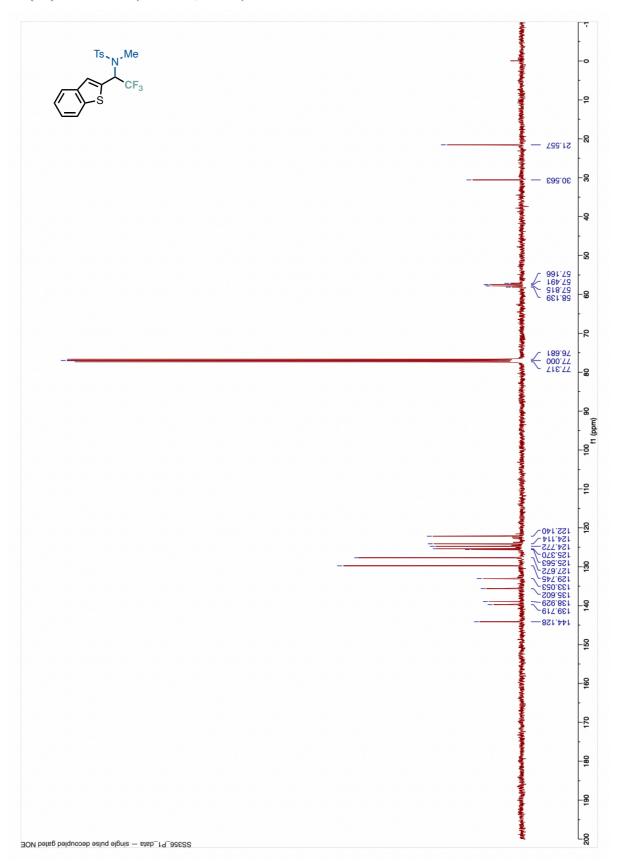
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3K}$ (101 MHz, CDCl₃)

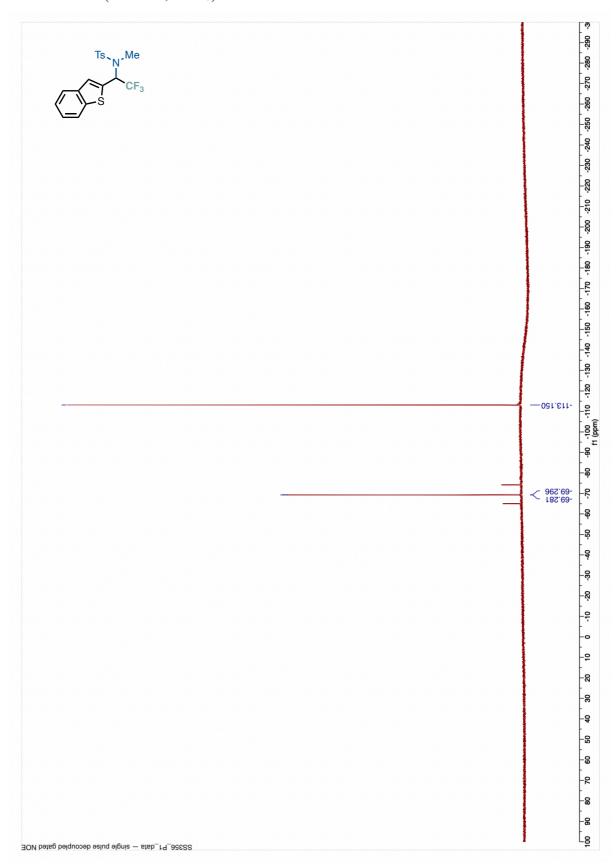


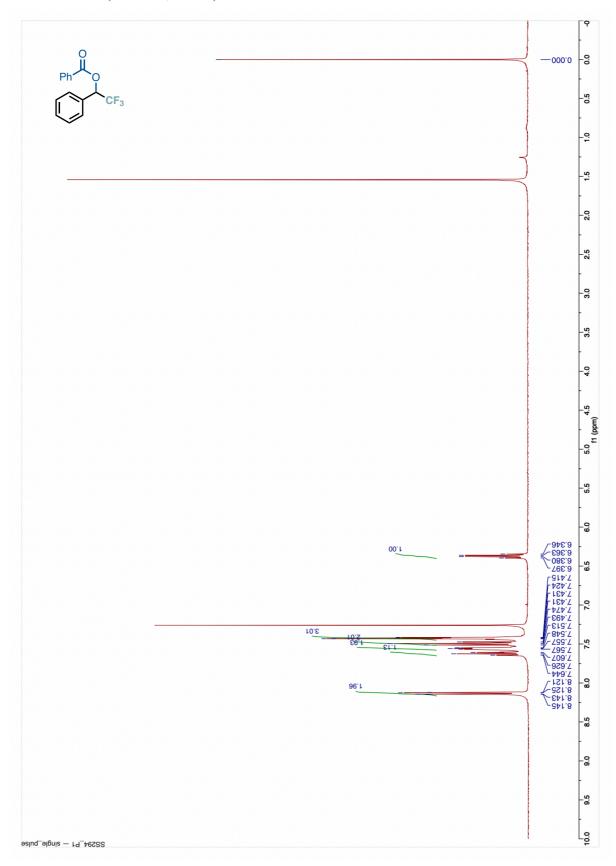




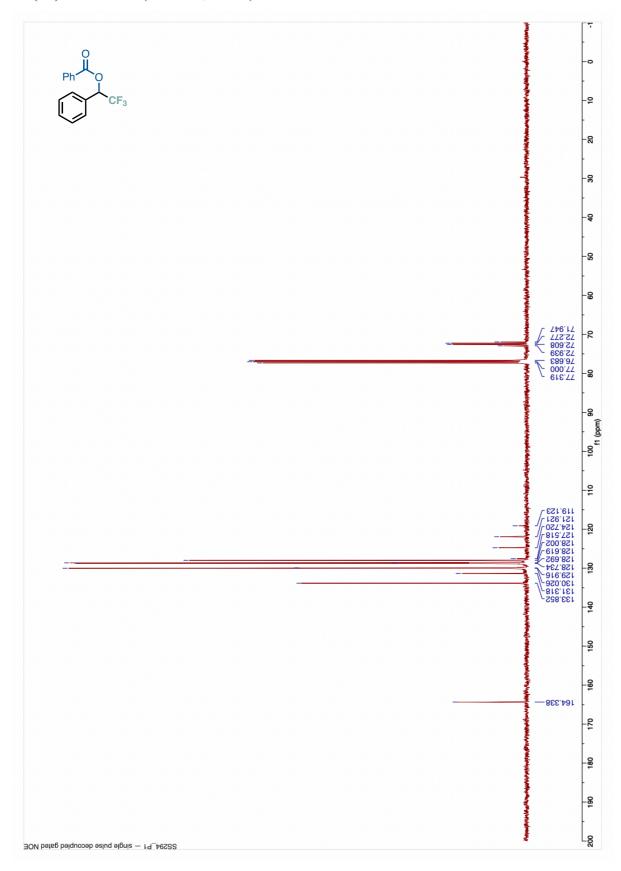
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3L}$ (101 MHz, CDCl₃)

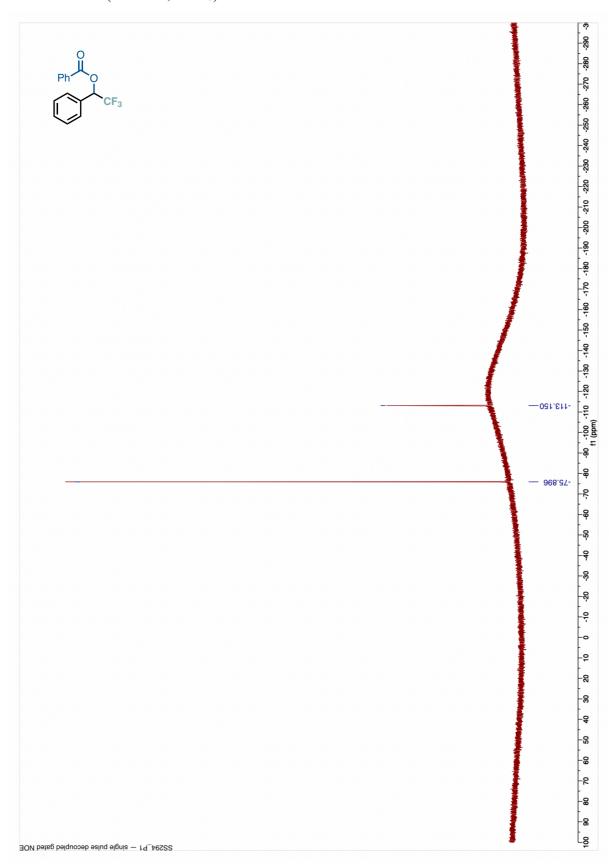


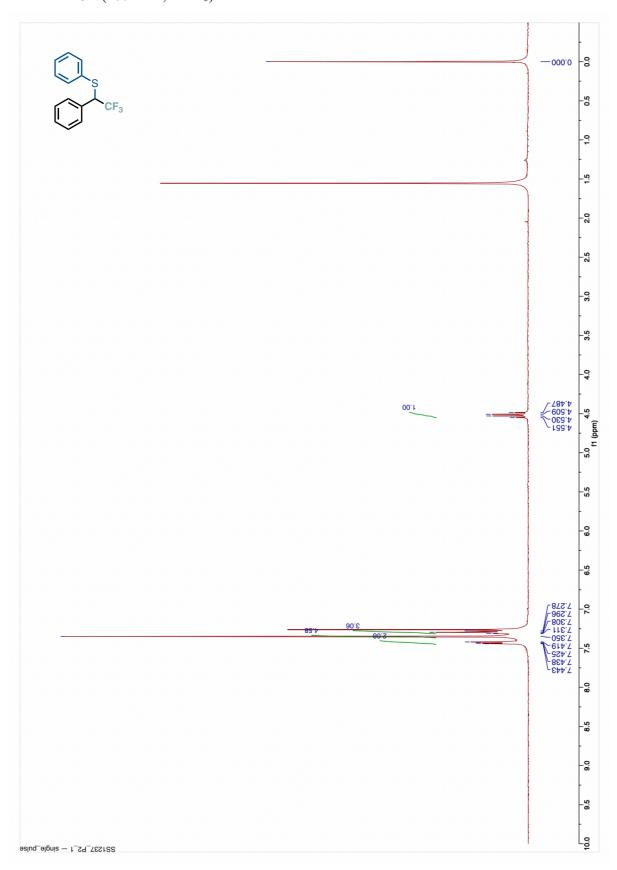




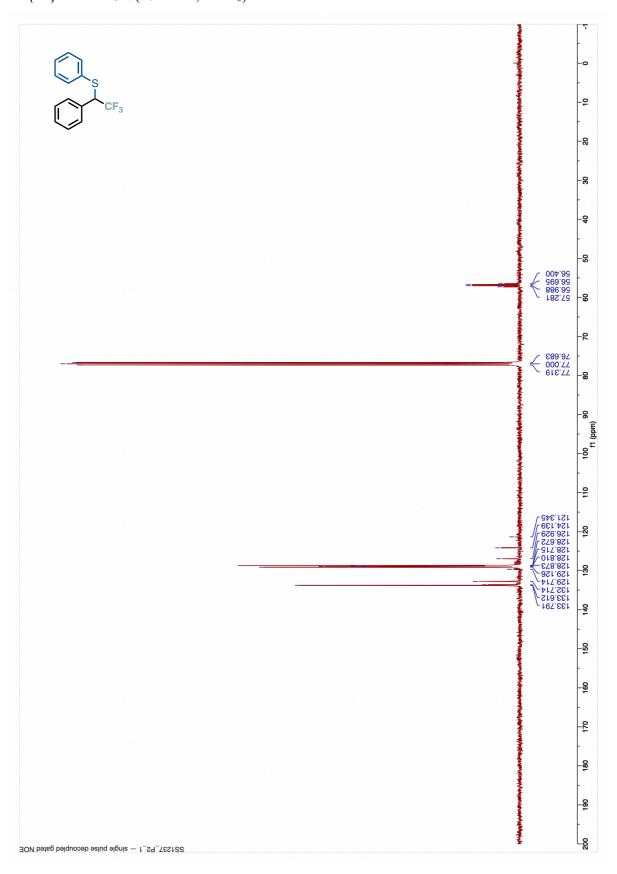
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\pmb{3M}$ (101 MHz, CDCl₃)

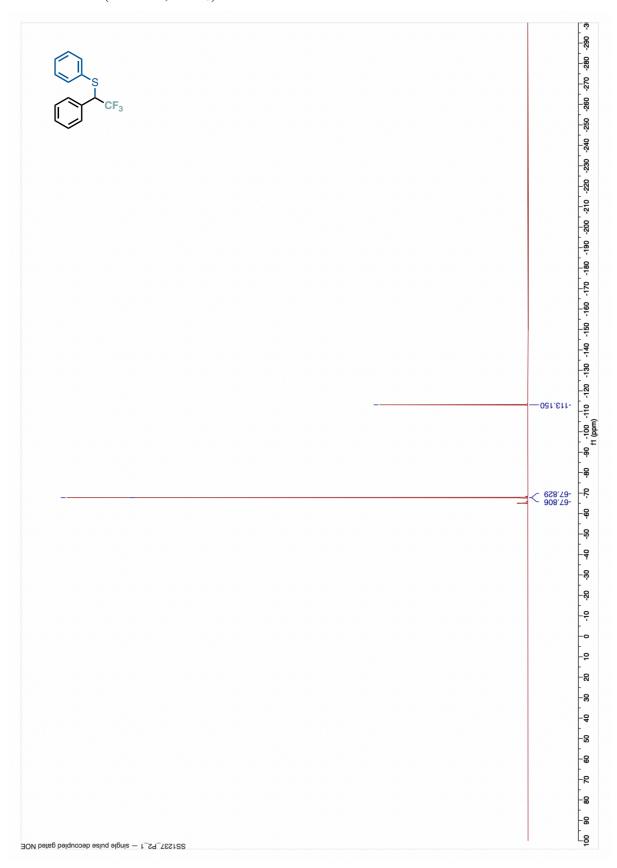


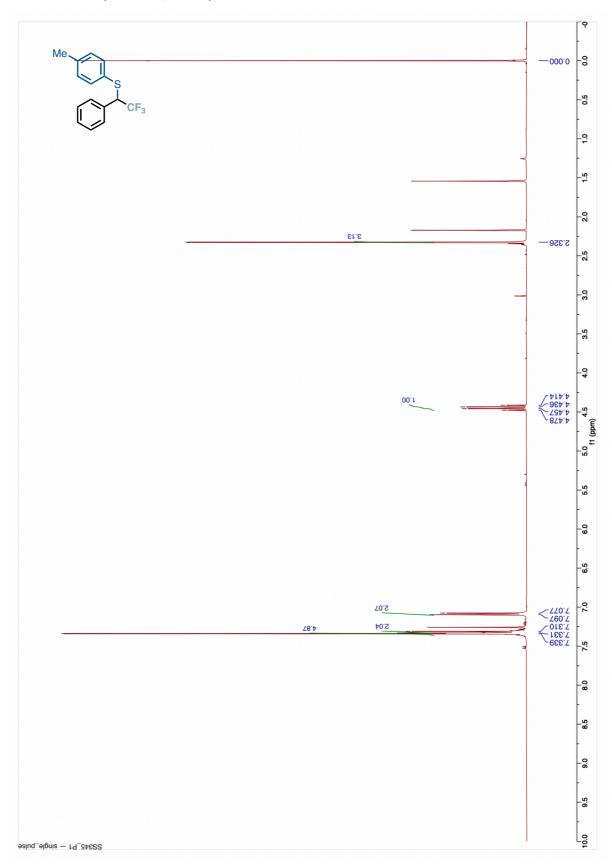




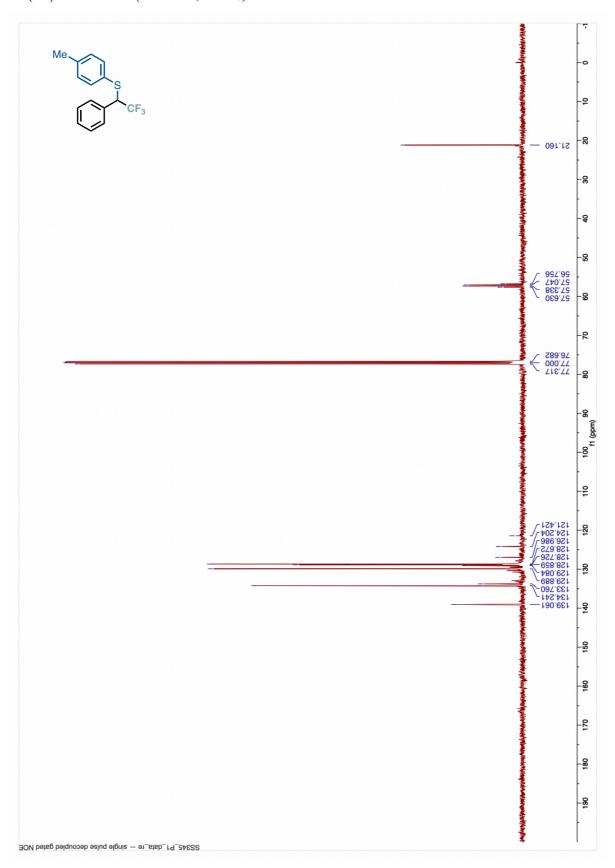
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3N}$ (101 MHz, CDCl₃)

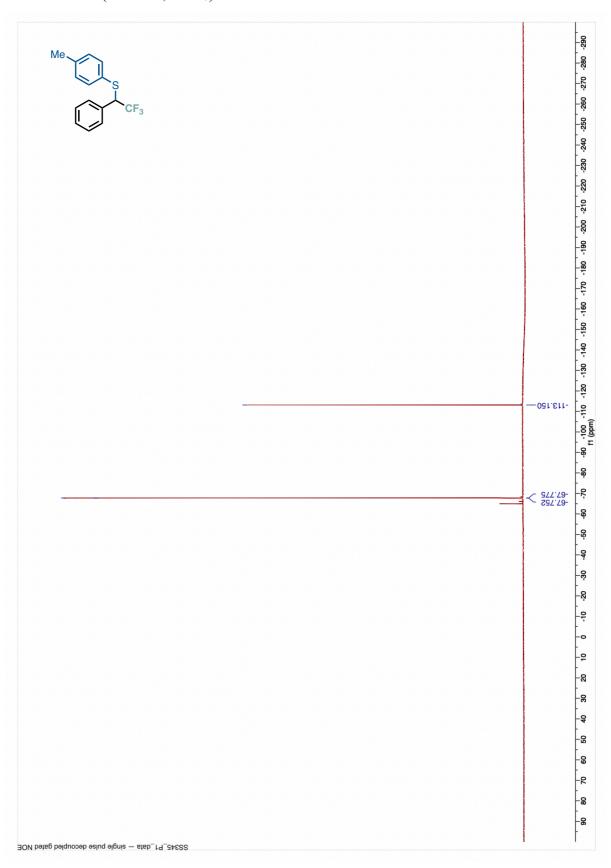


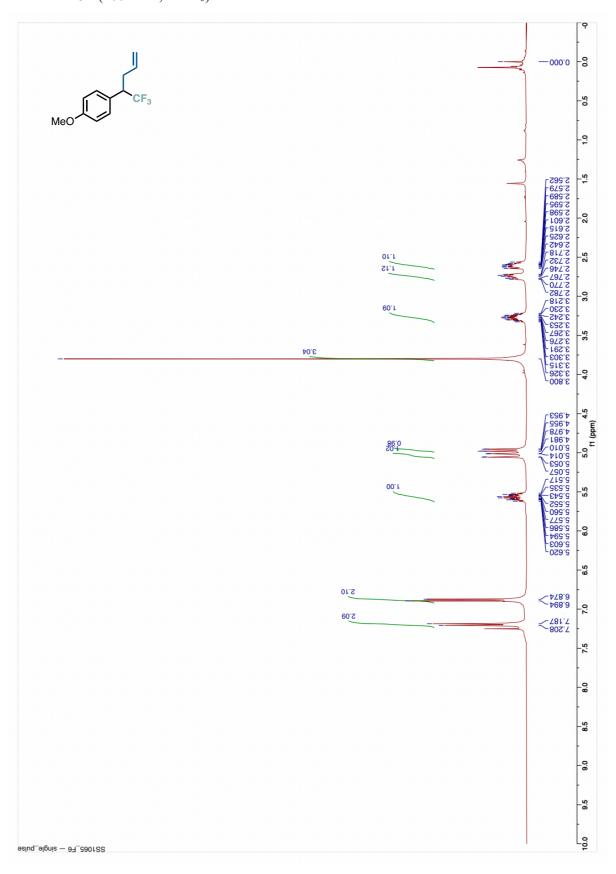




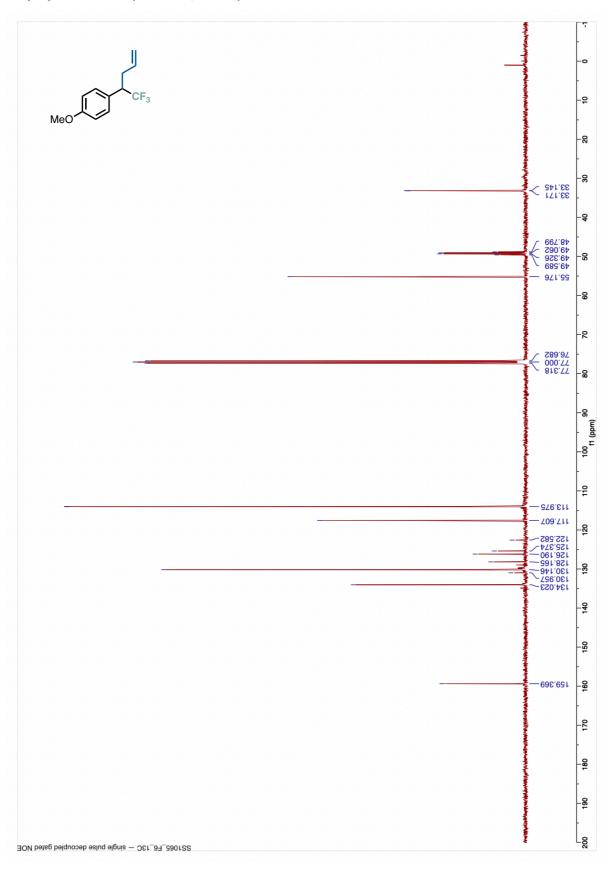
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{30}$ (101 MHz, CDCl₃)

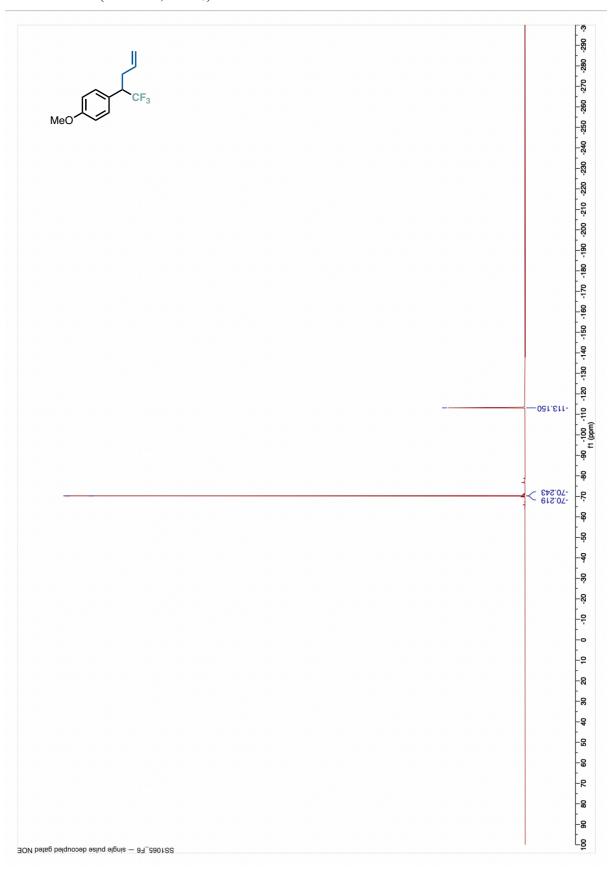


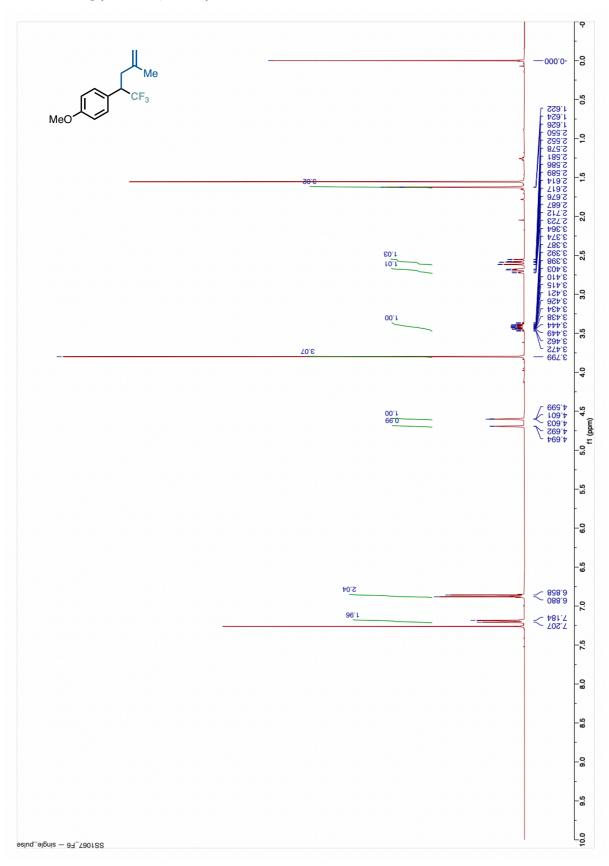




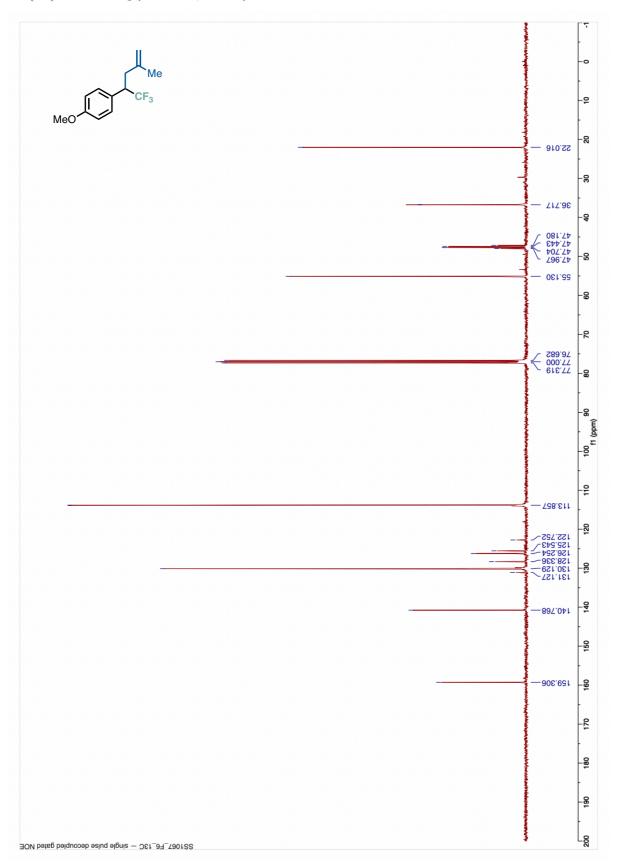
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3P}$ (101 MHz, CDCl₃)

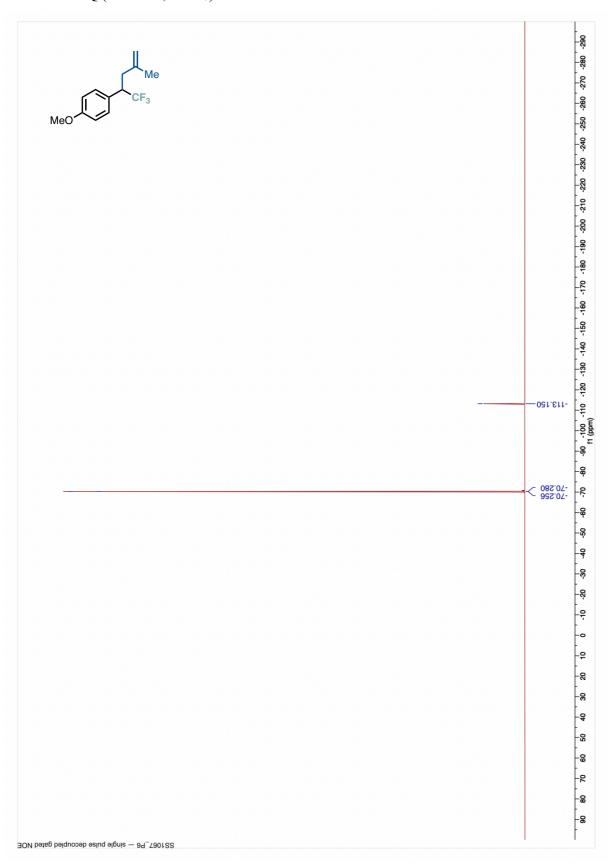




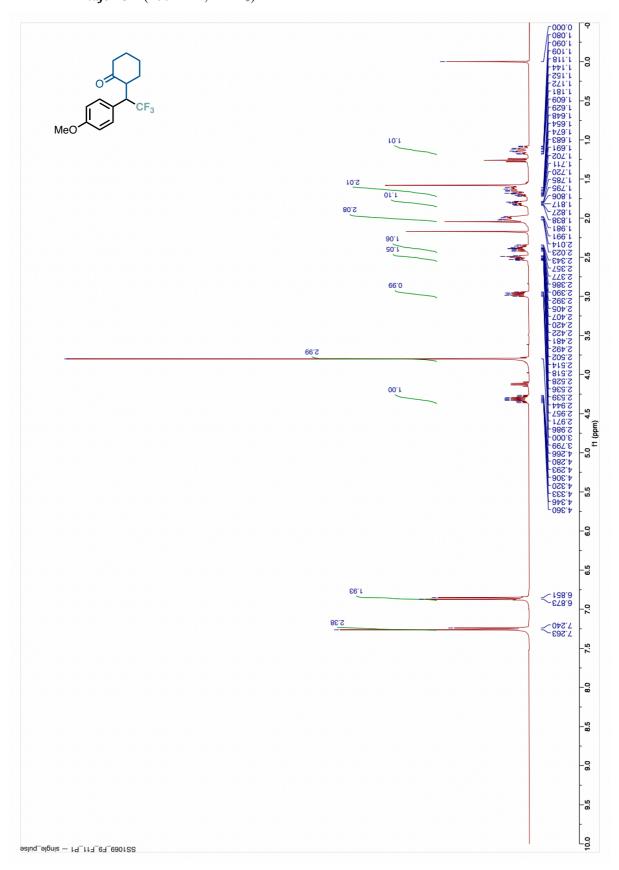


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3Q}$ (101 MHz, CDCl₃)

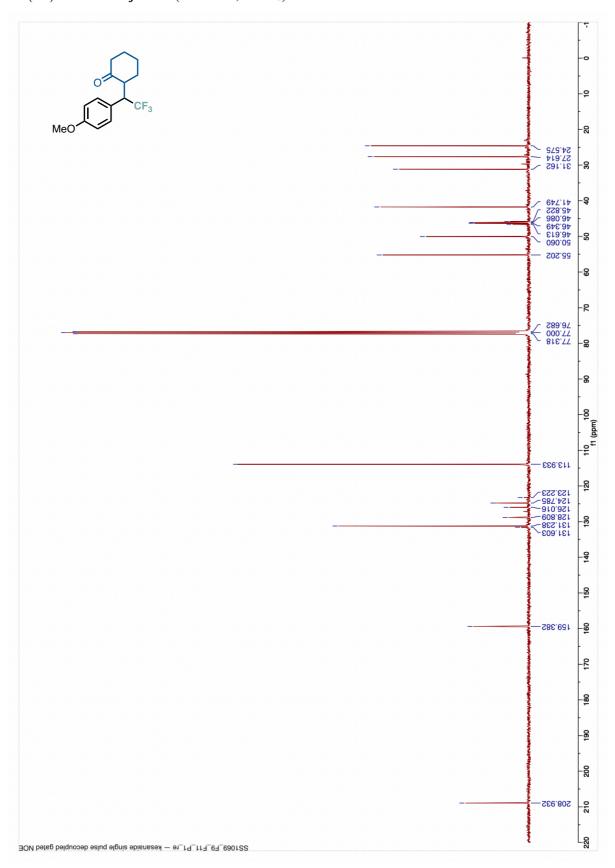


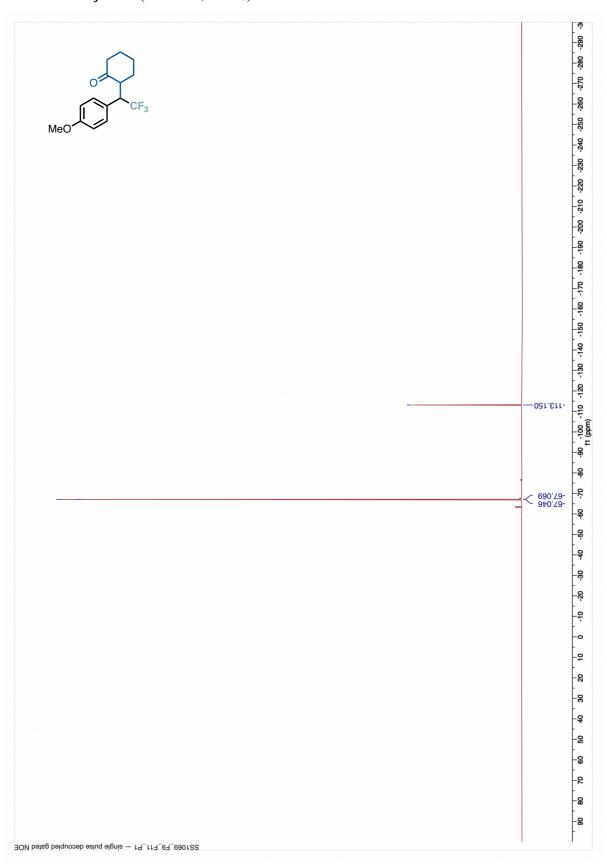


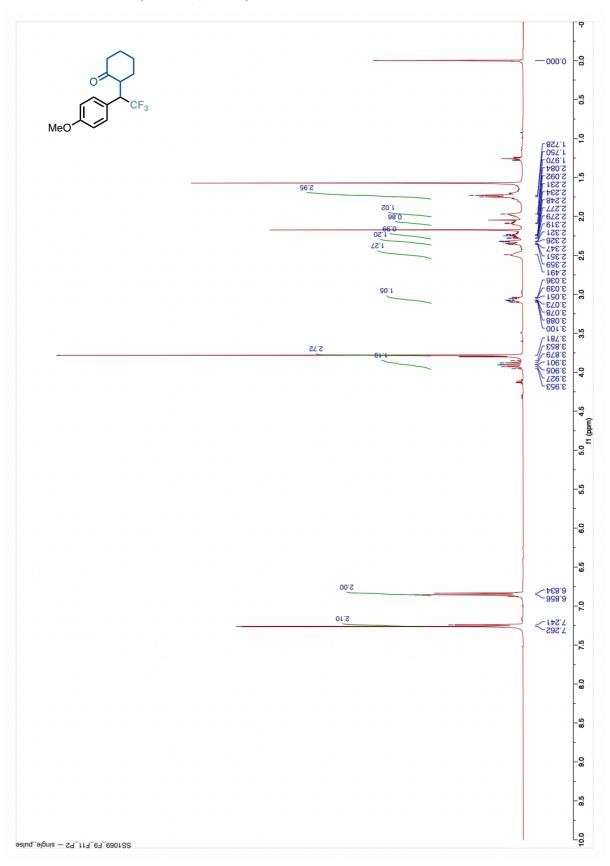
¹H NMR of *major-3R* (400 MHz, CDCl₃)



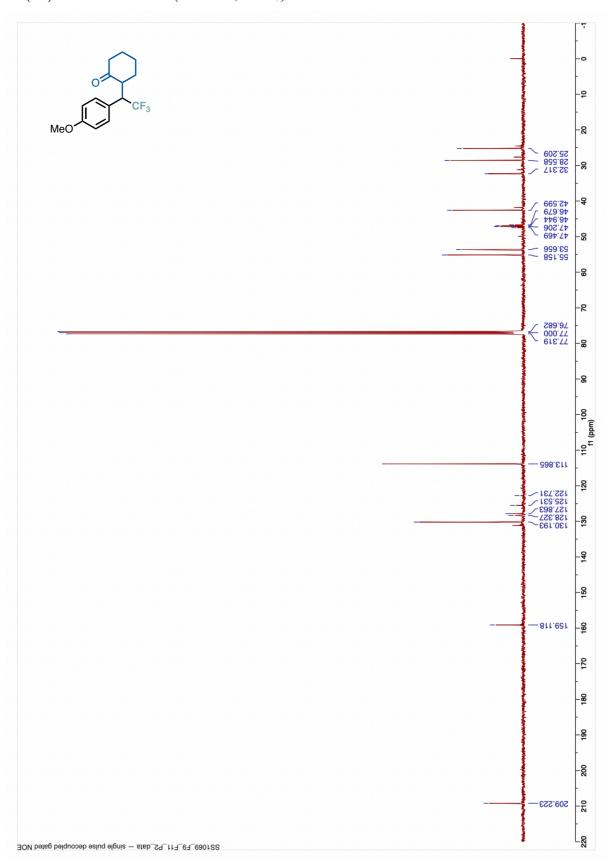
 $^{13}\text{C}\{^1\text{H}\}$ NMR of major-3R (101 MHz, CDCl₃)

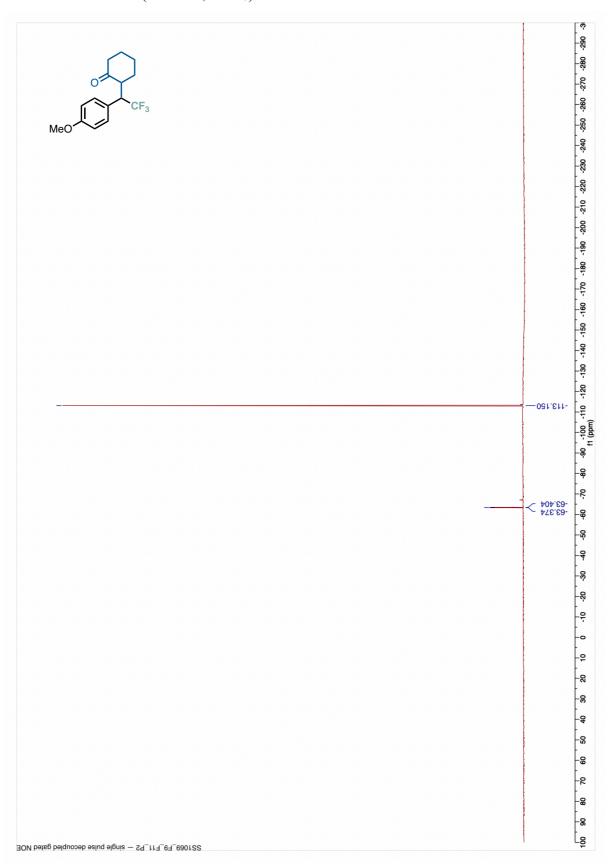


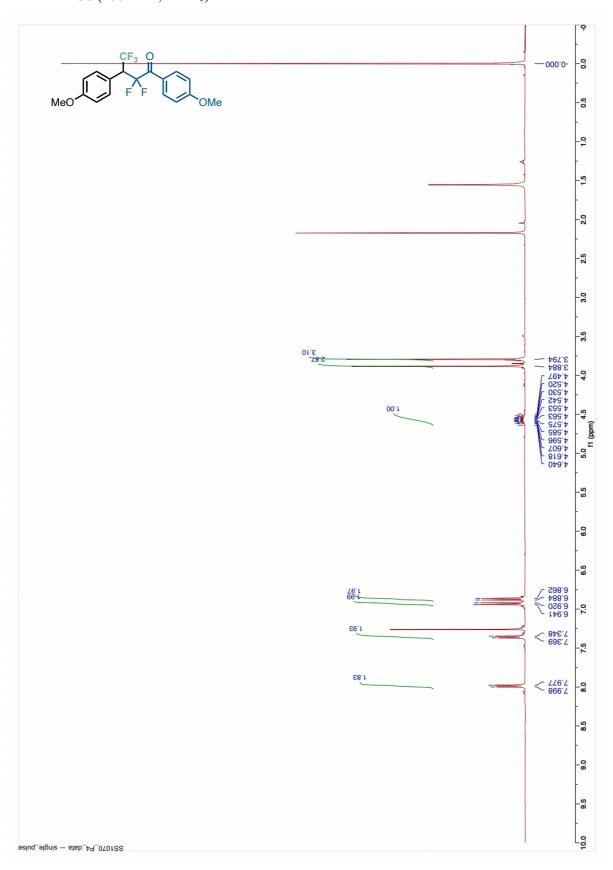




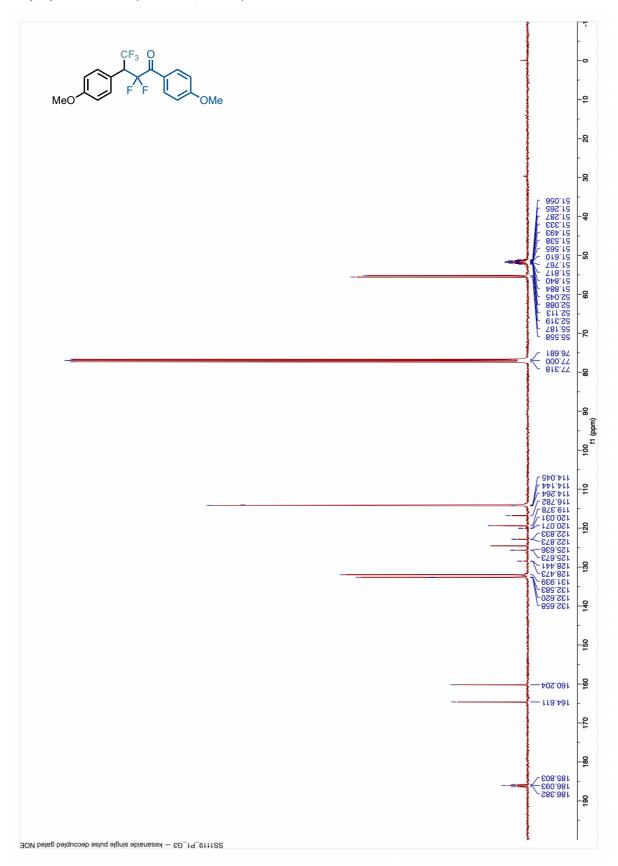
 $^{13}\text{C}\{^1\text{H}\}$ NMR of minor-3R (101 MHz, CDCl₃)

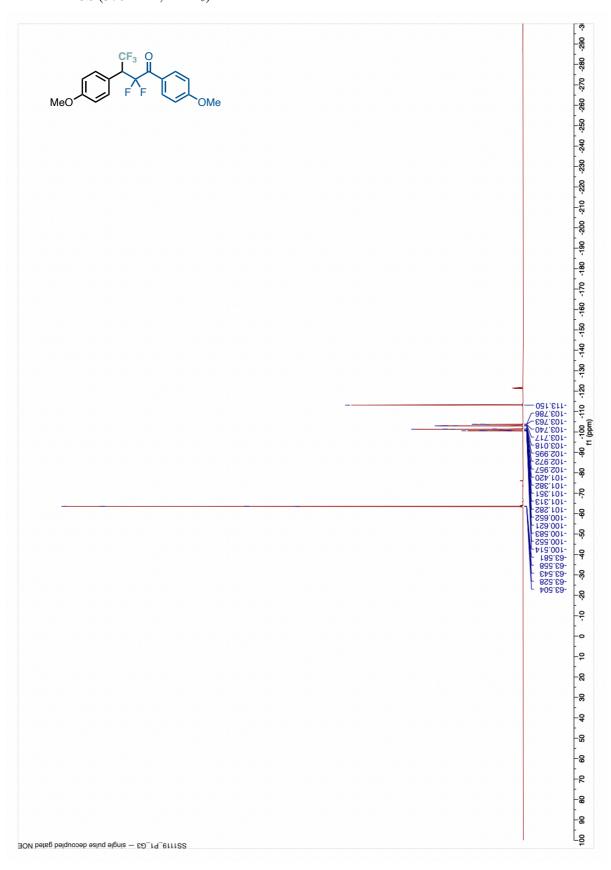


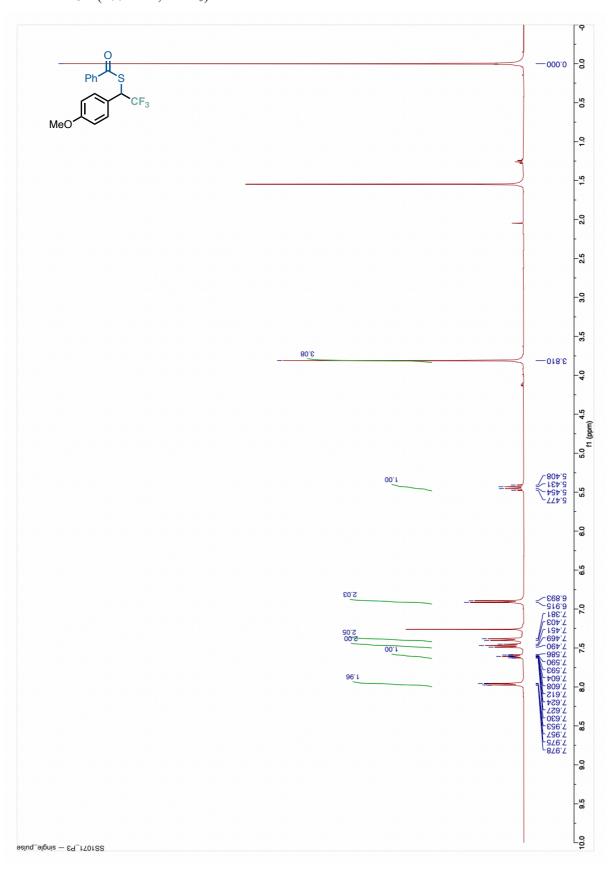




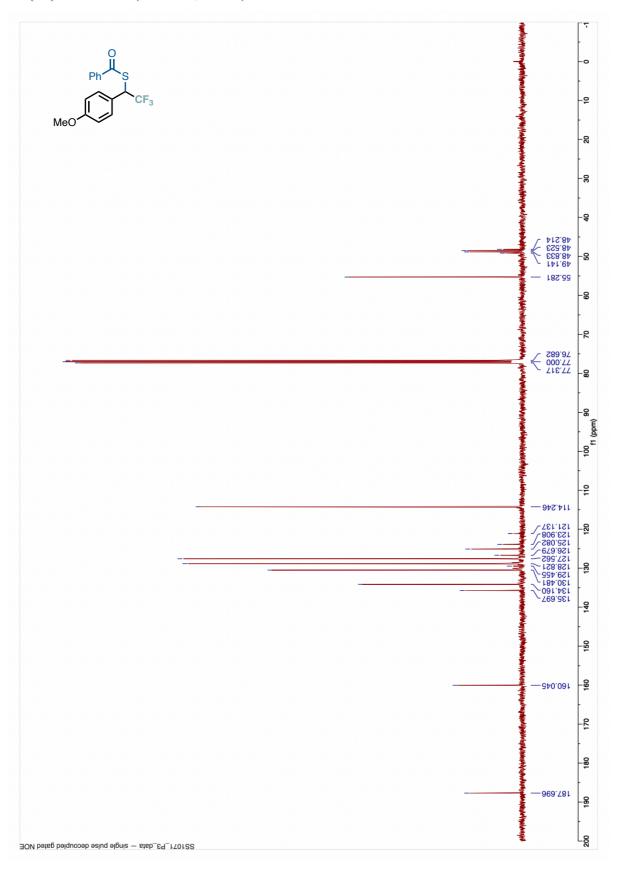
 $^{13}C\{^{1}H\}$ NMR of **3V** (101 MHz, CDCl₃)

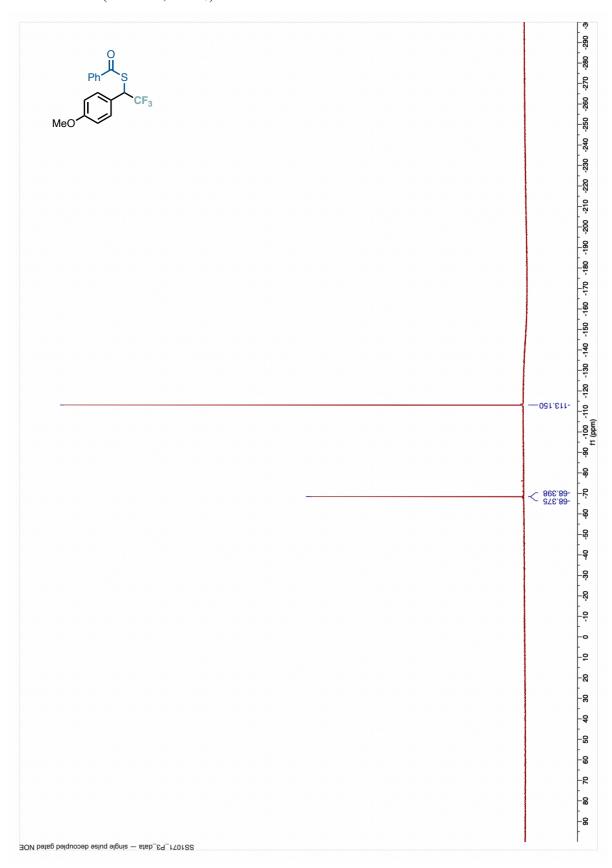


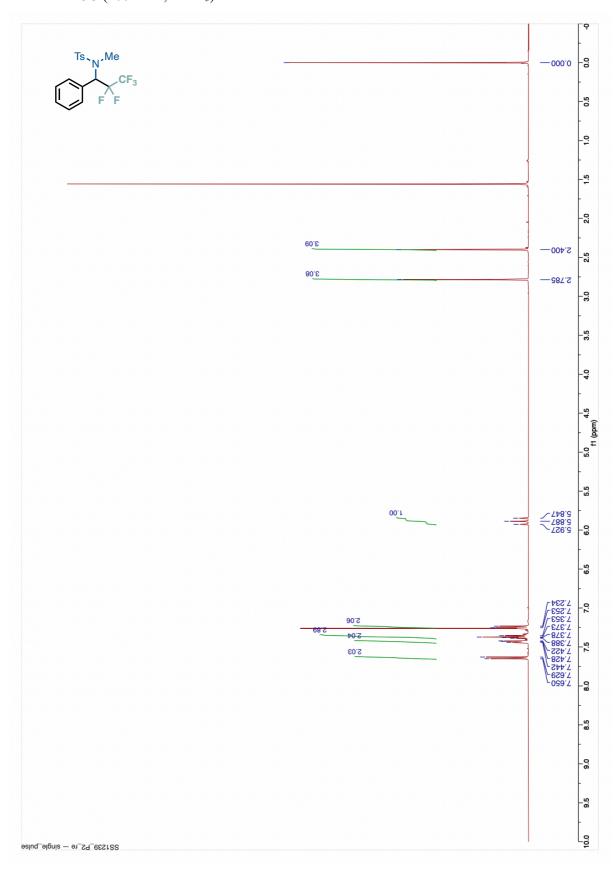




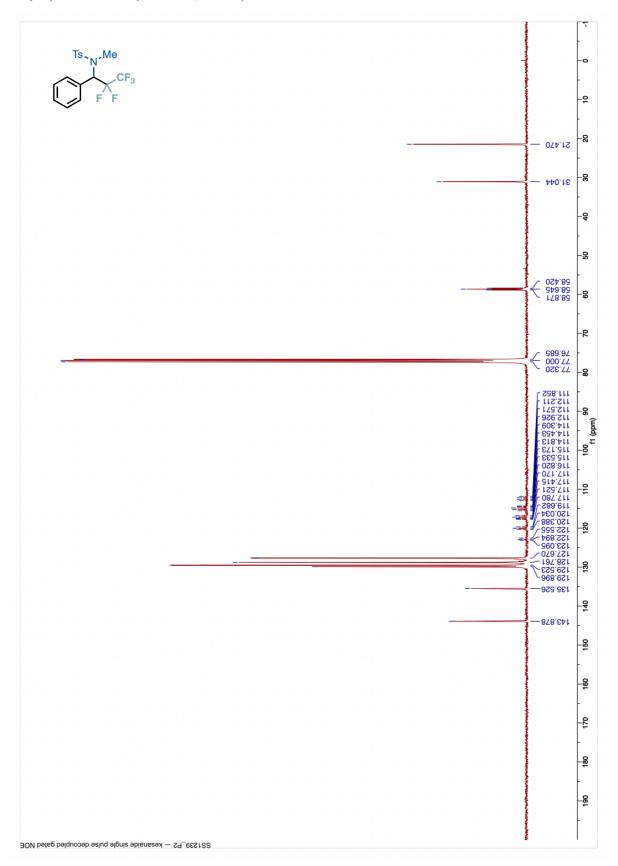
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3T}$ (101 MHz, CDCl₃)

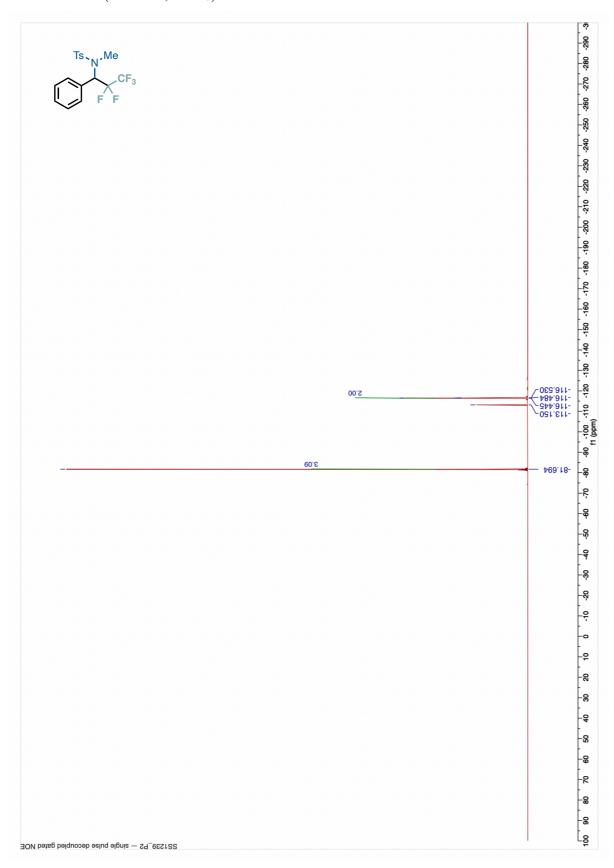


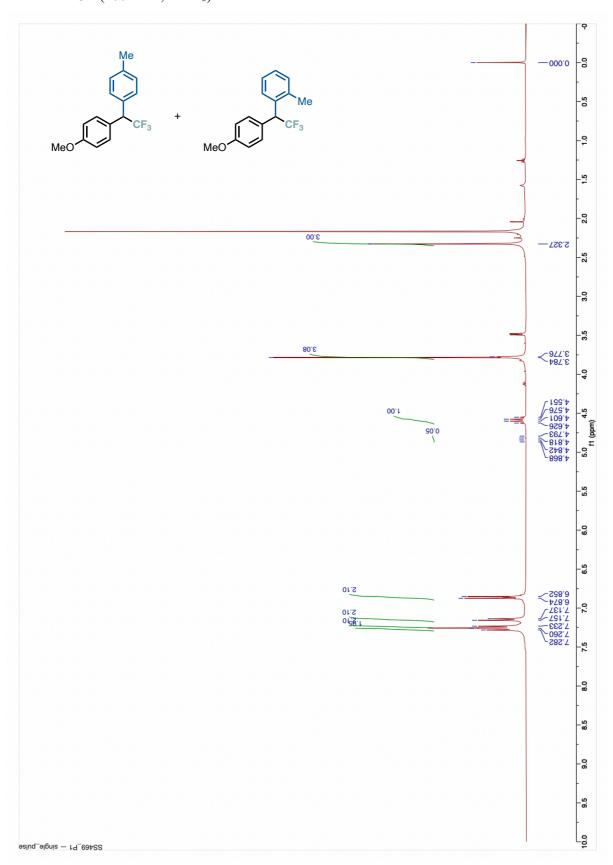




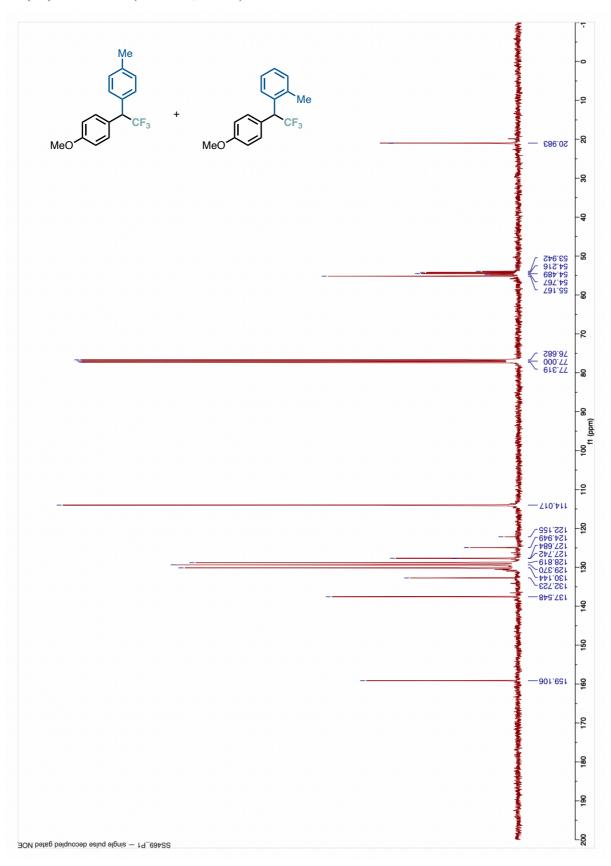
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{3}\boldsymbol{U}$ (101 MHz, CDCl₃)

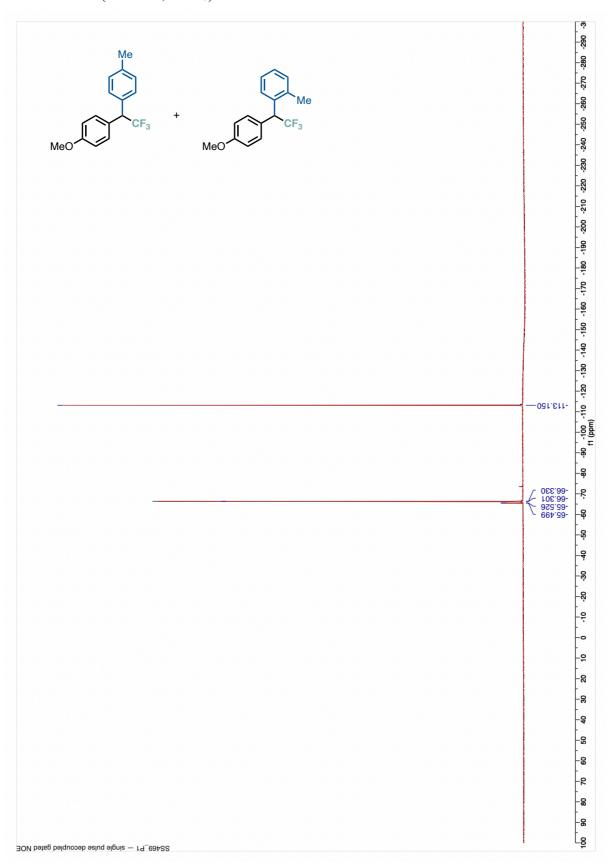




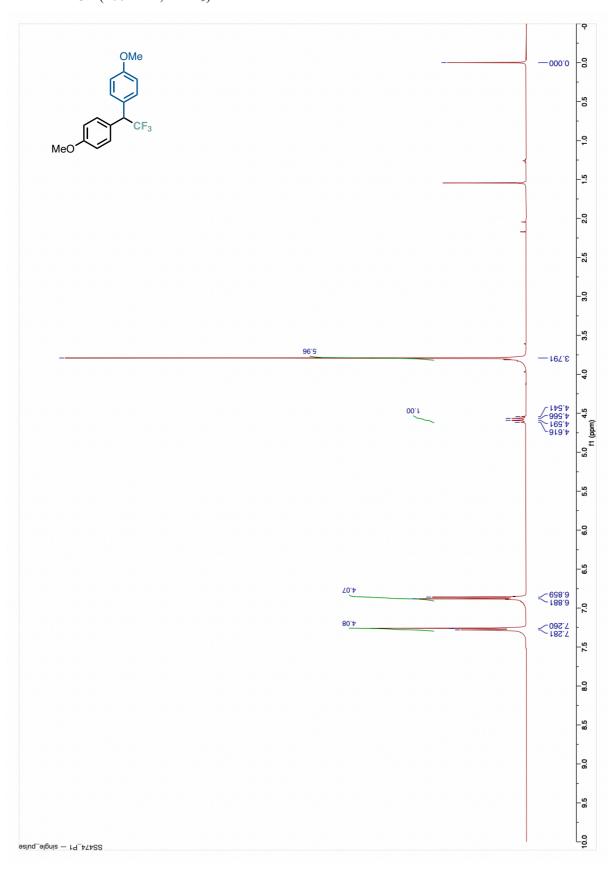


 $^{13}\text{C}\{^1\text{H}\}$ NMR of 5A (101 MHz, CDCl₃)

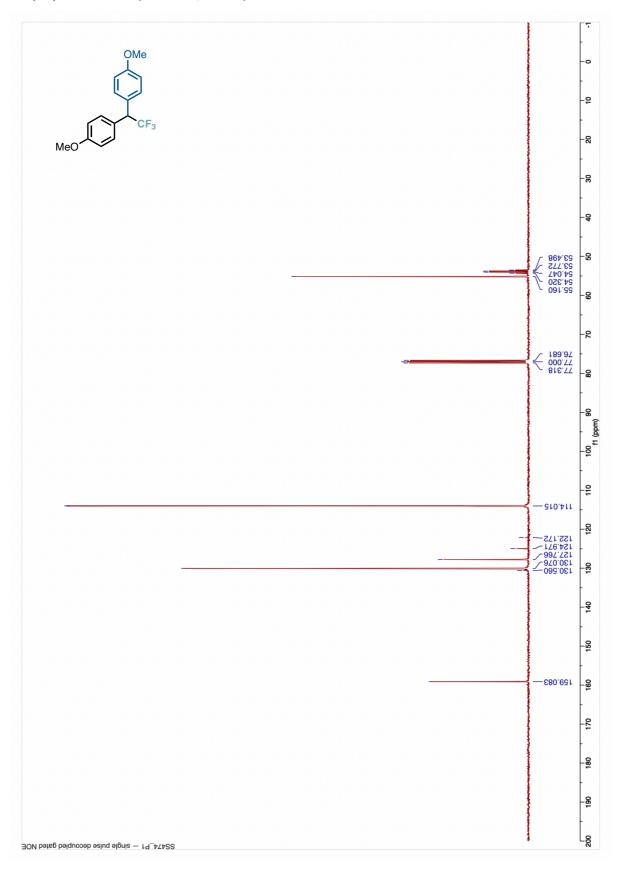


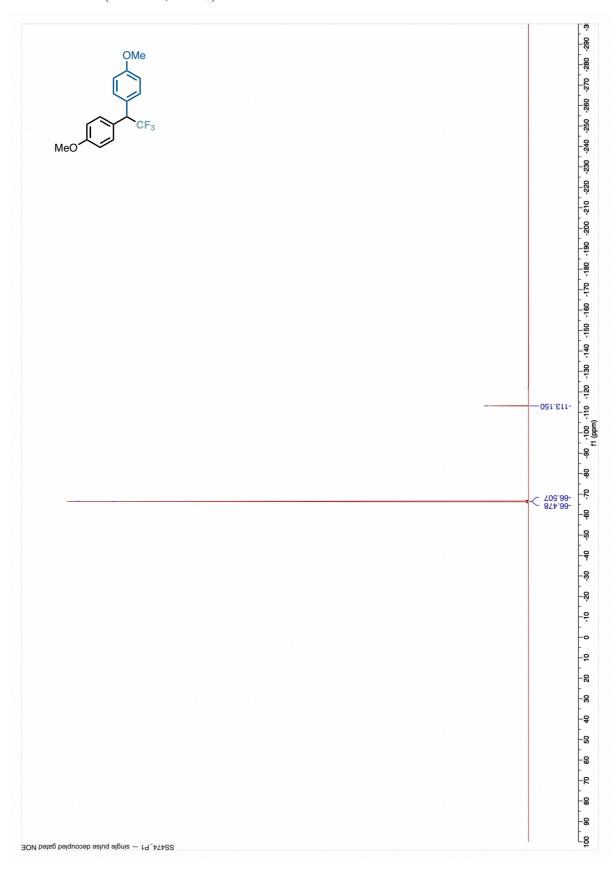


1 H NMR of **5B** (400 MHz, CDCl₃)

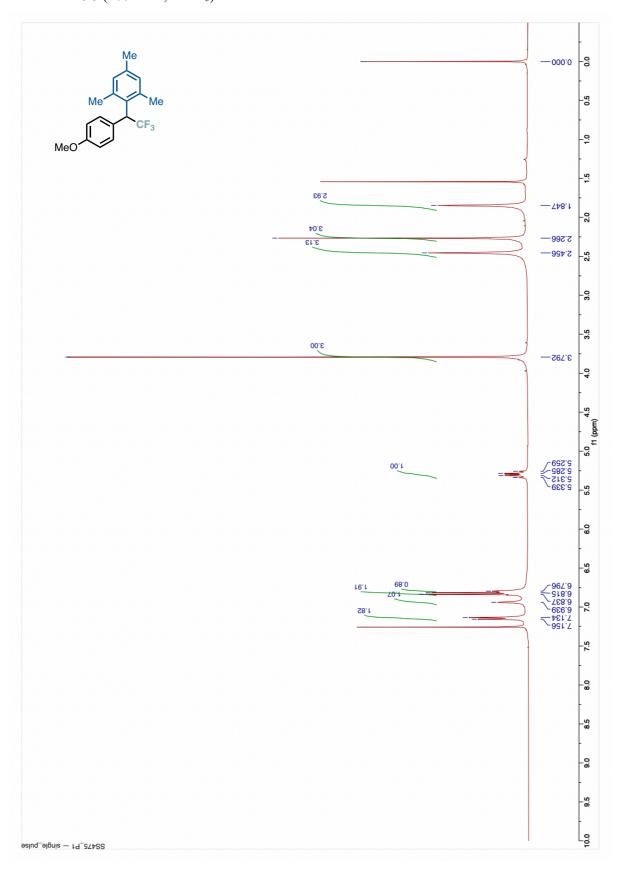


 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{5B}$ (101 MHz, CDCl₃)

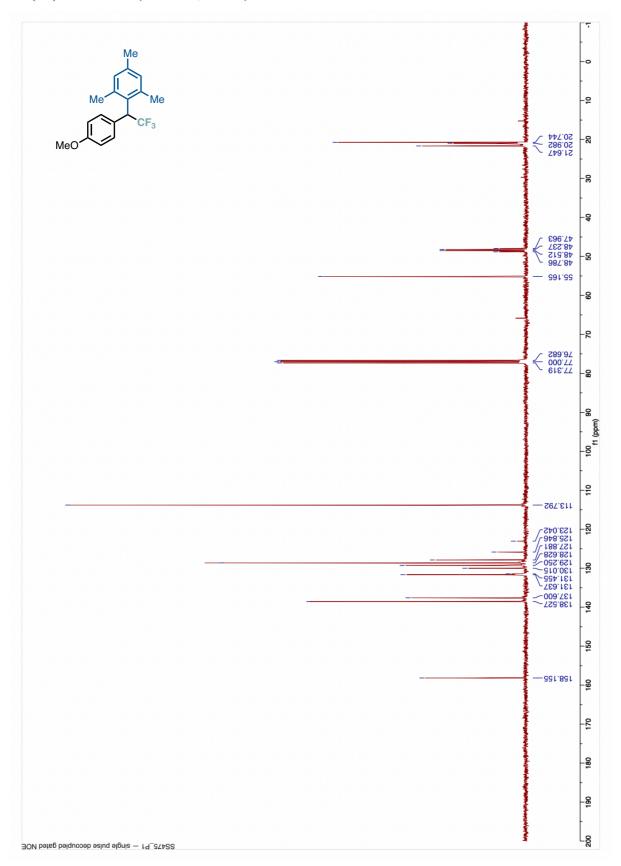


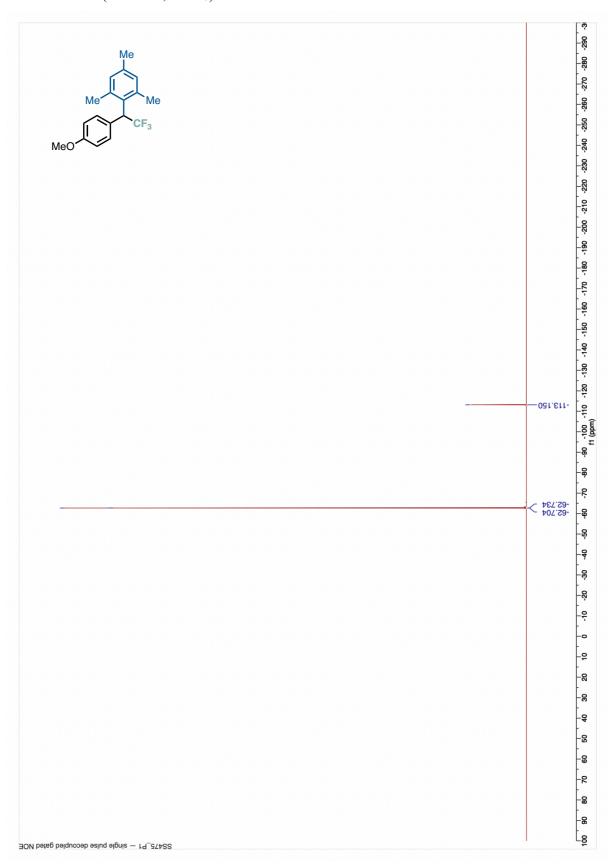


¹H NMR of **5C** (400 MHz, CDCl₃)

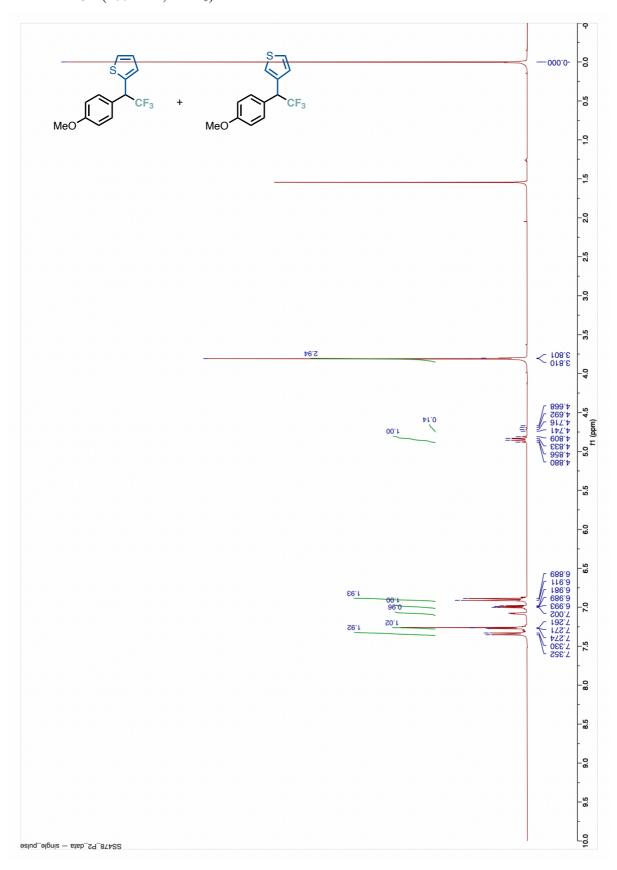


 $^{13}\text{C}\{^1\text{H}\}$ NMR of 5C (101 MHz, CDCl₃)

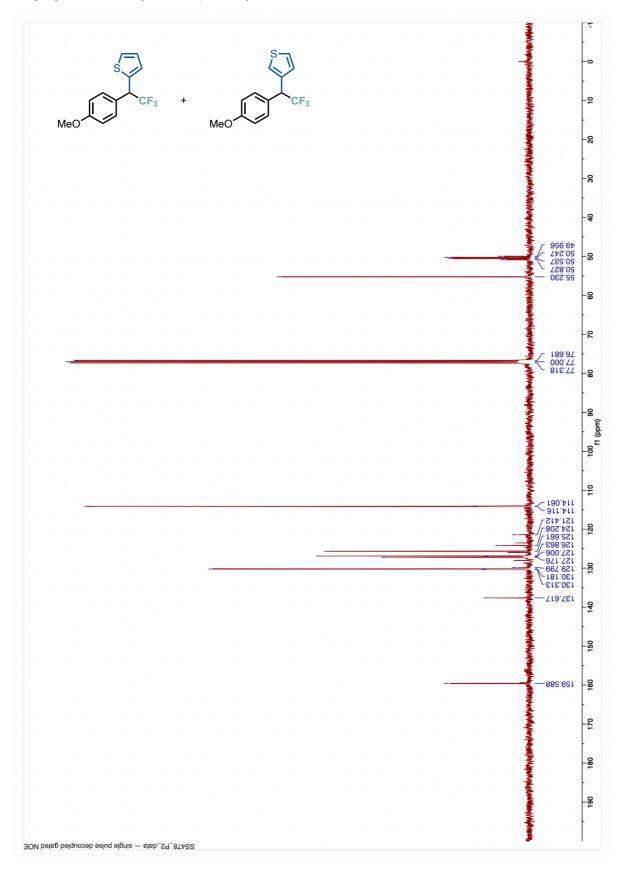


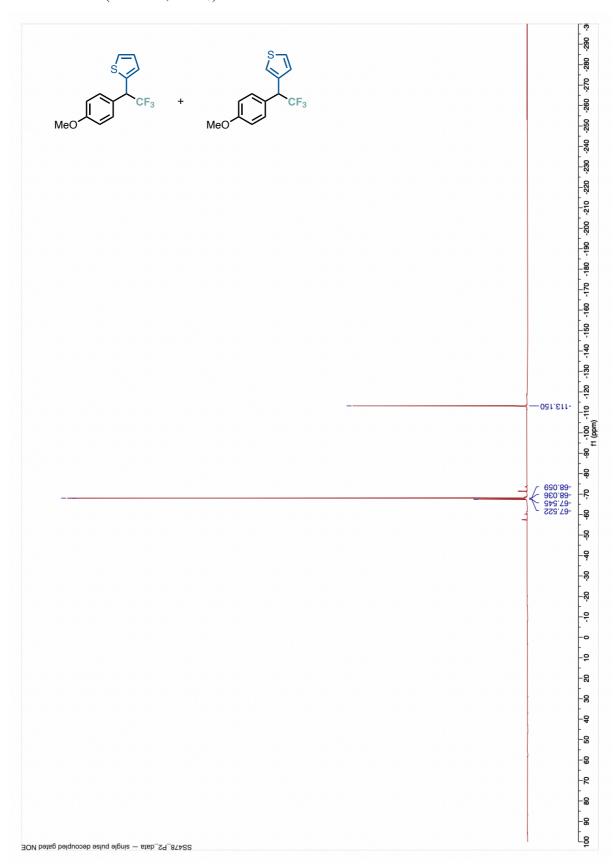


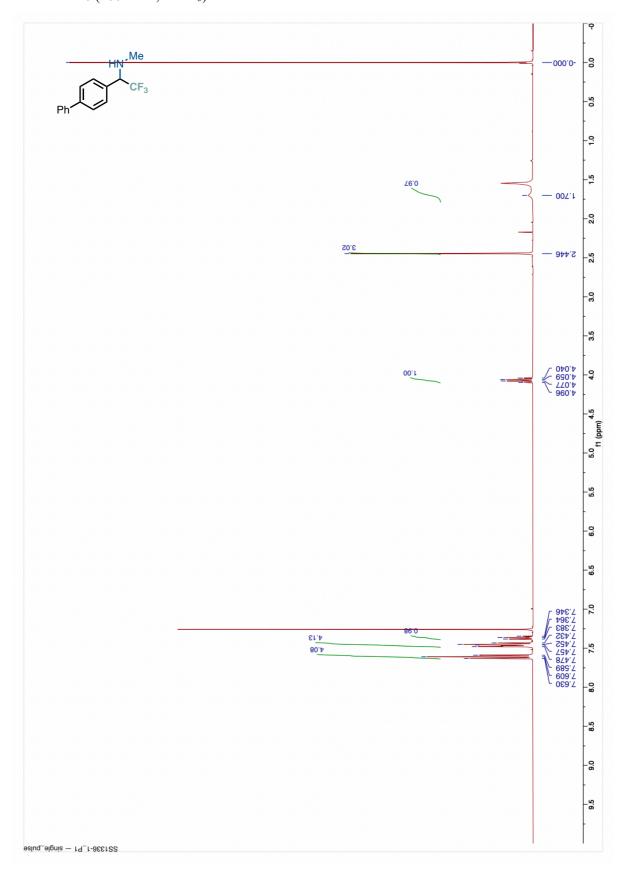
$^{1}\text{H NMR of }\textbf{5D} \ (400 \ \text{MHz}, \text{CDCl}_{3})$



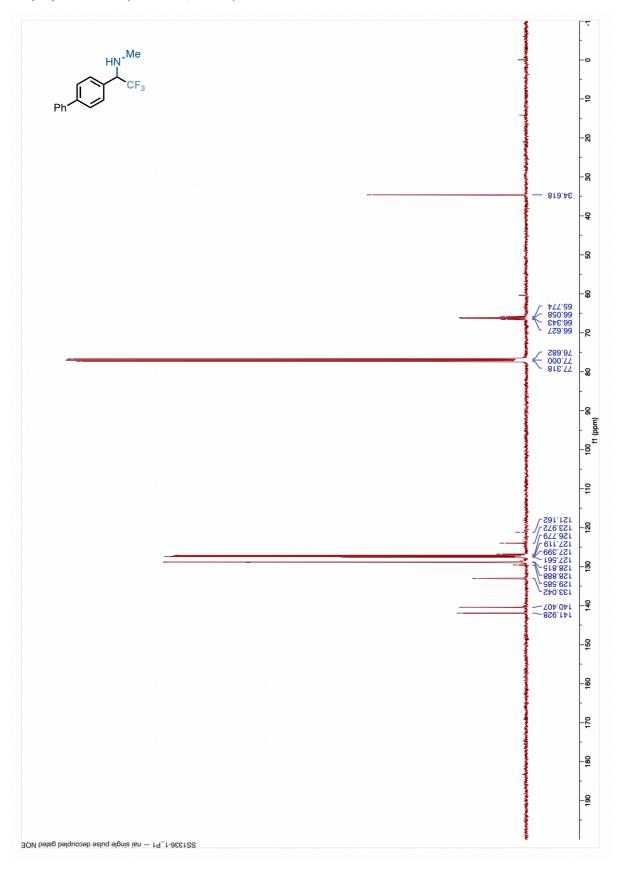
 $^{13}\text{C}\{^1\text{H}\}$ NMR of $\boldsymbol{5D}$ (101 MHz, CDCl₃)

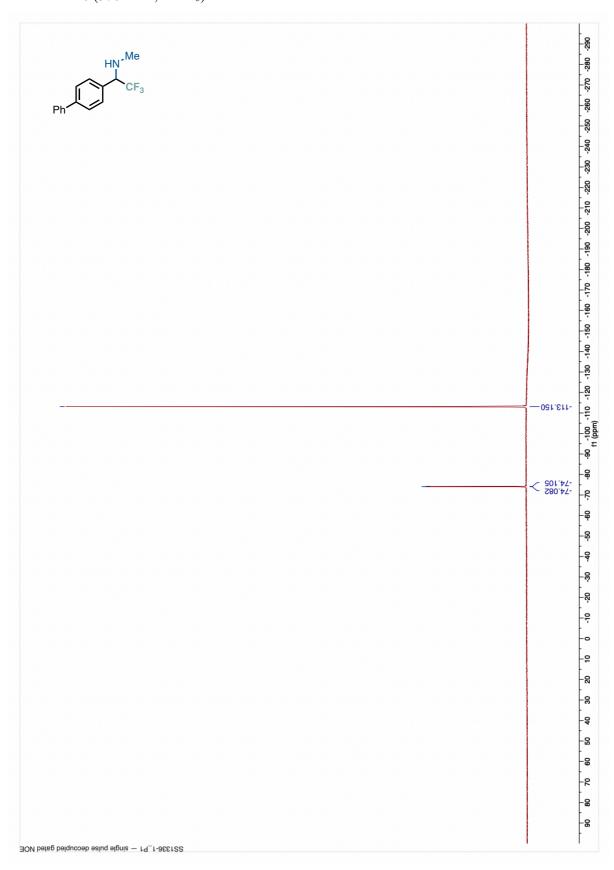


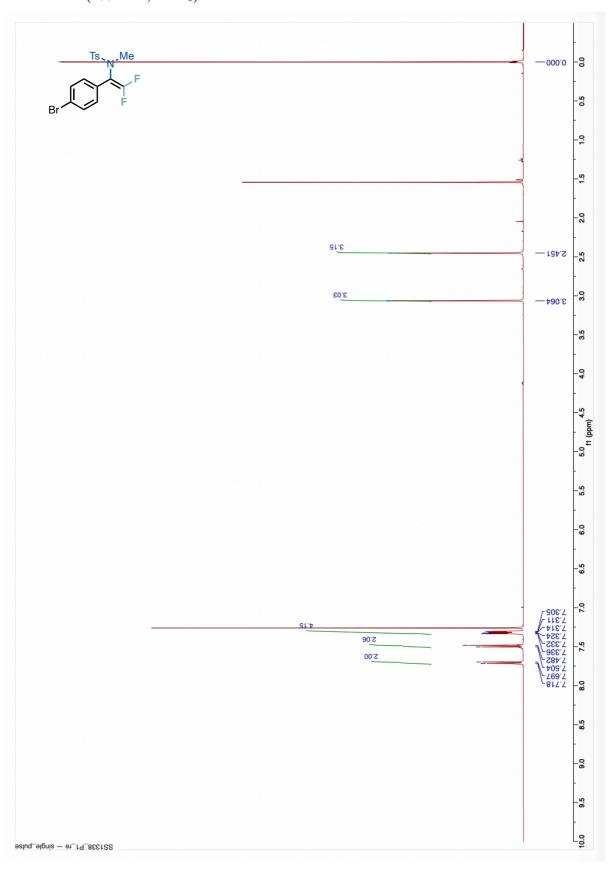




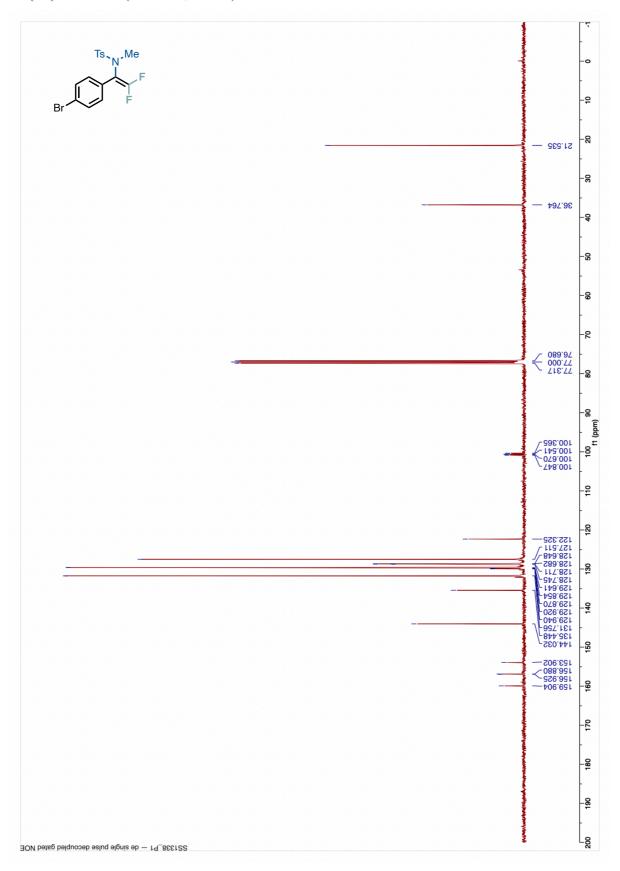
 $^{13}\text{C}\{^1\text{H}\}$ NMR of 6 (101 MHz, CDCl₃)

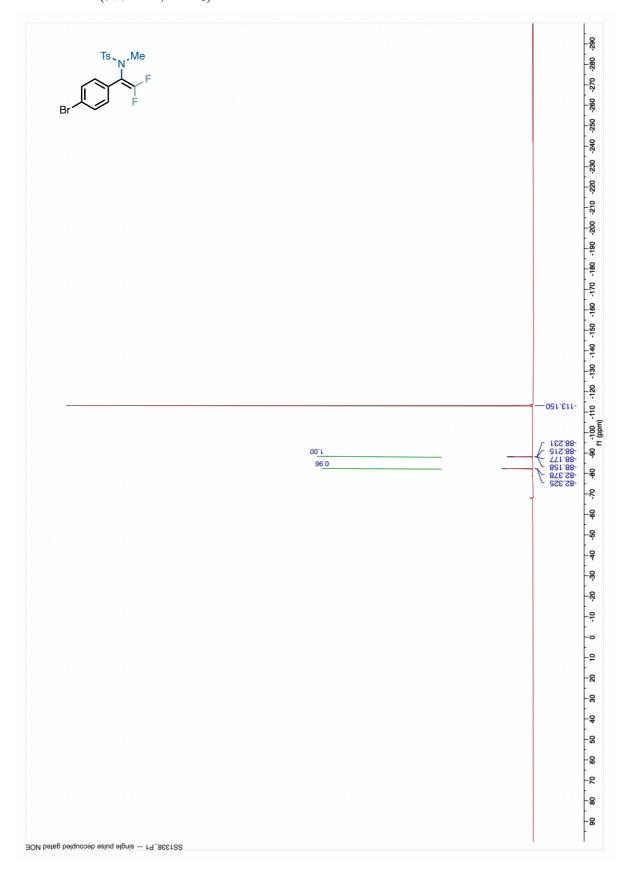


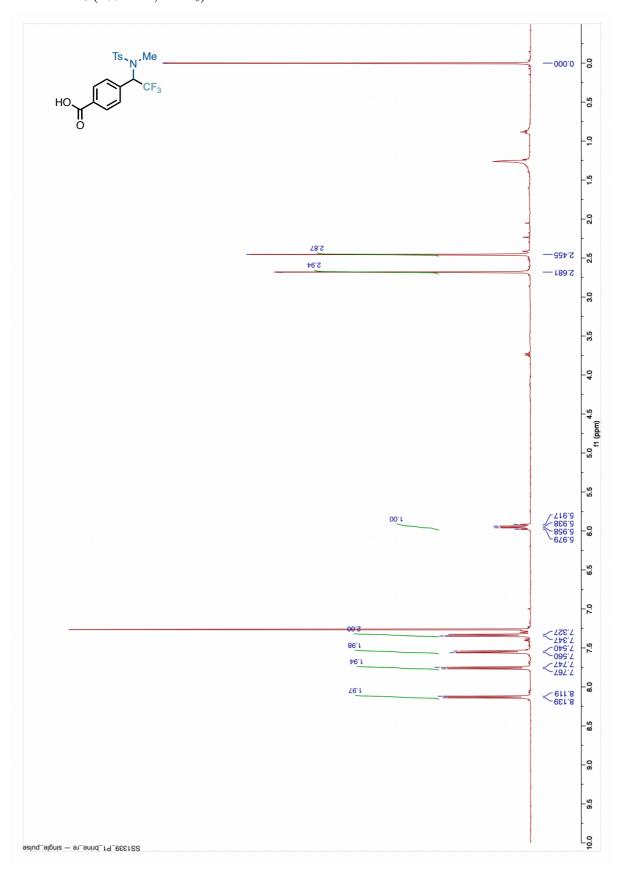




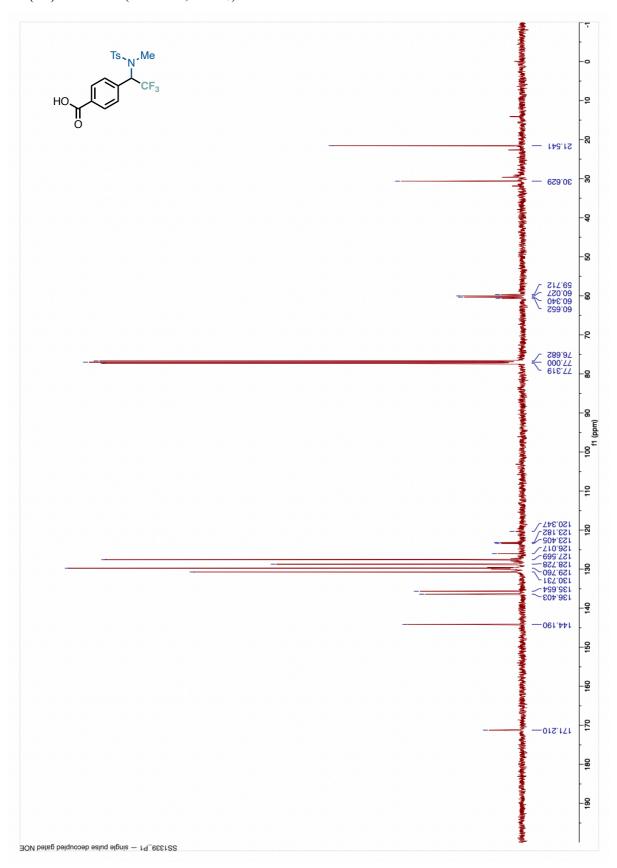
 $^{13}\text{C}\{^1\text{H}\}$ NMR of 7 (101 MHz, CDCl₃)

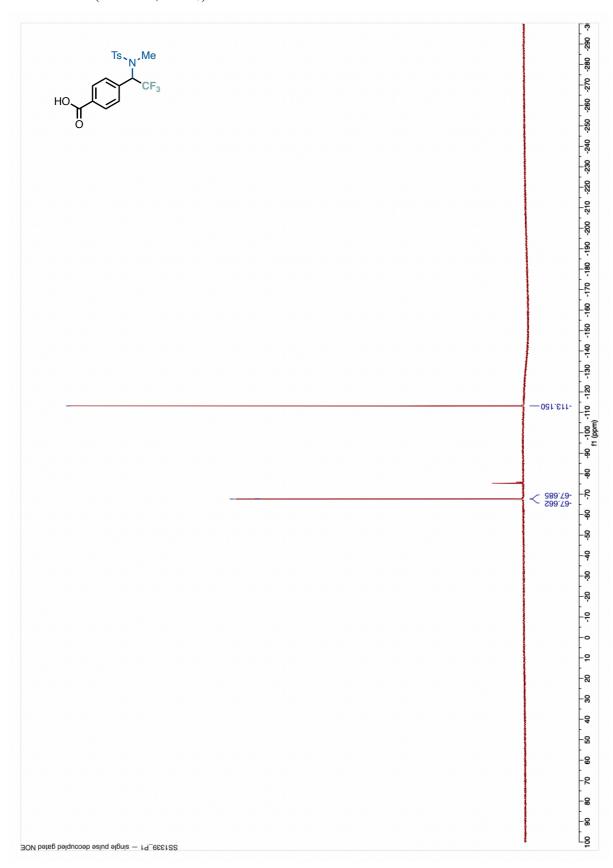


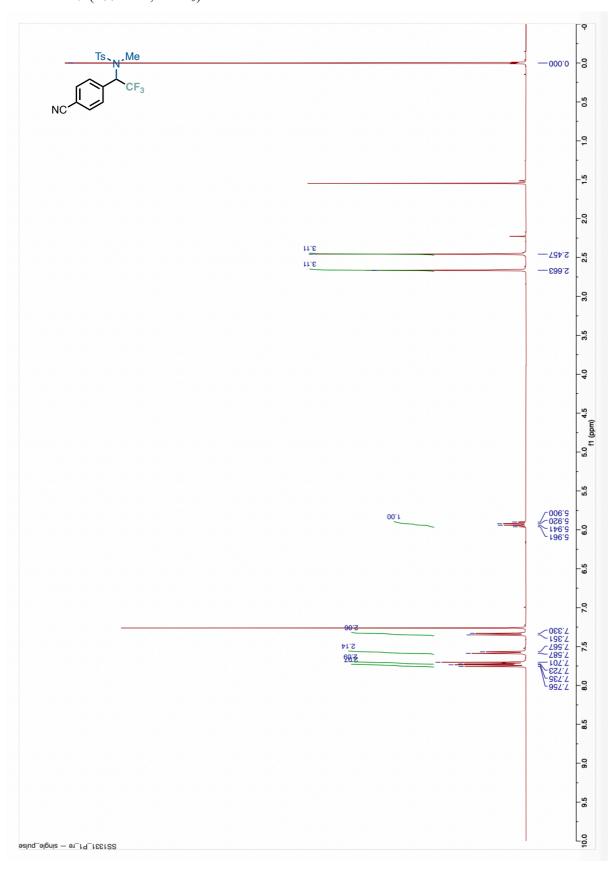




 $^{13}\text{C}\{^1\text{H}\}$ NMR of 8 (101 MHz, CDCl₃)







¹³C{¹H} NMR of **9** (101 MHz, CDCl₃)

