

Photoredox-catalyzed Giese reaction for the synthesis of γ -nitro alcohols

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Supporting Information

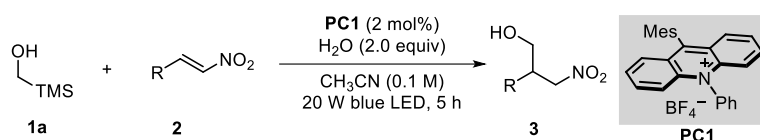
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1. General information

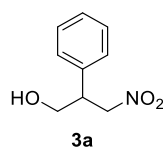
All commercial reagents were purchased and used as received unless mentioned otherwise. All visible-light-induced catalytic reactions were conducted in Schlenk seal tubes (purchased from Sythware Glass) at room temperature under an argon atmosphere and under irradiation with 20W blue LEDs lamp Kessil PR160L-456 nm. Inert atmosphere techniques were carried out through Schlenk system and Standard Glovebox. Reactions were monitored by thin-layer chromatography (TLC) (purchased from Yantai Xinnuo New Materials Technology Co., LTD). Column chromatography was performed on silica gel (300–400 mesh) using petroleum ether-EtOAc (PE/EA) (v/v). ^1H NMR (400 MHz) and ^{13}C NMR (101 MHz) spectra were recorded in DMSO- d_6 and CDCl_3 . ^1H NMR chemical shifts are reported in ppm relative to tetramethylsilane (TMS), with the solvent resonance employed as the internal standard (DMSO- d_6 at 2.50 ppm and CDCl_3 at 7.26 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, brs = broad singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz) and integration. ^{13}C NMR chemical shifts are reported in ppm from tetramethylsilane (TMS) with the solvent resonance as the internal standard (DMSO- d_6 at 39.52 ppm and CDCl_3 at 77.16 ppm). HRMS was recorded on the Agilent 6545 LC/Q-TOF mass spectrometer. Melting points were recorded on a OptiMelt MPA 1000. Cyclic voltammograms were collected with a CH Instruments Model 760E Series Electrochemical Analyzer/Workstation containing platinum wire working electrode, platinum wire counter electrode, SCE reference electrode (Sweep rate: 50 mV/s). Electrochemical potentials were obtained with a standard set of conditions to main internal consistency. Fluorescence studies were carried out using Techcomp Fluorescence Spectrophotometer (Model: FL970). The β -nitrostyrene^[1-2] were prepared following the procedures reported in literatures.

2. General experimental procedure for the synthesis of compounds 3



To a flame dried reaction tube equipped with a magnetic stir bar was charged with **2** (0.2 mmol, 1.0 equiv), **PC1** (0.004 mmol, 2 mol %), and degassed CH₃CN (2 mL, 0.1 M for **2**) under an argon atmosphere. To this mixture, **1a** (0.4 mmol, 2.0 equiv) and H₂O (0.4 mmol, 2.0 equiv) were added sequentially via syringe. The reaction mixture was then subjected to three cycles of degassing (vacuum/argon) and irradiated at room temperature for 5 hours using a 20 W blue Kessil LED (456 nm) positioned approximately 3 cm away. A high-speed fan was employed to maintain ambient temperature throughout the irradiation period. Upon completion, the solvent was removed under reduced pressure. The crude residue was then purified via flash column chromatography on silica gel to give the products **3**.

3-nitro-2-phenylpropan-1-ol (3a)



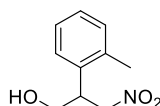
yellow oil; 25.7 mg, 71% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.29 (t, *J* = 7.2 Hz, 2H), 7.23 (t, *J* = 7.1 Hz, 1H), 7.17 (t, *J* = 5.9 Hz, 2H), 4.81 (dd, *J* = 12.8, 6.8 Hz, 1H), 4.62 (dd, *J* = 12.8, 8.0 Hz, 1H), 3.85 (dd, *J* = 11.0, 5.4 Hz, 1H), 3.76 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.70 – 3.59 (m, 1H), 1.65 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 137.1, 129.3, 128.2, 127.9, 77.3, 64.4, 46.4.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₉H₁₀NO₃ 180.0666; found: 180.0663.

3-nitro-2-(o-tolyl)propan-1-ol (3b)



3b

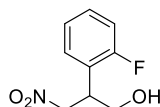
yellow oil; 19.9 mg, 51% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.13 (d, *J* = 4.2 Hz, 3H), 7.07 (d, *J* = 7.4 Hz, 1H), 4.81 (dd, *J* = 12.9, 7.0 Hz, 1H), 4.62 (dd, *J* = 12.9, 7.8 Hz, 1H), 4.05 – 3.92 (m, 1H), 3.80 (dd, *J* = 11.1, 5.3 Hz, 1H), 3.71 (dd, *J* = 11.1, 7.1 Hz, 1H), 2.34 (s, 3H), 1.65 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 136.9, 135.2, 131.3, 127.9, 126.7, 125.9, 77.1, 63.9, 41.4, 19.7.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₀H₁₃NNaO₃ 218.0788; found: 218.0791.

2-(2-fluorophenyl)-3-nitropropan-1-ol (3c)



3c

yellow oil; 20.1 mg, 51% yield; eluent: PE/EA = 5/1;

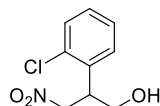
¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.15 (m, 2H), 7.11 – 6.97 (m, 2H), 4.85 (dd, *J* = 12.9, 6.4 Hz, 1H), 4.71 (dd, *J* = 13.0, 7.6 Hz, 1H), 3.99 – 3.78 (m, 3H), 1.67 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 161.0 (d, *J* = 247.4 Hz, 1C), 129.9 (d, *J* = 8.1 Hz, 1C), 129.6 (d, *J* = 4.0 Hz, 1C), 124.8 (d, *J* = 4.0 Hz, 1C), 124.0 (d, *J* = 14.1 Hz, 1C), 116.3 (d, *J* = 22.2 Hz, 1C), 76.0, 63.1, 41.0.

¹⁹F NMR (376 MHz, CDCl₃) δ -117.0.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₉H₉FNO₃ 198.0572; found: 198.0567.

2-(2-chlorophenyl)-3-nitropropan-1-ol (3d)



3d

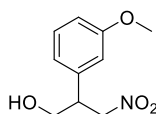
yellow oil; 25.4 mg, 59% yield; eluent: PE/EA = 5/1;

¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 6.3 Hz, 1H), 7.31 – 7.21 (m, 3H), 4.92 (dd, *J* = 13.0, 7.2 Hz, 1H), 4.79 (dd, *J* = 13.0, 7.2 Hz, 1H), 4.36 – 4.22 (m, 1H), 3.98 (dd, *J* = 11.1, 4.9 Hz, 1H), 3.89 (dd, *J* = 11.2, 6.6 Hz, 1H), 1.72 (br s, 1H).

¹³C NMR (400 MHz, CDCl₃) δ 134.6, 134.3, 130.5, 129.3, 128.3, 127.5, 75.8, 62.8, 42.4.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀ClNNaO₃ 238.0241; found: 238.0244.

2-(3-methoxyphenyl)-3-nitropropan-1-ol (3e)



3e

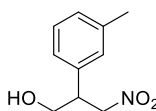
yellow oil; 24.9 mg, 59% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.15 (m, 1H), 6.83 – 6.66 (m, 3H), 4.78 (dd, *J* = 12.9, 6.9 Hz, 1H), 4.60 (dd, *J* = 12.8, 8.0 Hz, 1H), 3.83 (dd, *J* = 11.1, 5.5 Hz, 1H), 3.78 – 3.68 (m, 4H), 3.65–3.55 (m, 1H), 1.72 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 160.1, 138.7, 130.3, 119.9, 114.1, 113.2, 77.2, 64.3, 55.4, 46.4.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₀H₁₃NNaO₄ 234.0737; found: 234.0735.

3-nitro-2-(m-tolyl)propan-1-ol (3f)



3f

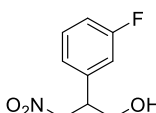
yellow oil; 22.6 mg, 58% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.13 (m, 1H), 7.04 (d, *J* = 7.6 Hz, 1H), 6.95 (d, *J* = 7.5 Hz, 2H), 4.78 (dd, *J* = 12.7, 6.9 Hz, 1H), 4.60 (dd, *J* = 12.8, 8.0 Hz, 1H), 3.82 (dd, *J* = 11.1, 5.6 Hz, 1H), 3.74 (dd, *J* = 11.0, 6.9 Hz, 1H), 3.67 – 3.53 (m, 1H), 2.27 (s, 3H), 1.70 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 139.0, 137.0, 129.1, 129.0, 128.6, 124.8, 77.3, 64.4, 46.4, 21.6.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₀H₁₃NNaO₃ 218.0788; found: 218.0790.

2-(3-fluorophenyl)-3-nitropropan-1-ol (3g)



3g

yellow oil; 22.2 mg, 56% yield; eluent: PE/EA = 4/1;

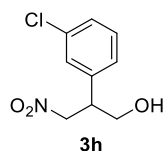
¹H NMR (400 MHz, CDCl₃) δ 7.33 (q, *J* = 7.3 Hz, 1H), 7.09 – 6.92 (m, 3H), 4.88 (dd, *J* = 13.0, 6.6 Hz, 1H), 4.68 (dd, *J* = 12.9, 8.2 Hz, 1H), 3.93 (dd, *J* = 11.0, 5.3 Hz, 1H), 3.82 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.77 – 3.66 (m, 1H), 1.81 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 163.1 (d, *J* = 248.5 Hz, 1C), 139.7 (d, *J* = 7.1 Hz, 1C), 130.8 (d, *J* = 8.1 Hz, 1C), 123.6 (d, *J* = 3.0 Hz, 1C), 115.2 (d, *J* = 21.2 Hz, 1C), 115.0 (d, *J* = 21.2 Hz, 1C), 77.0, 64.1, 46.0.

¹⁹F NMR (376 MHz, CDCl₃) δ -111.7.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₉H₉FNO₃ 198.0572; found: 198.0569.

2-(3-chlorophenyl)-3-nitropropan-1-ol (3h)



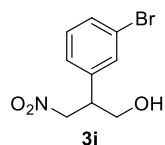
yellow oil; 25.8 mg, 60% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.21 (dd, *J* = 11.3, 6.4 Hz, 3H), 7.07 (t, *J* = 4.1 Hz, 1H), 4.81 (dd, *J* = 12.9, 6.5 Hz, 1H), 4.60 (dd, *J* = 12.9, 8.2 Hz, 1H), 3.85 (dd, *J* = 11.0, 5.2 Hz, 1H), 3.75 (dd, *J* = 11.1, 6.5 Hz, 1H), 3.61 (dt, *J* = 12.1, 6.6 Hz, 1H), 1.72 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 139.2, 135.0, 130.48, 128.5, 128.1, 126.1, 76.9, 64.0, 46.0.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀ClNNaO₃ 238.0241; found: 238.0249.

2-(3-bromophenyl)-3-nitropropan-1-ol (3i)



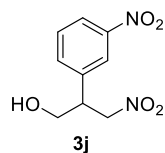
yellow oil; 26.2 mg, 50% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, *J* = 17.7 Hz, 2H), 7.23 – 7.07 (m, 2H), 4.80 (dd, *J* = 13.0, 6.7 Hz, 1H), 4.60 (dd, *J* = 13.0, 8.2 Hz, 1H), 3.85 (dd, *J* = 11.0, 5.3 Hz, 1H), 3.74 (dd, *J* = 11.0, 6.7 Hz, 1H), 3.60 (dt, *J* = 14.0, 6.7 Hz, 1H), 1.74 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 139.5, 131.4, 131.0, 130.8, 126.6, 123.2, 76.9, 64.0, 46.0.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀BrNNaO₃ 281.9736; found: 281.9743.

3-nitro-2-(3-nitrophenyl)propan-1-ol (3j)



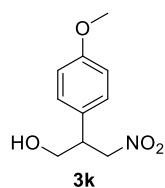
yellow solid; 26.1 mg, 58% yield; m.p. 85.5-85.9 °C; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 8.27 – 8.08 (m, 2H), 7.64 (d, *J* = 6.4 Hz, 1H), 7.56 (t, *J* = 8.2 Hz, 1H), 4.96 (dd, *J* = 13.1, 6.2 Hz, 1H), 4.76 (dd, *J* = 13.2, 8.4 Hz, 1H), 4.01 (dd, *J* = 10.7, 4.9 Hz, 1H), 3.95 – 3.80 (m, 2H), 1.81 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 148.7, 139.6, 134.4, 130.2, 123.3, 122.9, 76.7, 63.7, 45.8.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀N₂NaO₅ 249.0482; found: 249.0484.

2-(4-methoxyphenyl)-3-nitropropan-1-ol (3k)



yellow oil; 27.2 mg, 64% yield; eluent: PE/EA = 4/1;

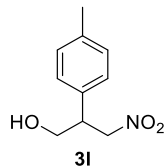
¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 8.6 Hz, 2H), 6.81 (d, *J* = 8.7 Hz, 2H), 4.77 (dd, *J* = 12.6, 6.8 Hz, 1H), 4.57 (dd, *J* = 12.6, 8.1 Hz, 1H), 3.81 (dd, *J* = 11.0, 5.6 Hz, 1H), 3.76 – 3.68 (m, 4H),

3.67 – 3.53 (m, 1H), 1.68 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 159.4, 128.9, 114.6, 77.6, 64.5, 55.4, 45.8.

HRMS (ESI) m/z: Calcd. for [M+K]⁺ C₁₀H₁₃KNO₄ 250.0476; found: 250.0487.

3-nitro-2-(p-tolyl)propan-1-ol (3l)



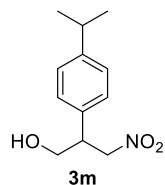
yellow oil; 23.8 mg, 61% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.10 (d, *J* = 8.0 Hz, 2H), 7.05 (d, *J* = 8.0 Hz, 2H), 4.79 (dd, *J* = 12.7, 6.8 Hz, 1H), 4.59 (dd, *J* = 12.7, 8.1 Hz, 1H), 3.83 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.74 (dd, *J* = 11.0, 6.9 Hz, 1H), 3.67 – 3.53 (m, 1H), 2.26 (s, 3H), 1.58 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 138.0, 133.9, 130.0, 127.7, 77.4, 64.5, 46.1, 21.2.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₀H₁₃NNaO₃ 218.0788; found: 218.0786.

2-(4-isopropylphenyl)-3-nitropropan-1-ol (3m)



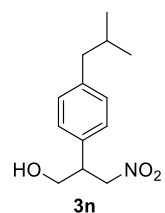
yellow solid; 26.6 mg, 60% yield; m.p. 60.0-60.5 °C; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.18 – 7.04 (m, 4H), 4.79 (dd, *J* = 12.7, 7.0 Hz, 1H), 4.61 (dd, *J* = 12.8, 7.9 Hz, 1H), 3.84 (dd, *J* = 11.1, 5.5 Hz, 1H), 3.75 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.68 – 3.57 (m, 1H), 2.82 (hept, *J* = 6.9 Hz, 1H), 1.58 (br s, 1H), 1.19 – 1.13 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 148.9, 134.2, 127.8, 127.3, 77.4, 64.5, 46.1, 33.9, 24.0.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₂H₁₇NNaO₃ 246.1101; found: 246.1098.

2-(4-isobutylphenyl)-3-nitropropan-1-ol (3n)



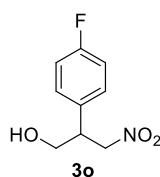
yellow oil; 23.1 mg, 49% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.06 (s, 4H), 4.79 (dd, *J* = 12.7, 6.9 Hz, 1H), 4.60 (dd, *J* = 12.7, 7.9 Hz, 1H), 3.83 (dd, *J* = 11.1, 5.5 Hz, 1H), 3.74 (dd, *J* = 11.1, 6.8 Hz, 1H), 3.66 – 3.56 (m, 1H), 2.37 (d, *J* = 7.2 Hz, 2H), 1.82 – 1.72 (m, 6.8 Hz, 1H), 1.65 (br s, 1H), 0.86 – 0.78 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 141.8, 134.2, 130.0, 127.5, 77.4, 64.5, 46.1, 45.1, 30.3, 22.5.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₃H₁₉NNaO₃ 260.1257; found: 260.1255.

2-(4-fluorophenyl)-3-nitropropan-1-ol (3o)



yellow oil; 25.4 mg, 64% yield; eluent: PE/EA = 4/1;

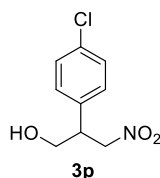
¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.11 (m, 2H), 6.97 (t, *J* = 8.6 Hz, 2H), 4.80 (dd, *J* = 12.8, 6.6 Hz, 1H), 4.58 (dd, *J* = 12.8, 8.3 Hz, 1H), 3.83 (dd, *J* = 11.0, 5.4 Hz, 1H), 3.74 (dd, *J* = 11.0, 6.9 Hz, 1H), 3.67 – 3.57 (m, 1H), 1.73 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 162.5 (d, *J* = 248.5 Hz, 1C), 132.9 (d, *J* = 4.0 Hz, 1C), 129.5 (d, *J* = 9.1 Hz, 1C), 116.2 (d, *J* = 22.2 Hz, 1C), 77.4, 64.3, 45.7.

¹⁹F NMR (376 MHz, CDCl₃) δ -113.9.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₉H₉FO₃ 198.0572; found: 198.0562.

2-(4-chlorophenyl)-3-nitropropan-1-ol (3p)



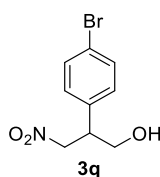
yellow oil; 25.8 mg, 60% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 8.5 Hz, 2H), 7.12 (d, *J* = 8.4 Hz, 2H), 4.80 (dd, *J* = 12.9, 6.5 Hz, 1H), 4.58 (dd, *J* = 12.9, 8.3 Hz, 1H), 3.83 (dd, *J* = 11.0, 5.3 Hz, 1H), 3.73 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.65 – 3.56 (m, 1H), 1.75 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 135.7, 134.1, 129.4, 129.3, 77.1, 64.1, 45.8.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀ClNNaO₃ 238.0241; found: 238.0247.

2-(4-bromophenyl)-3-nitropropan-1-ol (3q)



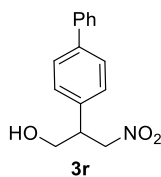
yellow oil; 29.5 mg, 57% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 8.3 Hz, 2H), 7.06 (d, *J* = 8.3 Hz, 2H), 4.80 (dd, *J* = 12.9, 6.6 Hz, 1H), 4.59 (dd, *J* = 12.9, 8.3 Hz, 1H), 3.84 (dd, *J* = 11.0, 5.3 Hz, 1H), 3.74 (dd, *J* = 10.9, 6.7 Hz, 1H), 3.66 – 3.53 (m, 1H), 1.66 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 136.2, 132.4, 129.6, 122.2, 77.0, 64.1, 45.9.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₀BrNNaO₃ 281.9736; found: 281.9746.

2-([1,1'-biphenyl]-4-yl)-3-nitropropan-1-ol (3r)



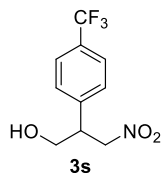
yellow solid; 32.3 mg, 63% yield; m.p. 131.1-131.6 °C; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.53 – 7.45 (m, 4H), 7.36 (t, *J* = 7.5 Hz, 2H), 7.28 (t, *J* = 7.3 Hz, 1H), 7.23 (d, *J* = 7.9 Hz, 2H), 4.83 (dd, *J* = 12.8, 6.7 Hz, 1H), 4.65 (dd, *J* = 12.8, 8.1 Hz, 1H), 3.87 (dd, *J* = 10.9, 5.4 Hz, 1H), 3.78 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.73 – 3.62 (m, 1H), 1.68 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 141.1, 140.4, 135.9, 128.9, 128.2, 127.8, 127.6, 127.1, 77.2, 64.3, 46.0.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₅H₁₅NNaO₃ 280.0944; found: 280.0946.

3-nitro-2-(4-(trifluoromethyl)phenyl)propan-1-ol (3s)



yellow oil; 34.2 mg, 69% yield; eluent: PE/EA = 4/1;

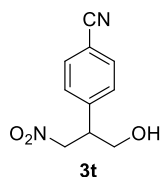
¹H NMR (400 MHz, CDCl₃) δ 7.62 (d, *J* = 7.9 Hz, 2H), 7.39 (d, *J* = 8.0 Hz, 2H), 4.92 (dd, *J* = 13.0, 6.4 Hz, 1H), 4.72 (dd, *J* = 12.9, 8.3 Hz, 1H), 3.95 (dd, *J* = 10.8, 5.1 Hz, 1H), 3.85 (dd, *J* = 11.0, 6.6 Hz, 1H), 3.82 – 3.74 (m, 1H), 1.84 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 141.3, 130.5 (q, *J* = 32.3 Hz, 1C), 128.4, 126.2 (q, *J* = 4.0 Hz, 1C), 124.0 (q, *J* = 273.7 Hz, 1C), 76.8, 64.0, 46.1.

¹⁹F NMR (376 MHz, CDCl₃) δ -62.7.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₁₀H₉F₃NO₃ 248.0540; found: 248.0538.

4-(1-hydroxy-3-nitropropan-2-yl)benzonitrile (3t)



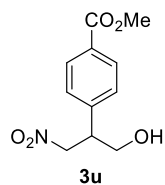
yellow oil; 20.5 mg, 50% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, *J* = 8.3 Hz, 2H), 7.40 (d, *J* = 8.4 Hz, 2H), 4.93 (dd, *J* = 13.1, 6.3 Hz, 1H), 4.72 (dd, *J* = 13.1, 8.5 Hz, 1H), 3.96 (dd, *J* = 10.8, 5.0 Hz, 1H), 3.85 (dd, *J* = 10.8, 6.4 Hz, 1H), 3.82 – 3.73 (m, 1H), 1.81 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 142.8, 132.9, 128.9, 118.5, 112.2, 76.6, 63.7, 46.2.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₁₀H₉N₂O₃ 205.0619; found: 205.0616.

methyl 4-(1-hydroxy-3-nitropropan-2-yl)benzoate (3u)



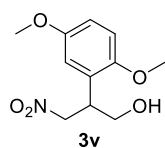
yellow oil; 19.3 mg, 40% yield; eluent: PE/EA = 2/1;

¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 8.4 Hz, 2H), 7.33 (d, *J* = 8.5 Hz, 2H), 4.92 (dd, *J* = 13.0, 6.4 Hz, 1H), 4.71 (dd, *J* = 13.0, 8.2 Hz, 1H), 3.94 (dd, *J* = 10.8, 5.0 Hz, 1H), 3.90 (s, 3H), 3.84 (dd, *J* = 10.7, 6.6 Hz, 1H), 3.81 – 3.73 (m, 1H), 1.91 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 166.8, 142.4, 130.4, 130.0, 128.0, 64.0, 52.4, 46.3.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₁H₁₃NNaO₅ 262.0686; found: 262.0691.

2-(2,5-dimethoxyphenyl)-3-nitropropan-1-ol (3v)



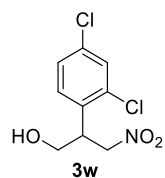
yellow solid; 26.4 mg, 55% yield; m.p. 69.0-69.7 °C; eluent: PE/EA = 3/1;

¹H NMR (400 MHz, CDCl₃) δ 6.80 – 6.63 (m, 3H), 4.79 (dd, *J* = 12.8, 6.8 Hz, 1H), 4.70 (dd, *J* = 12.8, 7.0 Hz, 1H), 3.90 (dt, *J* = 12.3, 6.6 Hz, 1H), 3.83 (d, *J* = 4.4 Hz, 2H), 3.73 (s, 3H), 3.68 (s, 3H), 1.82 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 153.8, 151.5, 126.2, 115.9, 112.9, 112.1, 76.2, 63.2, 56.1, 55.8, 42.1.

HRMS (ESI) m/z: Calcd. for [M+H]⁺ C₁₁H₁₆NO₅ 242.1023; found: 242.1026.

2-(2,4-dichlorophenyl)-3-nitropropan-1-ol (3w)



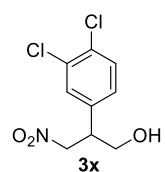
yellow oil; 27.8 mg, 56% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.38 (s, 1H), 7.18 (s, 2H), 4.83 (dd, *J* = 13.0, 7.0 Hz, 1H), 4.69 (dd, *J* = 13.0, 7.4 Hz, 1H), 4.22 – 4.10 (m, 1H), 3.89 (dd, *J* = 11.1, 4.7 Hz, 1H), 3.79 (dd, *J* = 11.1, 6.4 Hz, 1H), 1.73 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 135.0, 134.5, 133.3, 130.3, 129.2, 127.8, 75.6, 62.6, 41.9.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₉Cl₂NNaO₃ 271.9852; found: 271.9853.

2-(3,4-dichlorophenyl)-3-nitropropan-1-ol (3x)



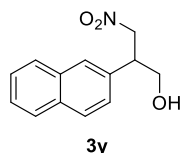
yellow solid; 23.2 mg, 48% yield; m.p. 66.5-67.0 °C; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 8.0 Hz, 1H), 7.16 (td, *J* = 8.2, 6.0 Hz, 1H), 7.05 (ddd, *J* = 11.1, 8.4, 1.3 Hz, 1H), 5.01 (ddd, *J* = 13.3, 6.5, 2.0 Hz, 1H), 4.87 (dd, *J* = 13.4, 7.7 Hz, 1H), 4.57 – 4.43 (m, 1H), 4.03 – 3.89 (m, 2H), 1.73 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 137.5, 133.3, 132.4, 131.1, 123.0, 127.3, 76.8, 63.8, 45.4.

HRMS (ESI) m/z: Calcd. for [M+K]⁺ C₉H₉Cl₂KNO₃ 287.9591; found: 287.9593.

2-(naphthalen-2-yl)-3-nitropropan-1-ol (3y)



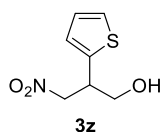
yellow solid; 27.0 mg, 58% yield; m.p. 96.0-96.3 °C; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.86 – 7.67 (m, 3H), 7.61 (s, 1H), 7.49 – 7.34 (m, 2H), 7.26 (d, *J* = 8.4 Hz, 1H), 4.87 (dd, *J* = 12.7, 6.4 Hz, 1H), 4.71 (dd, *J* = 12.7, 7.7 Hz, 1H), 3.90 (dd, *J* = 10.5, 5.0 Hz, 1H), 3.86 – 3.71 (m, 2H), 1.70 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 134.4, 133.5, 133.0, 129.1, 127.9, 127.8, 127.0, 126.7, 126.5, 125.5, 77.2, 64.3, 46.5.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₁₃H₁₃NNaO₃ 254.0788; found: 254.0787.

3-nitro-2-(thiophen-2-yl)propan-1-ol (3z)



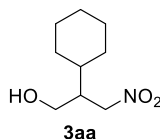
yellow oil; 21.6 mg, 58% yield; eluent: PE/EA = 3/1;

¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 5.2 Hz, 1H), 7.06 – 6.91 (m, 2H), 4.87 (dd, *J* = 12.8, 6.8 Hz, 1H), 4.67 (dd, *J* = 12.8, 7.7 Hz, 1H), 4.08 – 4.01 (m, 1H), 3.96 (dd, *J* = 11.1, 4.9 Hz, 1H), 3.87 (dd, *J* = 11.1, 6.3 Hz, 1H), 1.77 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 139.2, 127.4, 126.0, 125.3, 77.7, 64.4, 41.8.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₇H₈NO₃S 186.0230; found: 186.0227.

2-cyclohexyl-3-nitropropan-1-ol (3aa)



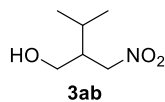
yellow oil; 21.7 mg, 58% yield; eluent: PE/EA = 5/1;

¹H NMR (400 MHz, CDCl₃) δ 4.56 (dd, *J* = 12.4, 7.5 Hz, 1H), 4.48 (dd, *J* = 12.3, 5.7 Hz, 1H), 3.78 (dd, *J* = 11.2, 4.5 Hz, 1H), 3.75 – 3.67 (m, 1H), 3.66 (dd, *J* = 11.1, 7.0 Hz, 1H), 2.33 – 2.18 (m, 1H), 1.75 (d, *J* = 11.3 Hz, 3H), 1.67 (d, *J* = 10.9 Hz, 3H), 1.51 – 1.41 (m, 1H), 1.22 – 1.18 (m, 1H), 1.18 – 1.10 (m, 1H), 1.09 – 0.95 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 75.7, 61.2, 58.6, 45.4, 37.3, 30.4, 30.1, 26.5, 26.4.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₉H₁₇NNaO₃ 210.1101; found: 210.1101.

3-methyl-2-(nitromethyl)butan-1-ol (3ab)



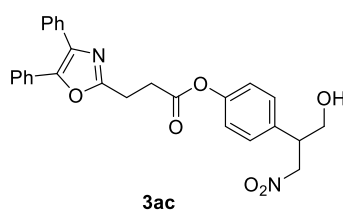
yellow oil; 17.4 mg, 59% yield; eluent: PE/EA = 5/1;

¹H NMR (400 MHz, CDCl₃) δ 4.56 (dd, *J* = 12.4, 7.4 Hz, 1H), 4.47 (dd, *J* = 12.4, 5.5 Hz, 1H), 3.79 (dd, *J* = 11.3, 4.5 Hz, 1H), 3.71 – 3.60 (m, 1H), 2.29 – 2.18 (m, 1H), 1.89 – 1.75 (m, 1H), 1.66 (br s, 1H), 1.04 – 0.91 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 75.7, 61.3, 46.0, 27.3, 19.9, 19.7.

HRMS (ESI) m/z: Calcd. for [M-H]⁻ C₆H₁₂NO₃ 146.0823; found: 146.0820.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl 3-(4,5-dimethyloxazol-2-yl)propanoate (3ac)



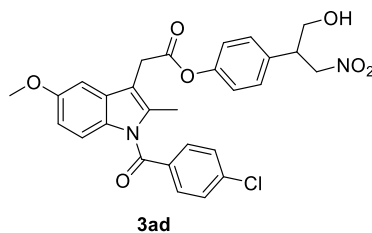
yellow oil; 35.8 mg, 38% yield; eluent: PE/EA = 2/1;

¹H NMR (400 MHz, CDCl₃) 7.64 (dd, *J* = 8.1, 1.7 Hz, 2H), 7.58 (dd, *J* = 7.8, 1.9 Hz, 2H), 7.41 – 7.37 (m, 2H), 7.37 – 7.33 (m, 4H), 7.22 (d, *J* = 8.6 Hz, 2H), 7.08 (d, *J* = 8.5 Hz, 2H), 4.85 (dd, *J* = 12.9, 6.7 Hz, 1H), 4.63 (dd, *J* = 12.9, 8.0 Hz, 1H), 3.86 (dd, *J* = 11.1, 5.3 Hz, 1H), 3.76 (dd, *J* = 11.0, 6.8 Hz, 1H), 3.68 (dt, *J* = 13.4, 6.7 Hz, 1H), 3.29 (t, *J* = 7.2 Hz, 2H), 3.16 (t, *J* = 7.2 Hz, 2H), 1.68 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 170.8, 161.5, 150.4, 145.8, 135.2, 134.9, 132.4, 129.0, 128.8, 128.73, 128.70, 128.3, 128.0, 126.6, 122.3, 64.2, 45.8, 31.3, 23.6.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₂₇H₂₄N₂NaO₆ 495.1527; found: 495.1530.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)acetate (3ad)



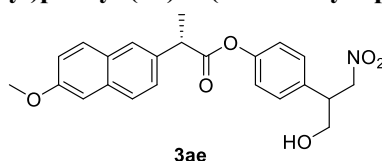
yellow solid; 55.1 mg, 51% yield; m.p. 121.3-121.9 °C; eluent: PE/EA = 2/1;

¹H NMR (400 MHz, CDCl₃) δ 7.67 (d, *J* = 8.2 Hz, 2H), 7.47 (d, *J* = 8.2 Hz, 2H), 7.21 (d, *J* = 8.3 Hz, 2H), 7.08 – 7.00 (m, 3H), 6.89 (d, *J* = 9.0 Hz, 1H), 6.69 (dd, *J* = 9.0, 2.5 Hz, 1H), 4.83 (dd, *J* = 12.9, 6.6 Hz, 1H), 4.63 (dd, *J* = 12.9, 8.2 Hz, 1H), 3.90 (s, 2H), 3.87 – 3.79 (m, 4H), 3.75 (dd, *J* = 10.9, 6.8 Hz, 1H), 3.71 – 3.62 (m, 1H), 2.44 (s, 3H), 1.83 (br s, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 169.5, 168.5, 156.2, 150.4, 139.5, 136.4, 134.9, 133.9, 131.3, 131.0, 130.6, 129.3, 129.0, 122.2, 115.2, 111.93, 111.88, 101.3, 77.1, 64.2, 55.9, 45.8, 30.6, 13.6.6

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₂₈H₂₅ClN₂NaO₇ 559.1242; found: 559.1250.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl (2S)-2-(6-methoxynaphthalen-2-yl)propanoate (3ae)



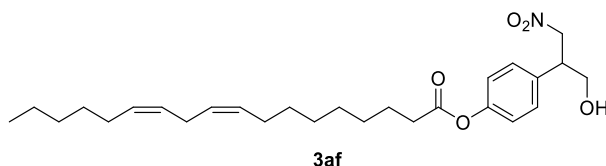
yellow solid; 31.7 mg, 39% yield, >20:1 dr; m.p. 105.2-105.8 °C; eluent: PE/EA = 2/1;

¹H NMR (400 MHz, CDCl₃) δ 7.74 (t, *J* = 8.0 Hz, 3H), 7.49 (d, *J* = 8.6 Hz, 1H), 7.22 – 7.11 (m, 4H), 6.96 (d, *J* = 8.7 Hz, 2H), 4.81 (dd, *J* = 12.9, 6.6 Hz, 1H), 4.60 (ddd, *J* = 12.8, 8.0, 2.4 Hz, 1H), 4.14 – 4.04 (m, 1H), 3.92 (s, 3H), 3.81 (dd, *J* = 11.0, 5.2 Hz, 1H), 3.76 – 3.68 (m, 1H), 3.68 – 3.57 (m, 1H), 1.86 (br s, 1H), 1.69 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.4, 157.9, 150.6, 135.0, 134.7, 134.0, 129.4, 129.1, 128.9, 127.6, 126.3, 126.2, 122.2, 119.3, 105.7, 77.1, 64.2, 55.5, 45.8, 45.6, 18.5.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₂₃H₂₃NNaO₆ 432.1418; found: 432.1421.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl (9Z,12Z)-octadeca-9,12-dienoate (3af)



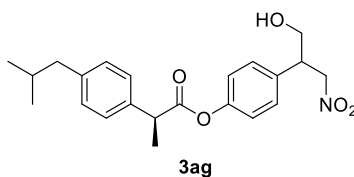
yellow oil; 44.5 mg, 48% yield; eluent: PE/EA = 4/1;

¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, *J* = 8.6 Hz, 2H), 7.06 (d, *J* = 8.5 Hz, 2H), 5.36 (tt, *J* = 10.9, 5.6 Hz, 4H), 4.85 (dd, *J* = 12.9, 6.5 Hz, 1H), 4.64 (dd, *J* = 12.9, 8.0 Hz, 1H), 3.83 (dd, *J* = 10.9, 5.2 Hz, 1H), 3.72 (ddd, *J* = 22.9, 12.0, 7.2 Hz, 2H), 2.78 (t, *J* = 6.4 Hz, 2H), 2.55 (t, *J* = 7.5 Hz, 2H), 2.09 – 2.01 (m, 4H), 1.78 – 1.69 (m, 2H), 1.45 – 1.28 (m, 15H), 0.89 (t, *J* = 6.7 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 172.6, 134.7, 130.3, 130.1, 128.9, 128.2, 128.0, 122.3, 64.2, 45.8, 34.5, 31.6, 29.7, 29.5, 29.3, 29.21, 29.17, 27.31, 27.29, 25.7, 25.0, 22.7, 14.2.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₂₇H₄₁NNaO₅ 482.2877; found: 482.2882.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl (2S)-2-(4-isobutylphenyl)propanoate (3ag)



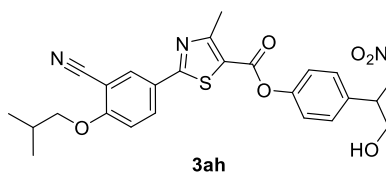
yellow oil; 39.4 mg, 51% yield, >20:1 dr; eluent: PE/EA = 2/1;

¹H NMR (400 MHz, CDCl₃) δ 7.29 (d, *J* = 8.0 Hz, 2H), 7.17 (dd, *J* = 15.8, 8.0 Hz, 4H), 6.98 (d, *J* = 8.4 Hz, 2H), 4.82 (dd, *J* = 12.9, 6.4 Hz, 1H), 4.61 (dd, *J* = 12.7, 8.2 Hz, 1H), 3.94 (q, *J* = 7.1 Hz, 1H), 3.85 – 3.75 (br s, 1H), 3.66 (td, *J* = 15.5, 13.2, 8.9 Hz, 2H), 2.48 (d, *J* = 7.2 Hz, 2H), 2.08 – 1.94 (m, 1H), 1.87 (dp, *J* = 13.6, 6.8 Hz, 1H), 1.60 (d, *J* = 7.1 Hz, 3H), 0.94 – 0.89 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 173.6, 150.5, 141.1, 137.1, 134.7, 129.7, 128.9, 127.3, 122.1, 77.1, 64.2, 45.8, 45.3, 45.1, 30.3, 22.5, 18.5.

HRMS (ESI) m/z: Calcd. for [M+Na]⁺ C₂₂H₂₇NNaO₅ 408.1781; found: 408.1786.

4-(1-hydroxy-3-nitropropan-2-yl)phenyl 2-(3-cyano-4-isobutoxyphenyl)-4-methylthiazole-5-carboxylate (3ah)



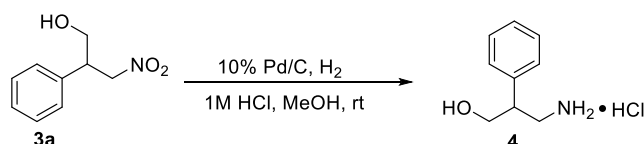
yellow solid; 41.6 mg, 42% yield; m.p. 132.5-133.2 °C; eluent: PE/EA = 1/1;

¹H NMR (400 MHz, CDCl₃) δ 8.21 (d, *J* = 2.3 Hz, 1H), 8.11 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.32 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.6 Hz, 2H), 7.03 (d, *J* = 9.0 Hz, 1H), 4.90 (dd, *J* = 12.9, 6.5 Hz, 1H), 4.69 (dd, *J* = 12.9, 8.2 Hz, 1H), 3.94 – 3.88 (m, 3H), 3.81 (dd, *J* = 10.9, 6.8 Hz, 1H), 3.78 – 3.69 (m, 1H), 2.92 – 2.70 (m, 4H), 2.27 – 2.14 (m, 1H), 1.10 – 1.07 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 168.5, 163.4, 162.8, 160.5, 150.0, 135.3, 132.8, 132.3, 129.1, 125.8, 122.4, 120.4, 115.5, 112.8, 103.1, 75.9, 64.2, 45.9, 28.3, 19.2, 17.8.

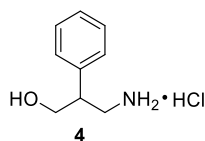
HRMS (ESI) m/z: Calcd. for [M+H]⁺ C₂₅H₂₆N₃O₆S 496.1537; found: 496.1538.

3. General experimental procedure for the synthesis of compound 4



A mixture of compound **3a** (0.2 mmol, 1.0 equiv), 10 wt% Pd/C (10 mg), and 1 M HCl in methanol (2 mL, 0.1 M) was stirred under a hydrogen atmosphere for 12 hours. The reaction mixture was then filtered through a pad of Celite, and the filtrate was concentrated under reduced pressure to afford compound **4**.

3-amino-2-phenylpropan-1-ol hydrochloride (4)



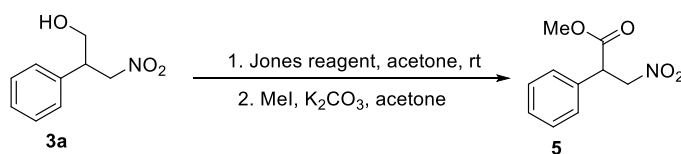
yellow solid; 36.4 mg, 97% yield; m.p. 274.4-275.2 °C;

¹H NMR (400 MHz, DMSO-*d*₆) δ 8.13 (s, 3H), 7.37 – 7.22 (m, 5H), 5.25 – 5.12 (m, 1H), 3.66 – 3.53 (m, 2H), 3.31 – 3.20 (m, 1H), 3.14 – 3.05 (m, 1H), 3.03 – 3.91 (m, 1H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 139.7, 128.5, 128.1, 127.0, 63.5, 45.5, 41.1.

HRMS (ESI) m/z: Calcd. for [M+H]⁺ C₉H₁₄NO 152.1070; found: 152.1069.

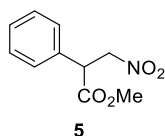
4. General experimental procedure for the synthesis of compound 5



A solution of compound **3a** (0.2 mmol, 1.0 equiv) in acetone (2 mL, 0.1 M) was cooled to 0 °C.

Jones reagent (0.4 mmol, 2.0 equiv) was added dropwise, and the mixture was stirred while warming to room temperature. Reaction progress was monitored by TLC. Upon completion, the reaction was quenched with isopropanol (added dropwise), and the mixture was filtered through a Celite pad to remove chromium salts. The filtrate was concentrated under reduced pressure, and the residue was dissolved in DMF (1 mL). Potassium carbonate (0.2 mmol, 1.0 equiv) and iodomethane (1 mmol, 5.0 equiv) were added sequentially, and the reaction was again monitored by TLC. After completion, the mixture was diluted with ethyl acetate, washed with brine, and the organic phase was concentrated under reduced pressure. The crude product was purified by flash column chromatography (silica gel, eluent: PE:EA = 15:1) to yield compound **5**.

methyl 3-nitro-2-phenylpropanoate (**5**)



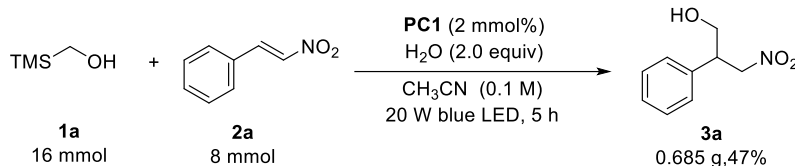
yellow oil; 17.3 mg, 41% yield; eluent: PE/EA = 15/1;

¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 13.1 Hz, 3H), 7.19 – 7.14 (m, 2H), 5.01 (dd, *J* = 14.6, 10.0 Hz, 1H), 4.45 (dd, *J* = 14.7, 5.1 Hz, 1H), 4.35 (dd, *J* = 9.9, 5.1 Hz, 1H), 3.63 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.2, 133.3, 129.5, 128.9, 128.0, 75.9, 53.0, 48.8.

HRMS (ESI) *m/z*: Calcd. for [M+Na]⁺ C₁₀H₁₁NNaO₄ 232.0580; found: 232.0585.

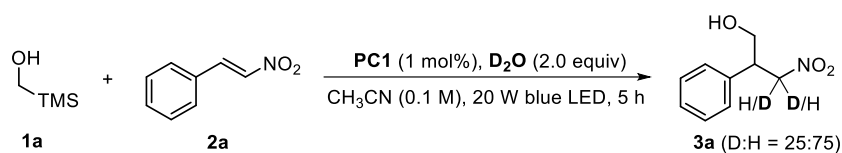
5. Scale-up reaction



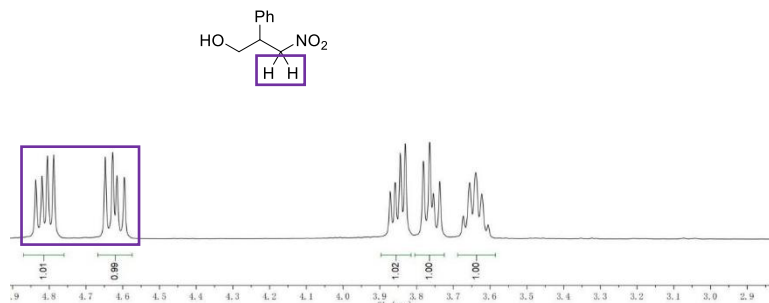
To a flame dried reaction tube equipped with a magnetic stir bar was charged with **2a** (8.0 mmol, 1.0 equiv), **PC1** (0.16 mmol, 2 mol %), and degassed CH₃CN (80 mL, 0.1 M for **2a**) under an argon atmosphere. To this mixture, **1a** (16.0 mmol, 2.0 equiv) and H₂O (16.0 mmol, 2.0 equiv) were added sequentially via syringe. The reaction mixture was then subjected to three cycles of degassing (vacuum/argon) and irradiated at room temperature for 5 hours using a 20 W blue Kessil LED (456 nm) positioned approximately 3 cm away. A high-speed fan was employed to maintain ambient temperature throughout the irradiation period. Upon completion, the solvent was removed under reduced pressure. The crude residue was then purified via flash column chromatography on silica gel to give the products **3**, 0.685 g, 47% yield.

6. Deuterium labelling experiment

To identify the proton source in the proposed mechanism, control experiments were conducted using D₂O in place of H₂O, while keeping all other parameters constant. ¹H NMR analysis of the crude product confirmed the incorporation of deuterium, as shown in the spectrum below.



^1H NMR of **3a** (H)



^1H NMR of Crude **3a** (D)

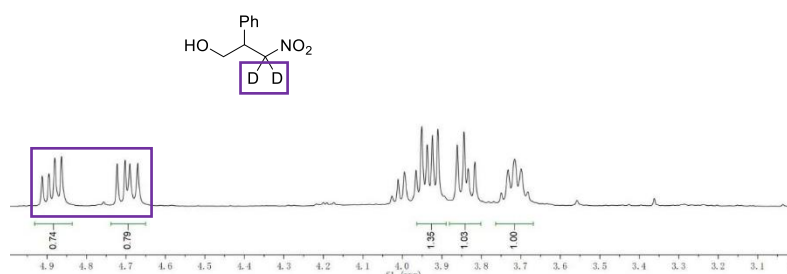


Figure S1. ^1H NMR analysis for deuterium labelling experiment

7. Radical trapping experiments

To a flame dried reaction tube equipped with a magnetic stir bar was charged with **2a** (0.2 mmol, 1.0 equiv), **PC1** (0.004 mmol, 2 mol %), a radical quencher (0.6 mmol, 3.0 equiv), and degassed CH_3CN (2 mL, 0.1 M for **2a**) under an argon atmosphere. To this mixture, **1a** (0.4 mmol, 2.0 equiv) and H_2O (0.4 mmol, 2.0 equiv) were added sequentially via syringe. The reaction mixture was then subjected to three cycles of degassing (vacuum/argon) and irradiated at room temperature using a 20 W blue Kessil LED (456 nm) positioned approximately 3 cm away. A high-speed fan was employed to maintain ambient temperature throughout the 5 hours irradiation period. Upon completion, the crude reaction mixture was analyzed by ^1H NMR and HRMS. The results (Table S1) indicated significant suppression of **3a** formation. Notably, trace amounts of the 1,1-diphenylethylene adduct **6**, TEMPO adduct **7**, and BHT adduct **8** were detected in the HRMS analysis of the crude mixture (Figure S2). These observations support the generation of a (trimethylsilyl)oxy methyl radical and are consistent with a radical addition pathway.

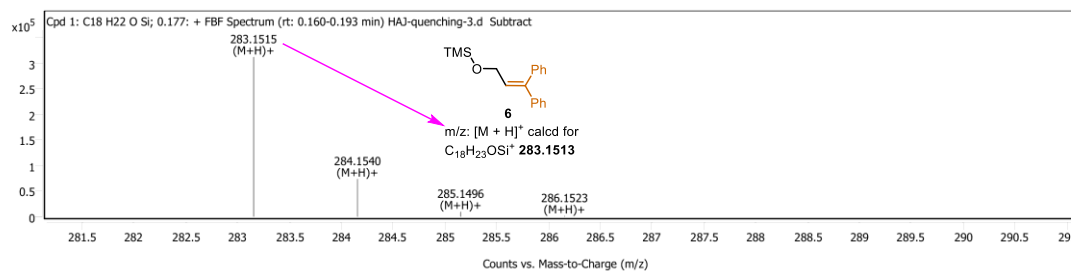
Table S1 Radical trapping experiments

<chem>CC(C)(C)O</chem> (1a) + <chem>O=[N]C=Cc1ccccc1</chem> (2a) $\xrightarrow[\text{CH}_3\text{CN (0.1 M), 20 W blue LED, 5 h}]{\text{PC1 (1 mol\%), H}_2\text{O (2.0 equiv) radical quencher (3.0 equiv)}}$ <chem>OCC(C)(O)[N+](=O)[O-]</chem> (3a)			
radical quencher	TEMPO	BHT	1,1-Diphenylethylene
Yield	0%	trace	25%

Compound Details

Cpd. 1: C₁₈H₂₂O Si

Compound Spectra



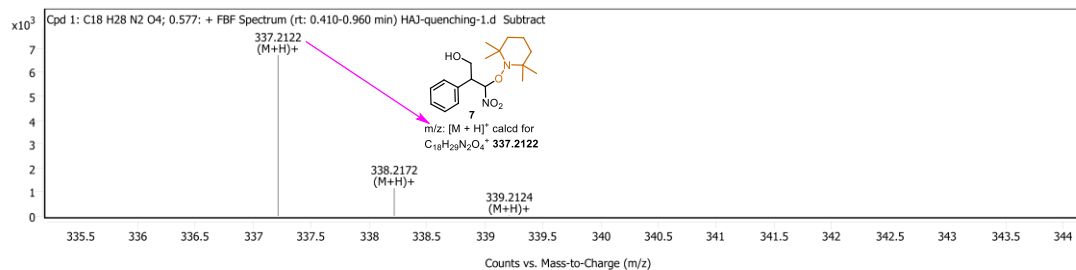
Spectrum Peaks

m/z	m/z (Calc)	Diff (ppm)	Abund	Height %	Height % (Calc)	Ion Species	Z
283.1515	283.1513	0.79	312709	100.00	100.00	(M+H)+	1
284.1540	284.1539	0.51	73929	23.64	24.85	(M+H)+	1
285.1496	285.1522	-8.96	10002	3.20	6.41	(M+H)+	1
286.1523	286.1537	-4.76	1821	0.58	0.92	(M+H)+	1

Compound Details

Cpd. 1: C₁₈H₂₈N₂O₄

Compound Spectra



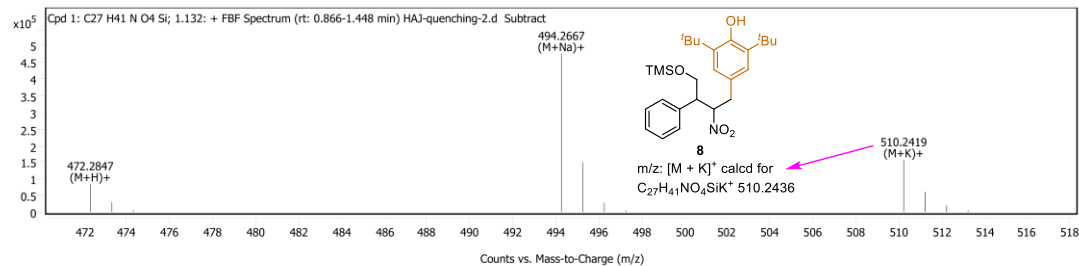
Spectrum Peaks

m/z	m/z (Calc)	Diff (ppm)	Abund	Height %	Height % (Calc)	Ion Species	Z
337.2122	337.2122	0.06	6697	100.00	100.00	(M+H)+	1
338.2172	338.2154	5.54	1164	17.37	20.68	(M+H)+	1
339.2124	339.2179	-16.29	32	0.47	2.85	(M+H)+	1

Compound Details

Cpd. 1: C₂₇H₄₁N O₄ Si

Compound Spectra



Spectrum Peaks

m/z	m/z (Calc)	Diff (ppm)	Abund	Height %	Height % (Calc)	Ion Species	Z
472.2847	472.2878	-6.39	84990	100.00	100.00	(M+H)+	1
473.2880	473.2905	-5.46	30155	35.48	35.28	(M+H)+	1
474.2910	474.2904	1.36	5788	6.81	10.11	(M+H)+	1
494.2667	494.2697	-6.13	476978	100.00	100.00	(M+Na)+	1
495.2694	495.2725	-6.23	149946	31.44	35.27	(M+Na)+	1
496.2709	496.2723	-2.88	28288	5.93	10.11	(M+Na)+	1
497.2723	497.2737	-2.87	5479	1.15	1.94	(M+Na)+	1
510.2419	510.2436	-3.44	157416	100.00	100.00	(M+K)+	1
511.2445	511.2464	-3.85	60616	38.51	35.28	(M+K)+	1
512.2469	512.2444	4.92	20172	12.81	17.33	(M+K)+	1
513.2444	513.2459	-2.95	5398	3.43	4.48	(M+K)+	1

Figure S2. HRMS analysis for radical trapping experiments

8. Stern-Volmer Fluorescence Quenching Experiments

Fluorescence quenching studies were carried out using Techcomp FL970 fluorescence spectrophotometer. Photocatalyst PC1 (9-Mesityl-10-phenylacridin-10-ium tetrafluoroborate) and different concentrations of added quenchers were prepared in dry CH₃CN in quartz cuvettes. CH₃CN was degassed with a stream of argon gas for 30 min. For the quenching experiments, the

concentration of **PC1** was 5.0×10^{-5} M. The solutions were excited at 440 nm in CH₃CN solution and the emission intensity was observed at ~540 nm. Plots were derived according to the Stern–Volmer equation and K_{sv} was calculated. Stern-Volmer equation: $I_0/I = 1 + K_{sv}[S]$. Where I_0 is the luminescence intensity of the photocatalyst in the absence of a quencher, I is the intensity of the photocatalyst in the presence of quenchers, $[S]$ is the concentration of added quencher, and K_{sv} is the Stern–Volmer quenching constant. All emission spectra were recorded after each addition of the quencher. The steady decrease of the emission intensity of the catalyst solution with the gradual increase of the amount of **1a** or **2a** as presented in Figure S1 and Figure S2 supports that reaction mechanism occurring through reductive quenching cycle.

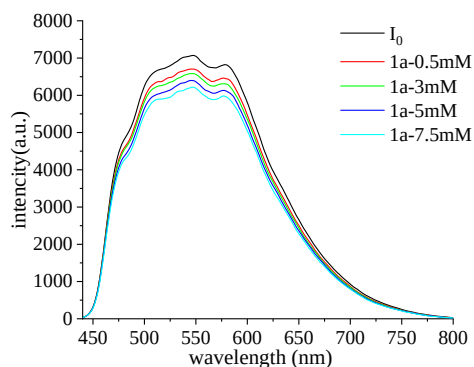


Figure S3. Fluorescence quenching by **1a**

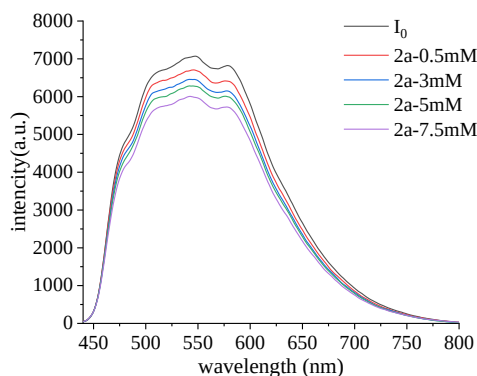


Figure S4. Fluorescence quenching by **2a**

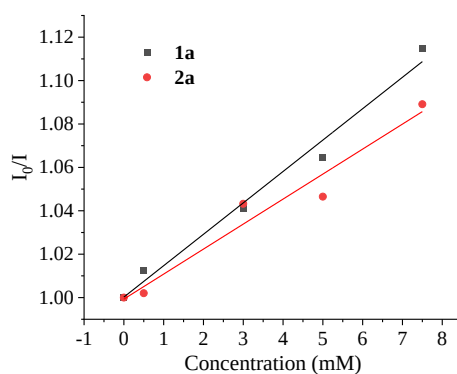


Figure S5. Stern-Volmer plot

9. Electrochemical measurements

Cyclic Voltammetry was performed using a CH Instruments 760E. The redox potentials (vs SCE) were determined using a 5.0 mM solution of the compounds in 0.1 M solution of $n\text{Bu}_4\text{NPF}_6$ (purged CH_3CN with Ar). Measurements employed a glassy carbon working electrode, saturated calomel reference electrode, and a platinum wire counter electrode. The potential was scanned at a scan rate 50 mV/s. A background of the electrolyte solution was subtracted from the voltammogram. Reduction was measured by scanning potential in the positive direction. The glassy carbon electrode was polished between each scan.

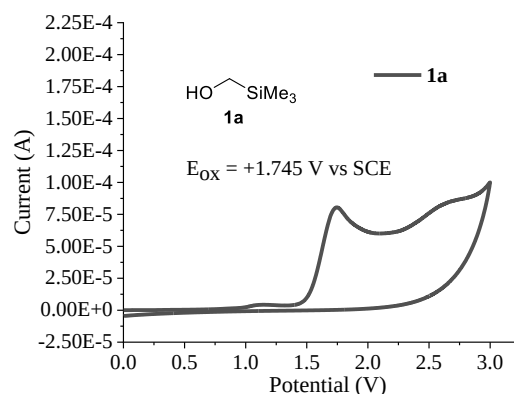


Figure S6. Cyclic voltammogram study for **1a**

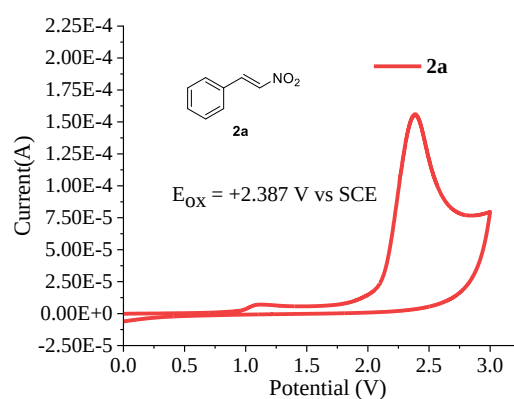


Figure S6. Cyclic voltammogram study for **2a**

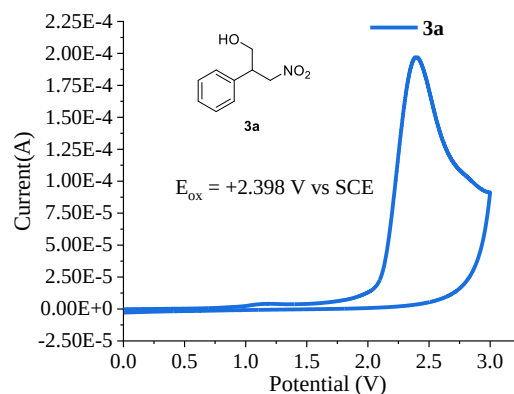
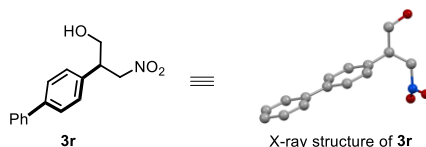


Figure S8. Cyclic voltammogram study for **3a**

10. X-ray crystal structure of **3r**

Single crystals of compound **3r** were prepared through dissolving the sample in *n*-pentane/EtOAc at room temperature and crystallizing by slow evaporation of solvent. A suitable crystal was selected for structure determination on an Xcalibur, Eos, Gemini diffractometer. The crystal was kept at 293(2) K during data collection. Using Olex2^[3], the structure was solved with the ShelXS^[4] structure solution program using Direct Methods and refined with the ShelXL^[5] refinement package using Least Squares minimisation.



ORTEP of **3r** (at 50% level)

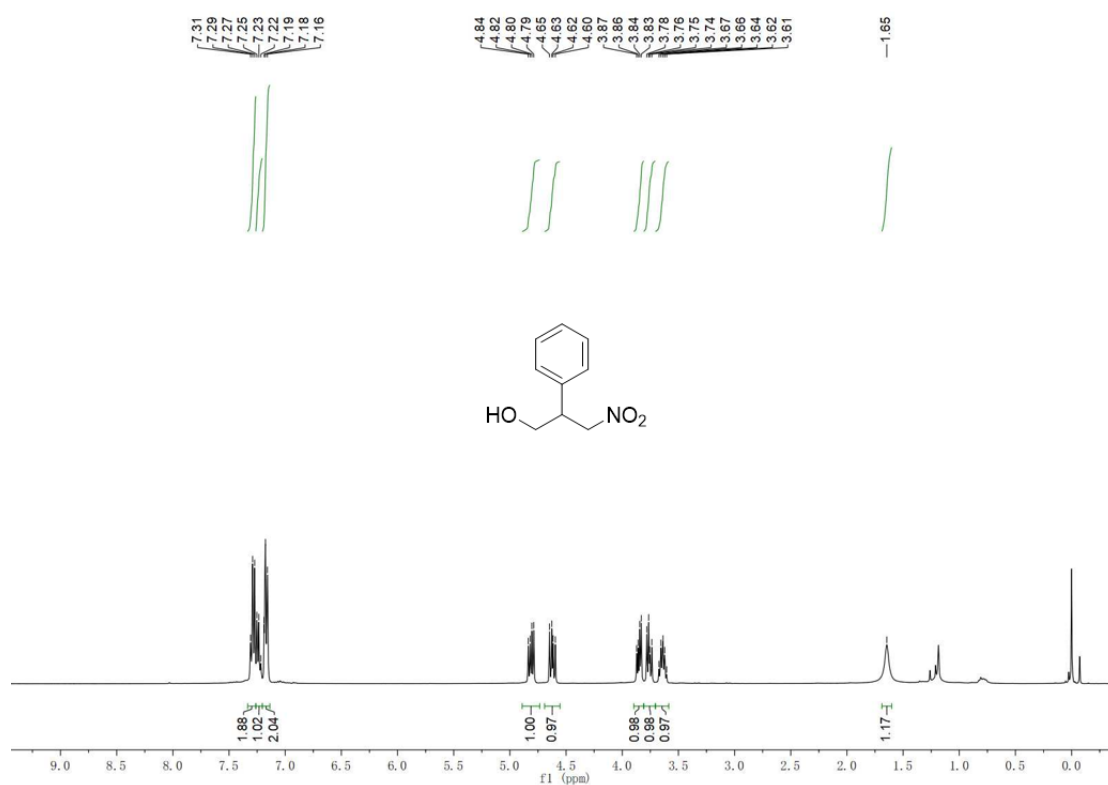
Identification code	3r
Empirical formula	C ₁₅ H ₁₅ NO ₃
Formula weight	257.28
Temperature/K	293(2)
Crystal system	monoclinic
Space group	P2 ₁
a/Å	7.6525(6)
b/Å	5.5652(5)
c/Å	15.5161(11)
α /°	90
β /°	90.246(6)
γ /°	90
Volume/Å ³	660.79(9)
Z	2
ρ _{calc} /cm ³	1.293
μ /mm ⁻¹	0.740
F(000)	272.0
Crystal size/mm ³	0.18 × 0.11 × 0.07
Radiation	Cu K α (λ = 1.54184)
2 θ range for data collection/°	11.406 to 142.186
Index ranges	-9 ≤ h ≤ 9, -5 ≤ k ≤ 6, -18 ≤ l ≤ 18
Reflections collected	6931
Independent reflections	2249 [R _{int} = 0.0349, R _{sigma} = 0.0336]
Data/restraints/parameters	2249/40/184
Goodness-of-fit on F ²	1.059
Final R indexes [I ≥ 2 σ (I)]	R ₁ = 0.0578, wR ₂ = 0.1659
Final R indexes [all data]	R ₁ = 0.0686, wR ₂ = 0.1814
Largest diff. peak/hole / e Å ⁻³	0.24/-0.15

11. References

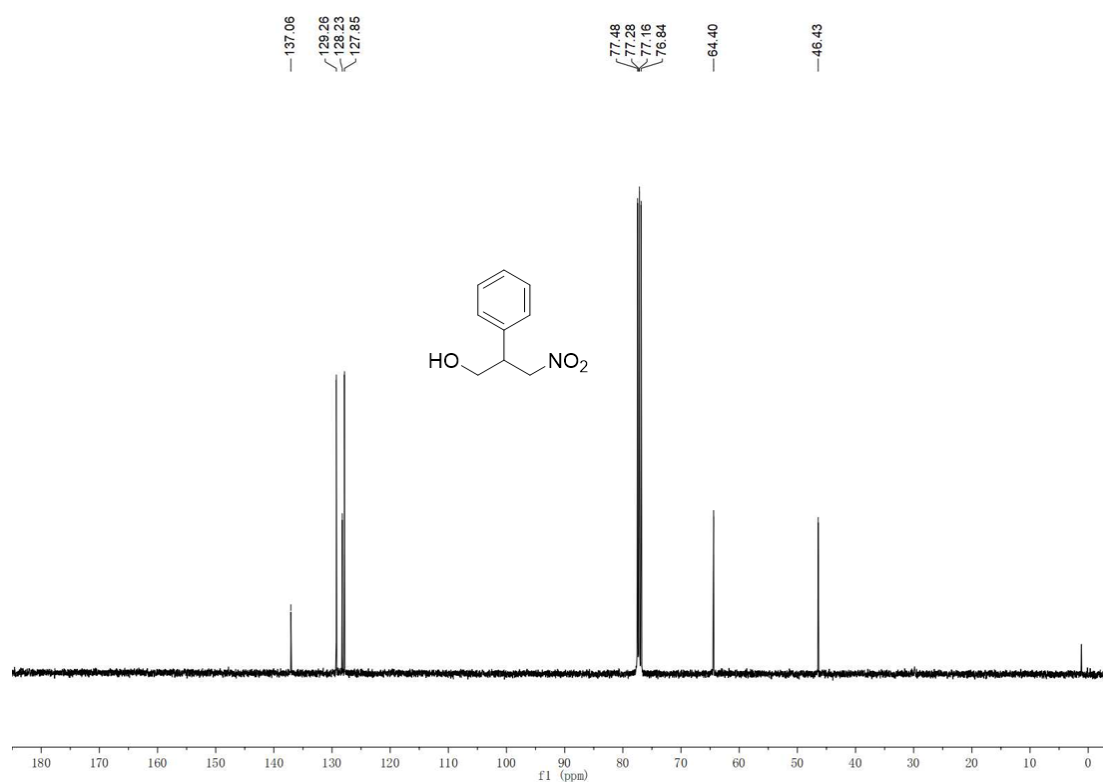
- [1] S. P. Midya, S. Mondal, A. S. M. Islam, A. Rashid, S. Mondal, A. Paul and P. Ghosh, *Org. Lett.*, 2022, **24**, 4438-4443.
- [2] S. Cai, X. Zhao, X. Wang, Q. Liu, Z. Li and D. Z. Wang, *Angew. Chem. Int. Ed.*, 2012, **51**, 8050-8053.
- [3] O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, *J. Appl. Cryst.*, 2009, **42**, 339-341.
- [4] G. M. Sheldrick, *Acta Cryst.*, 2008, **A64**, 112-122.
- [5] G. M. Sheldrick, *Acta Cryst.*, 2015, **C71**, 3-8.

12. ^1H , ^{13}C NMR spectra for compounds 3, 4, 5

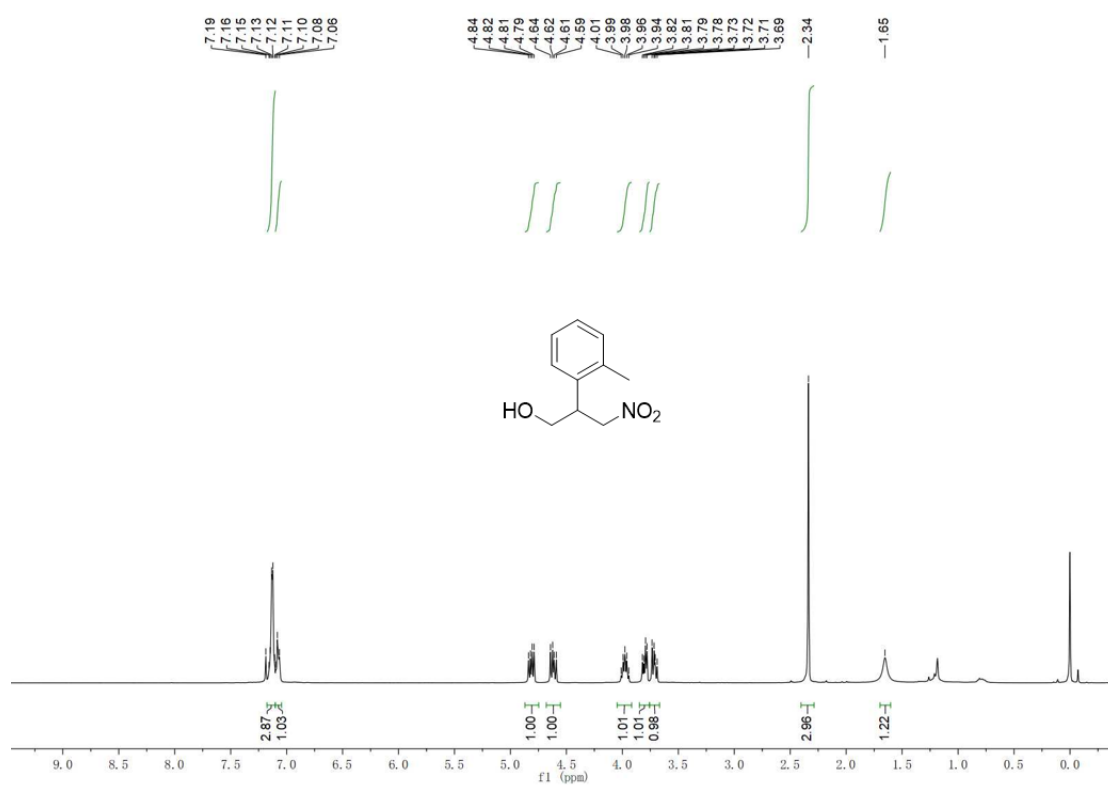
^1H NMR (400 MHz, CDCl_3) of **3a**



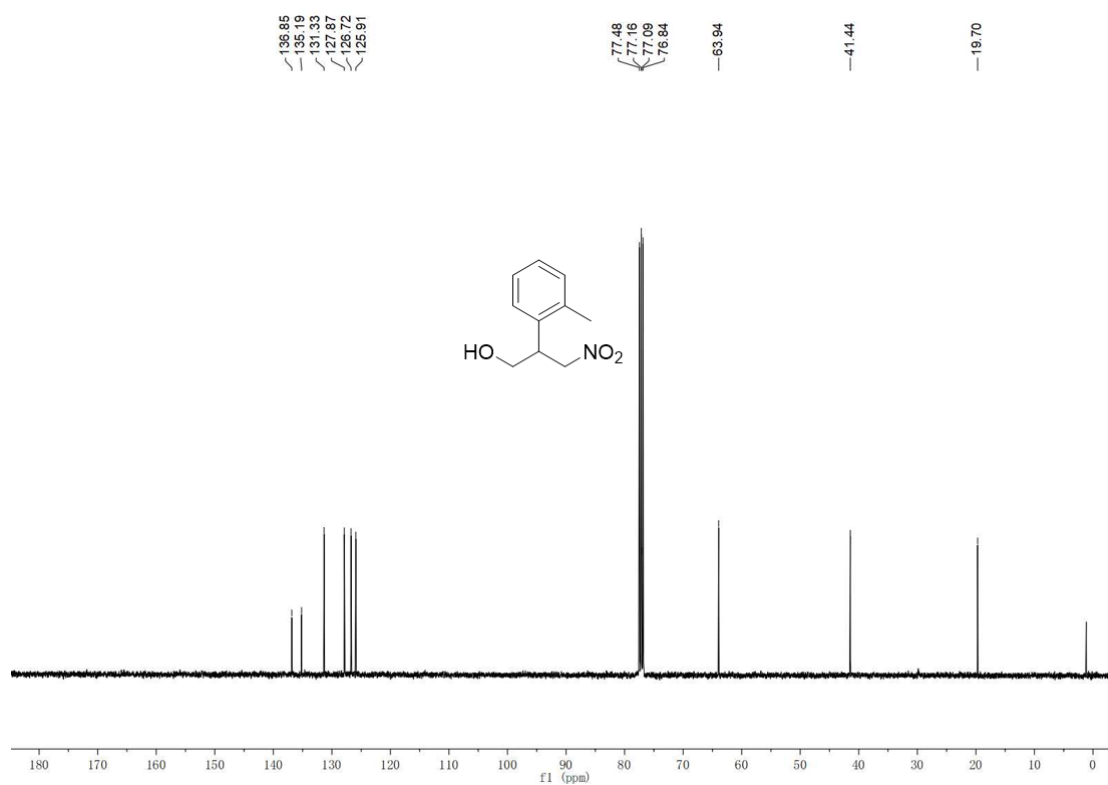
^{13}C NMR (101 MHz, CDCl_3) of **3a**



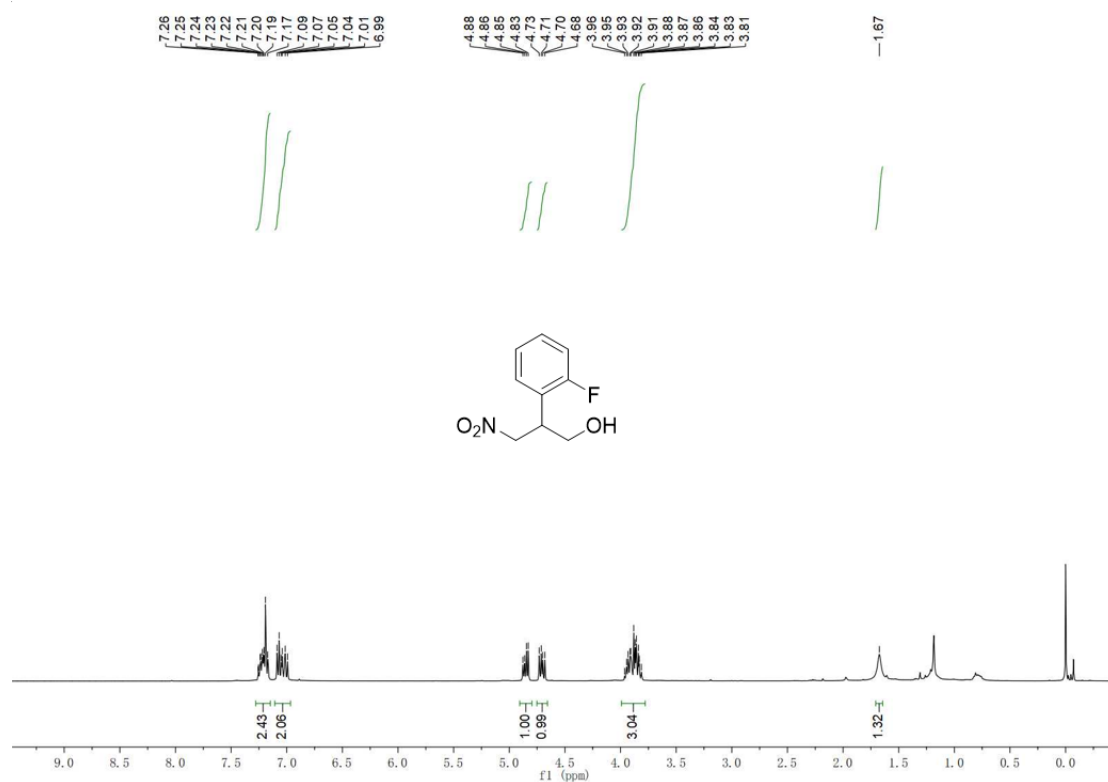
¹H NMR (400 MHz, CDCl₃) of **3b**



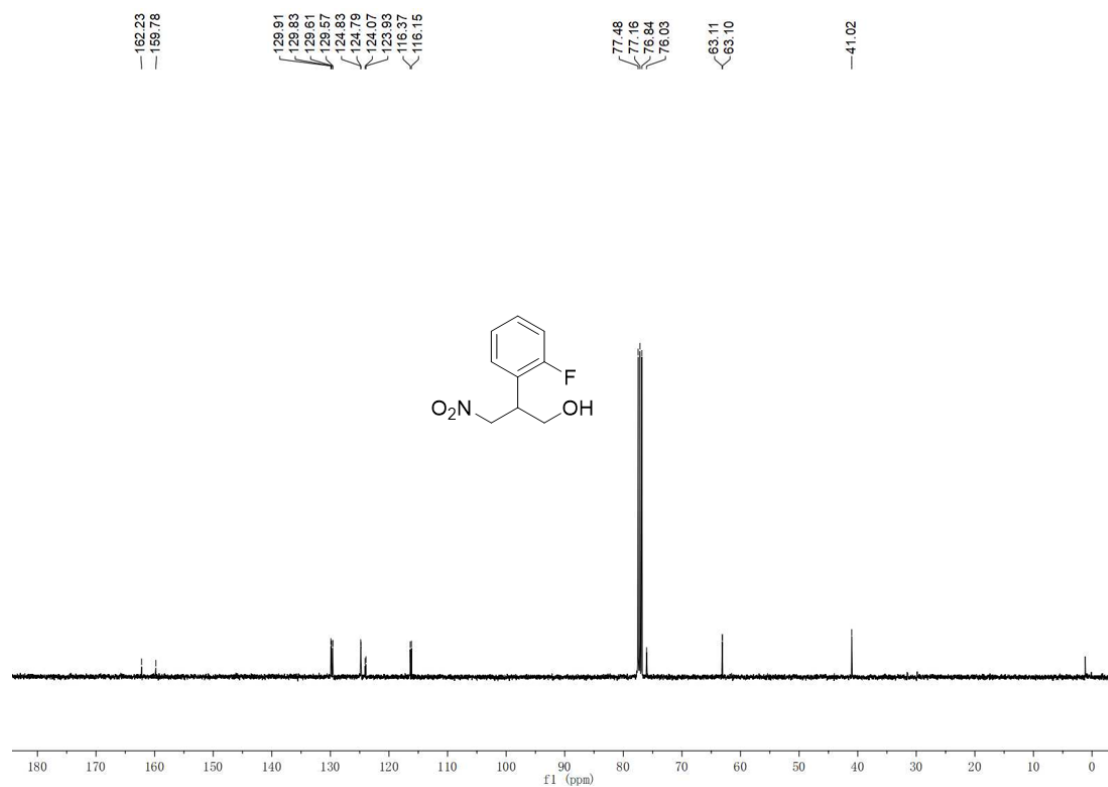
¹³C NMR (101 MHz, CDCl₃) of **3b**



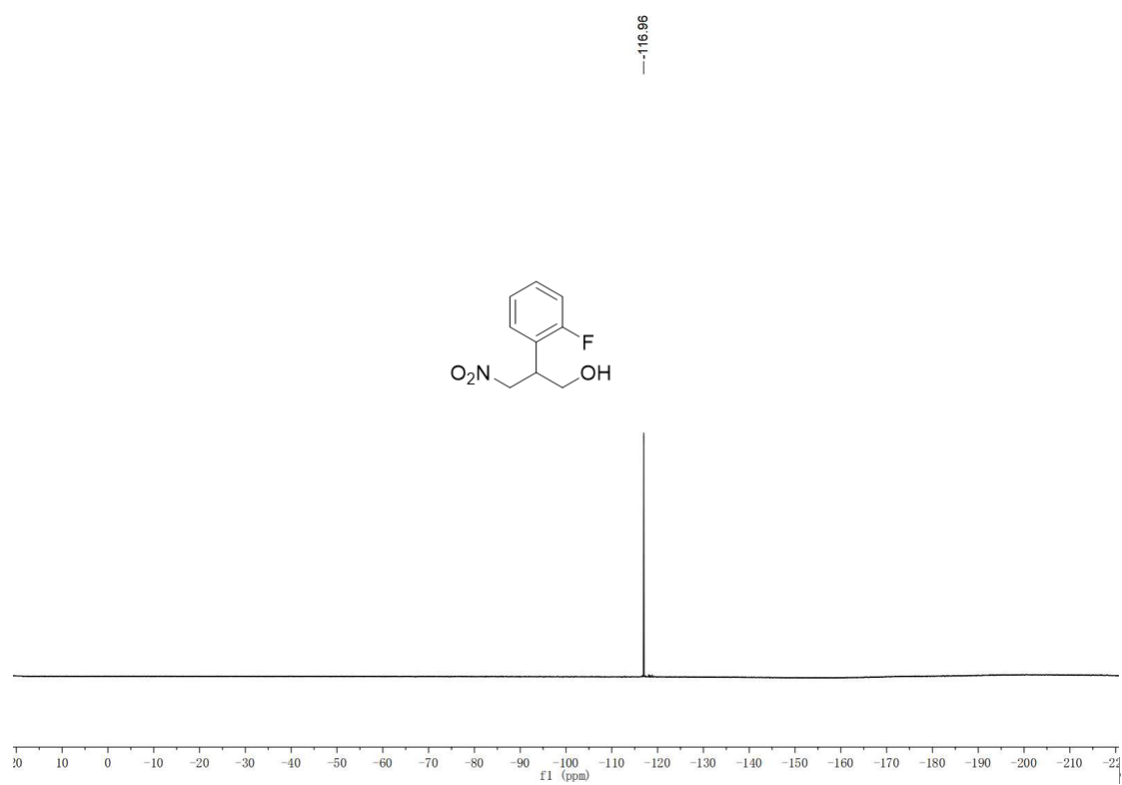
¹H NMR (400 MHz, CDCl₃) of **3c**



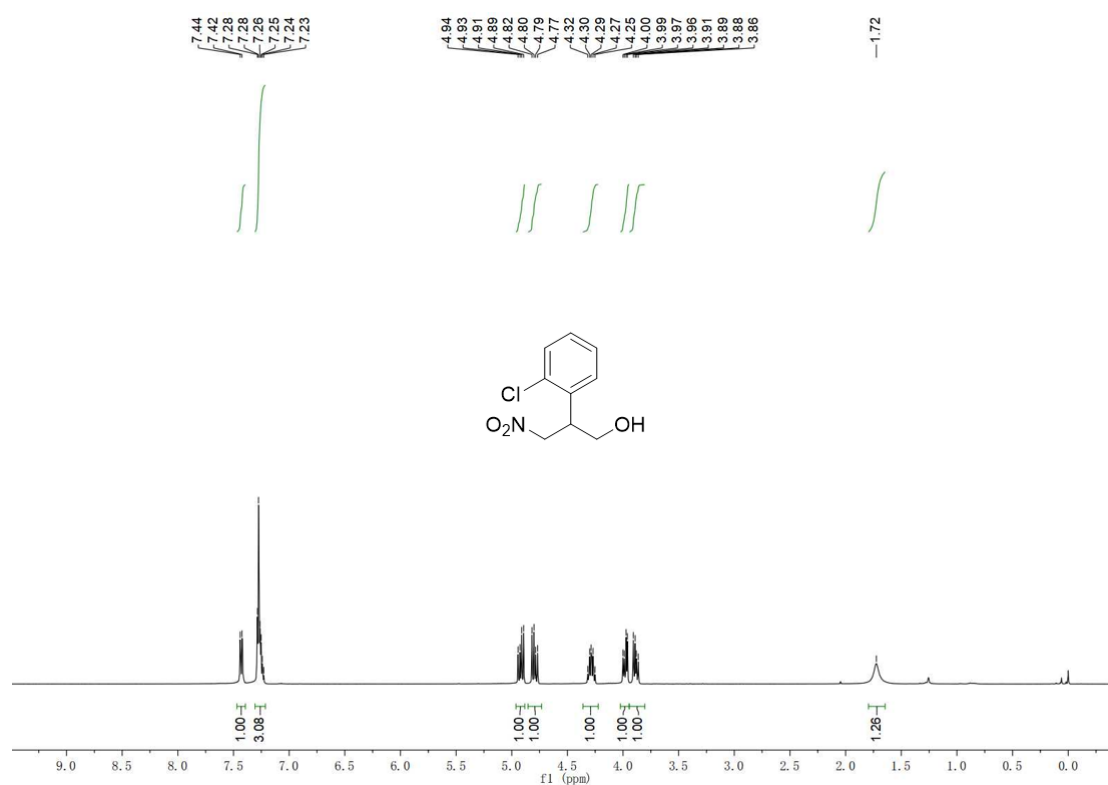
¹³C NMR (101 MHz, CDCl₃) of **3c**



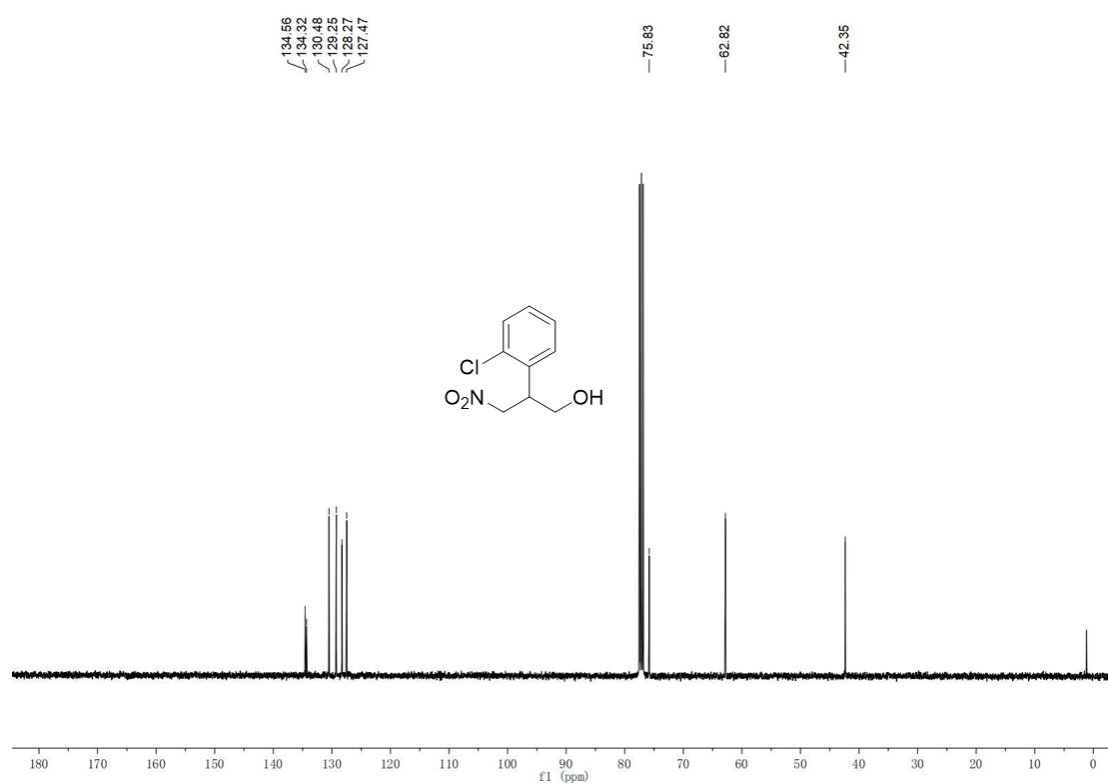
^{19}F NMR (376 MHz, CDCl_3) of **3c**



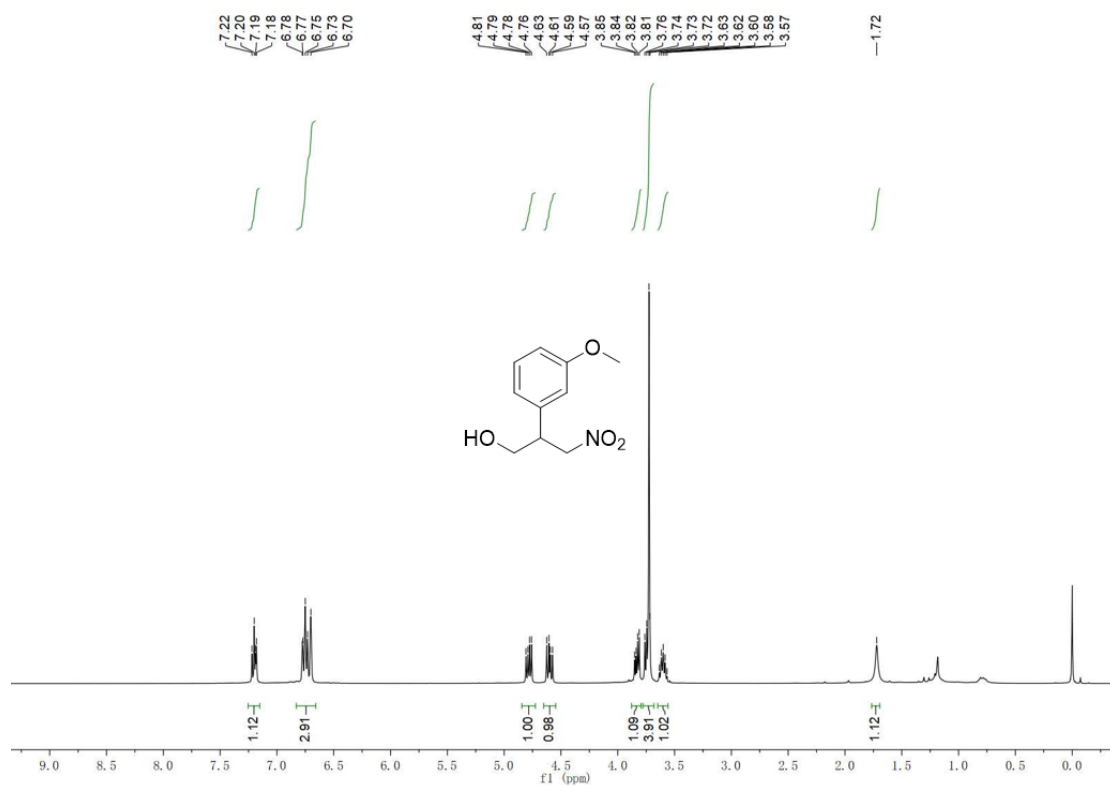
¹H NMR (400 MHz, CDCl₃) of **3d**



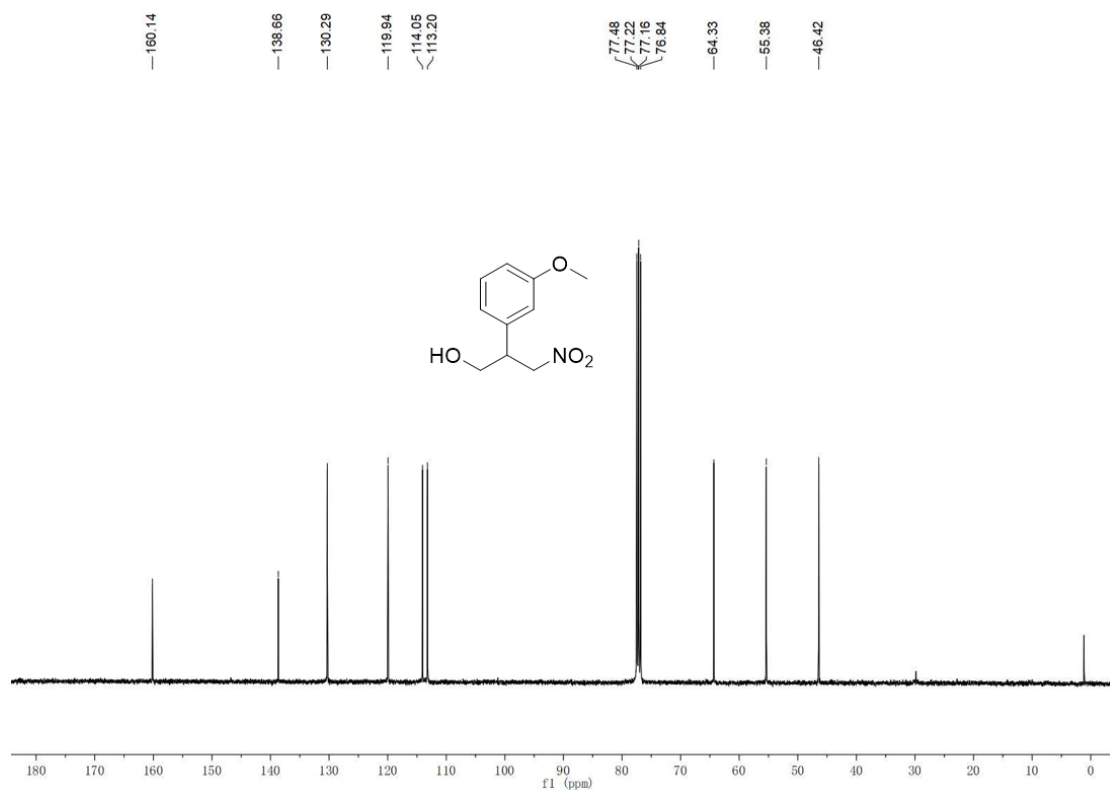
¹³C NMR (101 MHz, CDCl₃) of **3d**



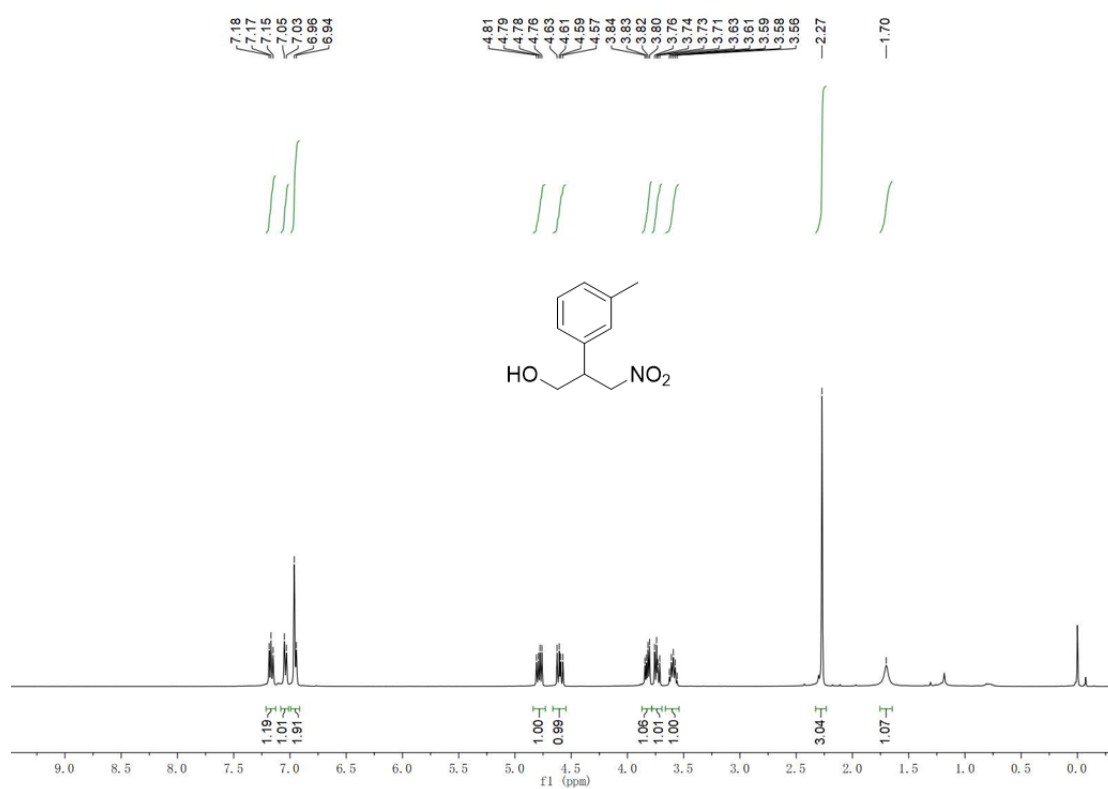
¹H NMR (400 MHz, CDCl₃) of **3e**



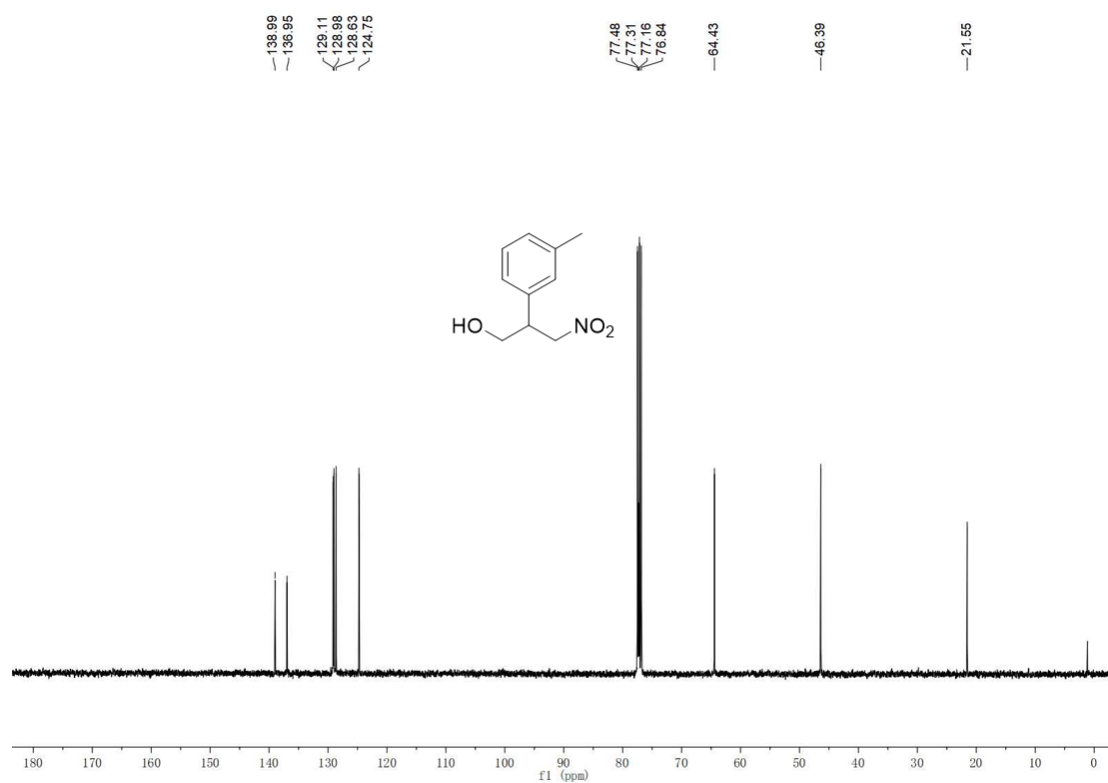
¹³C NMR (101 MHz, CDCl₃) of **3e**



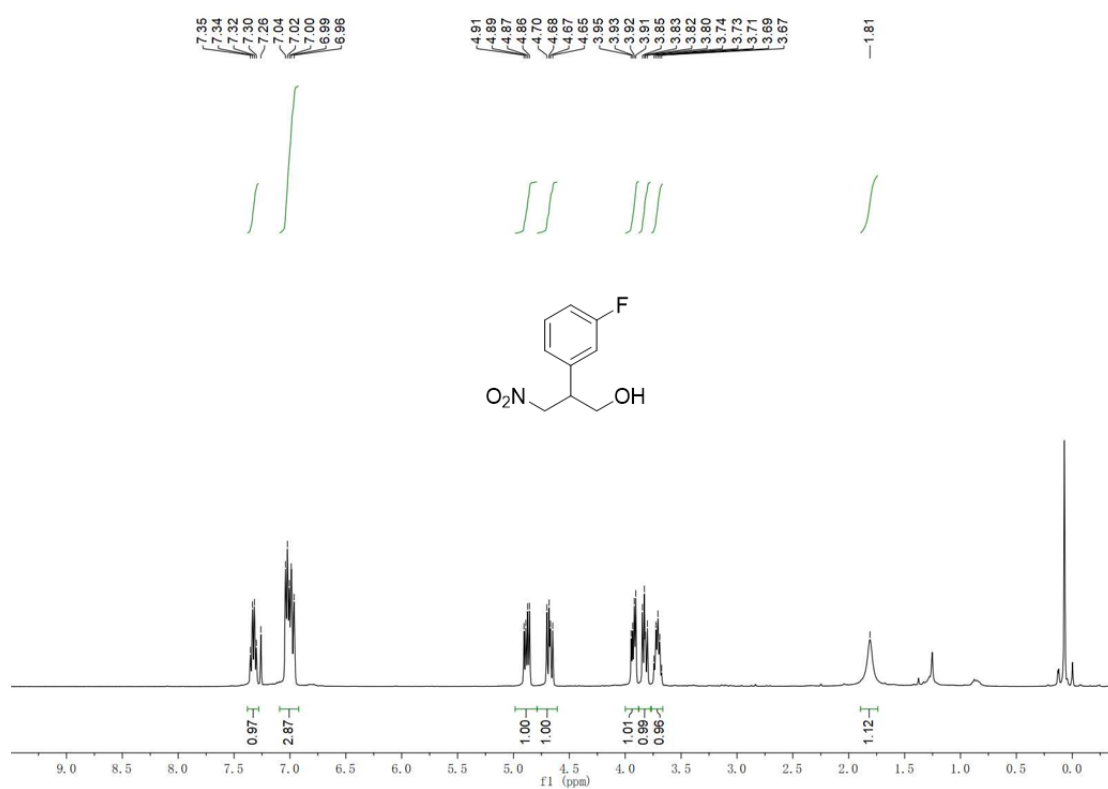
¹H NMR (400 MHz, CDCl₃) of **3f**



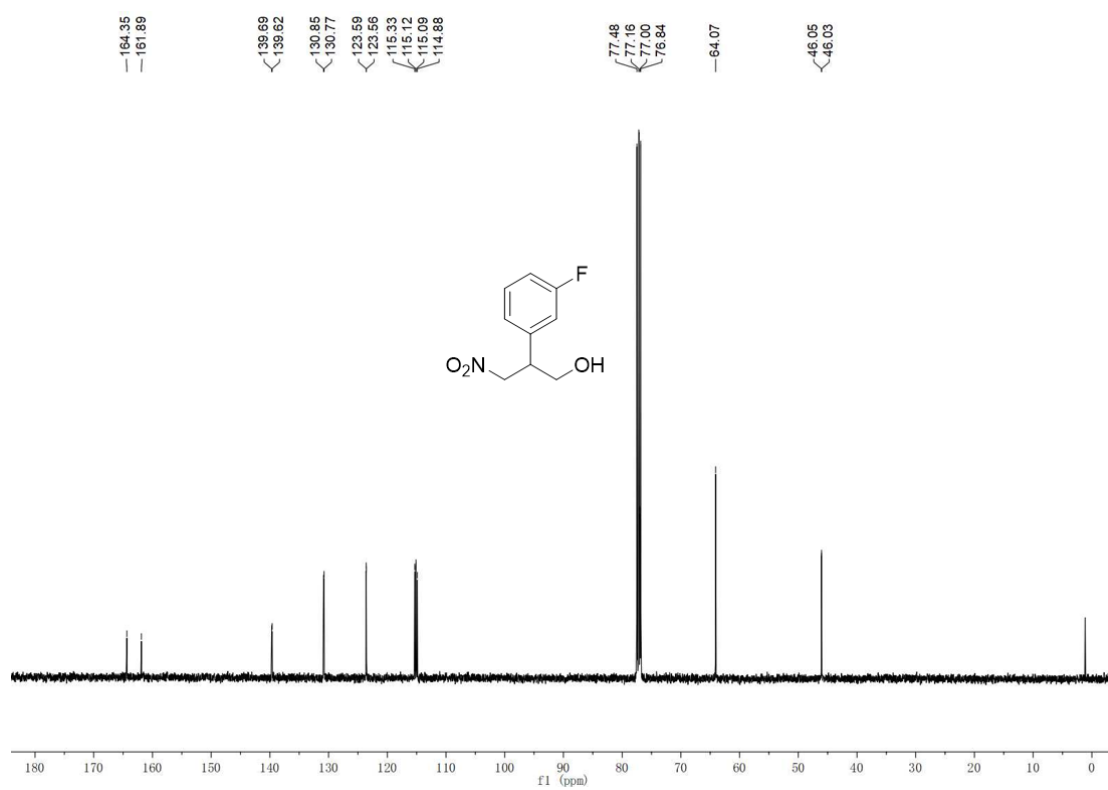
¹³C NMR (101 MHz, CDCl₃) of **3f**



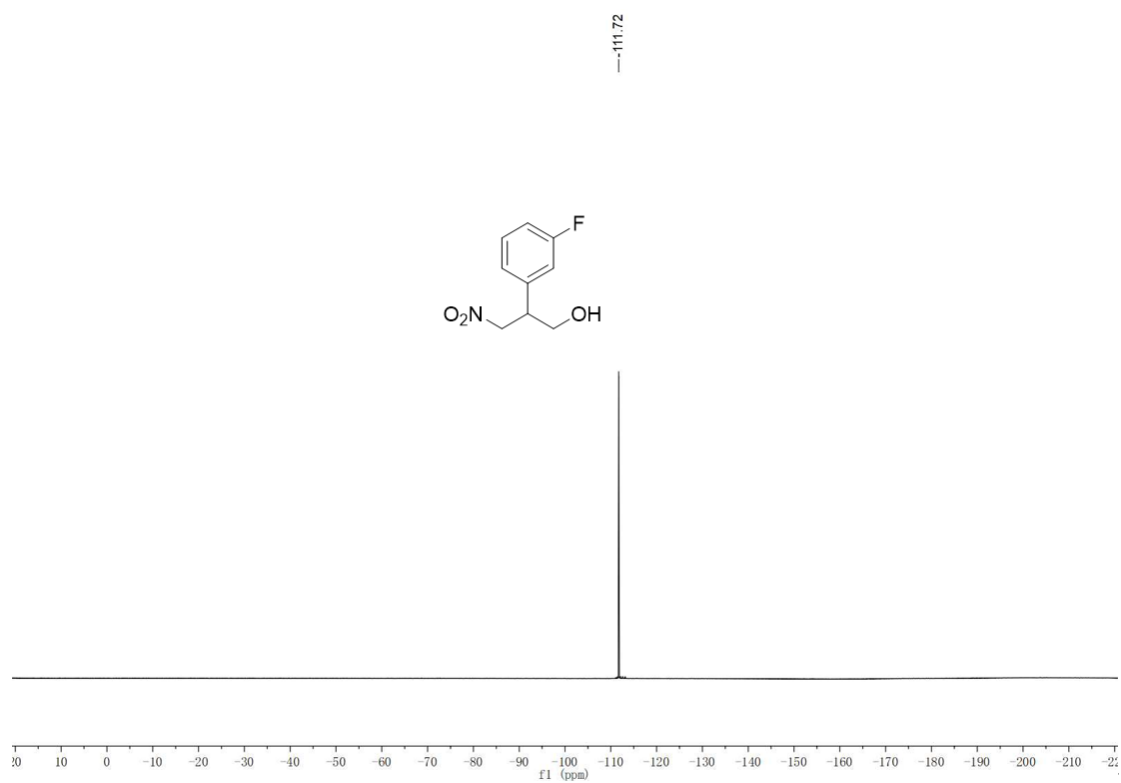
^1H NMR (400 MHz, CDCl_3) of **3g**



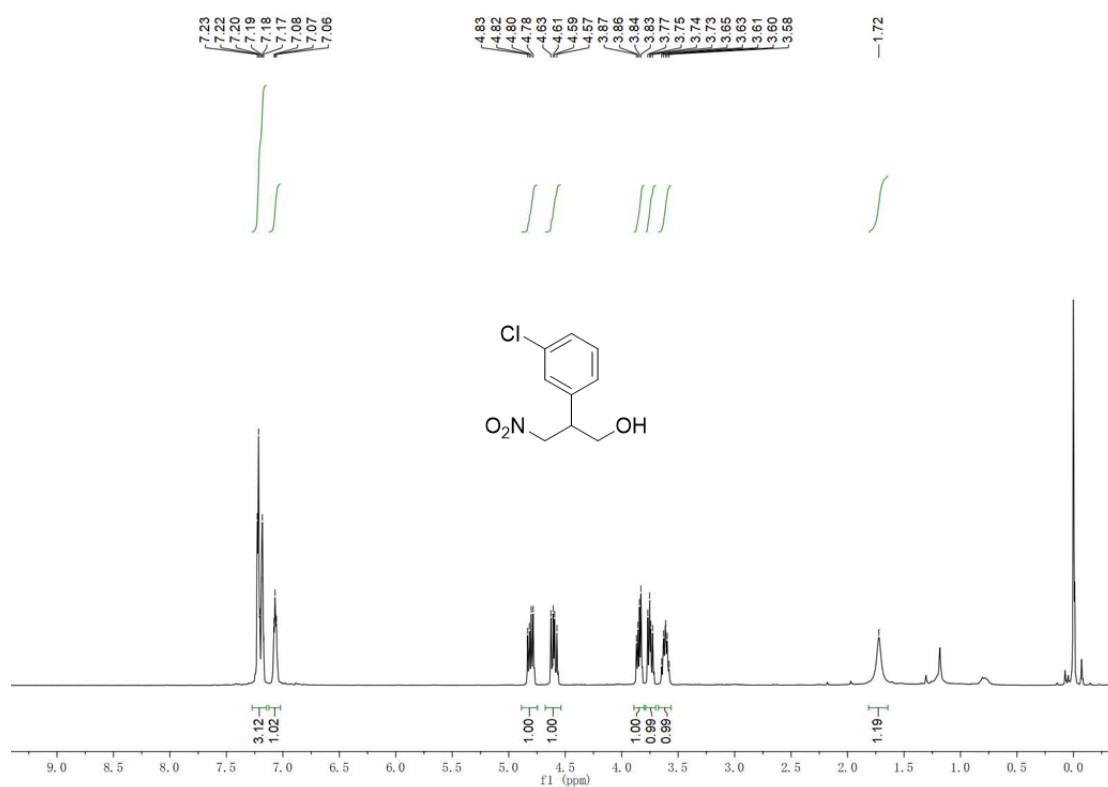
^{13}C NMR (101 MHz, CDCl_3) of **3g**



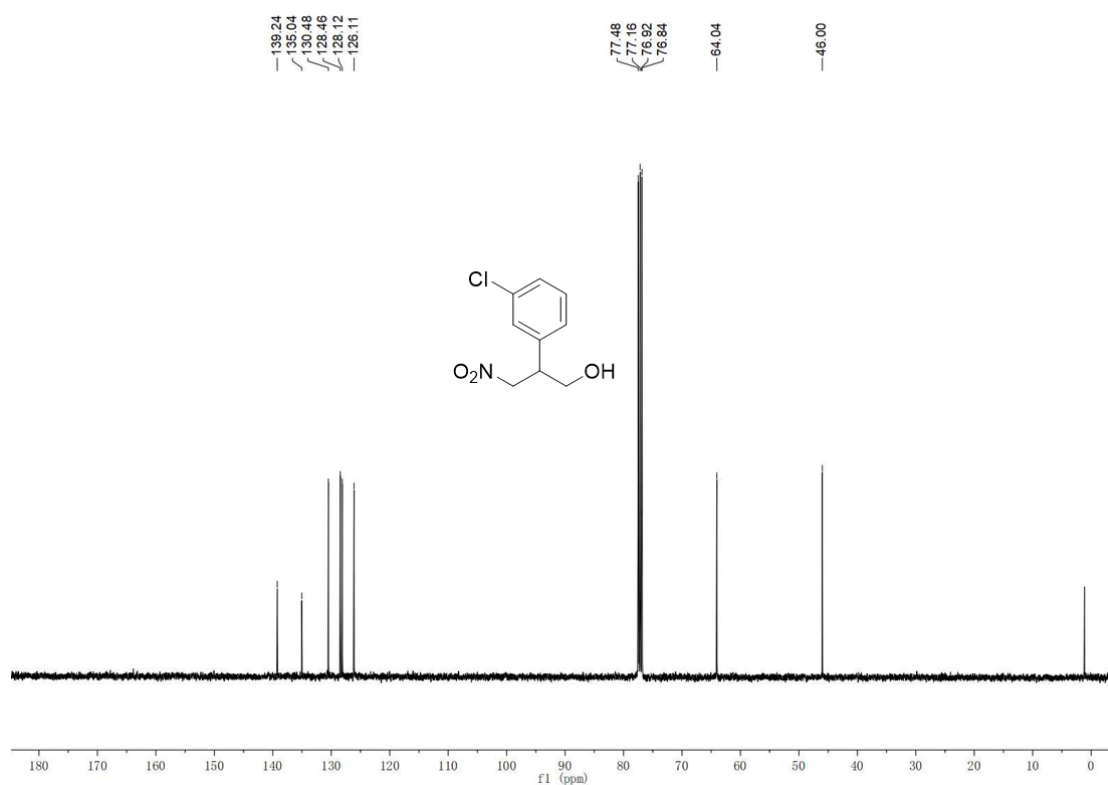
^{19}F NMR (376 MHz, CDCl_3) of **3g**



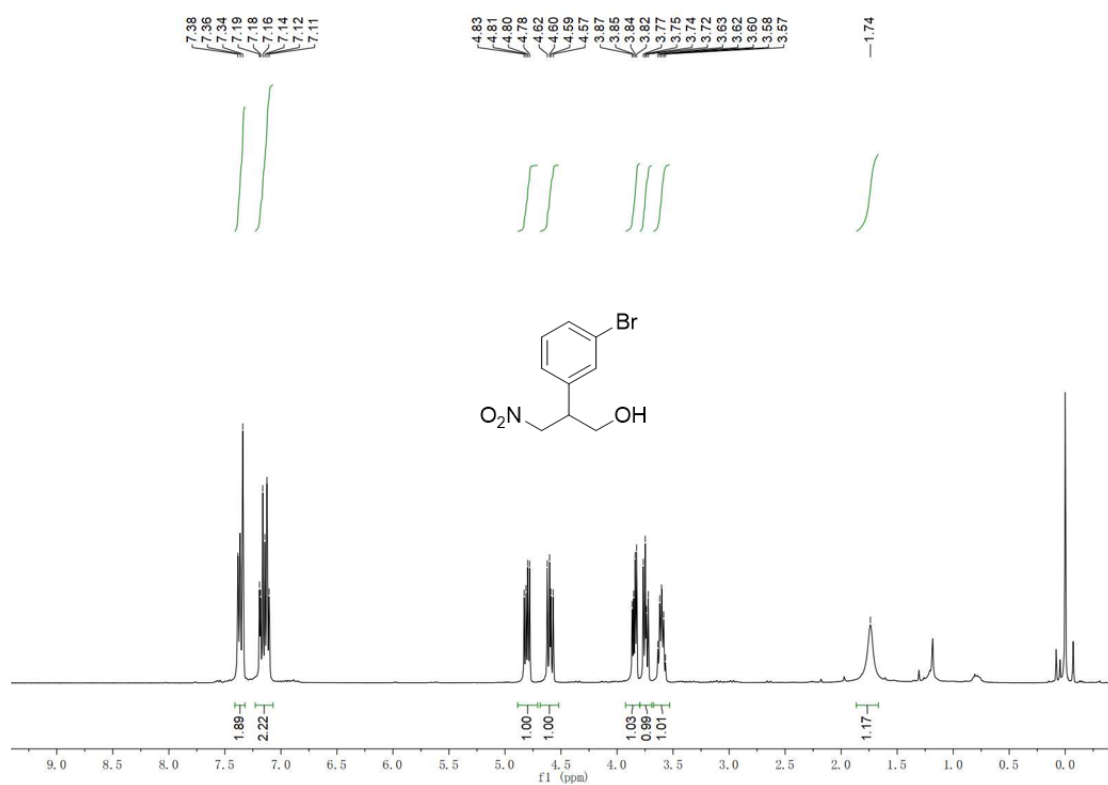
¹H NMR (400 MHz, CDCl₃) of **3h**



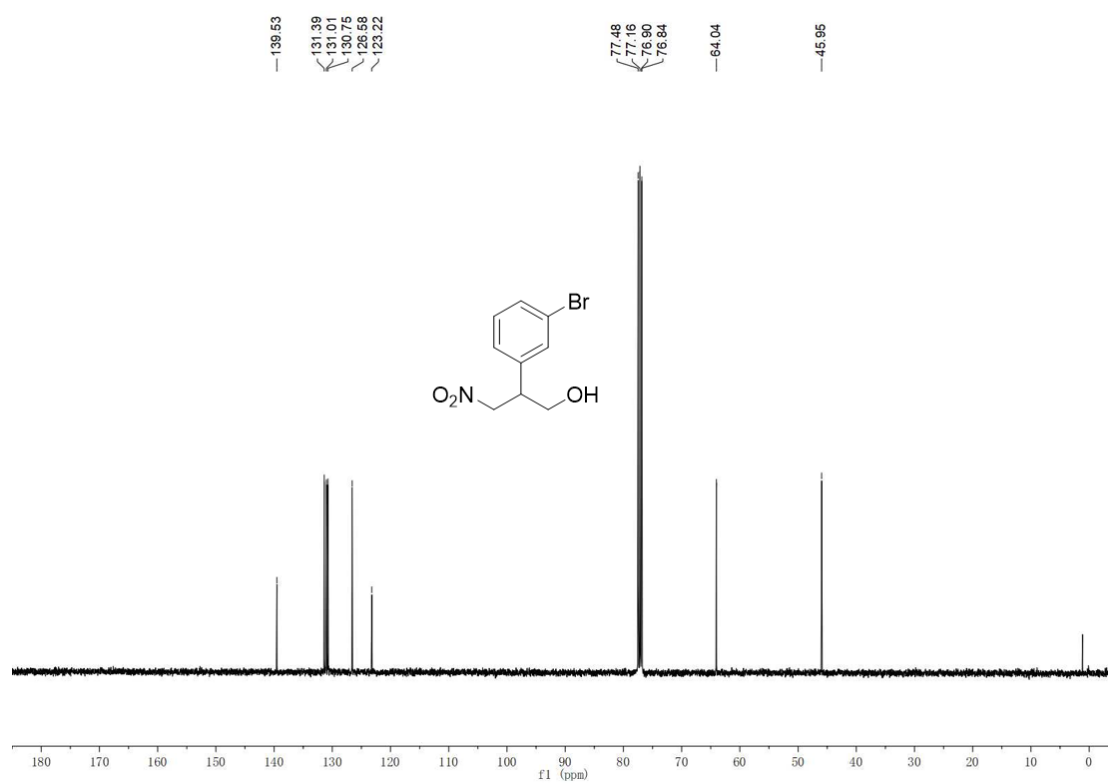
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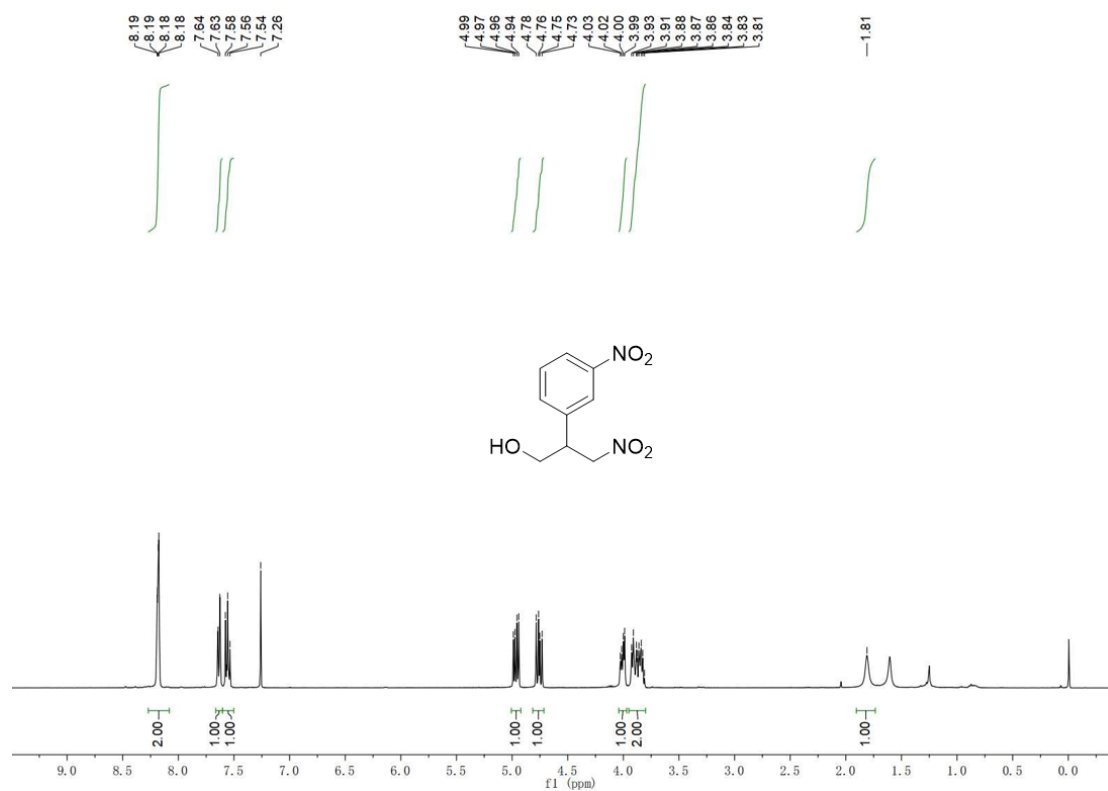
¹H NMR (400 MHz, CDCl₃) of **3i**



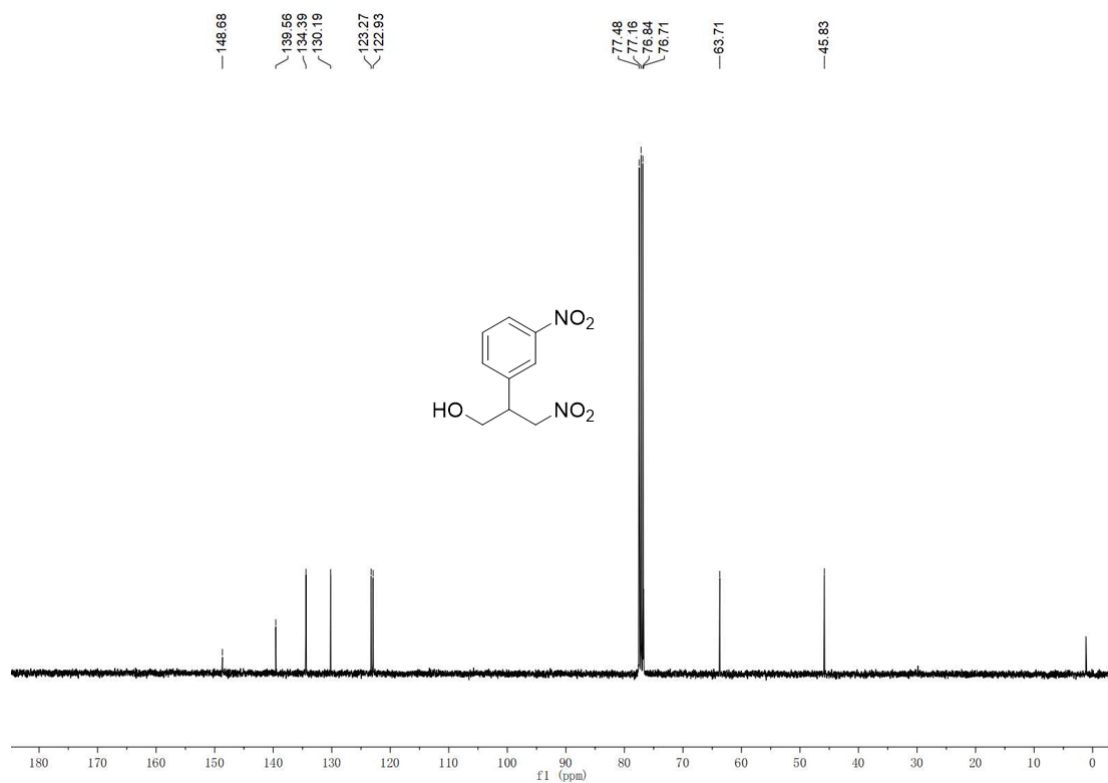
¹³C NMR (101 MHz, CDCl₃) of **3i**



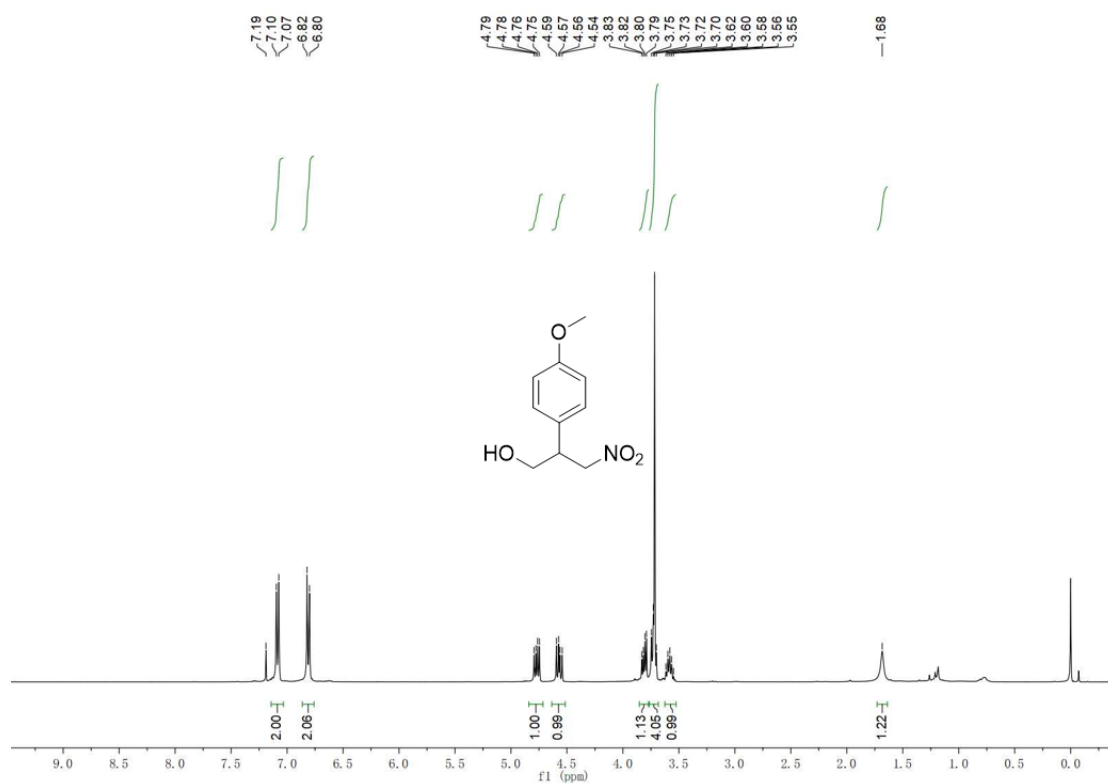
¹H NMR (400 MHz, CDCl₃) of **3j**



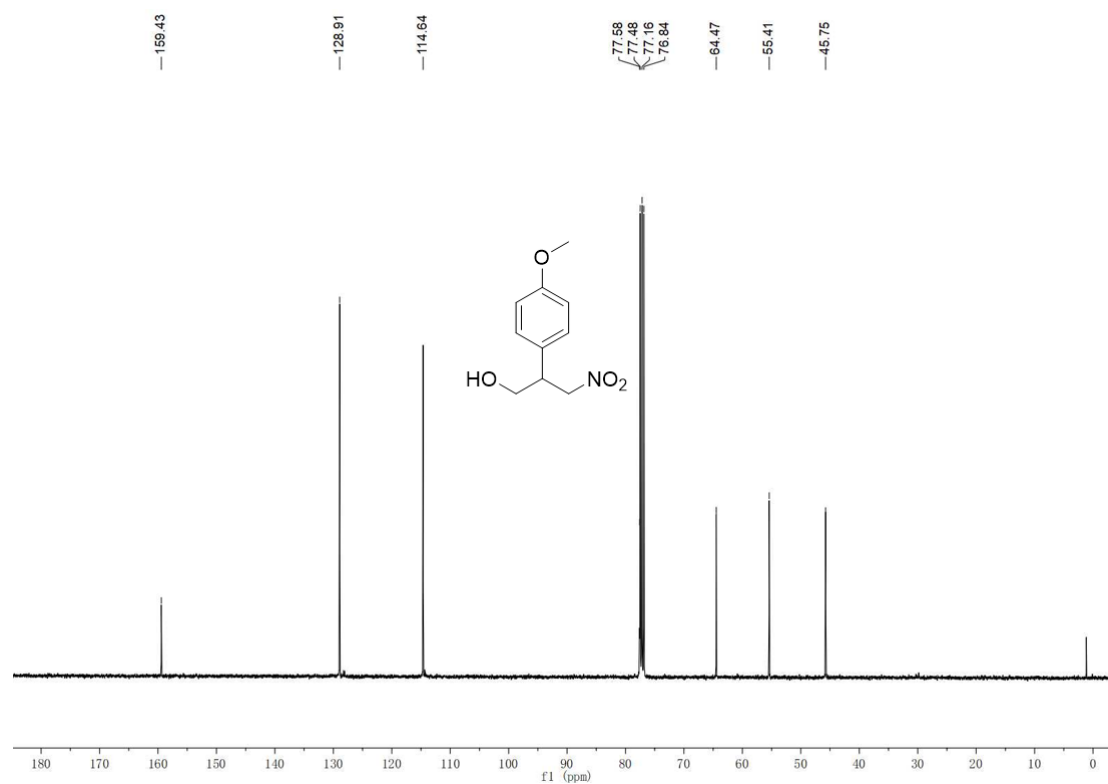
¹³C NMR (101 MHz, CDCl₃) of **3j**



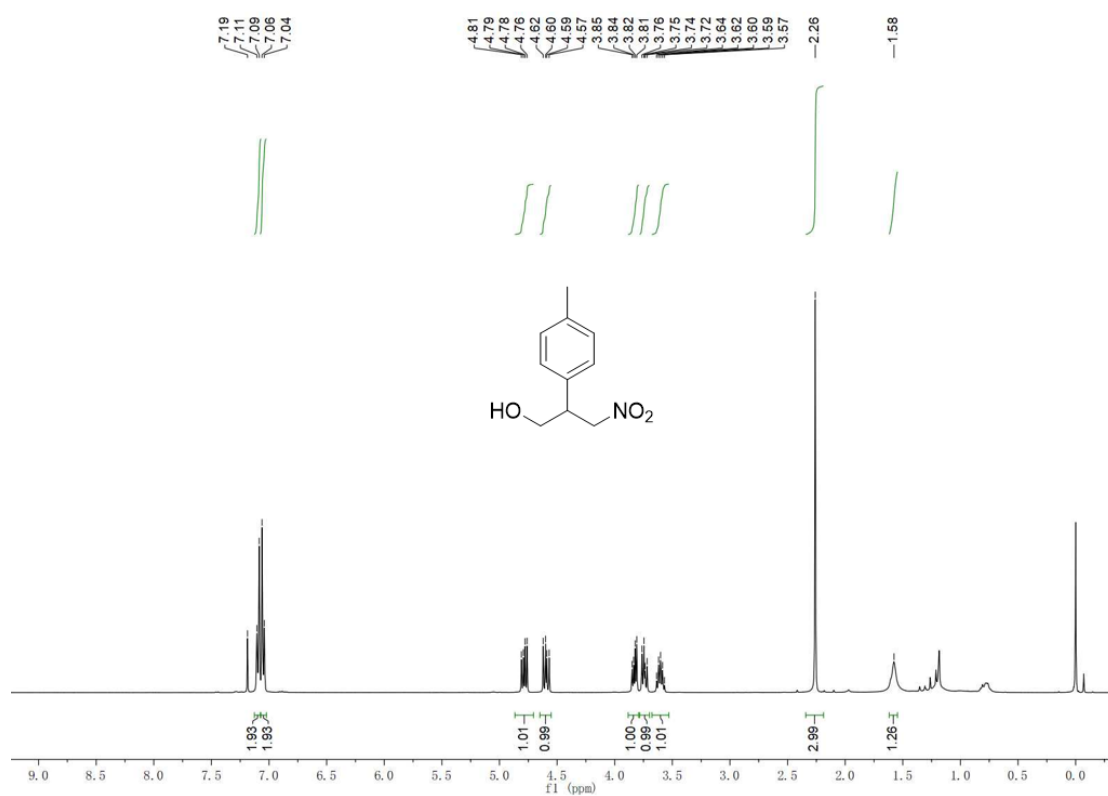
¹H NMR (400 MHz, CDCl₃) of **3k**



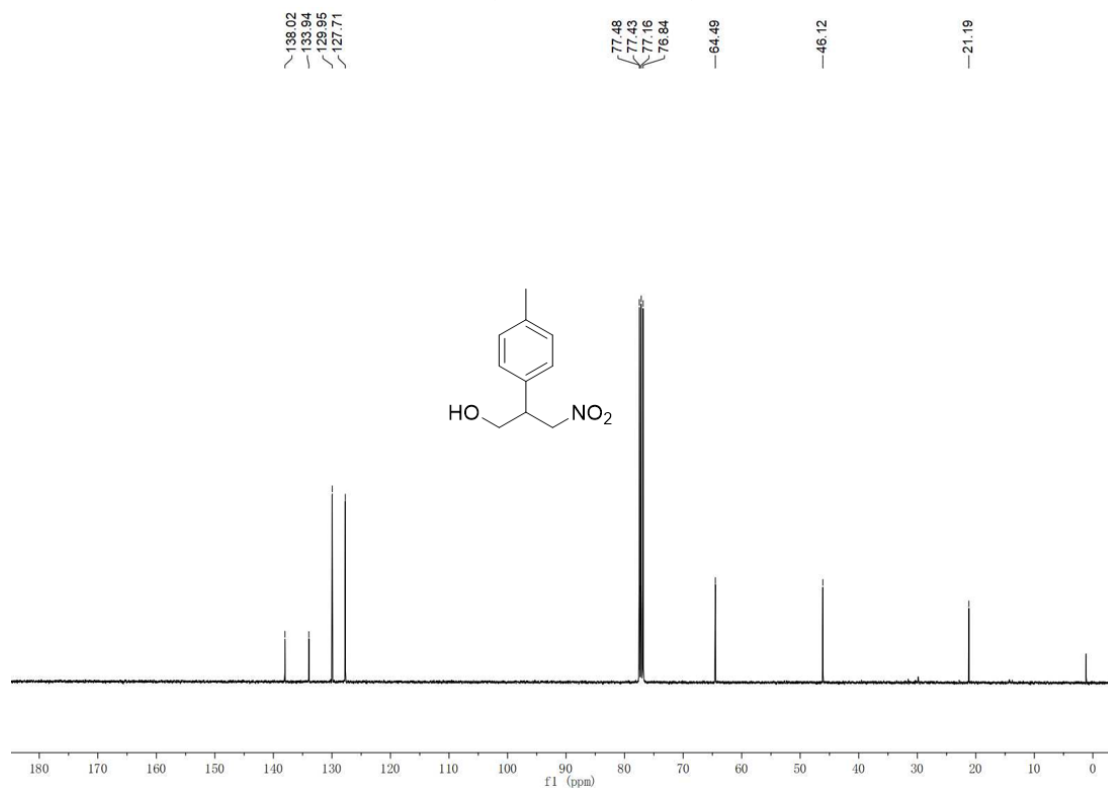
¹³C NMR (101 MHz, CDCl₃) of **3k**



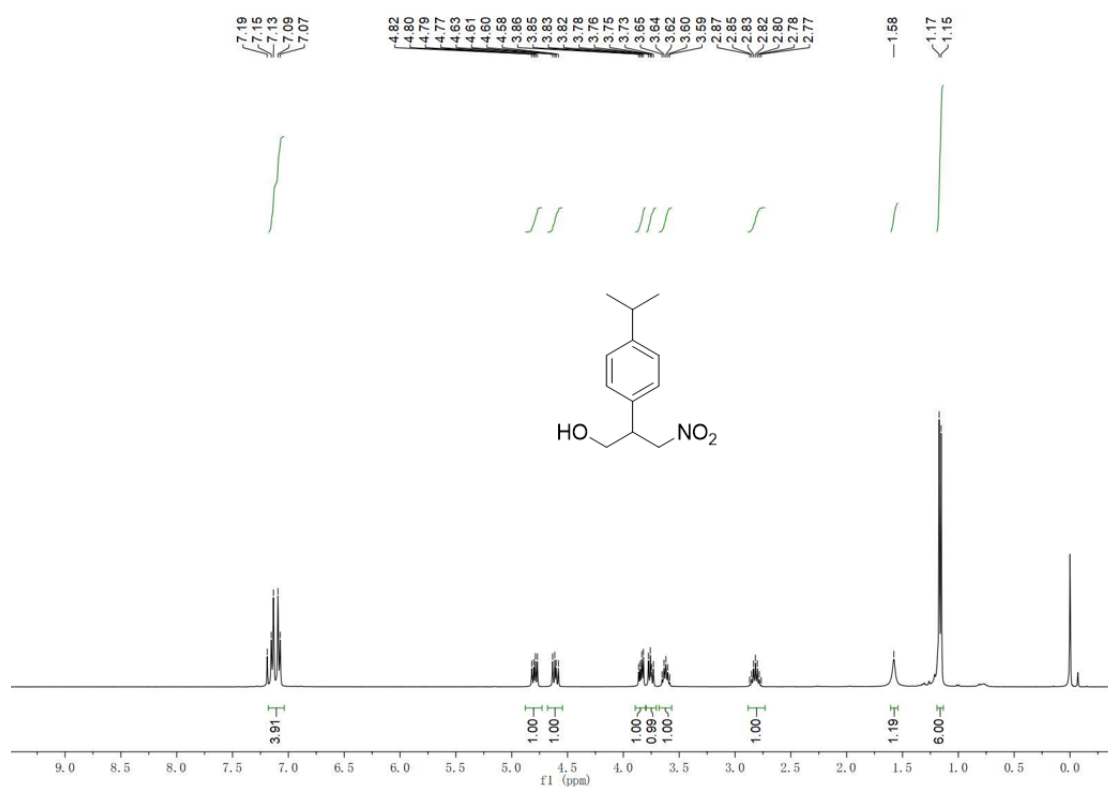
¹H NMR (400 MHz, CDCl₃) of **31**



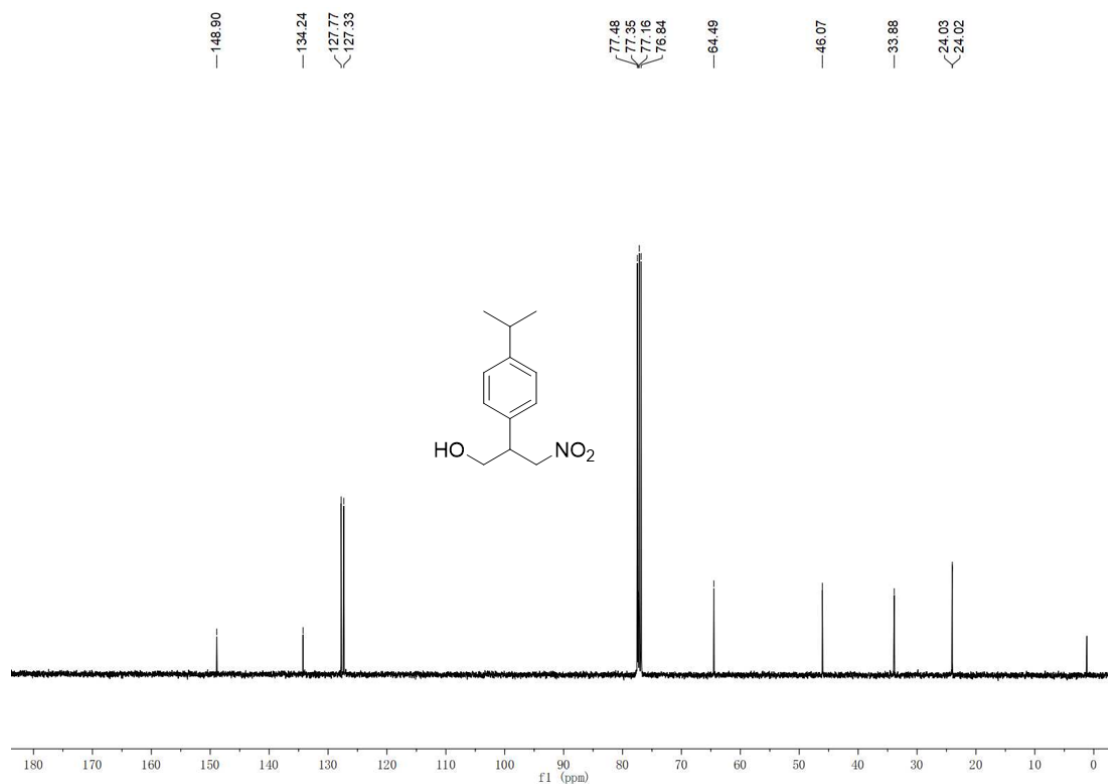
¹³C NMR (101 MHz, CDCl₃) of **31**



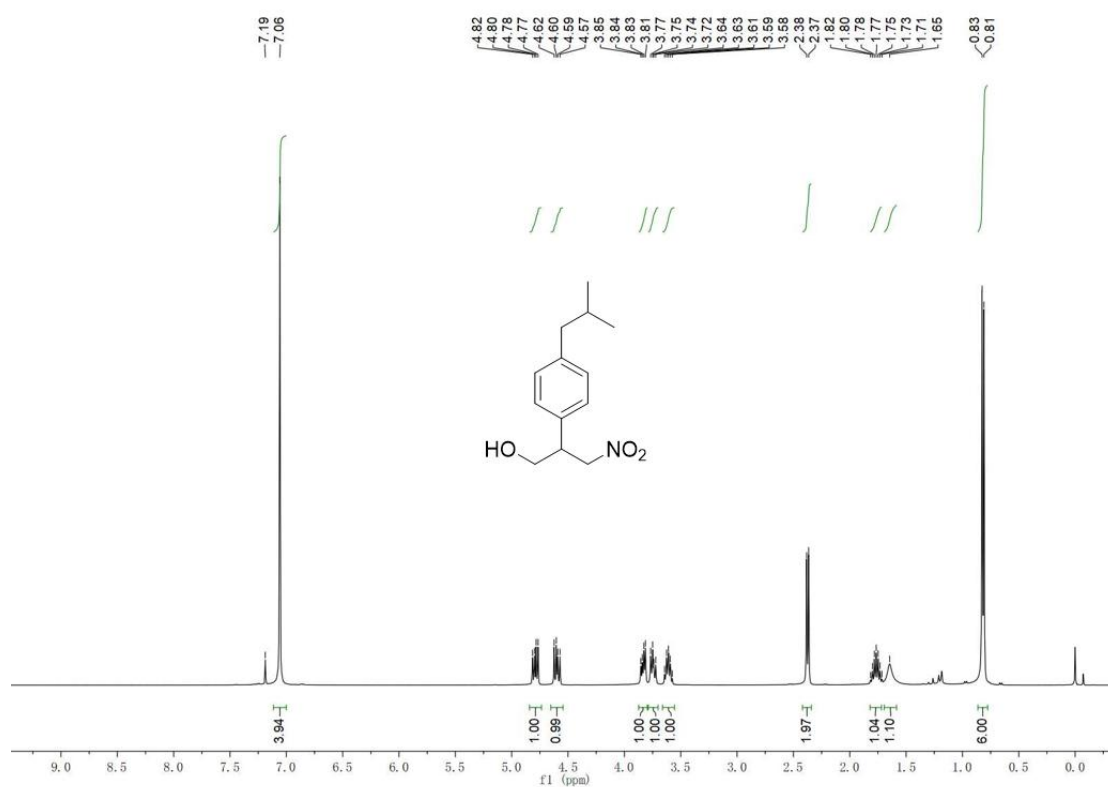
¹H NMR (400 MHz, CDCl₃) of **3m**



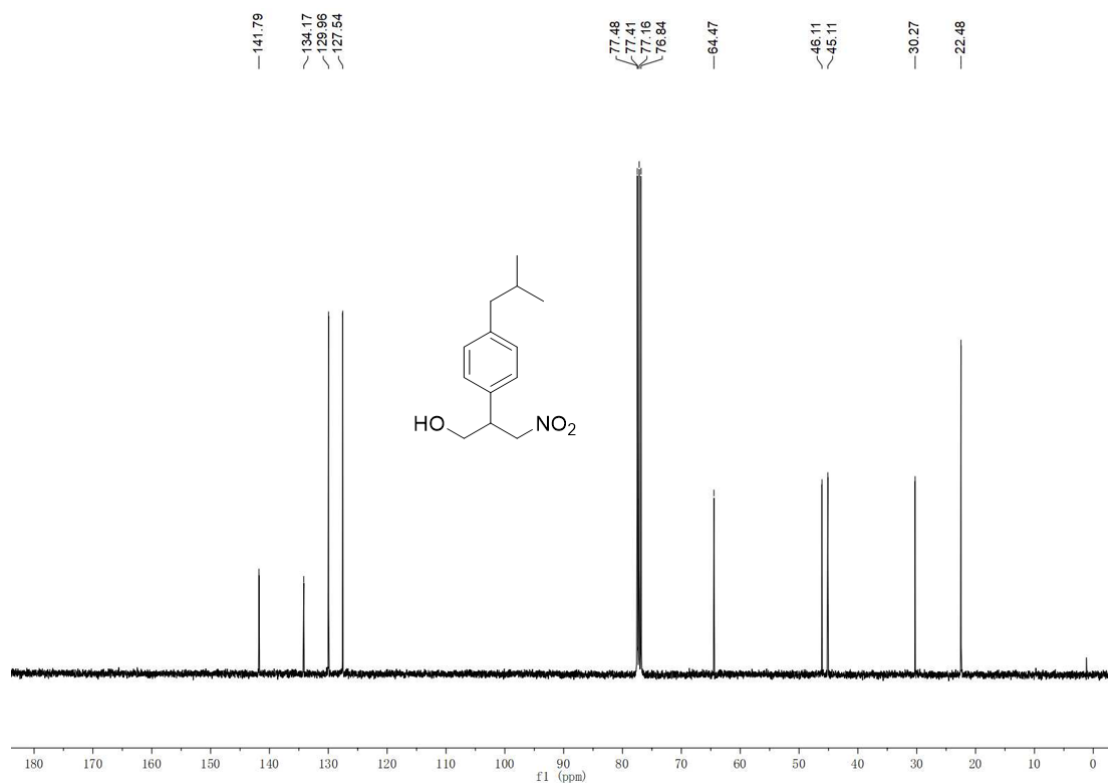
¹³C NMR (101 MHz, CDCl₃) of **3m**



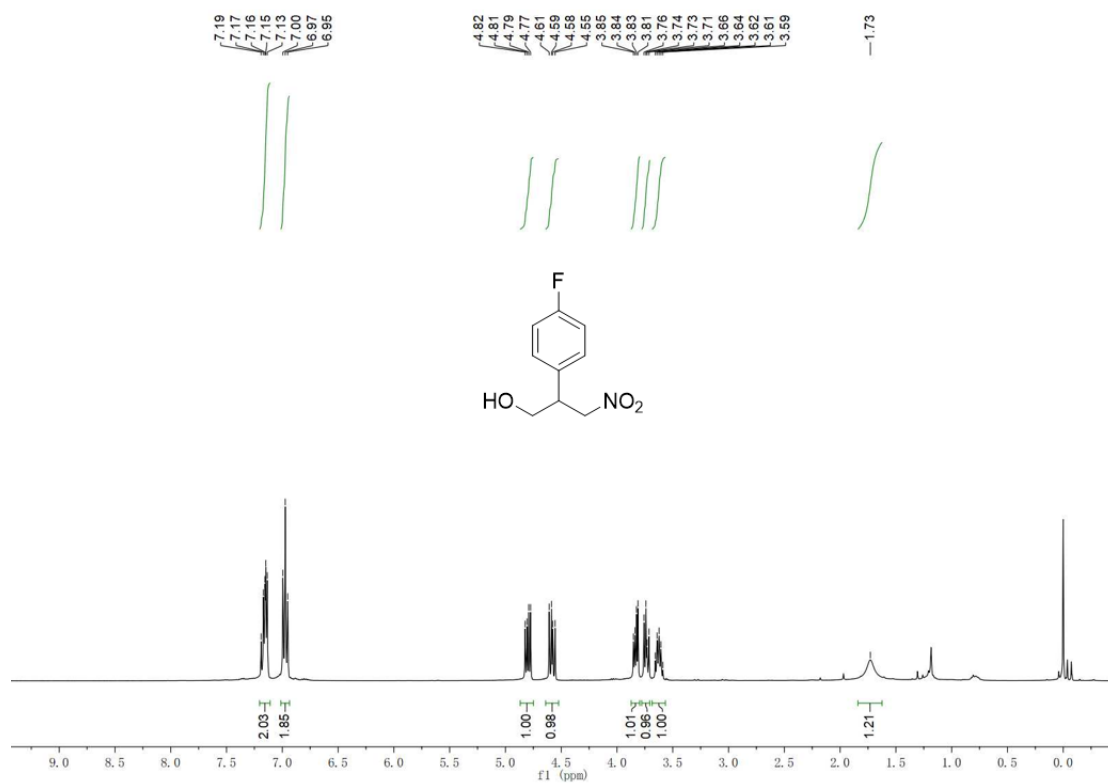
^1H NMR (400 MHz, CDCl_3) of **3n**



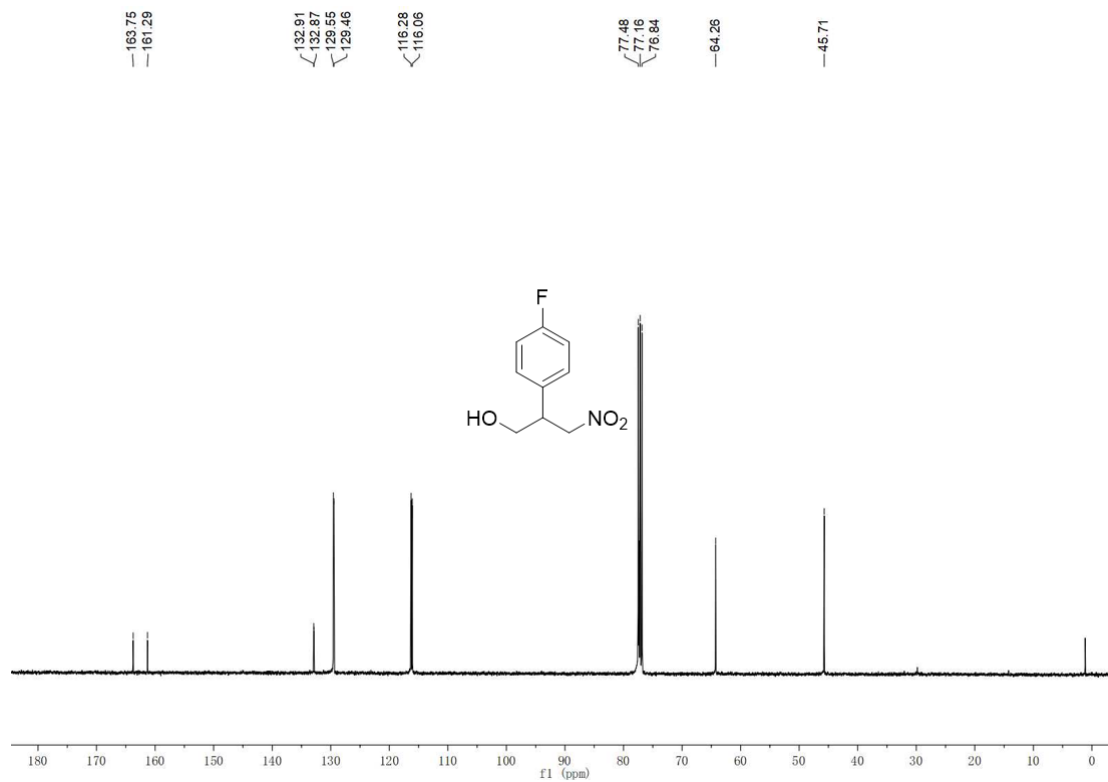
^{13}C NMR (101 MHz, CDCl_3) of **3n**



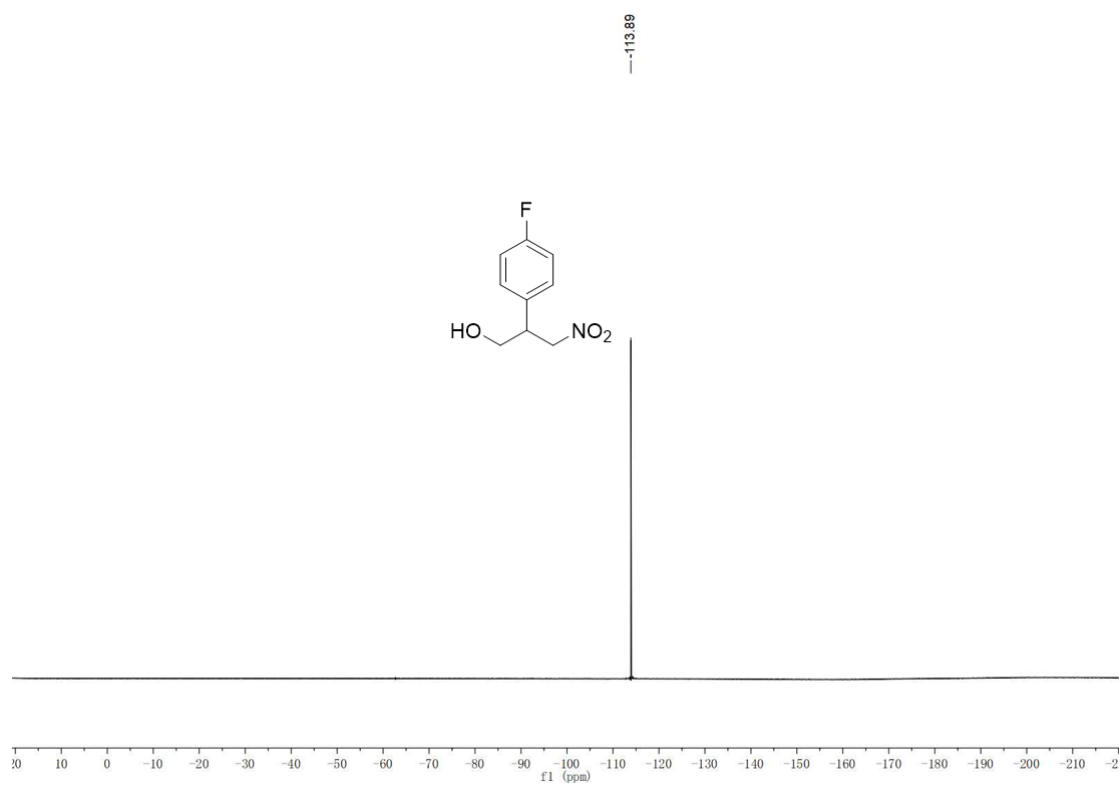
¹H NMR (400 MHz, CDCl₃) of **3o**



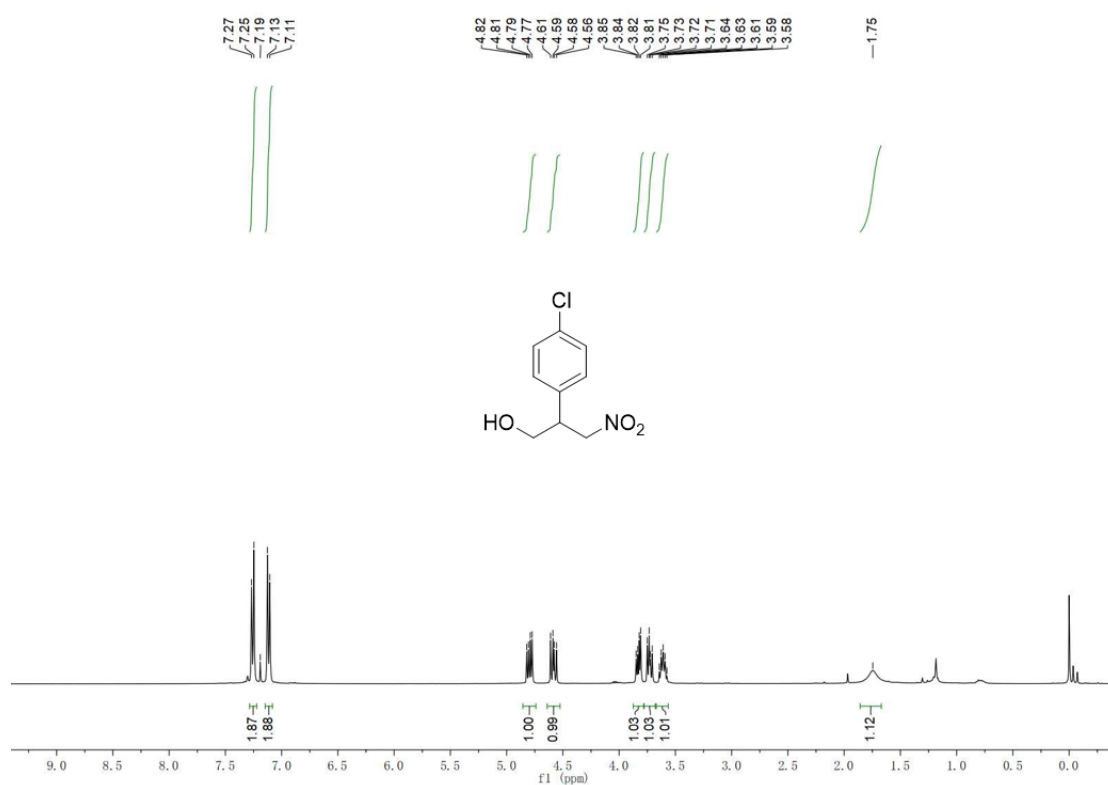
¹³C NMR (101 MHz, CDCl₃) of **3o**



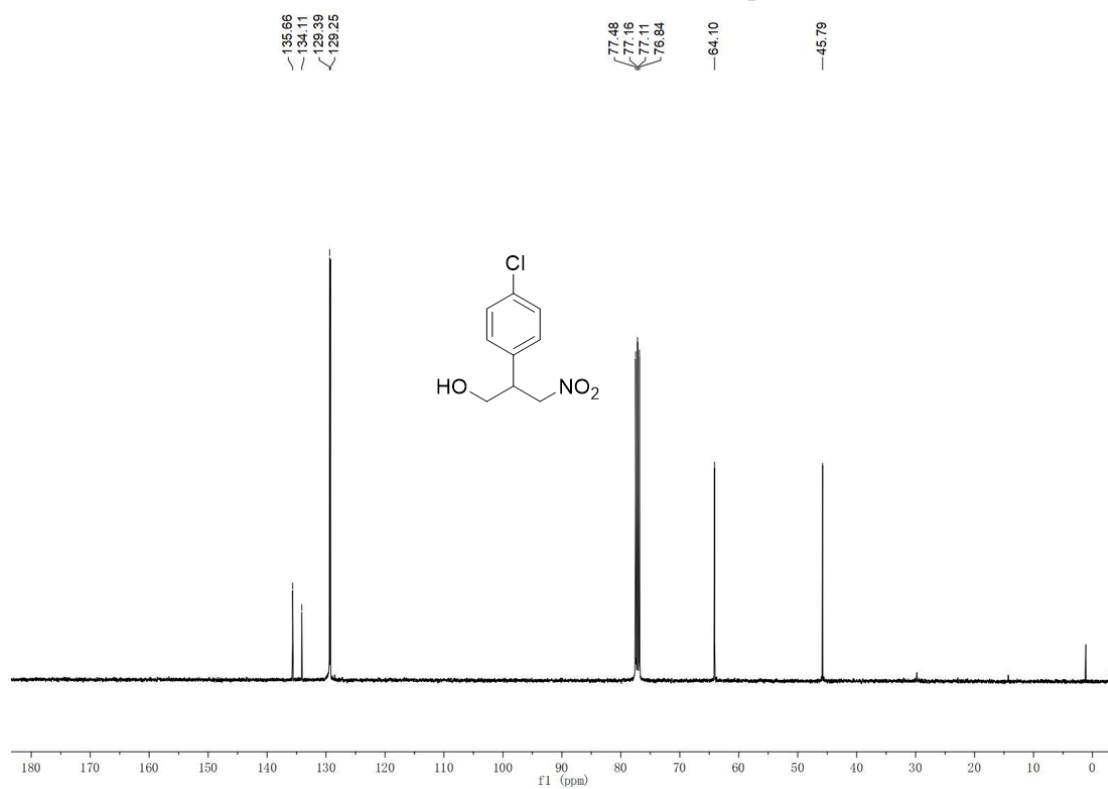
^{19}F NMR (376 MHz, CDCl_3) of **3o**



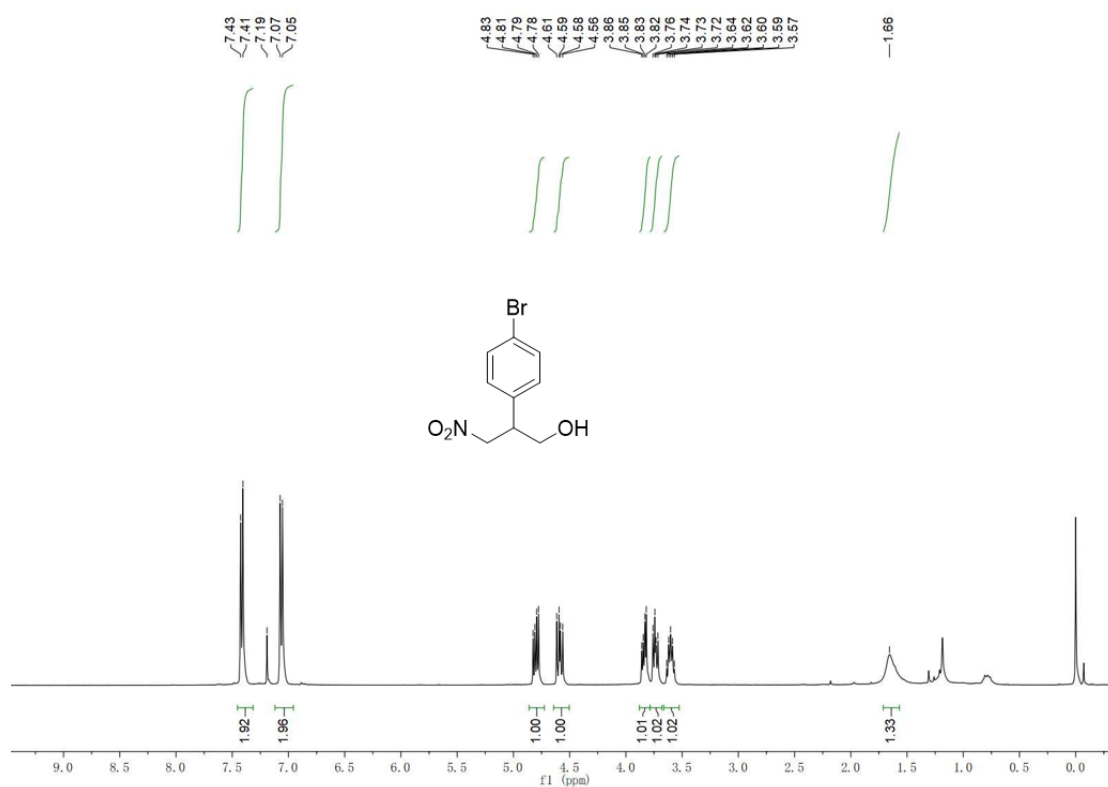
¹H NMR (400 MHz, CDCl₃) of **3p**



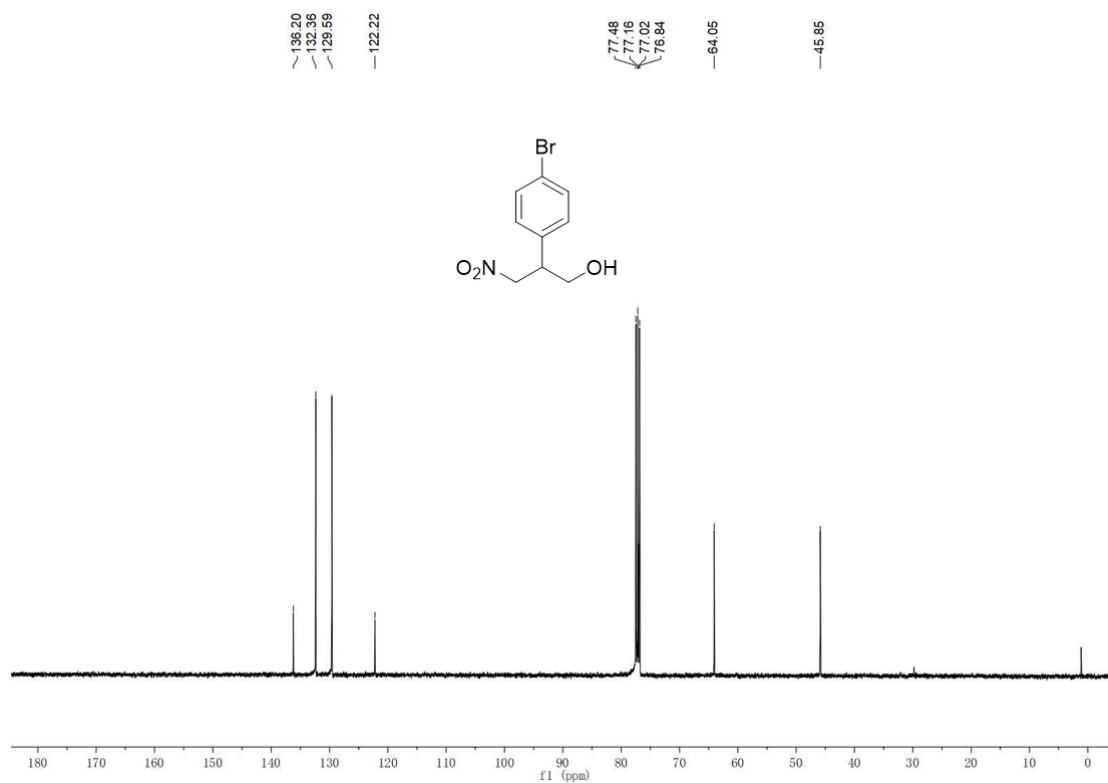
¹³C NMR (101 MHz, CDCl₃) of **3p**



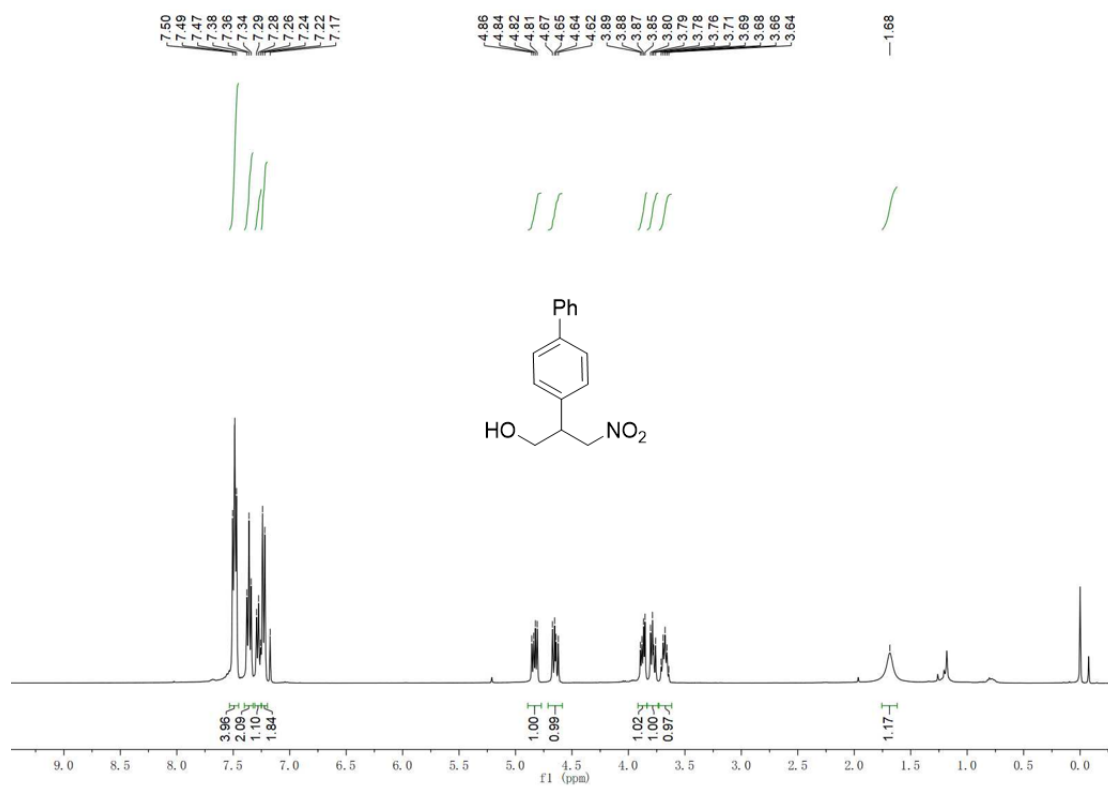
^1H NMR (400 MHz, CDCl_3) of **3q**



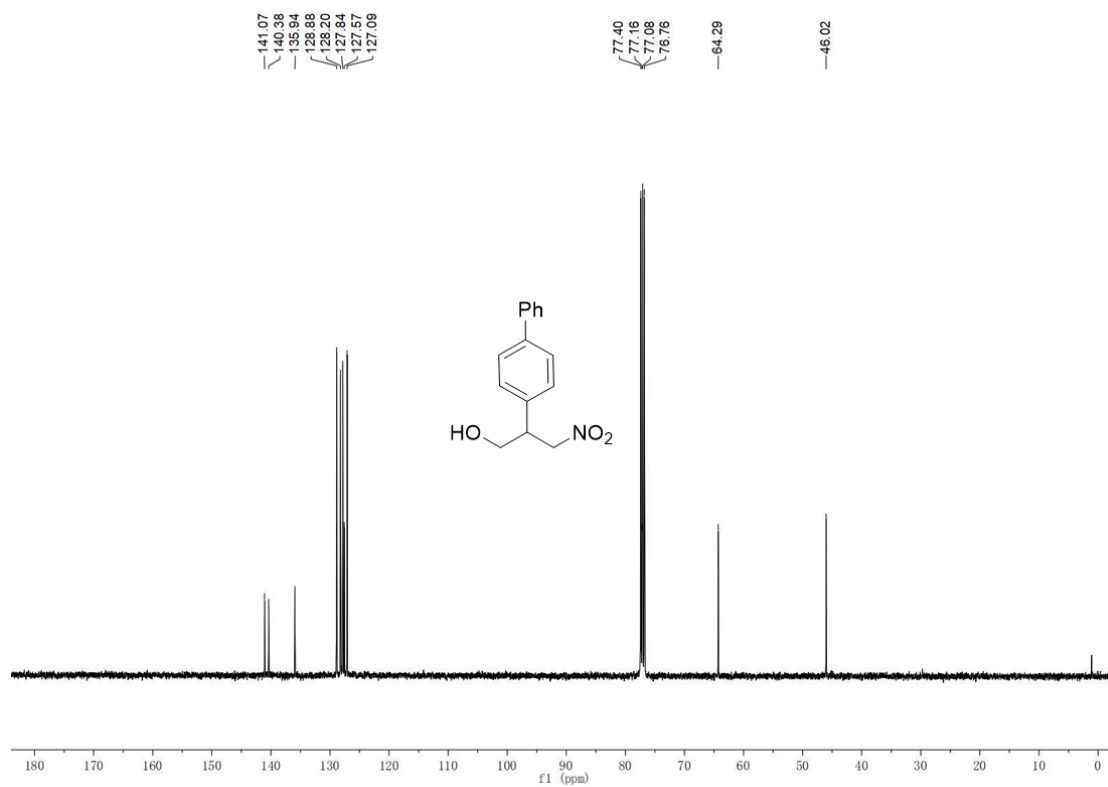
^{13}C NMR (101 MHz, CDCl_3) of **3q**



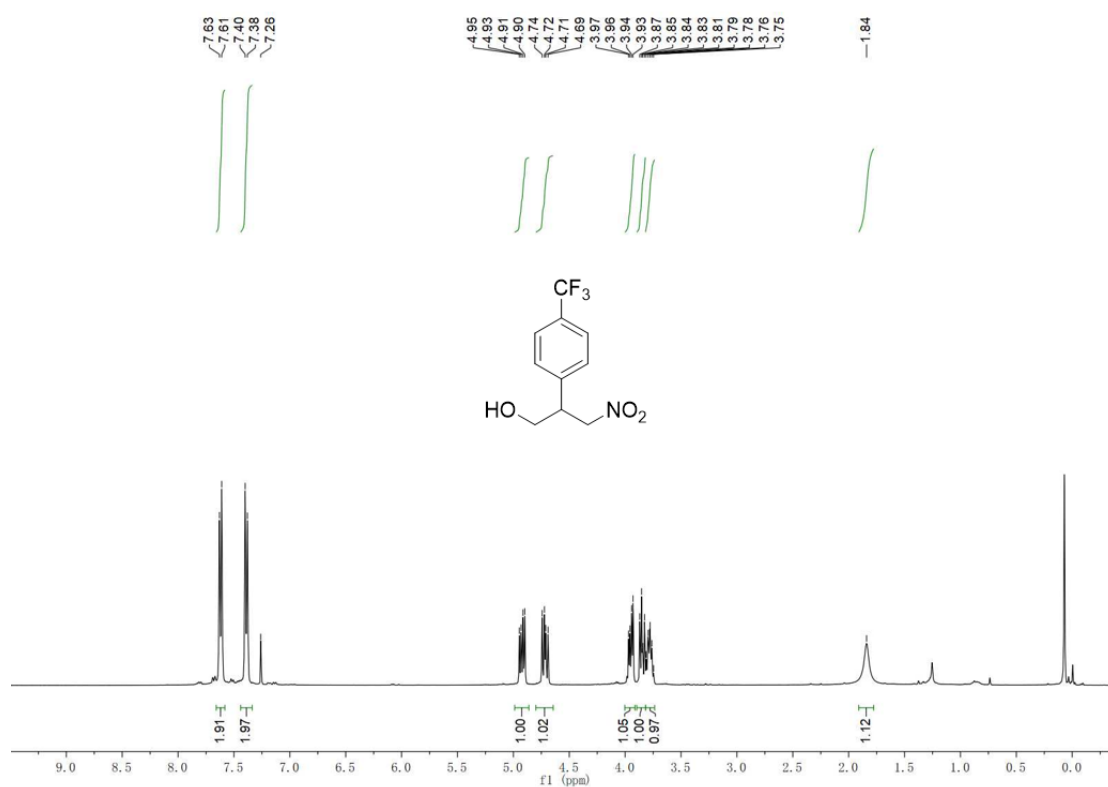
¹H NMR (400 MHz, CDCl₃) of **3r**



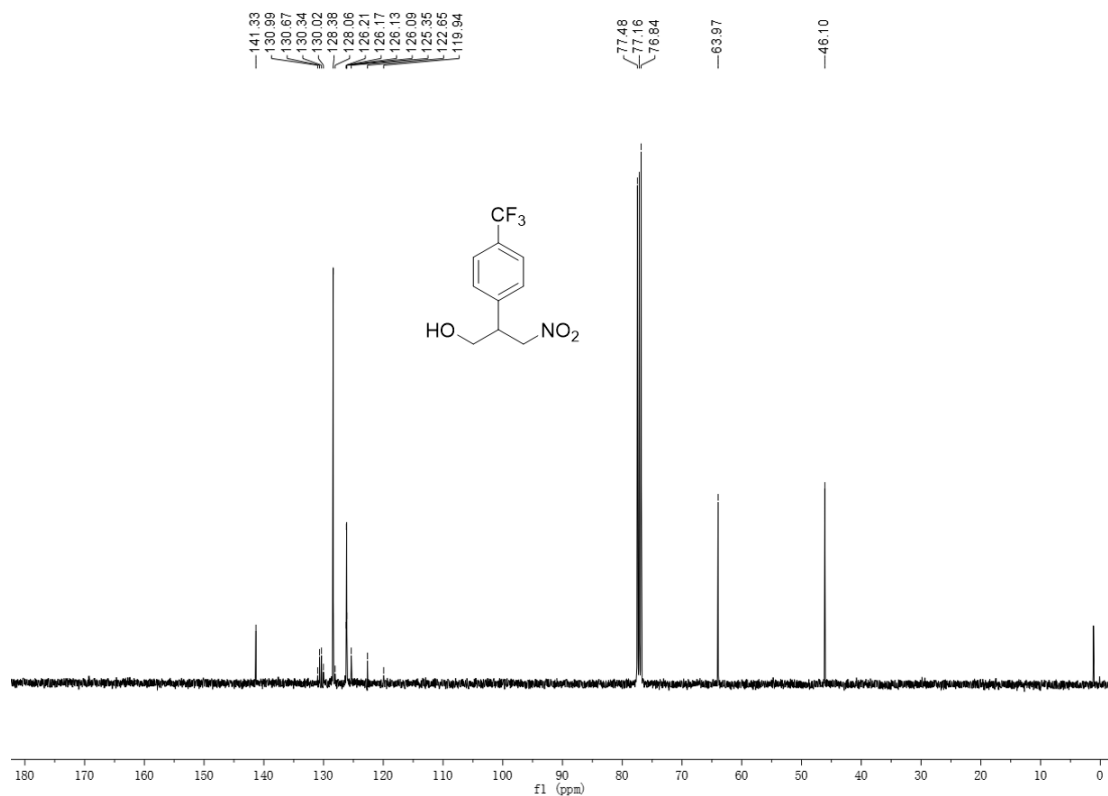
¹³C NMR (101 MHz, CDCl₃) of **3r**



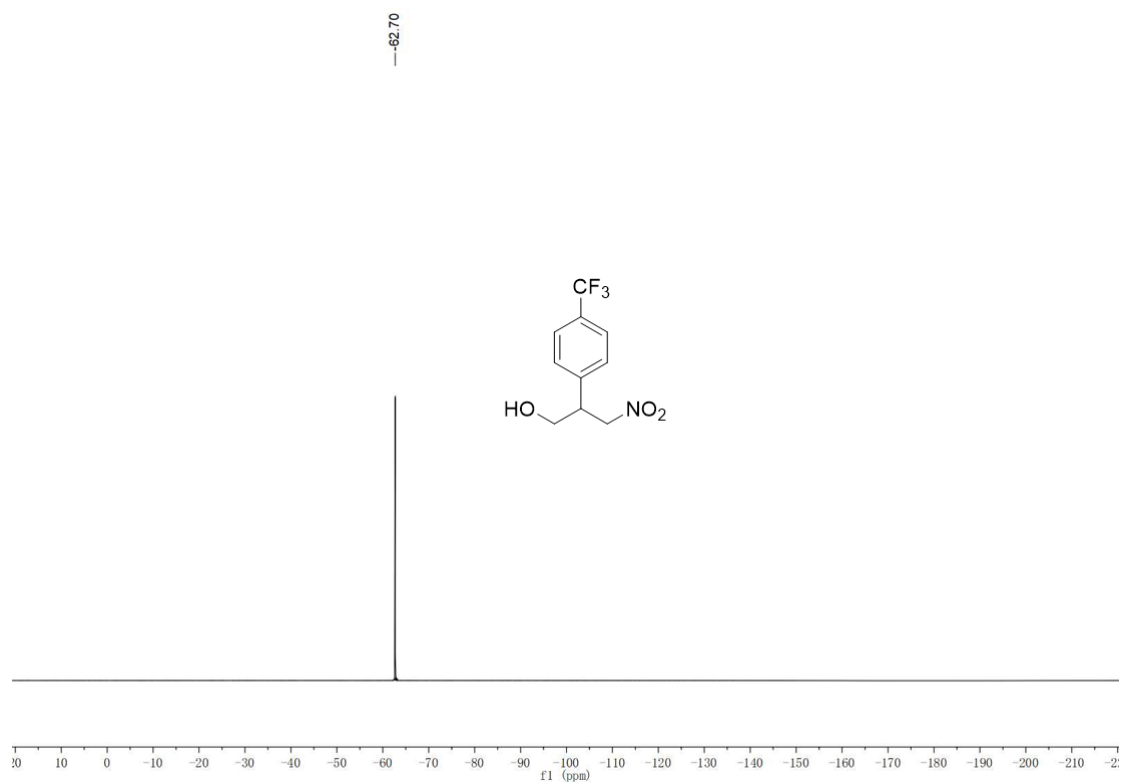
¹H NMR (400 MHz, CDCl₃) of **3s**



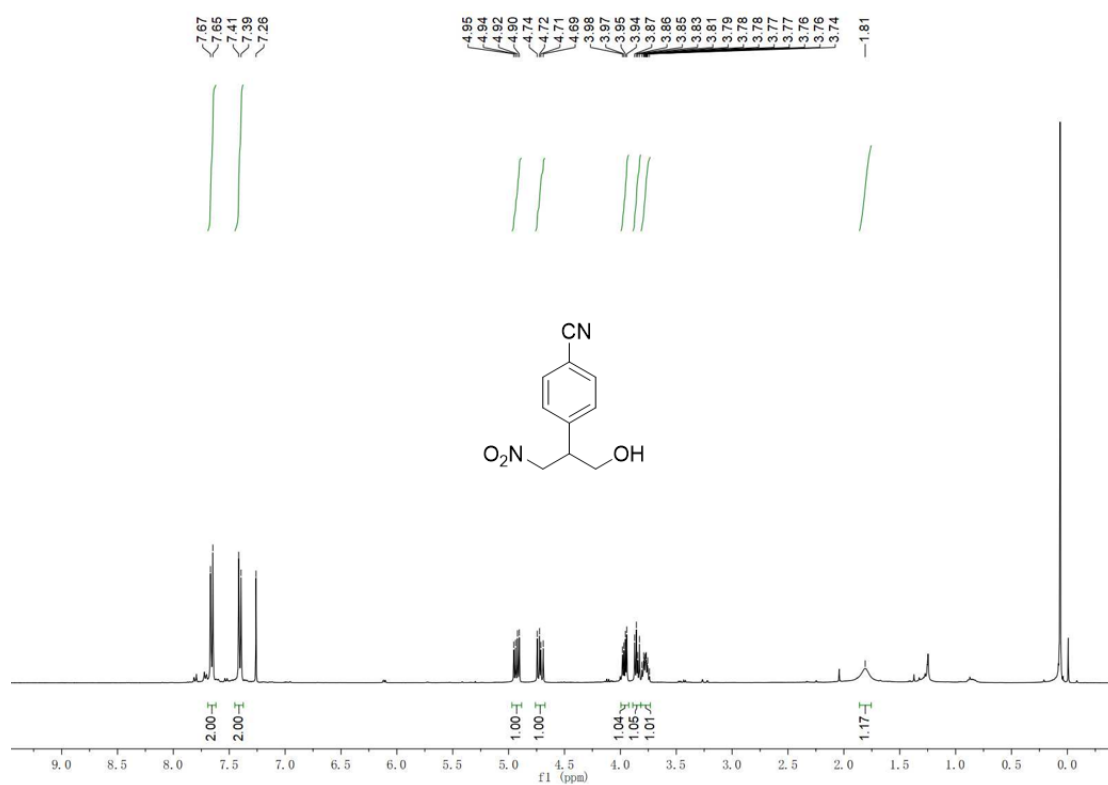
¹³C NMR (101 MHz, CDCl₃) of **3s**



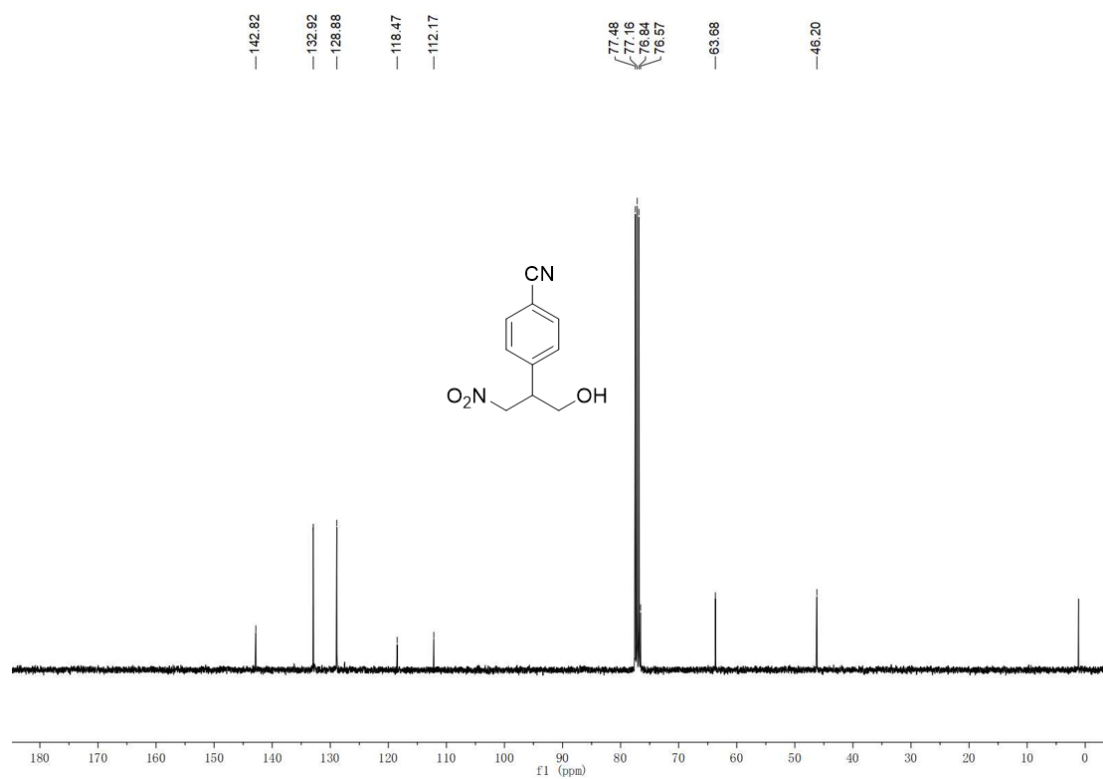
^{19}F NMR (376 MHz, CDCl_3) of **3s**



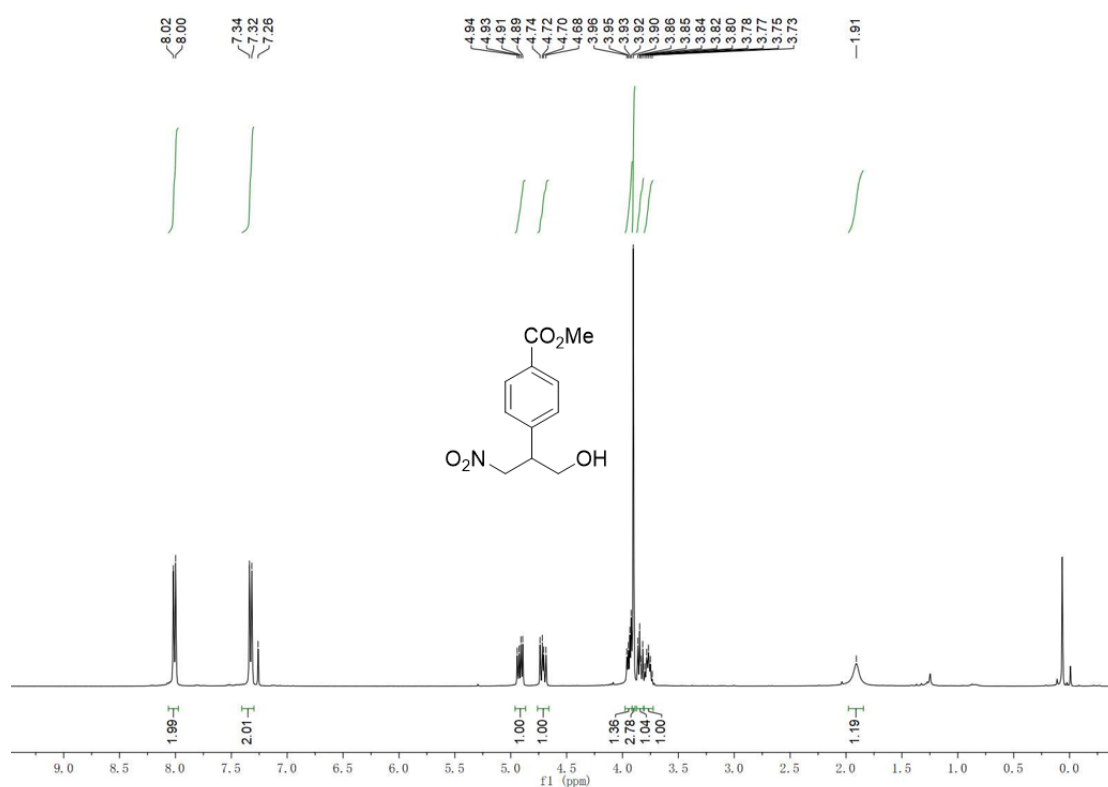
¹H NMR (400 MHz, CDCl₃) of **3t**



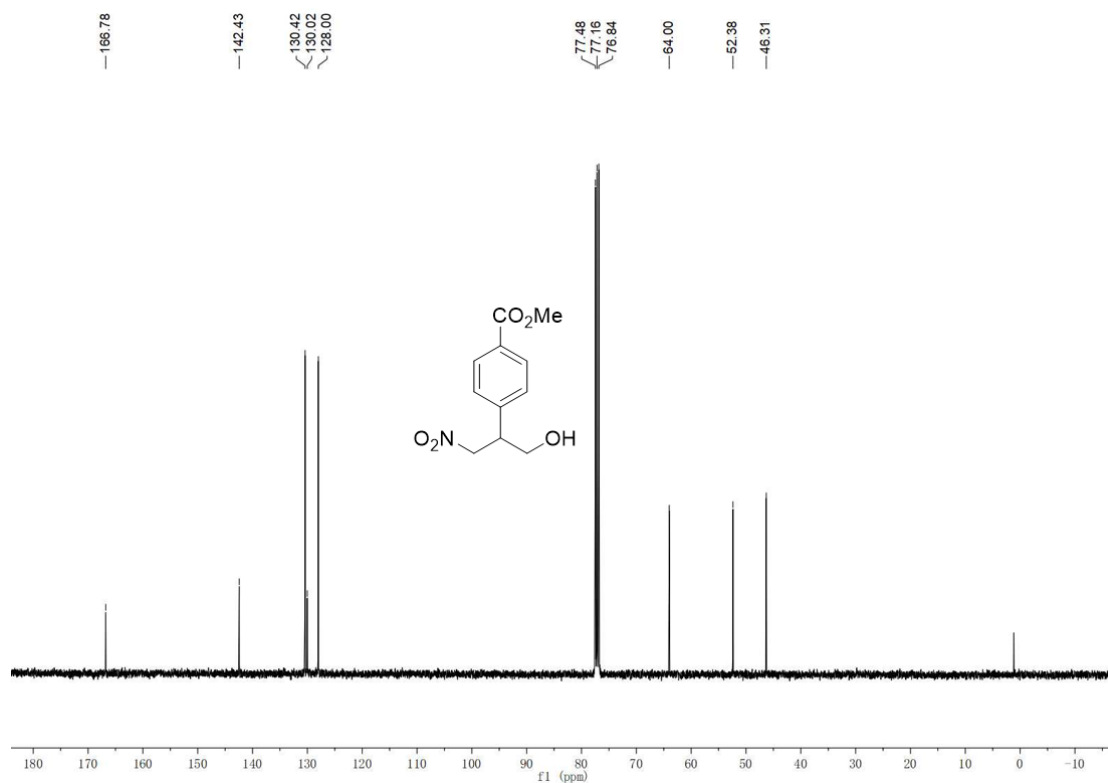
¹³C NMR (101 MHz, CDCl₃) of **3t**



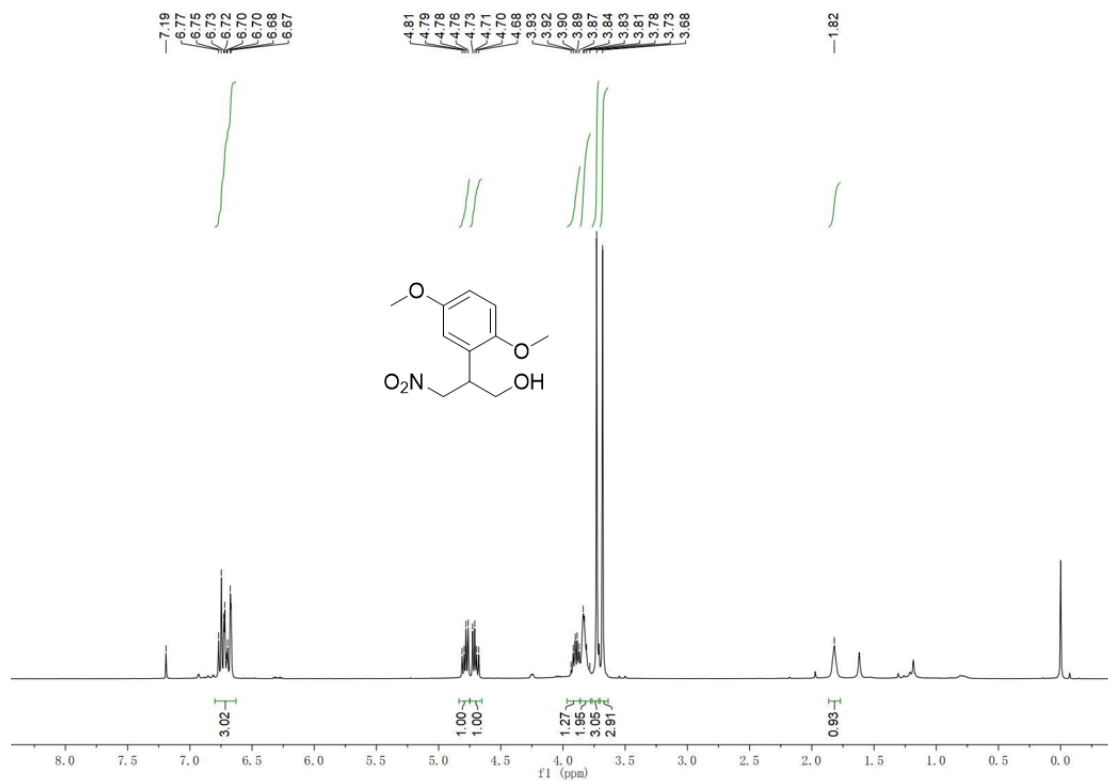
^1H NMR (400 MHz, CDCl_3) of **3u**



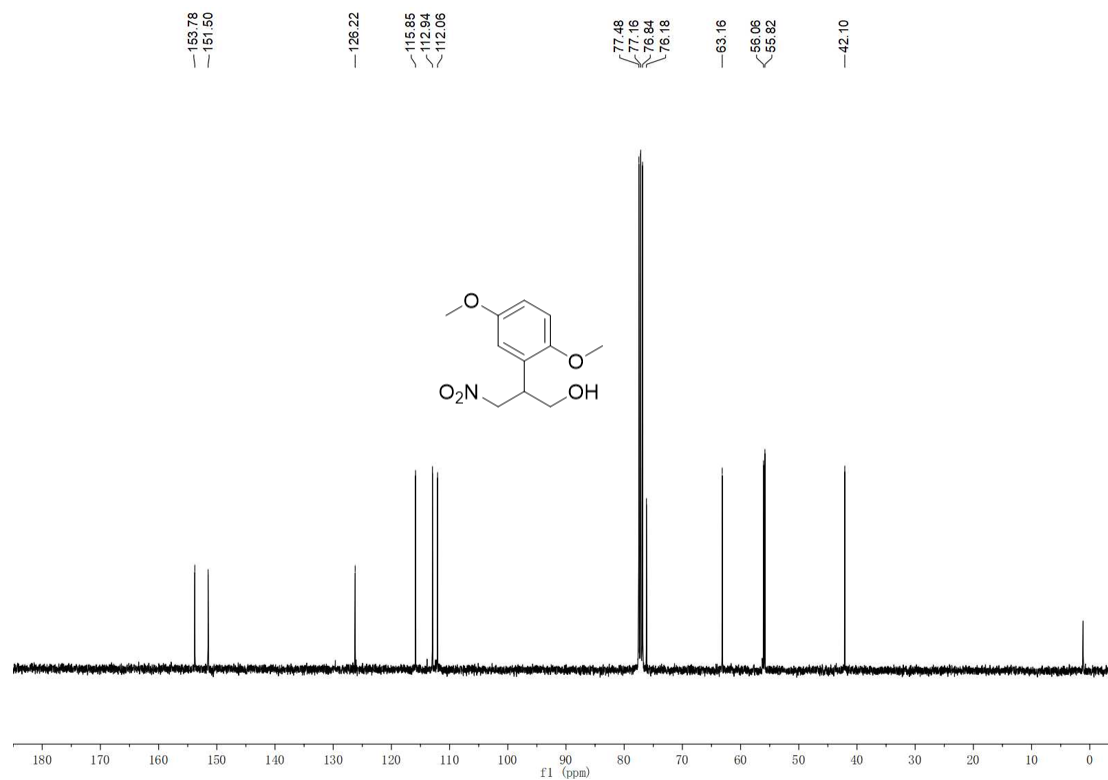
^{13}C NMR (101 MHz, CDCl_3) of **3u**



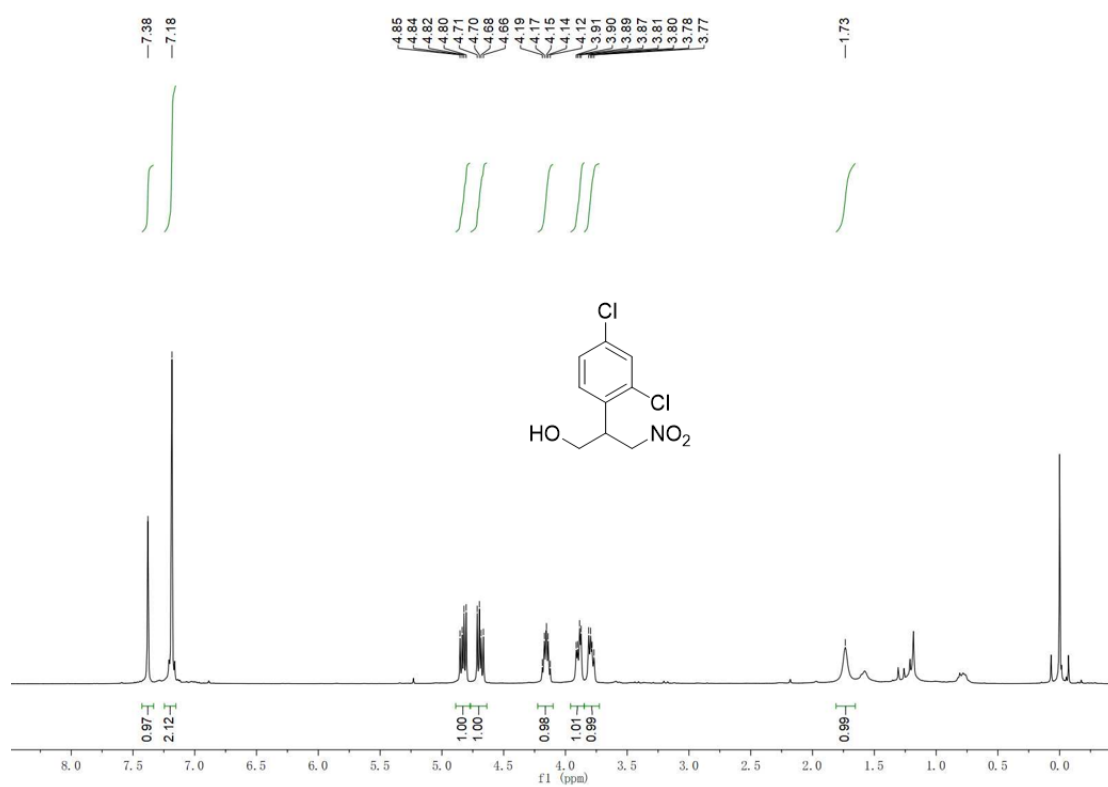
¹H NMR (400 MHz, CDCl₃) of **3v**



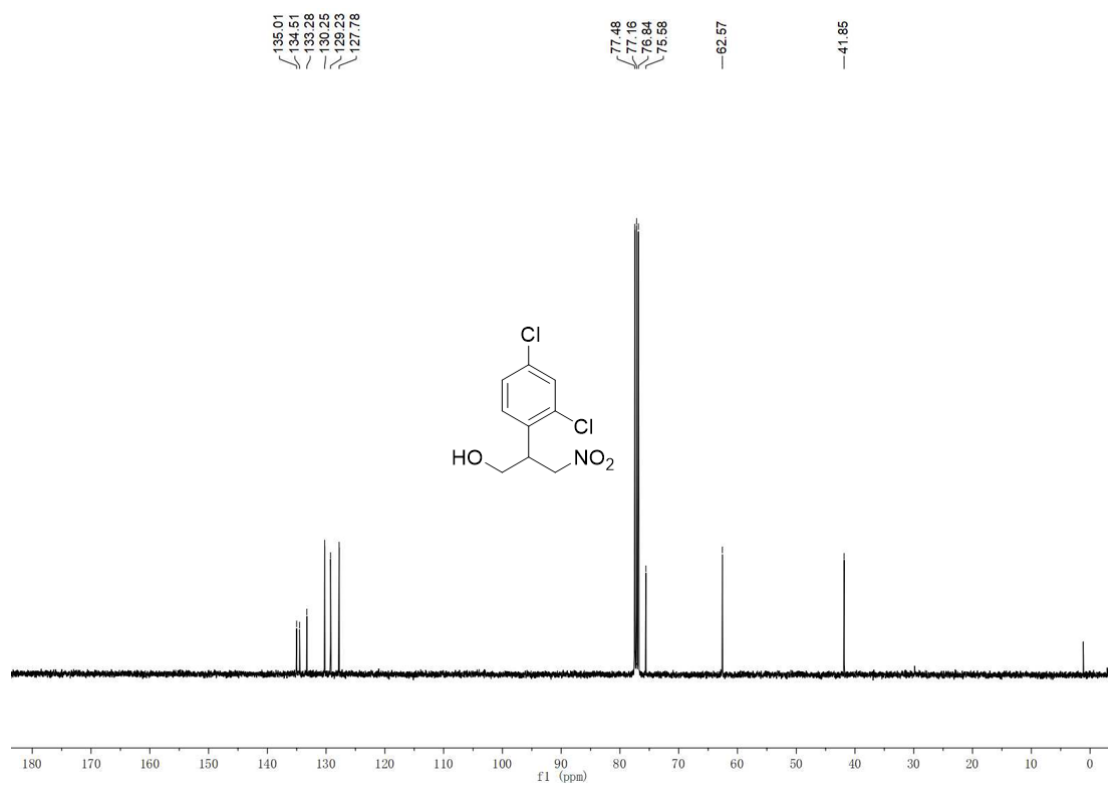
¹³C NMR (101 MHz, CDCl₃) of **3v**



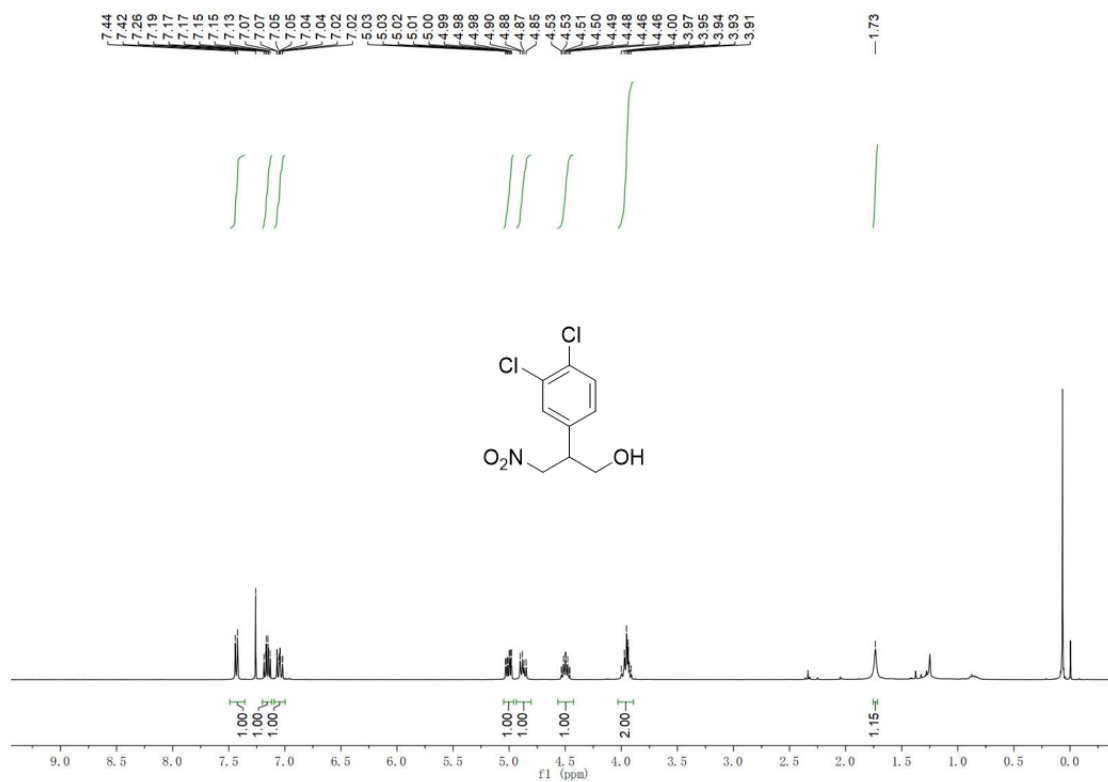
^1H NMR (400 MHz, CDCl_3) of **3w**



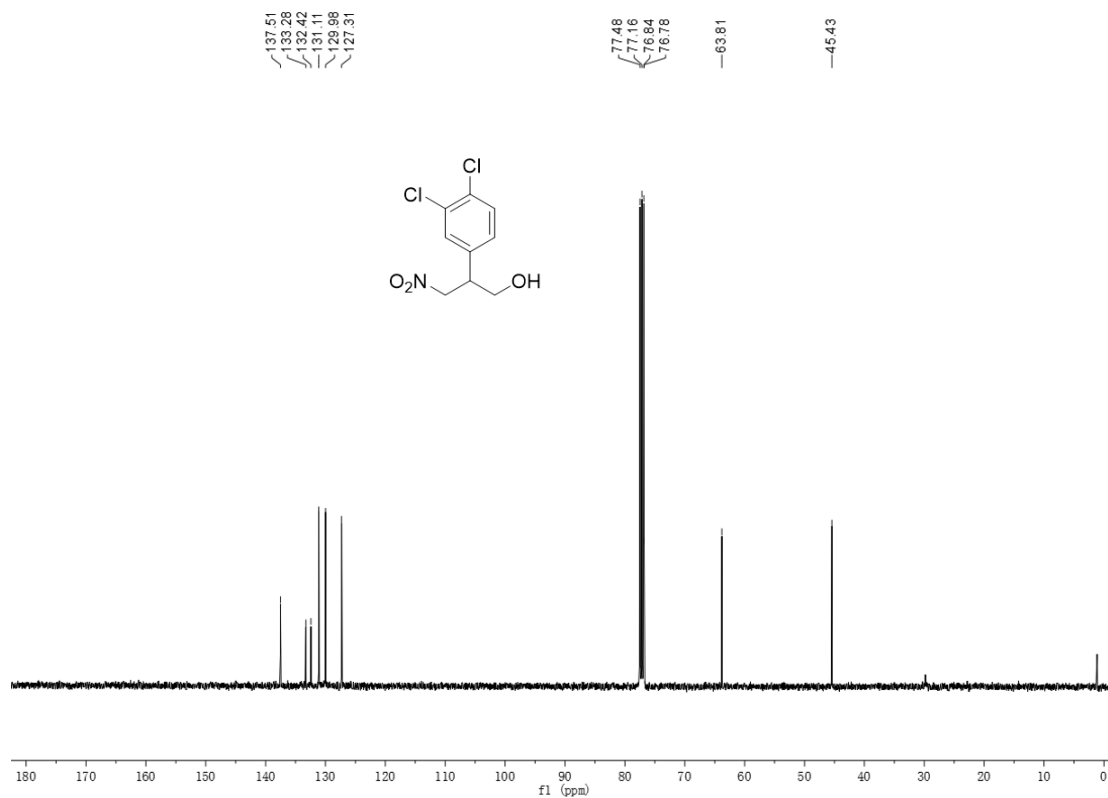
^{13}C NMR (101 MHz, CDCl_3) of **3w**



¹H NMR (400 MHz, CDCl₃) of **3x**



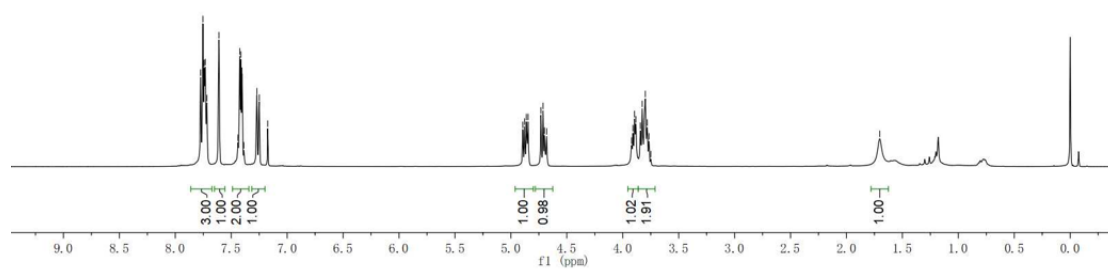
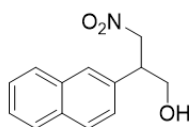
¹³C NMR (101 MHz, CDCl₃) of **3x**



7.77
7.75
7.73
7.72
7.72
7.61
7.44
7.42
7.41
7.40
7.39
7.27
7.25
7.17

4.89
4.88
4.86
4.85
4.83
4.77
4.70
4.68
3.92
3.91
3.89
3.88
3.84
3.83
3.80
3.78
3.77
3.75

—1.70

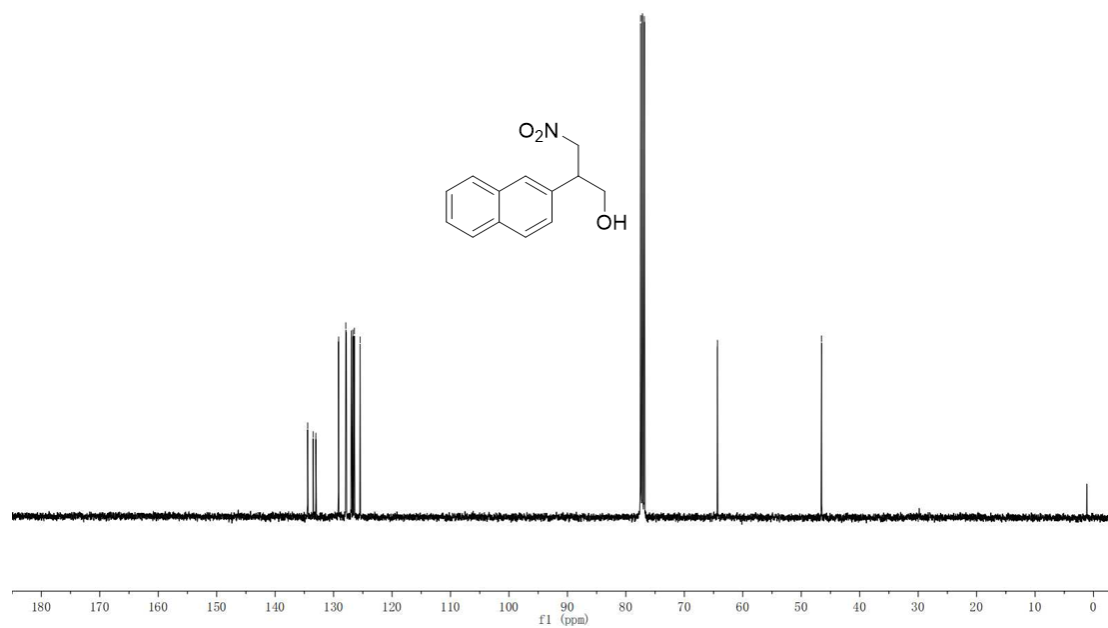


134.43
133.49
133.03
129.14
127.91
127.82
126.98
126.70
126.46
125.46

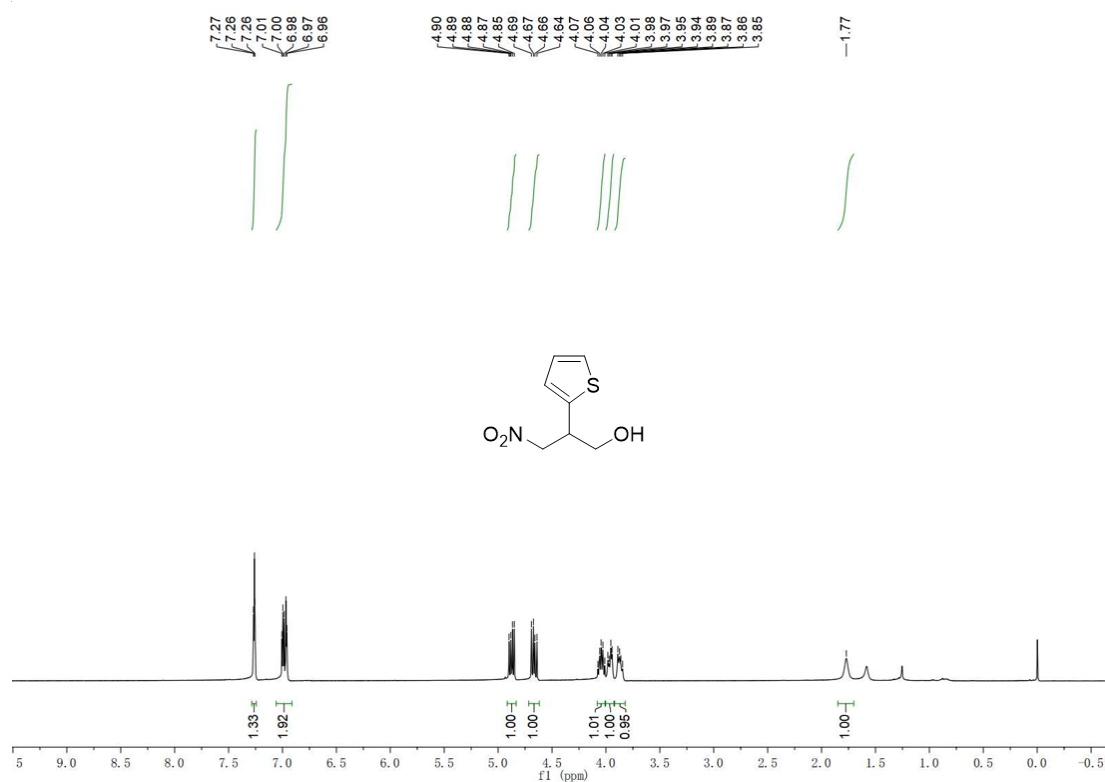
77.48
77.23
77.16
76.84

— 64.33

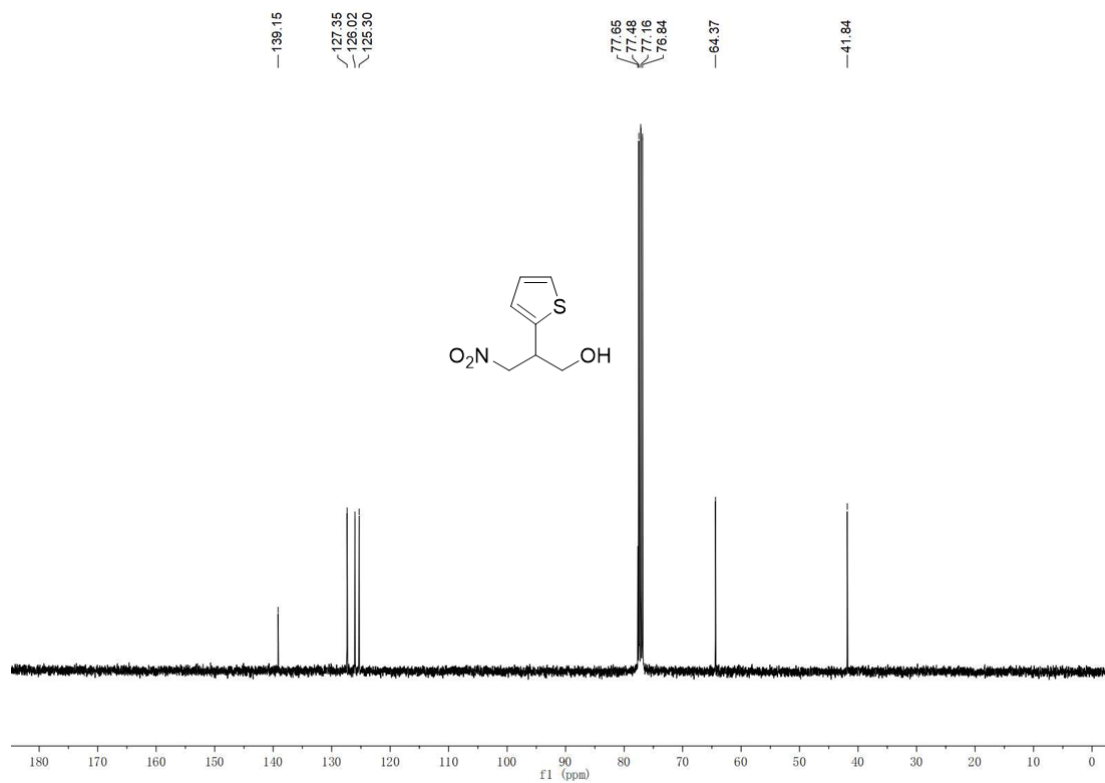
46.53



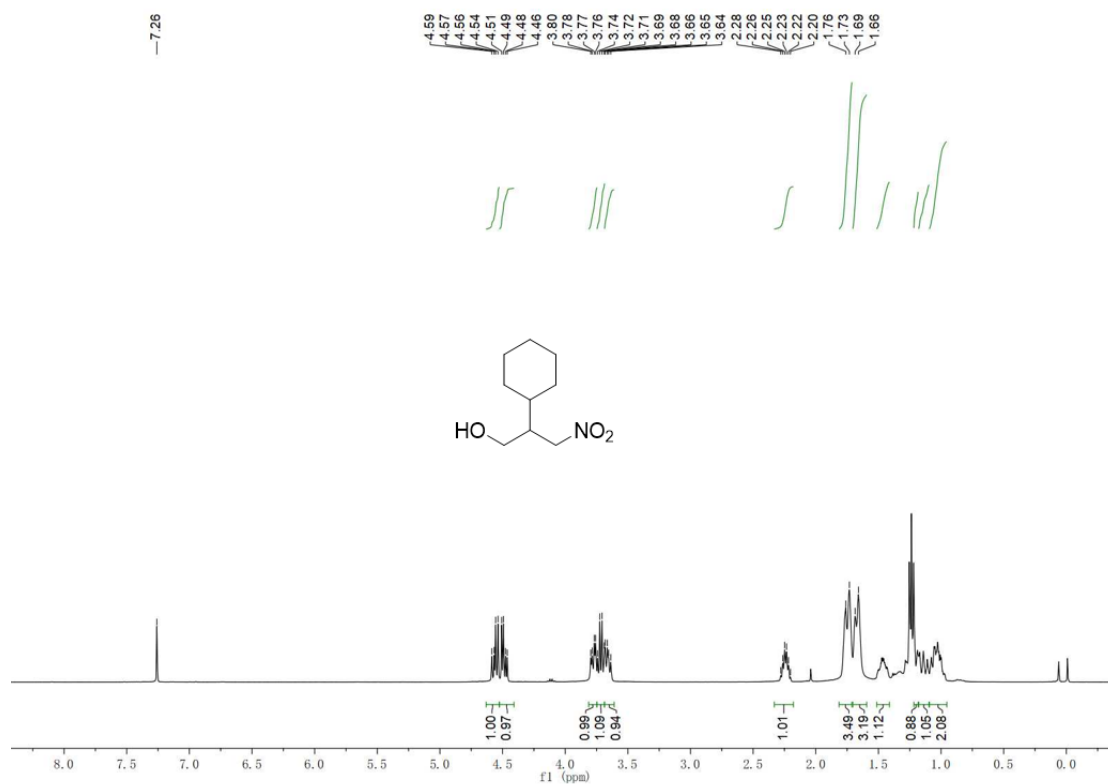
¹H NMR (400 MHz, CDCl₃) of **3z**



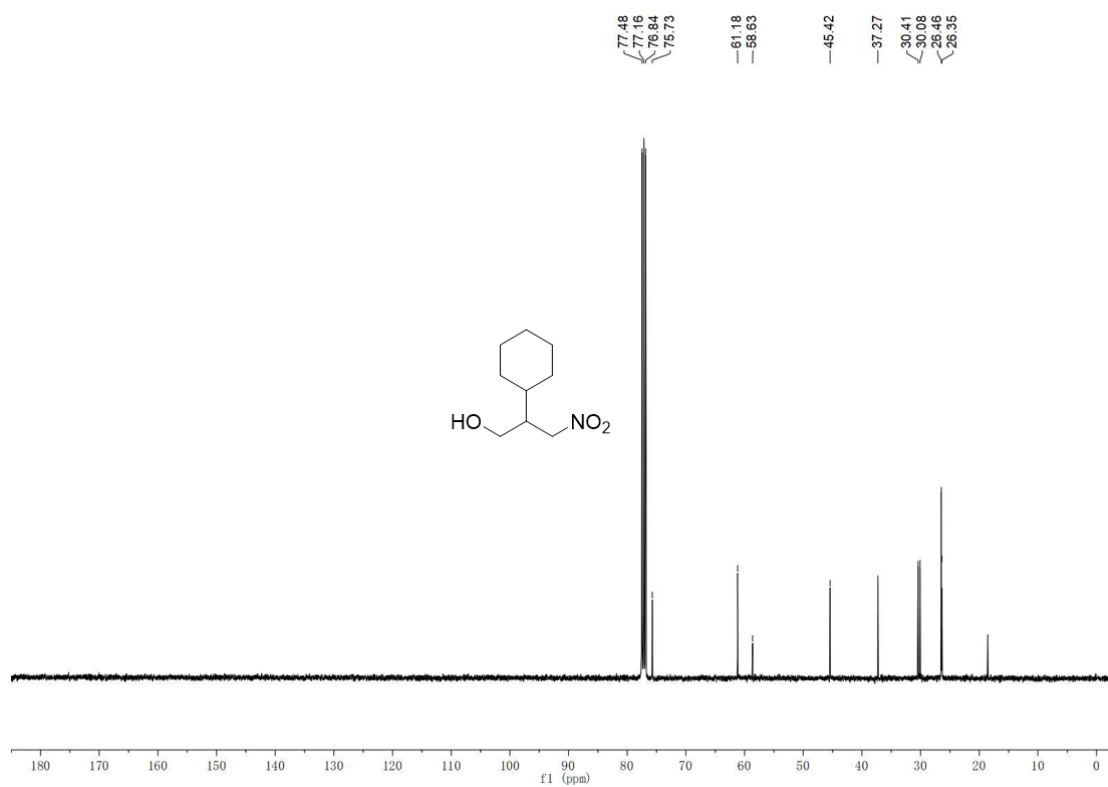
¹³C NMR (101 MHz, CDCl₃) of **3z**



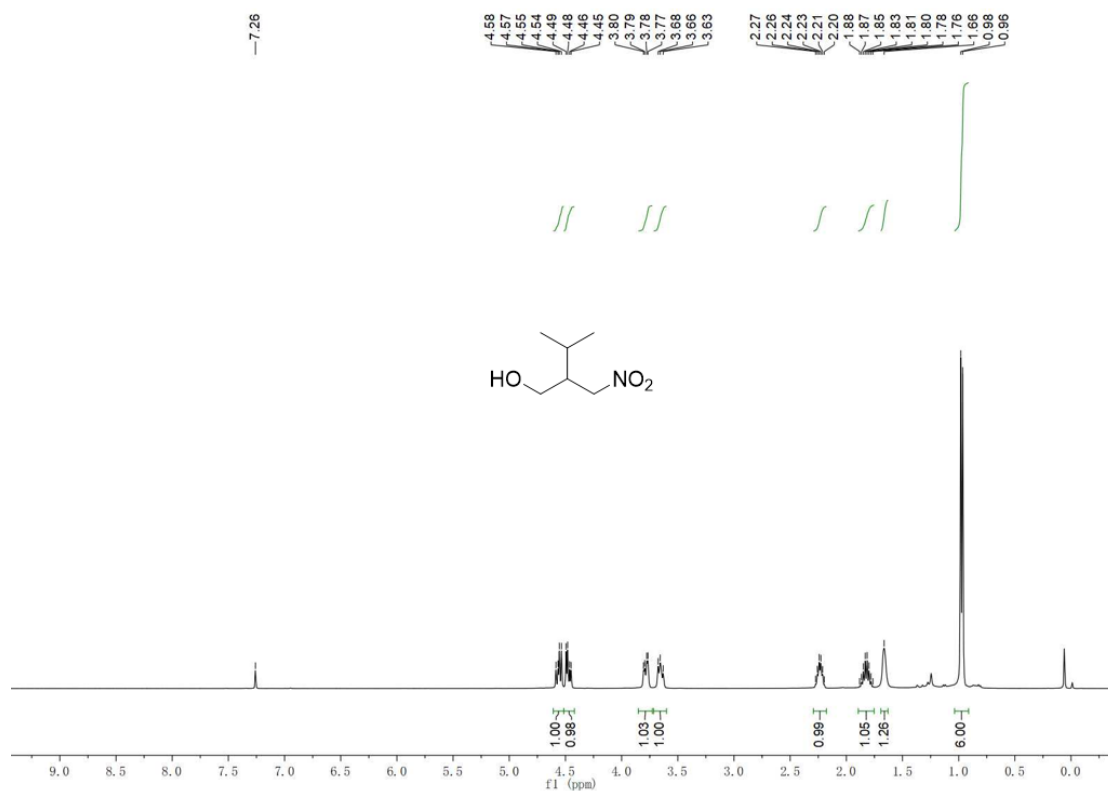
¹H NMR (400 MHz, CDCl₃) of **3aa**



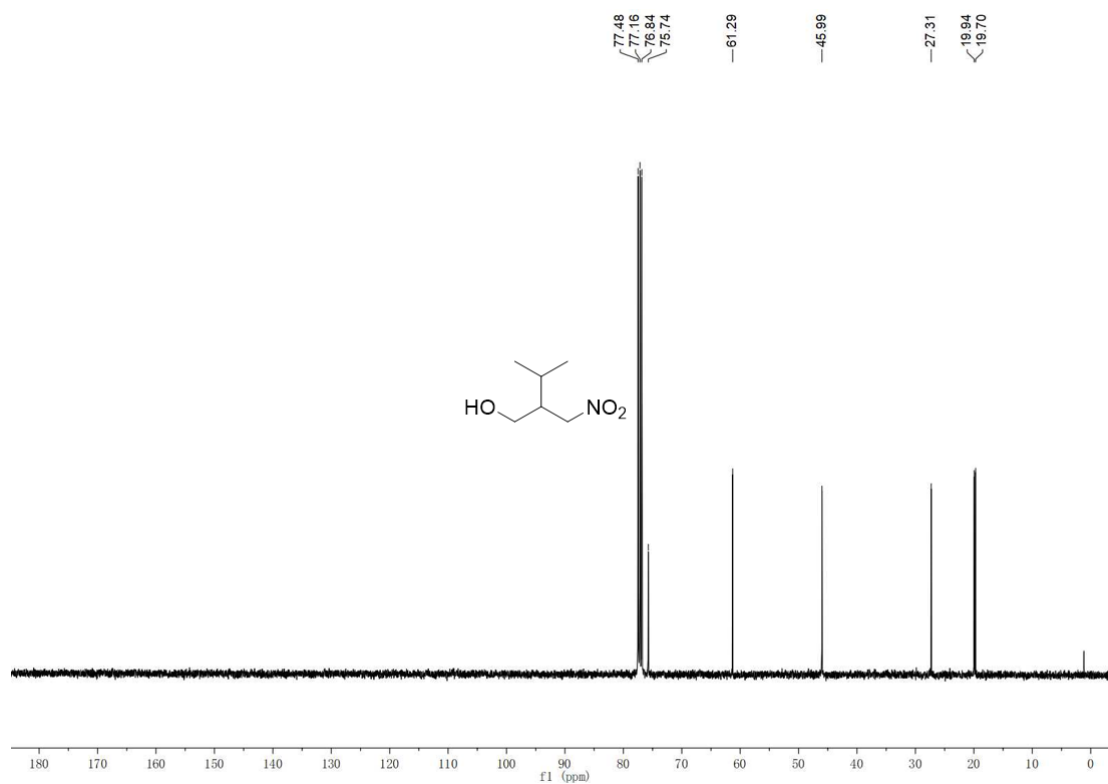
¹³C NMR (101 MHz, CDCl₃) of **3aa**



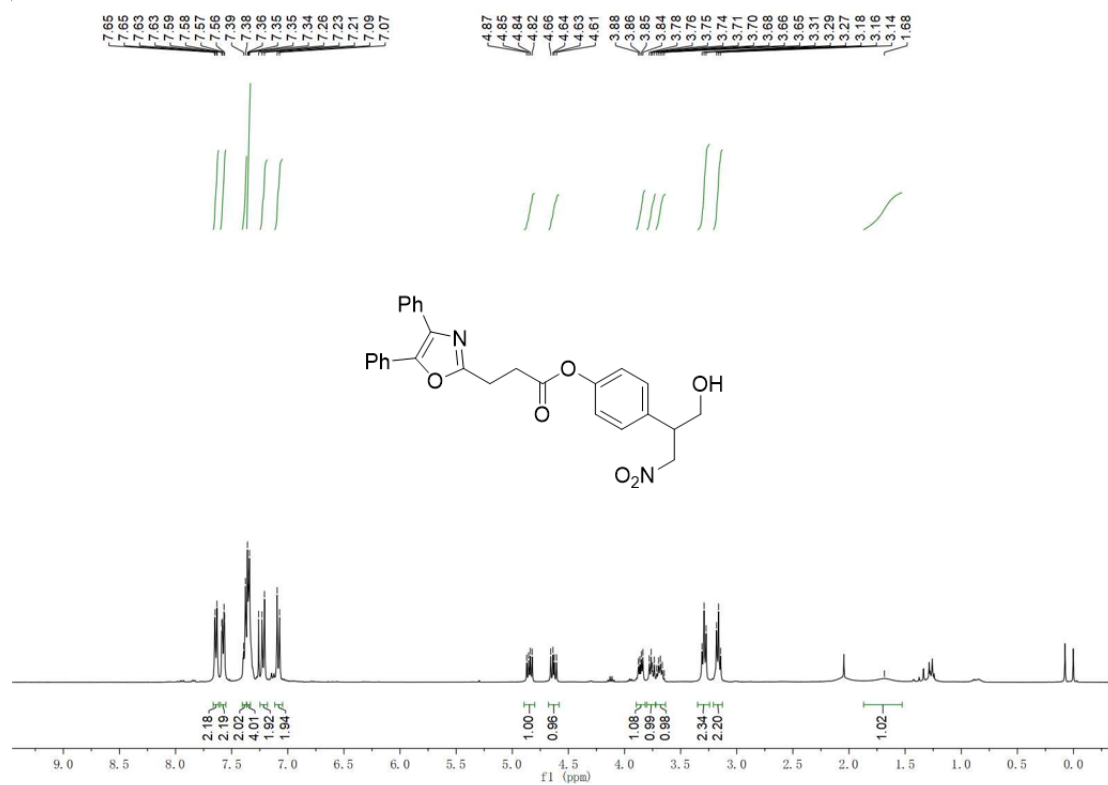
¹H NMR (400 MHz, CDCl₃) of **3ab**



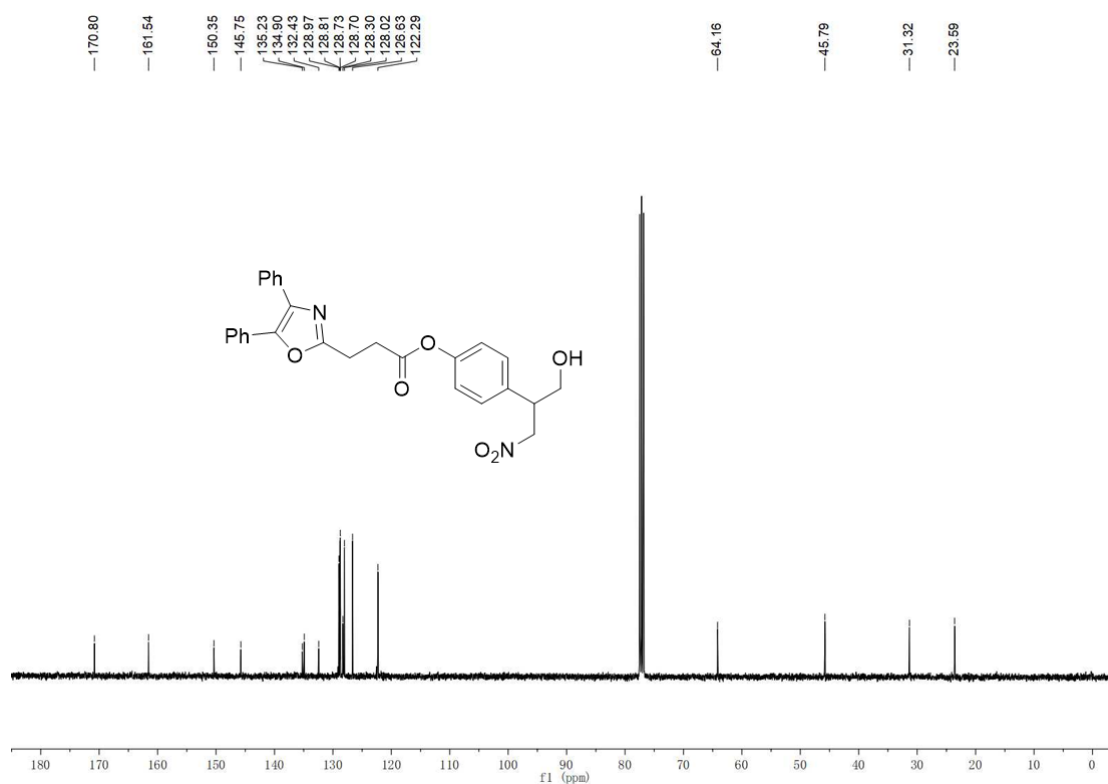
¹³C NMR (101 MHz, CDCl₃) of **3ab**



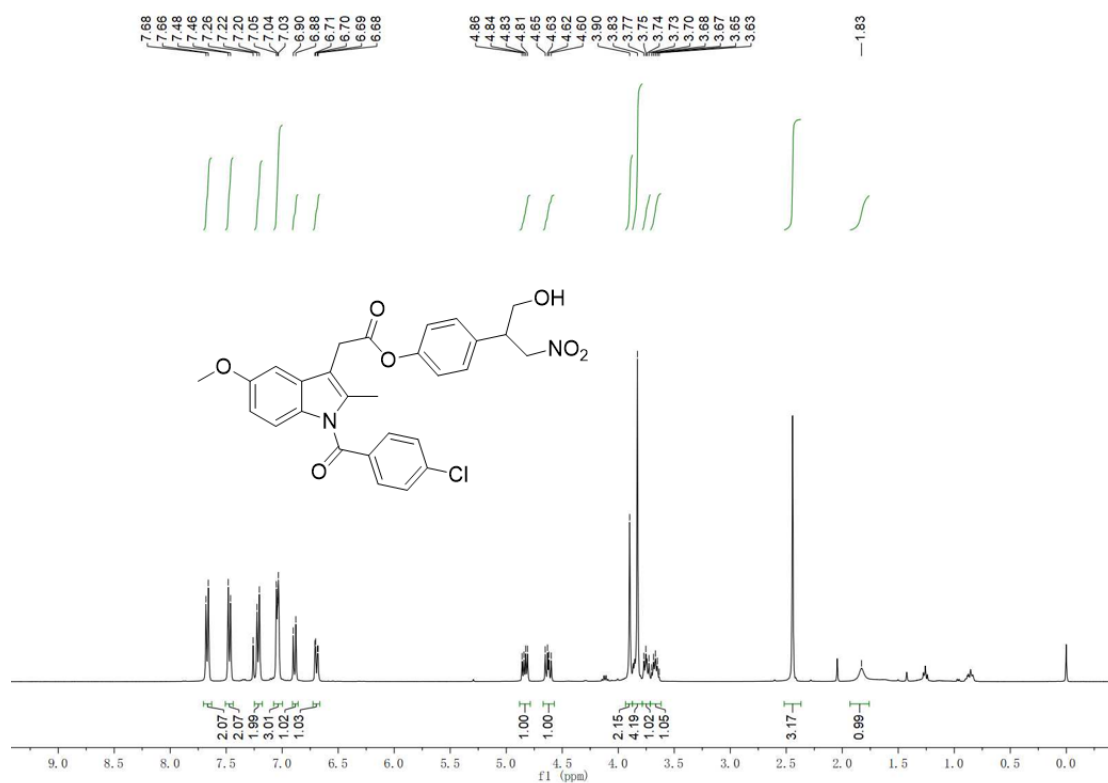
¹H NMR (400 MHz, CDCl₃) of **3ac**



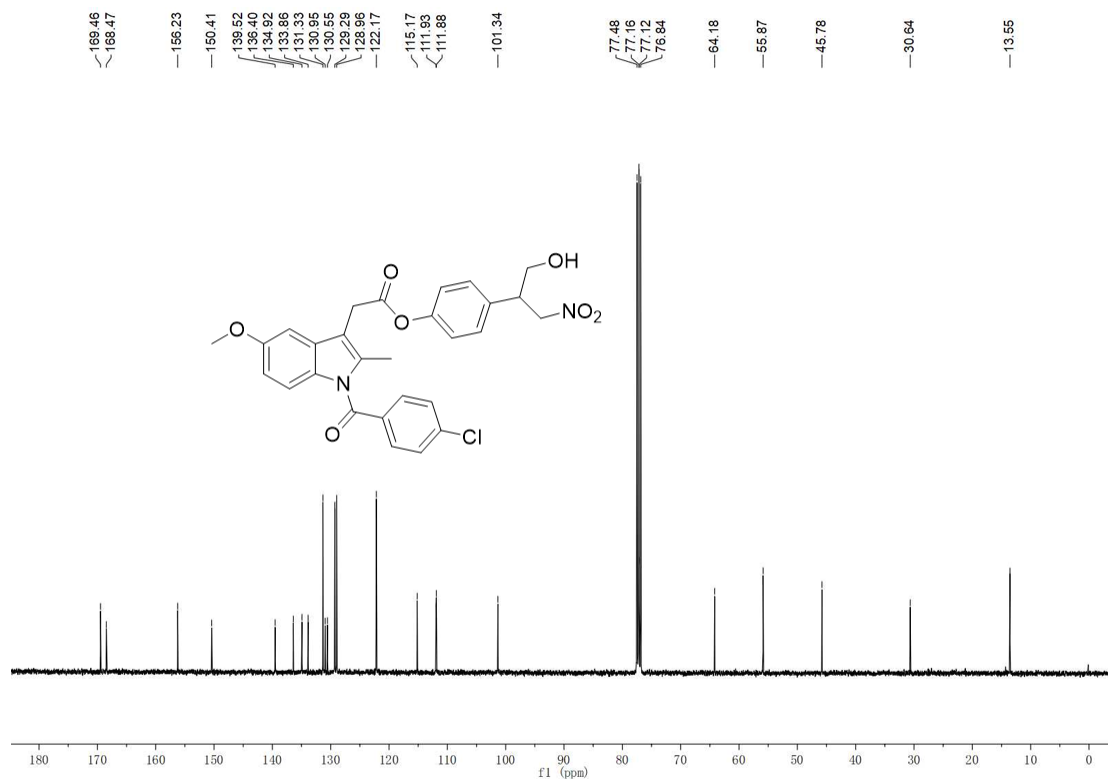
¹³C NMR (101 MHz, CDCl₃) of **3ac**



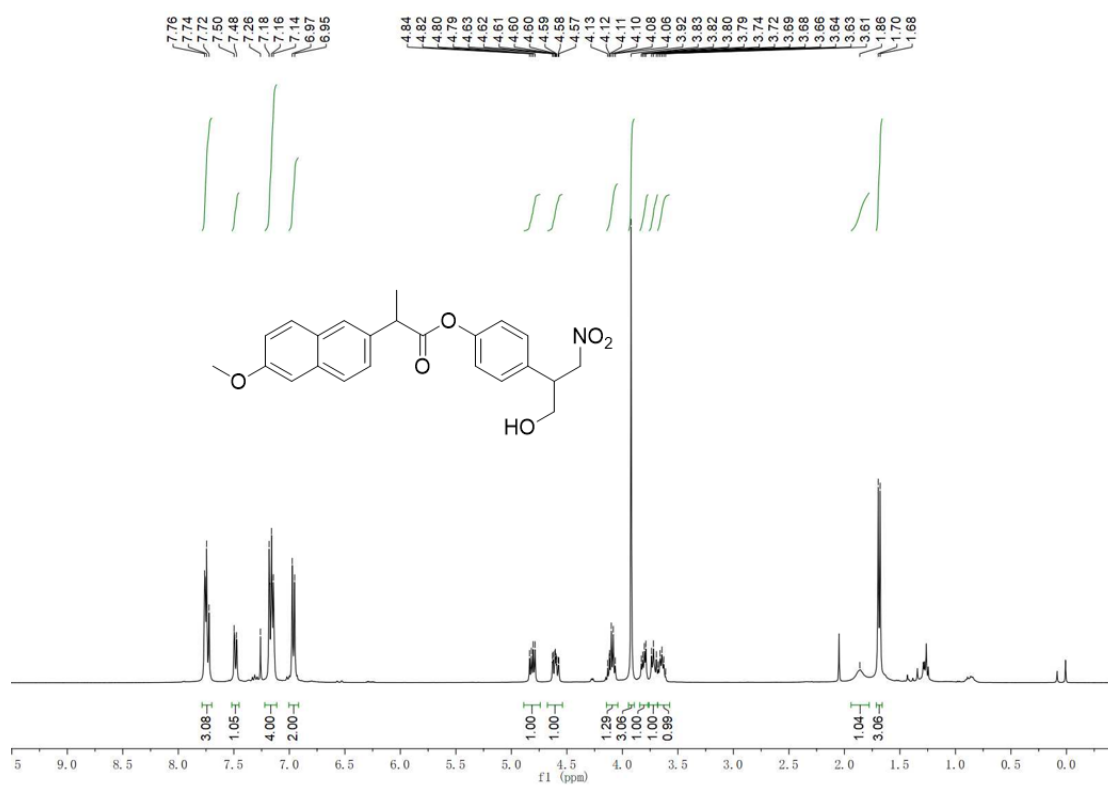
¹H NMR (400 MHz, CDCl₃) of **3ad**



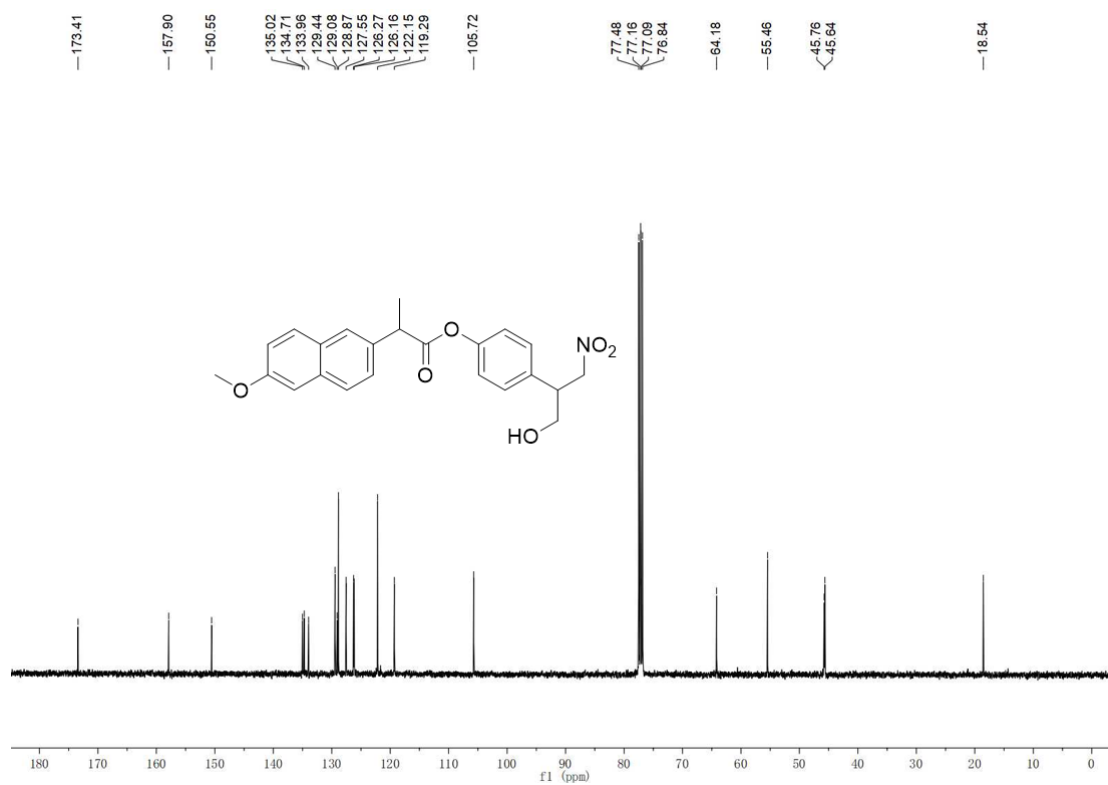
¹³C NMR (101 MHz, CDCl₃) of **3ad**



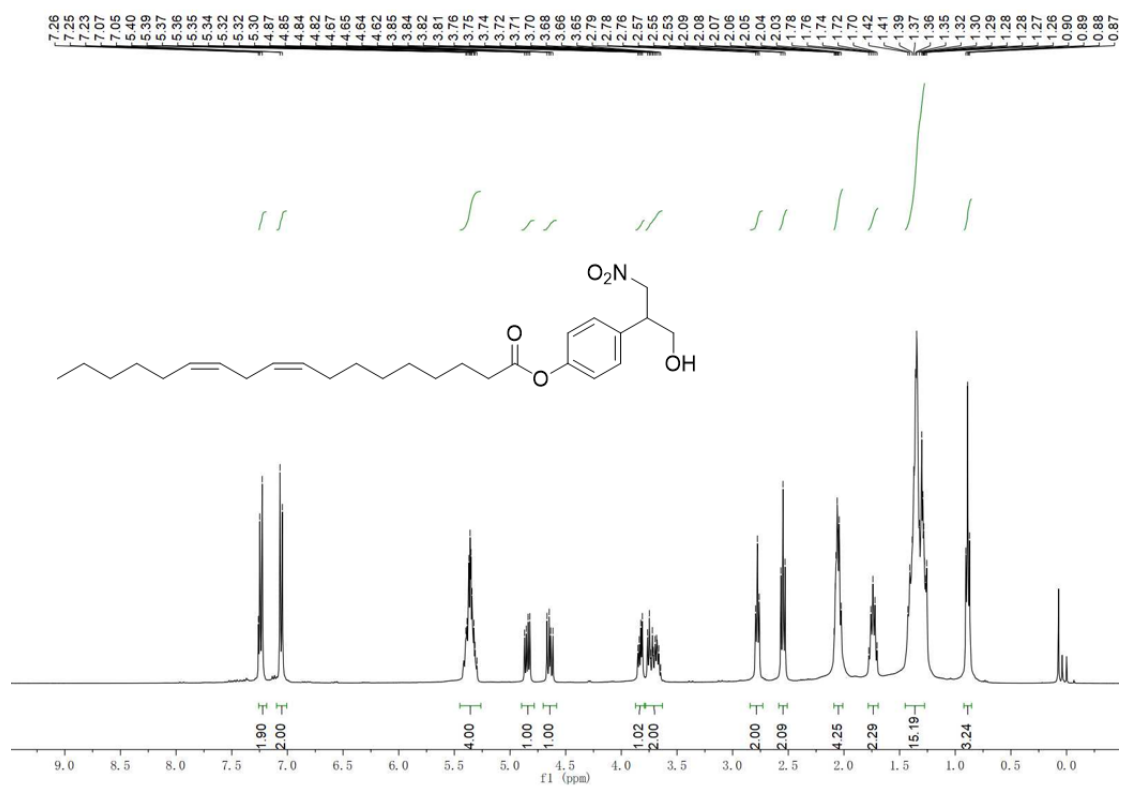
¹H NMR (400 MHz, CDCl₃) of **3ae**



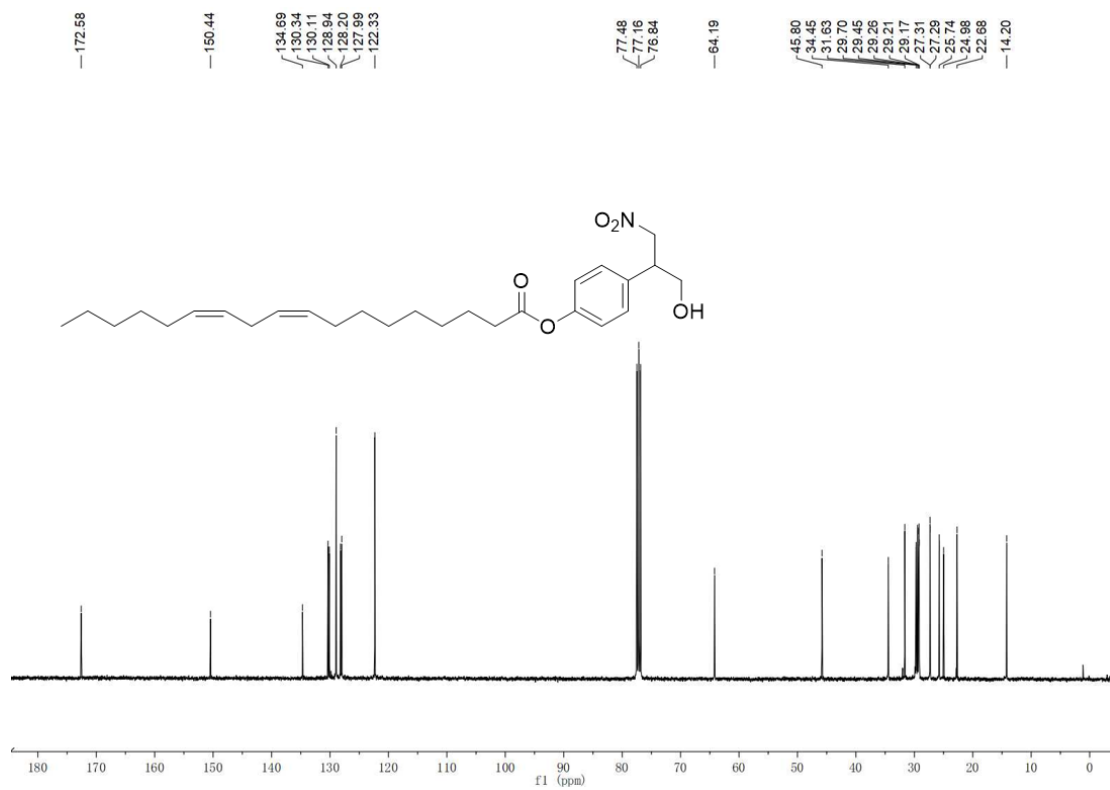
¹³C NMR (101 MHz, CDCl₃) of **3ae**



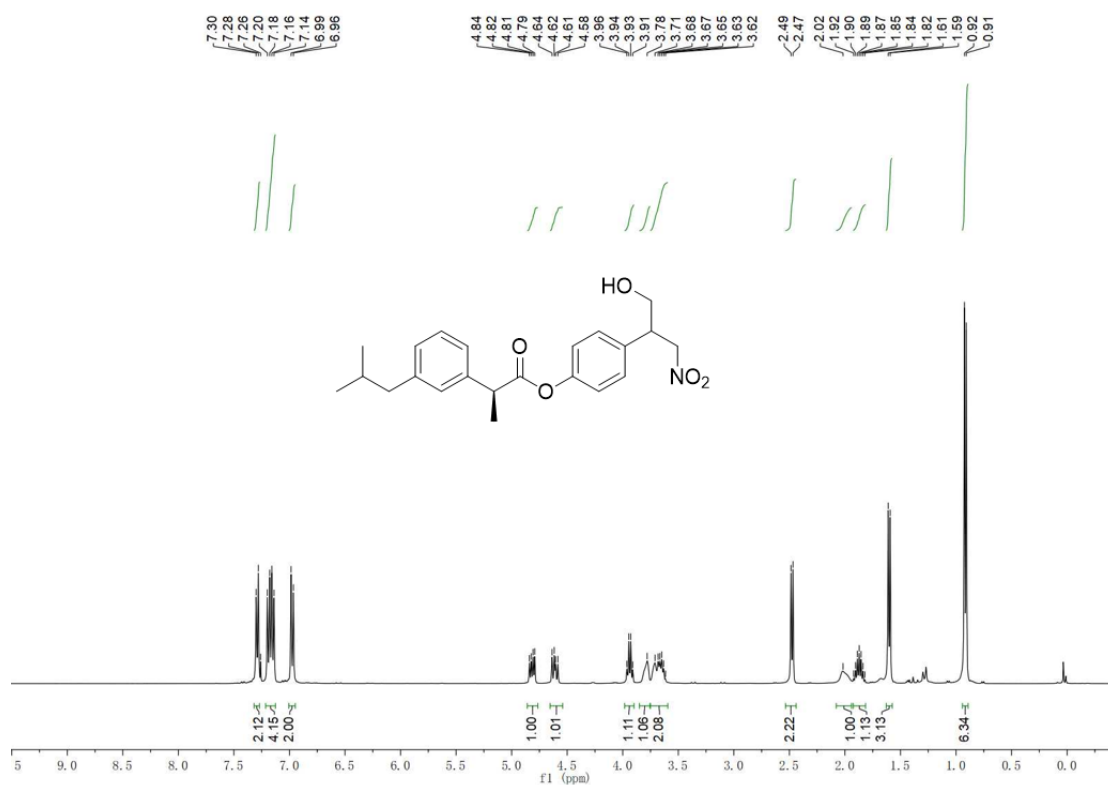
¹H NMR (400 MHz, CDCl₃) of **3af**



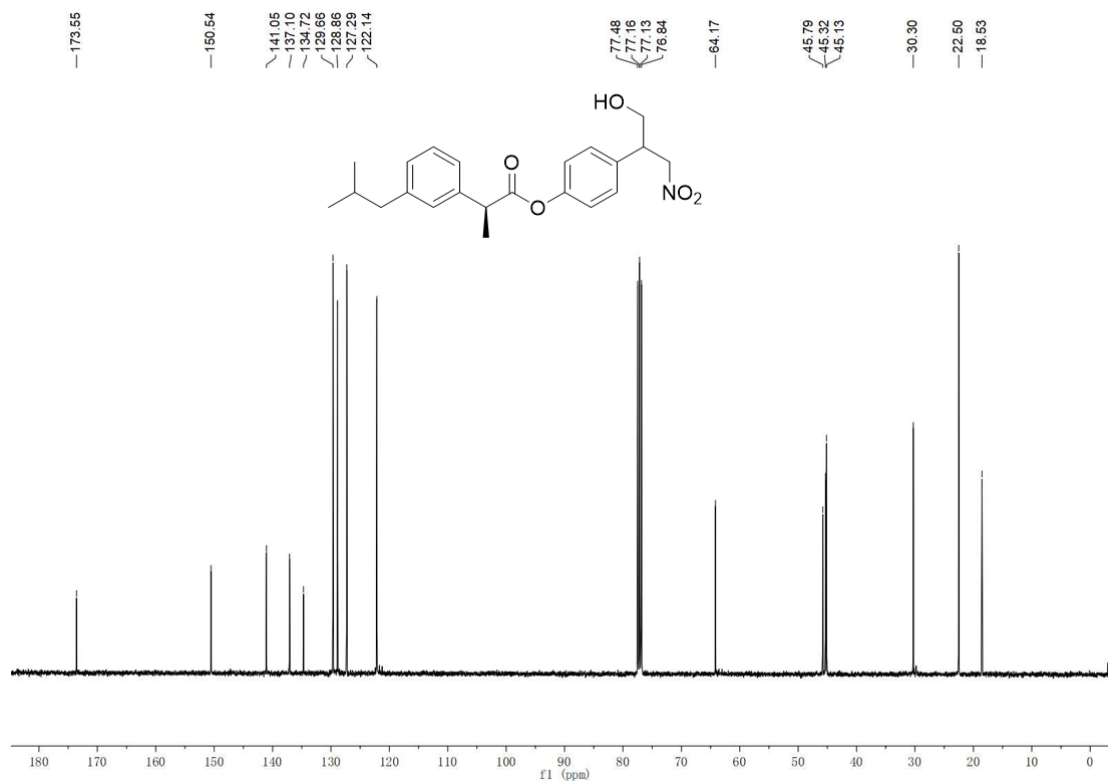
¹³C NMR (101 MHz, CDCl₃) of **3af**



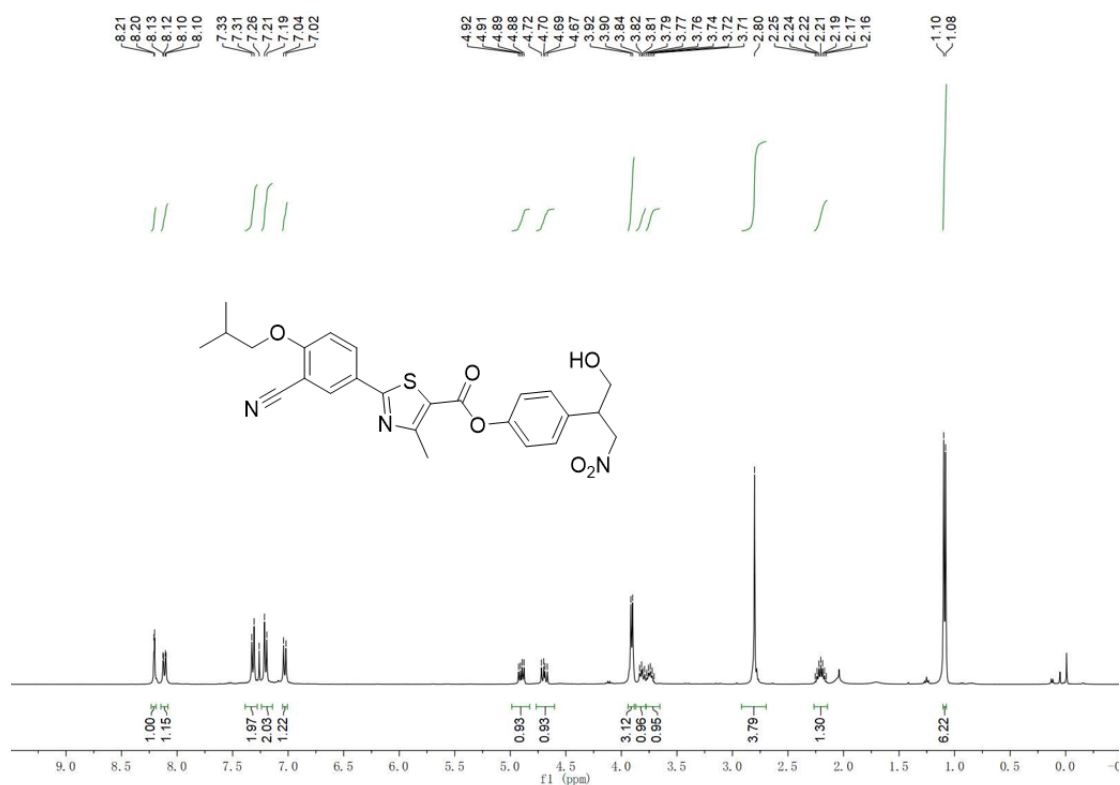
¹H NMR (400 MHz, CDCl₃) of **3ag**



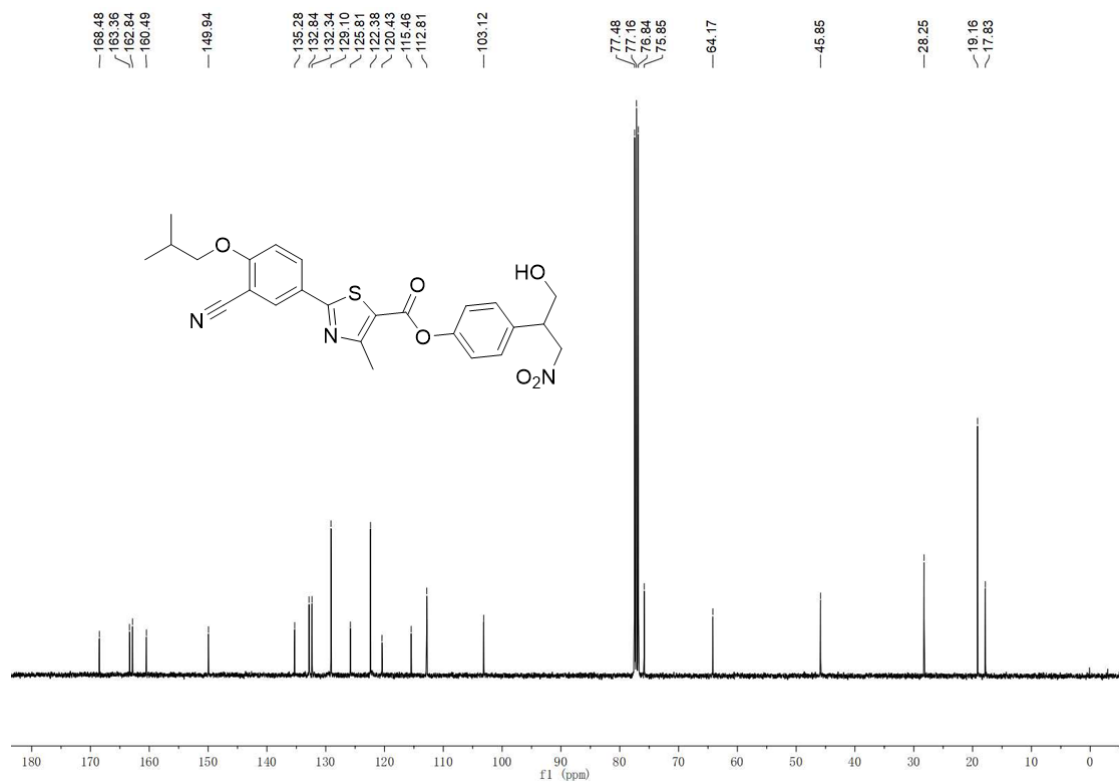
¹³C NMR (101 MHz, CDCl₃) of **3ag**



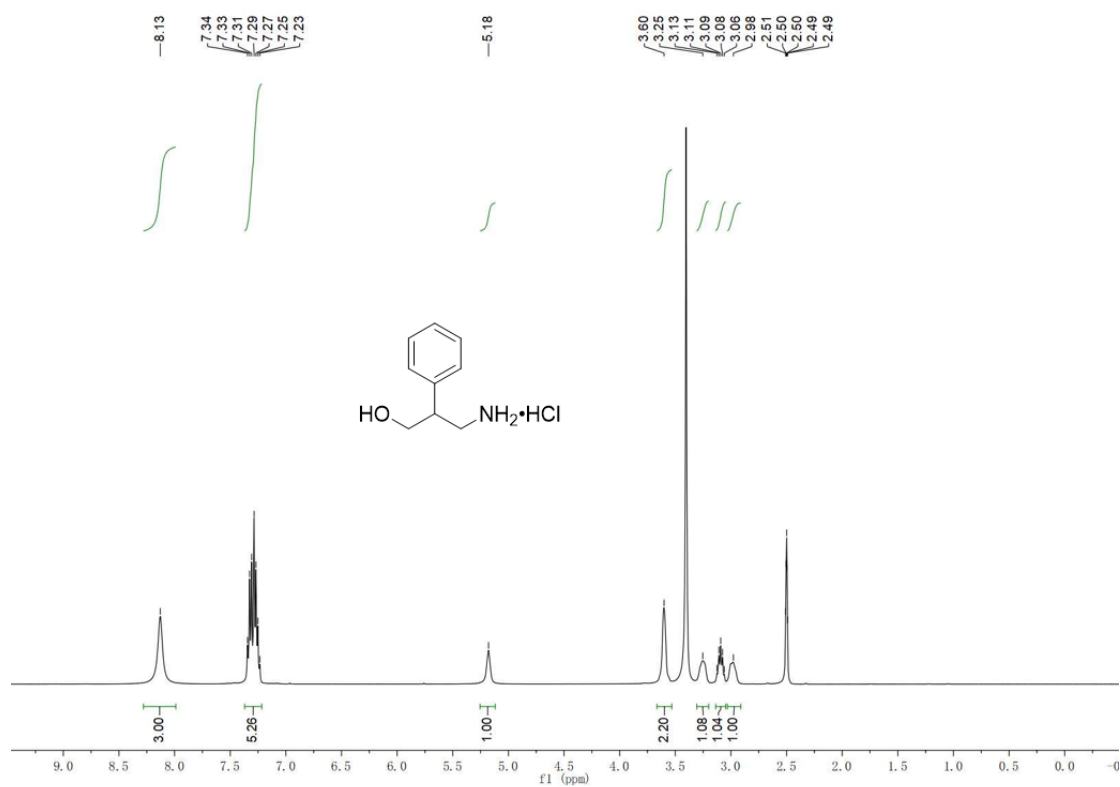
¹H NMR (400 MHz, CDCl₃) of **3ah**



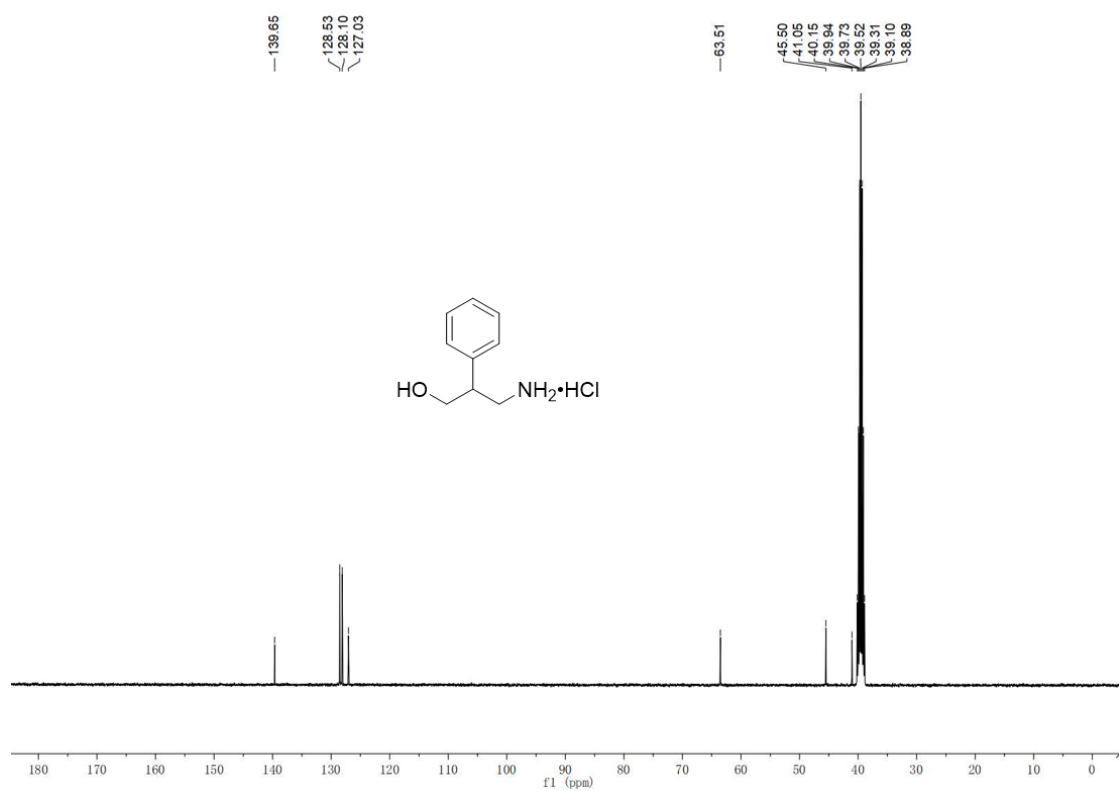
¹³C NMR (101 MHz, CDCl₃) of **3ah**



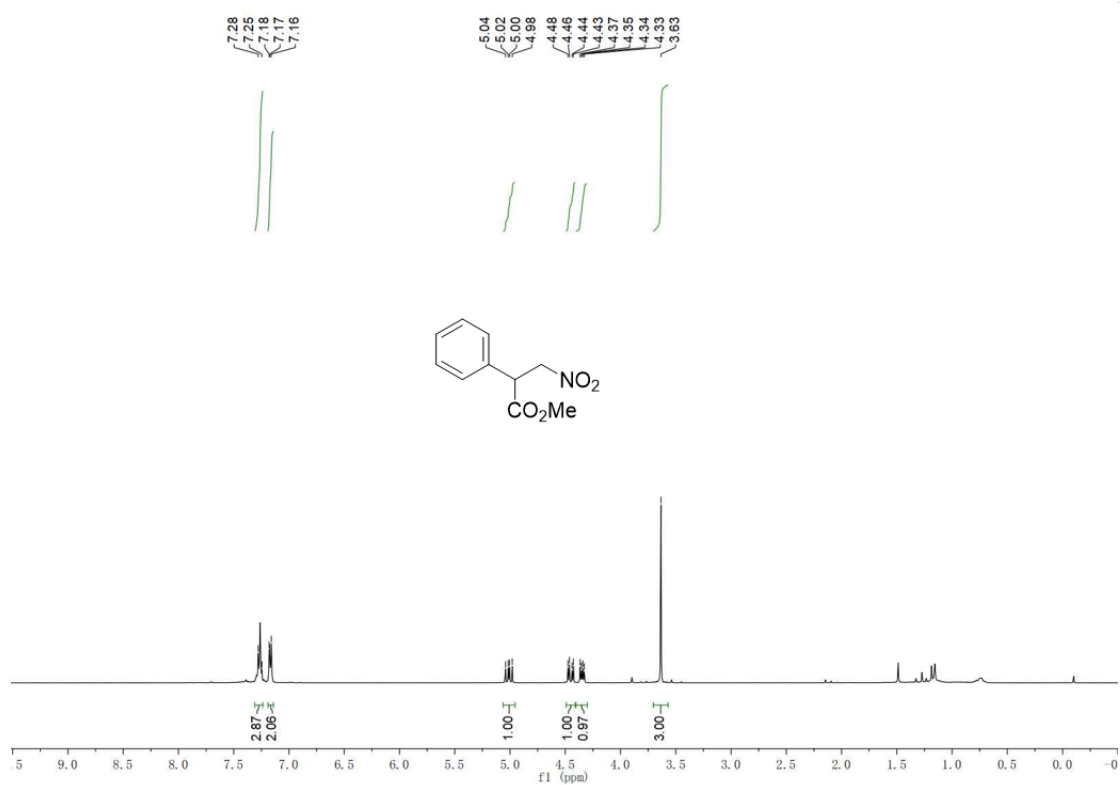
¹H NMR (400 MHz, DMSO-d₆) of **4**



¹³C NMR (101 MHz, DMSO-d₆) of **4**



¹H NMR (400 MHz, CDCl₃) of **5**



¹³C NMR (101 MHz, CDCl₃) of **5**

