

SUPPLEMENTARY INFORMATION

Solvated Electron-Driven Hydroamination of Olefins

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1.0 General materials and methods

Unless otherwise noted, commercial reagents, catalysts, and solvents were purchased from Sigma-Aldrich and utilized without further purification. We used solely commercial suppliers for olefins and amines except 3,4-dimethoxystyrene, and 2,6-dimethoxystyrene which were synthesized according to standard procedures. A water bath was used to concentrate organic solutions under reduced pressure on a Büchi rotary evaporator. Utilizing flash chromatography on silica gel, products were chromatographically purified (Interchim, 230-400 mesh). Analtech Uniplate 0.25 mm silica gel plates were utilized for preparative thin layer chromatography (PTLC) for purification and visualized using a UV 254 nm light source. The UV-Vis absorption spectra were measured using Shimadzu UV-2450 Spectrometer scanned between 200 and 800 nm wavelengths. Branson CPX2800H Digital Heated Ultrasonic Cleaner was employed for sonication.

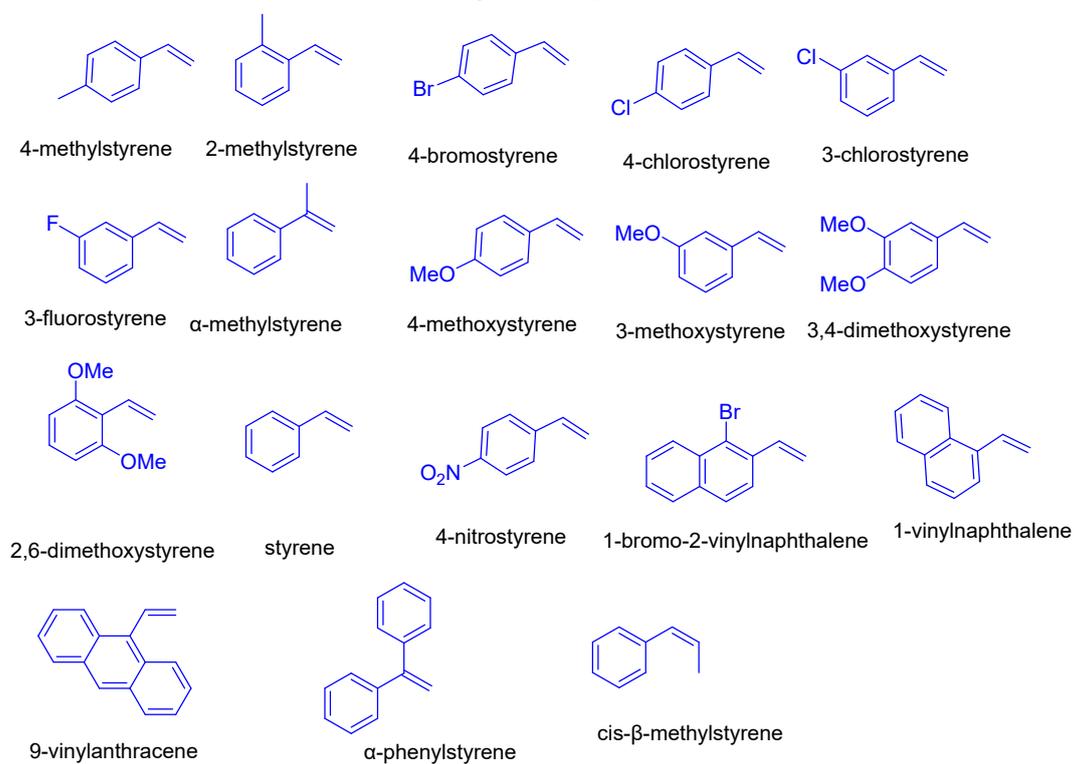
Bruker UltraShield Plus 400 (75 MHz) or 500 MHz (125 MHz) instruments were used to record the ^1H and ^{13}C NMR spectra, which are internally referred to the residual protio solvent signals (CDCl_3 referenced at 7.26 and 77.0 ppm respectively). For ^1H NMR, the following information is provided: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, m = multiplet, dd = doublet of doublets, and br = broad and its various combinations), coupling constant (Hz), and integration. No specific terminology is utilized for equivalent carbons when reporting ^{13}C NMR data; instead, data are provided in terms of chemical shift. High resolution mass spectra were obtained at the University of Connecticut mass spectrometry facilities on Agilent Technologies 6220 Time Of-Flight LC/MS with electrospray ionization method. Quantitative analysis of the material was done using ^1H NMR with 1,4-dinitrobenzene as the internal standard or by isolated yields.

For ^1H NMR quantification, a stock solution of the internal standard was prepared to a concentration of 0.05 M by dissolving 1 mmol of 1,4-dinitrobenzene (168 mg) in 20 mL of chloroform from which aliquots described in the relevant sections were taken and added to the sample for NMR quantification. Where peaks were not distinct, the materials were rather isolated, and yields were quantified.

Note: It is important to note that sonication was employed in place of using a stir bar for mixing because stir bars reacted with the solvated electrons and thus rendered the reaction inefficient. This observation has also been reported in literature.^{1,2} Gratifyingly, sonication also increased the efficiency of our reaction compared to when stir bars were employed by enabling better mixing and granulation of lithium. However, there is an inherent risk due to the proximity of lithium metal within the reactor and the water bath. Those replicating this reaction should proceed with caution.

2.0 Successful substrate scope

Vinylarene scope



Amine scope

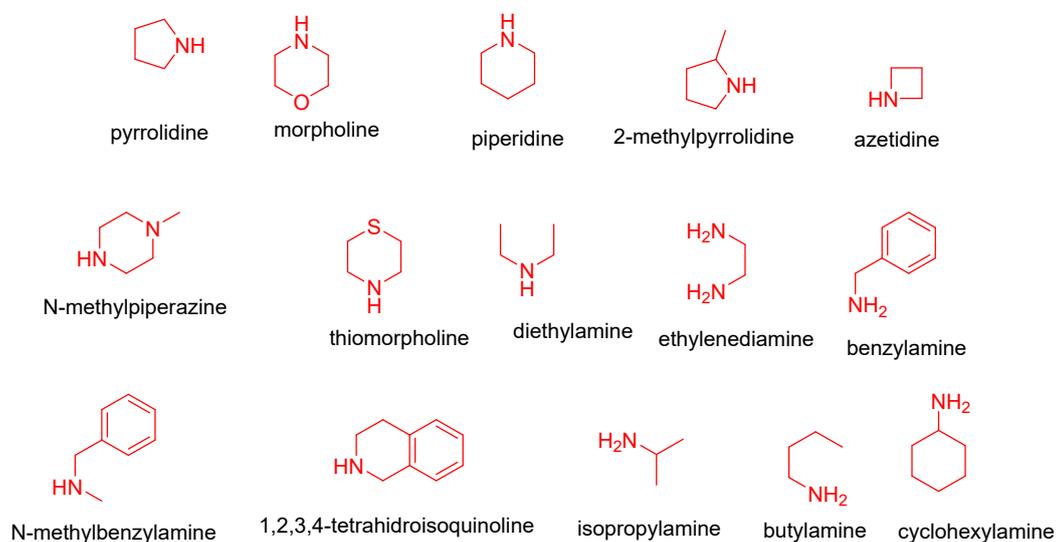


Figure S1: Successful substrate scope

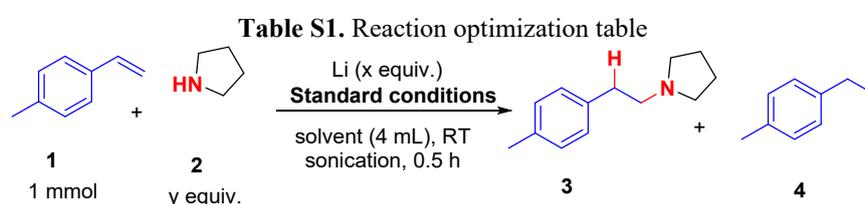
3.0 General procedure for reaction optimization studies

In a flame-dried two-neck round-bottom flask equipped with a septum and connected to a Schlenk line, lithium metal (designated amount, x equiv) and, when applicable, potassium iodide (0.5 equiv; see Table S1, entry 1) were added and dried under high vacuum for 10 minutes. The flask was then purged and backfilled with nitrogen.

Under an inert atmosphere, 4 mL of anhydrous solvent was introduced, followed by 4-methylstyrene (1 mmol, 0.13 mL) and pyrrolidine (y equiv), added via syringe through the septum. The resulting mixture was sonicated at room temperature for 30 minutes under nitrogen.

After sonication, the reaction mixture was cooled to 0 °C in an ice-water bath and quenched by the dropwise addition of 20 mL deionized water. The aqueous phase was extracted with diethyl ether (3 × 20 mL), and the combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure using a rotary evaporator.

The crude product was analyzed, and yields were determined after purification by preparative thin-layer chromatography (prep TLC) using 1,4-dinitrobenzene as internal standard. Optimization results are summarized in Table S1.



entry	Deviation from standard conditions	Lithium (x equiv.)	Amine (y equiv.)	Conv. (%)	Yield (%)	
					3	4
1	THF + 0.5 mmol KI	1.4	2.5	>99	64	36
2	THF	1	1.5	>99	77	23
3	Triethylamine	1	1.5	79	53	47
4	Et ₂ O	1	1.5	77	95	5
5	1,4-dioxane	1	1.5	43	91	9
6	Pentane	1	1.5	65	61	39
7	Toluene	1	1.5	80	63	37
8	Ethyl acetate	1	1.5	S.R	-	-
9	chloroform	1	1.5	S.R	-	-
10	2-MeTHF	1	1.5	>99	>99	trace

4.0 General optimized procedure

Reactions were conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) which is dried extensively under vacuum (about 10 minutes), purged, and then filled

up with nitrogen. Then 4 mL of 2-methyltetrahydrofuran followed by 1 mmol of olefin (**1**) and 1.5 mmol of amine (**2**) were added using a syringe and needle through the septum end. The reaction mixture was sonicated until sonication confirmed completion of the reaction (0.5 h – 2h). Afterwards, the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water. Extraction was done using 3 x 20 mL of diethyl ether. The organic phase was then dried over sodium sulfate and solvent evaporated using a rotary evaporator.

5.0 Reaction scope

5.1 Successful vinylarene scope

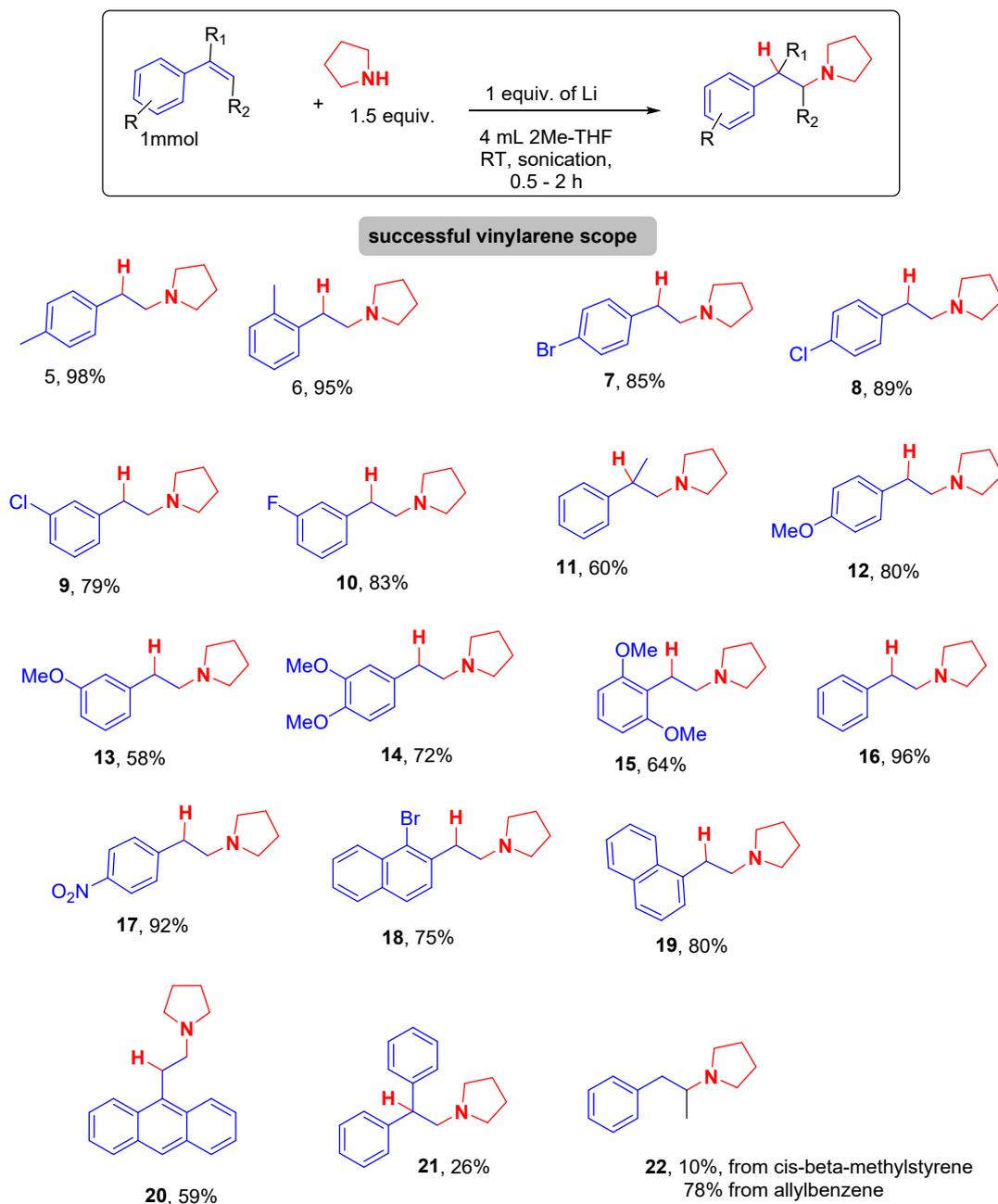


Figure S2. Vinylarene scope for the hydroamination of olefins promoted by solvated electrons. The reaction was carried out under sonication at room temperature under inert conditions for 0.5 – 2 h with 1 mmol of vinylarene and 1.5 equiv. of pyrrolidine (**2**). Yields were determined after isolation by preparative thin layer chromatography.

5.2 Unsuccessful vinylarene scope

Our substrate scope exploration extended to *meta*- and *para*-substituted nitrostyrenes. While 4-nitrostyrene underwent efficient hydroamination, 3-nitrostyrene was unreactive, likely due to the electronic influence of the *meta*-nitro group, which may hinder single-electron transfer (SET) initiation. β -Substituted styrenes such as β -methylstyrene and 1,2-dihydronaphthalene underwent complete reduction of the olefinic bond, underscoring the role of steric hindrance in diverting the reaction away from the hydroamination pathway and favoring reduction.

Sterically encumbered substrates such as 2,6-dimethylstyrene failed to react, further emphasizing the sensitivity of the hydroamination pathway to steric bulk. Substrates bearing reactive functional groups—including 3- and 4-trifluoromethylstyrene, 4-cyanostyrene, and 4-vinylbenzaldehyde—exhibited competing reactivity at the side-chain functionalities, leading to diminished or complex product mixtures.

In the case of heteroaryl olefins, 2- and 4-vinylpyridine were prone to rapid polymerization under the reaction conditions and were not investigated further. Finally, 4-vinylbenzoic acid remained unreactive, consistent with our prior observations regarding acidic substrates.² The strongly basic nature of the reducing system likely deprotonates such substrates to form their conjugate bases, which act as quenchers and suppress both the hydroamination and reduction pathways.

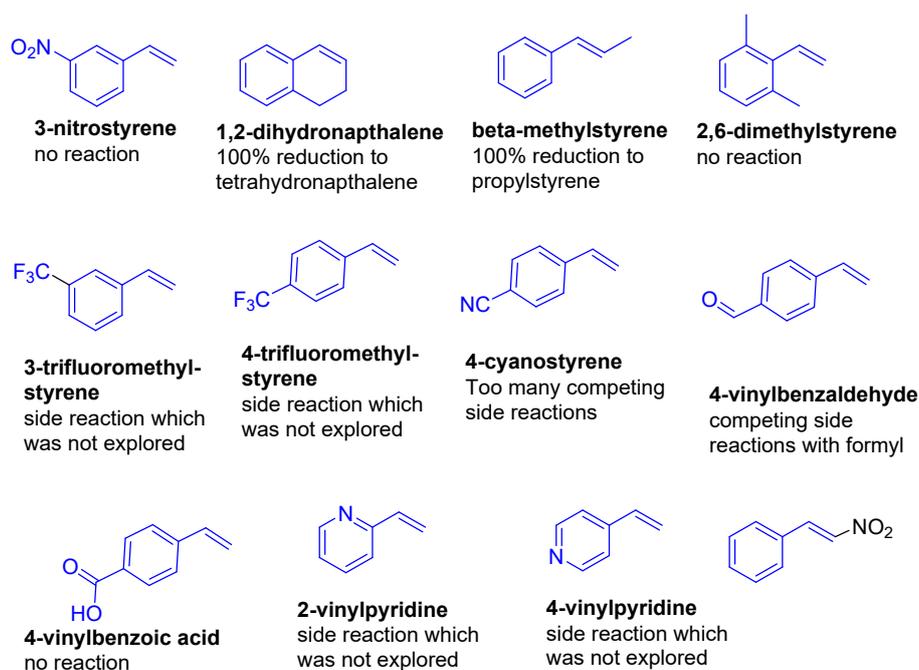


Figure S3. Unsuccessful vinylarene scope including a brief discussion of our observation and conclusion.

5.3 Competing reactivity with 3-fluorostyrene

To probe the impact of solvent environment on chemoselectivity, we examined the reaction of 3-fluorostyrene under standard conditions in both THF and 2-methyltetrahydrofuran (2-MeTHF). In THF, multiple pathways were observed, including hydroamination, hydrodefluorination, and undesired side-chain amination, resulting in a complex mixture. In contrast, reactions performed in 2-MeTHF showed exclusive formation of the desired hydroamination product (compound 10), demonstrating that solvent polarity and coordination can significantly modulate chemoselectivity in this system. These results underscore the role of 2-MeTHF in suppressing undesired pathways and promoting cleaner conversions.

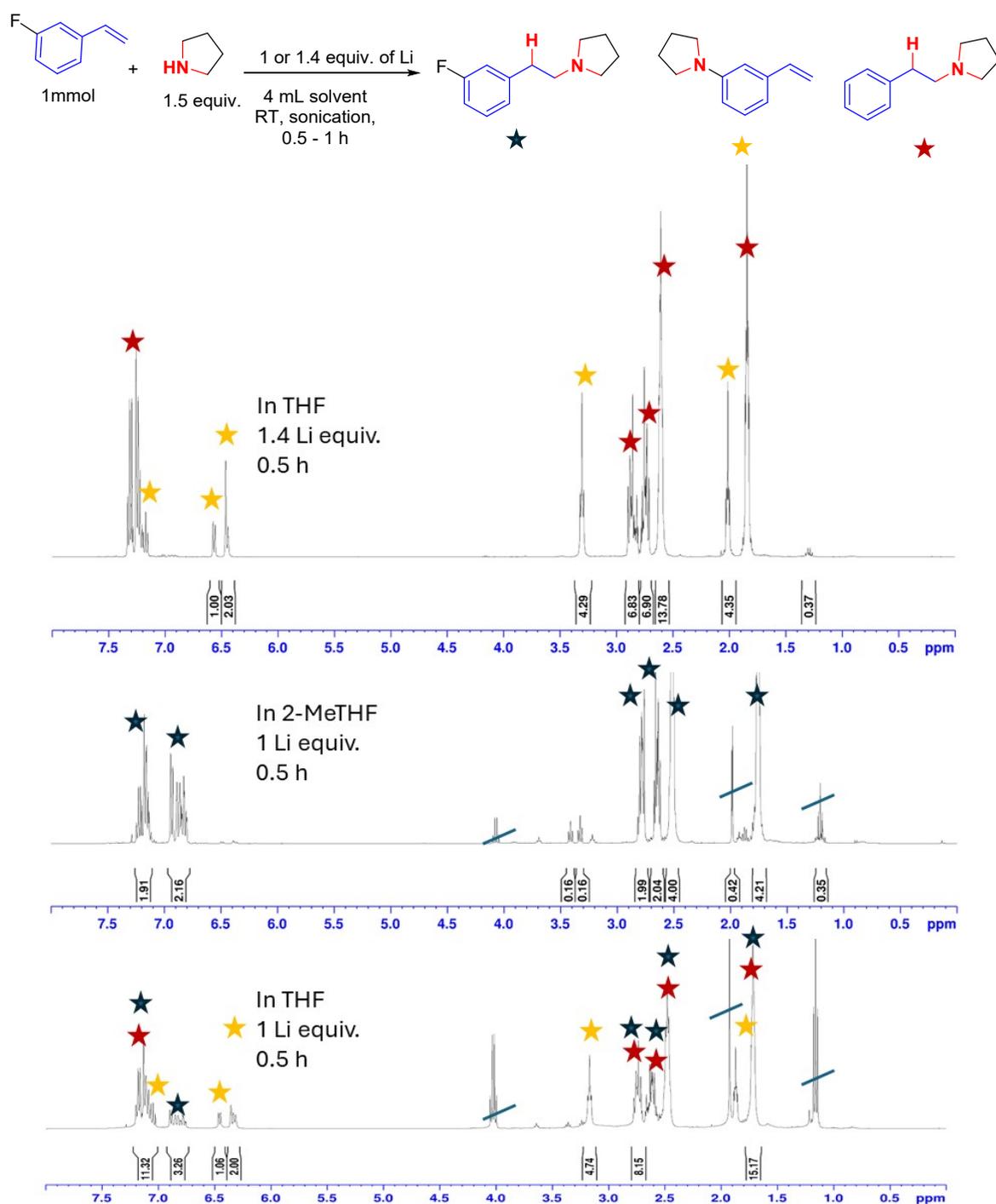


Figure S4. Competing selectivity observed during the reaction of 3-fluorostyrene in THF and 2-MeTHF. Top: In THF (1.4 equiv. Li, 0.5 h), a mixture of products arising from hydroamination, hydrodefluorination, and side-chain

amination was detected. Middle: In 2-MeTHF (1 equiv. Li, 0.5 h), selective formation of the hydroamination product (compound 10) was observed. Bottom: In THF (1 equiv. Li, 0.5 h), similar competing pathways were again evident, confirming the reduced selectivity in this solvent. Colored stars indicate distinct product signals corresponding to each pathway. Crossed out peaks are ethylacetate traces from extraction procedure.

5.4 Successful amine scope

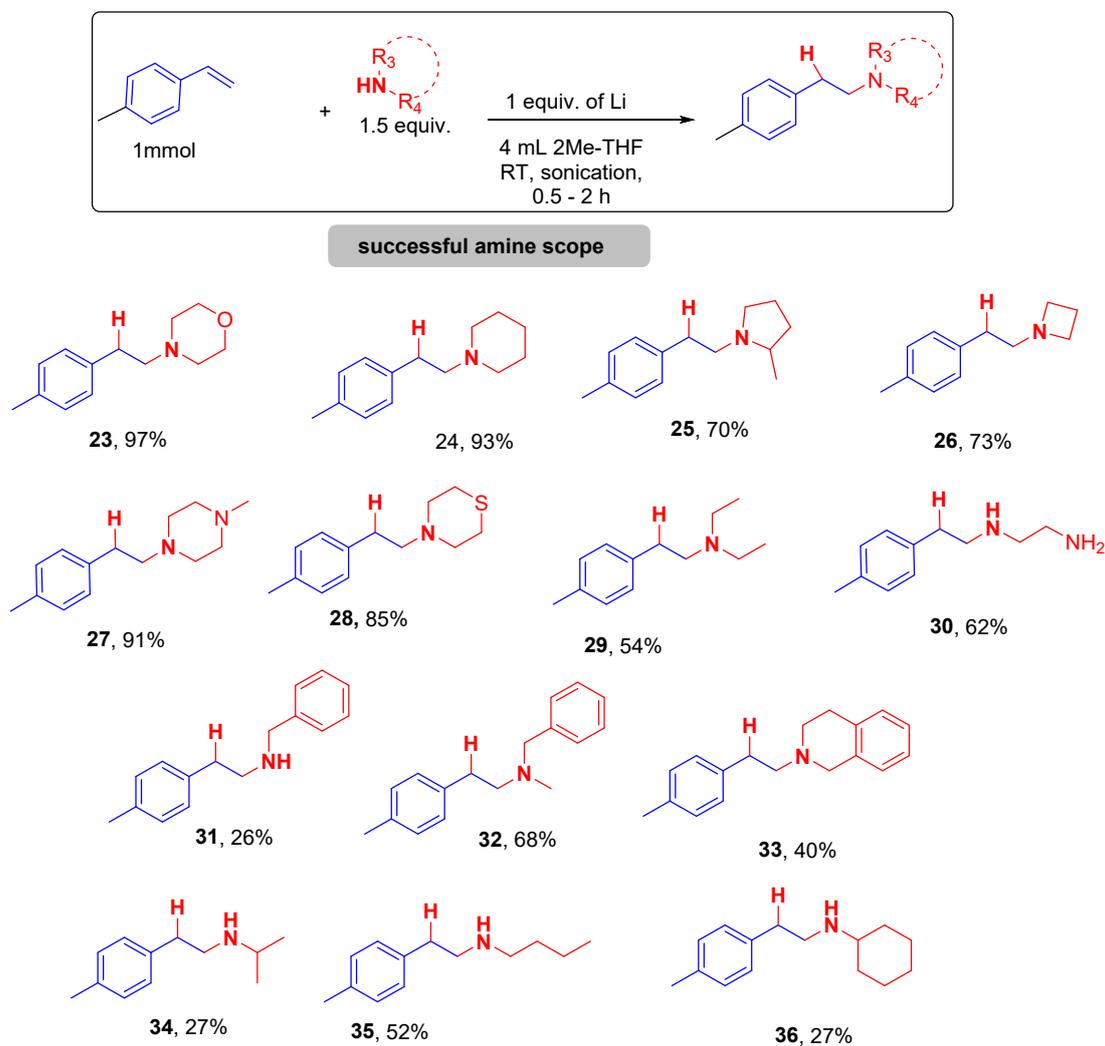


Figure S5. Amine scope for the hydroamination of olefins promoted by solvated electrons. The reaction was carried out under sonication at room temperature under inert conditions for 0.5 – 2 h with 1 mmol of 4-methylstyrene (**1**) and 1.5 equiv. of amines. Yields were determined after isolation by preparative thin layer chromatography.

5.5 Unsuccessful Aniline

To evaluate the substrate scope with less nucleophilic amines, we investigated the hydroamination of 4-methylstyrene with aniline under the optimized lithium-mediated conditions. As shown in Figure S6, 72% conversion of the starting material was observed; however, no hydroamination product was detected by ^1H NMR. Instead, the major products were the reduced alkane (20%) and the dimerized compound (80%), arising from radical coupling of the benzylic intermediates. The low nucleophilicity and high oxidation potential of aniline likely prevent effective nitrogen-centered addition, diverting the reaction toward reductive and radical termination pathways. These results emphasize that the hydroamination proceeds efficiently only with aliphatic amines capable of stabilizing the amide radical intermediate.

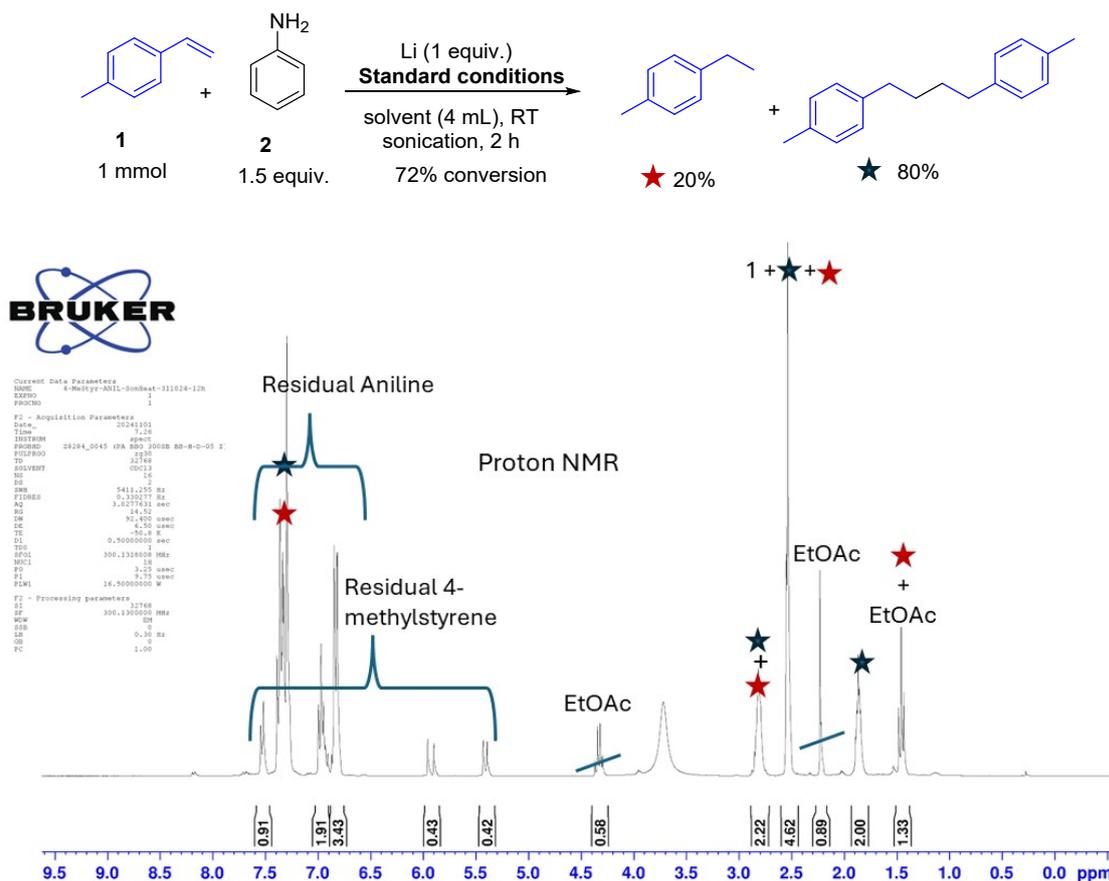


Figure S6. Reactivity of 4-methylstyrene (1) with aniline (2) under standard conditions. The reaction exhibited 72% conversion of starting material but failed to yield the desired hydroamination product. Instead, a mixture of reduced and dimerized products was obtained, consistent with lithium-mediated reduction and radical coupling pathways. Reaction conditions: 1 mmol of 4-methylstyrene (1), 1.5 equiv. aniline (2), 1 equiv. Li, 4 mL 2-MeTHF, RT, sonication, 2 h, under inert atmosphere. Yields determined by ^1H NMR using 1,4-dinitrobenzene as internal standard.

6.0 Mechanistic Studies

6.1 Radical quenching experiment with TEMPO

Reaction was conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg), and 1 mmol of TEMPO (156.25 mg) which are dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of tetrahydrofuran followed by 1 mmol of 4-methylstyrene (0.13 mL) and 1.5 mmol of pyrrolidine (0.13 mL) were added using a syringe and needle through the septum end. The reaction mixture was sonicated until sonicated for 0.5 h. Afterwards, the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water. Extraction was done using 3 x 20 mL of ethyl acetate. The organic phase was then dried over sodium sulfate and solvent evaporated using a rotary evaporator.

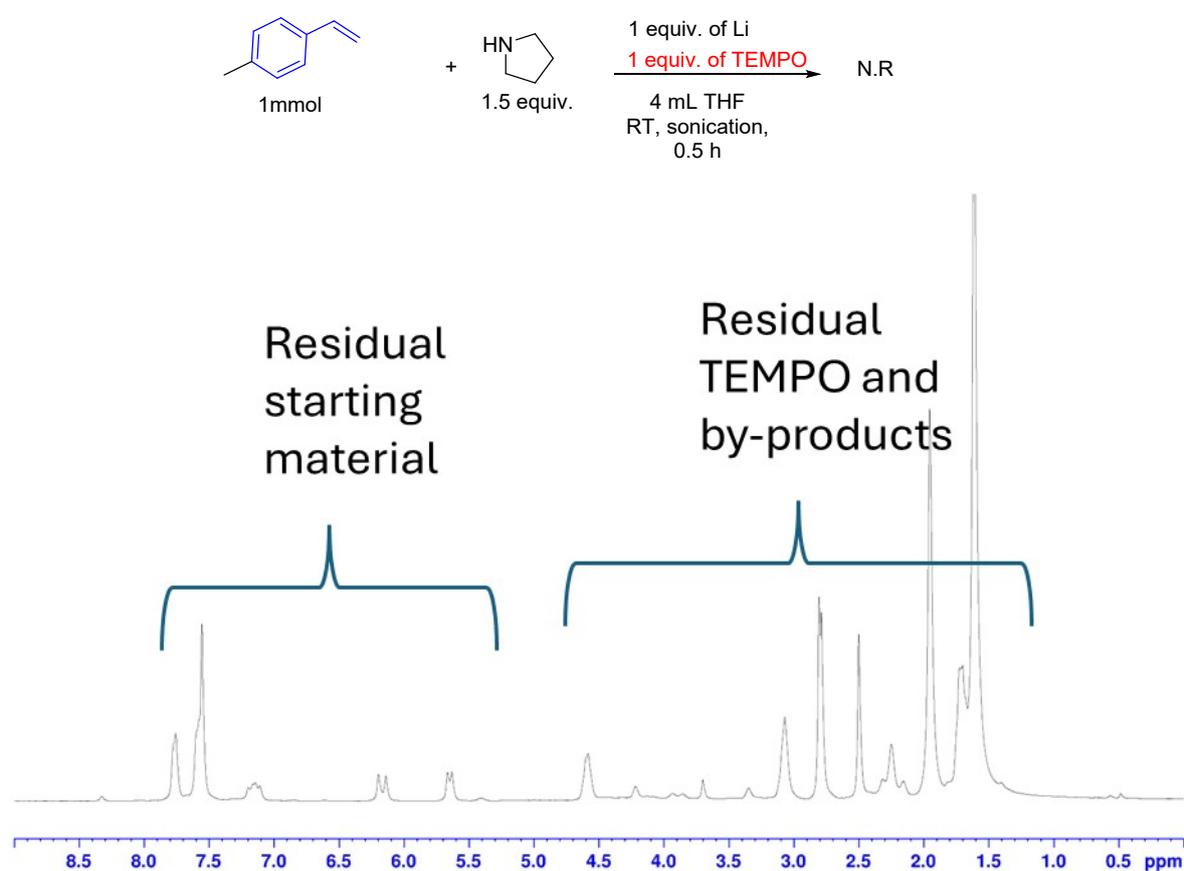


Figure S7. Radical quenching experiment using TEMPO. No desired product peaks were observed, indicating effective quenching of the reaction. The ¹H NMR spectrum shows residual 4-methylstyrene along with signals corresponding to TEMPO and its derivatives, confirming the suppression of the radical pathway.

6.2 D₂O Quenching experiment

Reaction was conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) which is dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of tetrahydrofuran followed by 1 mmol of 4-methylstyrene (0.13 mL) and 1.5 mmol of pyrrolidine (0.13 mL) were added using a syringe and needle through the septum end. The reaction mixture was sonicated until sonicated for 0.5 h. Afterwards, the mixture was cooled to 0 °C in

an ice bath and then quenched with 20 mL of deuterated water (D_2O). Extraction was done using 3 x 20 mL of ethyl acetate. The organic phase was then dried over sodium sulfate and solvent evaporated using a rotary evaporator.

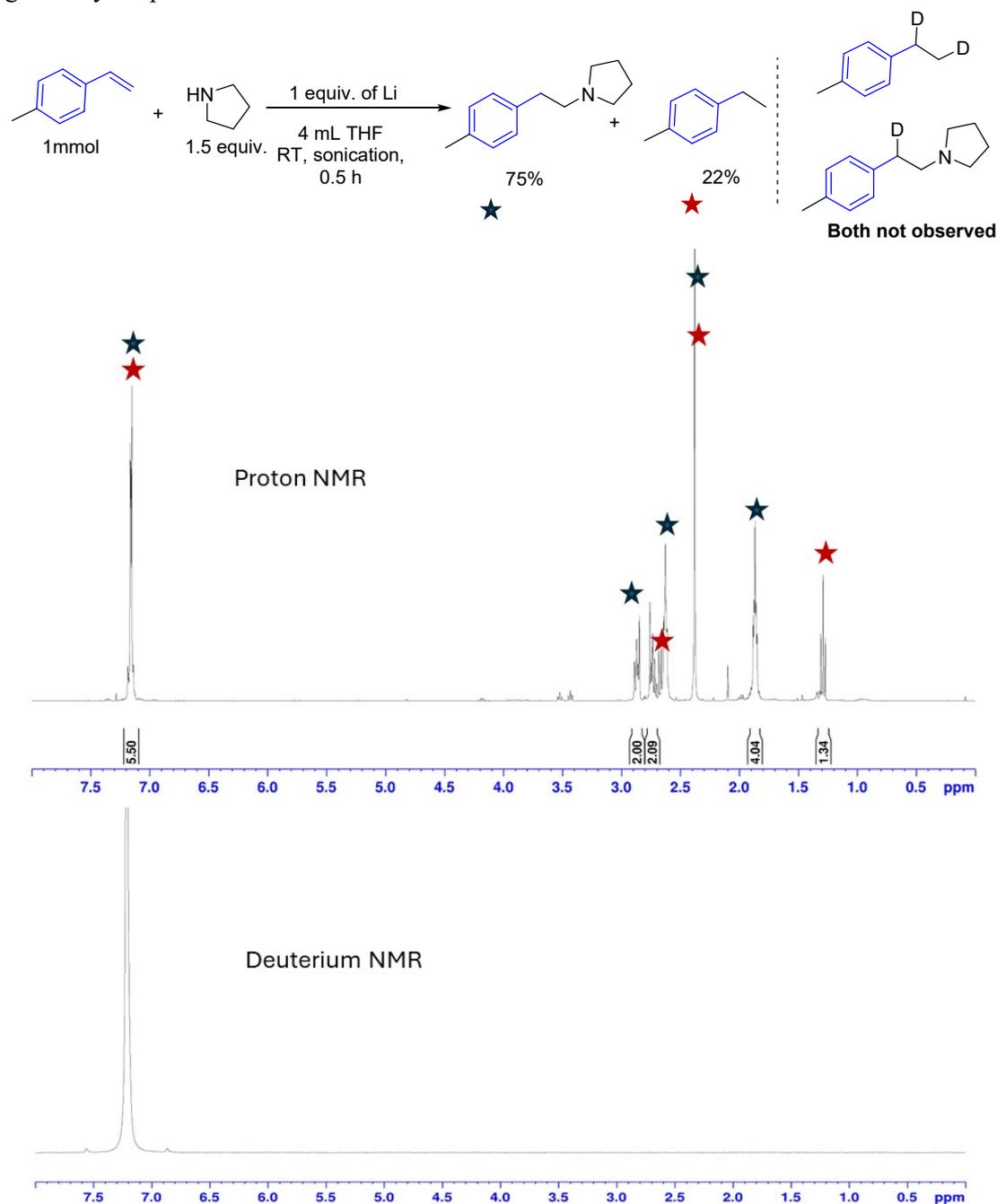


Figure S8. D_2O quenching experiment. The 1H NMR spectrum (top) spectra shows no evidence of deuterium incorporation in either product, as indicated by unchanged proton integrations. The 2H NMR spectrum (bottom) confirms this result, showing no detectable deuterium signals.

6.3 Deuterated solvent experiment

Reaction was conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) which is dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of deuterated tetrahydrofuran (d_8 -THF) followed by 1 mmol of 4-methylstyrene (0.13 mL) and 1.5 mmol of piperidine (0.13 mL) were added using a syringe and needle

through the septum end. The reaction mixture was sonicated for 0.5 h. Afterwards; the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water (D₂O). Extraction was done using 3 x 20 mL of ethyl acetate. The organic phase was then dried over sodium sulfate and solvent evaporated using a rotary evaporator.

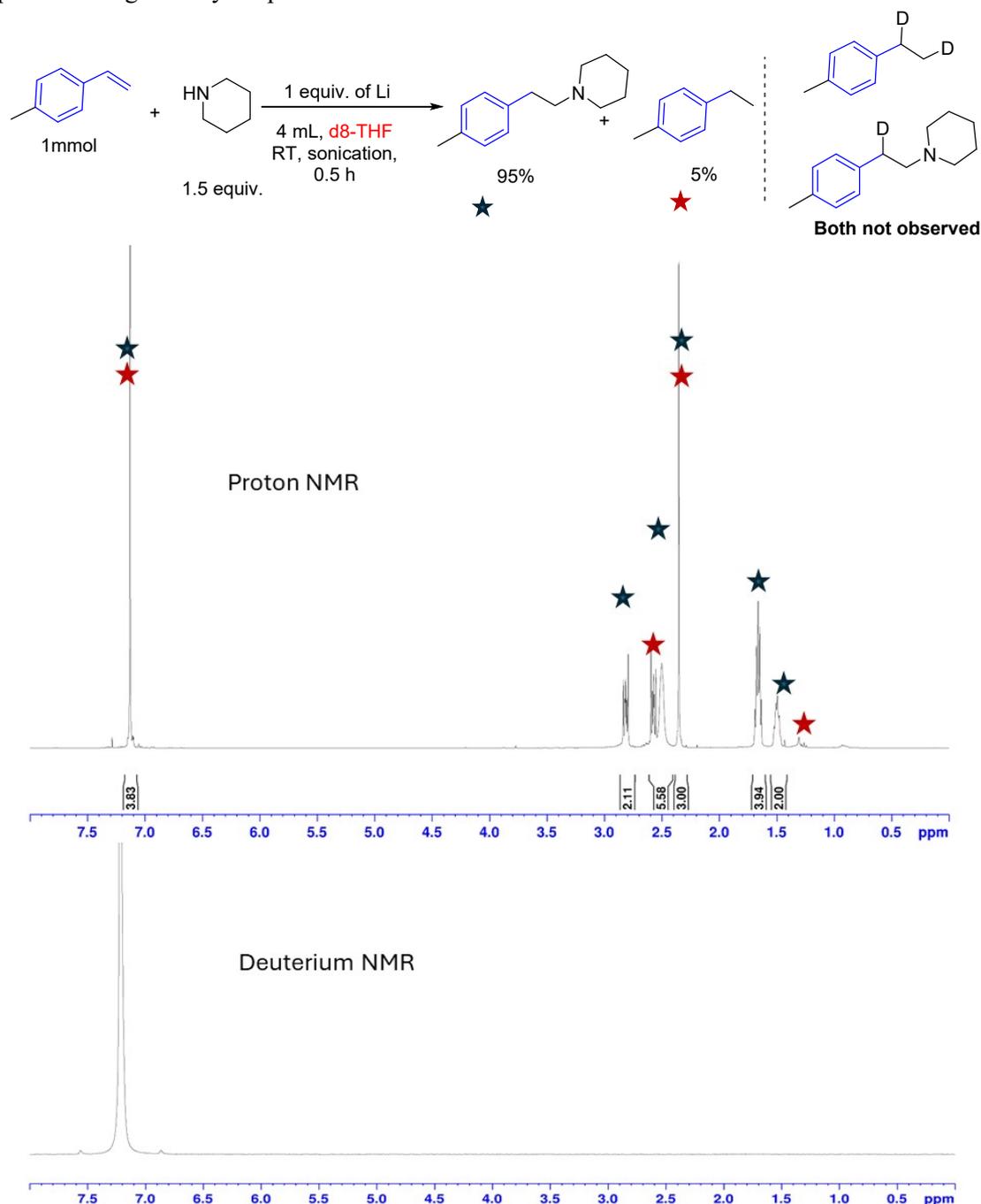


Figure S9. Deuterated solvent experiment. The ¹H NMR spectrum (top) spectra shows no evidence of deuterium incorporation in either product, as indicated by unchanged proton integrations. The ²H NMR spectrum (bottom) confirms this result, showing no detectable deuterium signals.

6.4 Deuterated piperidine experiment

Synthesis of N-deuterated piperidine

In a flame-dried, nitrogen-flushed round-bottom flask equipped with a magnetic stir bar, piperidine (10 mmol, 0.85 mL) was dissolved in anhydrous tetrahydrofuran (THF, 10 mL) and cooled to 0 °C in an ice bath. To this solution, n-butyllithium (n-BuLi, 12 mmol, 4.8 mL of a 2.5 M solution in hexanes)

was added dropwise over 2 minutes under inert atmosphere. The reaction mixture was stirred at 0 °C for 30 minutes to ensure complete deprotonation of the amine. After this period, excess D₂O (10 mL) was added dropwise to quench the reaction and introduce deuterium at the nitrogen center. The mixture was extracted with diethyl ether (3 x 10 mL) and the combined organic layers were dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was used without further purification. The ¹H and ²H NMR (Figure S9) confirm successful deuterium incorporation, as evidenced by the attenuation/appearance of the N-H/N-D signals in the ¹H and ²H NMR spectrum, respectively.

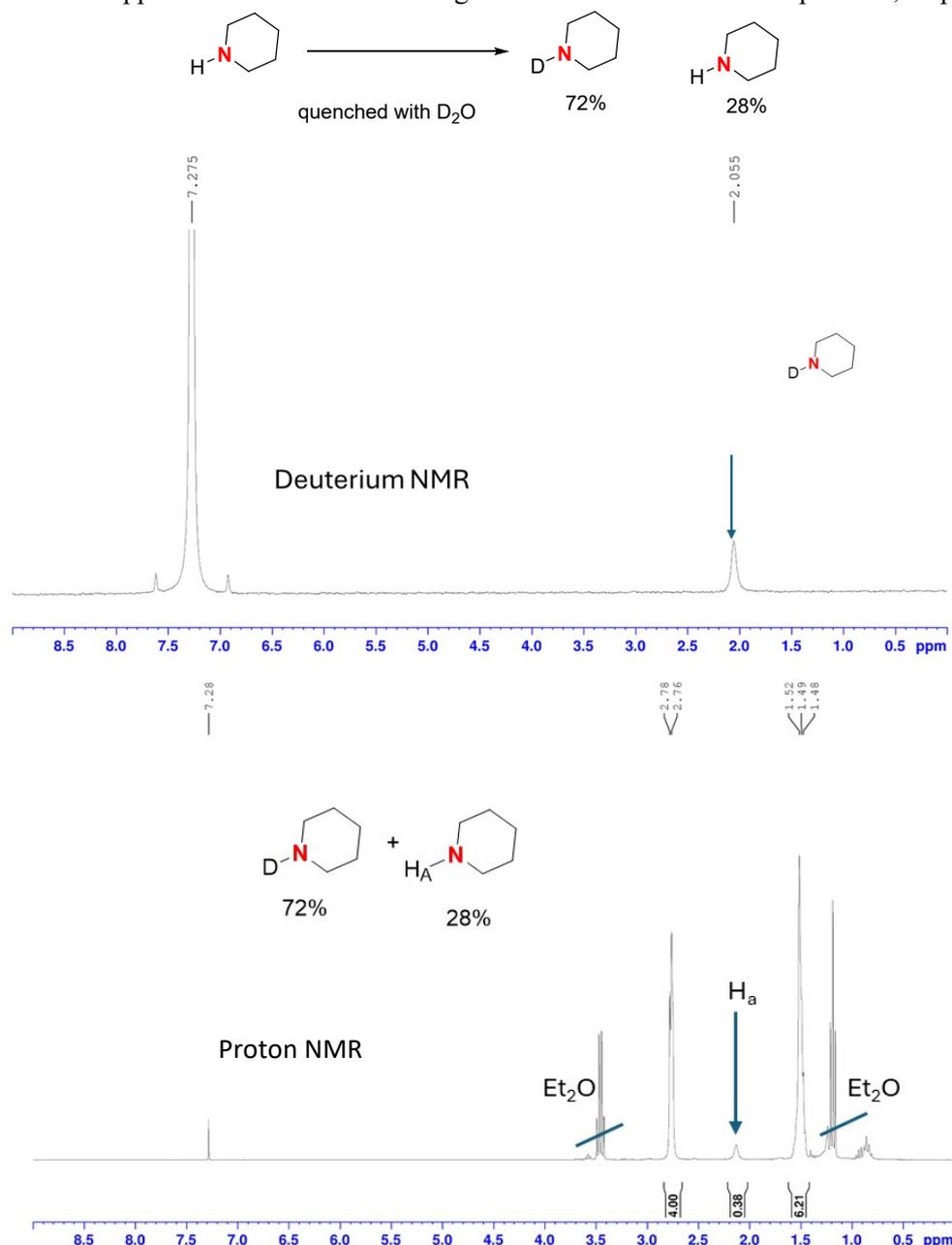


Figure S10. ¹H (top) and ²H (bottom) NMR confirming the successful synthesis of N-deuterated piperidine. The distinct ²H signal at δ 2.08 ppm corresponds to the N–D proton, verifying deuterium incorporation at the nitrogen position.

Procedure for the deuterated piperidine experiments.

All reactions were conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) which is dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of deuterated tetrahydrofuran (d₈-THF) followed by 1 mmol of 4-

methylstyrene (0.13 mL) and 1.5 mmol of piperidine (0.13 mL) were added using a syringe and needle through the septum end. The reaction mixture was sonicated for 0.5 h. Afterwards; the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water (D₂O). Extraction was done using 3 x 20 mL of ethyl acetate. The organic phase was then dried over sodium sulfate and solvent evaporated using a rotary evaporator.

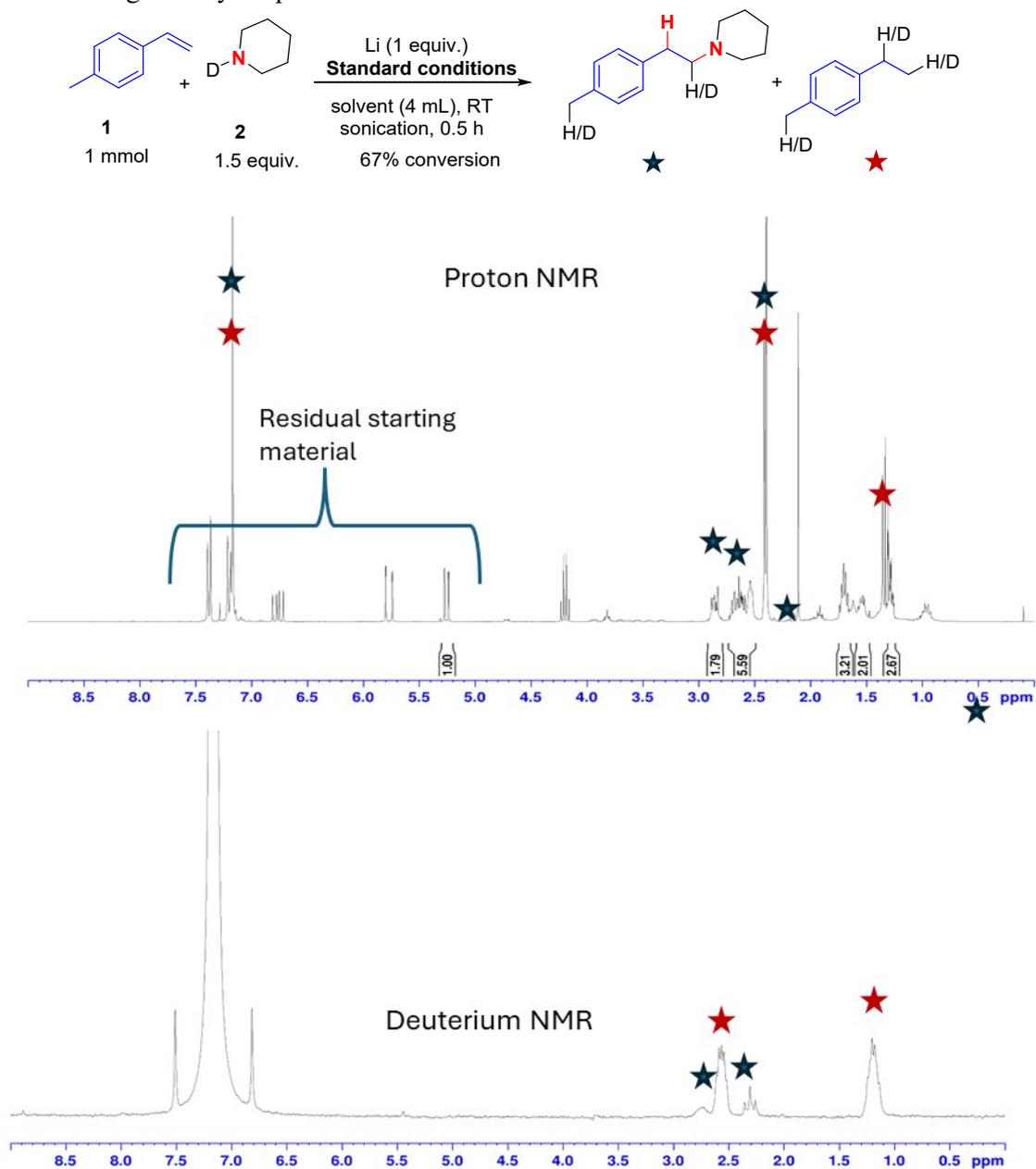
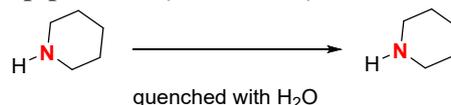


Figure S11. ²H NMR (top) and ¹H NMR (bottom) spectra of the crude reaction mixture from the lithium-mediated hydroamination of 4-methylstyrene with N-deuterated piperidine under standard conditions. Both spectra show deuterium incorporation in both products. Residual starting 4-methylstyrene shown in the ¹H NMR show incomplete conversion, consistent with a kinetic isotope effect resulting from the use of N-D labeled amines.

Control reaction with N-protio piperidine (30 minutes)



N-protio piperidine was synthesized following the same procedure as for N-deuterated piperidine (see Page S14), except deionized water was used for quenching in place of D₂O. This control experiment was designed to assess whether the observed incomplete conversion in the deuterated reaction stemmed from residual impurities in the synthesized amine, rather than a true kinetic isotope effect (KIE). The results help differentiate between effects arising from isotopic substitution versus those due to reagent quality or contamination.

Reaction of 4-methylstyrene with N-protio piperidine

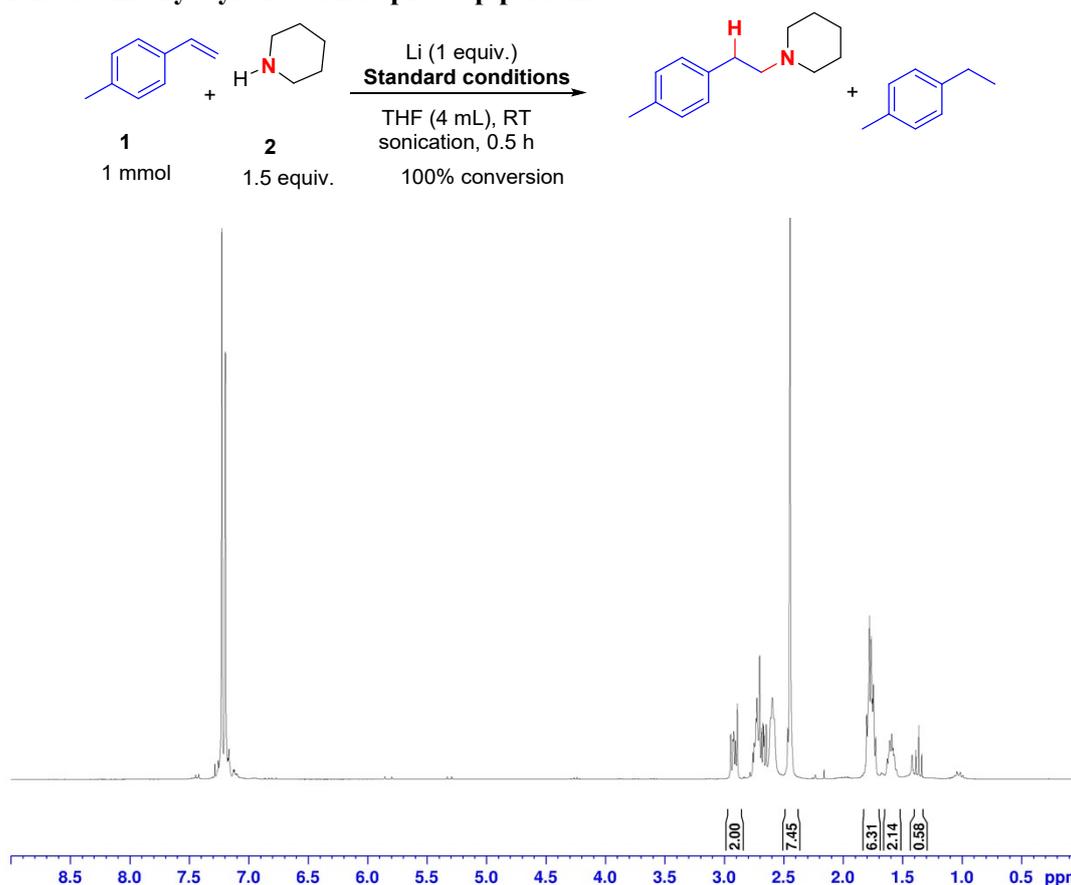


Figure S12. ¹H NMR spectra of the crude reaction mixture indicating full conversion of the 4-methylstyrene substrate.

N-deuterated piperidine experiments (1 hour)

To further probe the influence of reaction time on conversion, the hydroamination of 4-methylstyrene with N-deuterated piperidine was conducted under standard conditions for 1 hour. As shown in Figure S12, extending the reaction time led to complete consumption of the 4-methylstyrene substrate. These results confirm that the lower conversion observed in shorter reactions was due to the kinetic isotope effect (KIE) associated with the N–D bond, rather than incomplete reactivity or reagent instability.

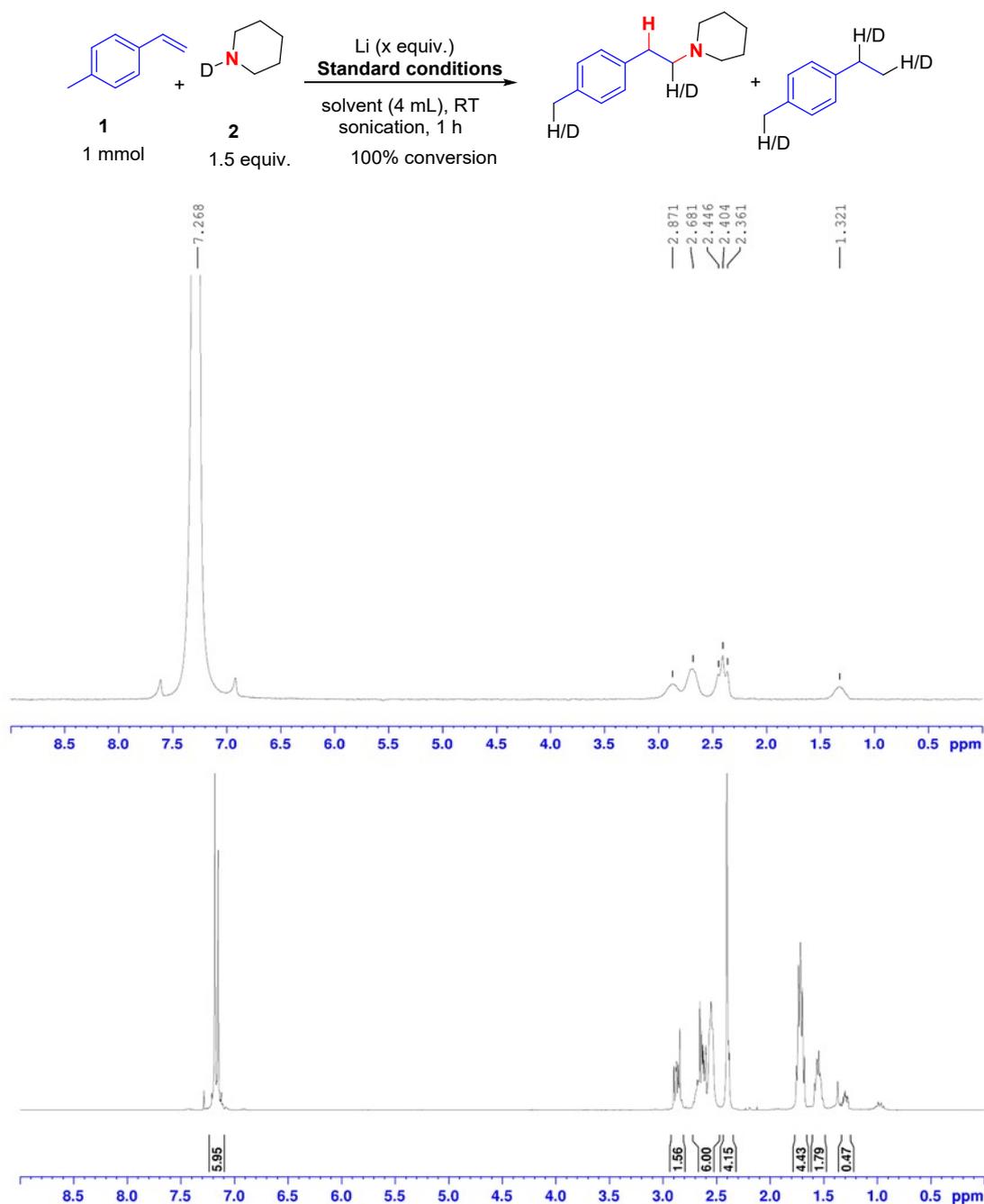


Figure S13. ^2H NMR (top) and ^1H NMR (bottom) spectra of the 1-hour reaction using N-deuterated piperidine. Complete conversion of 4-methylstyrene is observed in the ^1H NMR spectrum, with deuterium incorporation in both product regioisomers confirmed by distinct ^2H resonances, as previously described.

Isolated products

We successfully isolated both the hydroaminated and reduced products for characterization. The reduced product underwent dimerization to form the 1,4-di-p-tolylbutane, as confirmed by spectroscopic analysis. As shown in Figure S13, the ^2H NMR spectrum indicates deuterium incorporation at all benzylic positions, consistent with reduction followed by dimerization. The ^1H and ^{13}C NMR spectra further support the structure of the symmetric 1,4-di-p-tolylbutane derivative.

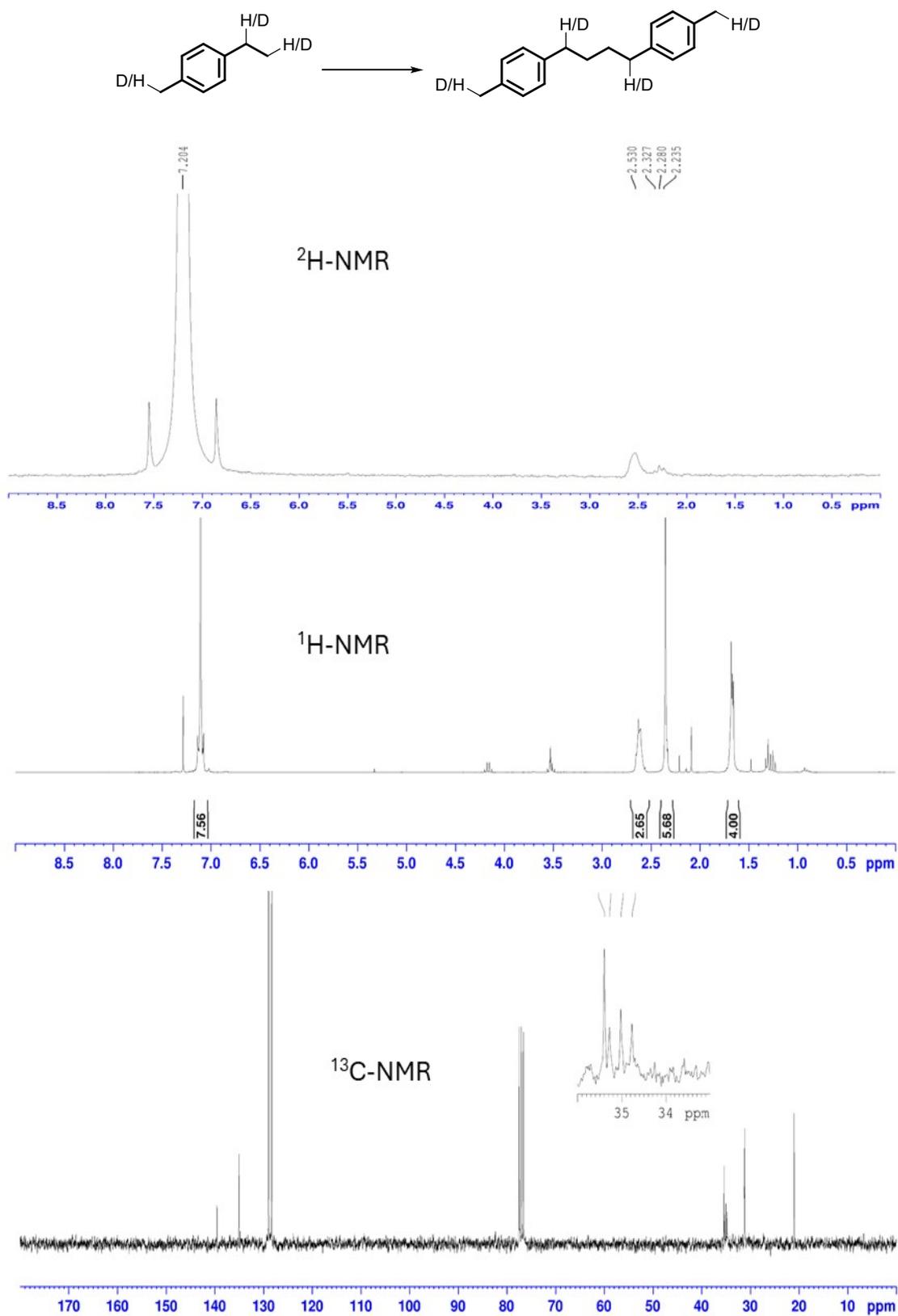


Figure S14. ^2H NMR (top) and ^1H NMR (middle) and ^{13}C NMR (bottom) spectra of the isolated reduced/dimerized product - 1,4-di-p-tolylbutane. The ^2H NMR confirms deuterium incorporation at all benzylic positions. The ^1H and ^{13}C spectra match the expected pattern for the deuterated dimer as shown.

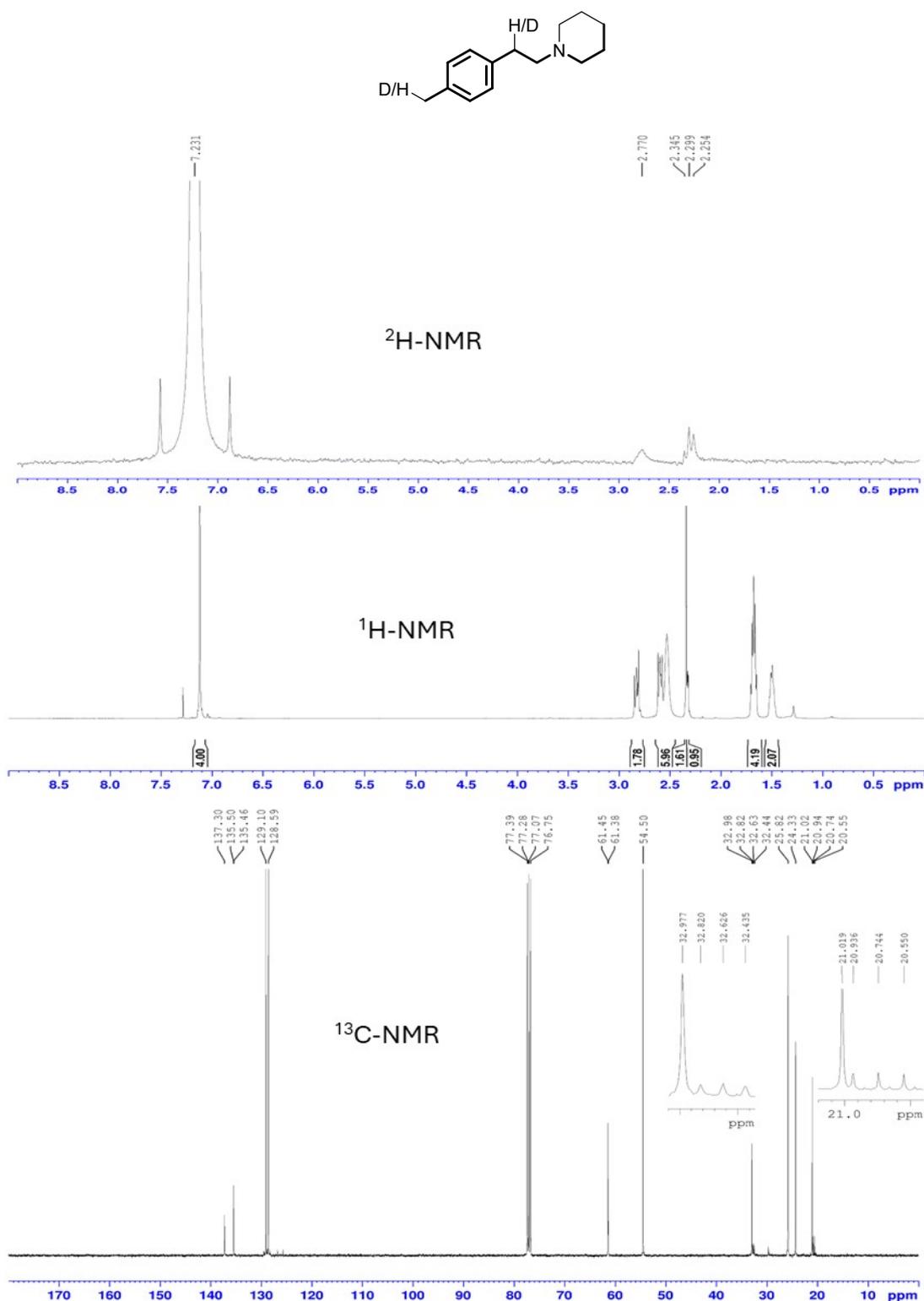


Figure S15. ^2H NMR (top) and ^1H NMR (middle) and ^{13}C NMR (bottom) spectra of the isolated reduced/dimerized product. The ^2H NMR confirms deuterium incorporation at all benzylic positions. The ^1H and ^{13}C spectra match the expected pattern for the deuterated dimer as shown.

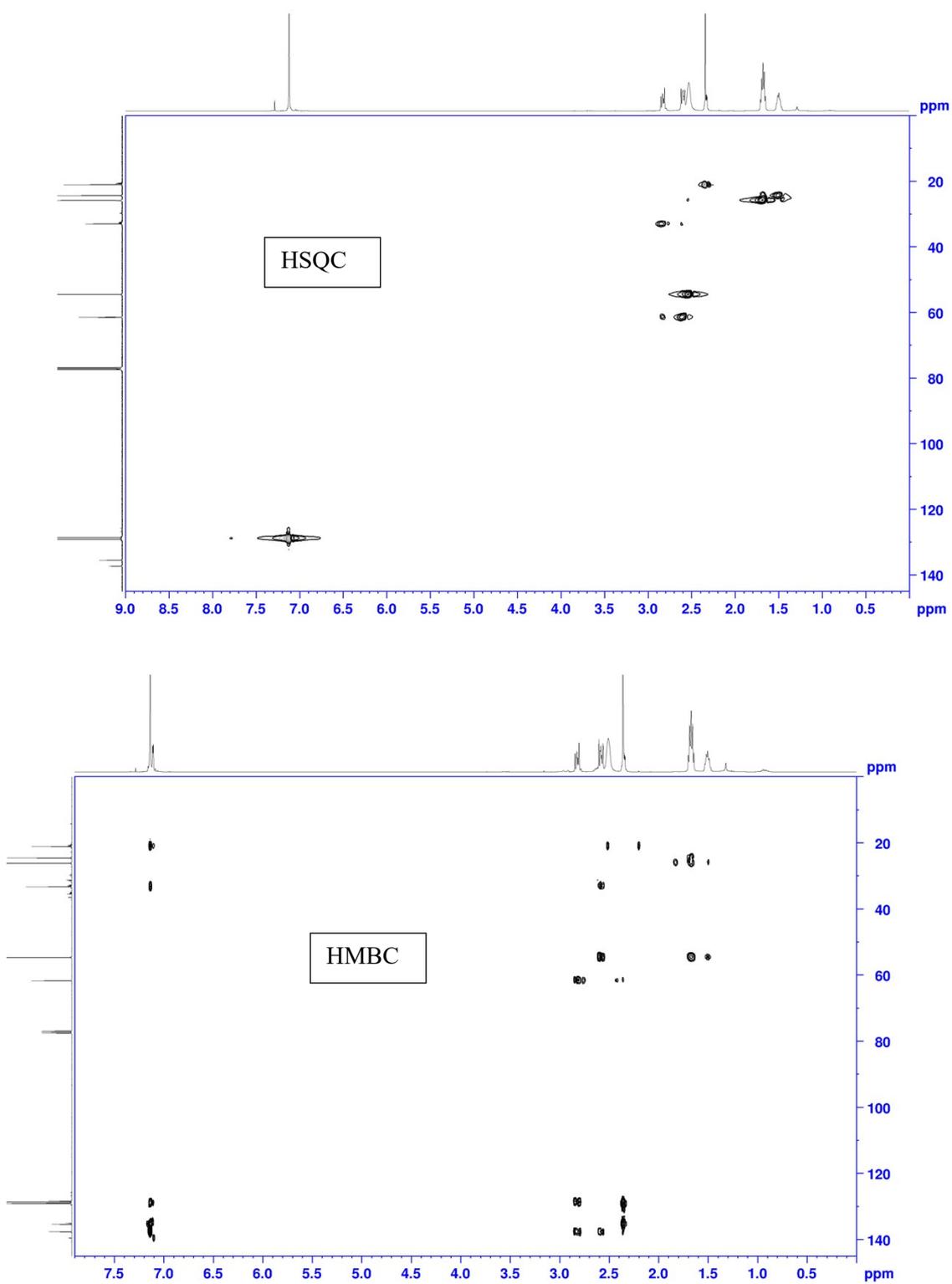


Figure S16. HSQC and HMBC NMR spectra which confirms the isolated deuterated hydroamination product.

6.5 Radical clock experiment

The reaction was conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) which is dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of tetrahydrofuran (THF) followed by 1 mmol of 4-methylstyrene (0.13 mL) and 1.5 mmol of cyclopropylamine (0.13 mL) were added using a syringe and needle through the septum end. The reaction mixture was sonicated for 0.5 h. Afterwards; the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water (D₂O). Extraction was done using 3 x 20 mL of diethyl ether, and the combined organic layers were dried over anhydrous sodium sulfate.

To prevent the loss of volatile propanal derivatives, the organic phase was analyzed directly without evaporation. An aliquot was removed and subjected to ¹H NMR analysis. The results (Figure S17) provide insight into whether ring-opening products indicative of radical intermediates were formed.

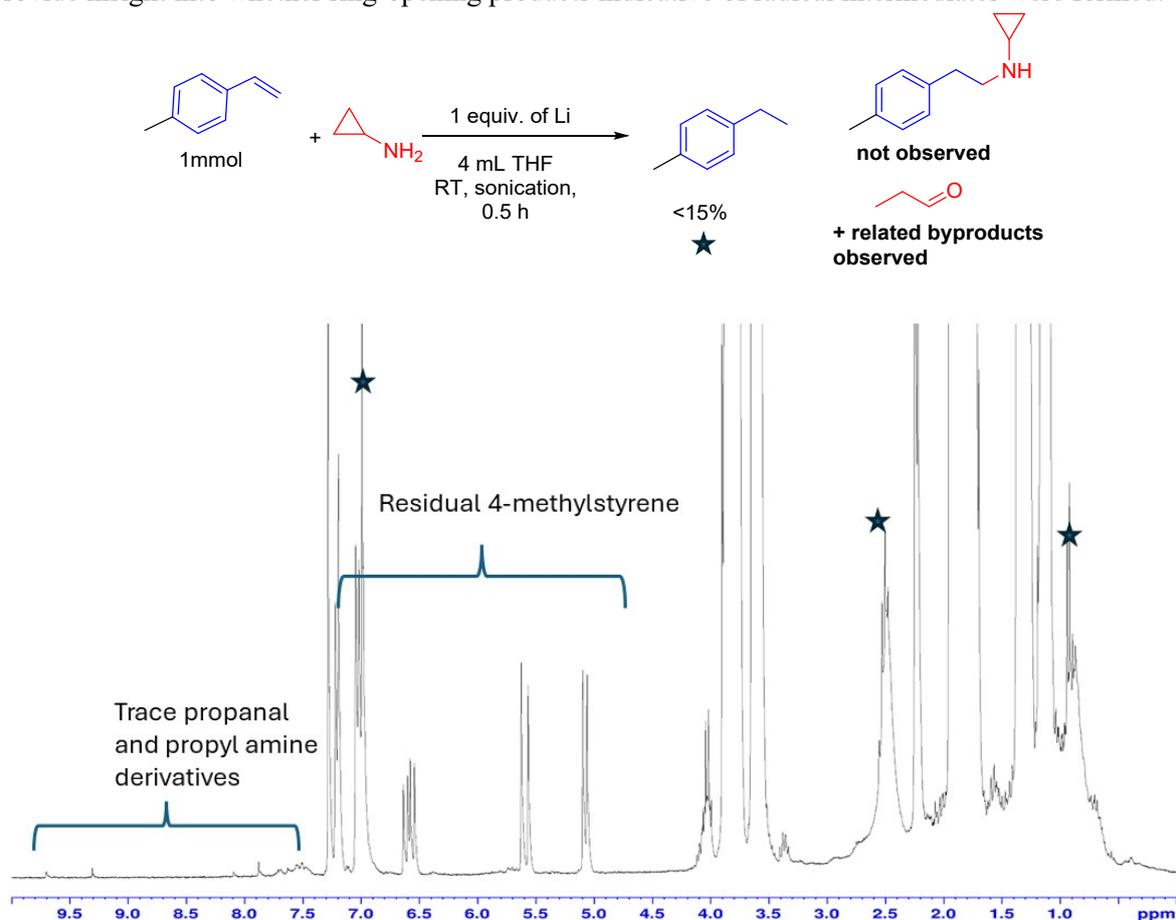


Figure S17. ¹H NMR spectrum of the crude reaction mixture from the radical clock experiment using cyclopropylamine. Prominent solvent peaks are observed due to the direct analysis of the non-evaporated organic phase. The spectrum shows exclusive formation of the reduced product, with no detectable signals corresponding to hydroamination product. However, trace ring opened propanal and propyl amine derivatives radical intermediacy under these conditions.

6.6 Amino acid experiment

The reaction was conducted at a concentration of 0.25 M. A two-necked round bottom flask that is sealed with a septum on one end and connected to a Schlenk line on the other is charged with 1 mmol of lithium (6.5 mg) and 1.5 equiv. of DL-proline (172.7 mg), which are dried extensively under vacuum (about 10 minutes), purged, and then filled up with nitrogen. Then 4 mL of tetrahydrofuran (THF)

followed by 1 mmol of 4-methylstyrene (0.13 mL) was added using a syringe and needle through the septum end. The reaction mixture was sonicated for 0.5 h. Afterwards; the mixture was cooled to 0 °C in an ice bath and then quenched with 20 mL of deionized water (D₂O). Extraction was done using 3 x 20 mL of ethyl acetate. The organic phase was then dried over sodium sulfate and evaporated under reduced pressure using a rotary evaporator.

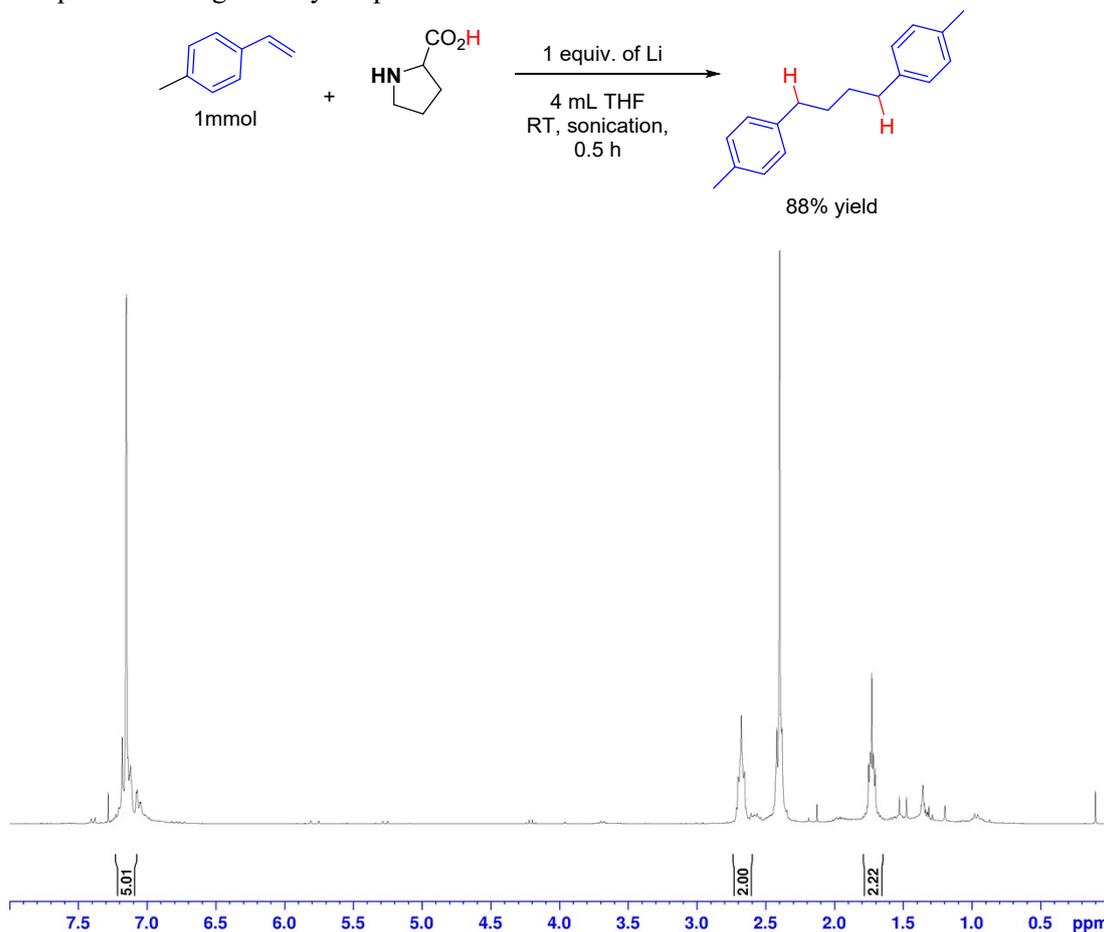
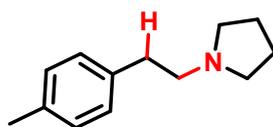


Figure S18. ¹H NMR spectrum of the crude reaction mixture from the reaction of 4-methylstyrene with DL-proline under standard conditions. The reaction expectedly yielded the dimer, 1,4-di-p-tolylbutane in 88% crude yield.

7.0 Characterization data



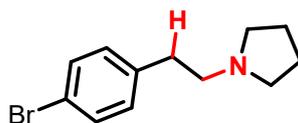
N-(2-(4-methylphenyl)ethyl)pyrrolidine, 5

Colorless oil. Yield = 98% (186 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.13 (s, 4H), 2.91-2.80 (m, 2H), 2.80-2.70 (m, 2H), 2.64 (br s, 4H), 2.36 (s, 3H), 1.85 (quin, $J = 3.4$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 137.2, 135.6, 129.1, 128.5, 58.4, 54.2, 35.1, 23.5, 21.0. The data matched what was reported in literature.³



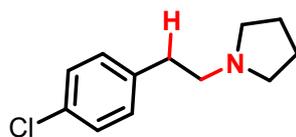
N-(2-(2-methylphenyl)ethyl)pyrrolidine, 6

Colorless oil. Yield = 95% (180 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.20-7.10 (m, 4H), 2.95-2.82 (m, 2H), 2.73-2.55 (m, 6H), 2.34 (s, 3H), 1.86 (quin, $J = 3.4$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 138.6, 136.1, 130.2, 129.2, 126.2, 126.0, 57.1, 54.3, 33.1, 23.5, 19.4. The data matched what was reported in literature.³



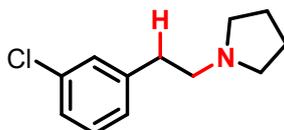
N-(2-(4-bromophenyl)ethyl)pyrrolidine, 7

Colorless oil. Yield = 85% (216 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.49-7.34 (m, 2H), 7.16-6.98 (m, 2H), 2.85-2.73 (m, 2H), 2.72-2.62 (m, 2H), 2.62-2.50 (m, 4H), 1.86-1.75 (m, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 142.9, 134.3, 129.8, 128.8, 127.1, 126.4, 58.2, 54.4, 35.7, 23.7. The data matched what was reported in literature.⁴



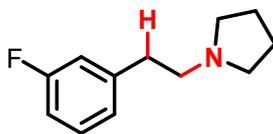
N-(2-(4-chlorophenyl)ethyl)pyrrolidine, 8

Colorless oil. Yield = 89% (187 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.25 (d, $J = 8.5$ Hz, 2H), 7.15 (d, $J = 8.5$ Hz, 2H), 2.87-2.75 (m, 2H), 2.75-2.63 (m, 2H), 2.63-2.51 (m, 4H), 1.88-1.72 (m, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 139.1, 131.7, 130.0, 128.4, 58.1, 54.2, 35.2, 23.5. The data matched what was reported in literature.⁵



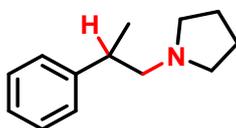
N-(2-(3-chlorophenyl)ethyl)pyrrolidine, 9

Colorless oil. Yield = 79% (166 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.27-7.02 (m, 4H), 2.92-2.76 (m, 2H), 2.77-2.64 (m, 2H), 2.64-2.50 (m, 4H), 1.88-1.74 (m, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 142.9, 134.3, 129.8, 128.8, 127.1, 126.4, 58.2, 54.4, 35.7, 23.7. The data matched what was reported in literature.³



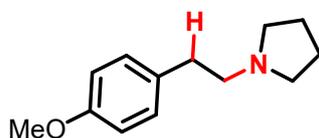
N-(2-(3-fluorophenyl)ethyl)pyrrolidine, 10

Yellow oil. Yield = 83% (160 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.26-7.11 (m, 2H), 6.96-6.79 (m, 2H), 2.84-2.73 (m, 2H), 2.70-2.59 (m, 2H), 2.57-2.46 (m, 4H), 1.81-1.69 (m, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 162.9 (d, $^1J_{\text{CF}} = 245.3$ Hz), 143.2 (d, $^3J_{\text{CF}} = 7.2$ Hz), 129.7 (d, $^3J_{\text{CF}} = 8.4$ Hz), 124.3 (d, $^4J_{\text{CF}} = 2.7$ Hz), 115.5 (d, $^2J_{\text{CF}} = 20.9$ Hz), 112.9 (d, $^2J_{\text{CF}} = 21.1$ Hz), 57.9, 54.3, 35.6, 23.5. The data matched what was reported in literature.⁶



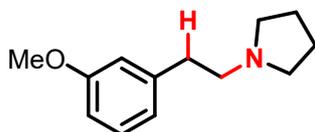
N-(2-phenylpropyl)pyrrolidine, 11

Yellow oil. Yield = 60% (114 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.37-7.20 (m, 5H), 3.02-2.85 (m, 1H), 2.81-2.65 (m, 1H), 2.60-2.36 (m, 5H), 1.76 (br s, 4H), 1.32 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 146.6, 128.3, 127.1, 126.1, 64.5, 54.5, 39.6, 23.5, 20.3. The data matched what was reported in literature.⁷



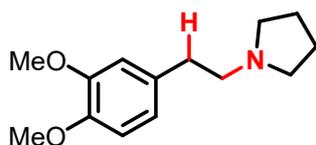
N-(2-(4-methoxyphenyl)ethyl)pyrrolidine, 12

Orange oil. Yield = 80% (164 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.15 (d, $J = 8.6$ Hz, 2H), 6.85 (d, $J = 8.6$ Hz, 2H), 3.81 (s, 3H), 2.86-2.74 (m, 2H), 2.74-2.64 (m, 2H), 2.63-2.53 (m, 4H), 1.83 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 157.9, 132.7, 129.5, 113.8, 58.7, 55.3, 54.3, 35.0, 23.5. The data matched what was reported in literature.⁸



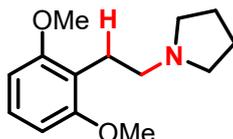
N-(2-(3-methoxyphenyl)ethyl)pyrrolidine, 13

Orange oil. Yield = 58% (119 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.21 (t, $J = 7.8$ Hz, 1H), 6.90-6.66 (m, 3H), 3.80 (s, 3H), 2.89-2.77 (m, 2H), 2.78-2.66 (m, 2H), 2.64-2.51 (m, 4H), 1.82 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 159.7, 142.2, 129.3, 121.0, 114.5, 111.3, 58.2, 55.1, 54.2, 35.9, 23.5. The data matched what was reported in literature.³



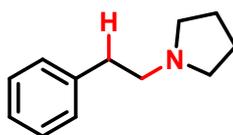
N-(2-(3,4-dimethoxyphenyl)ethyl)pyrrolidine, 14

Colorless oil. Yield = 72% (169 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 6.84-6.69 (m, 3H), 3.87 (s, 3H), 3.85 (s, 3H), 2.84-2.73 (m, 2H), 2.73-2.64 (m, 2H), 2.63-2.51 (m, 4H), 1.81 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 148.9, 147.4, 133.2, 120.5, 112.1, 111.3, 58.5, 55.9, 55.8, 54.2, 35.4, 23.5. The data matched what was reported in literature.⁹



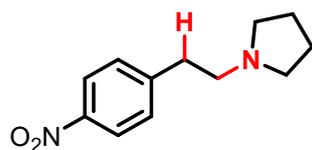
N-(2-(2,6-dimethoxyphenyl)ethyl)pyrrolidine, 15

Colorless oil. Yield = 64% (151 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.14 (t, $J = 8.3$ Hz, 1H), 6.55 (d, $J = 8.3$ Hz, 2H), 3.82 (s, 6H), 2.98-2.87 (m, 2H), 2.70-2.49 (m, 6H), 1.82 (quin, $J = 3.4$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 158.5, 126.9, 117.0, 103.7, 55.7, 55.4, 54.1, 23.5, 22.8. HRMS (ESI) calcd. for $\text{C}_{14}\text{H}_{21}\text{NO}_2$ $[\text{M}+\text{H}]^+$: 236.1645, found: 236.1675.



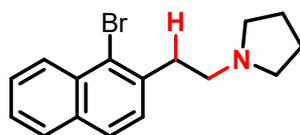
N-(2-phenylethyl)pyrrolidine, 16

Colorless oil. Yield = 96% (168 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.31-7.18 (m, 5H), 2.93-2.82 (m, 2H), 2.78-2.68 (m, 2H), 2.64-2.51 (m, 4H), 1.88-1.75 (m, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 140.5, 128.7, 128.4, 126.0, 58.4, 54.2, 35.9, 23.6. The data matched what was reported in literature.¹⁰



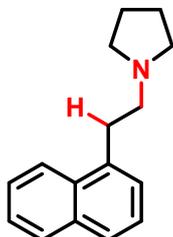
N-(2-(4-nitrophenyl)ethyl)pyrrolidine, 17

Colorless oil. Yield = 92% (203 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 8.19-8.06 (m, 2H), 7.36 (d, 2H), 2.98-2.86 (m, 2H), 2.77-2.69 (m, 2H), 2.63-2.47 (m, 4H), 1.79 (quin, $J = 3.4$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 148.5, 146.5, 129.4, 123.6, 57.3, 54.2, 35.6, 23.5. The data matched what was reported in literature.¹¹



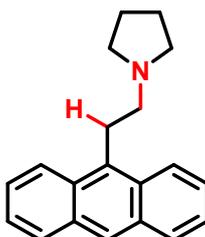
N-(2-(1-bromo-2-naphthalenyl)ethyl)pyrrolidine, 18

White solid. Yield = 75% (228 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 8.34 (d, $J = 8.5$ Hz, 1H), 7.82 (d, $J = 7.8$ Hz, 1H), 7.77 (d, $J = 8.4$ Hz, 1H) 7.65-7.55 (m, 1H), 7.55-7.45 (m, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 3.40-3.10 (m, 2H), 2.89-2.60 (m, 6H), 1.94-1.79 (br. s, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 137.9, 133.3, 132.6, 128.3, 128.0, 127.7, 127.34, 127.30, 125.9, 123.9, 56.2, 54.2, 37.0, 23.6. HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{18}^{79}\text{BrN}$ $[\text{M}+\text{H}]^+$: 304.0695, found: 304.0685



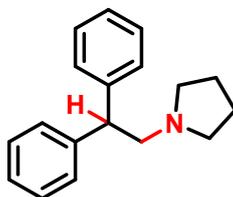
N-(2-(1-naphthalenyl)ethyl)pyrrolidine, 19

Colorless oil. Yield = 80% (180 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 8.14 (d, $J = 8.3$ Hz, 1H), 7.88 (d, $J = 7.7$ Hz, 1H), 7.76 (d, $J = 7.80$ Hz, 1H), 7.61-7.38 (m, 4H), 3.44-3.28 (m, 2H), 2.91-2.81 (m, 2H), 2.74-2.64 (m, 4H), 1.89 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 136.7, 133.9, 132.0, 128.8, 126.9, 126.5, 125.9, 125.6, 125.5, 123.8, 57.7, 54.3, 33.0, 23.6. HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{19}\text{N}$ $[\text{M}+\text{H}]^+$: 226.1590, found: 226.1591



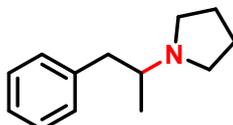
N-(2-(anthracene-9-yl)ethyl)pyrrolidine, 20

Colorless oil Yield = 59% (163 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 8.44-8.26 (m, 3H), 8.08-7.96 (m, 2H), 7.59-7.44 (m, 4H), 4.00-3.76 (m, 2H), 2.97-2.83 (m, 2H), 2.85-2.63 (m, 4H), 1.94 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 132.3, 131.7, 129.8, 129.3, 126.1, 125.7, 124.9, 124.3, 56.9, 54.3, 27.9, 23.6. The data matched what was reported in literature.³



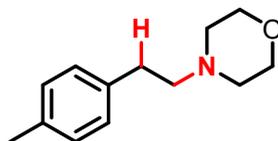
N-(2,2-diphenylethyl)pyrrolidine, 21

Colorless oil. Yield = 26% (65 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 8.19-8.06 (m, 2H), 7.39-7.17 (m, 10H), 4.24 (t, $J = 7.5$ Hz, 1H), 3.17 (d, $J = 7.5$ Hz, 2H), 2.63-2.44 (m, 4H), 1.75 (quin, $J = 3.3$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 144.2, 128.4, 128.1, 126.2, 61.9, 54.6, 51.1, 23.6. The data matched what was reported in literature.¹²



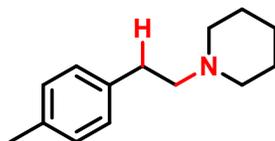
N-(1-methyl-2-phenylethyl)pyrrolidine, 22

Yellow oil. Yield = 10% (19 mg, from cis-beta-methylstyrene), 78% (148 mg, from allylbenzene). ^1H NMR (CDCl_3 , 400 MHz) δ 7.12 (m, 5H), 3.16 (dd, $J = 12.7$ Hz, 3.3 Hz, 1H), 2.82-2.63 (m, 4H), 2.64-2.49 (m, 1H), 2.52-2.35 (m, 1H), 1.84 (quin, $J = 3.3$ Hz, 4H), 1.01 (d, $J = 6.2$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 140.3, 129.4, 128.2, 125.9, 61.0, 51.4, 42.0, 23.5, 17.7. The data matched what was reported in literature.¹³



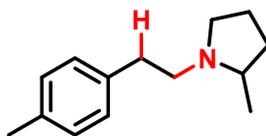
N-(4-methylphenethyl)morpholine, 23

Yellow oil. Yield = 97% (199 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.13 (br s, 4H), 3.78 (t, $J = 4.7$ Hz, 4H), 2.86-2.71 (m, 2H), 2.65-2.50 (m, 6H), 2.35 (s, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 137.1, 135.6, 129.1, 128.6, 67.0, 61.0, 53.8, 32.9, 21.0. The data matched what was reported in literature.¹⁴



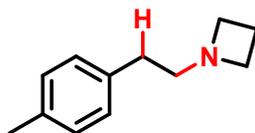
N-(4-methylphenethyl)piperidine, 24

Yellow oil. Yield = 93% (189 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.12 (br s, 4H), 2.85-2.77 (m, 2H), 2.61-2.43 (m, 6H), 2.35 (s, 3H), 1.72-1.59 (m, 4H), 1.54-1.43 (m, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 137.6, 135.3, 129.1, 128.6, 61.6, 54.6, 33.3, 26.1, 24.5, 21.0. The data matched what was reported in literature.¹⁵



N-(4-methylphenethyl)-2-methylpyrrolidine, 25

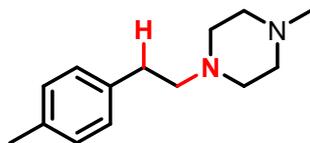
Yellow oil. Yield = 70% (142 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.13 (br s, 4H), 3.34-3.20 (m, 1H), 3.11-2.96 (m, 1H), 2.91-2.71 (m, 2H), 2.38-2.16 (m, 6H), 2.04-1.64 (m, 3H), 1.57-1.40 (m, 1H), 1.14 (d, $J = 6.1$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 137.6, 135.4, 129.1, 128.5, 60.1, 56.3, 54.0, 35.0, 32.7, 21.7, 21.0, 18.9. HRMS (ESI) calcd. for $\text{C}_{14}\text{H}_{21}\text{N}$ $[\text{M}+\text{H}]^+$: 204.1747, found: 204.1765



N-(4-methylphenethyl)azetidine, 26

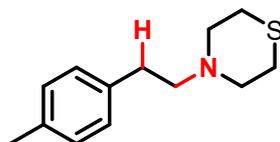
Yellow oil. Yield = 73% (128 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.13 (br s, 4H), 3.21 (t, $J = 7.0$ Hz, 4H), 2.62 (br s, 4H), 2.33 (s, 3H), 2.08 (quin, $J = 7.0$ Hz, 2H); ^{13}C NMR (CDCl_3 , 100

MHz) δ 137.1, 135.4, 129.0, 128.5, 61.8, 55.3, 34.0, 21.0, 17.8. HRMS (ESI) calcd. for $C_{12}H_{17}N$ $[M+H]^+$: 176.1434, found: 176.1415



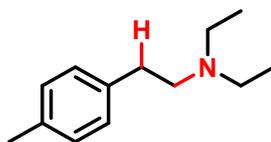
N¹-(4-methylphenethyl)-N²-methylpiperazine, 27

Yellow oil. Yield = 91% (199 mg). ¹H NMR (CDCl₃, 400 MHz) δ 7.11 (br s, 4H), 2.85-2.73 (m, 2H), 2.70-2.38 (m, 10H), 2.33 (s, 3H), 2.32 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.3, 135.5, 129.1, 128.6, 60.7, 55.2, 53.2, 46.1, 33.2, 21.0. The data matched what was reported in literature.¹⁴



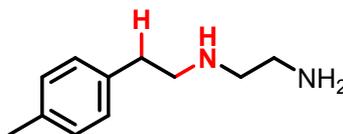
N-(4-methylphenethyl)thiomorpholine, 28

Yellow oil. Yield = 85% (188 mg). ¹H NMR (CDCl₃, 400 MHz) δ 7.13 (br s, 4H), 2.87-2.71 (m, 10H), 2.70-2.61 (m, 2H), 2.36 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.2, 135.5, 129.1, 128.6, 61.4, 55.0, 32.7, 28.1, 21.1. HRMS (ESI) calcd. for $C_{13}H_{19}NS$ $[M+H]^+$: 222.1311, found: 222.1308



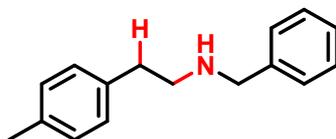
N-(4-methylphenethyl)diethylamine, 29

Yellow oil Yield = 54% (103 mg). ¹H NMR (CDCl₃, 400 MHz) δ 7.12 (br s, 4H), 2.78-2.68 (m, 4H), 2.64 (q, J = 7.2 Hz, 4H), 2.35 (s, 3H), 1.09 (t, J = 7.2 Hz, 6H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.7, 135.4, 129.1, 128.6, 55.1, 46.9, 32.9, 21.0, 11.8. The data matched what was reported in literature.¹⁶



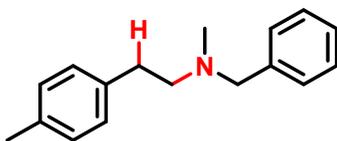
N-(4-methylphenethyl)ethylenediamine, 30

Yellow oil. Yield = 62% (111 mg). ¹H NMR (CDCl₃, 400 MHz) δ 7.12 (br s, 4H), 2.89 (t, J = 6.8 Hz, 2H), 2.79 (q, J = 5.8 Hz, 4H), 2.70 (t, J = 5.9 Hz, 2H), 2.34 (s, 3H), 1.43 (br s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.0, 135.6, 129.2, 128.6, 52.5, 51.2, 41.8, 36.0, 21.0. HRMS (ESI) calcd. for $C_{11}H_{18}N_2$ $[M + CO + H]^+$: 207.1492, found: 207.1486



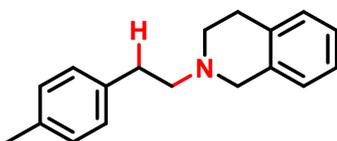
N-(4-methylphenethyl)benzylamine, 31

Yellow oil. Yield = 26% (59 mg). $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.37-7.23 (m, 5H), 7.12 (br s, 4H), 3.84 (s, 2H), 3.00-2.75 (m, 4H), 2.34 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 139.9, 136.7, 135.7, 129.2, 128.7, 128.4, 128.2, 127.0, 53.7, 50.5, 35.7, 21.0. The data matched what was reported in literature.⁵



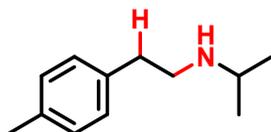
N-(4-methylphenethyl)-N-methylbenzylamine, 32

Yellow oil. Yield = 68% (163 mg). $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.38-7.27 (m, 5H), 7.14 (br s, 4H), 3.61 (s, 2H), 2.91-2.81 (m, 2H), 2.74-2.65 (m, 2H), 2.37 (s, 3H), 2.33 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 139.1, 137.5, 135.4, 129.07, 129.05, 128.7, 128.3, 127.0, 62.2, 59.4, 42.2, 33.5, 21.1. The data matched what was reported in literature.³



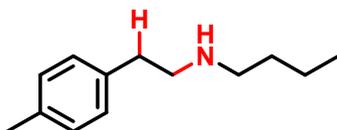
N-(4-methylphenethyl)-1,2,3,4-tetrahydroisoquinoline, 33

Yellow oil. Yield = 40% (101 mg). $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.25-7.14 (m, 7H), 7.14-7.04 (m, 1H), 3.78 (s, 2H), 3.04-2.91 (m, 4H), 2.91-2.77 (m, 4H), 2.39 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 137.3, 135.6, 134.8, 134.3, 129.2, 128.71, 128.67, 126.7, 126.2, 125.7, 60.5, 56.2, 51.1, 33.6, 29.2, 21.1. HRMS (ESI) calcd. for $\text{C}_{18}\text{H}_{21}\text{N}$ $[\text{M} + \text{H}]^+$: 252.1747, found: 252.1739



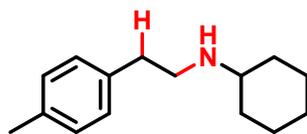
N-(4-methylphenethyl)isopropylamine, 34

Yellow oil. Yield = 27% (48 mg). $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.13 (br s, 4H), 2.96-2.79 (m, 5H), 2.50 (br s, 1H), 2.34 (s, 3H), 1.11 (d, $J = 6.3$ Hz, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 136.6, 135.7, 129.2, 128.6, 48.7, 48.6, 35.6, 22.5, 21.0. HRMS (ESI) calcd. for $\text{C}_{12}\text{H}_{19}\text{N}$ $[\text{M} + \text{H}]^+$: 178.1590, found: 178.1563



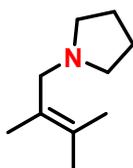
N-(4-methylphenethyl)butylamine, 35

Yellow oil. Yield = 52% (100 mg). $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.12 (br s, 4H), 2.96-2.77 (m, 4H), 2.66 (t, $J = 7.3$, 2H), 2.34 (s, 3H), 1.57-1.43 (m, 2H), 1.38-1.27 (m, 3H), 0.92 (t, $J = 7.2$ Hz, 6H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 136.8, 135.7, 129.2, 128.6, 51.1, 49.4, 35.6, 31.9, 21.0, 20.5, 14.0. HRMS (ESI) calcd. for $\text{C}_{13}\text{H}_{21}\text{N}$ $[\text{M} + \text{H}]^+$: 192.1747, found: 192.1741



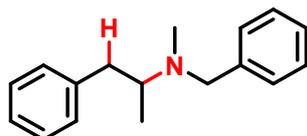
N-(4-methylphenethyl)cyclohexylamine, 36

Yellow oil. Yield = 27% (59 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.12 (br s, 4H), 2.98-2.89 (m, 2H), 2.89-2.80 (m, 2H), 2.58-2.46 (m, 1H), 2.34 (s, 3H), 1.93 (d, $J = 11.9$ Hz, 2H), 1.81-1.70 (m, 2H), 1.64 (d, $J = 11.4$ Hz, 1H), 1.34-1.08 (m, 6H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 136.6, 135.7, 129.2, 128.6, 56.8, 48.0, 35.6, 32.9, 26.0, 25.0, 21.0. HRMS (ESI) calcd. for $\text{C}_{15}\text{H}_{23}\text{N}$ [$\text{M} + \text{H}$] $^+$: 218.1909, found: 218.1899



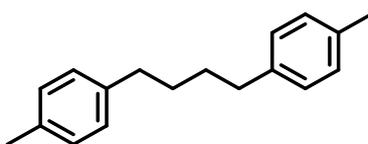
1-(2,3-Dimethyl-2-buten-1-yl)pyrrolidine, 37

Colorless oil. Yield = 46% (71 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 3.06 (s, 2H), 2.55-2.40 (m, 4H), 1.81-1.65 (m, 13 H). The data matched what was reported in literature.¹⁷



Benzphetamine, 38

White solid. Yield = 64% (153 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.43-7.19 (m, 10H), 3.69 (q, $J = 11.3$ Hz, 2H), 3.16-3.01 (m, 2H), 2.58 (q, $J = 8.3$ Hz, 1H), 2.33 (s, 3H), 1.09 (d, $J = 6.4$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 140.9, 140.2, 129.3, 129.1, 128.7, 128.2, 126.8, 125.8, 59.9, 57.8, 39.5, 36.9, 14.1. The data matched what was reported in literature.¹⁸



1,4-di-p-tolylbutane, 39

White solid. Yield = 88% (210 mg). ^1H NMR (CDCl_3 , 400 MHz) δ 7.16-7.05 (m, 8H), 2.62 (t, $J = 6.9$ Hz, 4H), 2.34 (s, 6H), 1.67 (quin, $J = 3.7$ Hz, 4H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 139.6, 135.0, 128.9, 128.3, 35.4, 31.3, 21.0. The data matched what was reported in literature.¹⁹

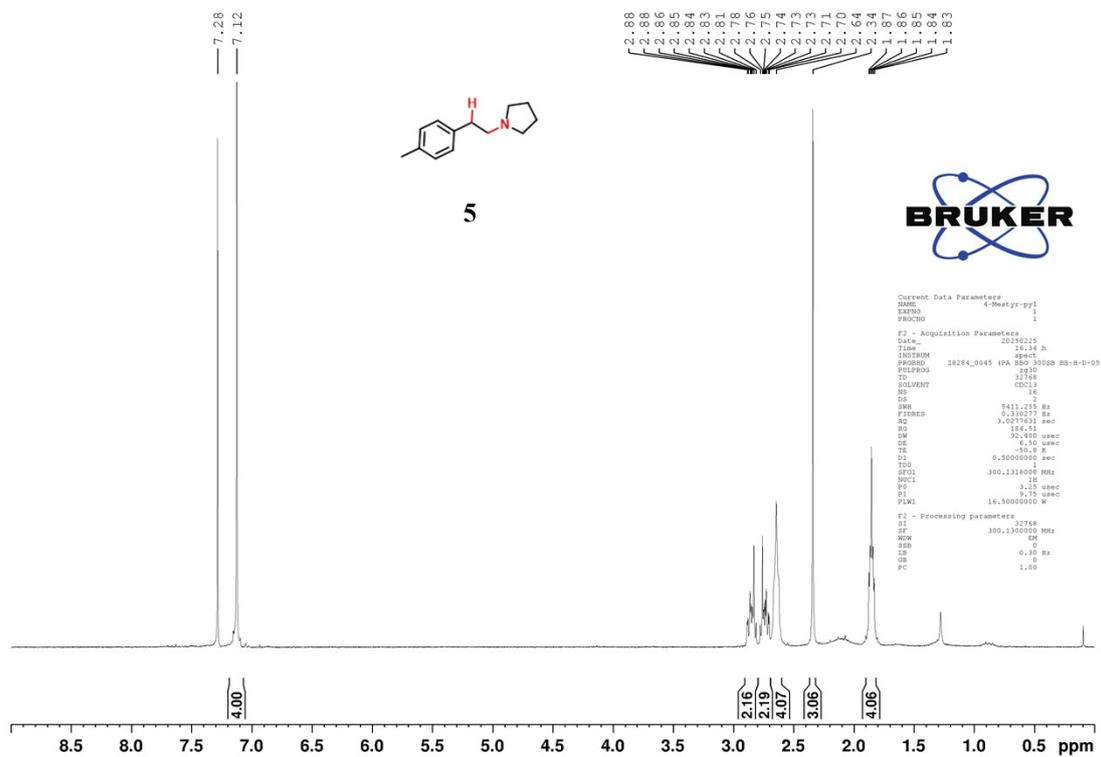
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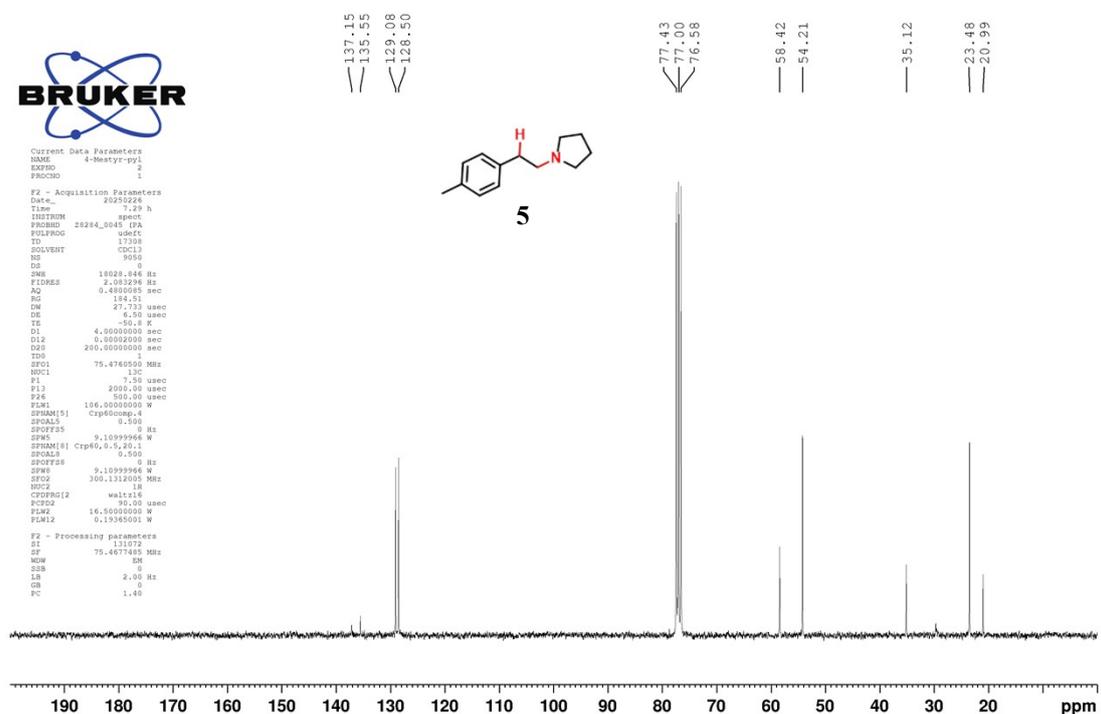
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¹H and ¹³C NMR Spectra

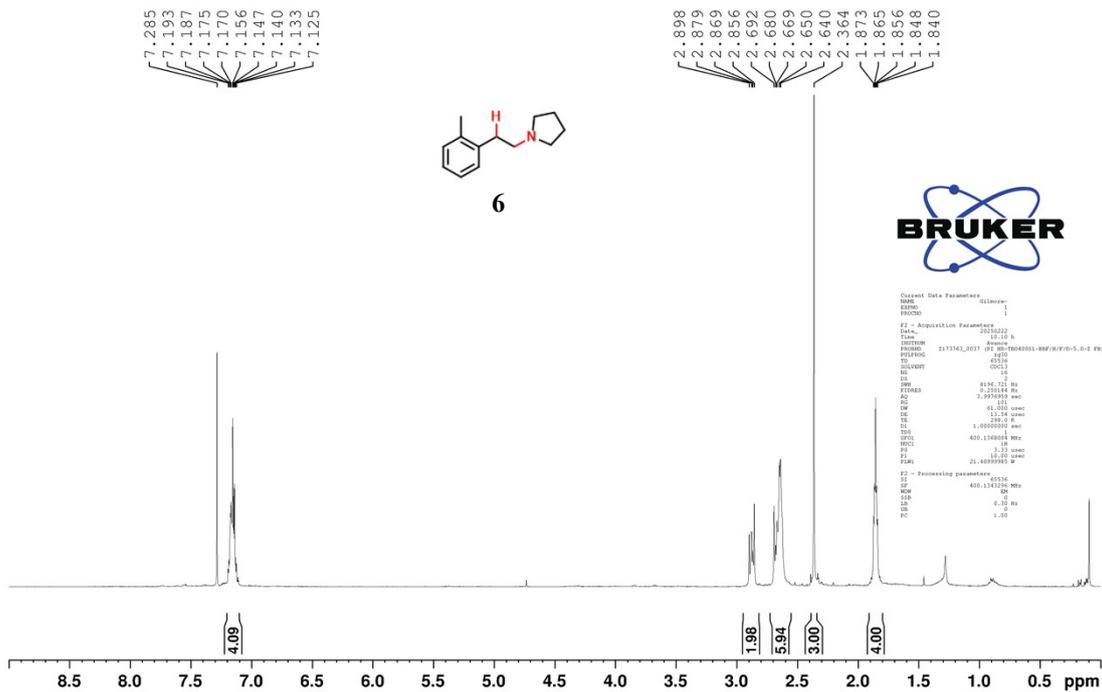
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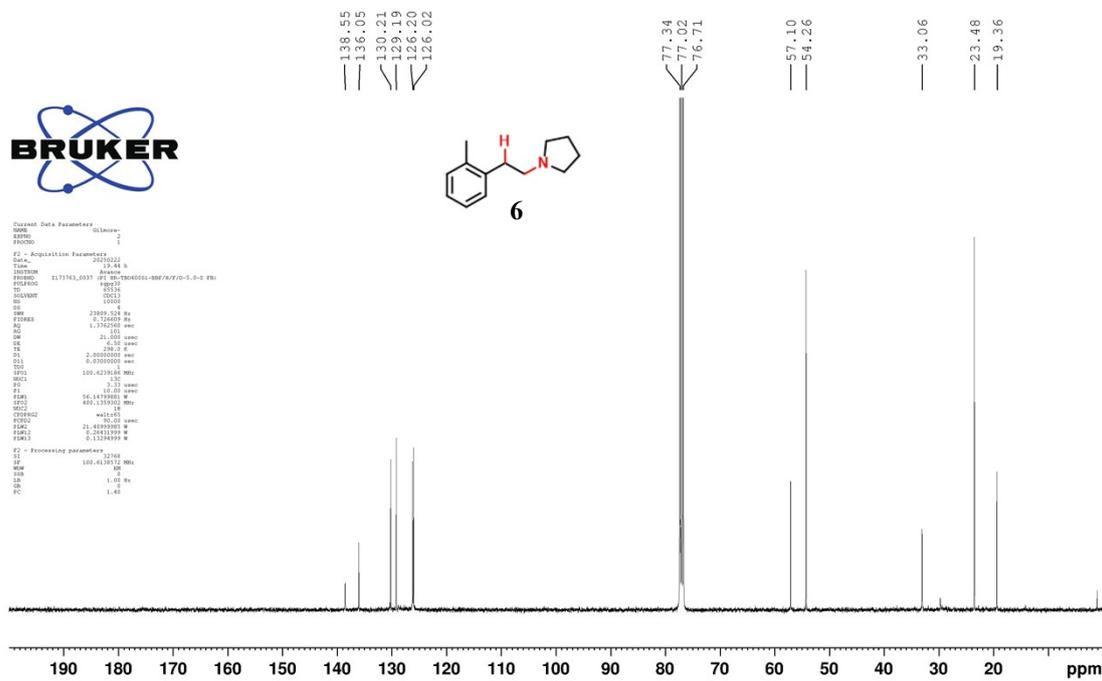
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N-(2-(2-methylphenyl)ethyl)pyrrolidine

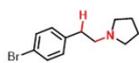


N-(2-(2-methylphenyl)ethyl)pyrrolidine



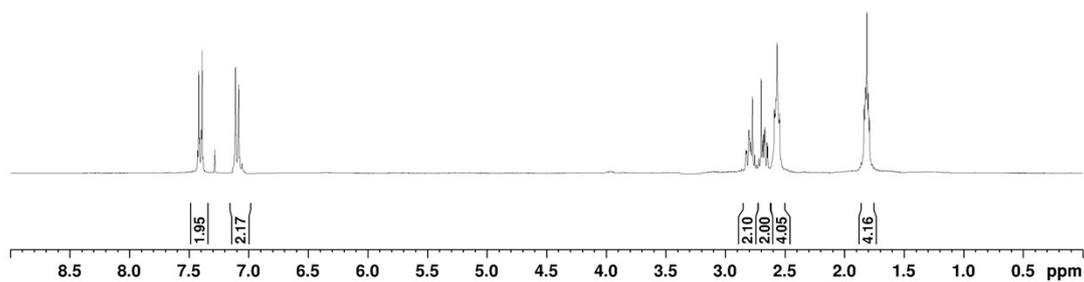
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7.11
7.08



7

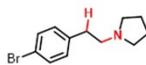
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2.58
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1.82
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1.80
1.79



N-(2-(4-bromophenyl)ethyl)pyrrolidine

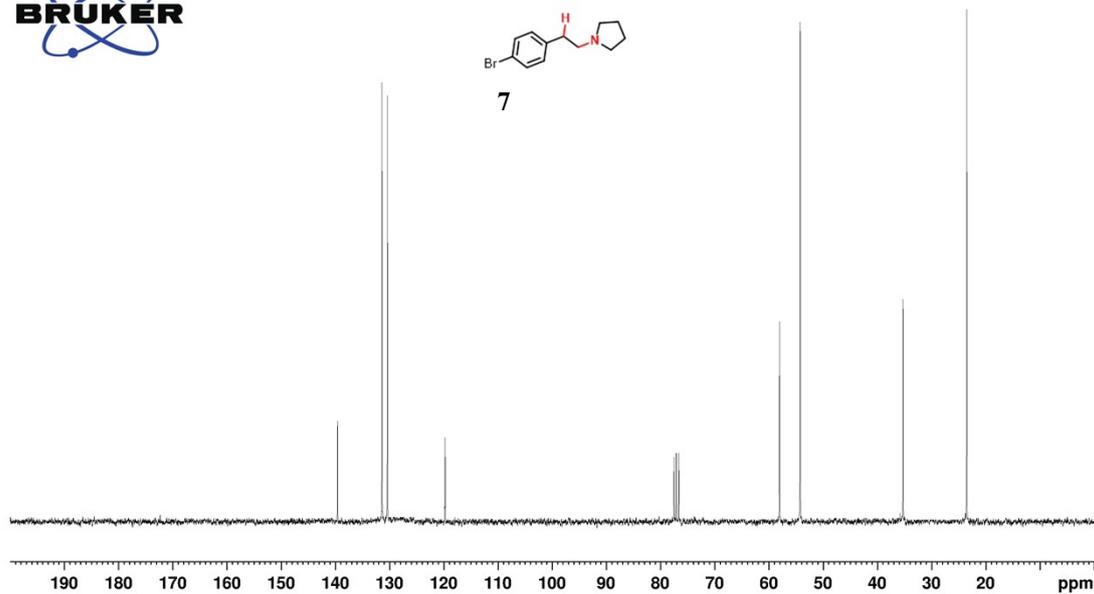


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7

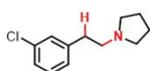
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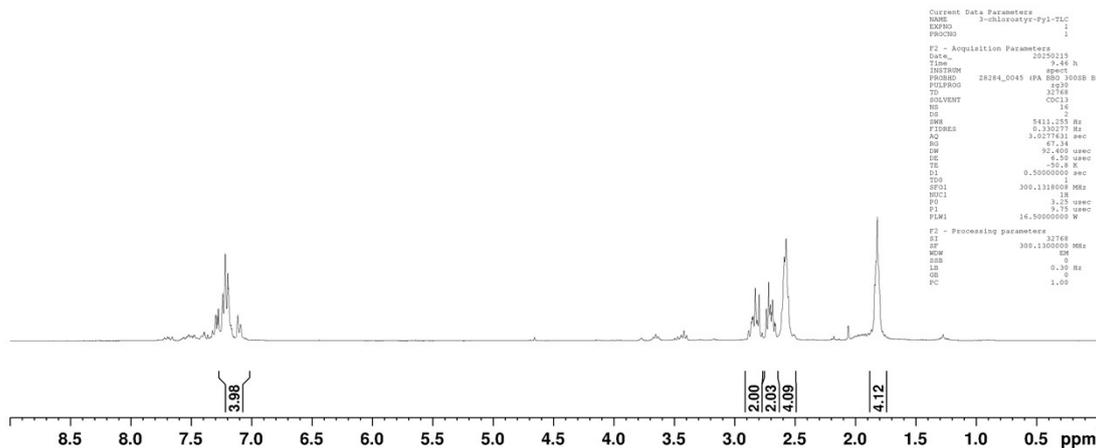
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7.09

2.66
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2.52



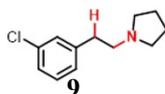
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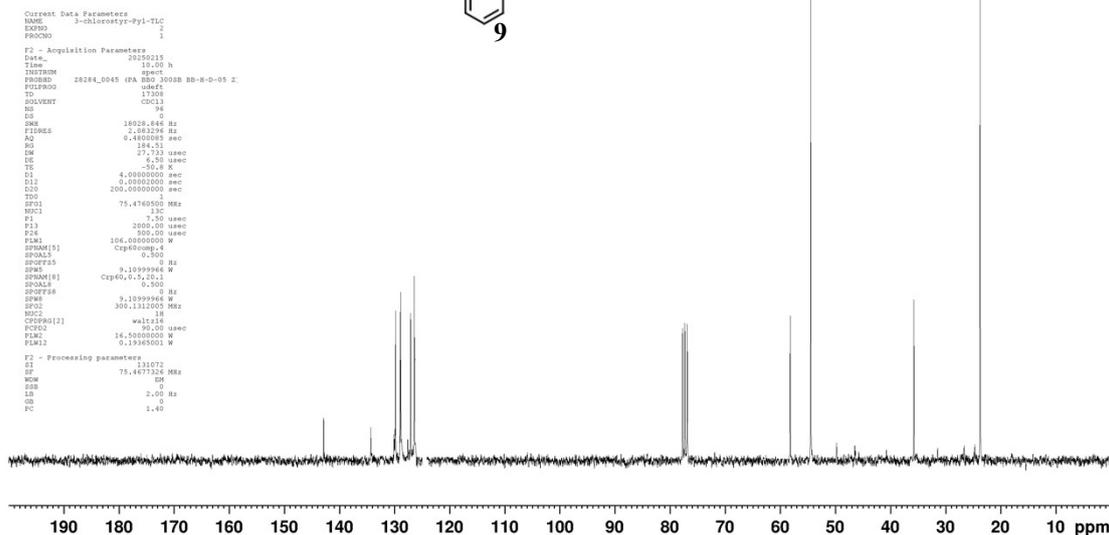
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142.84
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23.70



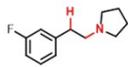
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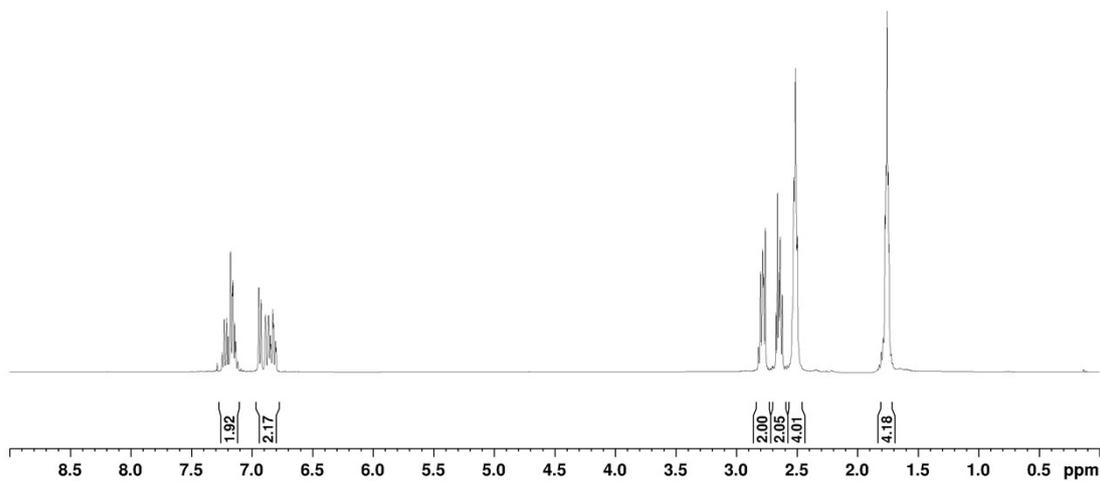
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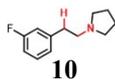


10



N-(2-(3-fluorophenyl)ethyl)pyrrolidine

164.12
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129.68
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115.37
112.56
112.75
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23.50



10

Current Data Parameters

NAME N-(2-(3-fluorophenyl)ethyl)pyrrolidine

EXPNO 101

PROCNO 1

F2 - Acquisition Parameters

Date_ 2023100

Time 01:19 h

INSTRUM spect

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PULPROG zgpg30

TD 32768

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AQ 0.360804 sec

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SE 20.400 usec

DE 6.50 usec

TE 300.2 K

D1 4.0000000 sec

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TD0

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MU1 1.0

F1 50.40 usec

F12 2000.00 usec

F14 800.00 usec

PL1 50.0029935 W

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PL14 11.9945989 W

PL16 0.5559001 W

PL18 0.5559001 W

PL20 0.5559001 W

PL22 0.5559001 W

PL24 0.5559001 W

PL26 0.5559001 W

PL28 0.5559001 W

PL30 0.5559001 W

PL32 0.5559001 W

PL34 0.5559001 W

PL36 0.5559001 W

PL38 0.5559001 W

PL40 0.5559001 W

PL42 0.5559001 W

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PL46 0.5559001 W

PL48 0.5559001 W

PL50 0.5559001 W

PL52 0.5559001 W

PL54 0.5559001 W

PL56 0.5559001 W

PL58 0.5559001 W

PL60 0.5559001 W

PL62 0.5559001 W

PL64 0.5559001 W

PL66 0.5559001 W

PL68 0.5559001 W

PL70 0.5559001 W

PL72 0.5559001 W

PL74 0.5559001 W

PL76 0.5559001 W

PL78 0.5559001 W

PL80 0.5559001 W

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PL88 0.5559001 W

PL90 0.5559001 W

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PL96 0.5559001 W

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PL100 0.5559001 W

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PL106 0.5559001 W

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PL126 0.5559001 W

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PL130 0.5559001 W

PL132 0.5559001 W

PL134 0.5559001 W

PL136 0.5559001 W

PL138 0.5559001 W

PL140 0.5559001 W

PL142 0.5559001 W

PL144 0.5559001 W

PL146 0.5559001 W

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PL150 0.5559001 W

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PL154 0.5559001 W

PL156 0.5559001 W

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PL160 0.5559001 W

PL162 0.5559001 W

PL164 0.5559001 W

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PL170 0.5559001 W

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PL176 0.5559001 W

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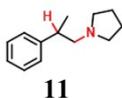
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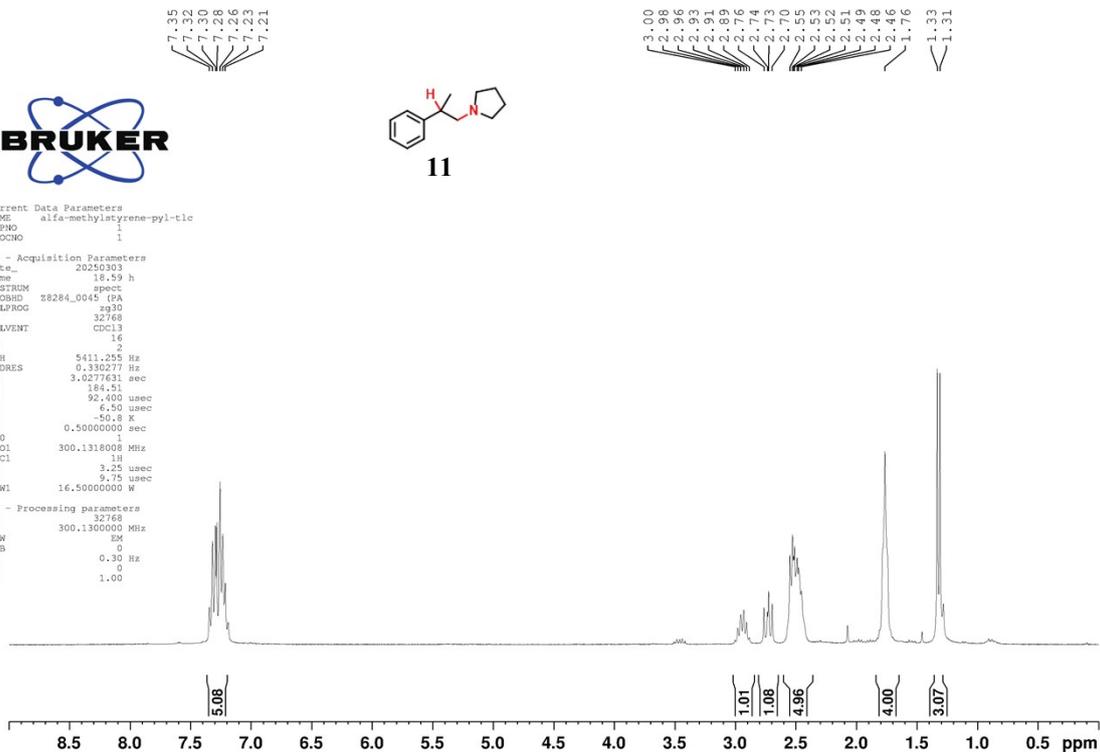
N-(2-phenylpropyl)pyrrolidine



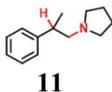
Current Data Parameters
 NAME alfa-methylstyrene-pyl-tlc
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20250303
 Time 18:19 h
 INSTRUM spect
 PROBHD 28284_0045 (PA)
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 5411.255 Hz
 FIDRES 0.330277 Hz
 AQ 3.0277631 sec
 RG 184.51
 DW 92.400 usec
 DE 6.50 usec
 TE -50.8 K
 D1 0.50000000 sec
 TDO 1
 SFO1 300.1318008 MHz
 NUC1 1H
 FO 312.5 usec
 P1 9.75 usec
 PLW1 16.5000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



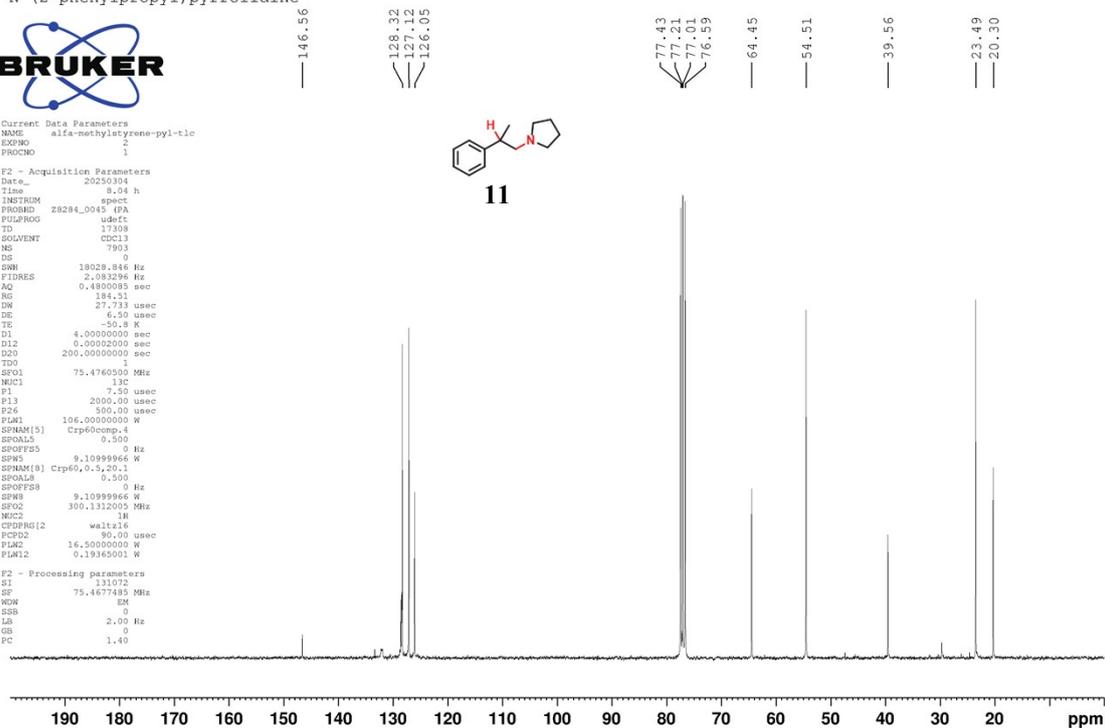
N-(2-phenylpropyl)pyrrolidine



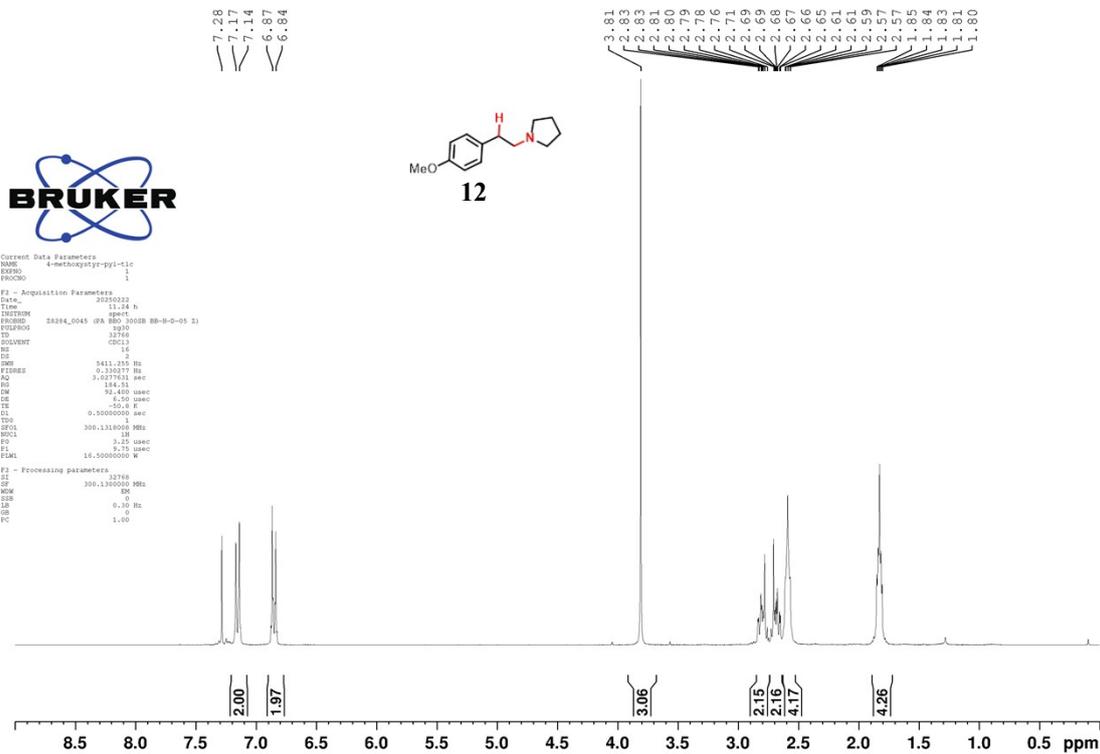
Current Data Parameters
 NAME alfa-methylstyrene-pyl-tlc
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20250304
 Time 0:04 h
 INSTRUM spect
 PROBHD 28284_0045 (PA)
 PULPROG waltz16
 TD 17308
 SOLVENT CDCl3
 NS 7903
 DS 0
 SWH 18028.846 Hz
 FIDRES 2.083296 Hz
 AQ 0.4800085 sec
 RG 184.51
 DW 27.753 usec
 DE 6.50 usec
 TE -50.8 K
 D1 4.00000000 sec
 D12 0.00002000 sec
 D20 200.00000000 sec
 TDO 1
 SFO1 75.4760500 MHz
 NUC1 13C
 P1 7.50 usec
 P13 2000.00 usec
 P26 300.00 usec
 PLW1 106.00000000 W
 SPNAM(5) Crp60comp_4
 SPOAL5 0.500
 SPOFFS5 0 Hz
 SPW5 9.10999966 W
 SPNAM(8) Crp60_0.5_20.1
 SPOAL8 0.500
 SPOFFS8 0 Hz
 SPW8 9.10999966 W
 SFO2 300.1312005 MHz
 NUC2 1H
 CPDPRG12 waltz16
 PCPD2 90.00 usec
 PLW2 16.50000000 W
 PLW12 0.19365001 W

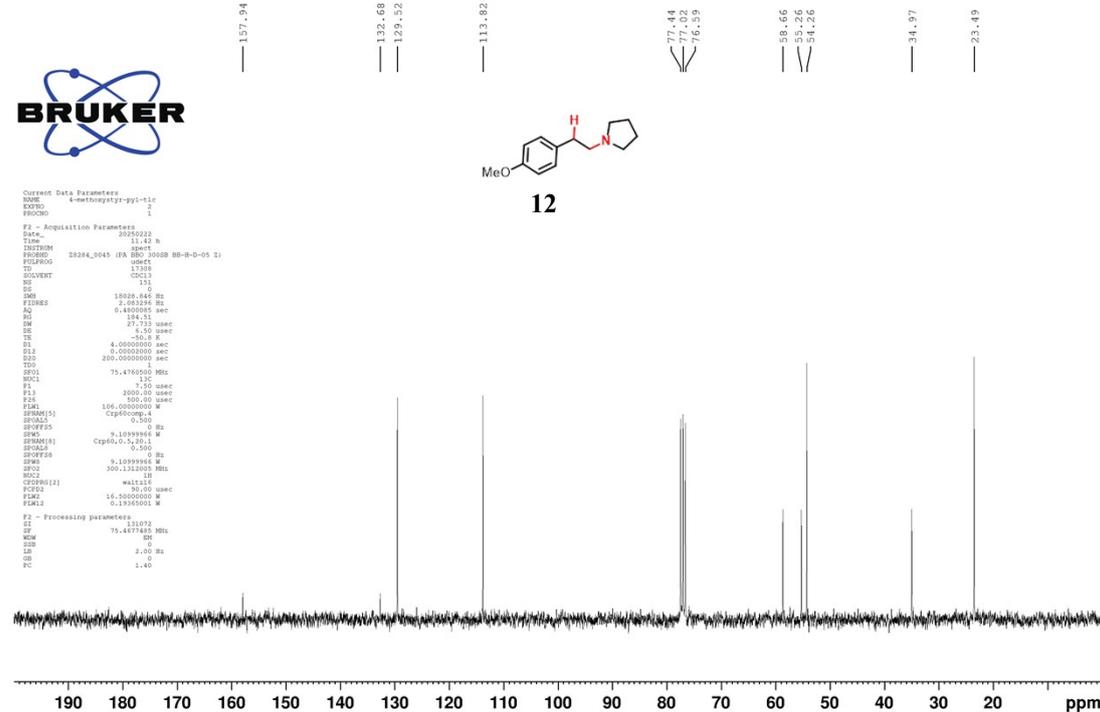
F2 - Processing parameters
 SI 131072
 SF 75.4677485 MHz
 WDW EM
 SSB 0
 LB 2.00 Hz
 GB 0
 PC 1.40



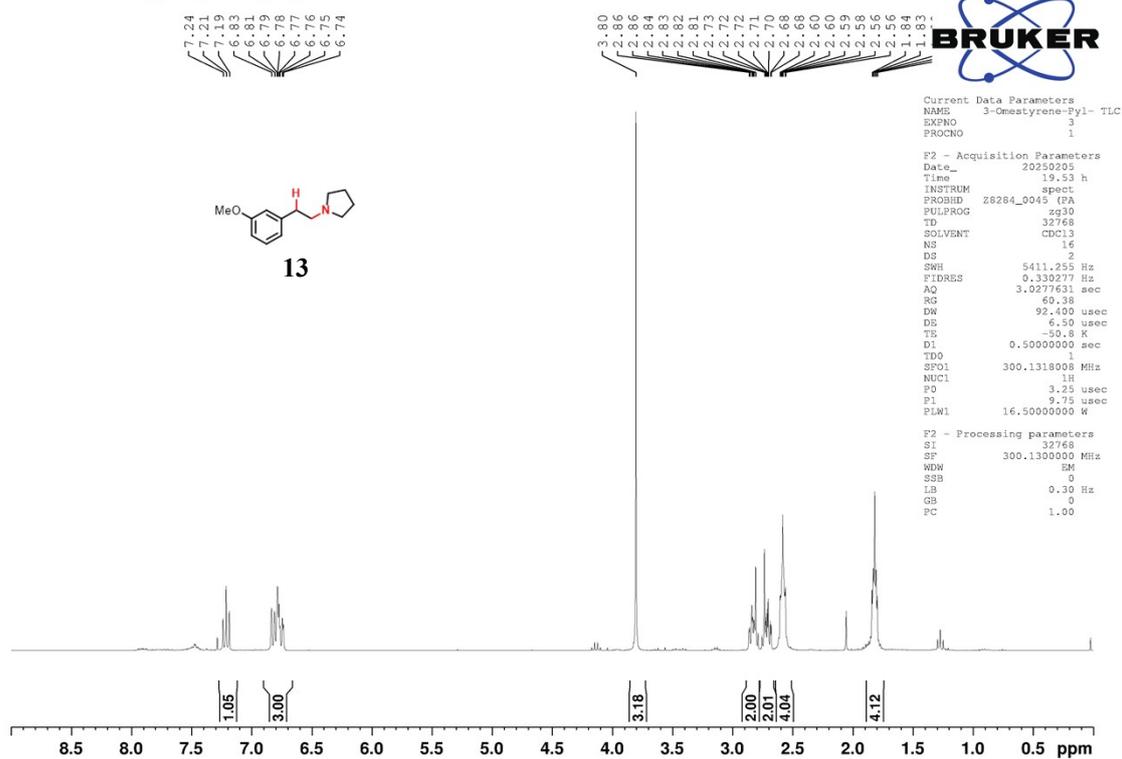
N-(2-(4-methoxyphenyl)ethyl)pyrrolidine



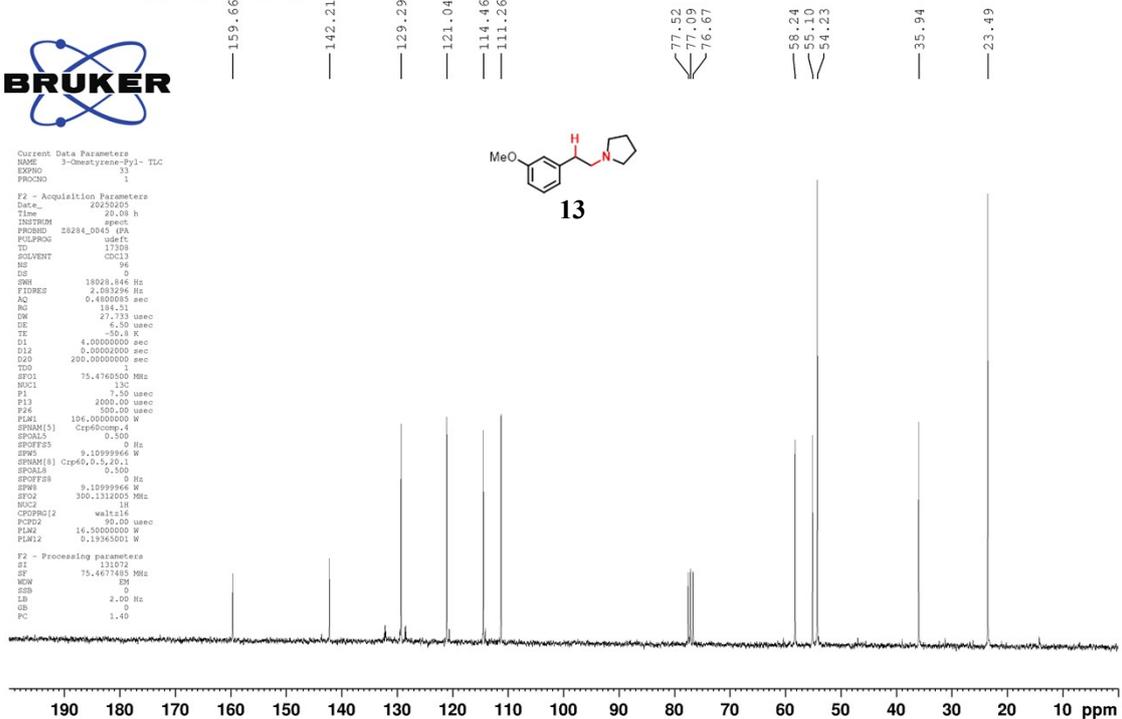
N-(2-(4-methoxyphenyl)ethyl)pyrrolidine



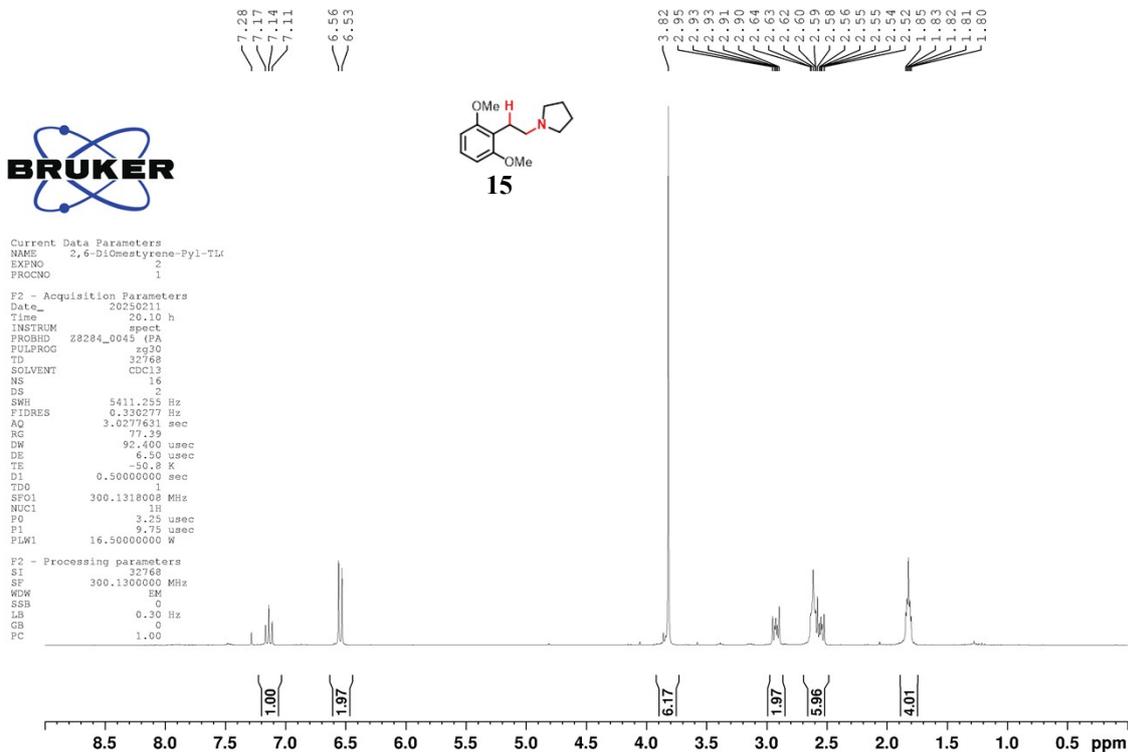
N-(2-(3-methoxyphenyl)ethyl)pyrrolidine



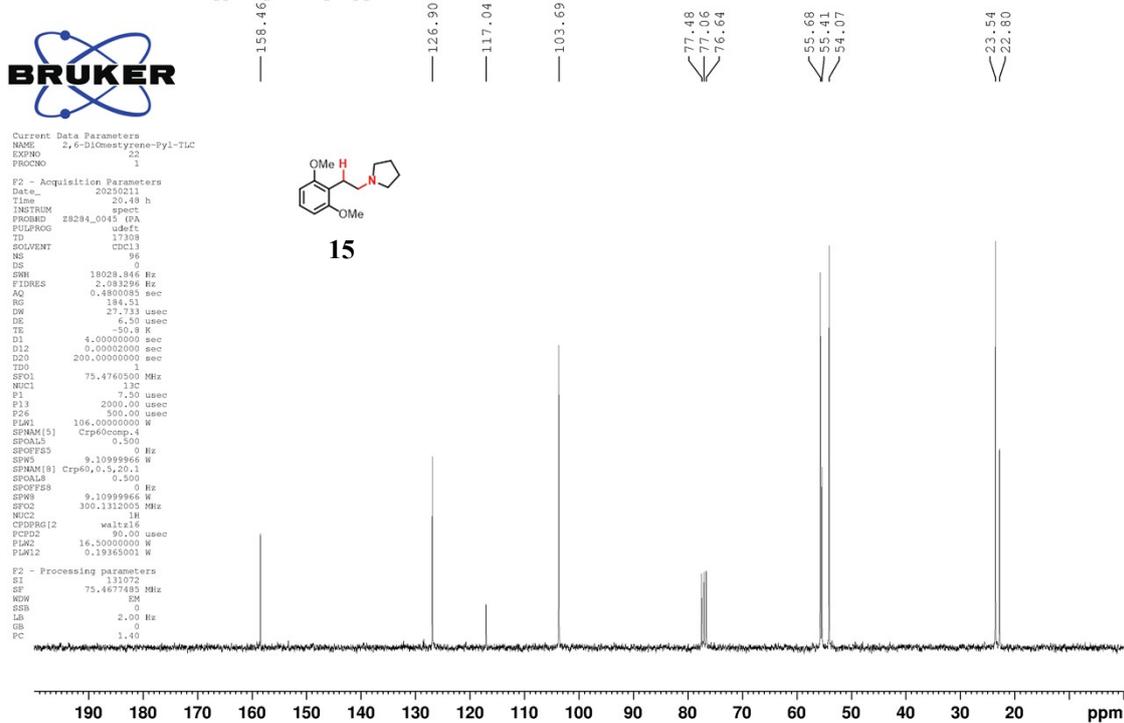
N-(2-(3-methoxyphenyl)ethyl)pyrrolidine



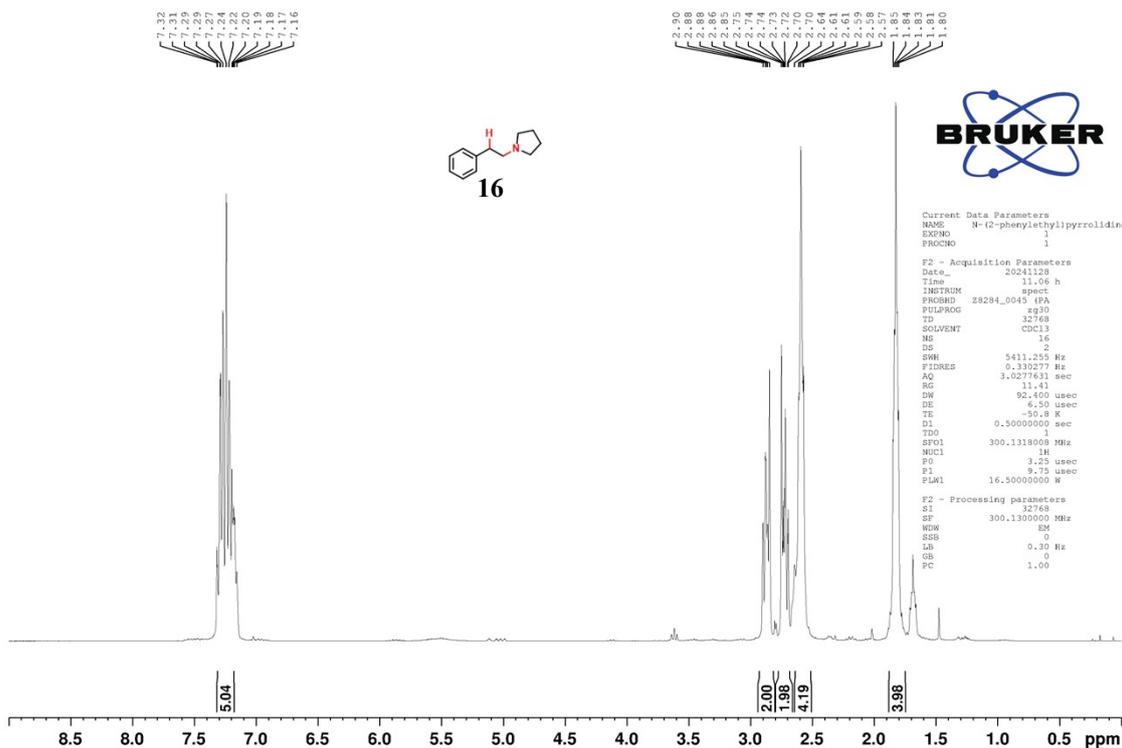
N-(2-(2,6-dimethoxyphenyl)ethyl)pyrrolidine



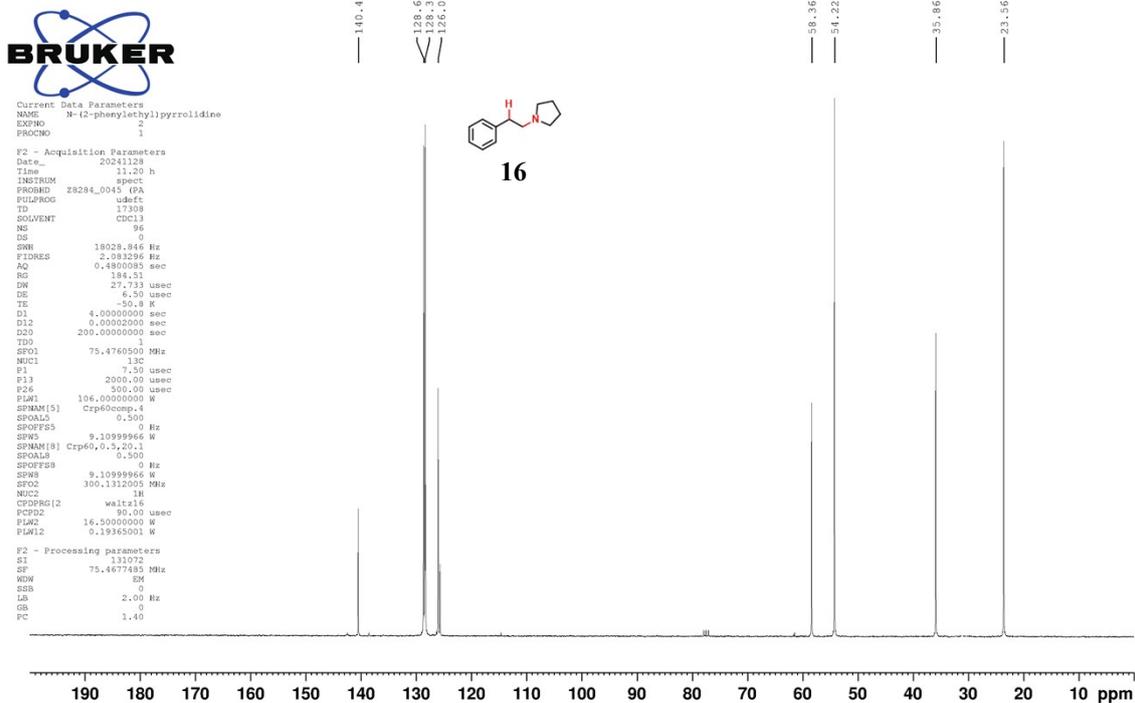
N-(2-(2,6-dimethoxyphenyl)ethyl)pyrrolidine



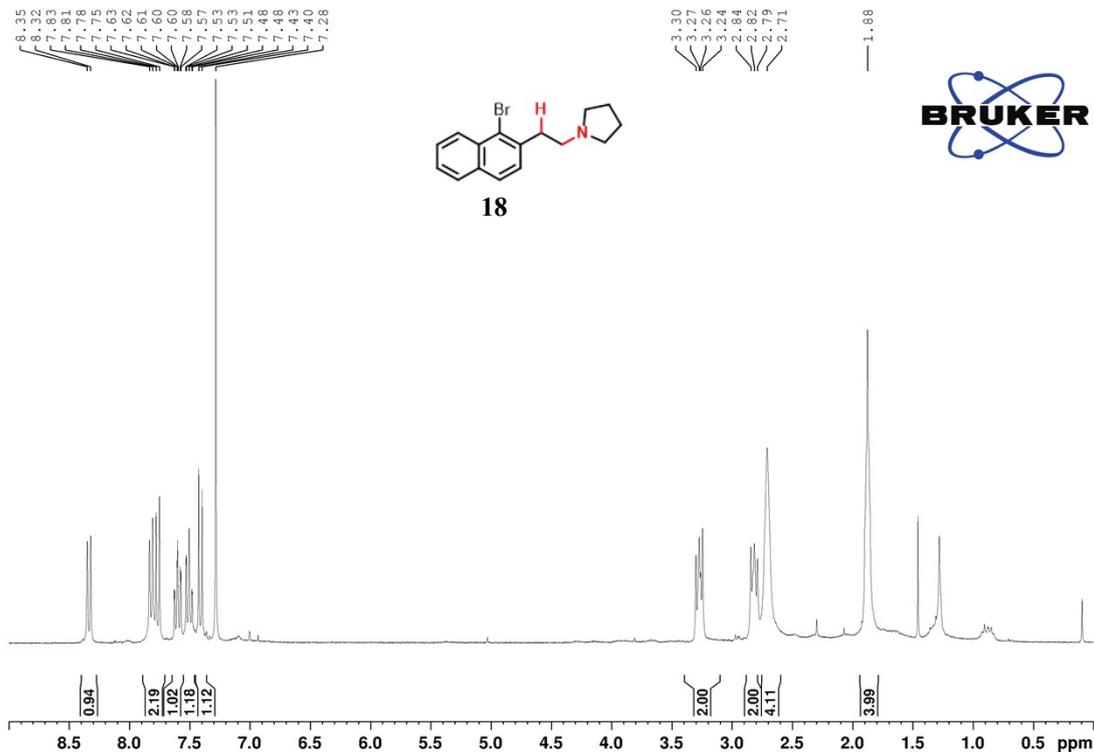
N-(2-phenylethyl)pyrrolidine



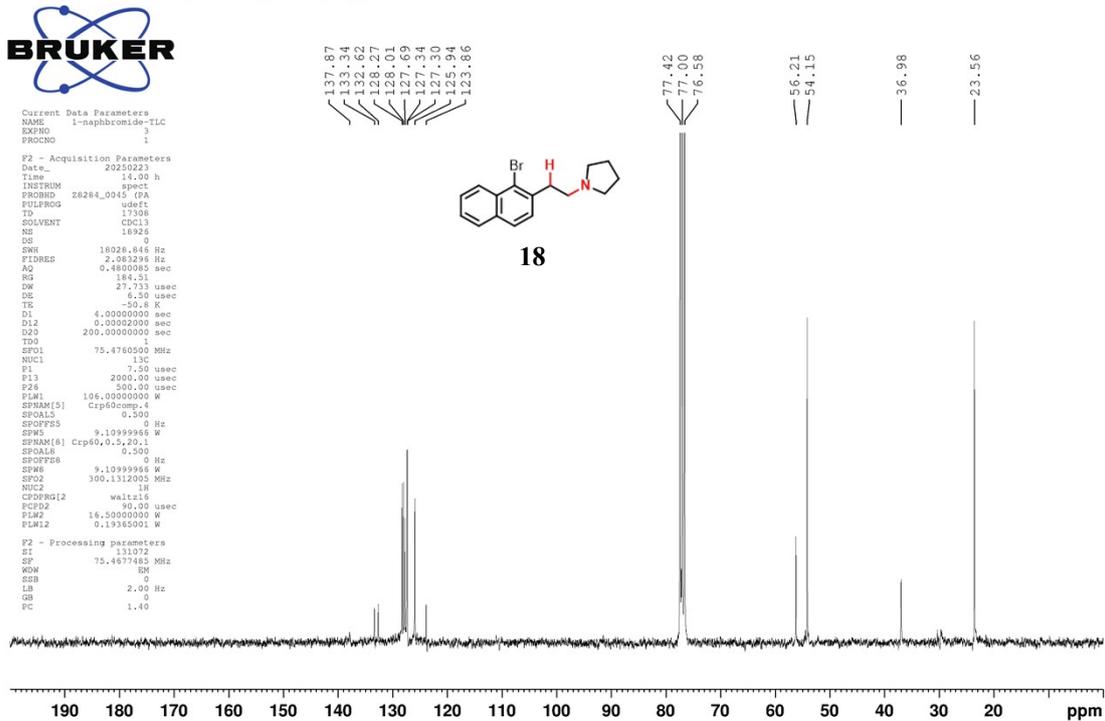
N-(2-phenylethyl)pyrrolidine



N-(2-(1-bromo-2-naphthalenyl)ethyl)pyrrolidine



N-(2-(1-bromo-2-naphthalenyl)ethyl)pyrrolidine



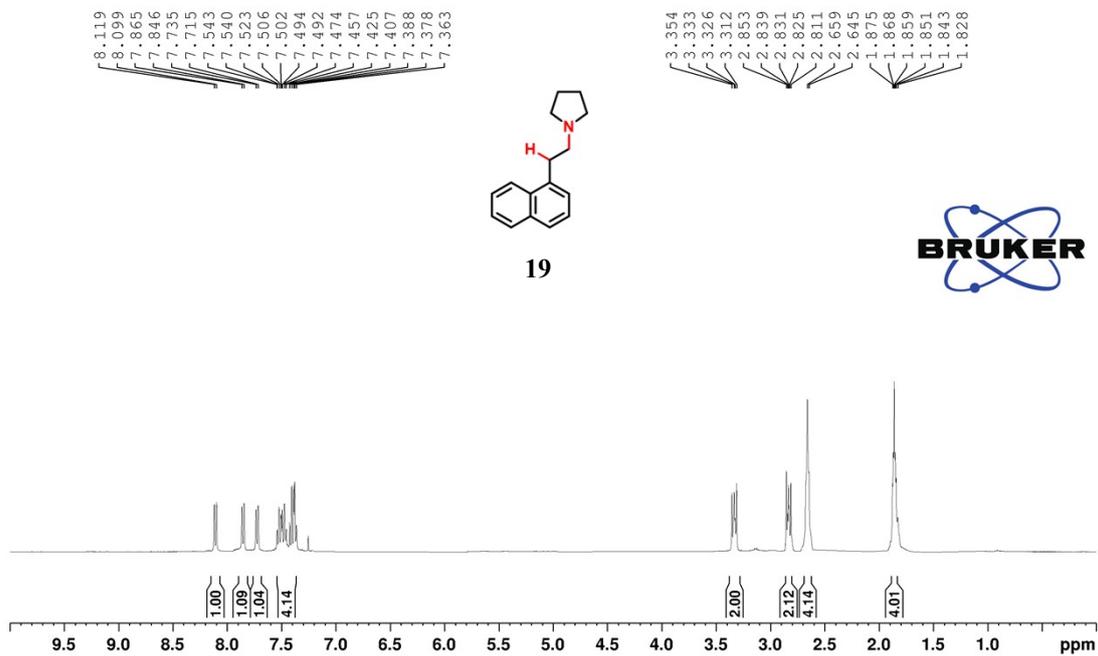
BRUKER

Current Data Parameters
NAME 1-naphbromide-TLC
EXPNO 1
PROCNO 1

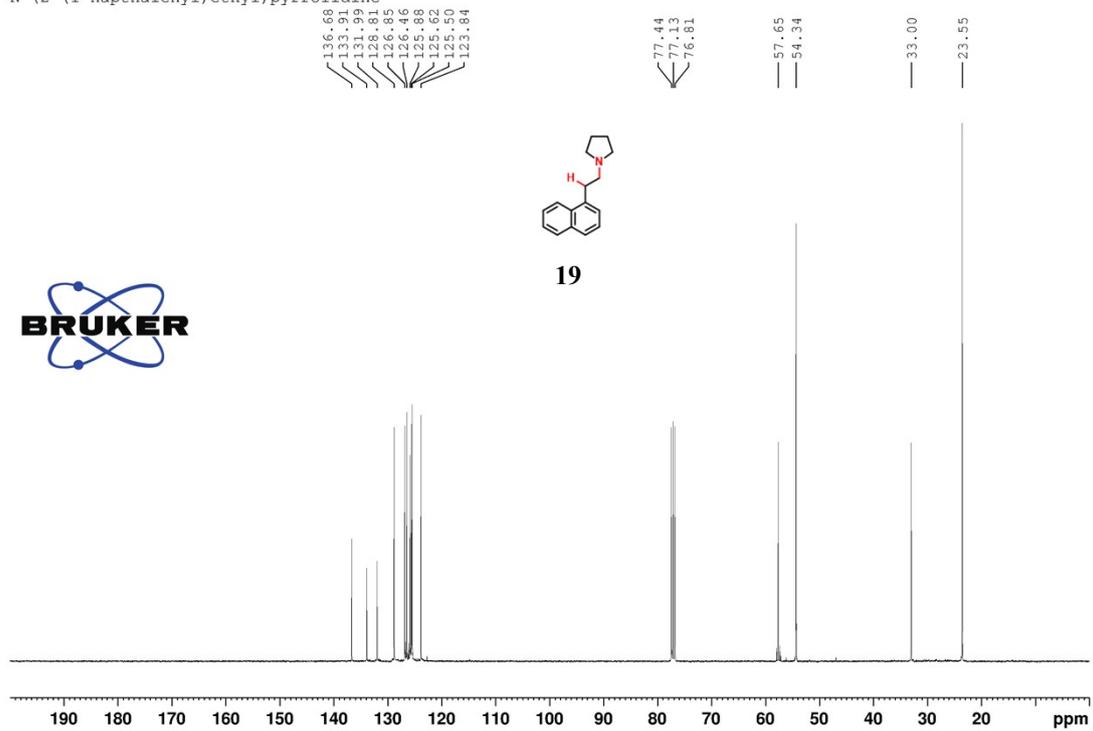
F2 - Acquisition Parameters
Date_ 20250223
Time 14:00 h
INSTRUM spect
PROBHD 28284_0946 GPC
PULPROG waltz16
TD 17308
SOLVENT CDCl3
NS 16926
DS 0
SWH 16026.846 Hz
FIDRES 2.083296 Hz
AQ 0.4600085 sec
RG 184.31
DW 27.733 usec
DE 6.50 usec
TE -50.8 K
D1 4.0000000 sec
D12 0.0000200 sec
D20 200.0000000 sec
TD0 1
SFO1 75.4760500 MHz
NUC1 13C
P1 7.50 usec
PL1 2000.00 usec
P2 500.00 usec
PL2 106.0000000 W
SFOAL5 Ccp60comp-4
SFOAL5 0.500
SFOFF5 0 Hz
SFOFF5 9.10999966 W
SFOFF5 Ccp60,0.5,20.1
SFOFF5 0.200
SFOFF5 0 Hz
SFOFF5 9.10999966 W
SFOFF5 300.1312005 MHz
NUC2 1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLM2 16.5000000 W
PLW2 0.19365001 W

F2 - Processing parameters
SI 131072
SF 75.4677485 MHz
WDW EM
SSB 0
LB 2.00 Hz
GB 0
PC 1.40

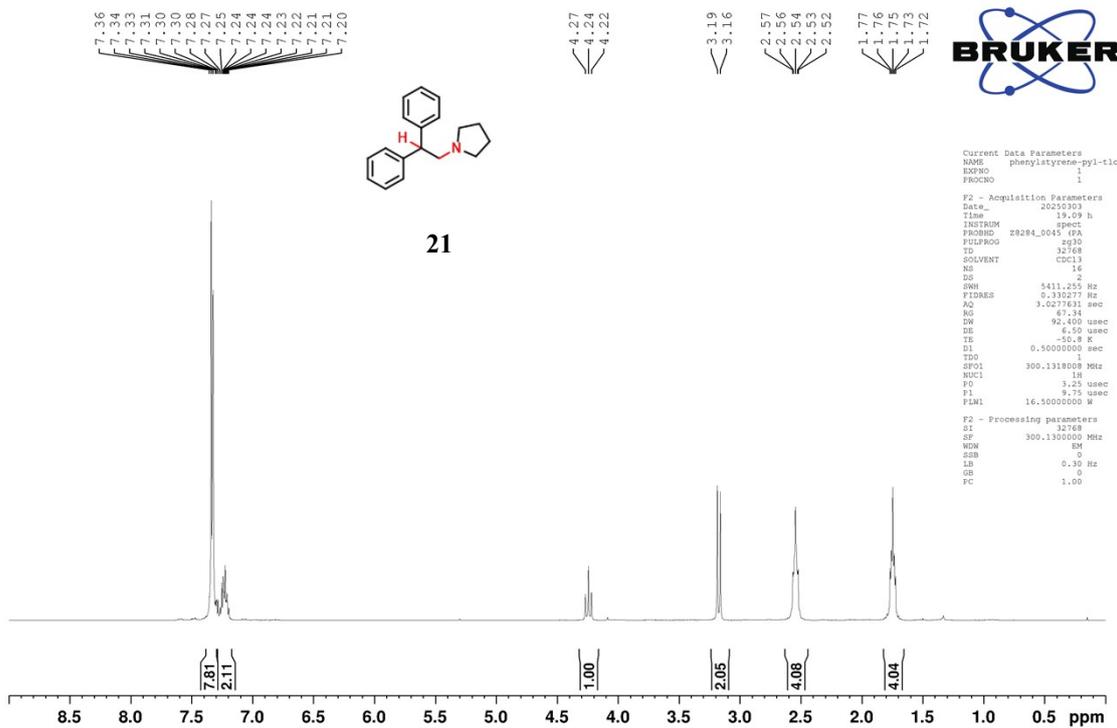
N-(2-(1-naphthalenyl)ethyl)pyrrolidine



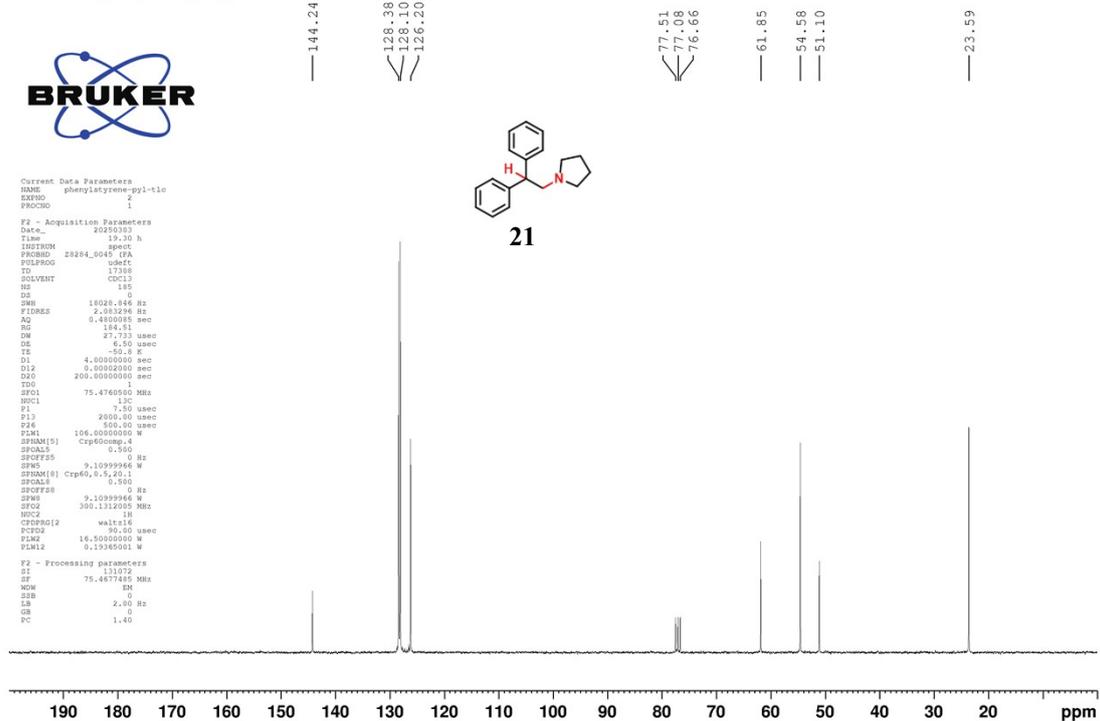
N-(2-(1-naphthalenyl)ethyl)pyrrolidine



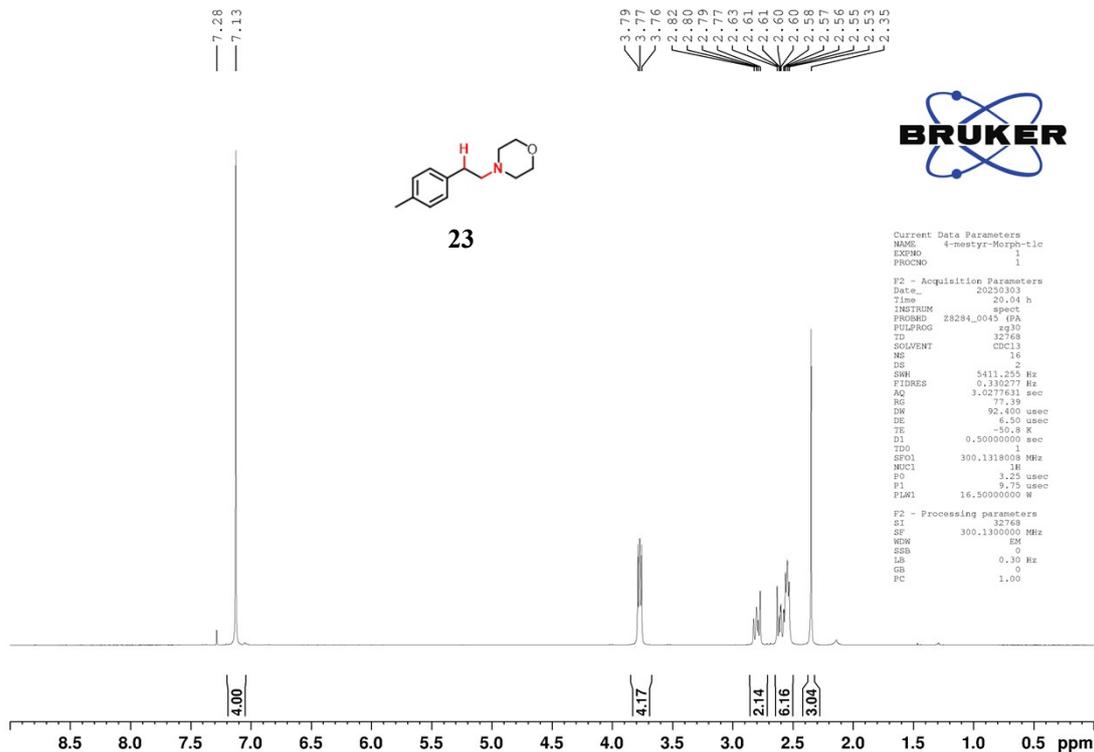
N-(2,2-Diphenylethyl)pyrrolidine



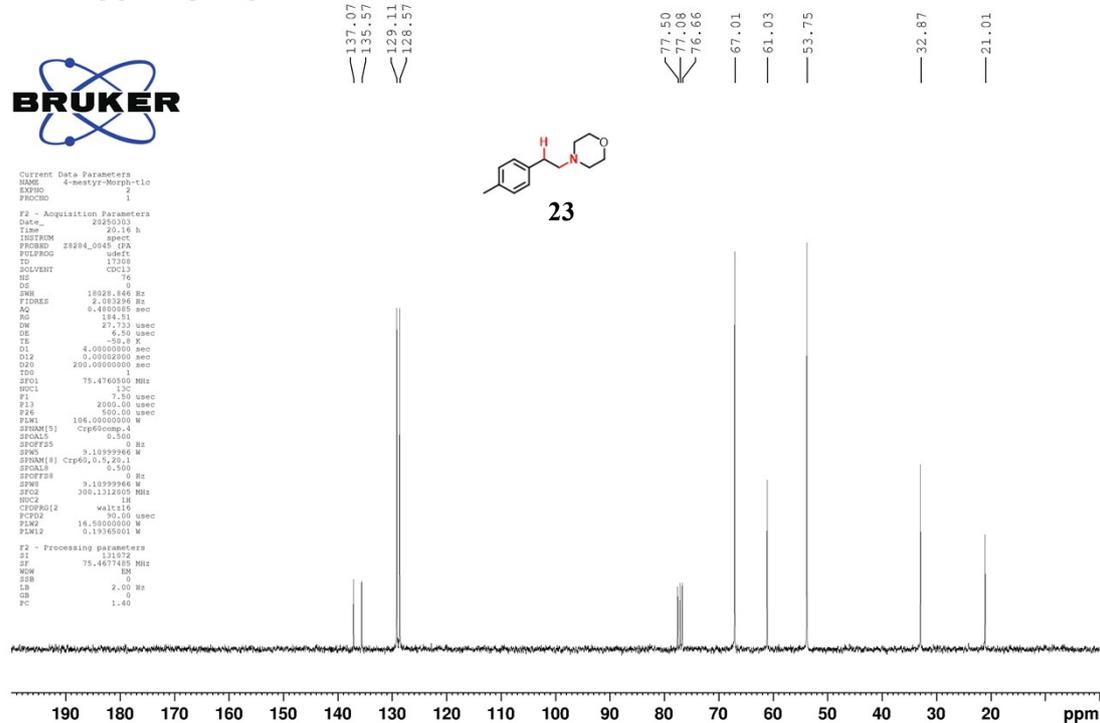
N-(2,2-diphenylethyl)pyrrolidine



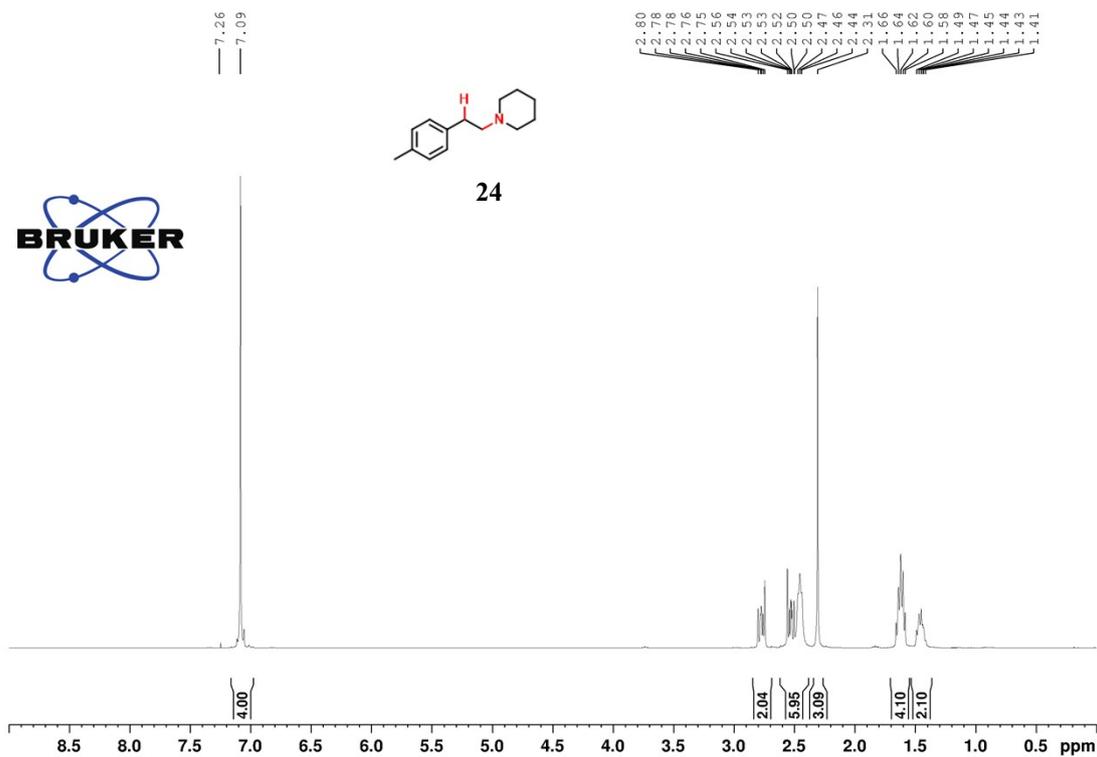
N-(4-methylphenethyl)morpholine



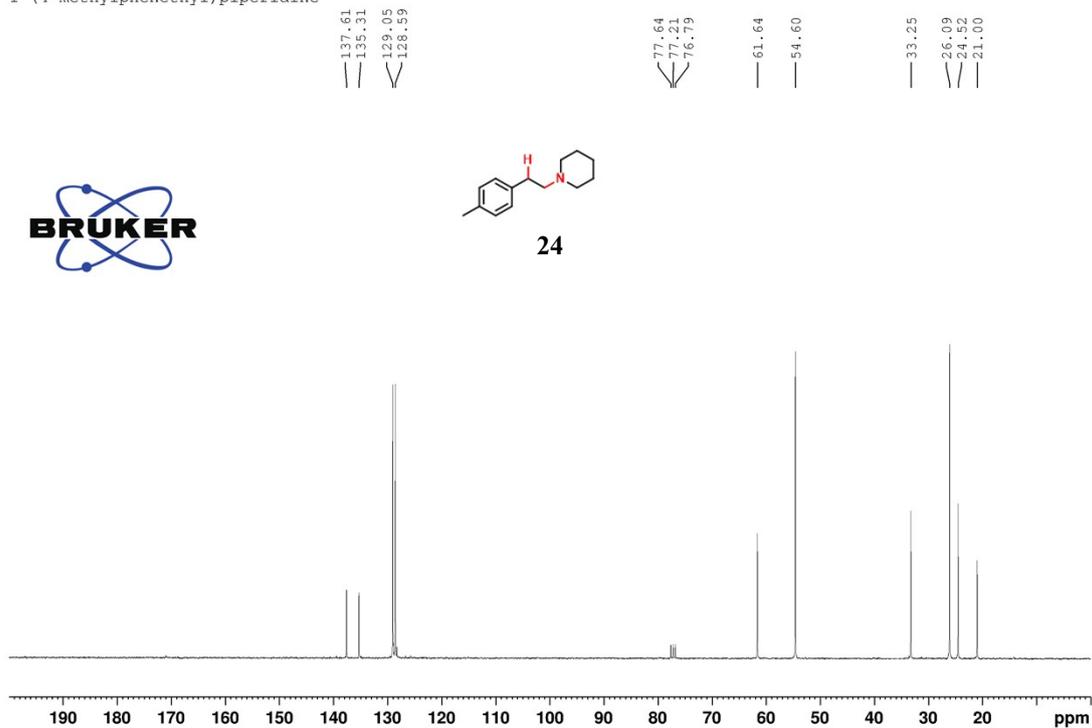
N-(4-methylphenethyl)morpholine



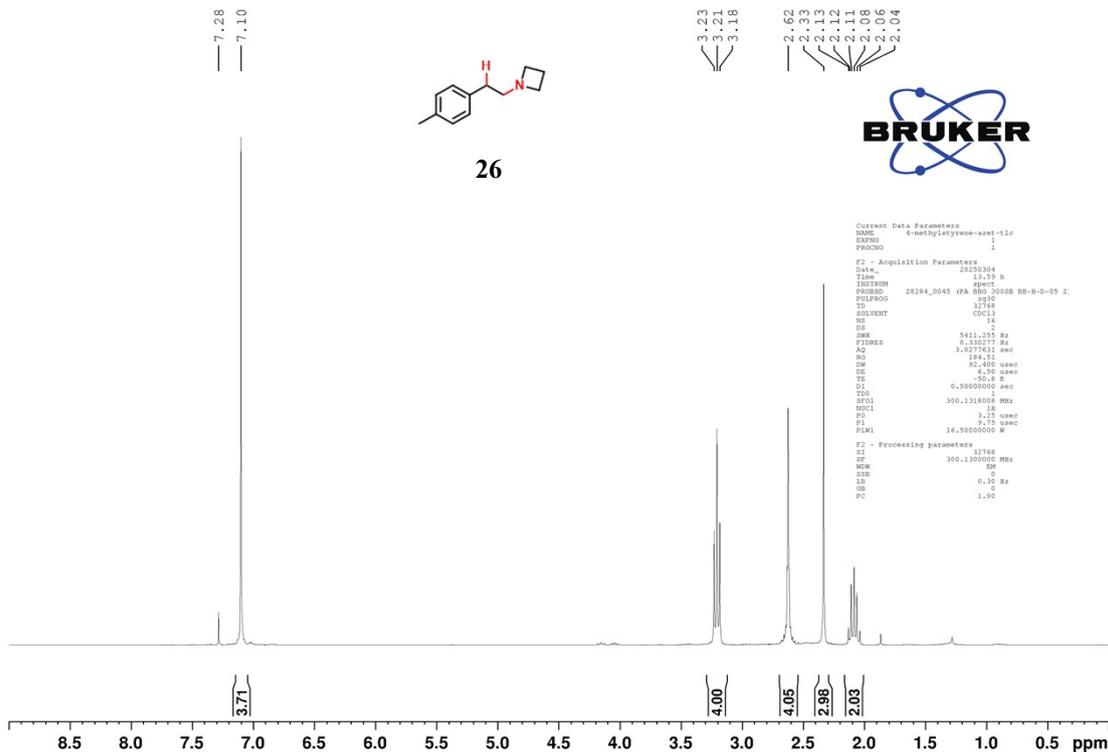
1-(4-methylphenethyl)piperidine



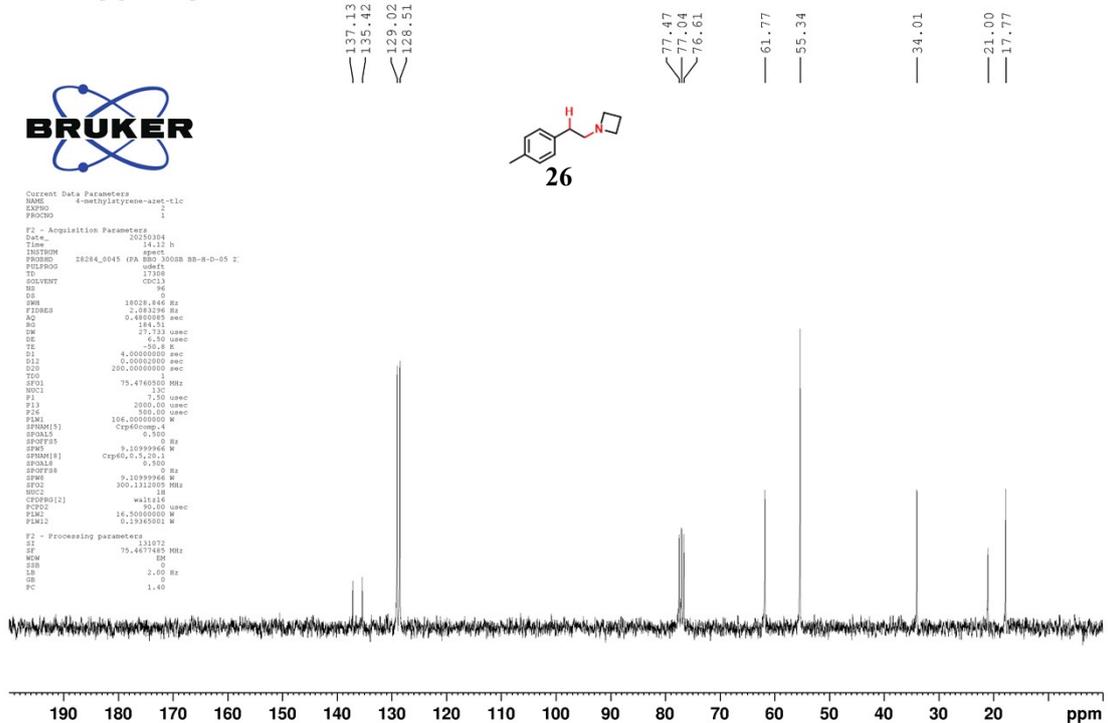
1-(4-methylphenethyl)piperidine



N-(4-methylphenethyl)azetidine



N-(4-methylphenethyl)azetidine



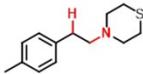
N-(4-methylphenethyl)thiomorpholine



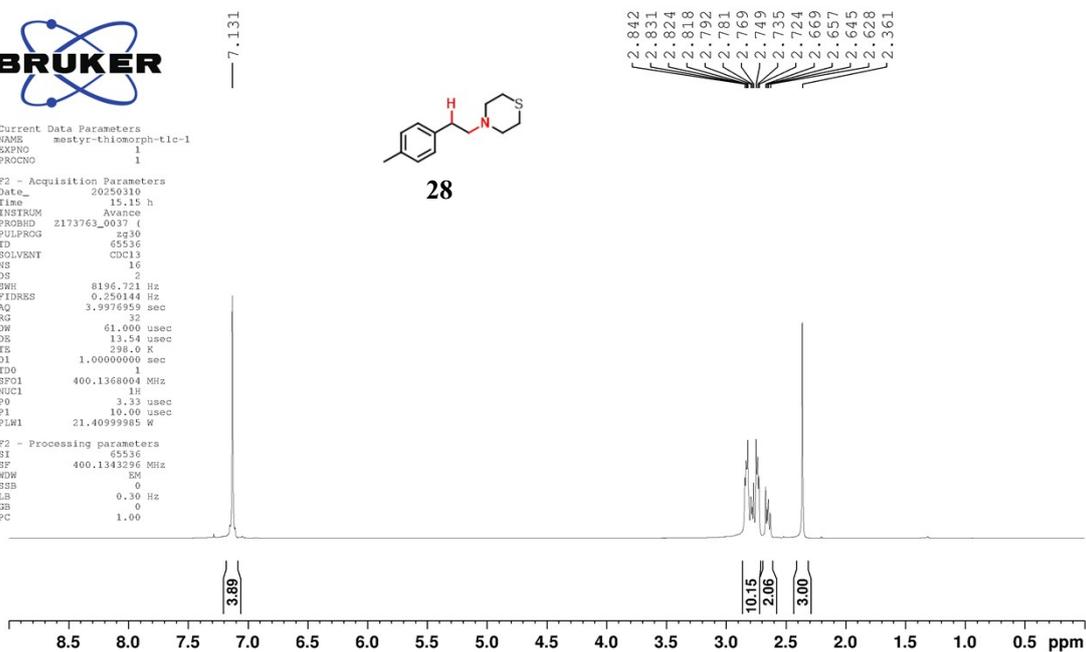
Current Data Parameters
 NAME mestyr-thiomorph-tic-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20250310
 Time 15.15 h
 INSTRUM Avance
 PROBHD z173763_0037 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8196.721 Hz
 FIDRES 0.250184 Hz
 AQ 3.9976959 sec
 RG 32
 DW 61.000 usec
 DE 13.54 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO
 SFO1 400.1368004 MHz
 NUC1 1H
 PD 3.33 usec
 P1 10.00 usec
 PLW1 21.4099985 W

F2 - Processing parameters
 SI 65536
 SF 400.1343296 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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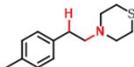
N-(4-methylphenethyl)thiomorpholine



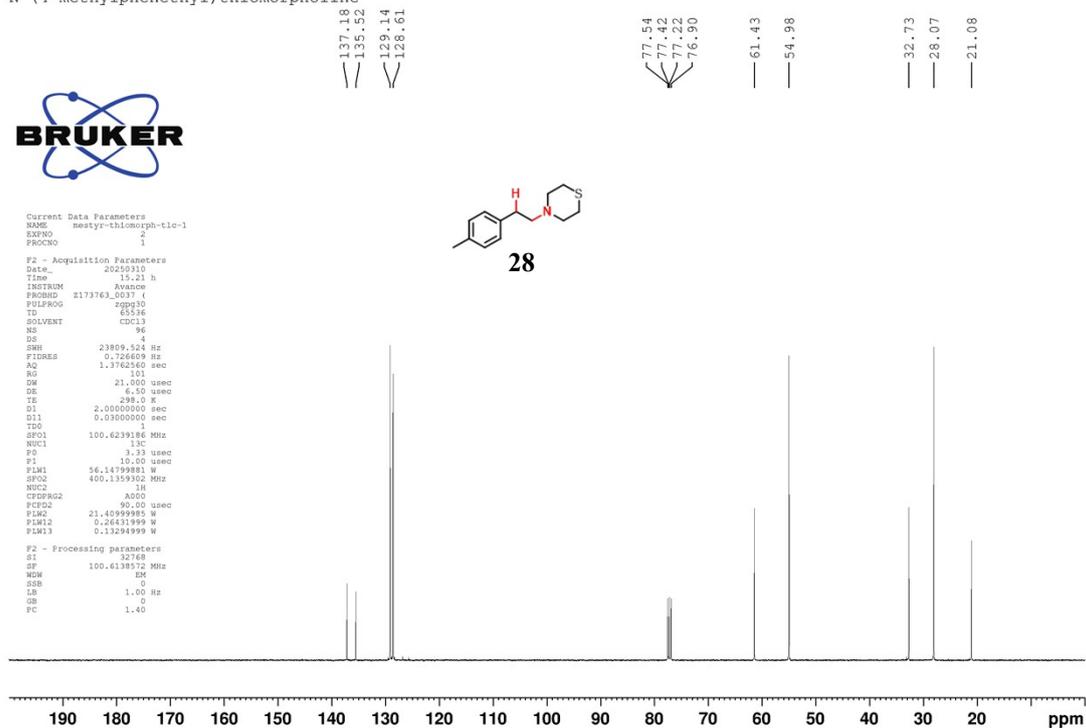
Current Data Parameters
 NAME mestyr-thiomorph-tic-1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20250310
 Time 15.21 h
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 PROBHD z173763_0037 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 4
 SWH 23809.524 Hz
 FIDRES 0.726609 Hz
 AQ 1.3762660 sec
 RG 101
 DW 21.000 usec
 DE 6.30 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO
 SFO1 100.6239186 MHz
 NUC1 13C
 PD 3.33 usec
 P1 10.00 usec
 PLW1 56.1479981 W
 SFO2 400.1359302 MHz
 NUC2 1H
 CFCPRG2 APO
 FCFD2 90.00 usec
 PLM2 21.4099985 W
 PLM12 0.56431999 W
 PLM13 0.13294999 W

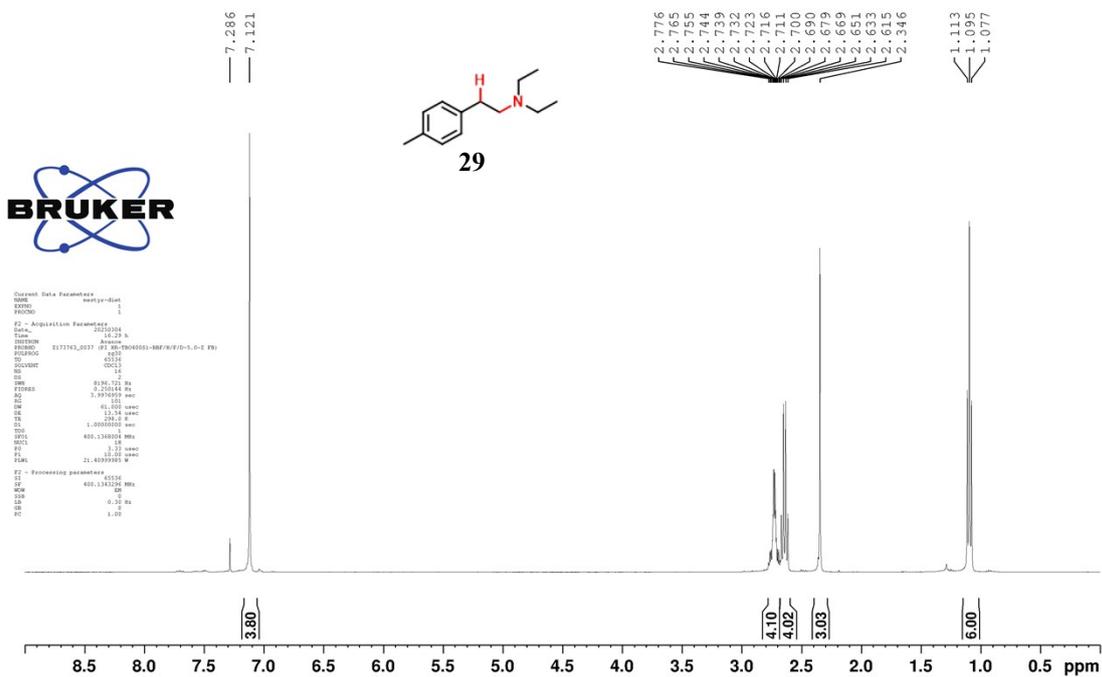
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 LB 1.00 Hz
 GB 0
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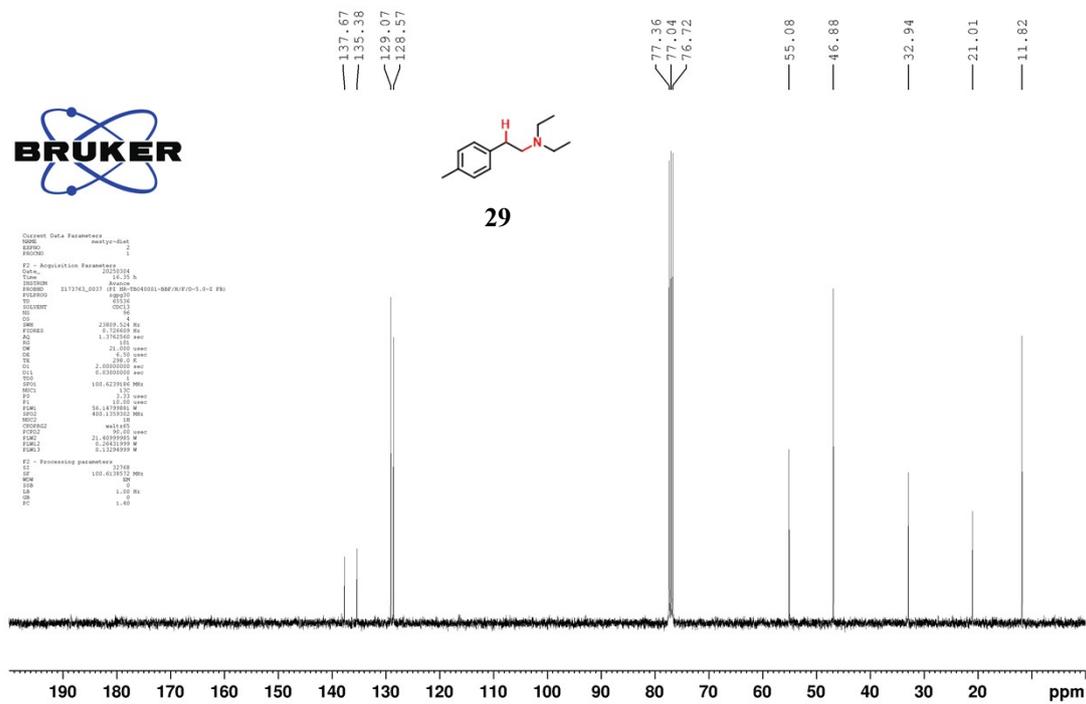
28



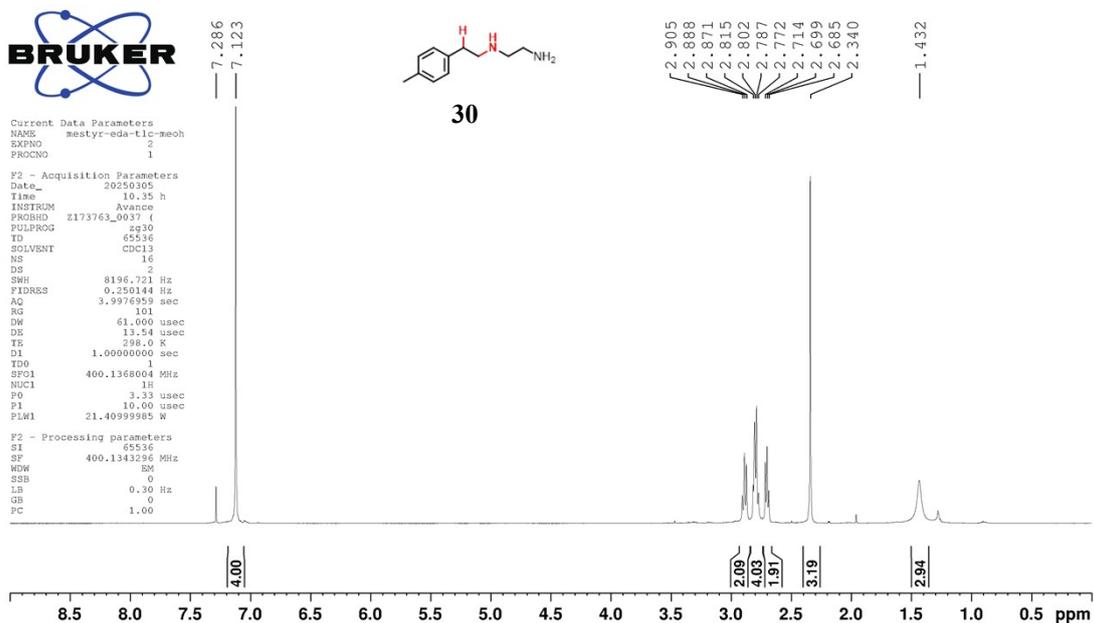
N-(4-methylphenethyl)diethylamine



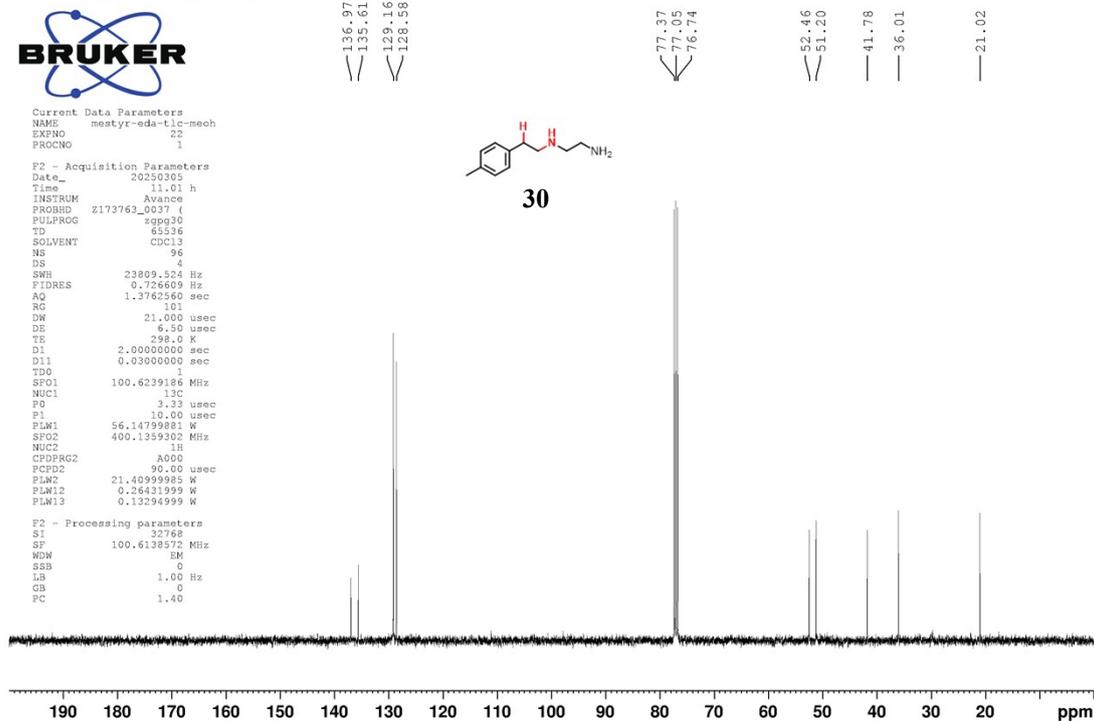
N-(4-methylphenethyl)diethylamine



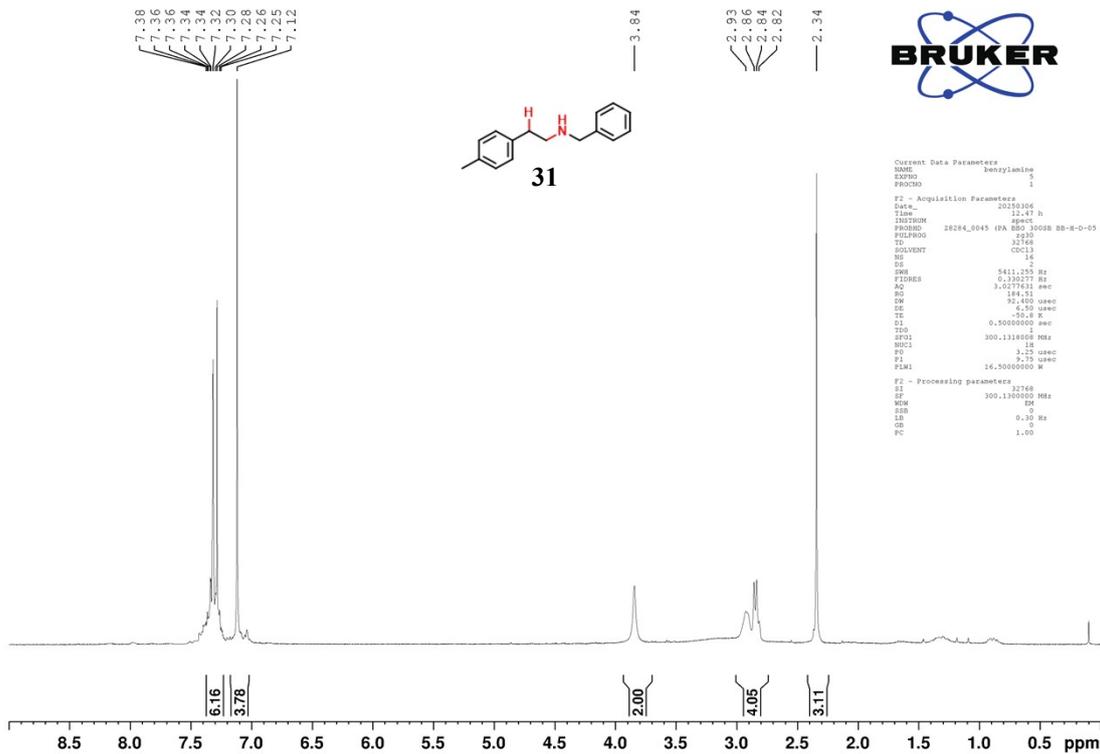
N-(4-methylphenethyl)ethylenediamine



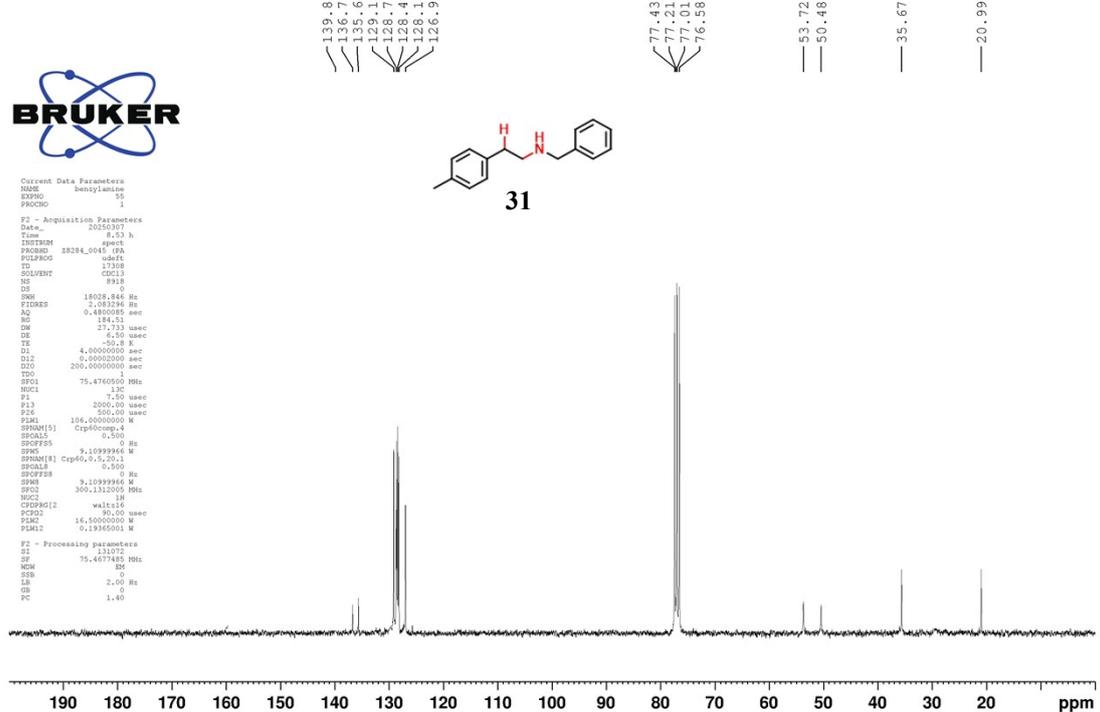
N-(4-methylphenethyl)ethylenediamine



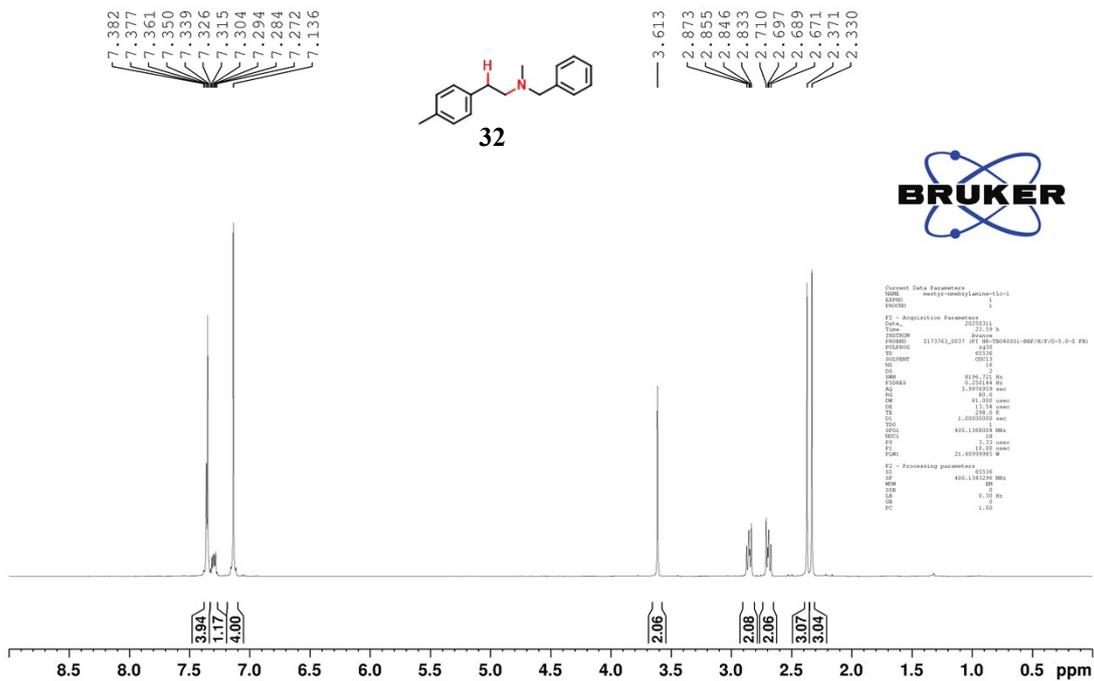
N-(4-methylphenethyl)benzylamine



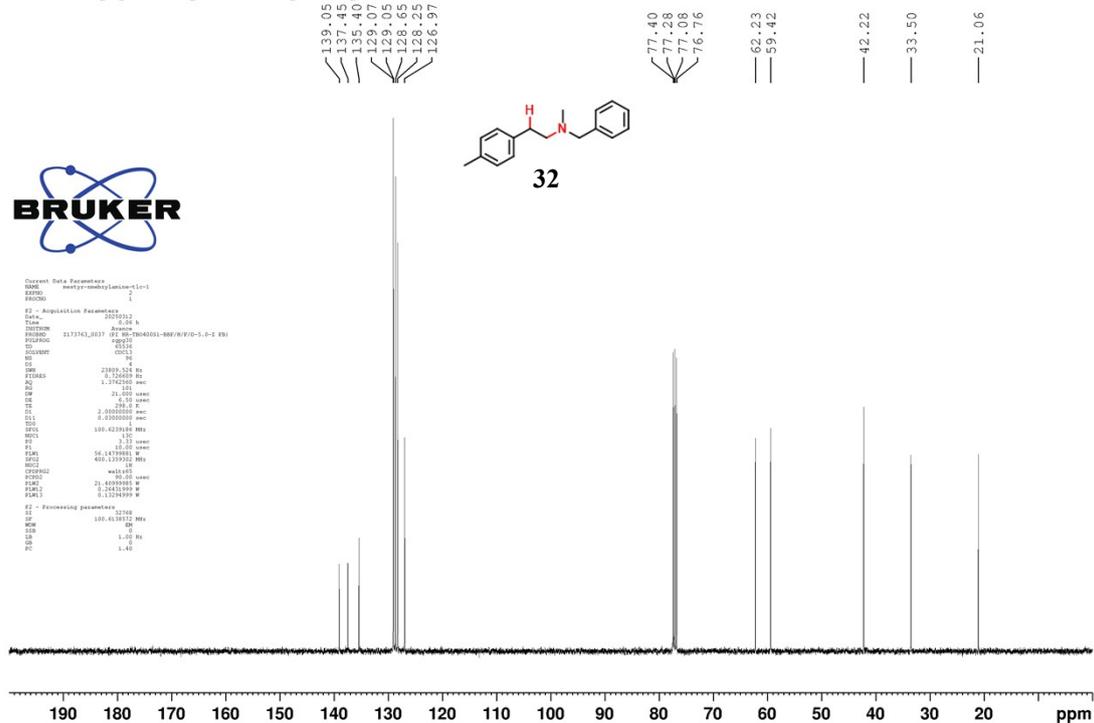
N-(4-methylphenethyl)benzylamine



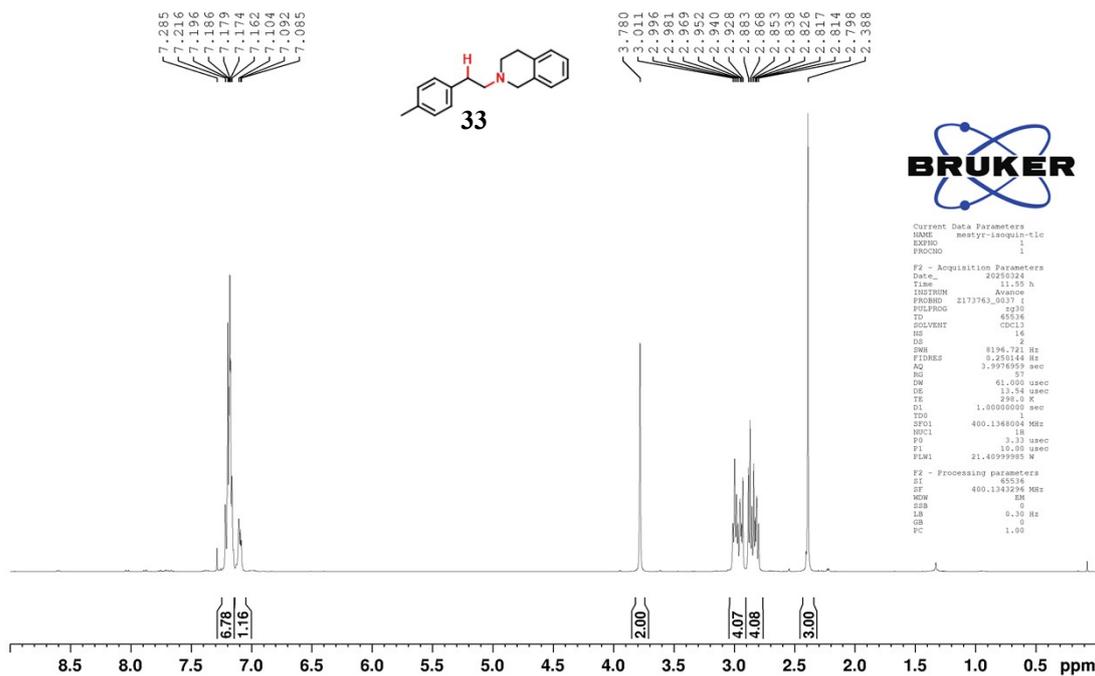
N-(4-methylphenethyl)-N-methylbenzylamine



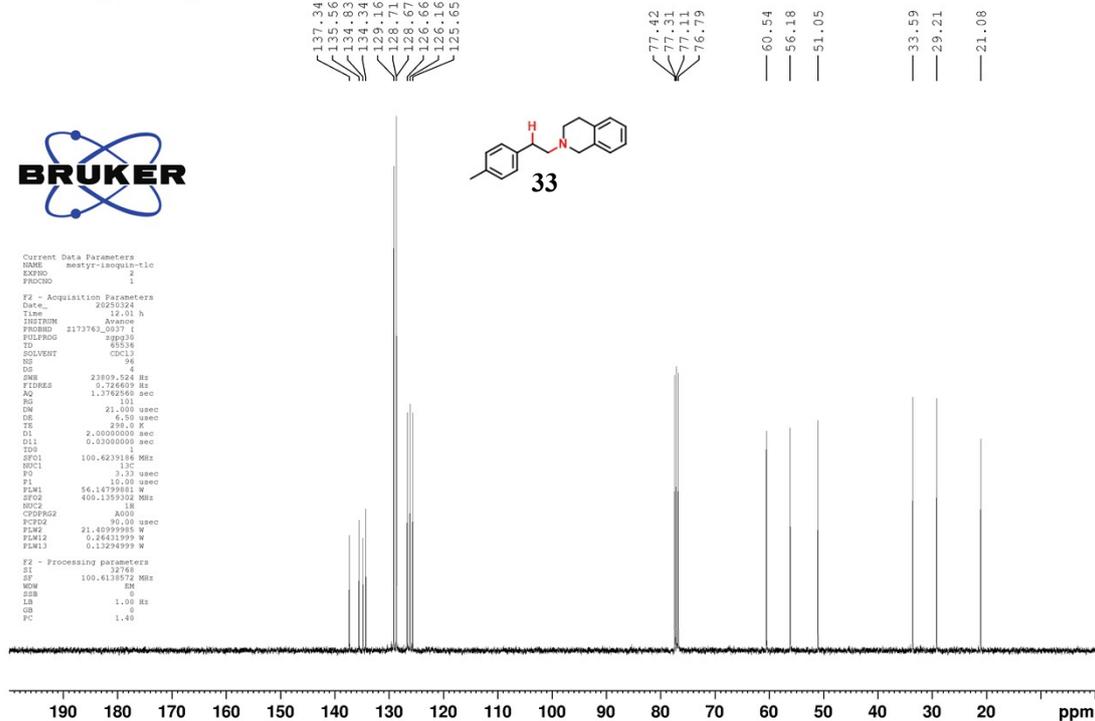
N-(4-methylphenethyl)-N-methylbenzylamine



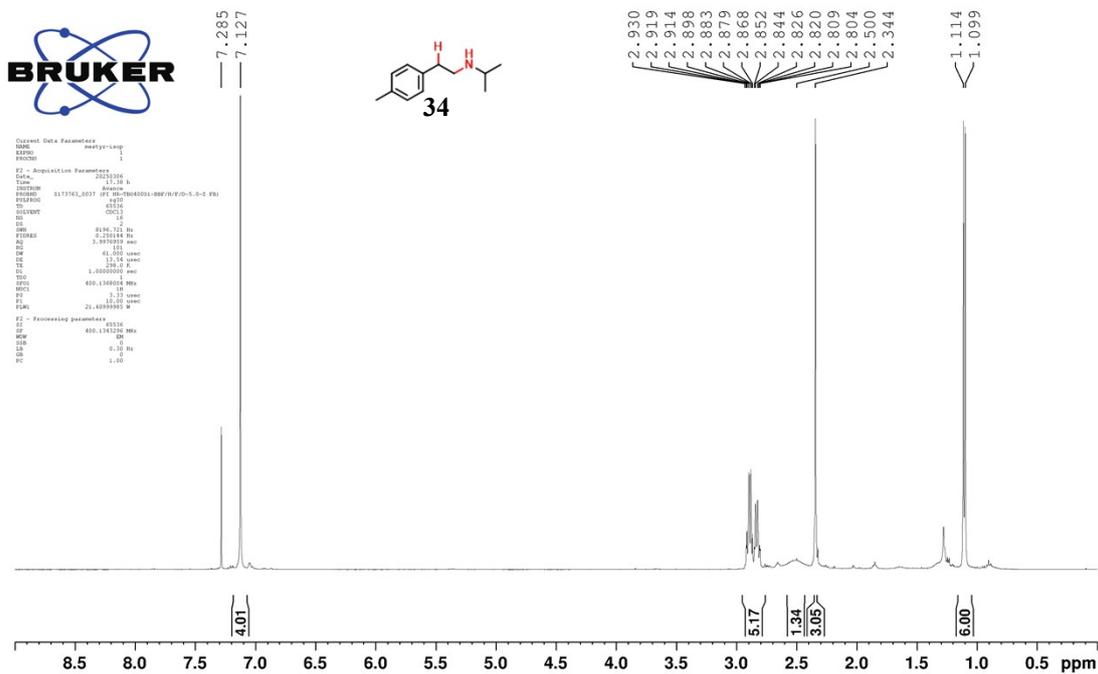
N-(4-methylphenethyl)-1,2,3,4-tetrahydroisoquinoline



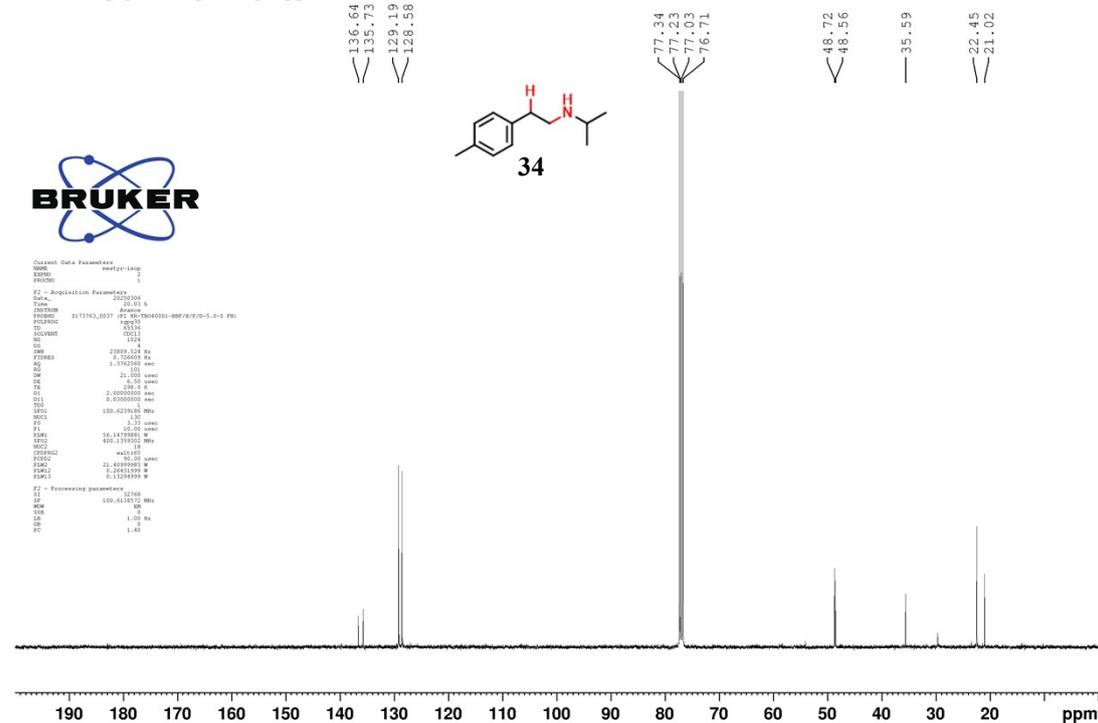
N-(4-methylphenethyl)-1,2,3,4-tetrahydroisoquinoline



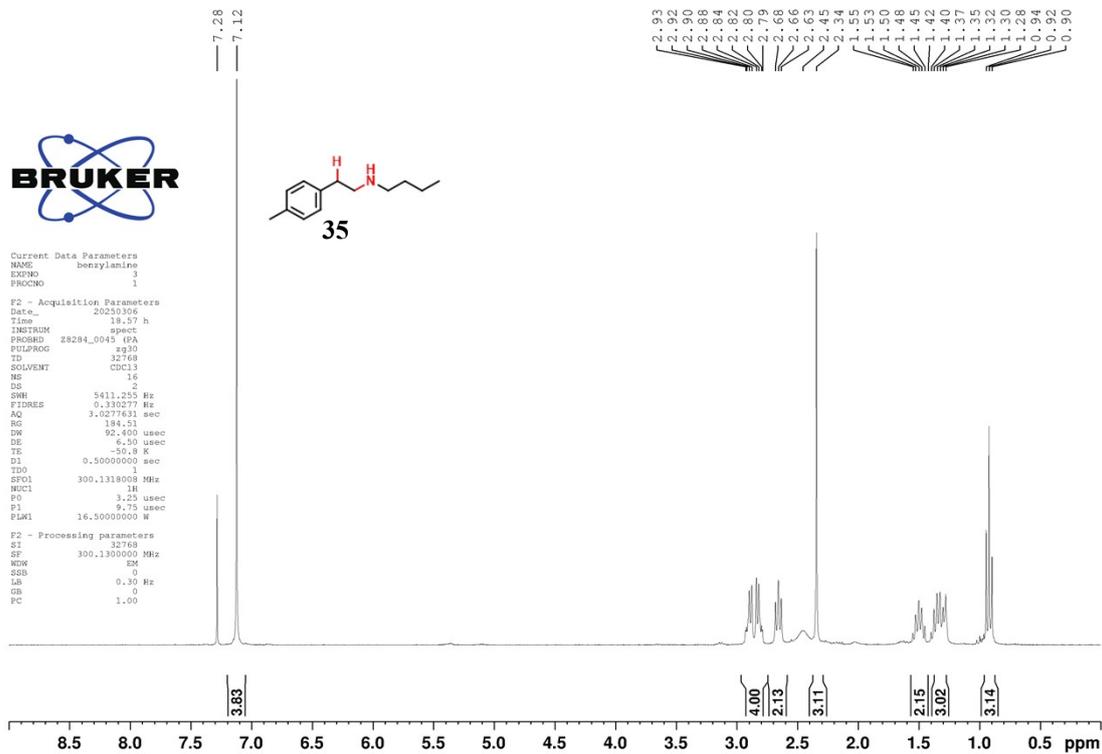
N-(4-methylphenethyl)isopropylamine



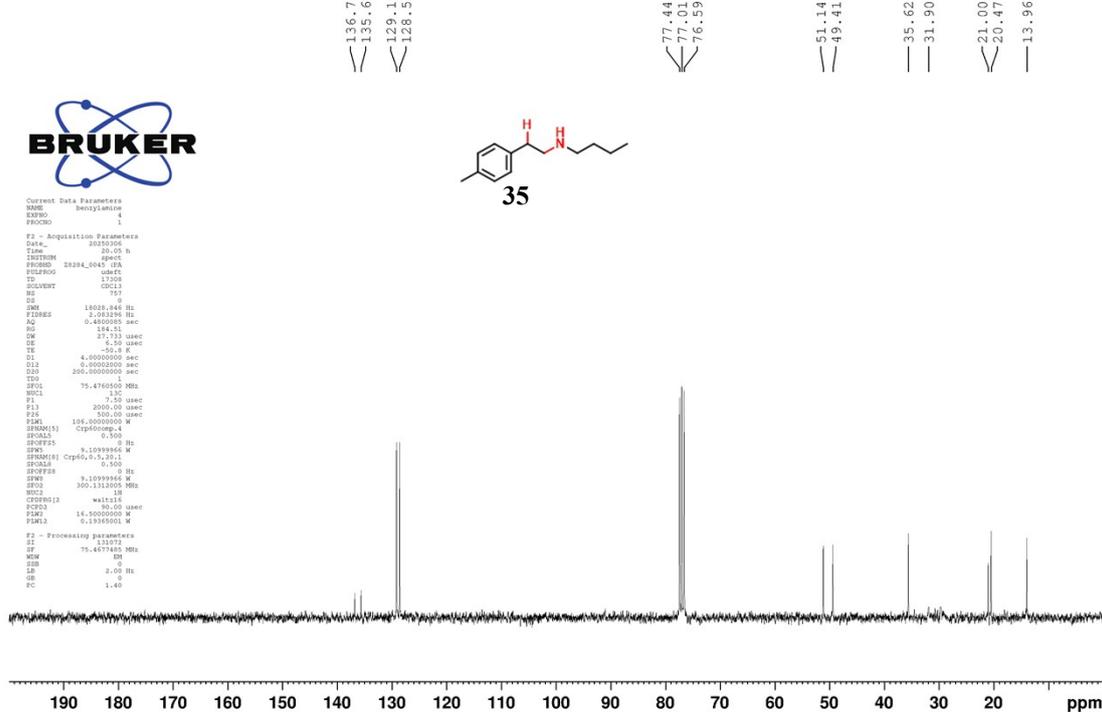
N-(4-methylphenethyl)isopropylamine



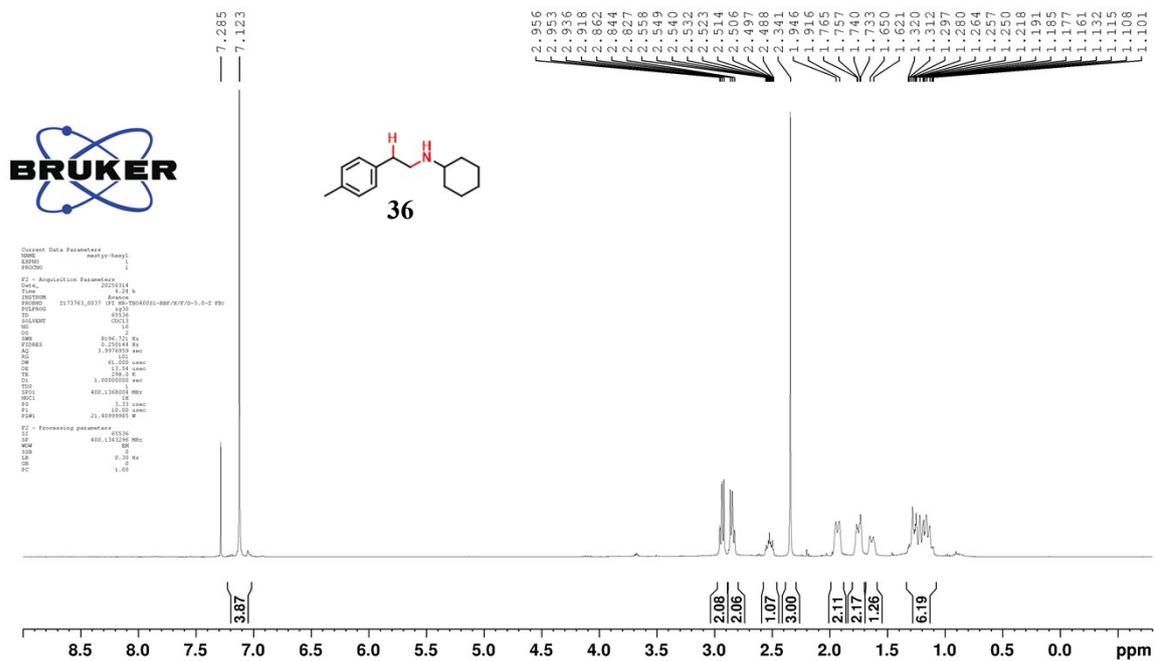
N-(4-methylphenethyl)butylamine



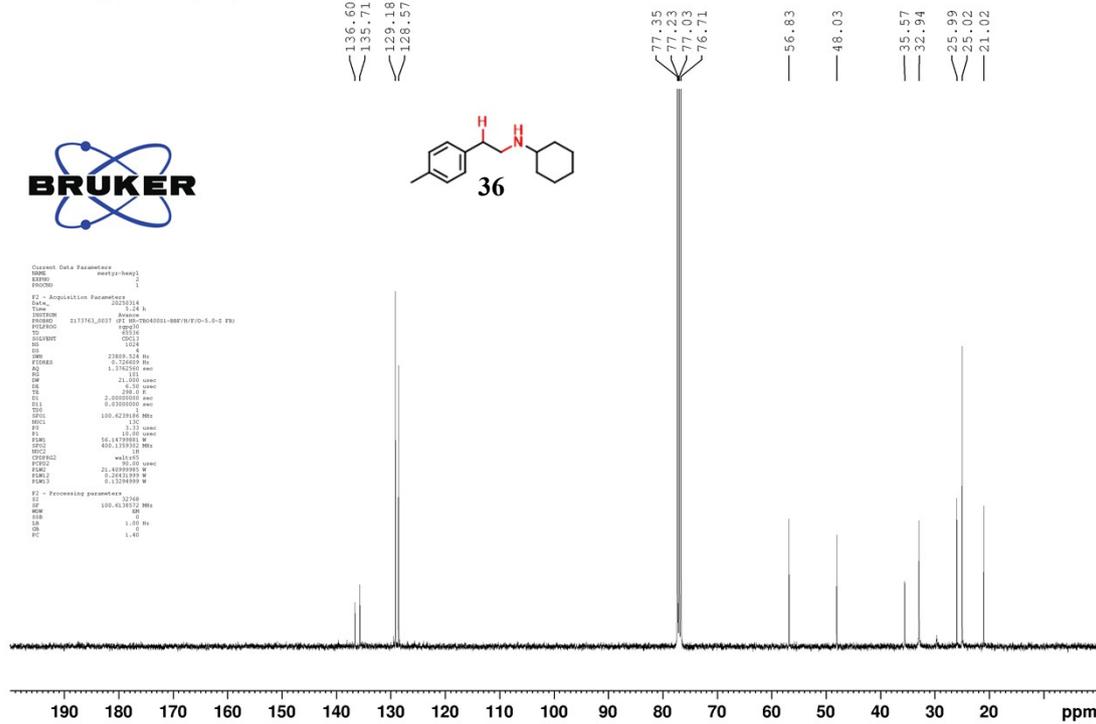
N-(4-methylphenethyl)butylamine



N-(4-methylphenethyl)cyclohexylamine



N-(4-methylphenethyl)cyclohexylamine



1-(2,3-Dimethyl-2-buten-1-yl)pyrrolidine



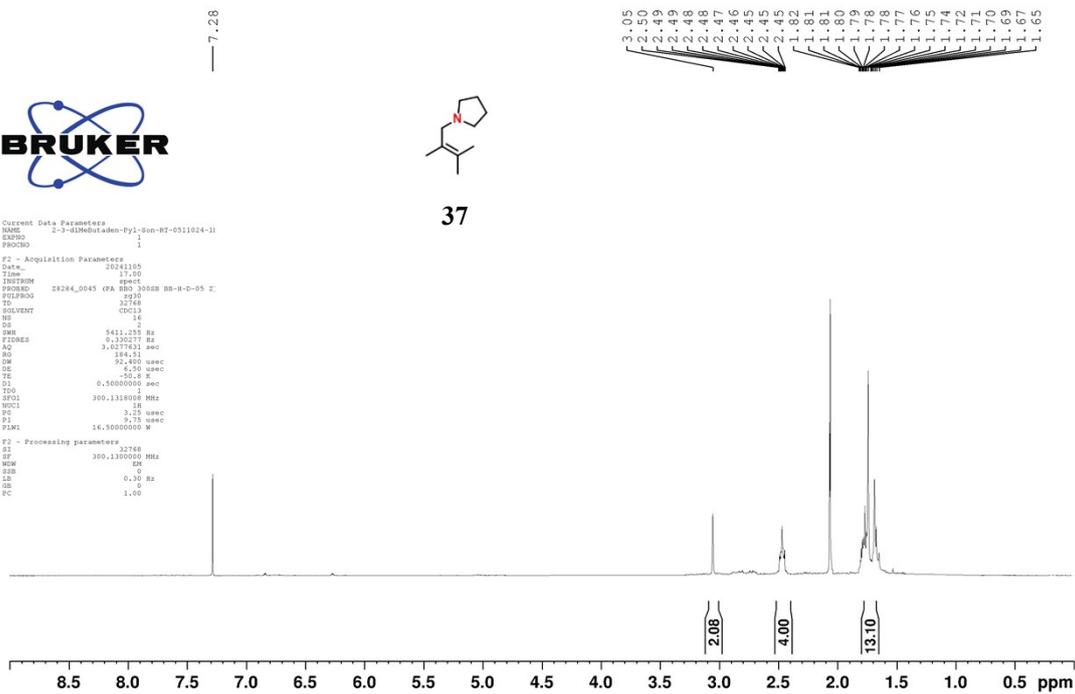
37

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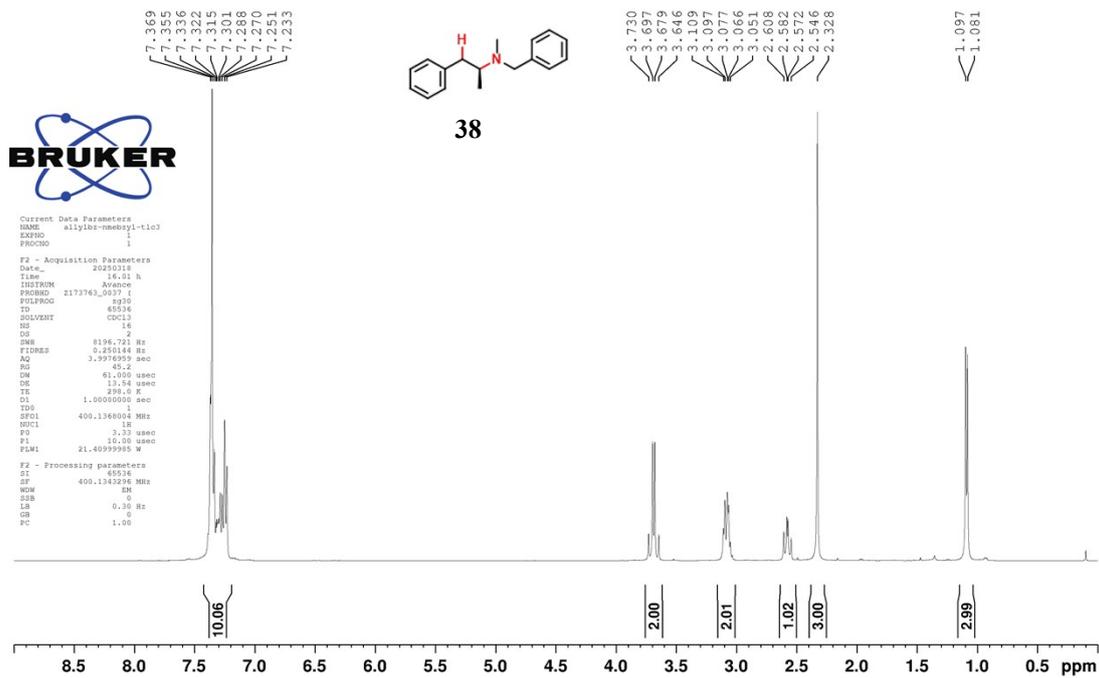
Current Data Parameters
NAME      2-3-dimethyladen-Pyl-Son-BT-0511024-11
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20241105
Time     17.00
INSTRUM  spect
PROBHD   28284_0049 (PA BBH 30628 BB-N-0-05 Z)
PULPROG  zgpg30
RG       327.68
SOLVENT  CDCl3
NS       14
DS       2
SWH      5411.100 Hz
FIDRES   0.1302377 Hz
AQ       1.4237611 sec
RG       384.51
DF       52.4800 usec
DE       6.50 usec
TE       29.68 K
DQ       0.5000000 sec
SFO1     300.136000 MHz
MPC1     18
PC       0.25 usec
PI       0.75 usec
PL1      16.50000000 w

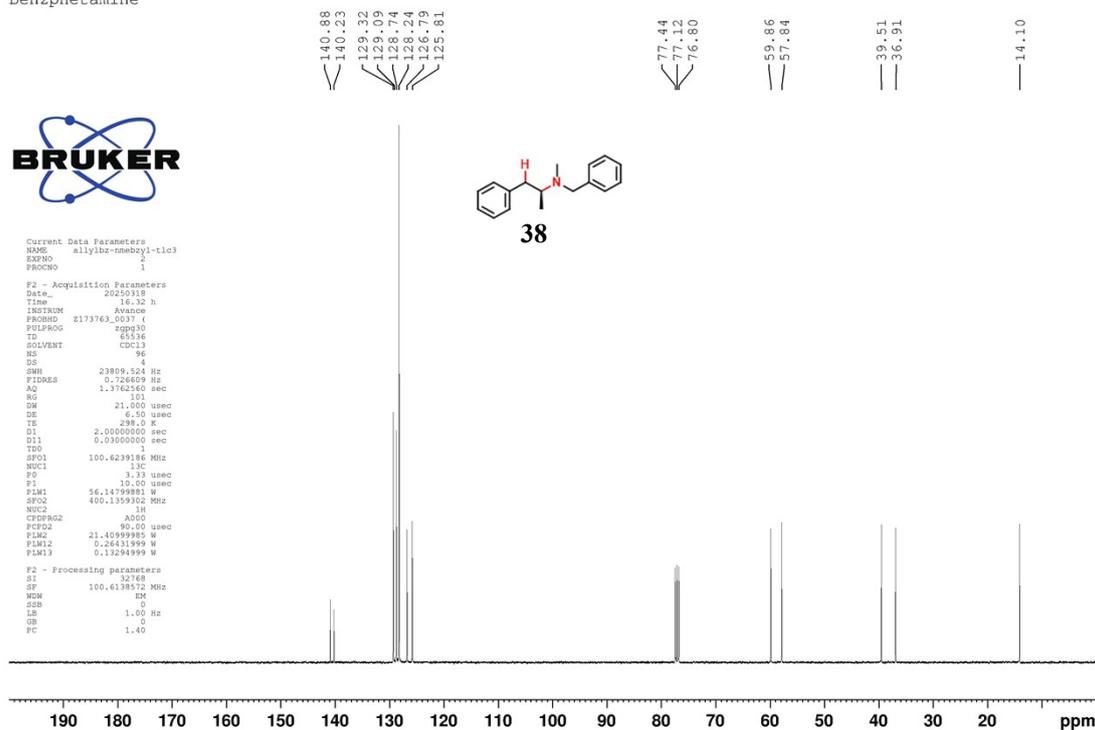
F2 - Processing parameters
SI       32768
SF       300.1360000 MHz
WDW      EM
SSB      0
LB       0.10 Hz
GB       0
AS       1.00
  
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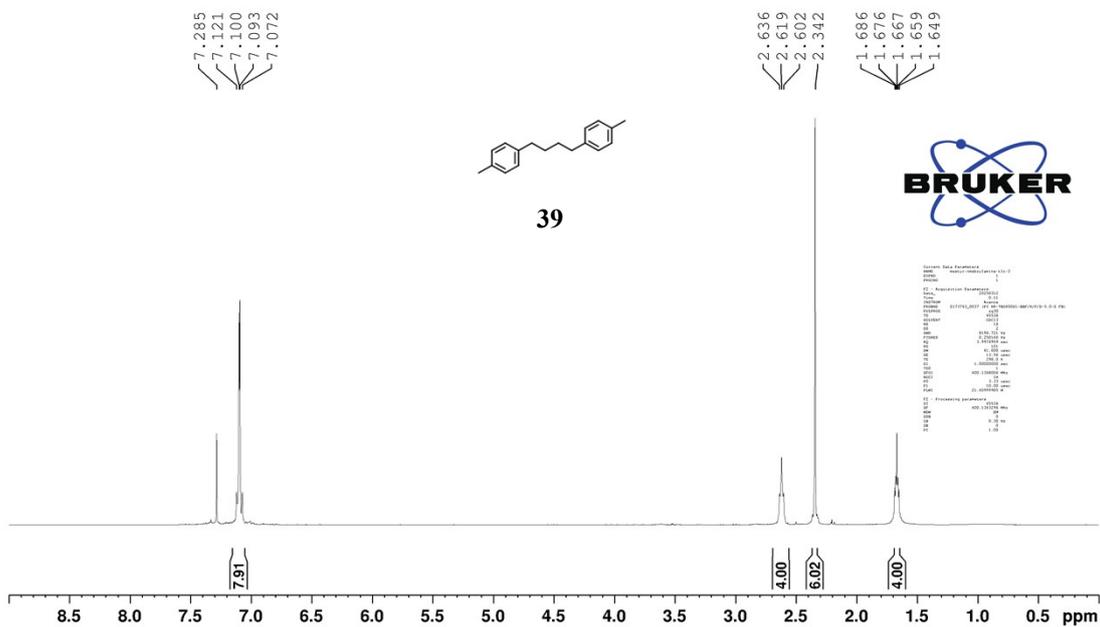
Benzphetamine



Benzphetamine



1,4-di-p-tolylbutane



1,4-di-p-tolylbutane

