

Synthesis and Design of Terbium-Activated CaWO₄ Nanocrystals for Fluorescence Sensing of Alkaline Phosphatase and Bioimaging Applications

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Optimization of synthesis parameters

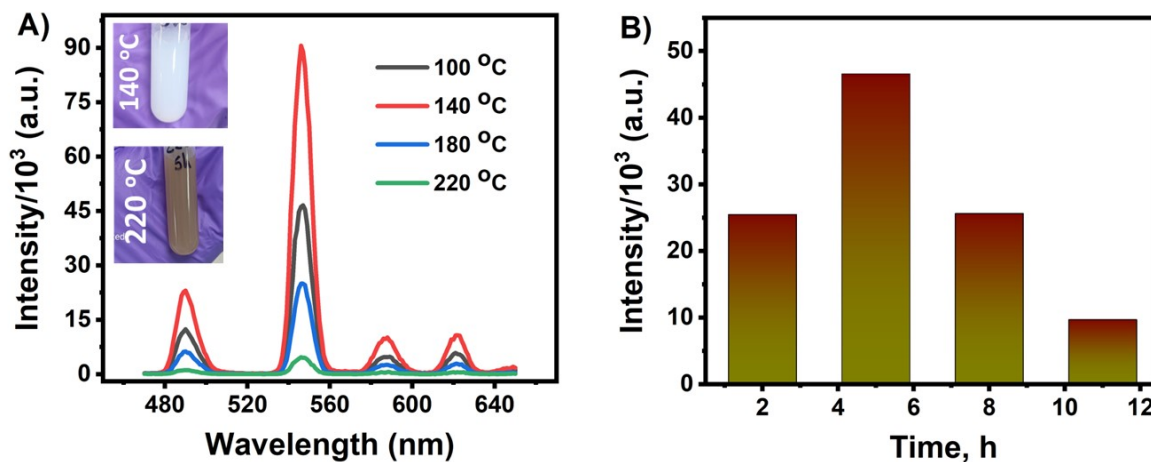


Fig. S1 Optimization of synthesis parameters A) temperature: insets are corresponding aqueous dispersion of PEG-Tb-CaWO₄ NCs and B) time.

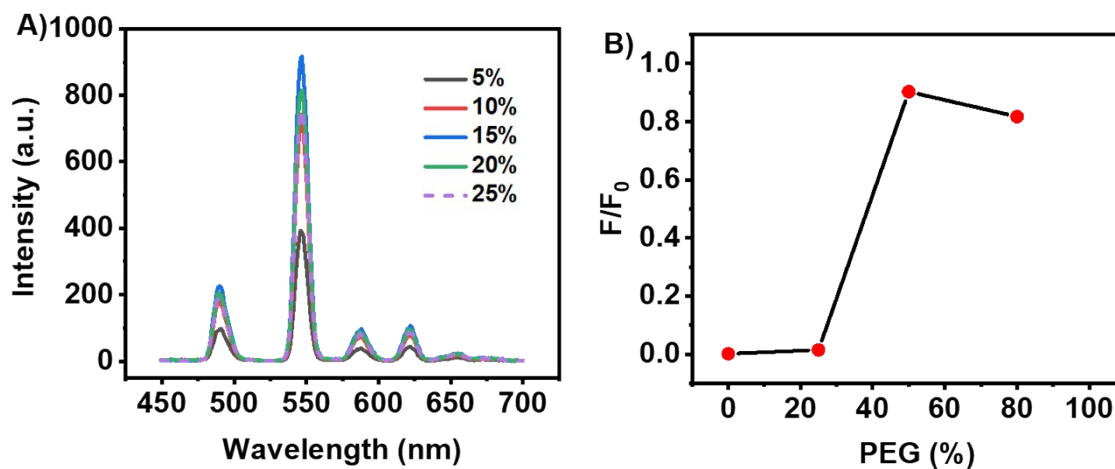


Fig. S2 Optimization of synthesis parameters A) %doping of Tb³⁺ and B) amount of PEG in the reaction medium

Characterization of the as-synthesized PEG-Tb-CaWO₄ NCs

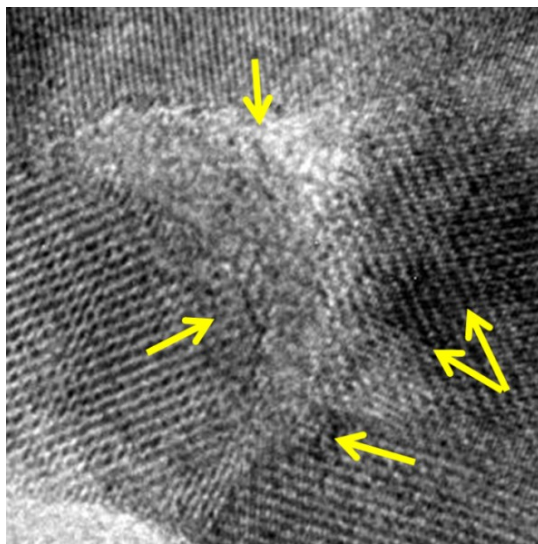


Fig. S3 HR-TEM image showing poly-crystalline nature of the PEG-Tb-CaWO₄ NCs.

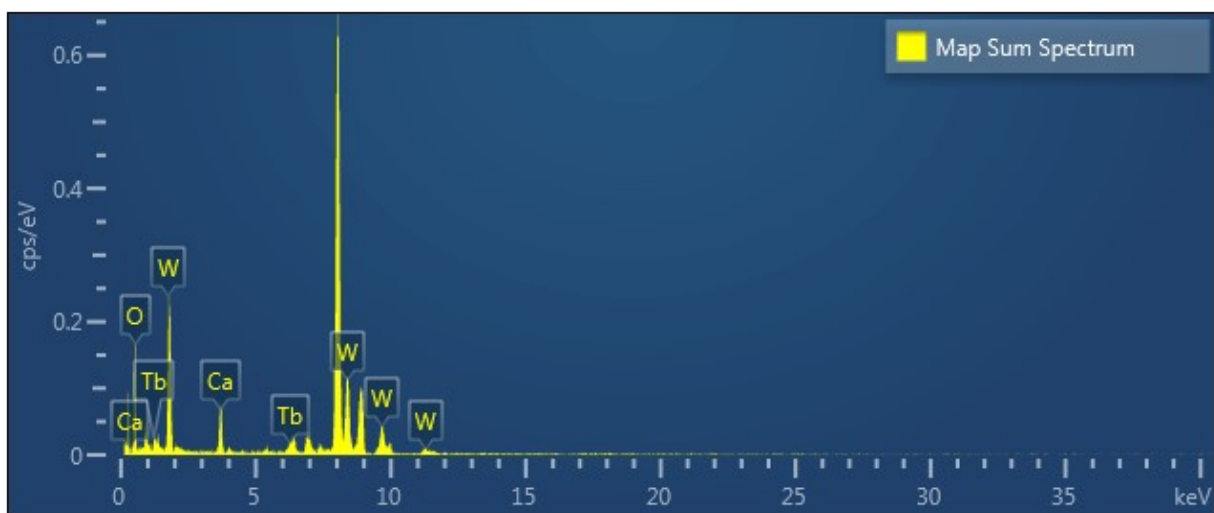


Fig. S4 Energy dispersive X-ray (EDS) spectrum of the PEG-Tb-CaWO₄ NCs.

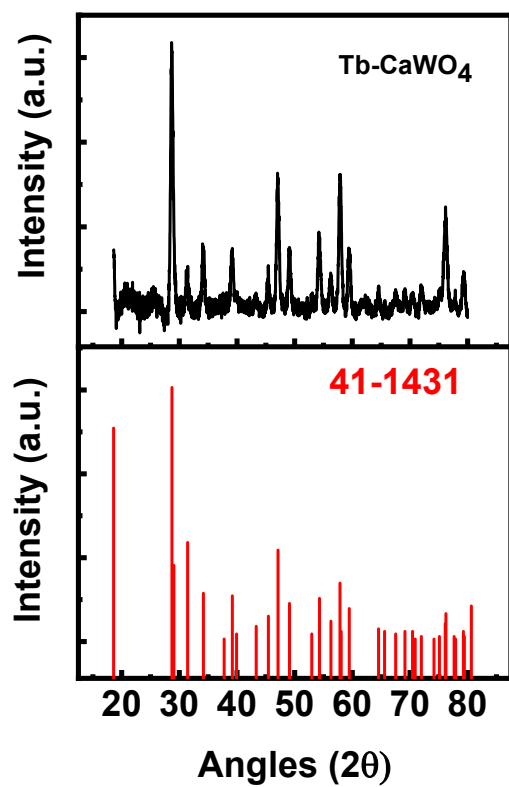


Fig. S5 XRD patterns of the Tb-CaWO₄ NCs and the standard diffraction pattern of CaWO₄ NCs.

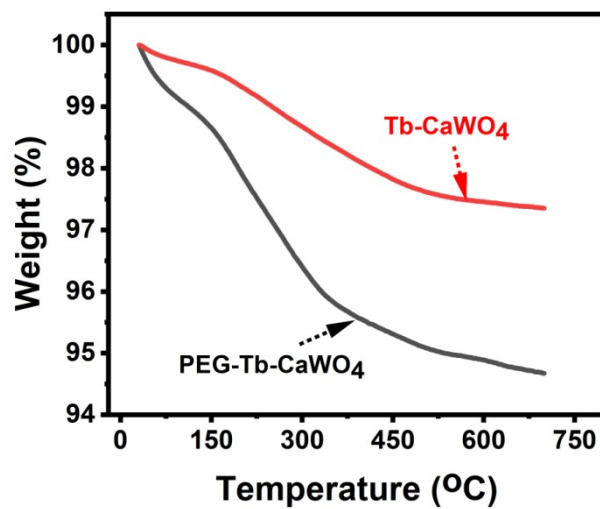


Fig. S6 Thermal gravimetric analysis (TGA) of bare and PEG coated Tb-CaWO₄ NCs.

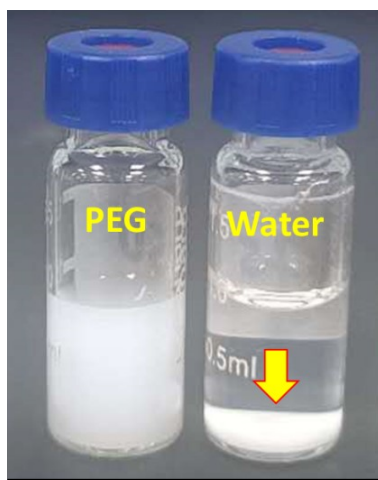


Fig. S7 A) Aqueous dispersion of PEG-coated (left) and uncoated (right) Tb-CaWO₄ NCs.

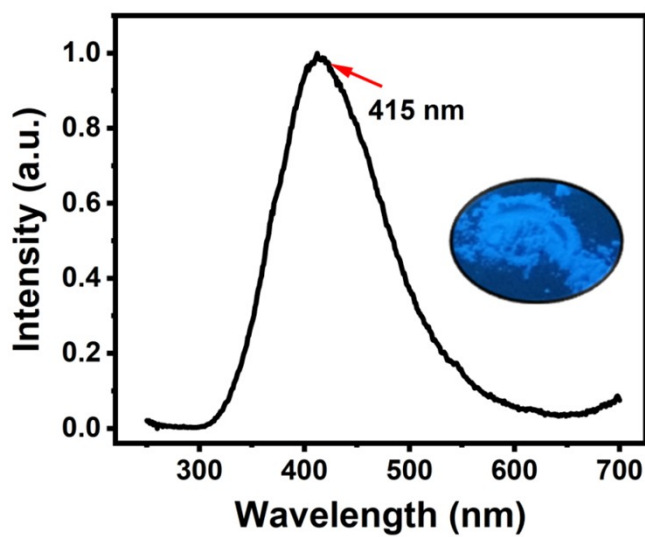


Fig. S8 Fluorescence emission of CaWO₄ and Inset: corresponding solid-state luminescence under excitation of 254 nm UV-light.

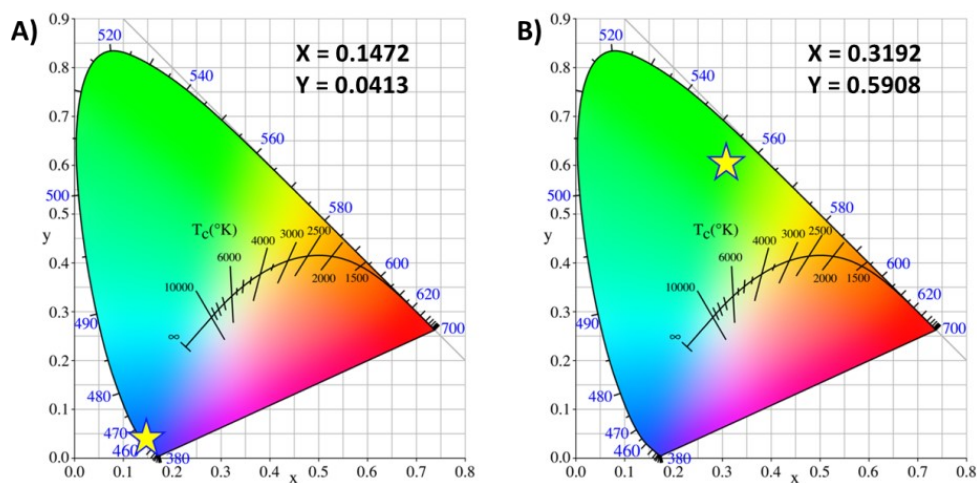


Fig. S9 CIE chromaticity diagram of A) CaWO_4 and B) PEG-Tb- CaWO_4 NCs. Insets show corresponding fluorescence emission coordinates.

Stability of PEG-Tb- CaWO_4 NCs

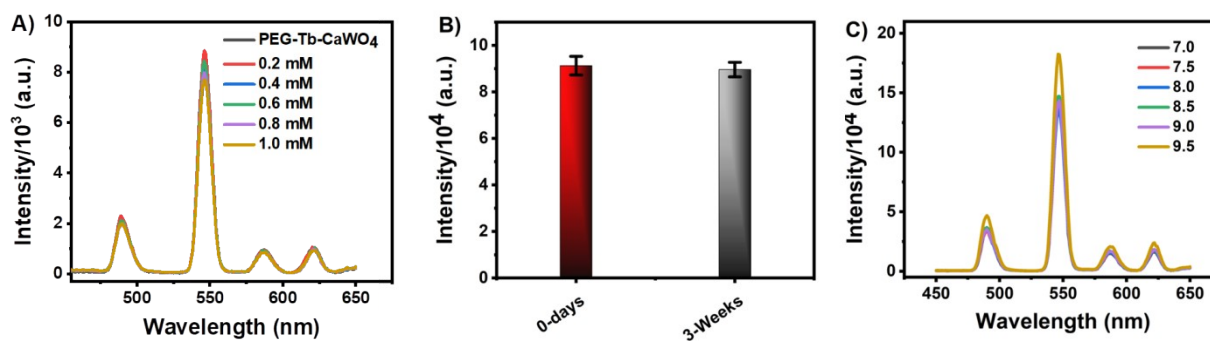


Fig. S10 Stability of the fluorescence intensity of PEG-Tb- CaWO_4 NCs against A) various ionic concentrations, B) storage time at 4°C, and C) various pH solutions.

Designing PEG-Tb-CaWO₄ NCs based sensor for ALP Detection

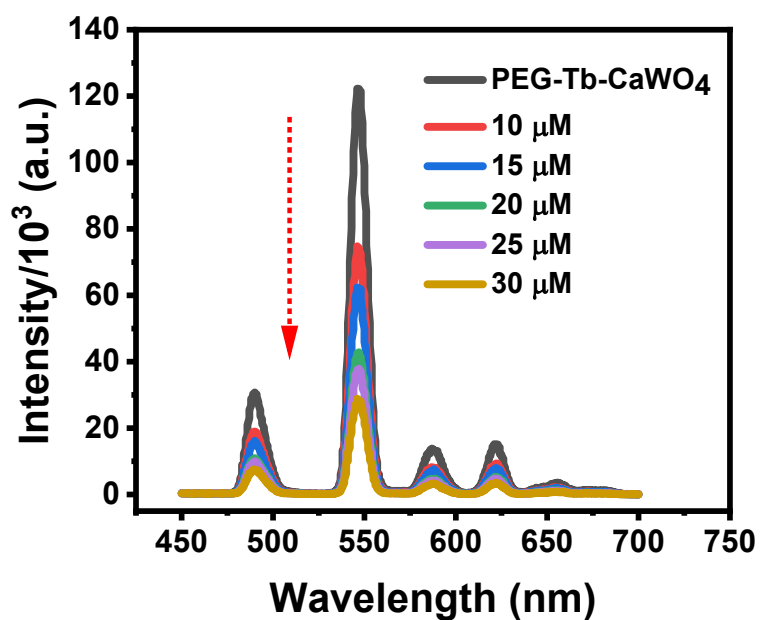


Fig. S11 Fluorescence emission intensity of PEG-Tb-CaWO₄ NCs with various concentrations of Ce⁴⁺ ions.

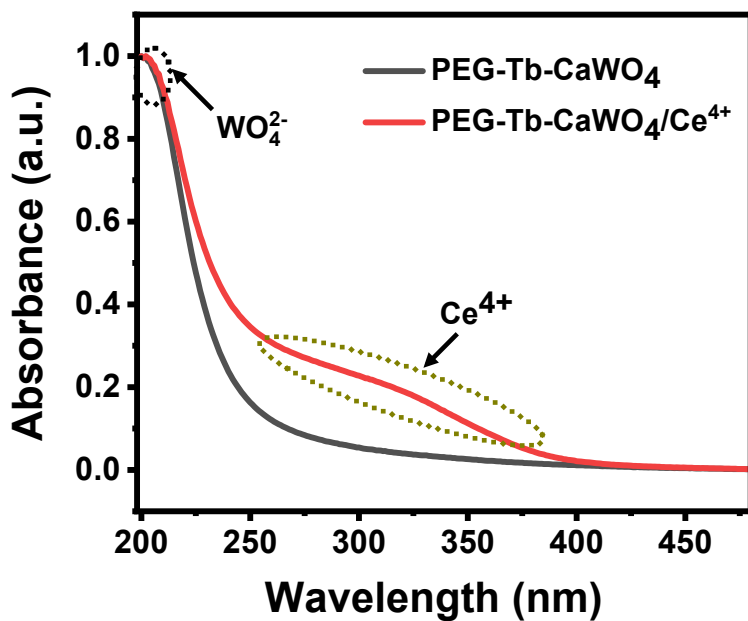


Fig. S12 UV-Visible absorption spectrum of PEG-Tb-CaWO₄ NCs with and without the addition of Ce⁴⁺ ions.

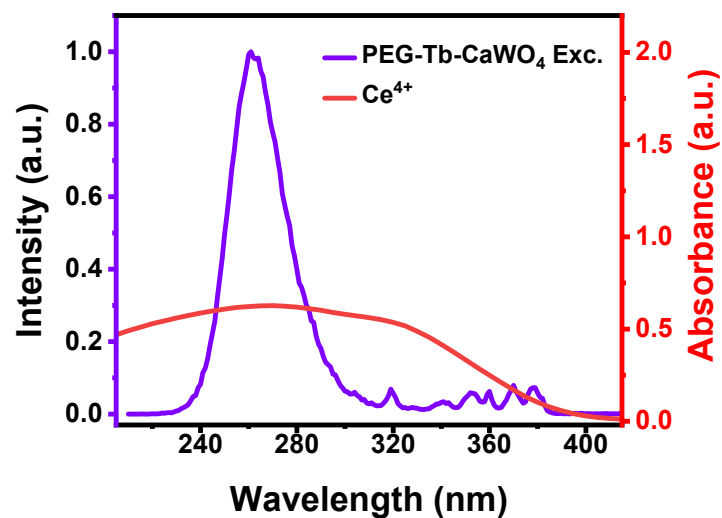


Fig. S13 Fluorescence excitation spectrum of PEG-Tb-CaWO₄ NCs and UV-visible absorption spectrum of Ce⁴⁺ ions.

Optimization of detection conditions

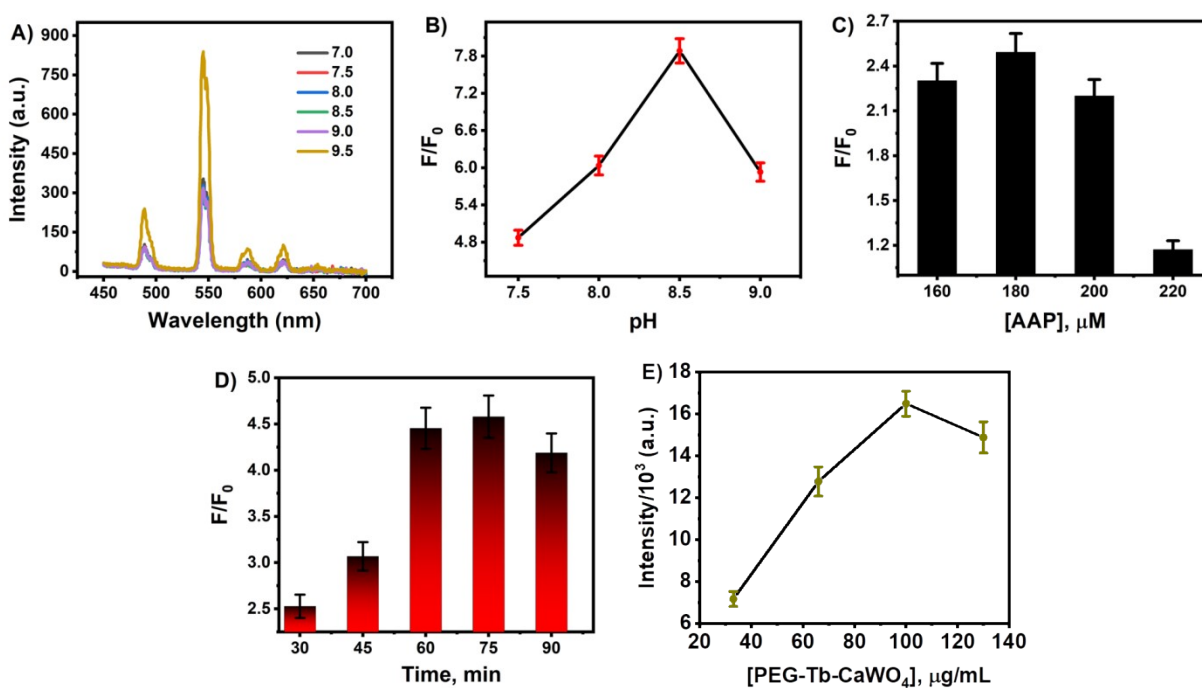


Fig. S14 Optimization of detection conditions A) Effect of pH on fluorescence quenching efficiency of Ce⁴⁺ ions in the absence of ALP. B) Effect of pH on the fluorescence recovery of the detection system in the presence of ALP. C) Concentration of AAP. D) Enzymatic reaction time. E) Amount of PEG-Tb-CaWO₄ NCs.

Optimization of ratio of AA-to-Ce⁴⁺

Redox reaction between Ce⁴⁺ ions and AA. ¹

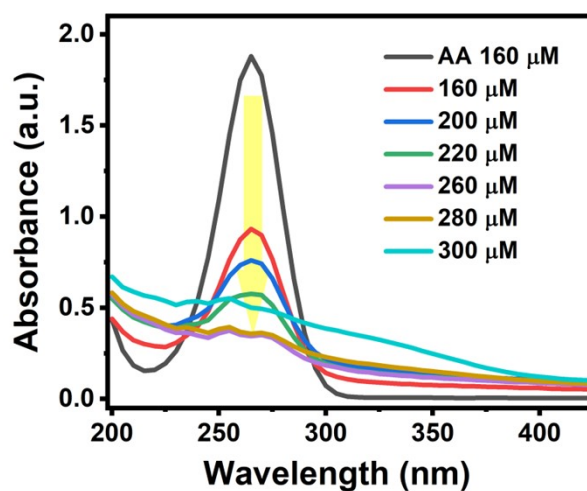
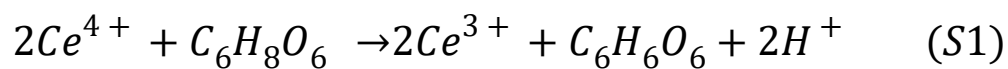


Fig. S15 UV-Vis absorption spectra showing optimization of the ratio (concentrations) of AA-to-Ce⁴⁺ ions.

Cell viability assay

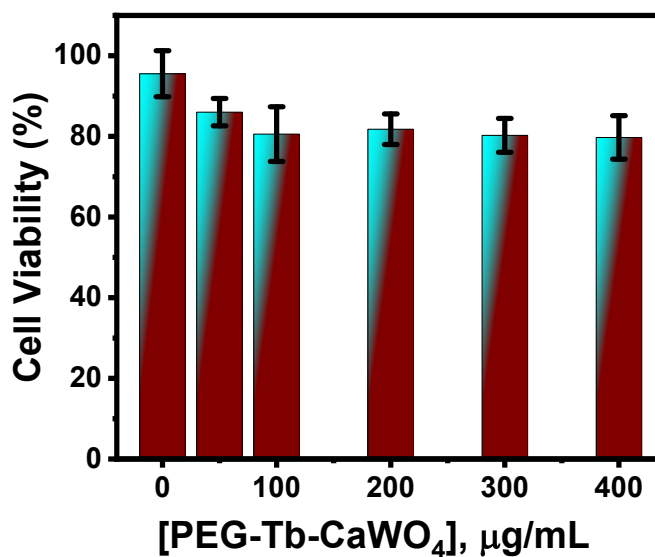


Fig. S16 MTT assay of HeLa cell viability after incubation with PBS (control) and PEG-Tb-CaWO₄ NCs at the concentration range of 0–400 μg/mL for 24 h.

Table S1. Comparison of ALP detection method

Method	Sensing Material	LOD (U/L)	Linear range (U/L)	Reference
Colorimetry	Fe/C NS	0.03	0.05 - 6.00	²
Colorimetry	Ag NPs	0.24	0.5 - 225	³
Electrochemistry	AuNPs/Ag ⁺	0.03	0.1 - 120	⁴
Electrochemistry	Luminol Si NPs	0.8	5 - 50	⁵
Fluorescence	Au/Ag NCs	0.193	0.5 - 10	⁶
Fluorescence	(PAA)-Ca ²⁺ (Ce ³⁺) NCs	0.62	5.0 - 200	⁷
Fluorescence	Au NCs-MnO ₂ NSs	1.5	5-800	⁸
Fluorescence	CQDs	5.5	18.2–1300	⁹
Fluorescence	E)-8-((4-methylbenzylidene) amino) naphthalen-1-amine (L)	50	400-3000	¹⁰
Fluorescence	Allosteric probe (AP)	12	20-150	¹¹
Fluorescence	Pyrococcus furiosus argonaute	2.7	10-1000	¹²
Fluorescence	PEG-Tb-CaWO ₄ NCs	0.5	6 - 40	This Work

Table S2. Determination of ALP in fetal bovine serum samples using the proposed method (n=3)

Sample	Added ALP (U/L)	Detected ALP (U/L)	Recovery (%)	RSD (%)
1	-	-	-	-
2	10	10.77 ± 0.02	107.68	1.61
3	16	13.83 ± 0.03	86.47	1.52
4	20	16.78 ± 0.05	83.89	2.11

References

1. N. K. Sahu, N. Shanta Singh, R. S. Ningthoujam and D. Bahadur, Ce^{3+} -Sensitized $\text{GdPO}_4\text{:Tb}^{3+}$ Nanorods: An Investigation on Energy Transfer, Luminescence Switching, and Quantum Yield, *ACS Photonics*, 2014, **1**, 337-346.
2. X. Chen, J. Chen, H.-Y. Zhang, F.-B. Wang, F.-F. Wang, X.-H. Ji and Z.-K. He, Colorimetric Detection of Alkaline Phosphatase on Microfluidic Paper-based Analysis Devices, *Chinese Journal of Analytical Chemistry*, 2016, **44**, 591-596.
3. S. M. Shaban, B.-S. Moon, D.-G. Pyun and D.-H. Kim, A colorimetric alkaline phosphatase biosensor based on p-aminophenol-mediated growth of silver nanoparticles, *Colloids and Surfaces B: Biointerfaces*, 2021, **205**, 111835.
4. J. Lei, J. Kang, J. Liu and G. Wang, A Novel Electrochemical Sensing Strategy Based on Poly (3,4-ethylenedioxythiophene): Polystyrene Sulfonate, AuNPs, and Ag^+ for Highly Sensitive Detection of Alkaline Phosphatase, *Nanomaterials*, 2022, **12**, 3392.
5. W. Qi, Y. Fu, M. Zhao, H. He, X. Tian, L. Hu and Y. Zhang, Electrochemiluminescence resonance energy transfer immunoassay for alkaline phosphatase using p-nitrophenyl phosphate as substrate, *Analytica Chimica Acta*, 2020, **1097**, 71-77.
6. M. Wang, X. Zhou, L. Cheng, M. Wang and X. Su, Lysozyme-Functionalized 5-Methyl-2-thiouracil Gold/Silver Nanoclusters for Luminescence Assay of Alkaline Phosphatase, *ACS Applied Nano Materials*, 2021, **4**, 9265-9273.
7. Y. Fan, Z. Ding, J. Chen, C. Li and Y. Han, Polyacrylic Acid- $\text{Ca}^{2+}(\text{Ce}^{3+})$ Nanoclusters as Fluorescent “Off–On–Off” Probes for Quantitative Detection of Pyrophosphate and Alkaline Phosphatase, *ACS Applied Nano Materials*, 2024, **7**, 5678-5688.
8. Y. Xue, C. Bao, H. Liu, F. Ma, M. Yang and X. Li, BSA-Assisted Synthesis of Au Nanoclusters/ MnO_2 Nanosheets for Fluorescence “Switch-On” Detection of Alkaline Phosphatase, 2025, **15**, 49.
9. Z. Qian, L. Chai, Q. Zhou, Y. Huang, C. Tang, J. Chen and H. Feng, Reversible Fluorescent Nanoswitch Based on Carbon Quantum Dots Nanoassembly for Real-Time Acid Phosphatase Activity Monitoring, *Analytical Chemistry*, 2015, **87**, 7332-7339.
10. S. Kiran, R. Khatik and R. Schirhagl, Smart probe for simultaneous detection of copper ion, pyrophosphate, and alkaline phosphatase in vitro and in clinical samples, *Analytical and Bioanalytical Chemistry*, 2019, **411**, 6475-6485.
11. J. Guo, M. Gao, Y. Song, L. Lin, K. Zhao, T. Tian, D. Liu, Z. Zhu and C. J. Yang, An Allosteric-Probe for Detection of Alkaline Phosphatase Activity and Its Application in Immunoassay, 2018, **6**, 618.
12. Z. Huang, L. Wei, Y. Zhou, Y. Li and Y. Chen, Guide DNA dephosphorylation-modulated *Pyrococcus furiosus* Argonaute fluorescence biosensor for the detection of alkaline phosphatase and aflatoxins B1, *Biosensors and Bioelectronics*, 2024, **265**, 116692.