### Supplemental Information

# Biocatalytic atroposelective synthesis of heterobiaryls and heterobiaryl *N*-oxides via dynamic kinetic resolution

Xinyue Hao,<sup>1,+</sup> Bin Wang,<sup>1,+</sup> Zhuangfei Tian,<sup>1</sup> Zhouchang Yao,<sup>1</sup> Tianzhang Qiao,<sup>2</sup> Ling Huang,<sup>1,\*</sup> and Haigen Fu<sup>1,\*</sup>

<sup>1</sup>NHC Key Laboratory of Biotechnology for Microbial Drugs, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing 100050, China. <sup>2</sup>Department of Chemistry and Chemical Biology, Cornell University, Ithaca, New York 14850, United States.

<sup>+</sup>These authors contributed equally to this work.

\*Address correspondence to Dr. Ling Huang. E-mail: huangling@imb.pumc.edu.cn \*Address correspondence to Prof. Haigen Fu. E-mail: fuhaigen@imb.pumc.edu.cn

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#### 1. General information

#### General

Unless otherwise noted, all chemicals and reagents for chemical reactions were obtained from commercial suppliers and used as received (Sigma-Aldrich, TCI, Admas, Innochem, Bidepharm). Silica gel chromatography purifications were carried out using Santai SepaFlash columns ( $40 \sim 63 \mu m$ , 60 Å). <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on JEOL ECZ-400S (400 MHz), Quantum-1 Plus (500 MHz), Bruker UltraShield Plus (600 MHz) instruments, and are internally referenced to residual proton signals in CDCl<sub>3</sub> (7.26 ppm for <sup>1</sup>H NMR and 77.16 ppm for <sup>13</sup>C NMR), DMSO-*d*<sub>6</sub> (2.50 ppm for <sup>1</sup>H NMR and 39.52 ppm for <sup>13</sup>C NMR) or CD<sub>3</sub>OD (3.31 ppm for <sup>1</sup>H NMR and 49.00 ppm for <sup>13</sup>C NMR).<sup>19</sup>F NMR spectra were reported unreferenced. <sup>1</sup>H NMR data are reported as follows: chemical shift ( $\delta$  ppm), multiplicity (s = singlet, brs = broad singlet, d = doublet, t = triplet, q = quartet, m = multiplet, dd = doublet of doublet, dt = doublet of triplet, ddd = doublet of doublet of doublet), coupling constant (Hz), and integration. High-resolution mass spectra (HRMS) were obtained on a Waters XEVO G2-XS using electrospray ionization time-of-flight (ESI-TOF).

#### Chromatography

Analytical high-performance liquid chromatography (HPLC) and Electron Spray Ionization (ESI) mass spectrometry were carried out using an Agilent 1200-6120 LCMS System. Yields and conversions were determined on a Poroshell C18 column (4.6 x 100 mm, 4.0 µm) against an internal standard 1,3,5-tribromobenzene (TBB) at 210 nm. Analytical chiral HPLC was carried out using an Agilent 1260 Infinity instrument equipped with ChiralPak AS-H, AD-H, OD-H, OJ-H (4.6 x 250 mm, 5.0 µm) columns with isopropanol and hexane as the mobile phase.

#### Cloning

pET28a (+) was used as a cloning and expression vector for all enzymes described in this study. Genes of imine-reductases were codon optimized for expression in *E. coli* and purchased from Sangon (Shanghai, China). All constructs were cloned directly in pET28a (+) between the *Ndel* and *Xhol* restriction sites. Cloned plasmids were transformed into *E. coli* DH5α cells for cell storage, and *E. coli* BL21 (DE3) chemically competent cells for protein expression.

#### **Protein and DNA Sequence**

(*S*)-Selective imine reductase from *Streptomyces sp.* GF3546 (*S-IRED-Ss*) GenBank accession number: WP\_319684944.1

### Wide-type S-IRED-Ss protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEALVARGAVLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKVIANLSSDTPDKAREAAKWAAKHGAKHLTGGVQVPPPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYRGEDAGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELL PYATMMTSMMPHFLELYAQHVDSADYPGDVDRLAMGAASVDHVLHTHQDAGVSTVLPAAVAEIFKA GMEKGFAENSFSSLIEVLKKPAV

### Wide-type S-IRED-Ss DNA sequence

ATGAGCAAACAGTCAGTTACGGTGATTGGTCTGGGTCCGATGGGTCAAAGCGATGGTCAATACCTT TCTGGATAATGGTCACGAAGTGACCGTGTGGAACCGTACGGCGTCAAAAGCAGAAGCTCTGGTG GCGCGCGGCGCAGTTCTGGCACCGACCGTCGAAGATGCTCTGAGCGCGCAATGAACTGATTGTTC TGTCTCTGACCGATTATGACGCCGTGTACGCAATCCTGGAACCGGTTACGGGCTCACTGTCGGGT AAAGTGATTGCAAACCTGAGCTCTGATACCCCGGACAAAGCGCGTGAAGCGGCCAAATGGGCAG CTAAACATGGTGCGAAACATCTGACCGGCGGTGTGCAGGTTCCGCCGCCGCTGATCGGCAAACC GGAAAGTTCCACCTATTACTCCGGTCCGAAAGATGTTTTTGACGCCCATGAAGATACCCTGAAAGT CCTGACGAACGCCGATTATCGTGGTGAAGATGCAGGTCTGGCCGCAATGTATTACCAGGCGCAAA TGACCATTTTCTGGACCACGATGCTGAGCTATTACCAGACGCTGGCTCTGGGCCAAGCGAATGGT GTTAGTGCTAAAGAACTGCTGCCGTATGCCACCATGATGACGTCCATGATGCCGCATTTTCTGGAA CTGTATGCTCAGCACGTCGATTCTGCGGACTATCCGGGTGATGTGGACCGTCTGGCGCATGGGCG CAGCTTCAGTCGATCACGTGCTGCATACCCACCAAGATGCGGGTGTTAGCACCGTCTGGCGATGGGCG CGCAGTGGCCGAAATCTTCAAAGCCGGTATGGAAAAAGGCTTTGCTGAAAATTCGTTCTCCTCT TGATTGAAGTCCTGAAAAAACCGGCAGTGTAA

## S-IRED-Ss-M9 protein sequence1

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQVPPPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPHFLELYAQHVDSADYPGDVDRLAMGAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

## S-IRED-Ss-M9 DNA sequence

ATGTCTAAACAGAGCGTTACCGTTATCGGCCTGGGTCCGATGGGTCAGGCGATGGTTAACACCT TCCTGGATAACGGCCACGAAGTTACCGTTTGGAACCGTACCGCGCTCTAAAGCGGAACCGCTGGT TGCTCGTGGCGCGCACCCTGGCGCCTACCGTTGAAGATGCGCTGAGCGCTAACGAACTGATCGT TCTGAGCCTGACCGATTACGATGCGGTGTACGCGATCCTGGAACCGGTTACCGGCAGCCTGAG CGGCAAAACCATCGTTAACCTGAGCAGCGATACCCCGGACAAAGCTCGTGAAATGGCGAAATGG GCGGCGAAACACGGCGCGAAACACCTGACCGGCGGCGTTCAGGTTCCGCCGCCGCTGATCGG TAAACCGGAAAGCAGCACCTACTACTCTGGCCCGAAAGATGTTTTCGATGCGCACGAAGATACC CTGAAAGTTCTGACCAACGCAGATTACCTGGGCCGAAGATCCGGGCCTGGCGGCAATGTACTACC AGGCGCAGATGACCATCTTCTGGACCACCATGCTGAGCTACTACCAGACCCTGGCGTAAGGTCA GGCTAACGGTGTGTCTGCAAAAGAACTGCTGCCGTACGCGACCATGACCACTCGTATGATCCG CATTTCCTGGAACTGTATGCACAGCACGTTGACAGCGCGGACTATCCGGGTGACGTTGATCGTC TGGCCATGGGTGCTGCCTCTGTTGATCATGTTCTGCATACCCACCACGGATGGCGGCGTTAGCAC CGTTCTGCCGGCTGCTGTTGCTGAGATCTTTAAAGCGGGCATGGAAAAAAAGGCTTCGCTGAAAAC TCTTTCAGCAGCCTGATCGAAGTTCTGAAAAAACCGGCGGTTTAA

### S-IRED-Ss-M12 protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQVPPPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPHFLELYAQHVDSADYPGDVDRLA<u>A</u>GAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

# S-IRED-Ss-M12 DNA sequence

ATGTCTAAACAGAGCGTTACCGTTATCGGCCTGGGTCCGATGGGTCAGGCGATGGTTAACACCTT CCTGGATAACGGCCACGAAGTTACCGTTTGGAACCGTACCGCGTCTAAAGCGGAACCGCTGGTT GCTCGTGGCGCGACCCTGGCGCCTACCGTTGAAGATGCGCTGAGCGGCTAACGAACTGATCGTTC TGAGCCTGACCGATTACGATGCGGTGTACGCGATCCTGGAACCGGTTACCGGCAGCCTGAGCGG CAAAACCATCGTTAACCTGAGCAGCGATACCCCGGACAAAGCTCGTGAAATGGCGAAATGGGCG GCGAAACACGGCGCGAAACACCTGACCGGCGGCGTTCAGGTTCCGCCGCCGCTGATCGGTAAA CCGGAAAGCAGCACCTACTACTCTGGCCCGAAAGATGTTTTCGATGCGCACGAAGATACCCTGAA AGTTCTGACCAACGCAGATTACCTGGGCGAAGATCCGGGCCTGGCGGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGAGGCTACTACCAGGCGCGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGACGCGACCATGACCACTCGTATGATGCCGCGCTAA CGGTGTGTCTGCAAAAGAACTGCTGCCGTACGCGACCATGACCACTCGTATGATGCCGCATTTCC TGGAACTGTATGCACAGCACGTTGACAGCGCGGACTATCCGGGTGACGTTGATGCTGCCGCC GGGTGCTGCCTCTGTTGATCATGTTCTGCATACCCACCAGGATGCGGGCGTTAGCACCGTTCGC CGGCTGCTGTTGCTGAGAACATCTTTAAAGCGGGCATGGAAAAAGGCTTCGCTGAAAACTCTTTCAGC AGCCTGATCGAAGTTCTGAAAAAACCGGCGGCATTAA

### S-IRED-Ss-M13 protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQVPPPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPH<u>W</u>LELYAQHVDSADYPGDVDRLA<u>A</u>GAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

# S-IRED-Ss-M13 DNA sequence

ATGTCTAAACAGAGCGTTACCGTTATCGGCCTGGGTCCGATGGGTCAGGCGATGGTTAACACCTT CCTGGATAACGGCCACGAAGTTACCGTTTGGAACCGTACCGCGCTAAAGCGGAACCGCTGGTT GCTCGTGGCGCGACCCTGGCGCCTACCGTTGAAGATGCGCTGAGCGCTAACGAACTGATCGTTC TGAGCCTGACCGATTACGATGCGGTGTACGCGATCCTGGAACCGGTTACCGGCAGCCTGAGCGG CAAAACCATCGTTAACCTGAGCAGCGATACCCCGGACAAAGCTCGTGAAATGGCGAAATGGGCG GCGAAACACGGCGCGAAACACCTGACCGGCGGCGTTCAGGTTCCGCCGCCGCTGATCGGTAAA CCGGAAAGCAGCACCTACTACTCTGGCCCGAAAGATGTTTTCGATGCGCACGAAGATACCCTGAA AGTTCTGACCAACGCAGATTACCTGGGCGAAGATCCGGGGCCTGGCGGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGAGCTACTACCAGACCCTGGCGTAAGATCACCAGGCG CTGGAACTGTATGCACAGCACCATGCTGCCGTACGCGACCATGACCATCGTTAGGTCAGGCTAA CGGTGTGTCTGCAAAAGAACTGCTGCCGTACGCGACCATGACCACTCGTATGATGCCGCAT<u>TGG</u> CTGGAACTGTATGCACAGCACGTTGACAGCGCGGACTATCCGGGTGACGTTGATCGTCTGGCCG CCGGTGCTGCTCTGTTGATCATGTTCTGCATACCCACCAGGATGCGGGCGTTAGCACCGTTCTG CCGGCTGCTGTTGCTGAGATCTTTAAAGCGGGCATGGAAAAAGGCTTCGCTGAAAACTCTTTCAG CAGCCTGATCGAAGTTCTGAAAAAACCGGCGGTTTAA

### S-IRED-Ss-M14 protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQV<u>H</u>PPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPH<u>W</u>LELYAQHVDSADYPGDVDRLA<u>A</u>GAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

# S-IRED-Ss-M14 DNA sequence

ATGTCTAAACAGAGCGTTACCGTTATCGGCCTGGGTCCGATGGGTCAGGCGATGGTTAACACCTT CCTGGATAACGGCCACGAAGTTACCGTTTGGAACCGTACCGCGTCTAAAGCGGAACCGCTGGTT GCTCGTGGCGCGACCCTGGCGCCTACCGTTGAAGATGCGCTGAGCGCTAACGAACTGATCGTTC TGAGCCTGACCGATTACGATGCGGTGTACGCGATCCTGGAACCGGTTACCGGCAGCCTGAGCGG CAAAACCATCGTTAACCTGAGCAGCGATACCCCGGACAAAGCTCGTGAAATGGCGAAATGGCGG GCGAAACACGGCGCGAAACACCTGACCGGCGGCGTTCAGGTT<u>CAT</u>CCGCCGCTGATCGGTAAA CCGGAAAGCAGCACCTACTACTCTGGCCCGAAAGATGTTTTCGATGCGCACGAAGATACCCTGAA AGTTCTGACCAACGCAGATTACCTGGGCGAAGATCCGGGCCTGGCGGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGAGGCTACTACCAGGCGCGCAATGTACTACCAGGCG CGGTGTGTCTGCAAAAGAACTGCTGCCGTACGCGACCATGACCACTCGTATGATGCCGCAT<u>TGG</u> CTGGAACTGTATGCACAGCACGTTGACAGCGCGGACCATGACCACTCGTATGATGCCGCAT<u>TGG</u> CCGGTGCTGCCTCTGTTGATCATGTTCTGCATACCCACCAGGATGCGGGCGTTAGCACCGTTCG CCGGCTGCTGCTGCTGAGATCTTTAAAGCGGGCATGGAAAAAGGCTTCGCTGAAAACTCTTTCAG CAGCCTGATCGAAGTTCTGAAAAAACCGGCGGGTTTAA

### S-IRED-Ss-M15 protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQV<u>V</u>PPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPHFLELYAQHVDSADYPGDVDRLA<mark>A</mark>GAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

# S-IRED-Ss-M15 DNA sequence

ATGTCTAAACAGAGCGTTACCGTTATCGGCCTGGGTCCGATGGGTCAGGCGATGGTTAACACCTT CCTGGATAACGGCCACGAAGTTACCGTTTGGAACCGTACCGCGTCTAAAGCGGAACCGCTGGTT GCTCGTGGCGCGACCTGGCGCCTACCGTTGAAGATGCGCGTGACCGGCTAACGAACTGATCGTTC TGAGCCTGACCGATTACGATGCGGTGTACGCGATCCTGGAACCGGTTACCGGCAGCCTGAGCGG CAAAACCATCGTTAACCTGAGCAGCGATACCCCGGACAAAGCTCGTGAAATGGCGAAATGGGCG GCGAAACACGGCGCGAAACACCTGACCGGCGGCGTTCAGGTTGTTCCGGCACGAAATGGCGAAA CCGGAAAGCAGCACCTACTACTCTGGCCCGAAAGATGTTTTCGATGCGCACGAAGATACCCTGAA AGTTCTGACCAACGCAGATTACCTGGGCGAAGATCCGGGCCTGGCGGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGAGGCTACTACCAGGCGCGCAATGTACTACCAGGCG CAGATGACCATCTTCTGGACCACCATGCTGACGCGACCATGACCACTCGTATGATGCCGCACTAA CGGTGTGTCTGCAAAAGAACTGCTGCCGTACGCGACCATGACCACTCGTATGATGCCGCATTTCC TGGAACTGTATGCACAGCACGTTGACAGCGCGGACTATCCGGGTGACGTTGATGCTGCCGCC GGGTGCTGCCTCTGTTGATCATGTTCTGCATACCCACCAGGATGCGGGCGTTAGCACCGTTCGC CGGCTGCTGTTGCTGAGATCTTTAAAGCGGGCCATGGAAAAAGGCTTCGCTGAAAACTCTTTCAGC AGCCTGATCGAAGTTCTGAAAAAACCGGCGGCGTTAA

### S-IRED-Ss-M16 protein sequence

MSKQSVTVIGLGPMGQAMVNTFLDNGHEVTVWNRTASKAEPLVARGATLAPTVEDALSANELIVLSL TDYDAVYAILEPVTGSLSGKTIVNLSSDTPDKAREMAKWAAKHGAKHLTGGVQV<u>V</u>PPLIGKPESSTYY SGPKDVFDAHEDTLKVLTNADYLGEDPGLAAMYYQAQMTIFWTTMLSYYQTLALGQANGVSAKELLP YATMTTRMMPHFLELYAQHVDSADYPGD<u>T</u>DRLA<u>A</u>GAASVDHVLHTHQDAGVSTVLPAAVAEIFKAG MEKGFAENSFSSLIEVLKKPAV

# S-IRED-Ss-M16 DNA sequence

#### **IREDs Protein Expression and Purification**

(*S*)-Selective imine reductase from *Streptomyces sp.* GF3546 (*S*-IRED-*Ss*) was produced in *E. coli* BL21 with a pET28a-*S*-IRED-*Ss* plasmid. Transformed glycerol stocks were used to initiate a 5 mL Luria-Bertani (LB) media with kanamycin (50 µg/mL) at 37 °C and 250 rpm overnight. Expression culture (500 mL in a 2 L baffled shake flask) containing kanamycin (50 µg/mL) was inoculated with 3 mL of the overnight culture and grown until the culture reached an OD<sub>600</sub> of 0.5 ~ 0.7 at 37 °C and 250 rpm. Flasks were chilled on ice for 10 min and protein expression was induced with 0.2 mM IPTG (20 °C, 24 h, 250 rpm). The cells were harvested by centrifugation (7,100 x g, 20 min, 4 °C), and frozen at -80 °C for further purification.

Frozen cells were thawed on ice and resuspended in buffer A (20 mM KPi, pH 7.0, 250 mM NaCl, 25 mM imidazole, with 1 mg/mL lysozyme, 0.1 mg/mL DNase I, 1 mM phenylmethylsulfonyl fluoride) to a final concentration of 2 mL/g of wet cells. The cells were disrupted by sonication (20 min, output 30 W, Ultrasonic Homogenizer JY92-IIN) under an ice bath. Lysates were centrifuged (24,200 x *g*, 1 h, 4 °C) to pellet insoluble materials. Proteins were purified by affinity chromatography with a nickel-NTA column. After washing with 10 column volumes of buffer A, enzymes were eluted with buffer B (20 mM KPi, pH 7.0, 250 mM NaCl, 250 mM imidazole) over 5 column volumes. Fractions containing IRED enzymes were pooled, concentrated, and subjected to three buffer exchanges into an imidazole-free storage buffer (100 mM KPi, pH 8.0). Concentrated enzymes were aliquoted, flash-frozen in liquid N<sub>2</sub>, and then stored at -20 °C until later use. Protein purity was assessed with SDS-PAGE. Protein concentrations were determined using absorbance at 280 nm with an extinction coefficient of 34,380 mM<sup>-1</sup> cm<sup>-1</sup>.

For the preparation of lyophilized cell-free extracts (CFE), frozen cells were thawed on ice and resuspended in Tris-HCl buffer (50 mM, pH 8.0) to a final concentration of 5 mL/g of wet cells. The cells were disrupted by sonication (20 min, output 30 W, Ultrasonic Homogenizer JY92-IIN) under an ice bath. Lysates were centrifuged (24,200 x g, 1 h, 4 °C) to pellet insoluble materials. The supernatant was transferred to a clean centrifuge tube, and then frozen instantly by liquid N<sub>2</sub>. Lyophilization was carried out on Alpha 1-2 LDplus (Christ, Germany).

The glucose dehydrogenase (GDH) used in this study was an engineered variant of glucose dehydrogenase from *Bacillus megaterium* (*Bm*GDH<sub>M6</sub>), which was obtained from published work by Zheng *et. al.*<sup>2</sup> The specific activity (69 U/mg) of the *Bm*GDH<sub>M6</sub> lyophilized CFE (thereafter named as *Bm*GDH) was assayed at 25 °C by monitoring the increase of the absorbance of NADPH at 340 nm using Agilent Cary 60 UV-Vis (Agilent, America).

#### 2. Supplemental experimental procedures

Table S1. The initial panel of enzymes screened for synthesis of 8-aryl quinoline 7.



Entry	Enzymes	Accession number	Source	Yield <sup>a</sup>	er <sup>b</sup>
1	IRED-1	WP_166633080	WP_166633080 Streptomyces sp. GC420		n.d.¢
2	IRED-2	WP_150514620.1	Streptomyces spectabilis	3	n.d.
3	IRED-3	WP_205760981.1	Luteolibacter luteus	3	n.d.
4	IRED-4	WP_088993565.1	Micromonospora echinaurantiaca	4	n.d.
5	IRED-5	WP_179754454.1	Microlunatus parietis	1	n.d.
6	S-IRED-Ss	WP_319684944.1	Streptomyces sp.	20	65:35
7	S-IRED-Ss-M9 <sup>d</sup>		Streptomyces sp.	30	93:7
8	R-IRED-Ss	WP_015610874.1	Streptomyces sp.	0	n.d.
9	IRED-8	WP_189823582	Streptomyces finlayi	4	n.d.
10	IRED-9	WP_011731218	Mycolicibacterium smegmatis	16	82:18
11	IRED-10	WP_207513941	Longitalea luteola	3	n.d.
12	<i>Ad</i> RedAm	XP_045278909.1	Ajellomyces dermatitidis	0	n.d.
13	<i>Af</i> RedAm	XP_748217.1	Aspergillus fumigatus	5	n.d.
14	No IRED			0	n.d.
15	No turnover system			0	n.d.

Reaction conditions: substrate **5a** (5  $\mu$ mol, 1 equiv), benzylamine **6a** (15  $\mu$ mol, 3 equiv), NADP<sup>+</sup> (0.1  $\mu$ mol, 2 mol%), glucose (15  $\mu$ mol, 3 equiv), *Bm*GDH (1 mg/mL), purified IREDs (entry 1~11) or reductive aminases (entry 12~13) (0.2 mol% based on **5a**) in Tris-HCl buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000  $\mu$ L. <sup>a</sup> Yield (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene (TBB). <sup>b</sup> Enantiomeric ratio (er, *S:R*) determined by HPLC on a chiral stationary phase. <sup>c</sup> n.d., not determined. <sup>d</sup> *S*-IRED-*Ss*-M9 is an engineered variant with nine mutations (A41P/V48T/V88T/A90V/A103M/R158L/A162P/M207T/S209R) based on *S*-IRED-*Ss*.



Table S2. The initial panel of enzymes screened for synthesis of biaryl pyridine N-oxide 29.

Entry	Enzymes	Accession number	Source	Yield <sup>a</sup>	er <sup>b</sup>
1	IRED-1	WP_166633080	Streptomyces sp. GC420	0	n.d.¢
2	IRED-2	WP_150514620.1	Streptomyces spectabilis	6	n.d.
3	IRED-3	WP_205760981.1	Luteolibacter luteus	0	n.d.
4	IRED-4	WP_088993565.1	Micromonospora echinaurantiaca	0	n.d.
5	IRED-5	WP_179754454.1	Microlunatus parietis	0	n.d.
6	S-IRED-Ss	WP_319684944.1	Streptomyces sp.	12	99:1
7	S-IRED-Ss-M9 <sup>d</sup>		Streptomyces sp.	11	99:1
8	R-IRED-Ss	WP_015610874.1	Streptomyces sp.	0	n.d.
9	IRED-8	WP_189823582	Streptomyces finlayi	0	n.d.
10	IRED-9	WP_011731218	Mycolicibacterium smegmatis	0	n.d.
11	IRED-10	WP_207513941	Longitalea luteola	0	n.d.
12	<i>Ad</i> RedAm	XP_045278909.1	Ajellomyces dermatitidis	0	n.d.
13	<i>Af</i> RedAm	XP_748217.1	Aspergillus fumigatus	0	n.d.
14	No IRED			0	n.d.
15	No turnover system			0	n.d.

Reaction conditions: substrate **28a** (5 µmol, 1 equiv), benzylamine **6a** (15 µmol, 3 equiv), NADP<sup>+</sup> (0.1 µmol, 2 mol%), glucose (15 µmol, 3 equiv), *Bm*GDH (1 mg/mL), purified IREDs (entry 1~11) or reductive aminases (entry 12~13) (0.2 mol% based on **28a**) in Tris-HCl buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000 µL. <sup>*a*</sup> Yield (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. <sup>*b*</sup> Enantiomeric ratio (er, *S:R*) determined by HPLC on a chiral stationary phase. <sup>*c*</sup> n.d., not determined. <sup>*d*</sup> S-IRED-*Ss*-M9 is an engineered variant with nine mutations (A41P/V48T/V88T/A90V/A103M/R158L/A162P/M207T/S209R) based on *S*-IRED-*Ss*.

#### Directed evolution of S-IRED-Ss-M9

#### Site Saturation Mutagenesis of S-IRED-Ss-M9

Site saturation mutagenesis primers were designed using the PCR protocol from Kille *et al.*<sup>3</sup> Plasmid encoding *S*-IRED-*Ss*-M9 was used as the template, and PCR products were digested with *DpnI*, and then used to directly transform *E. coli* BL21 (DE3) chemically competent cells and plated on LB agar plates containing kanamycin (50 µg/mL). Sites examined based on *S*-IRED-*Ss*-M9 were V119, P122, P123, Y169, Q172, M173, F176, W177, M210, F214, V231, and M236 (Fig. S1).

#### Site Saturation Library Construction and Protein Expression.

Single colonies were picked with sterile toothpicks and used to inoculate 400  $\mu$ L of Luria-Bertani (LB) media containing kanamycin (50  $\mu$ g/mL) in deep-well 96-well plates, and cultured overnight (37 °C, 250 rpm). Wells A2, B4, C6, D8, E10, F12, G2, and H4 were embedded controls of the parent protein for engineering. Well H12 was used as blank control. A glycerol stock plate of the library was prepared by mixing sterilized glycerol solution (30% *v*/*v*, 50  $\mu$ L/well) with the overnight cell cultures (50  $\mu$ L/well). The library was sealed and stored at -80 °C. The expression cultures (1000  $\mu$ L of LB media containing 50  $\mu$ g/mL of kanamycin) in 96-well plates were inoculated by adding 50  $\mu$ L of the overnight cultures, and grown for 4 hours (37 °C, 250 rpm). Protein expression was induced with 0.2 mM IPTG (20 °C, 24 h, 250 rpm) in the 96-well plate. Cells were harvested by centrifugation at 7,100 x *g*, 20 min, 4 °C, and frozen at -20 °C overnight for later screening.

#### General procedure for screening in 96-well plates.

The cell libraries were thawed and resuspended with lysis buffer (150 µL/well) and allowed to shake for 1 hour at 30 °C. The crude cell lysates were centrifuged (7,100 x *g*, 20 min, 4 °C) and the supernatants (50 µL/well) were transferred into an optical 96-well plate with clear flat bottom for screening assay immediately, 130 µL/well of Tris-HCl buffer (100 mM, pH 8.0) and 20 µL/well of reaction mixture containing NADPH (0.3 mM), substrates (0.63 mM of **5a** or **28a** and 1.9 mM of benzylamine **6a**) and 6% DMSO. The reaction progress was monitored by measuring the depletion of the NADPH absorption at 340 nm for 30 min by Microplate Reader (Biotek Synergy HTX, America). <u>Recipe for lysis buffer</u>. Tris-HCl buffer (100 mM, pH 8.0) containing DNase I (0.1 mg/mL), lysozyme (1 mg/mL), and PMSF (2 mM).

Mutations that displayed an improved initial rate of consuming NADPH than that of the parent enzyme were selected for further confirmation. Promising mutations were subjected to cultivation in shaking flasks and purified *S*-IRED-*Ss* mutants were used in the standard reaction condition for further confirmation of yield and er. The DNA plasmids of hits were identified by gene sequencing.

Standard reaction condition to confirm hits: substrate 5a or 28a (5  $\mu$ mol, 1 equiv), benzylamine 6a (15  $\mu$ mol, 3 equiv), NADP<sup>+</sup> (0.1  $\mu$ mol, 2 mol%), glucose (15  $\mu$ mol, 3 equiv), *Bm*GDH (1 mg/mL), purified *S*-IRED-*Ss* variants (0.2 mol%) in Tris-HCI buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature for 24 h, the final total volume is 1000  $\mu$ L. For yield determination, the reaction was quenched with 3 mL of acetonitrile and 100  $\mu$ L of 5 mg/mL 1,3,5-tribromobenzene (TBB) in acetonitrile

as the internal standard. The mixture was shaken for 30 min, centrifuged at 13,400 x g for 5 min, and the supernatant was filtered and retained for LCMS analysis (30-95% ACN/H<sub>2</sub>O with 0.1% formic acid, 8 minutes) for yield calculation. After LCMS analysis, the supernatant was concentrated under reduced pressure, extracted with EtOAc, and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting crude residue was dissolved in 30% isopropanol/hexanes (v/v) for chiral HPLC analysis.

For the first part of protein engineering, we focused on 8-aryl quinoline aldehyde **5a** as substrate, three rounds of iterative site-saturation mutagenesis (ISM) have been conducted using S-IRED-Ss-M9 as the starting point, yielding a mutant **S-IRED-Ss-M14 (M236A/F214W/P122H)** as the final biocatalyst for efficient and asymmetric synthesis of 8-aryl quinoline amine **7**.

For the second part of protein engineering, we focused on biaryl pyridine *N*-oxide aldehyde **28a** as substrate, three rounds of ISM have been conducted using *S*-IRED-*Ss*-M9 as the starting point, yielding a mutant *S*-IRED-*Ss*-M16 (M236A/P122V/V231T) as the final biocatalyst for efficient and asymmetric synthesis of biaryl pyridine *N*-oxide amine **29**.



**Fig. S1**. Structure of S-IRED-Ss-WT (PDB:40QY) and twelve amino acid residues (V119, P122, P123, Y169, Q172, M173, F176, W177, M210, F214, V231, and M236.) were examined during directed evolution. Pymol was used for graph preparation.



**Fig. S2**. Iterative site-saturation mutagenesis of *S*-IRED-*Ss*-M9 for the synthesis of *N*-heterobiaryl amine **7**. (A) The first round of site-saturation mutagenesis of *S*-IRED-*Ss*-M9. Sites Y169, Q172, M173, F176, W177, M210, and M236 were examined (Salmon balls). (B) The second round of engineering of *S*-IRED-*Ss*-M9. Sites P122, F176, M210 and F214 were examined (Salmon balls) using *S*-IRED-*Ss*-M12 (M236A, purple ball) as parent protein. (C) The third round of engineering of *S*-IRED-*Ss*-M13 (M236A/F214W, purple balls) as parent protein. (D) Final engineered *S*-IRED-*Ss*-M14 variant (M236A/F214W/P122H, purple balls) based on *S*-IRED-*Ss*-M9.

M	е СНО	S-IRED-Ss variants (0.2 mol%) NADP <sup>+</sup> /GDH/glucose		l Sn
	6a	Tris-HCl (100 mM, pH 8.0) 6% DMSO, rt, 24 h		
	rac- <b>5a</b>		7	
Entry	S-IRED variants	Mutations	Yield <sup>a</sup>	er <sup>b</sup>
1	S-IRED-Ss-WT	none	20%	65:35
2	S-IRED-Ss-M9	441P/V48T/V88T/A90V/A103M/R	30%	93:7
		158L/A162P/M207T/S209R		
3	S-IRED-Ss-M12	M9+M236A	37%	99:1
4	S-IRED-Ss-M13	M12+F214W	82%	99:1
5	S-IRED-Ss-M14	M13+P122H	98%	99:1

Table S3. Summary of the site-saturation mutagenesis results for heterobiaryl amine 7.

Reaction conditions: **5a** (5  $\mu$ mol, 1 equiv), benzylamine **6a** (15  $\mu$ mol, 3 equiv), NADP<sup>+</sup> (0.1  $\mu$ mol, 2 mol%), glucose (15  $\mu$ mol, 3 equiv), *Bm*GDH (1 mg/mL), purified *S*-IRED variants (0.2 mol%) in Tris-HCI buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000  $\mu$ L. <sup>*a*</sup> Yield (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. <sup>*b*</sup> Enantiomeric ratio (er, *S*:*R*) determined by HPLC on a chiral stationary phase.

Me	S-IF	RED- <i>Ss</i> variants (0.2 mol%) NADP <sup>+</sup> /GDH/glucose	Me H Bn
Ĺ	6a T	ris-HCl (100 mM, pH 8.0) 6% DMSO, rt, 24 h	
rac	-5a		7
Entry	S IPED. So veriente	Initial velocity <sup>a</sup>	Specific activity <sup>b</sup>
Enuy	S-IRED-38 Variants	(µmol/min)	(U/mg)
1	S-IRED-Ss-WT	$0.0022 \pm 0.0007$	0.0070 ± 0.0023
2	S-IRED-Ss-M9	0.0014 ± 0.0005	0.0050 ± 0.0018
3	S-IRED-Ss-M12	0.0048 ± 0.0005	0.0155 ± 0.0019
4	S-IRED-Ss-M13	0.0101 ± 0.0001	0.0325 ± 0.0003
5	S-IRED-Ss-M14	0.0667 ± 0.0022	0.2150 ± 0.0073
6	M14 lyophilized CFE	0.0466 ± 0.0020	0.0116 ± 0.0005

Table S4. The specific activity of S-IRED-Ss variants for the model reaction of heterobiaryl amine 7.

Reaction conditions: **5a** (5 µmol, 1 equiv), benzylamine **6a** (15 µmol, 3 equiv), NADP<sup>+</sup> (0.1 µmol, 2 mol%), glucose (15 µmol, 3 equiv), *Bm*GDH (1 mg/mL), *S*-IRED-*Ss* variants (0.2 mol%) or lyophilized CFE of *S*-IRED-*Ss*-M14 (4.0 mg/mL) in Tris-HCl buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature, the final total volume is 1000 µL. Yield (triplicate) of **7** was determined *via* LCMS relative to an internal standard (TBB). **Note**: <sup>*a*</sup> Initial rate of the enzymatic reaction was calculated over the period of 0–90 min for the purified *S*-IRED-*Ss*-M14 and its lyophilized CFE. <sup>*b*</sup> For calculation of specific activity, one unit (µmol/min) was defined as the amount of biocatalyst required for forming 1 µmol of product **7** per minute.



**Fig. S3**. Iterative site-saturation mutagenesis of *S*-IRED-*Ss*-M9 for the synthesis of biaryl pyridine *N*-oxide **29**. (A) The first round of site-saturation mutagenesis of *S*-IRED-*Ss*-M9. Sites Y169, Q172, M173, F176, W177, M210, and M236 were examined (Salmon balls). (B) The second round of engineering of *S*-IRED-*Ss*-M9. Sites P122, F176, M210 and F214 were examined (Salmon balls) using *S*-IRED-*Ss*-M9. Sites V12, M173, M12 (M236A, purple ball) as parent protein. (C) The third round of engineering of *S*-IRED-*Ss*-M15 (M236A/P122V, purple balls) as parent protein. (D) Final engineered *S*-IRED-*Ss*-M16 variant (M236A/P122V/V231T, purple balls) based on *S*-IRED-*Ss*-M9.

Me	СНО СНО	S-IRED-Ss variants (0.2 mol%) NADP <sup>+</sup> /GDH/glucose	Me	H N Bn
		Tris-HCl (100 mM, pH 8.0) 6% DMSO, rt, 24 h		
	rac- <b>28a</b>		29	
Entry	S-IRED variants	Mutations	Yield <sup>a</sup>	er <sup>b</sup>
1	S-IRED-Ss-WT	none	12%	99:1
2	S-IRED-Ss-M9	A41P/V48T/V88T/A90V/A103M/R	11%	99:1
		158L/A162P/M207T/S209R		
3	S-IRED-Ss-M12	M9+M236A	32%	99:1
4	S-IRED-Ss-M15	M12+P122V	92%	99:1
5	S-IRED-Ss-M16	M15+V231T	98%	99:1

Table S5. Summary of site-saturation mutagenesis results for synthesis of biaryl pyridine N-oxide 29.

Reaction conditions: **28a** (5  $\mu$ mol, 1 equiv), benzylamine **6a** (15  $\mu$ mol, 3 equiv), NADP<sup>+</sup> (0.1  $\mu$ mol, 2 mol%), glucose (15  $\mu$ mol, 3 equiv), *Bm*GDH (1 mg/mL), purified *S*-IRED variants (0.2 mol%) in Tris-HCI buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000  $\mu$ L. <sup>*a*</sup> Yield (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. <sup>*b*</sup> Enantiomeric ratio (er, *S*:*R*) determined by HPLC on a chiral stationary phase.

Me	CHO S-IRED-S	s variants (0.2 mol%) <sup>D+</sup> /GDH/glucose M	e H <sub>N</sub> <sub>Bn</sub>
	<b>6a</b> Tris-HC	l (100 mM, pH 8.0) DMSO, rt, 24 h	
rac	-28a		29
Entry	S IRED Se variante	Initial velocity <sup>a</sup>	Specific activity <sup>b</sup>
Linu y	S-INED-55 Valiants	(µmol/min)	(U/mg)
1	S-IRED-Ss-WT	0.0006± 0.0002	0.0020 ± 0.0007
2	S-IRED-Ss-M9	0.0004 ± 0.0005	0.0013 ± 0.0002
3	S-IRED-Ss-M12	0.0010 ± 0.0001	0.0031 ± 0.0004
4	S-IRED-Ss-M15	0.0126 ± 0.0009	0.0408 ± 0.0030
5	S-IRED-Ss-M16	0.0237 ± 0.0011	0.0764 ± 0.0035
6	M16 lyophilized CFE	0.0158 ± 0.0013	0.0511 ± 0.0003

Table S6. The specific activity of S-IRED-Ss variants for the model reaction of pyridine N-oxide 29.

Reaction conditions: **28a** (5 µmol, 1 equiv), benzylamine **6a** (15 µmol, 3 equiv), NADP<sup>+</sup> (0.1 µmol, 2 mol%), glucose (15 µmol, 3 equiv), *Bm*GDH (1 mg/mL), purified *S*-IRED-*Ss* variants (0.2 mol%) or lyophilized CFE of *S*-IRED-*Ss*-M16 (4.5 mg/mL) in Tris-HCI buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature, the final total volume is 1000 µL. Yield (triplicate) of **29** was determined *via* LCMS relative to an internal standard (TBB). **Note**: <sup>a</sup> Initial rate of the enzymatic reaction was calculated over the period of 0–120 min for the purified *S*-IRED-*Ss*-WT and *S*-IRED-*Ss*-M9, the period of 0–90 min for *S*-IRED-*Ss*-M12 and M15 variants or the period of 0–60 min for the purified *S*-IRED-*Ss*-M16 and its lyophilized CFE. <sup>b</sup> For calculation of specific activity, one unit (µmol/min) was defined as the amount of biocatalyst required for forming 1 µmol of product **29** per minute.



**Fig. S4**. SDS-PAGE analysis of the purified *S*-IRED-*Ss* variants for the synthesis of 8-aryl quinoline. M: protein marker; lane 1, *S*-IRED-*Ss*-WT; lane 2, *S*-IRED-*Ss*-M9; lane 3, *S*-IRED-*Ss*-M12; lane 4, *S*-IRED-*Ss*-M13; lane 5, *S*-IRED-*Ss*-M14.



**Fig. S5**. SDS-PAGE analysis of the purified *S*-IRED-*Ss* variants for the synthesis of biaryl pyridine *N*-oxide. M: protein marker; lane 1, *S*-IRED-*Ss*-WT; lane 2, *S*-IRED-*Ss*-M9; lane 3, *S*-IRED-*Ss*-M12; lane 4, *S*-IRED-*Ss*-M15; lane 5, *S*-IRED-*Ss*-M16.

#### Enzymatic atroposelective DKR for the synthesis of N-heterobiaryl amines



#### General procedure 1 for analytical scale reaction

A screw vial (4 mL) was charged with *Bm*GDH (200  $\mu$ L, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (54  $\mu$ L, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP+ (15  $\mu$ L, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), S-IRED-Ss-M14 (0.2 mol% or 0.4 mol% catalyst loading), aldehyde (30  $\mu$ L, 167 mM stock in DMSO, 5  $\mu$ mol, 1 equiv) and amine (30  $\mu$ L, 500 mM stock in DMSO, 15  $\mu$ mol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 1000  $\mu$ L with 6% DMSO ( $\nu/\nu$ ) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 3 mL of acetonitrile and 100  $\mu$ L of 5 mg/mL 1,3,5-tribromobenzene (TBB) in acetonitrile as the internal standard. The mixture was shaken for 30 min, centrifuged (13,400 x g, 5 mins), and the supernatant was filtered and retained for LCMS analysis for yield calculation. After LCMS analysis, the supernatant was concentrated under reduced pressure, extracted with EtOAc, the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting crude residue was dissolved in 10% isopropanol/hexanes ( $\nu/\nu$ ) for chiral HPLC analysis.

#### General procedure 2 for preparative scale reaction using purified enzyme

A round bottle (100 mL) was charged with *Bm*GDH (4 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (1.08 mL, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP+ (0.3 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), S-IRED-*Ss*-M14 protein (0.2 mol% catalyst loading), aldehyde (0.6 mL, 167 mM stock in DMSO, 0.1 mmol, 1 equiv) and amine (0.6 mL, 500 mM stock in DMSO, 0.3 mmol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 20 mL with 6% DMSO (*v*/*v*) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 20 mL of acetonitrile. The mixture was shaken for 30 min, filtered, concentrated, and extracted with EtOAc (3 × 20 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure to provide the crude product, which was purified by Flash chromatography (EtOAc/Petroleum ether, 10~20%, *v*/*v*).

#### General procedure 3 for preparative scale reaction using lyophilized CFE

A round bottle (250 mL) was charged with *BmGDH* (40 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (10.8 mL, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP<sup>+</sup> (3 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), lyophilized S-IRED-*Ss*-M16 CFE (800 mg), aldehyde substrate (6 mL, 167 mM stock in DMSO, 1 mmol, 1 equiv) and amine substrate (6 mL, 500 mM stock in DMSO, 3 mmol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 200 mL with 6% DMSO (*v*/*v*) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 200 mL of acetonitrile. The mixture was shaken for 30 min, filtered, concentrated, and extracted with EtOAc (3 × 100 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure to provide the crude product, which was purified by Flash chromatography (EtOAc/Petroleum ether, 10~20%, *v*/*v*).

#### (S)-N-benzyl-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (7)



Prepared according to the general procedure 1 using substrate **5a** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Yields**: run 1: 98%, run 2: 99%, average yield 98%.

0.1-mmol-scale preparative reaction was conducted according to the general procedure 2 using **5a** (0.1 mmol, 1 equiv) and **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%). **Isolated yield**: 89% (colorless amorphous, 30 mg).

1.0-mmol-scale preparative reaction was conducted according to the general procedure 3 using **5a** (1 mmol, 1 equiv) and **6a** (3 mmol, 3 equiv) catalyzed by lyophilized *S*-IRED-*Ss*-M14 CFE (4 mg/mL). **Isolated yield**: 65% (colorless amorphous, 220 mg).

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 8.75 min. **Absolute configuration** of the enzymatic product **7** was assigned as **S** by X-ray crystallography.



(S)-N-(2-Fluorobenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (8)



Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and (2-fluorophenyl)methanamine **6b** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Yields**: run 1: 95%, run 2: 98%, average yield 96%.

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 7.15 min,  $t_R$  (minor) = 8.37 min.



(S)-N-(3-Methyl-2-(quinolin-8-yl)benzyl)-1-(m-tolyl)methanamine (9)



Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and *m*-tolylmethanamine **6c** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%).

Yields: run 1: 97%, run 2: 97%, average yield 97%.

**Enantioselectivity**: 96:4 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 8.10 min,  $t_R$  (minor) = 9.00 min.



(S)-N-(3-Methoxybenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (10)



Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and (3-methoxyphenyl)methanamine **6d** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Yields**: run 1: 91%, run 2: 83%, average yield 87%.

**Enantioselectivity**: 95:5 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 10.79 min,  $t_R$  (minor) = 12.62 min.



(S)-N-(3-Bromobenzyl)-1-(3-methyl-2-(quinolin-8-yl)phenyl)methanamine (11)



Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and (3-bromophenyl)methanamine **6e** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Yields**: run 1: 97%, run 2: 99%, average yield 98%.

**Enantioselectivity**: 92:8 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 7.34 min,  $t_R$  (minor) = 8.87 min.





(S)-N-(4-Methoxybenzyl)-1-(3-methyl-2-(quinolin-8-yl)phenyl)methanamine (12)

Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and (4-methoxyphenyl)methanamine **6f** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Yields**: run 1: 91%, run 2: 95%, average yield 93%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **5a** (0.1 mmol, 1 equiv), (4-methoxyphenyl)methanamine **6f** (0.3 mmol, 3 equiv) catalyzed by lyophilized *S*-IRED-*Ss*-M14 (0.2 mol%). **Isolated yield**: 68% (colorless amorphous, 25 mg).

**Enantioselectivity**: 97:3 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 10.97 min,  $t_R$  (minor) = 12.83 min.





(S)-N-[3-Methyl-2-(quinolin-8-yl)benzyl]-1-[4-(trifluoromethyl)phenyl]methanamine (13)

Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and (4-(trifluoromethyl)phenyl)methanamine **6g** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.4 mol%). **Yields**: run 1: 80%, run 2: 82%, average yield 81%.

**Enantioselectivity**: 94:6 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 6.67 min,  $t_R$  (minor) = 9.45 min.





(S)-N-[3-Methyl-2-(quinolin-8-yl)benzyl)-1-(pyridin-3-yl)methanamine (14)



Prepared according to the general procedure 1 using substrate **5a** (5  $\mu$ mol, 1 equiv) and pyridin-3ylmethanamine **6h** (15  $\mu$ mol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 88%, run 2: 89%, average yield 88%.

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 15.09 min,  $t_R$  (minor) = 18.16 min.



	#	Time	Туре	Area	Height	Width	Area%	Symmetry
ſ	1	15.088	MF	57285.9	1786.9	0.5343	97.682	0.59
	2	18.164	MF	1359.5	37.2	0.6099	2.318	0.819





Prepared according to the general procedure 1 using substrate **5a** (5 µmol, 1 equiv) and furan-2ylmethanamine **6i** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%).

Yields: run 1: 94%, run 2: 97%, average yield 96%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **5a** (0.1 mmol, 1 equiv) and furan-2-ylmethanamine **6i** (0.3 mmol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%). **Isolated yield**: 82% (yellow amorphous, 27 mg).

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 7.59 min,  $t_R$  (minor) = 8.89 min.



(S)-N-[3-Methyl-2-(quinolin-8-yl)benzyl]prop-2-yn-1-amine (16)



Prepared according to the general procedure 1 using substrate **5a** (5 µmol, 1 equiv) and prop-2-yn-1amine **6j** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.4 mol%).

Yields: run 1: 70%, run 2: 72%, average yield 71%.

**Enantioselectivity**: 82:18 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 7.55 min,  $t_R$  (minor) = 8.88 min.



(R)-N-Benzyl-1-[3-chloro-2-(quinolin-8-yl)phenyl]methanamine (17)



Prepared according to the general procedure 1 using substrate **5b** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 98%, run 2: 99%, average yield 98%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **5b** (0.1 mmol, 1 equiv) and benzylamine **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%). **Isolated yield**: 86% (yellow solid, 31 mg).

**Enantioselectivity**: 99:1 er. Obtained from acetylated derivatives of racemic reference and the enzymatic product, respectively. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 34.12 min.



#### (R)-3-[(benzylamino)methyl]-2-(quinolin-8-yl)benzonitrile (18)



Prepared according to the general procedure 1 using substrate **5c** (5  $\mu$ mol, 1 equiv) and benzylamine **6a** (15  $\mu$ mol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 97%, run 2: 98%, average yield 98%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **5c** (0.1 mmol, 1 equiv) and benzylamine **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%). **Isolated yield**: 88% (yellow solid, 31 mg).

**Enantioselectivity**: 86:14 er. Obtained from *N*-Boc protected derivatives of racemic reference and the enzymatic product, respectively. Chiral HPLC method: AD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 13.67 min,  $t_R$  (major) = 15.31 min.


(R)-N-Benzyl-1-[2-(quinolin-8-yl)-3-(trifluoromethyl)phenyl]methanamine (19)



Prepared according to the general procedure 1 using substrate **5d** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by S-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 91%, run 2: 89%, average yield 90%.

**Enantioselectivity**: 71:29 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 19.43 min,  $t_R$  (major) = 20.80 min.





(R)-N-Benzyl-1-[3-methoxy-2-(quinolin-8-yl)phenyl]methanamine (20)



Prepared according to the general procedure 1 using substrate **5e** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 74%, run 2: 76%, average yield 75%.

**Enantioselectivity**: 97:3 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 18.30 min,  $t_R$  (minor) = 19.96 min.





(S)-N-Benzyl-1-[1-(quinolin-8-yl)naphthalen-2-yl]methanamine (21)



Prepared according to the general procedure 1 using substrate **5f** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 60%, run 2: 61%, average yield 60%.

**Enantioselectivity**: 94:6 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 19.00 min,  $t_R$  (minor) = 20.83min.





(S)-N-Benzyl-1-[2-(5-chloroquinolin-8-yl)-3-methylphenyl]methanamine (22)



Prepared according to the general procedure 1 using substrate **5g** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 75%, run 2: 81%, average yield 78%.

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 7.80 min.



(S)-N-Benzyl-1-[3-methyl-2-(2-methylquinolin-8-yl)phenyl]methanamine (23)



Prepared according to the general procedure 1 using substrate **5h** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 78%, run 2: 79%, average yield 78%.

**Enantioselectivity**: 91:9 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 6.56 min,  $t_R$  (minor) = 8.78 min.



#	Time	Туре	Area	Height	Width	Area%	Symmetry
1	6.558	MF	4985	427.4	0.1944	91.067	0.812
2	8.78	MF	489	26.4	0.3082	8.933	0.786

# (S)-N-Benzyl-1-[3-methyl-2-(quinoxalin-5-yl)phenyl]methanamine (24)



Prepared according to the general procedure 1 using substrate **5i** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 94%, run 2: 93%, average yield 94%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **5i** (0.1 mmol, 1 equiv) and benzylamine **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%). **Isolated yield**: 81% (yellow amorphous, 27 mg).

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 2% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 31.24 min.



(S)-N-[2-(Benzo[d]thiazol-4-yl)-3-methylbenzyl]-1-phenylmethanamine (25)



Prepared according to the general procedure 1 using substrate **5j** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by S-IRED-Ss-M14 (0.2 mol%).

Yields: run 1: 85%, run 2: 84%, average yield 84%.

**Enantioselectivity**: 92:8 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 8.96 min,  $t_R$  (major) = 10.68 min.



(S)-N-[2-(Benzo[d]oxazol-4-yl)-3-methylbenzyl]-1-phenylmethanamine (26)



Prepared according to the general procedure 1 using substrate **5k** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%) or *S*-IRED-*Ss*-C8 (0.2 mol%). *S*-IRED-*Ss*-M14

Yields: run 1: 68%, run 2: 69%, average yield 68%. Enantioselectivity: 60:40 er.

S-IRED-Ss-C8 (M236A/P122Y/F214W based on S-IRED-Ss-M9)

Yields: run 1: 78%, run 2: 85%, average yield 82%. Enantioselectivity: 96:4 er.

Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 9.44 min,  $t_R$  (major) = 12.71 min.





(S)-N-Benzyl-1-[3-methyl-2-(1-methyl-1H-benzo[*d*]imidazol-4-yl)phenyl]methanamine (27)

Prepared according to the general procedure 1 using substrate **5I** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M14 (0.2 mol%).

Yields: run 1: 93%, run 2: 94%, average yield 93%.

**Enantioselectivity**: 99:1 er. Obtained from *N*-Boc protected derivatives of racemic reference and the enzymatic product, respectively. Chiral HPLC method: AD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 9.81 min.



#### (A) Other tested amines



**Fig. S6.** (A) Amines not well accepted by S-IRED-*Ss*-M14. (B) Unsuccessful ketone substrate. Reaction conditions: **5a** (5 µmol, 1 equiv), amines (15 µmol, 3 equiv), NADP<sup>+</sup> (0.1 µmol, 2 mol%), glucose (15 µmol, 3 equiv), *Bm*GDH (1 mg/mL), S-IRED-*Ss*-M14 (0.2 mol%) in Tris-HCl buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000 µL. [a] Conversion (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. (C) Imine as substrate. The imine (>98% yield, monitored by NMR) was formed in situ over 2 hour from **5a** (1 equiv), BnNH<sub>2</sub> (3 equiv), and HOAc (0.5 equiv), then directly used in the enzymatic reaction.

# Enzymatic atroposelective DKR for the synthesis of heterobiaryl N-oxide amines



### General procedure 4 for analytical scale reaction

A screw vial (4 mL) was charged with *Bm*GDH (200  $\mu$ L, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (54  $\mu$ L, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP+ (15  $\mu$ L, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), S-IRED-*Ss*-M16 protein (0.2 mol% or 0.4 mol% catalyst loading), aldehyde (30  $\mu$ L, 167 mM stock in DMSO, 5  $\mu$ mol, 1 equiv) and amine (30  $\mu$ L, 500 mM stock in DMSO, 15  $\mu$ mol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 1000  $\mu$ L with 6% DMSO (*v*/*v*) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 3 mL of acetonitrile and 100  $\mu$ L of 5 mg/mL 1,3,5-tribromobenzene (TBB) in acetonitrile as the internal standard. The mixture was shaken for 30 min, centrifuged (13,400 x g, 5 mins), and the supernatant was filtered and retained for LCMS analysis for yield calculation. After LCMS analysis, the supernatant was concentrated under reduced pressure, extracted with EtOAc, the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The resulting crude residue was dissolved in 30% isopropanol/hexanes (*v*/*v*) for chiral HPLC analysis.

# General procedure 5 for preparative scale reaction using purified enzyme

A round bottle (100 mL) was charged with *Bm*GDH (4 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (1.08 mL, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP+ (0.3 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), S-IRED-*Ss*-M16 protein (0.2 mol% catalyst loading), aldehyde (0.6 mL, 167 mM stock in DMSO, 0.1 mmol, 1 equiv) and amine (0.6 mL, 500 mM stock in DMSO, 0.3 mmol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 20 mL with 6% DMSO (*v*/*v*) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 20 mL of acetonitrile. The mixture was shaken for 30 min, filtered, concentrated, and extracted with EtOAc (3 × 20 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure to provide the crude product, which was purified by Flash chromatography (MeOH/DCM, 5~10%, *v*/*v*).

#### General procedure 6 for preparative scale reaction using lyophilized CFE

A round bottle (250 mL) was charged with *BmGDH* (40 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), glucose (10.8 mL, 50 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), NADP+ (3 mL, 5 mg/mL stock solution in 100 mM Tris-HCl buffer pH 8.0), lyophilized S-IRED-*Ss*-M16 CFE (800 mg), aldehyde substrate (6 mL, 167 mM stock in DMSO, 1 mmol, 1 equiv) and amine substrate (6 mL, 500 mM stock in DMSO, 3 mmol, 3 equiv). Tris-HCl buffer (100 mM pH 8.0) was added to bring the total volume to 200 mL with 6% DMSO (*v*/*v*) as cosolvent. The vial was sealed and placed on a shaker at 200 rpm at room temperature for 24 hours. Upon completion, the reaction was quenched with 200 mL of acetonitrile. The mixture was shaken for 30 min, filtered, concentrated, and extracted with EtOAc (3 × 100 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure to provide the crude product, which was purified by Flash chromatography (MeOH/DCM, 5~10%, *v*/*v*).

(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}pyridine 1-oxide (29)



Prepared according to the general procedure 4 using substrate **28a** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.2 mol%). **Yields**: run 1: 98%, run 2: 97%, average yield 98%.

0.1-mmol-scale reaction was conducted according to the general procedure 5 using **28a** (0.1 mmol, 1 equiv) and **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 86% (white solid, 26 mg).

1.0-mmol-scale preparative reaction was conducted according to the general procedure 6 using **28a** (1 mmol, 1 equiv) and **6a** (3 mmol, 3 equiv) catalyzed by lyophilized *S*-IRED-*Ss*-M16 CFE (4 mg/mL). **Isolated yield**:76% (white solid, 231 mg).

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 15.82 min. **Absolute configuration** of the enzymatic product **29** was assigned as **S** by X-ray crystallography.





(S)-2-{2-{[(2-Fluorobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (30)

Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and (2-fluorophenyl)methanamine **6b** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.2 mol%). **Yields**: run 1:89%, run 2: 95%, average yield 92%.

**Enantioselectivity**: 97:3 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 8.67 min,  $t_R$  (major) = 10.99 min.



(S)-2-{2-Methyl-6-{[(3-methylbenzyl)amino]methyl}phenyl}pyridine 1-oxide (31)



Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and *m*-tolylmethanamine **6c** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 64%, run 2: 64%, average yield 64%.

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 20% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 18.75 min,  $t_R$  (major) = 34.21 min.



(S)-2-{2-{[(3-Methoxybenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (32)



Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and (3-methoxyphenyl)methanamine **6d** (15  $\mu$ mol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Yields**: run 1: 90%, run 2: 89%, average yield 90%.

Preparative enzymatic reaction was conducted according to the general procedure 5 using **28a** (0.1 mmol, 1 equiv) and **6d** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 75% (yellow solid, 25 mg).

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 17.14 min.



(S)-2-{2-{[(3-Bromobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (33)



Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and (3-bromophenyl)methanamine **6e** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.2 mol%).

Yields: run 1: 81%, run 2: 84, average yield 82%.

Preparative enzymatic reaction was conducted according to the general procedure 5 using **28a** (0.1 mmol, 1 equiv) and **6e** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 65% (yellow solid, 25 mg).

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 10.87 min.



(S)-2-{2-{[(4-Methoxybenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (34)



Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and (4-methoxyphenyl)methanamine **6f** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.2 mol%). **Yields**: run 1: 59%, run 2: 62%, average yield 61%.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 16.57 min.





(S)-2-{2-{[(4-Fluorobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (35)

Prepared according to the general procedure 4 using substrate **28a** (5 μmol, 1 equiv) and (4-fluorophenyl)methanamine **6k** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 88%, run 2: 82, average yield 85%.

**Enantioselectivity**: 98:2 er. Obtained from *N*-Boc protected derivatives of racemic reference and the enzymatic product, respectively Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 9.61 min,  $t_R$  (major) = 14.46 min.



(S)-2-{2-{[4-Chlorobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (36)



Prepared according to the general procedure 4 using substrate **28a** (5 µmol, 1 equiv) and (4-chlorophenyl)methanamine **6I** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.4 mol%).

Yields: run 1: 71%, run 2: 74%, average yield 72%.

**Enantioselectivity**: 97:3 er. Obtained from *N*-Boc protected derivatives of racemic reference and the enzymatic product, respectively Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 12.70 min,  $t_R$  (major) = 13.99 min.



(S)-2-{2-Methyl-6-{[(pyridin-3-ylmethyl)amino]methyl}phenyl}pyridine 1-oxide (37)



Prepared according to the general procedure 4 using substrate **28a** (5 µmol, 1 equiv) and pyridin-3ylmethanamine **6h** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 90%, run 2: 83%, average yield 86%.

Preparative enzymatic reaction was conducted according to the general procedure 5 using **28a** (0.1 mmol, 1 equiv) and **6h** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 79% (yellow solid, 25 mg).

**Enantioselectivity**: 94:6 er. Chiral HPLC method: OD-H column, 210 nm, 20% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 24.38 min,  $t_R$  (minor) = 28.39 min.





(S)-2-{2-methyl-6-{[(thiophen-3-ylmethyl)amino]methyl}phenyl}pyridine 1-oxide (38)

Prepared according to the general procedure 4 using substrate **28a** (5  $\mu$ mol, 1 equiv) and thiophen-3ylmethanamine **6m** (15  $\mu$ mol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.2 mol%).

Yields: run 1: 90%, run 2: 83%, average yield 87%.

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 12.79 min.



(S)-2-{2-Methyl-6-[(prop-2-yn-1-ylamino)methyl]phenyl}pyridine 1-oxide (39)



Prepared according to the general procedure 4 using substrate **28a** (5 µmol, 1 equiv) and prop-2-yn-1amine **6j** (15 µmol, 3 equiv) catalyzed by S-IRED-Ss-M16 (0.4 mol%).

Yields: run 1: 79%, run 2: 72%, average yield 76%.

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 8.72 min,  $t_R$  (major) = 10.19 min.



(R)-2-{2-[(benzylamino)methyl]-6-chlorophenyl}pyridine 1-oxide (40)



Prepared according to the general procedure 1 using substrate **28b** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by S-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 78%, run 2: 79%, average yield 78%.

**Enantioselectivity**: 97:3 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 11.56 min,  $t_R$  (major) = 25.25 min.



(*R*)-2-{2-[(Benzylamino)methyl]-6-methoxyphenyl}pyridine 1-oxide (**41**)



Prepared according to the general procedure 4 using substrate **28c** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 88%, run 2: 90%, average yield 89%.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 9.81 min,  $t_R$  (major) = 20.47 min.





(S)-2-{2-[(Benzylamino)methyl]naphthalen-1-yl}pyridine 1-oxide (42)



Prepared according to the general procedure 4 using substrate **28d** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 78%, run 2: 79%, average yield 78%.

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 9.30 min.





(S)-2-{2-[(benzylamino)methyl]-6-methylphenyl}-4-methylpyridine 1-oxide (43)



Prepared according to the general procedure 4 using substrate **28e** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 70%, run 2: 82%, average yield 76%.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 18.45 min.



(S)-2-{2-[(benzylamino)methyl]-6-methylphenyl}-4-chloropyridine 1-oxide (44)



Prepared according to the general procedure 4 using substrate **28f** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.4 mol%).

Yields: run 1: 60%, run 2: 60%, average yield 60%.

**Enantioselectivity**: 86:14 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 12.63 min,  $t_R$  (major) = 17.86 min.



1,,		10		12		14		· · ·	
	#	Time	Туре	Area	Height	Width	Area%	Symmetry	
	1	12.627	MM	2970.2	60.2	0.8226	14.420	0.323	
	2	17.861	MM	17626.9	375.8	0.7818	85.580	0.792	

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(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}-5-fluoropyridine 1-oxide (45)



Prepared according to the general procedure 4 using substrate **28g** (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 91%, run 2: 93%, average yield 92%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **28g** (0.1 mmol, 1 equiv) and **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 75% (white solid, 24 mg).

**Enantioselectivity**: 95:5 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 8.07 min,  $t_R$  (minor) = 11.88 min.

Absolute configuration of the enzymatic product 45 was assigned as S by X-ray crystallography.



(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}-5-methylpyridine 1-oxide (46)



Prepared according to the general procedure 4 using substrate **28h** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 78%, run 2: 78%, average yield 78%.

Preparative enzymatic reaction was conducted according to the general procedure 2 using **28h** (0.1 mmol, 1 equiv) and **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 69% (white solid, 22 mg).

**Enantioselectivity**: >99:1 er. Chiral HPLC method: OD-H column, 210 nm, 20% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 9.94 min.



(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}-5-(trifluoromethyl)pyridine 1-oxide (47)



Prepared according to the general procedure 4 using substrate **28i** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 75%, run 2: 82%, average yield 78%.

**Enantioselectivity**: 89:11 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 6.13 min,  $t_R$  (minor) = 9.06 min.



(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}-6-methylpyridine 1-oxide (48)



Prepared according to the general procedure 4 using substrate **28j** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 85%, run 2: 89%, average yield 87%.

Preparative enzymatic reaction was conducted according to the general procedure 5 using **28j** (0.1 mmol, 1 equiv) and **6a** (0.3 mmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%). **Isolated yield**: 69% (white solid, 22 mg).

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 20% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 9.72 min,  $t_R$  (major) = 10.75 min.

Absolute configuration of the enzymatic product 48 was assigned as S by X-ray crystallography.



(S)-1-{2-[(Benzylamino)methyl]phenyl}isoquinoline 2-oxide (49)



Prepared according to the general procedure 4 using substrate **28k** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.4 mol%).

Yields: run 1: 36%, run 2: 41%, average yield 39%.

Note: Waters symmetry silica C18 column (4.6 x 250 mm, 5.0 µm) was used.

**Enantioselectivity**: 98:2 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 14.73 min,  $t_R$  (major) = 43.82 min.





Fig. S7. Kinetic resolution of the ortho-tetrasubstituted substrate rac-28I using S-IRED-Ss-M16.

(S)-2-{2-[(Benzylamino)methyl]-6-methylphenyl}-3-methylpyridine 1-oxide (50)



Prepared according to the general procedure 4 using substrate *rac-28I* (5 µmol, 1 equiv) and benzylamine **6a** (15 µmol, 3 equiv) catalyzed by S-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 49%, run 2: 48%, average yield 48%.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 9.02 min,  $t_R$  (major) = 11.58 min.



(R)-2-(2-Formyl-6-methylphenyl)-3-methylpyridine 1-oxide (28I)



Prepared according to the general procedure 4 using substrate **28I** (5 μmol, 1 equiv) and benzylamine **6a** (15 μmol, 3 equiv) catalyzed by *S*-IRED-*Ss*-M16 (0.2 mol%).

Yields: run 1: 49%, run 2: 48%, average yield 48%.

Substrate enantioselectivity: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 30% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 15.55 min.









**Fig. S8.** (A) Amines not well accepted by S-IRED-Ss-M16. Reaction conditions: **28a** (5 µmol, 1 equiv), amines (15 µmol, 3 equiv), NADP<sup>+</sup> (0.1 µmol, 2 mol%), glucose (15 µmol, 3 equiv), *Bm*GDH (1 mg/mL), S-IRED-Ss-M16 (0.2 mol%) in Tris-HCI buffer (100 mM, pH 8.0) with 6% DMSO as cosolvent at room temperature (rt) for 24 h, the final total volume is 1000 µL. [a] Conversion (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. (B) Imine as substrate. The imine *rac-28m* (>99% yield, monitored by NMR) was formed in situ over 2 hour from **28a** (1 equiv), BnNH<sub>2</sub> (3 equiv), and HOAc (0.5 equiv), then directly used in the enzymatic reaction.
# Derivatization and application of enzymatic products Derivatization of enzymatic product (*S*)-7



To a solution of (*S*)-7 (obtained from enzymatic reaction catalyzed by *S*-IRED-*Ss*-M14, 200 mg, 0.6 mmol, 1.0 equiv) in MeOH/DCM (6 mL, 2:1) under N<sub>2</sub> atmosphere was added NiCl<sub>2</sub>•6H<sub>2</sub>O (56 mg, 0.24 mmol, 0.4 equiv) and NaBH<sub>4</sub> (179 mg, 4.8 mmol, 8.0 equiv) at 0 °C, and the reaction was stirred for another 3 hours until (*S*)-7 was consumed. Then the reaction was quenched with H<sub>2</sub>O (7 mL) and extracted with DCM (7 mL × 3), the organic phase was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated under vacuum, and purified through chromatography (PE/EA= 2:1) to give the axially chiral diamine product **51** (yellow oil, 122 mg, 60% yield, 99:1 er).

To a solution of **51** (60 mg, 0.18 mmol, 1.0 equiv) in MeOH (2 mL) was added *p*-toluenesulfonic acid monohydrate (0.1 mg, 5.3 µmol, 0.03 equiv) and a solution of formaldehyde (37% in water, 17 µL, 0.21 mmol, 1.2 equiv) under N<sub>2</sub> atmosphere at room temperature, and the mixture was stirred vigorously for 8 hours. Then the reaction was concentrated and purified through chromatography (PE/EA= 20:1) to give the product **52** (yellow oil, 45 mg, 72% yield, 97:3 er).

(S)-N-Benzyl-1-[3-methyl-2-(1,2,3,4-tetrahydroquinolin-8-yl)phenyl]methanamine (51)



Yellow oil, 122 mg, 60% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.34 (dd, *J* = 7.5, 1.5 Hz, 1H), 7.28 – 7.25 (m, 2H), 7.24 – 7.22 (m, 2H), 7.22 – 7.19 (m, 1H), 7.17 (dd, *J* = 8.0, 1.5 Hz, 2H), 6.98 (m, 1H), 6.72 (dd, *J* = 7.5, 2.0 Hz, 1H), 6.67 (t, *J* = 7.5 Hz, 1H), 3.68 (d, *J* = 13.2 Hz, 1H), 3.62 – 3.58 (m, 2H), 3.52 (d, *J* = 12.8 Hz, 1H), 3.12 (m, 1H), 3.02 (m, 1H), 2.84 – 2.73 (m, 3H), 2.03 (s, 3H), 1.87 – 1.78 (m, 1H), 1.78 – 1.69 (m, 1H).

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 141.7, 139.7, 138.8, 138.0, 137.8, 129.4, 128.9, 128.4, 128.2, 127.9, 127.8, 127.2, 127.0, 124.2, 121.7, 117.0, 53.4, 52.0, 42.0, 27.5, 22.1, 20.3.

HRMS: m/z calcd for C<sub>24</sub>H<sub>27</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 343.2174, found 343.2178.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 6.79 min,  $t_R$  (minor) = 7.32 min.



(S)-6-Benzyl-11-methyl-2,3,6,7-tetrahydro-1*H*,5*H*-benzo[5,6][1,3]diazocino[7,8,1-*ij*]quinoline (52)



Yellow oil, 45 mg, 72% yield.

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 (m, 2H), 7.33 (m, 2H), 7.28 (m, 1H), 7.23 (m, 2H), 6.98 (d, J = 7.5 Hz, 1H), 6.96 (m, 1H), 6.67 – 6.60 (m, 2H), 4.37 (d, J = 13.5 Hz, 1H), 3.96 (d, J = 12.0 Hz, 1H), 3.76 (d, J = 13.5 Hz, 1H), 3.66 (d, J = 12.0 Hz, 1H), 3.44 (d, J = 13.5 Hz, 1H), 3.35 (m, 2H), 3.20 – 3.15 (m, 1H), 2.88 (m, 1H), 2.82 (m, 1H), 2.23 (s, 3H), 2.04 (d, J = 14.5 Hz, 1H), 2.01 – 1.95 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ 145.7, 144.1, 139.5, 135.9, 132.7, 130.5, 129.0, 128.8, 128.7, 128.6, 128.4, 128.3, 127.0, 127.0, 126.5, 56.7, 55.4, 52.4, 29.4, 22.8, 22.2.

HRMS: m/z calcd for  $C_{25}H_{27}N_2$  [M+H]<sup>+</sup>: 355.2174, found 355.2174.

**Enantioselectivity**: 97:3 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 6.82 min,  $t_R$  (minor) = 7.38 min.







To a stirred solution of enzymatic product (*S*)-7 (50 mg, 0.15 mmol, 1.0 equiv) in DCM (10 mL) was added Ac<sub>2</sub>O (0.75 mmol, 5 equiv) and K<sub>2</sub>CO<sub>3</sub> (207 mg, 1.5 mmol, 10 equiv), the reaction mixture was stirred over 16 hours at room temperature (rt). After completion of the reaction, the reaction mixture was quenched by saturated NaHCO<sub>3</sub> solution (20 mL), then extracted with EtOAc (3 × 20 mL). The combined organic layers were washed with brine (50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated to provide a crude product, which was further purified by flash chromatography (EtOAc/ Hexanes, 5%, v/v) to provide the pure acetamide product **53** (white solid, 43 mg, 75% yield, 99:1 er).

To a stirred solution of acetamide **53** (40 mg, 0.11 mmol, 1.0 equiv) in dry DCM (5 mL) under ice-bath was added the *meta*-chloroperbenzoic acid (*m*-CPBA, 57 mg, 0.33 mmol, 3 equiv) and then stirred at 0 °C for 8 hours. After completion of the reaction, the reaction mixture was quenched by saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution (5 mL). The resulting reaction mixture was concentrated under reduced pressure and diluted with water (10 mL), then extracted with EtOAc (3 × 20 mL). The combined organic layers were washed with brine (50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated to provide a crude product, which was further purified by flash chromatography (EtOAc/Petroleum ether, 10%, *v*/*v*) to provide the pure *N*-oxide product **54** (yellow amorphous, 25 mg, 60% yield, 99:1 er).

(S)-N-Benzyl-N-(3-methyl-2-(quinolin-8-yl)benzyl)acetamide (53)



White solid, 43 mg, 75% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.86 (dd, J = 4.2, 1.8 Hz, 1H), 8.79 (dd, J = 4.4, 1.8 Hz, 0.54H\*), 8.20 (dd, J = 8.3, 1.8 Hz, 1H), 8.16 (d, J = 8.2 Hz, 0.58H\*), 7.84 (d, J = 6.7 Hz, 1H), 7.80 (d, J = 6.7 Hz, 0.52H\*), 7.60 – 7.56 (m, 1H), 7.54 (d, J = 7.7 Hz, 0.65H\*), 7.49 – 7.43 (m, 1.6H\*), 7.43 – 7.36 (m, 2H), 7.36 – 7.31 (m, 2H), 7.29 (d, J = 7.5 Hz, 0.77H\*),7.14 (dd, J = 13.8, 6.7 Hz, 4H+2H\*), 7.05 – 7.01 (m, 2H), 6.80 – 6.76 (m, 1H\*), 4.56 (d, J = 14.6 Hz, 1H), 4.48 (d, J = 14.6 Hz, 1H), 4.39 (d, J = 15.1 Hz, 0.59H\*), 4.30 (d, J = 17.1 Hz, 0.59H\*), 4.23 (d, J = 15.1 Hz, 0.56H\*), 4.17 (d, J = 17.1 Hz, 0.61H\*), 3.96 (s, 2H), 1.94 (s, 3H), 1.91 (s, 1.55H\*), 1.89 (s, 3H), 1.75 (s, 1.75H\*). \*Indicates the peaks of rotamer (2:1).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 171.5, 150.8, 137.6, 137.5, 136.6, 135.6, 134.9, 130.3, 129.3, 128.6, 128.5, 128.3, 128.3, 128.2, 127.2, 126.6, 126.1, 123.0, 121.4, 121.0, 49.8, 48.2, 21.4, 20.5.

HRMS: m/z calcd for C<sub>26</sub>H<sub>25</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 381.1967, found 381.1972.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OJ-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 23.12 min.



(S)-8-{2-[(N-Benzylacetamido)methyl]-6-methylphenyl}quinoline 2-oxide (54)



Yellow amorphous, 25 mg, 60% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.99 (dt, *J* = 4.1, 1.8 Hz, 1H), 8.94 – 8.90 (m, 0.5H\*), 8.34 (dt, *J* = 8.3, 1.8 Hz, 1H), 8.29 (d, *J* = 8.4 Hz, 0.53H\*), 7.98 (dd, *J* = 8.2, 1.6 Hz, 1H), 7.93 (dt, *J* = 8.2, 1.6 Hz, 0.64H\*), 7.85 – 7.78 (m, 2H+1H\*), 7.74 – 7.65 (m, 2H+1H\*), 7.62 (dd, *J* = 3.1, 1.6 Hz, 1H\*), 7.60 (dd, *J* = 3.1, 1.6 Hz, 2H), 7.59 (t, *J* = 2.5 Hz, 1H), 7.57 (d, *J* = 1.5 Hz, 0.48H\*), 7.56 – 7.51 (m, 1H+1H\*), 7.46 (d, *J* = 7.8 Hz, 2H), 7.44 – 7.41 (m, 0.7H), 7.41 – 7.38 (m, 0.42H\*), 7.19 – 7.15 (m, 2H), 6.91 (dd, *J* = 5.1, 2.8 Hz, 1H\*), 4.70 (d, *J* = 14.6 Hz, 1H), 4.62 (d, *J* = 14.6 Hz, 1H), 4.53 (d, *J* = 15.1 Hz, 0.62H\*), 4.44 (d, *J* = 17.2 Hz, 0.61H\*), 4.37 (d, *J* = 15.1 Hz, 0.65H\*), 4.30 (d, *J* = 17.2 Hz, 0.58H\*), 4.10 (s, 2H), 2.07 (s, 3H), 2.05 (d, *J* = 1.6 Hz, 1.77H\*), 2.02 (s, 3H), 1.88 (s, 1.77H). \*Indicates the peaks of rotamer (2:1). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  171.58, 150.80, 137.54, 136.61, 134.87, 132.26, 132.18, 130.31, 129.31, 128.68, 128.58, 128.45, 128.33, 128.25, 128.16, 127.18, 126.62, 126.08, 123.00, 121.45, 49.77, 48.22, 21.34, 20.50.

HRMS: m/z calcd for  $C_{26}H_{25}N_2O_2$  [M+H]<sup>+</sup>: 397.1916, found 397.1917.

**Enantioselectivity**: 99:1 er. Chiral HPLC method: OD-H column, 210 nm, 10% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (major) = 12.01 min,  $t_R$  (minor) = 14.43 min.



# Asymmetric allylation of *p*-anisaldehyde catalyzed by enzymatic product (S)-29



**General procedure:** Adapted from the method by Hayashi *et al.*<sup>4</sup> To a solution of *p*-anisaldehyde (**55**, 30 mg, 0.22 mmol, 1 equiv) and heterobiaryl *N*-oxide amine (**29**) in anhydrous acetonitrile (1 mL) was added *N*,*N*-diisopropylethylamine (0.12 mL, 0.66 mmol, 3.0 equiv) under N<sub>2</sub> atmosphere. The solution was cooled to -45°C before allyl trichlorosilane **56** was added dropwise. The reaction was stirred at -45°C for 48 h before quenched with 1 M NaOH aqueous solution (2 mL), and stirred for another 30 mins at room temperature. Then the reaction mixture was extracted with EtOAc (2 mL x 3), washed with brine, dried with Na<sub>2</sub>SO<sub>4</sub>, and concentrated for further HPLC analysis. The final product was purified through chromatography (PE/EA=10:1) to give a yellow oil.

Entry	Catalyst	Allyl trichlorosilane	Yield	er <sup>b</sup>
1	1 mol% <i>rac-<b>29</b></i>	1.2 equiv	9%ª	n.d.
2	5 mol% <i>rac-</i> 29	1.2 equiv	58% <sup>a</sup>	n.d.
3	10 mol% <i>rac-<b>29</b></i>	1.2 equiv	72% <sup>a</sup>	n.d.
4	10 mol% <i>rac</i> - <b>29</b>	1.6 equiv	82% <sup>c</sup>	50:50
5	10 mol% ( <i>S</i> )- <b>29</b>	1.6 equiv	80% <sup>c</sup>	73:27

**Table S7**. Allylation of *p*-anisaldehyde catalyzed by heterobiaryl *N*-oxide **29**. <sup>*a*</sup> Yield (average of duplicate) determined *via* LCMS relative to an internal standard 1,3,5-tribromobenzene. <sup>*b*</sup> Enantiomeric ratio determined by HPLC on a chiral stationary phase. <sup>*c*</sup> Isolated yield after chromatography.

(S)-1-(4-Methoxyphenyl)but-3-en-1-ol (57)



Yellow oil, 31 mg, 80% isolated yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.28 (d, *J* = 8.6 Hz, 2H), 6.88 (d, *J* = 8.7 Hz, 2H), 5.85 – 5.74 (m, 1H), 5.19 – 5.09 (m, 2H), 4.68 (t, *J* = 6.6 Hz, 1H), 3.80 (s, 3H), 2.53 – 2.46 (m, 2H), 2.10 (brs, 1H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  159.1, 136.2, 134.7, 127.2, 118.3, 113.9, 73.1, 55.4, 43.9.

The NMR spectra is in agreement with published data.<sup>4</sup>

Height

120.5

Туре

1842.3

12.235 FM

Width

0.2547

Area% Sy 26.571 73.429

0.88

**Enantioselectivity**: 73:27 er. Chiral HPLC method: OD-H column, 210 nm, 5% isopropanol/hexanes, flow rate 1.0 mL/min, room temperature,  $t_R$  (minor) = 12.24 min,  $t_R$  (major) = 14.53 min. **Absolute configuration** of the major product of **57** is assigned as **S** by comparison with the previously reported chiral HPLC data.<sup>5</sup>



# Synthesis of racemic heterobiaryl substrates



# **General procedure A**

To a stirred solution of aldehyde **S-1** (5 mmol, 1.0 equiv) in 1,2-dimethoxyethane (DME, 20 mL) and  $H_2O$  (10 mL) was added boronic acid **S-2** (7.5 mmol, 1.5 equiv),  $Na_2CO_3$  (1.59 g, 15 mmol, 3.0 equiv) and Pd(PPh<sub>3</sub>)<sub>4</sub> (577 mg, 0.5 mmol, 0.1 equiv), the reaction mixture was heated at reflux overnight. After completion of the reaction, the solvent was removed under reduced pressure. The resulting mixture was extracted with EtOAc (25 mL × 3), and then organic layers were collected and dried over  $Na_2SO_4$ , and concentrated to provide the crude product, which was purified by flash chromatography (EtOAc/Petroleum ether, 5~10%, v/v) to give products **5a-5h**.

#### **General procedure B**

To a stirred solution of **S-3** (5 mmol, 1.0 equiv) in 1,2-dimethoxyethane (DME, 20 mL) and H<sub>2</sub>O (10 mL) was added trifluoromethanesulfonate **S-4** (4 mmol, 0.8 equiv), Na<sub>2</sub>CO<sub>3</sub> (1.59 g, 15 mmol, 3.0 equiv.) and Pd(PPh<sub>3</sub>)<sub>4</sub> (577 mg, 0.5 mmol, 0.1 equiv), the reaction mixture was heated at reflux overnight. After completion of the reaction, the solvent was removed under reduced pressure. The resulting mixture was extracted with EtOAc (25 mL × 3), and then organic layers were collected and dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated to provide the crude product, which was purified by flash chromatography (EtOAc/Petroleum ether,  $5\sim10\%$ , v/v) to give products **5**i.

## **General procedure C**

To a stirred solution of **S-3** (5 mmol, 1.0 equiv) in 1,2-dimethoxyethane (DME, 20 mL) and H<sub>2</sub>O (10 mL) was added **S-5** (7.5 mmol, 1.5 equiv.), Na<sub>2</sub>CO<sub>3</sub> (1.59 g, 15 mmol, 3.0 equiv.) and Pd(PPh<sub>3</sub>)<sub>4</sub> (577 mg, 0.5 mmol, 0.1 equiv.), the reaction mixture was heated at reflux overnight. After completion of the reaction, the solvent was removed under reduced pressure. The resulting mixture was extracted with EtOAc (25 mL × 3), and then organic layers were collected and dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated to provide the crude product, which was further purified by flash chromatography (EtOAc/Petroleum ether,  $5\sim10\%$ , v/v) to give products **5j-5I**.

3-Methyl-2-(quinolin-8-yl)benzaldehyde (5a)



Yellow solid. 1087 mg, 88% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.50 (s, 1H), 8.87 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.24 (d, *J* = 8.3 Hz, 1H), 7.94 (t, *J* = 8.8 Hz, 2H), 7.64 (t, *J* = 7.6 Hz, 1H), 7.58 (d, *J* = 7.2 Hz, 2H), 7.48 (t, *J* = 7.7 Hz, 1H), 7.42 (dd, *J* = 8.3, 4.2 Hz, 1H), 2.01 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 192.7, 150.9, 146.9, 143.3, 138.1, 136.4, 135.5, 135.0, 131.6, 128.6, 128.5, 128.0, 126.1, 124.8, 121.4, 20.1.

The NMR spectra is in agreement with published data.<sup>6</sup>

HRMS: m/z calcd for C<sub>17</sub>H<sub>14</sub>NO [M+H]<sup>+</sup>: 248.1075, found 248.1076.

3-Chloro-2-(quinolin-8-yl)benzaldehyde (5b)



White solid. 1161 mg, 87% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.45 (d, *J* = 5.1 Hz, 1H), 8.87 (s, 1H), 8.26 (t, *J* = 6.8 Hz, 1H), 8.05 – 7.96 (m, 2H), 7.79 (t, *J* = 6.7 Hz, 1H), 7.71 – 7.67 (m, 2H), 7.54 (q, *J* = 7.2 Hz, 1H), 7.47 – 7.43 (m, 1H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  191.4, 151.1, 146.9, 142.0, 136.7, 136.4, 135.4, 134.8, 134.1, 132.1, 129.4, 129.3, 128.4, 126.0, 125.7, 121.7.

The NMR spectra is in agreement with published data.<sup>6</sup>

HRMS: m/z calcd for C<sub>16</sub>H<sub>11</sub>CINO [M+H]<sup>+</sup>: 268.0529, found 268.0531.

3-Formyl-2-(quinolin-8-yl)-benzonitrile (5c)



Yellow solid. 464 mg, 36% yield.

<sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ 9.45 (d, *J* = 3.4 Hz, 1H), 8.87 – 8.82 (m, 1H), 8.58 – 8.54 (m, 1H), 8.32 (dd, *J* = 7.9, 3.7 Hz, 1H), 8.27 – 8.22 (m, 2H), 8.05 – 7.94 (m, 1H), 7.89 – 7.80 (m, 2H), 7.67 – 7.61 (m, 1H).

<sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>) δ 190.2, 151.1, 145.9, 145.7, 137.8, 136.7, 135.3, 132.6, 131.9, 131.2, 130.2, 129.1, 127.9, 126.2, 122.0, 117.3, 114.8.

HRMS: m/z calcd for C<sub>17</sub>H<sub>11</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 259.0871, found 259.0869.

2-(Quinolin-8-yl)-3-(trifluoromethyl)benzaldehyde (5d)



White solid. 400 mg, 27% yield.

<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.22 (s, 1H), 8.73 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.44 (dd, *J* = 8.3, 1.8 Hz, 1H), 8.19 - 8.13 (m, 2H), 8.12 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.81 (t, *J* = 7.8 Hz, 1H), 7.77 - 7.73 (m, 1H), 7.72 - 7.61 (m, 1H), 7.52 (dd, *J* = 8.3, 4.2 Hz, 1H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  191.0, 151.0, 147.5, 142.1, 136.3, 136.2, 133.3, 131.7, 131.4 (q, *J* = 5.1 Hz), 130.7 (q, *J* = 29.8 Hz), 130.3, 129.5, 128.4, 128.0, 125.6, 123.7 (q, *J* = 274.3 Hz), 121.6, 121.0. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>)  $\delta$  -58.1.

HRMS: m/z calcd for C<sub>17</sub>H<sub>11</sub>F<sub>3</sub>NO [M+H]<sup>+</sup>: 302.0793, found 302.0798.

3-Methoxy-2-(quinolin-8-yl)benzaldehyde (5e)

MeO сно

Yellow solid. 750 mg, 57% yield.

<sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.35 (s, 1H), 8.81 – 8.76 (m, 1H), 8.46 (dd, *J* = 8.3, 2.0 Hz, 1H), 8.10 (dd, *J* = 7.4, 2.3 Hz, 1H), 7.77 – 7.67 (m, 2H), 7.66 – 7.58 (m, 1H), 7.59 – 7.53 (m, 2H), 7.48 (d, *J* = 8.2 Hz, 1H), 3.67 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 192.2, 157.4, 150.6, 147.2, 136.2, 135.8, 132.9, 132.4, 132.3, 129.0, 128.5, 128.2, 125.8, 121.1, 118.9, 116.3, 56.0.

HRMS: m/z calcd for  $C_{17}H_{14}NO_2$  [M+H]<sup>+</sup>: 259.0871, found 259.0869.

1-(Quinolin-8-yl)-2-naphthaldehyde (5f)



Yellow solid. 1288 mg, 91% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.71 (s, 1H), 8.78 (s, 1H), 8.26 (d, *J* = 8.3 Hz, 1H), 8.19 (d, *J* = 8.6 Hz, 1H), 8.01 (d, *J* = 8.2 Hz, 2H), 7.94 (d, *J* = 8.3 Hz, 1H), 7.74 (d, *J* = 7.0 Hz, 1H), 7.69 (t, *J* = 7.6 Hz, 1H), 7.56 (t, *J* = 7.5 Hz, 1H), 7.40 (q, *J* = 7.4, 5.4 Hz, 2H), 7.33 (t, *J* = 7.7 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 192.6, 151.0, 147.8, 144.5, 136.3, 134.9, 133.0, 132.6, 131.9, 129.0, 128.7, 128.6, 128.4, 128.3, 127.7, 126.7, 125.8, 122.2, 121.6.

The NMR spectra is in agreement with published data.<sup>6</sup>

HRMS: m/z calcd for C<sub>20</sub>H<sub>14</sub>NO [M+H]<sup>+</sup>: 284.1075, found 284.1072.

2-(5-Chloroquinolin-8-yl)-3-methylbenzaldehyde (5g)



Yellow solid. 829 mg, 59% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.50 (s, 1H), 8.90 (dd, *J* = 4.3, 1.7 Hz, 1H), 8.68 (dd, *J* = 8.6, 1.7 Hz, 1H), 7.93 (d, *J* = 7.7 Hz, 1H), 7.74 (d, *J* = 7.7 Hz, 1H), 7.58 (d, *J* = 7.5 Hz, 1H), 7.56 – 7.45 (m, 3H), 1.99 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 192.3, 151.4, 147.5, 142.3, 138.1, 136.0, 135.6, 135.1, 133.3, 131.9, 131.1, 128.3, 126.6, 126.3, 125.3, 122.3, 20.1.

The NMR spectra is in agreement with published data.<sup>6</sup>

HRMS: m/z calcd for C17H13CINO [M+H]+: 282.0686, found 282.0694.

3-Methyl-2-(2-methylquinolin-8-yl)-benzaldehyde (5h)



Yellow solid. 692 mg, 53% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.45 (s, 1H), 8.08 (d, *J* = 8.5 Hz, 1H), 7.92 (d, *J* = 7.8 Hz, 1H), 7.86 (d, *J* = 7.5 Hz, 1H), 7.59 - 7.51 (m, 3H), 7.45 (s, 1H), 7.28 (s, 1H), 2.54 (s, 3H), 2.02 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 193.2, 159.5, 146.6, 143.8, 138.2, 136.1, 135.6, 135.3, 135.2, 131.6, 128.3, 127.5, 126.6, 125.0, 124.3, 122.3, 25.7, 20.3.

HRMS: m/z calcd for  $C_{18}H_{16}NO [M+H]^+$ : 262.1232, found 262.1236.

3-Methyl-2-(quinoxalin-5-yl)benzaldehyde (5i)



Yellow amorphous. 460 mg, 74% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.49 (s, 1H), 8.88 (d, *J* = 1.8 Hz, 1H), 8.79 (d, *J* = 1.8 Hz, 1H), 8.24 (d, *J* = 8.5 Hz, 1H), 7.94 (d, *J* = 7.8 Hz, 1H), 7.92 - 7.85 (m, 1H), 7.67 (d, *J* = 8.5 Hz, 1H), 7.59 (d, *J* = 7.6 Hz, 1H), 7.51 (t, *J* = 7.7 Hz, 1H), 1.98 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 192.1, 145.4, 145.3, 143.2, 142.0, 141.5, 138.2, 137.1, 135.6, 135.2, 131.9, 130.2, 129.7, 128.5, 125.6, 20.1.

HRMS: m/z calcd for  $C_{16}H_{13}N_2O$  [M+H]<sup>+</sup>: 249.1028, found 249.1029.

2-(Benzo[d]thiazol-4-yl)-3-methylbenzaldehyde (5j)



Colorless amorphous. 405 mg, 32% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.57 (s, 1H), 8.97 (s, 1H), 8.06 (d, *J* = 8.2 Hz, 1H), 7.94 (d, *J* = 7.8 Hz, 1H), 7.61 - 7.52 (m, 2H), 7.48 (t, *J* = 7.7 Hz, 1H), 7.38 (d, *J* = 7.3 Hz, 1H), 2.07 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 192.4, 154.7, 152.0, 142.2, 137.8, 135.7, 134.6, 134.4, 132.2, 128.4, 128.2, 125.4, 125.1, 122.2, 20.1.

HRMS: m/z calcd for C15H12NOS [M+H]+: 254.0640, found 254.0652.

2-(Benzo[d]oxazol-4-yl)-3-methylbenzaldehyde (5k)



Colorless amorphous. 436 mg, 37% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.60 (s, 1H), 8.09 (s, 1H), 7.92 (d, *J* = 7.9 Hz, 1H), 7.68 (dd, *J* = 8.2, 1.0 Hz, 1H), 7.57 (d, *J* = 7.3 Hz, 1H), 7.53 – 7.44 (m, 2H), 7.25 (d, *J* = 7.4 Hz, 1H), 2.11 (s, 3H) <sup>13</sup>C NMR (126 MHz, CDCl3)  $\delta$  192.3, 153.1, 149.9, 140.2, 139.3, 138.0, 135.7, 134.7, 129.6, 128.5, 126.7, 125.6, 125.3, 111.0, 20.1.

HRMS: m/z calcd for  $C_{15}H_{12}NO_2$  [M+H]<sup>+</sup>: 238.0868, found 238.0868.

3-Methyl-2-(1-methyl-1*H*-benzo[*d*]imidazol-4-yl)benzaldehyde (5I)



White solid. 543 mg, 53% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.54 (s, 1H), 7.90 (d, *J* = 7.8 Hz, 1H), 7.85 (s, 1H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.47 (d, *J* = 8.1 Hz, 1H), 7.41 (dt, *J* = 15.4, 7.7 Hz, 2H), 7.14 (d, *J* = 7.2 Hz, 1H), 3.89 (s, 3H), 2.10 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 193.0, 144.1, 142.8, 142.1, 138.1, 135.6, 134.8, 134.6, 128.6, 128.0, 124.7, 124.23, 122.8, 109.6, 31.3, 20.1.

HRMS: m/z calcd for C<sub>16</sub>H<sub>15</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 254.1184, found 254.1187.



# **General procedure**

To a stirred solution of **S-6** (5 mmol, 1.0 equiv.) in 1,2-dimethoxyethane (DME, 20 mL) and H<sub>2</sub>O (10 mL) was added 2-bromopyridine 1-oxide **S-7** (4 mmol, 0.8 equiv.), Na<sub>2</sub>CO<sub>3</sub> (1.59 g, 15 mmol, 3.0 equiv.) and Pd(PPh<sub>3</sub>)<sub>4</sub> (577 mg, 0.5 mmol, 0.1 equiv.), the reaction mixture was heated at reflux overnight. After completion of the reaction, the solvent was removed under reduced pressure. The resulting mixture was extracted with Dichloromethane (DCM) (25 mL × 3), and then organic layers were collected and dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated to provide the crude product, which was further purified by flash chromatography (MeOH/DCM, 5%, v/v) to give products **28a-28m**.

2-(2-Formyl-6-methylphenyl)-pyridine 1-oxide (28a)



White solid. 650 mg, 61% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.76 (s, 1H), 8.40 – 8.36 (m, 1H), 7.84 (dd, *J* = 7.6, 1.9 Hz, 1H), 7.60 – 7.51 (m, 2H), 7.38 – 7.32 (m, 2H), 7.26 – 7.23 (m, 1H), 2.19 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 191.0, 147.2, 140.3, 138.4, 136.0, 134.7, 134.1, 129.9, 128.2, 128.2, 125.7, 125.4, 19.2.

HRMS: m/z calcd for C13H10NO2 [M+H]+: 214.0868, found 214.0862.

2-(2-Chloro-6-formylphenyl)pyridine 1-oxide (28b)

White solid. 950 mg, 41% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.72 (s, 1H), 8.40 – 8.35 (m, 1H), 7.93 (d, *J* = 7.6 Hz, 1H), 7.78 (d, *J* = 8.0 Hz, 1H), 7.60 (t, *J* = 7.9 Hz, 1H), 7.44 – 7.37 (m, 3H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 189.3, 144.9, 140.3, 136.6, 135.2, 135.1, 132.9, 131.2, 130.1, 128.7, 128.4, 126.4, 125.9.

HRMS: m/z calcd for C12H9CINO2 [M+H]+: 234.0322, found 234.0327.

2-(2-Formyl-6-methoxyphenyl)-pyridine 1-oxide (28c)

Yellow solid. 954 mg, 83% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 9.74 (s, 1H) 8.38 – 8.33 (m, 1H), 7.65 – 7.57 (m, 2H), 7.40 – 7.32 (m, 3H), 7.32 – 7.24 (m, 1H), 3.82 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl3) δ 190.5, 157.4, 144.4, 140.1, 135.9, 131.3, 129.4, 125.6, 125.3, 123.0, 121.5, 116.8, 56.3.

HRMS: m/z calcd for  $C_{13}H_{12}NO_3$  [M+H]<sup>+</sup>: 230.0817, found 230.0802.

2-(2-FormyInaphthalen-1-yl)-pyridine 1-oxide (28d)



White solid. 710 mg, 57% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.87 (s, 1H), 8.43 (d, *J* = 6.3 Hz, 1H), 8.09 – 8.01 (m, 2H), 7.93 (d, *J* = 8.2 Hz, 1H), 7.61 (t, *J* = 7.5 Hz, 1H), 7.53 – 7.36 (m, 5H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 190.7, 145.4, 140.5, 136.3, 135.4, 131.9, 131.1, 130.5, 129.6, 129.2, 128.7, 127.9, 126.3, 125.8, 125.3, 123.1.

The NMR spectra is in agreement with published data.<sup>5</sup>

HRMS: m/z calcd for C<sub>16</sub>H<sub>12</sub>NO<sub>2</sub> [M-H]<sup>-</sup>: 250.0868, found 250.0871.

2-(2-Formyl-6-methylphenyl)-4-methylpyridine 1-oxide (28e)

Me M

Yellow solid. 420 mg, 37% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.74 (s, 1H), 8.27 (d, *J* = 6.6 Hz, 1H), 7.83 (dd, *J* = 7.5, 1.7 Hz, 1H), 7.60 - 7.47 (m, 2H), 7.16 (dd, *J* = 6.7, 2.6 Hz, 1H), 7.05 (d, *J* = 2.6 Hz, 1H), 2.38 (s, 3H), 2.20 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 191.1, 146.3, 139.6, 138.4, 137.0, 136.0, 134.7, 134.4, 129.8, 128.7, 127.8, 126.5, 20.5, 19.3.

HRMS: m/z calcd for C<sub>14</sub>H<sub>14</sub>NO<sub>2</sub> [M+H]<sup>+</sup>: 228.1025, found 228.1033.

4-Chloro-2-(2-formyl-6-methylphenyl)-4-chloro-pyridine 1-oxide (28f)

White solid. 327 mg, 26% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.79 (d, *J* = 1.3 Hz, 1H), 8.25 (d, *J* = 7.0 Hz, 1H), 7.79 (dd, *J* = 6.5, 2.5 Hz, 1H), 7.61 – 7.47 (m, 2H), 7.30 (dd, *J* = 7.0, 3.0 Hz, 1H), 7.20 (d, *J* = 3.0 Hz, 1H), 2.19 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 190.8, 148.5, 140.7, 138.4, 136.1, 134.6, 132.2, 131.2, 130.2, 129.5, 127.6, 125.8, 19.2.

HRMS: m/z calcd for C<sub>13</sub>H<sub>11</sub>CINO<sub>2</sub> [M+H]<sup>+</sup>: 248.0748, found 248.0783.

4-Fluoro-2-(2-formyl-6-methylphenyl) pyridine 2-oxide (28g)



White solid. 693 mg, 60% yield.

<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.34 (s, 1H), 8.35 (dd, *J* = 5.0, 2.3 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 1H), 7.34 (d, *J* = 7.5 Hz, 1H), 7.28 (d, *J* = 7.7 Hz, 2H), 7.26 – 7.20 (m, 1H), 7.18 – 7.11 (m, 1H), 1.75 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  197.0, 165.0 (d, *J* = 249.5 Hz), 148.4 (d, *J* = 3.9 Hz), 143.6, 141.0, 139.9, 138.6, 136.8 (d, *J* = 9.7 Hz), 134.9, 134.1 (d, *J* = 11.6 Hz), 133.5 (d, *J* = 9.6 Hz), 133.0, 118.8 (d, *J* = 20.1 Hz), 23.9.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -114.8.

HRMS: m/z calcd for C<sub>13</sub>H<sub>11</sub>FNO<sub>2</sub> [M+H]<sup>+</sup>: 232.0774, found 232.0769.

2-(2-Formyl-6-methylphenyl)-5-methylpyridine 2-oxide (28h)



White solid. 590 mg, 52% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.72 (s, 1H), 8.22 (s, 1H), 7.81 (d, *J* = 7.5 Hz, 1H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.50 (t, *J* = 7.6 Hz, 1H), 7.16 (d, *J* = 7.9 Hz, 1H), 7.12 (d, *J* = 8.0 Hz, 1H), 2.37 (s, 3H), 2.17 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  191.1, 144.0, 140.1, 138.5, 136.4, 135.9, 134.7, 134.3, 129.7, 127.7, 127.5, 126.8, 19.2, 18.3.

HRMS: m/z calcd for  $C_{14}H_{14}NO_2[M+H]^+$ : , found 228.1025, found 228.1028.

2-(2-Formyl-6-methylphenyl)-5-(trifluoromethyl)-pyridine 1-oxide (28i)



Yellow solid. 717 mg, 51% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.81 (s, 1H), 8.61 (s, 1H), 7.85 – 7.77 (m, 1H), 7.63 – 7.57 (m, 2H), 7.52 (dd, J = 8.3, 1.6 Hz, 1H), 7.35 (d, J = 8.1 Hz, 1H), 2.18 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 190.9, 151.0, 138.4, 137.7 (q, *J* = 3.8 Hz), 136.3, 134.6, 131.6, 130.43, 129.7 (q, *J* = 34.9 Hz), 128.02, 122.0 (q, *J* = 273.0 Hz), 121.4, 19.2.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -63.1.

HRMS: m/z calcd for C14H11F3NO2 [M+H]+: 282.0742, found 282.0745.

2-(2-Formyl-6-methylphenyl)-6-methylpyridine 2-oxide (28j)



White solid. 545 mg, 48% yield.

<sup>1</sup>H NMR (500 MHz, DMSO-*a*<sub>6</sub>)  $\delta$  9.57 (s, 1H), 7.80 (d, *J* = 7.6 Hz, 1H), 7.67 (d, *J* = 7.6 Hz, 1H), 7.61 – 7.54 (m, 2H), 7.39 (d, *J* = 7.8 Hz, 1H), 7.35 (t, *J* = 8.4 Hz, 1H), 2.41 (s, 3H), 2.07 (s, 3H).

<sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>) δ 191.4, 148.6, 145.2, 137.8, 135.7, 135.5, 134.3, 129.2, 126.5, 126.3, 126.0, 124.2, 18.8, 17.7.

HRMS: m/z calcd for C14H14NO2 [M+H]+: 228.1025, found 228.1028.

1-(2-Formylphenyl)isoquinoline 1-oxide (28k)



Red solid. 988 mg, 79% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.73 (s, 1H), 8.23 (d, *J* = 7.2 Hz, 1H), 8.10 (dd, *J* = 7.7, 1.4 Hz, 1H), 7.82 (d, *J* = 8.1 Hz, 1H), 7.78 (td, *J* = 7.5, 1.4 Hz, 1H), 7.73 (d, *J* = 7.2 Hz, 1H), 7.69 (t, *J* = 7.6 Hz, 1H), 7.55 (ddd, *J* = 8.2, 6.9, 1.2 Hz, 1H), 7.46 (td, *J* = 7.7, 7.0, 1.3 Hz, 1H), 7.42 (d, *J* = 7.6 Hz, 1H), 7.27 (d, *J* = 9.1 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 190.4, 143.8, 136.9, 135.3, 134.2, 132.6, 131.2, 130.2, 130.1, 129.6, 129.5, 129.0, 128.6, 127.2, 124.8, 124.1.

The NMR spectra is in agreement with published data.<sup>5</sup>

HRMS: m/z calcd for  $C_{16}H_{12}NO_2$  [M+H]<sup>+</sup>: 250.0868, found 250.0873.

2-(2-formyl-6-methylphenyl)-3-methylpyridine 1-oxide (28I)



White solid. 864 mg, 76% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.74 (s, 1H), 8.26 (d, *J* = 6.2 Hz, 1H), 7.85 (d, *J* = 7.6 Hz, 1H), 7.61 (d, *J* = 7.6 Hz, 1H), 7.55 (t, *J* = 7.6 Hz, 1H), 7.29 - 7.19 (m, 2H), 2.12 (s, 3H), 1.93 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  191.2, 147.1, 138.0, 137.7, 136.5, 136.4, 134.1, 133.4, 129.8, 129.2,

127.5, 124.6, 19.0, 18.7.

HRMS: m/z calcd for C14H14NO2 [M+H]+: 228.1033, found 228.1025.

# Synthesis of heterobiaryl reference compounds



## **General procedure**

To a stirred solution of aldehyde **5a–5j** and **28a–28m** (0.4 mmol, 1.0 equiv) in dry DCM (10 mL) was added amine (0.5 mmol, 1.2 equiv) and HOAc (0.04 mmol, 0.1 equiv), the reaction mixture was stirred overnight. After completion of the reaction, the solvent was removed under reduced pressure to give the crude product (Schiff base) which was used for the next step without further purification. To the solution of Schiff base in THF/MeOH (2:1, 15 mL) under ice-bath was added BH<sub>3</sub> (1 M in THF, 0.8 mmol, 2.0 equiv) or NaBH<sub>3</sub>CN (4 mmol, 10 equiv), and then the reaction mixture was stirred for at 0 °C for 5 hours. After completion of the reaction, the reaction mixture was quenched by saturated NH<sub>4</sub>Cl solution (20 mL). The resulting reaction mixture was concentrated under reduced pressure and diluted with water (20 mL), then extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with brine (50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated to provide a crude product, which was further purified by flash chromatography to provide the racemic heterobiaryl amine products (**7–27** and **29–50**).

N-Benzyl-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (7)



Colorless amorphous. 64 mg, 47% yield.

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  8.73 (dd, *J* = 4.1, 1.8 Hz, 1H), 8.21 (dd, *J* = 8.2, 1.8 Hz, 1H), 7.86 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.59 (t, *J* = 7.5 Hz, 1H), 7.51 (d, *J* = 6.9 Hz, 1H), 7.45 (d, *J* = 7.6 Hz, 1H), 7.40 – 7.32 (m, 2H), 7.27 (d, *J* = 7.6 Hz, 1H), 7.15 (dd, *J* = 4.9, 1.9 Hz, 3H), 6.99 – 6.94 (m, 2H), 3.56 (d, *J* = 13.2 Hz, 1H), 3.49 – 3.44 (m, 2H), 3.42 (d, *J* = 13.2 Hz, 1H), 2.37 (s, 1H), 1.88 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 150.6, 146.8, 140.3, 139.6, 139.3, 138.4, 136.9, 136.3, 130.5, 128.6, 128.6, 128.1, 127.9, 127.7, 127.7, 126.7, 126.6, 126.3, 121.1, 53.1, 51.0, 20.7.

HRMS: m/z calcd for C<sub>24</sub>H<sub>23</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 339.1861, found 339.1861.

N-(2-Fluorobenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (8)



White solid. 16 mg, 10% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.68 (d, *J* = 5.2 Hz, 1H), 8.15 – 8.08 (m, 1H), 7.76 (t, *J* = 6.6 Hz, 1H), 7.50 (q, *J* = 7.4 Hz, 1H), 7.45 – 7.38 (m, 2H), 7.32 – 7.26 (m, 2H), 7.19 (t, *J* = 6.4 Hz, 1H), 7.07 – 6.98 (m, 1H), 6.97 – 6.89 (m, 1H), 6.86 – 6.74 (m, 2H), 3.51 (dd, *J* = 13.6, 4.8 Hz, 1H), 3.43 (dd, *J* = 13.8, 4.9 Hz, 1H), 3.39 – 3.34 (m, 2H), 2.35 (brs, 1H), 1.81 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 150.7, 146.8, 139.8, 139.3, 137.6 (d, J = 1.26 Hz, 1H), 137.1, 136.5, 130.7, 130.2, 129.0, 128.7, 128.5 (d, J = 2.52 Hz, 1H), 128.4, 128.0, 127.8 (d, J = 2.52 Hz, 1H), 127.0, 126.4, 123.9, 121.2, 115.1 (d, J = 3.78 Hz, 1H), 115.0, 51.8, 46.2, 20.8.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) *δ* -116.4.

HRMS: m/z calcd for  $C_{24}H_{22}FN_2$  [M+H]<sup>+</sup>: 357.1767, found 357.1783.

N-(3-Methyl-2-(quinolin-8-yl)benzyl)-1-(m-tolyl)methanamine (9)



White solid. 16 mg, 10% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.80 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.20 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.85 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.62 - 7.56 (m, 1H), 7.52 (dd, *J* = 7.0, 1.6 Hz, 1H), 7.42 (d, *J* = 6.3 Hz, 1H), 7.40 - 7.31 (m, 2H), 7.30 - 7.23 (m, 1H), 7.04 (t, *J* = 7.5 Hz, 1H), 6.95 (d, *J* = 7.5 Hz, 1H), 6.83 (s, 1H), 6.75 (d, *J* = 7.5 Hz, 1H), 3.49 - 3.41 (m, 3H), 3.37 (d, *J* = 13.1 Hz, 1H), 2.25 (s, 3H), 1.90 (s, 3H), 1.83 (brs, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  150.6, 146.8, 140.2, 139.6, 139.3, 138.4, 137.7, 137.0, 136.4, 130.6, 128.8, 128.7, 128.7, 128.1, 127.8, 127.8, 127.4, 126.8, 126.4, 125.0, 121.2, 53.1, 52.0, 21.4, 20.7. HRMS: m/z calcd for C<sub>25</sub>H<sub>25</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 353.2018 , found 353.2014.

N-(3-Methoxybenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (10)



Colorless amorphous. 76 mg, 52% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.73 (dd, *J* = 4.3, 1.8 Hz, 1H), 8.18 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.84 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.58 (t, *J* = 7.6 Hz, 1H), 7.50 (dd, *J* = 7.1, 1.5 Hz, 1H), 7.43 (d, *J* = 7.5 Hz, 1H), 7.38 – 7.29 (m, 2H), 7.24 (s, 1H), 7.05 (t, *J* = 7.8 Hz, 1H), 6.68 (dd, *J* = 8.0, 2.6 Hz, 1H), 6.63 (s, 1H), 6.53 (d, *J* = 7.6 Hz, 1H), 3.73 (s, 3H), 3.50 (d, *J* = 13.3 Hz, 1H), 3.43 (s, 2H), 3.38 (d, *J* = 13.2 Hz, 1H), 1.87 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  159.7, 150.8, 146.6, 139.4, 139.0, 137.2, 136.7, 131.0, 129.5, 129.3, 128.7, 128.0, 128.0, 127.4, 126.6, 121.4, 120.8, 113.8, 113.3, 55.4, 52.4, 51.3, 20.8. HRMS: m/z calcd for C<sub>25</sub>H<sub>25</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 369.1967, found 369.1967.

N-(3-Bromobenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (11)



Yellow amorphous. 21 mg, 13% yield.

<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  8.50 (dd, *J* = 4.1, 2.2 Hz, 1H), 7.68 – 7.54 (m, 2H), 7.54 – 7.51 (m, 3H), 7.50 – 7.47 (m, 1H), 7.47 – 7.39 (m, 6H), 4.26 (d, *J* = 13.0 Hz, 1H), 4.14 – 4.08 (m, 2H), 4.09 (d, *J* = 10.0 Hz, 1H), 3.68 (d, *J* = 13.1 Hz, 1H), 2.05 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 148.0, 141.2, 137.5, 132.9, 132.8, 132.8, 132.6, 132.3, 131.1, 130.9, 130.9, 130.4, 129.9, 129.1, 128.9, 126.9, 123.1, 50.1, 50.1, 20.4.

HRMS: m/z calcd for  $C_{24}H_{22}BrN_2$  [M+H]<sup>+</sup>: 417.0966, found 417.0940.

N-(4-Methoxybenzyl)-1-[3-methyl-2-(quinolin-8-yl)phenyl]methanamine (12)



Yellow amorphous. 15 mg, 10% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.44 (d, *J* = 4.2 Hz, 1H), 8.30 (d, *J* = 6.5 Hz, 1H), 7.93 (d, *J* = 8.2 Hz, 1H), 7.64 (t, *J* = 7.6 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 2H), 7.44 – 7.37 (m, 2H), 7.32 (d, *J* = 7.6 Hz, 1H), 6.87 (d, *J* = 8.2 Hz, 2H), 6.72 (d, *J* = 8.7 Hz, 2H), 5.35 (brs, 1H), 3.76 (s, 3H), 3.75 (d, *J* = 6.1 Hz, 1H), 3.72 (d, *J* = 4.4 Hz, 1H), 3.58 (d, *J* = 12.6 Hz, 1H), 3.36 (d, *J* = 13.0 Hz, 1H), 1.78 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  159.8, 150.9, 146.1, 139.5, 138.1, 137.8, 137.7, 132.4, 131.8, 131.1,

130.4, 129.0, 128.7, 128.6, 128.0, 127.1, 125.5, 121.8, 114.3, 55.4, 51.2, 51.0, 20.9.

HRMS: m/z calcd for  $C_{25}H_{25}N_2O$  [M+H]<sup>+</sup>: 369.1967 , found 369.1968.

*N*-[3-methyl-2-(quinolin-8-yl)benzyl]-1-[4-(trifluoromethyl)phenyl]methanamine (13)



Yellow amorphous. 17 mg, 10% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.64 (d, *J* = 4.1 Hz, 1H), 8.11 (d, *J* = 7.8 Hz, 1H), 7.75 (d, *J* = 8.1 Hz, 1H), 7.48 (t, *J* = 7.6 Hz, 1H), 7.39 (d, *J* = 6.9 Hz, 1H), 7.33 (d, *J* = 7.6 Hz, 1H), 7.29 – 7.24 (m, 4H), 7.18 (d, *J* = 11.3 Hz, 2H), 6.93 (d, *J* = 7.9 Hz, 2H), 3.50 (d, *J* = 13.9 Hz, 1H), 3.41 – 3.29 (m, 3H), 1.98 (brs, 1H), 1.78 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 150.6, 146.8, 143.8, 139.4 (2), 137.3, 136.6, 130.7, 129.2, 128.7, 128.2, 128.0, 127.9, 127.1, 126.5, 125.1 (q, *J* = 3.7 Hz), 124.4 (q, *J* = 277.8 Hz), 121.3, 52.4, 51.7, 20.7. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -58.1.

HRMS: m/z calcd for  $C_{25}H_{22}F_3N_2$  [M+H]<sup>+</sup>: 407.1735 , found 407.1750.

N-(pyridin-3-ylmethyl)-1-[2-(quinolin-8-yl)-3-methylphenyl]methanamine (14)



White amorphous. 72 mg, 53% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.78 (d, *J* = 3.7 Hz, 1H), 8.18 (d, *J* = 5.3 Hz, 1H), 7.83 (d, *J* = 6.5 Hz, 1H), 7.56 (t, *J* = 7.6 Hz, 1H), 7.49 (d, *J* = 7.0 Hz, 1H), 7.44 (d, *J* = 7.6 Hz, 1H), 7.37 – 7.32 (m, 2H), 7.25 (d, *J* = 5.3 Hz, 1H), 7.09 (q, *J* = 6.9 Hz, 1H), 6.95 (t, *J* = 7.5 Hz, 1H), 6.92 – 6.80 (m, 2H), 3.55 (d, *J* = 13.8 Hz, 1H), 3.48 (d, *J* = 13.8 Hz, 1H), 3.43 – 3.37 (m, 2H), 1.95 (brs, 1H), 1.88 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 150.7, 149.6, 148.4, 146.7, 139.4, 139.3, 137.5, 137.3, 136.6, 135.8, 135.0, 130.6, 129.1, 128.7, 127.9, 127.0, 126.5, 123.3, 121.3, 51.9, 50.2, 20.7.

HRMS: m/z calcd for  $C_{23}H_{22}N_3$  [M+H]<sup>+</sup>: 340.1814, found 340.1812.

1-(Furan-2-yl)-N-(3-methyl-2-(quinolin-8-yl)benzyl)methanamine (15)



Yellow amorphous. 16 mg, 10% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.83 (s, 1H), 8.21 (d, *J* = 8.2 Hz, 1H), 7.86 (d, *J* = 8.1 Hz, 1H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.51 (d, *J* = 7.1 Hz, 1H), 7.43 – 7.37 (m, 2H), 7.34 (t, *J* = 7.6 Hz, 1H), 7.26 (d, *J* = 7.2 Hz, 1H), 7.13 (s, 1H), 6.12 (s, 1H), 5.82 (s, 1H), 3.48 (d, *J* = 14.3 Hz, 1H), 3.43 – 3.33 (m, 3H), 1.93 (brs, 1H), 1.89 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 153.8, 150.7, 146.8, 141.5, 139.4, 139.3, 138.1, 137.0, 136.4, 130.6, 128.8, 128.7, 127.8, 127.8, 126.8, 126.4, 121.2, 109.9, 106.6, 51.5, 45.4, 20.8. HRMS: m/z calcd for C<sub>22</sub>H<sub>21</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 329.1654, found 329.1655.

N-(prop-2-yn-1-yl)-1-[2-(quinolin-8-yl)-3-methylphenyl]methanamine (16)

Me

Yellow amorphous. 94 mg, 33% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.07 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.43 (dd, *J* = 8.3, 1.8 Hz, 1H), 8.09 (d, *J* = 8.2 Hz, 1H), 7.83 (t, *J* = 7.6 Hz, 1H), 7.75 (dd, *J* = 7.0, 1.5 Hz, 1H), 7.65 – 7.60 (m, 2H), 7.56 (t, *J* = 7.6 Hz, 1H), 7.50 – 7.47 (m, 1H), 3.69 (d, *J* = 12.5 Hz, 1H), 3.61 (d, *J* = 12.5 Hz, 1H), 3.36 (dd, *J* = 16.9, 2.5 Hz, 1H), 3.29 (dd, *J* = 16.9, 2.4 Hz, 1H), 2.12 (s, 3H), 1.98 (t, *J* = 2.4 Hz, 1H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  150.6, 146.9, 139.4, 139.4, 137.8, 137.1, 136.5, 130.7, 128.9, 128.8,

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) o 150.6, 146.9, 139.4, 139.4, 137.8, 137.1, 136.5, 130.7, 128.9, 128.8, 127.9, 127.7, 126.9, 126.4, 121.3, 81.6, 70.7, 50.9, 37.4, 20.8.

HRMS: m/z calcd for  $C_{20}H_{19}N_2$  [M+H]<sup>+</sup>: 287.1548, found 287.1553.

*N*-Benzyl-1-[3-chloro-2-(quinolin-8-yl)phenyl]methanamine (**17**)



Yellow amorphous. 104 mg, 72% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.82 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.21 (dd, *J* = 8.2, 1.8 Hz, 1H), 7.90 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.66 – 7.51 (m, 3H), 7.49 (dd, *J* = 8.0, 1.3 Hz, 1H), 7.43 – 7.36 (m, 2H), 7.23 – 7.12 (m, 3H), 7.05 – 6.96 (m, 2H), 3.53 (d, *J* = 13.2 Hz, 1H), 3.50 – 3.39 (m, 3H), 1.82 (brs, 1H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  150.7, 146.6, 141.0, 140.0, 138.3, 137.4, 136.4, 134.4, 130.9, 128.9, 128.4, 128.2, 128.1, 128.0, 127.9, 127.8, 127.5, 126.7, 126.2, 121.3, 53.1, 51.7. HRMS: m/z calcd for C<sub>23</sub>H<sub>20</sub>ClN<sub>2</sub> [M+H]<sup>+</sup>: 359.1315, found 359.1302.

N-Benzyl-1-[2-(quinolin-8-yl)-3-(trifluoromethyl)phenyl]methanamine (18)



Yellow amorphous. 85 mg, 54% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.78 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.24 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.95 (dd, *J* = 6.1, 3.6 Hz, 1H), 7.90 (d, *J* = 6.5 Hz, 1H), 7.72 (d, *J* = 7.7 Hz, 1H), 7.67 – 7.61 (m, 2H), 7.54 (t, *J* = 7.8 Hz, 1H), 7.42 (dd, *J* = 8.3, 4.2 Hz, 1H), 7.23 – 7.13 (m, 3H), 7.01 (dd, *J* = 7.2, 2.4 Hz, 2H), 3.54 (d, *J* = 13.3 Hz, 1H), 3.52 – 3.44 (m, 3H), 2.08 (brs, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  150.9, 146.5, 143.3, 140.5, 139.7, 136.6, 136.3, 133.5, 131.6, 131.0, 129.4, 128.7, 128.4, 128.4, 128.0, 127.0, 126.3, 121.6, 118.5, 114.3, 53.3, 51.0. HRMS: m/z calcd for C<sub>24</sub>H<sub>20</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 350.1657, found 350.1669. N-Benzyl-1-[2-(quinolin-8-yl)-3-(trifluoromethyl)phenyl]methanamine (19)



Yellow amorphous. 105 mg, 54% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.76 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.20 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.89 (dd, *J* = 7.5, 2.1 Hz, 1H), 7.83 (d, *J* = 7.7 Hz, 1H), 7.76 (d, *J* = 9.3 Hz, 1H), 7.57 (qd, *J* = 7.5, 3.1 Hz, 3H), 7.37 (dd, *J* = 8.3, 4.2 Hz, 1H), 7.22 - 7.11 (m, 3H), 7.03 - 6.95 (m, 2H), 3.50 (d, *J* = 13.3 Hz, 1H), 3.45 - 3.29 (m, 3H), 2.04 (brs, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 150.5, 147.2, 140.8, 140.0, 138.1, 136.4 (×2), 132.5, 131.1, 129.7 (q, *J* = 29.2 Hz), 128.6, 128.3, 128.2, 128.0 127.9, 126.8, 125.8, 124.94 (q, *J* = 5.2 Hz), 124.2 (d, *J* = 274.3 Hz), 121.2, 53.3, 51.1.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -58.4.

HRMS: m/z calcd for  $C_{24}H_{20}F_3N_2$  [M+H]<sup>+</sup>: 393.1579, found 393.1582.

*N*-Benzyl-1-[3-methoxy-2-(quinolin-8-yl)phenyl]methanamine (20)



Yellow solid. 120 mg, 34% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.79 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.19 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.85 (dd, *J* = 7.3, 2.4 Hz, 1H), 7.59 (q, *J* = 6.5, 5.9 Hz, 2H), 7.44 (t, *J* = 8.0 Hz, 1H), 7.36 (dd, *J* = 8.2, 4.2 Hz, 1H), 7.24 (d, *J* = 7.7 Hz, 1H), 7.16 (q, *J* = 5.3 Hz, 3H), 7.00 (dd, *J* = 5.3, 3.0 Hz, 3H), 3.64 (s, 3H), 3.53 (d, *J* = 13.2 Hz, 1H), 3.48 – 3.40 (m, 3H), 1.94 (brs, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 157.4, 150.4, 147.2, 140.2, 140.1, 136.6, 136.4, 131.3, 128.8, 128.6, 128.6, 128.2, 128.0, 127.8, 126.7, 126.2, 121.7, 121.0, 110.0, 56.0, 53.1, 51.4.

HRMS: m/z calcd for  $C_{24}H_{23}N_2O$  [M+H]<sup>+</sup>: 355.1810, found 355.1829.

N-Benzyl-1-[1-(quinolin-8-yl)naphthalen-2-yl]methanamine (21)



Yellow amorphous. 67 mg, 46 % yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.69 (dd, *J* = 4.2, 1.8 Hz, 1H), 8.26 (dd, *J* = 8.3, 1.8 Hz, 1H), 7.96 (t, *J* = 7.6 Hz, 2H), 7.90 (d, *J* = 8.2 Hz, 1H), 7.80 (d, *J* = 8.5 Hz, 1H), 7.66 (t, *J* = 7.5 Hz, 1H), 7.64 – 7.60 (m, 1H), 7.41 (t, *J* = 7.5 Hz, 1H), 7.38 (dd, *J* = 8.3, 4.1 Hz, 1H), 7.26 – 7.19 (m, 1H), 7.17 (h, *J* = 3.5 Hz, 3H), 7.11 (d, *J* = 8.5 Hz, 1H), 7.04 – 6.98 (m, 2H), 3.64 – 3.58 (m, 3H), 3.50 (d, *J* = 13.3 Hz, 1H), 2.29 (brs, 1H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  150.7, 147.6, 140.3, 138.4, 136.4, 136.3, 136.2, 133.5, 132.9, 131.8, 128.6, 128.2, 128.1, 127.9, 127.5, 126.7, 126.6, 126.3, 125.8, 125.3, 121.3, 53.4, 51.9. HRMS: m/z calcd for C<sub>27</sub>H<sub>23</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 375.1861, found 375.1862.

N-Benzyl-1-[2-(5-chloroquinolin-8-yl)-3-methylphenyl]methanamine (22)



Yellow amorphous. 108 mg, 86% yield .

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.94 (dd, J = 4.2, 1.7 Hz, 1H), 8.74 (dd, J = 8.6, 1.8 Hz, 1H), 7.75 (d, J = 7.7 Hz, 1H), 7.57 (dd, J = 8.6, 4.2 Hz, 1H), 7.55 – 7.51 (m, 2H), 7.46 (t, J = 7.6 Hz, 1H), 7.39 – 7.29 (m, 1H), 7.26 – 7.20 (m, 3H), 7.06 – 6.99 (m, 2H), 3.55 (d, J = 13.4 Hz, 1H), 3.51 (d, J = 12.8 Hz, 1H), 3.45 (d, J = 12.9 Hz, 1H), 1.99 (s, 3H), 1.69 (brs, 1H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 151.1, 147.4, 140.2, 138.9, 138.5, 136.9, 133.1, 130.8, 130.2, 128.7, 128.2, 128.0, 127.9, 126.8, 126.7, 126.5, 121.9, 53.2, 51.6, 20.7.

HRMS: m/z calcd for C<sub>24</sub>H<sub>22</sub>ClN<sub>2</sub> [M+H]<sup>+</sup>: 373.1472, found 373.1458.

N-Benzyl-1-[3-methyl-2-(2-methylquinolin-8-yl)phenyl]methanamine (23)



Yellow amorphous. 92 mg, 65 % yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.14 (d, *J* = 8.4 Hz, 1H), 7.88 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.63 – 7.48 (m, 3H), 7.43 (t, *J* = 7.5 Hz, 1H), 7.34 (d, *J* = 7.5 Hz, 1H), 7.29 (d, *J* = 8.4 Hz, 1H), 7.27 – 7.18 (m, 3H), 7.07 – 7.00 (m, 2H), 3.60 (d, *J* = 13.1 Hz, 1H), 3.56 – 3.51 (m, 2H), 3.42 (d, *J* = 13.1 Hz, 1H), 2.49 (s, 3H), 2.04 (brs, 1H), 2.00 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 159.0, 146.4, 140.4, 139.9, 139.0, 138.6, 137.1, 136.2, 130.5, 128.4, 128.1, 127.9, 127.3, 127.2, 126.8, 126.8, 126.5, 125.3, 122.0, 53.3, 52.4, 25.4, 20.8.

HRMS: m/z calcd for C<sub>25</sub>H<sub>25</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 353.2018, found 353.2029.

N-Benzyl-1-[3-methyl-2-(quinoxalin-5-yl)phenyl]methanamine (24)



Yellow amorphous. 52 mg, 38% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.82 (d, *J* = 1.7 Hz, 1H), 8.67 (d, *J* = 1.8 Hz, 1H), 8.15 (d, *J* = 8.5 Hz, 1H), 7.85 – 7.78 (m, 1H), 7.60 (d, *J* = 7.1 Hz, 1H), 7.54 (d, *J* = 7.6 Hz, 1H), 7.38 (t, *J* = 7.6 Hz, 1H), 7.27 (d, *J* = 7.5 Hz, 1H), 7.14 (dd, *J* = 5.0, 1.9 Hz, 3H), 6.97 (dd, *J* = 6.5, 2.9 Hz, 2H), 3.58 (d, *J* = 13.3 Hz, 1H), 3.49 (d, *J* = 13.3 Hz, 1H), 3.44 – 3.41 (m, 2H), 1.85 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 145.071, 145.0, 143.2, 141.6, 139.7, 138.7, 137.9, 137.4, 137.2, 131.3, 130.0, 129.4, 129.1, 128.4, 128.3, 128. 1, 127.1, 126.9, 52.8, 51.1, 20.8.

HRMS: m/z calcd for  $C_{23}H_{22}N_3$  [M+H]<sup>+</sup>: 340.1814, found 340.1842.

N-benzyl-1-[2-(Benzo[d]thiazol-4-yl) 3-methylbenzyl]methanamine (25)



White solid. 16 mg, 10% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.83 (s, 1H), 7.99 (d, *J* = 8.1 Hz, 1H), 7.53 – 7.47 (m, 2H), 7.38 – 7.30 (m, 2H), 7.27 (d, *J* = 7.4 Hz, 1H), 7.23 – 7.14 (m, 3H), 7.07 (d, *J* = 9.4 Hz, 2H), 3.63 (d, *J* = 13.2 Hz, 1H), 3.54 (d, *J* = 13.1 Hz, 1H), 3.51 – 3.48 (m, 2H), 1.95 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 154.1, 151.8, 138.5, 138.1, 136.8, 135.4, 134.3, 129.0, 128.3, 128.2, 128.0, 127.3, 127.0, 126.8, 126.0, 125.8, 121.3, 53.1, 51.7, 20.8.

HRMS: m/z calcd for  $C_{22}H_{21}N_2S$  [M+H]<sup>+</sup>: 345.1425, found 345.1440.

N-[2-(benzo[d]oxazol-4-yl)-3-methylbenzyl]-1-phenylmethanamine (26)



Yellow amorphous. 108 mg, 82% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.53 (dd, *J* = 8.1, 3.4 Hz, 1H), 7.44 (q, *J* = 2.0 Hz, 1H), 7.35 - 7.29 (m, 1H), 7.17 (t, *J* = 5.5 Hz, 2H), 7.15 - 7.05 (m, 3H), 7.01 (dd, *J* = 7.2, 3.4 Hz, 1H), 6.98 (t, *J* = 5.3 Hz, 2H),

6.73 (q, *J* = 2.2 Hz, 1H), 3.73 (d, *J* = 3.6 Hz, 1H), 3.43 (dd, *J* = 17.4, 3.6 Hz, 3H), 1.92 (s, 3H), 1.69 (brs, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  152.6, 145.2, 140.3, 139.0, 137.4, 135.7, 128.8, 128.5, 128.3, 128.2, 128.0, 127.7, 127.1, 126.78, 125.4, 123.8, 123.2, 120.6, 106.9, 53.1, 51.6, 20.5. HRMS: m/z calcd for C<sub>22</sub>H<sub>21</sub>N<sub>2</sub>O [M+H]<sup>+</sup>:329.1654, found 329.1668.

N-Benzyl-1-[3-methyl-2-(1-methyl-1H-benzo[a]imidazol-4-yl]phenylmethanamine (27)



Yellow amorphous. 63 mg, 10% yield.

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.58 (s, 1H), 7.51 (d, *J* = 7.5 Hz, 1H), 7.44 – 7.38 (m, 2H), 7.40 – 7.33 (m, 1H), 7.33 (d, *J* = 9.6 Hz, 2H), 7.31 – 7.25 (m, 4H), 7.10 (dd, *J* = 7.0, 1.3 Hz, 1H), 4.13 (brs, 1H), 3.94 (d, *J* = 13.0 Hz, 1H), 3.85 (s, 3H), 3.79 (d, *J* = 13.0 Hz, 1H), 3.64 (d, *J* = 5.0 Hz, 1H), 3.62 (d, *J* = 5.0 Hz, 1H), 2.00 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 144.0, 141.8, 138.4, 137.9, 134.8, 130.8, 129.2, 128.8, 128.7, 128.6, 128.1, 128.1, 127.4, 127.1, 124.1, 123.6, 109.4, 51.2, 51.0, 31.5, 21.0.

HRMS: m/z calcd for  $C_{23}H_{24}N_3$  [M+H]<sup>+</sup>: 342.1970, found 342.1971.

2-{2-[(Benzylamino)methyl]-6-methylphenyl}pyridine 2-oxide (29)



Colorless amorphous. 87 mg, 72% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.21 (d, *J* = 5.2 Hz, 1H), 7.65 – 7.52 (m, 3H), 7.50 – 7.43 (m, 2H), 7.40 – 7.30 (m, 4H), 7.24 – 7.19 (m, 2H), 4.08 (d, *J* = 13.1 Hz, 1H), 3.95 (d, *J* = 12.9 Hz, 1H), 3.80 (d, *J* = 13.1 Hz, 1H), 3.61 (d, *J* = 12.9 Hz, 1H), 2.07 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 148.1, 141.3, 137.6, 132.9, 132.6, 130.9, 130.7, 130.3, 130.2, 130.1, 129.8, 129.5, 128.9, 127.0, 50.8, 49.8, 20.4.

HRMS: m/z calcd for  $C_{20}H_{21}N_2O$  [M+H]<sup>+</sup>: 305.1654, found 305.1628.

2-{2-{[(2-Fluorobenzyl)amino]methyl}-6-methylphenyl}pyridine 2-oxide (30)



Colorless amorphous. 84 mg, 79% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.28 (dd, *J* = 6.9, 3.3 Hz, 1H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.55 (t, *J* = 7.5 Hz, 1H), 7.46 (q, *J* = 7.5 Hz, 2H), 7.38 – 7.26 (m, 4H), 7.14 (t, *J* = 6.9 Hz, 1H), 7.01 (t, *J* = 9.1 Hz, 1H), 4.15 – 4.07 (m, 1H), 4.01 – 3.92 (m, 2H), 3.60 (d, *J* = 12.8 Hz, 1H), 2.06 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  161.9, 160.3, 148.2, 141.2, 137.3, 132.5, 132.4, 132.4, 131.7 (d, *J* = 8.2 Hz), 131.1, 130.7, 130.0 (d, *J* = 2.6 Hz), 132.9, 128.6, 126.6, 125.4 (d, *J* = 3.7 Hz), 118.6 (d, *J* = 14.5 Hz), 115.8 (d, *J* = 21.4 Hz), 50.1, 44.0, 20.3.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -116.4.

HRMS: m/z calcd for  $C_{20}H_{20}N_2OF$  [M+H]<sup>+</sup>: 323.1560, found 323.1569.

2-{2-Methyl-6-{[(3-methylbenzyl)amino]methyl}phenyl}pyridine 2-oxide (31)



Yellow amorphous. 92 mg, 72% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.29 (dd, *J* = 5.2, 2.5 Hz, 1H), 7.32 (q, *J* = 7.7, 7.2 Hz, 2H), 7.28 – 7.13 (m, 4H), 7.16 – 7.06 (m, 1H), 7.04 – 6.93 (m, 3H), 3.65 (d, *J* = 13.3 Hz, 1H), 3.59 (d, *J* = 13.2 Hz, 1H), 3.52 (d, *J* = 12.8 Hz, 1H), 3.43 (d, *J* = 13.1 Hz, 1H), 2.44 (brs, 1H), 2.27 (s, 3H), 2.05 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  149.4, 140.3, 139.7, 138.5, 138.0, 137.2, 132.5, 129.6, 129.4, 129.2, 128.3, 128.2, 127.8, 127.3, 125.6, 125.4, 125.2, 53.3, 51.4, 21.5, 19.8. HRMS: m/z calcd for C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 319.1810, found 319.1805.

2-{2-{[(3-methoxybenzyl)amino]methyl}-6-methylphenyl}pyridine 2-oxide (32)



Colorless amorphous. 120 mg, 90% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.23 (d, J = 6.5 Hz, 1H), 7.62 (t, J = 7.8 Hz, 1H), 7.55 – 7.42 (m, 3H), 7.41 (d, J = 7.6 Hz, 1H), 7.30 (dd, J = 7.9, 2.0 Hz, 1H), 7.23 (d, J = 7.9 Hz, 1H), 6.89 – 6.82 (m, 2H), 6.80 (d,

*J* = 7.5 Hz, 1H), 4.02 (d, *J* = 13.0 Hz, 1H), 3.93 (d, *J* = 12.9 Hz, 1H), 3.80 (d, *J* = 8.3 Hz, 1H), 3.77 (s, 3H), 3.56 (d, *J* = 12.8 Hz, 1H), 2.04 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 160.1, 148.1, 141.3, 137.5, 132.8, 132.4, 131.6, 130.9, 130.6, 130.3, 130.0, 129.1, 126.9, 122.0, 115.3, 115.3, 55.6, 50.7, 50.0, 20.4.

HRMS: m/z calcd for  $C_{22}H_{24}NO_2$  [M+H]<sup>+</sup>: 334.1807, found 334.1815.

2-{2-{[(3-Bromobenzyl)amino]methyl}-6-methylphenyl}pyridine 2-oxide (33)



Colorless amorphous. 79 mg, 52% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.25 (d, *J* = 6.4 Hz, 1H), 7.61 (t, *J* = 7.7 Hz, 1H), 7.55 (d, *J* = 4.7 Hz, 2H), 7.52 - 7.43 (m, 2H), 7.40 - 7.30 (m, 4H), 7.25 - 7.21 (m, 2H), 4.06 (d, *J* = 13.1 Hz, 1H), 3.92 (d, *J* = 12.8 Hz, 1H), 3.80 (d, *J* = 13.2 Hz, 1H), 3.64 (d, *J* = 12.8 Hz, 1H), 2.09 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 148.0, 141.2, 137.5, 132.9, 132.8, 132.8, 132.6, 132.3, 131.1, 130.9, 130.9, 130.4, 129.9, 129.1, 128.9, 126.9, 123.2, 50.1, 50.1, 20.4.

HRMS: m/z calcd for  $C_{22}H_{24}NO_2$  [M+H]<sup>+</sup>: 334.1807, found 334.1815.

2-{2-{[(4-methoxybenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (34)



Colorless amorphous. 78 mg, 58% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.26 (d, *J* = 6.5 Hz, 1H), 7.62 – 7.56 (m, 2H), 7.52 (t, *J* = 7.7 Hz, 1H), 7.43 (dd, *J* = 11.3, 7.9 Hz, 2H), 7.30 (d, *J* = 9.9 Hz, 1H), 7.16 (d, *J* = 8.2 Hz, 2H), 6.84 (d, *J* = 8.4 Hz, 2H), 4.06 (d, *J* = 13.0 Hz, 1H), 3.96 (d, *J* = 12.9 Hz, 1 H), 3.81 (s, 3H), 3.77 (d, *J* = 12.1 Hz, 1H), 3.59 (d, *J* = 13.0 Hz, 1H), 2.07 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 160.5, 148.2, 141.6, 137.6, 132.8, 132.6, 131.6, 130.8, 130.5, 130.2, 128.8, 126.7, 122.3, 114.8, 114.3, 55.6, 50.2, 49.5, 20.5.

HRMS: m/z calcd for  $C_{21}H_{23}N_2O_2$  [M+H]<sup>+</sup>: 335.1760, found 335.1763.

2-{2-{[(4-Fluorobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (35)



Colorless amorphous. 44 mg, 34% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.44 – 8.38 (m, 1H) , 8.16 (d, *J* = 6.4 Hz, 1H), 7.56 (t, *J* = 7.8 Hz, 1H), 7.47 (d, *J* = 7.6 Hz, 1H), 7.42 – 7.37 (m, 2H), 7.33 (d, *J* = 7.8 Hz, 1H), 7.25 (d, *J* = 7.8 Hz, 1H), 7.22 – 7.15 (m, 2H), 6.91 (t, *J* = 8.4 Hz, 2H), 3.96 – 3.87 (m, 2H), 3.76 (d, *J* = 13.1 Hz, 1H), 3.50 (d, *J* = 12.7 Hz, 1H), 1.96 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  164.1, 162.1, 147.9, 141.0, 137.4, 132.7, 132.3, 132.1 (d, *J* = 8.5 Hz), 130.8 (d, *J* = 21.2 Hz), 130.2, 129.8, 129.2, 129.1, 126.8, 126.4 (d, *J* = 3.3 Hz), 116.2 (d, *J* = 21.7 Hz), 49.9, 20.3.

<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -111.0.

HRMS: m/z calcd for C<sub>20</sub>H<sub>20</sub>N<sub>2</sub>OF [M+H]<sup>+</sup>: 323.1560, found 323.1565.

2-{2-{[(4-Chlorobenzyl)amino]methyl}-6-methylphenyl}pyridine 1-oxide (36)



Colorless amorphous. 69 mg, 51% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.44 – 8.38 (m, 1H), 7.46 (d, *J* = 4.6 Hz, 2H), 7.40 (dd, *J* = 5.7, 3.1 Hz, 2H), 7.36 (d, *J* = 5.2 Hz, 2H), 7.31 (d, *J* = 8.5 Hz, 3H), 1H NMR (500 MHz, Chloroform-d)  $\delta$  8.44 – 8.38 (m, 1H), 7.46 (d, J = 4.6 Hz, 2H), 7.29 – 7.27 (m, 2H), 3.82 (d, J = 13.5 Hz, 1H), 7.27 (d, *J* = 8.2 Hz, 2H), 3.82 (d, *J* = 13.5 Hz, 1H), 3.74 (d, *J* = 13.4 Hz, 1H), 3.62 (d, *J* = 12.8 Hz, 1H), 3.57 (d, *J* = 13.0 Hz, 1H), 2.62 (brs, 1H), 2.17 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CD<sub>3</sub>OD) δ 148.1, 141.0, 137.7, 135.3, 132.7, 132.2, 131.5, 131.4, 130.8, 130.4, 130.1, 129.6, 129.2, 129.0, 126.9, 49.7, 49.6, 19.0.

HRMS: m/z calcd for C<sub>20</sub>H<sub>20</sub>N<sub>2</sub>OCI [M+H]<sup>+</sup>: 339.1246, found 339.1277.

2-{2-Methyl-6-{[(pyridin-3-ylmethyl)amino]methyl}phenyl}pyridine 2-oxide (37)



Colorless amorphous. 84 mg, 69% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.38 (d, J = 6.6 Hz, 2H), 8.27 – 8.22 (m, 1H), 7.54 (d, J = 7.5 Hz, 1H), 7.32 – 7.19 (m, 5H), 7.16 – 7.07 (m, 2H), 4.11 (brs, 1H), 3.69 (d, J = 13.5 Hz, 1H), 3.62 (d, J = 13.5 Hz, 1H), 3.51 – 3.43 (m, 2H), 2.00 (s, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 150.0, 149.2, 148.8, 140.5, 137.3, 137.0, 136.3, 134.1, 132.6, 130.0, 129.7, 128.3, 127.7, 126.3, 125.5, 123.5, 51.4, 50.2, 19.9.

HRMS: m/z calcd for  $C_{19}H_{20}N_3O$  [M+H]<sup>+</sup>:360.1606, found 360.1613.

2-{2-Methyl-6-{[(thiophen-3-ylmethyl)amino]methyl}phenyl}pyridine 1-oxide (38)



Colorless amorphous. 32 mg, 26% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.36 – 8.31 (m, 1H), 7.42 – 7.34 (m, 2H), 7.29 (dd, *J* = 5.9, 2.8 Hz, 2H), 7.25 – 7.18 (m, 3H), 7.08 – 7.04 (m, 1H), 6.97 (d, *J* = 4.9 Hz, 1H), 3.73 (d, *J* = 13.6 Hz, 1H), 3.68 (d, *J* = 13.7 Hz, 1H), 3.55 (d, *J* = 12.9 Hz, 1H), 3.48 (d, *J* = 12.9 Hz, 1H), 2.42 (brs, 1H), 2.09 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  148.2, 141.4, 137.5, 132.7, 132.5, 131.0, 130.8, 130.6, 130.4, 130.1, 128.9, 128.3, 127.6, 127.5, 126.9, 49.8, 44.9, 20.4.

HRMS: m/z calcd for  $C_{18}H_{19}N_2OS$  [M+H]<sup>+</sup>:311.1218, found 311.1210.

2-{2-Methyl-6-[(prop-2-yn-1-ylamino)methyl]phenyl}pyridine 2-oxide (39)



White solid. 31 mg, 31% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.43 (dd, *J* = 6.5, 1.3 Hz, 1H), 7.63 – 7.56 (m, 1H), 7.53 (ddd, *J* = 7.7, 6.4, 2.1 Hz, 1H), 7.42 (d, *J* = 4.6 Hz, 2H), 7.39 – 7.33 (m, 2H), 6.41 (brs, 1H), 3.75 (d, *J* = 12.6 Hz, 1H), 3.70 (d, *J* = 12.6 Hz, 1H), 3.45 (dd, *J* = 16.9, 2.5 Hz, 1H), 3.35 (dd, *J* = 17.0, 2.5 Hz, 1H), 2.27 (t, *J* = 2.4 Hz, 1H), 2.08 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 148.9, 141.3, 137.5, 133.8, 132.5, 131.4, 130.4, 129.2, 128.9, 128.8, 126.3, 75.2, 50.0, 36.5, 20.2.

HRMS: m/z calcd for C<sub>16</sub>H<sub>17</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 253.1341, found 253.1349.

2-{2-[(benzylamino)methyl]-6-chlorophenyl}pyridine 2-oxide (40)



Colorless amorphous. 48 mg, 32% yield.

<sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD)  $\delta$  8.45 (d, *J* = 6.5 Hz, 1H), 7.81 – 7.70 (m, 3H), 7.69 – 7.58 (m, 3H), 7.49 – 7.41 (m, 5H), 4.29 (d, *J* = 13.0 Hz, 1H), 4.25 (d, *J* = 13.0 Hz, 1H), 4.13 (d, *J* = 13.0 Hz, 1H), 3.74 (d, *J* = 13.1 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, CD<sub>3</sub>OD) δ 146.1, 140.9, 134.1, 133.2, 132.1, 131.9, 131.4, 131.0, 130.9, 130.4, 130.1, 129.9, 129.4, 129.1, 127.6, 50.6, 49.1.

HRMS: m/z calcd for C<sub>19</sub>H<sub>18</sub>ClN<sub>2</sub>O [M+H]<sup>+</sup>:325.1108 found 325.1129.

2-{2-[(Benzylamino)methyl]-6-methoxyphenyl}pyridine 2-oxide (41)



Colorless amorphous. 32 mg, 25% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.21 (d, *J* = 6.5 Hz, 1H), 7.62 – 7.55 (m, 2H), 7.47 – 7.41 (m, 1H), 7.39 – 7.31 (m, 4H), 7.28 (dd, *J* = 9.7, 5.0 Hz, 3H), 7.13 (d, *J* = 8.5 Hz, 1H), 4.09 (d, *J* = 13.0 Hz, 1H), 3.94 (d, *J* = 12.8 Hz, 1H), 3.85 (d, *J* = 12.9 Hz, 1H), 3.72 (s, 3H), 3.58 (d, *J* = 12.7 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 157.4, 145.8, 141.0, 132.5, 131.5, 130.7, 130.5, 130.3, 130.0, 129.8, 129.5, 126.7, 124.4, 121.4, 113.6, 56.2, 50.8, 49.6.

HRMS: m/z calcd for  $C_{20}H_{21}N_2O_2[M+H]^+:321.1603$ , found 321.1605.

2-{2-[(Benzylamino)methyl]naphthalen-1-yl}pyridine 2-oxide (42)



White solid. 87 mg, 64% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.69 (dd, J = 4.2, 1.8 Hz, 1H), 8.26 (dd, J = 8.3, 1.8 Hz, 1H), 7.99 – 7.93 (m, 2H), 7.90 (d, J = 8.2 Hz, 1H), 7.80 (d, J = 8.5 Hz, 1H), 7.69 – 7.59 (m, 2H), 7.44 – 7.35 (m, 2H),

7.23 (t, *J* = 7.7 Hz, 1H), 7.19 – 7.13 (m, 3H), 7.11 (d, *J* = 8.5 Hz, 1H), 7.01 (dd, *J* = 7.0, 2.5 Hz, 2H), 3.64 – 3.58 (m, 3H), 3.50 (d, *J* = 13.3 Hz, 1H), 2.29 (brs, 1H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  150.7, 147.6, 140.3, 138.4, 136.4, 136.3, 136.2, 133.5, 132.9, 131.8, 128.6, 128.2, 128.1, 127.9, 127.5, 126.7, 126.6, 126.3, 125.8, 125.3, 121.3, 53.4, 51.9. HRMS: m/z calcd for C<sub>23</sub>H<sub>21</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 341.1654, found 341.1658.

2-(2-((Benzylamino)methyl)-6-methylphenyl)-4-methylpyridine 1-oxide (43)



White solid. 31 mg, 24 % yield.

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.22 (d, *J* = 6.6 Hz, 1H), 7.40 – 7.32 (m, 2H), 7.30 – 7.22 (m, 5H), 7.24 – 7.18 (m, 1H), 7.08 (dd, *J* = 6.7, 2.6 Hz, 1H), 7.02 (d, *J* = 2.6 Hz, 1H), 3.76 (d, *J* = 13.3 Hz, 1H), 3.69 (d, *J* = 13.3 Hz, 1H), 3.58 (d, *J* = 13.0 Hz, 1H), 3.50 (d, *J* = 12.9 Hz, 1H), 2.35 (s, 3H), 2.10 (s, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  148.6, 139.7, 139.3, 137.9, 137.6, 137.2, 132.7, 129.6, 129.6, 128.7, 128.5, 128.5, 127.5, 127.2, 126.1, 53.1, 51.4, 20.5, 19.9.

HRMS: m/z calcd for  $C_{21}H_{23}N_2O$  [M+H]<sup>+</sup>: 319.1810, found 319.1810.

2-{2-[(Benzylamino)methyl]-6-methylphenyl}-4-chloropyridine 1-oxide (44)



White solid. 38 mg, 28 % yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.18 (d, *J* = 6.9 Hz, 1H), 7.37 – 7.29 (m, 2H), 7.27 – 7.14 (m, 8H), 3.69 (d, *J* = 13.3 Hz, 1H), 3.64 (d, *J* = 13.3 Hz, 1H), 3.54 (d, *J* = 12.8 Hz, 1H), 3.42 (d, *J* = 12.7 Hz, 1H), 2.17 (brs, 1H), 2.08 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 150.3, 140.7, 139.8, 138.6, 137.2, 131.5, 131.3, 129.9, 129.4, 128.4, 128.3, 128.1, 127.4, 127.0, 125.4, 53.5, 51.5, 19.7.

HRMS: m/z calcd for C<sub>20</sub>H<sub>20</sub>CIN<sub>2</sub>O [M+H]<sup>+</sup>:339.1264, found 339.1271.

2-{2-[(benzylamino)methyl]-6-methylphenyl}-5-fluoropyridine 1-oxide (45)



White solid. 33 mg, 26 % yield.

<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  8.54 (dd, *J* = 4.2, 2.3 Hz, 1H), 7.65 – 7.59 (m, 1H), 7.57 – 7.49 (m, 4H), 7.51 – 7.41 (m, 5H), 4.29 (d, *J* = 13.0 Hz, 1H), 4.15 (d, *J* = 8.7 Hz, 1H), 4.12 (d, *J* = 8.9 Hz, 1H), 3.71 (d, *J* = 12.9 Hz, 1H), 2.08 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CD<sub>3</sub>OD) δ 160.9, 159.2, 144.8, 138.1, 132.2, 132.0, 131.3 (d, J = 36.5 Hz), 130.9, 130.4, 130.0, 129.5 (d, J = 9.1 Hz), 129.3, 129.1, 129.0, 118.4 (d, J = 19.8 Hz), 50.5, 49.0, 18.8. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -119.1.

HRMS: m/z calcd for  $C_{20}H_{20}N_2OF$  [M+H]<sup>+</sup>: 323.1560, found 323.1534.

2-{2-[(Benzylamino)methyl]-6-methylphenyl}-5-methylpyridine 2-oxide (46)



White solid. 89 mg, 70% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.14 (s, 1H), 7.44 (d, *J* = 6.4 Hz, 1H), 7.38 (t, *J* = 7.6 Hz, 1H), 7.33 – 7.26 (m, 3H), 7.25 – 7.21 (m, 3H), 7.17 – 7.14 (m, 1H), 7.10 (d, *J* = 7.9 Hz, 1H), 5.14 (brs, 1H), 3.85 (d, *J* = 13.3 Hz, 1H), 3.72 (d, *J* = 13.3 Hz, 1H), 3.59 (d, *J* = 2.6 Hz, 2H), 2.35 (s, 3H), 2.07 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 146.0, 140.5, 137.3, 136.5, 136.3, 132.6, 130.2, 129.7, 128.9, 128.6, 128.2, 128.1, 127.6, 127.5, 52.5, 50.9, 20.0, 18.3.

HRMS: m/z calcd for  $C_{21}H_{23}N_2O$  [M+H]<sup>+</sup>: 319.1810, found 319.1810.

2-{2-[(Benzylamino)methyl]-6-methylphenyl}-5-(trifluoromethyl)pyridine 2-oxide (47)


Colorless amorphous. 65 mg, 44% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.44 (s, 1H), 7.58 – 7.51 (m, 2H), 7.47 (t, *J* = 7.7 Hz, 1H), 7.39 – 7.33 (m, 2H), 7.32 – 7.26 (m, 3H), 7.24 – 7.22 (m, 1H), 3.98 (d, *J* = 13.2 Hz, 1H), 3.79 (d, *J* = 4.5 Hz, 1H), 3.76 (d, *J* = 4.0 Hz, 1H), 3.54 (d, *J* = 12.8 Hz, 1H), 2.07 (s, 3H).

<sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  154.3, 145.4, 138.3, 138.2, 124.8 (q, *J* = 3.9 Hz), 136.2, 130.4, 128.4 (2), 128.0, 127.2 (q, *J* = 31.5 Hz), 126.9, 126.6, 125.9, 124.8 (q, *J* = 3.9 Hz), 124.2 (q, *J* = 271.8 Hz), 119.1, 116.1, 51.7, 50.5, 20.2.

<sup>19</sup>F NMR (565 MHz, CD<sub>3</sub>OD) δ -64.4.

HRMS: m/z calcd for  $C_{21}H_{20}F_3N_2O$  [M+H]<sup>+</sup>: 373.1528, found 373.1536.

2-{2-[(Benzylamino)methyl]-6-methylphenyl}-6-methylpyridine 2-oxide (48)



White solid. 63 mg, 61% yield.

<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  7.69 (d, *J* = 5.0 Hz, 2H), 7.56 – 7.51 (m, 3H), 7.47 – 7.39 (m, 5H), 4.24 – 4.10 (m, 3H), 3.79 (d, *J* = 12.9 Hz, 1H), 2.60 (s, 3H), 2.08 (s, 3H).

<sup>13</sup>C NMR (151 MHz, CD<sub>3</sub>OD) δ 151.9, 148.7, 138.2, 134.0, 132.7, 131.9, 131.1, 131.0, 130.6, 129.8, 129.5, 129.5, 128.1, 127.6, 50.8, 50.5, 19.5, 17.7.

HRMS: m/z calcd for C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 319.1810, found 319.1815.

1-{2-[(Benzylamino)methyl]phenyl}isoquinoline 1-oxide (49)



White solid. 63 mg, 46% yield.

<sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD)  $\delta$  8.38 (d, *J* = 7.2 Hz, 1H), 8.15 (dd, *J* = 13.5, 7.7 Hz, 2H), 7.89 – 7.84 (m, 2H), 7.80 – 7.65 (m, 3H), 7.41 (d, *J* = 8.1 Hz, 2H), 7.28 – 7.24 (m, 3H), 7.17 – 7.12 (m, 2H), 3.78 (d, *J* = 13.2 Hz, 1H), 3.63 (d, *J* = 13.2 Hz, 1H), 3.59 (d, *J* = 12.8 Hz, 1H), 3.56 (d, *J* = 12.8 Hz, 1H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  147.0, 136.5, 133.5, 131.5, 131.2, 131.1, 131.1,131.0, 130.9, 130.8, 130.5, 130.4, 130.2, 129.7, 129.5, 129.1, 127.5, 126.2, 125.3, 50.8, 49.8. HRMS: m/z calcd for C<sub>23</sub>H<sub>21</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 341.1654, found 341.1671. 2-{2-[(Benzylamino)methyl]-6-methylphenyl}-3-methylpyridine 1-oxide (50)



White solid. 66 mg, 52% yield.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.18 (d, *J* = 6.3 Hz, 1H), 7.43 (d, *J* = 7.7 Hz, 1H), 7.38 (t, *J* = 7.4 Hz, 1H), 7.29 (d, *J* = 19.3 Hz, 4H), 7.25 - 7.16 (m, 4H), 3.86 (d, *J* = 13.3 Hz, 1H), 3.73 (d, *J* = 13.2 Hz, 1H), 3.63 (d, *J* = 13.0 Hz, 1H), 3.52 (d, *J* = 12.9 Hz, 1H), 1.99 (s, 3H), 1.92 (s, 3H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 148.9, 138.3, 136.9, 136.7, 134.9, 131.8, 130.8, 129.8, 129.1 (2), 128.8, 128.7, 127.9, 124.8, 115.0, 52.1, 50.9, 19.4, 19.0.

HRMS: m/z calcd for C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O [M+H]<sup>+</sup>: 319.1810, found 319.1816.

### **Evaluation of Barriers to Rotation**

The racemization study for atropisomer (*S*)-7 (6 mg/ mL) was performed at 100 °C in toluene heated with an oil bath. Aliquots were taken at different time intervals and the decay in enantiomeric excess (*ee*) was monitored by chiral HPLC analysis. The slope of the first-order kinetic line (plot of ln(*ee*) vs time) gives the racemization constant ( $k_{rac} = 2 \times k_{enant}$ ). Eyring equation gives the enantiomerization barrier from the enantiomerization constant ( $k_{enant}$ ).

 $k_{rac} = 2.30 \times 10^{-7} \text{ s}^{-1} (\text{at } 100 \text{ °C});$   $k_{enant} = k_{rac}/2 = 1.15 \times 10^{-7} \text{ s}^{-1} (\text{at } 100 \text{ °C});$   $\Delta G^{\neq}_{enant} = 141.7 \text{ kJ/mol} = 33.8 \text{ kcal/mol} (\text{at } 100 \text{ °C});$ Half-life time  $t_{1/2} = \ln 2/k_{rac} = 3013683 \text{ s} = 837.1 \text{ h} (\text{at } 100 \text{ °C}).$ 

	Time (s)	ee (%)
	2820	96.4
	7860	96.5
N N	22440	95.9
	71220	94.8
(S)- <b>7</b>	93720	94.3
Solvent: Toluene	172260	92.8
Temperature: 100 °C	256080	91.1
	341460	89.1
	363840	88.7



Fig. S9. The plot of  $ln(ee) \vee T$  (s) for the decay in ee of compound (S)-7.

The racemization study for atropisomer (S)-**29** (6 mg/ mL) was performed at 100  $^{\circ}$ C in toluene heated with an oil bath. Aliquots were taken at different time intervals and the decay in enantiomeric excess (*ee*) was monitored by chiral HPLC analysis.

 $k_{rac} = 5.01 \times 10^{-6} \text{ s}^{-1} (\text{at } 100 \text{ °C});$   $k_{enant} = k_{rac}/2 = 2.51 \times 10^{-6} \text{ s}^{-1} (\text{at } 100 \text{ °C});$   $\Delta G^{\neq}_{enant} = 132.1 \text{ kJ/mol} = 31.6 \text{ kcal/mol} (\text{at } 100 \text{ °C});$ Half-life time  $t_{1/2} = \ln 2/k_{rac} = 138353 \text{ s} = 38.4 \text{ h} (\text{at } 100 \text{ °C}).$ 

	Time (s)	ee (%)
	0	100.0
	3240	96.0
$\operatorname{Me}^{r}$ $\operatorname{He}^{r}$ $\operatorname{He}^{r}$ $\operatorname{He}^{r}$ $\operatorname{He}^{r}$ $\operatorname{He}^{r}$	7500	93.9
	11220	92.3
(5)-29	14460	91.3
(3)-23	22980	86.4
Solvent: Toluene	52680	77.6
	67080	70.0
	89580	62.1



Fig. S10. The plot of ln(ee) v T (s) for the decay in ee of compound (S)-29.

### X-ray Crystallography

Experiment details. Low-temperature X-ray diffraction data was collected on a Rigaku XtaLAB Synergy diffractometer coupled to a Rigaku Hypix detector with Cu K $\alpha$  radiation ( $\lambda$  = 1.54184 Å), from a PhotonJet micro-focus X-ray source at 173 K. The diffraction images were processed and scaled using the CrysAlisPro software. Using Olex2, the structure was solved with the SHELXT structure solution program using Direct Methods and refined with the SHELXL. The residual values of the refinements are listed in Table S8-11. Crystallographic data for the structures reported in this Article have been deposited at the Cambridge Crystallographic Data Centre, under deposition numbers 2381733 (**29**), 2381730 (**45**), 2381732 (**48**), 2381731 (**53**). Copies of these data can be obtained free of charge via https://www.ccdc.cam.ac.uk/structures/.



Identification code hxy\_3 Empirical formula C<sub>26</sub>H<sub>24</sub>N<sub>2</sub>O 380.47 Formula weight Temperature/K 100.02(10) Crystal system orthorhombic P212121 Space group a/Å 9.50530 (10) b/Å 11.21780 (10) c/Å 18.29070 (10) α/° 90 β/° 90 γ/° 90 Volume/Å 1950.31 (3) Ζ 4 pcalcg/cm<sup>3</sup> 1.296 µ/mm-1 0.616 F(000) 808.0 Crystal size/mm<sup>3</sup> 0.16 × 0.14 × 0.13 Radiation Cu K $\alpha$  ( $\lambda$  = 1.54184) 2O range for data collection/° 9.248 to 155.172  $-11 \le h \le 12, -13 \le k \le 14, -22 \le l \le 23$ Index ranges 30036 **Reflections collected** 4013 [R<sub>int</sub> = 0.0215, R<sub>sigma</sub> = 0.0101] Independent reflections 4013/0/265 Data/restraints/parameters Goodness-of-fit on F2 1.063 Final R indexes  $[I \ge 2\sigma (I)]$  $R_1 = 0.0249$ ,  $wR_2 = 0.0652$ Final R indexes [all data] R<sub>1</sub> = 0.0250, wR<sub>2</sub> = 0.0652 Largest diff. peak/hole / e Å-3 0.18/-0.14 Flack parameter 0.04 (4)

Table S8. Crystal data and structure refinement for 53 (CCDC: 2381731).



Identification code hxy-5\_auto Empirical formula  $C_{20}H_{22}CI_2N_2O$ Formula weight 377.29 Temperature/K 295.24 (10) orthorhombic Crystal system P212121 Space group a/Å 7.8488 (3) b/Å 15.7366 (5) c/Å 15.8854 (6) α/° 90 β/° 90 γ/° 90 Volume/Å 1962.06 (12) Ζ 4 pcalcg/cm<sup>3</sup> 1.277 µ/mm-1 3.046 F(000) 792.0 Crystal size/mm3 0.14 × 0.1 × 0.08 Cu K $\alpha$  ( $\lambda$  = 1.54184) Radiation 2O range for data collection/° 7.908 to 150.468  $-7 \le h \le 9, -17 \le k \le 19, -19 \le l \le 18$ Index ranges **Reflections collected** 10412 Independent reflections 3909 [Rint = 0.0352, Rsigma = 0.0405] Data/restraints/parameters 3909/0/231 Goodness-of-fit on F2 1.063 R1 = 0.0573, wR2 = 0.1673 Final R indexes  $[I \ge 2\sigma (I)]$ Final R indexes [all data] R1 = 0.0653, wR2 = 0.1754 Largest diff. peak/hole / e Å-3 0.83/-0.57 Flack parameter -0.022 (9)

Table S9. Crystal data and structure refinement for (S)-29 HCl salt (CCDC: 2381733).





Identification code	HXY_2_autored	
Empirical formula	$C_{21}H_{24}CI_2N_2O$	
Formula weight	391.32	
Temperature/K	100.00 (10)	
Crystal system	orthorhombic	
Space group	P212121	
a/Å	7.90220 (10)	
b/Å	15.1936 (2)	
c/Å	16.9727 (2)	
α/°	90	
β/°	90	
γ/°	90	
Volume/Å	2037.79 (4)	
Z	4	
ρcalcg/cm <sup>3</sup>	1.276	
µ/mm-1	2.951	
F(000)	824.0	
Crystal size/mm3	0.13 × 0.09 × 0.05	
Radiation	Cu Kα (λ = 1.54184)	
2O range for data collection/°	7.81 to 155.222	
Index ranges	-9 ≤ h ≤ 9, -19 ≤ k ≤ 19, -21 ≤ l ≤ 15	
Reflections collected	27542	
Independent reflections	4146 [ $R_{int} = 0.0373$ , $R_{sigma} = 0.0175$ ]	
Data/restraints/parameters	4146/0/238	
Goodness-of-fit on F2	1.061	
Final R indexes [I>=2σ (I)]	R <sub>1</sub> = 0.0289, wR <sub>2</sub> = 0.0803	
Final R indexes [all data]	R <sub>1</sub> = 0.0292, wR <sub>2</sub> = 0.0806	
Largest diff. peak/hole / e Å <sup>-3</sup>	0.38/-0.29	
Flack parameter	-0.002 (4)	

Table S10. Crystal data and structure refinement for (S)-45 HCl salt (CCDC: 2381730).



Empirical formula $C_{20}H_{20.5}CIFN_2O_{1.25}$ Formula weight         363.33           Temperature/K         100.00 (10)           Crystal system         monoclinic           Space group         P21           a/A         10.2259 (2)           b/A         19.4327 (3)           c/A         10.2890 (3)           a/ <sup>o</sup> 90 $\beta'^o$ 117.079 (3) $\gamma'^o$ 90           Volume/A         1820.47 (8)           Z         4           pcalcg/cm <sup>3</sup> 1.326 $\mu/mm^{-1}$ 2.038           F(000)         762.0           Crystal size/mm3         0.1 × 0.07 × 0.05           Radiation         Cu Ka ( $\lambda = 1.54184$ )           20 range for data collection/°         9.102 to 149.978           Index ranges         -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12           Reflections collected         17287           Independent reflections         6613 [Rint = 0.0377, Rsigma = 0.0410]           Data/restraints/parameters         6613/1/488           Goodness-of-fit on F2         1.030           Final R indexes [al data]         R1 = 0.0372, wR2 = 0.0974           Final R indexes [al d	Identification code	hxy6-7	
Formula weight       363.33         Temperature/K       100.00 (10)         Crystal system       monoclinic         Space group       P2,         a/A       10.2259 (2)         b/A       19.4327 (3)         c/A       10.2890 (3)         a/°       90 $\beta''$ 117.079 (3) $\gamma''$ 90 $\gamma''$ 90         Volume/A       1820.47 (8)         Z       4         pcalcg/cm³       1.326         µ/mm-1       2.038         F(000)       762.0         Crystal size/mm3       0.1 × 0.07 × 0.05         Radiation       Cu Ka ( $k = 1.54184$ )         20 range for data collection/°       9.102 to 149.978         Index ranges       -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12         Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       6613/1/488         Goodness-of-fit on F2       1.030         Final R indexes [I>=20 (I)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [I=data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å <sup>3</sup> 0.37/-0.27 <td>Empirical formula</td> <td colspan="2">C<sub>20</sub>H<sub>20.5</sub>CIFN<sub>2</sub>O<sub>1.25</sub></td>	Empirical formula	C <sub>20</sub> H <sub>20.5</sub> CIFN <sub>2</sub> O <sub>1.25</sub>	
Temperature/K       100.00 (10)         Crystal system       monoclinic         Space group $P2_1$ a/A       10.2259 (2)         b/A       19.4327 (3)         c/A       10.2890 (3)         a/°       90 $\beta$ /°       117.079 (3) $\gamma$ /°       90         Volume/Å       1820.47 (8)         Z       4         pcalcg/cm³       1.326 $\mu/mn-1$ 2.038         F(000)       762.0         Crystal size/mm3       0.1 × 0.07 × 0.05         Radiation       Cu Kar ( $\lambda$ = 1.54184)         20 range for data collection/°       9.102 to 149.978         Index ranges       -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12         Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       6613/1/488         Goodness-of-fit on F2       1.030         Final R indexes [I > 2 $\sigma$ (I)]       R1 = 0.0372, wR2 = 0.0987         Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27         Flack parameter       -0.001 (7)	Formula weight	363.33	
Crystal system         monoclinic           Space group         P21           a/Å         10.2259 (2)           b/Å         19.4327 (3)           c/Å         10.2890 (3)           a/°         90           β/°         117.079 (3)           γ/°         90           Volume/Å         1820.47 (8)           Z         4           pcalcg/cm³         1.326           µ/mm-1         2.038           F(000)         762.0           Crystal size/mm3         0.1 × 0.07 × 0.05           Radiation         Cu Ka (λ = 1.54184)           20 range for data collection/°         9.102 to 149.978           Index ranges         -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12           Reflections collected         17287           Independent reflections         6613 [Rint = 0.0377, Rsigma = 0.0410]           Data/restraints/parameters         6613/1/488           Goodness-of-fit on F2         1.030           Final R indexes [I=2σ (I)]         R1 = 0.0372, wR2 = 0.0987           Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27           Flack parameter         -0.001 (7)	Temperature/K	100.00 (10)	
Space group $P_{1}$ $a/A$ $10.2259 (2)$ $b/A$ $19.4327 (3)$ $c/A$ $10.2890 (3)$ $a'^{\circ}$ $90$ $\beta'^{\circ}$ $117.079 (3)$ $\gamma'^{\circ}$ $90$ Volume/Å $1820.47 (8)$ $Z$ $4$ pcalcg/cm <sup>3</sup> $1.326$ $\mu/mm-1$ $2.038$ F(000) $762.0$ Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ Radiation       Cu K $\alpha$ ( $\lambda = 1.54184$ )         20 range for data collection/° $9.102$ to $149.978$ Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected $17287$ Independent reflections $6613$ [Rint = $0.0377$ , Rigma = $0.0410$ ]         Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [all data]       R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]       R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å <sup>3</sup> $0.37/-0.27$ Flack parameter $-0.001 (7)$	Crystal system	monoclinic	
$a/A$ 10.2259 (2) $b/A$ 19.4327 (3) $c/A$ 10.2890 (3) $a'^{\circ}$ 90 $\beta'^{\circ}$ 117.079 (3) $\gamma'^{\circ}$ 90Volume/A1820.47 (8)Z4pcalcg/cm <sup>3</sup> 1.326 $\mu/mm-1$ 2.038F(000)762.0Crystal size/mm30.1 × 0.07 × 0.05RadiationCu K $\alpha$ ( $k = 1.54184$ )20 range for data collection/°9.102 to 149.978Index ranges-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [l>=2 $\sigma$ (l)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27Flack parameter-0.001 (7)	Space group	P21	
b/Å19.4327 (3)c/Å10.2890 (3) $a'^{\circ}$ 90 $\beta'^{\circ}$ 117.079 (3) $\gamma'^{\circ}$ 90Volume/Å1820.47 (8)Z4pcalcg/cm³1.326 $\mu$ /mm-12.038F(000)762.0Crystal size/mm30.1 × 0.07 × 0.05RadiationCu Ka ( $\lambda$ = 1.54184)20 range for data collection/°9.102 to 149.978Index ranges-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [l>=2 $\sigma$ (1)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-³0.37/-0.27Flack parameter-0.001 (7)	a/Å	10.2259 (2)	
$c/Å$ 10.2890 (3) $a''$ 90 $\beta''$ 117.079 (3) $\gamma''$ 90Volume/Å1820.47 (8)Z4 $pcalcg/cm^3$ 1.326 $\mu/mm^-1$ 2.038F(000)762.0Crystal size/mm30.1 × 0.07 × 0.05RadiationCu Ka ( $\lambda$ = 1.54184)20 range for data collection/°9.102 to 149.978Index ranges-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [l>=2\sigma(l)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	b/Å	19.4327 (3)	
$a'^{\circ}$ 90 $\beta/^{\circ}$ 117.079 (3) $\gamma'^{\circ}$ 90Volume/A1820.47 (8)Z4pcalcg/cm <sup>3</sup> 1.326 $\mu/mm^{-1}$ 2.038F(000)762.0Crystal size/mm30.1 × 0.07 × 0.05RadiationCu Ka ( $\lambda$ = 1.54184)2Θ range for data collection/°9.102 to 149.978Index ranges-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [I)=2σ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27Flack parameter-0.001 (7)	c/Å	10.2890 (3)	
$\beta/^{\circ}$ 117.079 (3) $\gamma/^{\circ}$ 90Volume/Å1820.47 (8)Z4pcalcg/cm31.326 $\mu/mm-1$ 2.038F(000)762.0Crystal size/mm30.1 × 0.07 × 0.05RadiationCu Ka ( $\lambda$ = 1.54184)20 range for data collection/°9.102 to 149.978Index ranges-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [I>=2σ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	α/°	90	
$\gamma/^{\circ}$ 90         Volume/Å       1820.47 (8)         Z       4         pcalcg/cm <sup>3</sup> 1.326 $\mu/mm \cdot 1$ 2.038         F(000)       762.0         Crystal size/mm3       0.1 × 0.07 × 0.05         Radiation       Cu Ka ( $\lambda$ = 1.54184)         20 range for data collection/°       9.102 to 149.978         Index ranges       -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12         Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       66131/1/488         Goodness-of-fit on F2       1.030         Final R indexes [I>=2 $\sigma$ (I)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27         Flack parameter       -0.001 (7)	β/°	117.079 (3)	
Volume/Å $1820.47 (8)$ Z4pcalcg/cm³ $1.326$ µ/mm-1 $2.038$ F(000) $762.0$ Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ RadiationCu Ka ( $\lambda = 1.54184$ )20 range for data collection/° $9.102$ to $149.978$ Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected $17287$ Independent reflections $6613$ [Rint = $0.0377$ , Rsigma = $0.0410$ ]Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [l>= $2\sigma$ (l)]R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å-³ $0.37/-0.27$ Flack parameter $-0.001$ (7)	γ/°	90	
Z4 $pcalcg/cm^3$ 1.326 $\mu/mm-1$ 2.038 $F(000)$ 762.0Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ RadiationCu Ka ( $\lambda = 1.54184$ )20 range for data collection/°9.102 to 149.978Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [I>=2 $\sigma$ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	Volume/Å	1820.47 (8)	
pcalcg/cm³1.326 $\mu/mm - 1$ 2.038F(000)762.0Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ RadiationCu Ka ( $\lambda = 1.54184$ )20 range for data collection/°9.102 to 149.978Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [I >=2 $\sigma$ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-³0.37/-0.27Flack parameter-0.001 (7)	Z	4	
$\mu$ /mm-12.038F(000)762.0Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ RadiationCu Ka ( $\lambda = 1.54184$ )20 range for data collection/° $9.102$ to 149.978Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [I>=2 $\sigma$ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	pcalcg/cm <sup>3</sup>	1.326	
F(000)762.0Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ RadiationCu Ka ( $\lambda = 1.54184$ )20 range for data collection/° $9.102$ to 149.978Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected $17287$ Independent reflections $6613$ [Rint = $0.0377$ , Rsigma = $0.0410$ ]Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [l>= $2\sigma$ (l)]R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å-3 $0.37/-0.27$ Flack parameter $-0.001$ (7)	µ/mm-1	2.038	
Crystal size/mm3 $0.1 \times 0.07 \times 0.05$ Radiation       Cu Ka ( $\lambda = 1.54184$ )         20 range for data collection/° $9.102$ to 149.978         Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       6613/1/488         Goodness-of-fit on F2       1.030         Final R indexes [l>=2 $\sigma$ (l)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27         Flack parameter       -0.001 (7)	F(000)	762.0	
Radiation       Cu K $\alpha$ ( $\lambda$ = 1.54184)         20 range for data collection/°       9.102 to 149.978         Index ranges       -12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12         Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       6613/1/488         Goodness-of-fit on F2       1.030         Final R indexes [l>=2 $\sigma$ (l)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å-3       0.37/-0.27         Flack parameter       -0.001 (7)	Crystal size/mm3	0.1 × 0.07 × 0.05	
$2\Theta$ range for data collection/° $9.102$ to $149.978$ Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected $17287$ Independent reflections $6613$ [Rint = $0.0377$ , Rsigma = $0.0410$ ]Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [l>= $2\sigma$ (l)]R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å-3 $0.37/-0.27$ Flack parameter $-0.001$ (7)	Radiation	Cu Kα (λ = 1.54184)	
Index ranges $-12 \le h \le 7, -24 \le k \le 23, -12 \le l \le 12$ Reflections collected17287Independent reflections6613 [Rint = 0.0377, Rsigma = 0.0410]Data/restraints/parameters6613/1/488Goodness-of-fit on F21.030Final R indexes [l>=2 $\sigma$ (l)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	2⊖ range for data collection/°	9.102 to 149.978	
Reflections collected       17287         Independent reflections       6613 [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters       6613/1/488         Goodness-of-fit on F2       1.030         Final R indexes [I>= $2\sigma$ (I)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å-3       0.37/-0.27         Flack parameter       -0.001 (7)	Index ranges	-12 ≤ h ≤ 7, -24 ≤ k ≤ 23, -12 ≤ l ≤ 12	
Independent reflections $6613$ [Rint = 0.0377, Rsigma = 0.0410]         Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [I>= $2\sigma$ (I)]       R1 = 0.0372, wR2 = 0.0974         Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å-3 $0.37/-0.27$ Flack parameter $-0.001$ (7)	Reflections collected	17287	
Data/restraints/parameters $6613/1/488$ Goodness-of-fit on F2 $1.030$ Final R indexes [I>= $2\sigma$ (I)]       R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]       R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å <sup>-3</sup> $0.37/-0.27$ Flack parameter $-0.001$ (7)	Independent reflections	6613 [Rint = 0.0377, Rsigma = 0.0410]	
Goodness-of-fit on F2 $1.030$ Final R indexes [I>= $2\sigma$ (I)]       R1 = $0.0372$ , wR2 = $0.0974$ Final R indexes [all data]       R1 = $0.0391$ , wR2 = $0.0987$ Largest diff. peak/hole / e Å-3 $0.37/-0.27$ Flack parameter $-0.001$ (7)	Data/restraints/parameters	6613/1/488	
Final R indexes [I>= $2\sigma$ (I)]R1 = 0.0372, wR2 = 0.0974Final R indexes [all data]R1 = 0.0391, wR2 = 0.0987Largest diff. peak/hole / e Å-30.37/-0.27Flack parameter-0.001 (7)	Goodness-of-fit on F2	1.030	
Final R indexes [all data]       R1 = 0.0391, wR2 = 0.0987         Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27         Flack parameter       -0.001 (7)	Final R indexes [I>=2σ (I)]	R1 = 0.0372, wR2 = 0.0974	
Largest diff. peak/hole / e Å <sup>-3</sup> 0.37/-0.27           Flack parameter         -0.001 (7)	Final R indexes [all data]	R1 = 0.0391, wR2 = 0.0987	
Flack parameter   -0.001 (7)	Largest diff. peak/hole / e Å <sup>-3</sup>	0.37/-0.27	
	Flack parameter	-0.001 (7)	

Table 11. Crystal data and structure refinement for (S)-48 HCl salt (CCDC: 2381732).

## 3. NMR spectra





# 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.45 8.27 8.23 8.23 8.23 8.23 8.23 8.25 8.25 8.25 8.25 8.25 8.25 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 8.27 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.45 9.47 9.47 9.49 9.47 9.49 9.49 9.49 9.49 9.49 9.45 <l



## (1) 200 (200) (2) 200 (200)



### -9.22 -9.22 -9.22 -9.24









10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

### 







110 100 fl (ppm) 170 160 

### 9.50 8.890 8.890 8.890 8.893 8.893 8.893 8.893 7.173 7.755 7.775 7.7555 7.7555 7.7555 7.7555 7.7555 7.7555 7.7555 7.7555 7.



— 1.99









## 



— 1.98







110 100 f1 (ppm) 

### - 9.60 8.09 8.09 7.59 7.59 7.51 7.55



- 2.11



## 9.7 8.8 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 8.9 9.9 9.9 1.12 1.12 1.12 1.12 1.12 1.12 1.12 1.12 1.12 1.12









## 





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm)



f1 (ppm)

## $\overset{9.79}{\underset{9.79}{\times}}$

- 2.19



### 

- 1.75







10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 f1 (ppm)



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -3 f1 (ppm)

- 2.18 -- 9.81 сно Me ,ō 1.01<del>1</del> 3.16<del>1</del> 1.00<del>1</del> 1.03 2.04 1.05 ₹ 1.02 4 5.5 5.0 f1 (ppm) 10.0 10.5 9.5 9, 0 8.5 8.0 7.5 7.0 6.5 6.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0  $\begin{array}{c} 151.05\\ 137.71\\ 137.67\\ 137.67\\ 137.67\\ 137.67\\ 138.66\\ 138.67\\ 130.43\\ 130.43\\ 130.43\\ 130.43\\ 130.43\\ 1129.24\\ 1129.24\\ 1129.24\\ 1120.36\\ 1120.36\\ 1120.38\\$ - 190.93 — 19.18 `<u>с</u>но +\_0 Me  $CF_{2}$ 200 110 100 fl (ppm) 190 180 170 160 150 140 130 120 90 80 70 60 50 40 30 20 10



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 11 (ppm)









### 9,73 9,73 9,73 9,73 9,73 9,12 9,12 9,12 9,12 9,12 9,12 9,12 1,12






f1 (ppm) 





10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

#### 8.80 8.79 8.79 8.79 8.79 8.79 8.79 8.79 8.79 8.79 8.79 8.73 8.73 8.73 8.73 8.73 8.73 8.75 8.75 8.75 8.75 8.75 7.75









## 8.45 8.23 8.23 8.23 8.23 8.23 8.23 8.23 1.12 </tr









10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)



# $\begin{array}{c} 8.83 \\ 8.21 \\ 7.156 \\ 7.756 \\ 7.756 \\ 7.756 \\ 7.756 \\ 7.737 \\ 7.733 \\$







 $\frac{1}{70}$ f1 (ppm)



#### 8.77 8.77 8.77 8.77 8.77 8.77 8.77 8.77 8.77 8.73 8.77 8.73 8.77 8.73 8.74 8.73 8.74 7.799 8.74 7.799 8.74 7.799 8.77 7.799 8.77 7.799 7.755 7.



100 f1 (ppm) 200 190 180 170 160 150 140 130 90 80  $\frac{1}{70}$ 60 50 40 30 20 10 0 120 110



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)





#### 8.8.95 8.93 8.93 8.93 8.93 8.93 8.93 8.875 8.875 8.877 8.



## 8.15 8.15 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.887 7.557 7.557 7.555 <tr





- 8.83 - 8.80 8.00 7.598 7.598 7.598 7.598 7.502 7.512 7.512 7.512 7.512 7.512 7.512 7.512 7.512 7.512 7.512 7.521 7.521 7.521 7.521 7.522 7.523 7.522 7.523 7.522 7.523 7.522 7.523 7.523 7.522 7.522 7.522 7.522 7.723 7.728 7









#### 8.22 8.25 7.75 7.62 7.75 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73 7.73



#### 8.29 8.29 8.28 8.28 8.28 8.28 8.28 8.28 8.28 8.28 8.28 8.28 8.28 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.29 8.20 <li









10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 F1 (ppm)





230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -3 f1 (ppm)

## 8.26 8.24 8.24 8.25 8.25 8.26 9.27 1.12 <t















10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

## $\begin{array}{c} 8.42\\ 8.44\\ 8.44\\ 7.46\\ 7.35\\ 7.33\\ 7.12$






## 78.44 78.45 78





f1 (ppm) 

#### 8.21 8.21 7.55 7.56 7.56 7.55 7.75 7.55 7.75





## 8.2 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8.3 8







- 2.08



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

# 8.14 7.45 7.72 7.74 7.75 </tr









10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

## 7.55 7.568 7.55 7.568 7.55 7.568 7.55 7.568 7.55 7.568 7.57 7.458 7.545 7.445 7.445 7.441 7.441 7.441 7.441 7.441 7.4114 7.411 7.4114 7.411 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 7.4114 7.4116 <tr/t 1.416</td> 7.4114</td





190 180 170 160 f1 (ppm) 















## $\begin{array}{c} & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & \\$



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