# Supporting Information for

# Rhodium-catalyzed one-pot tandem reductive amination/asymmetric transfer hydrogenation of quinoxaline-2-carbaldehydes and anilines for the efficient synthesis of chiral vicinal diamines

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# **1. General information**

Unless otherwise noted, all experiments were carried out under nitrogen atmosphere and all commercially available chemicals including organic solvents were used as received from Aldrich, Acros or Strem without further purification. <sup>1</sup>H NMR and <sup>13</sup>C{<sup>1</sup>H} NMR spectra were recorded on a Bruker Model Advance DMX 400 Spectrometer (<sup>1</sup>H 400 MHz and <sup>13</sup>C{<sup>1</sup>H} 101 MHz, respectively), Bruker Model Advance DMX 500 Spectrometer (<sup>1</sup>H 500 MHz and <sup>13</sup>C{<sup>1</sup>H} 126 MHz, respectively) or Bruker Model Advance DMX 600 Spectrometer (<sup>1</sup>H 600 MHz and <sup>13</sup>C{<sup>1</sup>H} 151 MHz, respectively). Chemical shifts ( $\delta$ ) are given in ppm and are referenced to residual solvent peaks. Melting points were measured on X-4 melting point apparatus and are uncorrected. High resolution mass spectra (HRMS) were performed on a VG Autospec-3000 spectrometer. Column chromatography was performed with silica gel (200-300 mesh). All the quinoxaline-2-carbaldehyde substrates were prepared according to the literature methods.<sup>1</sup>

# 2. Optimization of reaction conditions



Table S1. Screening of catalysts for the RA/ATH of 1a and 2a.<sup>a</sup>

| Entry | Catalyst                            | $3a/4a/5a (\%)^b$ | ee of <b>3a</b> (%) <sup><i>c</i></sup> |
|-------|-------------------------------------|-------------------|---|
| 1     | ( <i>R</i> , <i>R</i> )-C1          | 91:9:             | 54                                      |
| 2     | ( <i>R</i> , <i>R</i> )- <b>C2</b>  | 76:24:            | 39                                      |
| 3     | ( <i>R</i> , <i>R</i> )-C3          | 93:7:             | 90                                      |
| 4     | ( <i>R</i> , <i>R</i> )- <b>C</b> 4 | 90:10:            | 46                                      |
| 5     | ( <i>R</i> , <i>R</i> )- <b>C5</b>  | 89:11:            | 89                                      |
| 6     | ( <i>R</i> , <i>R</i> )- <b>C6</b>  | 90:10:            | 72                                      |
| 7     | ( <i>R</i> , <i>R</i> )- <b>C7</b>  | 90:10:            | 59                                      |
| 8     | ( <i>R</i> , <i>R</i> )- <b>C8</b>  | 82:18:            | 88                                      |
| 9     | ( <i>R</i> , <i>R</i> )- <b>C9</b>  | 80:20:            | 87                                      |
| 10    | ( <i>R</i> , <i>R</i> )-C10         | 84:16:            | 89                                      |
| 11    | ( <i>R</i> , <i>R</i> )-C11         | 81:19:            | 88                                      |
| 12    | ( <i>R</i> , <i>R</i> )-C12         | 92:8:             | 77                                      |
| 13    | ( <i>R</i> , <i>R</i> )-C13         | 94:6:             | 79                                      |
| 14    | ( <i>R</i> , <i>R</i> )-C14         | 87:13:            | 67                                      |
| 15    | ( <i>R</i> , <i>R</i> )-C15         | 44:56:            | 23                                      |
| 16    | ( <i>R</i> , <i>R</i> )-C16         | 42:58:            | 25                                      |
| 17    | ( <i>R</i> , <i>R</i> )-C17         | 45:55:            | 22                                      |
| 18    | ( <i>R</i> , <i>R</i> )-C18         | 95:5:             | 66                                      |
| 19    | ( <i>R</i> , <i>R</i> )-C19         | 77:23:            | 64                                      |
| 20    | ( <i>R</i> , <i>R</i> )- <b>C20</b> | 98: :             | 78                                      |
| 21    | ( <i>R</i> , <i>R</i> )- <b>C21</b> | 74:26:            | 65                                      |
| 22    | ( <i>R</i> , <i>R</i> )- <b>C22</b> | 82:18:            | 78                                      |
| 23    | ( <i>R</i> , <i>R</i> )- <b>C23</b> | 22:78:            | 48                                      |
| 24    | ( <i>R</i> , <i>R</i> )- <b>C24</b> | 25:75:            | 33                                      |
| 25    | ( <i>R</i> , <i>R</i> )- <b>C25</b> | 34:66:            | 62                                      |
| 26    | ( <i>R</i> , <i>R</i> )- <b>C26</b> | :58:              |   |
| 27    | ( <i>R</i> , <i>R</i> )- <b>C27</b> | :58:              |   |

<sup>&</sup>lt;sup>*a*</sup>Reaction conditions: **1a** (0.2 mmol), **2a** (0.2 mmol), catalyst (1.0 mol %), HCO<sub>2</sub>H/Et<sub>3</sub>N (5:2) (2.4 mmol), THF (1.0 mL), r. t., N<sub>2</sub>, 16 h. <sup>*b*</sup>Determined by <sup>1</sup>H NMR analysis of crude product. <sup>*c*</sup>Determined by HPLC analysis with a chiral AD-H column.

| N<br>N<br>1a<br>+<br>PhNH2 | HO<br>( <i>R</i> , <i>R</i> )- <b>C3</b> (1 mol%)<br>HCO <sub>2</sub> H/NEt <sub>3</sub> (5/2)<br>solvent, r.t., 16 h | ,,_NHPh + NNNHPh + NH            | IPh +                                   |
|----------------------------|---|----------------------------------|---|
| 2a <sup>2</sup>            | 3a  | 4a                               | 5a                                      |
| Entry                      | Solvent   | <b>3a/4a/5a</b> (%) <sup>b</sup> | ee of <b>3a</b> (%) <sup><i>c</i></sup> |
| 1                          | iPrOH   | 70:30:                           | 64                                      |
| 2                          | HFIP  |                                  |   |
| 3                          | EtOAc   | 64:36:                           | 54                                      |
| 4                          | DME   | 26:74:                           | 88                                      |
| 5                          | THF   | 93:7:                            | 90                                      |
| 6                          | 1,4-dioxane   | 82:30:                           | 85                                      |
| 7                          | MTBE  | 88:12:                           | 83                                      |
| 8                          | dibutyl ether   | 80:20:                           | 84                                      |
| 9                          | MeOH  | 61:39:                           | 73                                      |
| 10                         | toluene   | 95:7:                            | 57                                      |
| 11                         | CHCl <sub>3</sub>   | 96:4:                            | 66                                      |
| 12                         | CH <sub>2</sub> Cl <sub>2</sub>   | 94:6:                            | 62                                      |

Table S2. Screening of solvents for the RA/ATH of 1a and 2a.<sup>a</sup>

<sup>*a*</sup>Reaction conditions: **1a** (0.2 mmol), **2a** (0.2 mmol), (*R*,*R*)-**C3** (1.0 mol %), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (2.4 mmol), solvent (1.0 mL), r.t., N<sub>2</sub>, 16 h. <sup>*b*</sup>Determined by <sup>1</sup>H NMR analysis of crude product. <sup>*c*</sup>Determined by HPLC analysis with a chiral AD-H column. HFIP: hexafluoroisopropanol; DME: dimethoxyethane; MTBE: methyl *tert*-butyl ether.

| N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>2 | $H_{2}$ CHO (R, I)<br>$H_{2}$ HCO<br>$H_{2}$ THF, a | R)-C3 (1 mol%)<br>DOH/NEt <sub>3</sub> (5/2)<br>Additive, r.t., 16 h<br>3a | NHPh + NHF        | oh + NNNNPh<br>5a                       |
|---|---|--|-------------------|---|
| Entry                                     | T (°C)  | Additive (mol%)  | $3a/4a/5a (\%)^b$ | ee of <b>3a</b> (%) <sup><i>c</i></sup> |
| 1   | r.t.  | AgOTf(2)   | 95:5:             | 90                                      |
| 2   | r.t.  | $AgSbF_{6}(2)$   | 94:6:             | 90                                      |
| 3   | r.t.  | Ag <sub>3</sub> PO <sub>4</sub> (2)  | 92:8:             | 92                                      |

Table S3. Screening of additives for the RA/ATH of 1a and 2a.<sup>a</sup>

| 4     | r.t. | $Ag_2SO_4(2)$                       | 90:10: | 85 |
|-------|------|-------------------------------------|--------|----|
| 5     | r.t. | KI (2)                              | 92:8:  | 78 |
| 6     | r.t. | Ag <sub>3</sub> PO <sub>4</sub> (2) | 96:4:  | 99 |
| 7     | 50   | Ag <sub>3</sub> PO <sub>4</sub> (2) | 99::   | 72 |
| 8     | r.t. | $Ag_{3}PO_{4}(1)$                   | 93:7:  | 91 |
| $9^d$ | r.t. | Ag <sub>3</sub> PO <sub>4</sub> (2) | 77:13: | 91 |

<sup>*a*</sup>Reaction conditions: **1a** (0.2 mmol), **2a** (0.2 mmol), (*R*,*R*)-**C3** (1.0 mol %), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (2.4 mmol), THF (1.0 mL), r.t., N<sub>2</sub>, 16 h. <sup>*b*</sup>Determined by <sup>1</sup>H NMR analysis of crude product. <sup>*c*</sup>Determined by HPLC analysis with a chiral AD-H column. <sup>*d*</sup>(*R*,*R*)-**C3** (0.5 mol%) was used.

| N<br>N<br>CHO<br>1a<br>+<br>PhNH <sub>2</sub><br>2a | (R,R)-C3 (1 mol%)<br>[H], THF, Ag <sub>3</sub> PO <sub>4</sub><br>r.t., 16 h<br>3a | N<br>N<br>4a      | NPh<br>5a                        |
|---|--|-------------------|----------------------------------|
| Entry   | [H]  | $3a/4a/5a (\%)^b$ | ee of <b>3a</b> (%) <sup>c</sup> |
| 1   | HCO <sub>2</sub> H/NEt <sub>3</sub> (1/1)  | 79:21:            | 89                               |
| 2   | HCO <sub>2</sub> H/NEt <sub>3</sub> (2/1)  | 82:18:            | 91                               |
| 3   | HCO <sub>2</sub> H/NEt <sub>3</sub> (5/2)  | 95:5:             | 99                               |
| 4   | HCO <sub>2</sub> H/NEt <sub>3</sub> (3/1)  | 85:15:            | 87                               |
| 5   | HCO <sub>2</sub> H   | 66:34:            | 44                               |
| $6^d$   | HCO <sub>2</sub> H/NEt <sub>3</sub> (5/2)  | 94:6: <b></b>     | 96                               |

Table S4. Screening of hydrogen donors for the RA/ATH of 1a and 2a.<sup>a</sup>

<sup>*a*</sup>Reaction conditions: **1a** (0.2 mmol), **2a** (0.2 mmol), THF (1.0 mL), (*R*,*R*)-**C3** (1.0 mol %), Ag<sub>3</sub>PO<sub>4</sub> (2.0 mol %), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (2.4 mmol), r.t., N<sub>2</sub>, 16 h. <sup>*b*</sup>Determined by <sup>1</sup>H NMR analysis of crude product. <sup>*c*</sup>Determined by HPLC analysis with a chiral AD-H column. <sup>*d*</sup>HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (2.0 mmol) was used.

| H<br>N<br>CHO<br>6a<br>+<br>PhNH <sub>2</sub><br>2a | catalyst (1 mol%)<br>Ag <sub>3</sub> PO <sub>4</sub> (2 mol%)<br>HCOOH/Et <sub>3</sub> N (5/2)<br>solvent, r.t., 16 h | H<br>N<br>N<br>H<br>7a | H<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N<br>N | H<br>N<br>N<br>NPh<br>7aa'       |
|---|---|------------------------|--|----------------------------------|
| Entry   | Catalyst  | Solvent                | <b>7a/7aa/7aa'</b> $(\%)^b$  | ee of <b>7a</b> (%) <sup>c</sup> |
| 1   | ( <i>R</i> , <i>R</i> )-C3  | THF                    | 88:12:   | 74                               |

Table S5. Screening of the reaction condition for RA/ATH of 6a and 2a.<sup>a</sup>

| 2                      | ( <i>R</i> , <i>R</i> )- <b>C8</b>  | THF               | 80:20:   | 74 |
|------------------------|-------------------------------------|-------------------|----------|----|
| 3                      | ( <i>R</i> , <i>R</i> )-C12         | THF               | 78:22:   | 46 |
| 4                      | ( <i>R</i> , <i>R</i> )-C16         | THF               | 75:25:   | 22 |
| 5                      | ( <i>R</i> , <i>R</i> )-C18         | THF               | 70:30:   | 36 |
| 6                      | ( <i>R</i> , <i>R</i> )- <b>C23</b> | THF               | 36:52:12 | 18 |
| 7                      | ( <i>R</i> , <i>R</i> )-C3          | IPA               | 79:21:   | 68 |
| 8                      | ( <i>R</i> , <i>R</i> )-C3          | DCE               | 89:11:   | 59 |
| 9                      | ( <i>R</i> , <i>R</i> )-C3          | 1,4-dioxane       | 85:15:   | 72 |
| 10                     | ( <i>R</i> , <i>R</i> )-C3          | toluene           | 90:10:   | 42 |
| 11                     | ( <i>R</i> , <i>R</i> )-C3          | CHCl <sub>3</sub> | 89:11:   | 37 |
| $12^d$                 | ( <i>R</i> , <i>R</i> )-C3          | THF               | 82:18:   | 74 |
| 13 <sup>e</sup>        | ( <i>R</i> , <i>R</i> )-C3          | THF               | 92:8:    | 69 |
| 14 <sup>f</sup>        | ( <i>R</i> , <i>R</i> )-C3          | THF               | 94:6:    | 62 |
| 15 <sup>g</sup>        | ( <i>R</i> , <i>R</i> )-C3          | THF               | 81:19:   | 70 |
| 16 <sup><i>h</i></sup> | ( <i>R</i> , <i>R</i> )-C3          | THF               | 80:20:   | 70 |
| $17^{i}$               | ( <i>R</i> , <i>R</i> )- <b>C3</b>  | THF               | 87:13:   | 72 |

<sup>*a*</sup>Reaction conditions: **1a** (0.2 mmol), **2a** (0.2 mmol), catalyst (1.0 mol %), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (2.4 mmol), Ag<sub>3</sub>PO<sub>4</sub> (2.0 mol %), Solvent (1.0 mL), r. t., N<sub>2</sub>, 16 h. <sup>*b*</sup>Determined by <sup>1</sup>H NMR analysis of crude product. <sup>*c*</sup>Determined by HPLC analysis with a chiral OJ-H column. <sup>*d*</sup>Reaction temperature 0 °C. <sup>*e*</sup>Reaction temperature 40 °C. <sup>*f*</sup>Reaction temperature 50 °C. <sup>*g*</sup>No Ag<sub>3</sub>PO<sub>4</sub> was used. <sup>*h*</sup>HCO<sub>2</sub>H/Et<sub>3</sub>N (1/1) (2.4 mmol) was used. <sup>*i*</sup>HCO<sub>2</sub>H/Et<sub>3</sub>N (3/1) (2.4 mmol) was used.

# 3. The general procedure



An oven-dried screw-capped pressure tube (25 mL) equipped with a magnetic stirrer bar was charged with quinoxaline-2-carbaldehyde 1 (0.2 mmol), amine 2 (0.2 mmol), Ag<sub>3</sub>PO<sub>4</sub> (1.67 mg, 0.004 mmol), (R,R)-C3 (1.40 mg, 0.002 mmol),

 $HCO_2H/Et_3N$  (5/2) (207.6 mg, 2.4 mmol, 0.2 mL) and THF (1.0 mL) under N<sub>2</sub> atmosphere in a glove box. Then the tube was capped and removed from the glovebox. The reaction mixture was stirred vigorously at room temperature for 16 h. The solvent was then removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (2.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 3.0 mL). The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate to provide product **3**.

# 4. Synthetic applications

#### 4.1 Scale-up reactions



To an oven-dried screw-capped pressure tube (100 mL) equipped with a magnetic stir bar were sequentially added quinoxaline-2-carbaldehyde (**1a**) (1.27 g, 8.0 mmol), aniline (**2a**) (0.75 g, 8.0 mmol), (*R*,*R*)-**C3** (55.44 mg, 0.08 mmol), Ag<sub>3</sub>PO<sub>4</sub> (67.0 mg, 0.16 mmol) and HCO<sub>2</sub>H/Et<sub>3</sub>N (8.06 mL, 96 mmol) and THF (40 mL) in a nitrogen atmosphere glovebox. Then the tube was capped and removed from the glovebox. The reaction mixture was allowed to stir vigorously at room temperature for 24 h. The solvent was then removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (100.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 50.0 mL). The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (5:1) to provide product **3a** (89%, 1.70 g, 99% ee).



To an oven-dried screw-capped pressure tube (100.0 mL) equipped with a magnetic stir bar were sequentially added quinoxaline-2-carbaldehyde (1a) (1.27 g, 8.0 mmol), 2,4,6-trimethylaniline (2aa) (1.08 g, 8.0 mmol), (*R*,*R*)-C3 (55.44 mg, 0.08 mmol), Ag<sub>3</sub>PO<sub>4</sub> (67.0 mg, 0.16 mmol) and HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (8.06 mL, 96 mmol) and THF (40.0 mL) in a nitrogen atmosphere glovebox. Then the tube was capped and removed from the glovebox. The reaction mixture was allowed to stir vigorously at room temperature for 24 h. The solvent was removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (100.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 50.0 mL). The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (20:1) to provide product **3aa** (87%, 1.96 g, 97% ee).





To a round bottom flask was added (*R*)-2,4,6-trimethyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (**3aa**) (140.7 mg, 0.5 mmol), PdCl<sub>2</sub> (106.4 mg, 0.6 mmol) and dry THF (3.0 mL). Then, the reaction mixture was stirred vigorously at 55 °C for 24 h. After cooling to room temperature, the resulting mixture was concentrated under vacuum to give an orange-yellow residue, which was recrystallized from CH<sub>2</sub>Cl<sub>2</sub>/hexane to afford the pure product **8** (201.8 mg, 88%).

8: Orange-yellow solid, 201.8 mg, isolated yield 88%; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)
δ 8.45 (d, *J* = 8.1 Hz, 1H), 8.11 (d, *J* = 4.2 Hz, 1H), 7.24 (dd, *J* = 11.1, 5.1 Hz, 1H),
7.04 - 6.98 (m, 1H), 6.73 - 6.66 (m, 3H), 6.57 (dd, *J* = 8.1, 1.4 Hz, 1H), 6.03 (d, *J* =

3.3 Hz, 1H), 4.22 (d, J = 12.3 Hz, 1H), 3.18 – 3.05 (m, 2H), 2.76 (dt, J = 12.6, 4.4 Hz, 1H), 2.55 (s, 3H), 2.29 (s, 3H), 2.09 (s, 3H); <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  140.57, 140.27, 135.46, 131.40, 131.31, 131.15, 129.37, 129.35, 127.33, 123.66, 117.16, 115.69, 55.93, 55.75, 38.26, 20.63, 19.89, 18.83; HRMS (ESI) calcd. for C<sub>21</sub>H<sub>22</sub>N<sub>3</sub> [M+Na]<sup>+</sup>: 480.0196, found: 480.0201.



To an oven-dried Schlenk flask containing a magnetic stir bar was added 1-bromo-2-methoxynaphthalene (**9**) (59.3 mg, 0.25 mmol), 1-naphthylboronic acid (**10**) (51.6 mg, 0.30 mmol), **8** (1.1 mg, 0.0025 mmol),  $K_3PO_4$  (106.1 mg, 0.50 mmol) and *t*-AmylOH (1.0 mL) under a nitrogen atmosphere. The reaction mixture was stirred vigorously at 25 °C until the aryl bromide was consumed as indicated by TLC. After filtration and being concentrated, the resulting residue was purified by silical gel column chromatography using hexane/ethyl acetate as eluent to give the pure product **11** (62.0mg, 81%, 85% ee).

#### (*R*)-2-Methoxy-1,1'-binaphthalene $(11)^2$



White solid, 62.0 mg, isolated yield 81%;  $[\alpha]_D^{20} = -22.6$  (c = 1.0, CHCl<sub>3</sub>); Enantiomeric excess; 85%; HPLC (OD-H, elute: hexane/isopropanol = 99/1, flowing rate = 0.4 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 19.097 min (minor), t<sub>R2</sub> = 20.510

min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 – 7.81 (m, 3H), 7.77 (dt, J = 8.3, 1.0 Hz, 1H), 7.52 (dd, J = 8.3, 7.0 Hz, 1H), 7.38 – 7.31 (m, 3H), 7.26 – 7.04 (m, 5H), 3.65 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  154.75, 134.68, 134.40, 133.83, 133.08, 129.60, 129.15, 128.57, 128.36, 127.93, 127.87, 126.51, 126.31, 125.99, 125.82, 125.71, 125.64, 123.70, 123.35, 113.96, 56.89; HRMS (ESI) calcd. for C<sub>21</sub>H<sub>16</sub>NaO [M+Na]<sup>+</sup>: 307.1093, found: 307.1099.

#### 4.3 Synthesis of hindered N-heterocyclic carbene ligand 12



To a solution of **3aa** (281.40 mg, 1.0 mmol) dissolved in CH<sub>2</sub>Cl<sub>2</sub> (20.0 mL) was sequentially added BzCl (122  $\mu$ L, 1.05 mmol) and Et<sub>3</sub>N (0.26 mL, 2.0 mmol) at 0 °C. After stirring at room temperature for 1 h, the reaction mixture was poured into water (20.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 20.0 mL). The combined extracts were washed with brine, dried with anhydrous magnesium sulfate, and evaporated in vacuo. The crude residue was dissolved in toluene (16.0 mL), and HC(OEt)<sub>3</sub> (0.85 mL, 5 mmol), NH<sub>4</sub>BF<sub>4</sub> (0.42 g, 4 mmol) and 2 drops of HCO<sub>2</sub>H were added. The reaction mixture was heated at 90 °C for 24 h. After cooling to room temperature, the resulting solution was concentrated under vacuum, and the residue was purified by silica gel column chromatography (eluent: CH<sub>2</sub>Cl<sub>2</sub>/MeOH = 50:1, v/v) to give the white solid **12** (0.31 g, 64%).

# (*S*)-5-Benzoyl-2-mesityl-3,3a,4,5-tetrahydroimidazo[1,5-a]quinoxalin-2iumtetrafluoroborate (12)



White solid, m.p. 285-287 °C;  $[\alpha]_D^{20} = -24.8 \ (c = 1.0, \text{CHCl}_3);$ <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.61 (s, 1H), 7.26 – 7.17 (m, 3H), 7.08 (t, J = 7.6 Hz, 2H), 7.00 – 6.90 (m, 2H), 6.79 (d, J= 2.0 Hz, 1H), 6.71 – 6.56 (m, 3H), 4.35 – 4.13 (m, 2H), 3.57

 $(dp, J = 11.2, 3.8, 3.2 Hz, 1H), 3.39 (dd, J = 13.8, 3.0 Hz, 1H), 2.83 (dd, J = 12.3, 9.6 Hz, 1H), 2.19 (s, 3H), 2.14 (s, 3H), 2.02 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) <math>\delta$  172.49, 160.11, 138.48, 138.13, 136.57, 135.27, 135.22, 134.31, 130.50, 130.33, 130.16, 128.21, 127.72, 126.28, 122.82, 117.76, 117.41, 115.61, 54.04, 51.80, 40.34, 20.96, 18.73, 18.62; HRMS (ESI) calcd. for C<sub>26</sub>H<sub>26</sub>N<sub>3</sub>O<sup>+</sup> [M-BF<sub>4</sub>]<sup>+</sup>: 396.2070, found: 396.2074.

# 5. Mechanistic Studies

#### 5.1 Control experiments



An oven-dried screw-capped pressure tube (25.0 mL) equipped with a magnetic stirrer bar was charged with **4a** (47.1 mg, 0.2 mmol), Ag<sub>3</sub>PO<sub>4</sub> (1.7 mg, 0.004 mmol), (*R*,*R*)-**C3** (1.4 mg, 0.002 mmol), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (207.6 mg, 2.4 mmol, 0.2 mL) and THF (1.0 mL) under N<sub>2</sub> atmosphere in a glove box. The reaction mixture was stirred vigorously at room temperature for 16 h. The solvent was then removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (2.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> ( $3 \times 3.0$  mL). The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (3/1) to provide product **3a** (44.0 mg, 92%, 99% ee).



An oven-dried screw-capped pressure tube (25.0 mL) equipped with a magnetic stirrer bar was charged with **5a** (46.7 mg, 0.2 mmol), Ag<sub>3</sub>PO<sub>4</sub> (1.7 mg, 0.004 mmol), (*R*,*R*)-**C3** (1.4 mg, 0.002 mmol), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (207.6 mg, 2.4 mmol, 0.2 mL) and THF (1.0 mL) under N<sub>2</sub> atmosphere in a glove box. The reaction mixture was stirred vigorously at room temperature for 16 h. The solvent was then removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (2.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 3.0 mL). The combined organic phases were dried over

anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (3/1) to provide product **3a** (44.0 mg, 92%, 99% ee).



#### 5.2 HRMS study





An oven-dried screw-capped pressure tube (25.0 mL) equipped with a magnetic stirrer bar was charged with 2-methylquinoxaline (13) (28.8 mg, 0.2 mmol), Ag<sub>3</sub>PO<sub>4</sub> (1.7 mg, 0.004 mmol), (R,R)-C3 (1.4 mg, 0.002 mmol), HCO<sub>2</sub>H/Et<sub>3</sub>N (5/2) (207.6 mg, 2.4 mmol, 0.2 mL) and THF (1.0 mL) under N<sub>2</sub> atmosphere in a glove box. The reaction mixture was stirred vigorously at room temperature for 16 h. The solvent was then removed under reduced pressure, and the residue was basified with the saturated aqueous NaHCO<sub>3</sub> (2.0 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 3.0 mL). The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and

concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (3/1) to provide product **14** (26.7 mg, 90% ee).

#### (S)-2-methyl-1,2,3,4-tetrahydroquinoxaline (14)<sup>3</sup>

Pale yellow oil, 26.7 mg, isolated yield 90%;  $[\alpha]_D^{20} = -30.4$  (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 90%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 10.322 min (minor), t<sub>R2</sub> = 12.092 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.60 (dd, J = 5.8, 3.4 Hz, 2H), 6.51 (dt, J = 5.8, 3.6 Hz, 2H), 3.71 – 3.43 (m, 3H), 3.32 (dd, J = 10.7, 2.9 Hz, 1H), 3.04 (dd, J = 10.7, 8.2 Hz, 1H), 1.19 (d, J = 6.3 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  133.67, 133.29, 118.80, 114.58, 114.53, 48.36, 45.82, 20.01 (one carbon was not detected due to overlap of resonances); HRMS (ESI) calcd. for C<sub>9</sub>H<sub>13</sub>N<sub>2</sub> [M+H]<sup>+</sup>: 149.1073, found: 149.1075.

#### 5.4 Verification of 3,4-dihydroquinoxalin-2(1H)-one configuration



To a 25 mL round-bottom flask in an ice-water bath was sequentially add **7a** (0.2 mmol, 50.7 mg), dry THF (10.0 mL) and LiAlH<sub>4</sub> (0.5 mmol, 19.0 mg). The reaction mixture was then vigorously stirred and heated at 90°C for 16 hours. After cooling to room temperature, the excess LiAlH<sub>4</sub> was quenched with sat. aq. NH<sub>4</sub>Cl. The resulting hydroxide precipitate was filtered, and the filtrate was dried over anhydrous sodium sulfate and concentrated under vacuum. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate (3/1) to provide product **3a** (82%, 39.2 mg, 67% ee). The structure of the product was determined to be (*S*) configuration through HPLC analysis.



| peak | Ret. Time | Area% | Area     |
|------|-----------|-------|----------|
| 1    | 43.088    | 16.58 | 6885339  |
| 2    | 47.793    | 83.42 | 34630701 |

# 6. Analytic data of products

# (S)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)aniline (3a)



Pale yellow oil, 43.5 mg, isolated yield 91%;  $[\alpha]_D^{20} = -85.6$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 99%; HPLC (AD-H elute: hexane/isopropanol = 90/10, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 49.357 min (minor),

 $t_{R2}$  = 53.587 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.24 – 7.15 (m, 2H), 6.74 (td, J = 7.3, 1.1 Hz, 1H), 6.69 – 6.58 (m, 4H), 6.54 (qt, J = 5.3, 2.7 Hz, 2H), 4.04 – 3.56 (m, 4H), 3.42 (dd, J = 10.9, 3.1 Hz, 1H), 3.35 – 3.21 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 148.25, 133.40, 132.93, 129.49, 119.17, 118.95, 117.93, 114.93, 114.68, 113.09,

49.69, 47.60, 44.16; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>18</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 240.1495, found: 240.1499.

#### (S)-4-Methyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3b)



Pale yellow oil, 48.6 mg, isolated yield 96%;  $[\alpha]_D^{20} = -$ 84.8 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 98%; HPLC (OD-H elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

41.170 min (minor),  $t_{R2} = 57.908$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.97 – 6.87 (m, 2H), 6.56 – 6.48 (m, 4H), 6.44 (ddt, J = 6.6, 2.8, 1.6 Hz, 2H), 3.98 – 3.42 (m, 4H), 3.32 (dd, J = 10.8, 3.1 Hz, 1H), 3.24 – 3.08 (m, 3H), 2.17 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  145.96, 133.41, 132.98, 129.97, 127.18, 119.13, 118.90, 114.88, 114.65, 113.25, 49.72, 47.96, 44.22, 20.50; HRMS (ESI) calcd. for C<sub>22</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 254.1652, found: 254.1656.

## (S)-4-Ethyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3c)



Pale yellow oil, 50.8 mg, isolated yield 95%;  $[\alpha]_D{}^{20} = -$ 89.8 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess; 93%; HPLC (OD-H, elute: hexane/isopropanol = 70/30, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 20.244$  min (minor),  $t_{R2} = 28.931$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.08 – 7.04 (m, 2H), 6.64 (dt, J = 6.5, 2.5 Hz, 4H), 6.55 (dtd, J = 6.6, 3.2, 1.5 Hz, 2H), 3.91 – 3.43 (m, 4H), 3.41 (dd, J = 10.8, 3.1 Hz, 1H), 3.32 – 3.19 (m, 3H), 2.58 (q, J = 7.6 Hz, 2H), 1.23 (t, J = 7.6 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.14, 133.82, 133.39, 132.97, 128.76, 119.09, 118.86, 114.86, 114.63, 113.22, 49.67, 47.89, 44.19, 28.01, 16.09; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 268.1808, found: 268.1810.

#### (S)-4-Isopropyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3d)



Pale yellow oil, 49.5 mg, isolated yield 88%.  $[\alpha]_D{}^{20} = -$ 84.1 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 93%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 13.1 min (minor), t<sub>R2</sub> = 15.1 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.14 – 7.04 (m, 2H), 6.67 – 6.58 (m, 4H), 6.57 – 6.50 (m, 2H), 4.03 – 3.57 (m, 4H), 3.42 (dd, J = 10.8, 3.1 Hz, 1H), 3.35 – 3.18 (m, 3H), 2.84 (p, J = 6.9 Hz, 1H), 1.24 (d, J = 6.9 Hz, 6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.22, 138.53, 133.41, 132.98, 127.33, 119.11, 118.88, 114.87, 114.63, 113.14, 49.67, 47.88, 44.22, 33.28, 24.37; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 282.1965, found: 282.1966

# (S)-4-(*tert*-Butyl)-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3e)



Pale yellow oil, 56.1 mg, isolated yield 95%, 95% ee.  $[\alpha]_D^{20} = -88.3$  (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (OJ-H, elute: hexane/isopropanol = 60/40, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1}$  =85.043 min (major),  $t_{R2}$  = 135.878 min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.25 – 7.21 (m, 2H), 6.65 – 6.59 (m, 4H), 6.55 – 6.50 (m, 2H), 3.72 – 3.66 (m, 1H), 3.42 (dd, *J* = 10.8, 3.1 Hz, 1H), 3.33 – 3.19 (m, 3H), 1.29 (s, 9H) (three active hydrogens were not detected); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  145.83, 140.82, 133.41, 132.98, 126.27, 119.16, 118.93, 114.91, 114.68, 112.86, 49.69, 47.85, 44.24, 34.01, 31.66; HRMS (ESI) calcd. for C<sub>19</sub>H<sub>26</sub>N<sub>3</sub><sup>+</sup> [M+H]<sup>+</sup>: 296.2121, found: 296.2123.

#### (S)-4-Benzyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3f)



Pale yellow oil, 56.6 mg, isolated yield 86%;  $[\alpha]_D^{20} =$ -92.7 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 94%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 38.308$  min (minor),  $t_{R2} = 62.957$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.34 – 7.28 (m, 2H), 7.22 (dt, J = 7.9, 1.5 Hz, 3H), 7.08 – 7.02 (m, 2H), 6.66 – 6.59 (m, 4H), 6.57 – 6.51 (m, 2H), 4.07 – 3.59 (m, 6H), 3.40 (dd, J = 10.8, 3.1 Hz, 1H), 3.32 – 3.18 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.48, 142.06, 133.38, 132.92, 130.53, 129.92, 128.88, 128.47, 125.94, 119.10, 118.89, 114.87, 114.63, 113.23, 49.62, 47.75, 44.13, 41.11; HRMS (ESI) calcd. for C<sub>22</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 330.1965, found:

330.1967.

#### (S)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)-[1,1'-biphenyl]-4-amine (3g)



Pale yellow oil, 53.0 mg, isolated yield 84%;  $[\alpha]_D{}^{20} = -$ 91.2 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 93%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

21.7 min (major),  $t_{R2} = 34.8$  min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.61 – 7.58 (m, 2H), 7.51 – 7.43 (m, 4H), 7.34 – 7.30 (m, 1H), 6.76 – 6.73 (m, 2H), 6.69 – 6.65 (m, 2H), 6.58 – 6.55 (m, 2H), 4.04 – 3.54 (m, 4H), 3.43 – 3.21 (m, 4H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  147.62, 141.16, 133.35, 132.84, 130.68, 128.78, 128.08, 126.37, 126.26, 119.13, 118.91, 114.89, 114.66, 113.28, 49.59, 47.48, 44.00; HRMS (ESI) calcd. for  $C_{21}H_{22}N_3$  [M+H]<sup>+</sup>: 316.1808, found: 316.1806.

#### (S)-4-Methoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3h)



White solid, m.p. 46.7-48.9 °C, 50.1 mg, isolated yield 93%;  $[\alpha]_D^{20} = -84.8$  (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 96%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV

detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 52.990min (minor), t<sub>R2</sub> = 75.188 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.82 – 6.78 (m, 2H), 6.65 – 6.58 (m, 4H), 6.53 (dt, *J* = 6.9, 2.6 Hz, 2H), 3.98 - 3.53 (m, 7H), 3.41 (dd, *J* = 10.8, 3.1 Hz, 1H), 3.29 – 3.16 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  152.52, 142.44, 133.42, 132.99, 119.13, 118.91, 115.11, 114.88, 114.66, 114.44, 55.96, 49.83, 48.61, 44.28; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>20</sub>N<sub>3</sub>O[M+H]<sup>+</sup>: 270.1601, found: 270.1597.

# (*S*)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)-4-(4,4,5,5-tetramethyl-1,3,2dioxaborolan-2-yl)aniline (3i)



Yellow solid, m.p. 56.4-58.6 °C, 64.1 mg, isolated yield 88%;  $[\alpha]_D{}^{20} = -84.8$  (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: >99%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda$  = 254 nm), t<sub>R1</sub> = 32.793min (minor major), t<sub>R2</sub> =47.341 min (major); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.67 – 7.65 (m, 2H), 6.63 – 6.61 (m, 4H), 6.54 – 6.52 (m, 2H), 4.21 – 3.55 (m, 4H), 3.43 – 3.21 (m, 4H), 1.34 (s, 12H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  150.74, 136.54, 133.30, 132.78, 119.15, 118.91, 114.90, 114.65, 112.04, 83.37, 49.51, 46.96, 43.88, 24.95; HRMS (ESI) calcd. for C<sub>21</sub>H<sub>29</sub>BN<sub>3</sub>O<sub>2</sub> [M+H]<sup>+</sup>: 365.2275, found: 365.2280.

# (S)-4-Fluoro-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3j)



Pale yellow oil, 47.8 mg, isolated yield 93%, 93% ee.  $[\alpha]_D^{20} = -79.7 (c = 1.0, CHCl_3);$  enantiomeric excess: 93%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda$  =

254 nm),  $t_{R1} = 18.978$  min (minor),  $t_{R2} = 22.378$  min (major); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.92 – 6.88 (m, 2H), 6.64 – 6.57 (m, 4H), 6.55 – 6.53 (m, 2H), 4.17-3.52 (m, 4H), 3.41 (dd, J = 10.8, 3.1 Hz, 1H), 3.28 – 3.25 (m, 2H), 3.20 (dd, J = 12.8, 7.3 Hz, 1H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  156.11 (d, J = 235.4 Hz), 144.61, 133.39, 132,85, 119.19, 119.01, 115.95, 115.80, 114.83 (d, J = 36.6 Hz), 113.94 (d, J = 7.5 Hz), 49.71, 48.27, 44.12; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>17</sub>FN<sub>3</sub> [M+H]<sup>+</sup>: 258.1401, found: 258.1401.

#### (S)-4-Chloro-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3k)



Pale yellow oil, 49.7 mg, isolated yield 91%;  $[\alpha]_D^{20} = -$ 82.7 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (OD-H, elute: hexane/isopropanol = 90/10, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

82.808 min (minor),  $t_{R2} = 115.401$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.15 – 7.11 (m, 2H), 6.65 – 6.60 (m, 2H), 6.59 – 6.52 (m, 4H), 4.20 – 3.68 (m, 4H), 3.40 (dd, J = 10.9, 3.1 Hz, 1H), 3.30 – 3.18 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.83, 133.34, 132.75, 129.26, 122.41, 119.23, 119.05, 114.97, 114.73, 114.13, 49.61, 47.67, 43.98; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>17</sub>ClN<sub>3</sub>[M+H]<sup>+</sup>: 274.1106, found: 274.1104, 276.1074.

#### (S)-4-Bromo-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (31)



Pale yellow oil, 57.9 mg, isolated yield 91%;  $[\alpha]_D^{20} = -$ 78.7 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 98%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

22.4 min (minor),  $t_{R2} = 31.4$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.27 (s, 1H), 7.25 (d, J = 2.2 Hz, 1H), 6.65 – 6.59 (m, 2H), 6.56 – 6.49 (m, 4H), 4.23 – 3.52 (m, 4H), 3.40 (dd, J = 10.9, 3.1 Hz, 1H), 3.32 – 3.18 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ 147.26, 133.33, 132.73, 132.14, 119.27, 119.08, 115.00, 114.75, 114.64, 109.44, 49.60, 47.58, 43.97; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>17</sub>BrN<sub>3</sub>[M+H]<sup>+</sup>: 318.0600, found: 318.0603, 320.0583.

#### (S)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)-4-(trifluoromethyl)aniline (3m)



Pale yellow oil,50.4 mg, isolated yield 82%;  $[\alpha]_D^{20} = -$ 72.6 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 99%; HPLC (OD-H, elute: hexane/isopropanol = 90/10, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 72.338$  min (minor),  $t_{R2} = 109.730$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.42 (d, J = 8.6 Hz, 2H), 6.64 (dt, J = 7.7, 2.6 Hz, 4H), 6.57 – 6.53 (m, 2H), 4.35 (s, 1H), 3.98 – 3.46 (m, 3H), 3.42 – 3.24 (m, 4H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  150.73, 133.30, 132.62, 126.78 (q, J = 3.8 Hz), 125.04 (q, J = 270.2 Hz), 119.27, 119.20 (q, J = 32.6 Hz), 119.09, 115.00, 114.75, 112.11, 49.47, 47.01, 43.76; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>17</sub>F<sub>3</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 308.1369, found: 308.1368.

# Methyl (S)-4-(((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)amino)benzoate (3n)



Pale yellow oil,48.1 mg, isolated yield 81%;  $[\alpha]_D^{20} =$ -75,4 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: >99%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 63.089$  min (minor),  $t_{R2} = 90.132$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 – 7.82 (m, 2H), 6.67 – 6.49 (m, 6H), 4.45 (s, 1H), 3.86 (s, 6H), 3.47 –

3.21 (m, 4H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 167.36, 152.01, 133.25, 132.60, 131.72, 119.32, 119.10, 118.86, 115.02, 114.77, 111.77, 51.71, 49.49, 46.90, 43.75; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>20</sub>N<sub>3</sub>O<sub>2</sub> [M+H]<sup>+</sup>: 298.1550, found: 298.1553.

#### (S)-4-(((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)amino)benzonitrile (30)



Pale yellow oil, 48.6 mg, isolated yield 92%;  $[\alpha]_D{}^{20} = -$ 75.6 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 99%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

59.066 min (minor),  $t_{R2} = 84.434$  min (major); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.44 – 7.36 (m, 2H), 6.63 (dt, J = 7.7, 3.8 Hz, 2H), 6.61 – 6.57 (m, 2H), 6.55 (dq, J = 6.2, 3.7 Hz, 2H), 4.68 (d, J = 6.1 Hz, 1H), 3.83 (d, J = 159.5 Hz, 3H), 3.41 – 3.24 (m, 4H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  151.47, 133.85, 133.18, 132.45, 120.53, 119.36, 119.14, 115.04, 114.80, 112.48, 99.04, 49.38, 46.73, 43.58; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>17</sub>N<sub>4</sub> [M+H]<sup>+</sup>: 265.1448, found: 265.1451.

# (S)-2-Methyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3p)



Pale yellow oil, 42.0 mg, isolated yield 83%;  $[\alpha]_D^{20} = -78.4$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =50.565

min (minor),  $t_{R2} = 67.338$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.21 (td, J = 7.7, 1.6 Hz, 1H), 7.15 (dd, J = 7.3, 1.6 Hz, 1H), 6.80 – 6.67 (m, 4H), 6.62 – 6.55 (m, 2H), 4.04 – 3.62 (m, 4H), 3.48 – 3.28 (m, 4H), 2.22 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.05, 133.35, 132.90, 130.30, 127.23, 122.24, 119.02, 118.82, 117.36, 114.84, 114.58, 109.84, 49.35, 47.43, 44.09, 17.62; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>20</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 254.1652, found: 254.1648.

#### (S)-2-Isopropyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3q)



Pale yellow oil, 45.0 mg, isolated yield 80%;  $[\alpha]_D^{20} = -76.5$ (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 96%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 10.562 min (major), t<sub>R2</sub> = 11.991 min (minor); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.15 (dd, J = 7.7, 1.6 Hz, 1H), 7.10 (td, J = 7.7, 1.6 Hz, 1H), 6.75 (td, J = 7.4, 1.2 Hz, 1H), 6.66 (dd, J = 8.1, 1.2 Hz, 1H), 6.62 – 6.57 (m, 2H), 6.51 – 6.48 (m, 2H), 4.05 (d, J = 5.8 Hz, 1H), 3.84 (s, 1H), 3.70 (qd, J = 6.3, 3.1 Hz, 1H), 3.60 (s, 1H), 3.38 – 3.22 (m, 4H), 2.84 (p, J = 6.8 Hz, 1H), 1.24 (dd, J = 6.8, 1.1 Hz, 6H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  144.78, 133.36, 132.91, 132.53, 126.82, 125.16, 119.05, 118.89, 117.75, 114.93, 114.63, 110.59, 49.36, 47.67, 44.16, 27.33, 22.40; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 282.1965, found: 282.1962.

#### (S)-2-Fluoro-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3r)



Pale yellow oil, 43.2 mg, isolated yield 84%;  $[\alpha]_D^{20} = -86.3$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 93%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =13.058

min (major),  $t_{R2} = 18.106$ min (minor); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.05 – 7.01 (m, 2H), 6.78 (td, J = 8.3, 1.5 Hz, 1H), 6.71 – 6.64 (m, 3H), 6.57 – 6.55 (m, 2H), 4.26 – 4.20 (m, 1H), 3.96 (s, 1H), 3.71 – 3.68 (m, 2H), 3.42 (dd, J = 10.9, 3.1 Hz, 1H), 3.34 (dt, J = 13.1, 5.4 Hz, 1H), 3.30 – 3.25 (m, 2H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  151.68 (d, J = 238.3 Hz), 136.63 (d, J = 11.5 Hz), 133.31, 132.74, 124.72 (d, J = 3.7 Hz), 119.13, 118.88, 117.09 (d, J = 7.0 Hz), 114.90, 114.64 (d, J = 18.4 Hz), 114.63, 112.28 (d, J = 3.0 Hz), 49.62, 47.10, 43.89; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>17</sub>FN<sub>3</sub> [M+H]<sup>+</sup>: 258.1401, found: 258.1406.

# (S)-2-Methyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3s)



Pale yellow oil, 44.1 mg, isolated yield 87%;  $[\alpha]_D^{20} = -55.2$  (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 0.7 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

51.6 min (minor),  $t_{R2} = 67.9$  min (major); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.14 (td, J = 7.3, 1.3 Hz, 1H), 6.69 – 6.66 (m, 2H), 6.63 (d, J = 7.4 Hz, 1H), 6.57 (dt, J = 7.0, 2.5 Hz, 2H), 6.52 (d, J = 7.3 Hz, 2H), 3.96-3.92 (m, 2H), 3.71-3.67 (m, 2H), 3.41 (dd, J = 10.9,

3.1 Hz, 1H), 3.32-3.22 (m, 3H), 2.35 (s, 3H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 148.21, 139.20, 133.34, 132.90, 129.27, 119.03, 118.81, 118.72, 114.83, 114.59, 113.76, 110.16, 49.55, 47.46, 44.07, 21.70; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>20</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 254.1652, found: 254.1651.

#### (S)-3,5-Dimethyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3t)



Pale yellow oil, 50.8 mg, isolated yield 95%;  $[\alpha]_D^{20} = -$ 81.7 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 94%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

16.685 min (minor),  $t_{R2} = 18.233$  min (major) <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.66 – 6.62 (m, 2H), 6.55 (ddt, J = 6.5, 5.7, 2.9 Hz, 2H), 6.44 (s, 1H), 6.32 (d, J = 1.6 Hz, 2H), 4.06 – 3.54 (m, 4H), 3.42 (dd, J = 10.8, 3.1 Hz, 1H), 3.33 – 3.21 (m, 3H), 2.28 (s, 6H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  148.29, 139.14, 133.38, 132.95, 119.87, 119.09, 118.86, 114.88, 114.62, 110.99, 49.63, 47.55, 44.17, 21.60; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 268.1808, found: 268.1805.

#### (S)-3,5-Dimethoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3u)



Pale yellow oil, 47.9 mg, isolated yield 80%;  $[\alpha]_D{}^{20} =$ -85.8 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 96%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 16.7 \text{ min} (\text{minor})$ ,  $t_{R2} = 18.2 \text{ min} (\text{major})$ ; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta 6.64 - 6.61 (\text{m}, 2\text{H})$ , 6.53 (dt, J = 7.0, 2.5 Hz, 2H), 5.92 (t, J = 2.2 Hz, 1H), 5.85 (d, J = 2.1 Hz, 2H), 4.09 - 3.65 (m, 3H), 3.76 (s, 6H), 3.67 (tdd, J = 6.6, 5.2, 3.1 Hz, 1H), 3.38 (dd, J = 10.9, 3.1 Hz, 1H), 3.29 - 3.18 (m, 3H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ 161.88, 150.18, 133.35, 132.87, 119.09, 118.86, 114.86, 114.63, 91.88, 90.04, 55.27, 49.53, 47.47, 43.95; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub>O<sub>2</sub> [M+H]<sup>+</sup>: 300.1707, found: 300.1705.

(S)-3,5-Dimethoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3v)



Pale yellow oil, 69.0 mg, isolated yield 92%;  $[\alpha]_D^{20} = -$ 85.8 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 99%; HPLC (OJ-H, elute: hexane/isopropanol = 60/40, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 12.938$  min (major),  $t_{R2} = 18.399$  min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.17 (s, 1H), 6.97 (d, J = 1.6 Hz, 2H), 6.66 (dd, J = 5.8, 3.4 Hz, 2H), 6.59 – 6.53 (m, 2H), 4.55 (t, J = 5.8 Hz, 1H), 3.94 – 3.54 (m, 3H), 3.44 – 3.26 (m, 4H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  148.96, 133.24, 132.61 (q, J = 32.7 Hz), 132.44, 123.66 (q, J = 272.7 Hz), 119.49, 119.30, 115.19, 114.91, 112.15 (d, J = 4.4 Hz), 110.60(m), 49.45, 47.22, 43.62; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>16</sub>F<sub>6</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 376.1243, found: 376.1245. **(S)-3,5-Dimethoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3w)** 



Pale yellow oil, 44.4 mg, isolated yield 83%;  $[\alpha]_D^{20} = -$ 80.7 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

11.775 min (major),  $t_{R2} = 12.872$  min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.99 – 6.95 (m, 2H), 6.66 – 6.61 (m, 3H), 6.57 – 6.54 (m, 2H), 4.05 – 3.52 (m, 4H), 3.45 (dd, J = 10.9, 3.1 Hz, 1H), 3.39 – 3.26 (m, 3H), 2.28 (s, 3H), 2.17 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.78, 133.40, 132.99, 131.24, 127.48, 126.63, 122.46, 119.08, 118.86, 114.88, 114.63, 110.17, 49.49, 47.79, 44.23, 20.42, 17.61; HRMS (ESI) calcd. for  $C_{17}H_{22}N_3$  [M+H]<sup>+</sup>: 268.1808, found: 268.1812.

# (S)-2,6-Dimethyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3x)



Pale yellow oil, 48.6 mg, isolated yield 91%;  $[\alpha]_D^{20} = -75.8$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 99%; HPLC (AD-H, elute: hexane/isopropanol = 90/10, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 19.768 min (minor),

 $t_{R2} = 21.784 \text{ min (major)};^{1}\text{H NMR (400 MHz, CDCl_3)} \delta 6.91 (d, J = 7.5 Hz, 2H), 6.76 (t, J = 7.5 Hz, 1H), 6.56 - 6.49 (m, 2H), 6.48 - 6.39 (m, 2H), 3.78 - 3.48 (m, 4H), 3.29 (dd, J = 10.8, 3.1 Hz, 1H), 3.14 (dd, J = 10.8, 6.8 Hz, 1H), 3.05 - 2.92 (m, 2H), 2.21 (s, J = 10.8, 10.8) (m, 2H) (m, 2H)$ 

6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 145.68, 133.46, 133.02, 129.78, 129.07, 122.41, 119.05, 118.88, 114.94, 114.61, 51.55, 51.14, 44.39, 18.63; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 268.1808, found: 268.1805.

# (R)-2,6-Dimethoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3y)



Pale yellow oil, 55.6 mg, isolated yield 93%;  $[\alpha]_D{}^{20} = -78.9$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 92%; HPLC (AD-H, elute: hexane/isopropanol = 90/10, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 19.768

min (minor),  $t_{R2} = 21.784$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.85 (t, J = 8.3 Hz, 1H), 6.64 – 6.54 (m, 4H), 6.52-6.49 (m, 2H), 4.40 - 4.91 (m, 2H), 3.87 (s, 6H), 3.71 (s, br, 1H), 3.48 – 3.34 (m, 3H), 3.28 – 3.12 (m, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  151.32, 133.47, 133.44, 126.17, 120.64, 118.89, 118.47, 114.60, 114.49, 104.77, 55.97, 50.09, 50.00, 44.29; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub>O<sub>2</sub> [M+H]+: 300.1707, found: 300.1710.

#### (S)-2,6-Diisopropyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3z)



Pale yellow oil, 51.1 mg, isolated yield 79%;  $[\alpha]_D{}^{20} = -86.9$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 91%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 5.64 min (major),

 $t_{R2}$  = 4.63 min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.16 – 7.12 (m, 3H), 6.70 – 6.60 (m, 3H), 6.56 – 6.54 (m, 1H), 3.72 – 3.66 (m, 1H), 3.48 (dd, *J* = 10.8, 3.1 Hz, 1H), 3.35 – 3.26 (m, 3H), 3.08 – 2.96 (m, 2H), 1.29 (d, *J* = 6.9 Hz, 6H), 1.24 (d, *J* = 6.9 Hz, 6H) (three active hydrogens were not detected due to overlap); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 142.89, 142.79, 133.49, 133.00, 124.32, 123.72, 119.08, 118.95, 115.05, 114.66, 54.90, 51.06, 44.56, 27.73, 24.43, 24.20; HRMS (ESI) calcd. for C<sub>21</sub>H<sub>30</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 324.2434, found: 324.2434.

## (S)-2,4,6-Trimethyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3aa)



Pale yellow oil, 50.6 mg, isolated yield 90%;  $[\alpha]_D^{20} = -$ 83.2 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 97%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

25.481 min (major),  $t_{R2} = 27.658$  min (minor); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.91 (s, 2H), 6.70 – 6.66 (m, 2H), 6.61 – 6.60 (m, 1H), 6.58 – 6.55 (m, 1H), 4.17 (s, br, 1H), 3.66 – 3.61 (m, 1H), 3.43 (dd, J = 10.8, 3.1 Hz, 1H), 3.28 (dd, J = 10.8, 6.9 Hz, 1H), 3.12 – 3.03 (m, 2H), 2.34 (s, 6H), 2.31 (s, 3H), (two active hydrogens were not detected); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  143.01, 133.44, 133.05, 131.82, 130.08, 129.60, 118.93, 118.76, 114.83, 114.53, 51.80, 51.05, 44.43, 20.63, 18.38; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 282.1965, found: 282.1969.

#### (S)-3,4,5-Trimethoxy-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3ab)



Pale yellow oil, 62.5 mg, isolated yield 95%;  $[\alpha]_D^{20} =$ -84.6 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: >99%; HPLC (OD-H, elute: hexane/isopropanol = 60/40, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1}$  =27.638 min (minor),  $t_{R2}$  = 43.175 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.64 – 6.58 (m, 2H), 6.57 – 6.49 (m, 2H), 5.89 (s, 2H), 4.01- 3.74(m, 12H), 3.70 – 3.64 (m, 1H), 3.41 (dd, J = 10.8, 3.1 Hz, 1H), 3.32 – 3.18 (m, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  154.09, 145.15, 133.34, 132.84, 130.40, 119.17, 118.93, 114.91, 114.66, 90.67, 61.19, 56.06, 49.99, 48.05, 44.02; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>24</sub>N<sub>3</sub>O<sub>3</sub> [M+H]<sup>+</sup>: 330.1812, found: 330.1816.

## (S)-2,4,6-Triisopropyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3ac)



Pale yellow oil, 51.1 mg, isolated yield 79%;  $[\alpha]_D^{20} = -$ 82.1 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 82%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 5.996 \text{ min (major)}$ ,  $t_{R2} = 8.270 \text{ min (minor)}$ ; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.96 (s, 2H), 6.64 – 6.58 (m, 3H), 6.54 – 6.52 (m, 1H), 4.04 – 3.04 (m, 9H), 3.02 (dd, J = 11.9, 4.4 Hz, 1H), 2.95 – 2.85 (m, 2H), 1.26 (dd, J = 6.9, 2.8 Hz, 12H), 1.21 (d, J = 6.9 Hz, 6H).; <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  144.44, 142.78, 140.40, 133.55, 133.14, 121.62, 119.08, 118.91, 114.99, 114.65, 55.04, 51.13, 44.71, 34.10, 27.83, 24.54, 24.30, 24.28, 24.25; HRMS (ESI) calcd. for C<sub>24</sub>H<sub>36</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 366.2904, found: 366.2908.

# (S)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)naphthalen-2-amine (3ad)



Pale yellow oil, 53.2 mg, isolated yield 92%,  $[\alpha]_D^{20} = -$ 90.4 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 92%; HPLC (hexane : isopropanol = 70 : 30, flowing rate = 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =19.899 min (major), t<sub>R2</sub> = 22.135 min (minor); <sup>1</sup>H NMR (600 MHz,

CDCl<sub>3</sub>)  $\delta$  7.73 (d, *J* = 8.1 Hz, 1H), 7.67 (dd, *J* = 12.8, 8.5 Hz, 2H), 7.43 (ddd, *J* = 8.1, 6.7, 1.3 Hz, 1H), 7.29 – 7.26 (m, 1H), 6.92 – 6.87 (m, 2H), 6.70 – 6.67 (m, 2H), 6.58 – 6.55 (m, 2H), 4.27 – 3.56 (m, 4H), 3.41 – 3.26 (m, 4H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  145.85, 135.17, 133.33, 132.85, 129.10, 127.73, 127.66, 126.53, 126.00, 122.24, 119.11, 118.90, 118.06, 114.91, 114.66, 104.62, 49.46, 47.42, 44.02; HRMS (ESI) calcd. for C<sub>19</sub>H<sub>20</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 290.1652, found: 290.1658

## (S)-N-((1,2,3,4-Tetrahydroquinoxalin-2-yl)methyl)naphthalen-1-amine (3ae)



Pale yellow oil, 53.8 mg, isolated yield 93%;  $[\alpha]_D{}^{20} = -93.7$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 90%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 13.478 min (minor),

t<sub>R2</sub> = 17.233 min (major); <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 7.69 – 7.64 (m, 2H), 7.34 – 7.30 (m, 2H), 7.22 (tdt, J = 7.4, 3.5, 1.7 Hz, 1H), 7.14 (dt, J = 7.3, 2.7 Hz, 1H), 6.51 (tdd, J = 7.5, 4.7, 2.8 Hz, 3H), 6.39 (dd, J = 5.9, 3.4 Hz, 2H), 4.58 (t, J = 5.6 Hz, 1H), 3.75 (s, 1H), 3.63 (qq, J = 6.2, 3.1 Hz, 1H), 3.48 (s, 1H), 3.25 – 3.15 (m, 4H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 143.35, 134.38, 133.33, 132.95, 128.76, 126.62, 125.92, 124.91, 123.60, 120.02, 119.10, 118.90, 117.78, 114.96, 114.65, 104.57, 49.18, 47.62, 44.18; HRMS (ESI) calcd. for C<sub>19</sub>H<sub>20</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 290.1652, found: 290.1654.

(S)-1,1-Diphenyl-N-((1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)methanamine (3af)

Pale yellow oil, 54.0 mg, isolated yield 82%;  $[\alpha]_D^{20} = -$ 93.7 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 90%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

35.366 min (minor),  $t_{R2} = 46.663$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.43 – 7.40 (m, 4H), 7.33 (td, J = 7.6, 3.1 Hz, 4H), 7.27 – 7.24 (m, 2H), 6.66 – 6.50 (m, 4H), 4.87 (s, 1H), 3.53 – 3.32 (m, 5H), 3.13 (dd, J = 10.7, 7.2 Hz, 1H), 2.77 (dd, J = 11.8, 4.5 Hz, 1H), 2.65 (dd, J = 11.8, 8.3 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  143.93, 143.50, 133.40, 133.28, 128.69, 128.65, 127.37, 127.32, 127.25, 119.05, 118.65, 114.71, 114.63, 67.49, 51.23, 50.33, 44.79; HRMS (ESI) calcd. for C<sub>22</sub>H<sub>24</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 330.1965, found: 330.1968.

#### (S)-N-((6,7-Dimethyl-1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3ba)



Pale yellow oil, 49.2 mg, isolated yield 92%;  $[\alpha]_D^{20} = -$ 87.3 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 92%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

22.263 min (major),  $t_{R2} = 24.300$  min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.24 – 7.20 (m, 2H), 6.77 (tt, J = 7.3, 1.1 Hz, 1H), 6.71 – 6.65 (m, 2H), 6.38 (d, J = 1.8 Hz, 2H), 3.76 – 3.41 (m, 4H), 3.39 (dd, J = 10.9, 3.0 Hz, 1H), 3.33 – 3.21 (m, 3H), 2.15 (t, J = 1.2 Hz, 6H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  148.26, 131.12, 130.65, 129.41, 126.94, 126.67, 117.75, 116.72, 116.49, 113.01, 49.85, 47.40, 44.43, 18.99, 18.98; HRMS (ESI) calcd. for C<sub>17</sub>H<sub>22</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 268.1808, found: 268.1810.

# (S)-N-((6,7-Difluoro-1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3ca)



Pale yellow oil, 47.9 mg, isolated yield 87%;  $[\alpha]_D^{20} = -$ 78.5 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 95%; HPLC (AD-H, elute: hexane/isopropanol = 80/10, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

16.498 min (minor),  $t_{R2} = 18.8$  min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.24 – 7.17 (m, 2H), 6.76 (tt, *J* = 7.3, 1.1 Hz, 1H), 6.68 – 6.65 (m, 2H), 6.31 (ddd, *J* = 11.5, 7.7, 1.7

Hz, 2H), 3.78 (s, br, 3H), 3.62 (dddd, J = 7.9, 6.5, 5.0, 3.1 Hz, 1H), 3.36 (dd, J = 11.0, 3.1 Hz, 1H), 3.29 (dd, J = 13.1, 5.0 Hz, 1H), 3.23 – 3.17 (m, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  146.92, 143.26 (dd, J = 10.3, 5.4 Hz), 141.02 (dd, J = 11.6, 4.4 Hz), 128.38, 127.95 (dd, J = 7.8, 2.4 Hz), 127.50 (dd, J = 7.9, 2.5 Hz), 116.96, 111.94, 102.13 (d, J = 20.0 Hz), 101.87 (d, J = 20.6 Hz), 48.29, 46.19, 42.76; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>16</sub>F<sub>2</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 276.1307, found: 276.1304.

# (S)-N-((6,7-Dichloro-1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3da)

Pale yellow oil, 55.9 mg, isolated yield 91%;  $[\alpha]_D^{20} = -$ 79.7 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 94%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> =

31.657 min (major),  $t_{R2} = 37.644$  min (minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.24 – 7.19 (m, 2H), 6.77 (tt, J = 7.3, 1.1 Hz, 1H), 6.66 (dt, J = 7.7, 1.1 Hz, 2H), 6.52 (d, J = 1.8 Hz, 2H), 4.04 (s, br, 1H), 3.89 (s, br, 1H), 3.71 (s, br, 1H), 3.63 – 3.57 (m, 1H), 3.36 (dd, J = 11.0, 3.1 Hz, 1H), 3.28 (dd, J = 13.2, 5.0 Hz, 1H), 3.18 (dt, J = 11.2, 5.2 Hz, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  147.95, 133.04, 132.55, 129.52, 120.60, 120.45, 118.12, 115.10, 114.80, 113.05, 49.16, 47.28, 43.48; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>16</sub>Cl<sub>2</sub>N<sub>3</sub> [M+H]<sup>+</sup>: 308.0716, found: 308.0718, 312.0660.

#### (S)-N-((6,7-Dibromo-1,2,3,4-tetrahydroquinoxalin-2-yl)methyl)aniline (3ea)

Pale yellow oil, 67.1 mg, isolated yield 85%;  $[\alpha]_D^{20} = -$ 76.2 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 87%; HPLC (AD-H, elute: hexane/isopropanol = 80/20, flowing rate:

1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 21.664 min (minor), t<sub>R2</sub> = 23.526 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.24 – 7.18 (m, 2H), 6.77 (ddt, J = 8.4, 7.3, 1.1 Hz, 1H), 6.70 – 6.64 (m, 4H), 4.07 – 3.68 (m, 3H), 3.63 – 3.56 (m, 1H), 3.35 (dd, J = 11.0, 3.2 Hz, 1H), 3.28 (dd, J = 13.2, 5.0 Hz, 1H), 3.23 – 3.13 (m, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  147.93, 133.84, 133.35, 129.53, 118.16, 118.09, 117.78, 113.07, 111.72, 111.56, 49.13, 47.30, 43.41; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>16</sub>Br<sub>2</sub>N<sub>3</sub> [M+H]<sup>+</sup>:

395.9705, found: 395.9708, 399.9662.

# (S)-3-((Phenylamino)methyl)-3,4-dihydroquinoxalin-2(1H)-one (7a)



Pale yellow oil, 41.5 mg, isolated yield 82%;  $[\alpha]_D{}^{20} = -67.3$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 74%; HPLC (OJ-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 29.4 min (major),

 $t_{R2} = 37.4 \text{ min (minor)}; {}^{1}\text{H NMR (400 MHz, CDCl_3)} \delta 8.28 (s, 1H), 7.22 - 7.16 (m, 2H), 6.91 (td,$ *J*= 7.5, 1.6 Hz, 1H), 6.80 - 6.67 (m, 6H), 4.23 (s, 1H), 4.21-4.17 (m, 1H), 4.11 (s, 1H), 3.68 (dd,*J*= 13.6, 4.4 Hz, 1H), 3.51 (dd,*J* $= 13.5, 8.2 Hz, 1H); {}^{13}\text{C NMR (101 MHz, CDCl_3)} \delta 167.59, 147.57, 132.84, 129.58, 125.18, 124.28, 119.85, 118.46, 115.53, 114.62, 113.35, 55.43, 45.78; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>16</sub>N<sub>3</sub>O [M+H]<sup>+</sup>: 254.1288, found: 254.1290.$ 

#### (S)-3-(((4-Isopropylphenyl)amino)methyl)-3,4-dihydroquinoxalin-2(1H)-one (7b)



Pale yellow oil, 47.2 mg, isolated yield 80%;  $[\alpha]_D{}^{20} = -$ 67.3 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 72%; HPLC (OJ-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 254 nm), t<sub>R1</sub> = 23.872 min (major), t<sub>R2</sub> = 37.258 min

(minor); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.21 (s, 1H), 7.11 – 7.03 (m, 2H), 6.90 (ddd, *J* = 7.8, 5.2, 3.7 Hz, 1H), 6.76 (dd, *J* = 4.2, 1.0 Hz, 2H), 6.71 – 6.61 (m, 3H), 4.33 (s, 1H), 4.23 – 4.15 (m, 1H), 3.67 (dd, *J* = 13.5, 4.3 Hz, 1H), 3.49 (dd, *J* = 13.5, 8.3 Hz, 1H), 2.83 (hept, *J* = 6.8 Hz, 1H), 1.22 (d, *J* = 7.0 Hz, 6H) (one active hydrogen was not detected); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  168.25, 145.54, 139.04, 132.86, 127.40, 125.23, 124.21, 119.71, 115.74, 114.51, 113.46, 55.34, 46.04, 33.28, 24.34; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>22</sub>N<sub>3</sub>O [M+H]<sup>+</sup>: 296.1757, found: 296.175

#### (S)-3-(((4-Methoxyphenyl)amino)methyl)-3,4-dihydroquinoxalin-2(1H)-one (7c)



Pale yellow oil, 48.3 mg, isolated yield 85%;  $[\alpha]_D^{20} = -$ 70.2 (c = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 69%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda =$ 

254 nm),  $t_{R1} = 17.802 \text{ min} (\text{minor})$ ,  $t_{R2} = 22.677 \text{ min} (\text{major})$ ; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.95 (s, 1H), 6.90 (ddd, J = 7.7, 6.3, 2.6 Hz, 1H), 6.82 – 6.73 (m, 4H), 6.70 – 6.63 (m, 3H), 4.30 (s, 1H), 4.17 (ddd, J = 8.2, 4.3, 1.7 Hz, 1H), 3.85 (s, 1H), 3.75 (s, 3H), 3.62 (dd, J = 13.3, 4.4 Hz, 1H), 3.45 (dd, J = 13.3, 8.1 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  168.07, 152.81, 141.67, 132.87, 125.22, 124.20, 119.71, 115.66, 115.12, 114.81, 114.50, 55.91, 55.43, 46.81; HRMS (ESI) calcd. for C<sub>16</sub>H<sub>18</sub>N<sub>3</sub>O<sub>2</sub> [M+H]<sup>+</sup>: 284.1394, found: 284.1395.

#### (S)-3-((Mesitylamino)methyl)-3,4-dihydroquinoxalin-2(1H)-one (7d)



Colorless oil, 45.5 mg, isolated yield 77%, 46% ee.  $[\alpha]_D^{20}$ = -55.4 (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 46%; HPLC (OJ-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda$  = 254 nm), t<sub>R1</sub>

= 11.748 min (minor),  $t_{R2}$  = 13.783 min (major); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.20 (s, 1H), 6.91 (ddd, *J* = 7.8, 6.4, 2.5 Hz, 1H), 6.83 (s, 2H), 6.79 – 6.73 (m, 2H), 6.71 – 6.66 (m, 1H), 4.34 (s, 1H), 4.12 (ddd, *J* = 6.8, 4.5, 1.9 Hz, 1H), 3.53 – 3.05 (m, 3H), 2.28 (s, 6H), 2.24 (s, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  168.28, 142.45, 133.06, 132.28, 130.49, 129.71, 125.33, 124.15, 119.73, 115.67, 114.55, 56.94, 49.83, 20.69, 18.40; HRMS (ESI) calcd. for C<sub>18</sub>H<sub>22</sub>N<sub>3</sub>O [M+H]<sup>+</sup>: 296.1757, found: 296.1753

# (S)-6,7-Difluoro-3-((phenylamino)methyl)-3,4-dihydroquinoxalin-2(1H)-one (7e)



Yellow solid, 48.6 mg, isolated yield 84%;  $[\alpha]_D^{20} = -65.2$ (*c* = 1.0, CHCl<sub>3</sub>); enantiomeric excess: 70%; HPLC (OD-H, elute: hexane/isopropanol = 80/20, flowing rate: 1.0 mL/min, 25 °C, UV detection at  $\lambda = 254$  nm), t<sub>R1</sub> = 15.111

min (major),  $t_{R2} = 17.573$  min (minor); <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  10.41 (s, 1H),

7.11 – 7.05 (m, 2H), 6.70 (td, J = 8.1, 3.6 Hz, 2H), 6.64 – 6.59 (m, 2H), 6.56 (tt, J = 7.3, 1.1 Hz, 1H), 6.28 (d, J = 2.0 Hz, 1H), 5.61 (t, J = 6.1 Hz, 1H), 3.96 (ddd, J = 7.4, 4.1, 1.9 Hz, 1H), 3.35 (s, 1H), 3.24 (dt, J = 13.6, 7.1 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ )  $\delta$  166.69, 148.81, 145.23 (dd, J = 224.1, 12.7 Hz), 141.95 (dd, J = 220.4, 12.1 Hz), 130.97 (d, J = 6.8 Hz), 129.38, 122.27 (d, J = 6.3 Hz), 116.67, 112.79, 103.84 (d, J = 21.9 Hz), 102.35 (d, J = 21.9 Hz), 54.86, 45.27; HRMS (ESI) calcd. for C<sub>15</sub>H<sub>14</sub>F<sub>2</sub>N<sub>3</sub>O [M+H]<sup>+</sup>: 290.1099, found: 290.1095.

# 7. References

- Y. Chen, Y. -M. He, S. Zhang, T. Miao, and Q. -H. Fan, *Angew. Chem. Int. Ed.* 2019, 58, 3809 – 3813.
- 2. M. Koichi, M. Takashi, H. Manabu, Chem. Commun. 2004, 2082-2083.
- N. Arai, Y. Saruwatari, K. Isobe, Takeshi Ohkuma, *Adv. Synth. Catal.* 2013, 355, 2769–2774.

# 8. <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of the products

 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound  $\bm{3a}$  in CDCl\_3







# <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3c** in CDCl<sub>3</sub>



<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3d** in CDCl<sub>3</sub>





 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 3e in CDCl\_3
#### $^1H$ and $^{13}C\{^1H\}$ NMR spectra of compound 3f in CDCl\_3



## <sup>1</sup>H and <sup>13</sup>C {<sup>1</sup>H} NMR spectra of compound **3g** in CDCl<sub>3</sub>



# <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound 3g in CDCl<sub>3</sub>

H N N H N N N







### $^1H$ and $^{13}C\{^1H\}$ NMR spectra of compound 3i in CDCl\_3

<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3j** in CDCl<sub>3</sub>



### <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3k** in CDCl<sub>3</sub>









 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound  $\boldsymbol{3m}$  in CDCl\_3



 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound **3n** in CDCl<sub>3</sub>

<sup>1</sup>H and <sup>13</sup>C {<sup>1</sup>H} NMR spectra of compound **3o** in CDCl<sub>3</sub>



<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3p** in CDCl<sub>3</sub>



# <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3q** in CDCl<sub>3</sub>



### <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3r** in CDCl<sub>3</sub>





<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3s** in CDCl<sub>3</sub>



<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3t** in CDCl<sub>3</sub>

#### <sup>1</sup>H and <sup>13</sup>C $\{^{1}H\}$ NMR spectra of compound **3u** in CDCl<sub>3</sub>



 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 3v in CDCl\_3







<sup>1</sup>H and <sup>13</sup>C {<sup>1</sup>H} NMR spectra of compound 3w in CDCl<sub>3</sub>











<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound 3x in CDCl<sub>3</sub>

#### <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound 3y in CDCl<sub>3</sub>









 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 3aa in CDCl\_3



 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound  $\boldsymbol{3ab}$  in CDCl\_3

#### $^1H$ and $^{13}C\{^1H\}$ NMR spectra of compound $\boldsymbol{3ac}$ in $CDCl_3$

- 404 - 404 - 404 - 405 - 400 - 405



### <sup>1</sup>H and <sup>13</sup>C {<sup>1</sup>H} NMR spectra of compound **3ad** in CDCl<sub>3</sub>







### <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3af** in CDCl<sub>3</sub>





 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 3ba in CDCl\_3

## <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **3ca** in CDCl<sub>3</sub>











#### $^1H$ and $^{13}C\{^1H\}$ NMR spectra of compound **3ea** in CDCl\_3

6 79 6 6.77 6 6.67 6 6 7.24 7.23 7.23 7.23 7.23 7.23 7.23 7.20 7.20 7.20 7.20 7.20



<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound 7a in CDCl<sub>3</sub>

<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of compound **7b** in CDCl<sub>3</sub>





 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 7c in CDCl\_3





 $\begin{array}{c} 6689\\ 66992\\ 66992\\ 66992\\ 66976\\ 66977$ 








## <sup>1</sup>H and <sup>13</sup>C {<sup>1</sup>H} NMR spectra of compound **11** in CDCl<sub>3</sub>





 $^1H$  and  $^{13}C\{^1H\}$  NMR spectra of compound 14 in CDCl\_3



9. Copy of HPLC spectra of the racemic and chiral products





| peak | Ret. Time | Area% | Area      |
|------|-----------|-------|-----------|
| 1    | 41. 170   | 1.09  | 1212964   |
| 2    | 57. 908   | 98.91 | 110249267 |



| 1 | 21.685 | 50.10 | 78916480 |
|---|--------|-------|----------|
| 2 | 30.547 | 49.90 | 78603936 |















| peak | Ret. Time | Area% | Area     |
|------|-----------|-------|----------|
| 1    | 50.046    | 50.71 | 22221195 |
| 2    | 71.992    | 49.29 | 21596002 |











| 1 | 23.939 | 50.13 | 26763835 |
|---|--------|-------|----------|
| 2 | 33.463 | 49.87 | 26628296 |









| 1 | 58.082 | 50.08 | 8045229 |
|---|--------|-------|---------|
| 2 | 88.428 | 49.92 | 8020582 |





















| peak | Ret. Time | Area% | Area     |
|------|-----------|-------|----------|
| 1    | 12.720    | 50.21 | 19354056 |
| 2    | 17.773    | 49.79 | 19191038 |



| peak | Ret. Time | Area% | Area     |  |
|------|-----------|-------|----------|--|
| 1    | 12.938    | 99.32 | 65617075 |  |
| 2    | 18.399    | 0.68  | 450055   |  |





| peak | Ret. Time | Area% | Area     |
|------|-----------|-------|----------|
| 1    | 19.685    | 49.68 | 11823673 |
| 2    | 21.738    | 50.32 | 11975219 |

99.27

21.784



6471942





4.15







| 1 | 28.207 | 49.02 | 28363296 |  |
|---|--------|-------|----------|--|
| 2 | 40.732 | 50.98 | 29499005 |  |





| peak | Ret. Time | Area%  | Area    |
|------|-----------|--------|---------|
| 1    | 5. 996    | 91. 19 | 3858241 |
| 2    | 8. 270    | 8. 81  | 372824  |














|    | $\sim$ |            |    |    |    |    |    |    |
|----|--------|------------|----|----|----|----|----|----|
|    |        | — <b>—</b> |    |    |    |    |    |    |
|    |        |            |    |    |    |    |    |    |
|    |        |            |    |    |    |    |    |    |
| 20 | 24     | 28         | 32 | 36 | 40 | 44 | 48 | 52 |
|    |        |            |    |    |    |    |    |    |
|    |        |            |    |    |    |    |    |    |

| peak | Ret. Time | Area% | Area     |  |
|------|-----------|-------|----------|--|
| 1    | 31.657    | 2.99  | 1937312  |  |
| 2    | 37.644    | 97.01 | 62741581 |  |



| Ret. lime | Area%            | Area   |
|-----------|------------------|--|
| 21.664    | 6.42             | 2335235  |
| 23.526    | 93.58            | 34048016   |
|           | 21.664<br>23.526 | Ret.         11me         Area%           21.664         6.42           23.526         93.58 |















|   |         |       | ni cu    |
|---|---------|-------|----------|
| 1 | 19.015  | 49.86 | 52380122 |
| 2 | 20. 481 | 50.14 | 52663792 |





## 9.X-Ray Crystal Data for Compounds 3h and 3i

Single crystal of **3h** was obtained by slow diffusion of ether into a chloroform solution at room temperature.

Single crystal diffraction data for **3h** was collected at 253 K. All single crystal diffraction data were collected using an I $\mu$ S micro-focus sealed X-ray tube with Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å) on a Bruker D8 venture Kappa Duo diffractometer equipped with a PHOTON 100 detector. Low-temperature holding was achieved by a Cryostream Cooler (Oxford Cryosystems). All the data were collected 0.5 degree per step and using the  $\omega$  scan mode. Frames were integrated using the Bruker SAINT software. Semiempirical absorption correction was applied with the SADABS program.



Table 1 Crystal data and structure refinement for TX15347\_auto.

| •                                     | —  |
|---------------------------------------|--|
| Identification code                   | TX15347_auto   |
| Empirical formula                     | C <sub>16</sub> H <sub>19</sub> N <sub>3</sub> O     |
| Formula weight                        | 269.34   |
| Temperature/K                         | 169.99(10)   |
| Crystal system                        | monoclinic   |
| Space group                           | I2   |
| a/Å                                   | 11.61718(18)   |
| b/Å                                   | 5.70808(9)   |
| c/Å                                   | 21.0321(3)   |
| $\alpha/^{\circ}$                     | 90   |
| β/°                                   | 101.6542(15)   |
| γ/°                                   | 90   |
| Volume/Å <sup>3</sup>                 | 1365.92(4)   |
| Ζ                                     | 4  |
| $ ho_{calc}g/cm^3$                    | 1.310  |
| µ/mm <sup>-1</sup>                    | 0.666  |
| F(000)                                | 576.0  |
| Crystal size/mm <sup>3</sup>          | $0.27 \times 0.25 \times 0.03$                       |
| Radiation                             | Cu Ka ( $\lambda = 1.54184$ )                        |
| $2\Theta$ range for data collection/° | 8.082 to 154.532                                     |
| Index ranges                          | $-14 \le h \le 14, -6 \le k \le 6, -24 \le l \le 25$ |
| Reflections collected                 | 8987   |
| Independent reflections               | $2662 \ [R_{int} = 0.0209, R_{sigma} = 0.0191]$      |
|                                       |  |

| Data/restraints/parameters                  | 2662/1/183                    |
|---|-------------------------------|
| Goodness-of-fit on F <sup>2</sup>           | 1.067                         |
| Final R indexes [I>=2 $\sigma$ (I)]         | $R_1 = 0.0338, wR_2 = 0.0936$ |
| Final R indexes [all data]                  | $R_1 = 0.0344, wR_2 = 0.0944$ |
| Largest diff. peak/hole / e Å <sup>-3</sup> | 0.33/-0.33                    |
| Flack parameter                             | -0.12(9)                      |

Single crystal of **3i** was obtained by slow diffusion of ether into a chloroform solution at room temperature.

Single crystal diffraction data for **3i** was collected at 253 K. All single crystal diffraction data were collected using an I $\mu$ S micro-focus sealed X-ray tube with Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å) on a Bruker D8 venture Kappa Duo diffractometer equipped with a PHOTON 100 detector. Low-temperature holding was achieved by a Cryostream Cooler (Oxford Cryosystems). All the data were collected 0.5 degree per step and using the  $\omega$  scan mode. Frames were integrated using the Bruker SAINT software. Semiempirical absorption correction was applied with the SADABS program.



**3i** (CCDC 2431788)

Table 1 Crystal data and structure refinement for fx1252.

| Identification code          | fx1252                       |
|------------------------------|------------------------------|
| Empirical formula            | $C_{21}H_{28}BN_3O_2$        |
| Formula weight               | 365.27                       |
| Temperature/K                | 170.00(10)                   |
| Crystal system               | orthorhombic                 |
| Space group                  | P212121                      |
| a/Å                          | 10.3685(2)                   |
| b/Å                          | 13.5410(4)                   |
| c/Å                          | 14.1637(3)                   |
| α/°                          | 90                           |
| β/°                          | 90                           |
| γ/°                          | 90                           |
| Volume/Å <sup>3</sup>        | 1988.58(8)                   |
| Ζ                            | 4                            |
| $\rho_{calc}g/cm^3$          | 1.220                        |
| µ/mm <sup>-1</sup>           | 0.618                        |
| F(000)                       | 784.0                        |
| Crystal size/mm <sup>3</sup> | $0.6 \times 0.3 \times 0.2$  |
| Radiation                    | Cu Ka ( $\lambda$ = 1.54184) |
|                              |                              |

| 20 range for data collection/°       | 9.036 to 151.94                                       |
|--------------------------------------|---|
| Index ranges                         | $-8 \le h \le 12, -16 \le k \le 17, -17 \le l \le 17$ |
| Reflections collected                | 12470   |
| Independent reflections              | $4002 \ [R_{int} = 0.0338, R_{sigma} = 0.0306]$       |
| Data/restraints/parameters           | 4002/0/249  |
| Goodness-of-fit on F <sup>2</sup>    | 1.065   |
| Final R indexes $[I \ge 2\sigma(I)]$ | $R_1 = 0.0387, wR_2 = 0.1055$                         |
| Final R indexes [all data]           | $R_1 = 0.0417, wR_2 = 0.1084$                         |
| Largest diff. peak/hole / e Å-3      | 0.22/-0.21  |
| Flack parameter                      | -0.09(11)   |
|                                      |   |