

Dimethylamino-Iodine(III)/PPh₃-Mediated Synthesis of α -Ketoamides from Glyoxylic Acids

Dan Xiao,^a Jiaxin He,^a Kaiyue Yang,^a Zhijian Wang,^a Cong Li,^a and Yunfei Du^{a*}

^a *Tianjin Key Laboratory for Modern Drug Delivery & High-Efficiency, School of Pharmaceutical Science and Technology, Faculty of Medicine, Tianjin University, Tianjin 300072, China.*

Corresponding author' email: duyunfeier@tju.edu.cn

Supporting Information

List of Contents

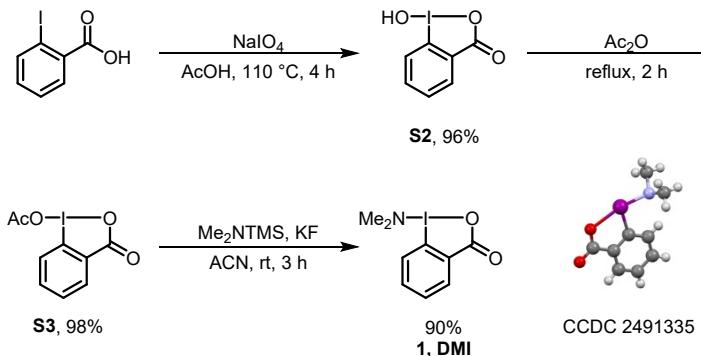
I.	General Information	S1
II.	General Procedure for the Preparation of DMI and Starting Materials	S2-S7
III.	General Procedure for Dimethylamination of Glyoxylic acids	S8-S19
IV.	General Procedure for Amination in One Pot	S20-S21
V.	Mechanistic Studies	S22-S23
VI.	X-Ray Diffraction Data of DMI	S24-S27
VII.	Reference	S28-S29
VIII.	NMR Spectra of Starting Materials and Products	S30-S129

I. General Information

¹H, ¹³C and ¹⁹F NMR spectra were recorded on a 400 MHz or 600 MHz spectrometer at 25 °C. Chemical shifts values are given in ppm and referred as the internal standard to TMS: 0.00 ppm. Chemical shifts were expressed in parts per million (δ) downfield from the internal standard tetramethylsilane, and were reported as s (singlet), d (doublet), t (triplet), q (quadruplet), dd (doublet of doublet), m (multiplet), etc. The coupling constants J , are reported in Hertz (Hz). High resolution mass spectrometry (HRMS) data were recorded on Q Exactive HF (Q Exactive™ HF/UltiMate™ 3000 RSLCnano) using electron spray ionization (ESI) in positive (or negative) mode. Melting points were determined with a Micromelting point apparatus. TLC plates were visualized by exposure to ultraviolet light.

Reagents and solvents were purchased as reagent grade and were used without further purification. All reactions were performed in standard glassware, heated at 70 °C for 3 h before used. Flash column chromatography was performed over silica gel (200-300 m) using a mixture of ethyl acetate (EA) and petroleum ether (PE).

II. General Procedure for the Preparation of DMI and Starting Materials



S3 was prepared according to the literature procedure.¹

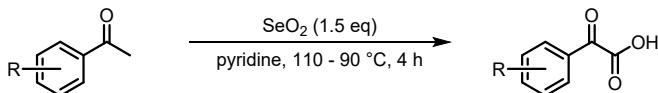
Preparation of S2: To a 500 mL round-bottomed flask equipped with a stirrer bar was added 2-iodobenzoic acid **S1** (8.0 g, 32.2 mmol, 1.0 equiv), 2-iodobenzamide NaIO₄ (7.24 g, 33.8 mmol, 1.0 equiv), and 30% (v:v) aq. AcOH (48 mL) under air. The mixture was vigorously stirred at 120 °C and refluxed. After stirring for 4 h, the reaction mixture was cooled to room temperature and diluted with cold water (180 mL), protecting it from light. The mixture was then filtered and further washed with ice water and cold acetone, air dried in the dark overnight to give the pure compound **S2** (8.5 g, 95%) as a white solid. It is a known compound¹ which is used directly for the next step.

Preparation of S3: To a 500 mL round-bottomed flask equipped with a stirrer bar was added compound **S2** (8.5 g, 32.2 mmol, 1.0 equiv) and acetic anhydride (30 mL). Then the reaction mixture was stirred and reflux at 110 °C. Until the solution turned clear (without suspension), reaction was cool down to room temperature and white crystals started to form. The crystallization was continued at -18 °C. The crystals were then collected and dried overnight under high vacuum to give compound **S3** (8.5 g, 86%) as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.25 (dd, *J* = 7.6, 1.6 Hz, 1H), 8.00 (dd, *J* = 8.3, 1.0 Hz, 1H), 7.96 – 7.90 (m, 1H), 7.71 (m, 1H), 2.25 (s, 3H). It is a known compound; the spectra data is in agreement with the reported one.¹

Preparation of DMI: To a 150 mL two-necked round-bottomed flask equipped with a stirrer bar were added compound **S3** (1.00 g, 3.28 mmol, 1.00 equiv), KF (19.1 mg, 0.33 mmol, 0.1 equiv) and ACN (30 mL) under N₂ atmosphere, followed by the careful addition of TMSNMe₂ (4.9 mol, 0.78 mL, 1.5 equiv). The reaction mixture was stirred at room temperature for 3 h in the dark. After the completion of the reaction, the precipitate was filtered and washed with Et₂O (5 mL x 3) to give product **DMI** (0.86 g) in 90% yield as a yellow solid; mp: 123-125 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.29 (dd, *J* = 7.5, 1.6 Hz, 1H), 7.93 (dd, *J* = 8.2, 1.0 Hz, 1H), 7.76 (m, 1H), 7.60 (td, *J* = 7.3, 1.0 Hz, 1H), 3.44 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.1, 133.5, 132.6, 132.6, 130.4, 125.2, 117.4, 77.2, 49.6. HRMS (ESI) calcd. for

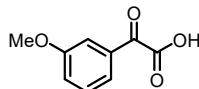
$C_9H_{11}INO_2^+$ [M + H⁺] 291.9756, found 291.9833.

Procedure A



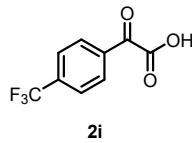
The α -keto acids were synthesized through the oxidation of corresponding methyl ketones with SeO₂ according to the literature procedure.²

In a dry, single-neck, 25-mL, round-bottom flask equipped with a stir bar and flushed with nitrogen, the substituted aryl-methylketone (5.0 mmol) and selenium dioxide (SeO₂, 0.835 g, 7.5 mmol, 1.5 equiv) were added followed by anhydrous pyridine (20 mL). The reaction mixture was heated in an oil bath to 110 °C for 1 h, and then the bath temperature was reduced to 90 °C. The mixture was stirred at this temperature (90 °C) for an additional 4 h, and progress of the reaction was monitored by TLC. After completion of the reaction, as determined by TLC, the solution containing precipitated selenium was filtered using a Buckner funnel, and the residue was washed with EA (50 mL). The combined filtrate was treated with 1N HCl (20 mL), the organic layer was separated, and the aqueous layer was extracted with EA (3 × 50 mL). The organic layers were combined and treated with 1N NaOH (50 mL), and the aqueous layer was separated. The organic layer was extracted with water (25 mL) and the combined aqueous layers were acidified using 1N HCl to about pH 1.5. The mixture was extracted with EA (3 × 50 mL), and the combined organic layers were dried (anhydrous Na₂SO₄) and concentrated on a rotary evaporator. The arylglyoxylic acid product was obtained and used directly for the next step.



2h

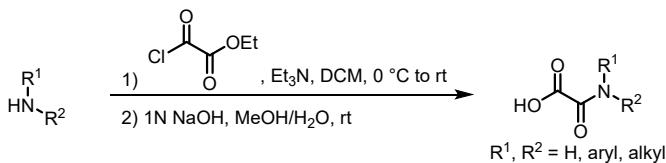
2-(3-Methoxyphenyl)-2-oxoacetic acid (**2h**) was obtained as a white solid (621 mg, 69% yield); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.57 – 7.48 (m, 2H), 7.41 (dd, *J* = 2.7, 1.5 Hz, 1H), 7.36 – 7.32 (m, 1H), 3.83 (s, 3H). It is a known compound; the spectra data is in agreement with the reported one.²



2i

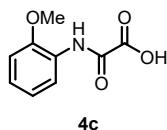
2-Oxo-2-(4-(trifluoromethyl)phenyl)acetic acid (**2i**) was obtained as a white solid (665 mg, 61% yield); ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.16 (d, J = 8.1 Hz, 2H), 7.96 (d, J = 7.7 Hz, 2H). It is a known compound; the spectra data is in agreement with the reported one.³

Procedure B

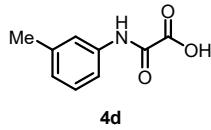


In a dry, single-neck, 25-mL, round-bottom flask equipped with a stir bar and flushed with nitrogen at 0 °C, to a solution of ethyl oxalyl chloride (1.0 g, 7.5 mmol, 1.5 equiv) in DCM (15 mL) was added a mixture of amino substrate (5.0 mmol) and Et_3N (0.76 g, 7.5 mmol, 1.5 equiv) in DCM (5 mL). The reaction mixture was stirred at room temperature for 5 h, and progress of the reaction was monitored by TLC. After completion of the reaction, as determined by TLC, the reaction was quenched by saturated aqueous NaHCO_3 at 0 °C and then was extracted with DCM (3×50 mL). The combined organic layers were dried (anhydrous Na_2SO_4) and concentrated on a rotary evaporator. The crude product was used directly for the next step.

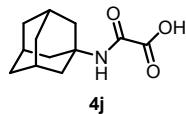
The crude product was dissolved in $\text{MeOH}/\text{H}_2\text{O}$ (20 mL, v/v = 1:1), followed by addition of aqueous 1N NaOH. The resulting mixture was stirred at room temperature for an additional 6 h and progress of the reaction was monitored by TLC. After completion of the reaction, as determined by TLC, the reaction was poured into H_2O (20 mL) and then extracted with EA (3×30 mL). The aqueous layer was acidified using 1N HCl to about pH 1.5. The mixture was extracted with EA (3×30 mL), and the combined organic layers were dried (anhydrous Na_2SO_4) and concentrated on a rotary evaporator. The arylglyoxylic acid product was obtained and used directly for the next step.



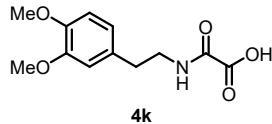
2-((2-Methoxyphenyl)amino)-2-oxoacetic acid (**4c**) was obtained as a white solid (741 mg, 76% yield over two steps); ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.62 (s, 1H), 8.07 (dt, J = 7.9, 1.2 Hz, 1H), 7.22 – 7.07 (m, 2H), 6.98 (td, J = 7.6, 1.6 Hz, 1H), 3.88 (s, 3H). ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 161.8, 155.6, 149.1, 125.7, 125.4, 120.5, 120.1, 111.2, 55.9. It is a known compound; the spectra data is in agreement with the reported one.⁴



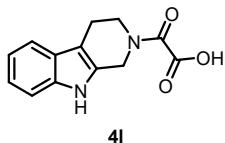
2-Oxo-2-(*m*-tolylamino)acetic acid (**4d**) was obtained as a yellow solid (653 mg, 73% yield over two steps); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.58 (s, 1H), 7.59 (d, *J* = 2.0 Hz, 1H), 7.56 – 7.51 (m, 1H), 7.22 (t, *J* = 7.8 Hz, 1H), 6.95 (d, *J* = 7.5 Hz, 1H), 2.29 (s, 3H). It is a known compound; the spectra data is in agreement with the reported one.⁴



2-((3s,5s,7s)-Adamantan-1-yl)amino)-2-oxoacetic acid (**4j**) was obtained as a yellow solid (792 mg, 71% yield over two steps); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.86 (s, 1H), 2.05 – 1.99 (m, 3H), 1.95 (d, *J* = 2.9 Hz, 6H), 1.62 (t, *J* = 3.0 Hz, 6H). HRMS (ESI) calcd. for C₁₂H₁₈NO₃⁺ [M + H⁺] 224.1208, found 224.1217.

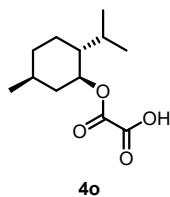


2-((3,4-Dimethoxyphenethyl)amino)-2-oxoacetic acid (**4k**) was obtained as a white solid (873 mg, 69% yield over two steps); ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.70 (t, *J* = 6.0 Hz, 1H), 6.84 (d, *J* = 8.2 Hz, 1H), 6.78 (d, *J* = 2.0 Hz, 1H), 6.69 (dd, *J* = 8.2, 2.0 Hz, 1H), 3.72 (d, *J* = 5.7 Hz, 6H), 3.40 – 3.29 (m, 2H), 2.71 (t, *J* = 7.3 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 159.8, 148.6, 147.2, 131.5, 120.4, 112.5, 111.9, 55.9, 55.3, 40.3, 34.1. HRMS (ESI) calcd. for C₁₂H₁₆NO₅⁺ [M + H⁺] 254.0950, found 254.0968.



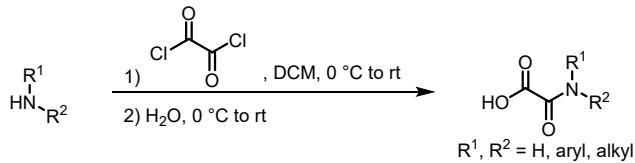
2-Oxo-2-(1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indol-2-yl)acetic acid (**4l**) was obtained as a yellow solid (952 mg, 78% yield over two steps); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.88 (s, 1H), 7.41 (dd, *J* = 7.8, 4.5 Hz, 1H), 7.33 (d, *J* = 8.0 Hz, 1H), 7.15 – 7.04 (m, 1H), 6.98 (td, *J* = 7.5, 2.3 Hz, 1H), 4.68 (m, 2H), 3.79 (m, 2H), 2.76 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 164.9, 164.8, 162.4, 161.9, 136.0, 136.0, 129.69,

129.6, 126.4, 126.4, 121.2, 121.1, 118.7, 118.7, 117.7, 117.6, 111.2, 111.2, 106.9, 106.3, 44.0, 43.5, 38.9, 21.5, 20.3. HRMS (ESI) calcd. for $C_{13}H_{13}N_2O_3^+ [M + H^+]$ 245.0848, found 245.0845.

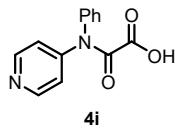


2-(((1S,2R,5S)-2-Isopropyl-5-methylcyclohexyl)oxy)-2-oxoacetic acid (**4o**) was obtained as a white solid (866 mg, 76% yield over two steps); 1H NMR (600 MHz, DMSO- d_6) δ 4.71 (td, J = 10.9, 4.4 Hz, 1H), 1.91 (dt, J = 11.8, 4.2 Hz, 1H), 1.80 (m, 1H), 1.65 (dt, J = 13.6, 3.3 Hz, 2H), 1.55 – 1.41 (m, 2H), 1.06 (m, 2H), 0.87 (dd, J = 14.6, 6.7 Hz, 7H), 0.73 (d, J = 7.0 Hz, 3H). ^{13}C NMR (151 MHz, DMSO- d_6) δ 159.7, 159.0, 75.9, 46.2, 33.5, 30.8, 25.9, 23.1, 21.7, 20.3, 16.2. HRMS (ESI) calcd. for $C_{12}H_{21}O_4^+ [M + H^+]$ 229.1362, found 229.1378.

Procedure C

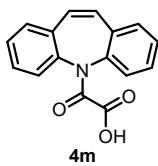


In a dry, single-neck, 25-mL, round-bottom flask equipped with a stir bar and flushed with nitrogen at 0 °C, to a solution of oxalyl chloride (0.95 g, 7.5 mmol, 1.5 equiv) in DCM (15 mL) was added a solution of amino substrate (5.0 mmol) in DCM (5 mL). The reaction mixture was stirred at room temperature for 2 h, and progress of the reaction was monitored by TLC. After completion of the reaction, as determined by TLC, the reaction was quenched by saturated H_2O (1 mL) at 0 °C. The mixture was then filtered and further washed with ice water, air dried overnight to give the desired compound.

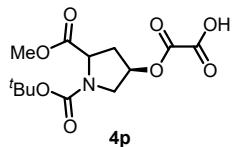


2-Oxo-2-(phenyl(pyridin-4-yl)amino)acetic acid (**4i**) was obtained as a yellow solid (799 mg, 66% yield over two steps); 1H NMR (600 MHz, DMSO- d_6) δ 8.83 – 8.79 (m, 2H), 7.77 – 7.72 (m, 2H), 7.62 – 7.57 (m, 3H), 7.55 (dd, J = 8.0, 1.9 Hz, 2H). ^{13}C NMR (151 MHz, DMSO- d_6) δ 163.2, 162.0, 153.7, 143.1,

136.8, 130.4, 130.3, 129.5, 118.9. HRMS (ESI) calcd. for $C_{13}H_{11}N_2O_3^+$ [M + H⁺] 243.0691, found 243.0709.

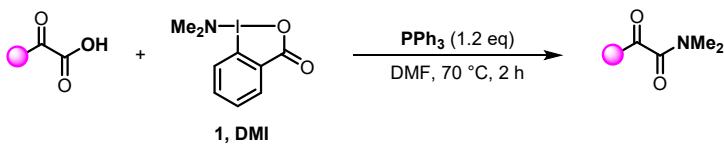


2-(5H-Dibenzo[b,f]azepin-5-yl)-2-oxoacetic acid (**4m**) was obtained as a white solid (1.02 g, 77% yield over two steps); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.50 (m, 6H), 7.45 – 7.39 (m, 2H), 7.06 (s, 2H). HRMS (ESI) calcd. for $C_{16}H_{12}NO_3^+$ [M + H⁺] 266.0739, found 266.0751.

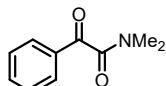


2-(((3*R*)-1-(*tert*-Butoxycarbonyl)-5-(methoxycarbonyl)pyrrolidin-3-yl)oxy)-2-oxoacetic acid (**4p**) was obtained as a white solid (919 mg, 58% yield over two steps); ¹H NMR (600 MHz, DMSO-*d*₆) δ 5.32 (dt, *J* = 7.0, 2.4 Hz, 1H), 4.32 – 4.21 (m, 1H), 3.67 (d, *J* = 12.4 Hz, 3H), 3.53 (dq, *J* = 12.4, 3.3, 2.5 Hz, 1H), 2.49 – 2.41 (m, 1H), 2.22 (m, 1H), 1.37 (d, *J* = 32.7 Hz, 9H), 1.10 (dd, *J* = 13.1, 6.2 Hz, 1H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 172.6, 172.1, 158.8, 158.7, 153.4, 152.7, 79.7, 79.6, 74.6, 74.0, 64.9, 57.3, 57.0, 52.0, 52.0, 51.6, 51.4, 35.4, 34.5, 27.9, 27.8, 27.8, 15.1. HRMS (ESI) calcd. for $C_{13}H_{20}NO_8^+$ [M + H⁺] 318.1111, found 318.1156.

III. General Procedure for Dimethylamination of Glyoxylic Acids

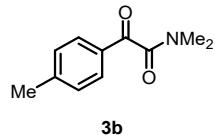


Glyoxylic acids (0.4 mmol), **DMI** (140 mg, 0.48 mmol, 1.2 equiv) and PPh_3 (126 mg, 0.48 mmol, 1.2 equiv) were placed in a 10 ml two-neck rounded bottom flask, and DMF (2.0 mL) was added. The reaction flask was placed in an oil bath and heated up to 70 °C. After substrate was fully consumed, the reaction was quenched with H_2O (10 mL) and extracted with EA (20 mL x 3). The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated in vacuo to afford the crude product, which was then purified by silica gel flash chromatography to give the corresponding products.

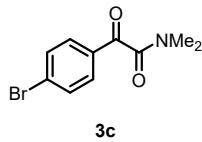


3a

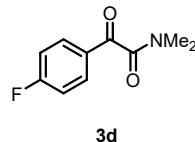
N,N-Dimethyl-2-oxo-2-phenylacetamide (**3a**) was isolated by column chromatography (PE/EA = 4:1) as a yellow oil (58 mg, 82% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.95 – 7.86 (m, 2H), 7.63 – 7.57 (m, 1H), 7.50 – 7.44 (m, 2H), 3.07 (s, 3H), 2.91 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 191.7, 166.9, 134.6, 132.9, 129.5, 128.9, 36.9, 33.8. It is a known compound; the spectra data is in agreement with the reported one.⁵



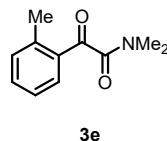
N,N-Dimethyl-2-oxo-2-(*p*-tolyl)acetamide (**3b**) was isolated by column chromatography (PE/EA = 4:1) as a yellow oil (63 mg, 83% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.85 – 7.81 (m, 2H), 7.31 – 7.28 (m, 2H), 3.10 (s, 3H), 2.94 (s, 3H), 2.43 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 191.5, 167.2, 145.9, 130.7, 129.8, 37.0, 33.9, 21.9. It is a known compound; the spectra data is in agreement with the reported one.⁶



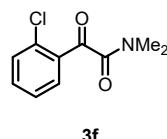
2-(4-Bromophenyl)-*N,N*-dimethyl-2-oxoacetamide (**3c**) was isolated by column chromatography (PE/EA = 3:1) as a yellow solid (72 mg, 70% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 8.6 Hz, 2H), 7.64 (d, *J* = 8.6 Hz, 2H), 3.10 (s, 3H), 2.95 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.5, 166.4, 132.4, 131.9, 131.0, 130.1, 37.0, 34.1. It is a known compound; the spectra data is in agreement with the reported one.⁵



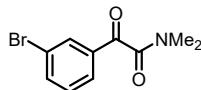
2-(4-Fluorophenyl)-*N,N*-dimethyl-2-oxoacetamide (**3d**) was isolated by column chromatography (PE/EA = 3:1) as a yellow oil (58 mg, 74% yield); ¹H NMR (400 MHz, CDCl₃) δ 8.10 – 7.90 (m, 2H), 7.17 (t, *J* = 8.6 Hz, 2H), 3.10 (s, 3H), 2.95 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.0, 167.5, 166.6, 165.8, 132.5, 132.4, 129.7, 116.4, 116.2, 37.0, 34.0. It is a known compound; the spectra data is in agreement with the reported one.⁵



N,N-Dimethyl-2-oxo-2-(o-tolyl)acetamide (**3e**) was isolated by column chromatography (PE/EA = 4:1) as a yellow oil (60 mg, 79% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.70 – 7.66 (m, 1H), 7.46 (dd, *J* = 7.5, 1.4 Hz, 1H), 7.30 (t, *J* = 7.4 Hz, 2H), 3.10 (s, 3H), 2.97 (s, 3H), 2.65 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 193.7, 167.8, 141.4, 133.6, 132.6, 132.5, 131.6, 126.1, 37.0, 34.0, 21.6. It is a known compound; the spectra data is in agreement with the reported one.⁶

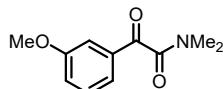


2-(2-Chlorophenyl)-*N,N*-dimethyl-2-oxoacetamide (**3f**) was isolated by column chromatography (PE/EA = 3:1) as a yellow oil (63 mg, 75% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.70 (dd, *J* = 1.7, 0.8 Hz, 1H), 7.36 (dd, *J* = 3.7, 0.8 Hz, 1H), 6.59 (dd, *J* = 3.6, 1.7 Hz, 1H), 3.07 (s, 3H), 3.02 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.1, 166.9, 134.3, 133.7, 133.4, 132.2, 130.7, 127.2, 37.0, 34.5. It is a known compound; the spectra data is in agreement with the reported one.⁵



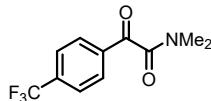
3g

2-(3-Bromophenyl)-*N,N*-dimethyl-2-oxoacetamide (**3g**) was isolated by column chromatography (PE/EA = 3:1) as a yellow solid (76 mg, 74% yield); ¹H NMR (400 MHz, CDCl₃) δ 8.08 (t, *J* = 1.8 Hz, 1H), 7.87 (dt, *J* = 7.7, 1.4 Hz, 1H), 7.75 (m, 1H), 7.38 (t, *J* = 7.9 Hz, 1H), 3.11 (s, 3H), 2.96 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.1, 166.2, 137.5, 134.9, 132.4, 130.5, 128.3, 123.2, 37.0, 34.1. It is a known compound; the spectra data is in agreement with the reported one.⁵



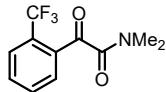
3h

2-(3-Methoxyphenyl)-*N,N*-dimethyl-2-oxoacetamide (**3h**) was isolated by column chromatography (PE/EA = 2:1) as a white solid (58 mg, 70% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.43 (m, 2H), 7.39 (t, *J* = 8.1 Hz, 1H), 7.16 (m, 1H), 3.84 (s, 3H), 3.09 (s, 3H), 2.93 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 191.6, 167.0, 160.1, 134.4, 130.0, 122.7, 121.5, 112.8, 55.5, 37.0, 33.9. It is a known compound; the spectra data is in agreement with the reported one.⁷



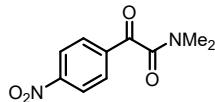
3i

N,N-Dimethyl-2-oxo-2-(4-(trifluoromethyl)phenyl)acetamide (**3i**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (89 mg, 91% yield); ¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 8.0 Hz, 2H), 7.79 – 7.71 (m, 2H), 3.13 (d, *J* = 2.2 Hz, 3H), 2.97 (d, *J* = 2.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.3, 166.2, 136.1, 136.0, 135.9, 135.7, 130.1, 126.1, 126.1, 126.1, 124.4, 37.1, 34.3. It is a known compound; the spectra data is in agreement with the reported one.⁶



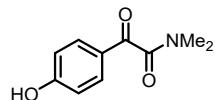
3j

N,N-Dimethyl-2-oxo-2-(4-(trifluoromethyl)phenyl)acetamide (**3j**) was isolated by column chromatography (PE/EA = 3:1) as a yellow oil (82 mg, 84% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.79 (m, 2H), 7.70 – 7.60 (m, 2H), 3.08 (d, J = 6.9 Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 190.4, 165.6, 134.4, 132.4, 131.9, 131.5, 127.3, 127.2, 127.1, 127.1, 124.7, 121.9, 37.0, 34.7. It is a known compound; the spectra data is in agreement with the reported one.⁶



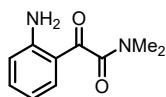
3k

N,N-Dimethyl-2-(4-nitrophenyl)-2-oxoacetamide (**3k**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (80 mg, 90% yield); ^1H NMR (400 MHz, CDCl_3) δ 8.34 (d, J = 8.8 Hz, 2H), 8.14 (d, J = 8.8 Hz, 2H), 3.15 (s, 3H), 3.00 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 189.2, 165.6, 151.1, 137.6, 130.8, 124.1, 77.2, 37.1, 34.3. It is a known compound; the spectra data is in agreement with the reported one.⁵



3l

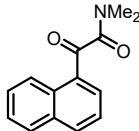
2-(4-Hydroxyphenyl)-*N,N*-dimethyl-2-oxoacetamide (**3l**) was isolated by column chromatography (PE/EA = 1:2) as a yellow oil (39 mg, 51% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.86 (s, 1H), 7.81 (d, J = 8.6 Hz, 2H), 6.97 (d, J = 8.6 Hz, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 191.3, 161.8, 132.6, 130.1, 116.2, 29.8, 1.2. HRMS (ESI) calcd. for $\text{C}_{10}\text{H}_{12}\text{NO}_3^+$ [M + H $^+$] 194.0739, found 194.0744.



3m

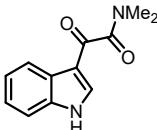
2-(2-Aminophenyl)-*N,N*-dimethyl-2-oxoacetamide (**3m**) was isolated by column chromatography (PE/EA

= 1:3) as a brown oil (53 mg, 69% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.42 (dd, J = 8.1, 1.6 Hz, 1H), 7.30 (m, 1H), 6.71 – 6.61 (m, 2H), 3.10 (s, 3H), 2.95 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 194.3, 167.4, 151.7, 136.0, 133.3, 117.2, 116.4, 114.3, 37.2, 34.0, 1.1. It is a known compound; the spectra data is in agreement with the reported one.⁸



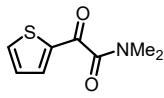
3n

N,N-Dimethyl-2-(naphthalen-1-yl)-2-oxoacetamide (**3n**) was isolated by column chromatography (PE/EA = 3:1) as a yellow oil (74 mg, 81% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.29 – 9.22 (m, 1H), 8.11 (d, J = 8.2 Hz, 1H), 7.99 (dd, J = 7.2, 1.2 Hz, 1H), 7.94 – 7.90 (m, 1H), 7.76 – 7.68 (m, 1H), 7.62 – 7.58 (m, 1H), 7.54 (dd, J = 8.1, 7.2 Hz, 1H), 3.16 (s, 3H), 3.02 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 194.2, 167.6, 135.9, 134.3, 134.1, 131.0, 129.3, 128.7, 128.5, 127.0, 125.8, 124.5, 37.2, 34.1. It is a known compound; the spectra data is in agreement with the reported one.⁵



3o

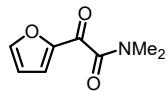
2-(1*H*-Indol-3-yl)-*N,N*-dimethyl-2-oxoacetamide (**3o**) was isolated by column chromatography (PE/EA = 1:1) as a yellow oil (66 mg, 76% yield); ^1H NMR (400 MHz, CDCl_3) δ 10.28 – 10.12 (m, 1H), 8.30 (d, J = 7.8 Hz, 1H), 7.71 (d, J = 3.1 Hz, 1H), 7.33 (dd, J = 8.1, 1.1 Hz, 1H), 7.29 (td, J = 7.6, 1.1 Hz, 1H), 7.25 – 7.22 (m, 1H), 3.07 (s, 3H), 3.02 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 186.2, 168.2, 136.9, 135.9, 125.4, 124.3, 123.3, 122.0, 114.4, 112.3, 37.7, 34.6. It is a known compound; the spectra data is in agreement with the reported one.⁹



3p

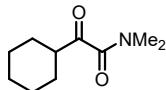
N,N-Dimethyl-2-oxo-2-(thiophen-2-yl)acetamide (**3p**) was isolated by column chromatography (PE/EA = 2:1) as a brown oil (66 mg, 86% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.81 (dd, J = 3.8, 1.2 Hz, 1H), 7.78

(dd, $J = 4.9, 1.2$ Hz, 1H), 7.17 (dd, $J = 4.9, 3.9$ Hz, 1H), 3.09 (s, 3H), 3.03 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 183.5, 165.8, 140.3, 136.4, 136.1, 128.6, 77.2, 37.3, 34.5. It is a known compound; the spectra data is in agreement with the reported one.⁶



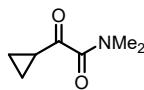
3q

2-(Furan-2-yl)-*N,N*-dimethyl-2-oxoacetamide (**3q**) was isolated by column chromatography (PE/EA = 1:1) as a yellow oil (52 mg, 78% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.70 (dd, $J = 1.7, 0.8$ Hz, 1H), 7.36 (dd, $J = 3.7, 0.8$ Hz, 1H), 6.59 (dd, $J = 3.6, 1.7$ Hz, 1H), 3.07 (s, 3H), 3.02 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 178.5, 165.4, 150.2, 148.6, 122.2, 112.8, 37.2, 34.5. It is a known compound; the spectra data is in agreement with the reported one.⁵



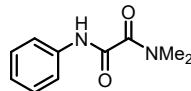
3r

2-Cyclohexyl-*N,N*-dimethyl-2-oxoacetamide (**3r**) was isolated by column chromatography (PE/EA = 6:1) as a yellow oil (49 mg, 67% yield); ^1H NMR (400 MHz, CDCl_3) δ 2.93 (d, $J = 12.7$ Hz, 7H), 1.93 – 1.84 (m, 2H), 1.80 – 1.70 (m, 2H), 1.68 – 1.60 (m, 1H), 1.32 – 1.12 (m, 5H). ^{13}C NMR (101 MHz, CDCl_3) δ 203.8, 167.4, 46.7, 36.8, 34.2, 27.0, 25.7, 25.2. HRMS (ESI) calcd. for $\text{C}_{10}\text{H}_{18}\text{NO}_2^+ [\text{M} + \text{H}^+]$ 184.1259, found 184.1257.



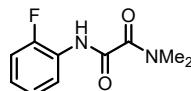
3s

2-Cyclopropyl-*N,N*-dimethyl-2-oxoacetamide (**3s**) was isolated by column chromatography (PE/EA = 6:1) as a yellow oil (49 mg, 67% yield); ^1H NMR (400 MHz, CDCl_3) δ 3.00 (d, $J = 1.8$ Hz, 6H), 2.35 (tt, $J = 7.9, 4.6$ Hz, 1H), 1.25 – 1.21 (m, 2H), 1.13 – 1.08 (m, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 200.9, 167.0, 37.1, 34.5, 19.5, 12.6. It is a known compound; the spectra data is in agreement with the reported one.¹⁰



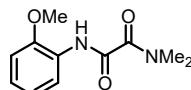
5a

N,N-Dimethyl-*N'*-phenyloxalamide (**5a**) was isolated by column chromatography (PE/EA = 1:1) as a yellow oil (48 mg, 63% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.24 (s, 1H), 7.60 (d, J = 8.0 Hz, 2H), 7.36 (t, J = 7.9 Hz, 2H), 7.16 (t, J = 7.4 Hz, 1H), 3.52 (s, 3H), 3.09 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.5, 158.4, 136.9, 129.1, 125.0, 119.8, 38.8, 37.9. It is a known compound; the spectra data is in agreement with the reported one.¹¹



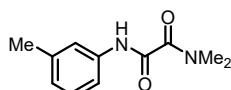
5b

N-(2-Fluorophenyl)-*N',N'*-dimethyloxalamide (**5b**) was isolated by column chromatography (PE/EA = 1:1) as a yellow oil (51 mg, 61% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.45 (s, 1H), 8.29 (td, J = 7.5, 1.4 Hz, 1H), 7.12 (m, 3H), 3.49 (s, 3H), 3.09 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.1, 158.5, 154.0, 151.6, 125.4, 125.3, 125.2, 124.5, 124.4, 121.3, 115.2, 115.0, 77.2, 38.7, 37.7. HRMS (ESI) calcd. for $\text{C}_{10}\text{H}_{12}\text{FN}_2\text{O}_2^+ [\text{M} + \text{H}^+]$ 211.0805, found 211.0809.



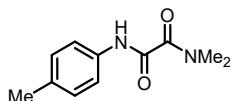
5c

N-(2-Methoxyphenyl)-*N',N'*-dimethyloxalamide (**5c**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (56 mg, 63% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.66 (s, 1H), 8.33 (dd, J = 8.1, 1.6 Hz, 1H), 7.07 (td, J = 7.8, 1.6 Hz, 1H), 6.94 (td, J = 7.8, 1.3 Hz, 1H), 6.87 (dd, J = 8.1, 1.4 Hz, 1H), 3.85 (s, 3H), 3.45 (s, 3H), 3.05 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 1161.9, 158.5, 148.7, 126.6, 124.8, 120.9, 119.6, 110.2, 55.7, 38.7, 37.5. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_3^+ [\text{M} + \text{H}^+]$ 223.1004, found 223.1009.



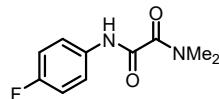
5d

N,N-Dimethyl-*N'*-(*m*-tolyl)oxalamide (**5d**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (58 mg, 70% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.28 (s, 1H), 7.48 – 7.44 (m, 1H), 7.41 – 7.34 (m, 1H), 7.22 (t, J = 7.8 Hz, 1H), 6.96 (m, 1H), 3.48 (s, 3H), 3.07 (s, 3H), 2.34 (d, J = 0.8 Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.8, 158.6, 139.1, 136.9, 129.0, 125.9, 120.6, 117.1, 77.4, 38.9, 37.8, 21.6. HRMS (ESI) calcd. for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_2^+$ [M + H $^+$] 207.1055, found 207.1061.



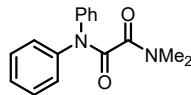
5d

N,N-Dimethyl-*N'*-(*p*-tolyl)oxalamide (**5e**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (49 mg, 60% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.29 (s, 1H), 7.52 – 7.45 (m, 2H), 7.14 (d, J = 8.1 Hz, 2H), 3.48 (s, 3H), 3.06 (s, 3H), 2.32 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.8, 158.4, 134.6, 134.3, 129.5, 119.8, 38.7, 37.6, 20.9. It is a known compound; the spectra data is in agreement with the reported one.¹¹



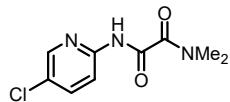
5e

N-(4-Fluorophenyl)-*N,N*-dimethyloxalamide (**5f**) was isolated by column chromatography (PE/EA = 1:1) as a brown oil (49 mg, 60% yield); ^1H NMR (400 MHz, CDCl_3) δ 9.30 (s, 1H), 7.60 – 7.53 (m, 2H), 7.09 – 7.00 (m, 2H), 3.51 (s, 3H), 3.09 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 161.4, 160.9, 158.3, 132.9, 132.9, 121.6, 121.5, 115.9, 115.7, 77.2, 38.8, 37.9. HRMS (ESI) calcd. for $\text{C}_{10}\text{H}_{12}\text{FN}_2\text{O}_2^+$ [M + H $^+$] 211.0805, found 211.0810.



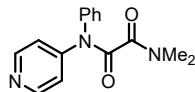
5f

N,N-Dimethyl-*N,N*-diphenyloxalamide (**5g**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (81 mg, 76% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.36 (m, 6H), 7.32 – 7.22 (m, 4H), 3.09 (s, 3H), 2.74 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 129.5, 129.1, 128.4, 127.8, 126.8, 125.9, 37.2, 33.5. HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{17}\text{N}_2\text{O}_2^+$ [M + H $^+$] 269.1212, found 269.1213.



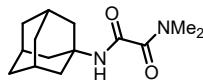
5h

N-(5-Chloropyridin-2-yl)-*N,N*'-dimethyloxalamide (**5h**) was isolated by column chromatography (PE/EA = 1:1) as a yellow oil (75 mg, 83% yield); ¹H NMR (400 MHz, CDCl₃) δ 9.79 (s, 1H), 8.29 (d, *J* = 2.6 Hz, 1H), 8.17 (d, *J* = 8.9 Hz, 1H), 7.68 (dd, *J* = 8.8, 2.6 Hz, 1H), 3.44 (s, 3H), 3.07 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 161.0, 159.1, 148.8, 147.1, 138.0, 127.6, 114.6, 38.7, 37.7. HRMS (ESI) calcd. for C₉H₁₁ClN₃O₂⁺ [M + H⁺] 228.0462, found 228.0465.



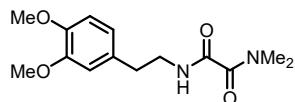
5i

N,N-Dimethyl-*N'*-phenyl-*N*-(pyridin-4-yl)oxalamide (**5i**) was isolated by column chromatography (PE/EA = 1:2) as a yellow oil (77 mg, 72% yield); ¹H NMR (400 MHz, CDCl₃) δ 8.55 (s, 2H), 7.51 – 7.42 (m, 3H), 7.30 (dt, *J* = 8.4, 2.8 Hz, 4H), 3.10 – 3.02 (m, 3H), 2.73 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 164.5, 163.1, 150.5, 148.3, 129.9, 37.2, 33.6. HRMS (ESI) calcd. for C₁₅H₁₆N₃O₂⁺ [M + H⁺] 270.1164, found 270.1168.



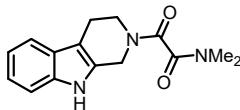
5j

N-((3s,5s,7s)-Adamantan-1-yl)-*N,N*'-dimethyloxalamide (**5j**) was isolated by column chromatography (PE/EA = 4:1) as a yellow oil (80 mg, 80% yield); ¹H NMR (400 MHz, CDCl₃) δ 6.99 (s, 1H), 3.35 (d, *J* = 1.7 Hz, 3H), 2.96 (s, 3H), 2.10 – 2.04 (m, 3H), 2.00 (d, *J* = 3.1 Hz, 6H), 1.66 (t, *J* = 3.1 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 162.7, 160.2, 52.0, 41.0, 38.5, 37.1, 36.2, 29.2. HRMS (ESI) calcd. for C₁₄H₂₃N₂O₂⁺ [M + H⁺] 251.1681, found 251.1683.



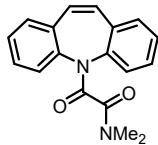
5k

N-(3,4-Dimethoxyphenethyl)-*N,N*'-dimethyloxalamide (**5k**) was isolated by column chromatography (PE/EA = 1:2) as a white oil (90 mg, 80% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.33 (s, 1H), 6.80 (d, *J* = 8.0 Hz, 1H), 6.77 – 6.70 (m, 2H), 3.86 (d, *J* = 11.3 Hz, 6H), 3.52 (q, *J* = 6.9 Hz, 2H), 3.37 (s, 3H), 2.99 (s, 3H), 2.79 (t, *J* = 7.1 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 162.0, 161.1, 149.0, 147.7, 130.9, 120.6, 111.8, 111.4, 55.9, 55.8, 40.6, 38.3, 37.0, 35.0, 29.7. HRMS (ESI) calcd. for C₁₄H₂₁N₂O₄⁺ [M + H⁺] 281.1423, found 281.1413.



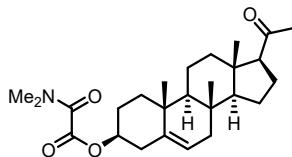
5l

N,N-Dimethyl-2-oxo-2-(1,3,4,9-tetrahydro-2*H*-pyrido[3,4-*b*]indol-2-yl)acetamide (**5l**) was isolated by column chromatography (PE/EA = 1:1) as a brown oil (93 mg, 86% yield); ¹H NMR (400 MHz, CDCl₃) δ 8.26 (s, 1H), 7.68 (m, 1H), 7.50 – 7.42 (m, 2H), 7.33 (t, *J* = 7.6 Hz, 1H), 7.17 (t, *J* = 7.6 Hz, 1H), 7.11 (m, 1H), 4.85 (t, *J* = 1.5 Hz, 2H), 4.60 (q, *J* = 1.5 Hz, 1H), 3.99 – 3.95 (m, 1H), 3.79 – 3.73 (m, 2H), 3.06 (s, 3H), 3.03 (s, 3H), 3.02 (s, 1H), 2.96 (d, *J* = 1.0 Hz, 2H), 2.89 – 2.80 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 1164.8, 164.6, 162.6, 136.3, 132.1, 132.1, 132.0, 128.6, 128.5, 126.7, 122.2, 122.1, 119.8, 118.1, 118.0, 111.1, 107.9, 44.5, 44.1, 39.6, 39.5, 37.3, 36.5, 33.9, 33.8, 29.7, 22.0, 20.8. HRMS (ESI) calcd. for C₁₅H₁₈N₃O₂⁺ [M + H⁺] 272.1321, found 272.1333.



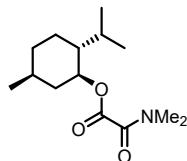
5m

N,N-Dimethyl-2-oxo-2-(1,3,4,9-tetrahydro-2*H*-pyrido[3,4-*b*]indol-2-yl)acetamide (**5m**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (85 mg, 73% yield); ¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.56 (m, 1H), 7.53 – 7.47 (m, 2H), 7.43 – 7.32 (m, 5H), 7.01 – 6.91 (m, 2H), 2.81 (s, 3H), 2.65 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 165.4, 163.9, 137.9, 137.6, 134.8, 133.5, 131.2, 129.7, 129.7, 129.6, 129.4, 129.0, 128.5, 128.2, 128.1, 37.2, 33.3. HRMS (ESI) calcd. for C₁₈H₁₇N₂O₂⁺ [M + H⁺] 293.1212, found 293.1243.



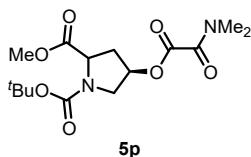
5n

(3*S*,8*S*,9*S*,10*R*,13*S*,14*S*,17*S*)-17-Acetyl-8,10,13-trimethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[a]phenanthren-3-yl 2-(dimethylamino)-2-oxoacetate (**5n**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (111 mg, 65% yield); ¹H NMR (400 MHz, CDCl₃) δ 5.44 – 5.40 (m, 1H), 4.86 – 4.76 (m, 1H), 3.02 (s, 3H), 2.98 (d, *J* = 1.6 Hz, 3H), 2.54 (t, *J* = 9.0 Hz, 1H), 2.47 – 2.41 (m, 2H), 2.22 – 2.15 (m, 1H), 2.13 (d, *J* = 1.6 Hz, 3H), 2.08 – 1.96 (m, 3H), 1.92 (dd, *J* = 13.5, 3.6 Hz, 1H), 1.76 – 1.65 (m, 3H), 1.62 (td, *J* = 11.5, 10.4, 5.5 Hz, 2H), 1.53 – 1.44 (m, 3H), 1.28 – 1.20 (m, 2H), 1.20 – 1.13 (m, 2H), 1.03 (s, 3H), 0.64 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 209.4, 139.0, 123.0, 75.9, 63.6, 56.8, 49.8, 43.9, 38.7, 37.7, 37.0, 36.9, 36.6, 34.0, 31.8, 31.7, 31.5, 27.4, 24.4, 22.8, 21.0, 19.2, 13.2. HRMS (ESI) calcd. for C₂₆H₄₀NO₄⁺ [M + H⁺] 430.2879, found 430.2885.



5o

(1*S*,2*R*,5*S*)-2-Isopropyl-5-methylcyclohexyl 2-(dimethylamino)-2-oxoacetate (**5o**) was isolated by column chromatography (PE/EA = 4:1) as a yellow oil (93 mg, 91% yield); ¹H NMR (400 MHz, CDCl₃) δ 4.84 (td, *J* = 10.9, 4.4 Hz, 1H), 2.96 (d, *J* = 11.1 Hz, 6H), 2.06 (dd, *J* = 11.5, 4.2 Hz, 1H), 1.93 (m, 1H), 1.74 – 1.64 (m, 2H), 1.55 – 1.41 (m, 2H), 1.15 – 1.00 (m, 2H), 0.89 (dd, *J* = 10.1, 6.8 Hz, 7H), 0.76 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 162.9, 162.0, 76.3, 46.7, 40.3, 36.9, 33.9, 33.8, 31.4, 25.9, 23.1, 21.8, 20.6, 15.9. HRMS (ESI) calcd. for C₁₄H₂₆NO₃⁺ [M + H⁺] 256.1834, found 256.1701.

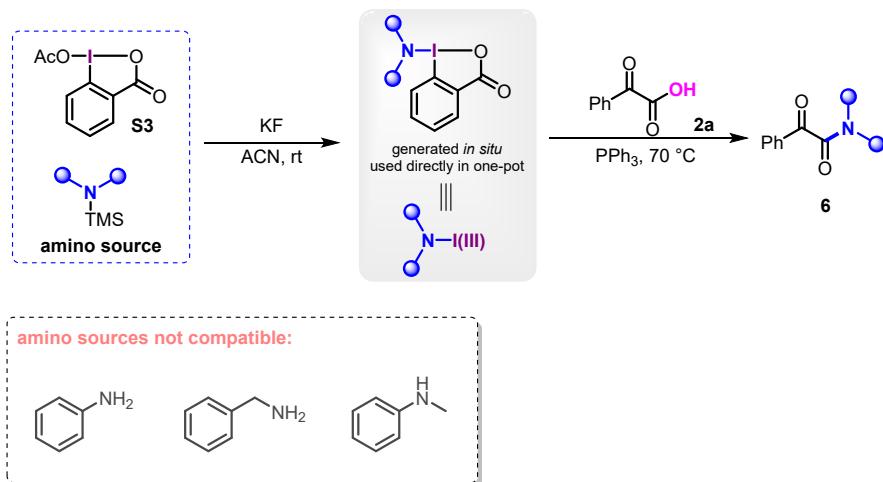


5p

1-(*tert*-Butyl) 2-methyl (4*R*)-4-(2-(dimethylamino)-2-oxoacetoxy)pyrrolidine-1,2-dicarboxylate (**5p**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (116 mg, 84% yield); ¹H NMR (400 MHz, CDCl₃) δ 5.43 (dd, *J* = 5.7, 2.9 Hz, 1H), 4.41 (dt, *J* = 34.2, 7.9 Hz, 1H), 3.78 – 3.65 (m, 5H), 2.97 (d, *J* = 5.6 Hz, 6H), 2.55 – 2.40 (m, 1H), 2.26 (m, 1H), 1.41 (d, *J* = 23.6 Hz, 9H). ¹³C NMR (101 MHz,

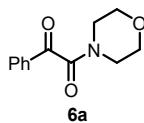
CDCl₃) δ 172.8, 172.6, 162.3, 162.2, 160.9, 80.7, 74.5, 73.7, 57.6, 57.3, 52.4, 52.2, 51.8, 51.7, 37.0, 36.9, 36.3, 35.3, 34.1, 29.6, 28.3, 28.2, 28.1. HRMS (ESI) calcd. for C₁₅H₂₅N₂O₇⁺ [M + H⁺] 345.1584, found 345.1576.

IV. General Procedure for Amination in One Pot

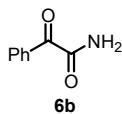


Scheme S1. Introduction of diverse amino functional groups in one-pot protocol.

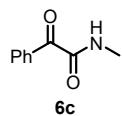
To a 50 mL two-necked round-bottomed flask equipped with a stirrer bar were added compound **S3** (100 mg, 0.33 mmol, 1.00 equiv), KF (1.9 mg, 0.033 mmol, 0.1 equiv) and ACN (5 mL) under N_2 atmosphere, followed by the careful addition of amino source (0.49 mmol, 1.5 equiv). The reaction mixture was stirred at room temperature for 3 h in the dark. Then benzoylformic acid (0.3 mmol) and PPh_3 (94 mg, 0.36 mmol, 1.2 equiv) were added to the reaction mixture. The reaction flask was placed in an oil bath and heated up to 70 °C. After substrate was fully consumed, the reaction was quenched with H_2O (10 mL) and extracted with EA (15 mL x 3). The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated in vacuo to afford the crude product, which was then purified by silica gel flash chromatography to give the corresponding products.



1-Morpholino-2-phenylethane-1,2-dione (**6a**) was isolated by column chromatography (PE/EA = 4:1) as a colorless oil (56 mg, 85% yield); ^1H NMR (400 MHz, CDCl_3) δ 7.98 – 7.90 (m, 2H), 7.64 (td, J = 7.5, 1.4 Hz, 1H), 7.54 – 7.48 (m, 2H), 3.82 – 3.74 (m, 4H), 3.67 – 3.58 (m, 2H), 3.40 – 3.33 (m, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 191.2, 165.5, 135.0, 133.1, 129.7, 129.2, 66.8, 66.8, 46.3, 41.7. It is a known compound; the spectra data is in agreement with the reported one.¹²

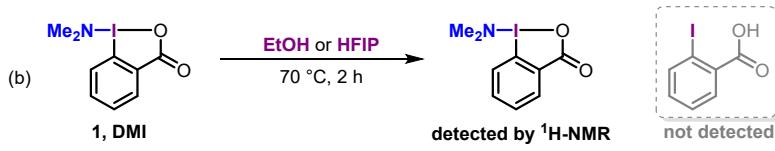
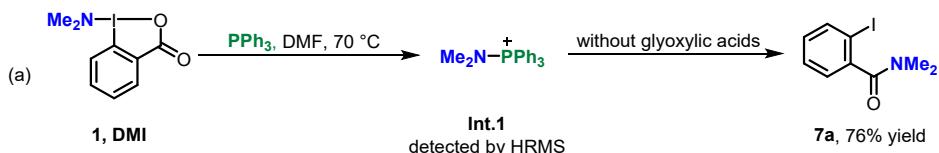


2-Oxo-2-phenylacetamide (**6b**) was isolated by column chromatography (PE/EA = 2:1) as a yellow oil (27 mg, 61% yield); ^1H NMR (400 MHz, CDCl_3) δ 8.31 (dd, J = 8.4, 1.4 Hz, 2H), 7.67 – 7.61 (m, 1H), 7.49 (dd, J = 8.5, 7.2 Hz, 2H), 6.96 (s, 1H), 5.82 (s, 1H). It is a known compound; the spectra data is in agreement with the reported one.¹³

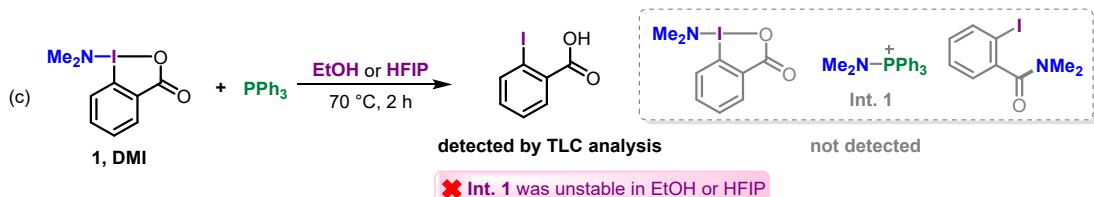


N-Methyl-2-oxo-2-phenylacetamide (**6c**) was isolated by column chromatography (PE/EA = 3:1) as a yellow oil (32 mg, 72% yield); ^1H NMR (600 MHz, CDCl_3) δ 8.33 (d, J = 7.8 Hz, 2H), 7.61 (t, J = 7.5 Hz, 1H), 7.47 (t, J = 7.6 Hz, 2H), 7.12 (s, 1H), 2.97 (d, J = 5.1 Hz, 3H). It is a known compound; the spectra data is in agreement with the reported one.¹⁴

V. Mechanistic Studies



✓ DMI was stable in EtOH and HFIP



✗ Int. 1 was unstable in EtOH or HFIP



Scheme S2. Control experiments

DMI (140 mg, 0.48 mmol, 1.2 equiv) and PPh_3 (126 mg, 0.48 mmol, 1.2 equiv) were placed in a 10 mL two-neck rounded bottom flask, and DMF (2.0 mL) was added. The reaction flask was placed in an oil bath and heated up to 70 °C. After 10 mins, Int.1 was detected by LC-MS analysis and when substrate was fully consumed, the reaction was quenched with H_2O (10 mL) and extracted with EA (20 mL x 3). The combined organic layer was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated in vacuo to afford the crude product, which was then purified by silica gel flash chromatography to give product **7a** with 76% yield. ^1H NMR (400 MHz, CDCl_3) δ 7.81 (dd, J = 8.0, 1.1 Hz, 1H), 7.38 (td, J = 7.6, 1.2 Hz, 1H), 7.21 (dd, J = 7.6, 1.7 Hz, 1H), 7.06 (td, J = 7.7, 1.7 Hz, 1H), 3.13 (s, 3H), 2.84 (s, 3H). It is a known compound; the spectra data is in agreement with the reported one.¹⁵

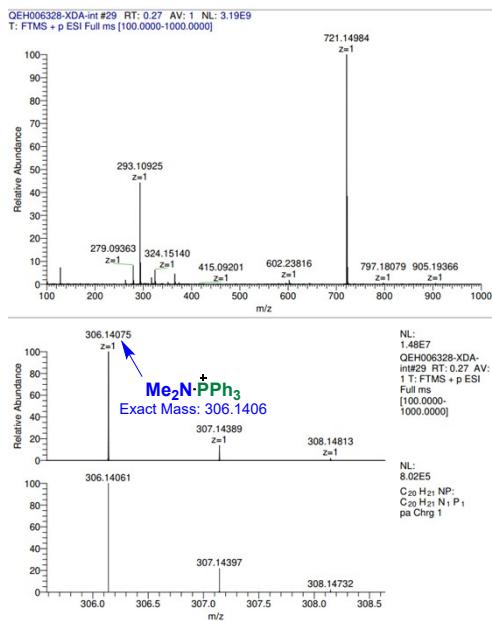


Figure S1 Analysis of reaction by HRMS.

VI. X-Ray Diffraction Data of DMI

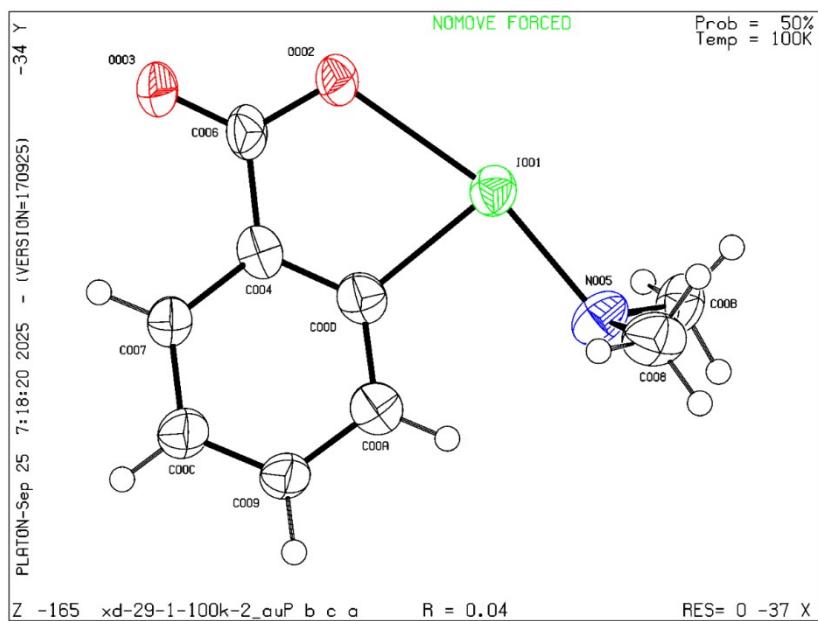


Figure 2 X-ray crystal structure of DMI with 40% ellipsoid probability

Table 1 Crystal data and structure refinement for DMI.

Identification code	DMI
Empirical formula	C ₁₁ H ₁₀ NOI
Formula weight	172.20
Temperature/K	99.98(10)
Crystal system	orthorhombic
Space group	Pbca
a/Å	7.9000(8)
b/Å	15.6486(13)
c/Å	15.661(2)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1936.0(4)
Z	8
ρ _{calcd} /cm ³	1.182
μ/mm ⁻¹	0.608
F(000)	728.0

Crystal size/mm ³	0.1 × 0.04 × 0.02
Radiation	Cu K α (λ = 1.54184)
2 Θ range for data collection/°	11.3 to 88.512
Index ranges	-3 ≤ h ≤ 7, -14 ≤ k ≤ 14, -13 ≤ l ≤ 9
Reflections collected	1697
Independent reflections	666 [$R_{\text{int}} = 0.0552$, $R_{\text{sigma}} = 0.0705$]
Data/restraints/parameters	666/194/121
Goodness-of-fit on F ²	1.037
Final R indexes [$ I >=2\sigma (I)$]	$R_1 = 0.0378$, $wR_2 = 0.0713$
Final R indexes [all data]	$R_1 = 0.0636$, $wR_2 = 0.0789$
Largest diff. peak/hole / e Å ⁻³	0.55/-0.44

Table 2 Fractional Atomic Coordinates (×10⁴) and Equivalent Isotropic Displacement Parameters (Å²×10³) for 2491335. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
I001	4077.3(12)	3101.3(5)	4308.0(6)	37.7(4)
O002	6396(10)	3168(6)	5262(6)	37(2)
O003	8526(12)	4018(5)	5653(7)	45(3)
C004	6792(18)	4415(8)	4471(10)	29(3)
N005	2386(15)	3302(7)	3328(8)	45(4)
C006	7310(20)	3849(10)	5193(11)	40(4)
C007	7731(18)	5150(8)	4254(10)	33(3)
C008	710(20)	3611(9)	3652(10)	54(5)
C009	5840(20)	5461(8)	3108(10)	41(4)
C00A	4880(19)	4731(9)	3310(10)	39(4)
C00B	2185(19)	2542(9)	2786(9)	46(4)
C00C	7270(20)	5656(9)	3557(10)	43(4)
C00D	5397(19)	4217(9)	3974(11)	40(4)

Table 3 Anisotropic Displacement Parameters (Å²×10³) for 2491335. The Anisotropic displacement factor exponent takes the form: -2π²[h²a²U₁₁+2hka²b²U₁₂+...].

Atom	U₁₁	U₂₂	U₃₃	U₂₃	U₁₃	U₁₂
I001	38.9(7)	41.8(6)	32.4(7)	-3.9(6)	-0.6(8)	-4.0(6)
O002	25(6)	45(5)	41(6)	3(5)	5(5)	-7(4)
O003	31(7)	53(6)	52(7)	6(6)	-4(6)	-14(5)

Table 3 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for 2491335. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^*{}^2U_{11} + 2hka^*b^*U_{12} + \dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
C004	33(8)	24(6)	32(8)	-16(6)	11(6)	4(5)
N005	46(7)	50(9)	38(9)	-9(6)	-7(6)	12(5)
C006	23(9)	45(8)	52(9)	1(7)	2(7)	-7(6)
C007	26(8)	28(7)	45(9)	-7(6)	0(8)	7(5)
C008	45(9)	52(9)	64(14)	8(9)	5(8)	17(9)
C009	42(9)	29(7)	53(11)	2(7)	-11(8)	7(7)
C00A	35(9)	35(8)	47(10)	-4(6)	7(7)	8(6)
C00B	45(11)	57(9)	35(10)	-10(8)	-2(8)	-7(8)
C00C	38(10)	34(8)	57(10)	0(7)	-8(8)	8(7)
C00D	36(9)	31(8)	52(10)	-2(6)	2(6)	5(6)

Table 4 Bond Lengths for DMI.

Atom	Atom	Length/ \AA	Atom	Atom	Length/ \AA
I001	O002	2.366(9)	C004	C00D	1.385(18)
I001	N005	2.059(12)	N005	C008	1.495(18)
I001	C00D	2.100(14)	N005	C00B	1.470(15)
O002	C006	1.294(16)	C007	C00C	1.397(17)
O003	C006	1.227(16)	C009	C00A	1.408(18)
C004	C006	1.50(2)	C009	C00C	1.362(19)
C004	C007	1.410(18)	C00A	C00D	1.376(18)

Table 5 Bond Angles for DMI.

Atom	Atom	Atom	Angle/ $^\circ$	Atom	Atom	Atom	Angle/ $^\circ$
N005	I001	O002	165.1(4)	O002	C006	C004	113.3(15)
N005	I001	C00D	90.5(6)	O003	C006	O002	124.5(15)
C00D	I001	O002	74.7(5)	O003	C006	C004	122.1(15)
C006	O002	I001	114.7(10)	C00C	C007	C004	120.8(15)
C007	C004	C006	121.3(15)	C00C	C009	C00A	120.8(15)
C00D	C004	C006	120.8(14)	C00D	C00A	C009	118.9(15)
C00D	C004	C007	117.8(15)	C009	C00C	C007	119.5(15)
C008	N005	I001	111.7(9)	C004	C00D	I001	116.2(12)
C00B	N005	I001	112.2(8)	C00A	C00D	I001	121.8(12)

C00B N005 C008 111.2(12)

C00A C00D C004 122.0(14)

Table 6 Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for 2491335.

Atom	x	y	z	U(eq)
H007	8691.8	5302.25	4585.34	40
H00A	194.72	3167.18	4007.23	81
H00B	-28.79	3740.02	3168.25	81
H00C	882.69	4128.97	3993.95	81
H009	5491.97	5822.45	2653.52	50
H00D	3890.31	4595.17	2992.44	47
H00E	3302.57	2315.55	2636.35	68
H00F	1576.28	2698.96	2263.36	68
H00G	1541.31	2105.96	3095.72	68
H00H	7941.51	6132.71	3397.51	51

Table 7 Atomic Occupancy for 2491335.

Atom	Occupancy
N005	1.01(3)

VII. Reference

(1) Yang Z.; Du F.-H.; Zhang C.; Du Y. Accessing aryl azides *via* copper powder-catalyzed cross-coupling of arylboronic acids with the hypervalent azido-iodine reagent. *Org. Chem. Front.* **2023**, *10*, 4131-4138.

(2) Bogonda G.; Kim H. Y.; Oh K. Direct Acyl Radical Addition to 2H-Indazoles Using Ag-Catalyzed Decarboxylative Cross-Coupling of α -Keto Acids. *Org. Lett.* **2018**, *20*, 2711-2715.

(3) Wang F.; Feng H.; Li H.; Miao T.; Cao T.; Zhang M. 1D $\text{Fe}_3\text{O}_4@\text{CuSiO}_3$ composites catalyzed decarboxylative A^3 -coupling for propargylamine synthesis. *Chin. Chem. Lett.* **2020**, *31*, 1558-1563.

(4) Dai Y.; Niu W.; Huang J.; Sun J.; Xu X. Photo-induced amidation/Smiles rearrangement of alkenes for synthesizing quaternary-carbon-containing succinyldiamides. *Org. Biomol. Chem.* **2025**, *23*, 1330-1337.

(5) Li D.; Wang M.; Liu J.; Zhao Q.; Wang L. Cu(ii)-catalyzed decarboxylative acylation of acyl C–H of formamides with α -oxocarboxylic acids leading to α -ketoamides. *Chem. Commun.* **2013**, *49*, 3640-3642.

(6) Wang Y.; Meng X.; Cai C.; Wang L.; Gong H. Radical Cross-Coupling Reaction Based on Hydrogen Atom Abstraction of DMF and Decarboxylation of α -Ketoacid under Electricity. *J. Org. Chem.* **2022**, *87*, 15042-15049.

(7) Yang D.-N.; Du Y.-N.; Wang P.; Han M.-Y. Brook-Oxidation Reaction of Acylsilanes: General Access to α -Ketoamides and α -Ketothioamides. *Org. Lett.* **2024**, *26*, 10020-10024.

(8) Choudhary S.; Mandal A.; Patra A.; Kant R.; Ghosh N. Copper/Zinc-Catalyzed Stitching of 2-Carbonylanilines with Bis(ynamides): Access to Pyrrolo 2,3-*b* quinolines and Its Photophysical Studies. *J. Org. Chem.* **2024**, *89*, 6274-6280.

(9) Pérez E. G.; Cassels B. K.; Eibl C.; Gündisch D. Synthesis and evaluation of *N*1-alkylindole-3-ylalkylammonium compounds as nicotinic acetylcholine receptor ligands. *Bioorg. Med. Chem.* **2012**, *20*, 3719-3727.

(10) Sarkar S.; Pal S.; Mukherjee A.; Santra S.; Zyryanov G. V.; Majee A. Visible-Light-Promoted Metal- and Photocatalyst-Free Reactions between Arylglyoxylic Acids and Tetraalkylthiuram Disulfides: Synthesis of α -Ketoamides. *J. Org. Chem.* **2024**, *89*, 1473-1482.

(11) Jayaram A.; Seenivasan V. T.; Govindan K.; Liu Y.-M.; Chen N.-Q.; Yeh T.-W.; Venkatachalam G.; Li C.-H.; Leung T.-F.; Lin W.-Y. Base-promoted triple cleavage of CCl_2Br : a direct one-pot synthesis of unsymmetrical oxalamide derivatives. *Chem. Commun.* **2024**, *60*, 3079-3082.

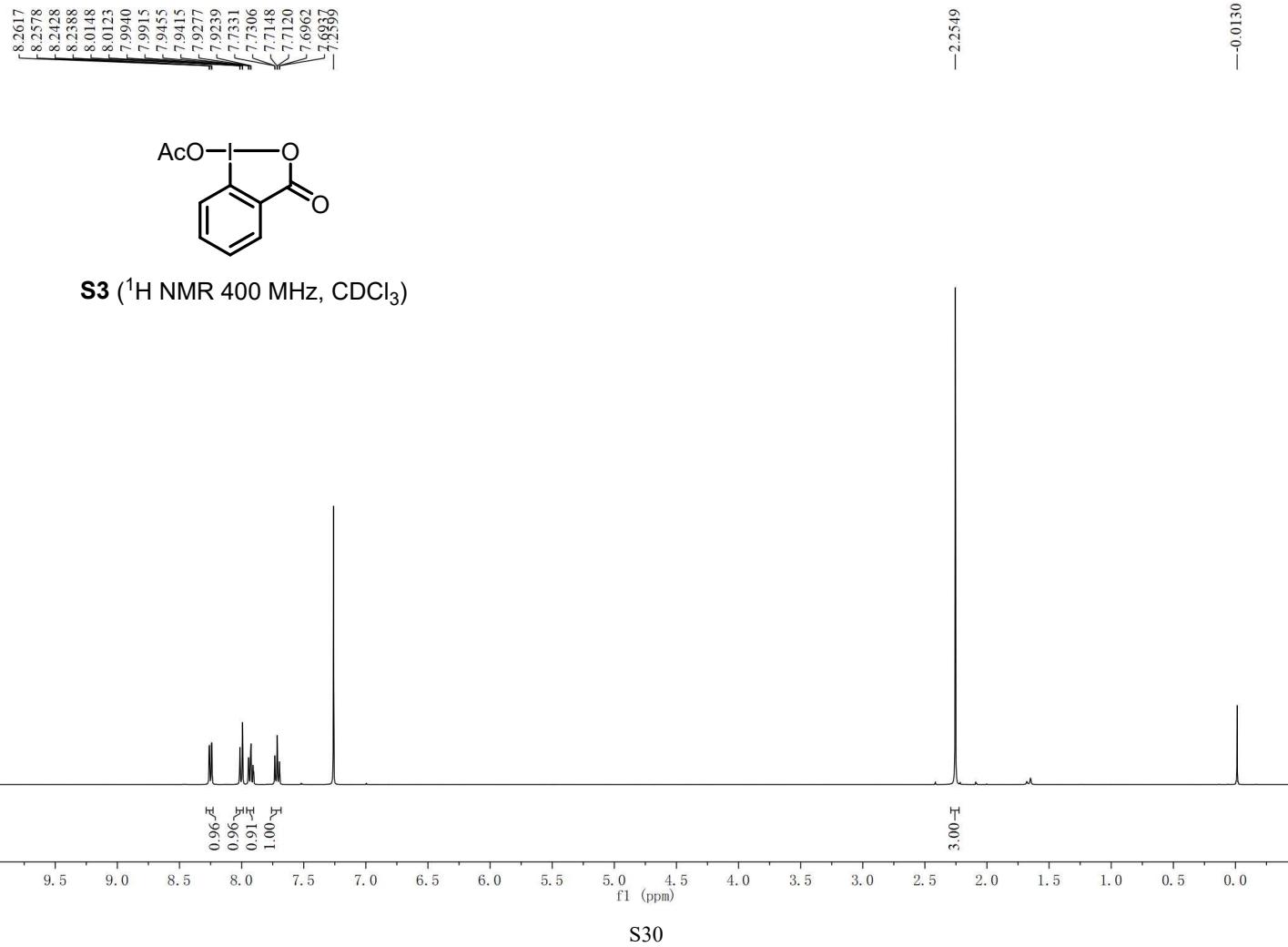
(12) Liu C.; Yang Z.; Guo S.; Zeng Y.; Zhu N.; Li X.; Fang Z.; Guo K. Copper–TEMPO-catalyzed synthesis of α -ketoamides via tandem $\text{sp}^3\text{C}-\text{H}$ aerobic oxidation and amination of phenethyl alcohol derivatives. *Org. Biomol. Chem.* **2016**, *14*, 8570-8575.

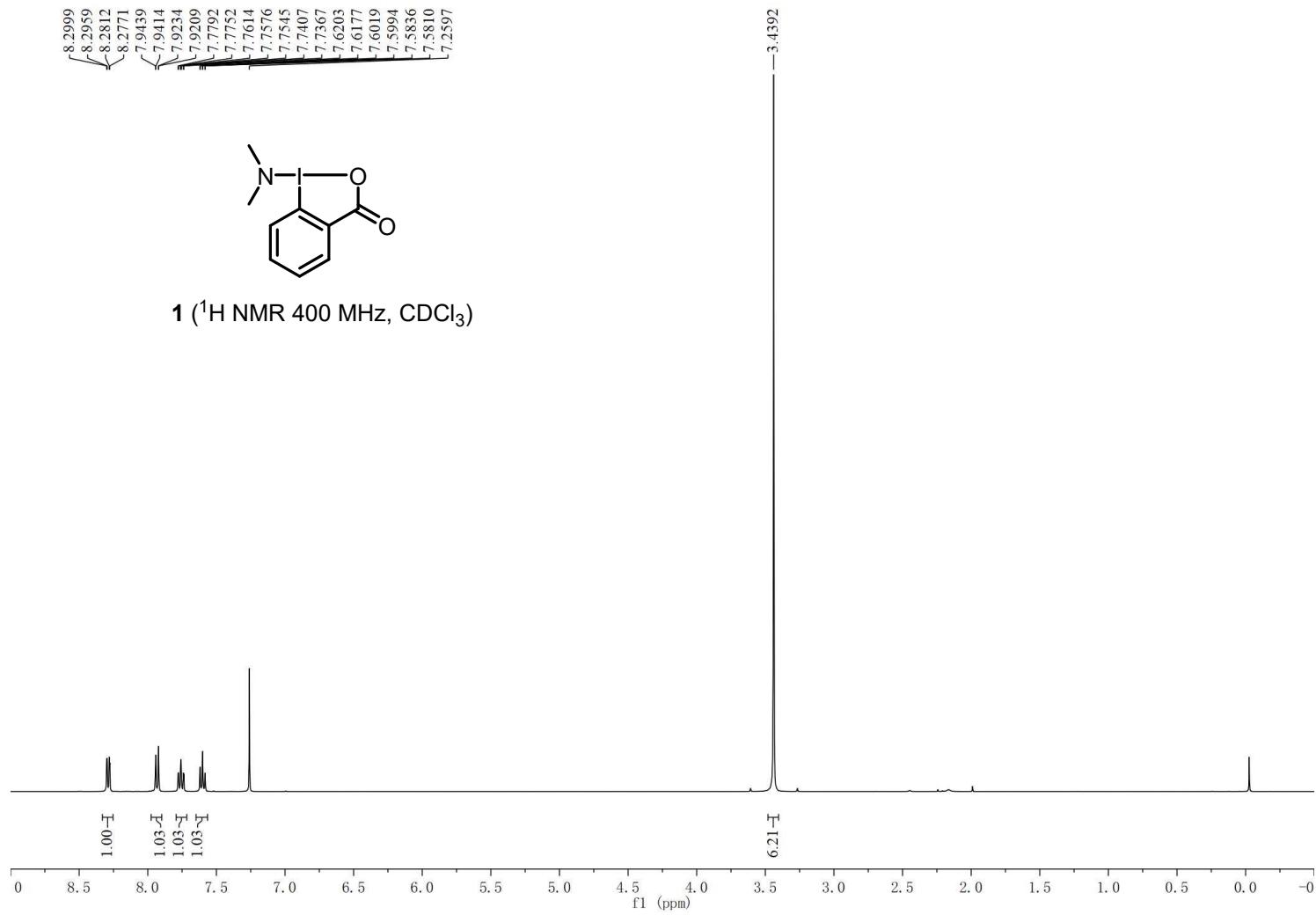
(13) Wang H.; Zhao Y.; Zheng Y.; Fang S.; Li J.; Wan X. Oxidative Coupling of Diazo and NH_4I : A Route to Primary Oxamates and α -Ketoamides. *J. Org. Chem.* **2020**, *85*, 3050-3058.

(14) Gu G.; Yang T.; Yu O.; Qan H.; Wang J.; Wen J.; Dang L.; Zhang X. Enantioselective Iridium-Catalyzed Hydrogenation of α -Keto Amides to α -Hydroxy Amides. *Org. Lett.* **2017**, *19*, 5920-5923.

(15) Sanz-Marco A.; Saavedra B.; Erbing E.; Malmberg J.; Johansson M. J.; Martin-Matute B. Selective C-H Iodination of Weinreb Amides and Benzamides through Iridium Catalysis in Solution and under

V. NMR Spectra of Starting Materials and Products





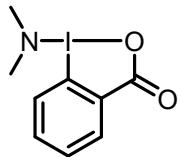
— 168.1197

133.4948
132.6125
132.5532
130.3624
— 125.1959

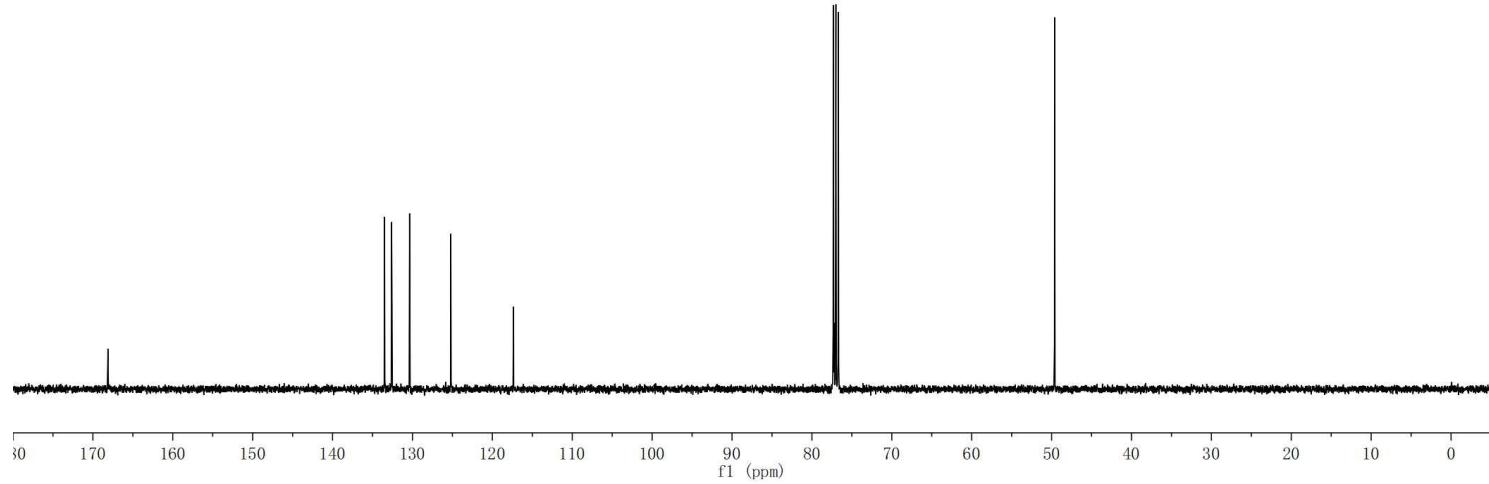
— 117.3522

77.3187
77.2020
76.9996
76.6835

— 49.6236

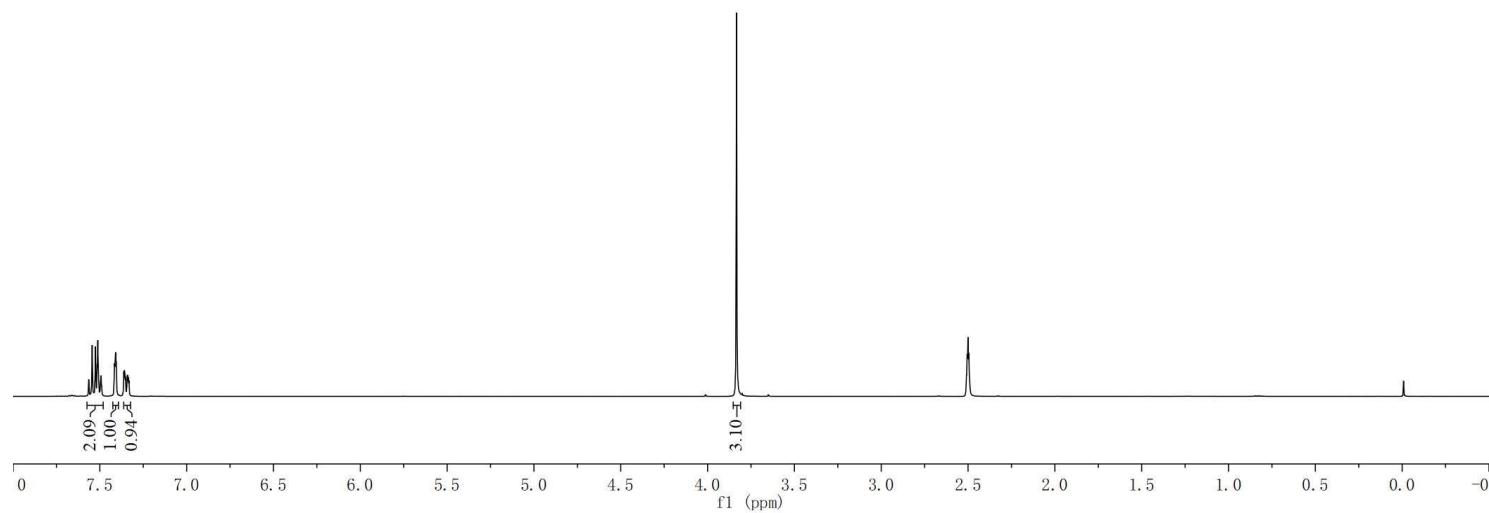


1 (^{13}C NMR 101 MHz, CDCl_3)



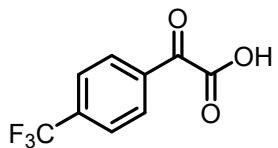


2h (^1H NMR 400 MHz, $\text{DMSO}-d_6$)

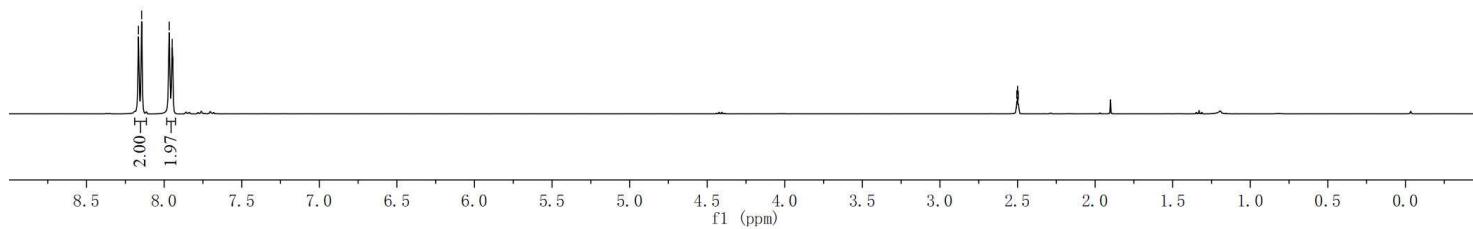


8.1655
8.1451
7.9674
7.9482
7.9138

2.5045
2.4998
2.4953



2i (^1H NMR 400 MHz, $\text{DMSO}-d_6$)

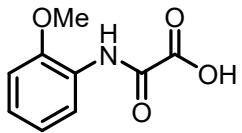


-9.6171

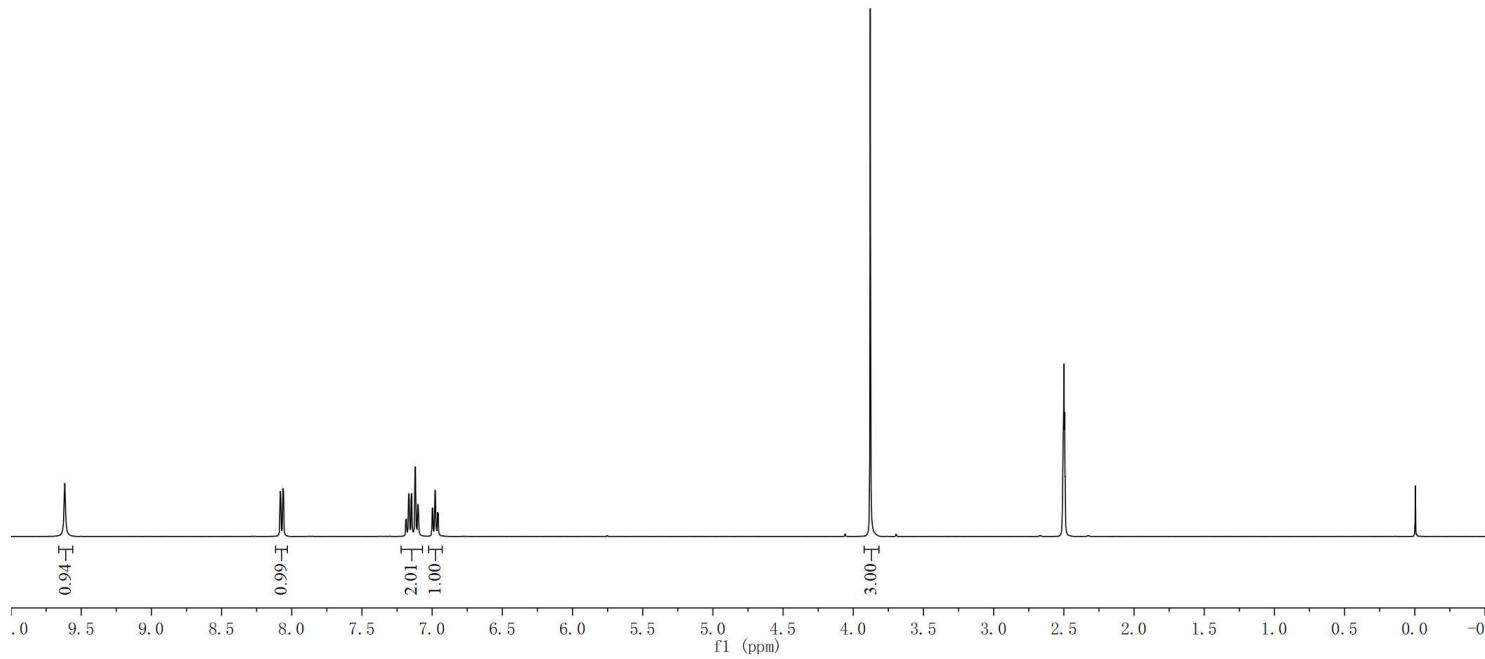
8.0835
8.0808
8.0777
8.0635
8.0611
8.0580
7.1839
7.1699
7.1673
7.1634
7.1492
7.1451
7.1228
7.1189
7.1021
7.0984
6.9993
6.9955
6.9803
6.9764
6.9614
6.9575

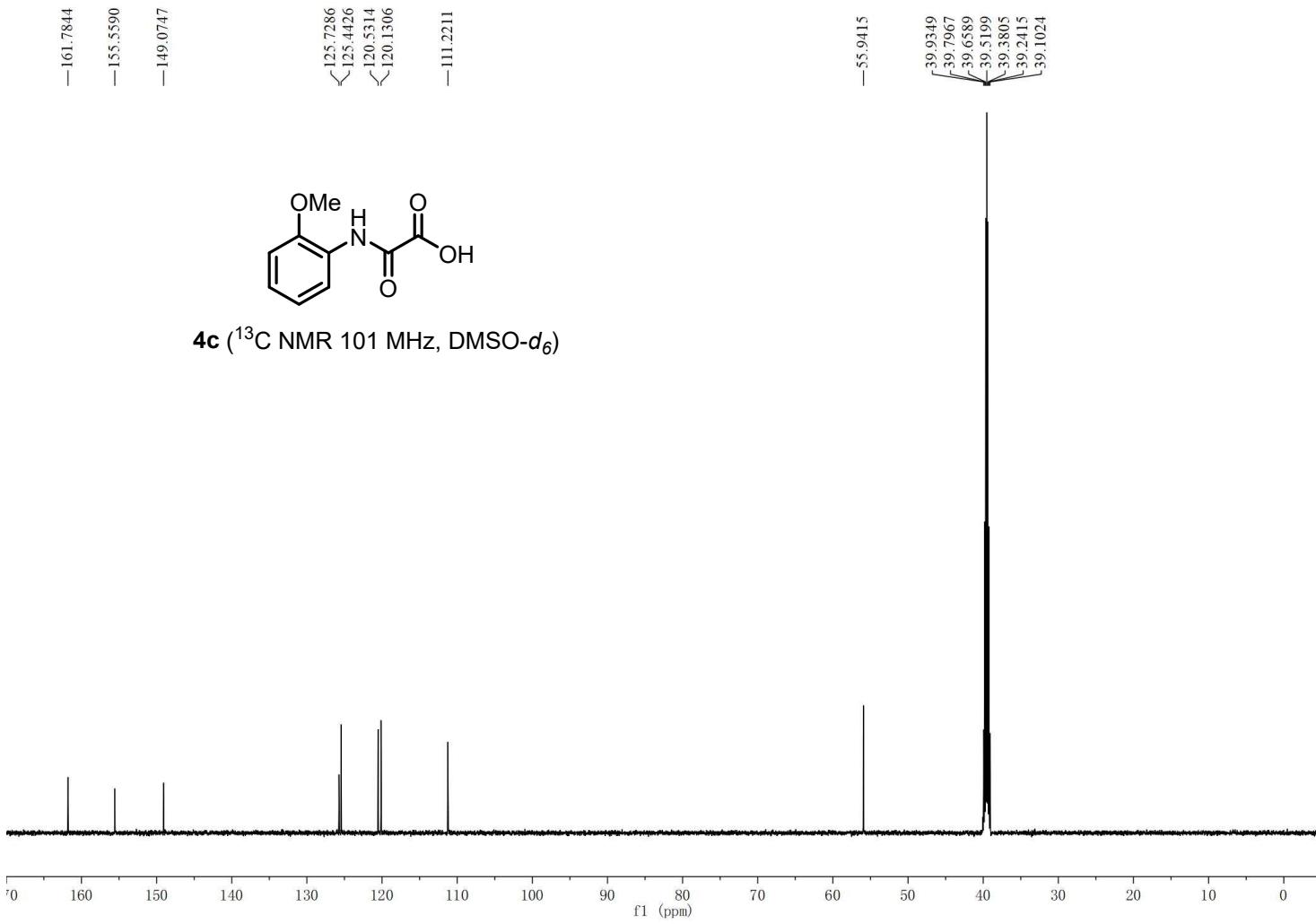
-3.8792

2.5087
2.5041
2.4996
2.4949
2.4902



4c (^1H NMR 400 MHz, $\text{DMSO}-d_6$)

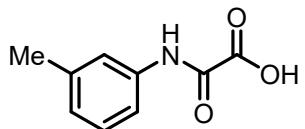




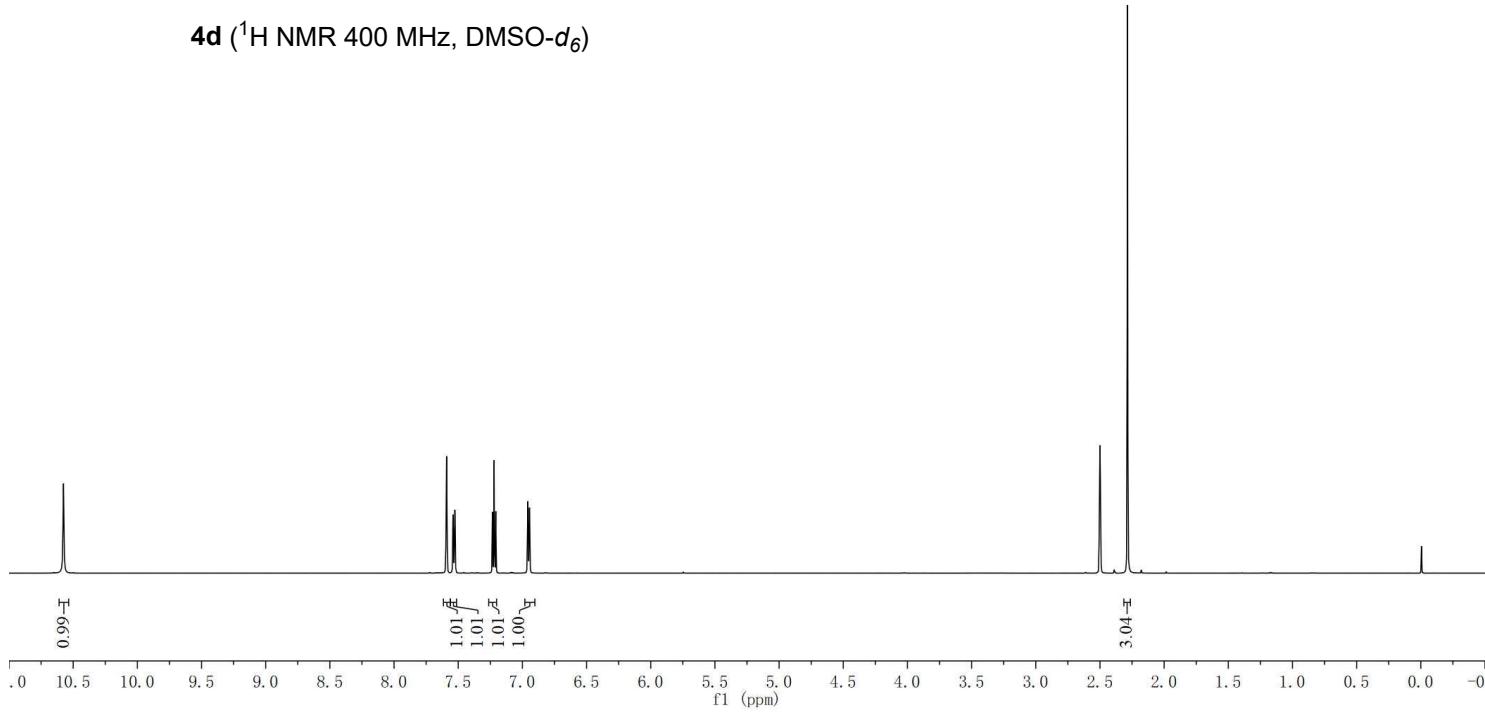
-10.5757

7.5941
7.5908
7.5875
7.5412
7.5376
7.5285
7.5263
7.5235
7.2335
7.2205
7.2074
6.9573
6.9448

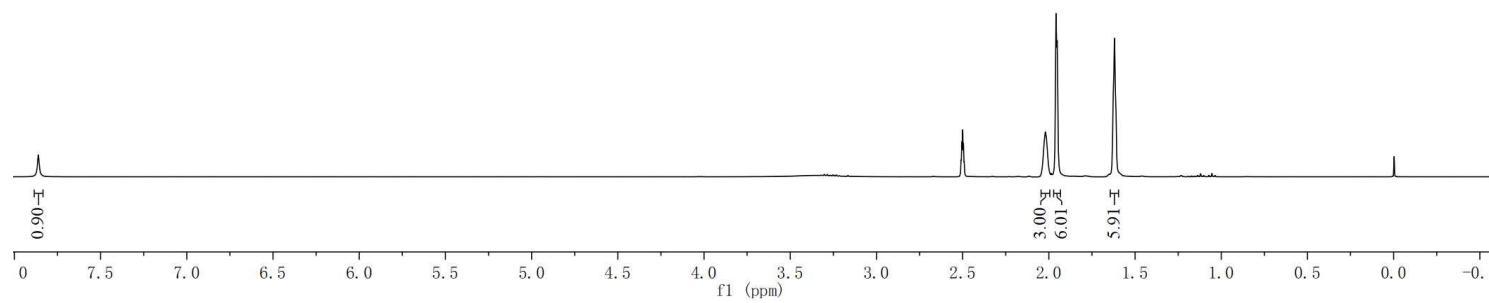
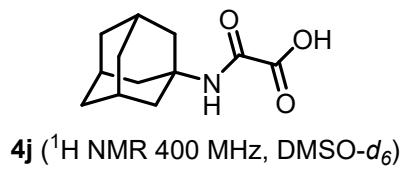
2.5059
2.5029
2.4998
2.4966
2.4934
2.2855



4d (^1H NMR 400 MHz, $\text{DMSO}-d_6$)



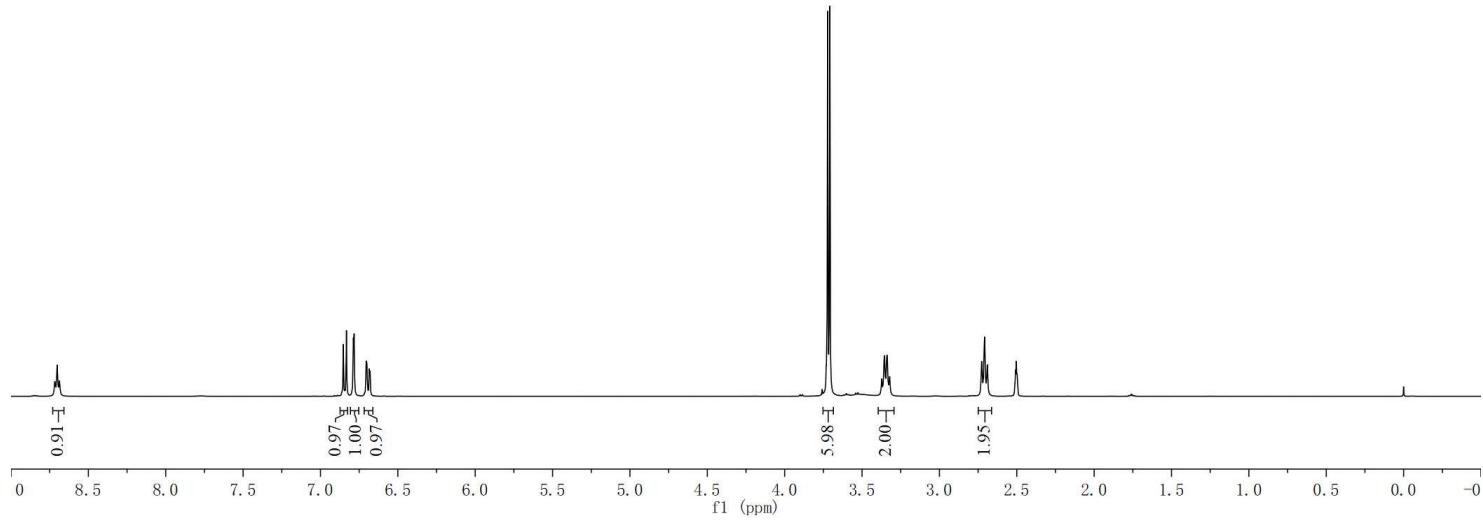
-7.8612

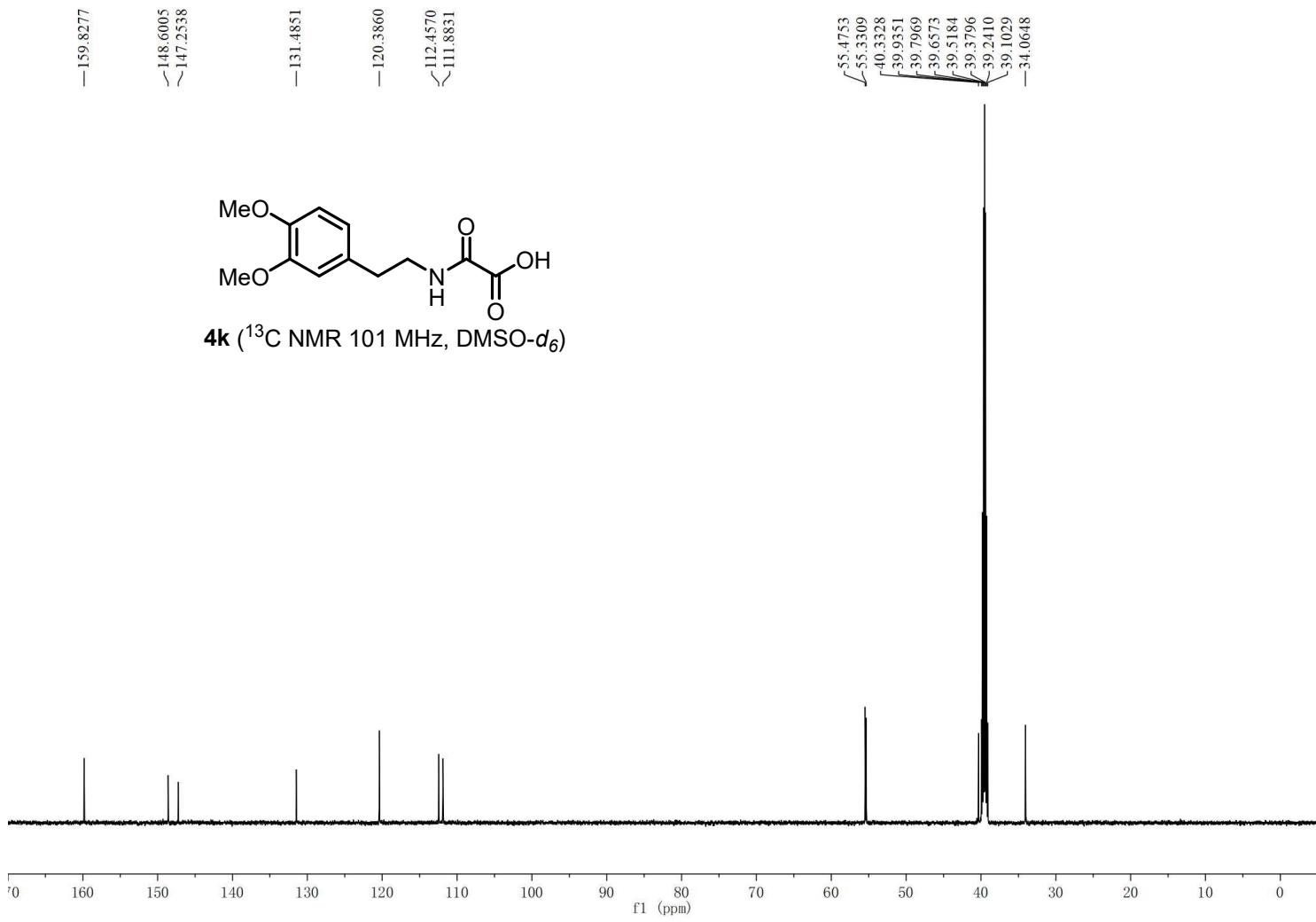


8.7168
8.7018
8.6866

6.8526
6.8321
6.7873
6.7523
6.7046
6.6997
6.6843
6.6792

3.7317
3.7226
3.7083
3.3731
3.3369
3.3529
3.3406
3.3387
3.3365
3.3215
2.7269
2.7086
2.6904
2.5127
2.5080
2.5034
2.4988
2.4941

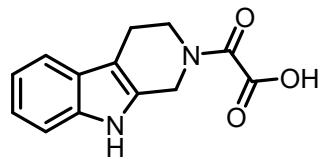




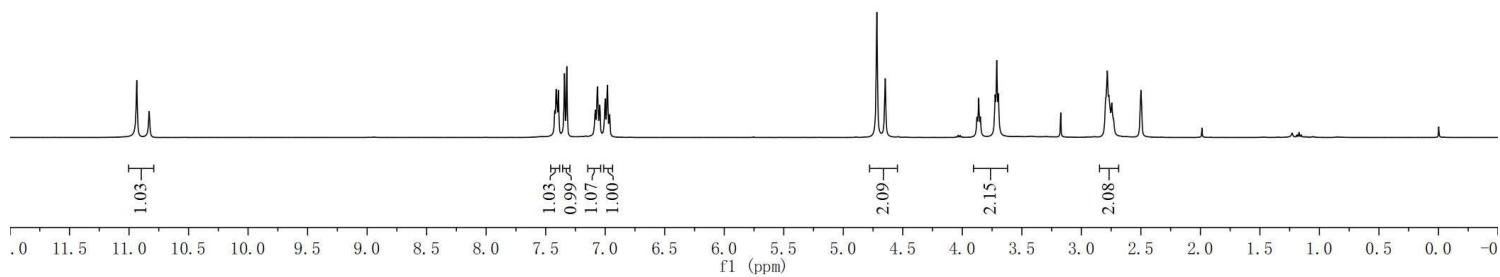
~ 10.9351
 ~ 10.8310

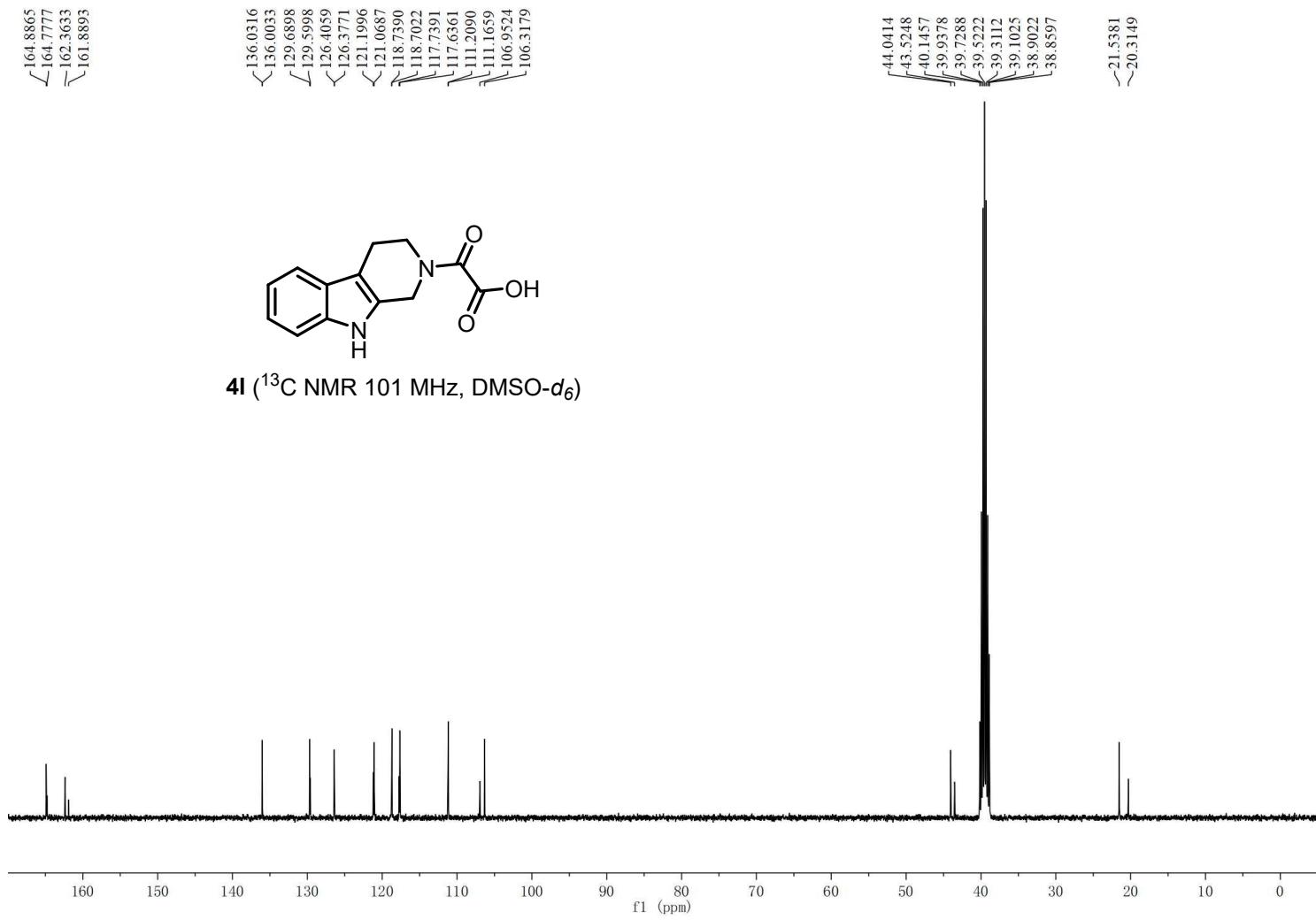
7.4237
7.4122
7.4038
7.3930
7.3423
7.3223
7.0883
7.0821
7.0710
7.0644
7.0512
7.0462
7.0425
7.0049
6.9990
6.9858
6.9801
6.9676
6.9619

4.7183
4.6479
3.8770
3.8626
3.8481
3.7247
3.7105
3.6963
2.7964
2.7823
2.7681
2.7591
2.7440
2.7291
2.5090
2.5044
2.4996
2.4949
2.4902

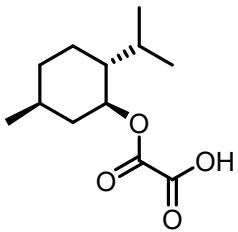


4l (^1H NMR 400 MHz, $\text{DMSO}-d_6$)

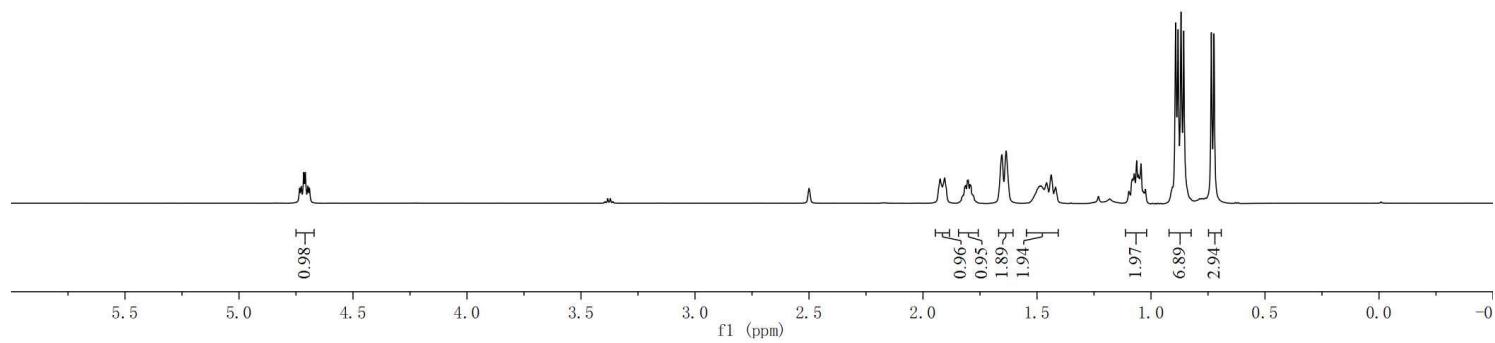




4.7344
-4.7271
-4.7161
-4.7088
-4.6980
-4.6907
-2.5062
-2.5031
-2.4997
-2.4963
-2.4933
-1.9324
-1.9253
-1.9177
-1.9116
-1.9049
-1.8980
-1.8173
-1.8128
-1.8057
-1.8012
-1.7941
-1.7896
-1.7824
-1.6634
-1.6577
-1.6523
-1.6393
-1.6349
-1.6297
-1.6243
-1.5075
-1.5024
-1.4980
-1.4925
-1.4870
-1.4815
-1.4766
-1.4725
-1.4618
-1.4568
-1.4376
-1.4232
-1.4183
-1.4131
-1.0976
-1.0857
-1.0821
-1.0739
-1.0631
-1.0555
-1.0504
-1.0438
-1.0332
-1.0242
-0.9117
-0.9065
-0.8920
-0.8813
-0.8682
-0.8565
-0.8434
-0.7333
0.7237



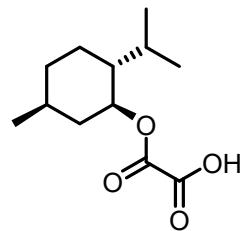
4o (^1H NMR 600 MHz, $\text{DMSO}-d_6$)



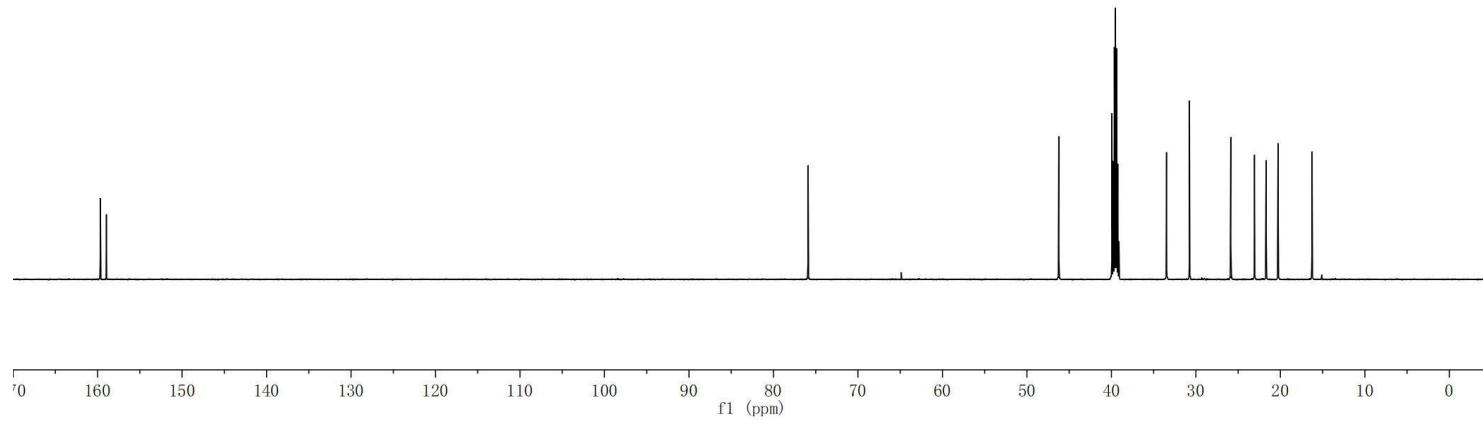
159.6676
158.9560

75.9000

46.2290
39.9413
39.7974
39.6582
39.5198
39.3809
39.2414
39.1021
33.4802
30.7617
25.8738
23.0664
21.6960
20.2762
16.2519

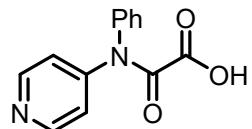


4o (^{13}C NMR 151 MHz, $\text{DMSO-}d_6$)

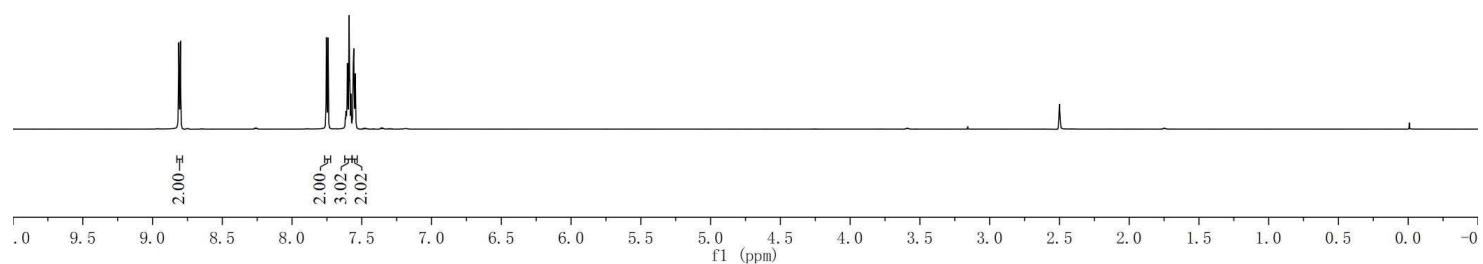


8.8118
 8.8090
 8.8025
 8.7997
 7.7539
 7.7511
 7.7446
 7.7417
 7.6142
 7.6036
 7.5992
 7.5946
 7.5914
 7.5886
 7.5856
 7.5827
 7.5780
 7.5607
 7.5572
 7.5533
 7.5503
 7.5470
 7.5443

2.5060
 2.5029
 2.4997
 2.4967
 2.4935



4i (^1H NMR 600 MHz, $\text{DMSO}-d_6$)



~163.1462

~162.0311

-143.0748

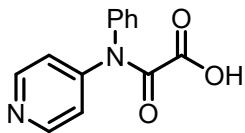
-136.8543

130.3601

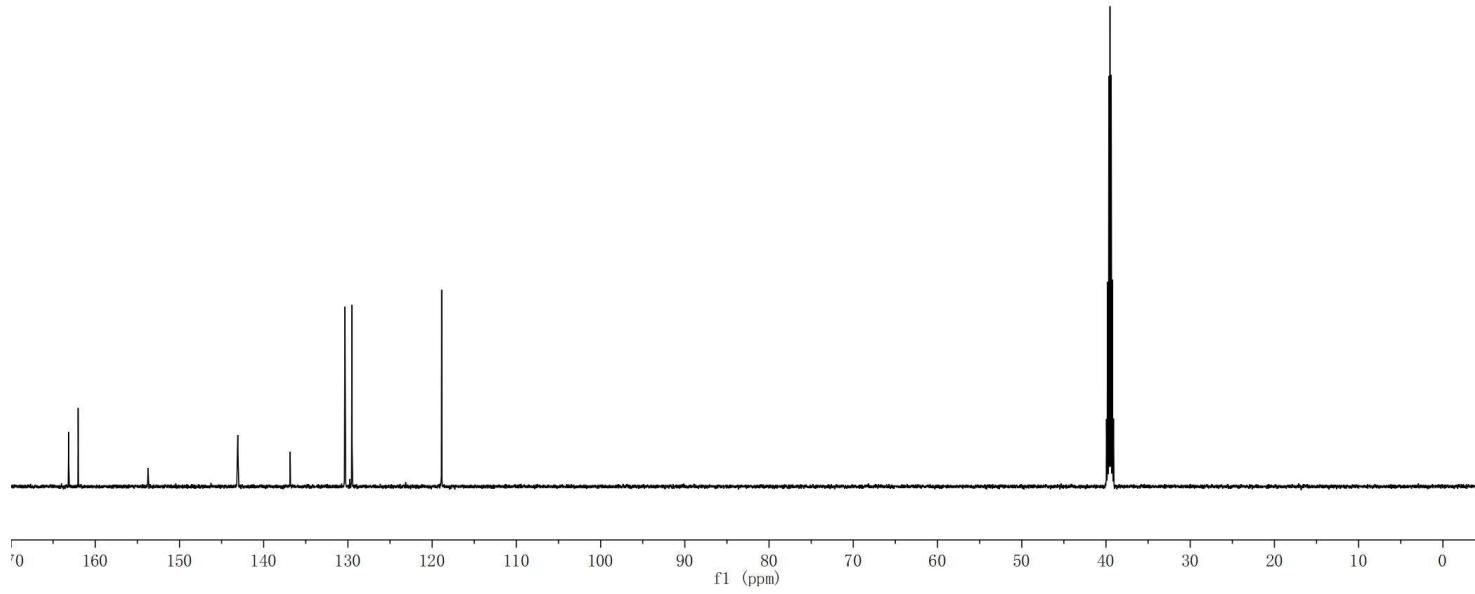
130.3047

129.5196

-118.8720



4i (^{13}C NMR 151 MHz, $\text{DMSO-}d_6$)

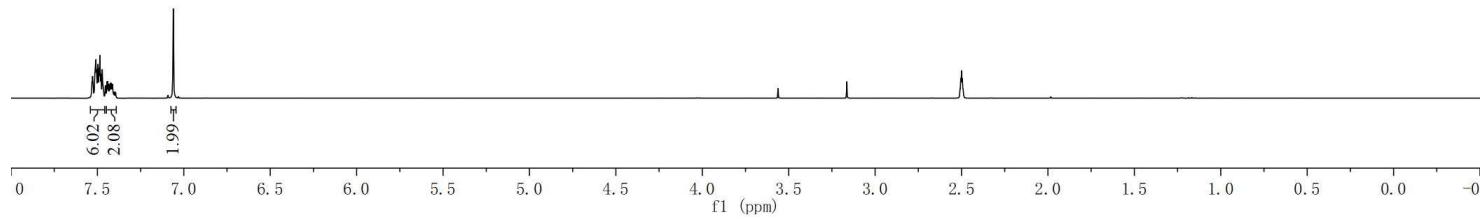


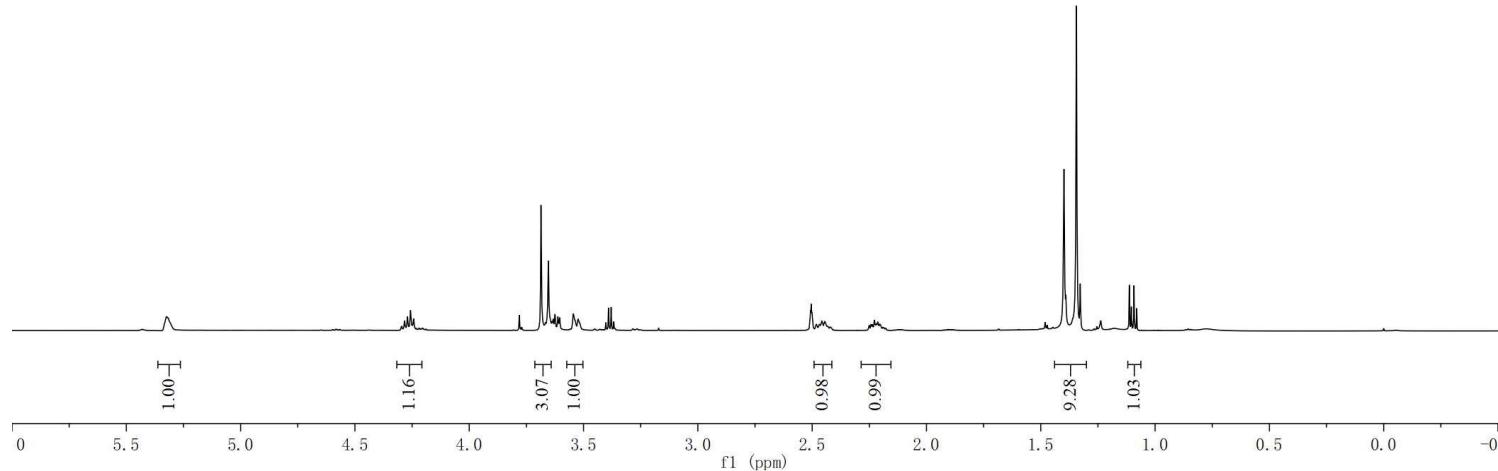
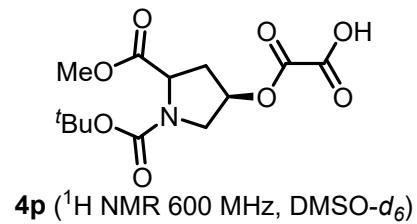
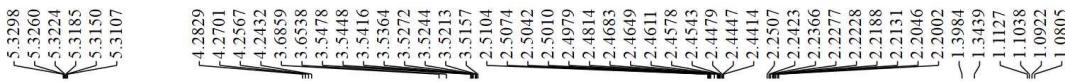
7.5357
7.5318
7.5281
7.5244
7.5152
7.5119
7.5092
7.5053
7.5021
7.4980
7.4944
7.4906
7.4860
7.4800
7.4718
7.4667
7.4536
7.4485
7.4441
7.4397
7.4349
7.4302
7.4261
7.4219
7.4185
7.4154
7.4123
7.4077
7.4038
7.3990
7.3943
7.0609

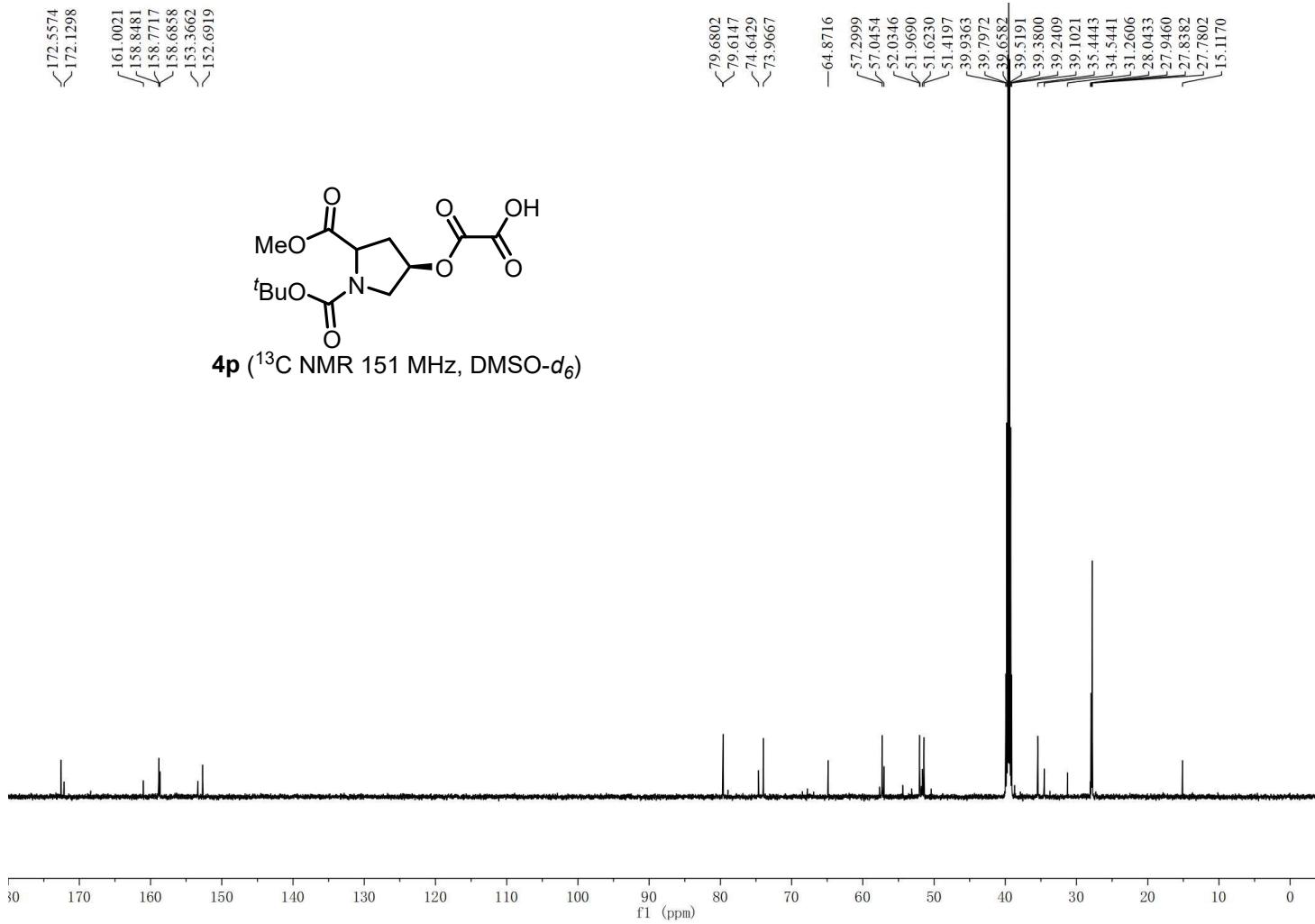
2.5089
2.5043
2.4997
2.4950
2.4902



4m (^1H NMR 400 MHz, $\text{DMSO}-d_6$)

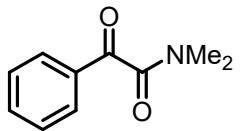




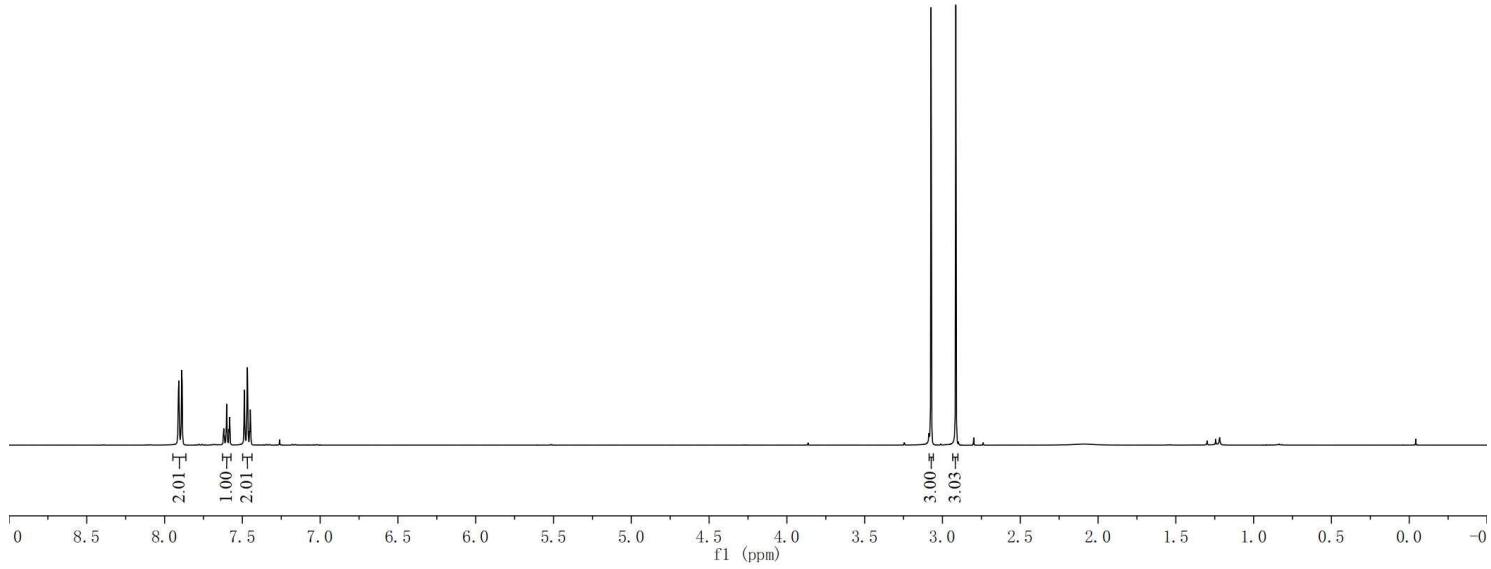


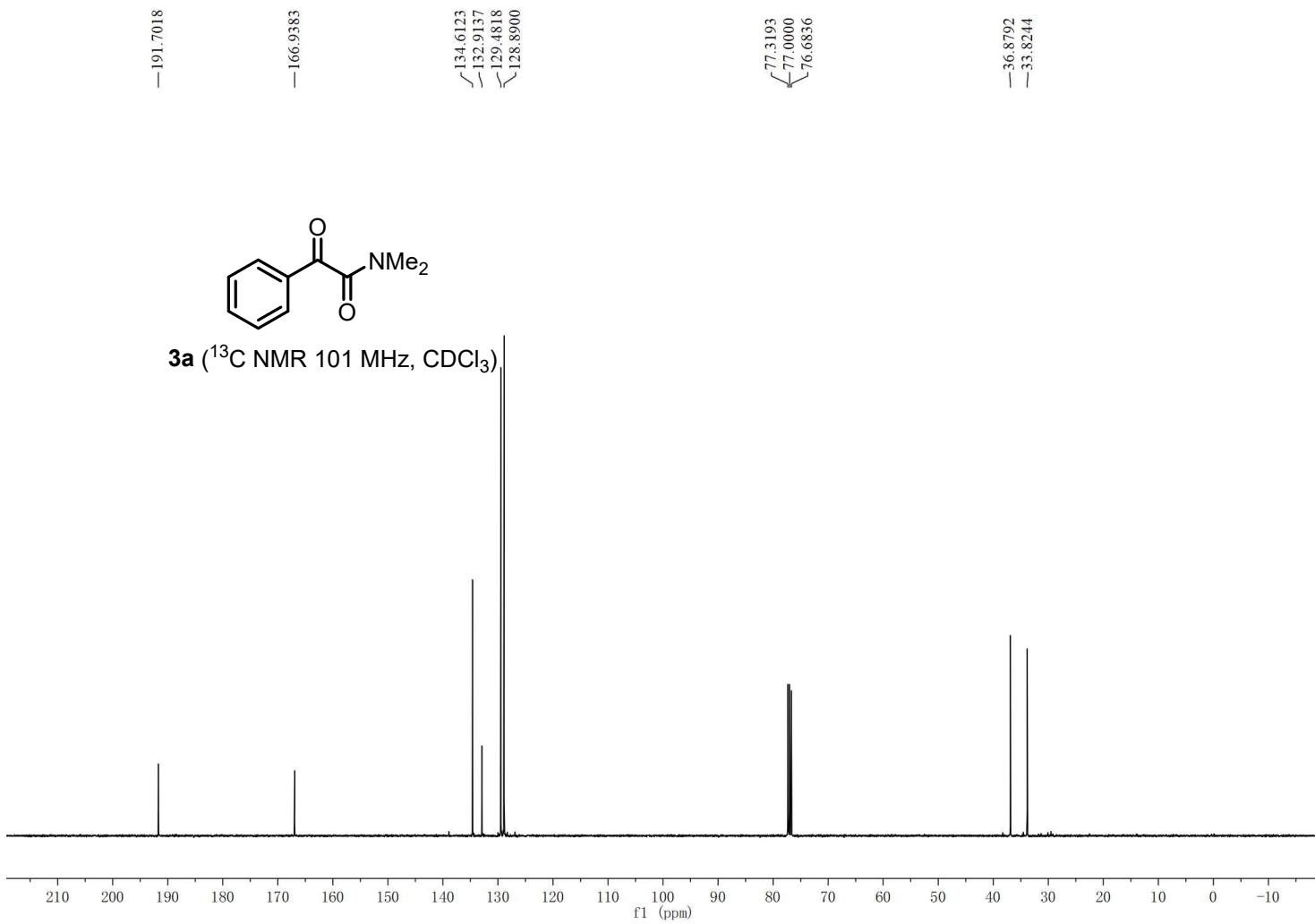
7.9156
7.9110
7.9080
7.9044
7.8954
7.8905
7.8867
7.6227
7.6194
7.6161
7.6056
7.6009
7.5964
7.5857
7.5823
7.5789
7.4917
7.4876
7.4835
7.4707
7.4676
7.4643
7.4533
7.4489
7.4459
7.2606

—3.0733
—2.9142

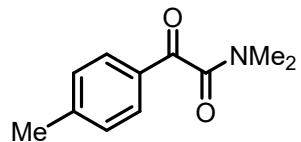


3a (^1H NMR 400 MHz, CDCl_3)

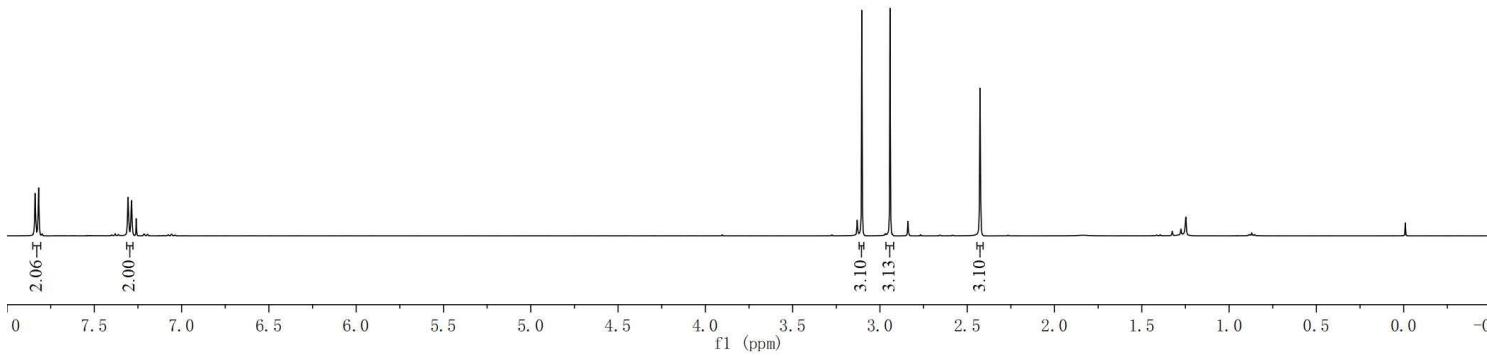


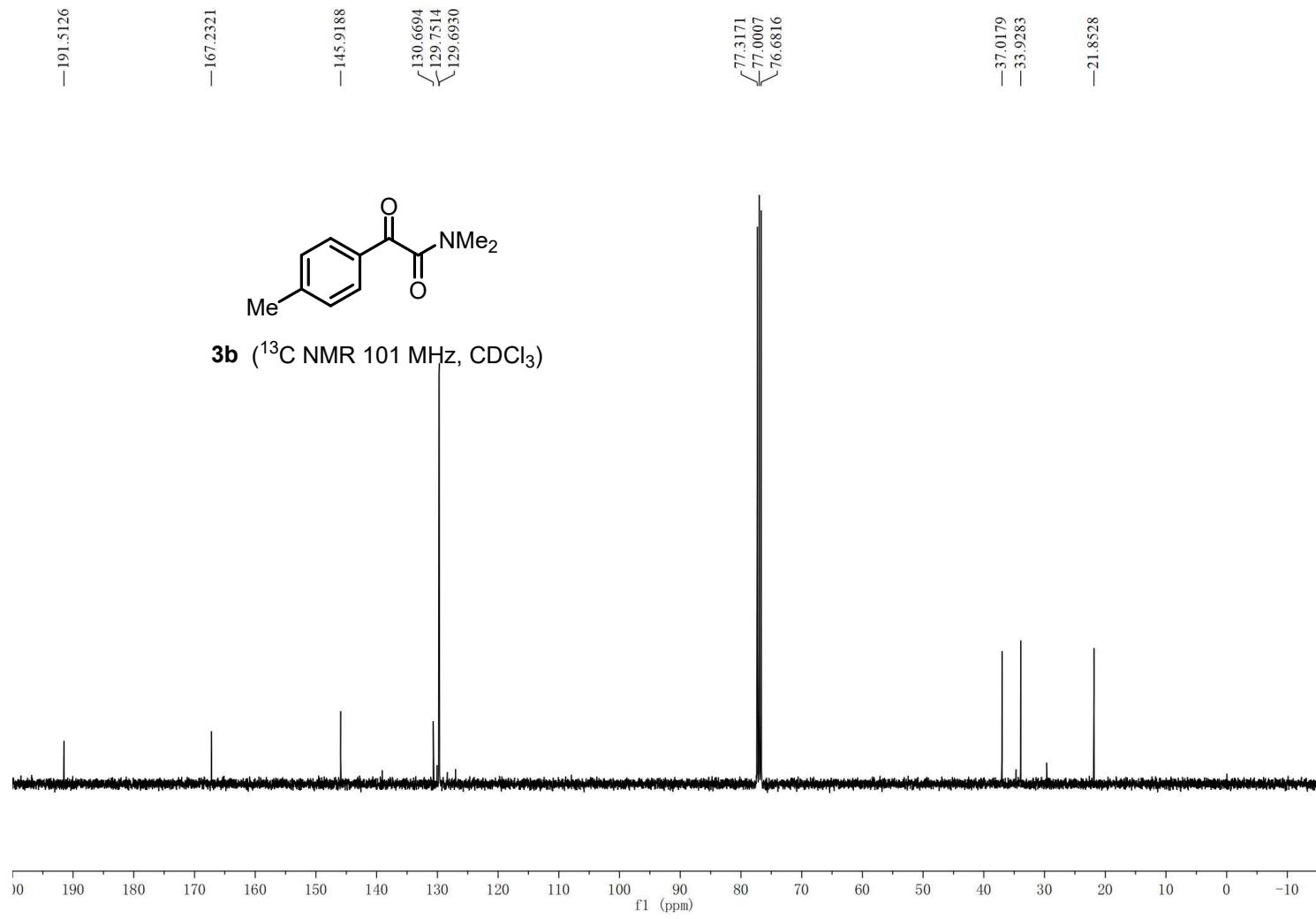


7.8396
7.8355
7.8236
7.8190
7.3085
7.3064
7.3032
7.2868
7.2598



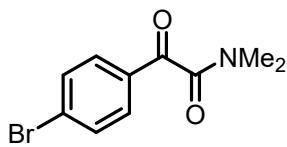
3b (^1H NMR 400 MHz, CDCl_3)



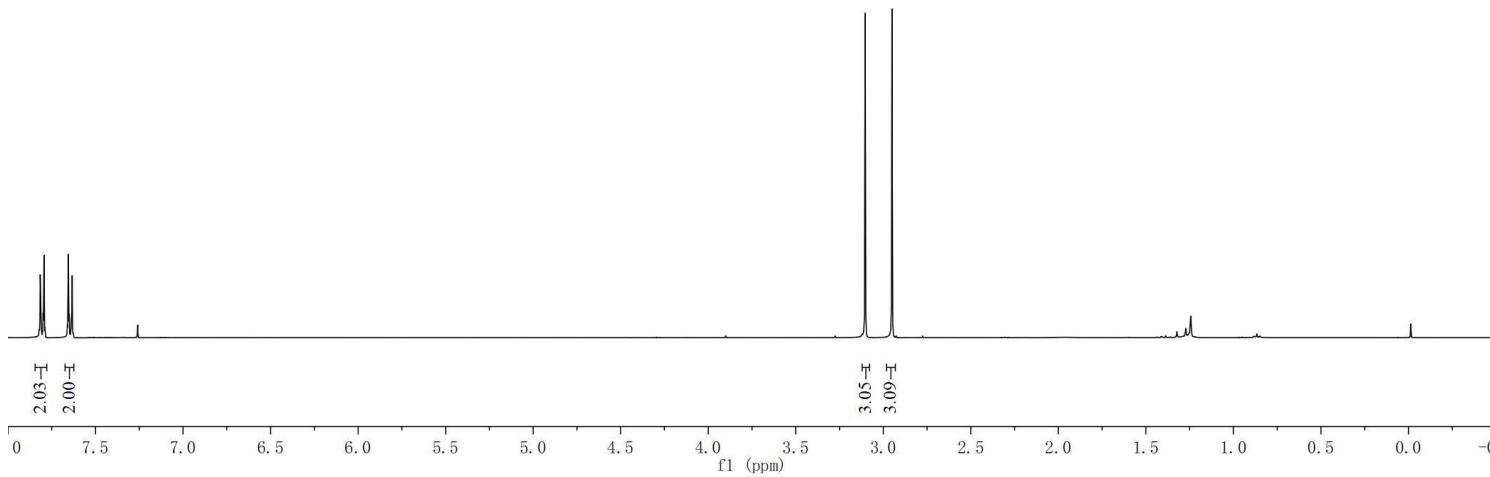


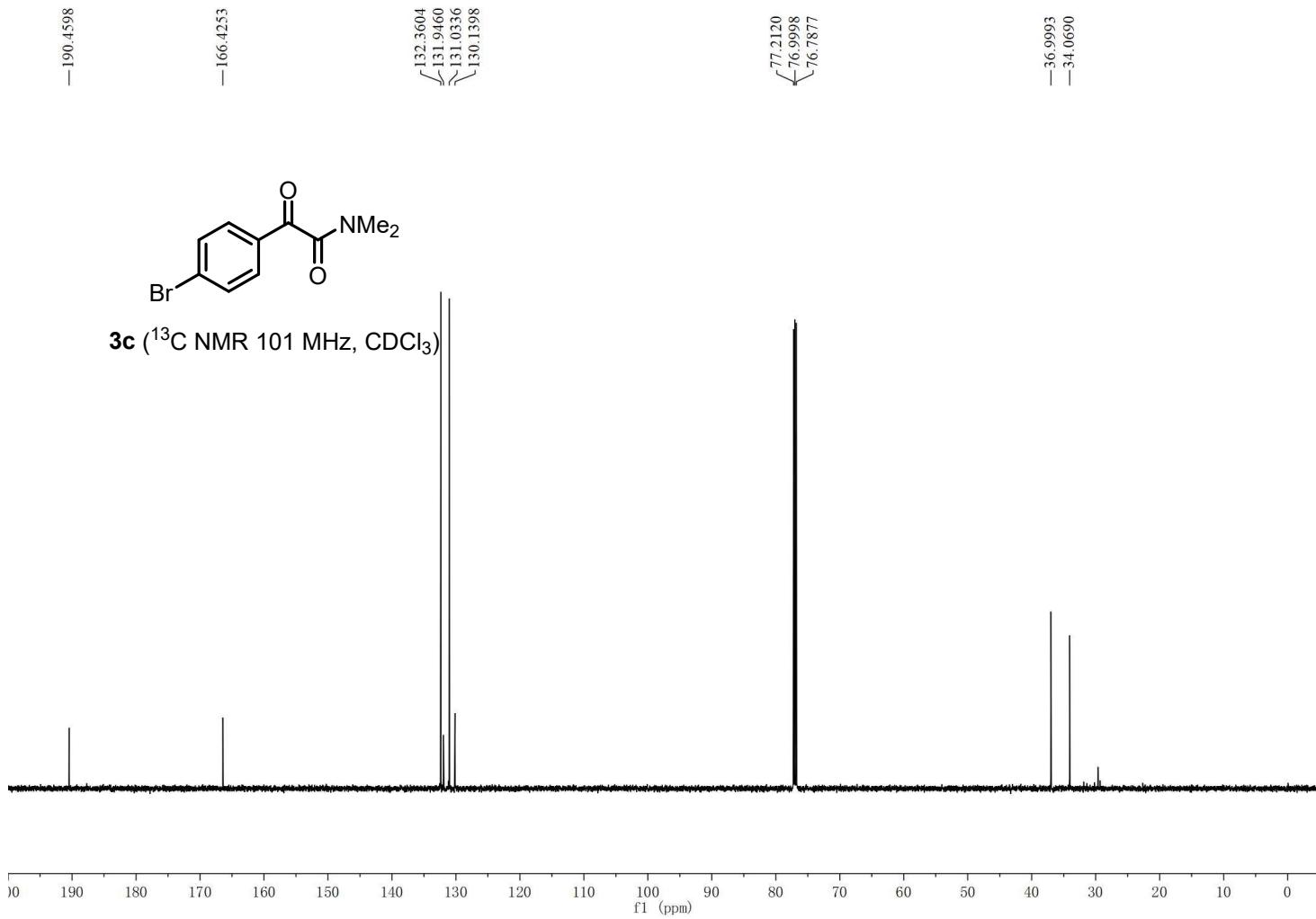
7.8158
7.7944
7.6552
7.6338

— 7.2596



3c (^1H NMR 400 MHz, CDCl_3)

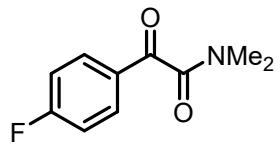




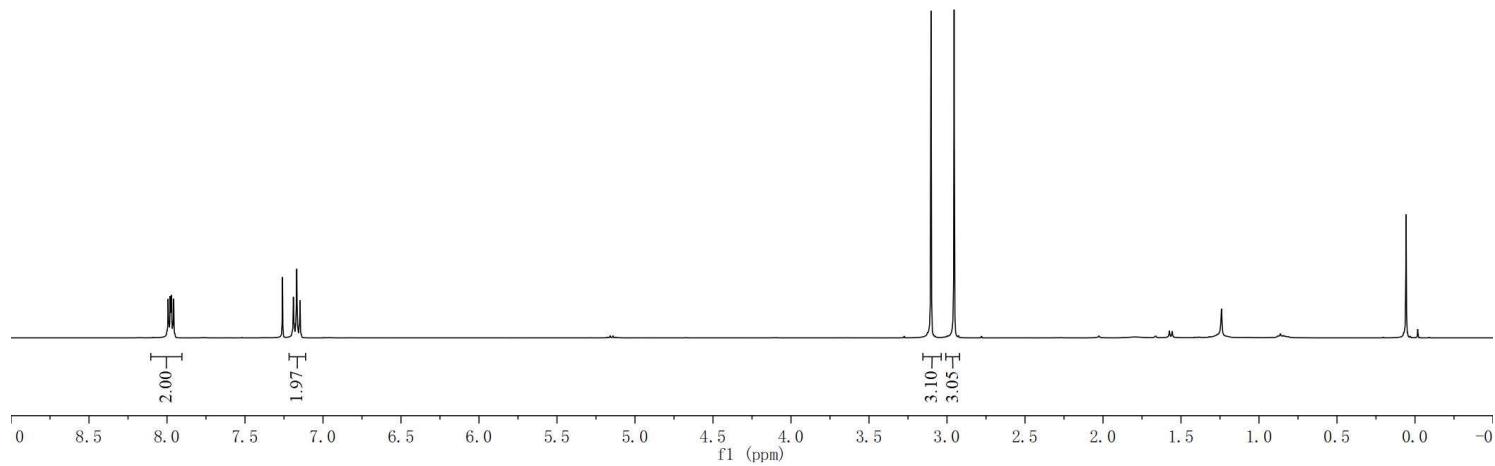
7.9929
7.9795
7.9751
7.9706
7.9573

7.2596
7.1900
7.1683
7.1471

-3.1024
-2.9545



3d (^1H NMR 400 MHz, CDCl_3)



— 189.9626

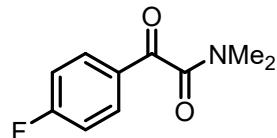
— 167.5084
— 166.6448
— 165.8009

— 132.4977
— 132.4323
— 129.6556

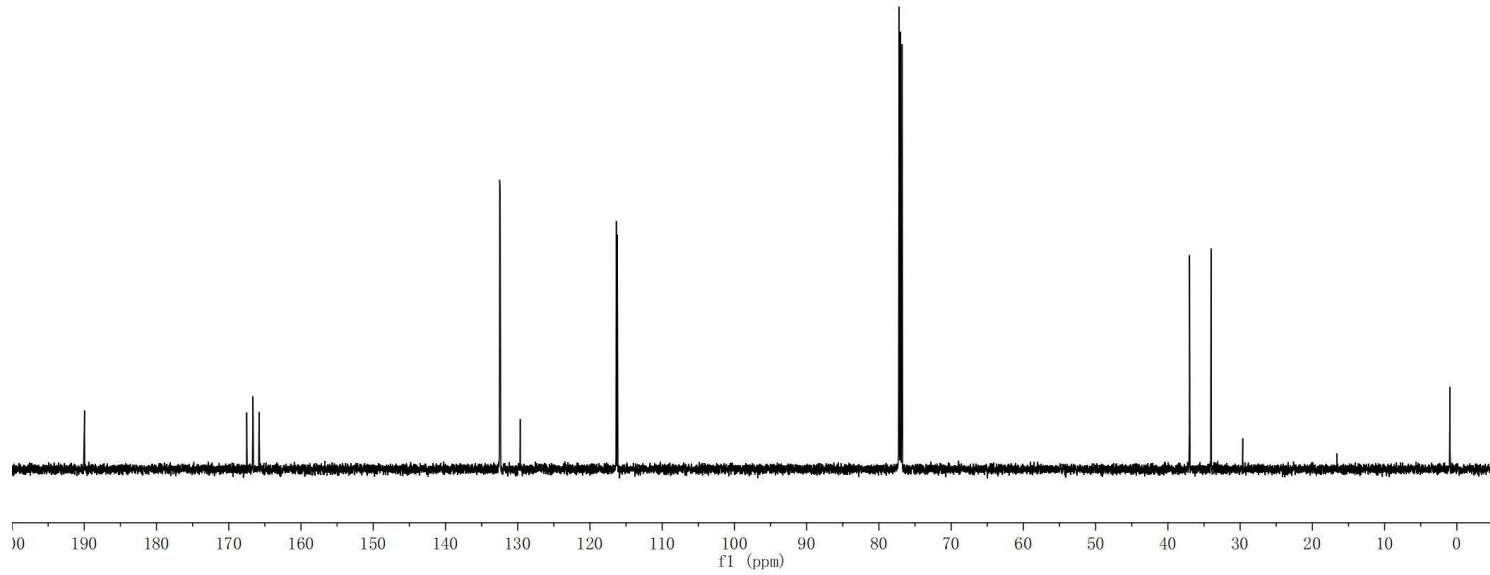
— 116.3549
— 116.2078

— 77.2116
— 76.9996
— 76.7877

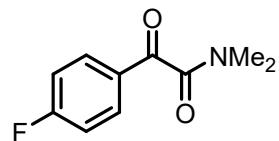
— 37.0058
— 34.0310



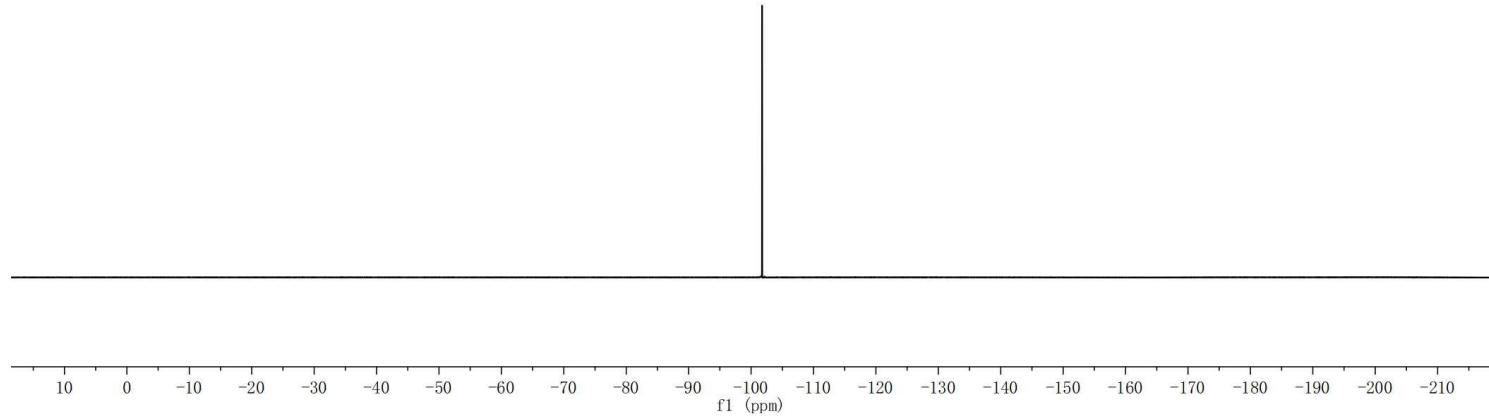
3d (^{13}C NMR 101 MHz, CDCl_3)



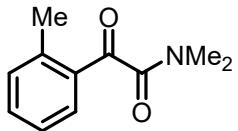
-101.755



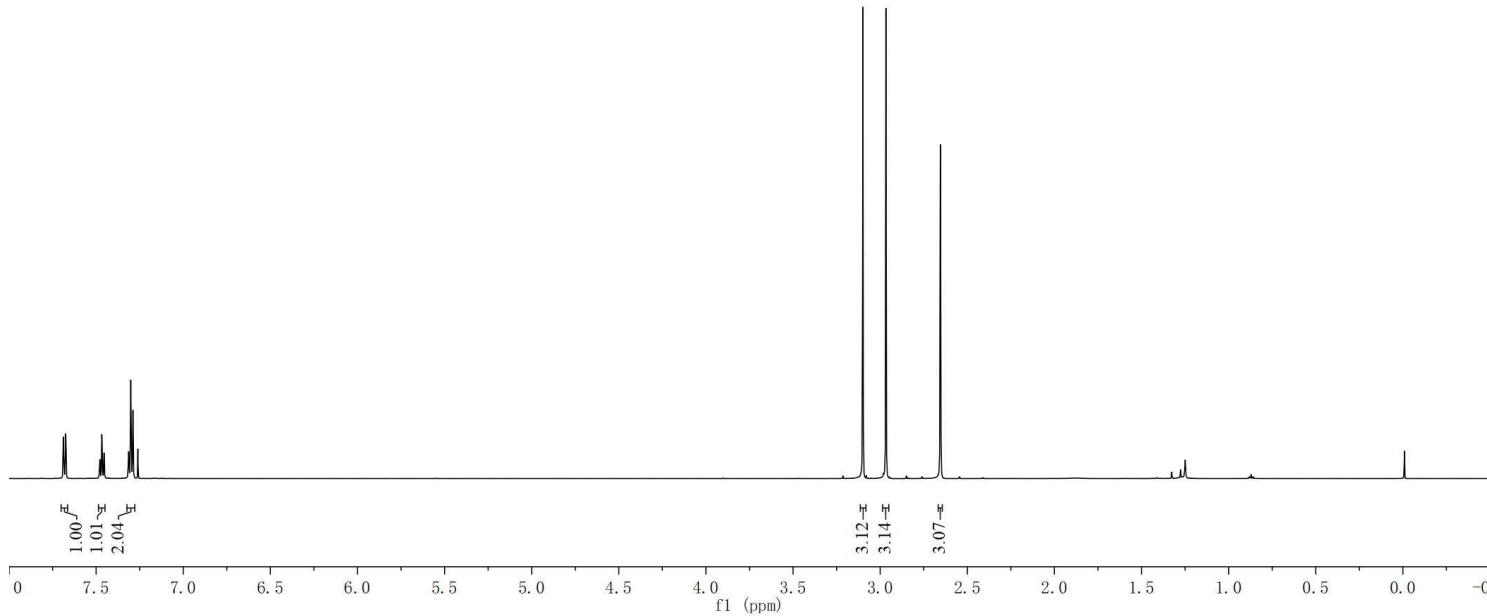
3d (¹⁹F NMR 376 MHz, CDCl₃)



7.6894
7.6867
7.6751
7.6730
7.4679
7.4655
7.4554
7.4531
7.3135
7.3013
7.2988
7.2604



3e (^1H NMR 400 MHz, CDCl_3)



—193.7083

—167.7826

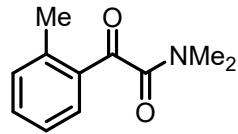
—141.3970
—133.5862
—132.5673
—132.5288
—131.5920
—126.1247

—77.2132
—77.0013
—76.7891

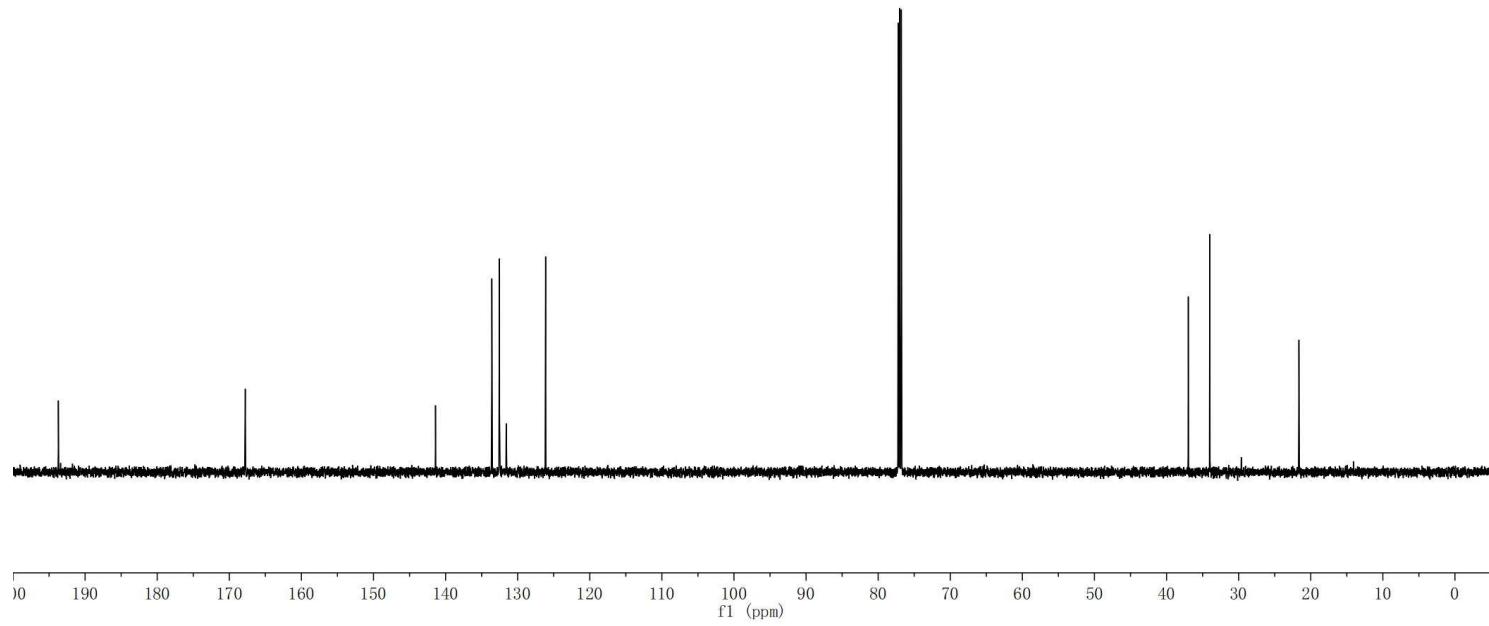
—36.9926

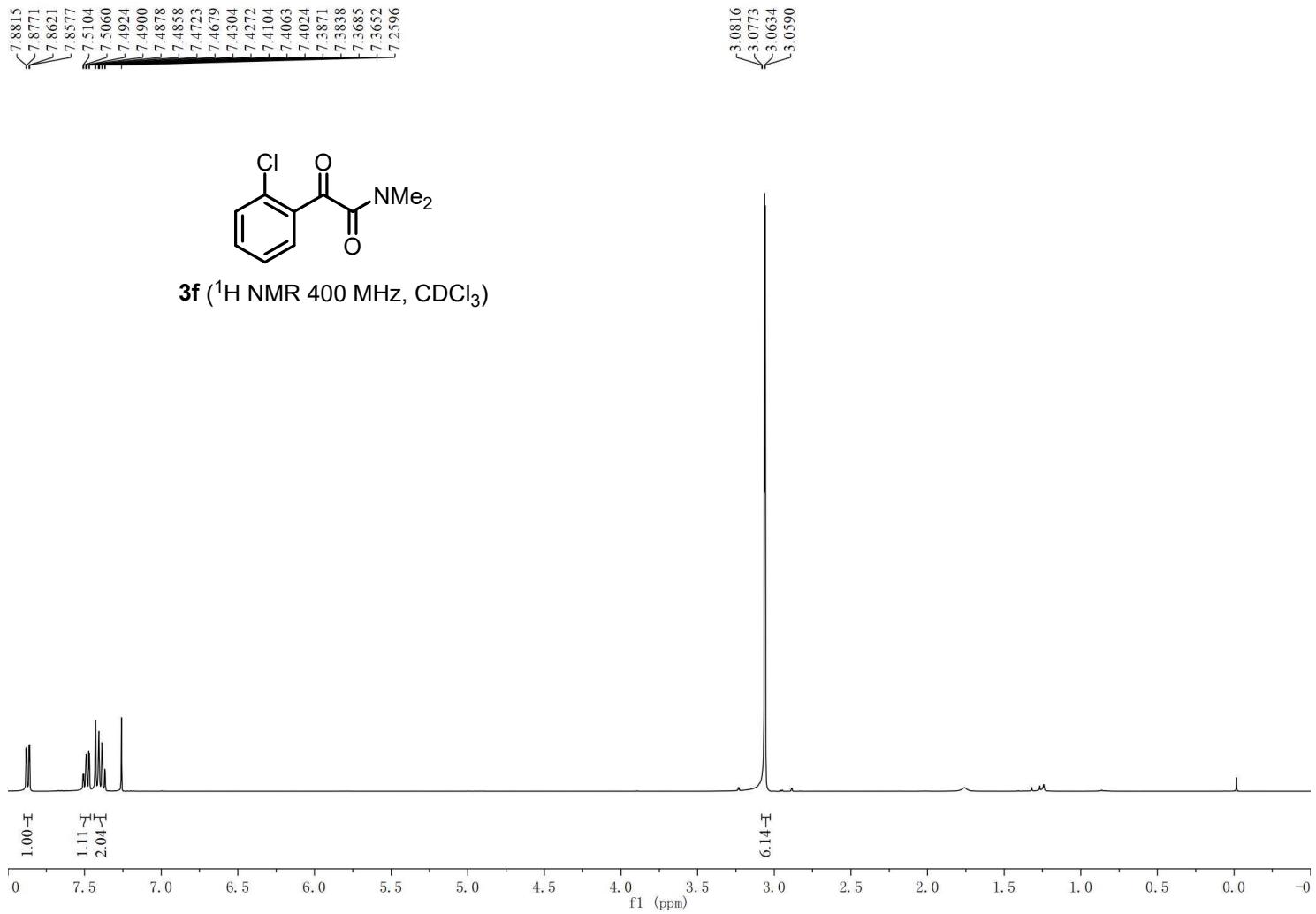
—34.0036

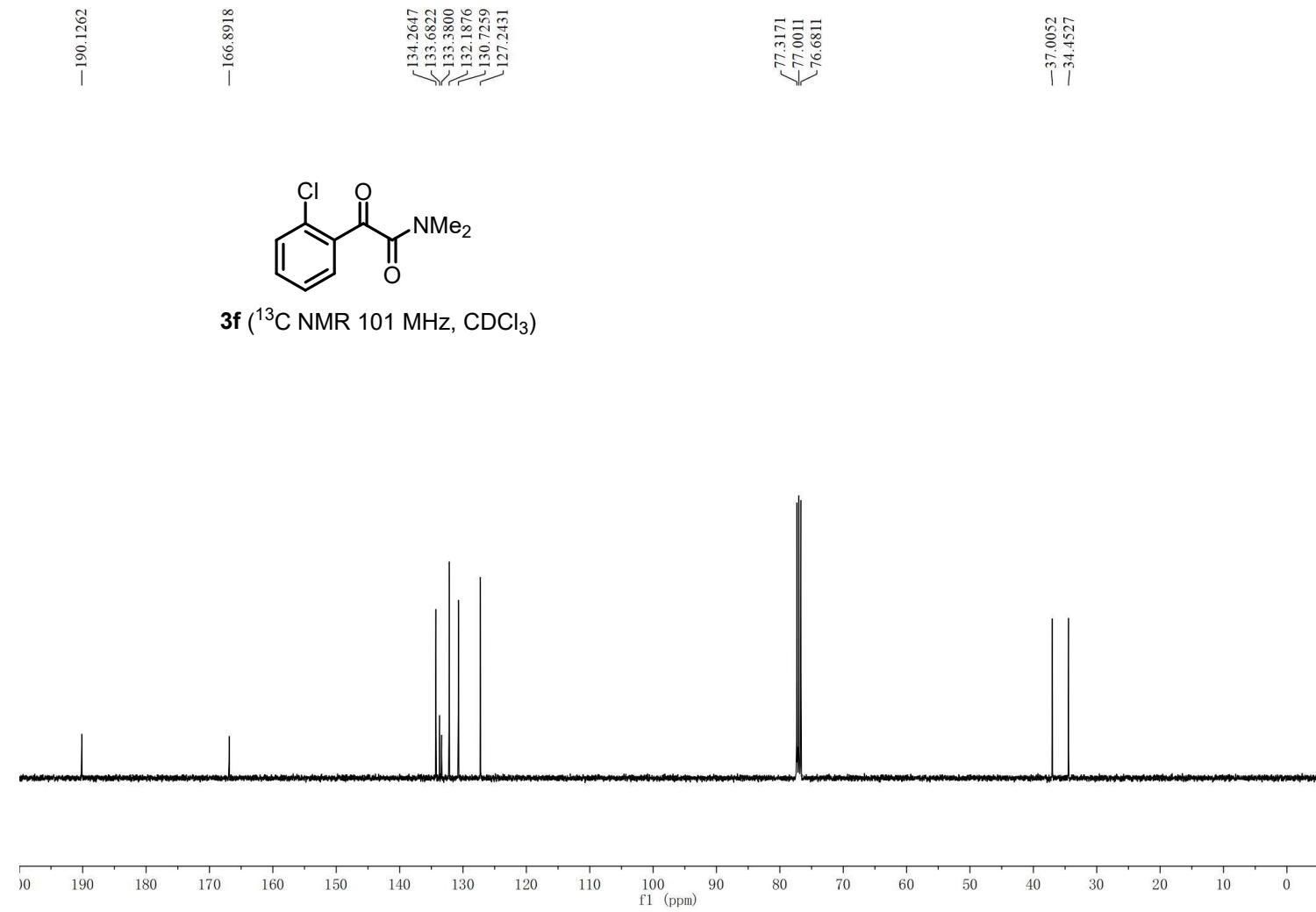
—21.6445

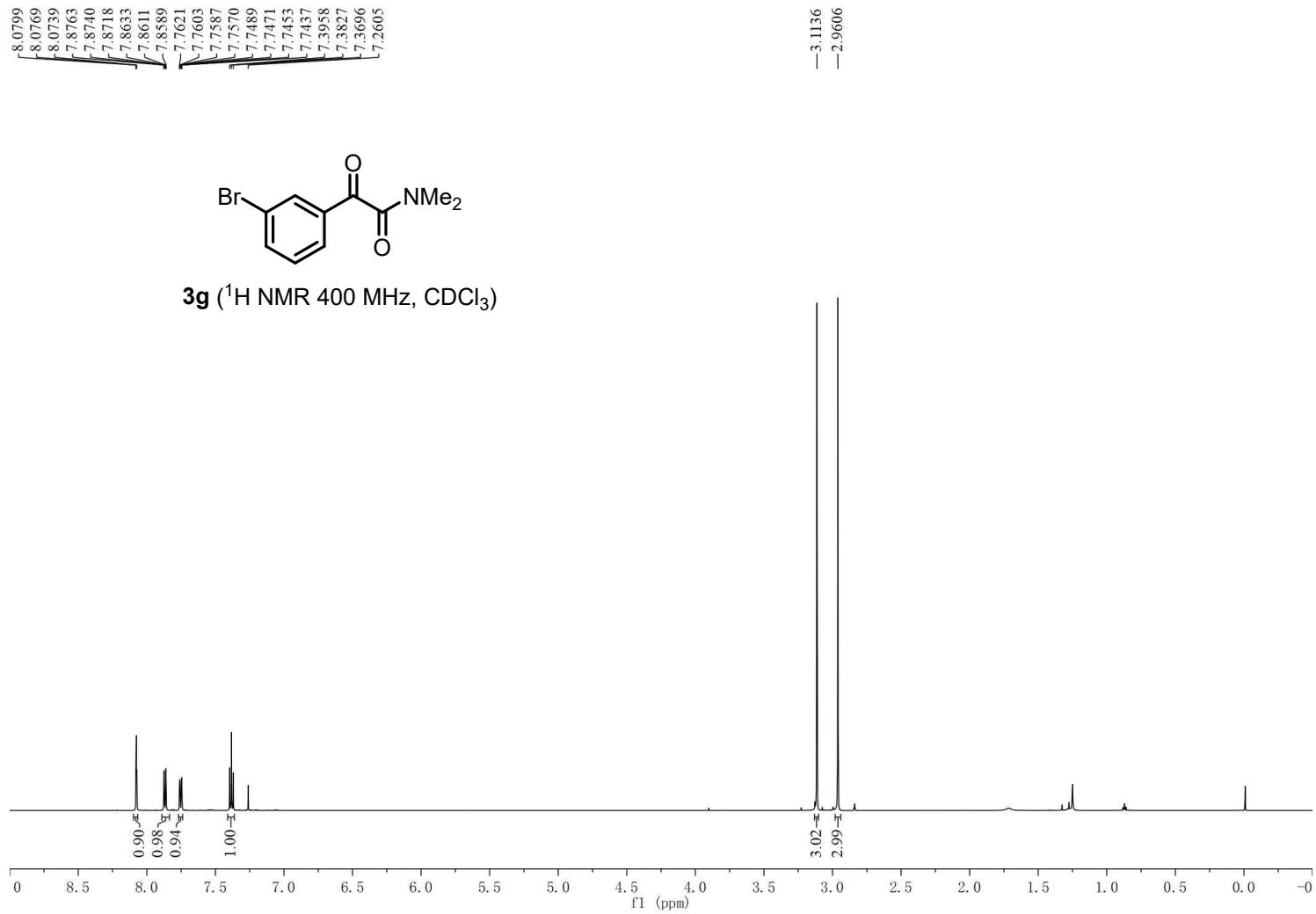


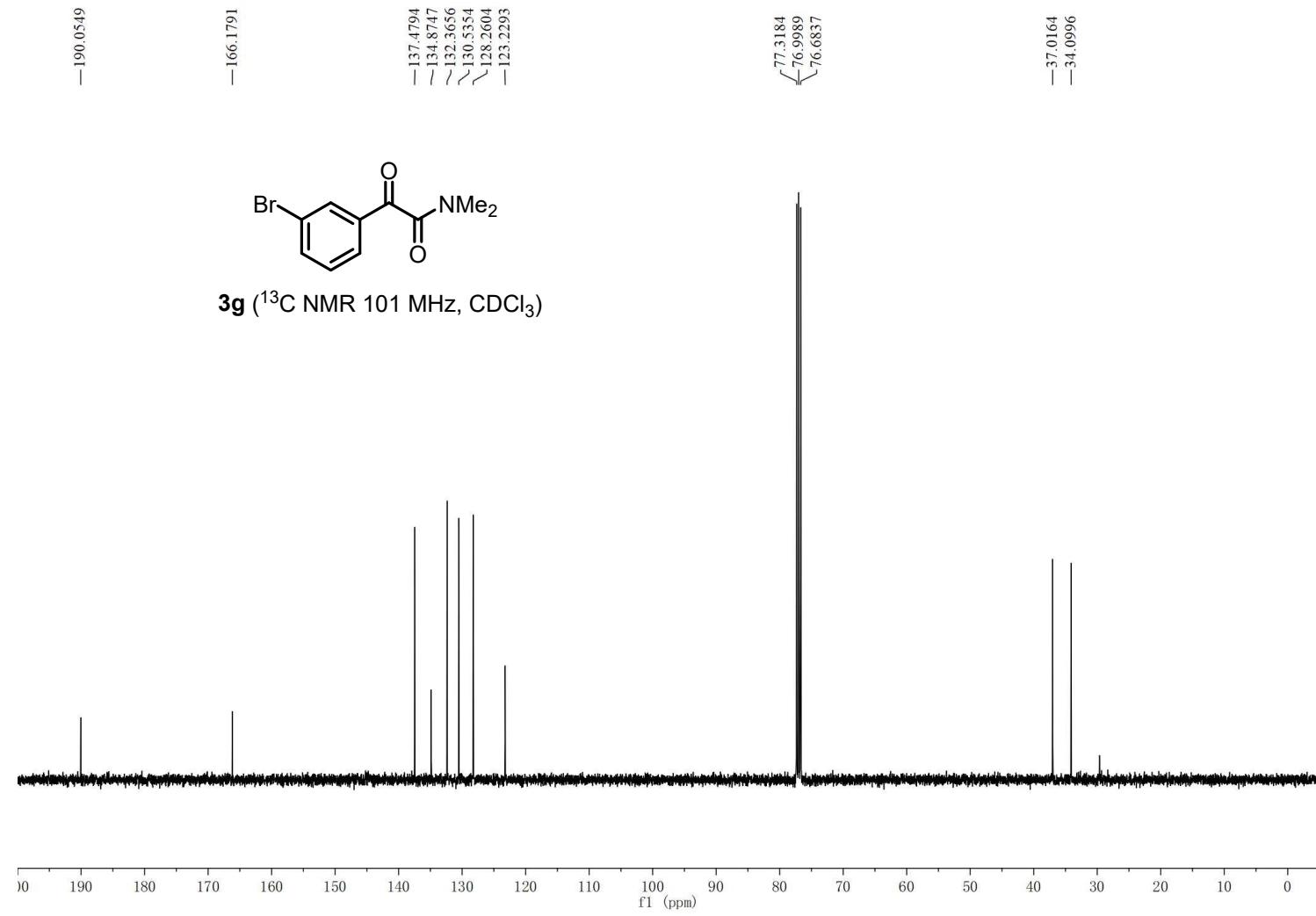
3e (^{13}C NMR 101 MHz, CDCl_3)





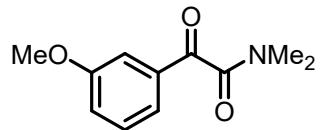




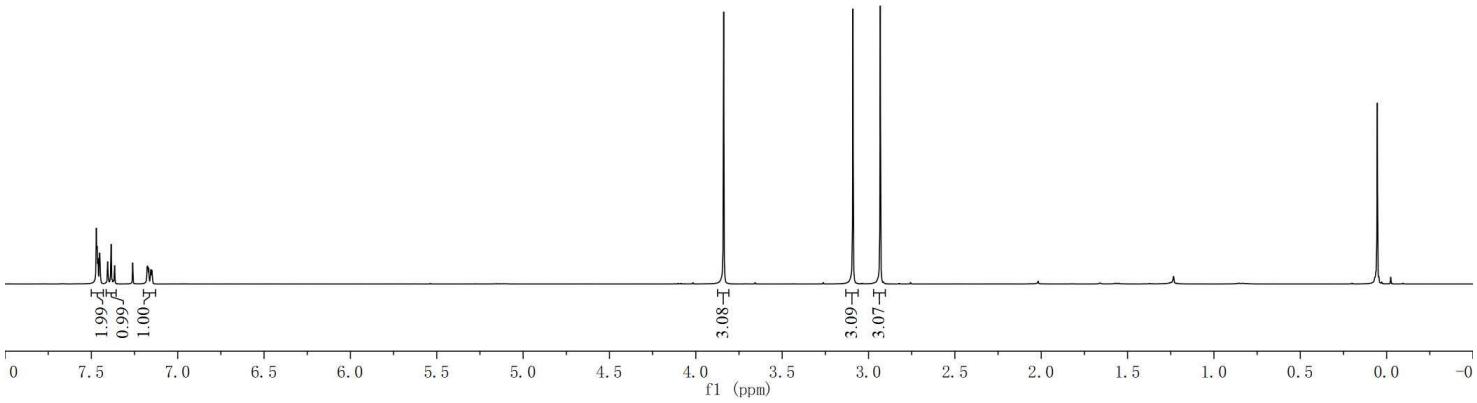


7.4713
7.4676
7.4652
7.4614
7.4542
7.4511
7.4477
7.4059
7.4025
7.3885
7.3853
7.3654
7.2606
7.1774
7.1736
7.1714
7.1678
7.1568
7.1541
7.1501
7.1474

—3.8388
—3.0903
—2.9320



3h (^1H NMR 400 MHz, CDCl_3)



—191.6153

—166.9574

—160.0695

—134.3803

—129.9850

~122.6999

~121.4698

—112.8292

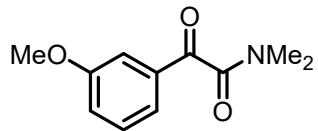
77.2116
76.9992
76.7877

—55.4626

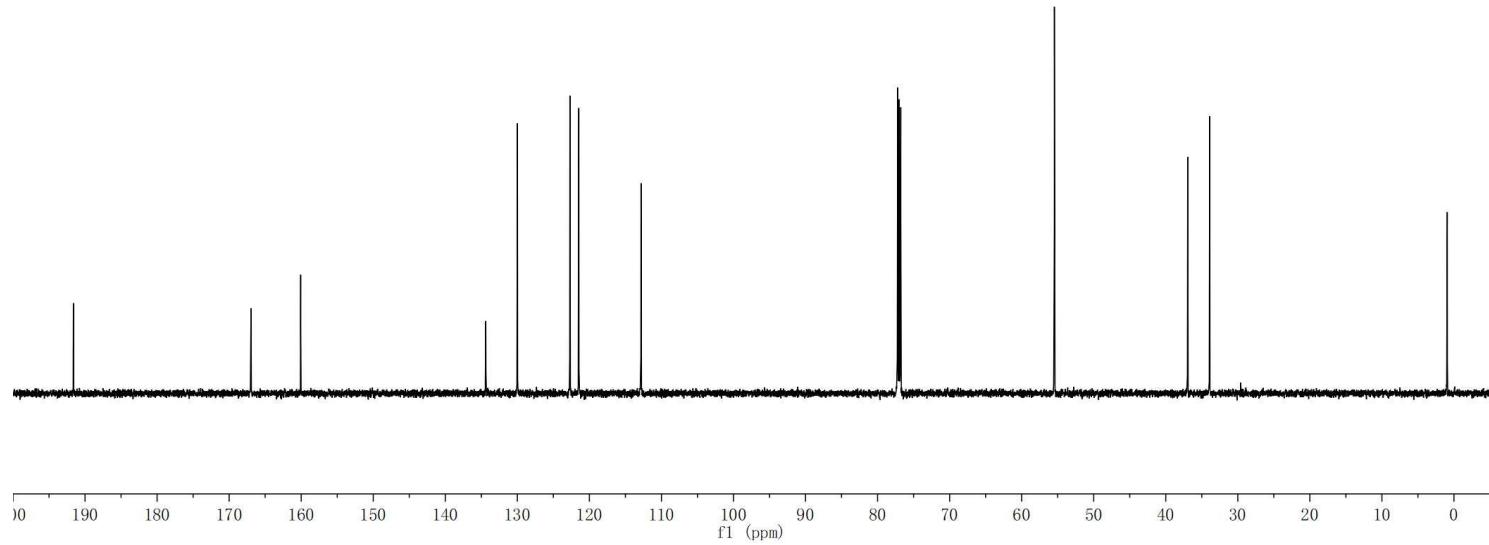
—36.9531

—33.9047

—0.9309

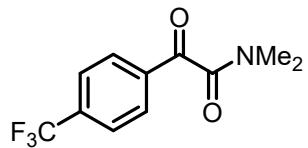


3h (¹³C NMR 101 MHz, CDCl₃)

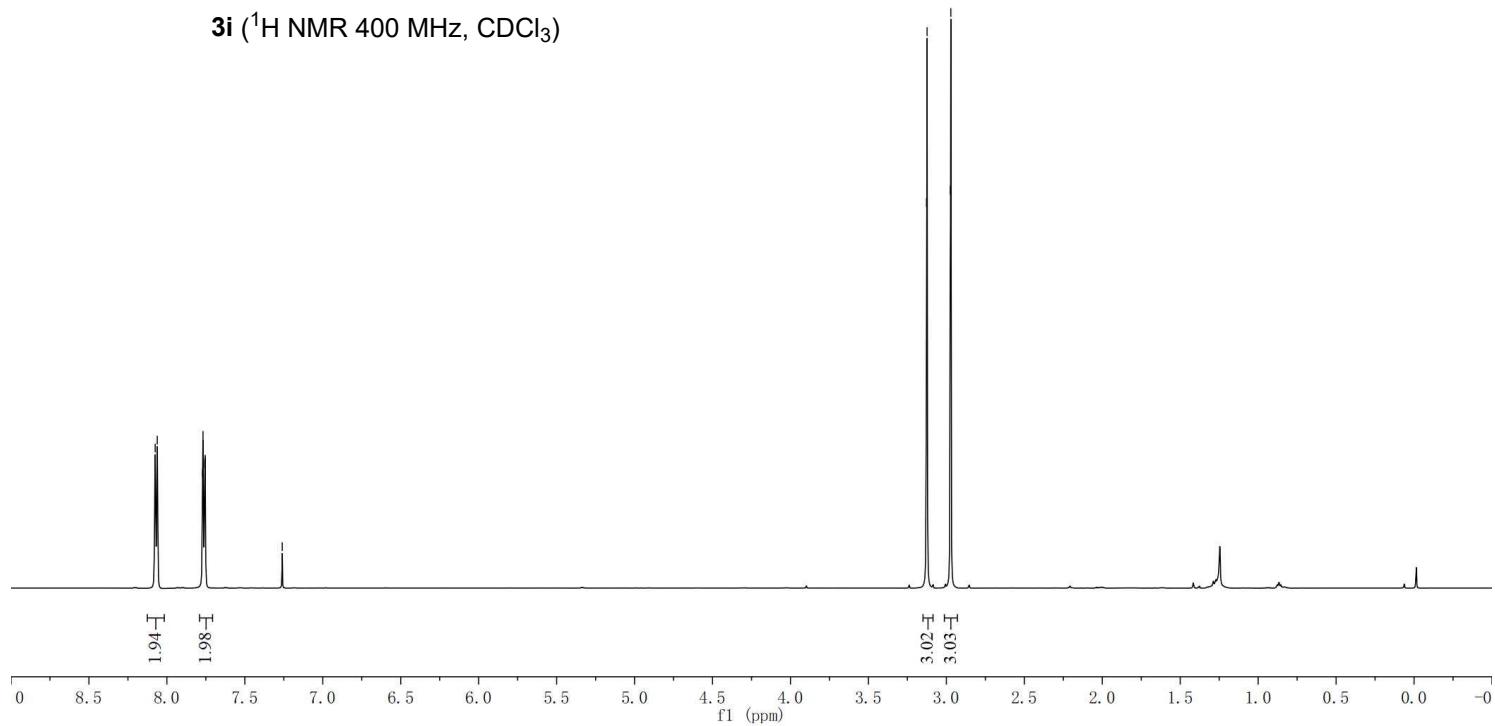


8.0751
8.0617
7.7716
7.7683
7.7574
7.7542
-7.2599

3.1274
3.1237
2.9746
2.9713



3i (^1H NMR 400 MHz, CDCl_3)

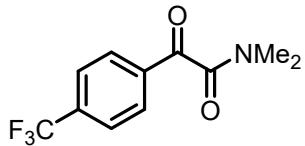


—190.3162

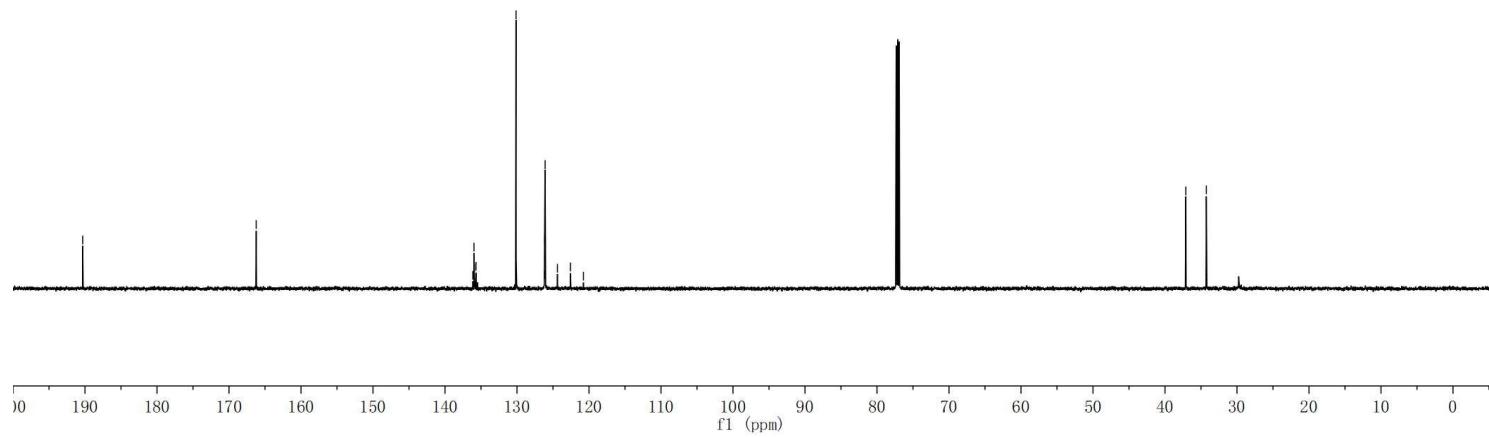
—166.2238

136.1306
135.9735
135.6973
130.1343
126.1938
126.1427
126.1184
126.0911
126.0668
124.3863
122.5804
120.7700

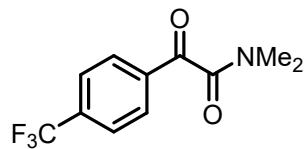
—37.1055
—34.2621



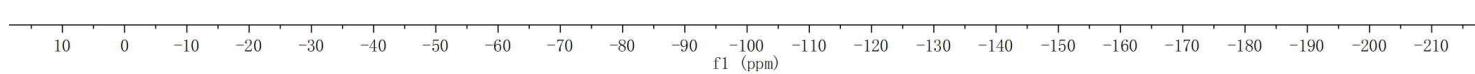
3i (^{13}C NMR 101 MHz, CDCl_3)



—63.3500

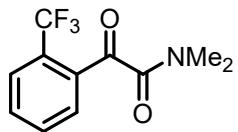


3i (¹⁹F NMR 376 MHz, CDCl₃)

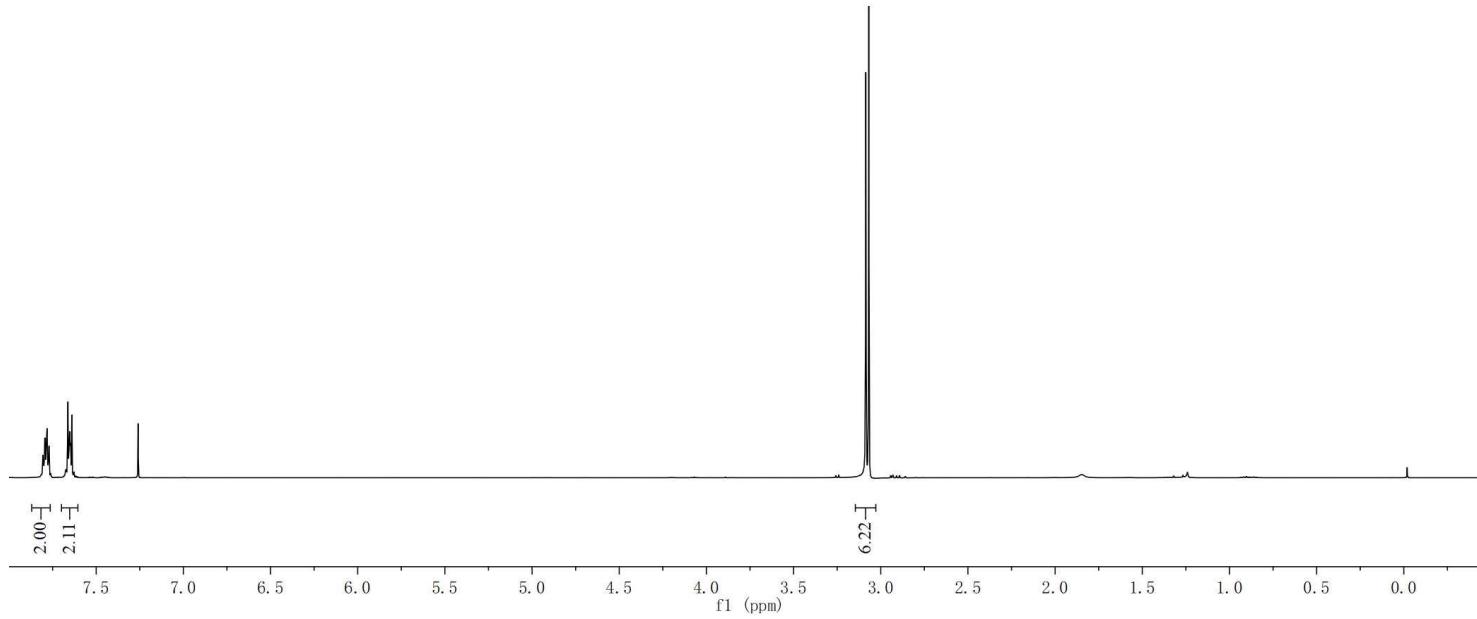


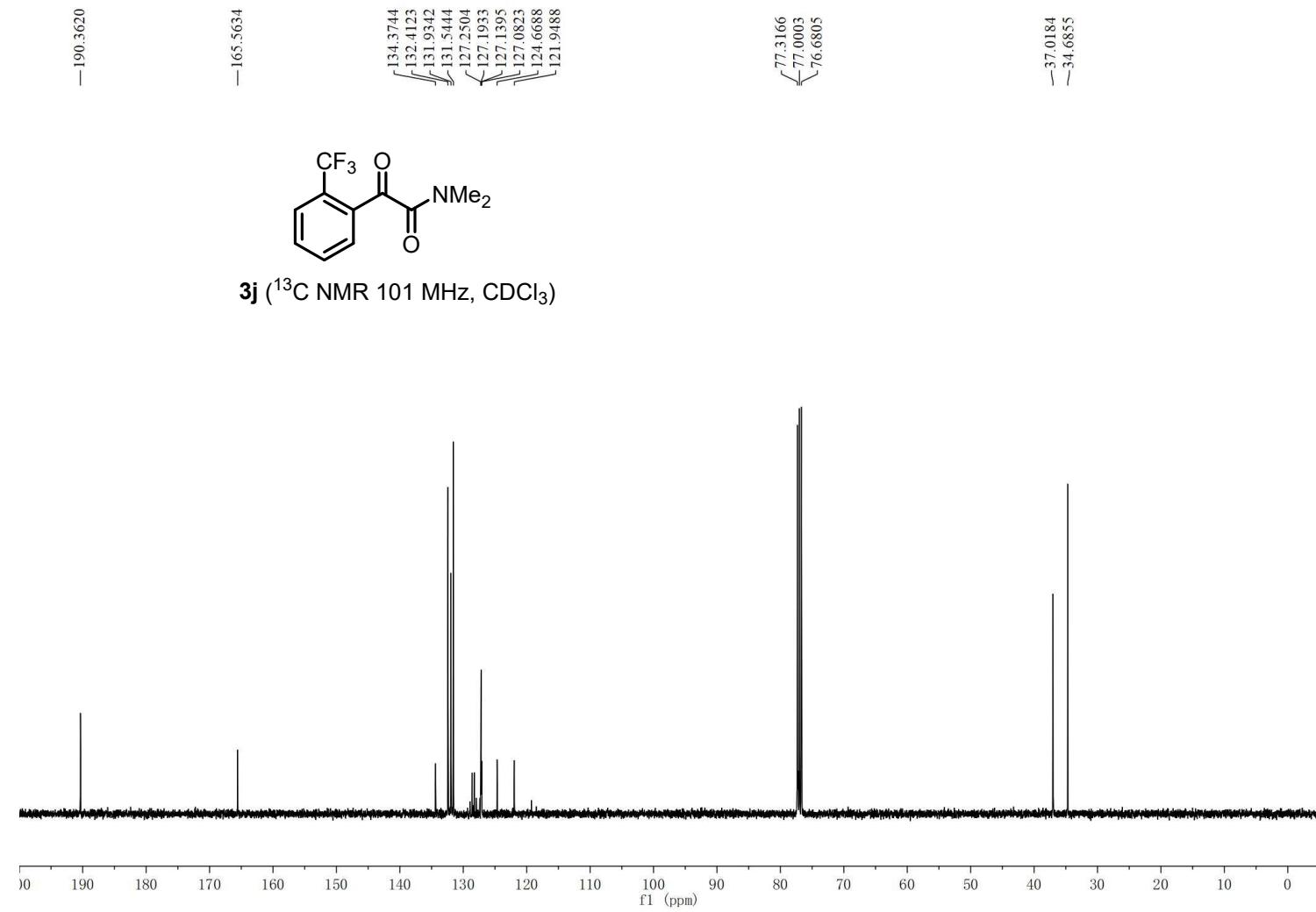
7.8072
7.8050
7.7981
7.7950
7.7917
7.7850
7.7817
7.7778
7.7754
7.7707
7.6622
7.6570
7.6534
7.6527
7.6497
7.6466
7.6396
7.2596

3.0858
3.0686

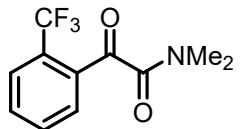


3j (^1H NMR 400 MHz, CDCl_3)

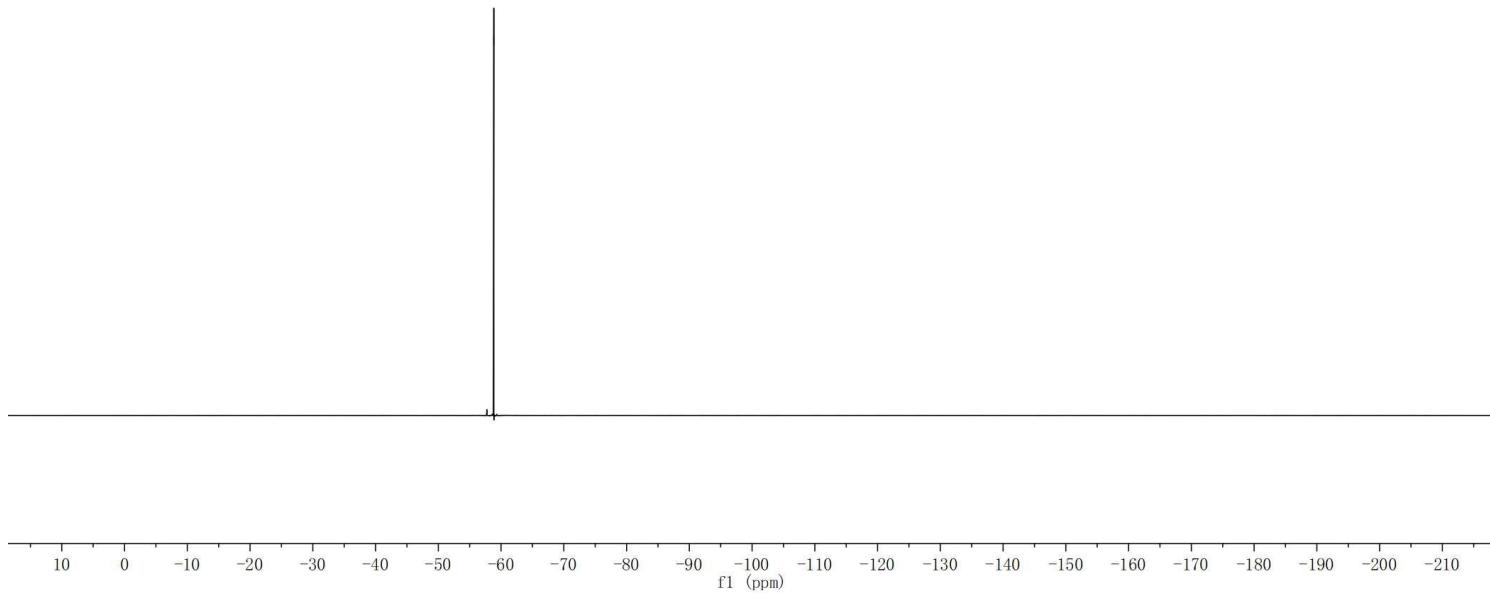




—
—58.8430



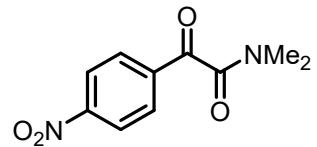
3j (^{19}F NMR 376 MHz, CDCl_3)



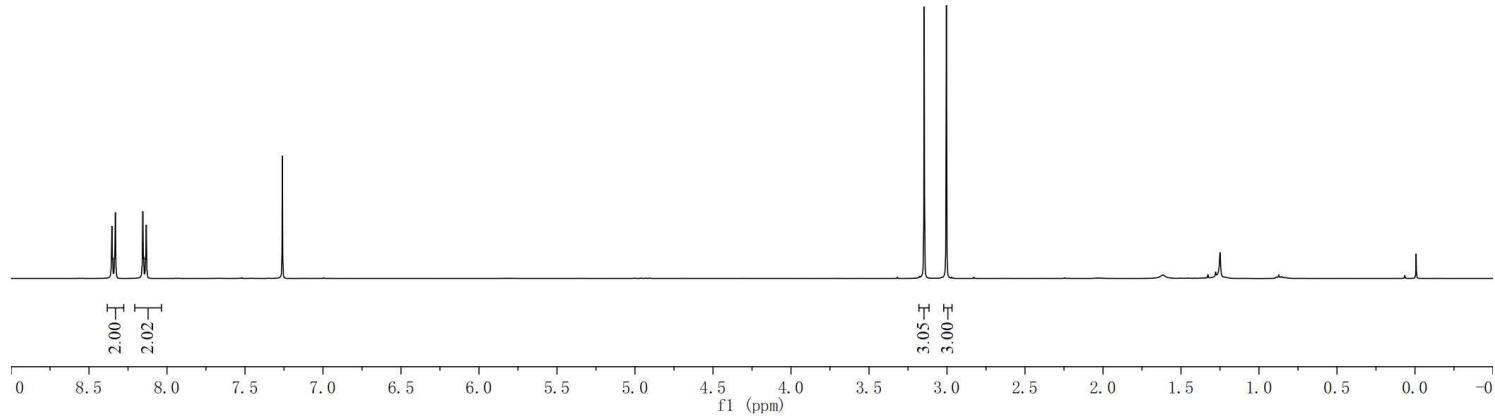
8.3533
8.3312
8.1551
8.1330

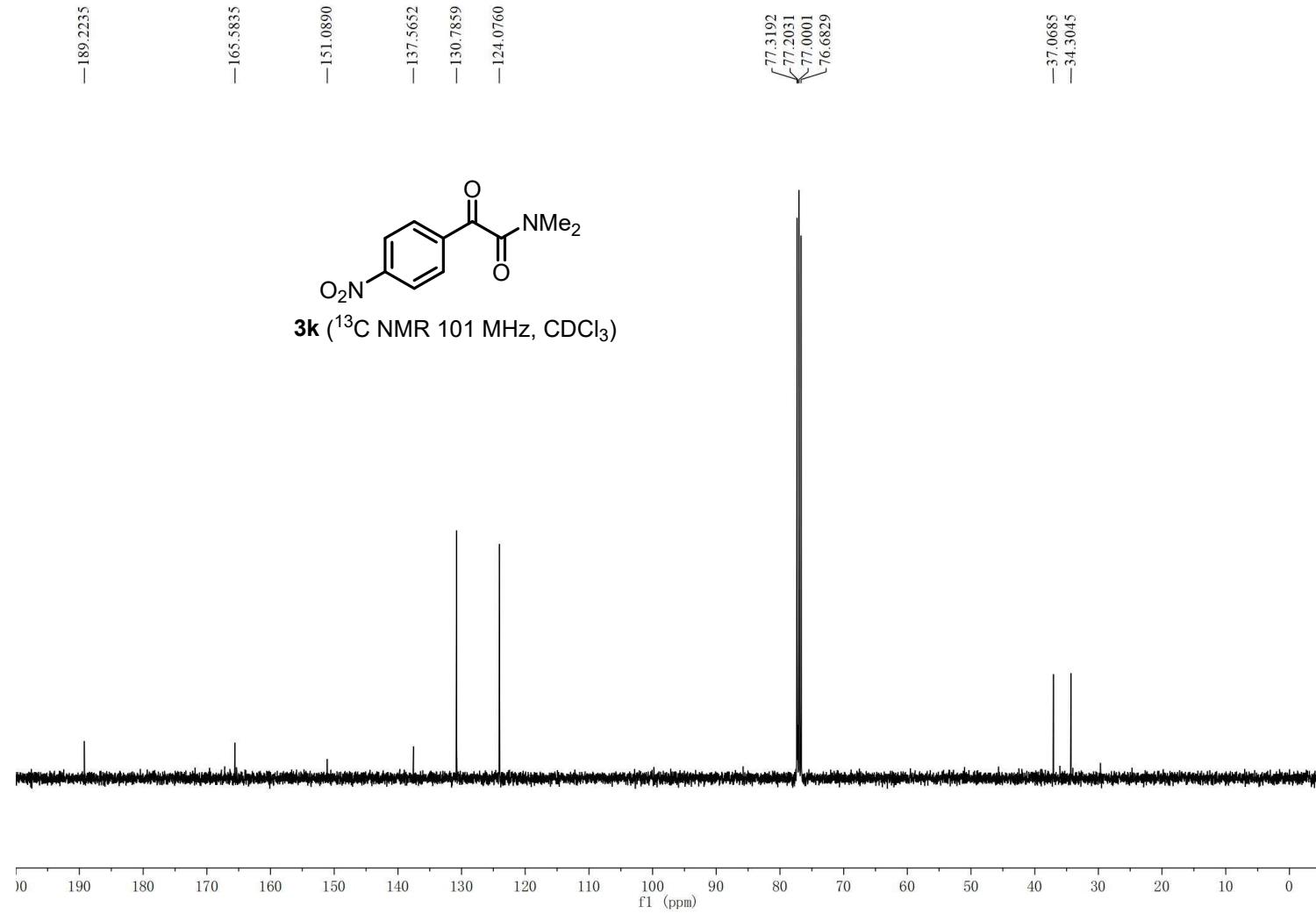
—7.2604

—3.1462
—3.0038



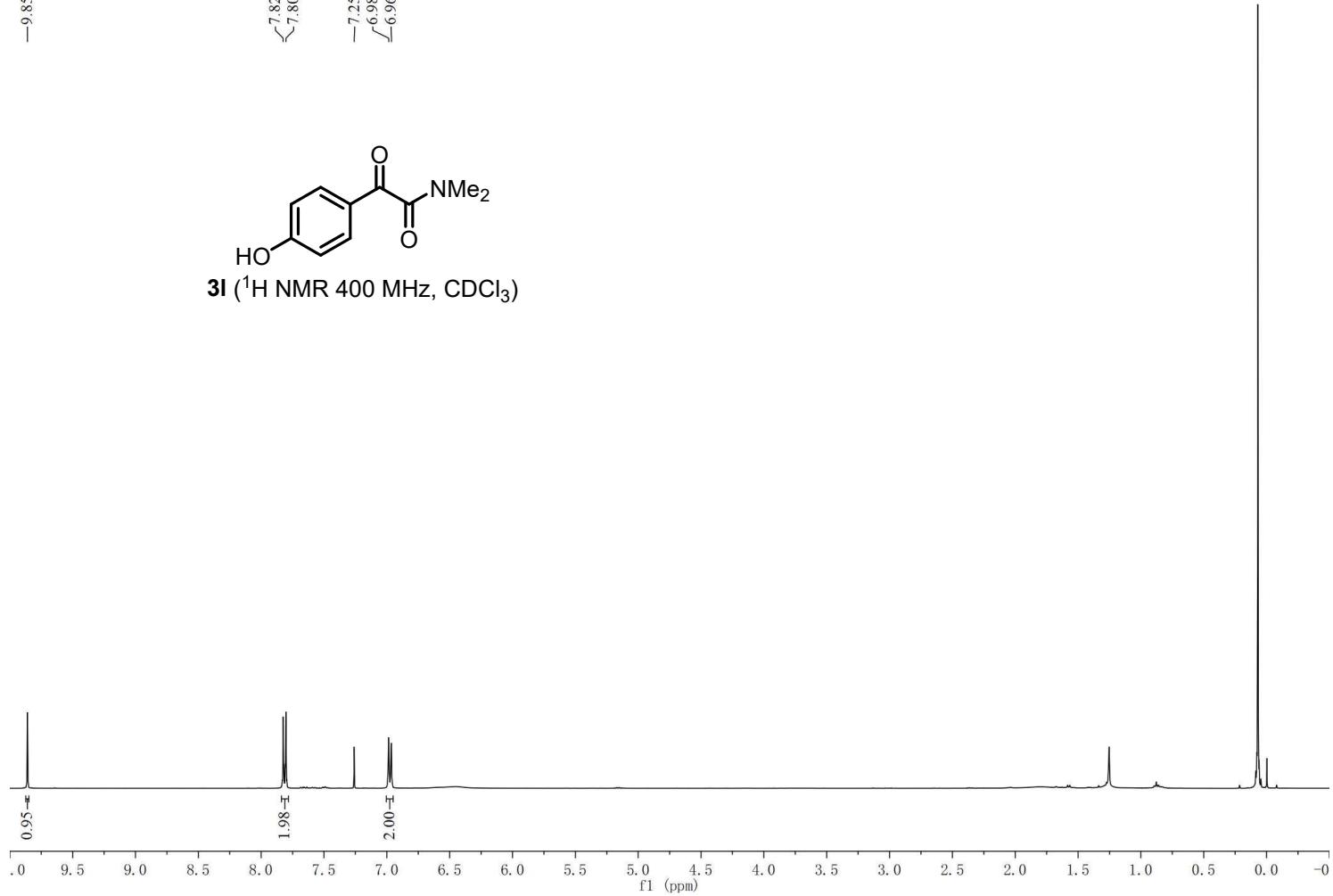
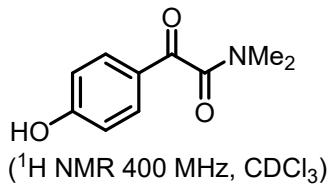
3k (^1H NMR 400 MHz, CDCl_3)

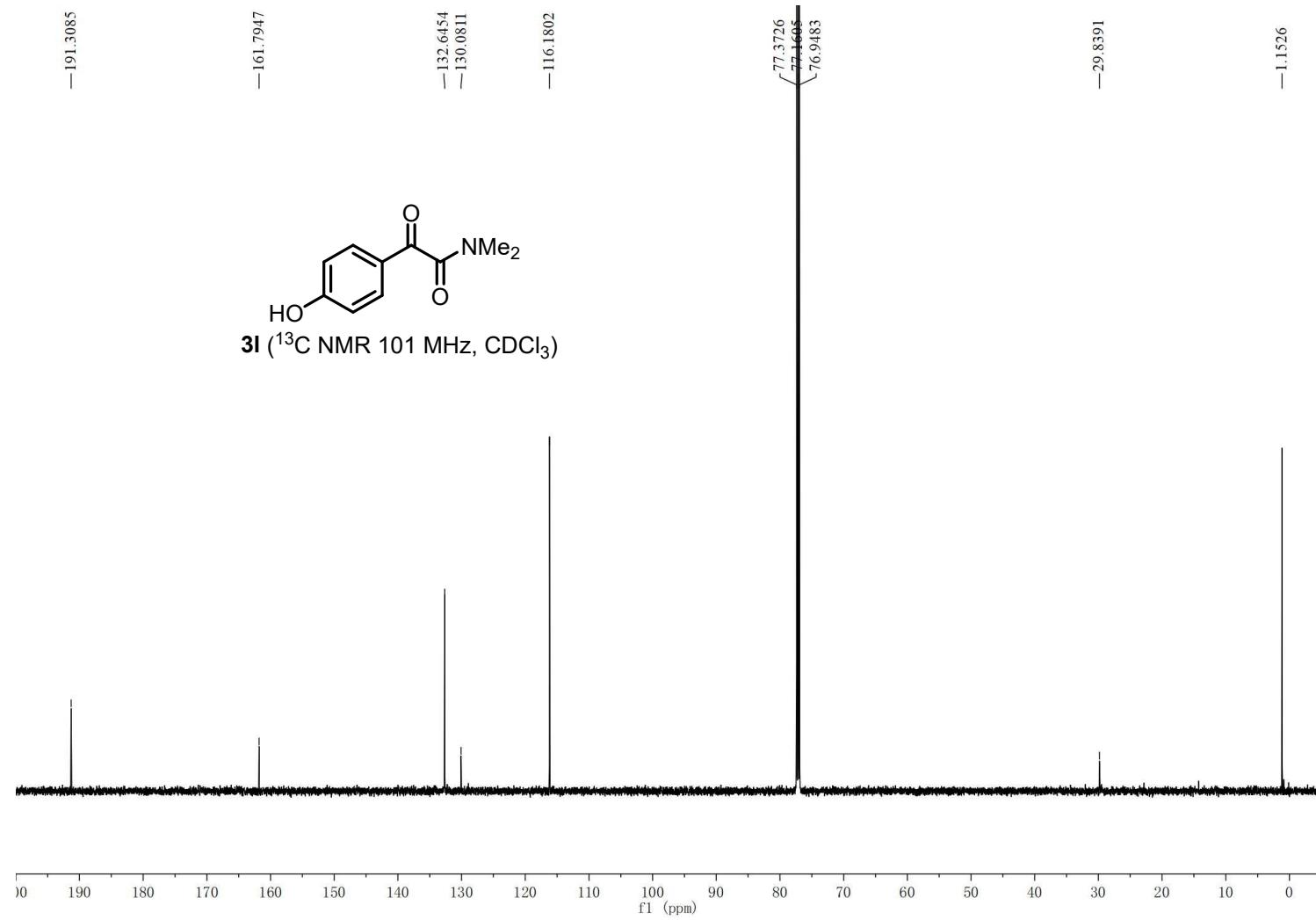




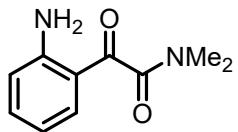
-9.8599

7.8247
7.8031
-7.2596
6.9846
6.9630

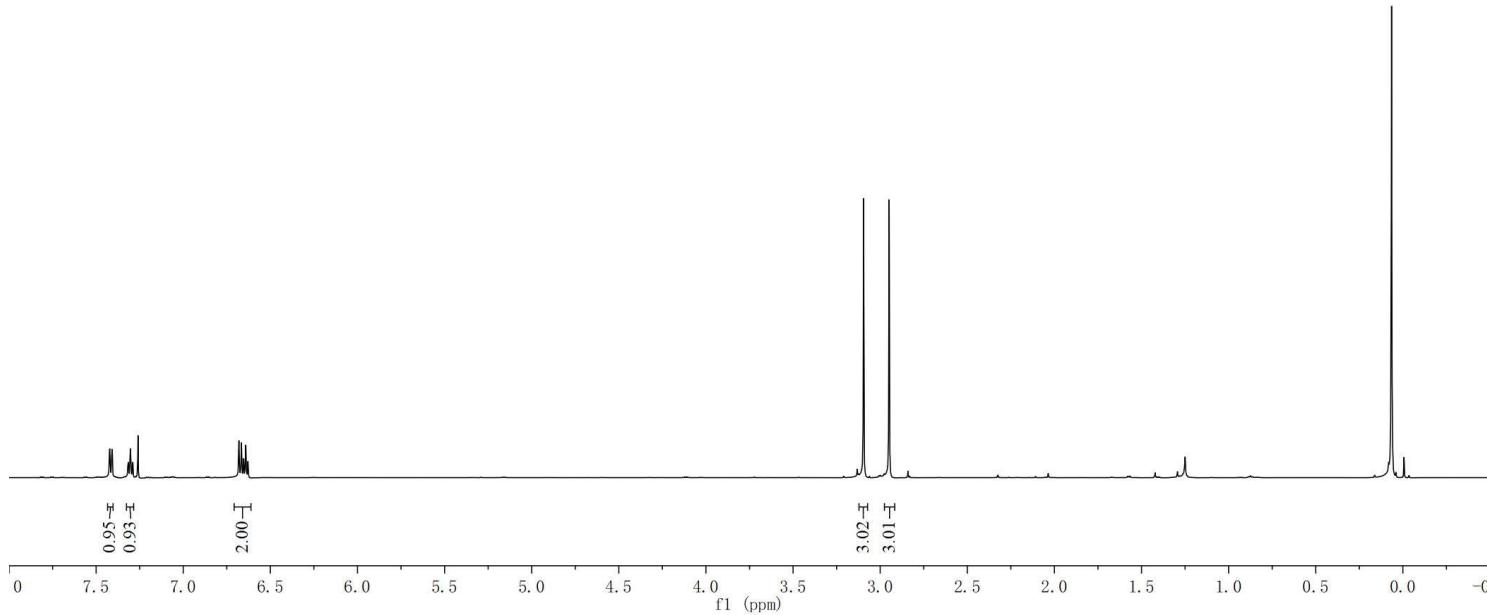


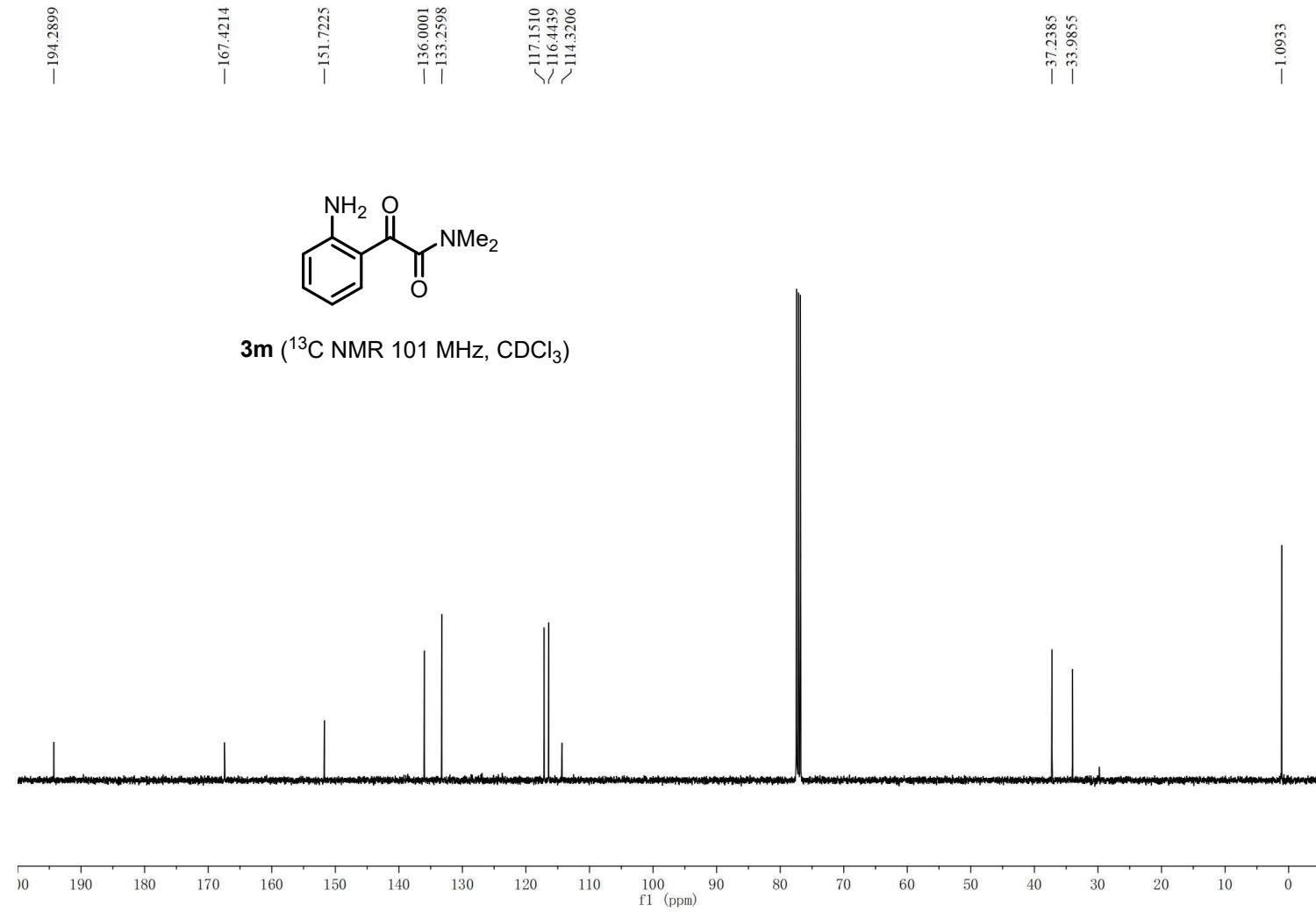


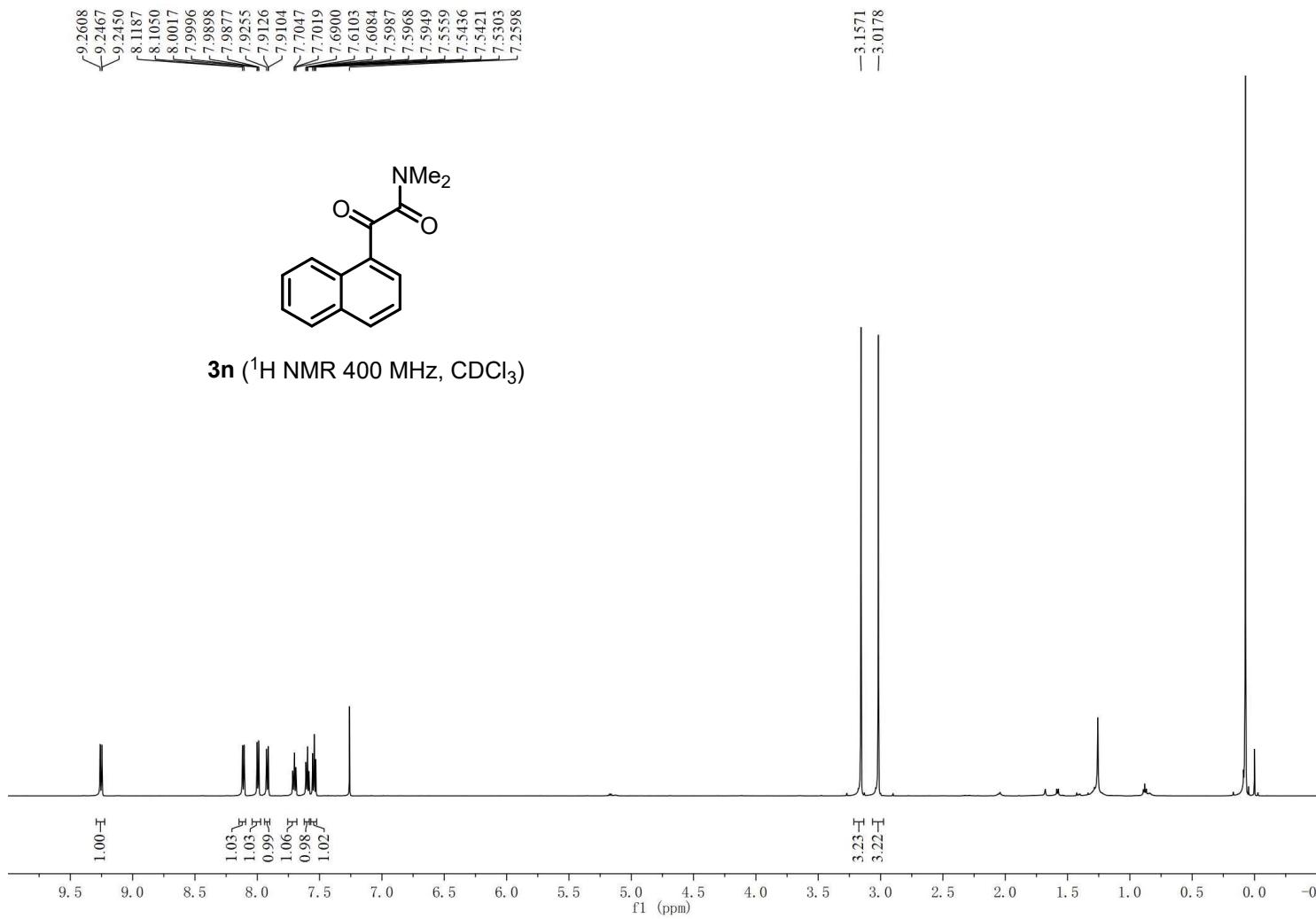
7.4235
7.4208
7.4101
7.4074
7.3172
7.3146
7.3056
7.3030
7.3004
7.2916
7.2889
7.2596
6.6803
6.6663
6.6547
6.6529
6.6431
6.6413
6.6392
6.6294
6.6275

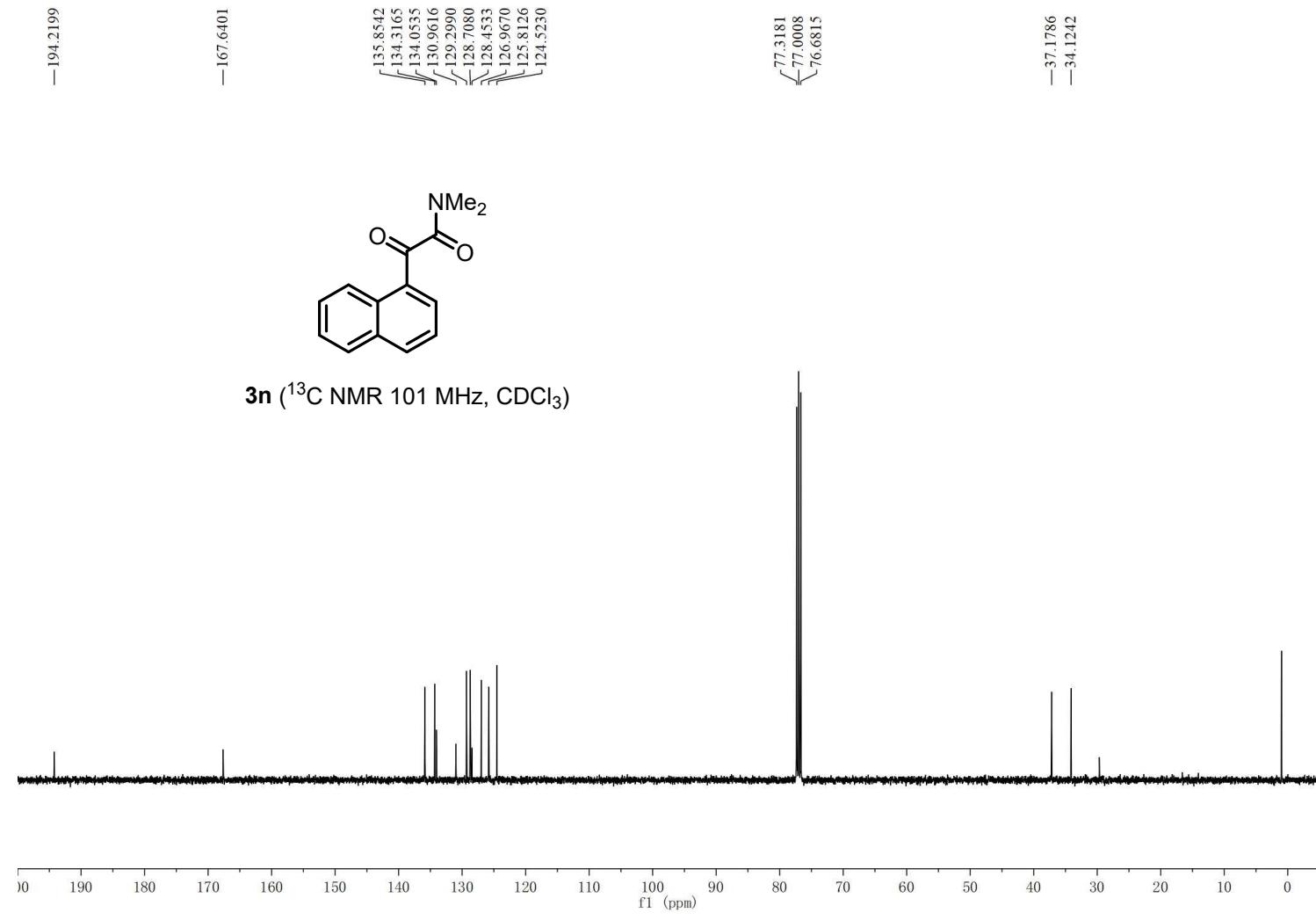


3m (^1H NMR 400 MHz, CDCl_3)



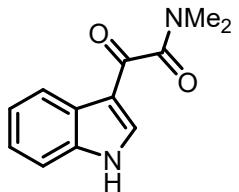




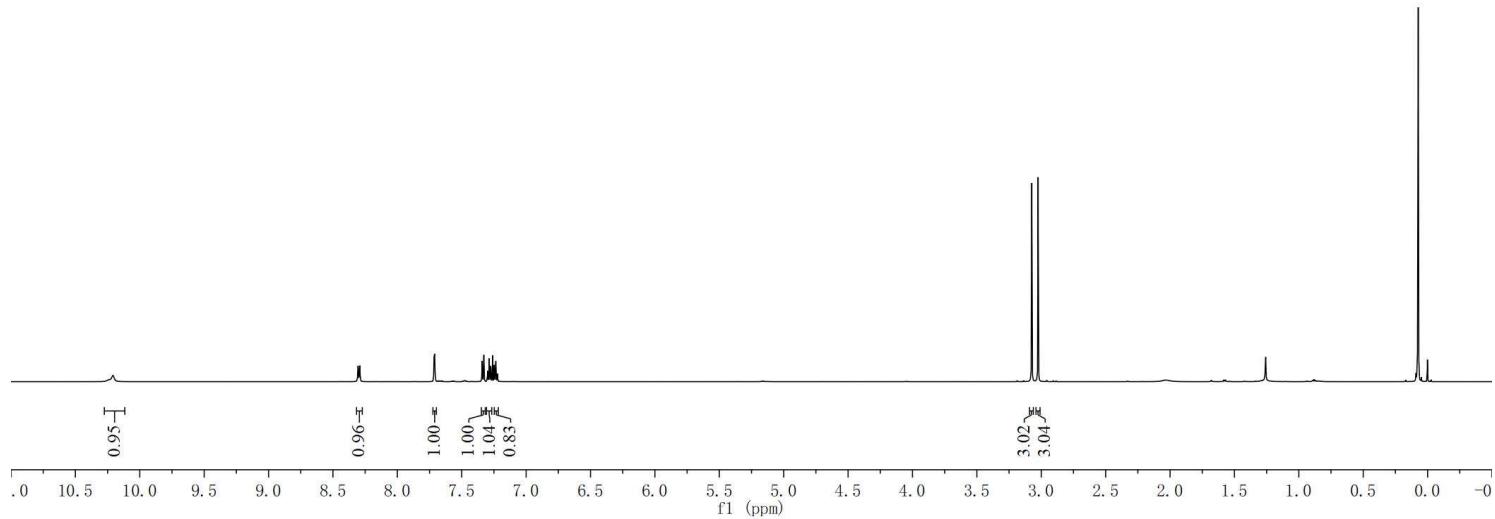


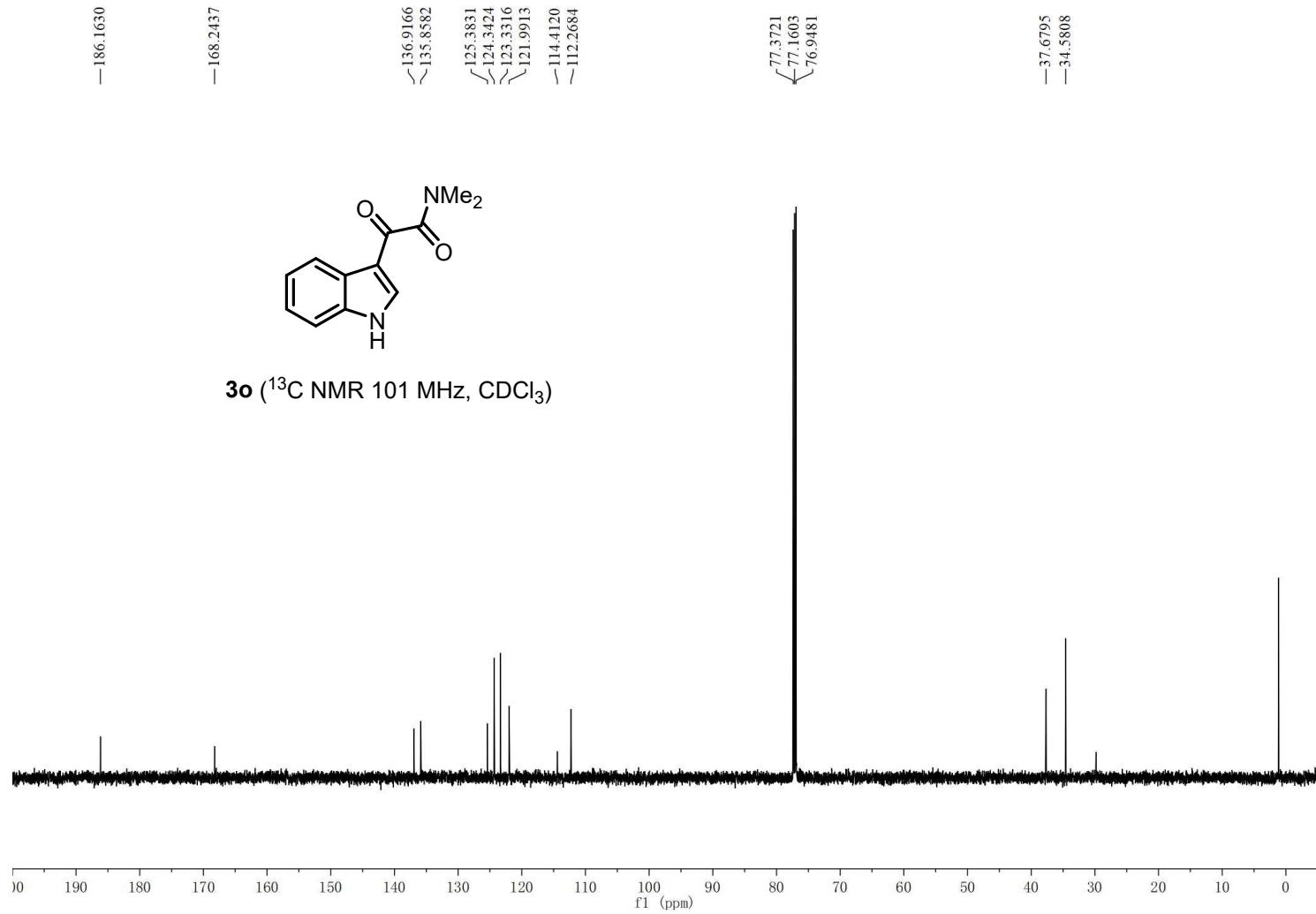
<-10.2115
<-10.2055

<-8.3052
<-8.2922
<-7.7154
<-7.7102
<-7.7059
<-7.3436
<-7.3420
<-7.3402
<-7.3286
<-7.3266
<-7.3003
<-7.2985
<-7.2882
<-7.2865
<-7.2753
<-7.2734
<-7.2605
<-7.2493
<-7.2471
<-7.2373
<-7.2354
<-7.2335
<-7.2237
<-7.2216



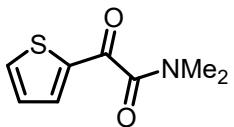
3o (^1H NMR 400 MHz, CDCl_3)



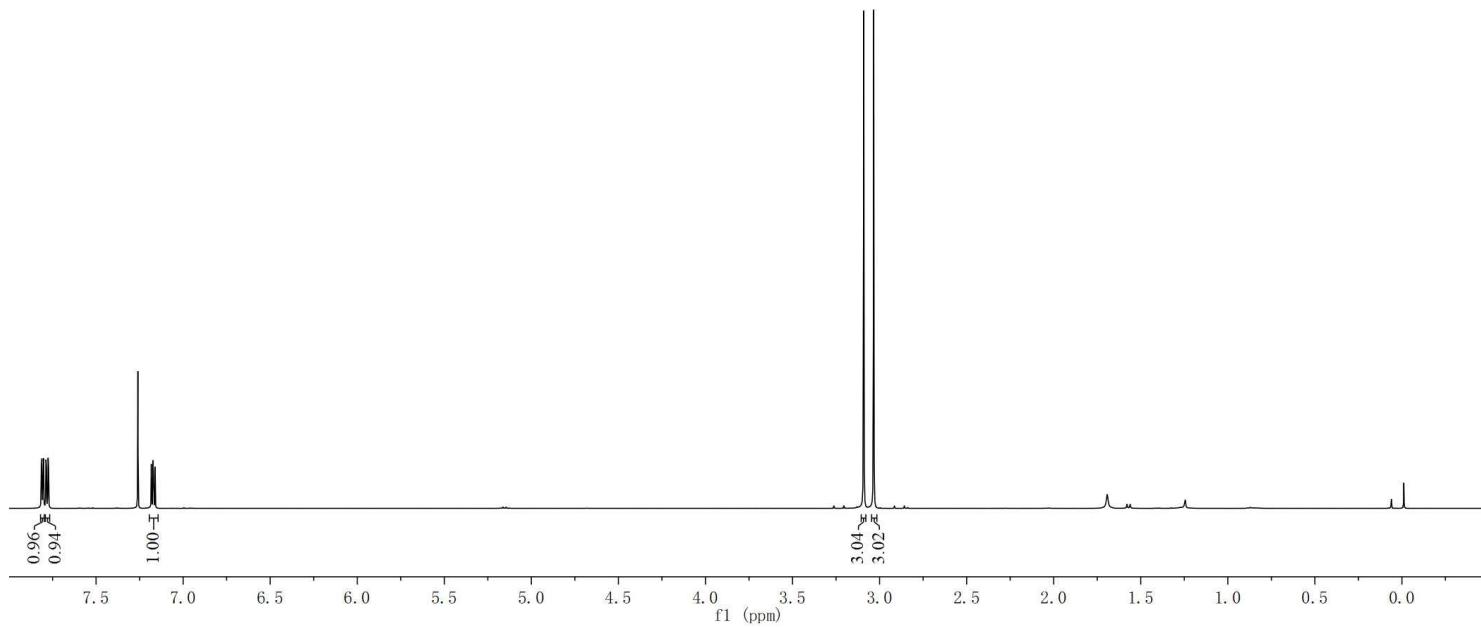


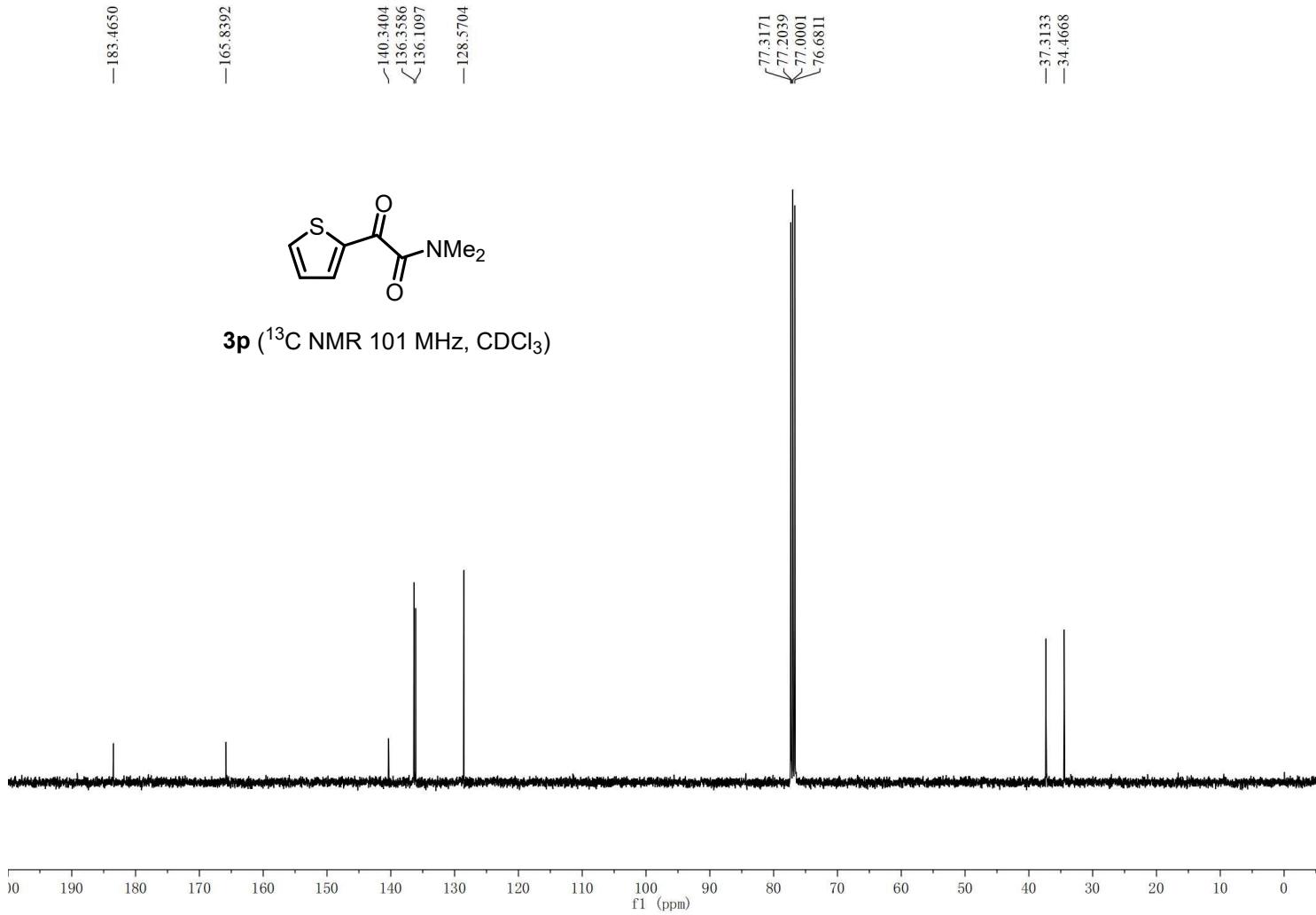
7.8141
7.8113
7.8046
7.8016
7.7879
7.7850
7.7757
7.7728
7.2599
7.1827
7.1730
7.1704
7.1608

~3.0910
~3.0339



3p (^1H NMR 400 MHz, CDCl_3)

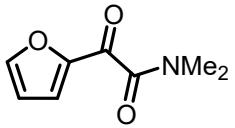




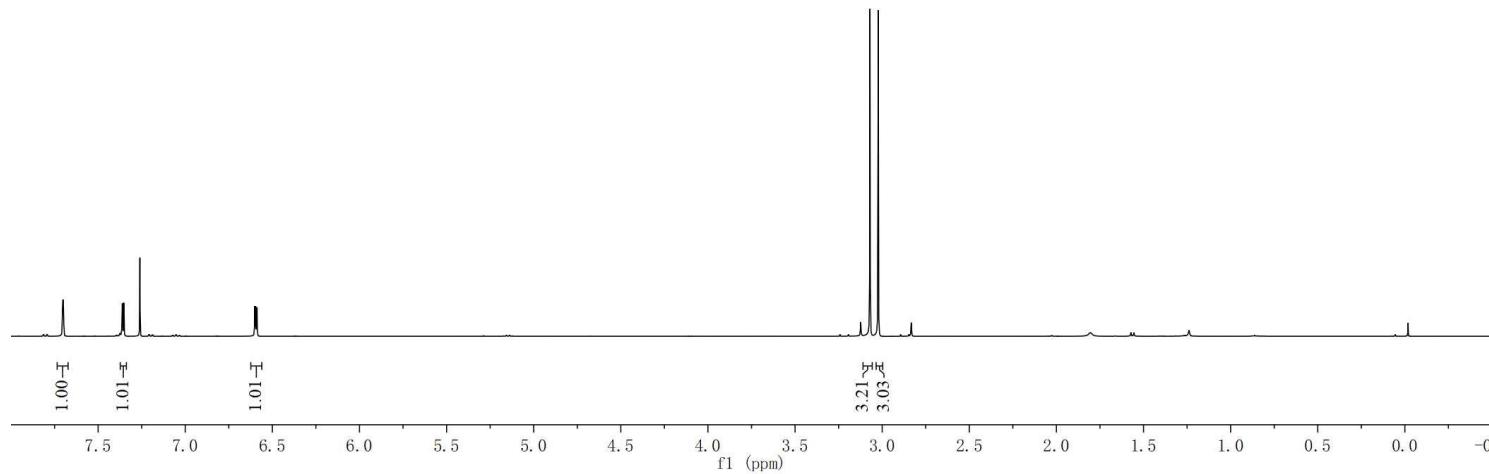
7.7026
7.7002
7.6989
7.3610
7.3537
7.3518
7.2603

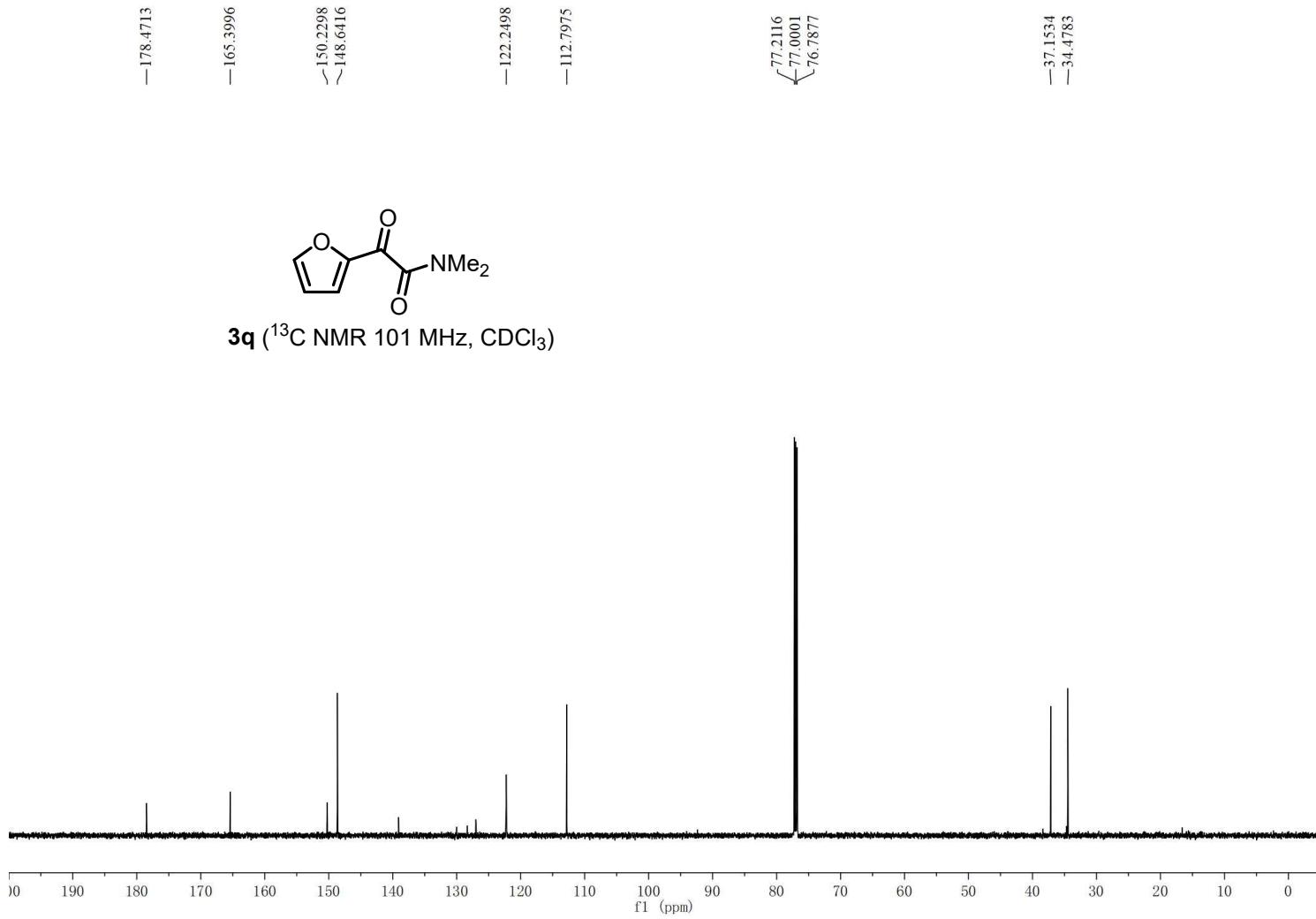
6.6012
6.5970
6.5921
6.5880

3.0701
3.0227



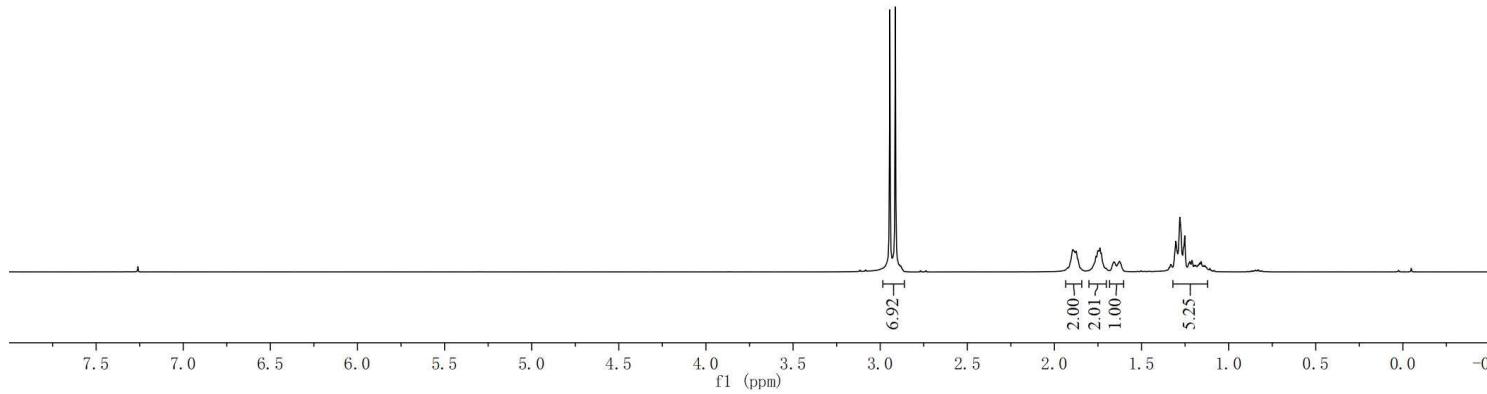
3q (^1H NMR 400 MHz, CDCl_3)

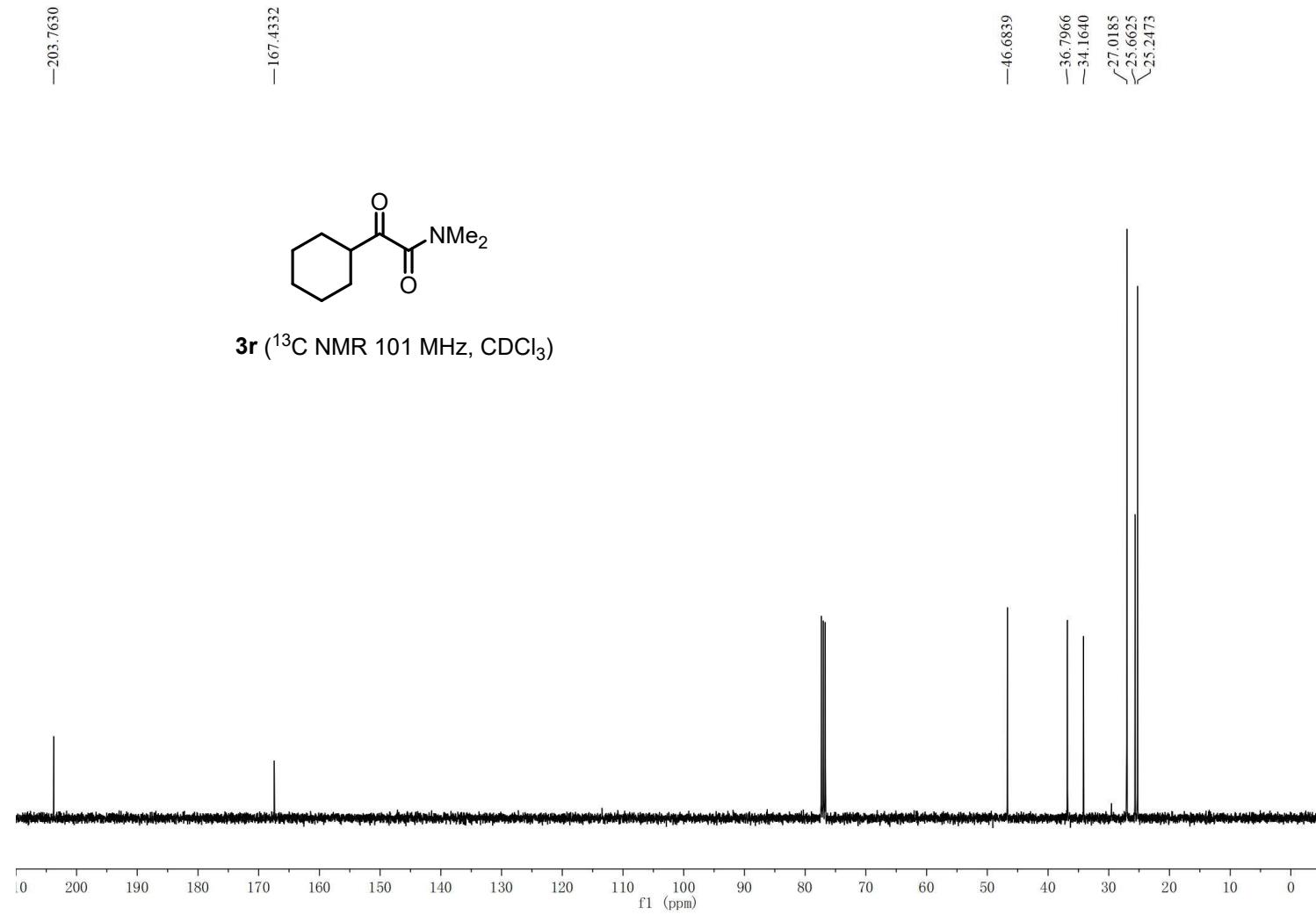




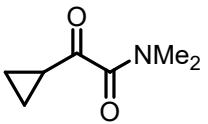


3r (¹H NMR 400 MHz, CDCl₃)

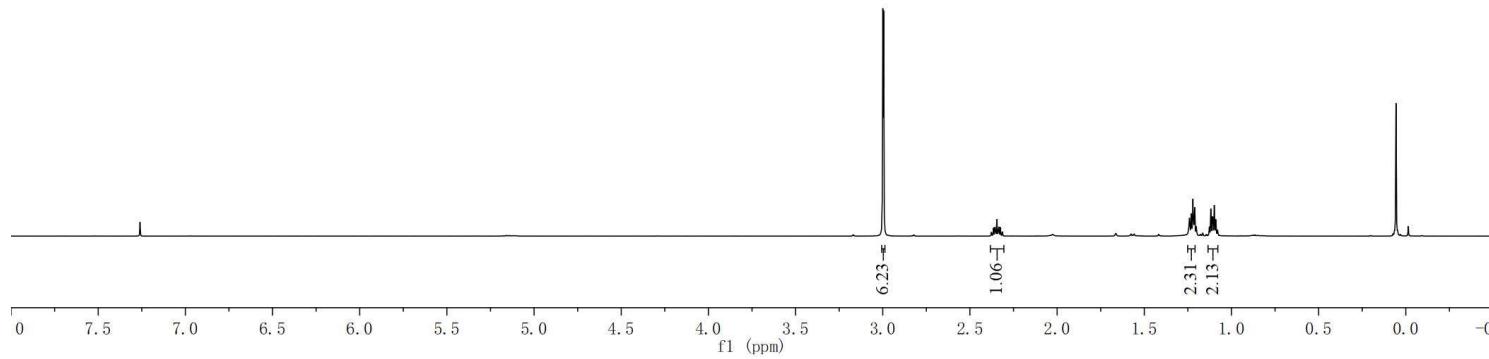




—7.2602



3s (^1H NMR 400 MHz, CDCl_3)



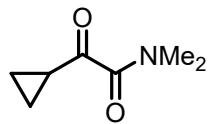
—200.9395

—167.0425

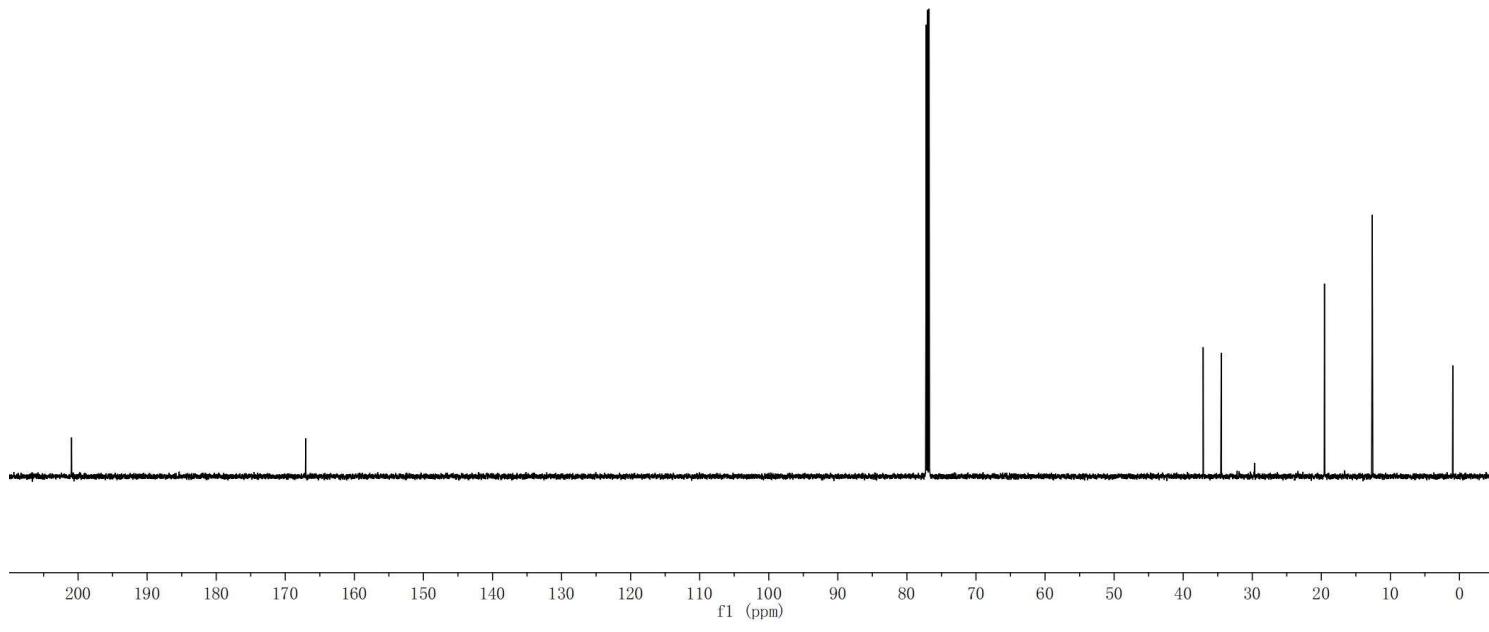
77.2129
77.0011
76.7888

37.1031
34.4873

—19.5465
—12.6225



3s (^{13}C NMR 101 MHz, CDCl_3)

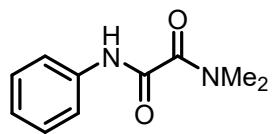


—9.2410

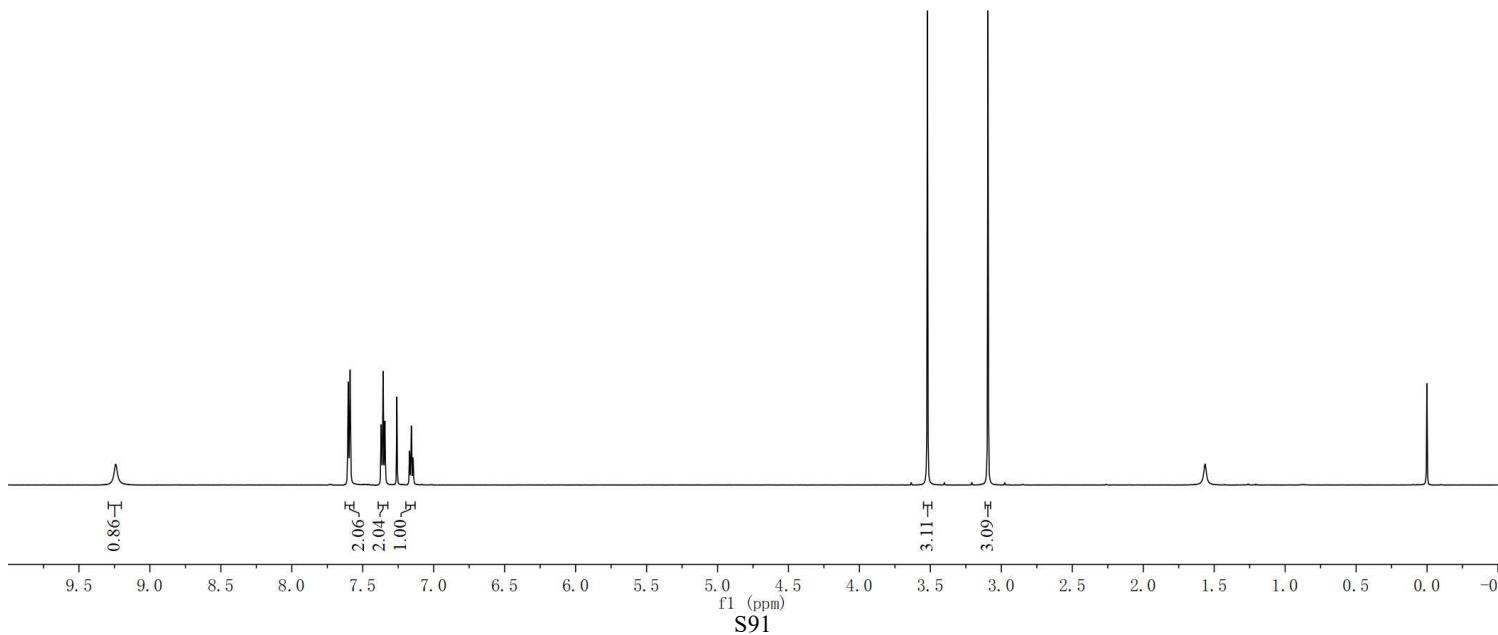
7.6024
7.5891
7.3704
7.3677
7.3668
7.3440
7.2596
7.1699
7.1576
7.1452

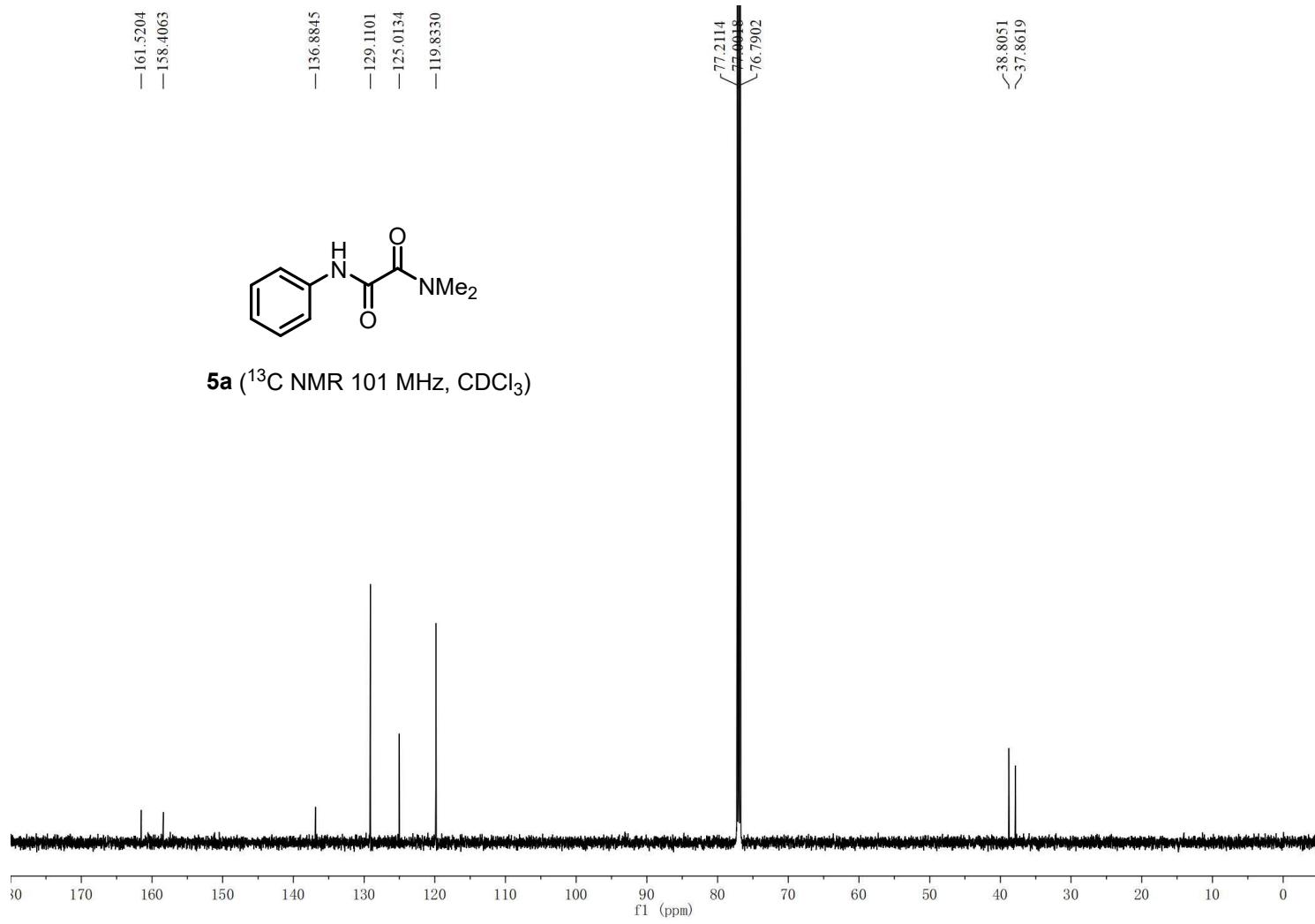
—3.5207

—3.0942



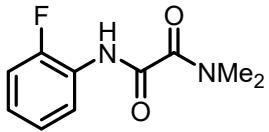
5a (^1H NMR 400 MHz, CDCl_3)



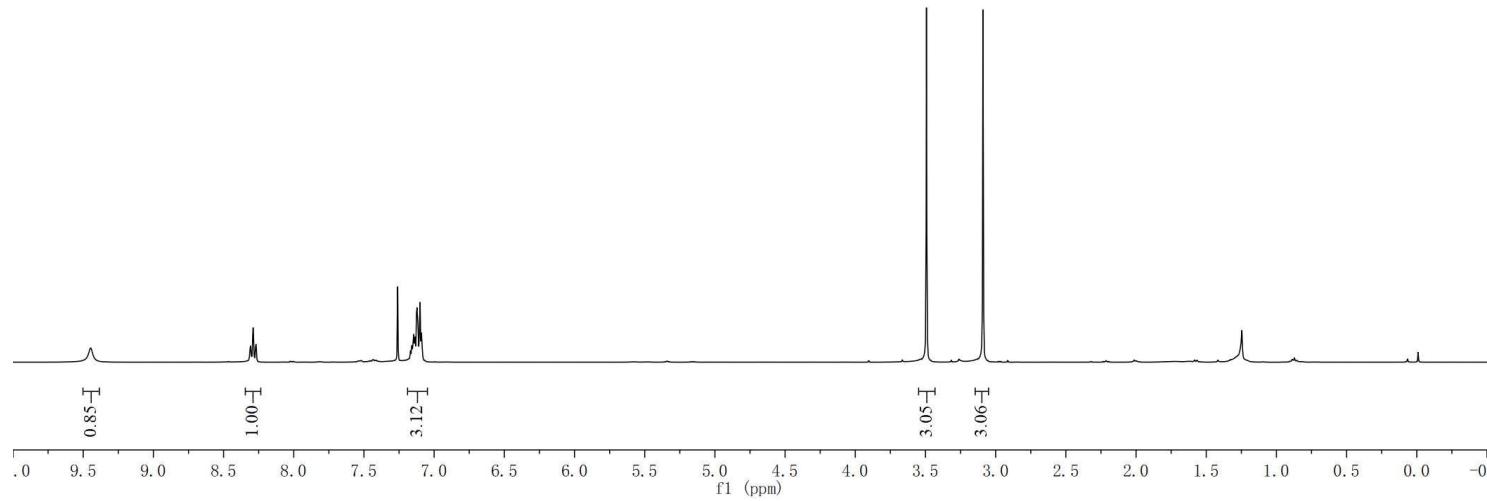


—9.4469

8.3121
8.3088
8.3056
8.2900
8.2867
8.2719
8.2681
7.2600
7.1686
7.1599
7.1572
7.1497
7.1456
7.1358
7.1259
7.1215
7.1160
7.1027
7.0999
7.0909
7.0857
7.0798



5b (^1H NMR 400 MHz, CDCl_3)

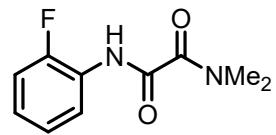


~161.0617
~158.4953
~154.0342
~151.5919

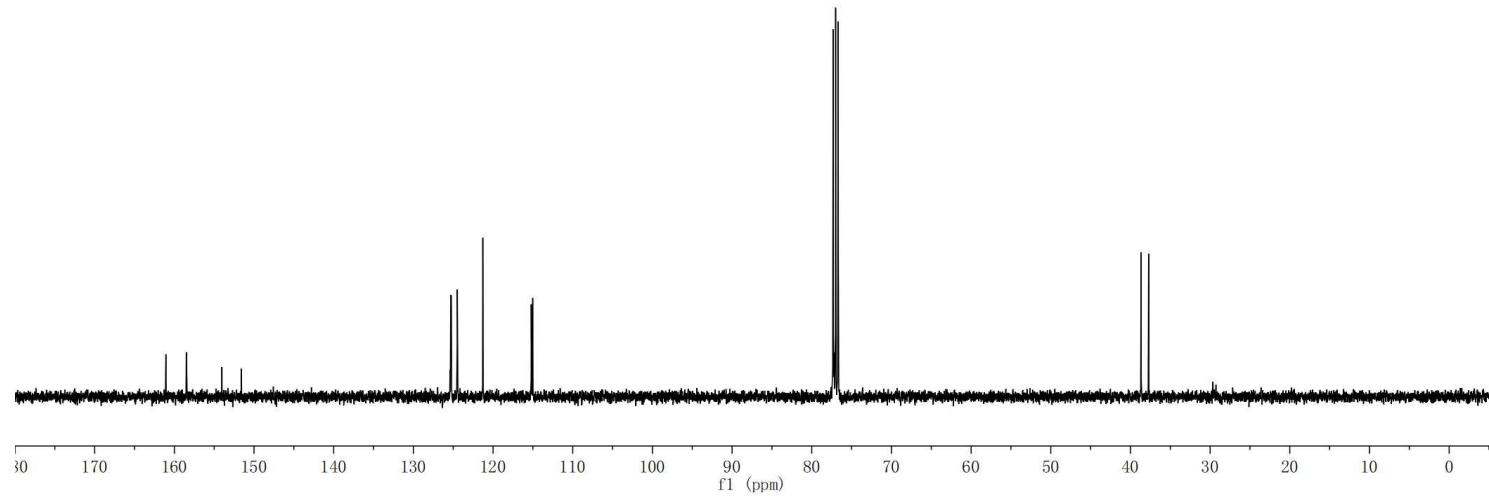
125.3977
125.3099
125.2372
124.4863
124.4492
121.2844
115.2097
115.0204

77.3169
77.1996
77.0002
76.6814

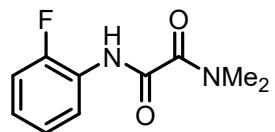
38.6666
37.7184



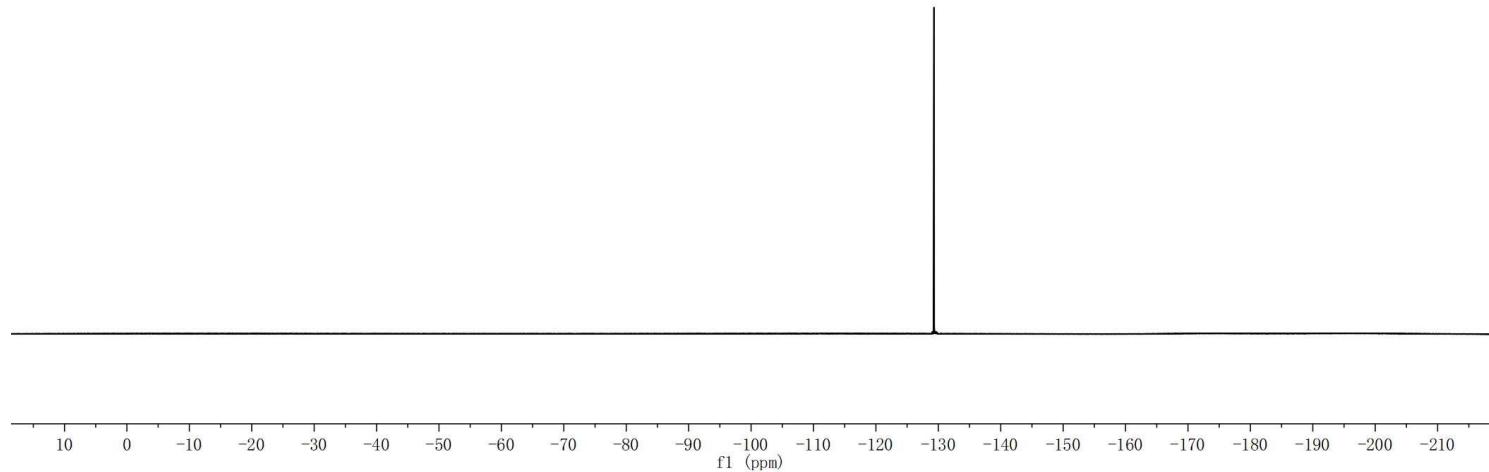
5b (^{13}C NMR 101 MHz, CDCl_3)



—
-129.316



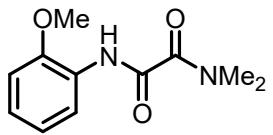
5b (^{19}F NMR 376 MHz, CDCl_3)



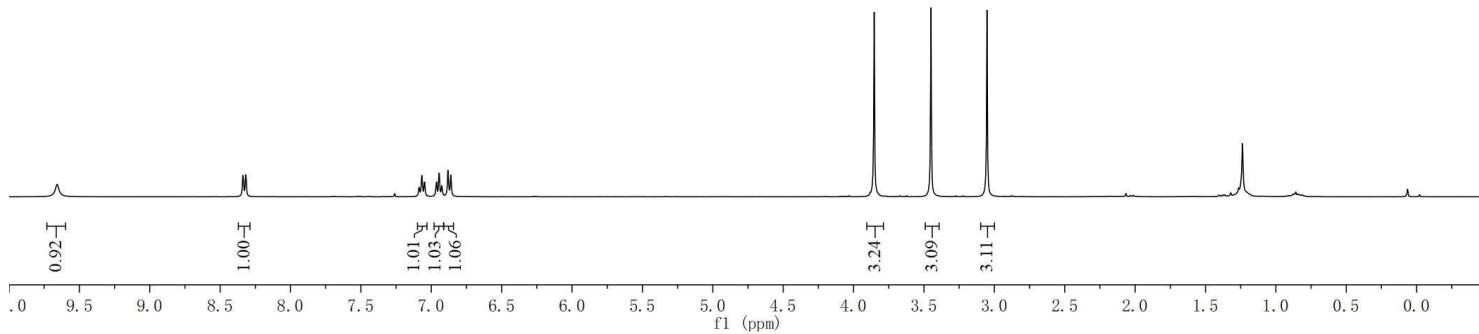
—9.6581

8.3396
8.3355
8.3194
8.3153
7.2599
7.0886
7.0845
7.0692
7.0649
7.0494
7.0452
6.9653
6.9620
6.9459
6.9424
6.9264
6.9230
6.8825
6.8793
6.8625
6.8589

—3.8536
—3.4499
—3.0521



5c (^1H NMR 400 MHz, CDCl_3)



—161.8621
—158.5284

—148.6784

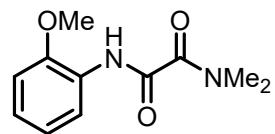
—126.6411
~124.7758
✓120.8736
—119.5764

—110.1783

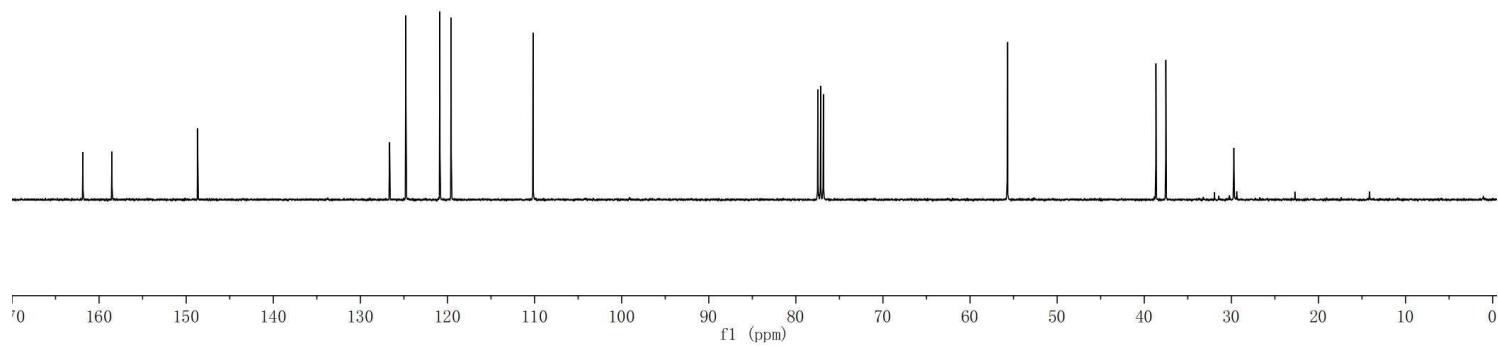
✓77.4785
✓77.1591
✓76.8422

—55.7180

✓38.6570
✓37.5197



5c (^{13}C NMR 101 MHz, CDCl_3)



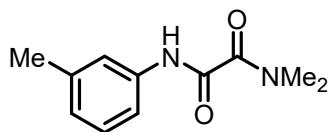
-9.2804

7.4660
7.4621
7.4599
7.4564
7.3814
7.3760
7.3615
7.3583
7.3554
7.2598
7.2407
7.2213
7.2016
6.9753
6.9729
6.9707
6.9686
6.9665
6.9564
6.9542
6.9518
6.9199
6.9480

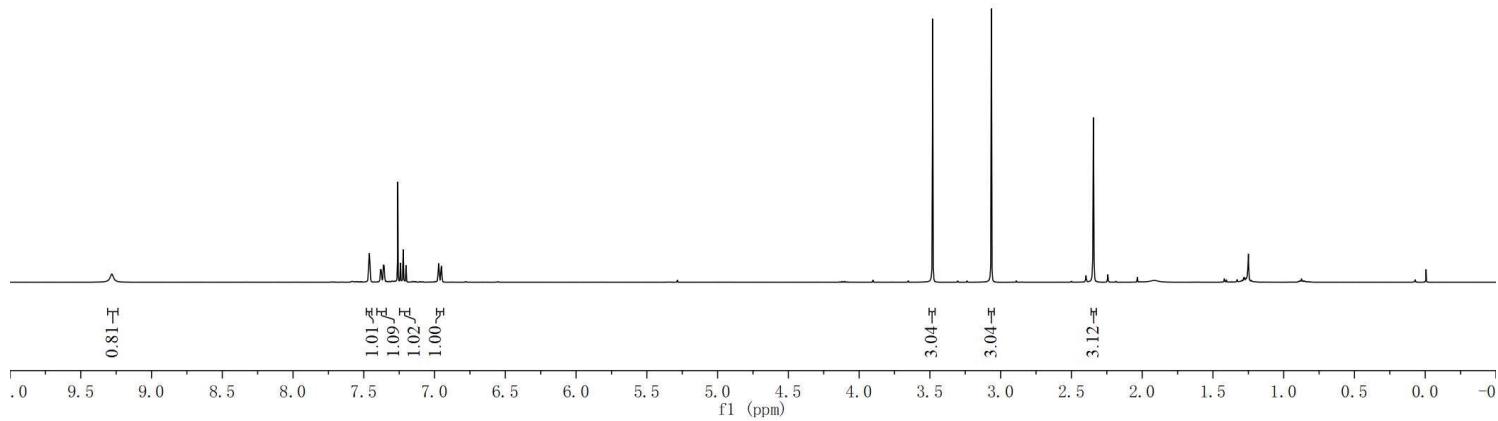
-3.4805

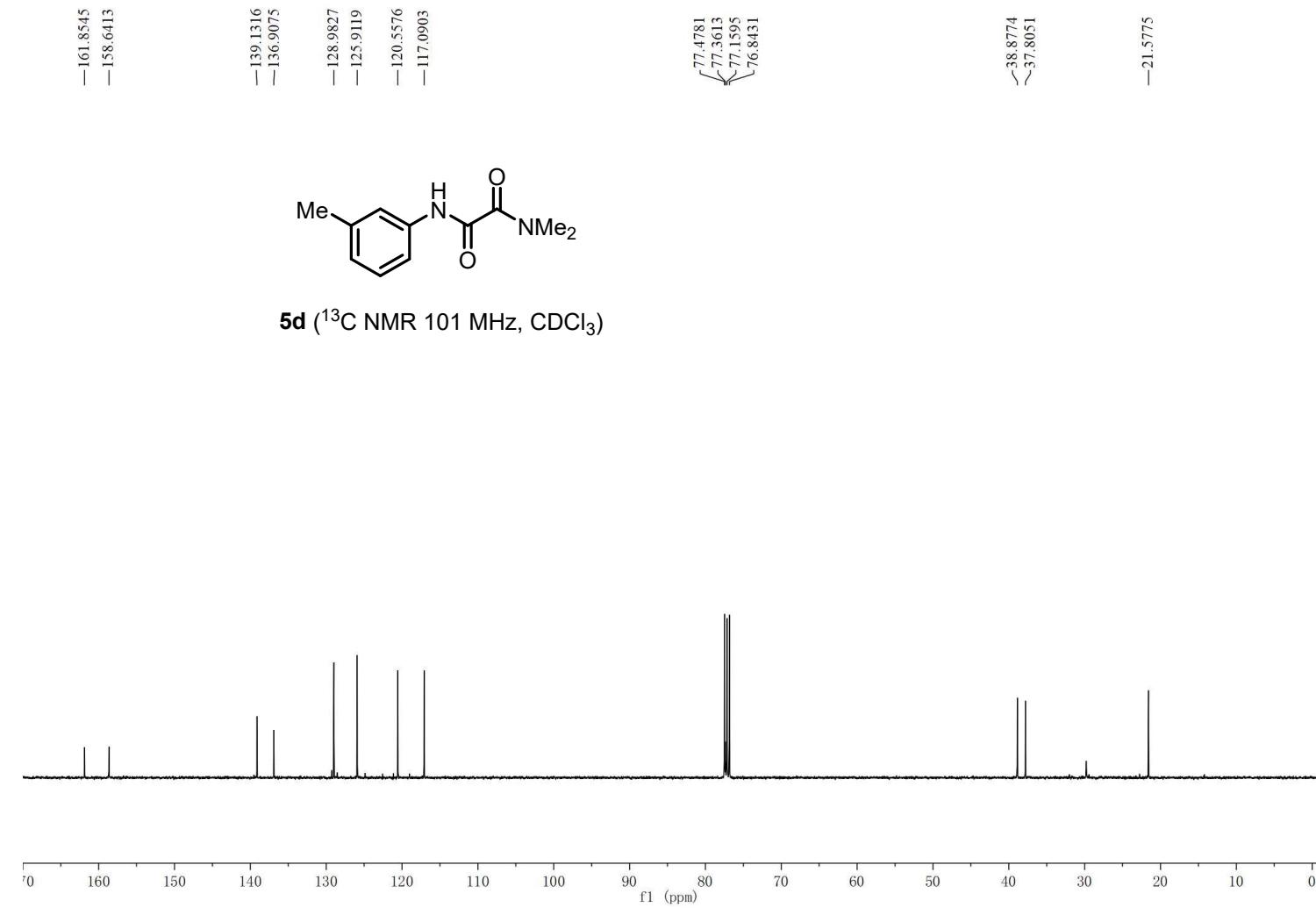
-3.0656

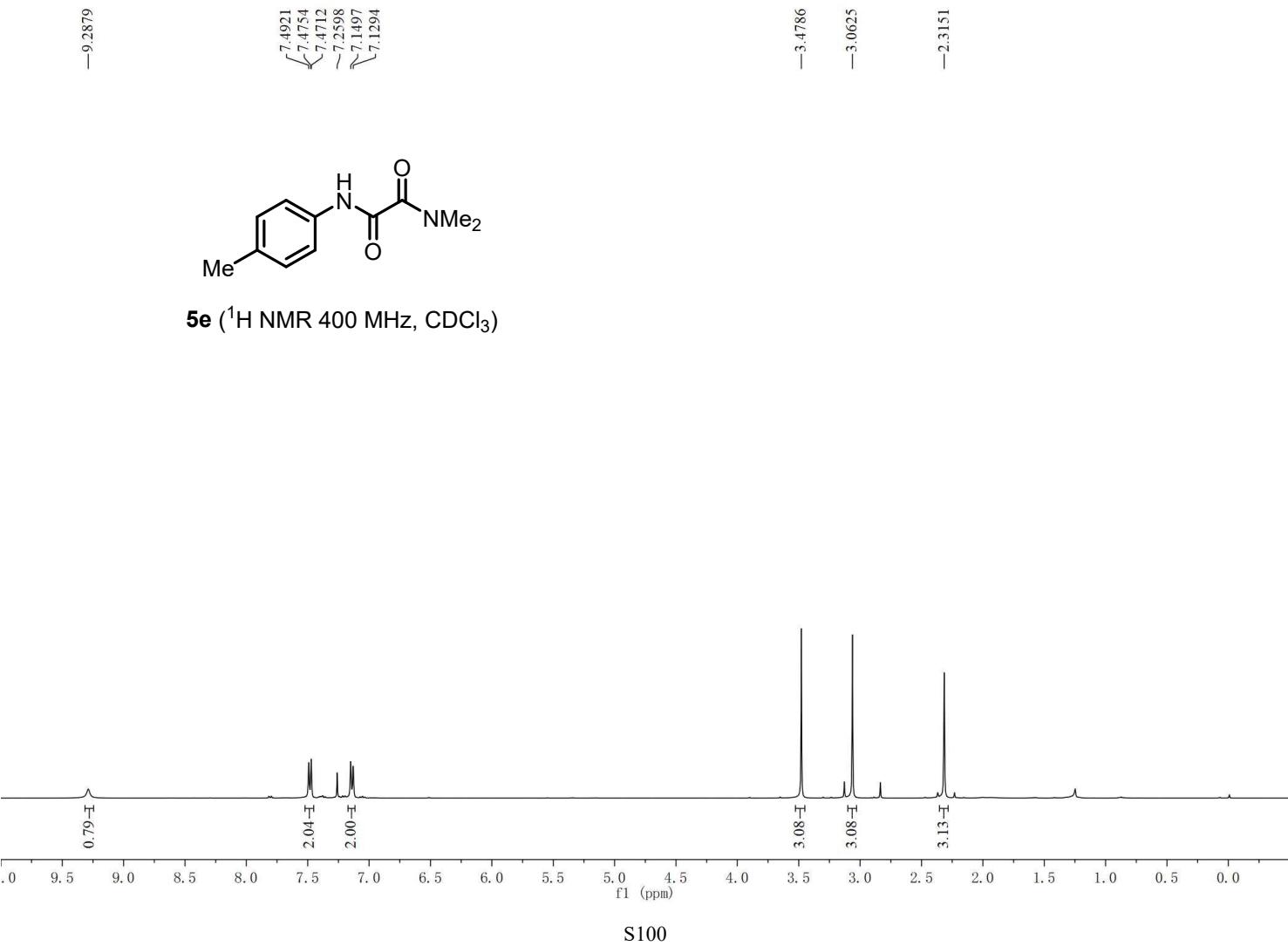
2.3451
2.3431



5d (^1H NMR 400 MHz, CDCl_3)







— 161.7567
— 158.4082

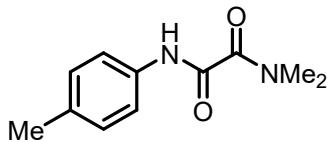
— 134.5994
— 134.2996
— 129.4944

— 119.7883

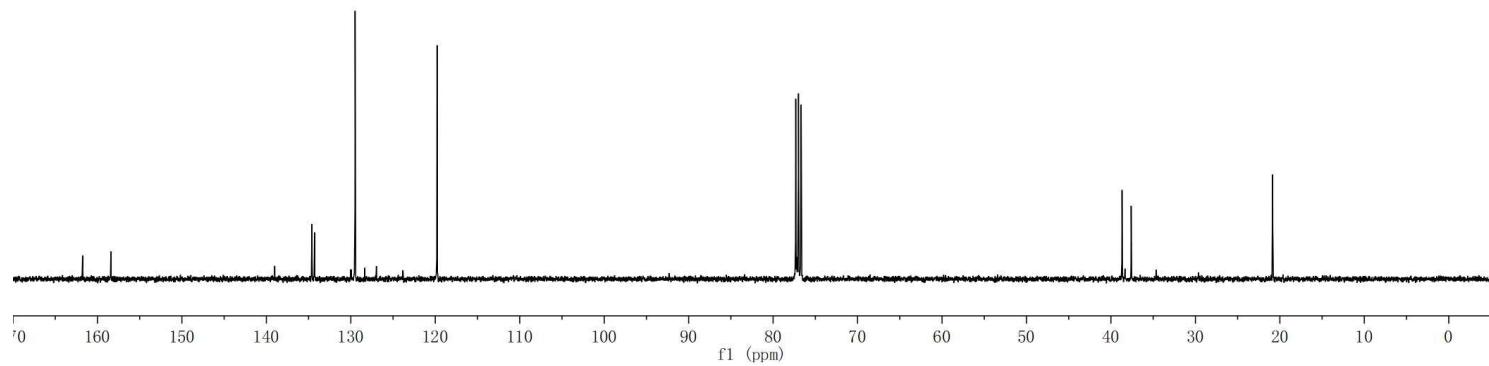
— 77.3197
— 76.9996
— 76.6835

— 38.6880
— 37.6004

— 20.8504



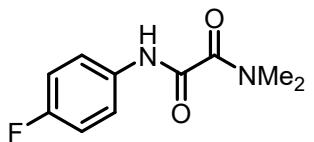
5e (^{13}C NMR 101 MHz, CDCl_3)



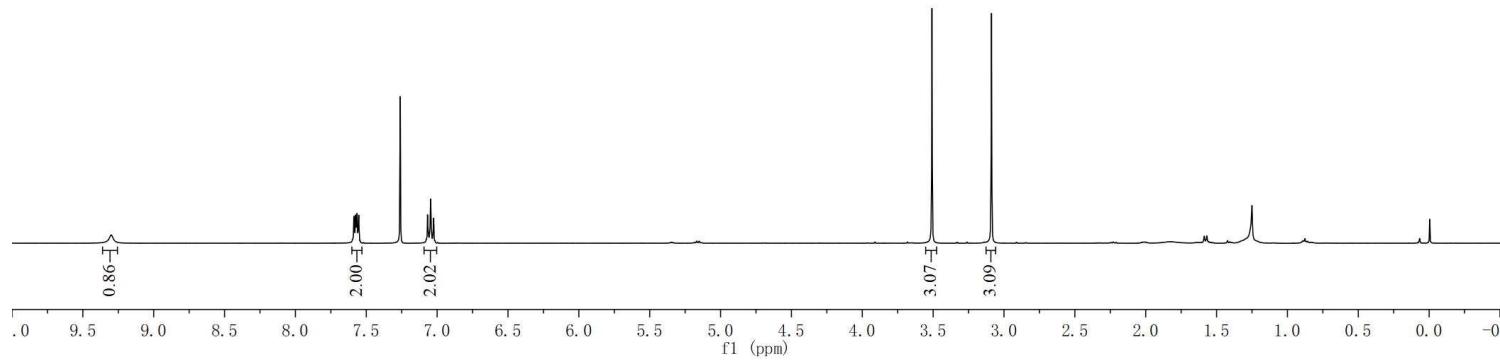
—9.2999

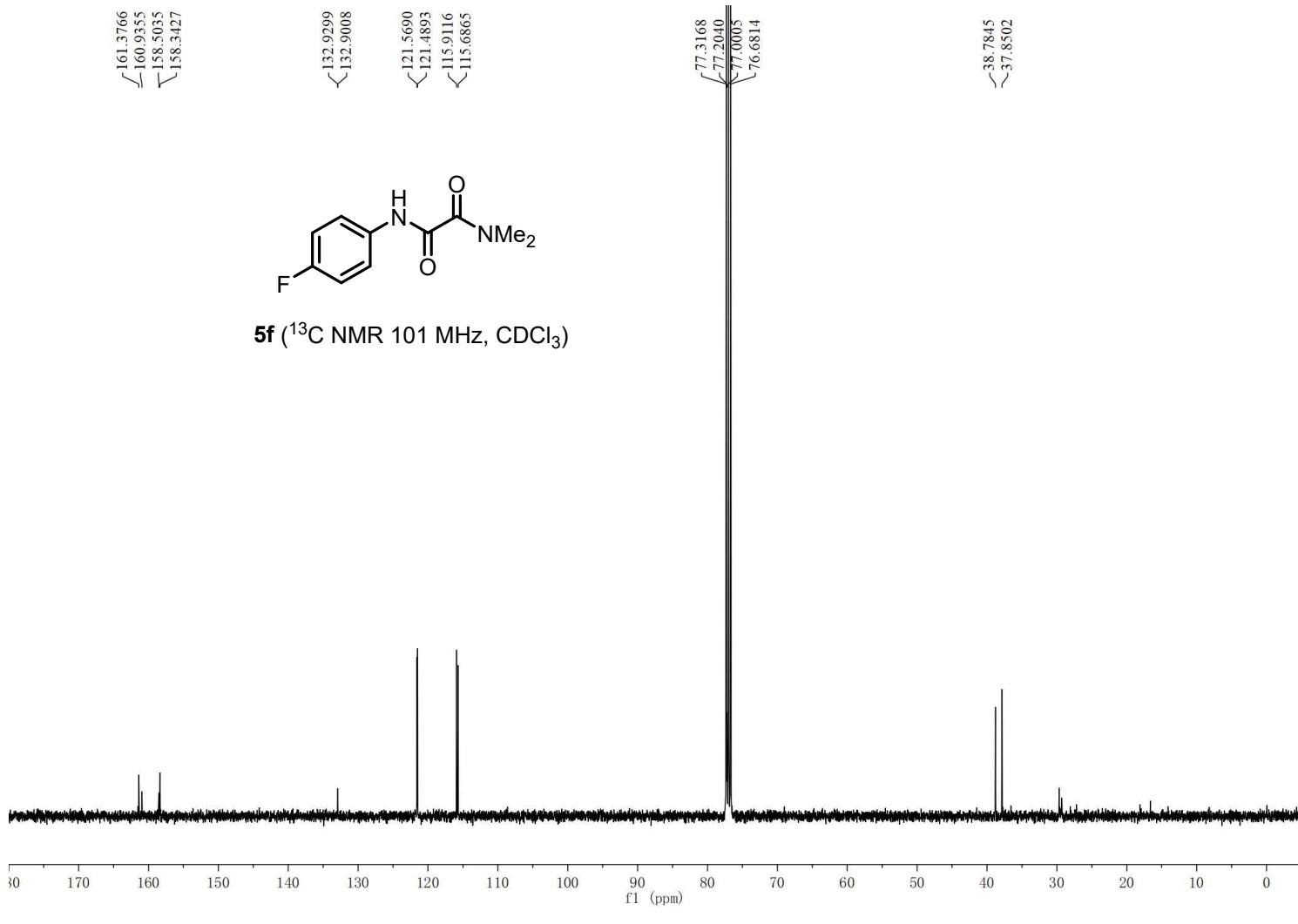
7.5877
7.5824
7.5759
7.5705
7.5652
7.5586
7.5533
7.2603
7.0676
7.0620
7.0560
7.0509
7.0469
7.0450
7.0415
7.0359
7.0300
7.0243

—3.5085
—3.0883

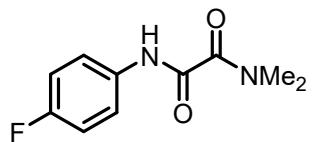


5f (^1H NMR 400 MHz, CDCl_3)

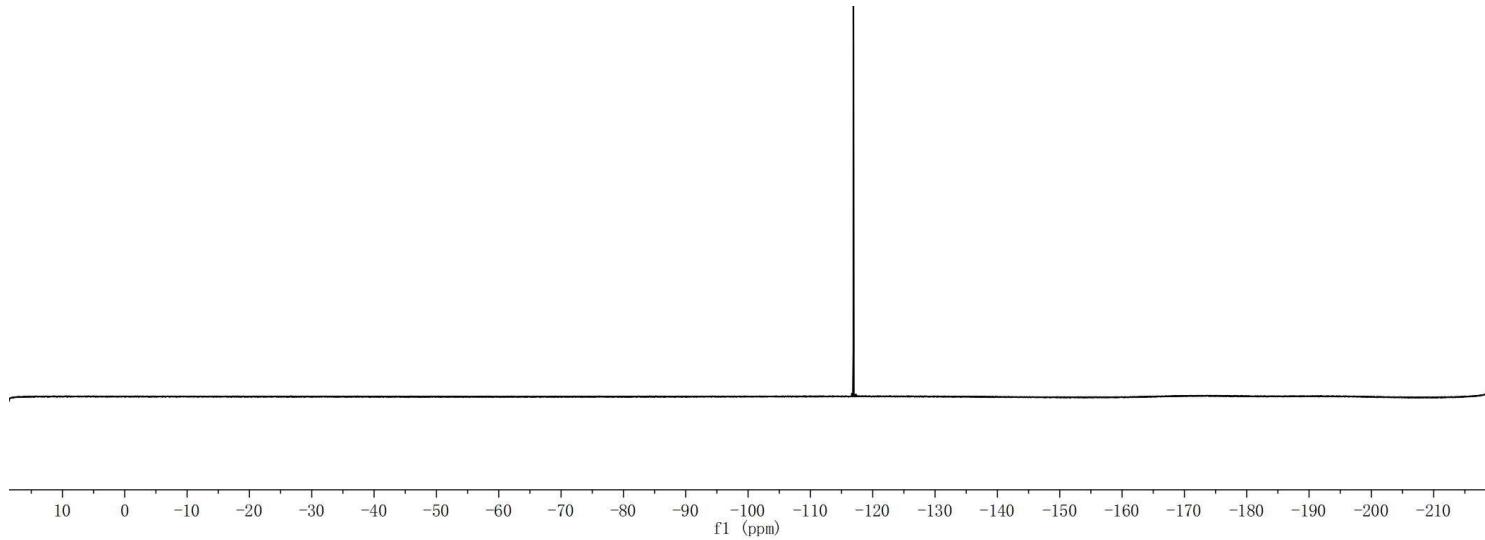


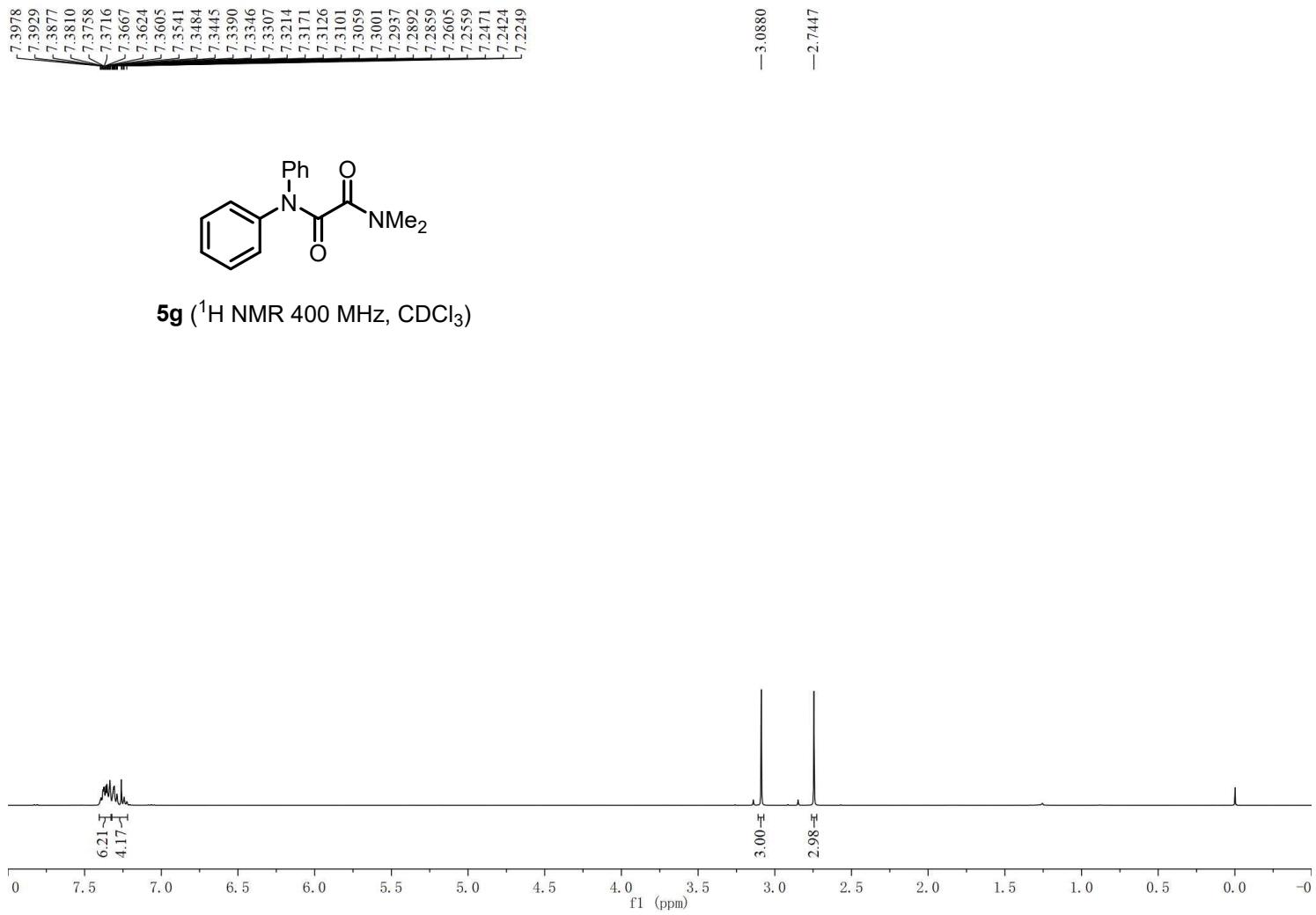


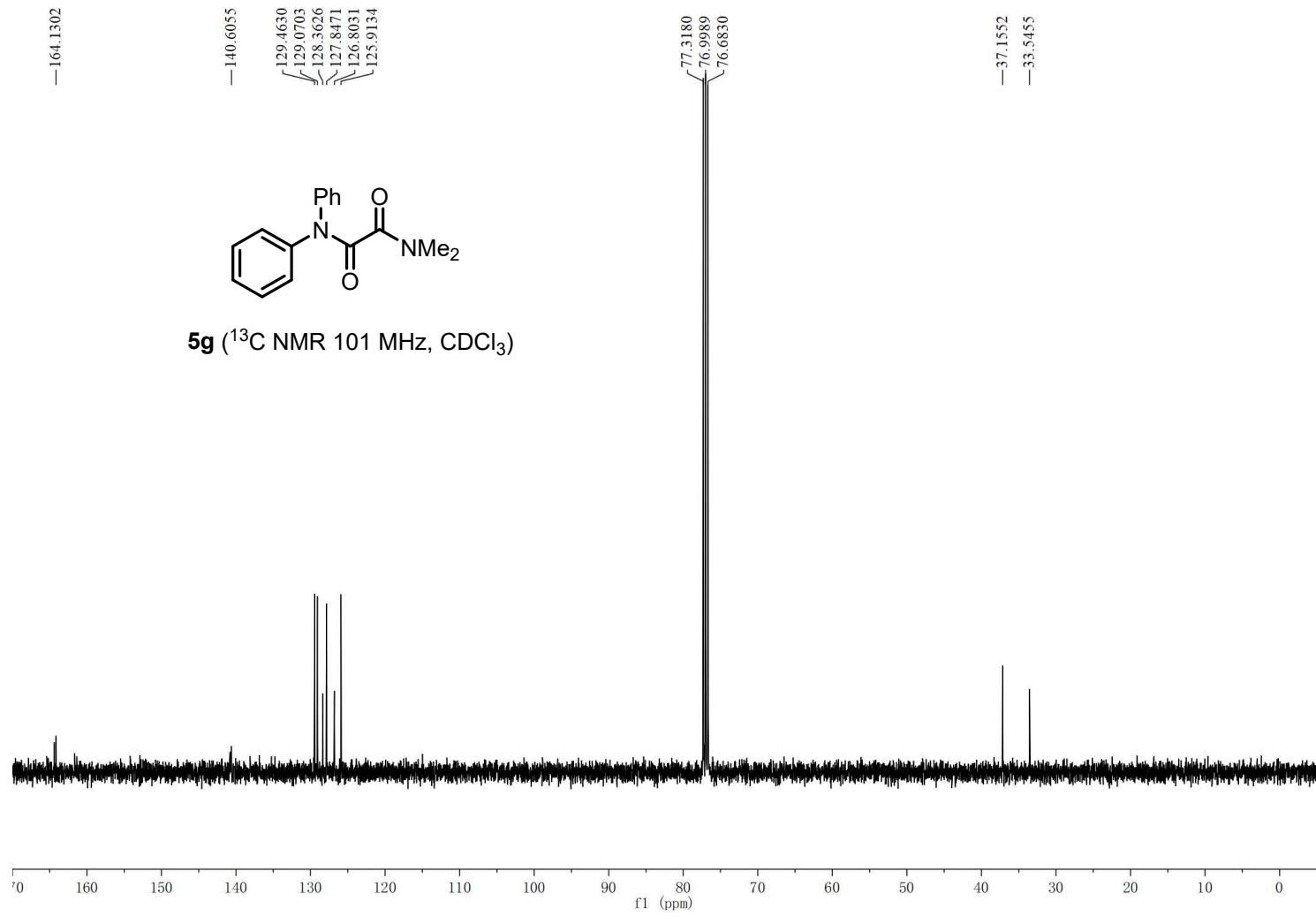
-116.895



5f (^{19}F NMR 376 MHz, CDCl_3)



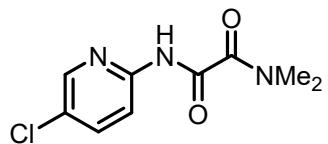




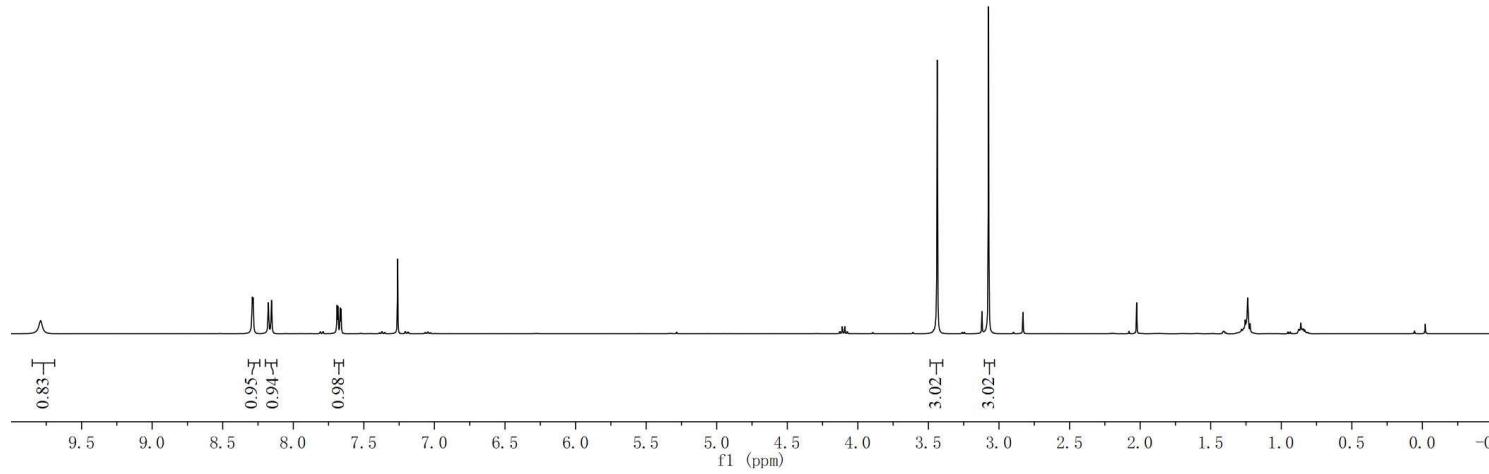
—9.7903

$$\begin{array}{r}
 8.2906 \\
 \swarrow 8.2842 \\
 \searrow 8.1764 \\
 8.1543 \\
 \nearrow 7.6900 \\
 \searrow 7.6835 \\
 \nearrow 7.6679 \\
 \searrow 7.6615 \\
 \hline
 -7.2605
 \end{array}$$

—3.4376



5h (^1H NMR 400 MHz, CDCl_3)



— 160.9926

— 159.1161

— 148.8315

— 147.1238

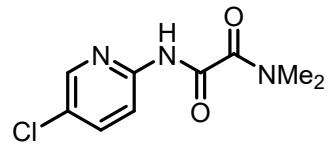
— 137.9625

— 127.5784

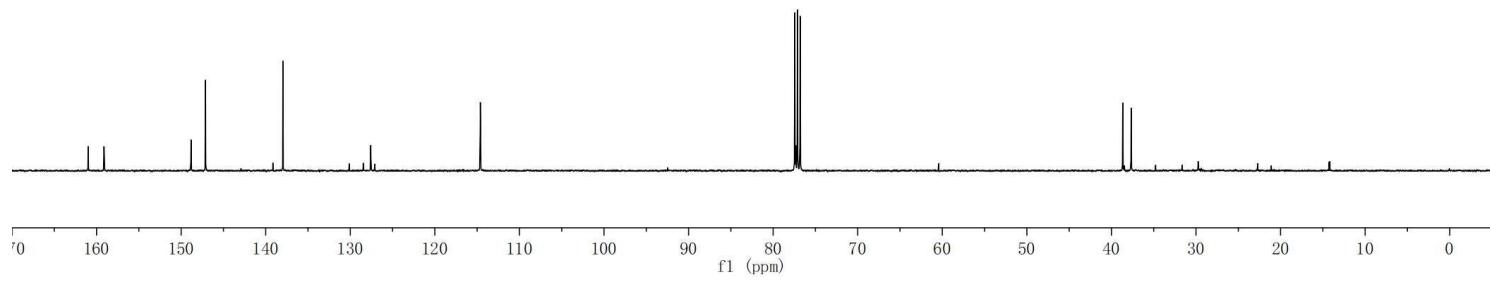
— 114.6247

— 38.6644

— 37.6659



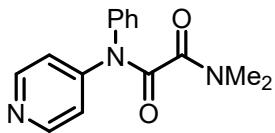
5h (^{13}C NMR 101 MHz, CDCl_3)



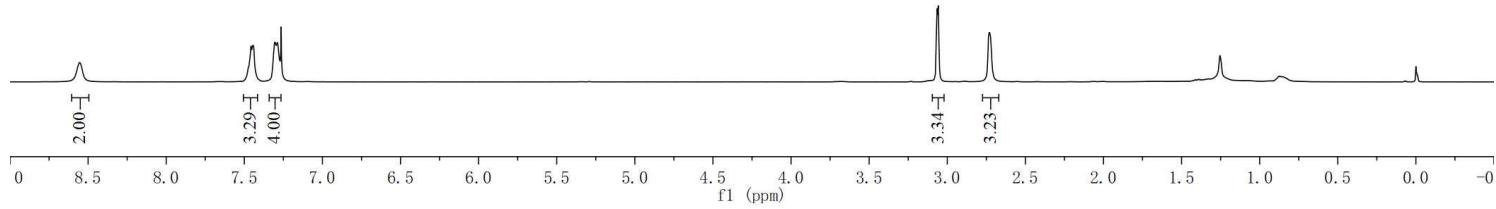
—8.5542

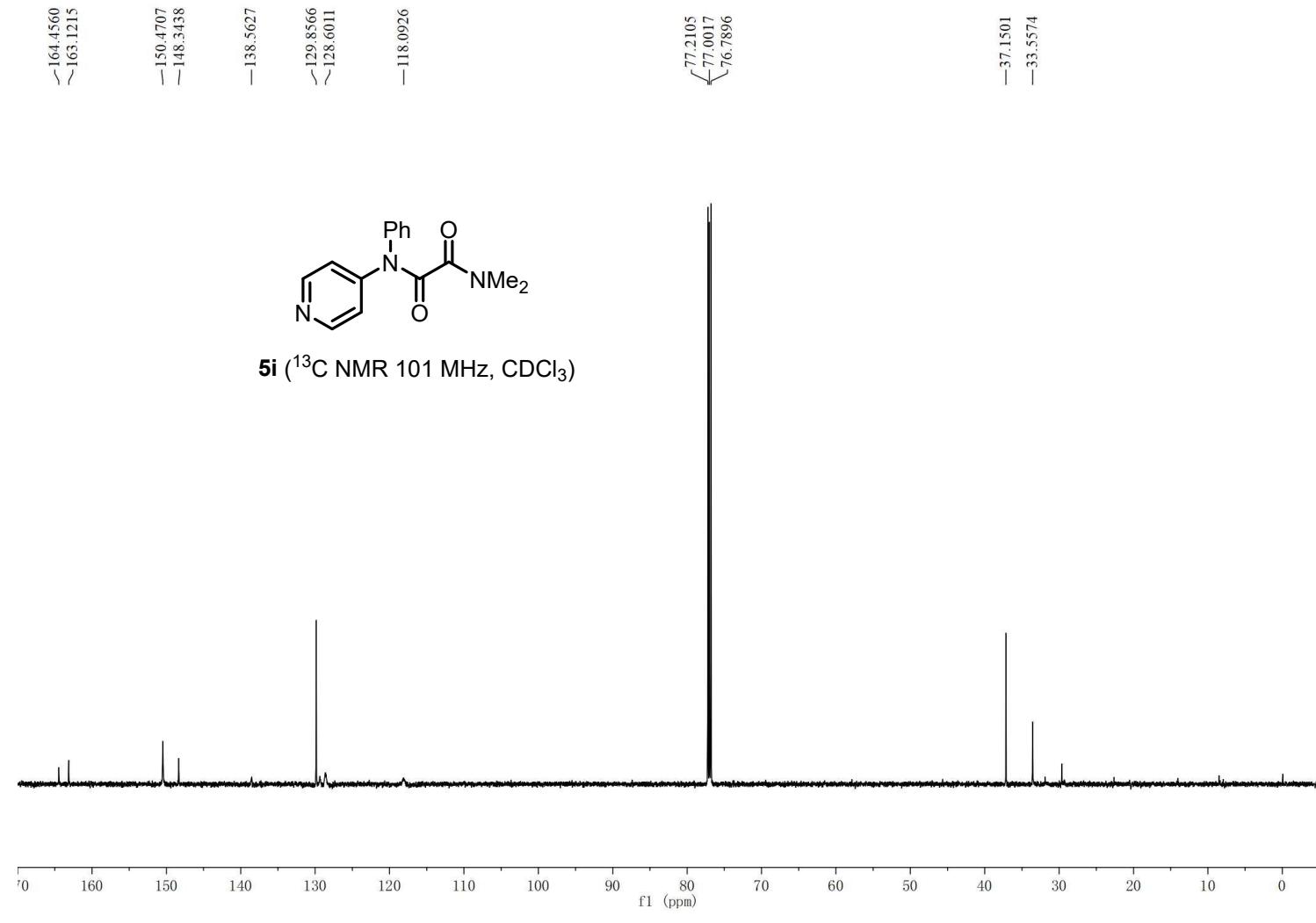
7.4760
7.4662
7.4579
7.4476
7.4396
7.3190
7.3131
7.3067
7.3007
7.2945
7.2872
7.2798
7.2643

3.0696
3.0645
3.0572
—2.7295

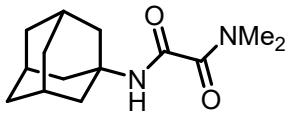


5i (^1H NMR 400 MHz, CDCl_3)

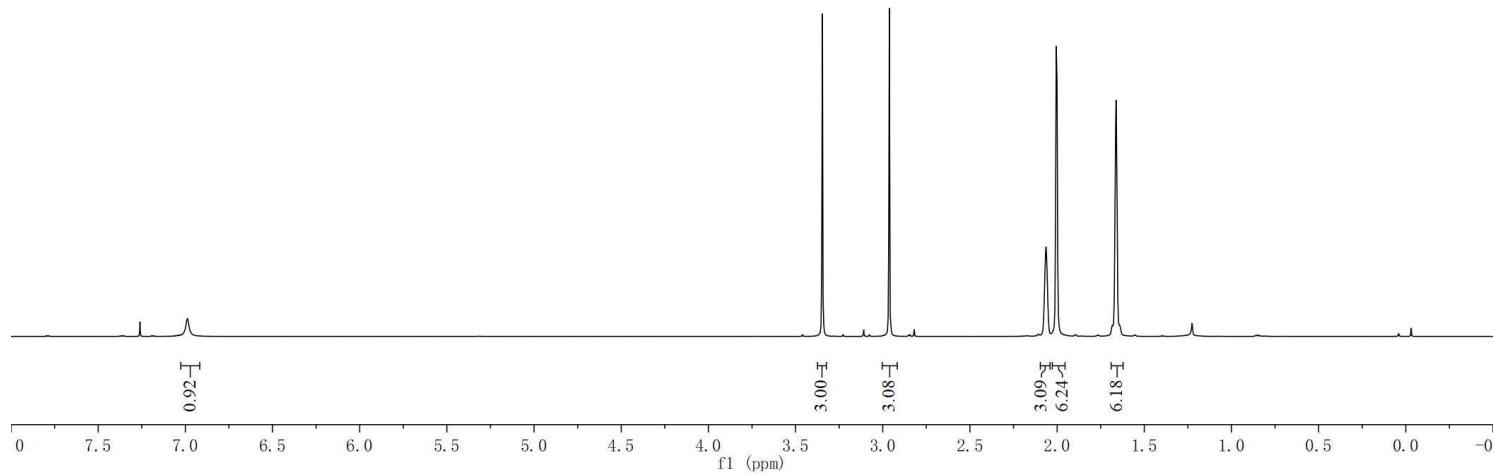




—7.2602
—6.9883



5j (^1H NMR 400 MHz, CDCl_3)



S111

— 162.6981

— 160.2325

77.3168

77.0006

76.6809

— 52.0058

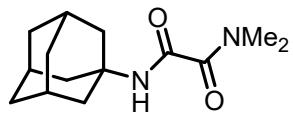
— 40.9707

— 38.4788

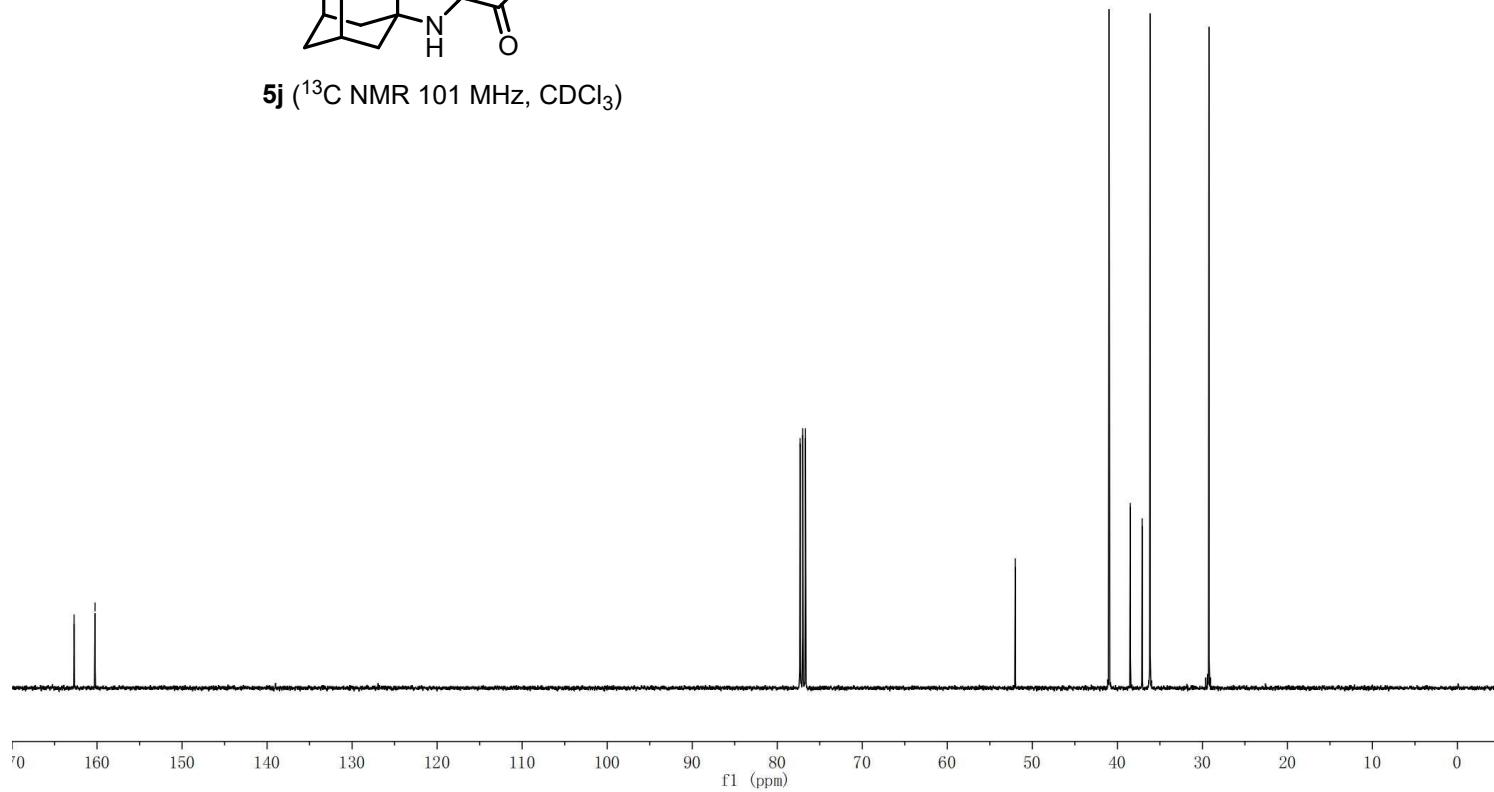
— 37.0842

— 36.1608

— 29.2341

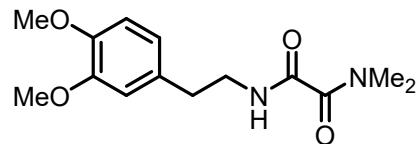


5j (^{13}C NMR 101 MHz, CDCl_3)

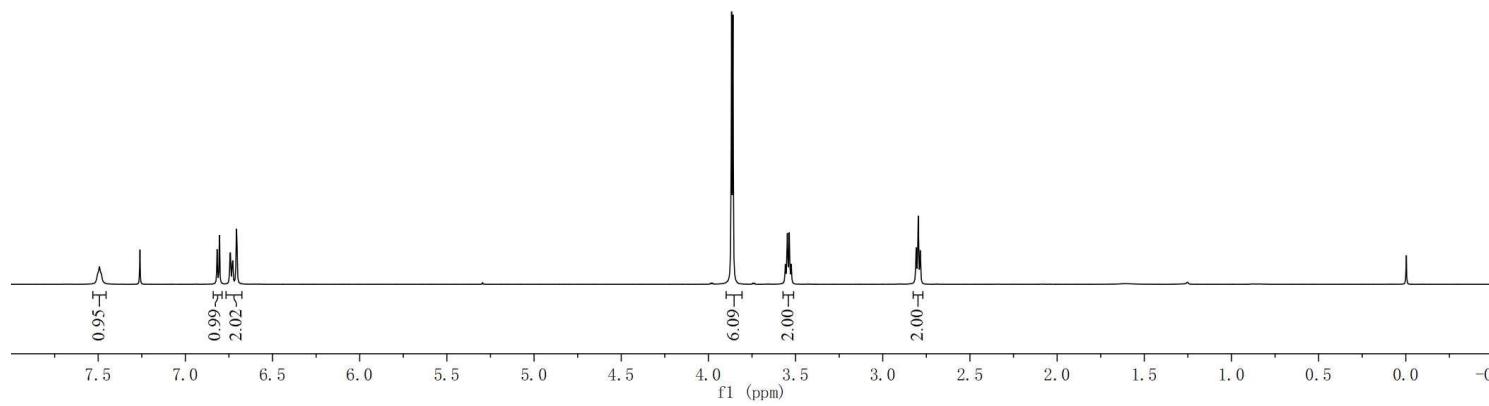


7.5033
7.4926
7.4819
7.2605
6.8178
6.8045
6.7444
6.7408
6.7310
6.7274
6.7072
6.7038

3.8673
3.8591
3.5588
3.5475
3.5362
3.5247
2.8085
2.7965
2.7846



5k (^1H NMR 400 MHz, CDCl_3)



~161.9885
~161.1413

~149.0162
~147.7371

—130.8804

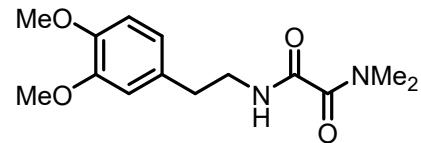
—120.6197

~111.8063
~111.3589

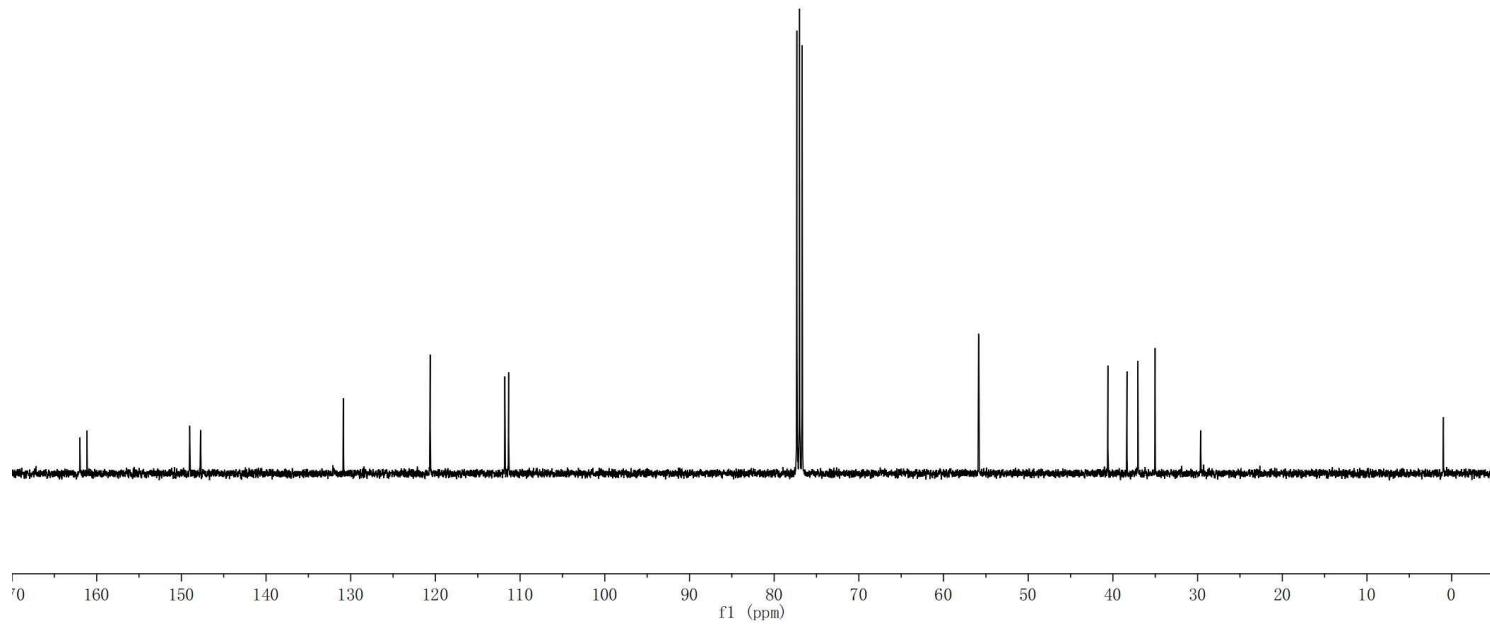
~77.3204
~77.2022
~77.0006
~76.6825

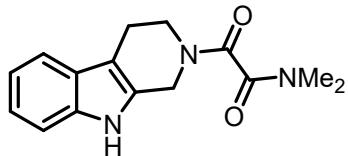
~55.8678
~55.8244

~40.5923
~38.3388
~37.0471
~35.0200
—29.6522

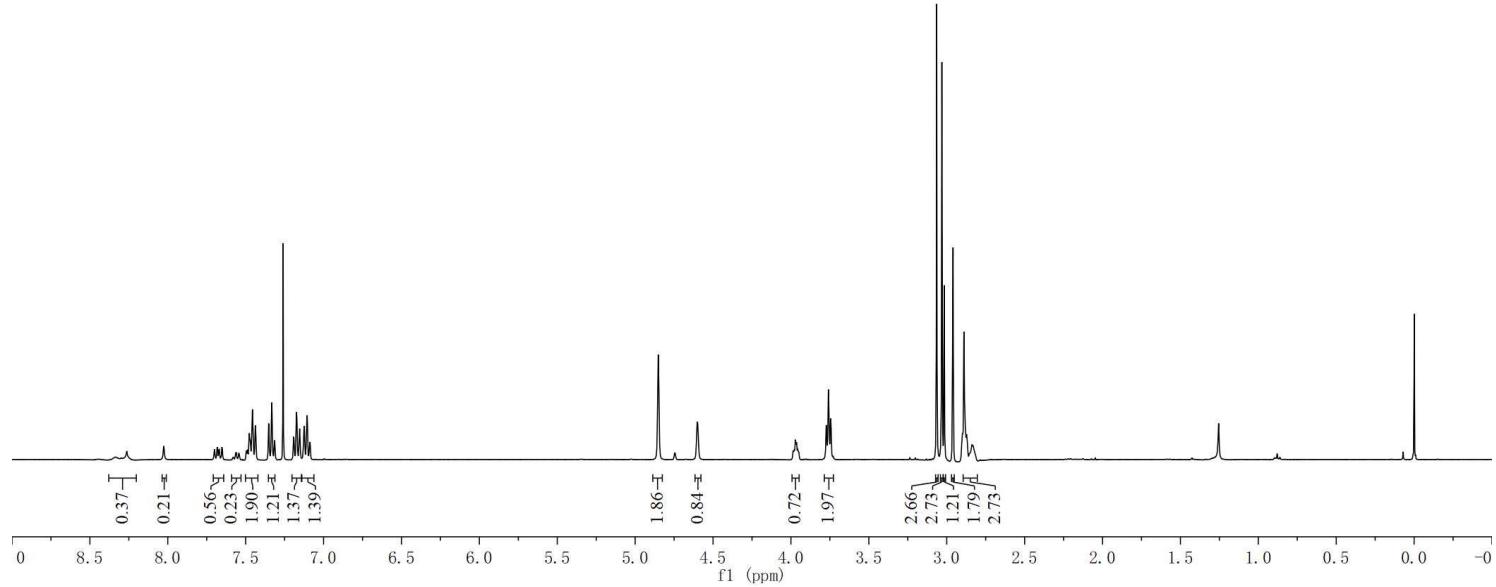


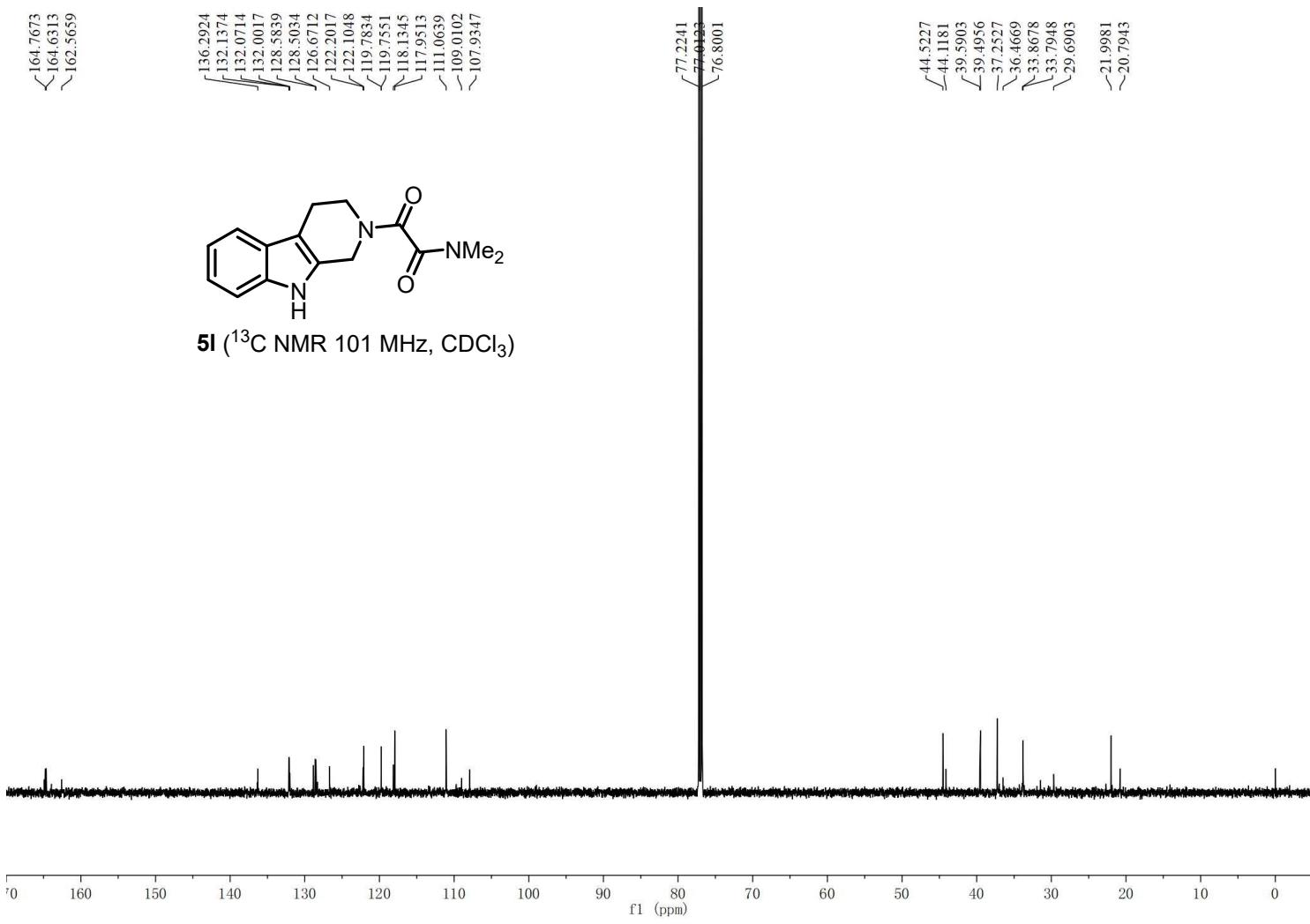
5k (^{13}C NMR 101 MHz, CDCl_3)





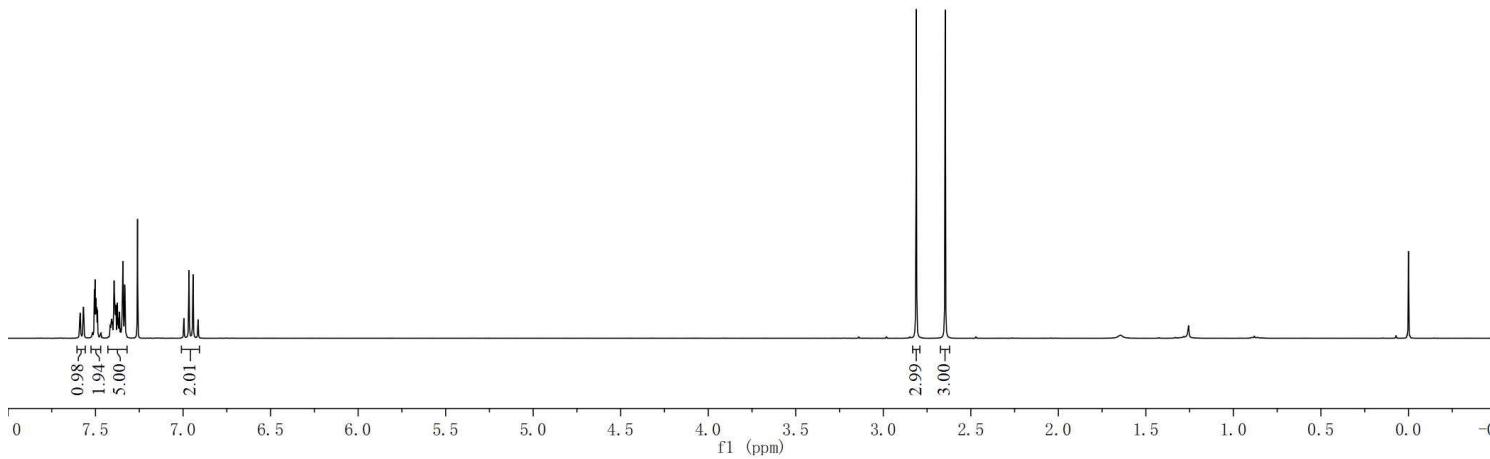
5I (^1H NMR 400 MHz, CDCl_3)

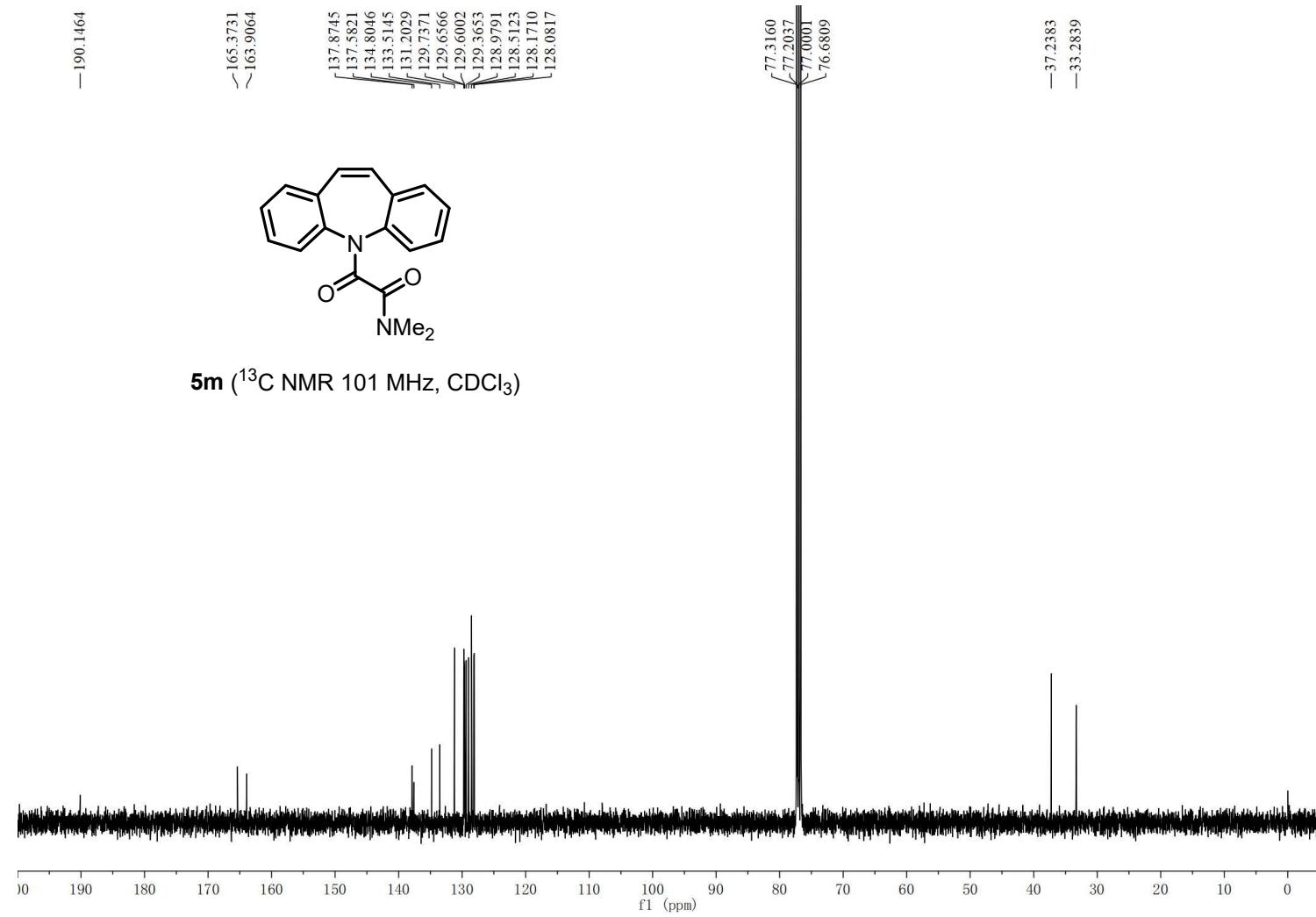


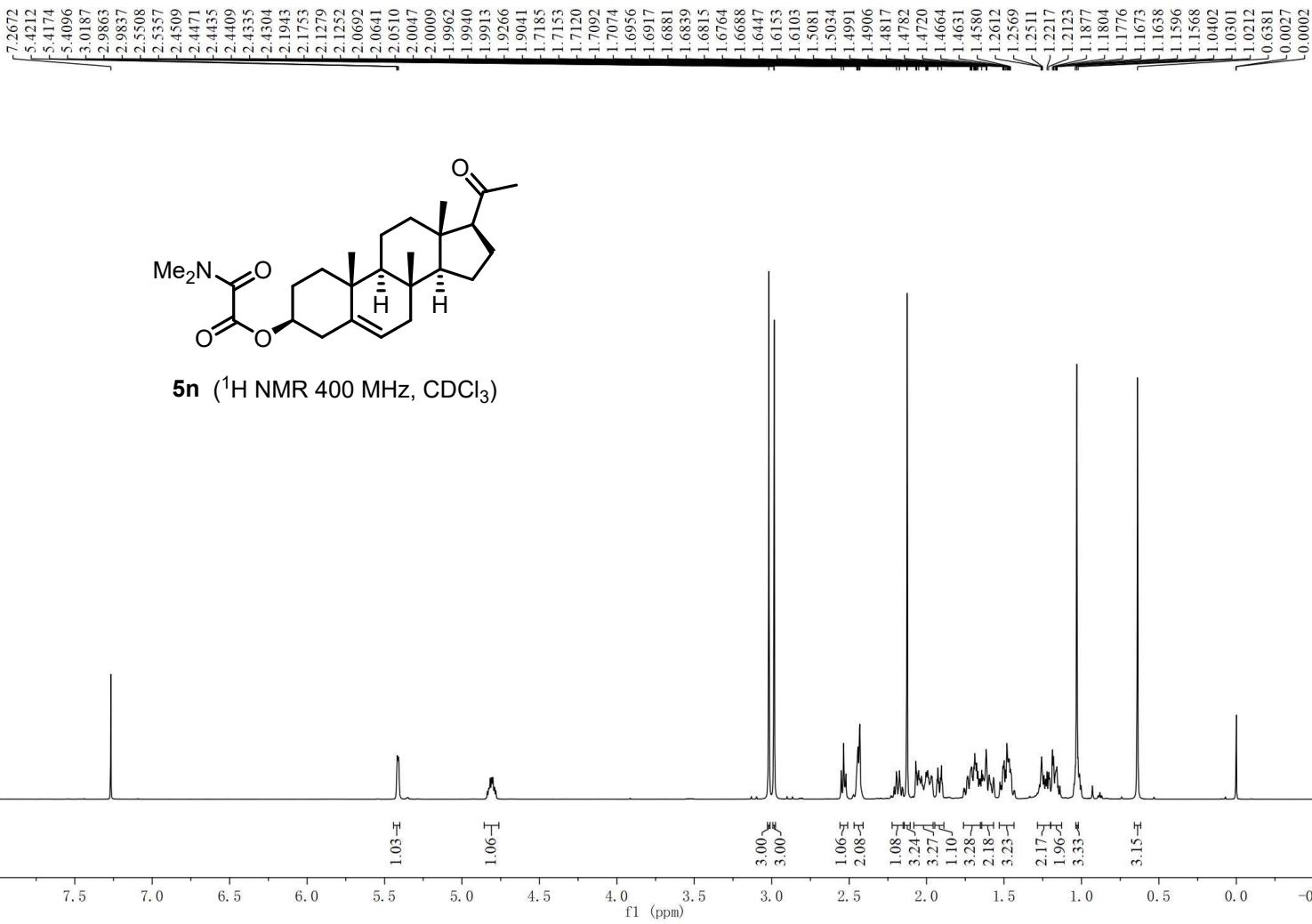




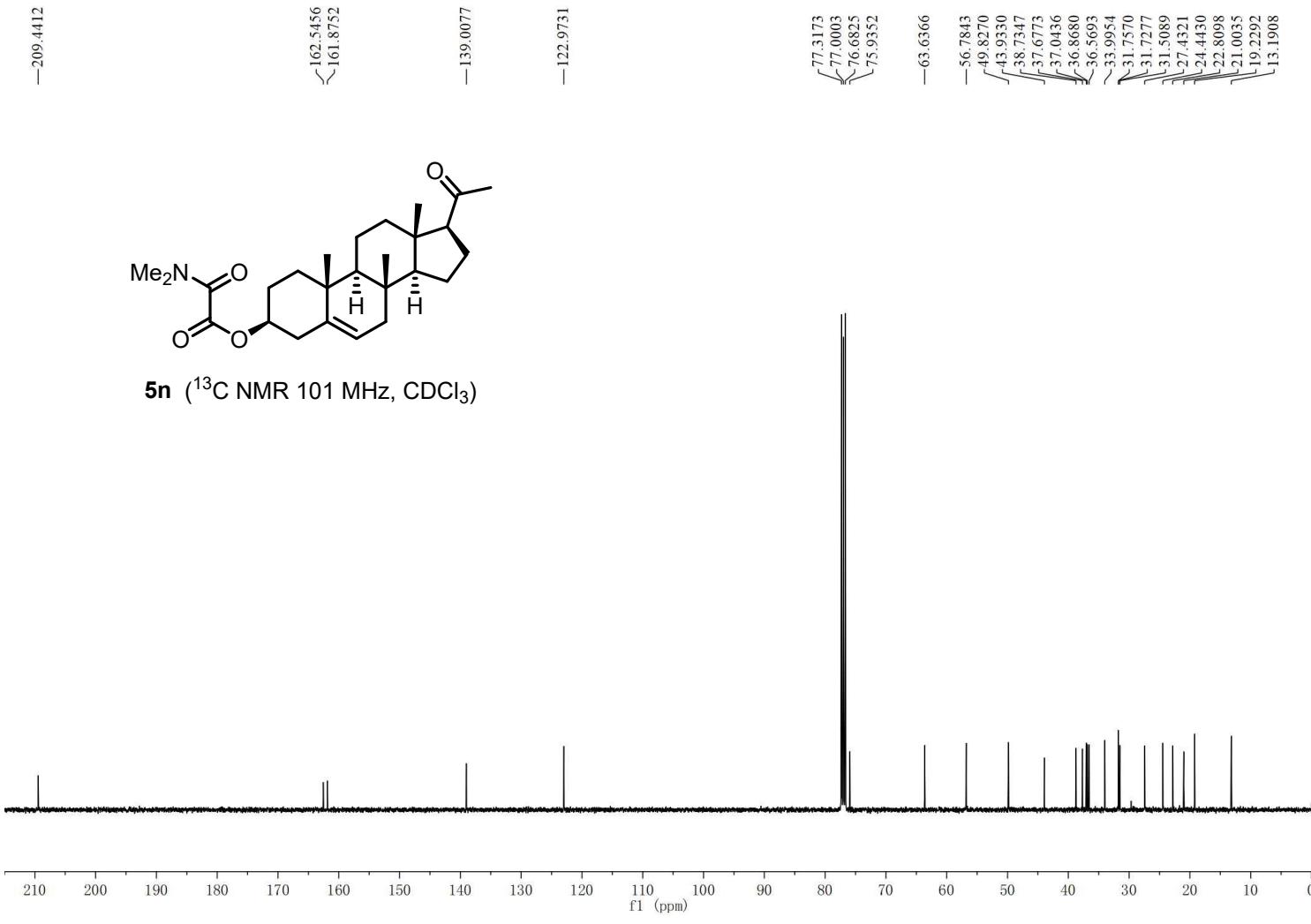
5m (^1H NMR 400 MHz, CDCl_3)



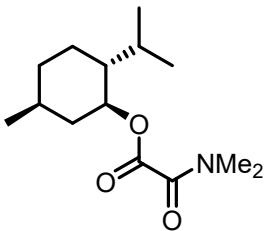




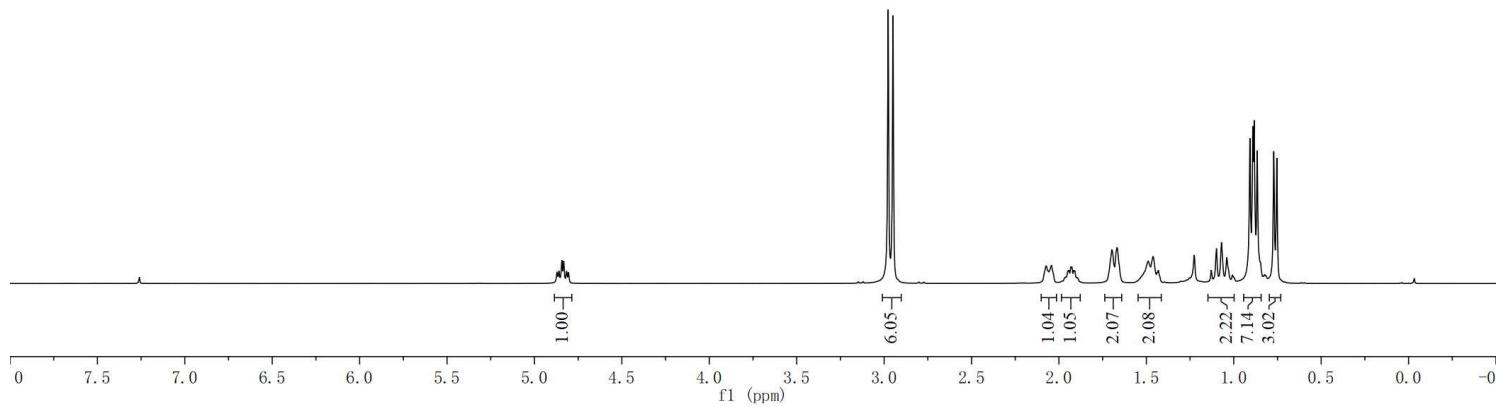
S119



—7.2602



5o (^1H NMR 400 MHz, CDCl_3)

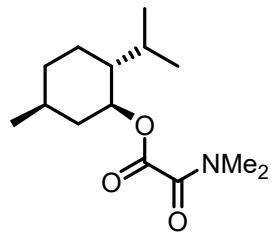


S121

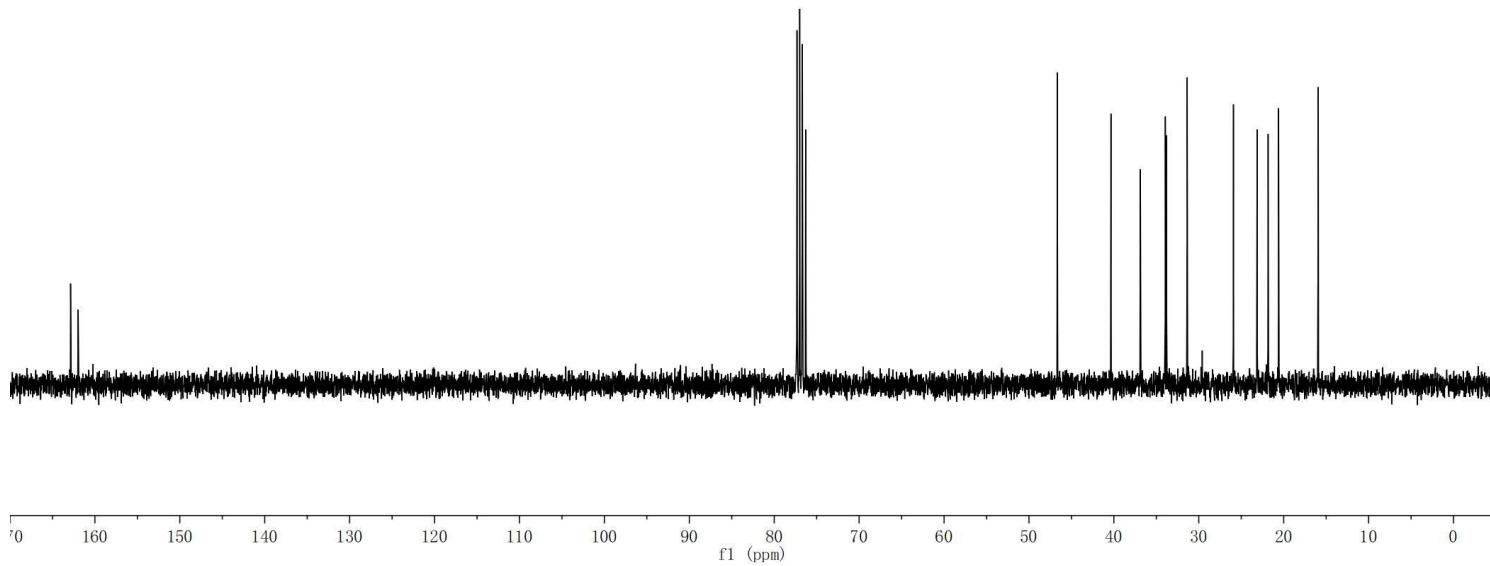
~162.8627
~161.9739

77.3199
76.9999
76.6849
76.3014

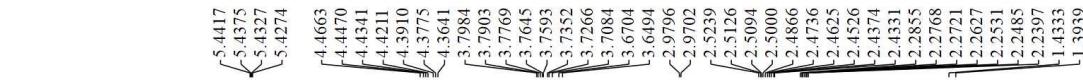
-46.6503
-40.3430
-36.8803
-33.9416
-33.8017
-31.3661
-25.9198
-23.1289
-21.8368
-20.6024
-15.9290



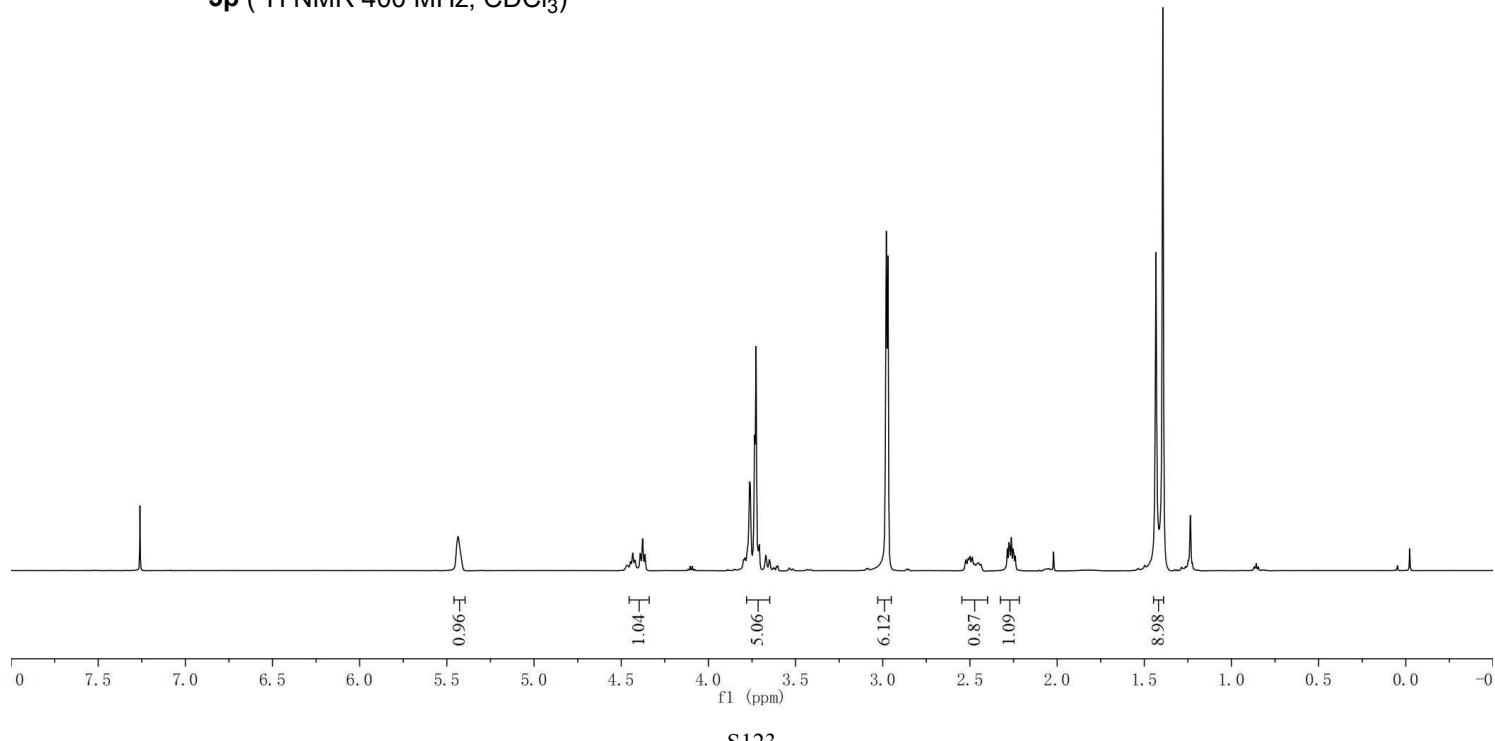
5o (^{13}C NMR 101 MHz, CDCl_3)



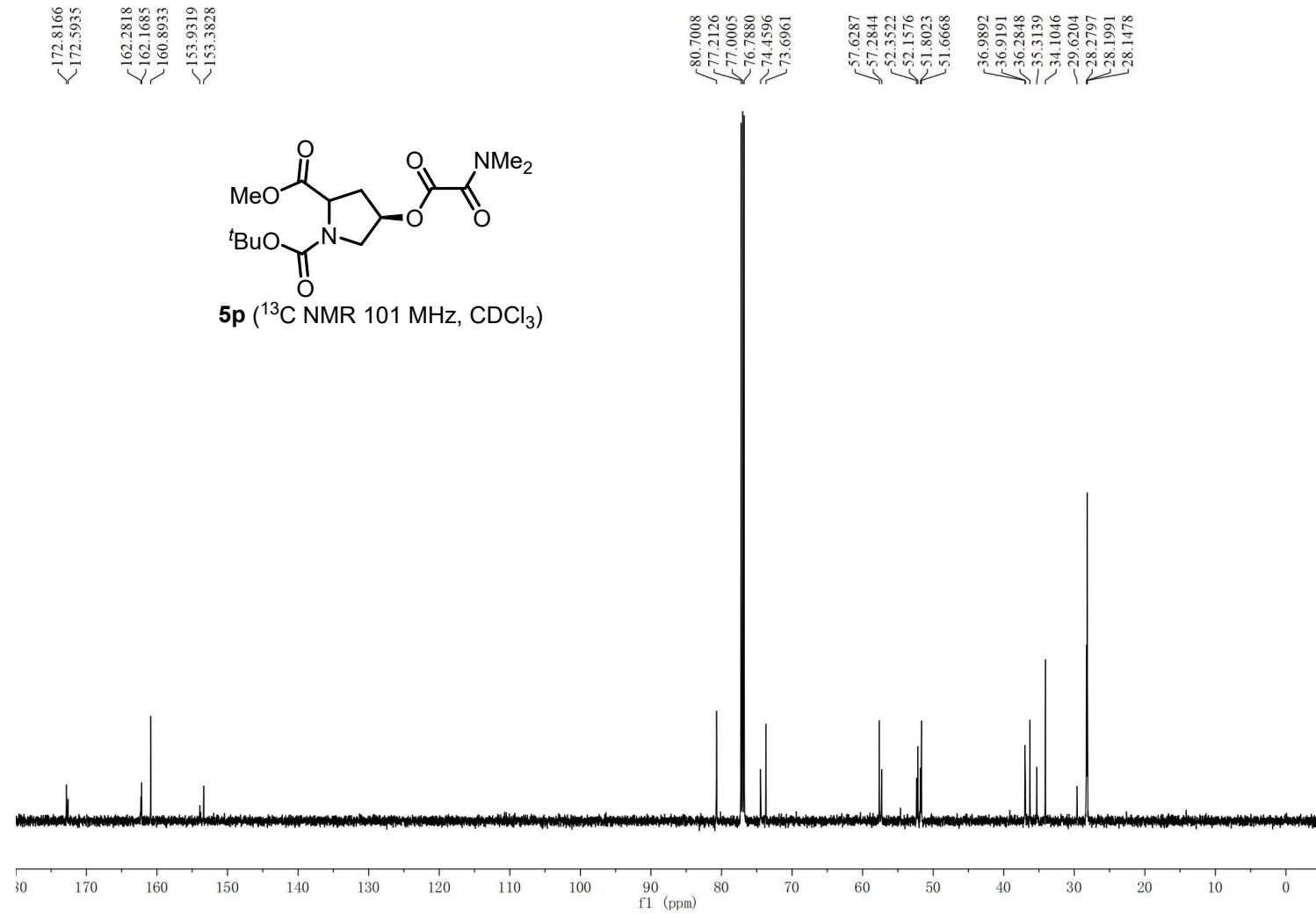
-7.2604



5p (¹H NMR 400 MHz, CDCl₃)

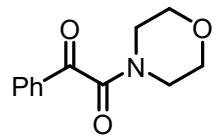


S123

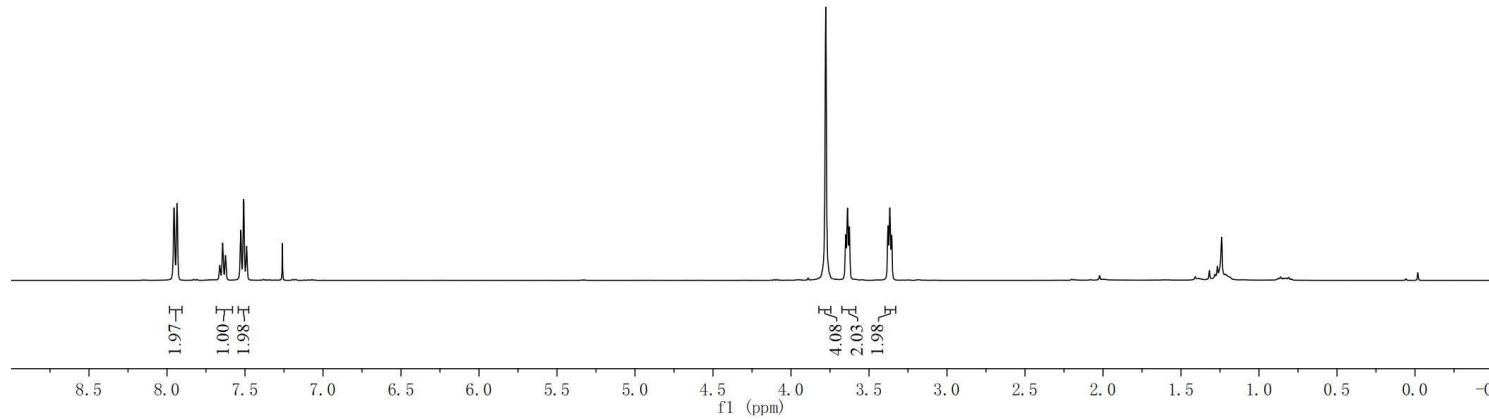


7.9550
7.9387
7.9348
7.9308
7.6630
7.6595
7.6439
7.6408
7.6257
7.6221
7.5288
7.5257
7.5084
7.4902
7.4872
7.2603

3.7783
3.7751
3.6506
3.6473
3.6375
3.6267
3.6233
3.3794
3.3761
3.3664
3.3554
3.3521



6a (^1H NMR 400 MHz, CDCl_3)



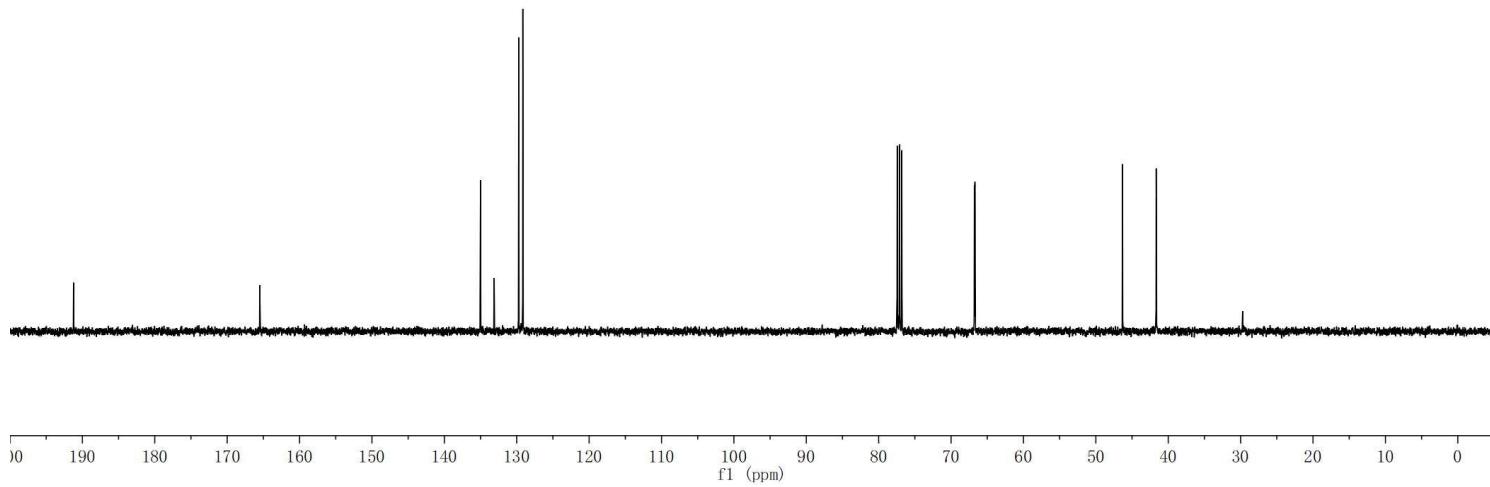
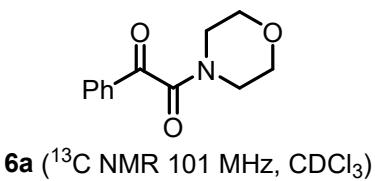
-191.2232

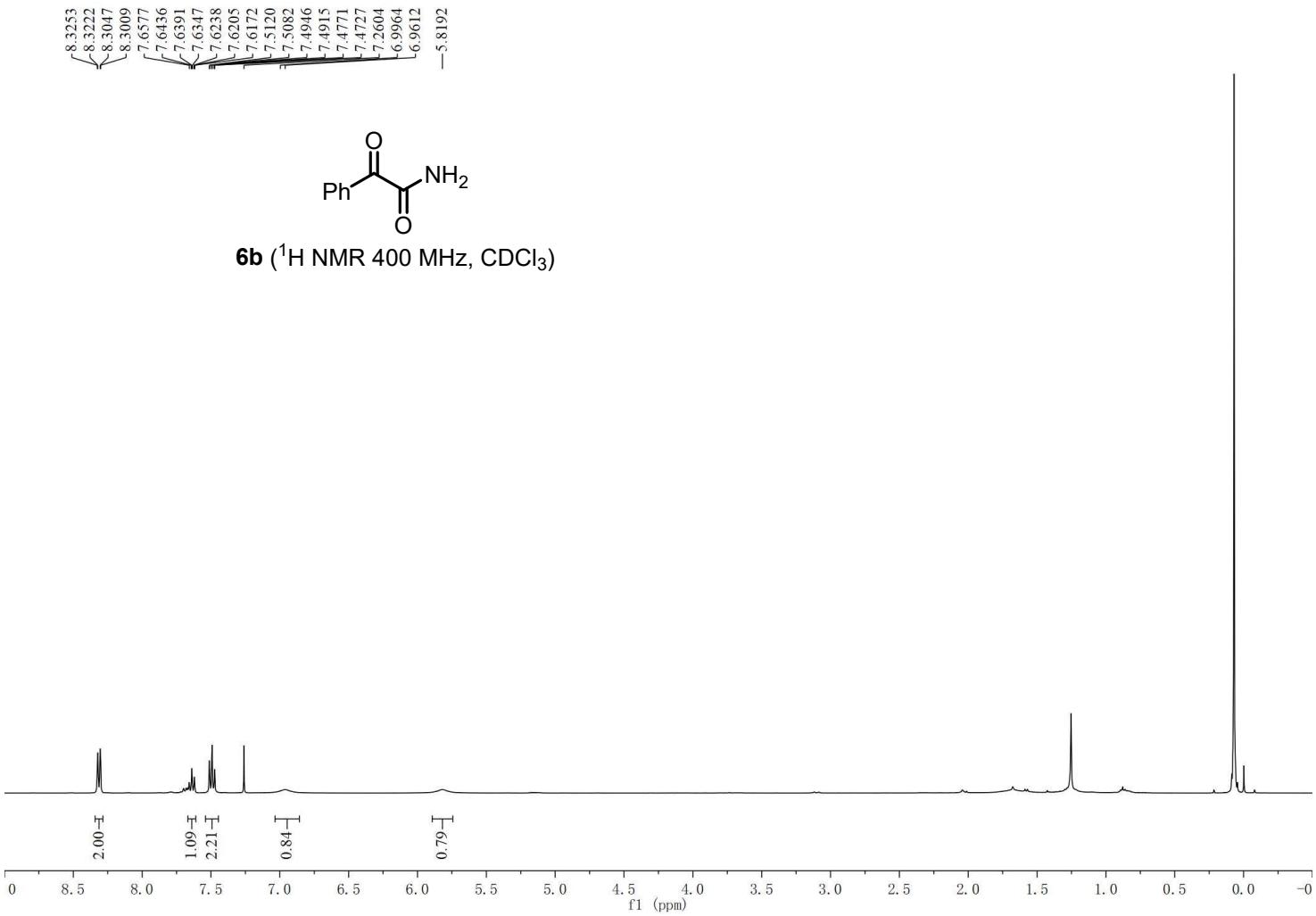
-165.5060

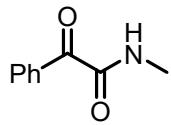
✓135.0019
✓133.1146
✓129.7223
✓129.1611

✓77.4386
✓77.3220
✓77.1195
✓76.8033
✓66.8797
✓66.7066

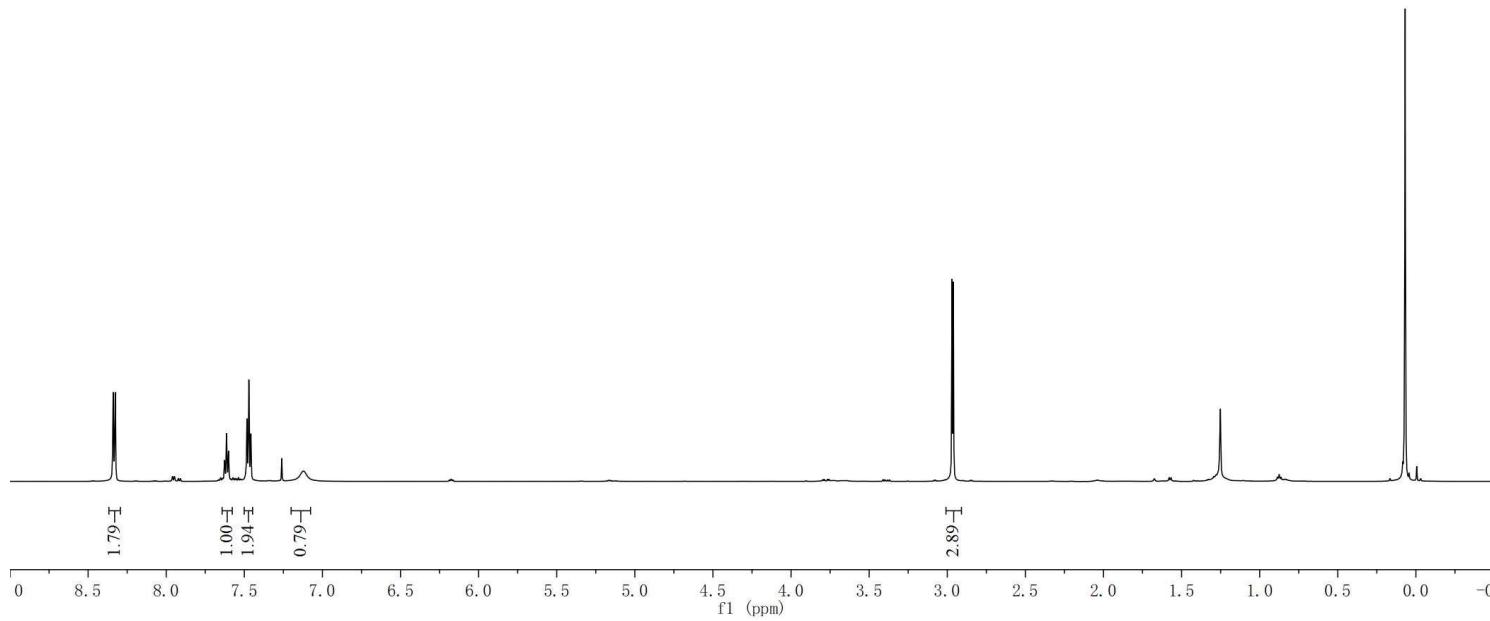
-146.3095
-41.6650



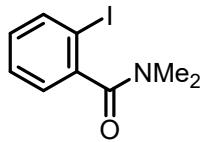




6c (^1H NMR 600 MHz, CDCl_3)



7.8237
7.8209
7.8036
7.8008
7.4027
7.3998
7.3838
7.3810
7.3651
7.3622
7.2605
7.2184
7.2142
7.1952
7.1952
7.0803
7.0761
7.0610
7.0568
7.0418
7.0376



7a (¹H NMR 400 MHz, CDCl₃)

