

On the impact of aromatic core fluorination in hydrogen-bonded liquid crystals

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1. Experimental

1.1. Characterization

Chemicals of analytical purity, sourced from commercial suppliers, were utilized as received. Solvents, when required, underwent drying using conventional techniques. Various spectral data were employed to confirm the purity and chemical composition of all synthesized compounds. Structure characterization of the produced materials was assessed by utilizing ¹H NMR, ¹³C NMR, and ¹⁹F NMR with Varian Unity 400 spectrometers in CDCl₃ solution. ¹H NMR and ¹³C NMR chemical shifts are reported in ppm referenced to tetramethylsilane. The residual proton signal of the deuterated solvent was used as internal standard for ¹H NMR and ¹³C NMR spectra. ¹⁹F NMR chemical shifts are reported in ppm referenced to trichlorofluoromethane as an external standard.

Infrared absorption spectra were measured in dry KBr with a Perkin-Elmer B25 spectrophotometer.

The assessment of mesophase phases and the determination of transition temperatures for the hydrogen-bonded compounds involved using a Nikon Optiphot-2 polarizing microscope in conjunction with a Mettler FP-82 HT hot stage and control unit. Enthalpies were determined by analyzing DSC thermograms obtained with a Perkin-Elmer DSC-7 instrument, employing a heating and cooling rate of 10 K min⁻¹.

The photoisomerization studies in solution were conducted using Ocean Optics HR 2000+ spectrophotometer, and absorption spectra were recorded at room temperature. The solutions in chloroform were taken in a 1cm quartz cuvette and covered to avoid the evaporation of the solvent. The solutions were irradiated with UV light of 1mW/cm² using Bluepoint LED Eco Hönle at a wavelength of 365 nm. A heat filter is inserted between the sample and the source to avoid the influence of UV heat on the sample. The *trans-cis-trans* photoisomerization in the LC phase was performed using of 1 mW/cm² Bluepoint LED Eco Hönle at a wavelength of 365 nm.

X-ray investigations were carried out with an Incoatec (Geesthacht, Germany) IpS microfocus source with a monochromator for CuK α radiation (λ = 0.154 nm), calibration with the powder pattern of Pb(NO₃)₂. A droplet of the sample was placed on a glass plate on a Linkam hot stage HFS-X350-GI (rate: 1 K/min). Exposure time was 5 min; the sample-detector distance was 9.00 cm for WAXS and 26.80 cm for SAXS. The diffraction patterns were recorded with a Vantec 500 area detector (Bruker AXS, Karlsruhe) and transformed into 1D plots using GADDS software.

1.2. Synthesis

1.2.1. Synthesis of B6F₂, B6F₃ and B6F₂₃

1.2.1.1. 2-Fluoro-4-n-hexyloxybenzoic acid, B6F₂

2-Fluoro-4-n-hexyloxybenzonitrile, BF₂

A suspension of 2-fluoro-4-hydroxybenzonitrile (2.74 g, 20 mmol, 1 eq.), 1-bromohexane (3.6 g, 22 mmol, 1.1 eq.), potassium carbonate (13.8 g, 100 mmol, 5 eq.) and a catalytic amount of *tetra-n*-butylammoniumiodide were refluxed in dry butanone for 5 hours. The solvent was

removed under vacuum and the crude product was purified by column chromatography (eluent: first hexane, then CHCl_3); yield 3.55 g (16 mmol, 80%); colourless liquid; $\text{C}_{13}\text{H}_{16}\text{FNO}$; $M = 221.27$ g/mol; ^1H NMR (400 MHz, CDCl_3) δ 7.49 (dd, $J = 8.7$ Hz, 7.5 Hz, 1H, Ar-H), 6.74 (dd, $J = 8.7$ Hz, 2.4 Hz, 1H, Ar-H), 6.69 (dd, $J = 11.1$ Hz, 2.3 Hz, 1H, Ar-H), 3.99 (t, $J = 6.5$ Hz, 2H, OCH_2CH_2), 1.84–1.75 (m, 2H, OCH_2CH_2), 1.49–1.40 (m, 2H, CH_2), 1.39–1.29 (m, 4H, CH_2), 0.91 (t, $J = 7.0$ Hz, 3H, CH_3); ^{19}F NMR (376 MHz, CDCl_3) δ –104.36 (dd, $J = 11.0$ Hz, 7.5 Hz).

2-Fluoro-4-n-hexyloxybenzoic acid, B6F₂.

A solution of 2-fluoro-4-*n*-hexyloxybenzonitrile (**BF₂**) (3.55 g, 16 mmol, 1 eq.) and potassium hydroxide (4.5 g, 80 mmol, 5 eq.) were refluxed in ethanol and water (2/1, v/v). Afterwards the solution is concentrated and acidified with concentrated hydrochloric acid and the resulting product was filtered off, washed with water and dried; yield 1.9 g (8 mmol, 50%); colourless solid; $\text{C}_{13}\text{H}_{17}\text{FO}_3$; $M = 240.27$ g/mol; Cr 110 °C N 120 °C Iso; ^1H NMR (400 MHz, CDCl_3) δ 7.96 (t, $J = 8.7$ Hz, 1H, Ar-H), 6.73 (dd, $J = 8.9$, 2.5 Hz, 1H, Ar-H), 6.64 (dd, $J = 12.9$, 2.4 Hz, 1H, Ar-H), 4.01 (t, $J = 6.6$ Hz, 2H, OCH_2CH_2), 1.87–1.73 (m, 2H, OCH_2CH_2), 1.48–1.44 (m, 2H, CH_2), 1.37–1.32 (m, 4H, CH_2), 0.91 (t, $J = 7.0$ Hz, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 168.6, 165.4, 165.0, 162.8, 134.0, 110.7, 102.5, 68.7, 31.4, 28.8, 25.5, 22.5, 13.9; ^{19}F NMR (378 MHz, CDCl_3) δ –105.02 (dd, $J = 12.8$ Hz, 8.6 Hz).

1.2.1.2. 3-Fluoro-4-n-hexyloxybenzoic acid, B6F₃

3-Fluoro-4-n-hexyloxybenzaldehyde, BF₃

A suspension of 3-fluoro-4-hydroxybenzaldehyde (2.8 g, 20 mmol, 1 eq.), 1-bromohexane (3.6 g, 22 mmol, 1.1 eq.), potassium carbonate (13.8 g, 100 mmol, 5 eq.) and a catalytic amount of tetra-*n*-butylammoniumiodide were refluxed in dry butanone for 5 hours. The solvent was removed under vacuum and the crude product was purified by column chromatography (eluent: first hexane, then CHCl_3); yield 3.1 g (14 mmol, 70%); colourless liquid; $\text{C}_{13}\text{H}_{17}\text{FO}_2$; $M = 224.27$ g/mol; ^1H NMR (400 MHz, CDCl_3) δ 9.85 (s, 1H, CHO), 7.64–7.58 (m, 2H, Ar-H), 7.05 (t, $J = 8.2$ Hz, 1H, Ar-H), 4.11 (t, $J = 6.6$ Hz, 2H, OCH_2CH_2), 1.90–1.82 (m, 2H, OCH_2CH_2), 1.53–1.43 (m, 2H, CH_2), 1.40–1.30 (m, 4H, CH_2), 0.91 (t, $J = 7.0$ Hz, 3H, CH_3); ^{19}F NMR (376 MHz, CDCl_3) δ –132.96 (dd, $J = 10.9$ Hz, $J = 7.1$ Hz).

3-Fluoro-4-n-hexyloxybenzoic acid, B6F₃

A mixture of the 3-fluoro-4-*n*-hexyloxybenzaldehyde (**BF₃**) (3.1 g, 14 mmol, 1 eq.), resorcinol (1.9 g, 17.5 mmol, 1.25 eq.) were dissolved under stirring at room temperature in *tert*-butanol (140 mL). Afterwards a solution of NaOCl₂ (7.3 g, 81 mmol, 5.8 eq.) and NaH₂PO₄·2H₂O (4.9 g, 42 mmol, 3 eq.) in distilled water (80 mL) were added dropwise. After finishing the reaction the solution was concentrated and acidified with concentrated hydrochloric acid and resulting product was filtered off, washed with water and dried; yield 2.0 g (8.4 mmol, 60%); colourless solid; C₁₃H₁₇FO₃; *M* = 240.27 g/mol; Cr 108 °C Iso; ¹H NMR (400 MHz, CDCl₃) δ 7.85 (ddd, *J* = 8.6, 1.9, 1.1 Hz, 1H, Ar-H), 7.78 (dd, *J* = 11.5, 2.0 Hz, 1H, Ar-H), 6.97 (t, *J* = 8.3 Hz, 1H, Ar-H), 4.08 (t, *J* = 6.6 Hz, 2H, OCH₂CH₂), 1.89–1.78 (m, 2H, OCH₂CH₂), 1.49–1.43 (m, 2H, CH₂), 1.36–1.31 (m, 4H, CH₂), 0.89 (t, *J* = 7.0 Hz, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 170.6, 153.0, 152.1, 150.5, 127.4, 117.8, 113.3, 69.3, 31.4, 28.9, 25.4, 22.5, 13.9; ¹⁹F NMR (378 MHz, CDCl₃) δ –133.86 (dd, *J* = 11.5 Hz, 8.1 Hz).

1.2.1.3. 2,3-Difluoro-4-n-hexyloxybenzoic acid, B6F₂₃

2,3-Difluoro-4-n-hexyloxybenzaldehyde, BF₂₃

Synthesized according to the procedure described above for **BF₃** from 2,3-difluoro-4-hydroxybenzaldehyde (0.8 g, 5.0 mmol) and 1-bromohexane (0.91 g, 5.5 mmol); purification by column chromatography (eluent: first hexane, then CHCl₃); yield 1.0 g (4.1 mmol, 82%); colourless oil; C₁₃H₁₆F₂O₂; *M* = 242.26 g/mol; ¹H NMR (500 MHz, CDCl₃) δ 10.19 (s, 1H, CHO), 7.60 (ddd, *J* = 9.1 Hz, 7.2 Hz, 2.2 Hz, 1H, Ar-H), 6.83 (ddd, *J* = 8.5 Hz, 6.9 Hz, 1.2 Hz, 1H, Ar-H), 4.12 (t, *J* = 6.6 Hz, 2H, OCH₂CH₂), 1.89–1.82 (m, 2H, OCH₂CH₂), 1.52–1.44 (m, 2H, CH₂), 1.39–1.31 (m, 4H, CH₂), 0.91 (t, *J* = 7.1 Hz, 3H, CH₃); ¹⁹F NMR (470 MHz, CDCl₃) δ –145.99 (ddd, *J* = 19.6 Hz, *J* = 7.2, *J* = 1.4 Hz), –159.00 (ddd, *J* = 19.6 Hz, *J* = 6.8, *J* = 2.1 Hz).

2,3-Difluoro-4-n-hexyloxybenzoic acid, B6F₂₃

Synthesized according to the above procedure described for **B6F₃** using 2,3-difluoro-4-*n*-hexyloxybenzaldehyde (**BF₂₃**) (1.0 g, 4.1 mmol), NaOCl₂ (2.2 g, 24 mmol), resorcinol (0.57 g, 5.1 mmol) and NaH₂PO₄·2H₂O (1.5 g, 12.3 mmol); yield 750 mg (2.9 mmol, 71%); Colourless solid; C₁₃H₁₆F₂O₃; *M* = 258.26 g/mol; Cr 108 °C Iso; ¹H NMR (400 MHz, CDCl₃) δ 7.77 (ddd, *J*

= 9.7, 7.7, 2.3 Hz, 1H, Ar-H), 6.79 (ddd, J = 8.9, 7.1, 1.7 Hz, 1H, Ar-H), 4.12 (t, J = 6.6 Hz, 2H, OCH₂CH₂), 1.93–1.77 (m, 2H, OCH₂CH₂), 1.59–1.27 (m, 6H, CH₂), 0.91 (t, J = 7.1 Hz, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 168.2, 153.3, 150.6, 140.2, 127.1, 110.7, 108.3, 69.9, 31.4, 28.8, 25.4, 22.4, 13.9; ¹⁹F NMR (378 MHz, CDCl₃) δ –132.50 (ddd, J = 19.3, 7.6, 1.5 Hz), –158.23 (ddd, J = 19.3, 7.0, 2.2 Hz).

1.2.2. Synthesis of AH and AF

The general procedure involved adding a solution of sodium nitrite (2.28 g, 33 mmol, 1.1 eq.) dissolved in 12 ml of water and 25 ml of a 10% potassium hydroxide aqueous solution to the corresponding fluoro-substituted phenol (30 mmol, 1.0 eq.). The solution was cooled to -20 °C using an acetone/dry ice mixture. Subsequently, a cooled solution of 4-aminopyridine (3.30 g, 36 mmol, 1.2 eq.) dissolved in 10 ml of water and 16 ml of concentrated HCl was added dropwise under vigorous stirring over 90 minutes. The reaction temperature was maintained at around -15 °C throughout the process. After the complete addition, the reaction mixture was stirred for an additional 1 hour, followed by the addition of sodium bicarbonate until no effervescence was observed. The resulting solid material was filtered off, washed with distilled water, dried under vacuum, and used without further purification for the next step.

1.2.3. Synthesis of AH_n and AF_n

Dissolving 2 mmol (1.0 eq.) of the specific 4-hydroxyphenylazopyridine derivatives **AH** or **AF** in 25 ml of DMF, along with the appropriate 1-bromoalkanes (2.4 mmol, 1.2 eq.), K₂CO₃ (6 mmol, 3 eq.), and a catalytic amount of KI, the reaction was agitated for 18 hours at 90°C. After bringing the reaction mixture to room temperature, it was poured into 100 ml of deionized water, resulting in suspension. The suspension was then extracted with ethyl acetate (3 x 50 mL). The mixed organic layers were washed with water and NaHCO₃, dried on anhydrous MgSO₄, and the solvent was removed under reduced pressure. The desired products were obtained from the crude material using column chromatography with ethyl acetate/*n*-hexane (2:8).

4-(4-Octyloxyphenylazo)pyridine, **AH8**

Orange solid. 80% Yield, m.p. 71 °C. ¹H NMR (600 MHz, CDCl₃) δ 8.77 (d, J = 6.1 Hz, 2H, Ar-H), 7.95 (d, J = 9.0 Hz, 2H, Ar-H), 7.67 (d, J = 6.1 Hz, 2H, Ar-H), 7.02 (d, J = 9.0 Hz, 2H,

Ar-H), 4.06 (t, $J = 6.6$ Hz, 2H, $-\text{OCH}_2\text{CH}_2$), 1.86–1.77 (m, 2H, $-\text{OCH}_2\text{CH}_2$), 1.56–1.20 (m, 10H, CH_2), 0.90 (t, $J = 7.0$ Hz, 3H, CH_3). ^{13}C NMR (151 MHz, CDCl_3) δ 162.9, 157.5, 151.1, 146.7, 125.6, 116.1, 114.9, 68.5, 31.8, 29.3, 29.2, 29.1, 25.9, 22.6, 14.1.

4-(4-Decyloxyphenylazo)pyridine, AH10

Orange solid. 65% Yield, m.p. 66 °C. ^1H NMR (600 MHz, CDCl_3) δ 8.77 (d, $J = 6.1$ Hz, 2H, Ar-H), 7.95 (d, $J = 8.9$ Hz, 2H, Ar-H), 7.67 (d, $J = 6.2$ Hz, 2H, Ar-H), 7.02 (d, $J = 8.9$ Hz, 2H, Ar-H), 4.06 (t, $J = 6.6$ Hz, 2H, $-\text{OCH}_2\text{CH}_2$), 1.87–1.78 (m, 2H, $-\text{OCH}_2\text{CH}_2$), 1.52–1.21 (m, 14H, CH_2), 0.89 (t, $J = 7.0$ Hz, 3H, CH_3). ^{13}C NMR (151 MHz, CDCl_3) δ 162.9, 157.5, 151.1, 146.7, 125.6, 116.1, 114.9, 68.5, 31.8, 29.6, 29.5, 29.3, 29.2, 29.1, 25.9, 22.6, 14.1.

4-(4-Dodecyloxyphenylazo)pyridine, AH12

Orange solid. 61% Yield, m.p. 73 °C. ^1H NMR (600 MHz, CDCl_3) δ 8.77 (d, $J = 6.1$ Hz, 2H, Ar-H), 7.95 (d, $J = 9.0$ Hz, 2H, Ar-H), 7.67 (d, $J = 6.1$ Hz, 2H, Ar-H), 7.02 (d, $J = 9.0$ Hz, 2H, Ar-H), 4.06 (t, $J = 6.6$ Hz, 2H, $-\text{OCH}_2\text{CH}_2$), 1.87–1.78 (m, 2H, $-\text{OCH}_2\text{CH}_2$), 1.53–1.21 (m, 18H, CH_2), 0.88 (t, $J = 7.0$ Hz, 3H, CH_3). ^{13}C NMR (151 MHz, CDCl_3) δ 162.9, 157.5, 151.0, 146.7, 125.6, 116.1, 114.9, 68.5, 31.9, 29.6, 29.6, 29.5, 29.5, 29.3, 29.2, 29.1, 25.9, 22.6, 14.1.

4-(4-Tetradecyloxyphenylazo)pyridine, AH14

Orange solid. 65% Yield, m.p. 69 °C. ^1H NMR (600 MHz, CDCl_3) δ 8.77 (d, $J = 5.9$ Hz, 2H, Ar-H), 7.95 (d, $J = 9.0$ Hz, 2H, Ar-H), 7.67 (d, $J = 6.0$ Hz, 2H, Ar-H), 7.02 (d, $J = 9.0$ Hz, 2H, Ar-H), 4.06 (t, $J = 6.6$ Hz, 2H, $-\text{OCH}_2\text{CH}_2$), 1.84–1.80 (m, 2H, $-\text{OCH}_2\text{CH}_2$), 1.56–1.12 (m, 22H, CH_2), 0.88 (t, $J = 7.0$ Hz, 3H, CH_3). ^{13}C NMR (151 MHz, CDCl_3) δ 162.9, 157.5, 151.0, 146.7, 125.6, 116.1, 114.9, 68.5, 31.9, 29.7, 29.6, 29.6, 29.5, 29.5, 29.3, 29.1, 25.9, 22.6, 14.1.

4-(3-Fluoro-4-octoxyphenylazo)pyridine, AF8

Orange crystals. 30% Yield, m.p. 58–59 °C. ^1H NMR (500 MHz, CDCl_3) δ 8.79 (dd, $J = 4.5, 1.6$ Hz, 2H, Ar-H), 7.82 (ddd, $J = 8.7, 2.3, 1.3$ Hz, 1H, Ar-H), 7.72 (dd, $J = 11.9, 2.3$ Hz, 1H, Ar-H), 7.67 (dd, $J = 4.5, 1.6$ Hz, 2H, Ar-H), 7.09 (t, $J = 8.6$ Hz, 1H, Ar-H), 4.14 (t, $J = 6.6$ Hz, 2H, $-\text{OCH}_2\text{CH}_2$), 1.93–1.82 (m, 2H, $-\text{OCH}_2\text{CH}_2$), 1.54–1.26 (m, 10H, CH_2), 0.89 (t, $J = 6.9$ Hz, 3H, CH_3). ^{13}C NMR (126 MHz, CDCl_3) δ 157.1, 153.9, 151.9, 151.3, 151.2, 151.1, 146.2, 124.3, 116.1, 113.4, 107.9, 69.6, 31.8, 29.3, 29.2, 29.0, 25.9, 22.6, 14.1. ^{19}F NMR (470 MHz, CDCl_3) δ -132.39 (dd, $J = 11.6, 8.7$ Hz).

4-(3-Fluoro-4-decoxyphenylazo)pyridine, AF10

Orange crystals. 35% Yield, m.p. 57-58 °C. ¹H NMR (500 MHz, CDCl₃) δ 8.79 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.82 (ddd, *J* = 8.7, 2.4, 1.2 Hz, 1H, Ar-H), 7.72 (dd, *J* = 11.9, 2.3 Hz, 1H, Ar-H), 7.67 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.09 (t, *J* = 8.6 Hz, 1H, Ar-H), 4.14 (t, *J* = 6.6 Hz, 2H, -OCH₂CH₂), 2.01–1.76 (m, 2H, -OCH₂CH₂), 1.60–1.18 (m, 14H, CH₂), 0.90–0.87 (m, 3H, CH₃). ¹³C NMR (126 MHz, CDCl₃) δ 157.1, 153.8, 151.9, 151.3, 151.2, 151.1, 146.2, 124.3, 116.1, 113.4, 107.9, 69.6, 31.8-22.6 (m, 8C), 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ - 132.40 (dd, *J* = 11.6, 8.6 Hz).

4-(3-Fluoro-4-dodecoxyphenylazo)pyridine, AF12

Orange crystals. 29% Yield, m.p. 66-67 °C. ¹H NMR (500 MHz, CDCl₃) δ 8.79 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.82 (ddd, *J* = 8.7, 2.2, 1.2 Hz, 1H, Ar-H), 7.72 (dd, *J* = 11.9, 2.3 Hz, 1H, Ar-H), 7.67 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.09 (t, *J* = 8.6 Hz, 1H, Ar-H), 4.14 (t, *J* = 6.6 Hz, 2H, -OCH₂CH₂), 1.94–1.79 (m, 2H, -OCH₂CH₂), 1.55–1.20 (m, 18H, CH₂), 0.88 (t, *J* = 6.9 Hz, 3H, CH₃). ¹³C NMR (126 MHz, CDCl₃) δ 157.1, 153.9, 151.9, 151.3, 151.2, 151.1, 146.2, 124.2, 116.1, 113.3, 107.9, 69.5, 31.9, 29.7, 29.6, 29.5, 29.4, 29.3, 29.3, 29.0, 25.8, 22.6, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ - 132.38 (dd, *J* = 11.6, 8.7 Hz).

4-(3-Fluoro-4-tetradecoxyphenylazo)pyridine, AF14

Orange crystals. 25% Yield, m.p. 68-69 °C. ¹H NMR (500 MHz, CDCl₃) δ 8.79 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.82 (ddd, *J* = 8.7, 2.3, 1.3 Hz, 1H, Ar-H), 7.72 (dd, *J* = 11.9, 2.3 Hz, 1H, Ar-H), 7.67 (dd, *J* = 4.5, 1.6 Hz, 2H, Ar-H), 7.09 (t, *J* = 8.6 Hz, 1H, Ar-H), 4.14 (t, *J* = 6.6 Hz, 2H, -OCH₂CH₂), 1.95–1.79 (m, 2H, -OCH₂CH₂), 1.56–1.15 (m, 22H, CH₂), 0.88 (t, *J* = 7.0 Hz, 3H, CH₃). ¹³C NMR (126 MHz, CDCl₃) δ 157.1, 153.8, 151.9, 151.3, 151.2, 151.1, 146.2, 124.3, 116.1, 113.4, 107.9, 69.6, 31.9, 29.7-29.3 (m, 8C), 29.0, 25.9, 22.7, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ - 132.38 (dd, *J* = 11.7, 8.6 Hz).

1.2.4. Preparation of final HBCLs supramolecules (HHHn - FFFn)

The target HBLCs (**HHHn - FFFn**) were formed by melting the acid (**B23H6**, **B2F6**, **B3F6** or **B23F6**) and each azopyridine derivatives (**AHn** or **AFn**) in equimolar ratios (1:1) together for few seconds with stirring, resulting in the formation of a homogeneous mixture. The resulting mixture was then allowed to cool to ambient temperature, yielding an orange solid substance. The formed solid was melted once again to ensure the complexation. All complexes exhibited

homogeneous melting and reproducible phase transition temperatures during DSC investigations, indicating the formation of stable hydrogen-bonded supramolecules (see **Figure 2** as examples for DSC traces).

2. NMR Charts

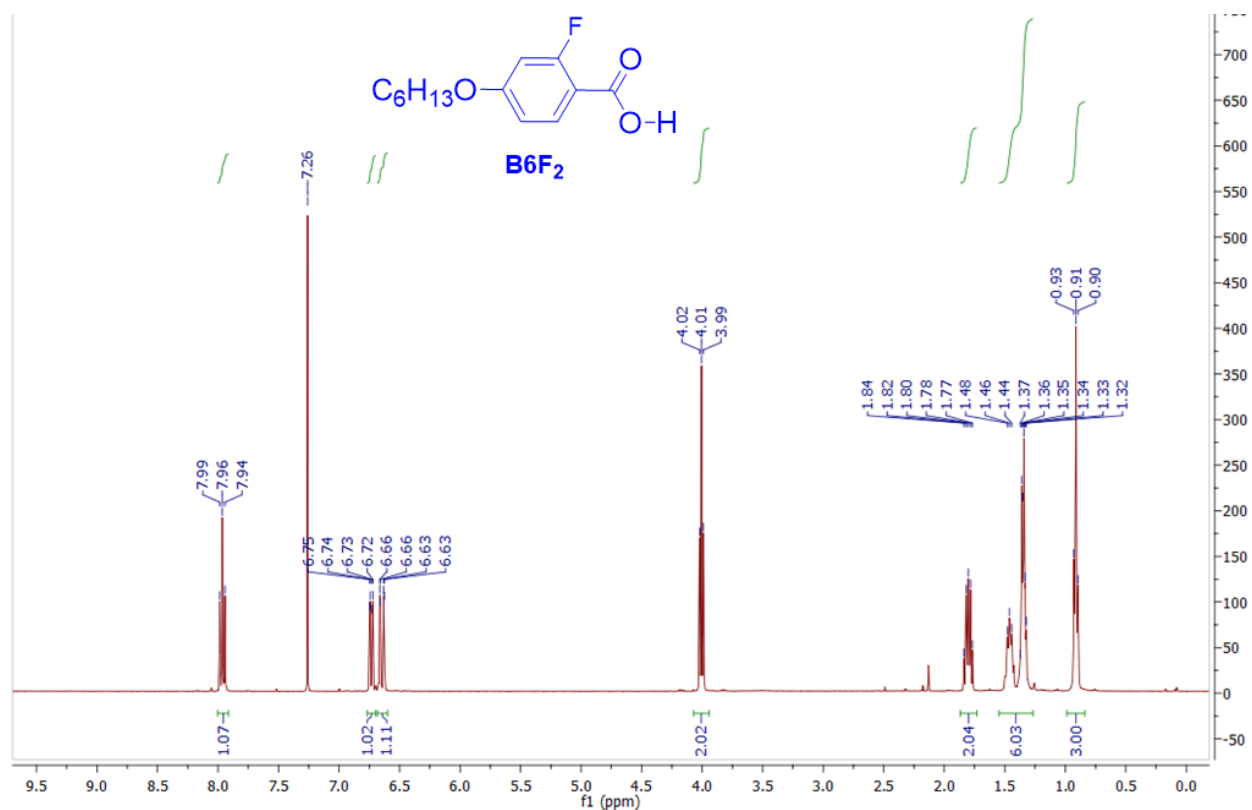


Figure S1a. ¹H NMR of compound **B6F₂**.

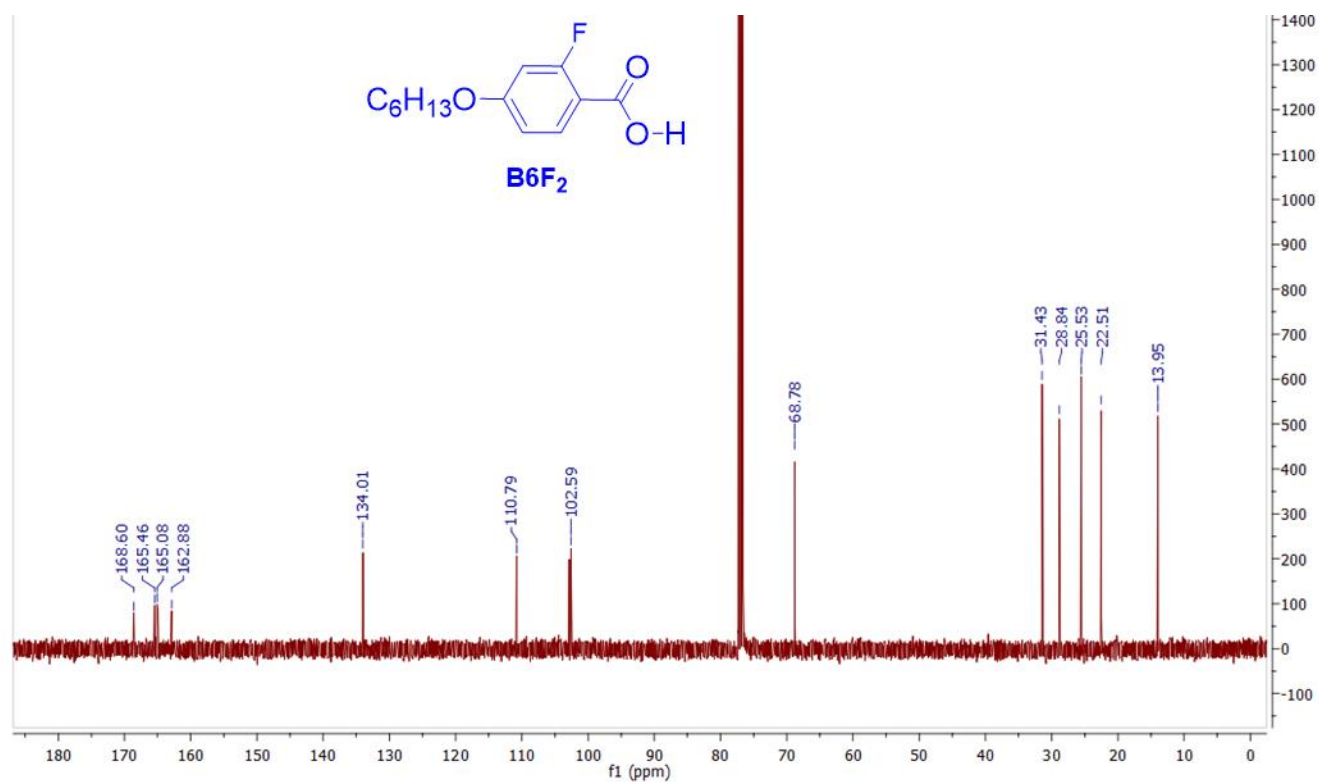


Figure S1b. ¹³C NMR of compound **B6F₂**.

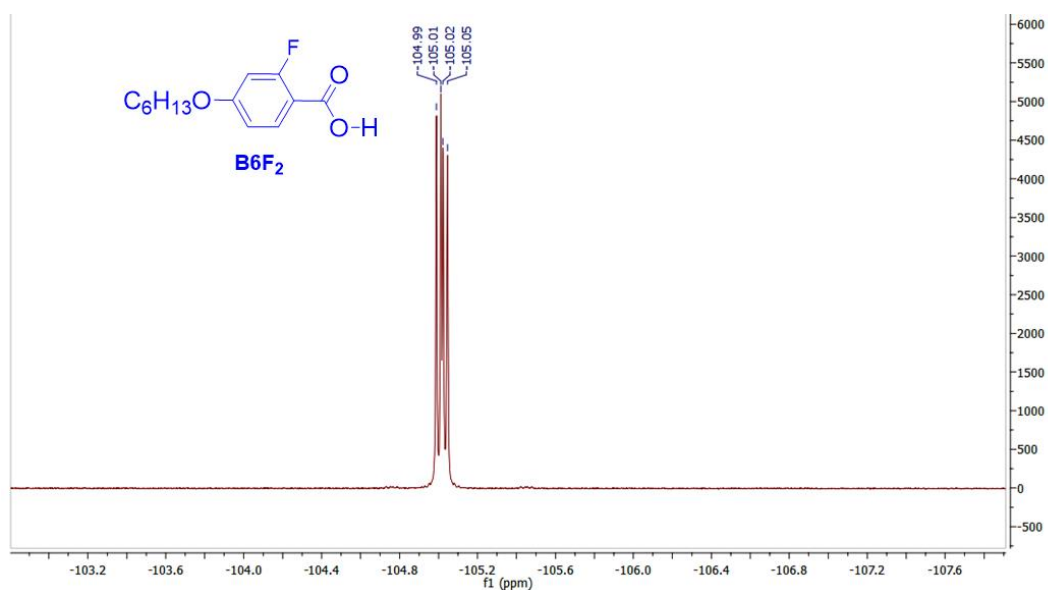


Figure S1c. ¹⁹F NMR of compound **B6F₂**.

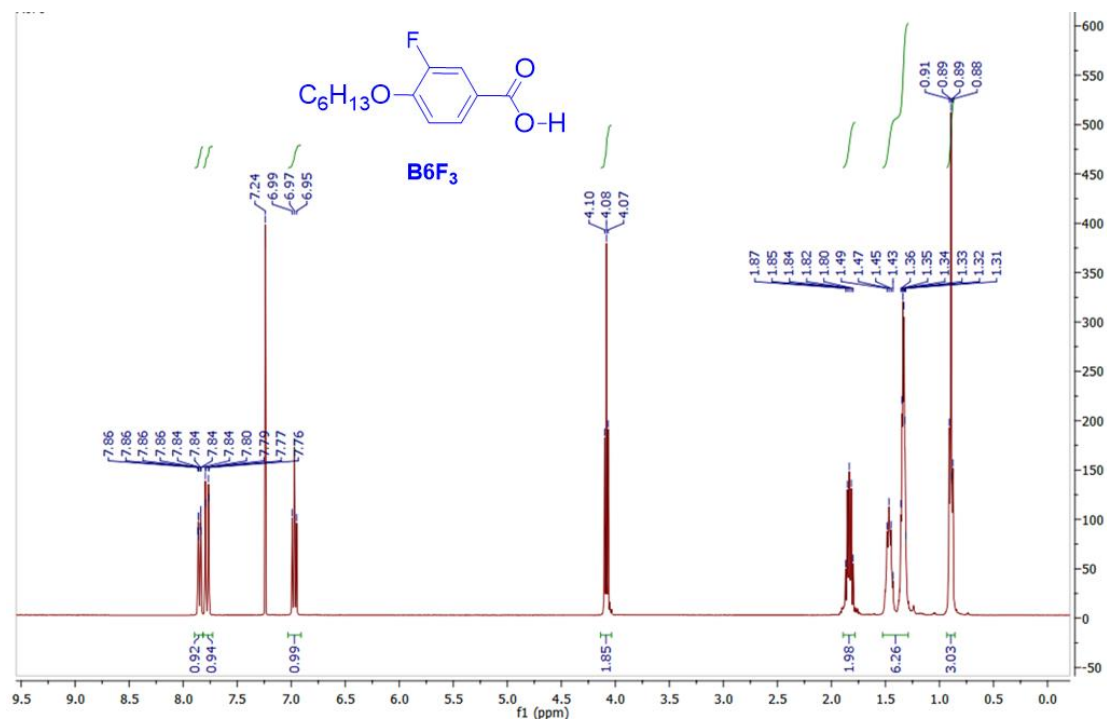


Figure S2a. ¹H NMR of compound **B6F₃**.

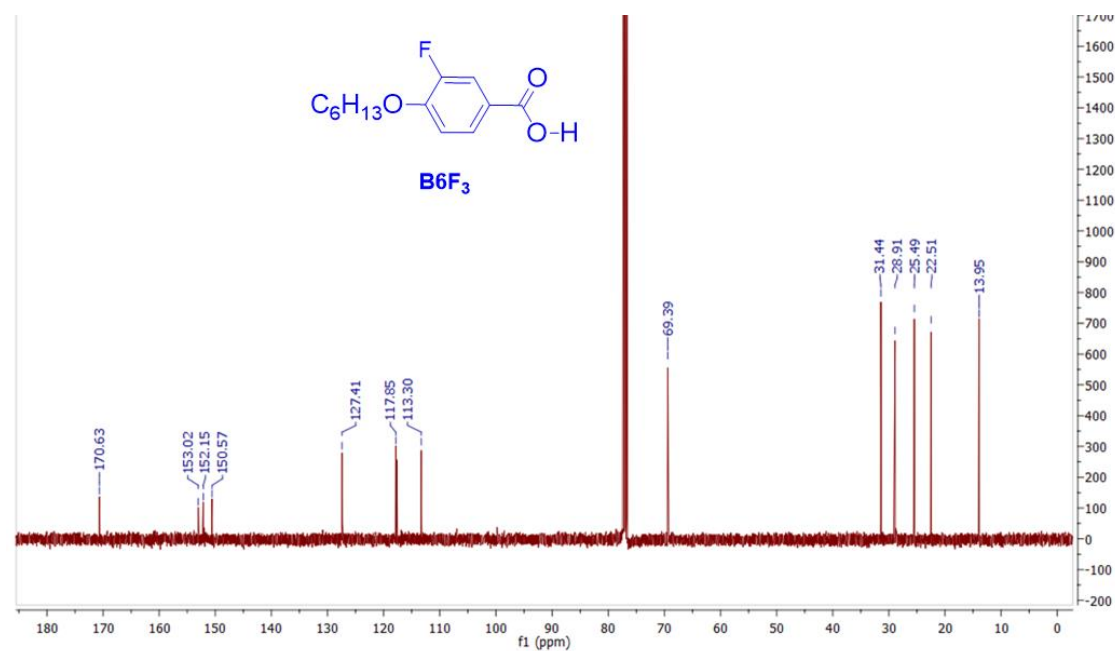


Figure S2b. ¹³C NMR of compound **B6F₃**.

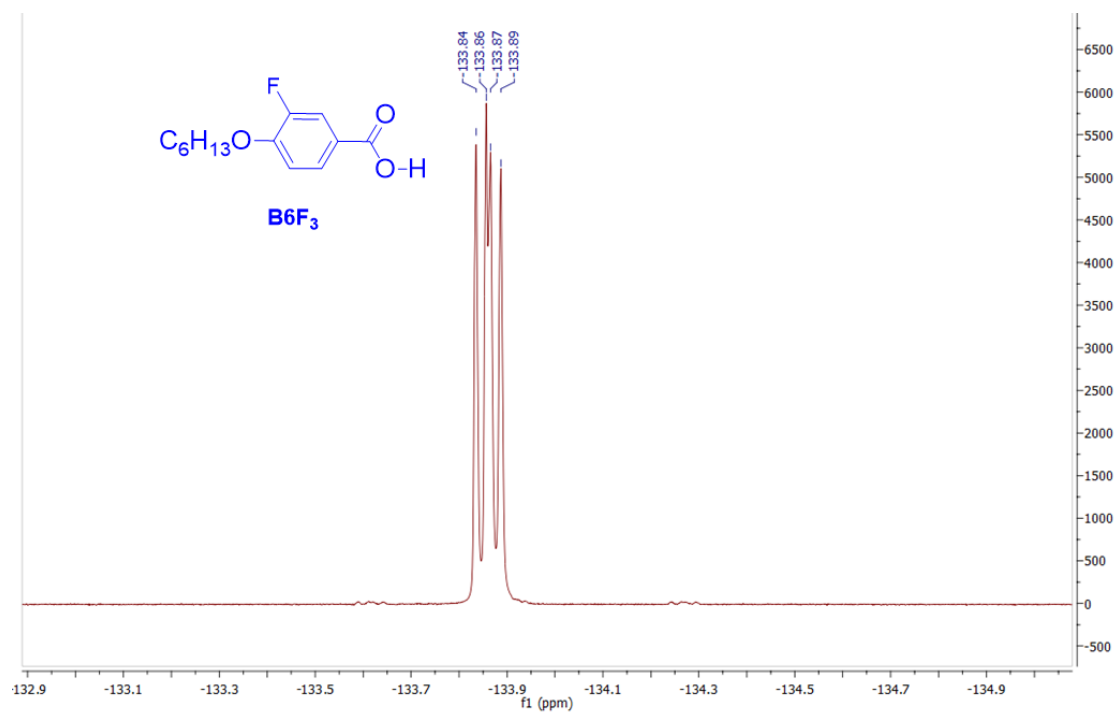


Figure S2c. ¹⁹F NMR of compound **B6F₃**.

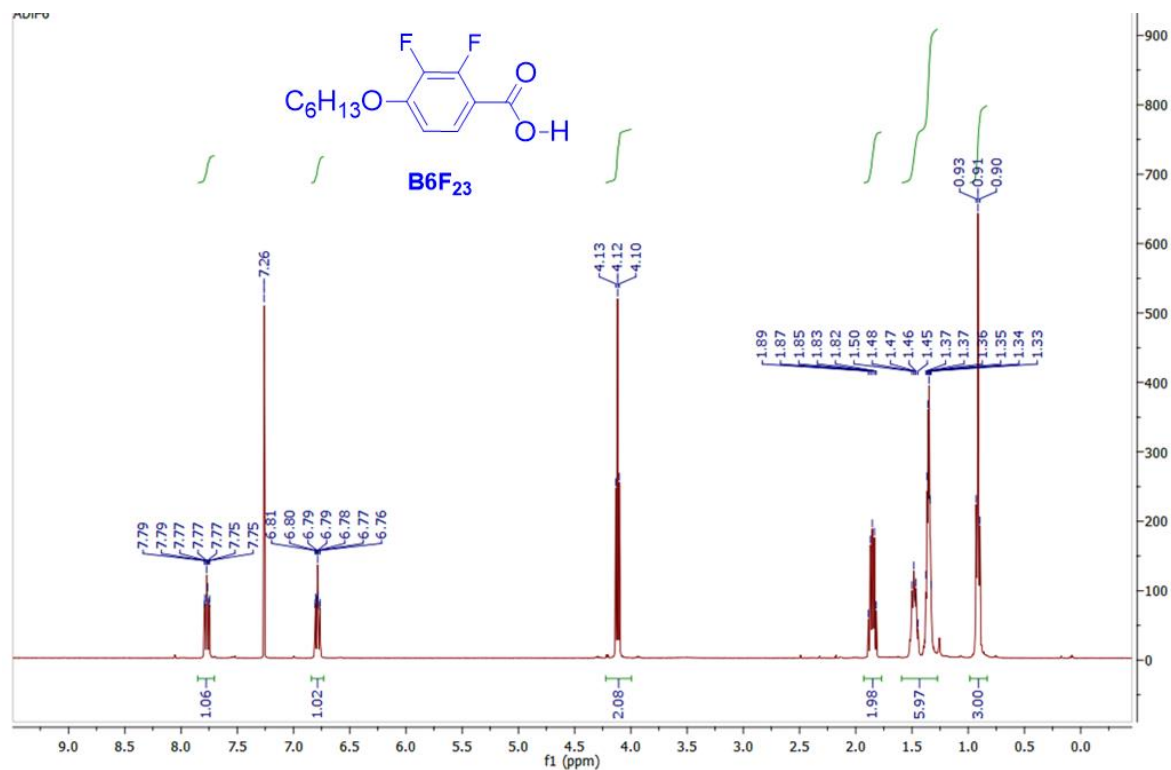


Figure S3a. ¹H NMR of compound **B6F₂₃**.

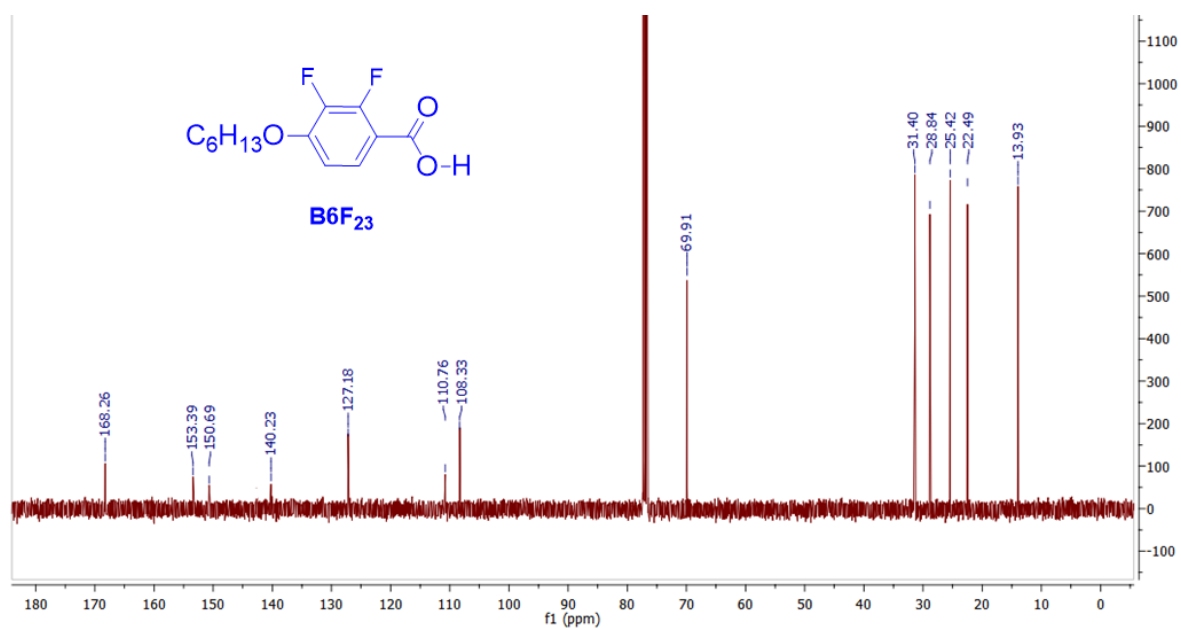


Figure S3b. ¹³C NMR of compound **B6F₂₃**.

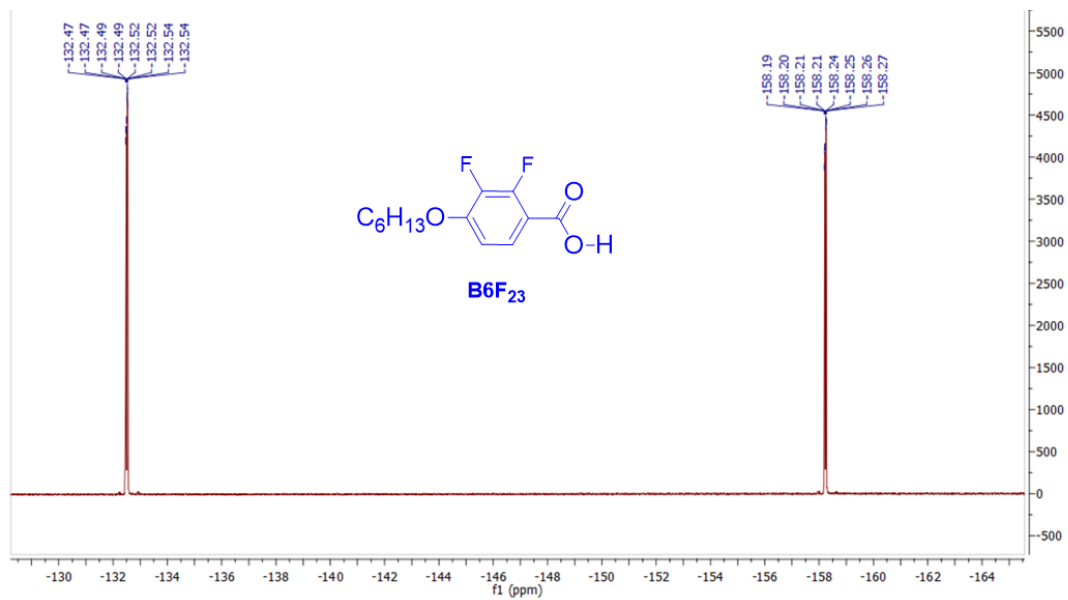


Figure S3c. ¹⁹F NMR of compound **B6F₂₃**.

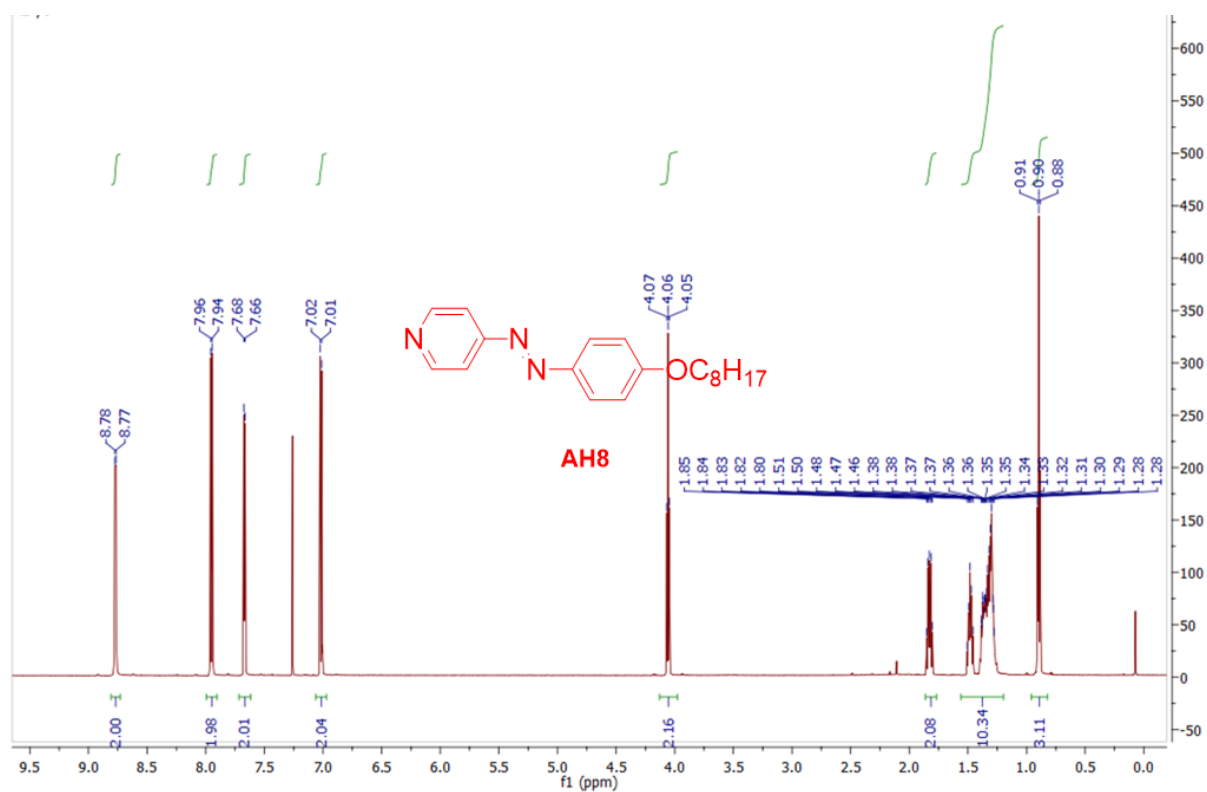


Figure S4a. ¹H NMR of compound AH8.

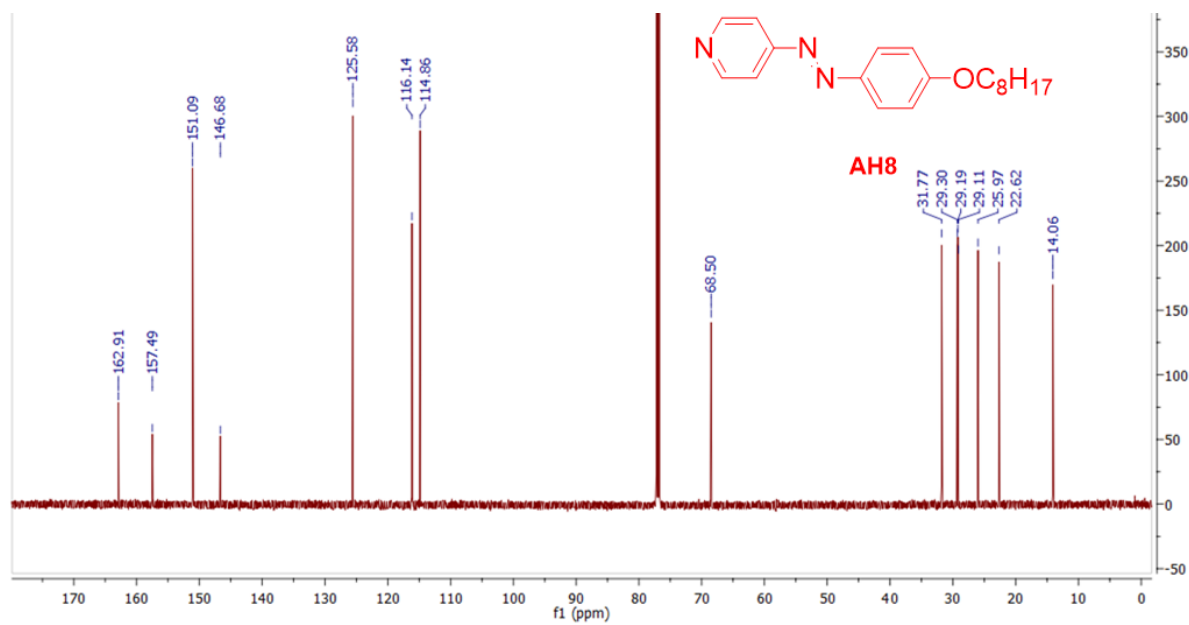


Figure S4b. ¹³C NMR of compound AH8.

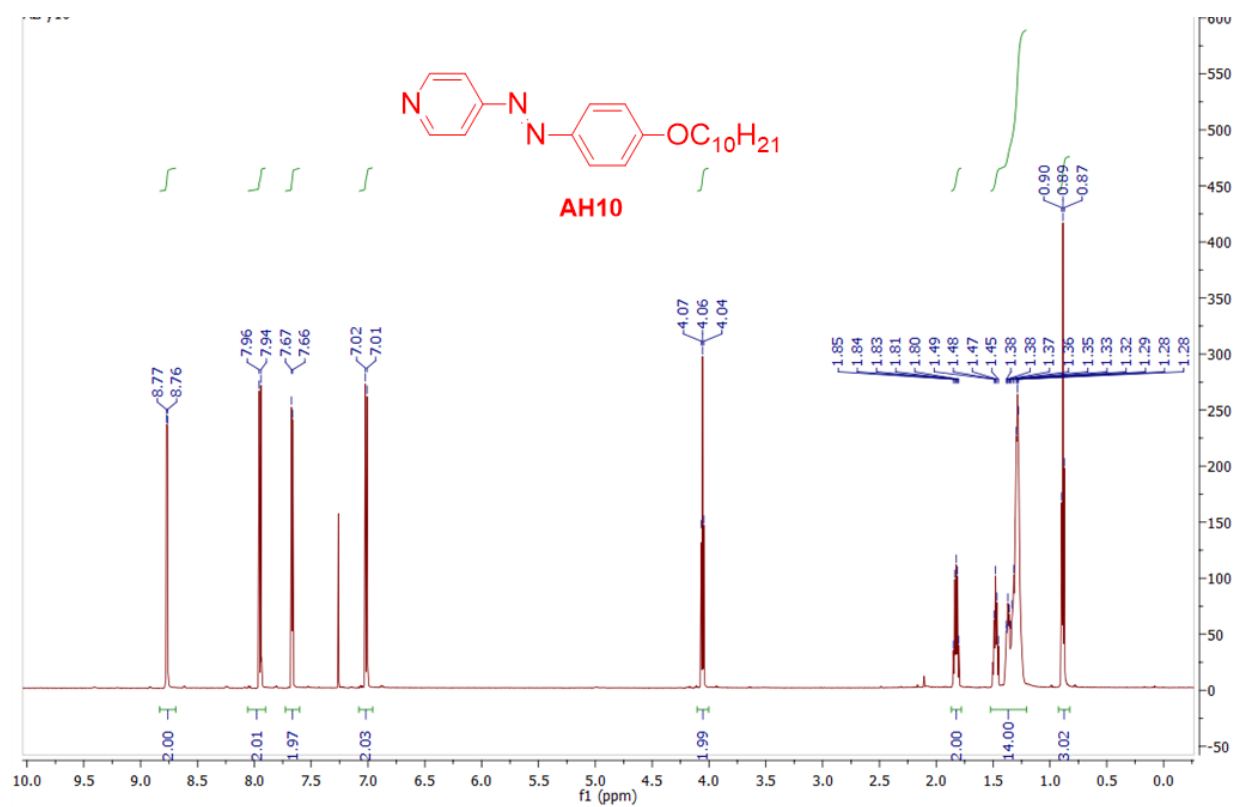


Figure S5a. ^1H NMR of compound AH10.

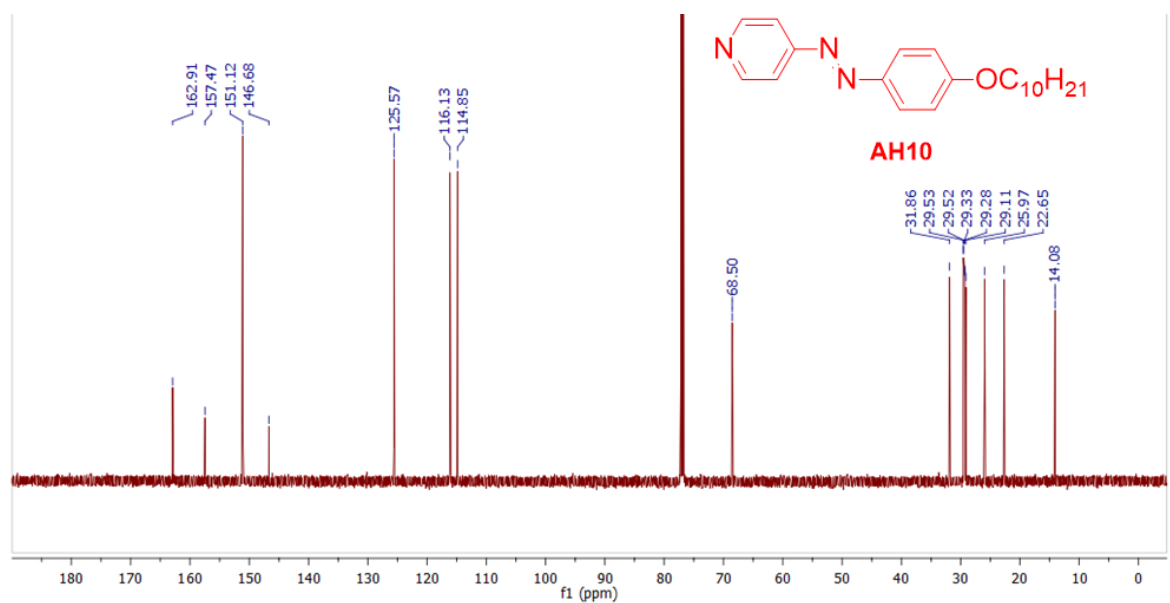


Figure S5b. ^{13}C NMR of compound AH10.

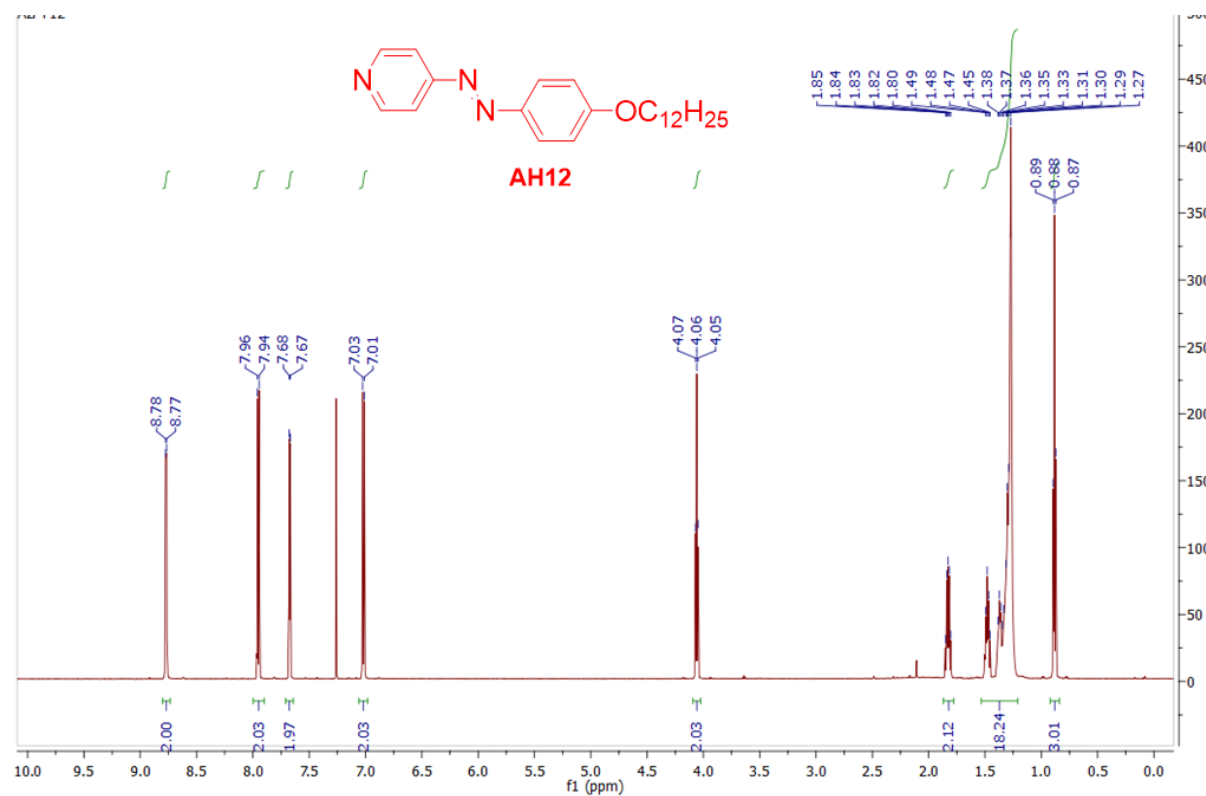


Figure S6a. ^1H NMR of compound AH12.

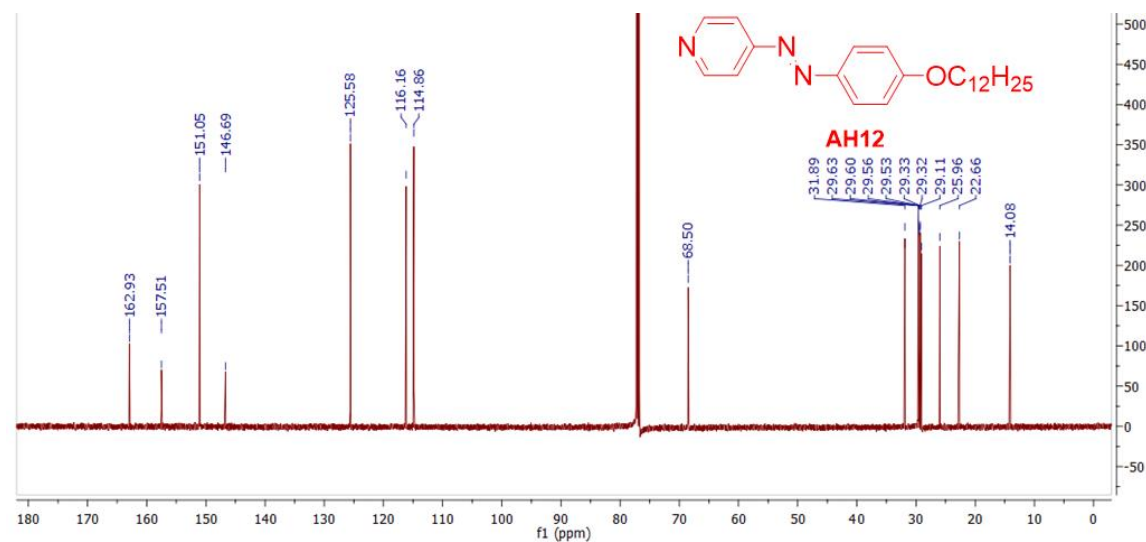


Figure S6b. ^{13}C NMR of compound AH12.

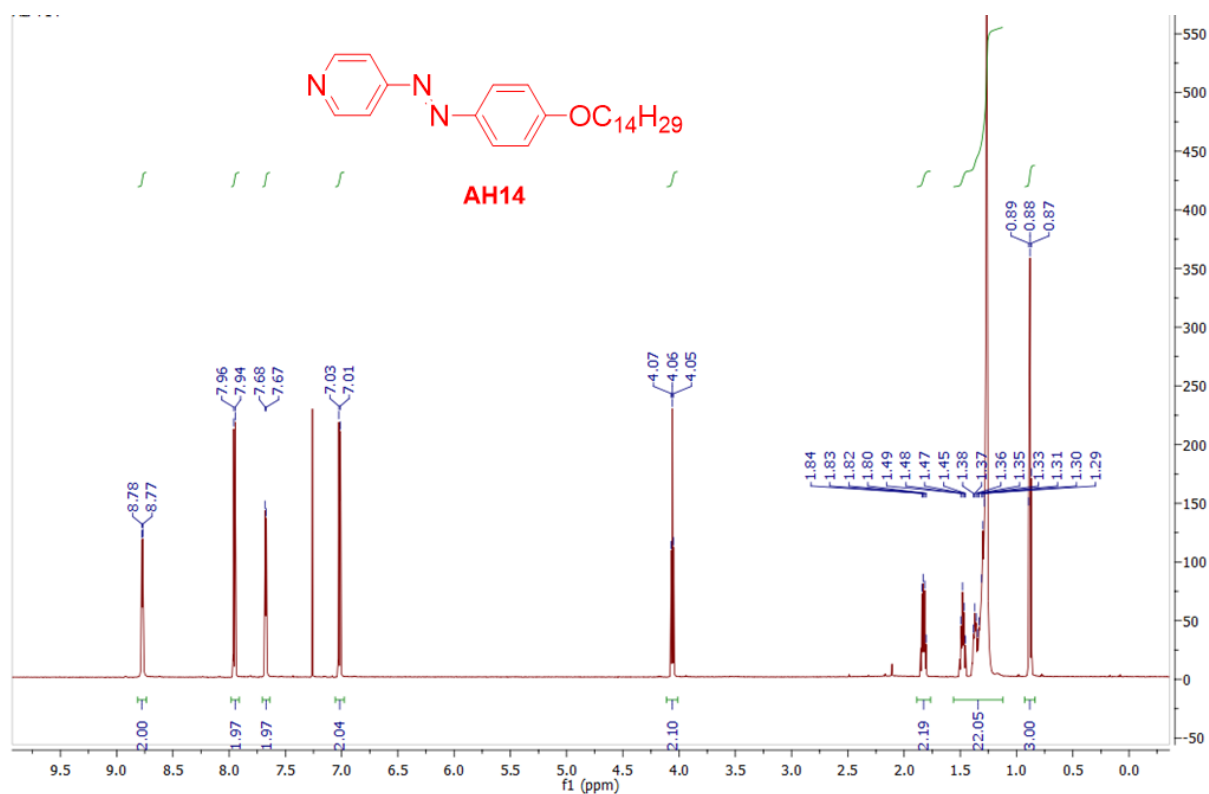


Figure S7a. ¹H NMR of compound AH14.

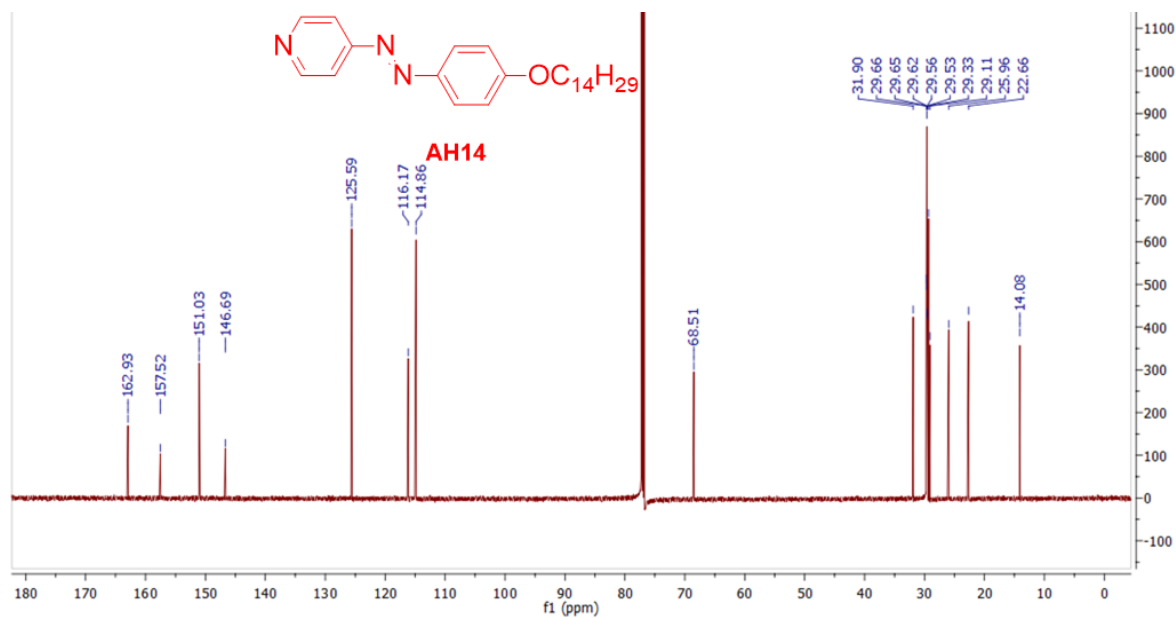


Figure S7b. ¹³C NMR of compound AH14.

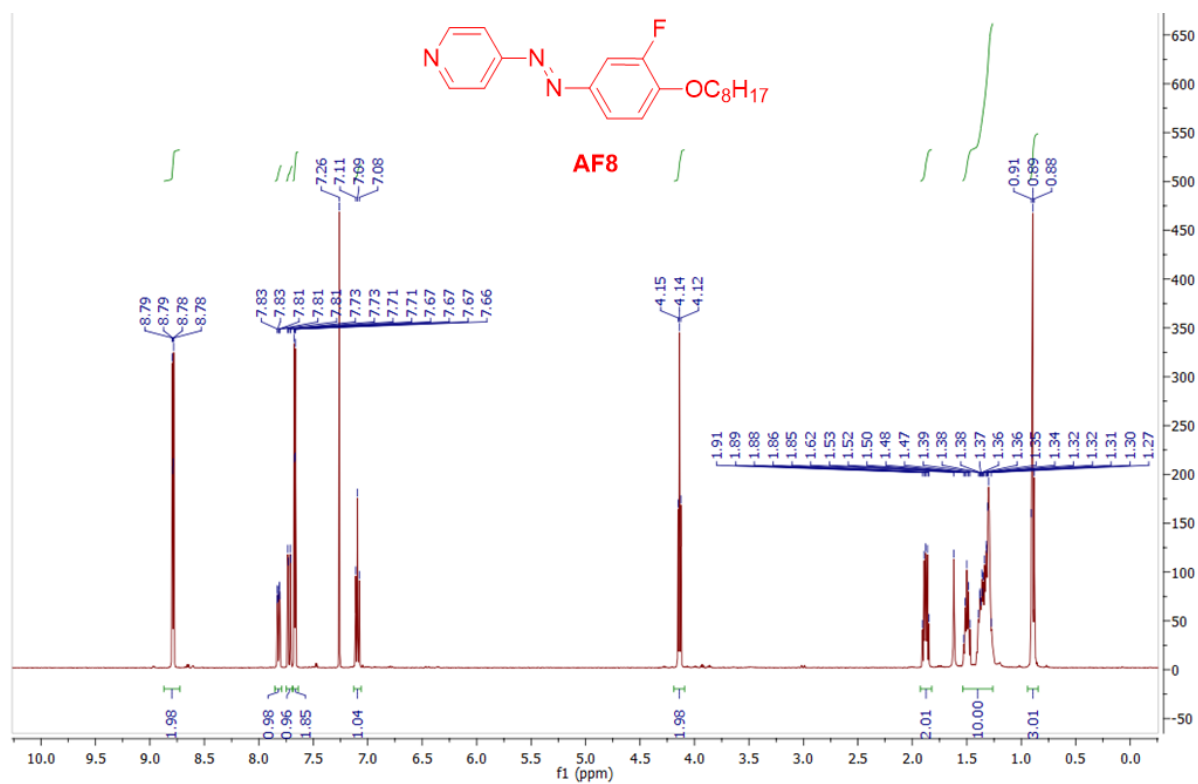


Figure S8a. ^1H NMR of compound AF8.

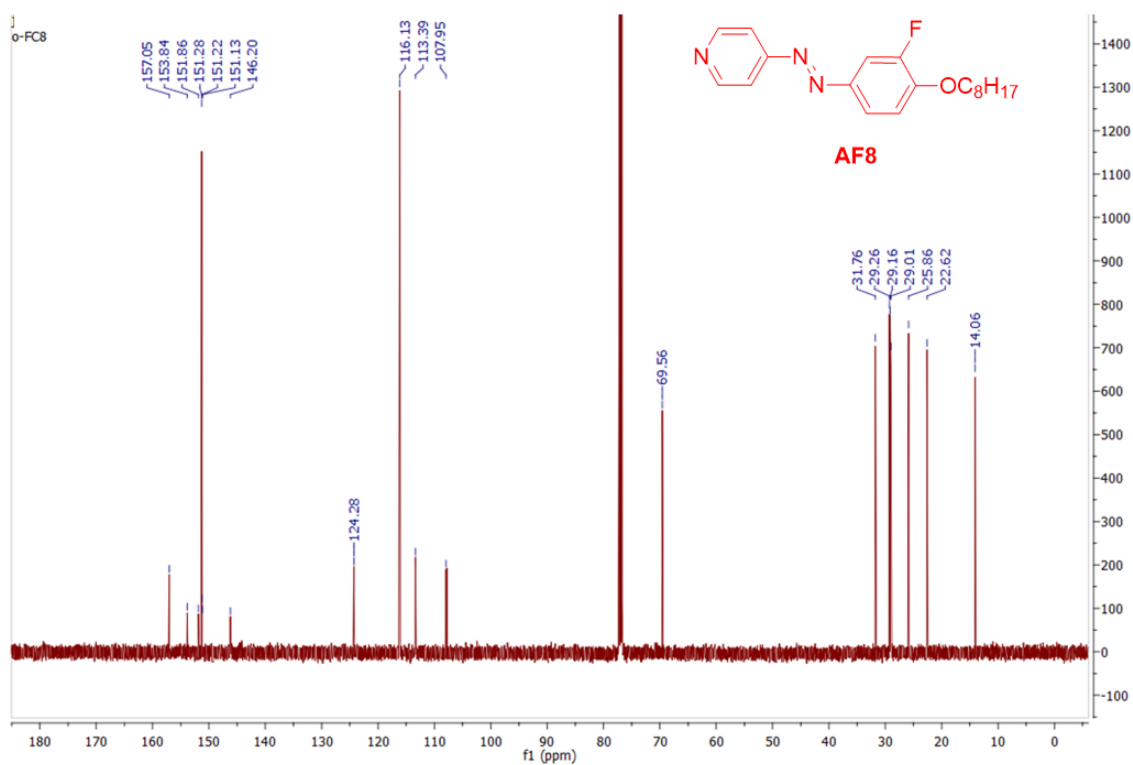


Figure S8b. ^{13}C NMR of compound AF8.

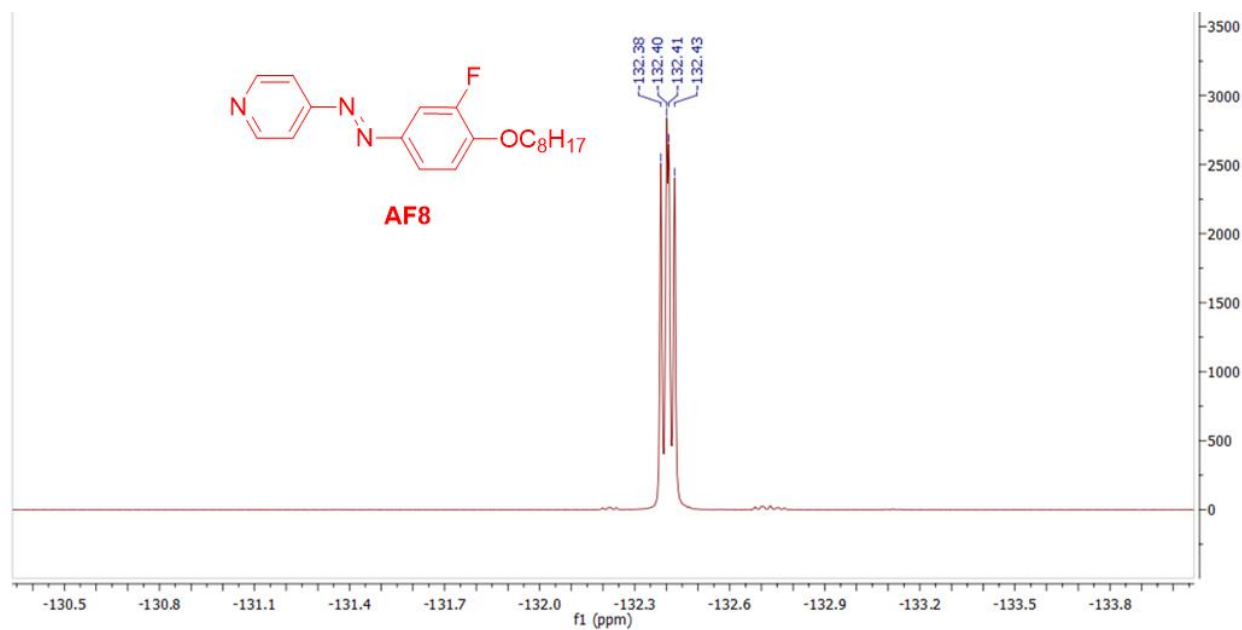


Figure S8c. ^{19}F NMR of compound AF8.

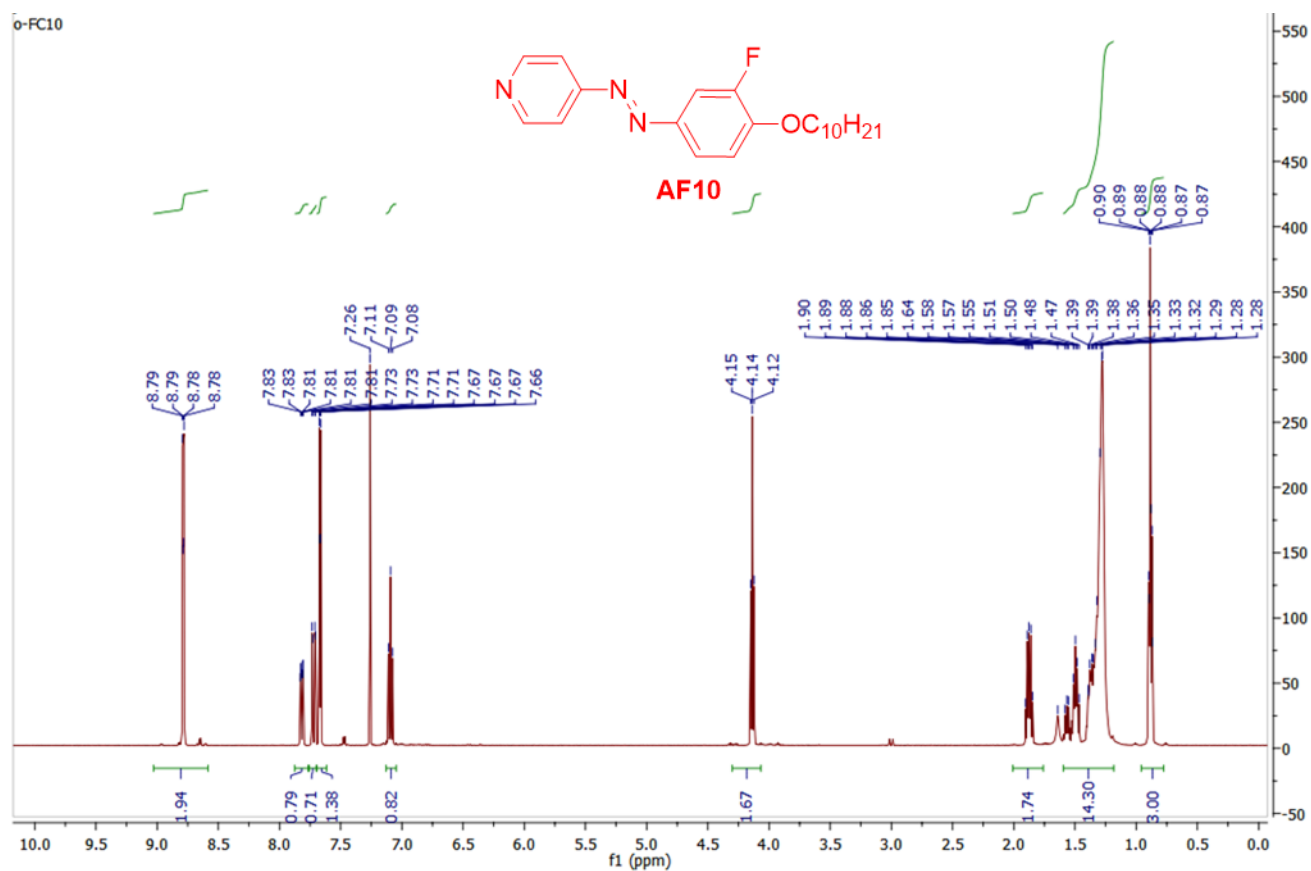


Figure S9a. ^1H NMR of compound AF10.

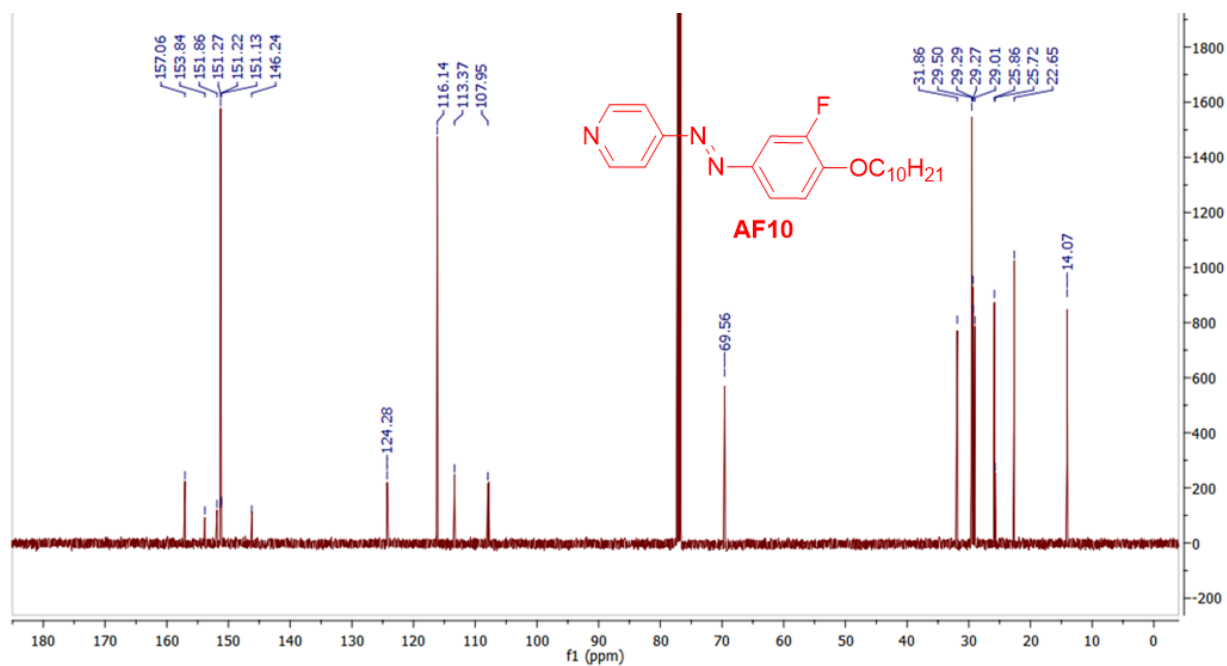


Figure S9b. ^{13}C NMR of compound AF10.

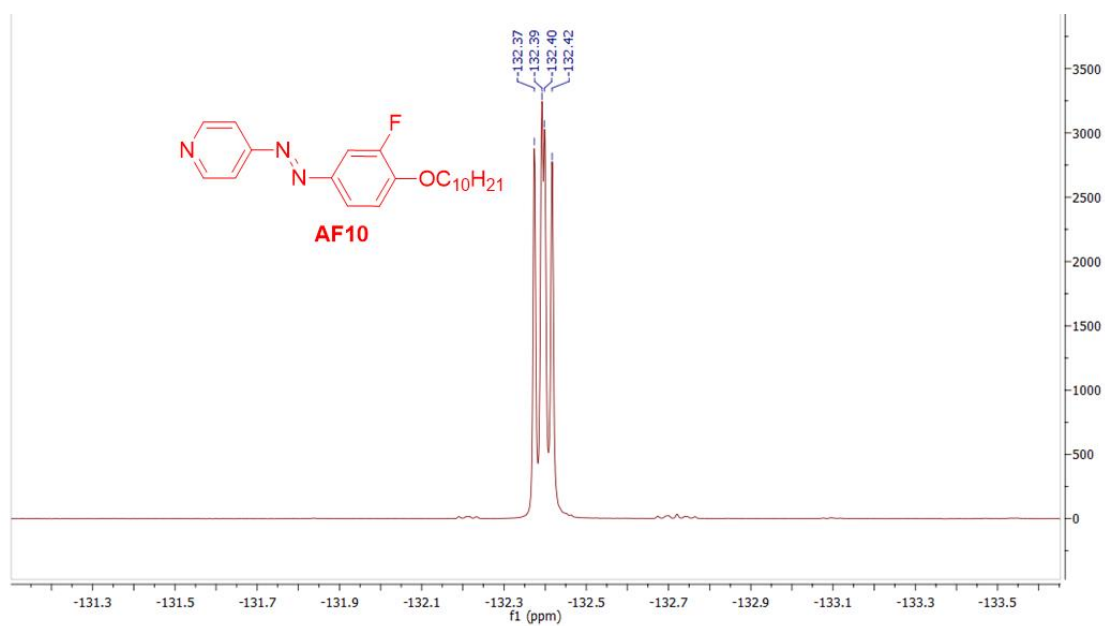


Figure S9c. ^{19}F NMR of compound AF10.

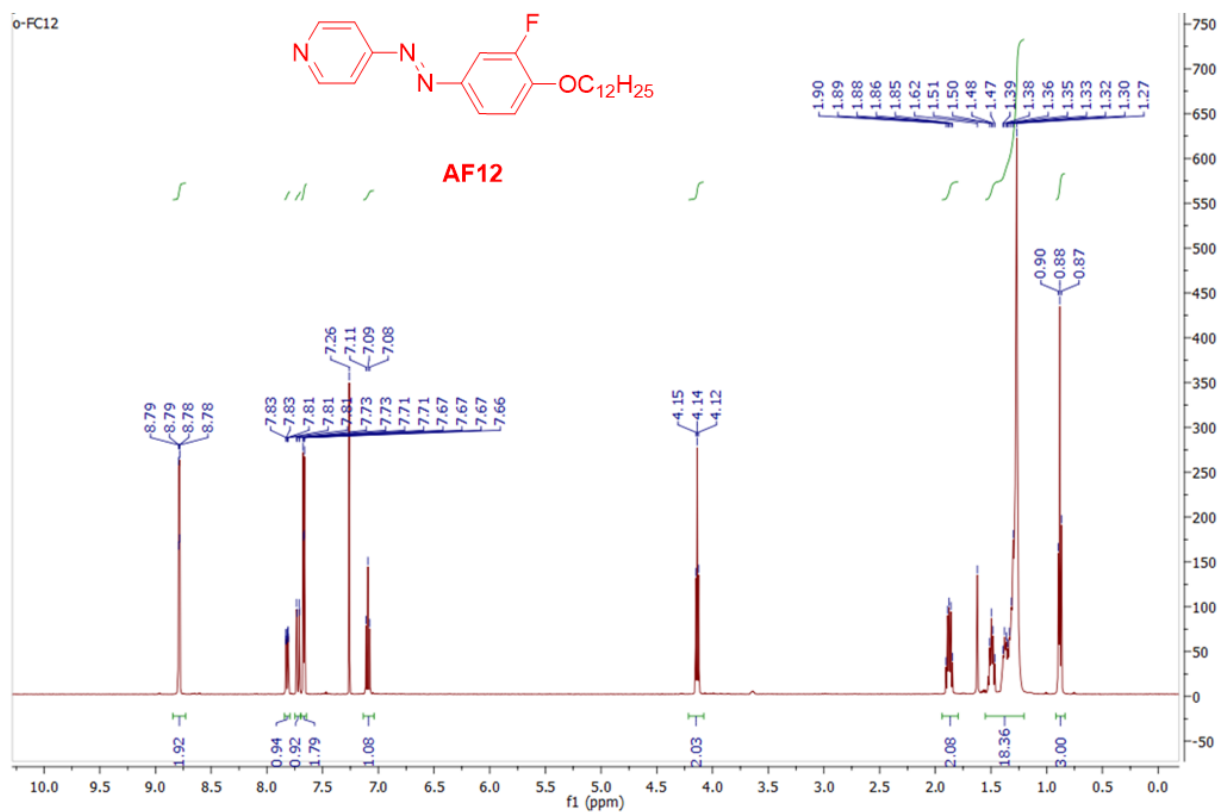


Figure S10a. ^1H NMR of compound AF12.

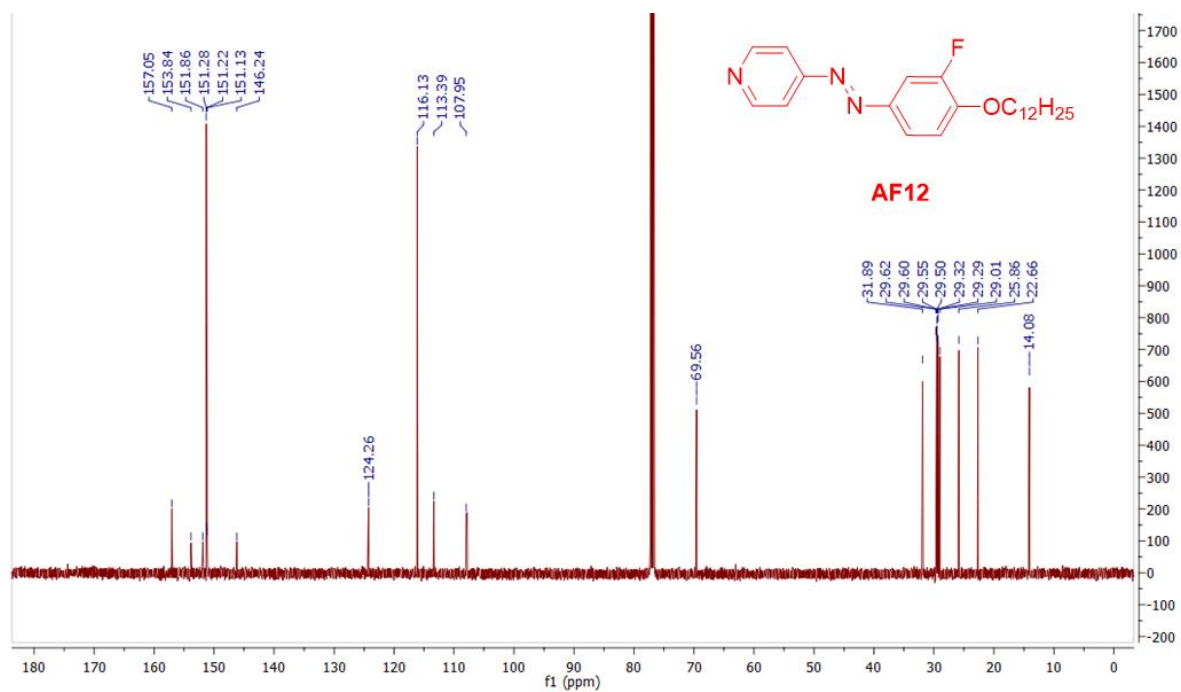


Figure S10b. ^{13}C NMR of compound AF12.

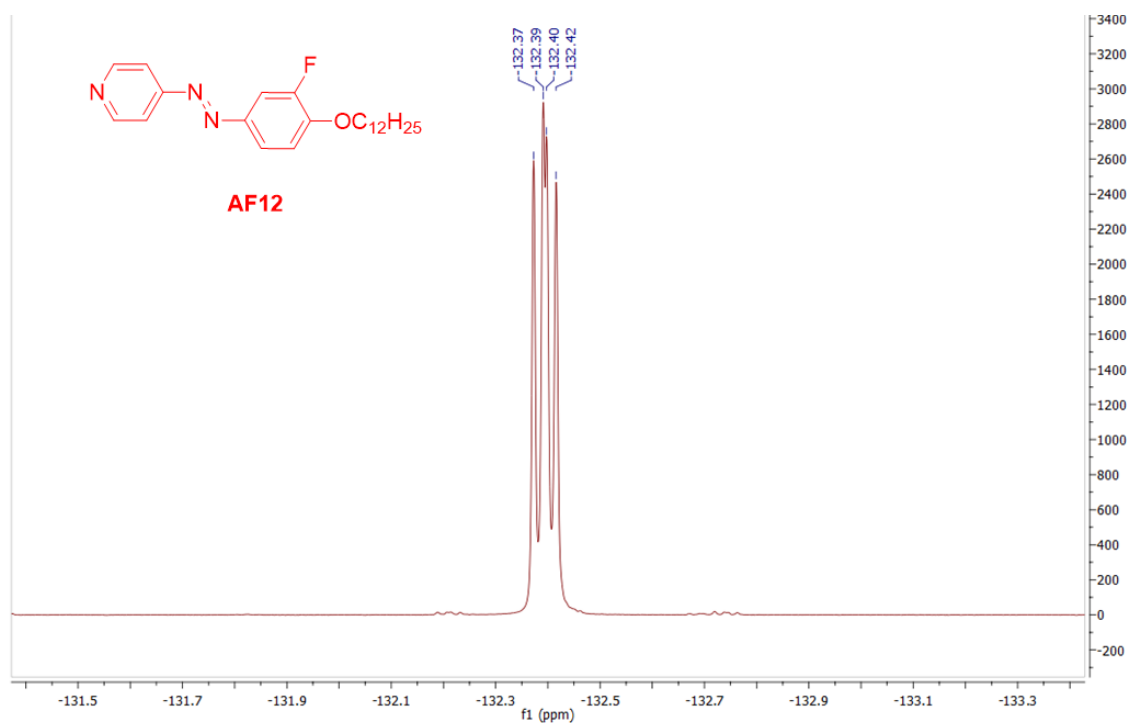


Figure S10c. ^{19}F NMR of compound AF12.

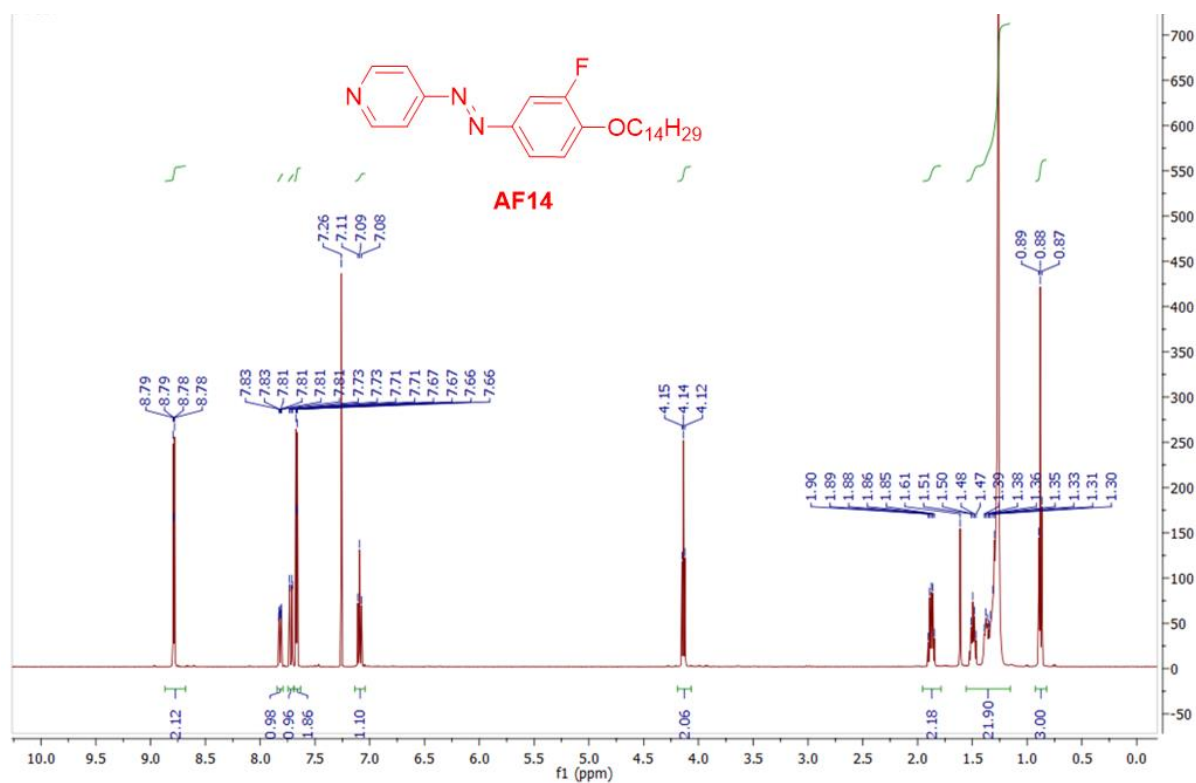


Figure S11a. ^1H NMR of compound AF14.

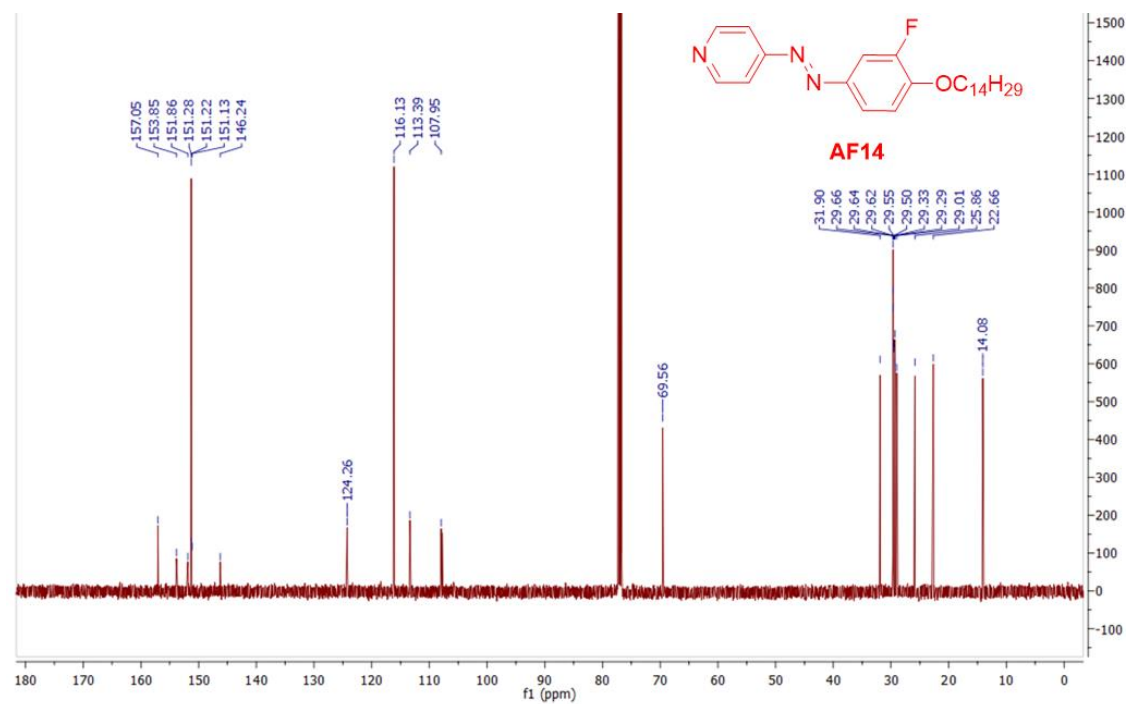


Figure S11b. ¹³C NMR of compound AF14.

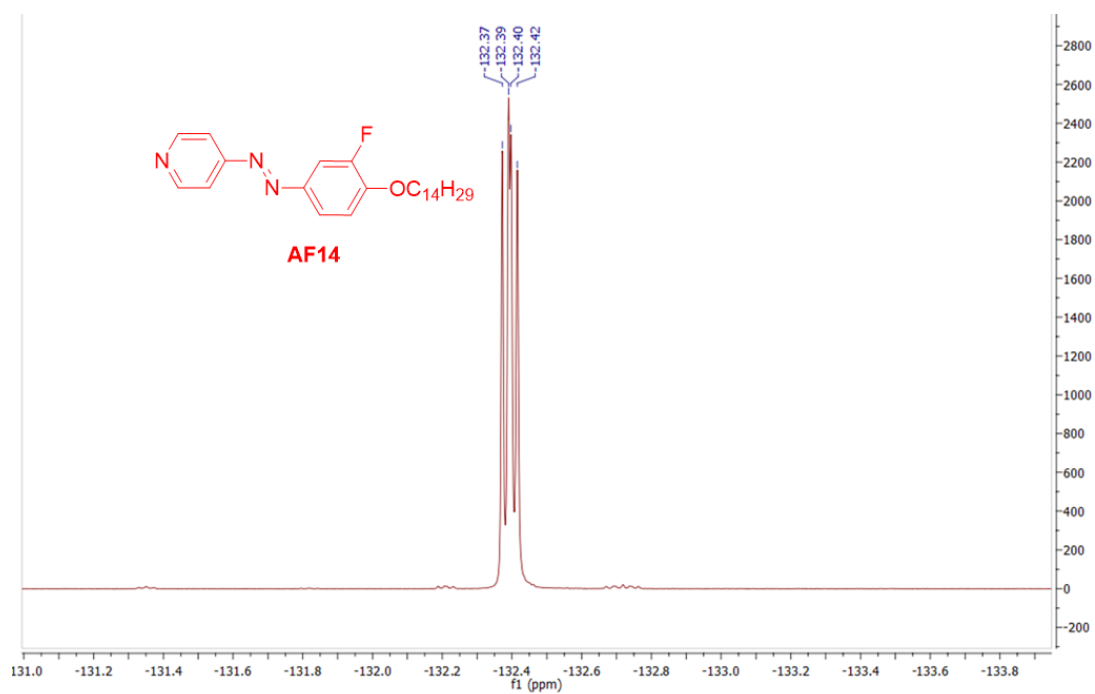


Figure S11c. ¹⁹F NMR of compound AF14.

3. IR Charts of hydrogen-bonded supramolecules

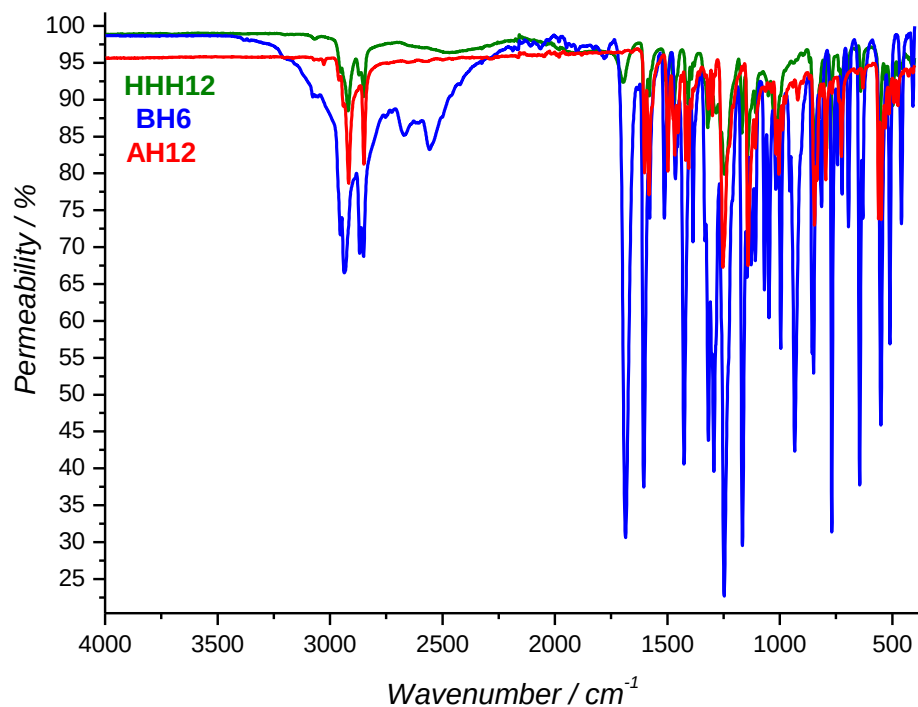


Figure S12a. FTIR spectra of the supramolecule **HHH12** (green) and its complementary components **B6H** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature.

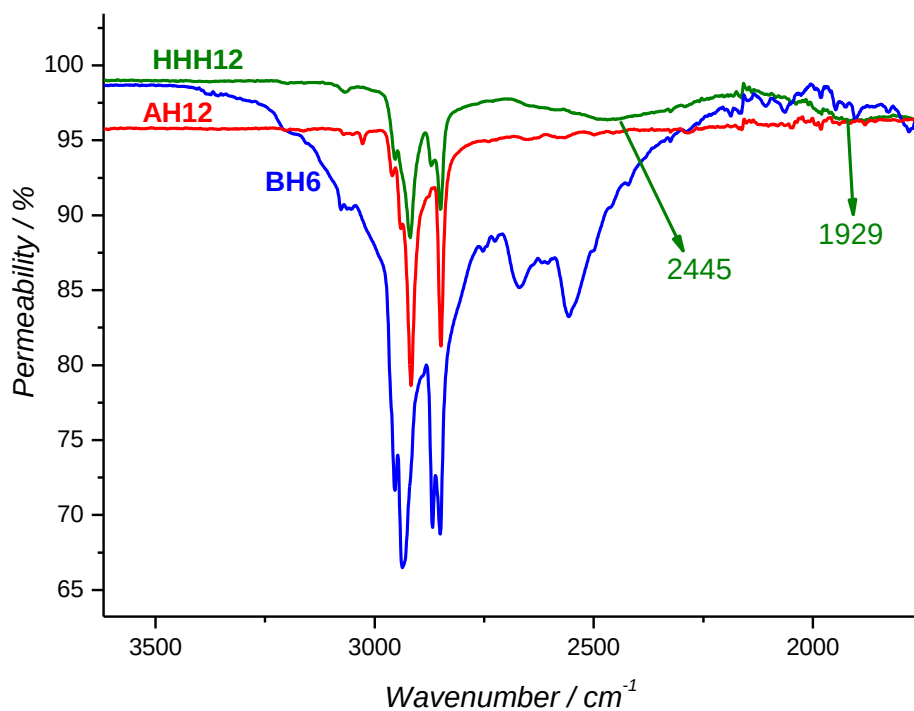


Figure S12b. FTIR spectra of the supramolecule **HHH12** (green) and its complementary components **B6H** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1750 cm⁻¹ and 3500 cm⁻¹

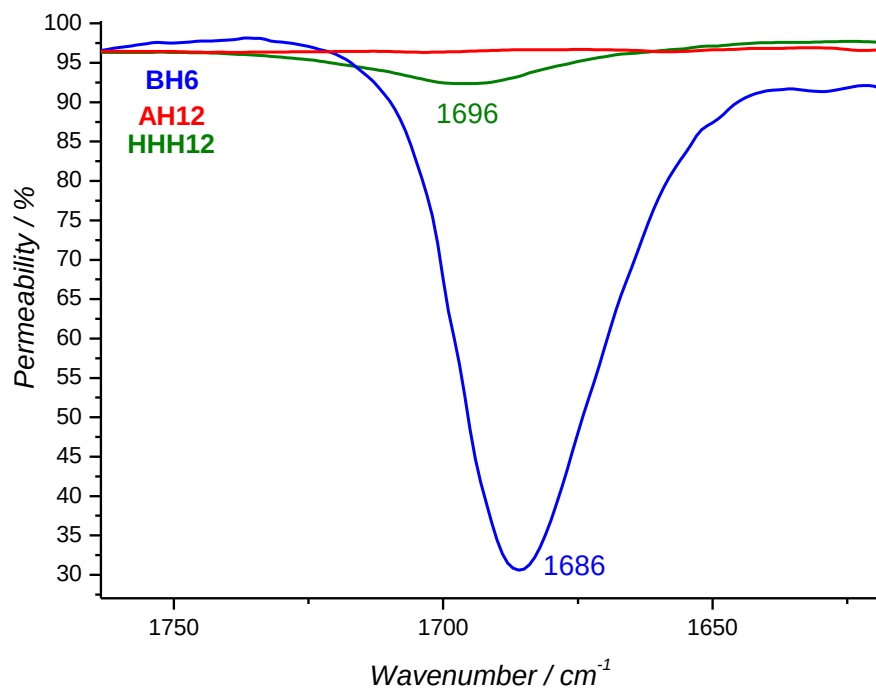


Figure S12c. FTIR spectra of the supramolecule **HHH12** (green) and its complementary components **B6H** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1620 cm^{-1} and 1750 cm^{-1}

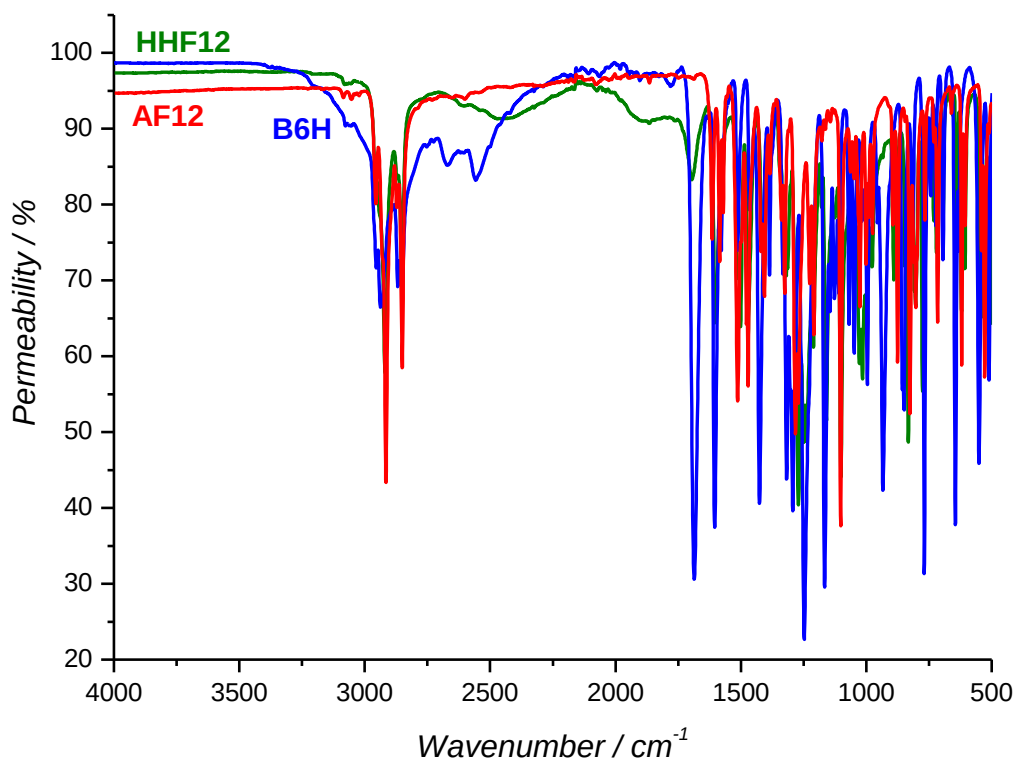


Figure S13a. FTIR spectra of the supramolecule **HHH12** (green) and its complementary components **B6H** (blue), **AF12** (red) in the crystalline state (KBr) at room temperature.

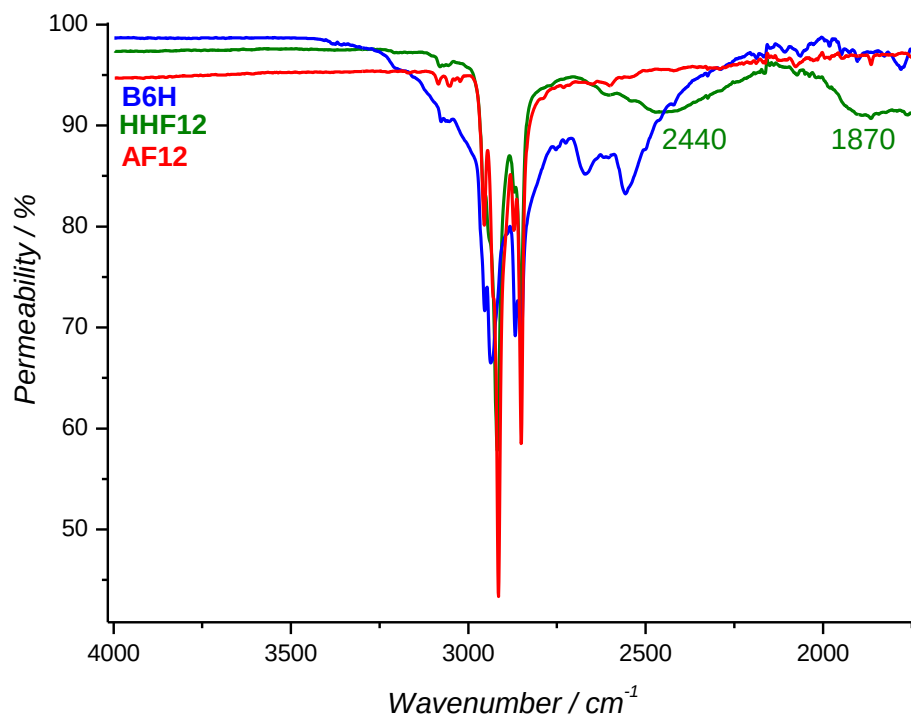


Figure S13b. FTIR spectra of the supramolecule **HHF12** (green) and its complementary components **B6H** (blue), **AF12** (red) in the crystalline state at room temperature enlarged area between 1750 cm^{-1} and 4000 cm^{-1}

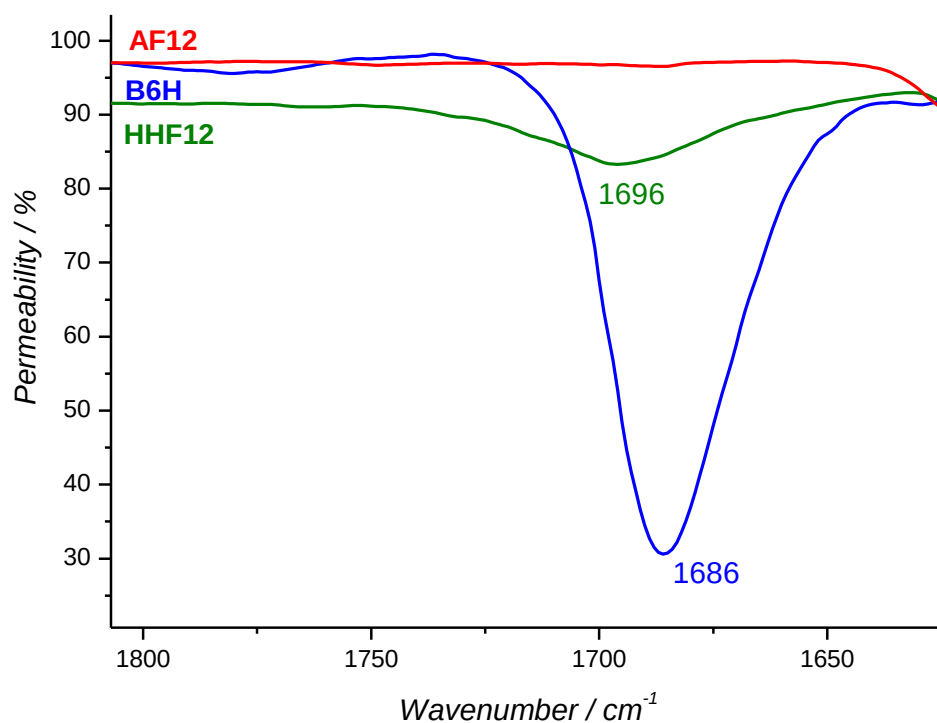


Figure S13c. FTIR spectra of the supramolecule **HHF12** (green) and its complementary components **B6H** (blue), **AF12** (red) in the crystalline state at room temperature enlarged area between 1600 cm^{-1} and 1800 cm^{-1}

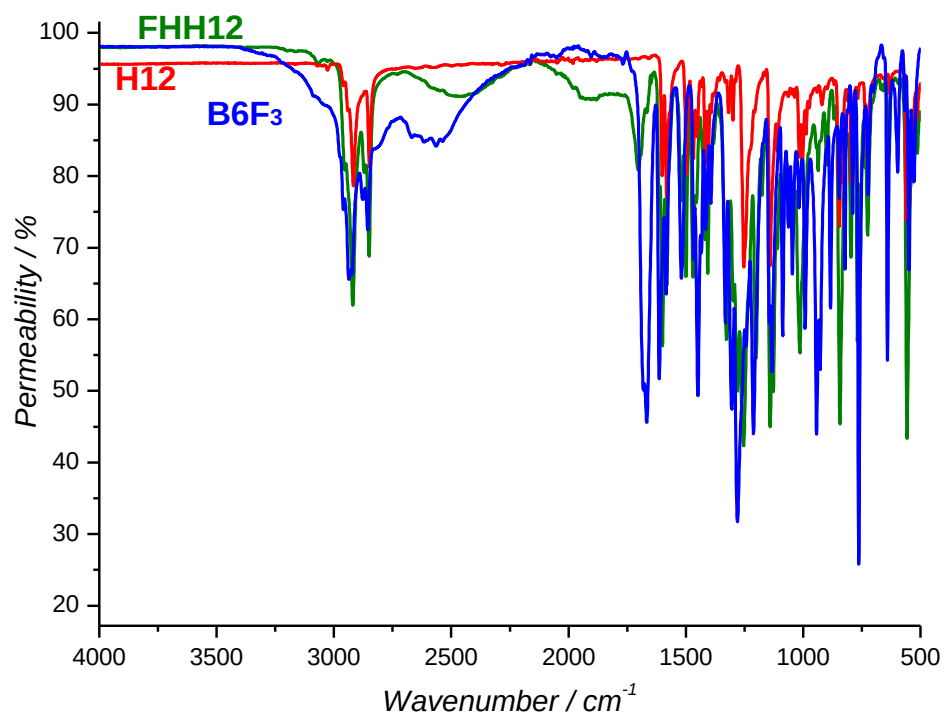


Figure S14a. FTIR spectra of the supramolecule **FHH12** (green) and its complementary components **B6F₃** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature.

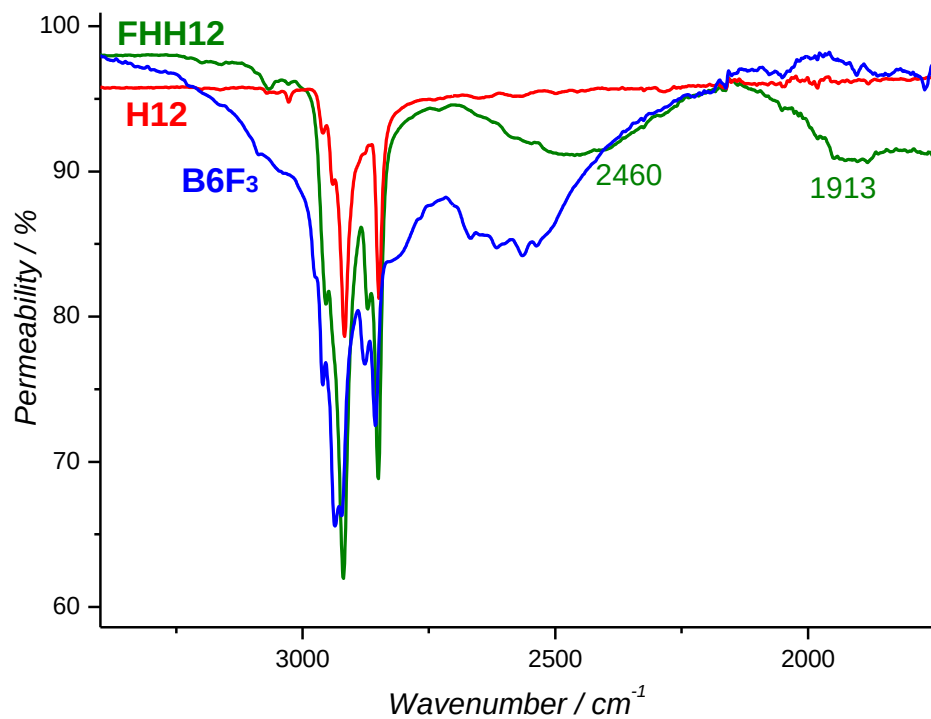


Figure S14b. FTIR spectra of the supramolecule **FHH12** (green) and its complementary components **B6F₃** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1750 cm^{-1} and 3500 cm^{-1}

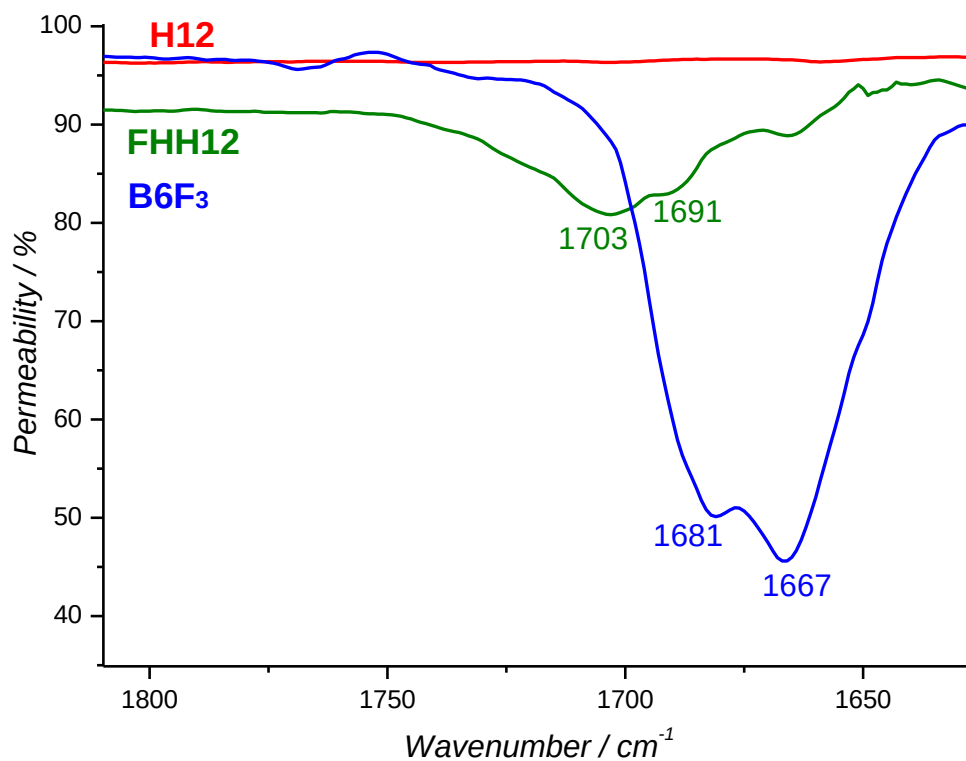


Figure S14c. FTIR spectra of the supramolecule **FHH12** (green) and its complementary components **B6F₃** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1625 cm⁻¹ and 1800 cm⁻¹

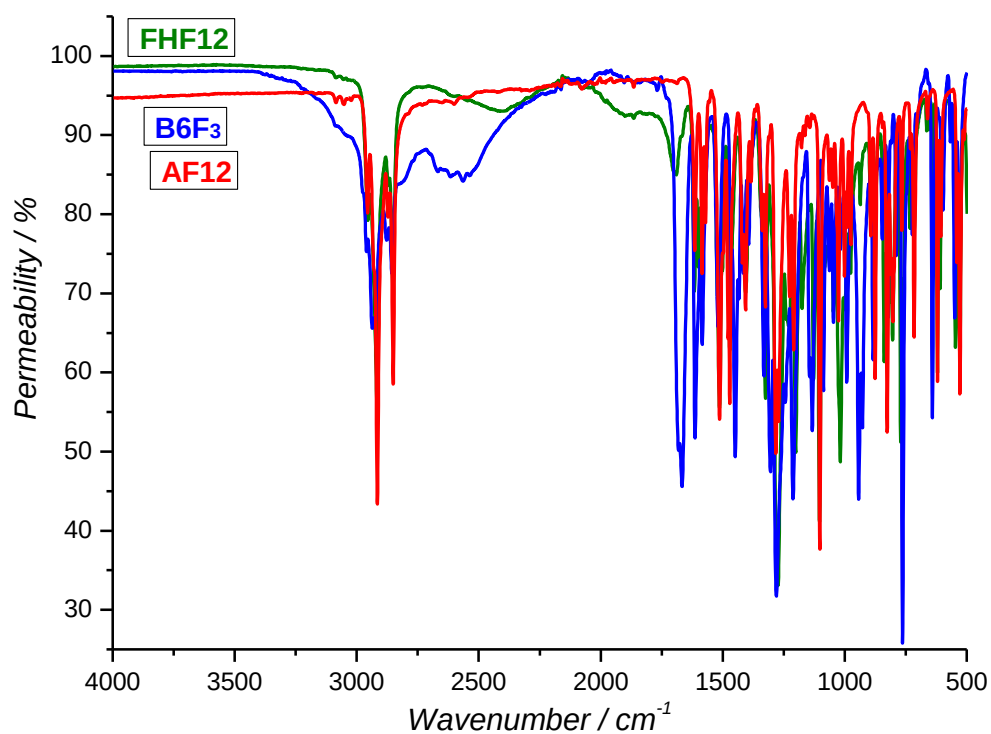


Figure S15a. FTIR spectra of the supramolecule **FHF12** (green) and its complementary components **B6F₃** (blue), **AF12** (red) in the crystalline state (KBr) at room temperature.

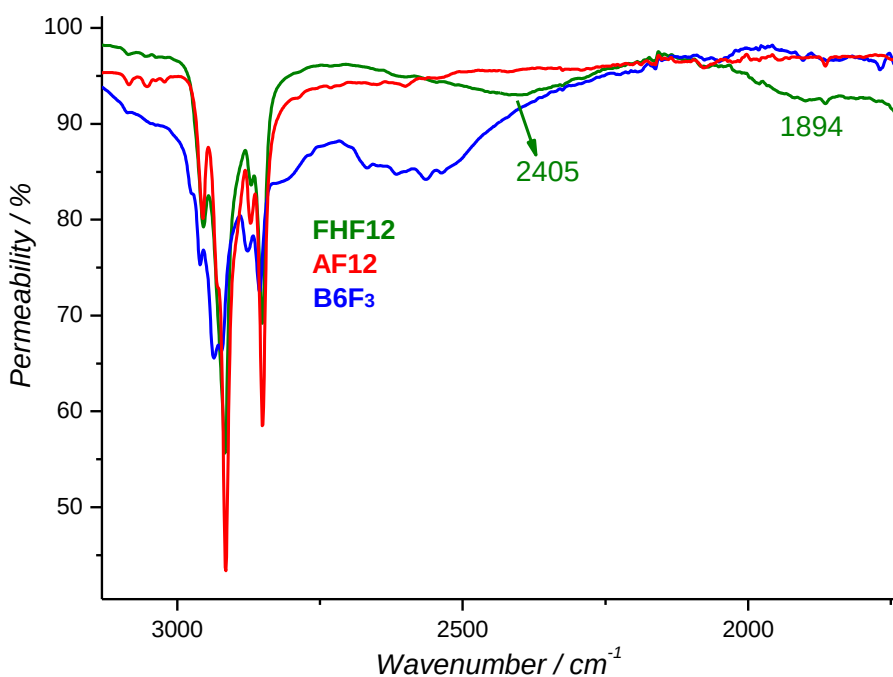


Figure S15b. FTIR spectra of the supramolecule FHF12 (green) and its complementary components B6F₃ (blue), AF12 (red) in the crystalline state at room temperature enlarged area between 1750 cm⁻¹ and 3000 cm⁻¹

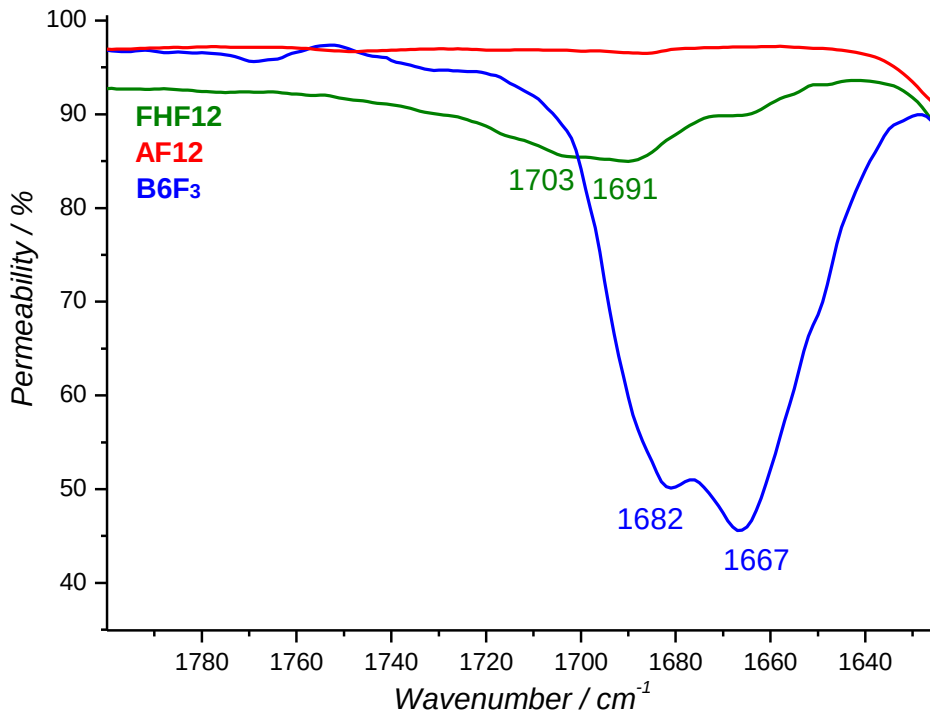


Figure S15c. FTIR spectra of the supramolecule FHF12 (green) and its complementary components B6F₃ (blue), AF12 (red) in the crystalline state at room temperature enlarged area between 1630 cm⁻¹ and 1790 cm⁻¹

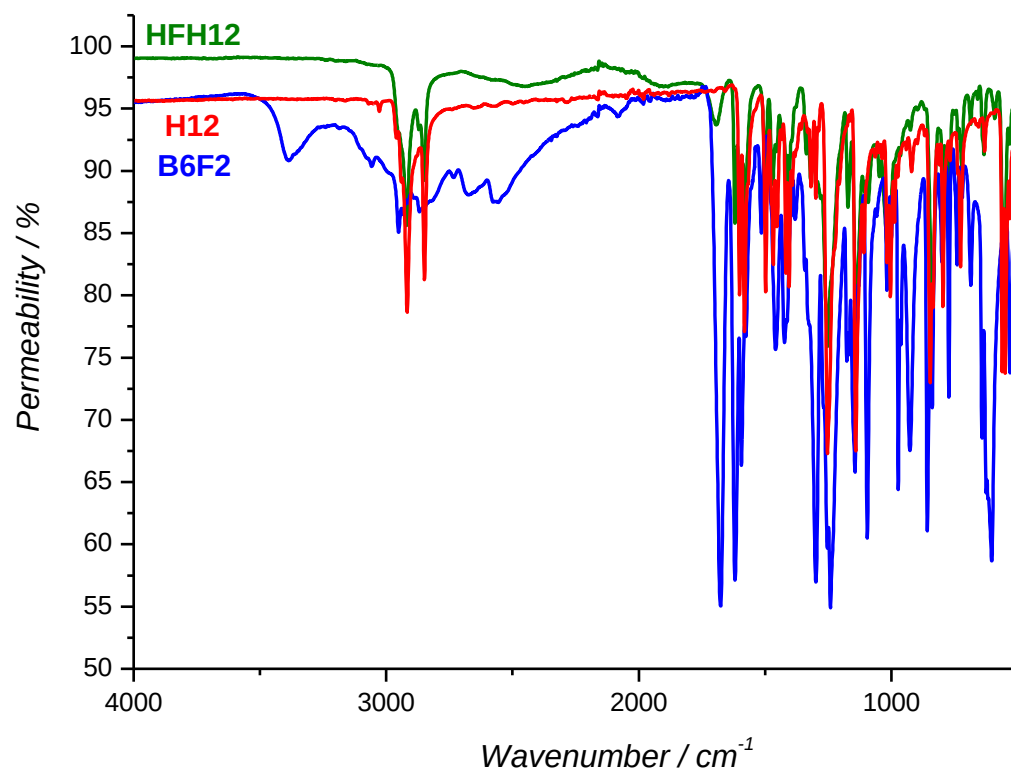


Figure S16a. FTIR spectra of the supramolecule **HFH12** (green) and its complementary components **B6F2** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature.

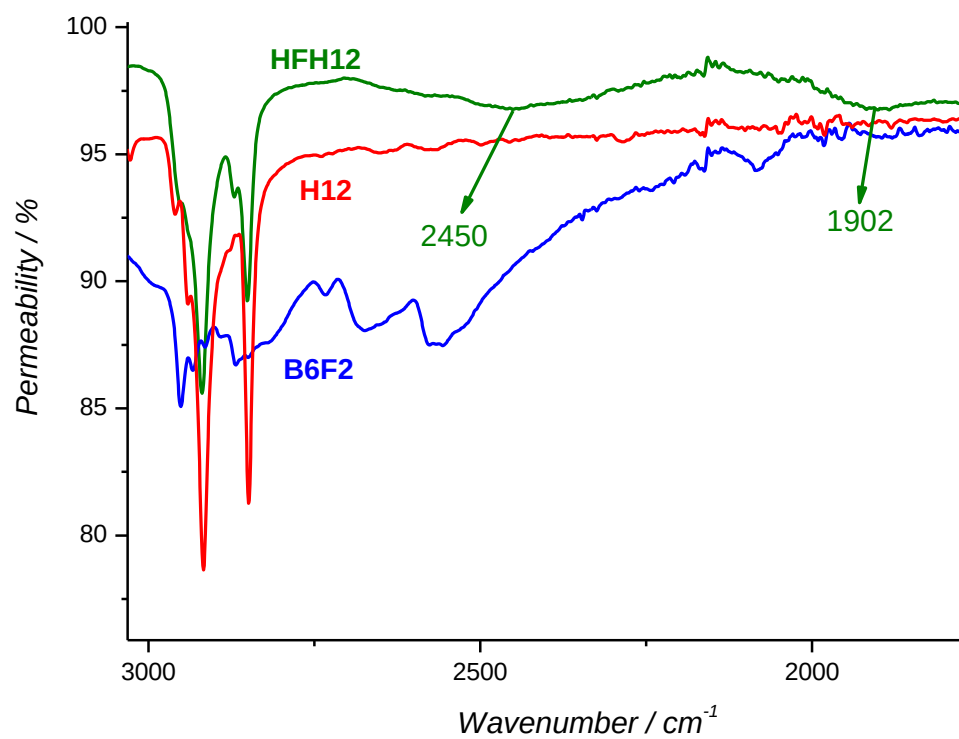


Figure S16b. FTIR spectra of the supramolecule **HFH12** (green) and its complementary components **B6F2** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1750 cm^{-1} and 3000 cm^{-1}

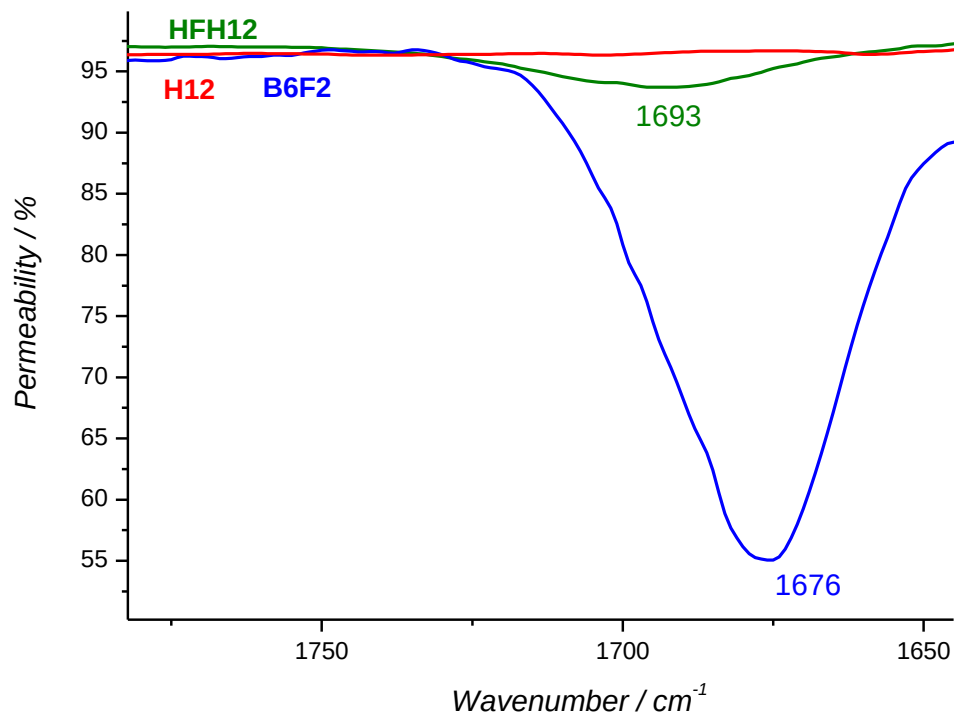


Figure S16c. FTIR spectra of the supramolecule **HFH12** (green) and its complementary components **B6F₂** (blue), **AH12** (red) in the crystalline state at room temperature enlarged area between 1650 cm⁻¹ and 1775 cm⁻¹

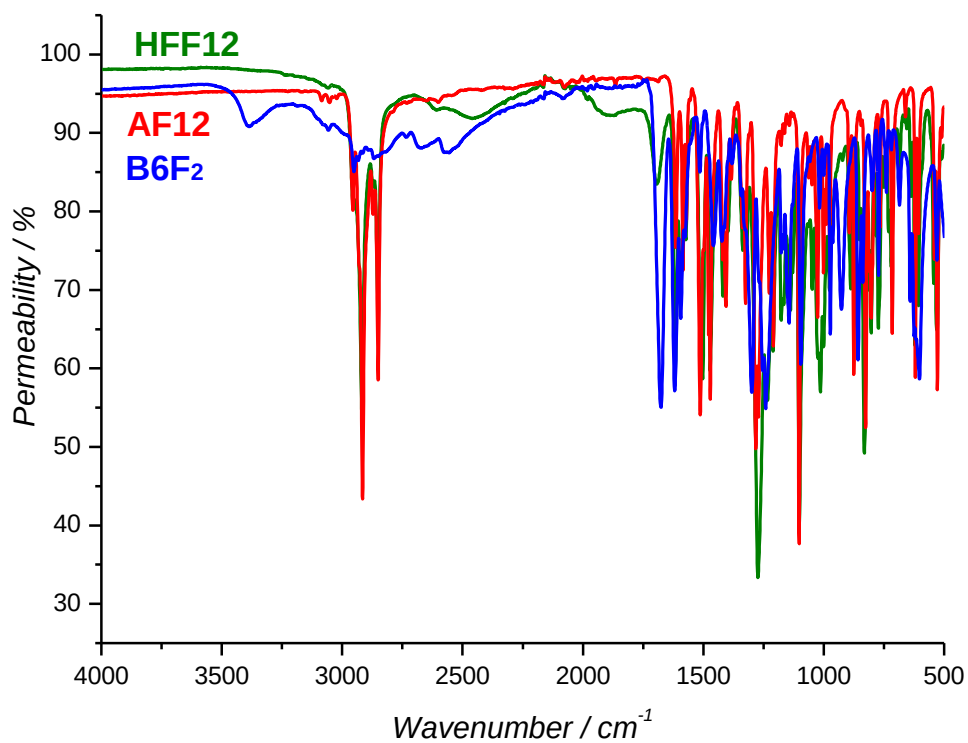


Figure S17a. FTIR spectra of the supramolecule **HFF12** (green) and its complementary components **B6F₂** (blue), **AF12** (red) in the crystalline state (KBr) at room temperature.

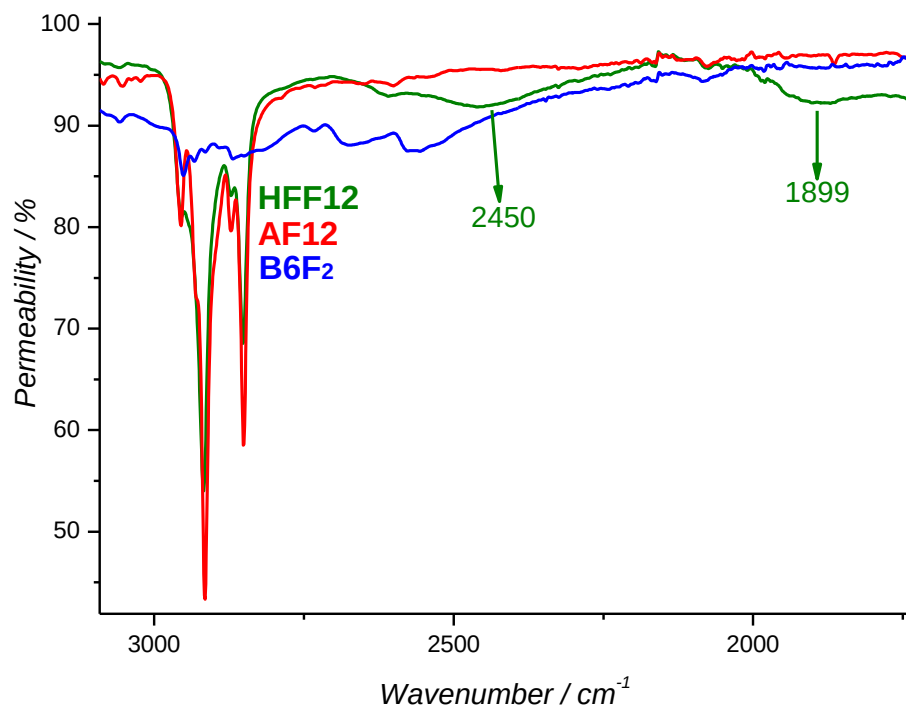


Figure S17b. FTIR spectra of the supramolecule HFF12 (green) and its complementary components **B6F₂** (blue), **AF12** (red) in the crystalline state at room temperature enlarged area between 1750 cm⁻¹ and 3000 cm⁻¹

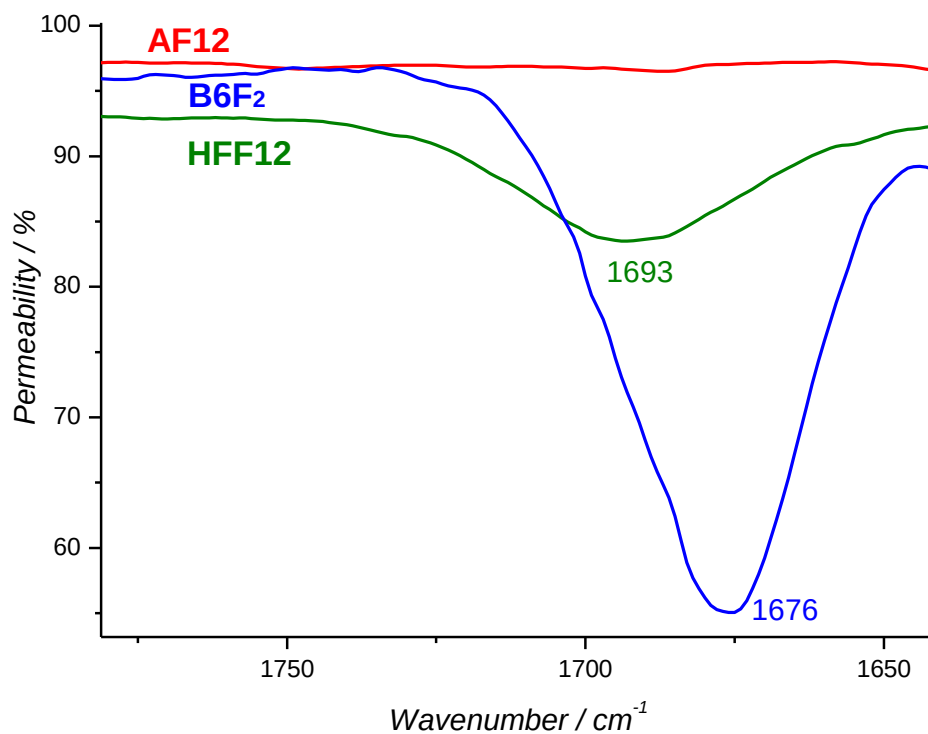


Figure S17c. FTIR spectra of the supramolecule HFF12 (green) and its complementary components **B6F₂** (blue), **AF12** (red) in the crystalline state at room temperature enlarged area between 1650 cm⁻¹ and 1775 cm⁻¹

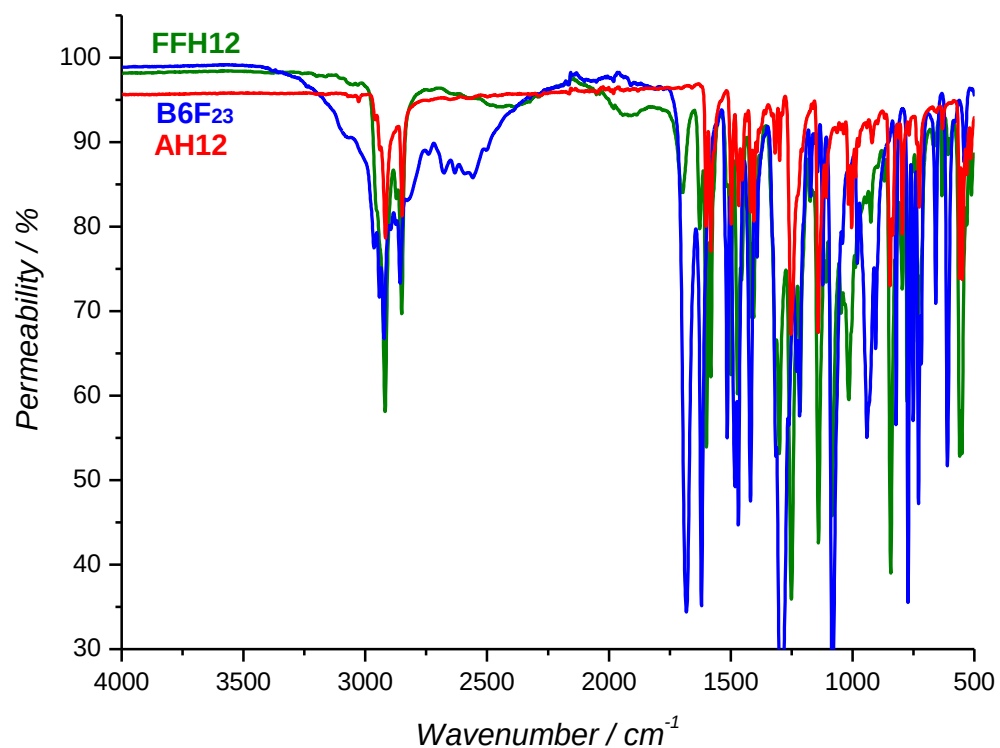


Figure S18a. FTIR spectra of the supramolecule **FFH12** (green) and its complementary components **B6F₂₃** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature.

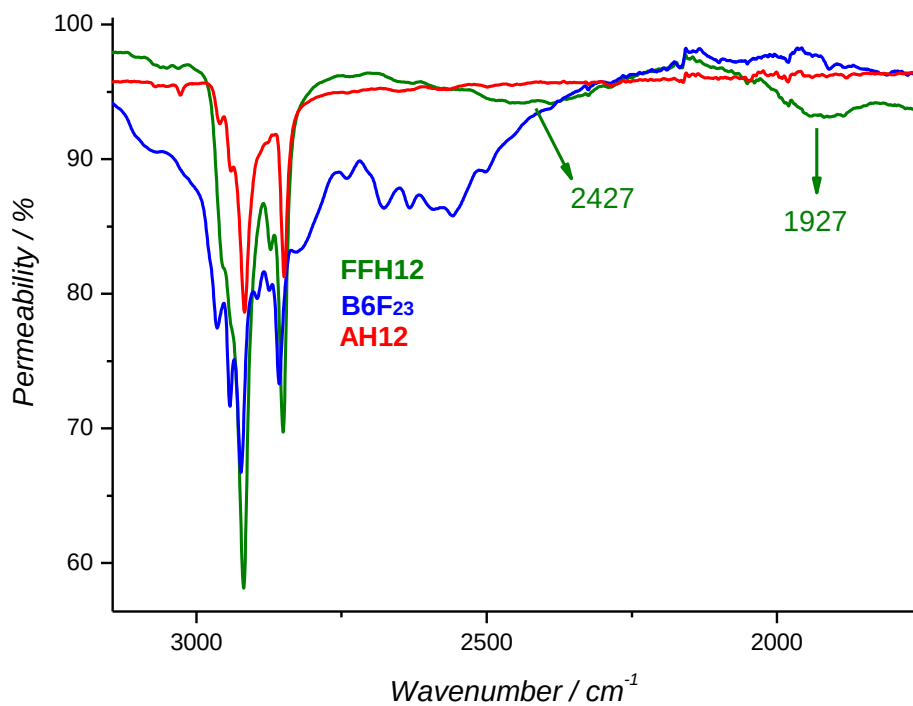


Figure S18b. FTIR spectra of the supramolecule **FFH12** (green) and its complementary components **B6F₂₃** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature enlarged area between 1750 cm^{-1} and 3000 cm^{-1} .

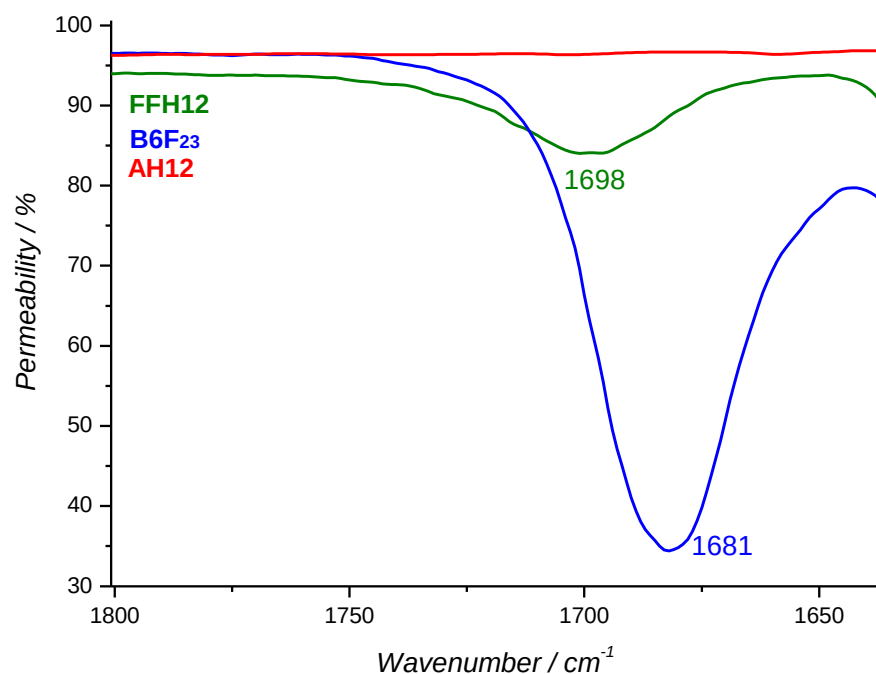


Figure S18c. FTIR spectra of the supramolecule **FFH12** (green) and its complementary components **B6F₂₃** (blue), **AH12** (red) in the crystalline state (KBr) at room temperature enlarged area between 1650 cm^{-1} and 1800 cm^{-1}

4. NMR Charts of hydrogen-bonded supramolecules

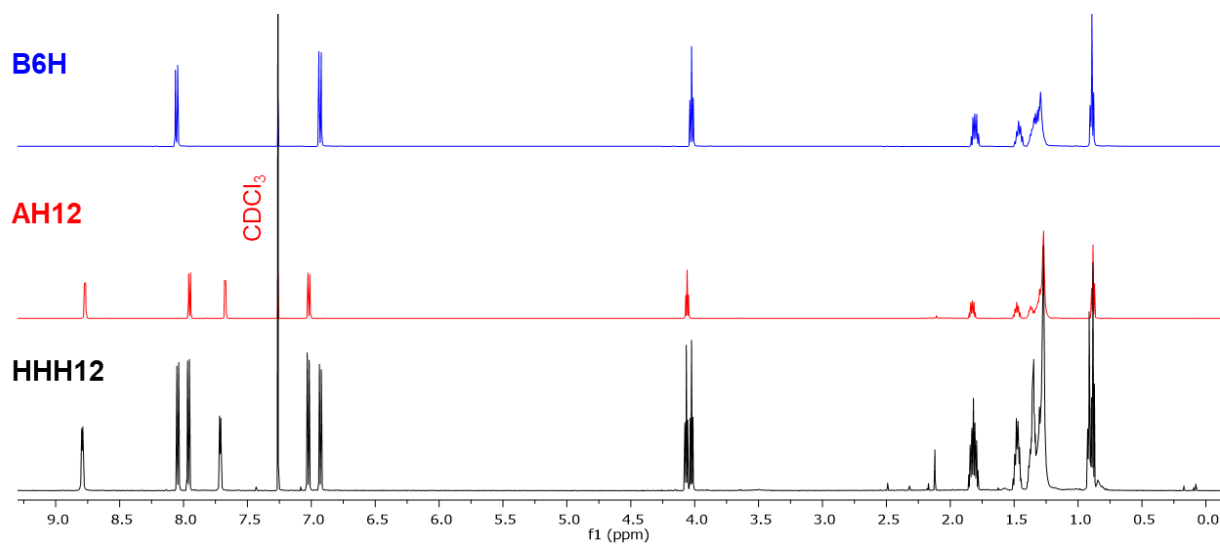


Figure S19. ^1H NMR spectra (500 MHz, CDCl_3) of the supramolecule **HHH12** (black) and its complementary components the azopyridine derivative **AH12** (red) and the benzoic acid derivative **B6H** (blue).

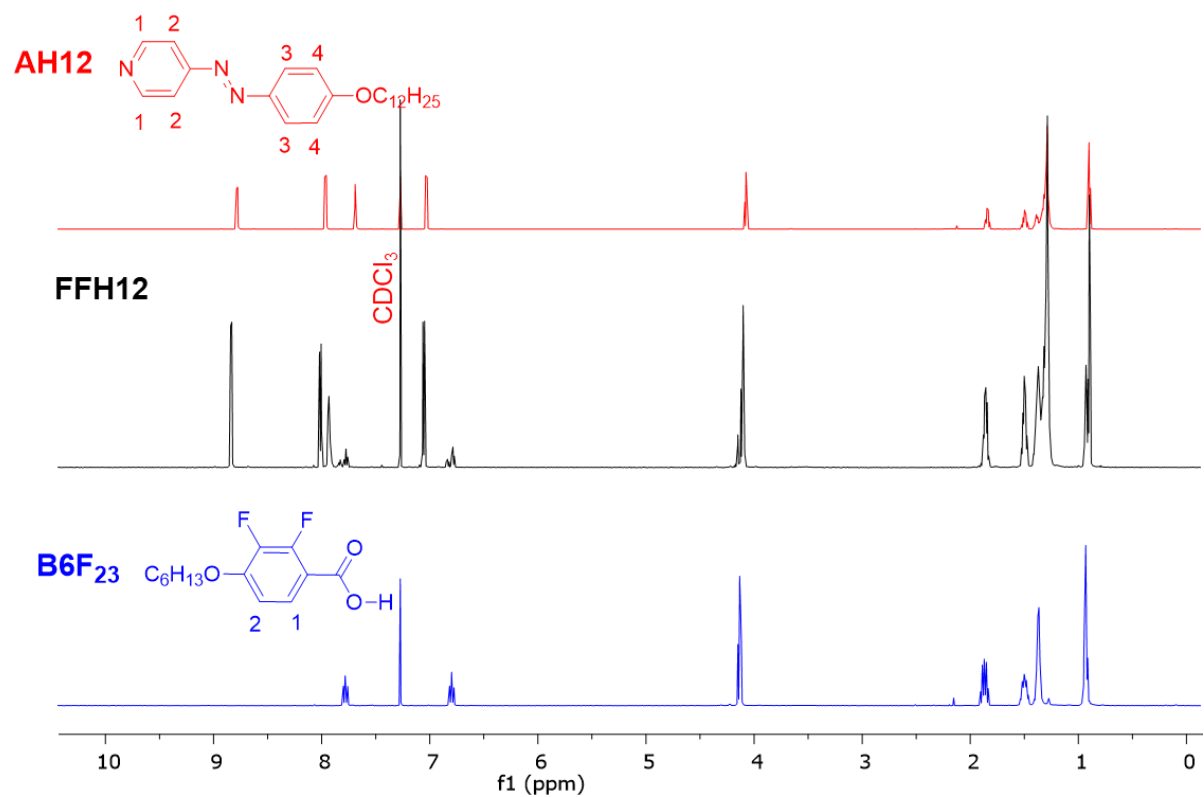


Figure S20. ¹H NMR spectra (500 MHz, CDCl₃) of the supramolecule **FFH12** (black) and its complementary components the azopyridine derivative **AH12** (red) and the benzoic acid derivative **B6F₂₃** (blue).

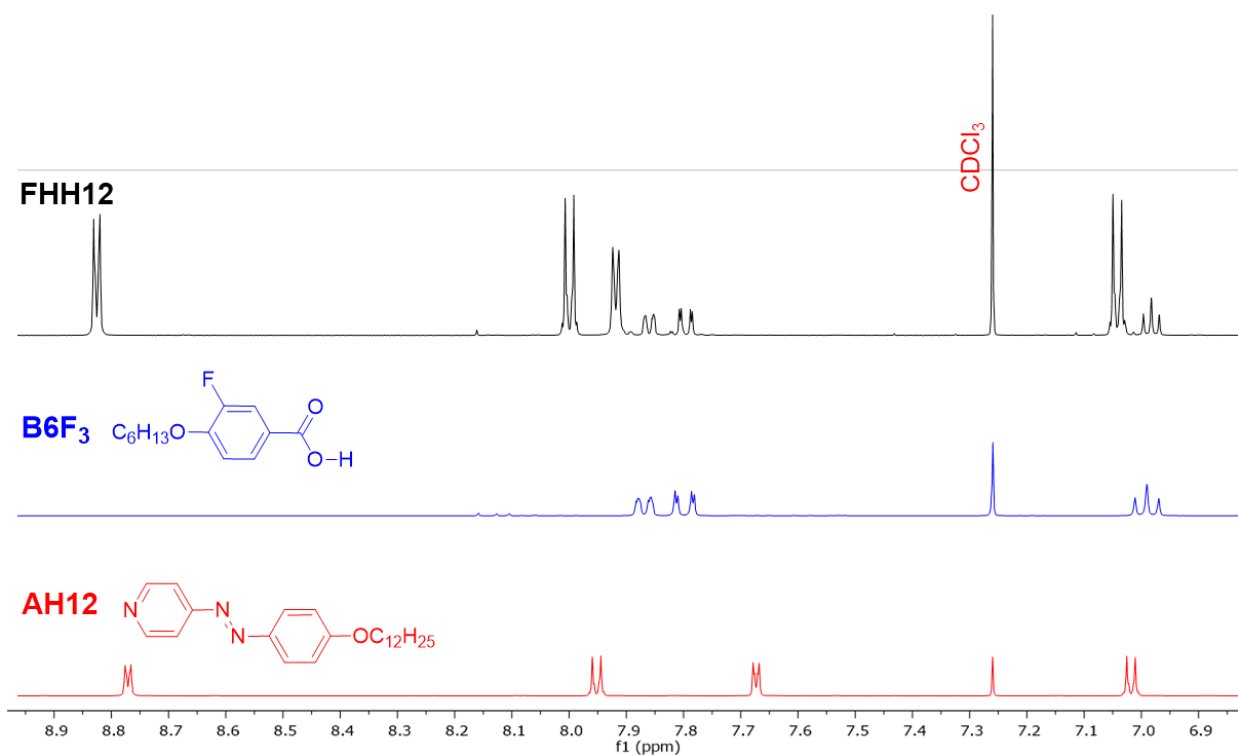


Figure S21a. ^1H NMR spectra (500 MHz, CDCl_3) in the aromatic region of the supramolecule **FHH12** (black) and its complementary components the azopyridine derivative **AH12** (red) and the benzoic acid derivative **B6F₃** (blue).

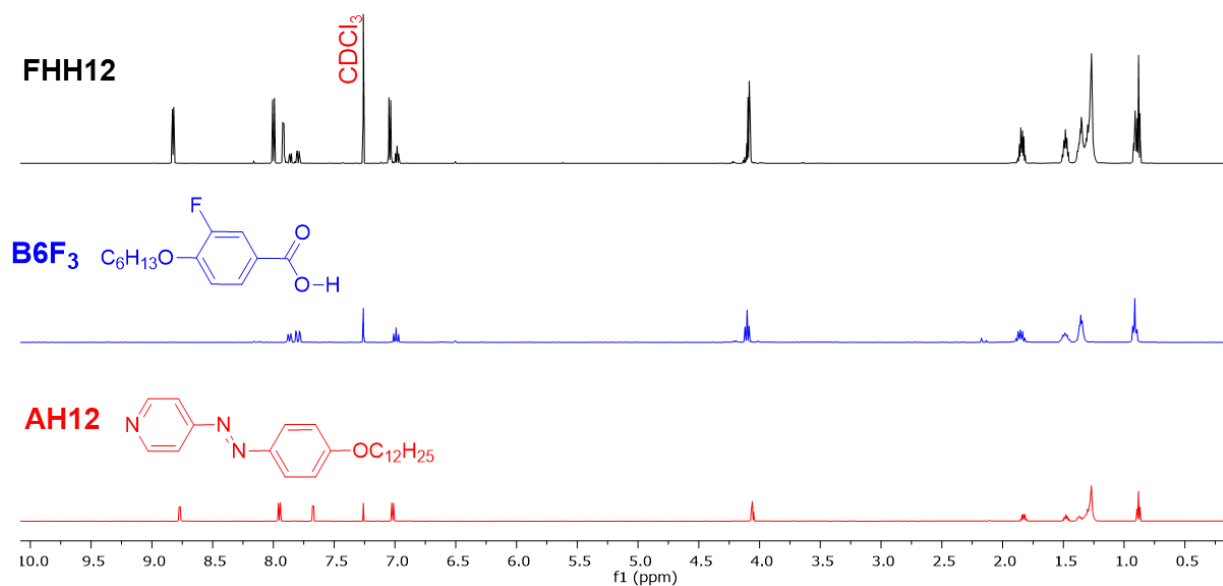
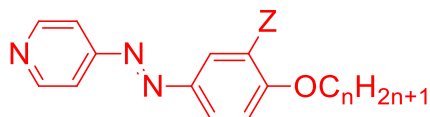


Figure S21b. ^1H NMR spectra (500 MHz, CDCl_3) of the supramolecule **FHH12** (black) and its complementary components the azopyridine derivative **AH12** (red) and the benzoic acid derivative **B6F₃** (blue).

5. Tables of transition temperatures

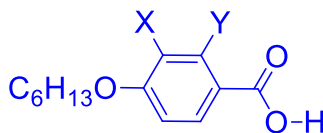
Table S1. Phase transition temperatures ($T/^{\circ}\text{C}$) of the azopyridine derivatives **AH n** and **AF n** .^{a,2}



Comp.	Z	Heating ($T/^{\circ}\text{C}$)
AH8	H	Cr 71 Iso
AH10	H	Cr 67 Iso
AH12	H	Cr 73 Iso
AH14	H	Cr 74 Iso
AF8	F	Cr 58 Iso
AF10	F	Cr 57 Iso
AF12	F	Cr 66 Iso
AF14	F	Cr 68 Iso

Notes: ^a Peak temperatures as determined from POM; abbreviations: Cr = crystalline solid; Iso = isotropic liquid.

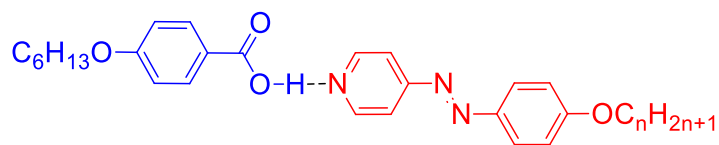
Table S2. Phase transition temperatures ($T/^{\circ}\text{C}$) of the benzoic acid derivatives **B6H**, **B6F₂**, **B6F₃** and **B6F₂₃**.^a



Comp.	X	Y	Heating ($T/^{\circ}\text{C}$)
B6H	H	H	Cr 104 N 134 Iso
B6F₂	H	F	Cr 110 N 120 Iso
B6F₃	F	H	Cr 108 Iso
B6F₂₃	F	F	Cr 108 Iso

Notes: ^a Peak temperatures as determined from POM; abbreviations: Cr = crystalline solid; N= nematic phase; Iso = isotropic liquid.

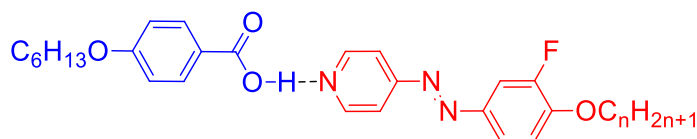
Table S3. DSC data scanned at the rate of 10 °C/min for different series of **HHHn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
HHH8	H: Cr 106 [56.2] SmA 124 [4.1] N 134 [7.0] Iso C: Iso 131 [-6.8] N 121 [-4.4] SmA 97 [-54.2] Cr
HHH10	H: Cr1 82 [18.9] Cr2 95 [48.0] SmA 128 [20.7] N 131 [20.7] ^c Iso C: Iso 128 [-19.5] N 125 [-19.5] ^c SmA 84 [-44.5] Cr
HHH12	H: Cr1 87 [30.3] Cr2 94 [38.5] SmC 110 [< 0.1] ^b SmA 131 [23.4] Iso C: Iso 127 [-22.3] SmA 108 [< 0.1] ^b SmC 84 [-41.0] Cr
HHH14	H: Cr 90 [40.8] SmC 116 [< 0.1] ^b SmA 131 [12.8] Iso C: Iso 129 [-12.5] SmA 114 [< 0.1] ^b SmC 83 [-17.2] Cr

N = nematic phase, SmA = Smectic A, SmC = Smectic C. ^bEnthalpy value is undetectable by DSC, ^ccannot be separated.

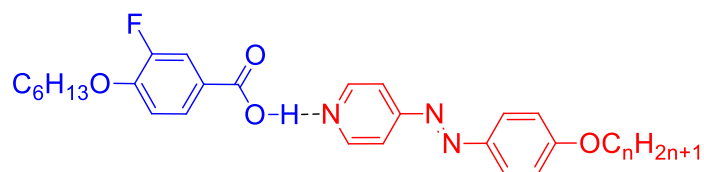
Table S4. DSC data scanned at the rate of 10 °C/min for different series of **HHFn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
HHF8	H: Cr 101 [57.9] N 118 [5.1] Iso C: Iso 115 [-5.6] N 84 [-53.3] Cr
HHF10	H: Cr 87 [55.5] SmC 95[1.1] N 113 [3.7] Iso C: Iso 110 [-4.8] N 92 [-1.5] SmC 70 [-55.5] Cr
HHF12	H: Cr 96 [49.3] SmC 112 [< 0.1] ^c N 114 [9.9] Iso C: Iso 110 [-11.7] N 102 [< 0.1] ^b SmC 73 [-50.9] Cr
HHF14	H: Cr 95 [56.3] SmC 111 [14.4] Iso C: Iso 108 [-13.8] SmC 74 [-64.1] Cr

For abbreviations see Table S1. ^bEnthalpy value is undetectable by DSC, ^ccannot be separated.

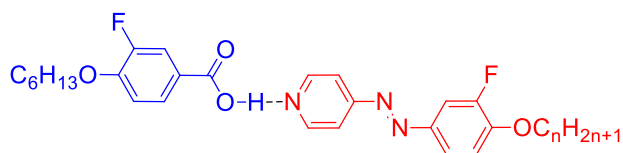
Table S5. DSC data scanned at the rate of 10 °C/min for different series of **FHHn** HBLCS^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
FHH8	H: Cr 89 [33.7] SmA 126 [15.6] Iso C: Iso 120 [-13.8] SmA 90 [< 0.01] ^b SmC 79 [-32.5] Cr
FHH10	H: Cr 85 [29.3] SmC 100 [< 0.01] ^b SmA 125 [15.8] Iso C: Iso 119 [-13.5] SmA 98 [< 0.01] ^b SmC 75 [-27.7] Cr
FHH12	H: Cr 80 [28.8] SmC 112 [< 0.1] ^b SmA 124 [19.7] Iso C: Iso 121 [-19.1] SmA 110 [< 0.1] ^b SmC 76 [-27.6] Cr
FHH14	H: Cr 62 [24.4] SmC 114 [< 0.01] ^b SmA 126 [5.9] Iso C: Iso 121 [-4.4] SmA 112 [< 0.01] ^b SmC 49 [-22.7] Cr

For abbreviations see Table S1. ^bEnthalpy value is undetectable by DSC.

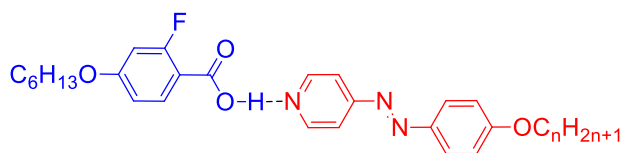
Table S6. DSC data scanned at the rate of 10 °C/min for different series of **FHF_n** HBLCS^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
FHF8	H: Cr 82 [53.3] SmC 90 [< 0.1] ^b SmA 100 [6.5] ^c N 105 [6.5] ^c Iso C: Iso 99 [-8.6] ^c N 88 [-8.6] ^c SmA 84 [< 0.1] SmC 67 [-52.8] Cr
FHF10	H: Cr 89 [72.7] SmC 98 [< 0.01] ^b SmA 107 [6.9] Iso C: Iso 100 [-9.6] ^c SmA 93 [-9.6] ^c SmC 77 [-71.6] Cr
FHF12	H: Cr 87 [54.8] SmC 90 [9.6] ^c SmA 110 [9.6] Iso C: Iso 102 [-10.6] ^c SmA 100 [-10.6] ^c SmC 72 [-53.9] Cr
FHF14	H: Cr 94 [66.3] SmC 100 [11.5] ^c SmA 111 [11.5] ^c Iso C: Iso 105 [-10.1] SmA 98 [-9.6] ^c SmC 79 [-67.5] Cr

For abbreviations see Table S1. ^bEnthalpy value is undetectable by DSC, ^ccannot be separated.

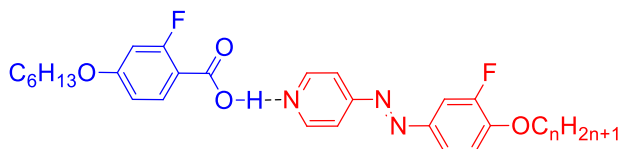
Table S7. DSC data scanned at the rate of 10 °C/min for different series of **HFHn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
HFH8	H: Cr 96 [62.7] N122 [9.9] Iso C: Iso 118 [-5.5] N 52 [-53.5] Cr
HFH10	H: Cr 85 [50.3] X 105 [1.9] N 119 [6.7] Iso C: Iso 115 [-6.9] N 102 [-2.7] X 67 [-26.7] Cr
HFH12	H: Cr 82 [64.7] X 112[18.2] ^c N 118 [18.2] ^c Iso C: Iso 116 [-17.9] ^c N 110 [-17.9] ^c X 74 [-29.5] Cr
HFH14	H: Cr 83 [62.3] X 114 [18.2] ^c N 118 [18.9] ^c Iso C: Iso 115 [-17.8] ^c N 112 [-17.8] ^c X 68 [-50.8] Cr

X = Unknown LC phase; for other abbreviations see Table S1. ^cEnthalpy value cannot be separated.

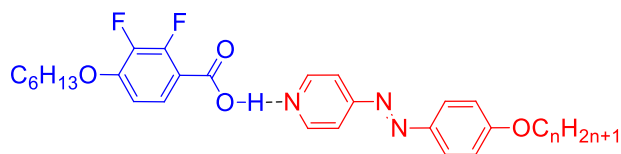
Table S8. DSC data scanned at the rate of 10 °C/min for different series of **HFFn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
HFF8	H: Cr 87 [38.8] N 105 [3.1] Iso C: Iso 103 [-2.8] N 70 [-19.6] Cr
HFF10	H: Cr 85 [39.8] N 100 [4.7] Iso C: Iso 99 [-4.5] N 52 [-49.2] Cr
HFF12	H: Cr 89 [74.0] N 104 [4.7] Iso C: Iso 101 [-5.8] N 72 [-0.4] Nx 51 [-48.2] Cr
HFF14	H: Cr 94 [66.3] SmC 100 [11.5] ^c SmA 111 [11.5] ^c Iso C: Iso 105 [-10.1] SmA 98 [-9.6] ^c SmC 79 [-67.5] Cr

Nx = Unknown nematic phase; for abbreviations see Table S1. ^bEnthalpy value cannot be separated.

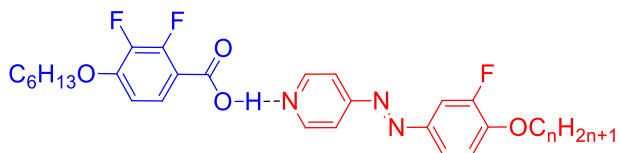
Table S9. DSC data scanned at the rate of 10 °C/min for different series of **FFHn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
FFH8	H: Cr 106 [47.2] SmC 115 [0.6] SmA 123 [13.7] ^c N 130 [13.7] ^c Iso C: Iso 128 [-14.5] ^c N 122 [-14.5] ^c SmA 113 [-0.7] SmC 100 [-46.9] Cr
FFH10	H: Cr 104 [45.4] SmC 122 [18.5] ^c SmA 128 [18.5] Iso C: Iso 125 [-18.1] SmA 120 [-18.1] ^c SmC 96 [-44.8] Cr
FFH12	H: Cr 98 [44.4] ^b SmC 129 [21.2] Iso C: Iso 126 [-22.1] SmC 87 [-43.0] Cr
FFH14	H: Cr 97 [46.7] SmC 130 [22.1] Iso C: Iso 126 [-21.3] SmC 83 [-43.5] Cr

For abbreviations see Table S1. ^bEnthalpy value is undetectable by DSC, ^ccannot be separated.

Table S10. DSC data scanned at the rate of 10 °C/min for different series of **FFFn** HBLCs^a.



Complex	Phase sequence (T/°C [ΔH (J/g)]) ^a
FFF8	H: Cr 93 [55.4] SmC 104 [16.2] ^c N 114 [16.2] ^c Iso C: Iso 111 [-14.7] ^c N 100 [-14.7] ^c SmC 83 [-51.5] Cr
FFF10	H: Cr 89 [47.8] SmC 107 [12.3] ^c N 114 [12.3] ^c Iso C: Iso 112 [-10.9] ^c N 104 [-10.9] ^c SmC 80 [-38.7] Cr
FFF12	H: Cr 91 [60.8] SmC 116 [18.4] Iso C: Iso 111 [-16.6] SmC 79 [-55.9] Cr
FFF14	H: Cr 95 [56.2] SmC 119 [16.9] Iso C: Iso 113 [-17.6] SmC 81 [-48.0] Cr

For abbreviations see Table S1. ^bEnthalpy value cannot be separated.

6. Additional DSC Plots

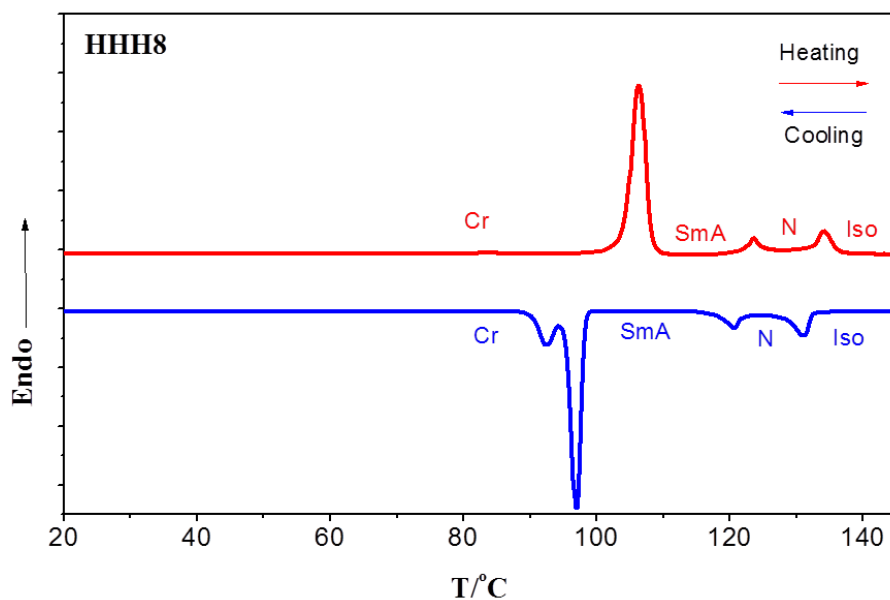


Figure S22. DSC Heating and cooling cycles of the HBLC **HHH8** at a scanning rate of 10 °C/min.

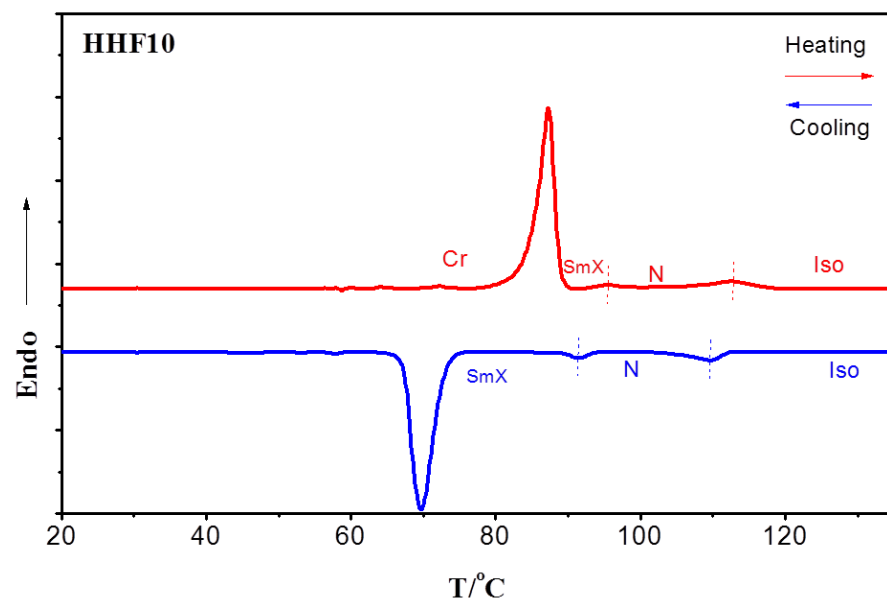


Figure S23. DSC Heating and cooling cycles of the HBLC **HHF10** at a scanning rate of 10 °C/min.

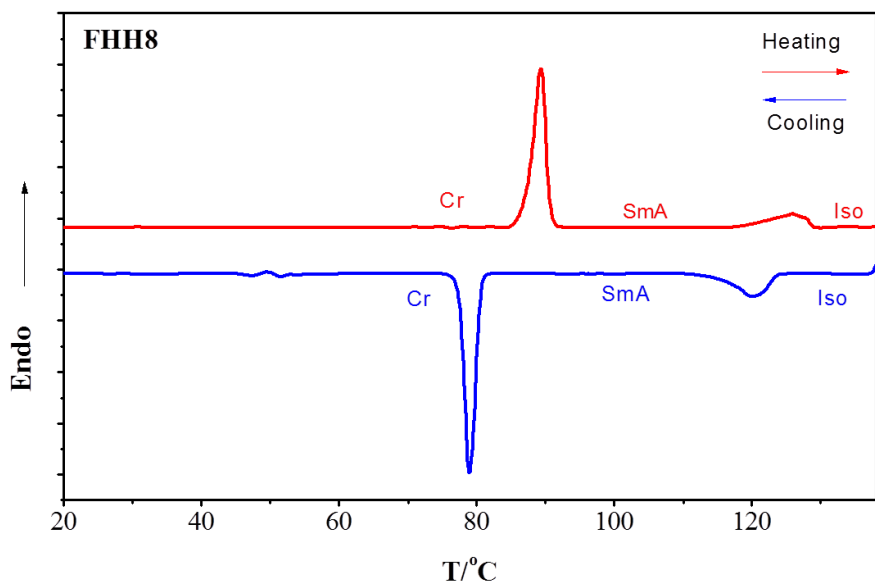


Figure S24. DSC Heating and cooling cycles of the HBLc **FHH8** at a scanning rate of 10 °C/min.

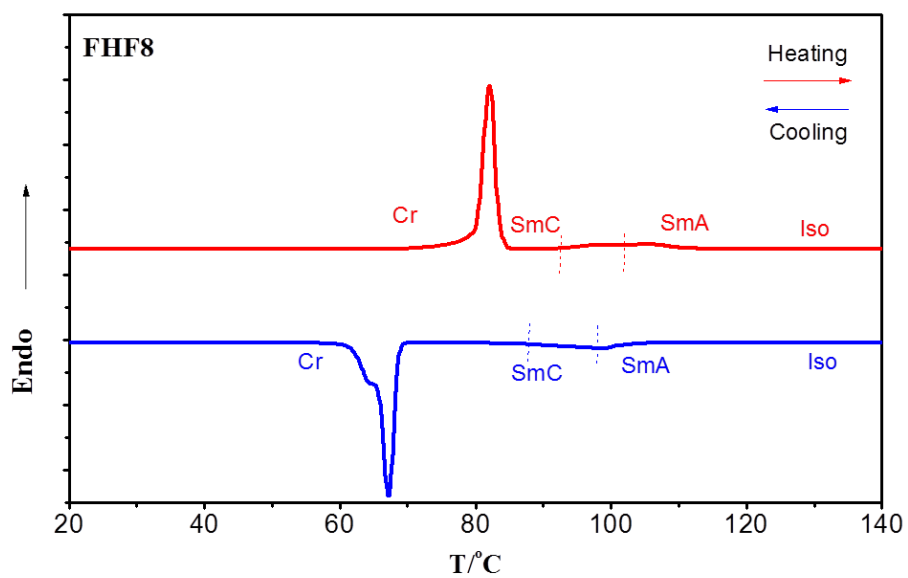


Figure S25. DSC Heating and cooling cycles of the HBLc **FHF8** at a scanning rate of 10 °C/min.

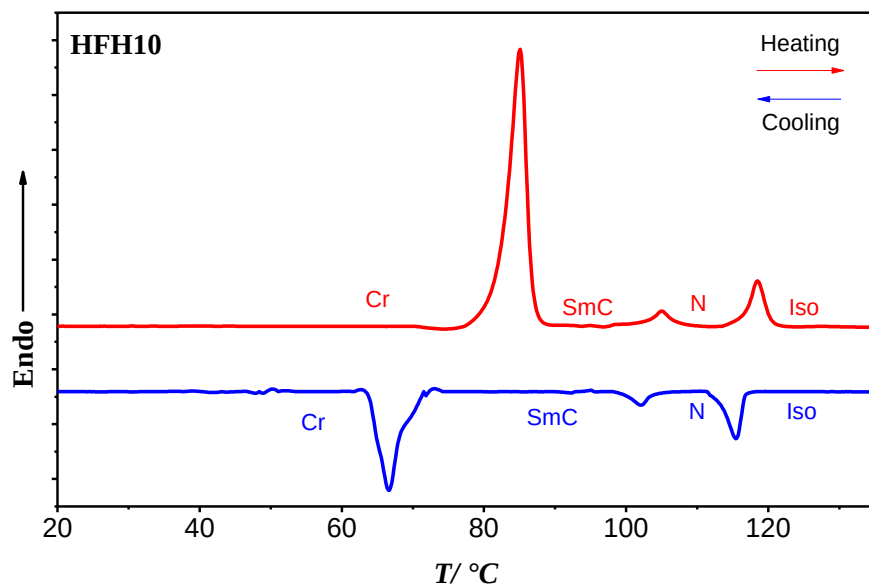


Figure S26. DSC Heating and cooling cycles of the HBLC **HFF10** at a scanning rate of 10 °C/min.

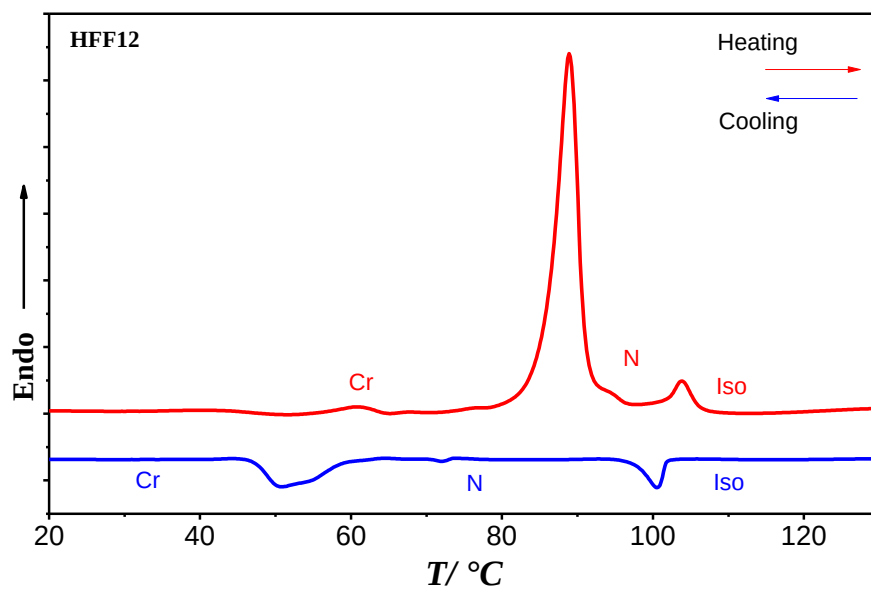


Figure S27. DSC Heating and cooling cycles of the HBLC **HFF12** at a scanning rate of 10 °C/min.

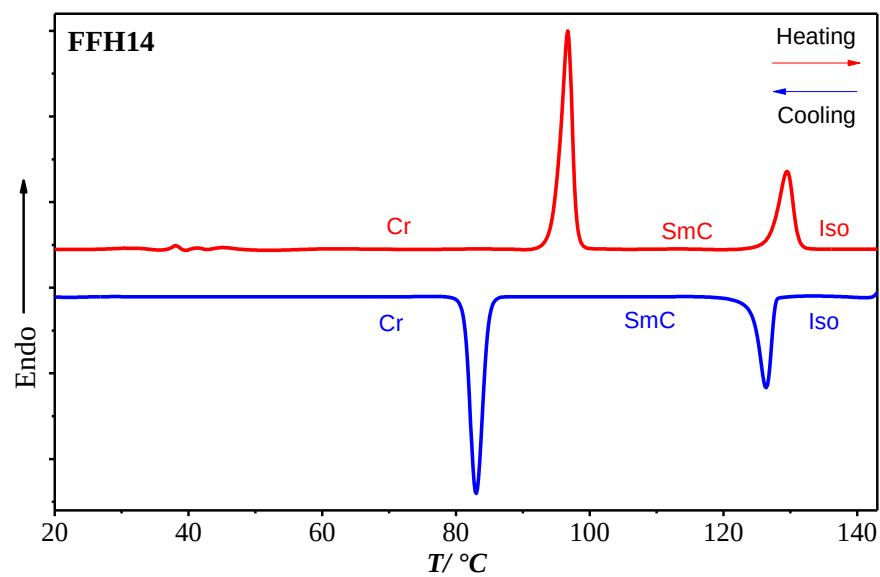


Figure S28. DSC Heating and cooling cycles of the HBLc **FFH14** at a scanning rate of 10 $^\circ\text{C}/\text{min}$.

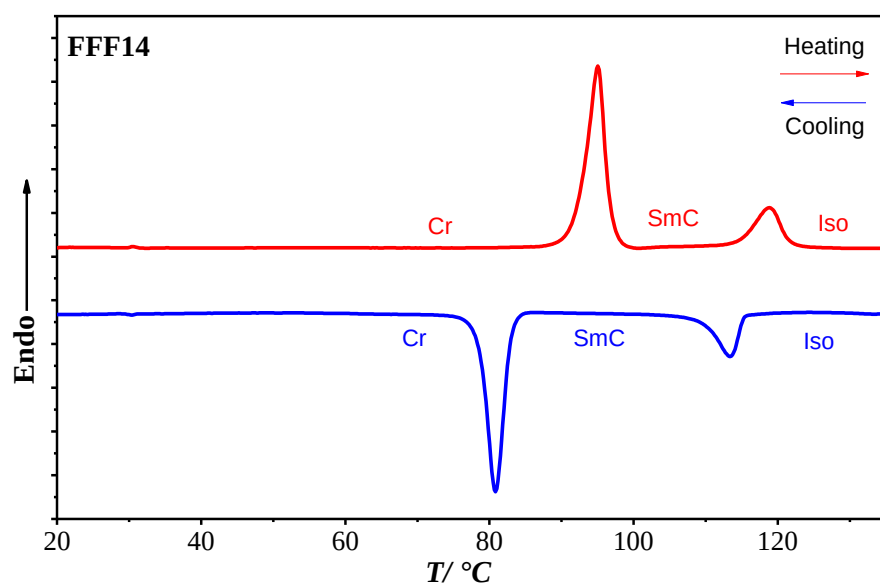


Figure S29. DSC Heating and cooling cycles of the HBLc **FFF14** at a scanning rate of 10 $^\circ\text{C}/\text{min}$.

7. Additional XRD data

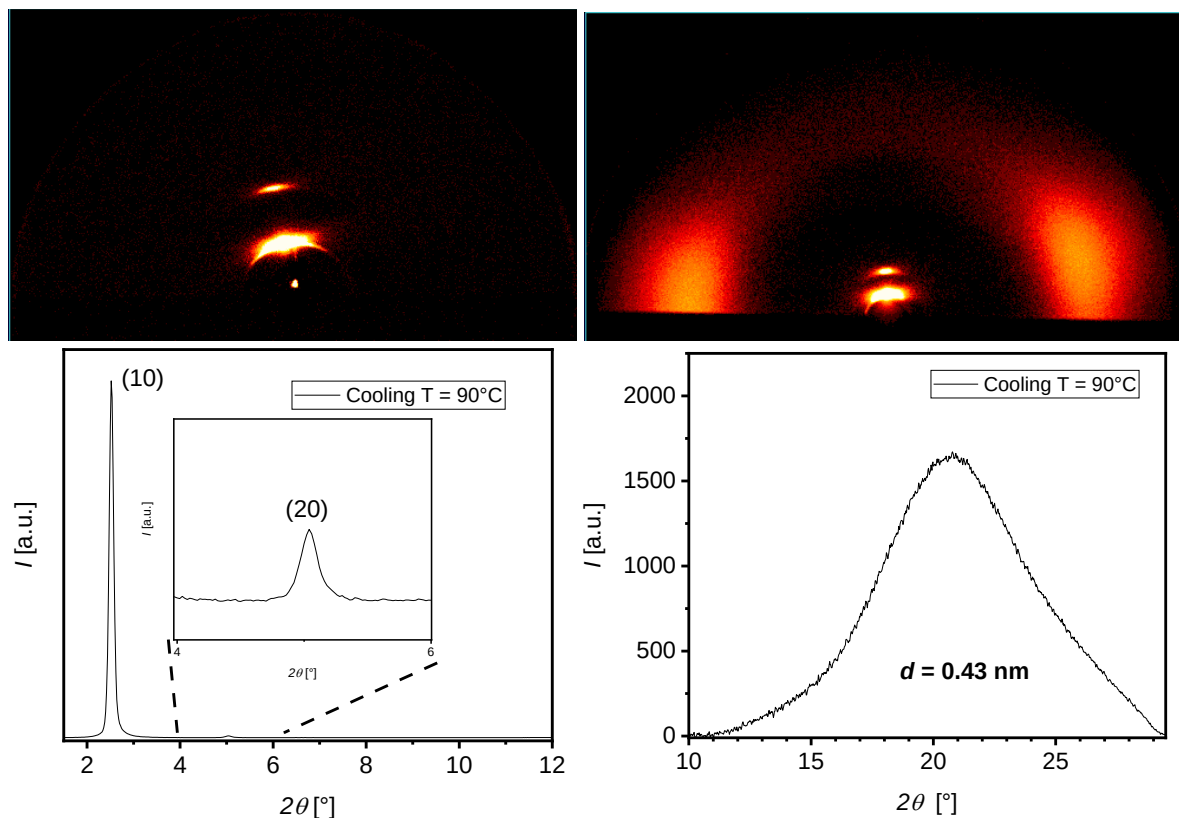


Figure S30. XRD investigations of **FHH8** on cooling in the SmC phase at $T = 90\text{ }^{\circ}\text{C}$. Right column in the small-angle region (SAXS) and left column in the WAXS region.

Table S11. XRD data of the supramolecule **FHH8** at $90\text{ }^{\circ}\text{C}$.

$2\theta\text{ [}^{\circ}\text{]}$	$d_{\text{obs}}\text{ [nm]}$	hkl
2.270	1.750	10
2.694	0.878	20
20.791	0.427	

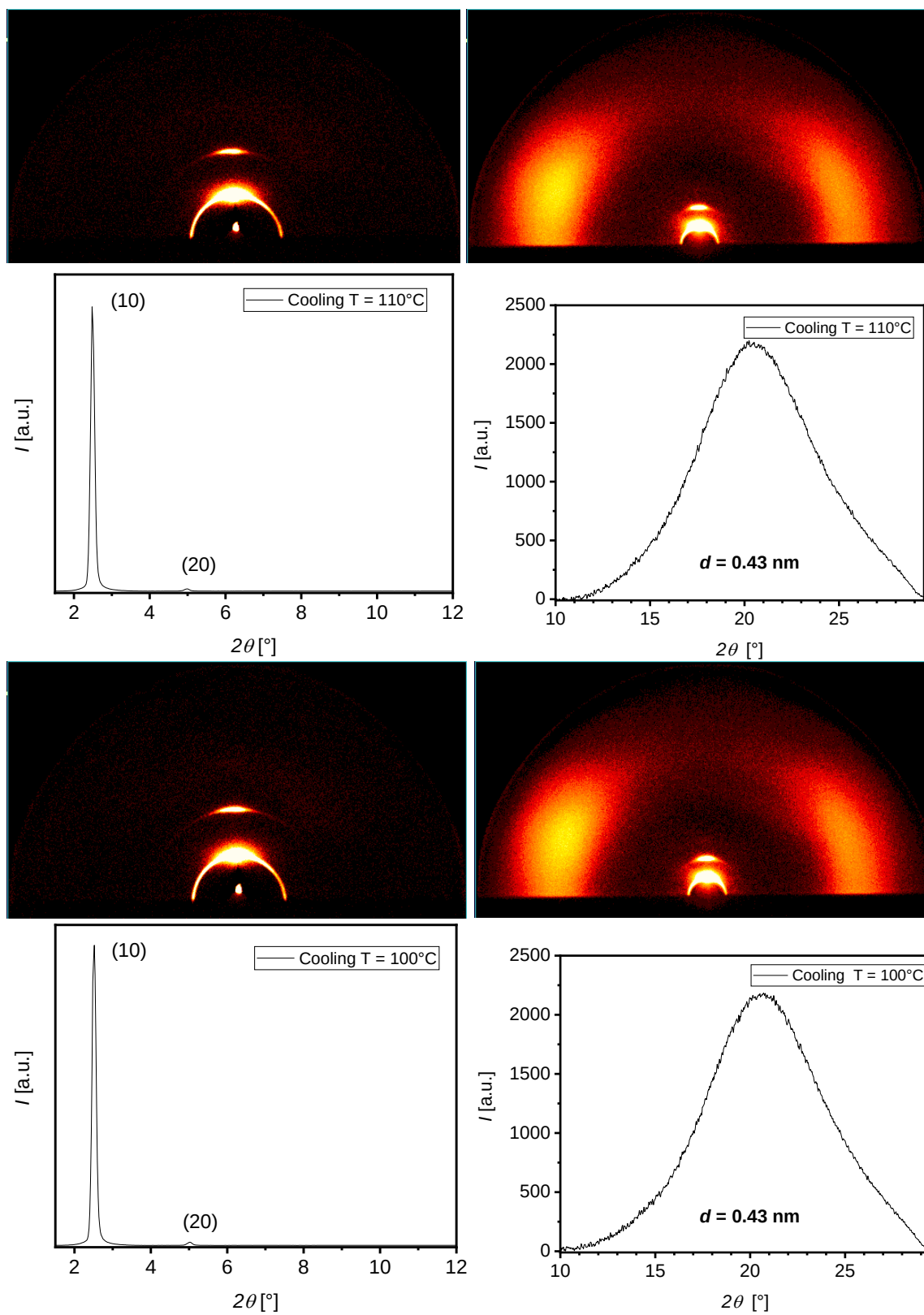


Figure S31. XRD investigations of **FFH12** on cooling in the SmC phase at $T = 110\text{ °C}$ and 100 °C . Right column in the small-angle region (SAXS) and left column in the WAXS region.

Table S12. XRD data of the supramolecule **FFH12** at 110 °C.

2θ [°]	d_{obs} [nm]	hkl
2.493	3.544	10
4.982	1.774	20
20.597	0.431	

Table S13. XRD data of the supramolecule **FFH12** at 100 °C.

2θ [°]	d_{obs} [nm]	hkl
2.514	3.514	10
5.018	1.761	20
20.743	0.428	

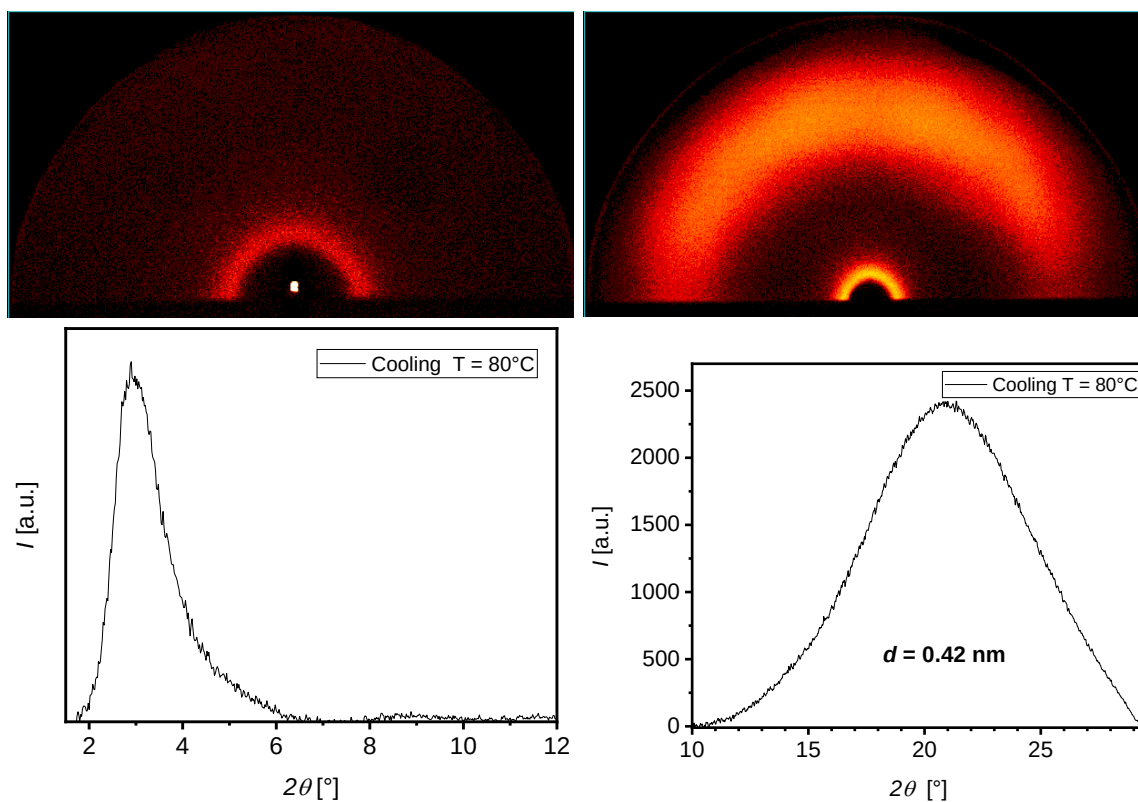


Figure S32. XRD investigations of **HFF8** on cooling in the nematic phase at $T = 80$ °C. Right column in the small-angle region (SAXS) and left column in the WAXS region.

8. References

- 1 M. Alaasar, J. C. Schmidt, A. F. Darweesh and C. Tschierske, *J. Mol. Liq.*, 2020, **310**, 113252.
- 2 M. Alaasar, S. Poppe, Q. Dong, F. Liu and C. Tschierske, *Chem. Commun.*, 2016, **52**, 13869–13872.