# **Supporting Information**

Homoleptic complexes of titanium(IV) fused with O^N^O Schiff Base derivatives: design, BSA-DNA interaction, molecular docking, DFT and cytotoxicity

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Figure.S1. <sup>1</sup>H NMR spectrum of ligand IS (400 MHz, DMSO-d<sub>6</sub>)



Figure.S2. <sup>13</sup>C NMR spectrum of ligand IS (400 MHz, DMSO-d<sub>6</sub>)







Figure.S4. ESI-MS spectrum of ligand IS



Figure.S5. <sup>1</sup>H NMR spectrum of ligand IN (400 MHz, DMSO-d<sub>6</sub>)



Figure.S6. <sup>13</sup>C NMR spectrum of ligand IN (400 MHz, DMSO-d<sub>6</sub>)







#### Figure.S8. ESI-MS spectrum of ligand IN



Figure.S9.<sup>1</sup>H NMR spectrum of ligand IO (400 MHz, DMSO-d<sub>6</sub>)



Figure.S10. <sup>13</sup>C NMR spectrum of ligand IO (400 MHz, DMSO-d<sub>6</sub>)







Figure.S12. ESI-MS spectrum of ligand IO



Figure.S13. <sup>1</sup>H NMR spectrum of ligand IF(400 MHz, DMSO-d<sub>6</sub>)



Figure.S14. <sup>13</sup>C NMR spectrum of ligand IF(400 MHz, DMSO-d<sub>6</sub>)











Figure.S17. <sup>1</sup>H NMR spectrum of ligand ICl (400 MHz, DMSO-d<sub>6</sub>)



Figure.S18. <sup>13</sup>C NMR spectrum of ligand ICl (400 MHz, DMSO-d<sub>6</sub>)







Figure.S20. ESI-MS spectrum of ligand ICl



Figure.S21. <sup>1</sup>H NMR spectrum of ligand IBr (400 MHz, DMSO-d<sub>6</sub>)



Figure.S22. <sup>13</sup>C NMR spectrum of ligand IBr (400 MHz, DMSO-d<sub>6</sub>)





Figure.S24. ESI-MS spectrum of ligand IF



Figure.S25. <sup>1</sup>H NMR spectrum of complex Ti-1-IS (400 MHz, DMSO-d<sub>6</sub>)



Figure.S26. <sup>13</sup>C NMR spectrum of complex Ti-1-IS (400 MHz, DMSO-d<sub>6</sub>)







Figure.S28. ESI-MS spectrum of complex Ti-1-IS



Figure.S29. HPLC spectrum of complex Ti-1-IS







Figure.S31. <sup>13</sup>C NMR spectrum of complex Ti-2-IN (400 MHz, DMSO-d<sub>6</sub>)



Figure.S32. FTIR spectrum of complex Ti-2-IN



Figure.S33. ESI-MS spectrum of complex Ti-2-IN (400 MHz, DMSO-d<sub>6</sub>)



Figure.S34. HPLC spectrum of complex Ti-2-IN



Figure.S35. <sup>1</sup>H NMR spectrum of complex Ti-3-IO (400 MHz, DMSO-d<sub>6</sub>)



Figure.S36. <sup>13</sup>C NMR spectrum of complex Ti-3-IO (400 MHz, DMSO-d<sub>6</sub>)







Figure.S38. ESI-MS spectrum of complex Ti-3-IO



Figure.839. HPLC spectrum of complex Ti-3-IO



Figure.S40. <sup>1</sup>H NMR spectrum of complex Ti-4-IF(400 MHz, DMSO-d<sub>6</sub>)



Figure.S41. <sup>13</sup>C NMR spectrum of complex Ti-4-IF(400 MHz, DMSO-d<sub>6</sub>)



Figure.S42. <sup>19</sup>F NMR spectrum of complex Ti-4-IF(400 MHz, DMSO-d<sub>6</sub>)







Figure.S44. ESI-MS spectrum of complex Ti-4-IF



Figure.S45. HPLC spectrum of complex Ti-4-IF







Figure.S47. <sup>13</sup>C NMR spectrum of complex Ti-5-ICl (400 MHz, DMSO-d<sub>6</sub>)



Figure.S48. FTIR spectrum of complex Ti-5-ICl



Figure.S49. ESI-MS spectrum of complex Ti-5-ICl



Figure.S50. HPLC spectrum of complex Ti-5-ICl



Figure.S51. <sup>1</sup>H NMR spectrum of complex Ti-6-IBr (400 MHz, DMSO-d<sub>6</sub>)



Figure.S52. <sup>13</sup>C NMR spectrum of complex Ti-6-IBr (400 MHz, DMSO-d<sub>6</sub>)







Figure.S54. ESI-MS spectrum of complex Ti-6-IBr



Figure.S55. HPLC spectrum of complex Ti-6-IBr



Fig.S57. Fluorescence Spectra of Ti(IV) complexes in DMSO: H<sub>2</sub>O (1:9)



Fig.S58. UV-Vis stability study of Ti(IV) complexes in 1:9 DMSO: H<sub>2</sub>O medium



Fig.S59. UV-Vis stability study of Ti(IV) complexes in GSH medium



Fig. S60. UV-visible spectra of Ti(IV) complexes for Lipophilicity study of complexes in octanol: water



Fig.S61. UV-Visible spectra of DNA Binding studies of Ti(IV) complexes



Fig.S62. Linear plots of DNA UV-binding studies of Ti(IV) complexes



Fig.S63. Fluorescence quenching spectra of DNA with increasing concentration of Ti(IV) complexes



Fig.S64. Stern-Volmer plots of I<sub>0</sub>/I vs. Ti(IV) complexes



Fig.S65. Scatchard plot of log([I<sub>0</sub>-I]/I) vs. log [Ti(IV) complexes]



Fig.S66. Viscosity Studies of Ti(IV) complexes



Fig.S67. Cyclic voltammetry DNA binding Studies of Ti(IV) complexes

Table S1: Oxidation and reduction peaks obtained from Cyclic Voltammograms of Ti(IV) complexes with CT-DNA(0–50  $\mu M)$ 

Complexes	Oxidation peak	Reduction peak
Ti-1-IS	-0.877, 0.533, 1.099	-0.860, -0.117, 1.327
Ti-2-IN	-1.328, -0.924, -0.233	-0.920, -0.335
Ti-3-IO	-0.892, -0.264	-0.900, -0.355, -0.665, 1.161
Ti-4-IF	-0.892, -0.417, 1.16	-0.892, -0.148, 0.859
Ti-5-ICl	-0.932, 0.425	-0.916, -0.218, 0.750
Ti-6-IBr	-0.908, 0.440, 1.239	-1.249, -0.908, -0.055



Fig.S68. Fluorescence quenching spectra of BSA with increasing concentration of Ti(IV) complexes



Fig.S69. Stern-Volmer plots of  $I_0/I$  vs. complex



Fig.S70. Scatchard plot of log([I<sub>0</sub>-I]/I) vs. log [complex]



Fig.S71. Synchronous spectra of BSA with increasing concentration of Ti(IV) complexes at  $\Delta\lambda$ =15 nm



Fig.S72. Stern-Volmer plots of  $I_0/I$  vs. complex of Synchronous spectra of BSA with increasing concentration of Ti(IV) complexes at  $\Delta\lambda$ =15 nm





Fig.S73. Synchronous spectra of BSA with increasing concentration of Ti(IV) complexes at  $\Delta\lambda$ =60 nm



Fig.S74. Stern-Volmer plots of  $I_0/I$  vs. complex of Synchronous spectra of BSA with increasing concentration of Ti(IV) complexes at  $\Delta\lambda$ =60 nm

Table S2; Stern-Volmer plots of  $I_0/I$  vs. complex of Synchronous spectra of BSA with increasing concentration of Ru(II) complexes at  $\Delta\lambda$ =15 nm and 60nm

Complexes	Ka	Ka
	Δλ=15nm	Δλ=60 nm
Ti-1-IS	0.1444	0.0334
Ti-2-IN	0.0150	0.0445
Ti-3-IO	0.0803	0.1882
Ti-4-IF	0.5071	0.6377
Ti-5-ICl	0.2911	0.1084
Ti-6-IBr	0.1376	0.3096



Fig.S75. Site marker fluorescence quenching studies of BSA+Ibuprofin with an increase in the concentration of Ti(IV) complexes



Fig.S76. Scatchard plot of  $\log([I_0-I]/I)$  vs log [complex] of site marker fluorescence quenching studies of BSA+Ibuprofin



Fig.S77. Site marker fluorescence quenching studies of BSA+Warfarin with an increase in the concentration of Ti(IV) complexes



Fig.S78. Scatchard plot of  $\log([I_0-I]/I)$  vs log [complex] of site marker fluorescence quenching studies of BSA+Warfarin

Table S3; The comparison of binding constants of the complexes Ti(IV) with BSA before and after the addition of site probe at 298 K. K<sub>b</sub> binding constant <sup>a</sup>

COMPLEXES	BSA	BSA+Ibuprofen	BSA + Warfarin
	Kb <sup>a</sup> (× 10 <sup>4</sup> M <sup>-1</sup> )	Kb <sup>a</sup> (× 10 <sup>4</sup> M <sup>-1</sup> )	Kb <sup>a</sup> (× 10 <sup>4</sup> M <sup>-1</sup> )
Ti-1-IS	0.040	0.034	0.048
Ti-2-IN	0.047	0.015	0.069
Ti-3-IO	0.036	0.050	0.058
Ti-4-IF	0.011	0.037	0.040
Ti-5-ICl	0.065	0.035	0.016
Ti-6-IBr	0.014	0.054	0.039



Fig.S79. Molecular docking of DNA with Ti-1-IS; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S80. Molecular docking of DNA with Ti-2-IN ; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S81. Molecular docking of DNA with Ti-3-IO ; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S82. Molecular docking of DNA with Ti-4-IF ; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S83. Molecular docking of DNA with Ti-5-ICl ; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S84.Molecular docking of DNA with Ti-6-IBr ; purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms

Complexes	Nucleotides	Docking scores in kcal/mol
Ti-1-IS	DC3, DG4, DT19, DC21, DG22	-9.3
Ti-2-IN	DG2, DG4, DC3, DC21, DT20	-10.3
Ti-3-IO	DA6, DG22	-8.4
Ti-4-IF	DC3, DG4, DC21, DG22	-9.7
Ti-5-ICl	DC21, DG22, DC3	-9.7
Ti-6-IBr	DG4,DA5,DA6,DA16, T19	-9.5

#### Table S4; Docking scores and binding sites of DNA with Ti(IV) complexes

#### Table S5; Docking scores and binding sites of BSA with Ti(IV) complexes

Complexes	Amino acid residues	Docking scores in kcal/mol
Ti-1-IS	GLU182, ARG185, PRO117, LYS116,	-10.8
	LEU115, LYS114, PRO516	
Ti-2-IN	SER109, LYS114, ARG427, GLU424,	-11.5
	THR421, PRO420	
Ti-3-IO	PRO420, VAL423, ILE522, GLU424,	-9.2
	SER109, PRO110, ARG144	
Ti-4-IF	GLU125, THR121, LEU122, LYS136,	-9.8
	GLU140, LEU115, LYS116, ASP118	
Ti-5-ICl	ASP118, THR121, LEU122, GLU125,	-9.9
	LYS132, LYS136, GLU140, PRO113,	
	LEU115, LYS116, LEU115, LYS116	
Ti-6-IBr	THR121, GLU125, LYS132, LYS136,	-9.7
	GLU140, LYS116, ASP118, LEU122	



Fig.S85. Molecular docking of BSA with Ti-1-IS purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S86. Molecular docking of BSA with Ti-2-IN purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S87. Molecular docking of BSA with Ti-3-IO purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S88. Molecular docking of BSA with Ti-4-IF purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S89. Molecular docking of BSA with Ti-5-ICl purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms



Fig.S90. Molecular docking of BSA with Ti-6-IBr purple color indicates carbon atoms, blue color indicates nitrogen, grey color indicates titanium(IV) ion, red color indicates oxygen atoms

S. NO	Code	O-Ti	N-Ti	O-Ti
1	Ti-1-IS	1.997	2.886	2.032
2	Ti-2-IN	1.997	2.887	2.031
3	Ti-3-IO	2.031	2.882	2.032
4	Ti-4-IF	1.998	2.885	2.030
5	Ti-5-ICl	2.001	2.886	2.031
6	Ti-6-IBr	2.000	2.885	2.029

## Table S6; Bond length (Å) of Ti(IV) complexes

### Table S7; Comparison of experimental and theoretical excitation spectral details

	Experimental	Theoretical Prediction			
Code	Abs (nm)	Abs. max (nm)	Oscillator strength (f)	Transition	Orbital Contribution
Ti-1-IS	339	389	0.325	S <sub>0</sub> →S9	H→L+4 82%, H-3→L 3%,
Ti-2-IN	341	378	0.233	$S_0 \rightarrow S_{10}$	H→L+4 46%, H-3→L 19%,
Ti-3-IO	340	412	0.298	$S_0 \rightarrow S_{12}$	H→L+3 68%, H-3→L 12%,
Ti-4-IF	343	406	0.282	$S_0 \rightarrow S_{10}$	H→L+5 48%, H-4→L 26%,
Ti-5-ICl	345	443	0.316	$S_0 \rightarrow S_{11}$	H→L+3 55%, H→L+4 21%,
Ti-6-IBr	343	625	0.238	$S_0 \rightarrow S_{10}$	H→L+5 78%, H-4→L 6%,

\*H – HOMO, L - LUMO

H→L+4 83%, H-4→L+2 4%



Fig.S91. DPPH assay of Ti(IV) complexes



Fig.S92. MTT assay of Ti(IV) complexes on HeLa cell line



Fig.S93. MTT assay of Ti(IV) complexes on MCF7 cell line

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