

## **Efficient oxidative degradation of organic pollutants in real industrial effluents using a green-synthesized magnetite supported on biochar catalyst**

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## Supplementary texts

### Text S1. Chemicals

Tetracycline ( $C_{22}H_{24}N_2O_8$ , 98%), atrazine ( $C_8H_{14}ClN_5$ , 98%), chlorpyrifos ( $C_9H_{11}Cl_3NO_3PS$ , 97%), methylene blue ( $C_{16}H_{18}ClN_3S$ , ACS reagent grade, 90%), potassium periodate ( $KIO_4$ , ACS reagent grade, 99.8%), sodium iodate ( $NaIO_3$ , ACS reagent grade,  $\geq 99\%$ ), L-Histidine ( $C_6H_9N_3O_2$ , ACS reagent grade,  $\geq 99.5\%$ ), iron (III) chloride hexahydrate ( $FeCl_3 \cdot 6H_2O$ , ACS reagent grade, 97%), and L-Ascorbic acid ( $C_6H_8O_6$ , ACS reagent grade,  $\geq 99\%$ ) were purchased from Sigma-Aldrich Co. (USA). Paracetamol ( $C_8H_9NO_2$ , 98%), hydrogen peroxide ( $H_2O_2$ , 30%), ethanol ( $C_2H_6O$ , HPLC grade, 99.8%), acetonitrile ( $C_2H_3N$ , HPLC grade, 99.9%), phosphoric acid ( $H_3PO_4$ , HPLC grade, 85%), sodium hydroxide ( $NaOH$ , ACS reagent grade), methanol ( $CH_4O$ , 99.9%), chloroform ( $CHCl_3$ , 99.5%), and phenol ( $C_6H_5OH$ , ACS reagent grade,  $\geq 98\%$ ) were supplied from Fischer Chemical Co. Ltd. (UK). Potassium persulphate ( $K_2S_2O_8$ , 99.5%), potassium peroxymonosulfate ( $KHSO_5$ , 99.9%), hydrochloric acid ( $HCl$ , analytical grade, 37%), sulfuric acid ( $H_2SO_4$ , analytical grade, 95-97%), sodium thiosulfate ( $Na_2O_3S_2$ , analytical grade,  $\geq 97\%$ ), and iron (II) sulfate heptahydrate ( $FeSO_4 \cdot 7H_2O$ , ACS reagent grade, 99.5%) were obtained from Merck Chemical Co. Ltd. (Germany).

### Text S2. Water lettuce powder preparation

The collected water lettuce samples were initially rinsed with running tap water to eliminate surface contaminants. Subsequently, the samples were air-dried under natural sunlight to reduce moisture content. The dried plants were then cut into small fragments and dried in a drying oven (ED53, BINDER, USA) at 105 °C for 24 h to enhance brittleness and facilitate uniform grinding. The resulting dried leaves were mechanically ground using an electrical blender into a fine powder with a particle size of less than 100  $\mu m$ . The produced powder was subjected to three consecutive washing cycles with ethanol, each followed by centrifugation (EBA 20, Hettich,

USA). Subsequently, the powder was treated with 1 M hydrochloric acid at a 1:10 (solid-to-liquid) volume ratio at room temperature for 24 h <sup>1</sup>. The acid-treated powder was then thoroughly rinsed with deionized water multiple times until the wash solution reached a neutral pH. The pH value was confirmed using a multiparameter meter (HQ440d, HACH, USA) and a pH probe (PHC301, HACH, USA). Finally, the produced fine powder was dried overnight at 60.0 °C to obtain the purified water lettuce powder.

### **Text S3. Guava leaves extract formulation**

The purchased guava leaves were thoroughly washed with tap water followed by rinsing with deionized water to remove surface impurities. The cleaned leaves were then air-dried at room temperature in the dark for 1 week, followed by oven drying at 60 °C overnight to ensure complete moisture removal <sup>2</sup>. The dried leaves were ground into a fine powder and stored in plastic bags for later use. Subsequently, 40 g of the powdered leaves were mixed with 1 L of deionized water and heated at 80 °C for 2 h using a magnetic stirrer hotplate (MSH-20D, Daihan Scientific, Vietnam) <sup>3</sup>. The resulting suspension was then filtered through Whatman No. 1 filter paper to obtain a clear plant extract with a concentration of 40 g/L. The extract was cooled to room temperature and stored at 4 °C for subsequent applications.

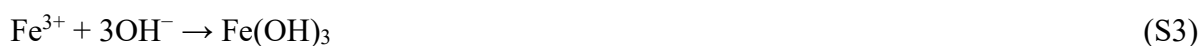
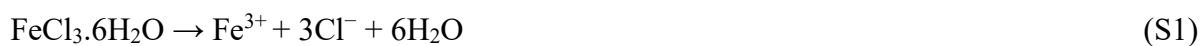
### **Text S4. Synthesis of the water lettuce biochar (WLBC)**

The WLBC was synthesized by pyrolyzing the pre-treated water lettuce powder at 600 °C for 2 h in a muffle furnace (AMF 25 N, Asahi-Rika, Japan) under a nitrogen atmosphere <sup>4</sup>. The resulting black powder was subsequently washed and centrifuged three times with ethanol, followed by three additional washes with deionized water to obtain purified WLBC <sup>5</sup>.

### **Text S5. Biosynthesis of magnetite nanoparticles**

In a 250 mL beaker, ferrous sulfate heptahydrate (4 g) and ferric chloride hexahydrate (8 g) were dissolved in deionized water (100 ml) under continuous stirring for 30 min at room

temperature <sup>6</sup>. Under a nitrogen atmosphere, 20 mL of the pre-prepared guava leaf extract was added dropwise, and the mixture was stirred at 70 °C for 45 min. The resulting black precipitate was magnetically separated, washed three times with deionized water and ethanol under centrifugation, and dried at 60 °C overnight to yield magnetite nanoparticles <sup>7</sup>. The reaction pathway is illustrated in Eqs. (S1–S5) <sup>8,9</sup>. The magnetized water lettuce biochar (MWLB) was prepared by introducing the synthesized WLBC into the reaction mixture before the addition of the plant extract.



#### **Text S6. Characterization techniques**

The functional groups were characterized using Fourier-transform infrared (FTIR) spectroscopy (FTIR-8400S, Shimadzu, Japan). The crystalline structure of the samples was determined using an X-ray diffraction (XRD) system (XRD 6000, Shimadzu, Japan). Surface morphology was examined through transmission electron microscopy (TEM) model (JEM 2100F, JEOL, Japan). Elemental distribution analysis was conducted using energy-dispersive X-ray spectroscopy (EDS) with an X-Max 80 mm<sup>2</sup> detector (Oxford Instruments, UK).

#### **Text S7. Analytical methods**

At 10-min intervals, 3 mL aliquots were withdrawn using disposable syringes, filtered through 0.22 µm syringe filters, and immediately quenched with 200 µL of 1 M sodium thiosulfate solution in centrifuge tubes to neutralize residual oxidizing iodine species <sup>10</sup>. The concentrations of tetracycline (TCN), paracetamol, chlorpyrifos, methylene blue, and atrazine were determined

using a UV-Vis spectrophotometer (Evolution 350, Thermo Fisher Scientific, USA) at 357, 243, 244, 660, and 265 nm, respectively <sup>5,11,12</sup>. Additionally, the concentration of periodate ions ( $\text{IO}_4^-$ ) and iodate ions ( $\text{IO}_3^-$ ) were determined using a high-performance liquid chromatography (HPLC) system (LC-2040C, Shimadzu, Japan), equipped with a diode array detector and an autosampler. Chromatographic separation was conducted on an Agilent XDB-C18 column ( $4.6 \times 250$  mm,  $5 \mu\text{m}$ ) with a mobile phase composed of 0.1% phosphoric acid and acetonitrile in a 70:30 (v/v) ratio. The analytical conditions were as follows: detection wavelength of 228 nm, injection volume of  $10 \mu\text{L}$ , flow rate of  $1.0 \text{ mL/min}$ , retention time of 2.3 min, and a column temperature maintained at  $30^\circ\text{C}$ . These parameters were applied in accordance with the method reported by Xiong et al. <sup>13</sup>. The concentrations of  $\text{IO}_4^-$  and  $\text{IO}_3^-$  were determined by constructing calibration curves using standard solutions of  $\text{KIO}_4$  and  $\text{NaIO}_3$ , respectively, within the range of  $0\text{--}500 \mu\text{M}$  <sup>14,15</sup>. TCN degradation by-products were identified by a liquid chromatography-mass spectroscopy (LC-MS) model (LCMS-2020, Shimadzu, Japan) under chromatographic conditions as described by Huang et al. <sup>16</sup>. Total organic carbon (TOC) was determined using the TOC-L CPH/CPN analyzer (Shimadzu, Japan). Total dissolved solids were quantified following Standard Method 2540-C <sup>17</sup>. Electrical conductivity was measured using a multimeter (HQ440d, HACH, USA) equipped with a conductivity probe (CDC401, HACH, USA). Removal efficiencies (RE) of pollutants, TOC mineralization rates, and oxidant decomposition ratios during the degradation processes were calculated using Eq. (S6) <sup>18</sup>. Degradation kinetics were assessed by fitting the experimental data to a pseudo-first-order kinetic model, as described in Eq. (S7) <sup>19</sup>. The coefficient of determination ( $R^2$ ) was used to evaluate the model's fit. Additionally, the catalyst's adsorption capacity ( $Q$ ,  $\text{mg/g}$ ) was determined using Eq. (S8) <sup>20</sup>.

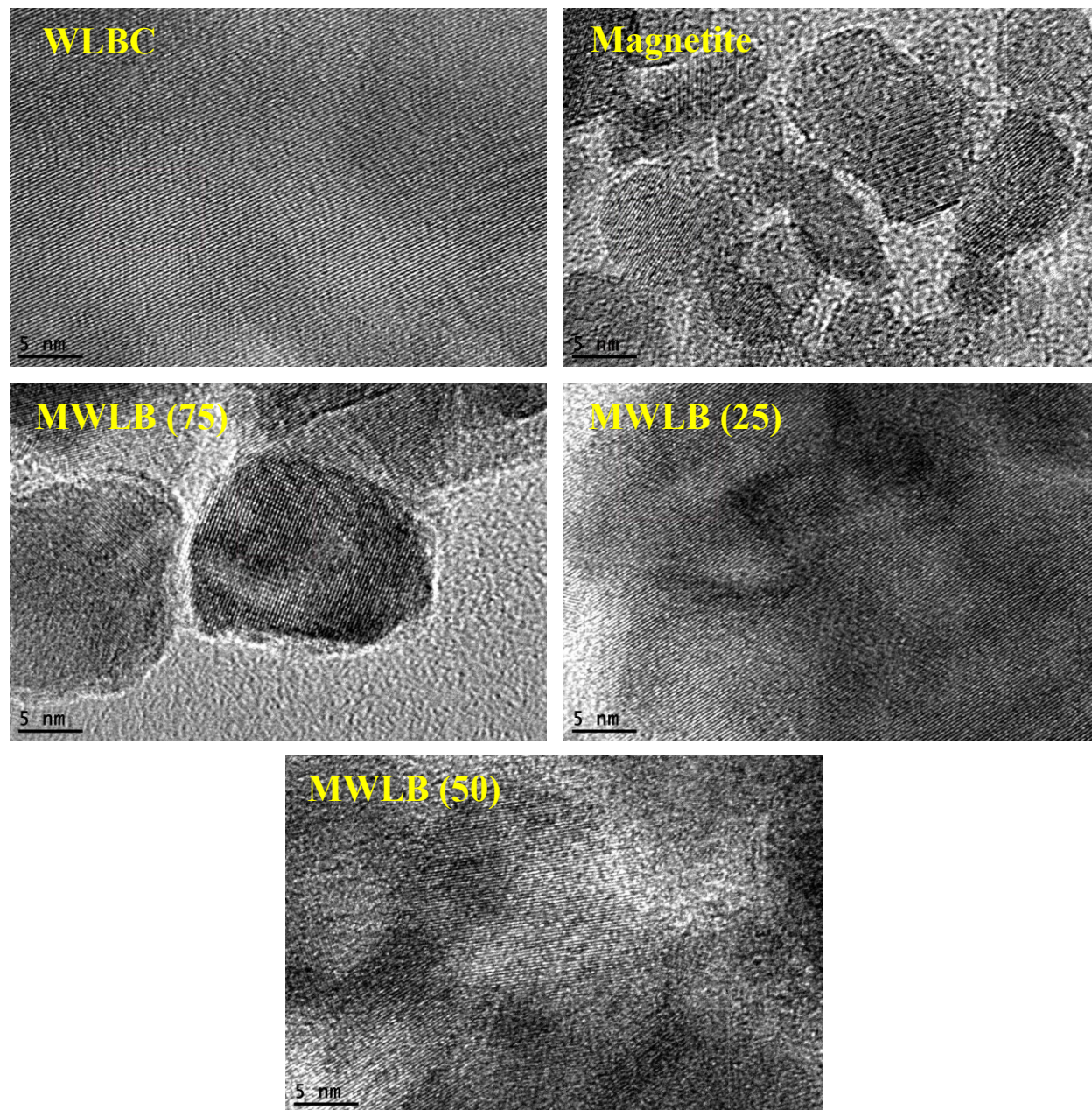
$$RE = \frac{C_o - C}{C_o} \times 100\% \quad (S6)$$

$$-\ln\left(\frac{C}{C_o}\right) = K_{obs} \cdot t \quad (S7)$$

$$Q = \frac{V(C_o - C)}{W} \quad (S8)$$

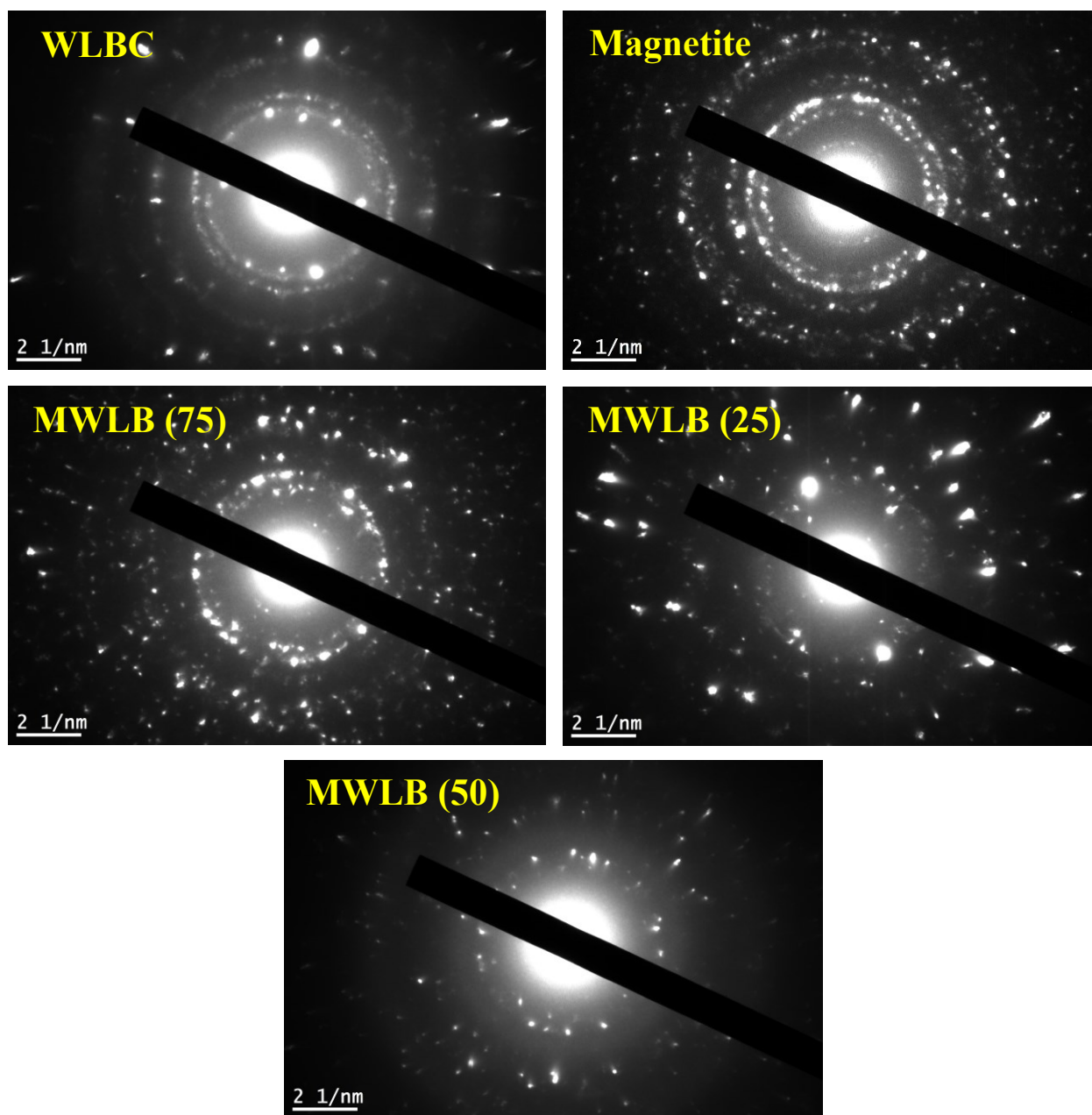
where  $C_o$  represents the pollutant initial concentration,  $C$  denotes the pollutant final concentration,  $K_{obs}$  refers to the kinetic rate constant ( $\text{min}^{-1}$ ),  $t$  corresponds the reaction time (min),  $V$  indicates to the reaction solution volume (L), and  $W$  is the mass of the catalyst (g).

## Supplementary figures



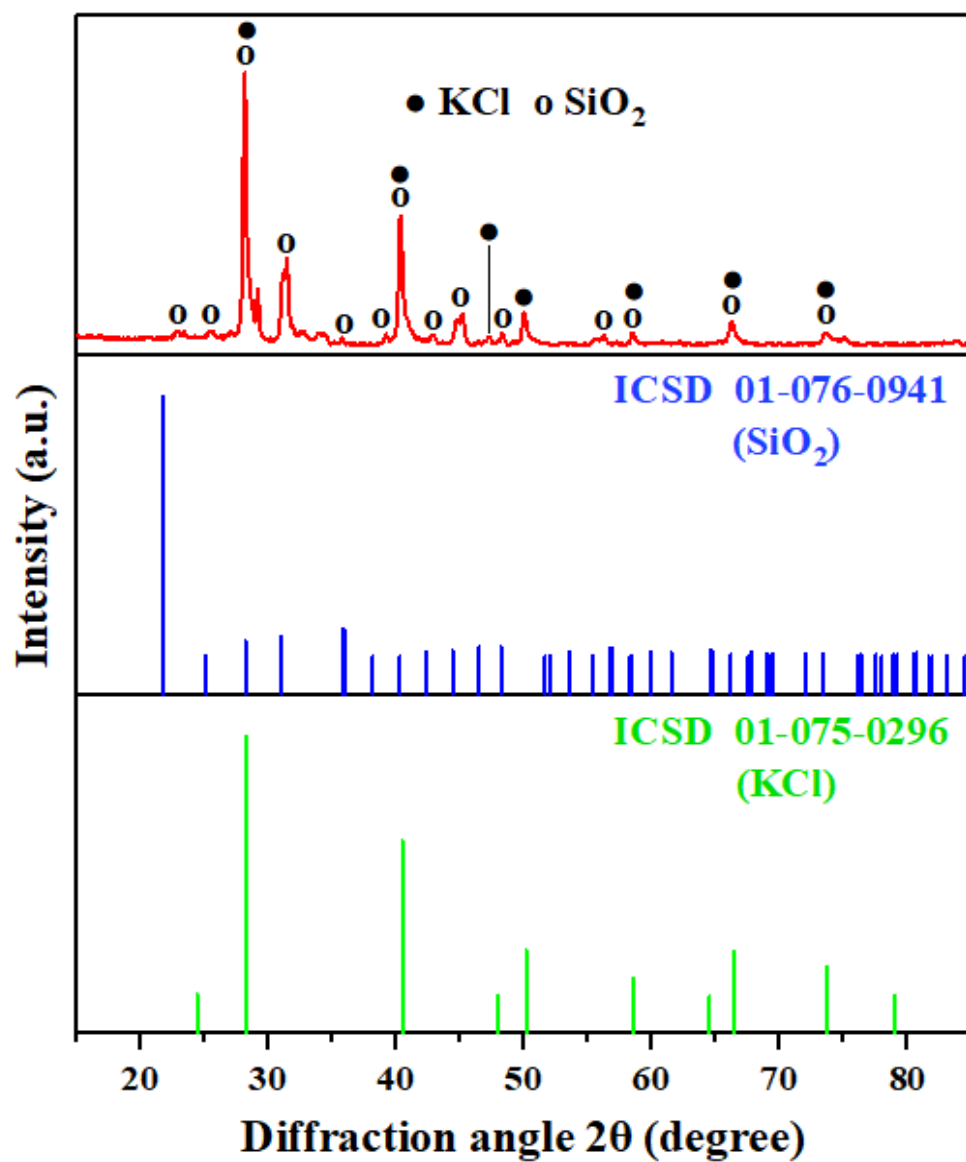
**Fig. S1.** High-resolution TEM (HRTEM) scans of the synthesized catalysts.



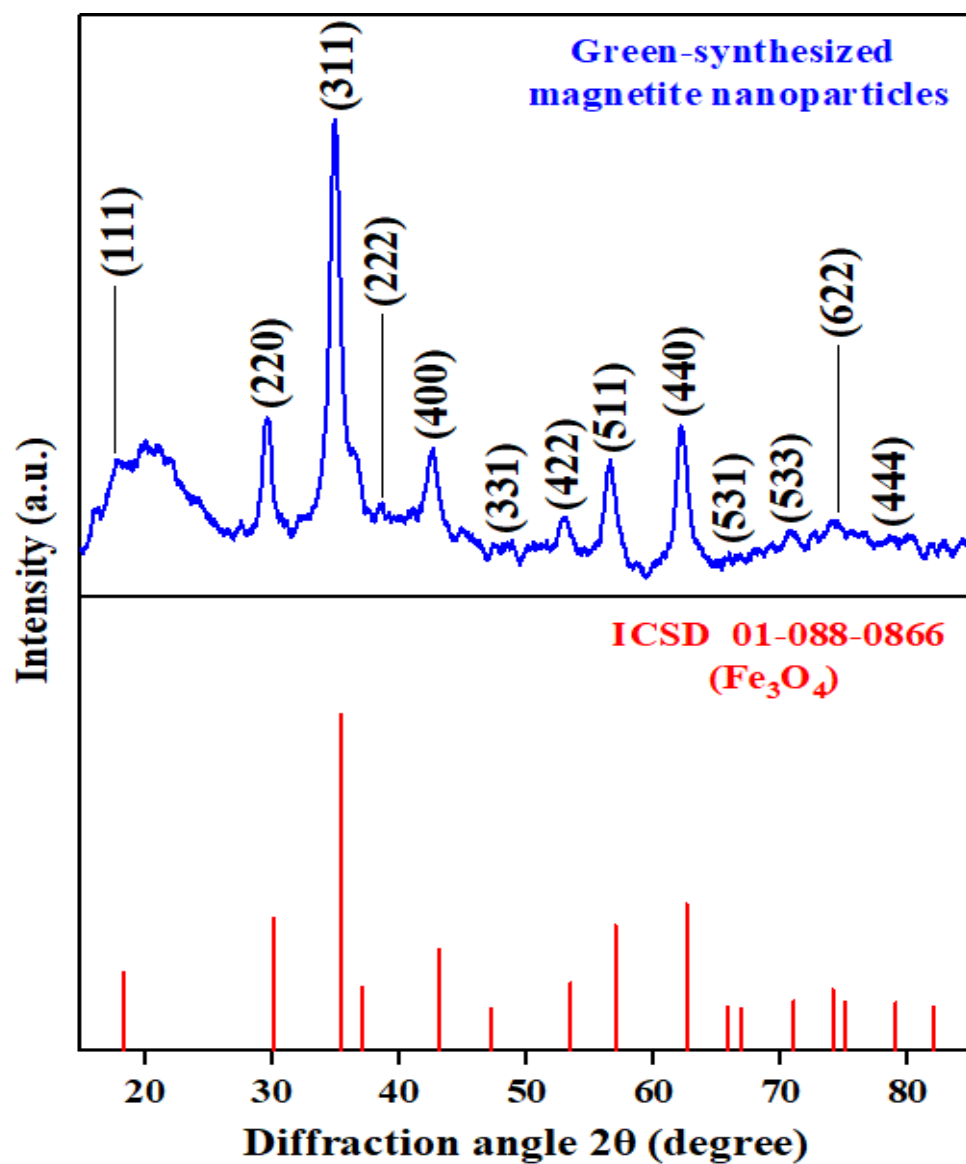


**Fig. S2.** Selected area electron diffraction (SAED) micrographs of the fabricated materials.

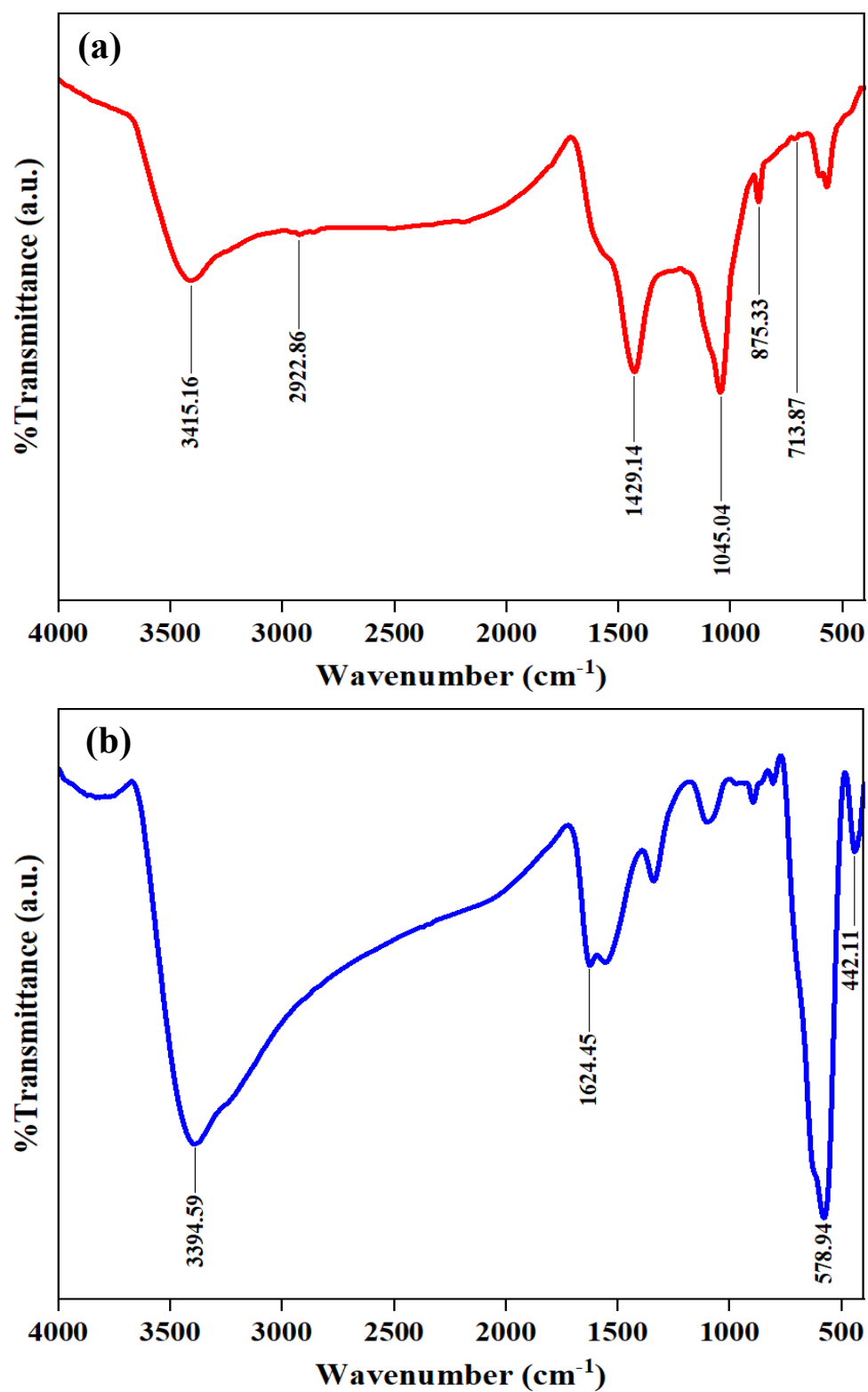




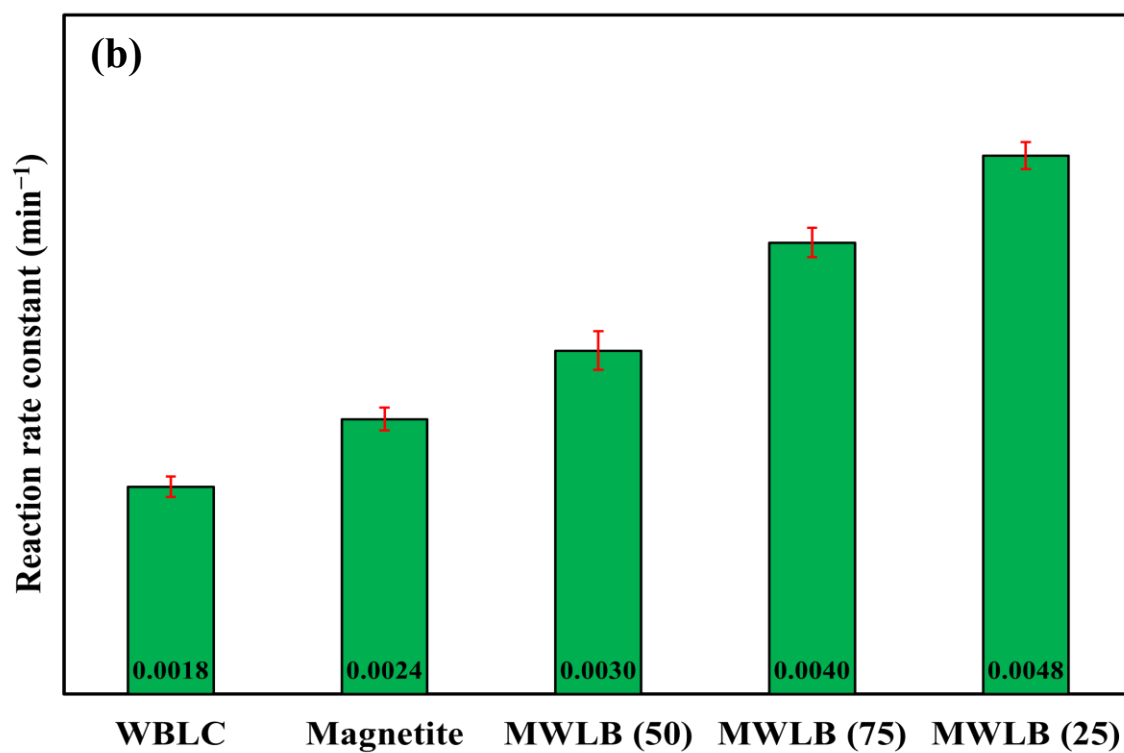
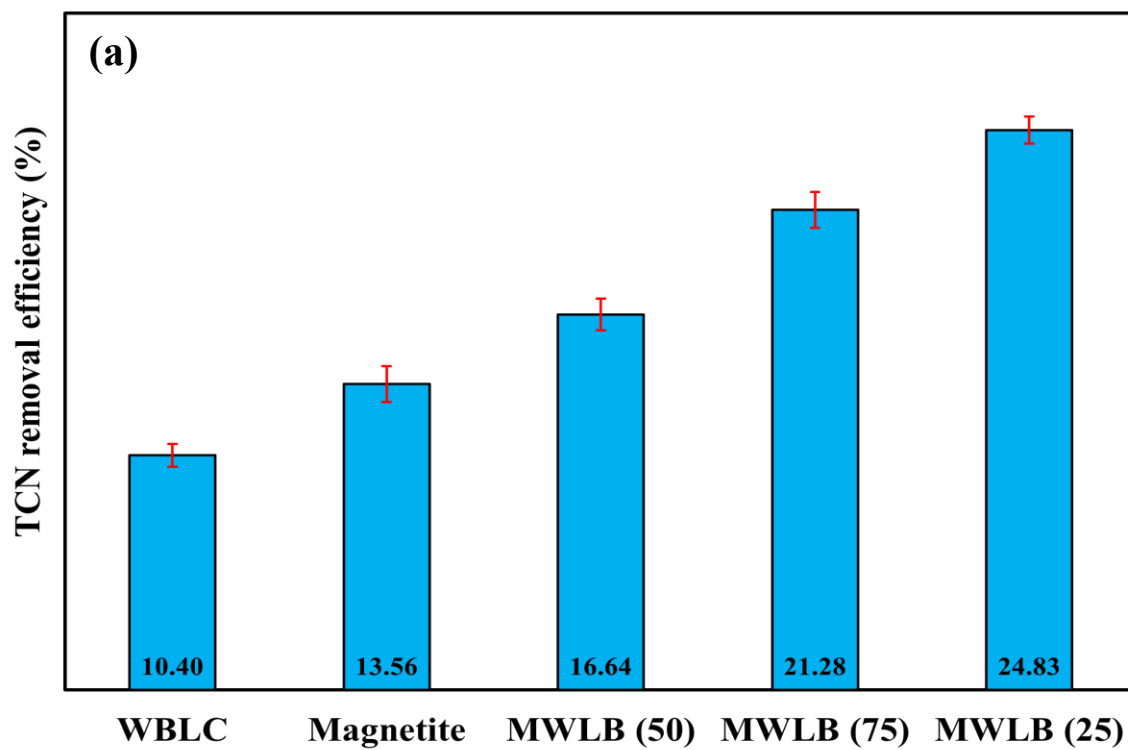
**Fig. S3.** XRD pattern the synthesized WLBC, compared to reference peaks from  $\text{SiO}_2$  and KCl standard cards.

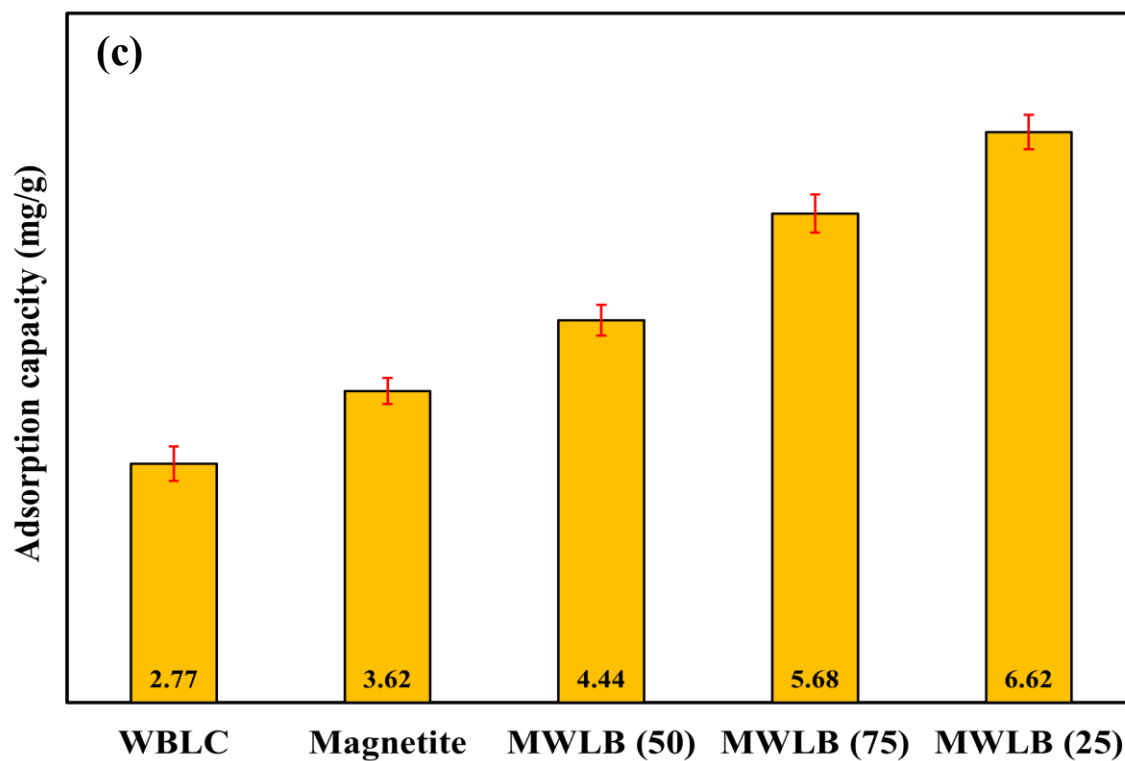


**Fig. S4.** XRD pattern of the green-synthesized magnetite nanoparticles reference to standard magnetite pattern (ICSD 01-088-0866).

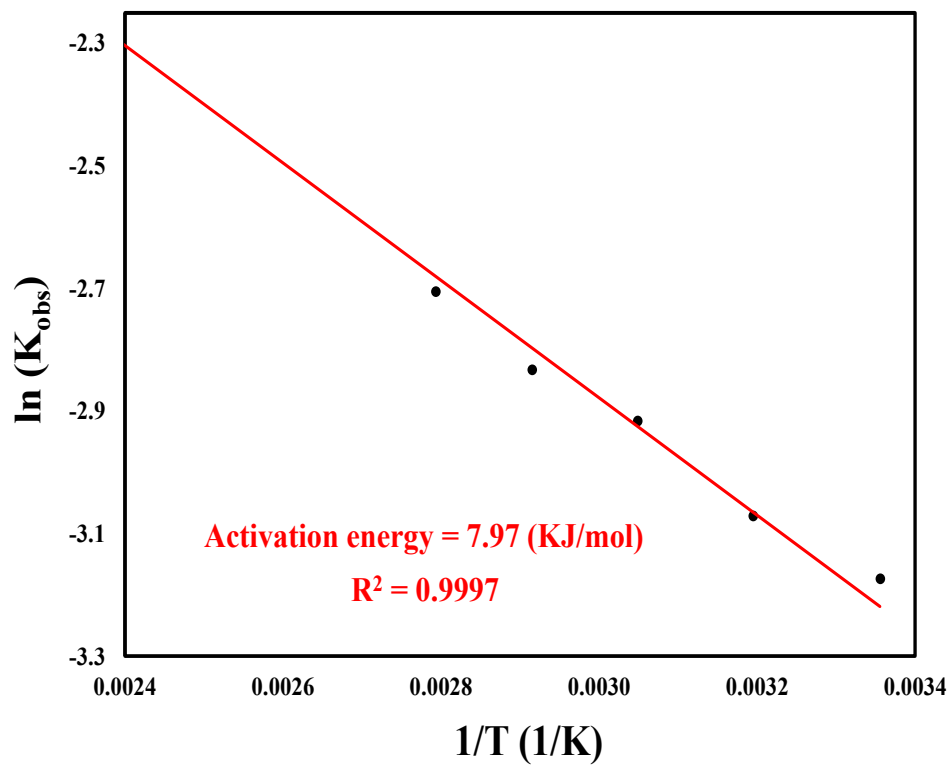


**Fig. S5.** FTIR spectra of (a) WLBC and (b) the green-synthesized magnetite nanoparticles.

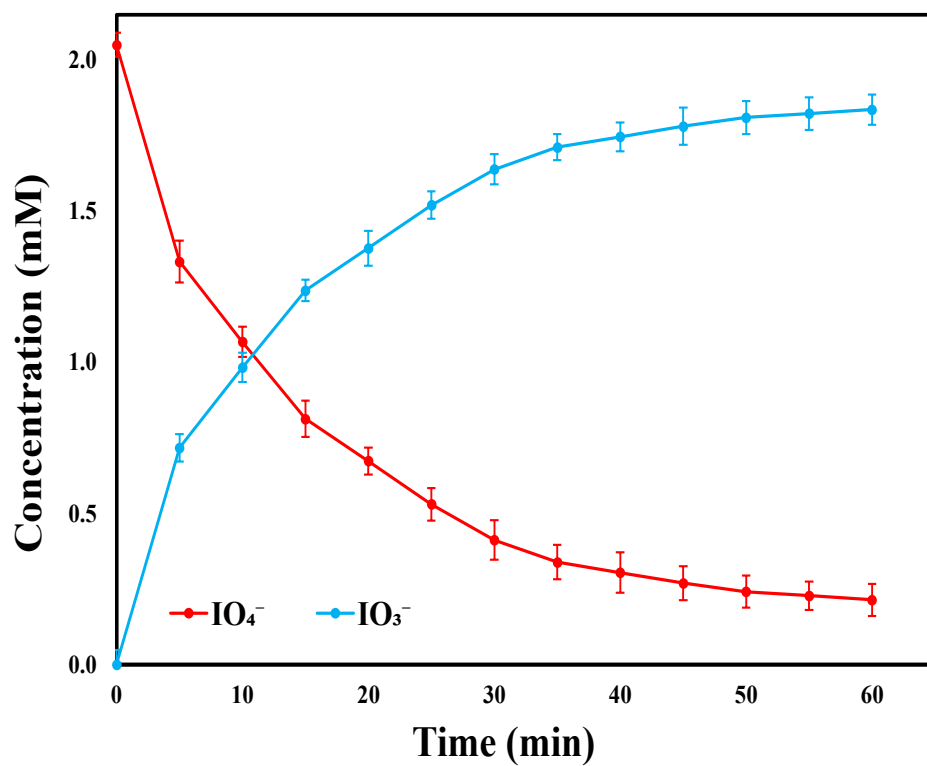




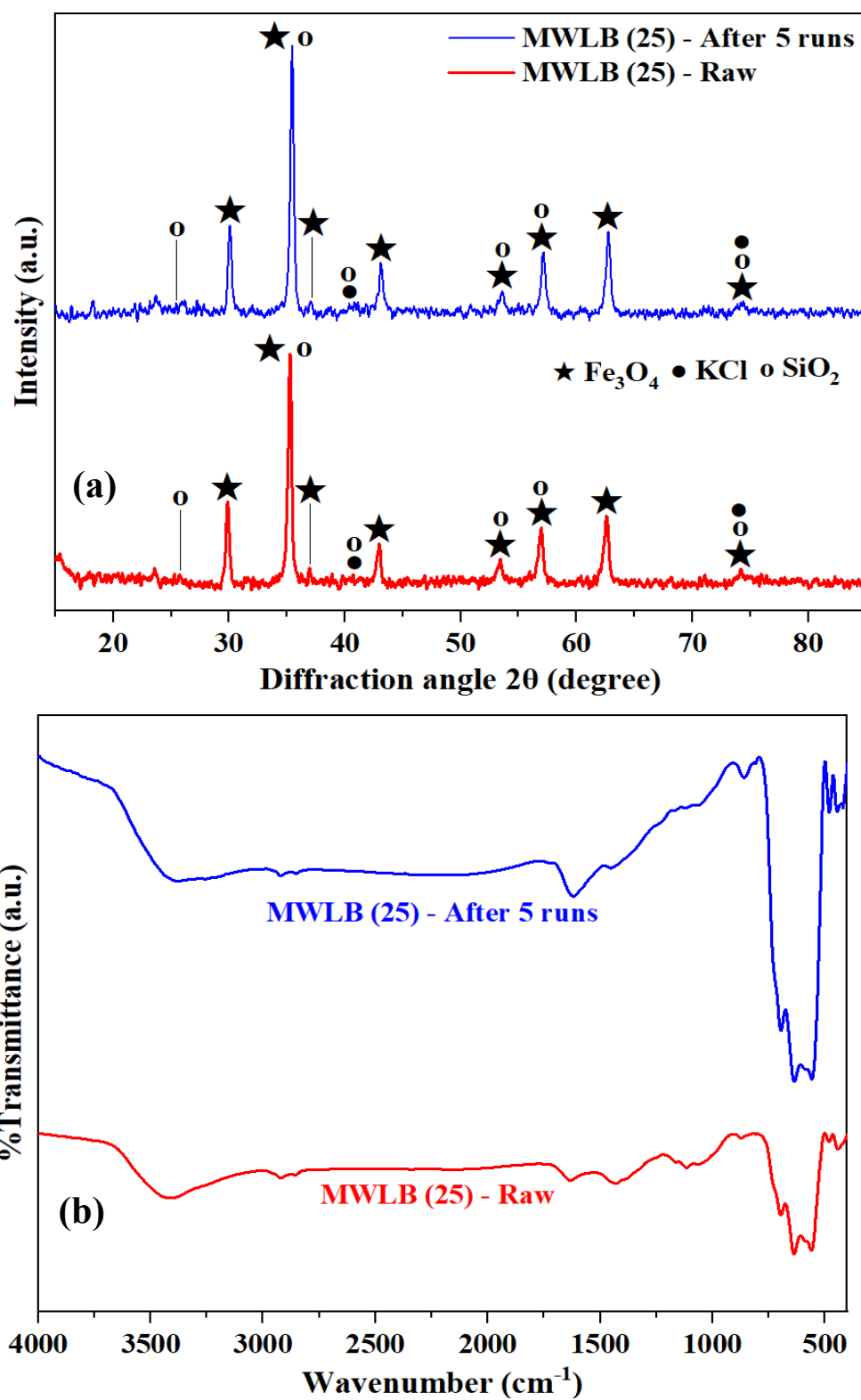
**Fig. S6.** Evaluation of the adsorption performance of the synthesized catalysts for TCN removal under the control conditions: (a) TCN degradation efficiency, (b) corresponding kinetic rate constants, and (c) calculated adsorption capacities.



**Fig. S7.** Activation energy for TCN degradation in the MWLB (25)/PI system.

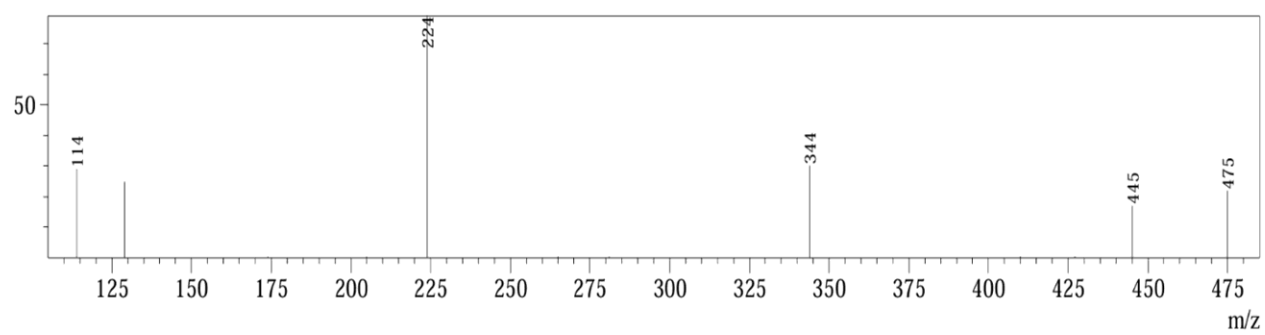


**Fig. S8.** Concentration of iodine species during TCN degradation in the MWLB (25)/PI system under the optimum conditions.



**Fig. S9.** (a) XRD patterns and (b) FTIR spectra of the MWLB (25) composite before and after five successive treatment cycles.





**Fig. S10.** The LC-MS spectra of TCN degradation in the MWLB (25)/PI system.

## Supplementary tables

**Table S1** Characterization of the real pharmaceutical industrial effluent.

Parameter	Concentration	Unit
TCN	428.36	mg/L
Total organic carbon	1372.29	mg/L
pH	7.48	—
Electrical conductivity	1466.85	μS/cm
Total dissolved solids	873.61	mg/L

**Table S2** XRD 2 $\theta$  peak positions of the synthesized composites, along with their corresponding miller indices from standard reference data.

The synthesized catalysts				Reference cards					
MWLB (50)	MWLB (25) - Raw	MWLB (25) - After 5 runs	MWLB (75)	SiO <sub>2</sub>		KCl		Fe <sub>3</sub> O <sub>4</sub>	
Diffraction peaks (2 $\theta$ )				2 $\theta$	hkl	2 $\theta$	hkl	2 $\theta$	hkl
18.45	18.34	18.52	18.33	—	—	—	—	18.31	111
21.83	21.76	21.9	21.71	21.81	101	—	—	—	—
24.72	24.52	24.69	24.52	—	—	24.49	111	—	—
25.39	25.11	25.16	25.1	25.17	110	—	—	—	—
28.05	28.26	28.31	28.36	28.24	111	28.38	200	—	—
30.16	30.29	30.66	30.27	—	—	—	—	30.12	220
31.06	31.18	31.24	31.32	31.09	102	—	—	—	—
35.57	35.89	35.44	35.26	35.9	200	—	—	35.48	311
36.04	36.22	36.43	36.18	36.02	112	—	—	37.11	222
37.11	37.21	37.36	37.24	—	—	—	—	37.11	222
38.46	38.41	38.48	38.22	38.19	201	—	—	—	—
40.55	40.41	40.47	40.43	40.35	210	40.52	220	—	—
42.61	42.34	42.44	42.55	42.4	211	—	—	43.11	400
43.22	43.27	43.51	43.43	—	—	—	—	43.12	400
44.44	44.36	44.5	44.68	44.46	202	—	—	—	—
46.54	46.52	46.73	46.49	46.46	113	—	—	—	—
47.84	47.75	47.82	47.69	—	—	47.96	311	47.21	331
48.29	48.36	48.47	48.55	48.28	212	—	—	—	—
50.37	50.35	50.47	50.23	—	—	50.19	222	—	—
51.59	51.54	51.59	51.76	51.68	220	—	—	—	—
52.16	52.03	52.17	52.12	52.01	004	—	—	53.5	422
53.74	53.41	53.57	53.46	53.59	203	—	—	53.5	422
55.53	55.63	55.69	55.6	55.41	104	—	—	—	—
56.62	56.96	56.57	56.89	56.91	213	—	—	57.07	511

58.53	58.68	58.71	58.64	58.33	310	58.66	400	—	—
59.89	59.75	59.75	59.99	59.94	311	—	—	—	—
61.67	61.73	61.77	61.71	61.56	302	—	—	—	—
62.85	62.6	62.74	62.53	—	—	—	—	62.62	440
64.63	64.79	64.87	64.59	64.61	312	64.58	331	—	—
65.77	65.83	65.93	65.79	—	—	—	—	65.86	531
66.54	66.79	66.75	66.67	66.20	223	66.4	420	66.90	442
67.79	67.76	67.76	67.98	67.81	214	—	—	—	—
68.83	68.99	68.95	68.85	69	321	—	—	—	—
69.13	69.3	69.32	69.25	69.44	105	—	—	—	—
71.12	71.27	71.28	71	—	—	—	—	71.05	620
72.21	72.18	72.22	72.09	72.1	313	—	—	—	—
73.79	73.39	73.33	73.62	73.36	322	73.71	422	—	—
74.13	74.17	74.17	74.11	—	—	—	—	74.5	533
75.14	75.15	75.12	75.15	—	—	—	—	75.09	622
76.25	76.42	76.43	76.51	76.4	224	—	—	—	—
77.61	77.65	77.67	77.77	77.95	205	—	—	—	—
78.92	78.85	78.73	78.97	78.90	410	—	—	—	—
79.01	79.13	79.3	79.14	79.18	304	79.05	511	79.06	444
80.55	80.58	80.49	80.65	80.44	323	—	—	—	—

**Table S3** FTIR peaks (cm<sup>-1</sup>) of the synthesized MWLB composites.

MWLB (50)	MWLB (25) - Raw	MWLB (25) - After 5 runs	MWLB (75)
—	3413.81	3377.75	3366.35
—	2921.61 2857.65	2920.83	2916.06 2857.15
1598.0	1632.16	1619.51	1616.06
1428.73	1431.28	1451.83	1432.91
1126.26	1161.42 1116.06 1062.52	—	1047.44
874.95	872.93	859.24	878.92
557.8 478.48 440.71 418.7	559.37 481.27 441.02 418.97	557.85 479.67 444.49 418.59	560.84 442.64

**Table S4** Comparison of activation energies for TCN degradation in different AOPs.

System	Temperature range (°C)	Activation energy (KJ/mol)	Reference
MWLB (25)/PI	25–85	7.97	This study
Ultrasound/persulfate	25–65	32.01	21
Heat/persulfate	40–70	70.48	22
Electro-Fenton process	15–35	32.2	23
Heat/PI	30–80	46.2	24
Fe, Co, and O co-doped g-C <sub>3</sub> N <sub>4</sub> /persulfate	15–35	23.88	25
N-doped carbon nanotubes encapsulated cobalt/PI	15–45	8.87	26

**Table S5** Comparative degradation performance of the MWLB (25)/PI system and various reported AOP systems employing different catalysts.

AOP system	Catalyst/light source	Operating conditions	Removal ratio	Reference
Photocatalysis	Fe <sub>3</sub> O <sub>4</sub> /rGO/TiO <sub>2</sub> nanocomposite Xenon lamp (150 W)	[TCH] <sub>o</sub> = 20 mg/L, [Catalyst] <sub>o</sub> = 0.6 g/L, pH = 6, T = 25 °C, and reaction time = 330 min.	93.1%	27
PI activation	Chalcopyrite (CuFeS <sub>2</sub> )	[TCH] <sub>o</sub> = 50 mg/L, [Catalyst] <sub>o</sub> = 0.3 g/L, [PI] <sub>o</sub> = 0.8 mM, pH = 8, and reaction time = 90 min.	89.5%	28
Photocatalysis	Fe <sub>3</sub> O <sub>4</sub> /graphitic carbon nitride (g-C <sub>3</sub> N <sub>4</sub> )/rGO Xenon lamp (300 W, 420 nm)	[TCH] <sub>o</sub> = 10 mg/L, [Catalyst] <sub>o</sub> = 0.1 g/L, pH = 6, T = 25 °C, and reaction time = 60 min.	86.7%	29
PI activation	Potassium ferrate modified biochar (Fe-BC)	[TCN] <sub>o</sub> = 20.3 mg/L, [Catalyst] <sub>o</sub> = 1.09 g/L, [PI] <sub>o</sub> = 3.29 mM, pH = 3, T = 25 °C, and reaction time = 150 min.	100%	30
Photocatalysis	Fe <sub>3</sub> O <sub>4</sub> /g-C <sub>3</sub> N <sub>4</sub> /MoO <sub>3</sub> nanocomposite Xenon lamp (300 W)	[TCN] <sub>o</sub> = 10 mg/L, [Catalyst] <sub>o</sub> = 1 g/L, pH = 7, T = 25 °C, and reaction time = 60 min.	98%	31



PI activation	MWLB (25)	[TCN] <sub>0</sub> = 16.52 mg/L, [Catalyst] <sub>0</sub> = 0.83 g/L, [PI] <sub>0</sub> = 2.05 mM, pH = 7, T = 25 °C, and reaction time = 60 min.	99.64%	This study
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