

Design and Synthesis of a Novel Chiral Triptycene-Based Imine Ligand for Enantioselective Epoxide Ring-Opening Reactions

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1. Experimental Section

1.1. Materials & Methods

All the solvents and reagents were purchased at the highest commercially quality and used without further purification. Melting points were obtained on a Thomas-Hoover apparatus in open capillaries and are uncorrected. FTIR spectra were recorded using Bruker alpha Eco-ATR spectrometer in the spectral region of 4000-650 cm^{-1} . ^1H -NMR spectra were recorded on Bruker Avance Neo 500 MHz spectrometer in CDCl_3 solvent. Spectroscopic data are represented as follows: chemical shift (ppm), multiplicity (s = singlet, d = doublet, t = triplet, dd = doublet of doublets, m = multiplet, br = broad singlet, ddd = doublet of doublet of doublet, dddd = doublet of doublet of doublet of doublet, td = triplet of doublet, dt = doublet of triplet, qd = quartet of doublet), integration, coupling constants in Hertz (Hz). ^{13}C NMR spectra were recorded at 125 MHz in CDCl_3 relative to trimethylsilane as internal standard. Powder X-ray diffraction (PXRD) spectra were recorded by using Panalytical's X'Pert Pro X-ray diffractometer. Field emission scanning electron microscope (FESEM) images and Energy-dispersive X-ray spectroscopy (EDS) data was obtained on a field emission scanning electron microscope (HITACHI Japan SU8010 Series). Crude products were isolated and purified by column chromatography over 60-120 mesh size silica gel using hexanes and ethyl acetate as eluents. The progress of the reaction was monitored by thin layer chromatography (TLC) on silica coated aluminum plates F₂₅₄ and visualized in UV chamber.

1.2. General Procedure for Epoxide Ring-Opening Reaction:

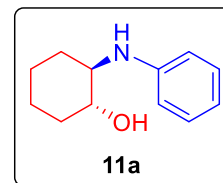
In a 25 mL round-bottom flask, epoxide (0.4 mmol, 1 equiv.) and 5 mol% of the triptycene-based cobalt catalyst (0.02 mmol) were dissolved in 1 mL of *tert*-butyl methyl ether. Subsequently, 5 mol% of an additive (0.02 mmol) was added to the reaction mixture. The mixture was stirred at room temperature, followed by the addition of aniline (0.4 mmol, 1 equiv.) or its derivatives. The reaction was allowed to proceed for the optimized reaction time, and progress was monitored by TLC. Upon completion, the solvent was evaporated under reduced pressure, and the crude product was purified by column chromatography using a gradient of ethyl acetate and hexanes (10:90 to 20:80) as the eluents to afford the pure product. The enantiomeric excess was determined by HPLC analysis using chiral columns Chiralpak IA, IB, ASH and phenomenex Amylose-1 ($\lambda = 254$ nm, IPA/hexanes used as the eluent).

1.3. Characterization Data

2-(phenylamino)cyclohexan-1-ol (11a): 92% yield, brown oil; **IR (ATR):**

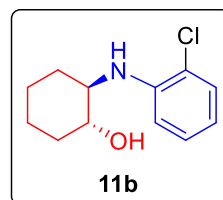
$\nu_{\max}$ 3349, 2958, 2923, 2853, 1611, 1461, 1278, 1261, 764, 750 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.20 (t, $J = 8\text{ Hz}$, 2H), 6.83-6.77 (m, 3H), 3.41

(td, $J_1 = 4.5\text{ Hz}$, $J_2 = 9.5\text{ Hz}$, 1H), 3.19-3.11 (m, 1H), 2.16-2.05 (m, 2H), 1.82-1.68 (m, 2H), 1.46-1.34 (m, 1H), 1.34-1.26 (m, 2H), 1.16-1.06 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 146.8, 129.5, 119.5, 115.4, 74.3, 61.2, 33.4, 31.4, 25.1, 24.4, ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 8.02 min, τ_{R} (major) = 8.6 min, 52% ee.

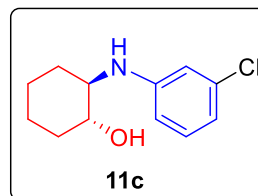


2-((2-chlorophenyl)amino)cyclohexan-1-ol (11b): 99% yield, Light brown oil **IR (ATR):** $\nu_{\max}$ 3401, 2929, 2856, 1596, 1515, 1325, 1032, 739, 694 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.28-7.25 (m, 1H), 7.15-7.11 (m, 1H), 6.89 (d, $J = 8\text{ Hz}$, 1H), 6.68 (td, $J_1 = 1.5\text{ Hz}$, $J_2 = 8\text{ Hz}$, 1H), 3.49 (td, $J_1 =$

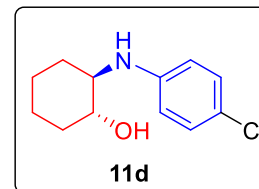
4.5 Hz , $J_2 = 9.5\text{ Hz}$, 1H), 3.25-3.17 (m, 1H), 2.17-2.04 (m, 2H), 1.83-1.71 (m, 2H), 1.46-1.37 (m, 1H), 1.36-1.29 (m, 2H), 1.22-1.12 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 143.4, 129.6, 128.0, 118.8, 115.6, 113.7, 74.5, 60.4, 33.3, 31.6, 25.1, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 6.4 min, τ_{R} (major) = 9.1 min, 20% ee.



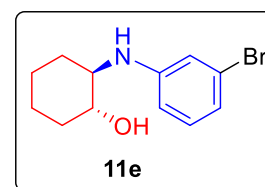
2-((3-chlorophenyl)amino)cyclohexan-1-ol (11c): 93% yield, light brown oil, **IR (ATR):** $\nu_{\max}$ 3347, 2931, 2857, 1596, 1503, 1483, 1280, 988, 938, 889, 682 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.07 (t, $J = 8.5\text{ Hz}$, 1H), 6.72-6.68 (m, 2H), 6.59-6.55 (m, 1H), 3.36 (td, $J_1 = 4.5\text{ Hz}$, $J_2 = 9.5\text{ Hz}$, 1H), 3.13-3.06 (m, 1H), 2.13-2.04 (m, 2H), 1.81-1.67 (m, 2H), 1.44-1.34 (m, 1H), 1.34-1.27 (m, 2H), 1.11-1.01 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 149.0, 135.2, 130.4, 118.4, 114.1, 112.8, 74.5, 60.2, 33.4, 31.6, 25.0, 24.4 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 9.1 min, τ_{R} (major) = 9.6 min, 62% ee.



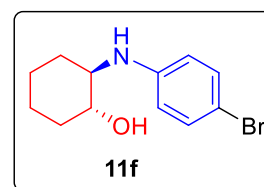
2-((4-chlorophenyl)amino)cyclohexan-1-ol (11d): 99% yield, brown solid ; **IR (ATR):** ν_{max} 3403, 2924, 2856, 1671, 1375, 951, 849, 697 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.13 (dt, $J_1 = 3 \text{ Hz}$, $J_2 = 10 \text{ Hz}$, 2H), 6.70-6.66 (m, 2H), 3.38 (td, $J_1 = 4.5 \text{ Hz}$, $J_2 = 9.5 \text{ Hz}$, 1H), 3.12-3.05 (m, 1H), 2.14-2.04 (m, 2H), 1.82-1.68 (m, 2H), 1.44-1.34 (m, 1H), 1.34-1.26 (m, 2H), 1.13-1.03 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 145.8, 129.3, 123.7, 116.1, 74.4, 61.0, 33.4, 31.4, 25.0, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 10.6 min, τ_{R} (major) = 12..2 min, 42% ee.



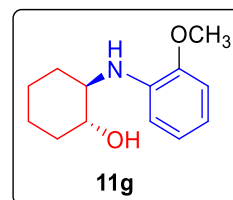
2-((3-bromophenyl)amino)cyclohexan-1-ol (11e): 94% yield, brown oil; **IR (ATR):** ν_{max} 3348, 2930, 2856, 1593, 1500, 1480, 1449, 1325, 1279, 759, 582 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.01 (t, $J = 8.5 \text{ Hz}$, 1H), 6.86-6.82 (m, 2H), 6.63-6.60 (m, 1H), 3.35 (td, $J_1 = 4 \text{ Hz}$, $J_2 = 9.5 \text{ Hz}$, 1H), 3.12-3.05 (m, 1H), 2.13-2.04 (m, 2H), 1.80-1.66 (m, 2H), 1.43-1.33 (m, 1H), 1.33-1.26 (m, 2H), 1.11-1.01 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 149.1, 130.7, 123.4, 121.2, 117.0, 113.2, 74.4, 60.2, 33.4, 31.6, 25.0, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 16.03 min, τ_{R} (major) = 17.4 min, 54% ee.



2-((4-bromophenyl) amino) cyclohexan-1-ol (11f): 98% yield, white crystal; mp: 75-77° C; **IR (ATR):** ν_{max} 3375, 2931, 2858, 1594, 1514, 1482, 1280, 1260, 766, 755, 505 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 7.23 (dt, $J_1 = 3.5 \text{ Hz}$, $J_2 = 10 \text{ Hz}$, 2H), 6.57 (dt, $J_1 = 3.5 \text{ Hz}$, $J_2 = 10 \text{ Hz}$, 2H), 3.36-3.30 (m, 1H), 3.08-3.01 (m, 1H), 2.11-2.02 (m, 2H), 1.78-1.66 (m, 2H), 1.42-1.32 (m, 1H), 1.31-1.24 (m, 1H), 1.08-0.97 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3) :** δ 145.8, 132.0, 116.0, 110.0, 74.4, 60.3, 33.4, 31.4, 24.9, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (major) = 12.3 min, τ_{R} (minor) = 16.2 min, 52% ee.

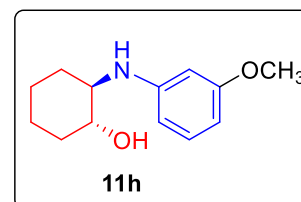


2-((2-methoxyphenyl) amino) cyclohexan-1-ol (11g): 85% yield, Greyish blue solid ; mp: 70-72° C; **IR (ATR):** ν_{max} 3383, 2926, 1600, 1521, 1451, 1345, 1224, 1048, 725 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3) :** δ 6.88 (td, $J_1 = 1 \text{ Hz}$, $J_2 = 7.5 \text{ Hz}$ 1H), 6.82-6.78 (m, 2H), 6.72 (td, $J_1 = 1.5$

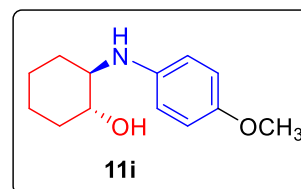


Hz, $J_2 = 7.5$ Hz, 1H), 3.84 (s, 1H), 3.46-3.40 (m, 1H), 3.18-3.13 (m, 1H), 2.17-2.07 (m, 2H), 1.83-1.69 (m, 2H), 1.47-1.38 (m, 1H), 1.38-1.26 (m, 1H), 1.14-1.04 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 147.5, 137.5, 121.3, 117.3, 111.5, 109.8, 74.4, 59.6, 55.4, 33.2, 31.5, 25.0, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_R (minor) = 7.8 min, τ_R (major) = 9.3 min, 16% ee.

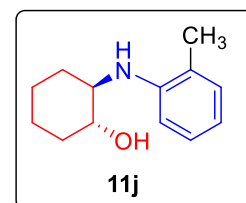
2-((3-methoxyphenyl)amino)cyclohexan-1-ol (11h): 82% yield, brown oil, IR (ATR): ν_{max} 3384, 2931, 2857, 1612, 1463, 1300, 1263, 1233, 1038, 829 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) : δ 7.07 (t, $J = 8$ Hz, 1H), 6.32-6.28 (m, 2H), 6.28-6.26 (m, 1H), 3.75 (s, 3H), 3.33-3.26 (m, 1H), 3.11-3.04 (m, 1H), 2.13-2.02 (m, 2H), 1.77-1.62 (m, 2H), 1.41-1.30 (m, 1H), 1.30-1.22 (m, 2H), 1.04-0.94 (m, 1H), ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 160.6, 149.2, 129.8, 107.0, 102.8, 100.0, 73.9, 59.6, 54.8, 33.2, 31.3, 24.6, 24.1 ppm. The ee was determined by HPLC using Chiralpak AS-H [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_R (major) = 13.9 min, τ_R (minor) = 16.2 min, 42% ee.



2-((4-methoxyphenyl) amino) cyclohexan-1-ol (11i): 87% yield, brown oil, IR (ATR): ν_{max} 3336, 2929, 2857, 1509, 1451, 1235, 1234, 1067, 1034, 820 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) : δ 6.77 (dt, $J_1 = 3.5$ Hz, $J_2 = 10$ Hz, 2H), 6.68 (dt, $J_1 = 3.5$ Hz, $J_2 = 10$ Hz, 2H), 3.73 (s, 3H), 3.35-3.28 (m, 1H), 3.01-2.94 (m, 1H), 2.13-2.03 (m, 2H), 1.78-1.65 (m, 2H), 1.43-1.30 (m, 1H), 1.29-1.20 (m, 2H), 1.95-0.95 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 153.0, 141.5, 116.5, 114.9, 74.3, 61.7, 55.8, 33.3, 31.5, 25.1, 24.3 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_R (major) = 25.4 min, τ_R (minor) = 28.3 min, 4% ee.



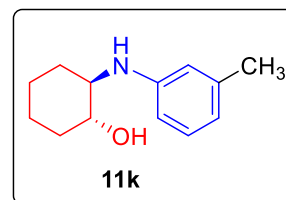
2-((2-methylphenyl) amino) cyclohexan-1-ol (11j): 82% yield, blackish blue oil, IR (ATR): ν_{max} 3400, 3016, 2929, 2856, 1605, 1508, 1448, 1315, 1258, 744 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) : δ 7.16-7.11 (m, 1H), 7.09 (d, $J = 7$ Hz, 1H), 6.82 (d, $J = 8$ Hz, 1H), 6.72 (td, $J_1 = 1$ Hz, $J_2 = 7.5$ Hz, 1H), 3.45 (td, $J_1 = 4$ Hz, $J_2 = 9.5$ Hz, 1H), 3.25-3.18 (m, 1H), 2.20-2.11 (m, 5H), 1.83-1.77 (m, 1H), 1.76-1.69 (m, 1H), 1.48-1.38 (m, 1H), 1.37-1.27 (m, 2H), 1.15-1.05 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 145.5, 130.6, 127.2, 123.4, 118.2, 112.0, 74.6, 60.2, 33.4, 31.8, 25.1, 24.4,



17.8 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (major) = 10.6 min, τ_R (minor) = 11.1 min, 2% ee.

2-(m-tolylamino)cyclohexan-1-ol (11k): 82% yield, light brown oil; **IR**

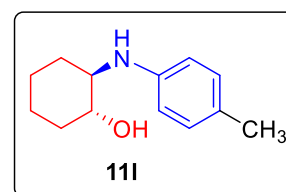
(ATR): $\nu_{\max}$ 3352, 3042, 2929, 2857, 1604, 1588, 1489, 1327, 767, 692 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** : δ 7.08 (t, J_1 = 7.5 Hz, 1H), 6.63-6.56 (m, 3H), 3.38 (td, J_1 = 4.5 Hz, J_2 = 9.5 Hz, 1H), 3.17-3.10 (m, 1H), 2.28 (s, 3H), 2.16-2.03 (m, 2H), 1.82-1.67 (m, 2H), 1.44-1.34 (m, 1H), 1.34-



1.27 (m, 1H), 1.14-1.03 (m, 1H) ppm; **¹³C NMR (125 MHz, CDCl₃)** : δ 147.1, 139.4, 129.4, 120.1, 115.9, 112.3, 74.4, 60.9, 33.3, 31.5, 25.1, 24.4, 21.7 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (minor) = 7.3 min, τ_R (major) = 7.7 min, 20% ee.

2-((4-methylphenyl) amino) cyclohexan-1-ol (11l): 78% yield, light

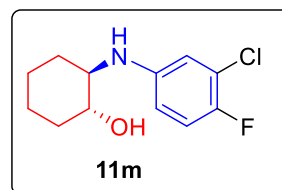
brown oil, **IR (ATR):** $\nu_{\max}$ 3383, 2929, 2858, 1617, 1518, 1450, 1302, 807 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** : δ 7.01 (d, J = 8 Hz, 2H), 6.69 (d, J = 8.5 Hz, 2H), 3.37 (td, J_1 = 4.5 Hz, J_2 = 9.5 Hz, 1H), 3.12-3.05 (m,



1H), 2.25 (s, 3H), 2.14-2.05 (m, 2H), 1.80-1.66 (m, 2H), 1.45-1.34 (m, 1H), 1.34-1.26 (m, 2H), 1.11-1.01 (m, 1H) ppm; **¹³C NMR (125 MHz, CDCl₃)** : δ 144.8, 130.0, 128.6, 115.4, 74.4, 61.3, 33.3, 31.4, 25.2, 24.4, 20.5 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (major) = 7.03 min, τ_R (minor) = 8.8 min, 10% ee.

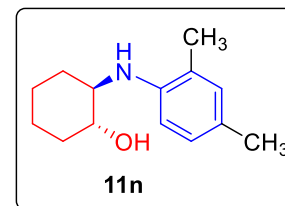
2-((3-chloro-4-fluorophenyl)amino)cyclohexan-1-ol (11m): 85%

yield, greenish oil, **IR (ATR):** $\nu_{\max}$ 3357, 2933, 2859, 1500, 1450, 1064, 1047, 804, 777, 732 cm⁻¹; **¹H NMR (500 MHz, CDCl₃)** : δ 6.88 (t, J = 9 Hz, 1H), 6.66 (dd, J_1 = 3 Hz, J_2 = 6 Hz, 1H), 6.50-6.45 (m,

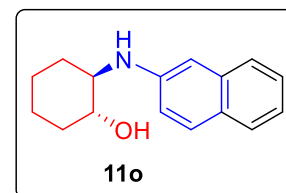


1H), 3.32-3.25 (m, 1H), 2.97-2.90 (m, 1H), 2.06-1.98 (m, 2H), 1.75-1.61 (m, 2H), 1.38-1.27 (m, 1H), 1.27-1.19 (m, 2H), 1.04-0.92 (m, 1H) ppm; **¹³C NMR (125 MHz, CDCl₃)** : δ 151.2 (d, J = 237.0 Hz), 144.8 (d, J = 1.8 Hz), 120.9 (d, J = 18.1 Hz), 116.7 (d, J = 21.5 Hz), 115.2, 113.6 (d, J = 5.8 Hz), 74.2, 60.6, 33.4, 31.3, 24.7, 24.2 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (major) = 6.9 min, τ_R (minor) = 7.5 min, 22% ee.

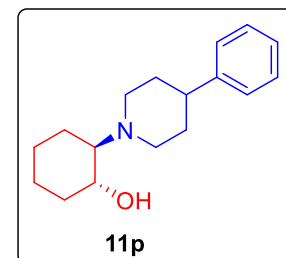
2-((2,4-dimethylphenyl)amino)cyclohexan-1-ol (11n): 75% yield, light green oil, **IR (ATR):** $\nu_{\max}$ 3398, 2929, 2856, 1619, 1510, 1266, 1039, 803, 731 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 6.99-6.92 (m, 2H), 6.74 (d, $J = 8$, 1H), 3.43 (td, $J_1 = 4.5$ Hz, $J_2 = 9.5$ Hz, 1H), 3.21-3.14 (m, 1H), 2.27 (s, 3H), 2.17-2.12 (m, 5H), 1.85-1.70 (m, 2H), 1.50-1.32 (m, 3H), 1.13-1.03 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 143.3, 131.3, 127.5, 127.3, 123.5, 112.3, 74.5, 60.4, 33.3, 31.8, 25.1, 24.4, 20.4, 17.7 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (95 : 5)]; flow rate 1.0 mL min^{-1} ; τ_R (major) = 8.7 min, τ_R (minor) = 9.4 min, 6% ee.



2-(naphthalen-1-ylamino)cyclohexan-1-ol (11o): 80% yield, brown solid, mp: 102-105 °C ;**IR (ATR):** $\nu_{\max}$ 3469, 3327, 2927, 2858, 1582, 1528, 1449, 1274, 790, 768, cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.89-7.85 (m, 1H), 7.84-7.80 (m, 1H), 7.51-7.44 (m, 2H), 7.39-7.28 (m, 2H), 6.82 (d, $J = 7.5$, 1H), 3.58-3.50 (m, 1H), 3.41-3.34 (m, 1H), 2.30-2.22 (m, 1H), 2.21-2.12 (m, 1H), 1.86-1.70 (m, 2H), 1.53-1.30 (m, 3H), 1.18-1.08 (m, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 142.8, 134.6, 128.9, 126.5, 125.9, 125.0, 124.3, 120.1, 118.5, 106.6, 74.6, 60.0, 33.5, 31.3, 25.0, 24.4 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (95 : 5)]; flow rate 1.0 mL min^{-1} ; τ_R (major) = 28.3 min, τ_R (minor) = 39.9 min, 14% ee.

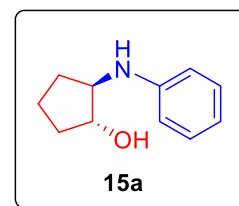


2-(4-phenylpiperidin-1-yl)cyclohexan-1-ol (11a): 80% yield, white solid; **IR (ATR):** $\nu_{\max}$ 3345, 2959, 2923, 2853, 1600, 1468, 1275, 1269, 760, 750 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.33-7.27 (m, 2H), 7.25-7.17 (m, 3H), 3.41 (dt, $J_1 = 4$ Hz, $J_2 = 9.5$ Hz, 1H), 2.95 (d, $J = 11$ Hz, 1H), 2.80-2.70 (m, 2H), 2.50 (tt, $J_1 = 4$ Hz, $J_2 = 12$, 1H), 2.30-2.20 (m, 2H), 2.17- 2.12 (m, 1H), 1.92-1.76 (m, 5H), 1.75-1.61 (m, 2H), 1.25-1.16 (m, 4H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 146.4, 128.5, 127.0, 126.3, 70.8, 68.8, 53.6, 45.6, 43.1, 34.5, 34.1, 33.5, 25.8, 24.3, 22.4 ppm. The ee was determined by HPLC using Chiralpak AS-H [hexane/iPrOH (97 : 3)]; flow rate 0.5 mL min^{-1} ; τ_R (major) = 19.1 min, τ_R (minor) = 20.8 min, 16% ee.

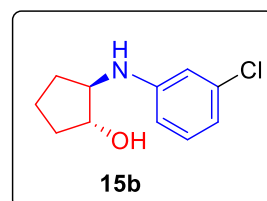


2-(phenylamino)cyclopentan-1-ol (15a): 85% yield, purple oil; **IR (ATR):** $\nu_{\max}$ 3345, 2956, 2872, 1601, 1498, 1316, 978, 748, 693 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.21-7.16 (m, 2H), 6.77-6.68 (m, 3H), 4.10 (q, $J = 4.5$ Hz, 1H), 3.64-3.59 (m, 1H), 2.30-2.22 (m, 1H), 2.04-1.95 (m,

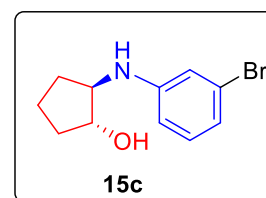
1H), 1.83-1.69 (m, 2H), 1.88-1.70 (m, 1H), 1.48-1.39 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 147.3, 129.5, 118.2, 113.9, 76.9, 62.7, 33.0, 31.1, 21.1 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min $^{-1}$; τ_{R} (major) = 9.8 min, τ_{R} (minor) = 11.3 min, 2% ee.



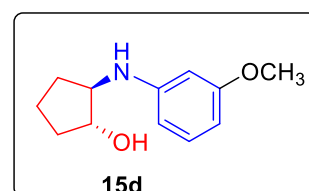
2-((3-chlorophenyl) amino) cyclopentan-1-ol (15b): 82% yield, dark maroon oil, IR (ATR): ν_{max} 3347, 2929, 2874, 1596, 1500, 1481, 1326, 987, 762, 682 cm $^{-1}$; ^1H NMR (500 MHz, CDCl_3) : δ 7.07 (t, J = 7.5 Hz, 1H), 6.69-6.64 (m, 2H), 6.53 (ddd, J_1 = 0.5, J_2 = 2, J_3 = 8 Hz, 1H), 4.05 (q, J = 4.5, 1H), 3.60-3.54 (m, 1H), 2.31-2.22 (m, 1H), 2.02-1.94 (m, 1H), 1.88-1.69 (m, 2H), 1.69-1.60 (m, 1H), 1.44-1.36 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 148.8, 135.2, 130.4, 117.6, 113.2, 111.9, 76.9, 62.2, 33.1, 31.2, 21.1 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min $^{-1}$; τ_{R} (major) = 10.4 min, τ_{R} (minor) = 11.4 min, 2% ee.



2-((3-bromophenyl) amino) cyclopentan-1-ol (15c): 79% yield, dark red oil, IR (ATR): ν_{max} 3337, 2926, 2871, 1594, 1498, 1479, 1325, 984, 760, 682 cm $^{-1}$; ^1H NMR (500 MHz, CDCl_3) : δ 7.01 (t, J = 8 Hz, 1H), 6.85-6.82 (m, 2H), 6.66 (ddd, J_1 = 1 Hz, J_2 = 2.5, J_3 = 7.5 Hz, 1H), 4.07 (q, J = 4.5 Hz, 1H), 3.59-3.54 (m, 1H), 2.30-2.21 (m, 1H), 2.03-1.94 (m, 1H), 1.88-1.70 (m, 2H), 1.69-1.60 (m, 1H), 1.46-1.36 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 148.7, 130.7, 123.4, 120.9, 116.4, 112.5, 76.9, 62.5, 33.1, 31.1, 21.1 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min $^{-1}$; τ_{R} (major) = 10.1 min, τ_{R} (minor) = 11.2 min, 2% ee.



2-((3-methoxyphenyl) amino) cyclopentan-1-ol (15d): 75% yield, dark brown oil, IR (ATR): ν_{max} 3353, 2955, 2835, 1600, 1495, 1261, 1209, 1160, 1038, 758 cm $^{-1}$; ^1H NMR (500 MHz, CDCl_3) : δ 7.09 (t, J = 8 Hz, 1H), 6.34-6.28 (m, 3H), 4.10 (q, J = 4.5 Hz, 1H), 3.77 (s, 3H), 3.62-3. (m, 1H), 2.29-2.21 (m, 1H), 2.03-1.95 (m, 1H), 1.88-1.69 (m, 2H), 1.68-1.60 (m, 1H), 1.48-1.40 (m, 1H) ppm; ^{13}C NMR (125 MHz, CDCl_3) : δ 161.0, 148.5, 130.2, 107.1, 103.6, 100.1, 76.9, 62.9, 55.3, 33.0, 31.1, 21.1 ppm. The ee was determined by HPLC using Chiralpak IB



[hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (major) = 14.6 min, τ_R (minor) = 17.8 min, 2% ee.

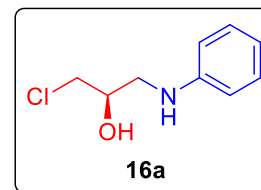
1-chloro-3-(phenylamino)propan-2-ol (16a): 87% yield, dark purple oil,

IR (ATR): $\nu_{\max}$ 3309, 2920, 1599, 1584, 1504, 1324, 748, 692 cm⁻¹; **¹H**

NMR (500 MHz, CDCl₃) : δ 7.24-7.19 (m, 2H), 6.84-6.80 (m, 1H), 6.77 (dd, J_1 = 1 Hz, J_2 = 8.5 Hz, 2H), 4.164.07 (m, 1H), 3.65 (qd, J_1 = 1 Hz, J_2

= 11.5 Hz, 2H), 3.40 (dd, J_1 = 4.5 Hz, J_2 = 13 Hz, 1H), 3.26 (dd, J_1 = 7.5 Hz, J_2 = 13 Hz, 1H) ppm;

¹³C NMR (125 MHz, CDCl₃) : δ 146.6, 129.6, 119.6, 114.5, 69.6, 48.3, 47.6 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (minor) = 19.4 min, τ_R (major) = 20.4 min, 8% ee.

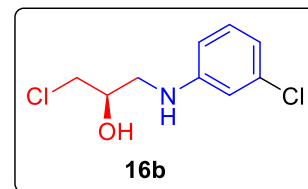


1-chloro-3-((3-chlorophenyl)amino)propan-2-ol (16b): 90% yield, greenish brown oil, **IR (ATR):** $\nu_{\max}$ 3332, 2917, 1596, 1504, 1484,

1250, 841, 763, 681 cm⁻¹; **¹H NMR (500 MHz, CDCl₃) :** δ 7.09 (t, J = 8 Hz, 1H), 6.71 (dd, J_1 = 1 Hz, J_2 = 8 Hz, 1H), 6.63 (s, 1H), 6.52 (dd,

J_1 = 1.5 Hz, J_2 = 8 Hz, 1H), 4.11-4.03 (m, 1H), 3.66 (ddd, J_1 = 4.5 Hz, J_2 = 11.5 Hz, J_3 = 28 Hz, 2H), 3.36 (dd, J_1 = 4 Hz, J_2 = 13 Hz, 1H), 3.22 (dd, J_1 = 7 Hz, J_2 = 13 Hz, 1H) ppm; **¹³C NMR**

(125 MHz, CDCl₃) : δ 149.0, 135.3, 130.5, 118.2, 113.1, 111.8, 69.9, 47.8, 47.0 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90: 10)]; flow rate 1.0 mL min⁻¹; τ_R (minor) = 36.8 min, τ_R (major) = 40.1 min, 28% ee.

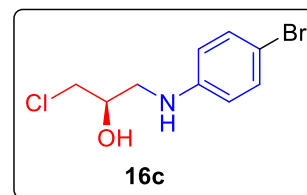


1-((4-bromophenyl)amino)-3-chloropropan-2-ol (16c): 88% yield, brown oil; **IR (ATR):** $\nu_{\max}$ 2916, 2850, 1730, 1594, 1492, 1261, 1179,

812, 750 cm⁻¹; **¹H NMR (500 MHz, CDCl₃) :** δ 7.29 (dt, J_1 = 3 Hz, J_2 = 10 Hz, 2H), 6.63 (dt, J_1 = 3 Hz, J_2 = 9.5 Hz, 2H), 4.14-4.08 (m, 1H),

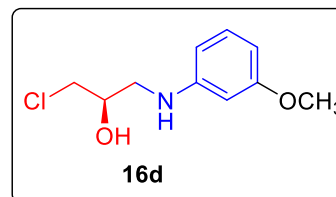
3.65 (qd, J_1 = 5 Hz, J_2 = 11.5 Hz, 2H), 3.36 (dd, J_1 = 4 Hz, J_2 = 13 Hz, 1H), 3.22 (dd, J_1 = 7.5 Hz, J_2 = 13.5 Hz, 1H) ppm; **¹³C NMR (125 MHz, CDCl₃) :** δ 145.6, 132.2, 115.8, 111.2, 69.5, 48.0,

47.4 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min⁻¹; τ_R (major) = 10.9 min, τ_R (minor) = 11.5 min, 26% ee.



1-((3-methoxyphenyl)amino)-3-chloropropan-2-ol (16d): 80%

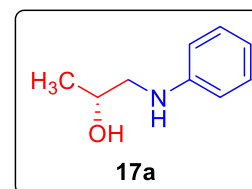
yield, green oil, **IR (ATR):** $\nu_{\max}$ 3308, 2919, 1600, 1584, 1504, 1324, 1171, 1074, 748, 692 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.11 (t, $J = 8$ Hz, 1H), 6.40-6.27 (m, 3H), 4.16-4.08 (m, 1H), 3.77



(s, 3H), 3.65 (qd, $J_1 = 5$ Hz, $J_2 = 11.5$ Hz, 2H), 3.39 (dd, $J_1 = 4.5$ Hz, $J_2 = 13.5$ Hz, 1H), 3.25 (dd, $J_1 = 7.5$ Hz, $J_2 = 13.5$ Hz, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 161.0, 130.4, 107.2, 104.5, 100.3, 69.7, 55.3, 47.7, 29.8 ppm. The ee was determined by HPLC using Chiralpak IB [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (major) = 21.0 min, τ_{R} (minor) = 24.1 min, 18% ee.

1-(phenylamino)propan-2-ol (17a): 88% yield, reddish brown oil, **IR**

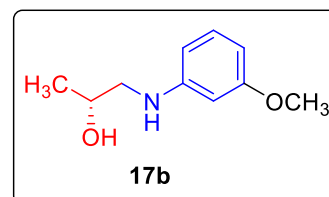
(ATR): $\nu_{\max}$ 3341, 3052, 2969, 2929, 1602, 1499, 1460, 1317, 1072, cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.25 (t, $J = 7$ Hz, 2H), 6.80 (t, $J = 6.5$ Hz, 1H), 6.70 (d, $J = 8$ Hz, 2H), 4.04 (s, 1H), 3.29-3.21 (m, 1H), 3.07-2.98



(m, 1H), 1.29 (d, $J = 6$ Hz, 1H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 148.2, 129.3, 117.9, 113.4, 66.4, 51.7, 20.9 ppm. The ee was determined by HPLC using phenomenex Amylose-1 [hexane/iPrOH (80 : 20)]; flow rate 1.0 mL min^{-1} ; τ_{R} (minor) = 8.9 min, τ_{R} (major) = 9.9 min, 38% ee.

1-((3-methoxyphenyl)amino)propan-2-ol (17b): 75% yield, dark

brown solid; **IR (ATR):** $\nu_{\max}$ 3341, 2958, 2923, 2853, 1611, 1461, 1278, 1261, 764, 750 cm^{-1} ; **^1H NMR (500 MHz, CDCl_3):** δ 7.08 (t, $J = 8.5$ Hz, 1H), 6.30 (dd, $J_1 = 2$ Hz, $J_2 = 8$ Hz, 1H), 6.25 (dd, $J_1 =$



1.5 Hz, $J_2 = 8$ Hz, 1H), 6.20 (t, $J = 2$ Hz, 1H), 4.03-3.94 (m, 1H), 3.75 (s, 3H), 3.17 (dd, $J_1 = 3$ Hz, $J_2 = 13$ Hz, 1H), 2.95 (dd, $J_1 = 8.5$ Hz, $J_2 = 13$ Hz, 1H), 1.22 (d, $J = 6.5$ Hz, 3H) ppm; **^{13}C NMR (125 MHz, CDCl_3):** δ 160.8, 149.7, 130.1, 106.4, 103.4, 99.3, 66.3, 55.1, 51.6, 20.8 ppm. The ee was determined by HPLC using Chiralpak IA [hexane/iPrOH (90 : 10)]; flow rate 1.0 mL min^{-1} ; τ_{R} (major) = 17.7 min, τ_{R} (minor) = 19.8 min, 46% ee.

1.4. EDS Profile of Catalysts:

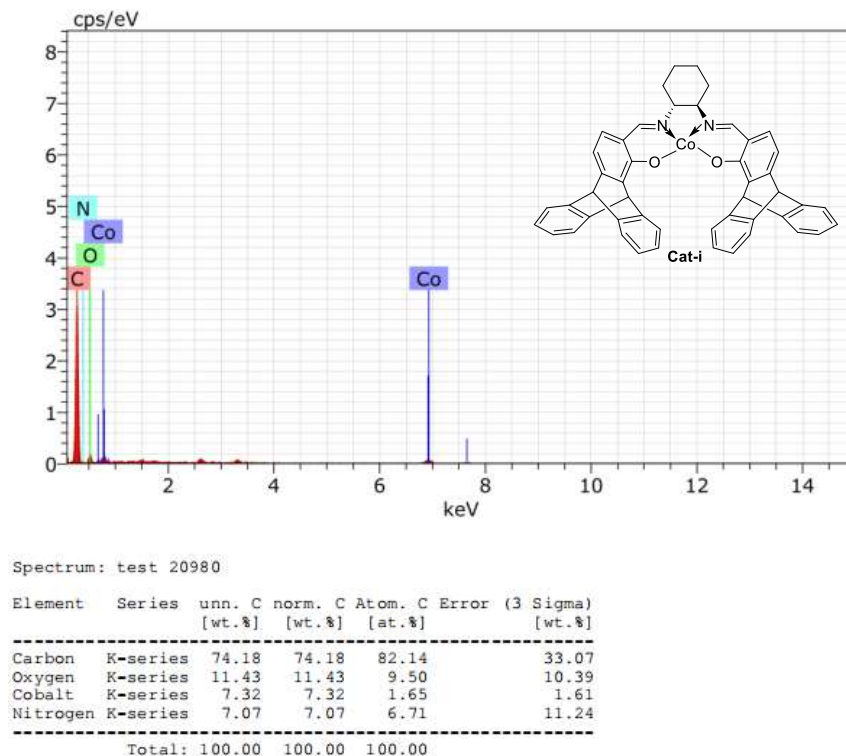


Figure S1: EDS spectrum of TpCo(II)(Cat-i).

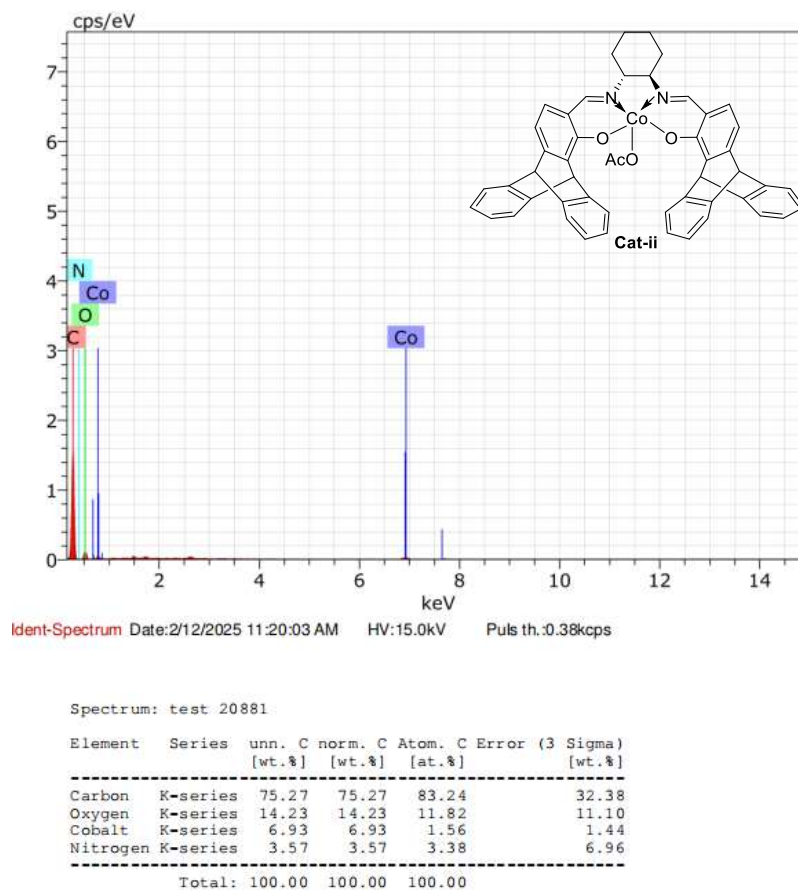


Figure S2: EDS spectrum of TpCo(III)-OAc (Cat-ii)

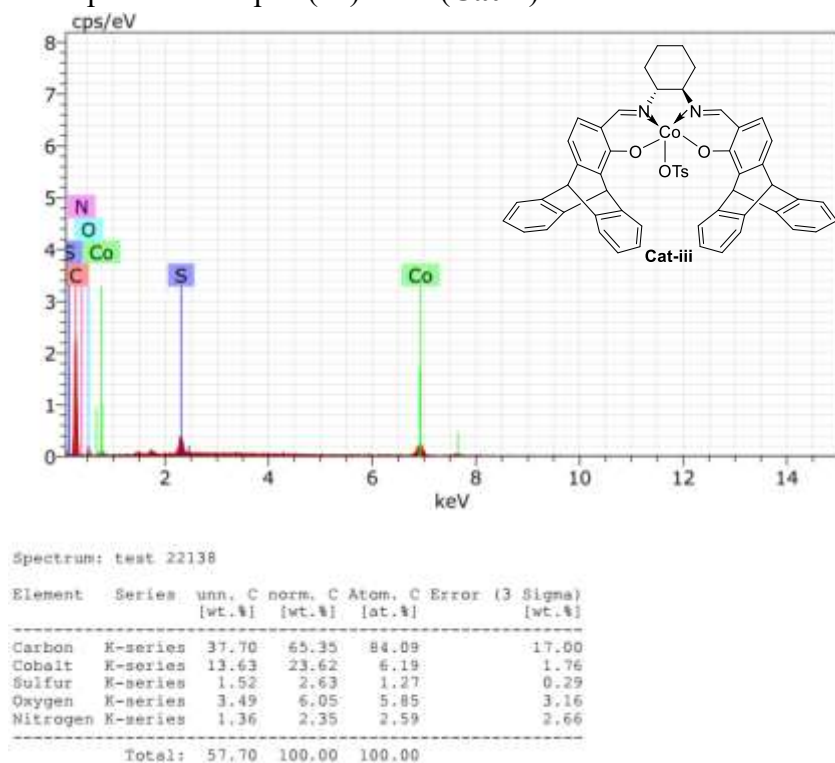


Figure S3: EDS spectrum of TpCo(III)-OTs (Cat-iii).

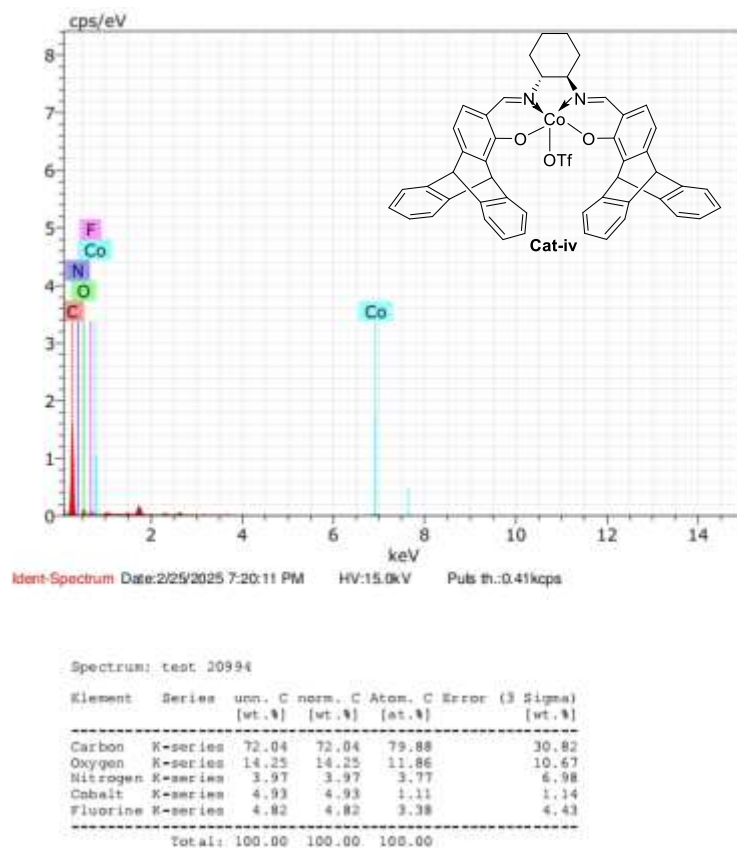
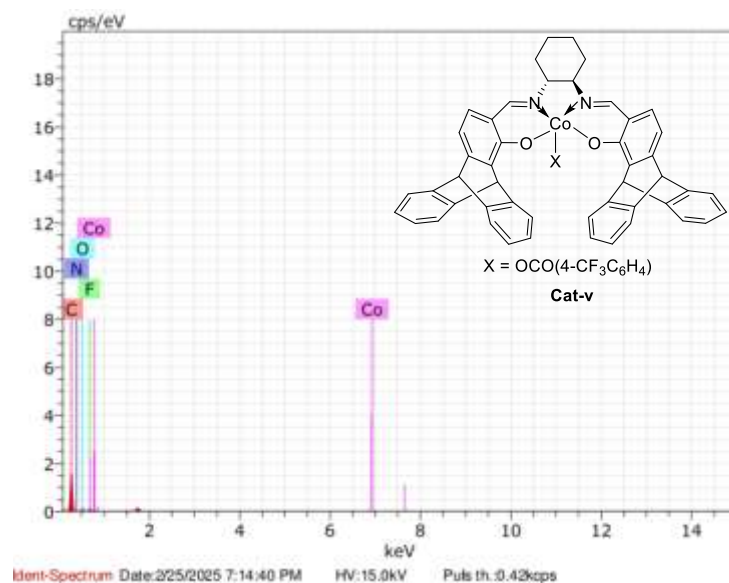


Figure S4: EDS spectrum of TpCo(III)-OTf (Cat-iv).



Spectrum: test_20992

Element	Series	unm. C [wt.%]	norm. C [wt.%]	Atom. C [at.%]	Error (3 Sigma) [wt.%]
Carbon	K-series	69.49	69.49	78.74	29.57
Fluorine	K-series	6.68	6.68	4.79	5.23
Nitrogen	K-series	3.89	3.89	3.78	6.48
Oxygen	K-series	13.06	13.06	11.11	9.59
Cobalt	K-series	6.87	6.87	1.59	1.32
Total:		100.00	100.00	100.00	

Figure S5: EDS spectrum of TpCo(III)-OCO(4-CF₃C₆H₄) (**Cat-v**).

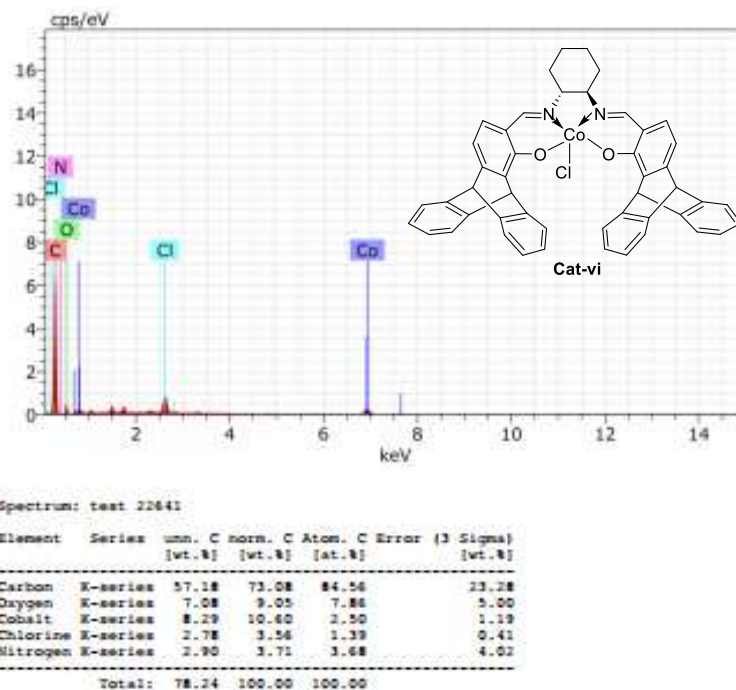


Figure S6: EDS spectrum of TpCo(III)-Cl (**Cat-vi**).

1.5. PXRD graphs of ligand 8 and cobalt catalysts.

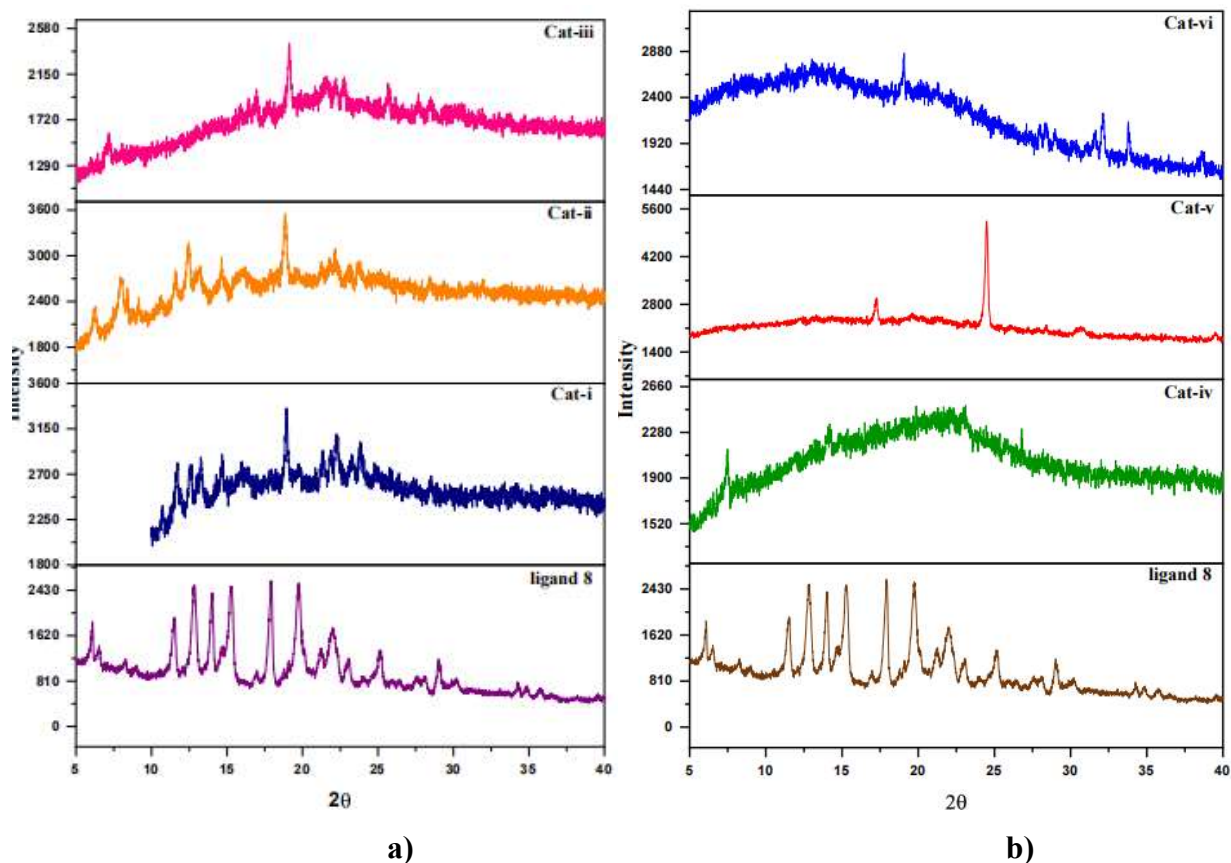


Figure S7: Comparison of PXRD graphs of ligand 8 **a)** with Cat-i - Cat-iii, **b)** Cat-iv - Cat-vi.

PXRD analysis was carried out for ligand 8 and all catalysts (Cat-i to Cat-Vi) to understand their structural characteristics. Ligand 8 exhibits a crystalline pattern, with several well-defined diffraction peaks in the 10-30° range. Prominent reflections at 11.5°, 12.9°, 13.9°, 15.3°, 17.8°, 19.7°, and 21.9° clearly indicate its crystalline nature. In Cat-i, these characteristic peaks become less intense and slightly shifted to 11.7°, 12.6°, 13.3°, 14.6°, and 18.9°, reflecting a transition to a semi-crystalline phase upon coordination with cobalt, thereby confirming successful complex formation. The PXRD profiles of Cat-ii to Cat-vi also display reduced peak intensity and partial broadening, suggesting semi-crystalline to moderately amorphous structural features across the series.

1.6. FESEM images of ligand **8**, Cat-i and Cat-ii:

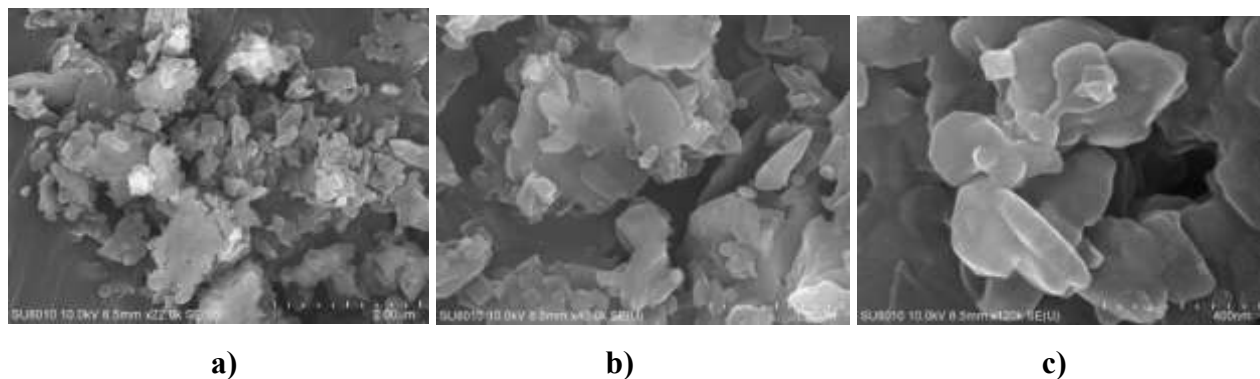


Figure S8: FESEM images of triptycene ligand **8** at different scales **a)** at 2μm, **b)** at 1 μm, **c)** at 400nm.

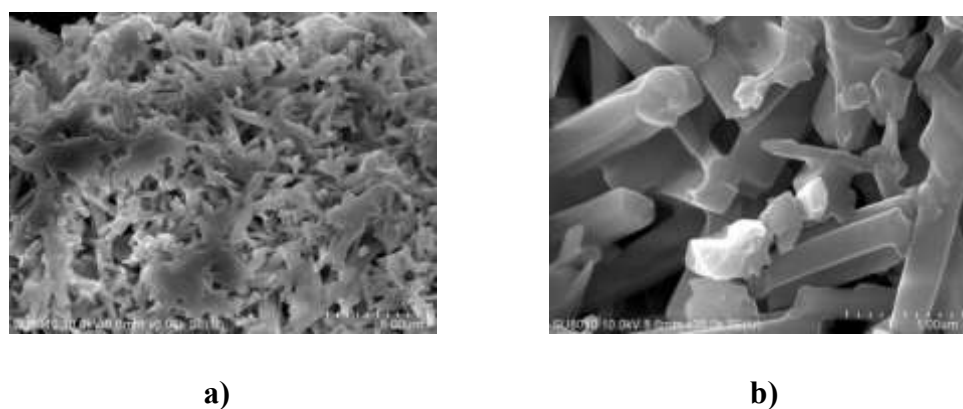


Figure S9: FESEM images of cat-i at different scales **a)** at 5μm **b)** at 1μm.

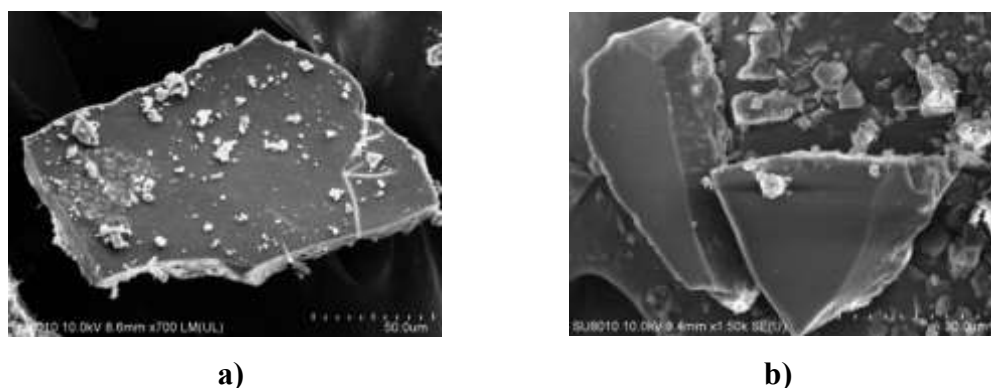
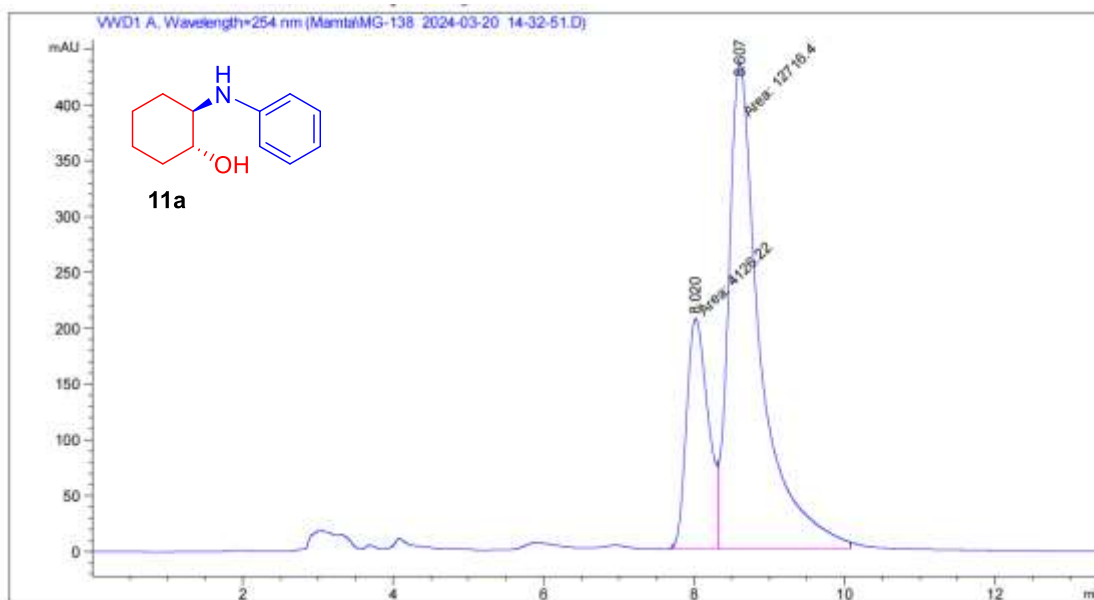


Figure S10: FESEM images of cat-ii at different scales **a)** at 50μm **b)** at 30μm.

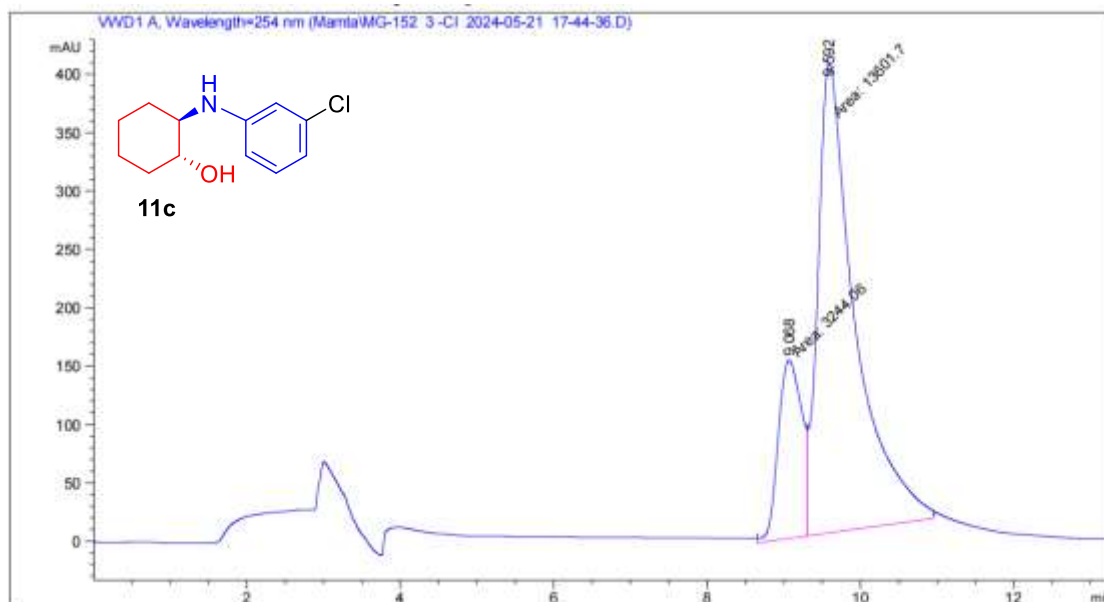
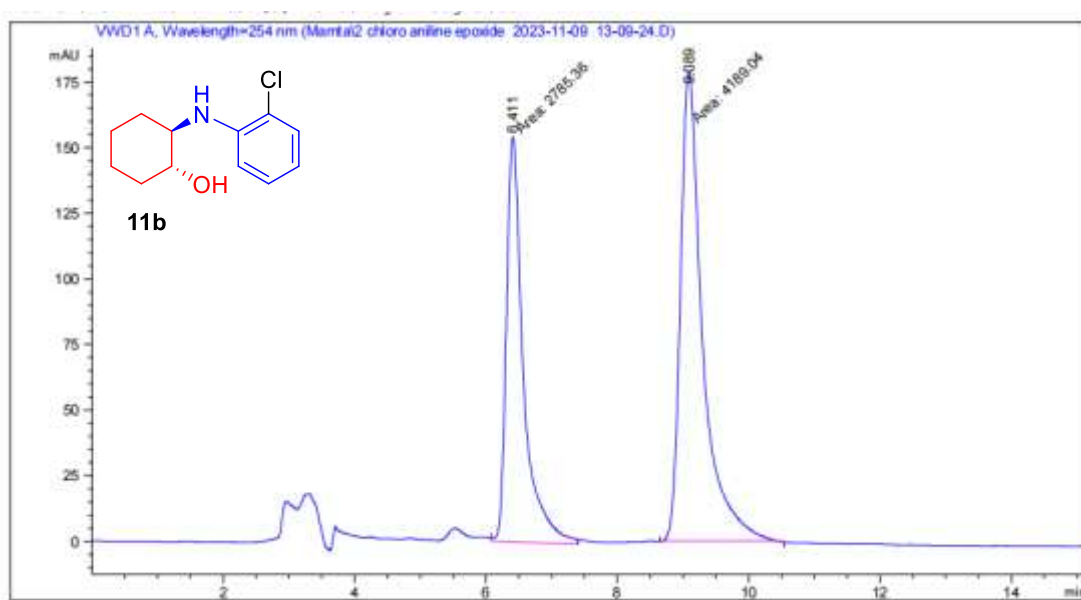
To study the structural morphology of the triptycene ligand **8** and the changes after coordination with the cobalt metal, SEM images of ligand **8**, **cat-i** and **cat-ii** were recorded and compared. The images of ligand **8** (figure S8) showed irregular, flaky particles that remain widely separated, indicating minimal interaction. In cat-i, the particles become closer and formed compact clusters (figure S9). The surface becomes denser, and the particles are no longer isolated, suggesting that cobalt coordination increased

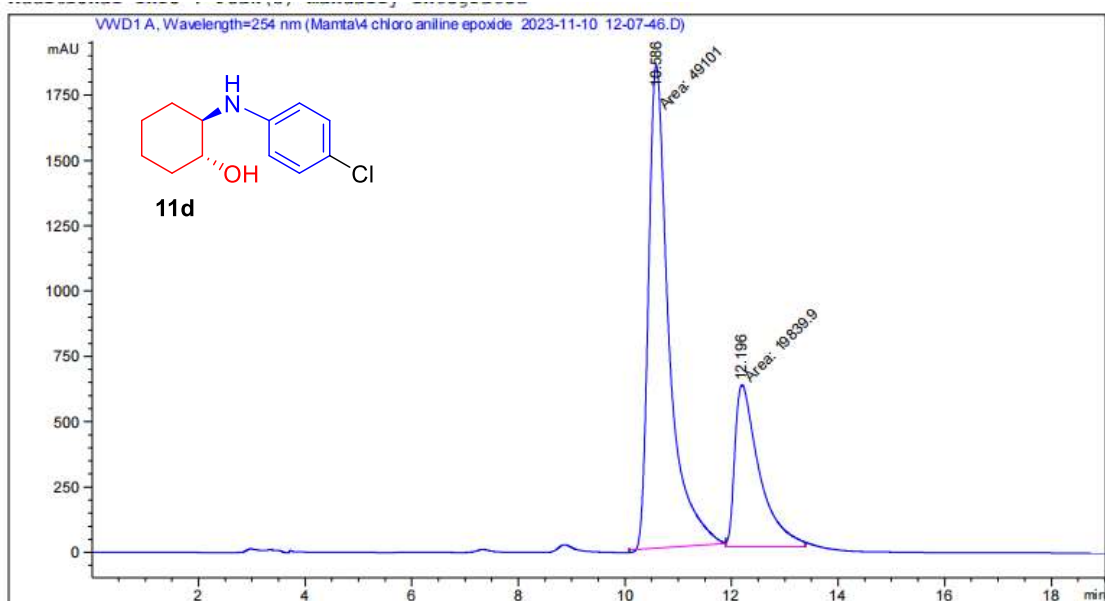
the attraction between the molecules and lead to aggregated structures. After introducing the axial ligand to obtain cat-ii, material developed into thin, sheet-like layers (figure S10). These sheets are broad, slightly overlapping, and give a clear indication of a more organized and continuous structure.

2. HPLC Chromatograms:

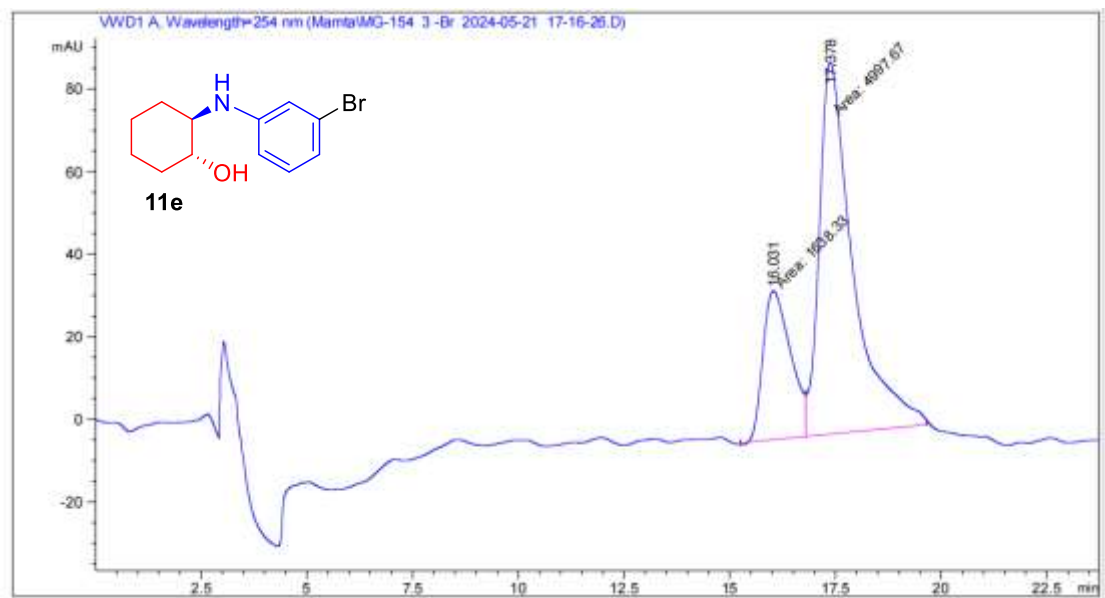


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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2	8.607	FM	0.4877	1.27164e4	434.57065	75.5013

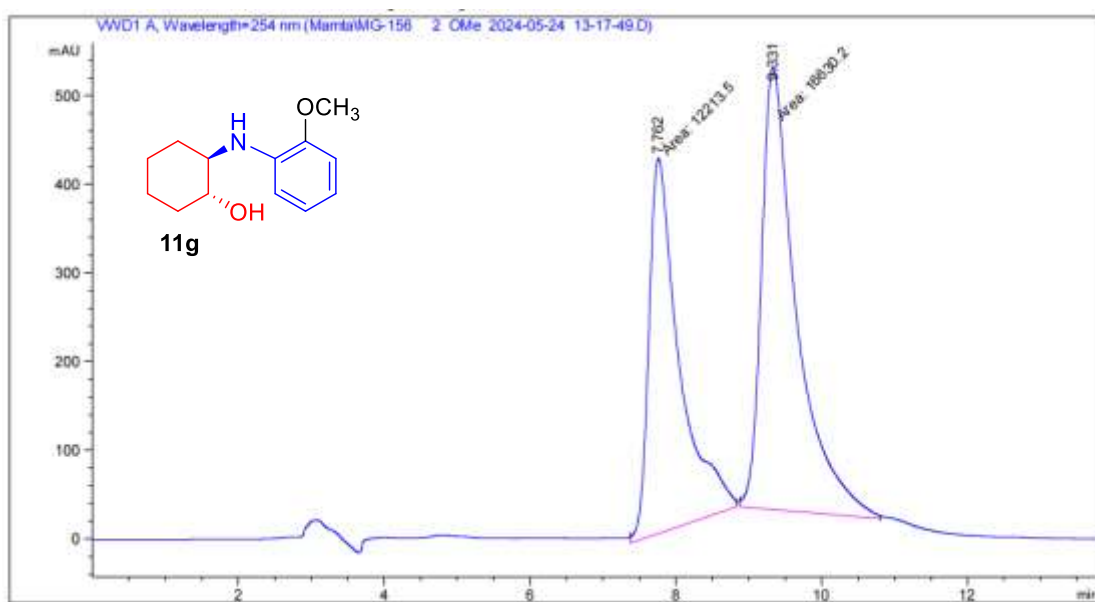
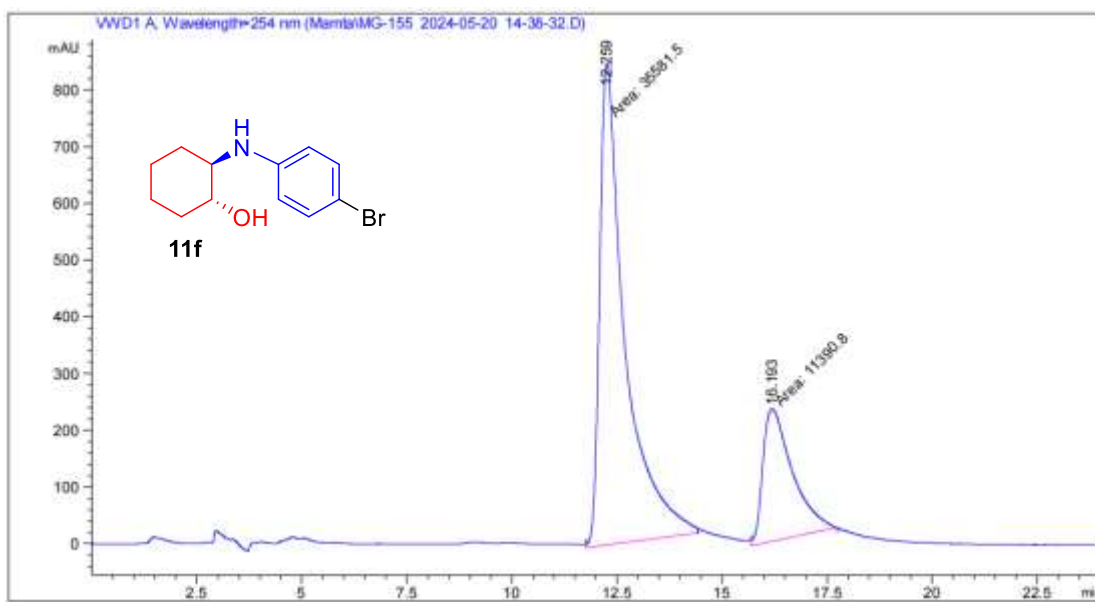


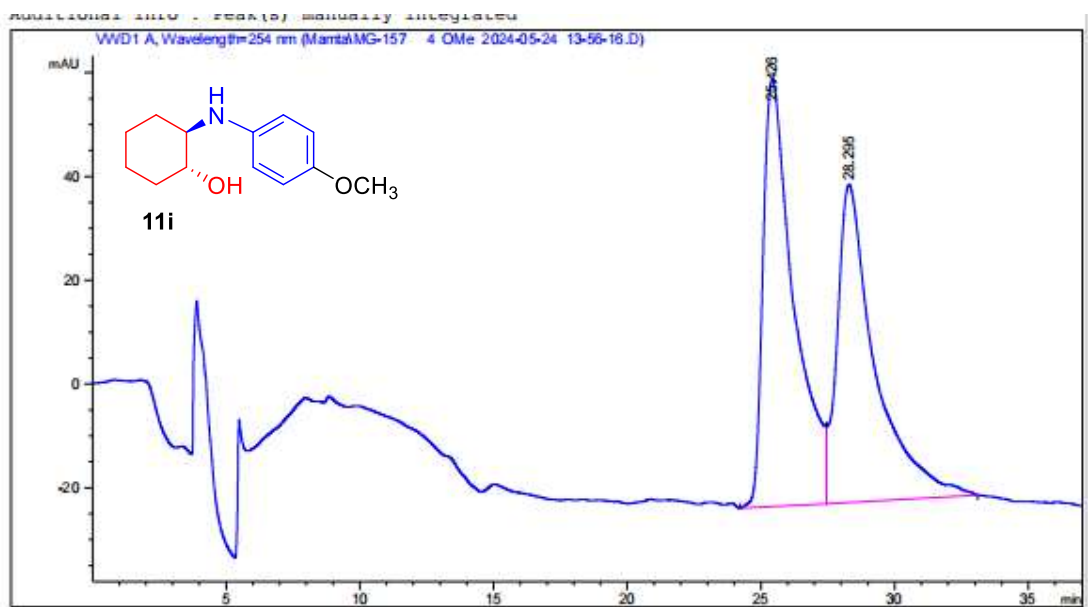
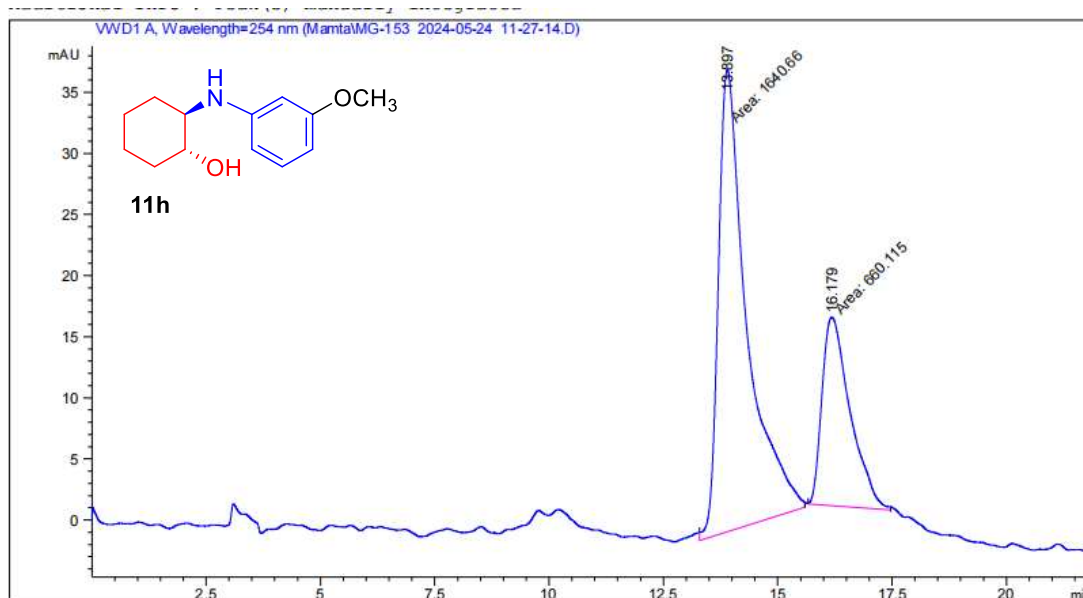


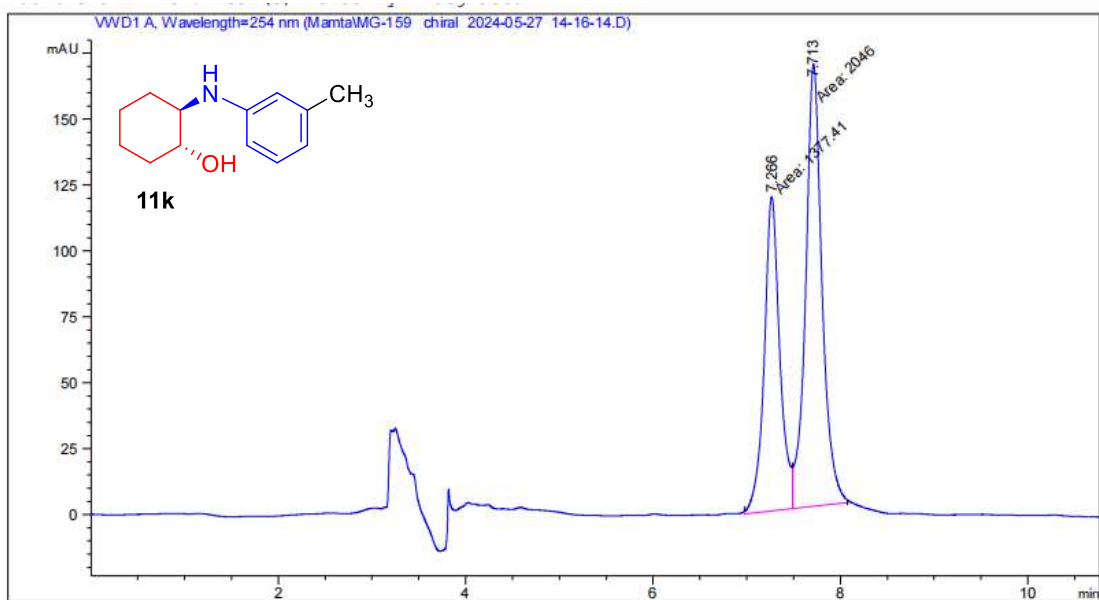
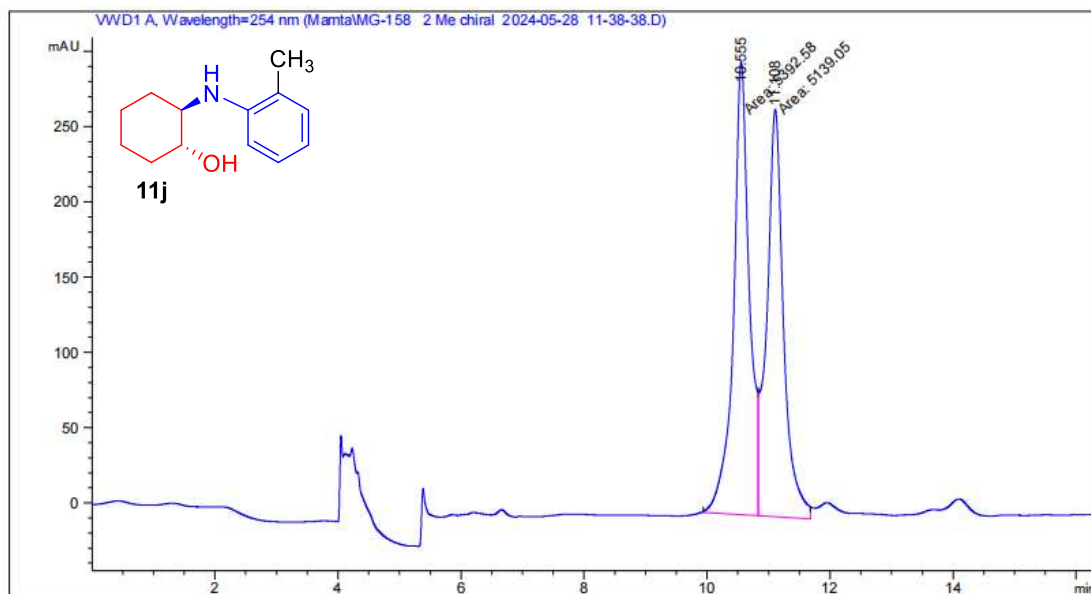
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.586	MM	0.4427	4.91010e4	1848.70093	71.2219
2	12.196	MM	0.5334	1.98399e4	619.96796	28.7781

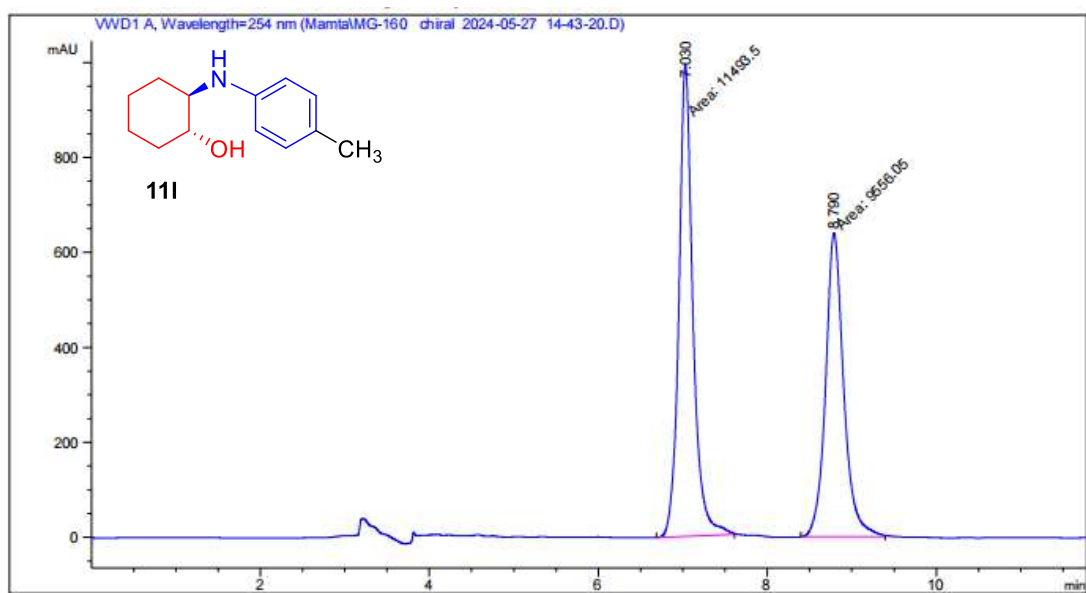


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.031	MF	0.7580	1638.32556	36.02132	24.6885
2	17.378	FM	0.9255	4997.66602	90.00153	75.3115

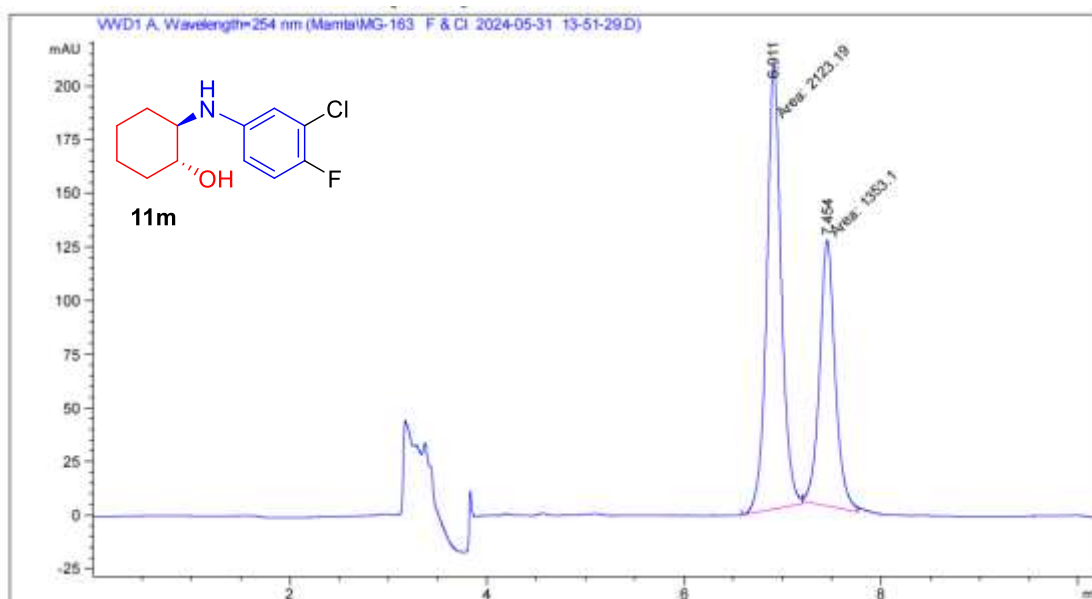




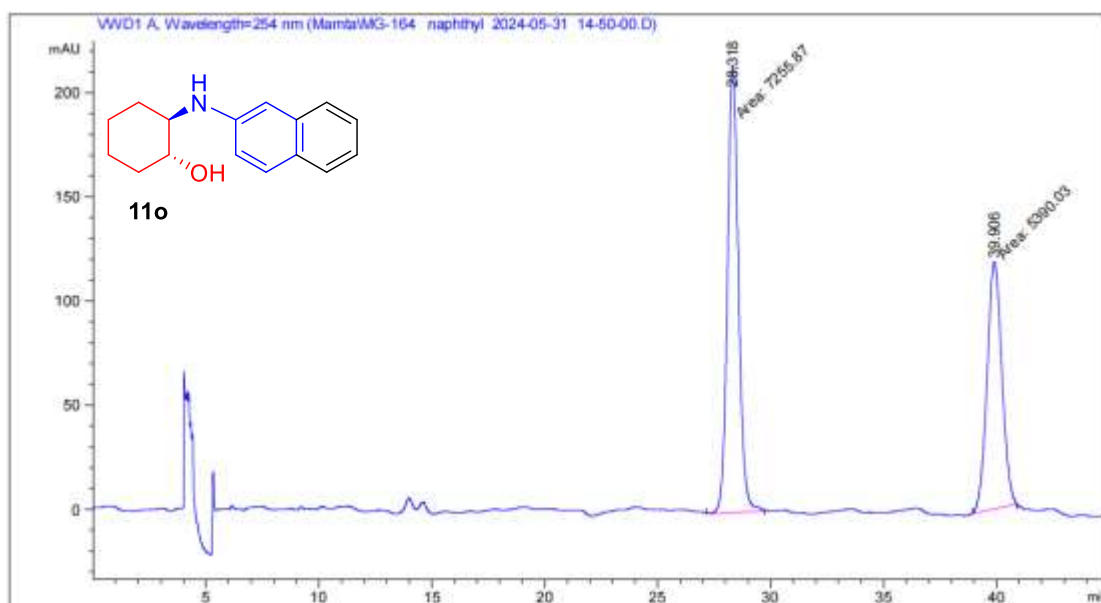
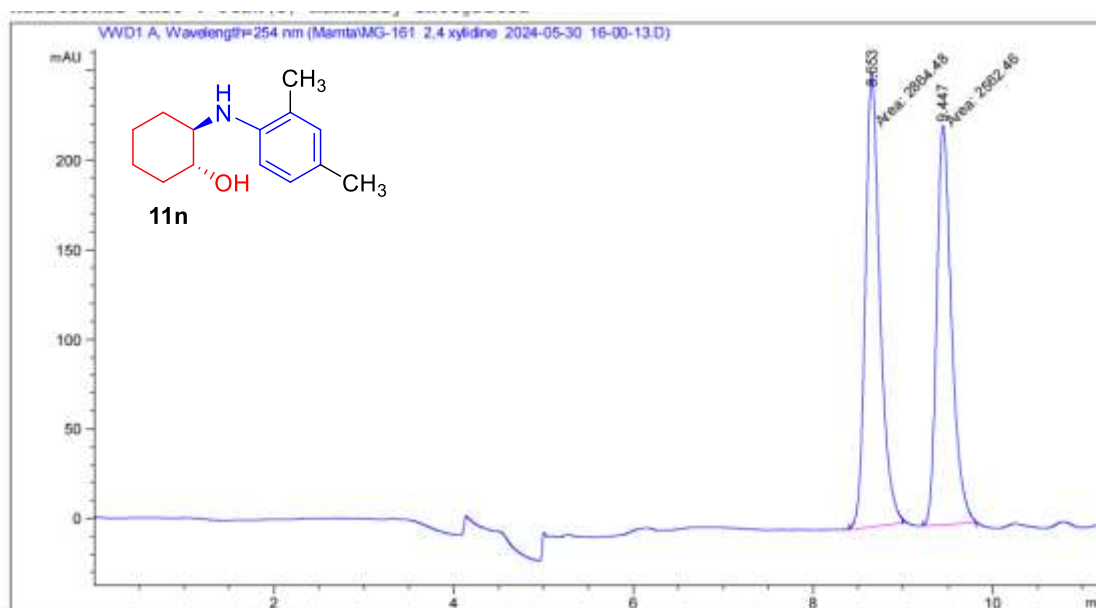


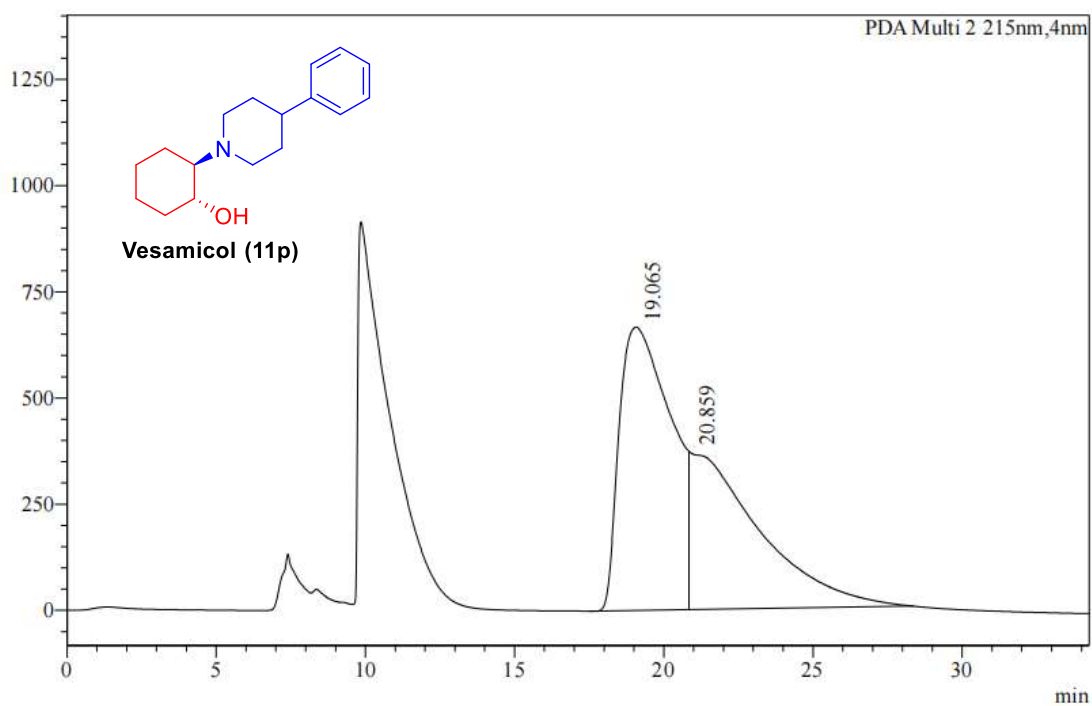


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.030	MM	0.1927	1.14935e4	993.99231	54.6021
2	8.790	MM	0.2490	9556.04590	639.69452	45.3979



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.911	MM	0.1711	2123.18530	206.78302	61.0762
2	7.454	MM	0.1821	1353.10229	123.81221	38.9238

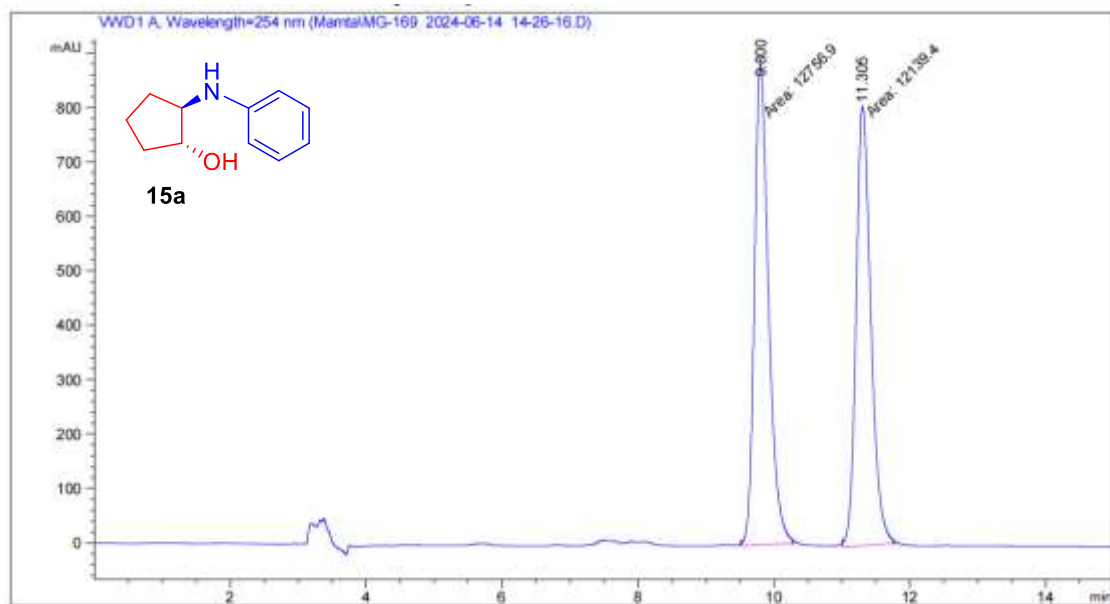




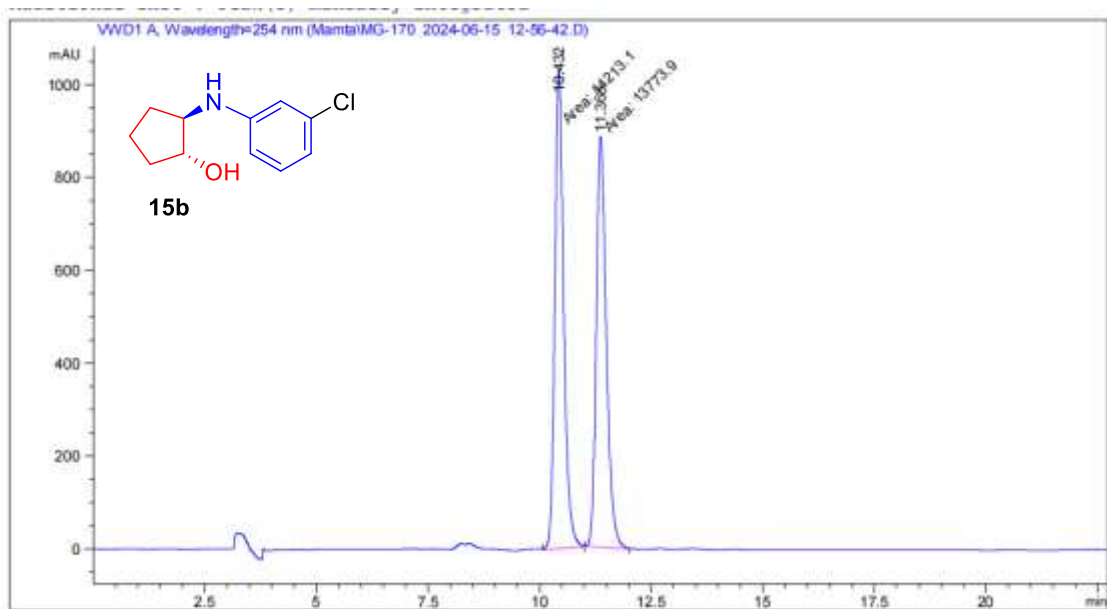
Peak Table

PDA Ch2 215nm

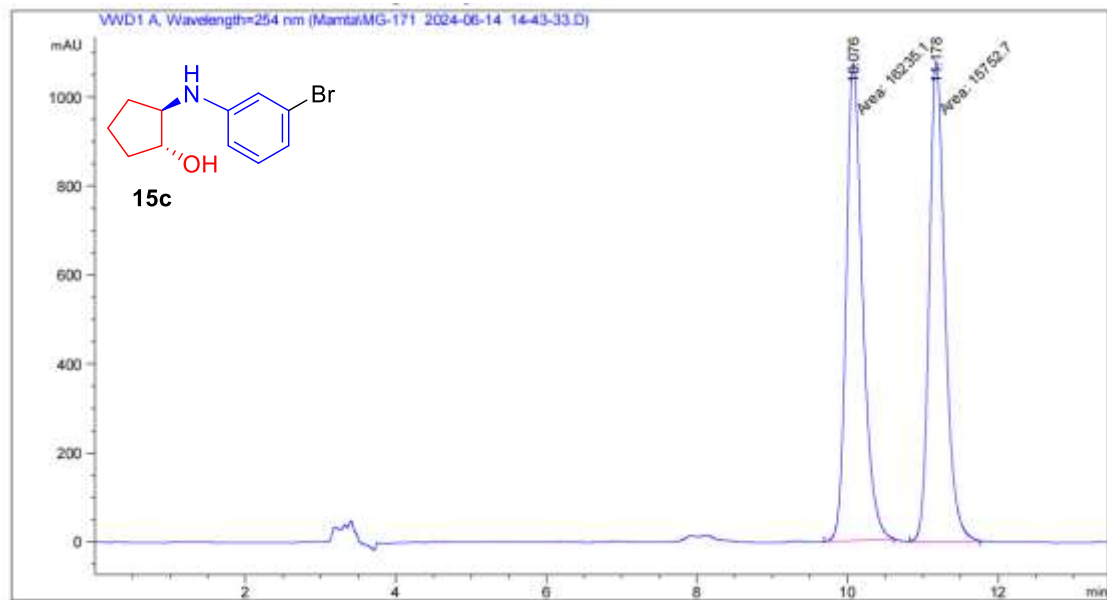
Peak#	Ret. Time	Area	Height	Area%
1	19.065	80913137	666941	57.502
2	20.859	59799588	371948	42.498
Total		140712725	1038888	100.000



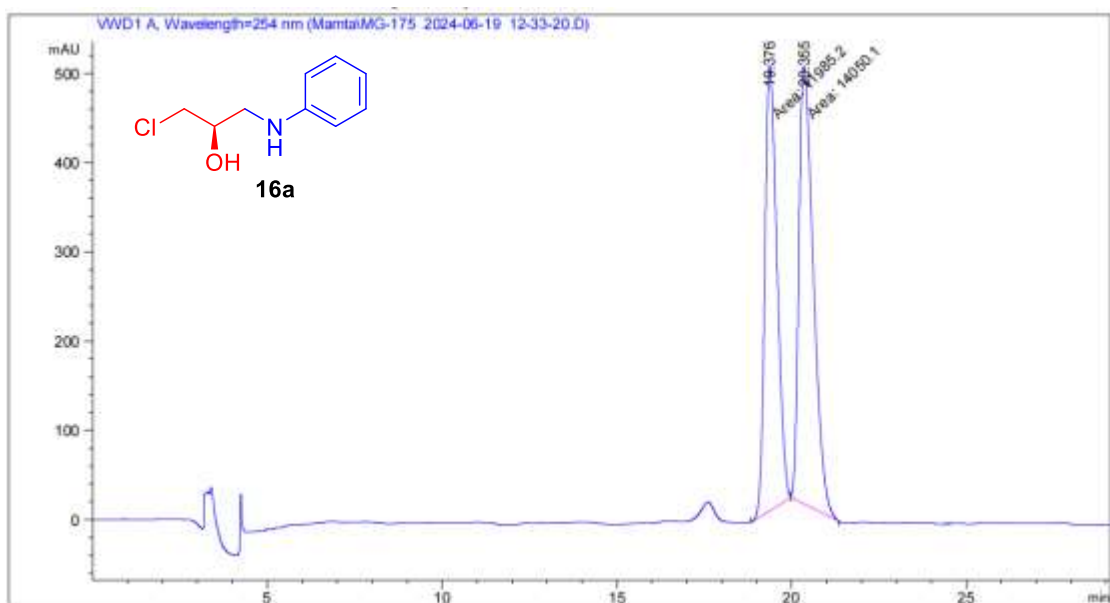
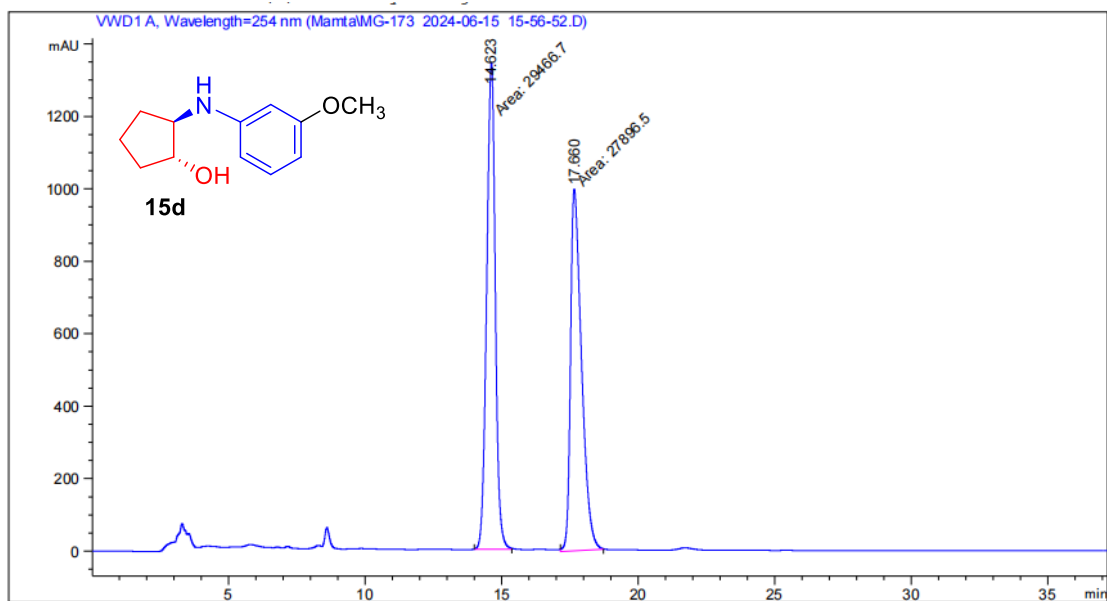
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.800	MM	0.2404	1.27569e4	884.34229	51.2402
2	11.305	MM	0.2507	1.21394e4	807.07159	48.7598

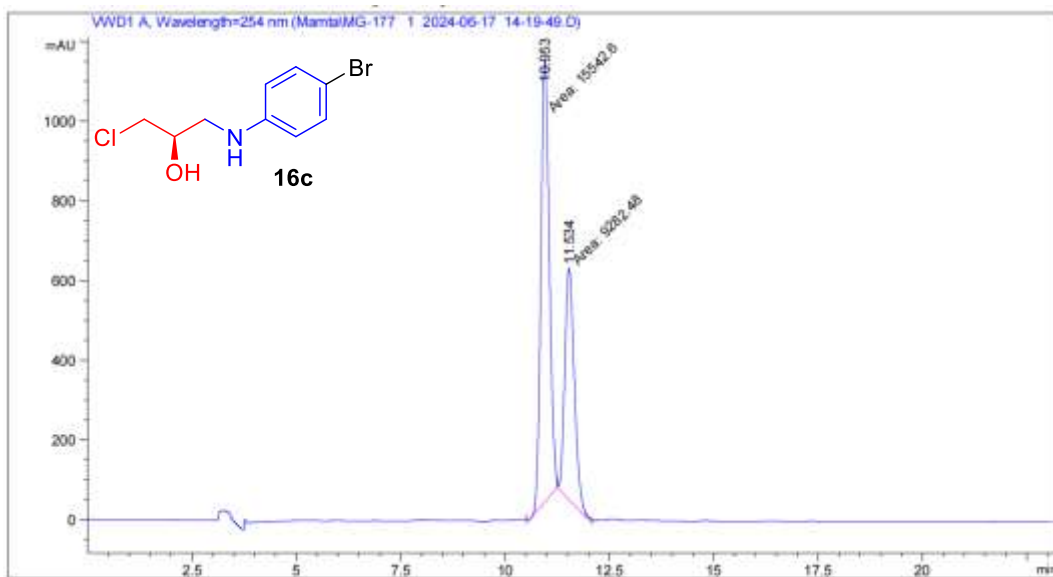
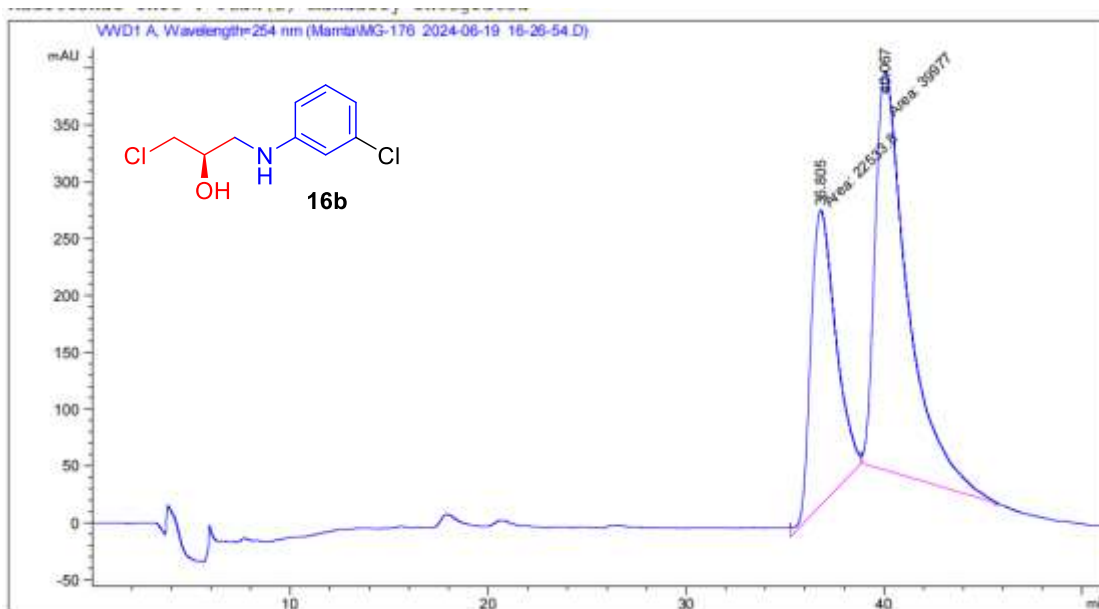


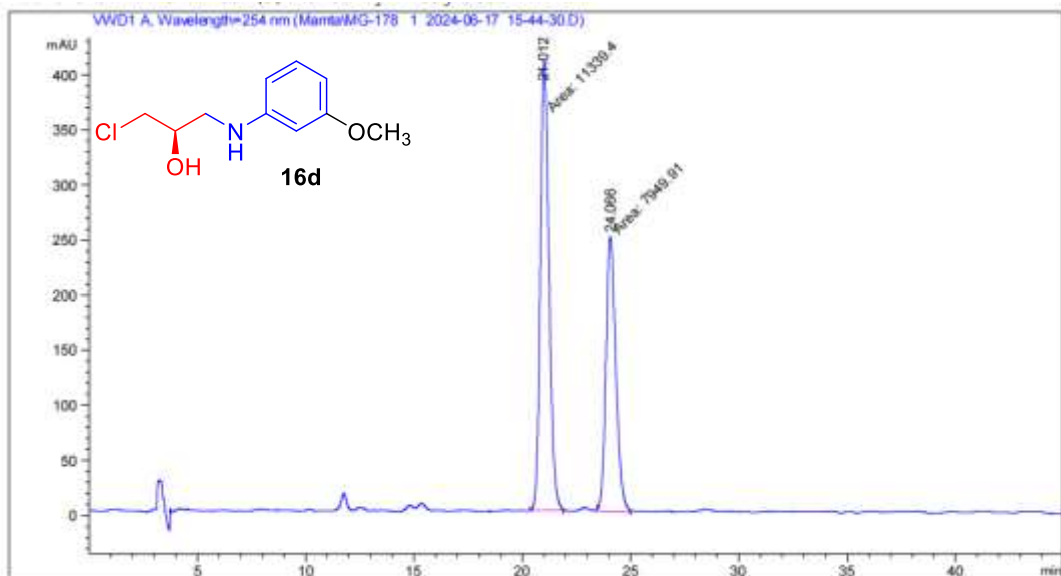
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.432	MM	0.2300	1.42131e4	1029.71936	50.7845
2	11.368	MM	0.2594	1.37739e4	885.14673	49.2155



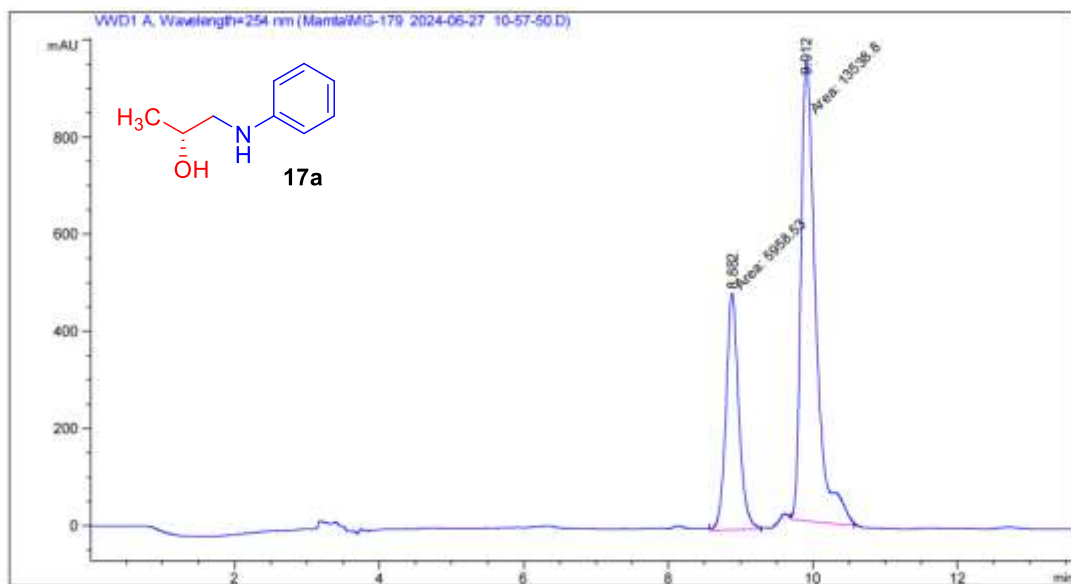
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.076	MM	0.2523	1.62351e4	1072.52429	50.7541
2	11.178	MM	0.2431	1.57527e4	1080.15857	49.2459



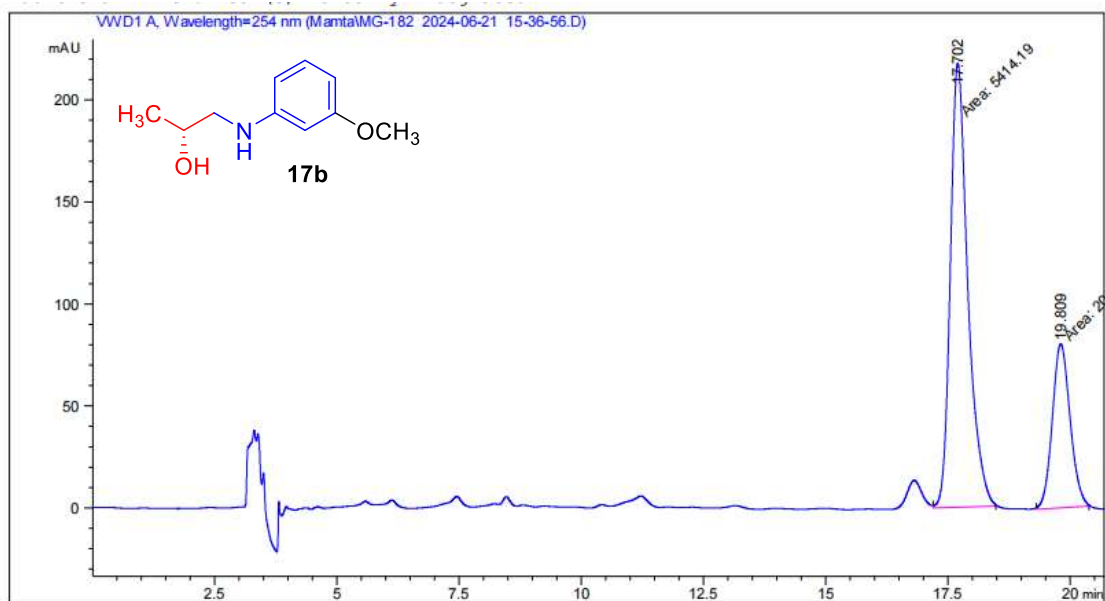




Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.012	MM	0.4630	1.13394e4	408.20551	58.7859
2	24.066	MM	0.5301	7949.91357	249.93777	41.2141



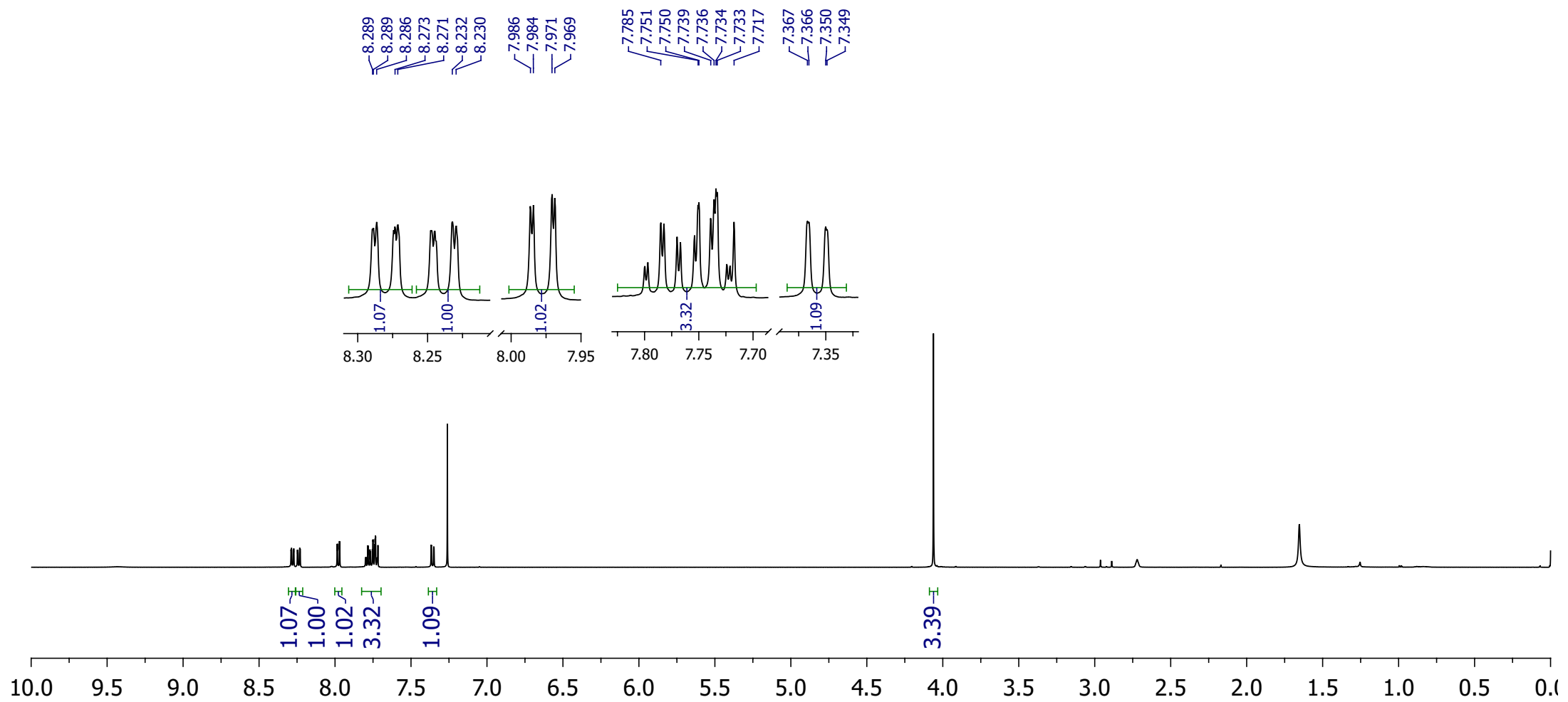
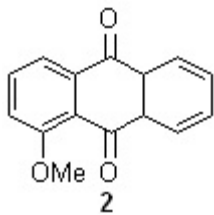
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.882	MM	0.2042	5958.52832	486.22195	30.5607
2	9.912	MM	0.2389	1.35388e4	944.72180	69.4393



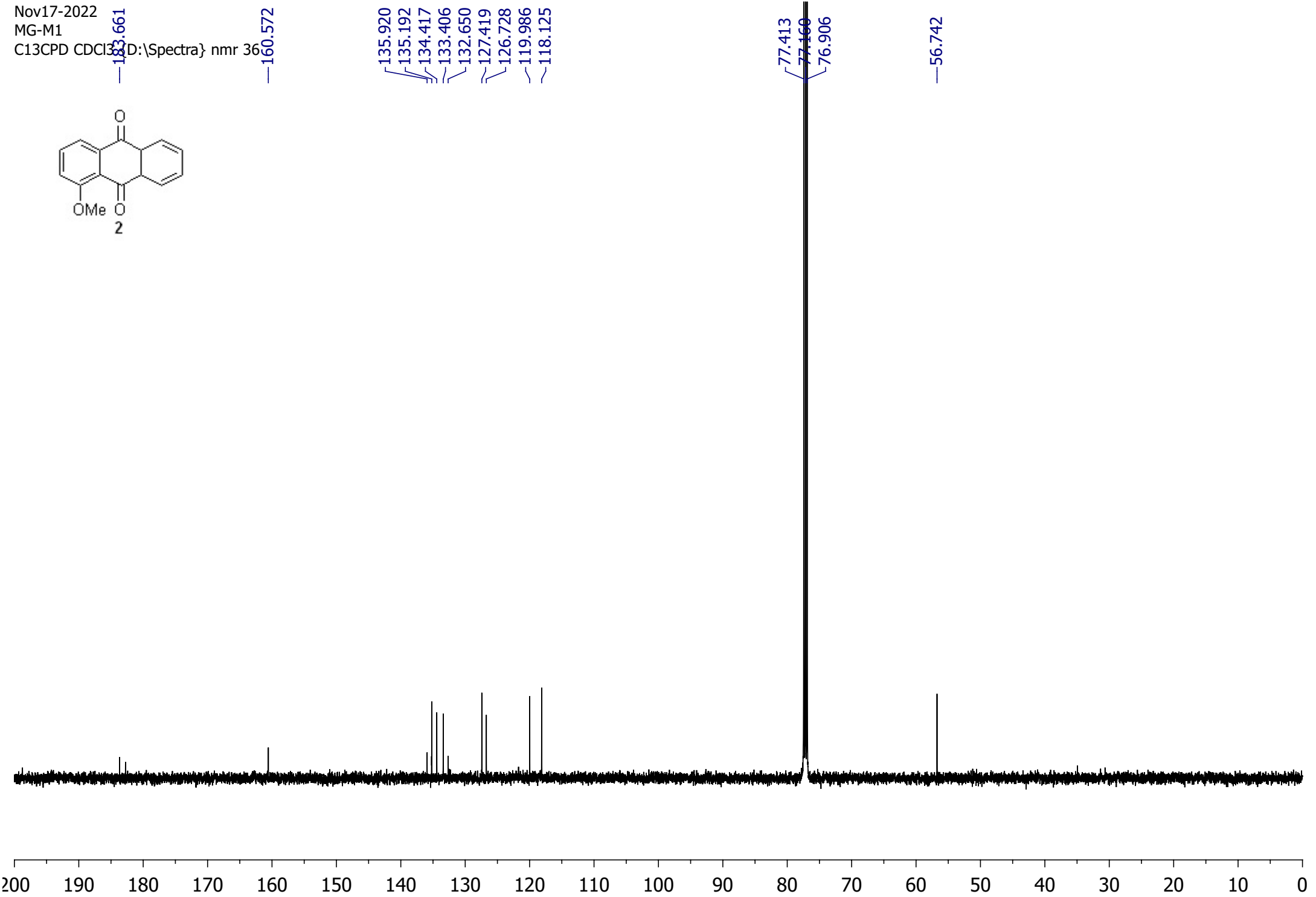
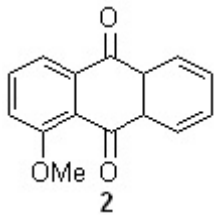
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.702	MM	0.4149	5414.18994	217.50145	72.6684
2	19.809	MM	0.4217	2036.35645	80.47579	27.3316

NMR Spectra of Triptycene Derivatives & Amino alcohol Products

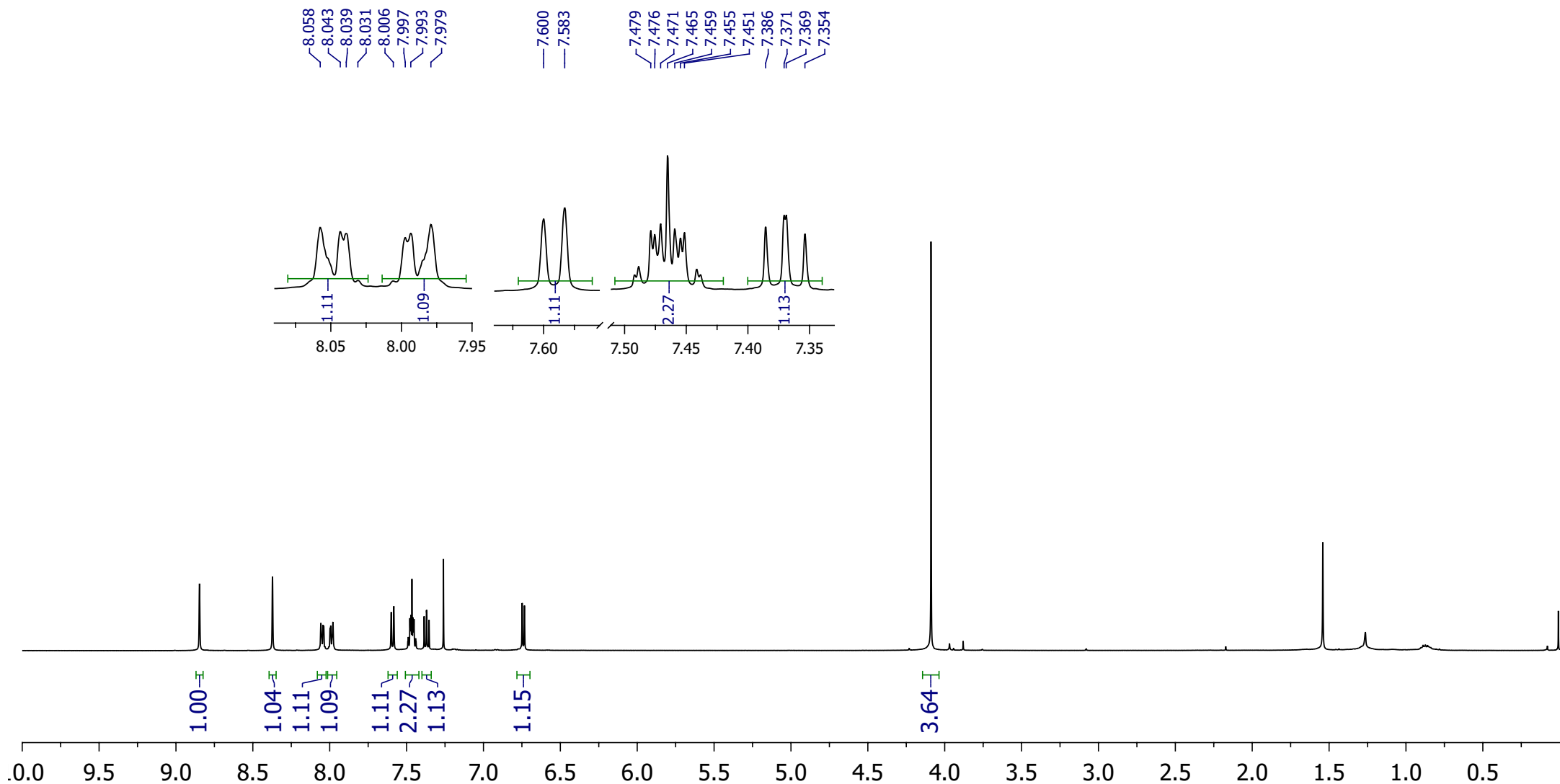
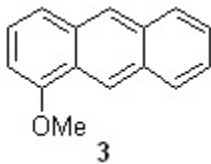
Nov 17, 2022
 MG-711
 1H_8scan CDCl3 3D Spectra, hmr 36



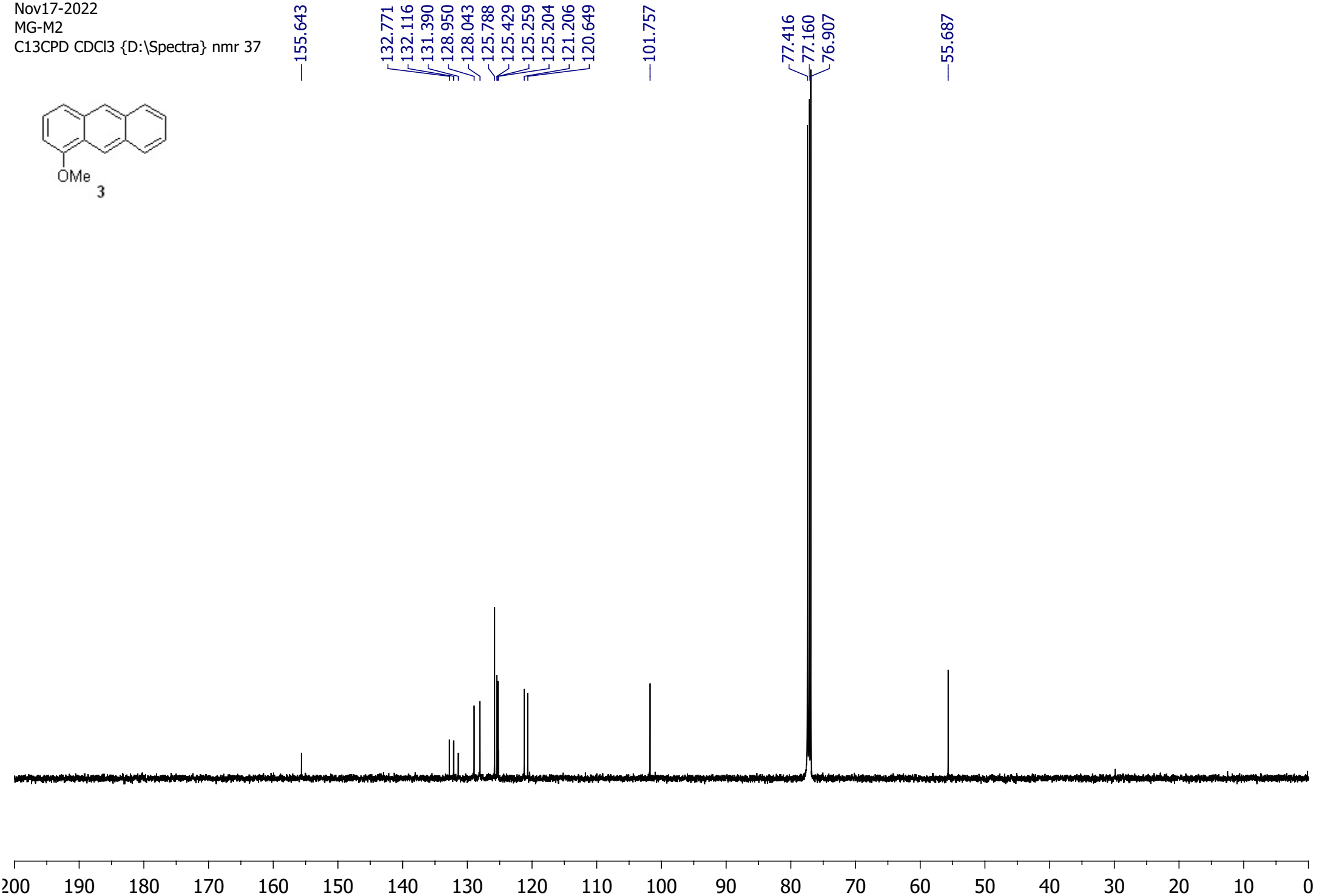
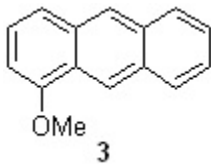
Nov17-2022
MG-M1
C13CPD CDCl3 D:\Spectra\ nmr 36



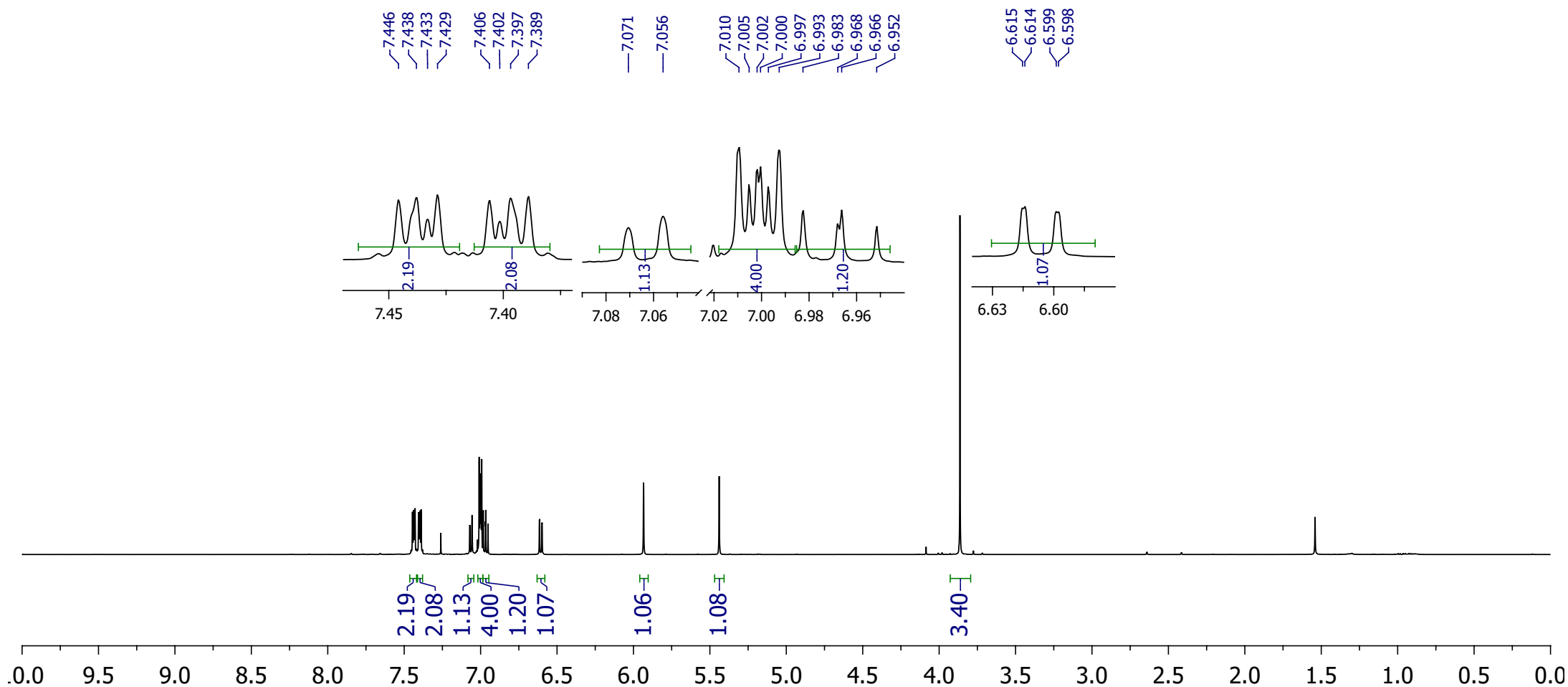
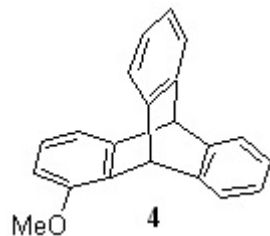
Nov17-2022
MG-M2
1H_8scan CDCl3 {D: Spectra} nmr 37



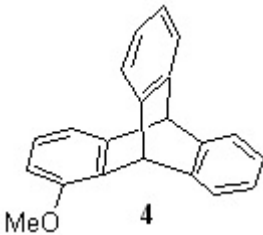
Nov17-2022
MG-M2
C13CPD CDCl3 {D:\Spectra} nmr 37



manta-2025-06-24T123531.00
 TP-QM
 1H_8scan_CDCl3(D:\Spectra\hmr 20



mamta - 2025-06-24T123531.606
TP-OMe
C13CPD CDCl3 {D:\Spectra} nmr 20



—154.571
—147.698
—145.933
—145.476

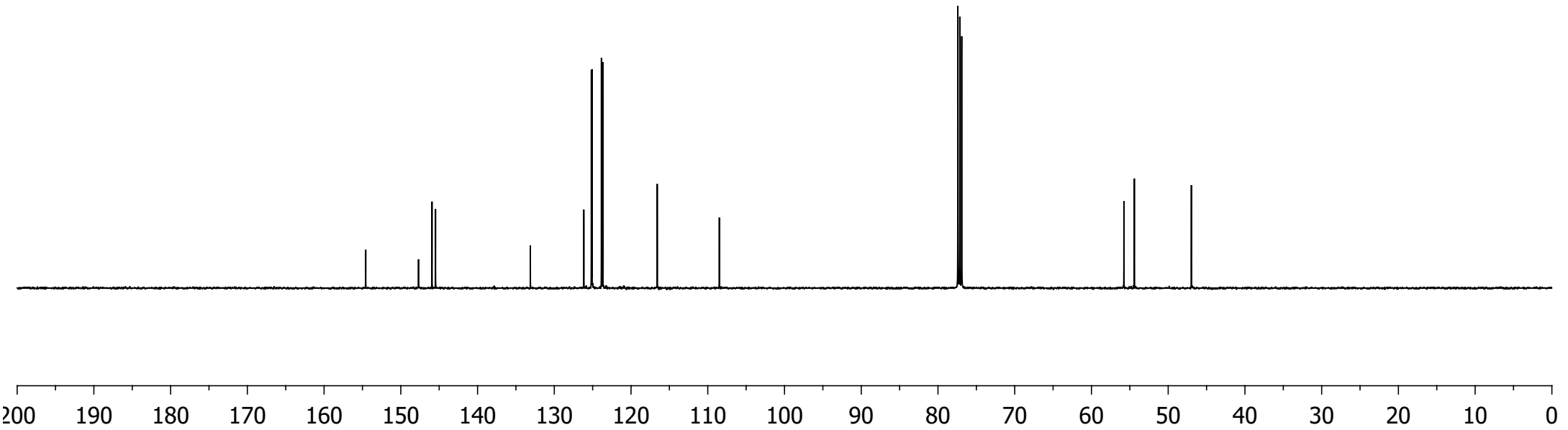
—133.117
—125.172
—125.092
—123.860
—123.689
—122.599

—108.496

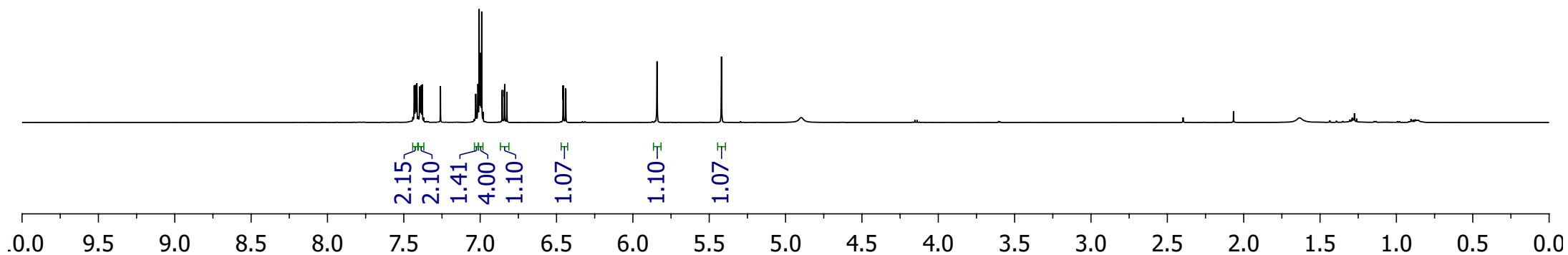
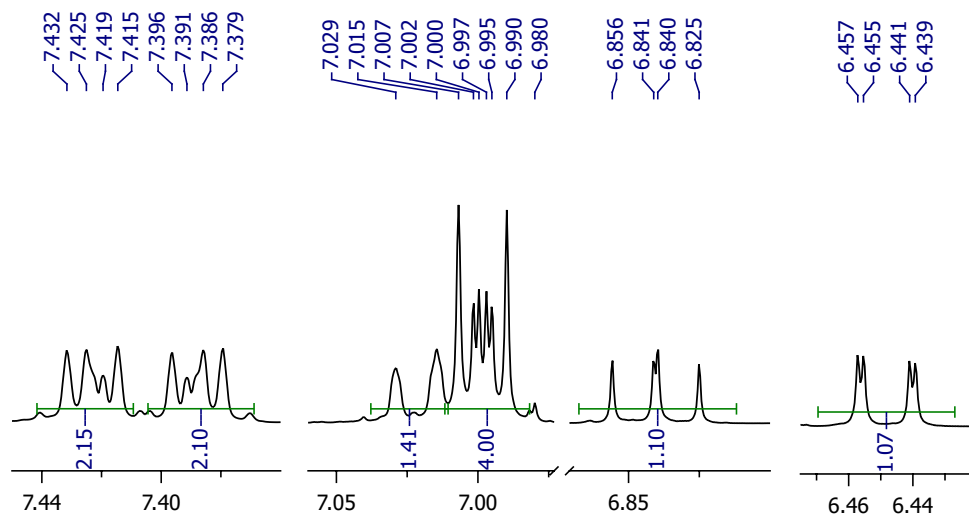
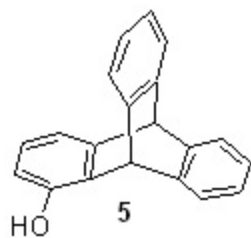
—77.413
—77.160
—76.905

—55.770
—54.407

—46.983



mamta_2025-06-24T12:35:31.606
 TP-01H
 1H_scan CDCl3 (D:\Spectra\ nmr 2)

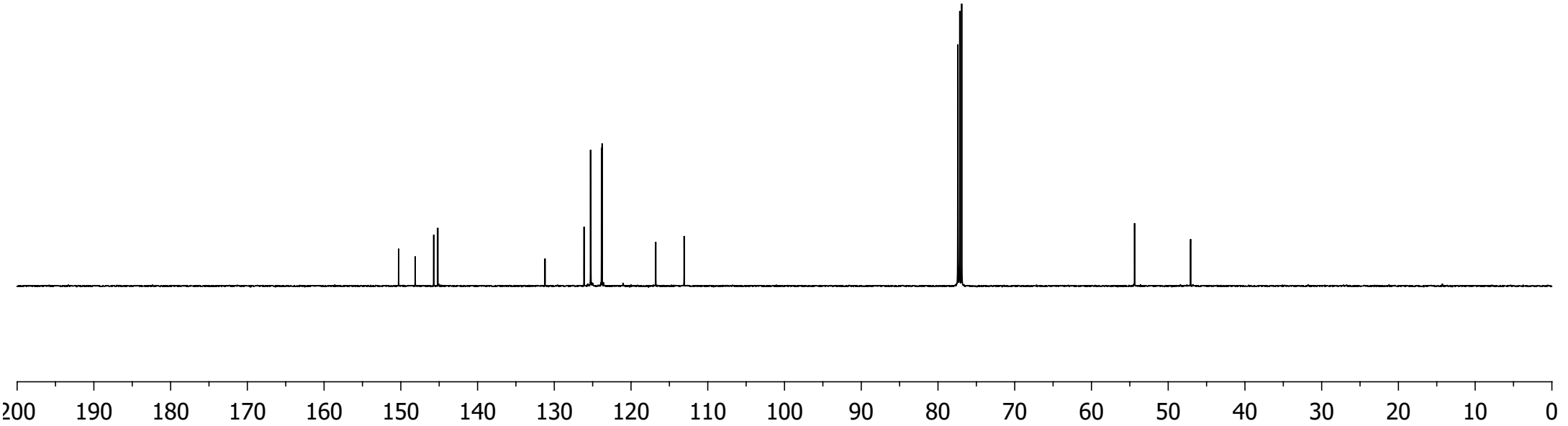
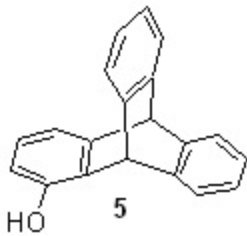


mamta - 2025-06-24T123531.606
TP-OH
C13CPD CDCl3 {D:\Spectra} nmr 21

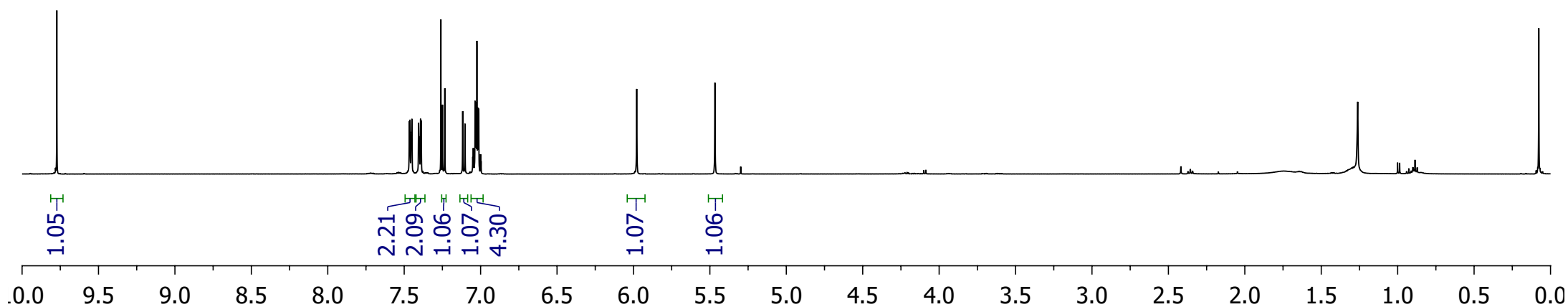
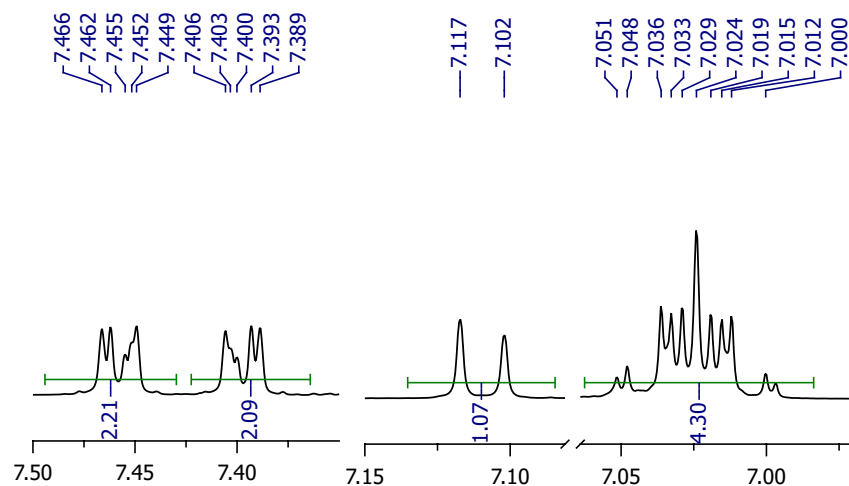
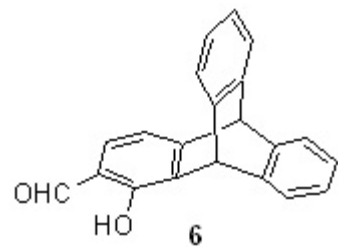
150.289
148.123
145.715
145.184
131.209
126.123
125.277
125.234
123.848
123.767
116.779
113.079

77.415
77.160
76.907

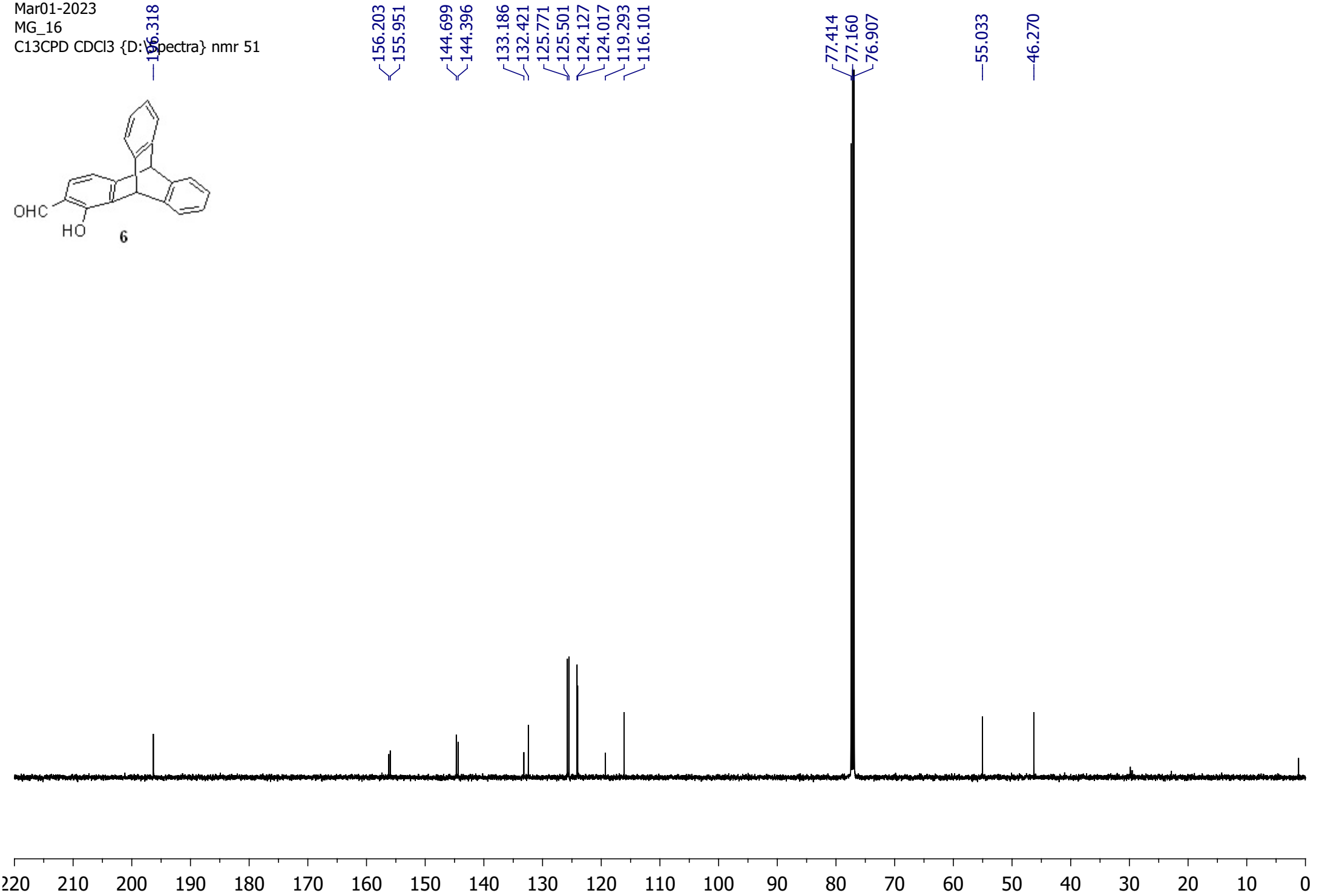
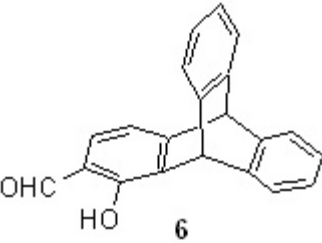
54.385
47.082



Mar 01-2023
 MG 16
 1H_NScan CDCl3 (D:\Spectra\mmr 51
 9.776
 7.462
 7.462
 7.455
 7.452
 7.449
 7.406
 7.403
 7.400
 7.393
 7.389
 7.260
 7.249
 7.234
 7.117
 7.102
 7.048
 7.036
 7.033
 7.029
 7.024
 7.019
 7.015
 7.012
 5.979
 5.467

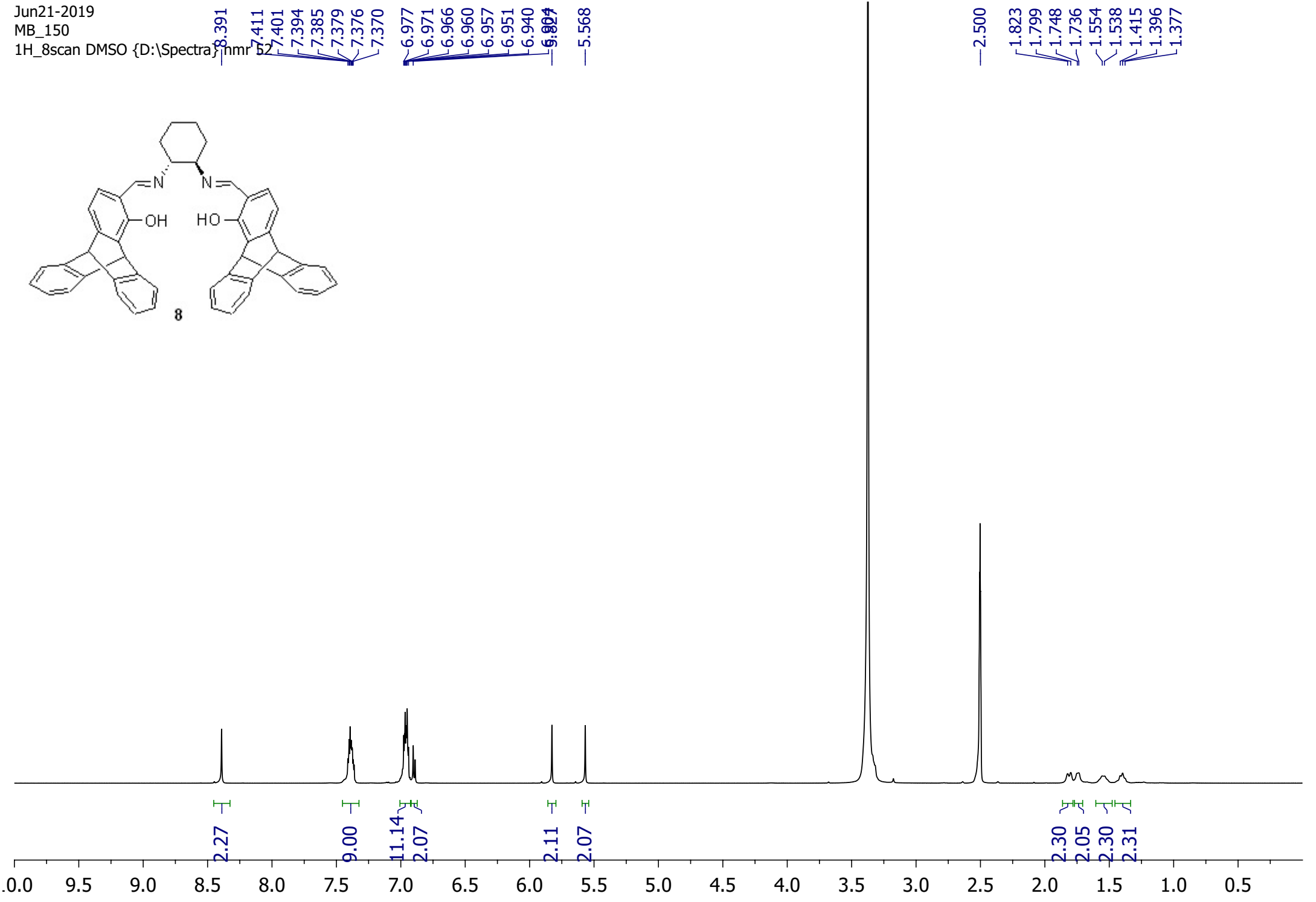
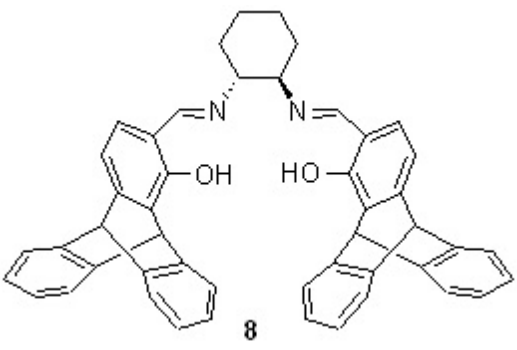


Mar01-2023
MG_16
C13CPD CDCl3 {D:\spectra} nmr 51

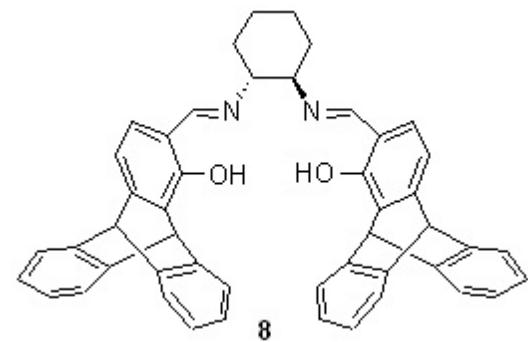


Jun21-2019
MB_150
1H_8scan DMSO {D:\Spectra\}

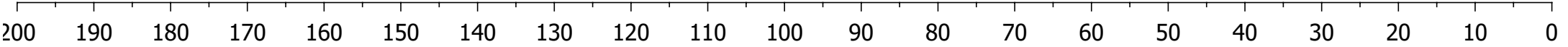
nmr 52
8.391
7.411
7.401
7.394
7.385
7.379
7.376
7.370
6.977
6.971
6.966
6.960
6.957
6.951
6.940
5.804
5.568



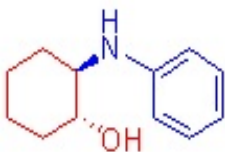
Jul11-2019
MB_150
C13CPD CDCl3 {D:\Spectra} nm 27



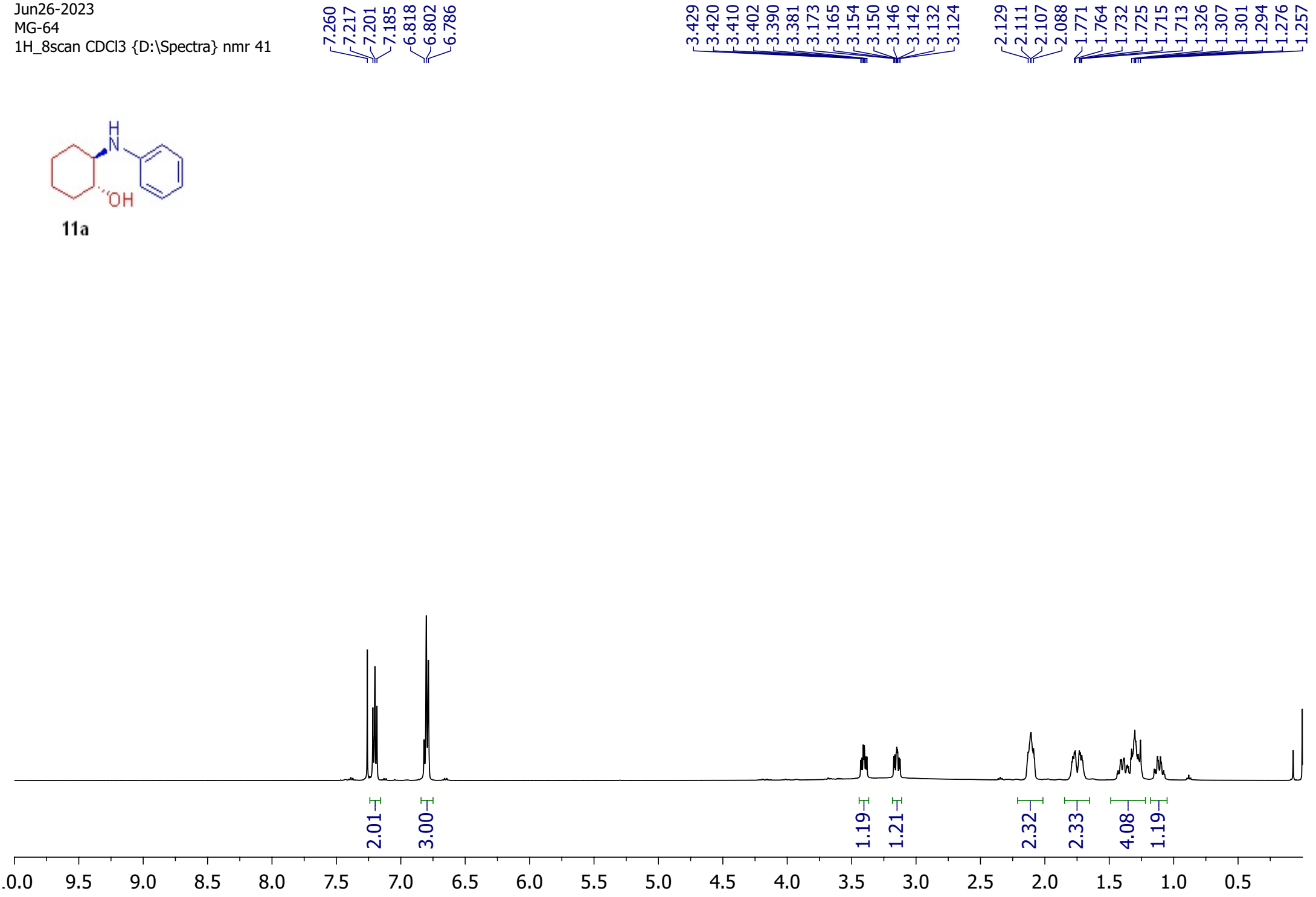
164.872
155.525
150.767
145.333
129.316
125.333
125.117
123.834
123.724
116.895
114.548
77.413
77.160
76.905
72.536
54.715
46.609
33.294
24.258



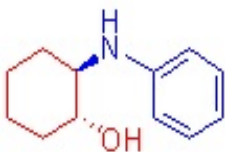
Jun26-2023
MG-64
1H_8scan CDCl3 {D:\Spectra} nmr 41



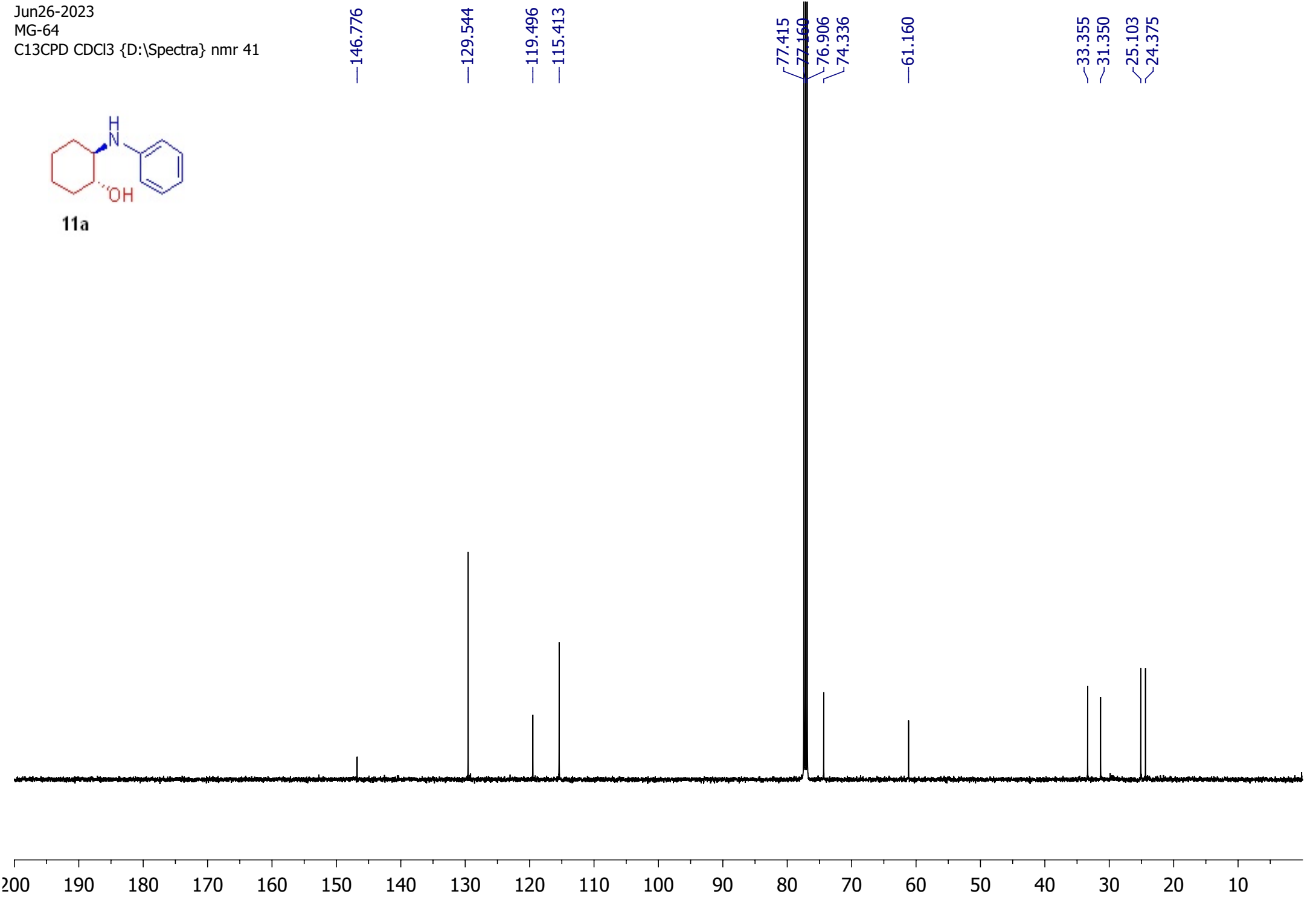
11a

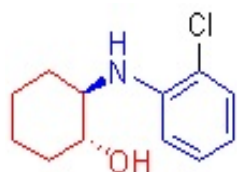


Jun26-2023
MG-64
C13CPD CDCl3 {D:\Spectra} nmr 41

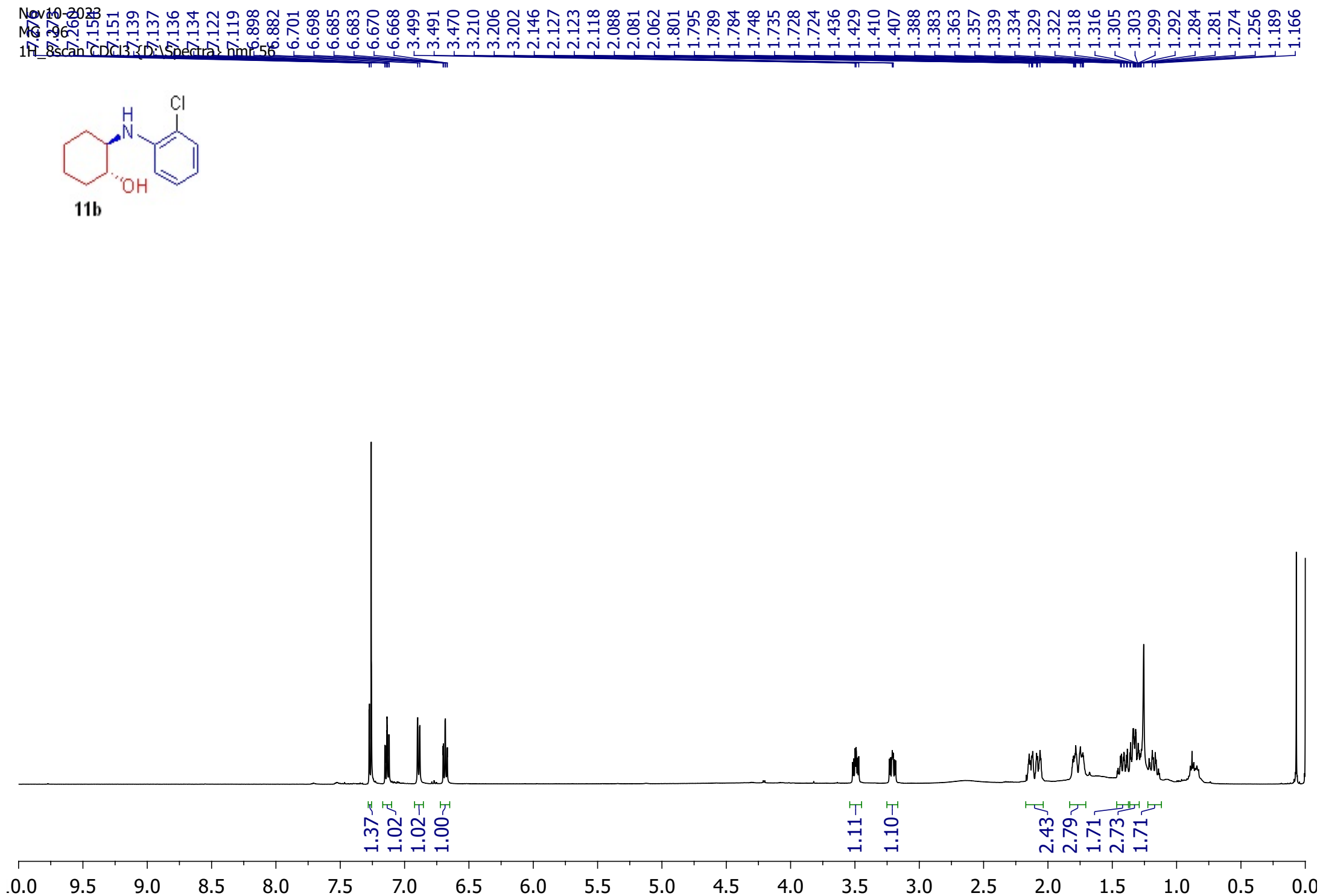


11a

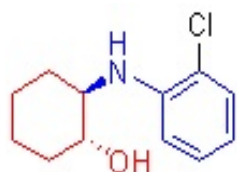




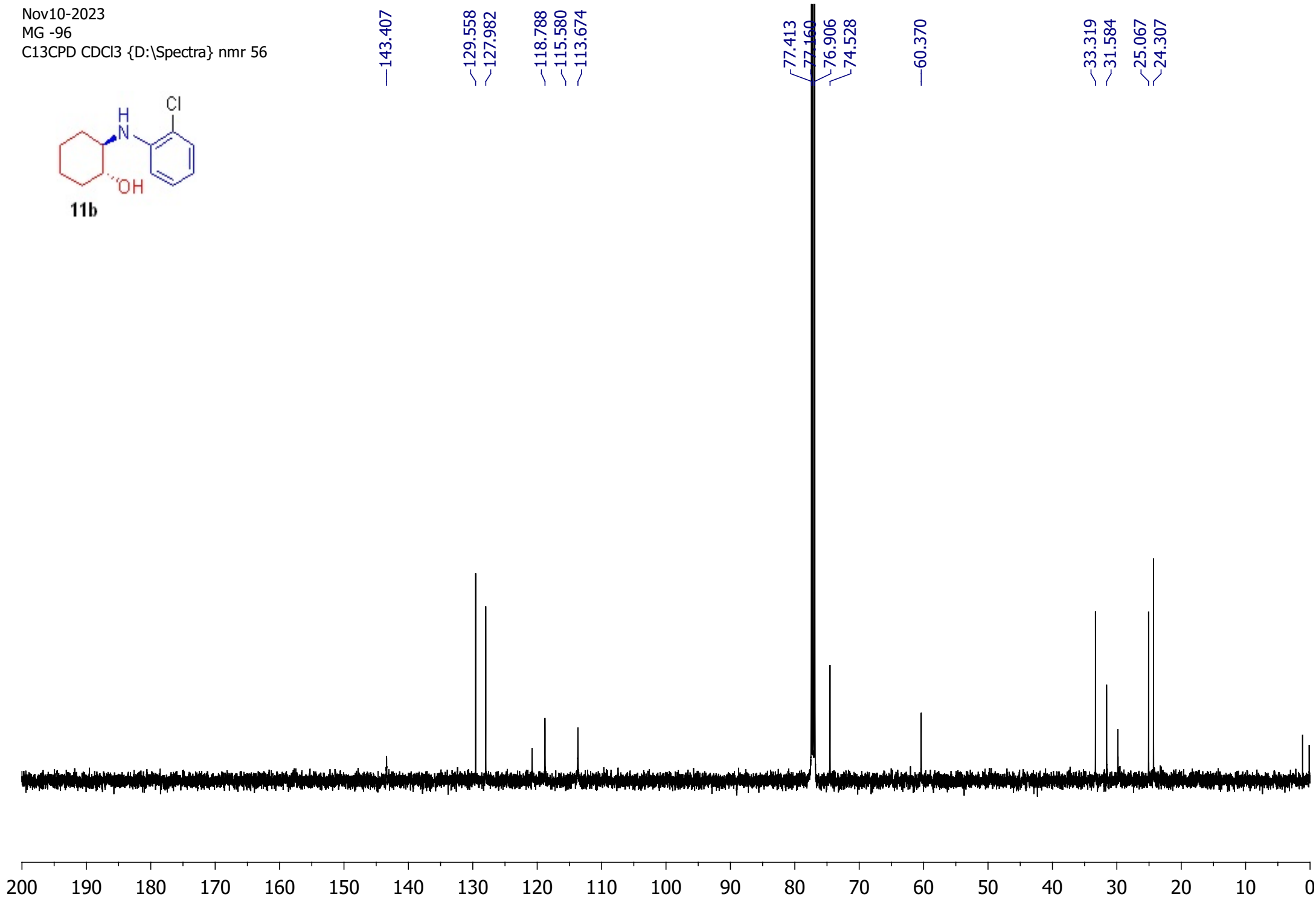
11b

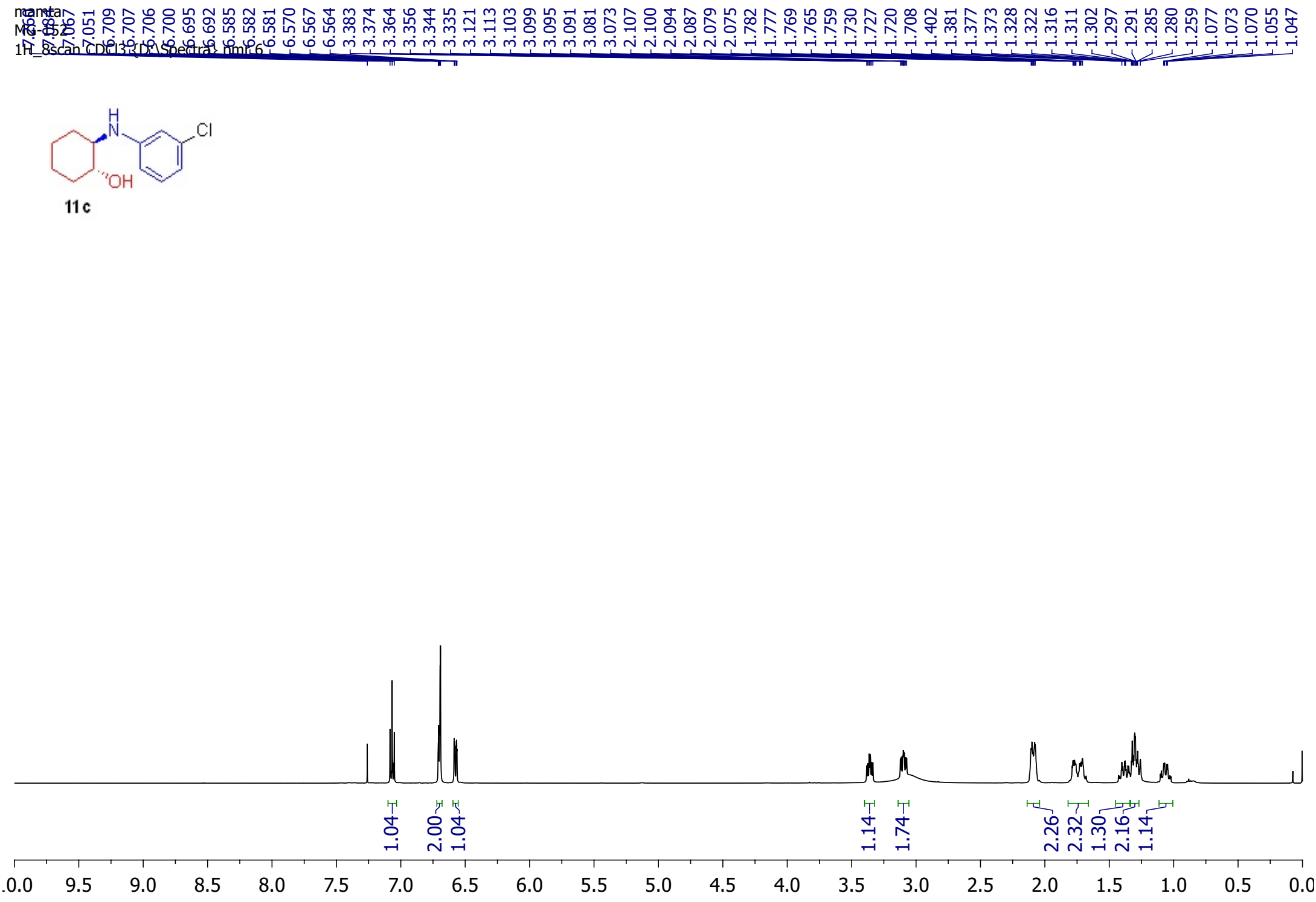


Nov10-2023
MG -96
C13CPD CDCl3 {D:\Spectra} nmr 56

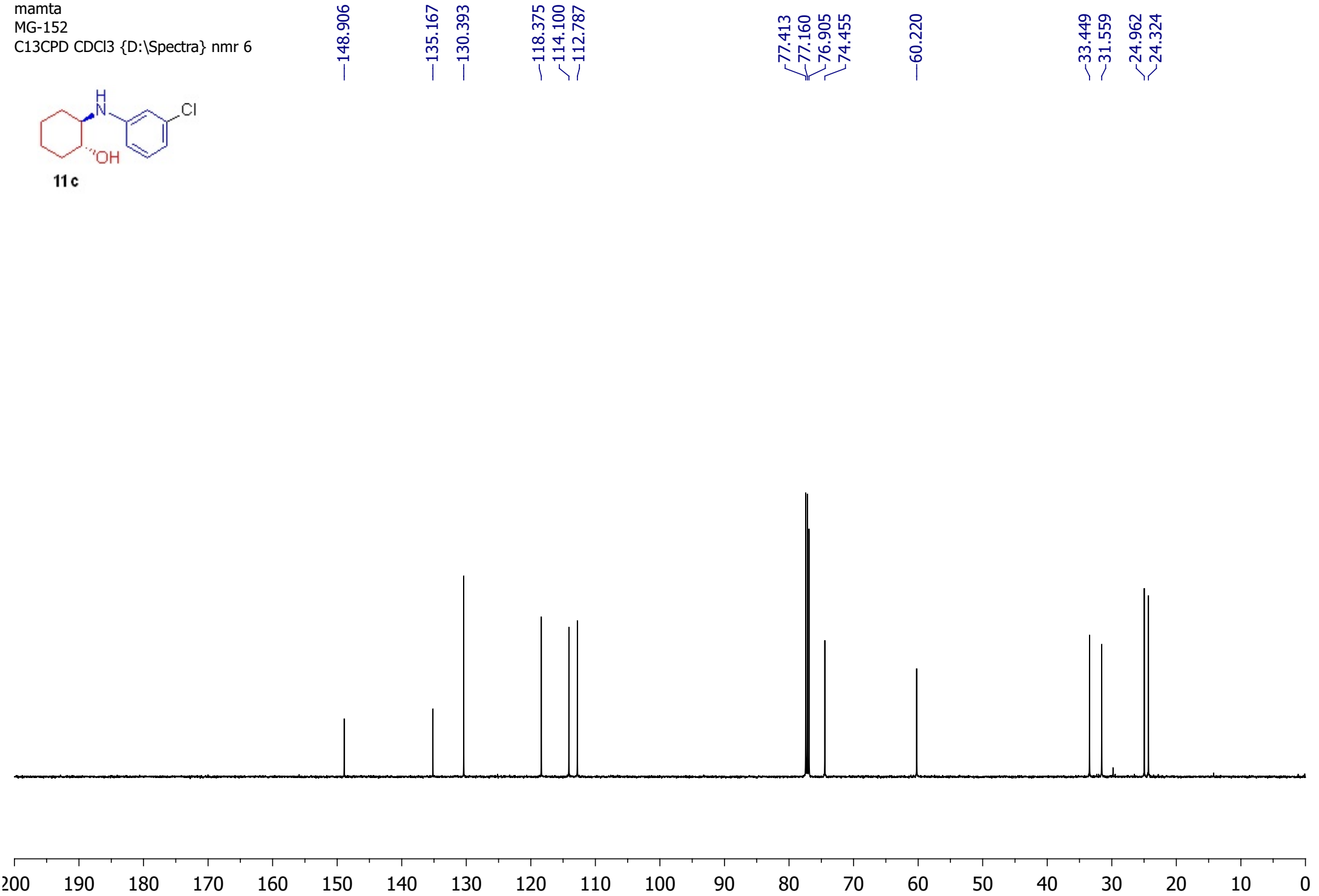


11b

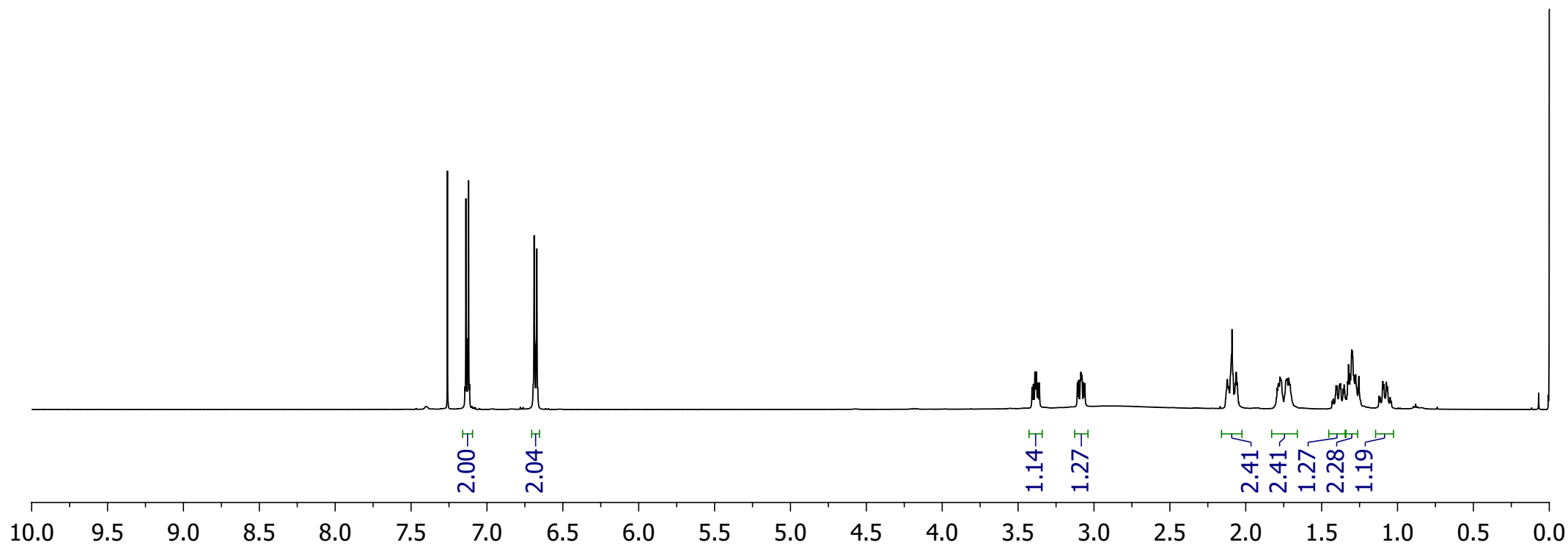
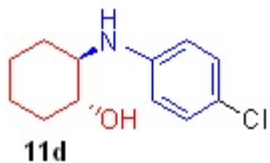




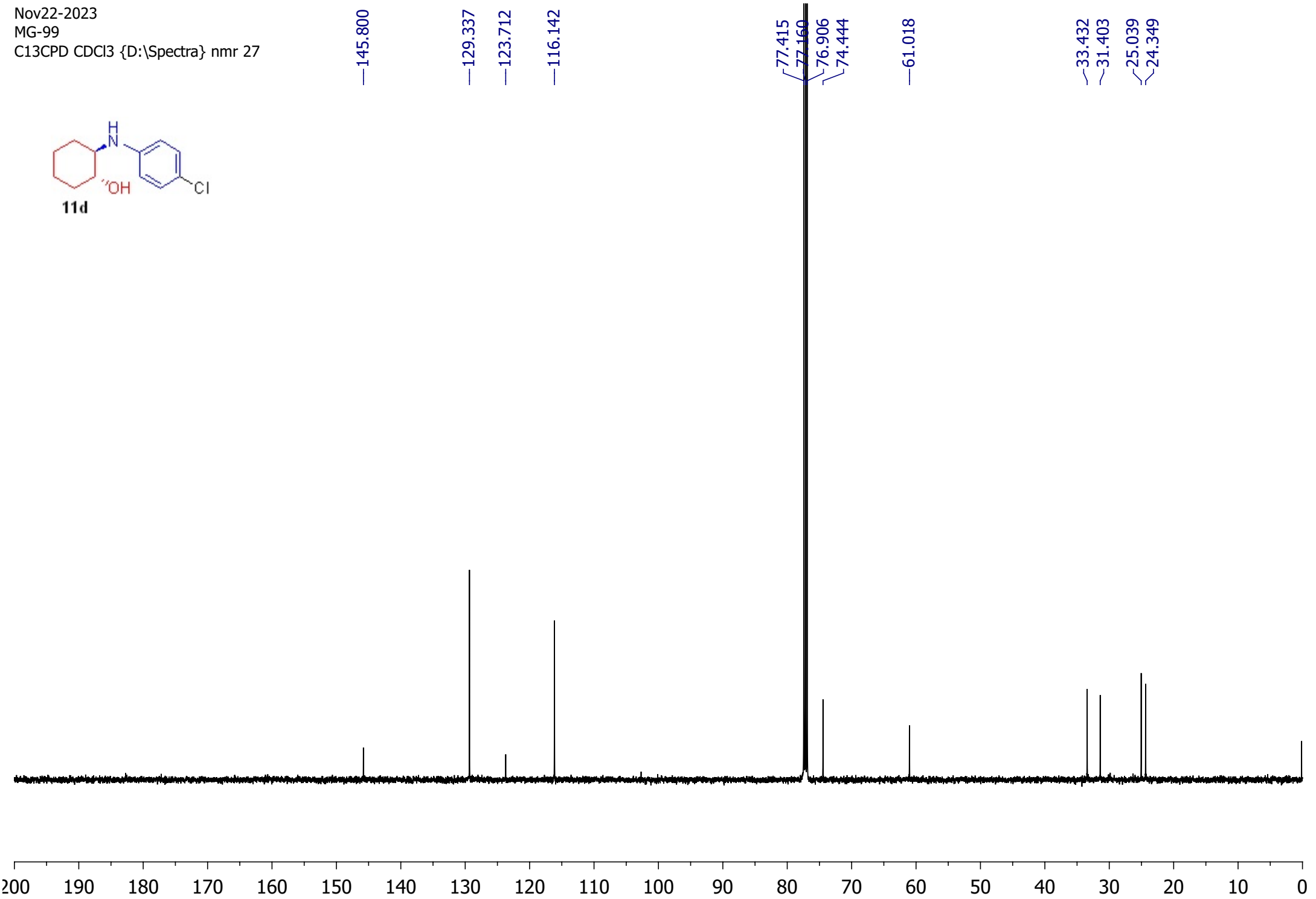
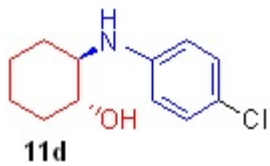
mamta
MG-152
C13CPD CDCl3 {D:\Spectra} nmr 6

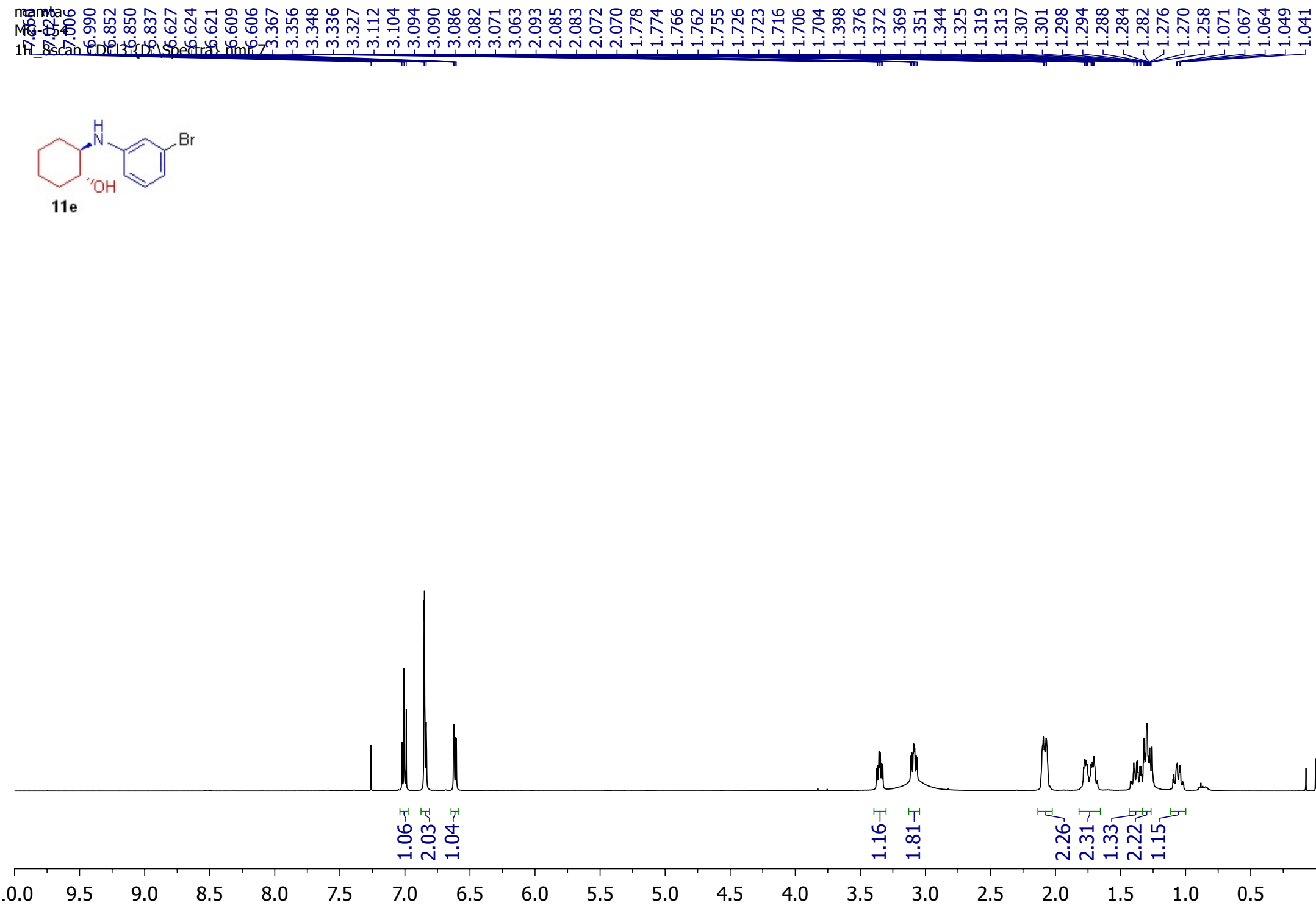


No. 23-0023
Date 7/13/23
Mod 7.99
1H NMR (CDCl₃) Spectrum
7.138, 7.121, 7.115, 6.689, 6.671, 3.399, 3.389, 3.381, 3.369, 3.360, 3.109, 3.101, 3.090, 3.086, 3.083, 3.079, 3.068, 3.060, 2.121, 2.112, 2.093, 2.090, 2.083, 2.069, 2.063, 2.056, 1.785, 1.782, 1.774, 1.770, 1.736, 1.731, 1.727, 1.717, 1.708, 1.405, 1.398, 1.383, 1.379, 1.376, 1.351, 1.328, 1.323, 1.317, 1.312, 1.310, 1.301, 1.297, 1.295, 1.288, 1.284, 1.281, 1.275, 1.255, 1.097, 1.092, 1.089, 1.074, 1.066

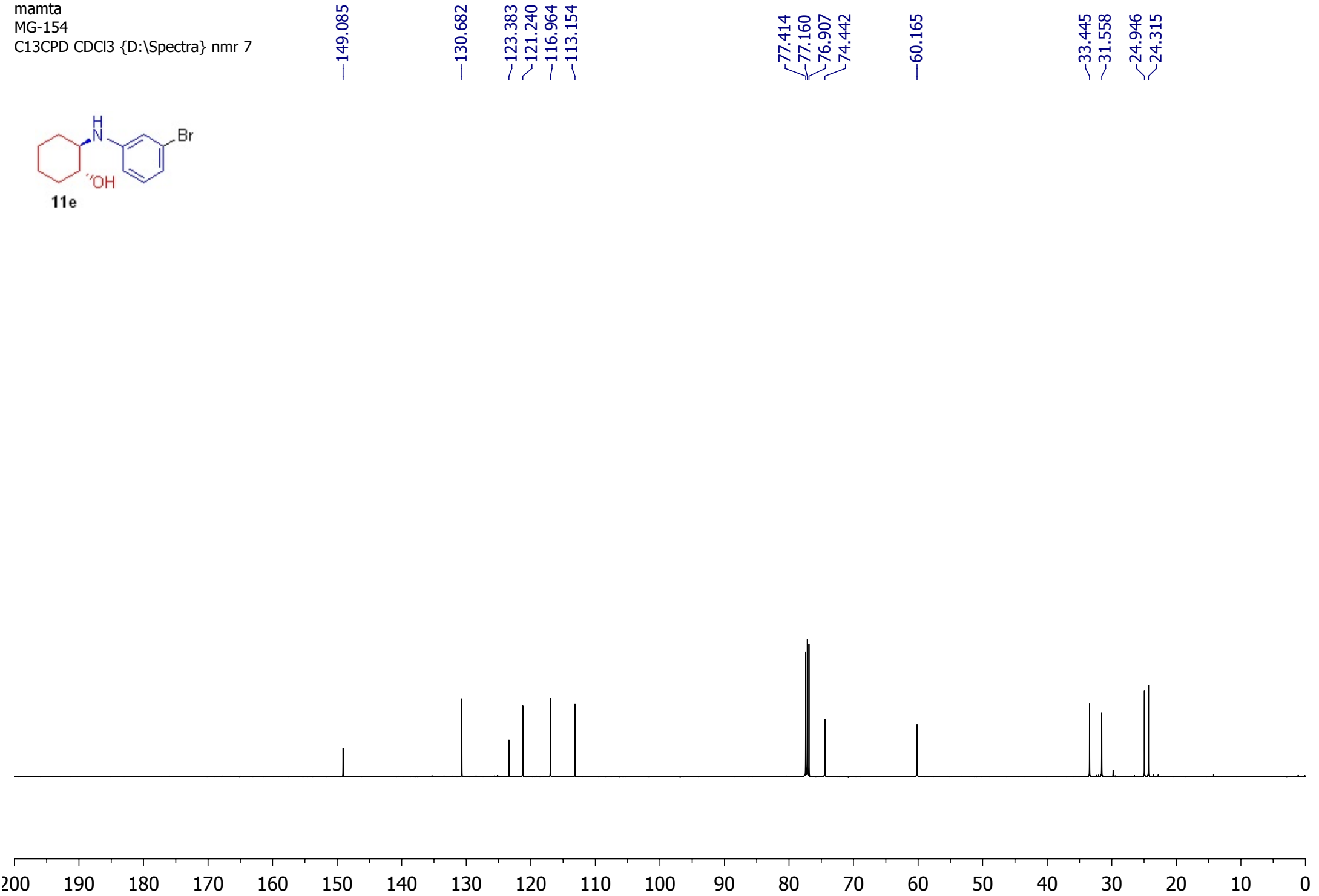
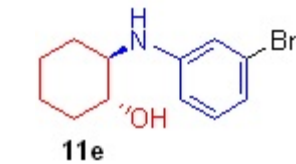


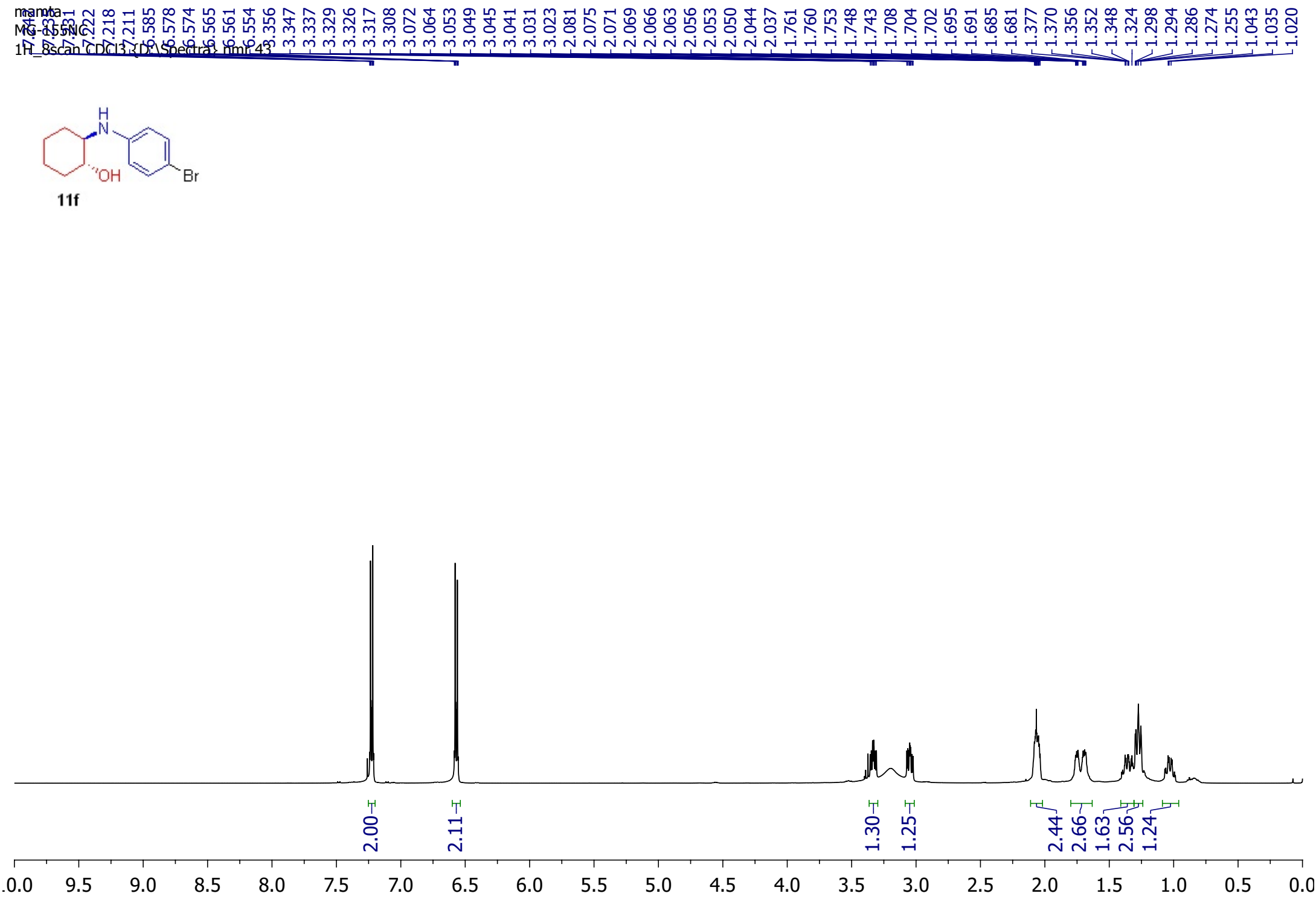
Nov22-2023
MG-99
C13CPD CDCl3 {D:\Spectra} nmr 27



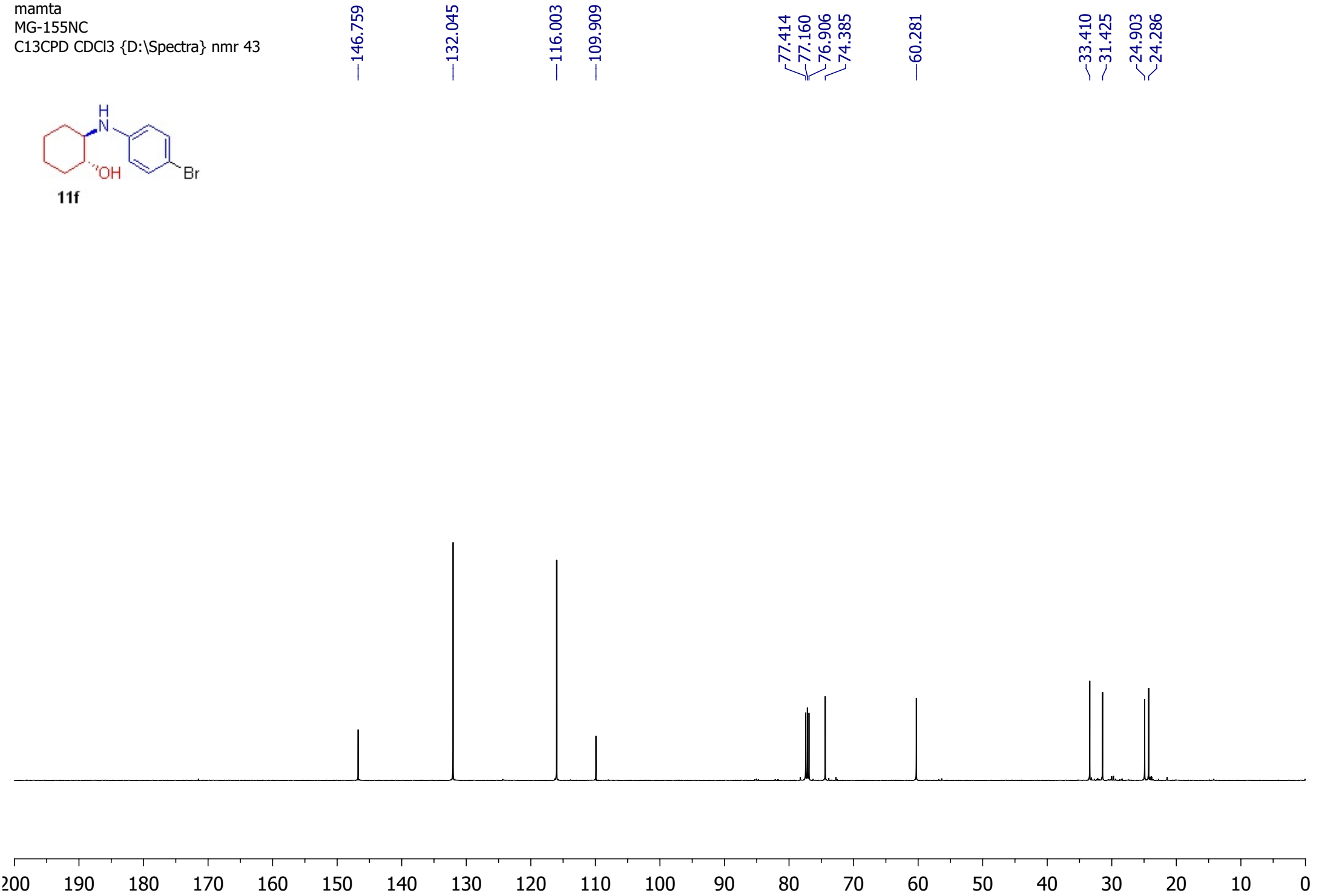
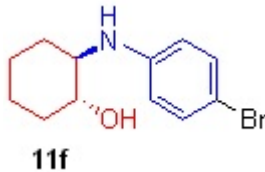


mamta
MG-154
C13CPD CDCl3 {D:\Spectra} nmr 7

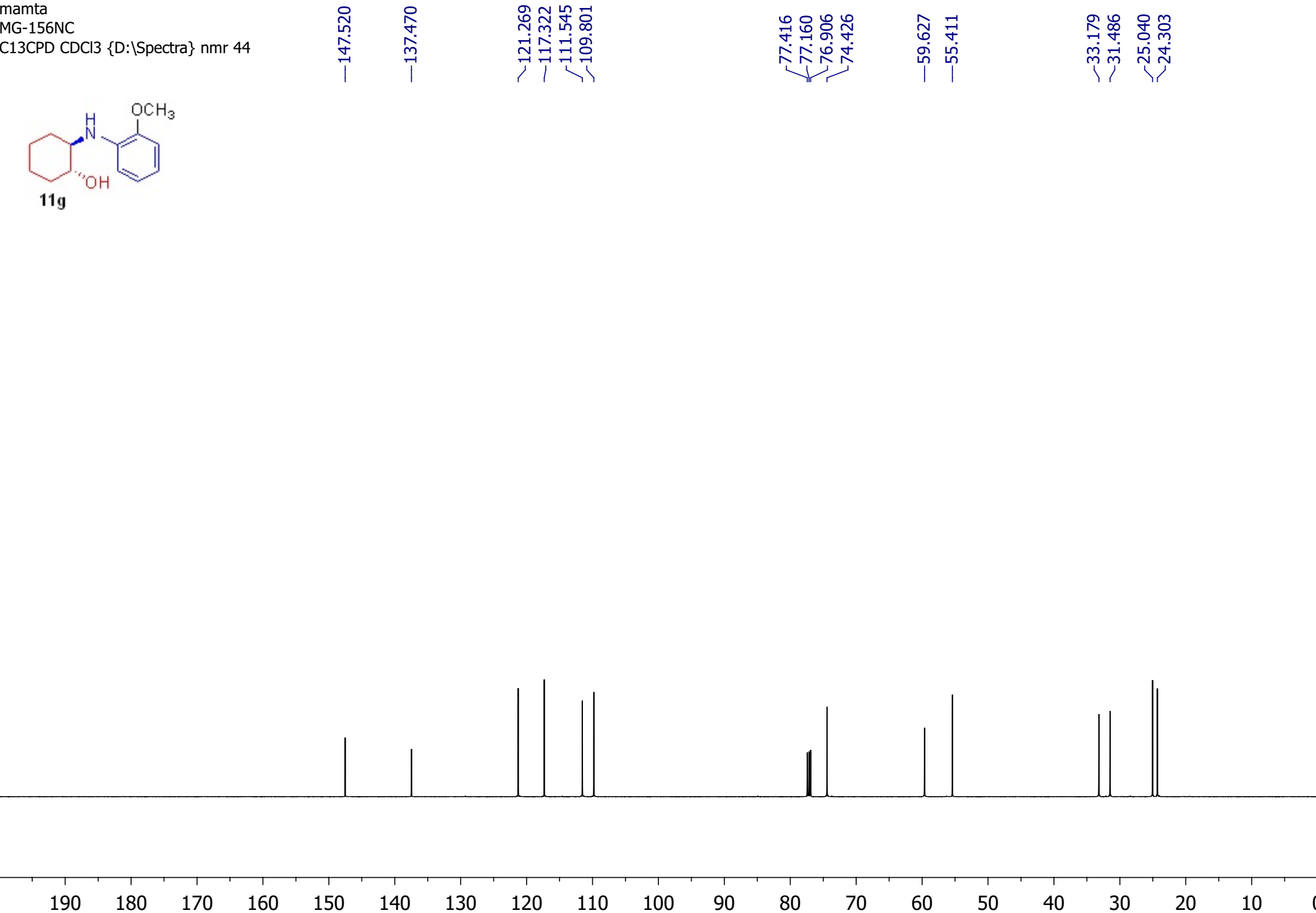
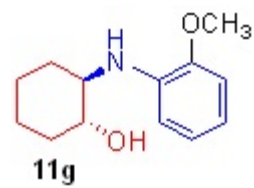


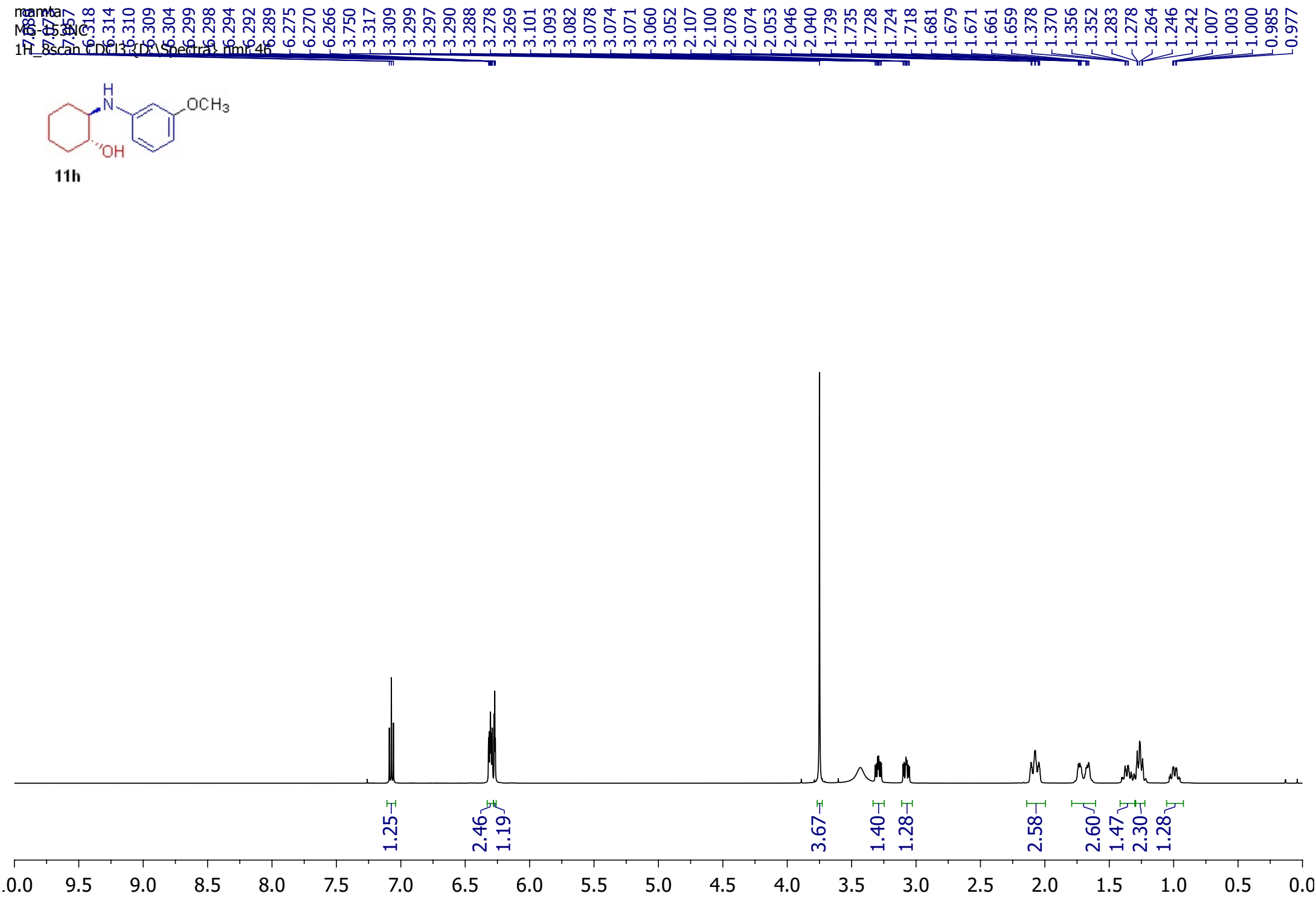


mamta
MG-155NC
C13CPD CDCl3 {D:\Spectra} nmr 43

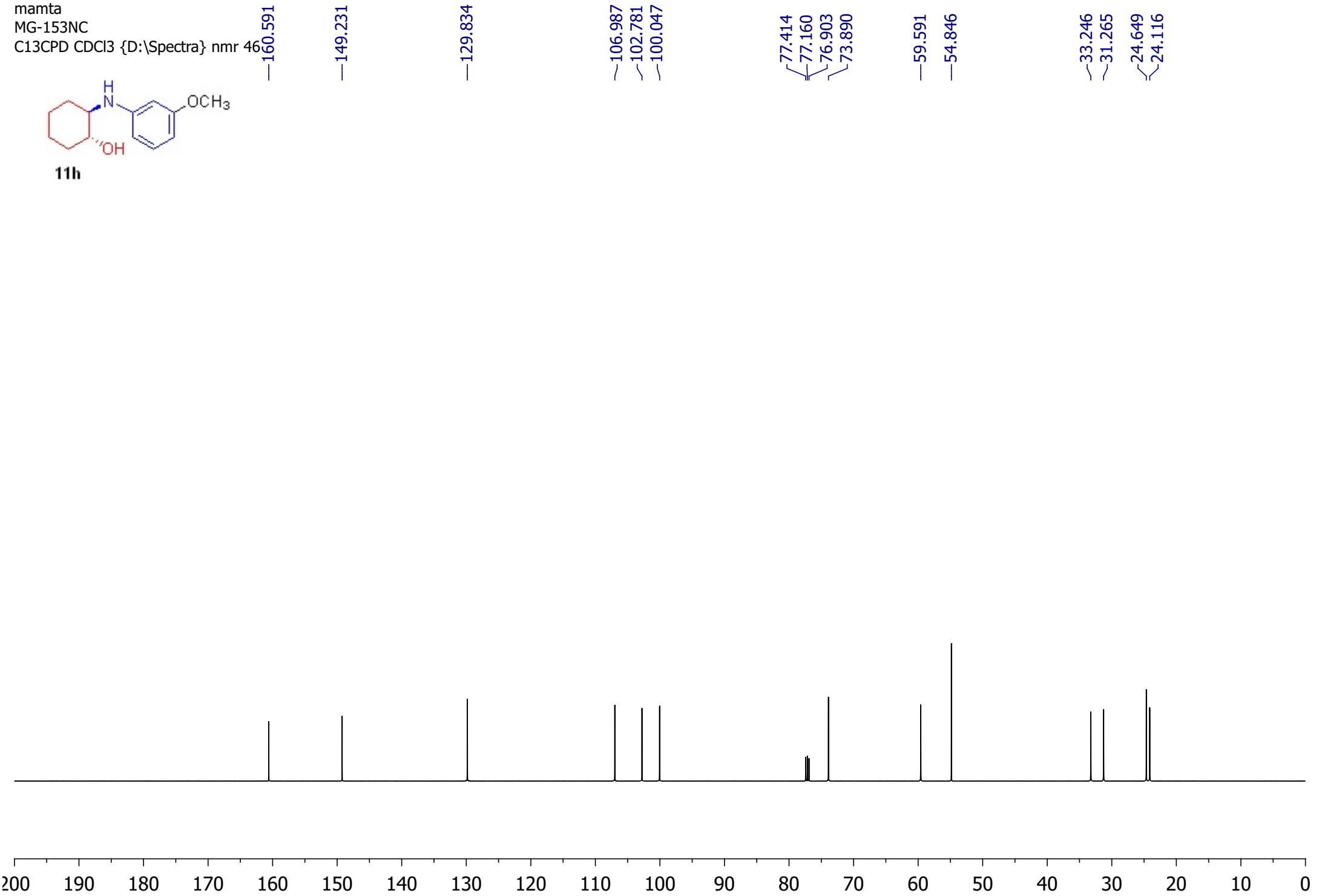


mamta
MG-156NC
C13CPD CDCl3 {D:\Spectra} nmr 44

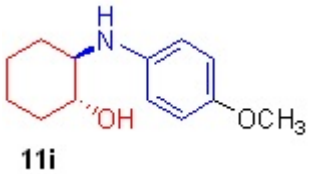




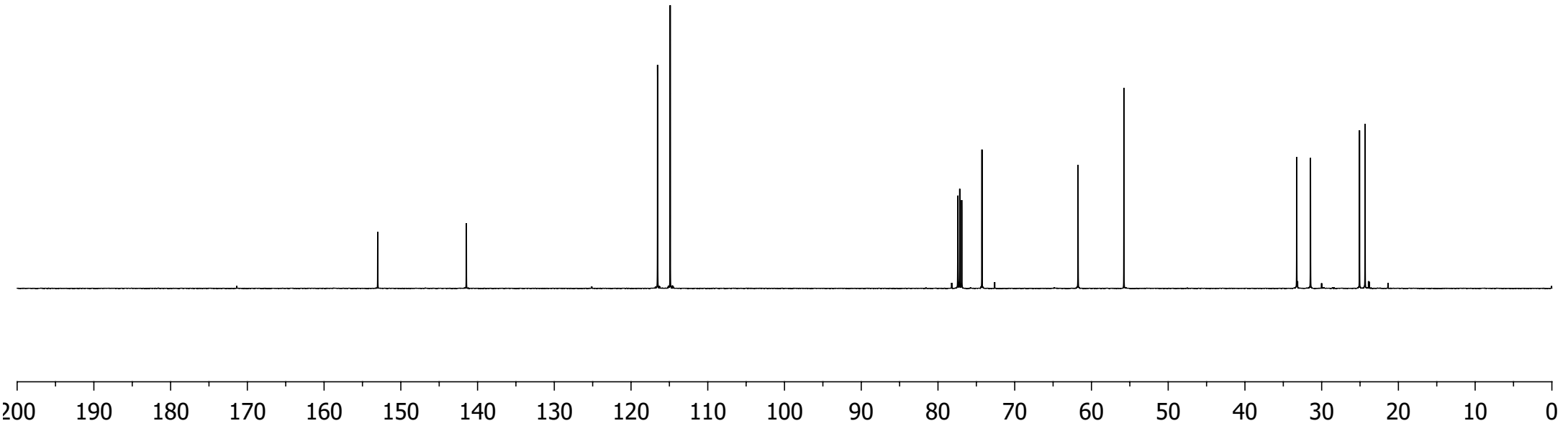
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MG-153NC
C13CPD CDCl3 {D:\Spectra} nmr 46

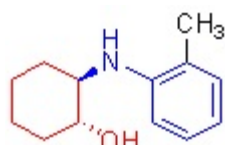


mamta
MG-157NC
C13CPD CDCl3 {D:\Spectra} nmr 45

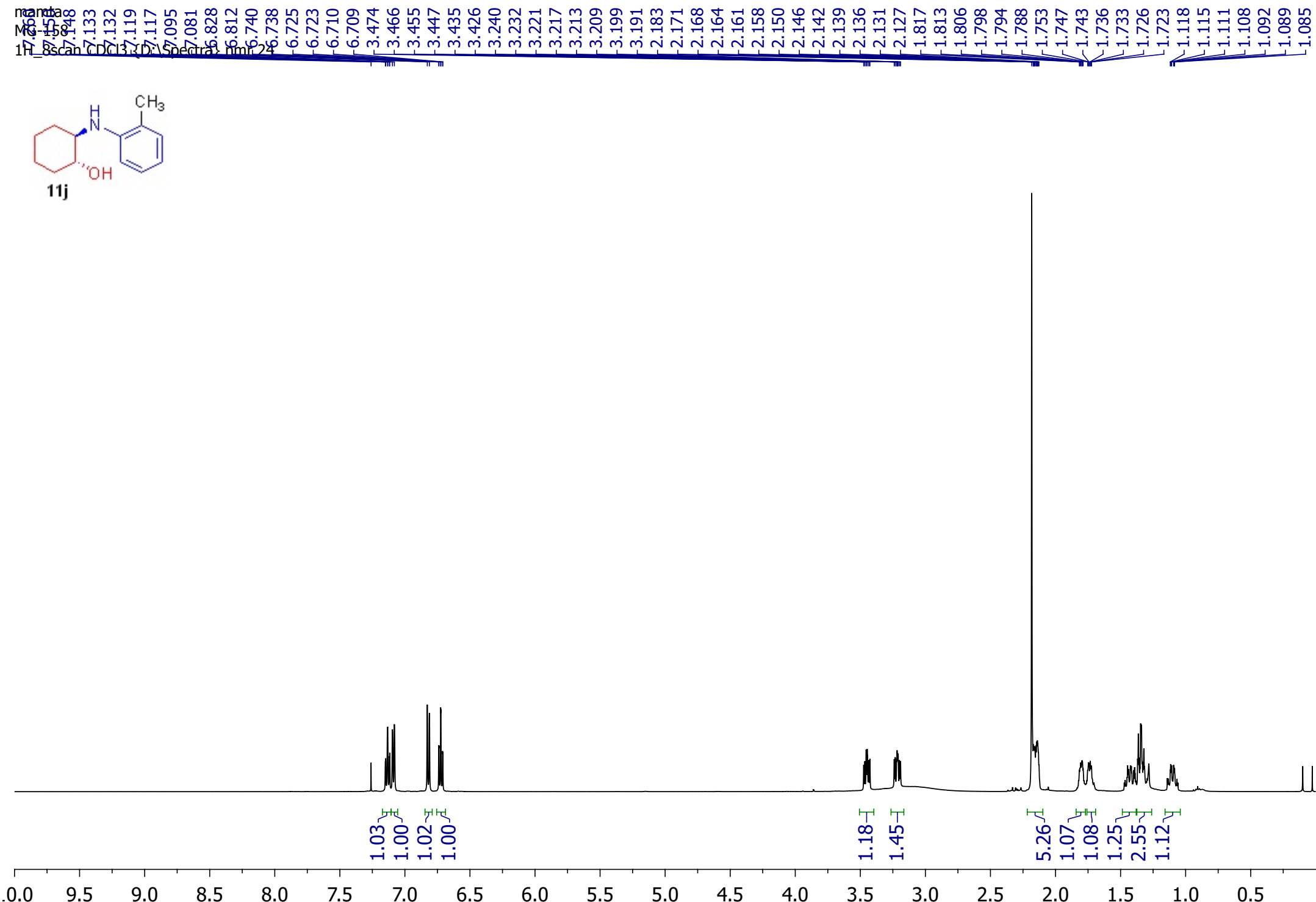


—153.006 —141.463 —116.539
 —114.897 77.414
 —77.160 76.906
 —74.268 —61.747
 —55.771 33.251
 —31.459 25.070
 —24.342

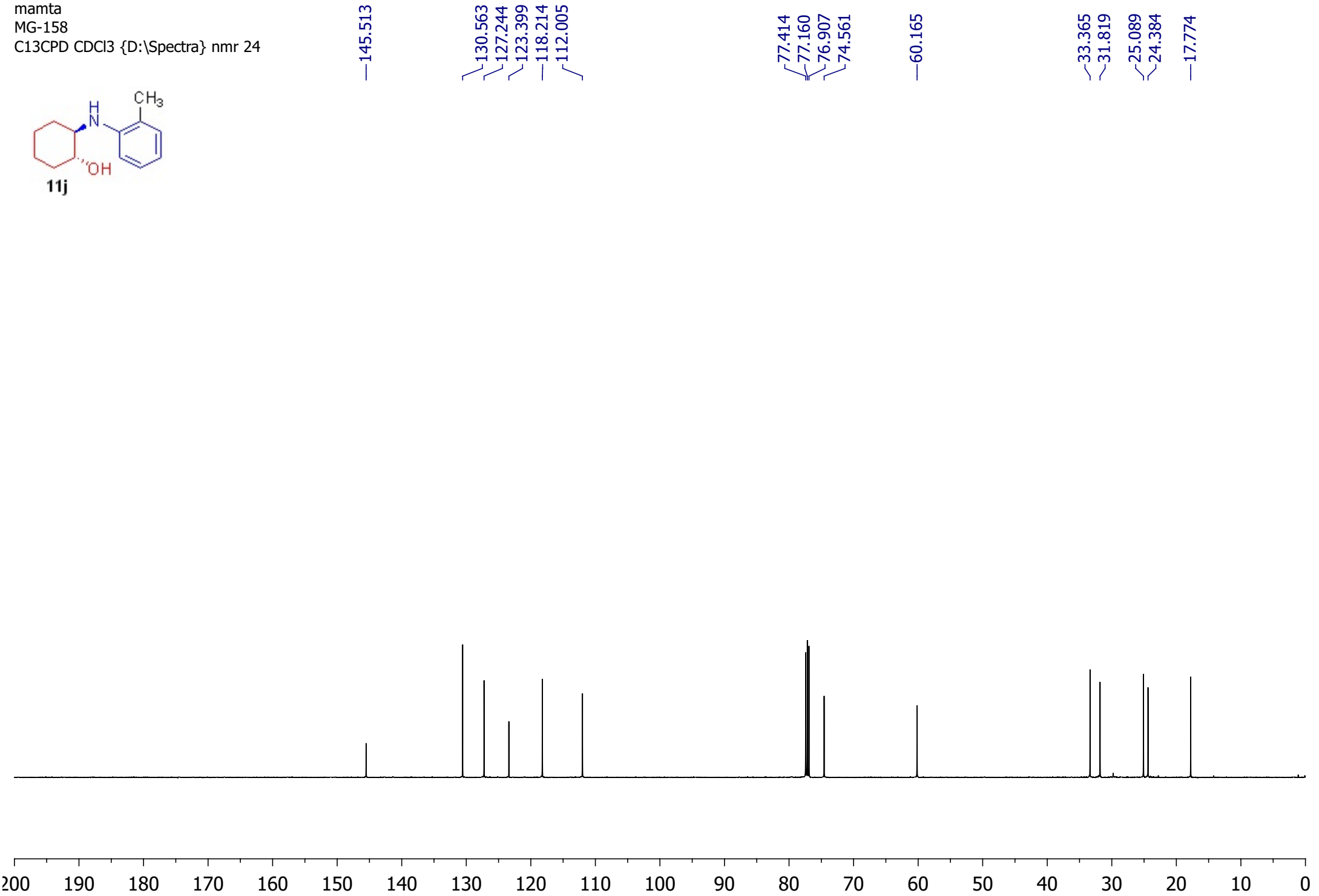
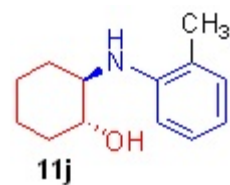




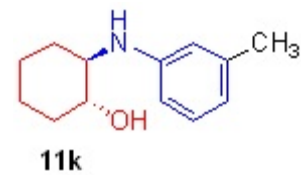
11j



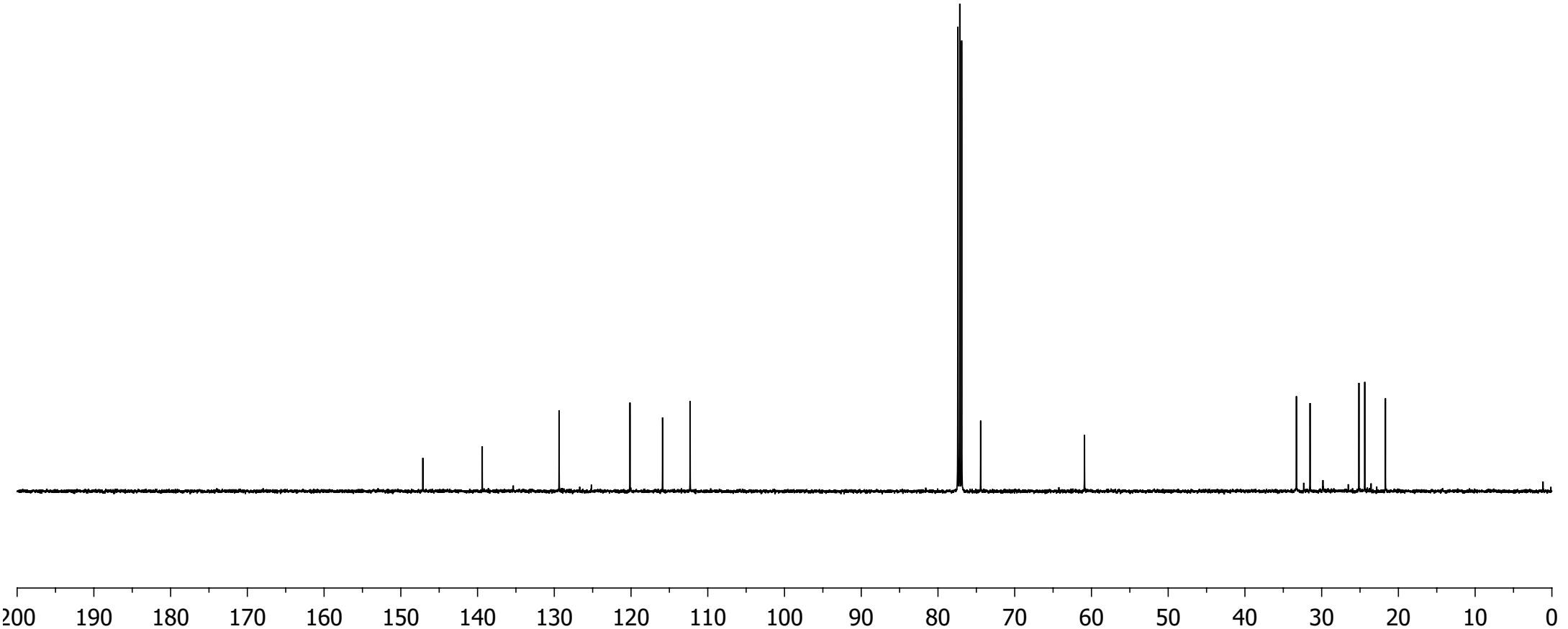
mamta
MG-158
C13CPD CDCl3 {D:\Spectra} nmr 24

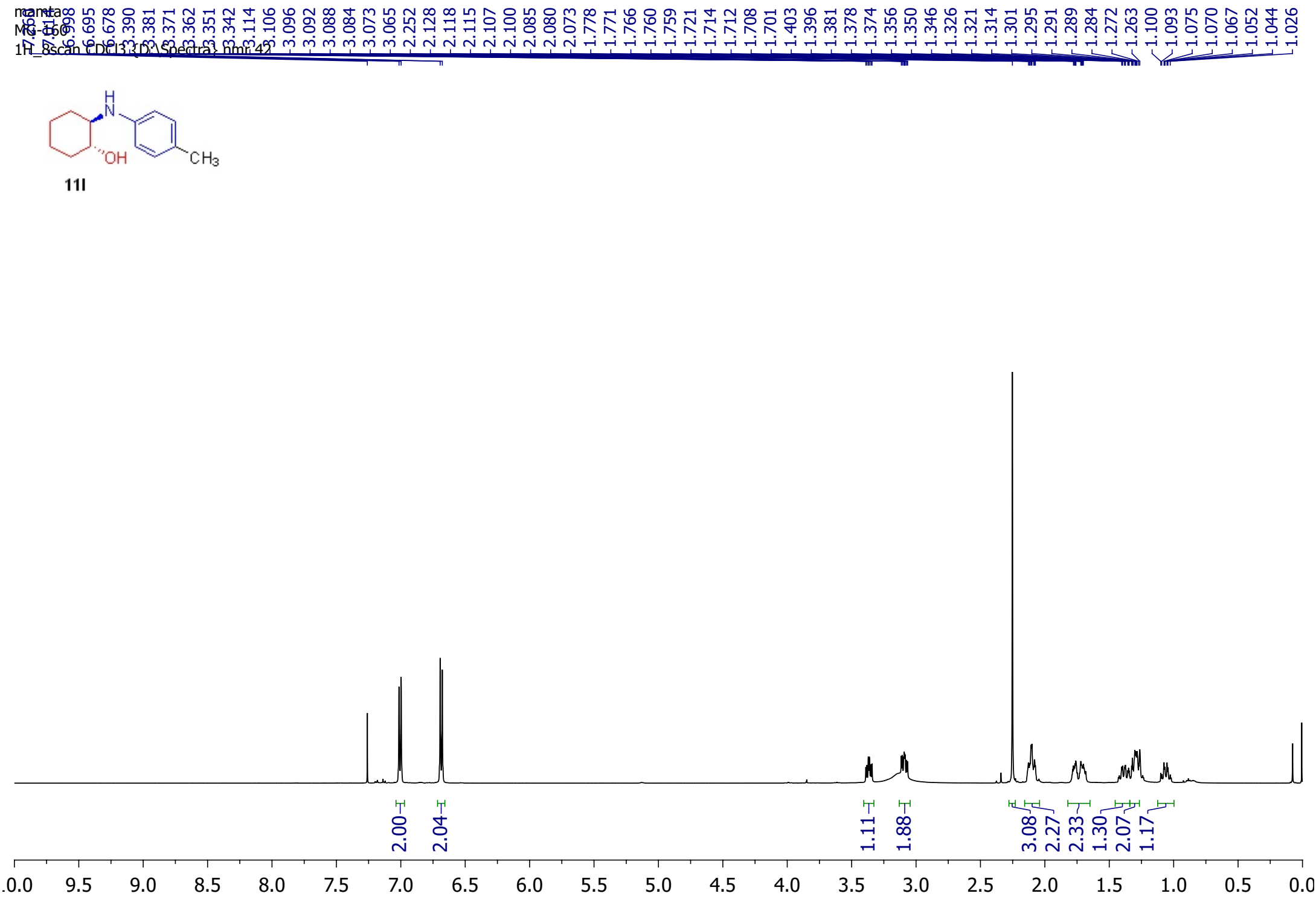


mamta
MG-159
C13CPD CDCl3 {D:\Spectra} nmr 41

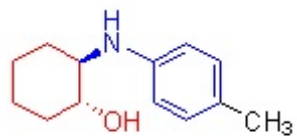


—147.131 —139.390 —129.371 ~120.137
~115.892 ~112.300 {77.413
77.160
76.906
74.441 —60.906 ~33.292
~31.513 ~25.142
~24.386 ~21.691

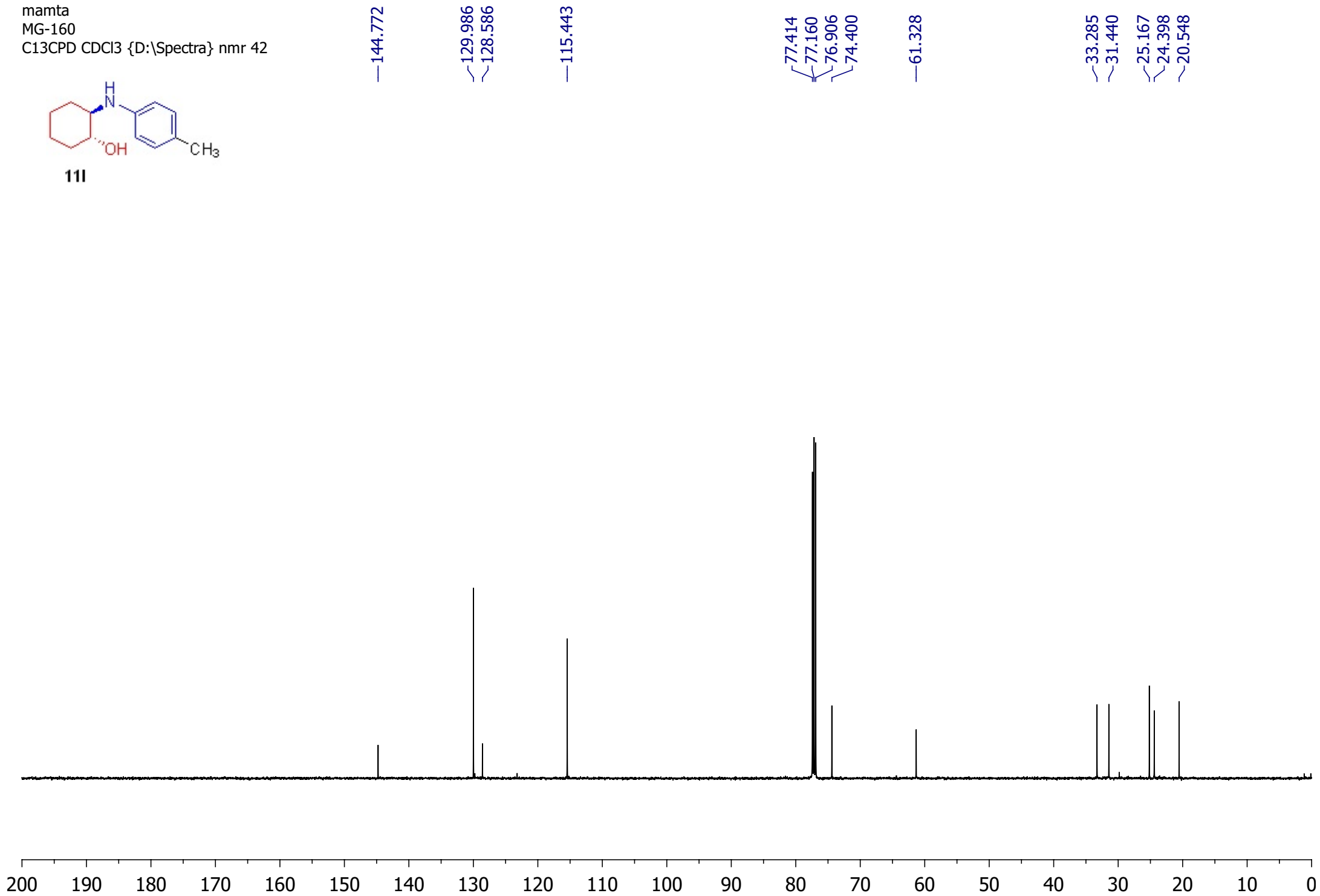


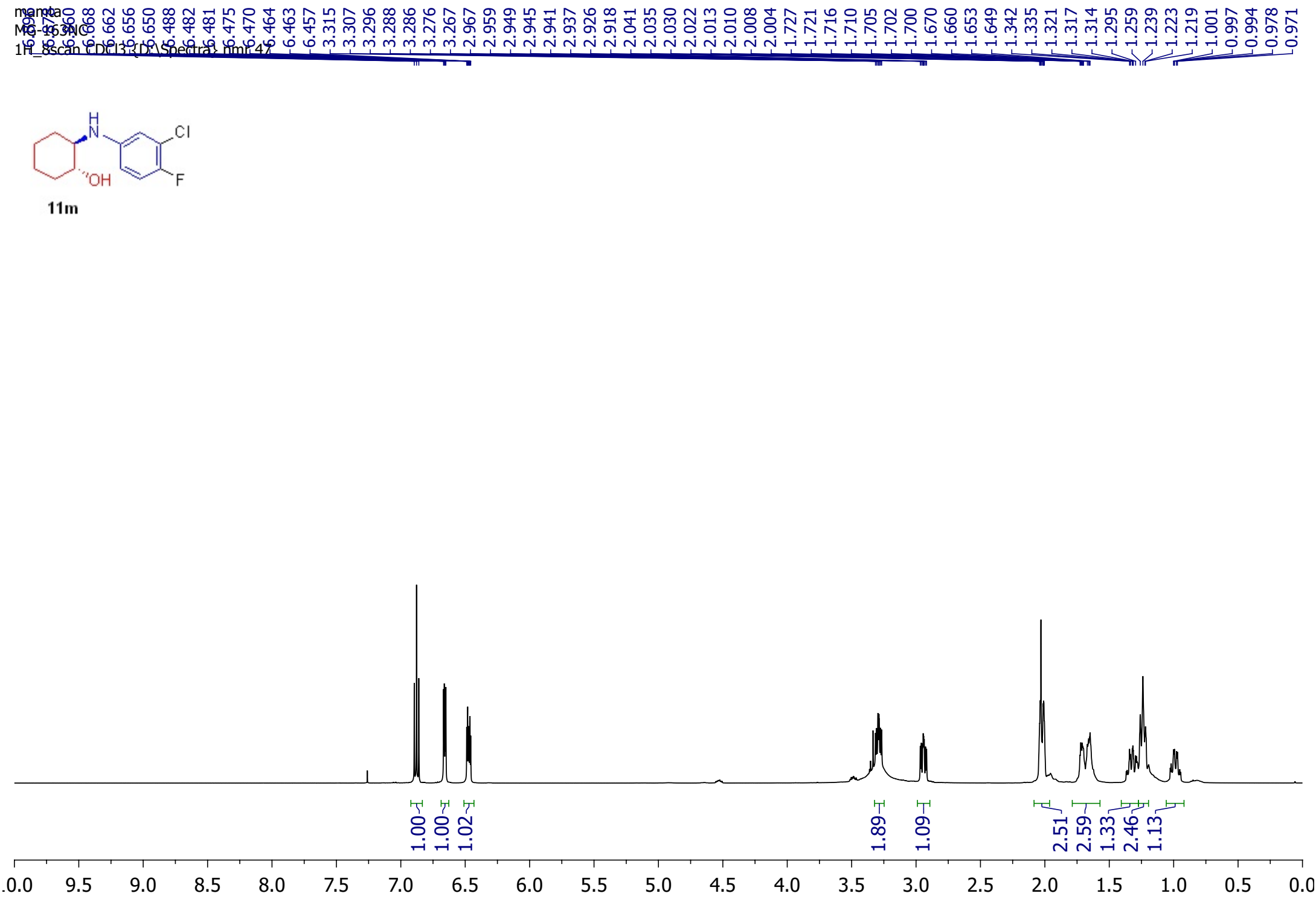


mamta
MG-160
C13CPD CDCl3 {D:\Spectra} nmr 42

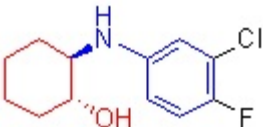


11l





mamta
MG-163NC
C13CPD CDCl3 {D:\Spectra} nmr 47



11m

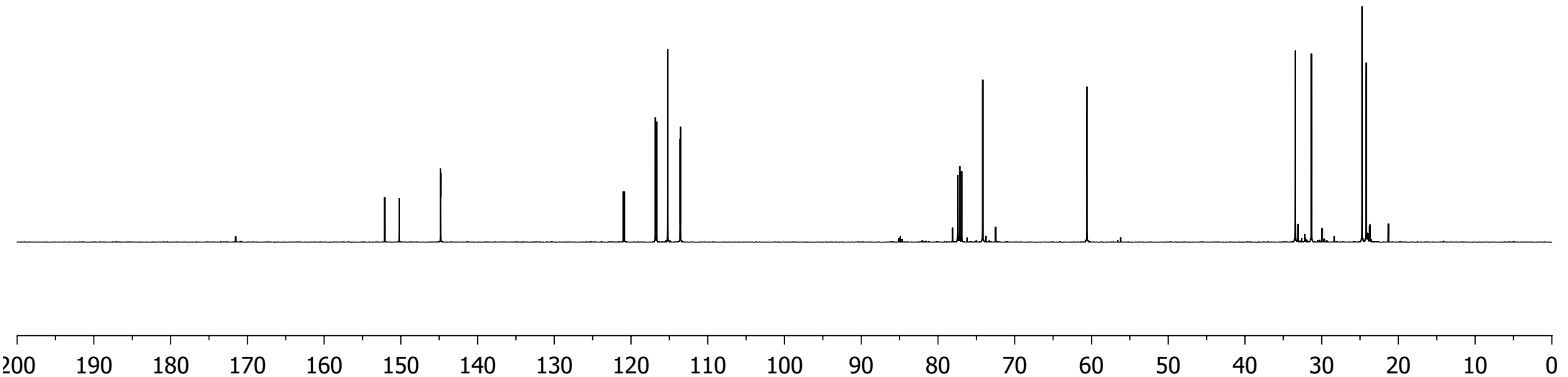
152.105
150.209
144.824
144.809

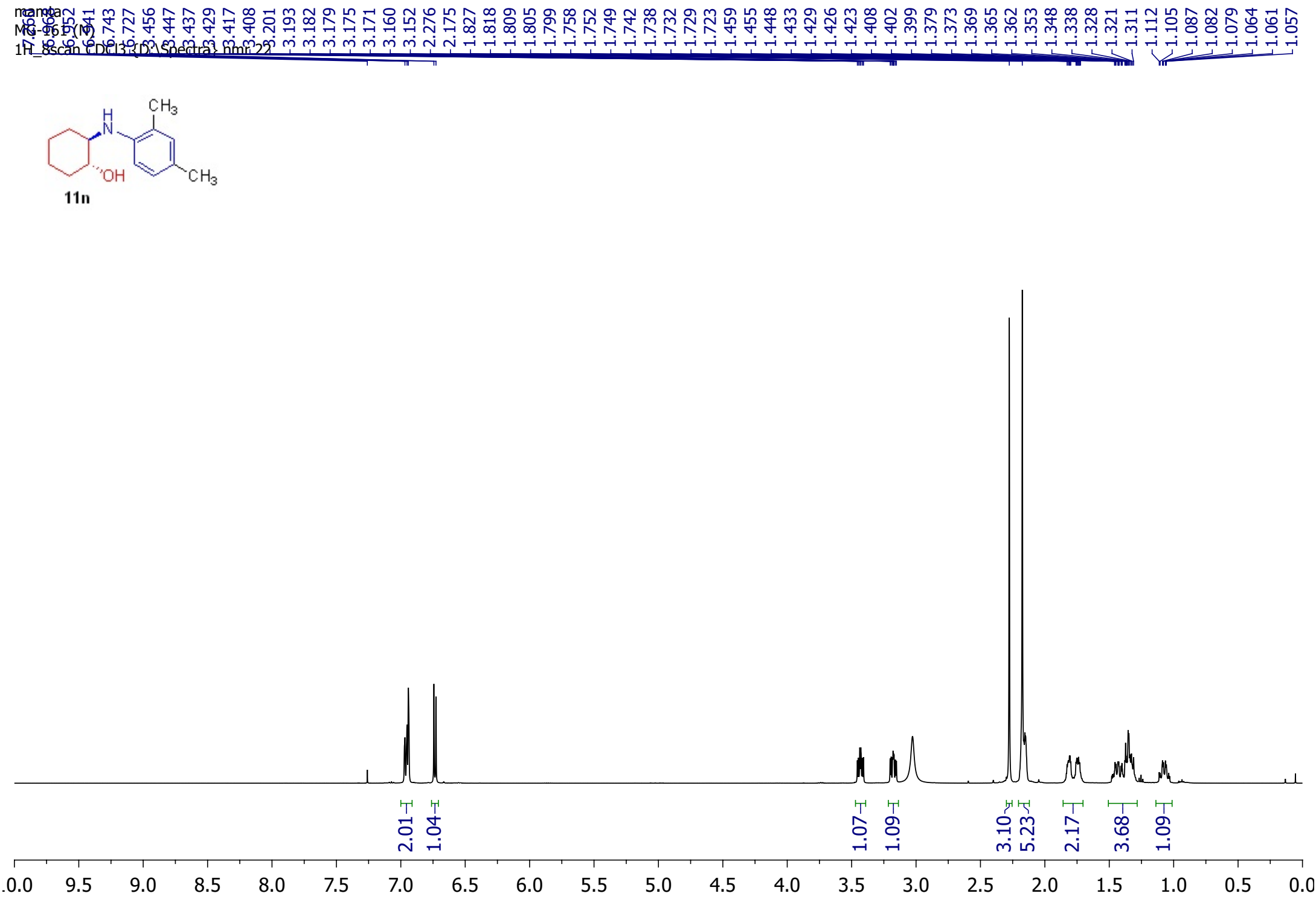
121.000
120.855
116.839
116.667
115.214
113.596
113.549

77.414
77.160
76.906
74.171

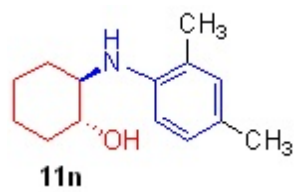
60.590

33.447
31.332
24.746
24.201





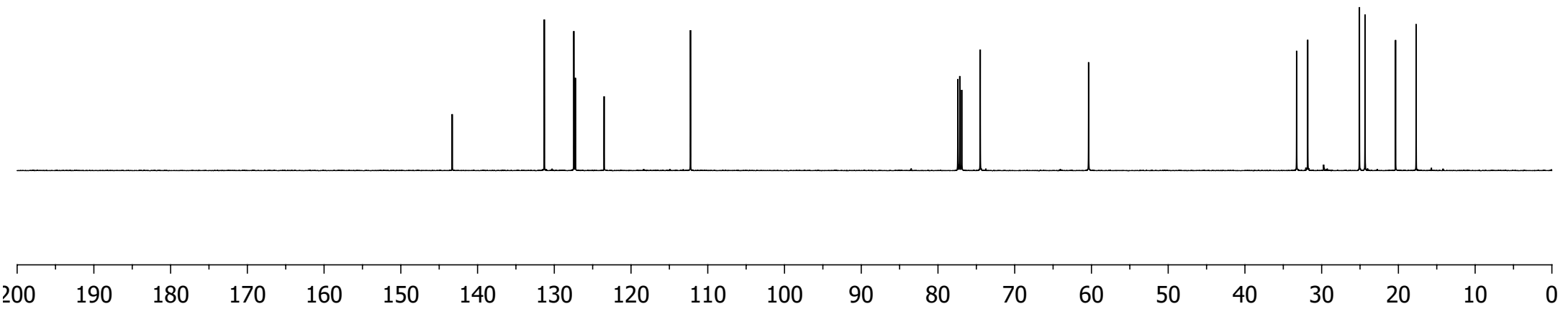
mamta
MG-161 (N)
C13CPD CDCl3 {D:\Spectra} nmr 22



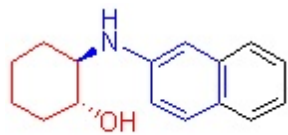
—143.303
~131.299
~127.452
~127.256
~123.510
—112.255

77.413
77.160
76.905
74.493
—60.372

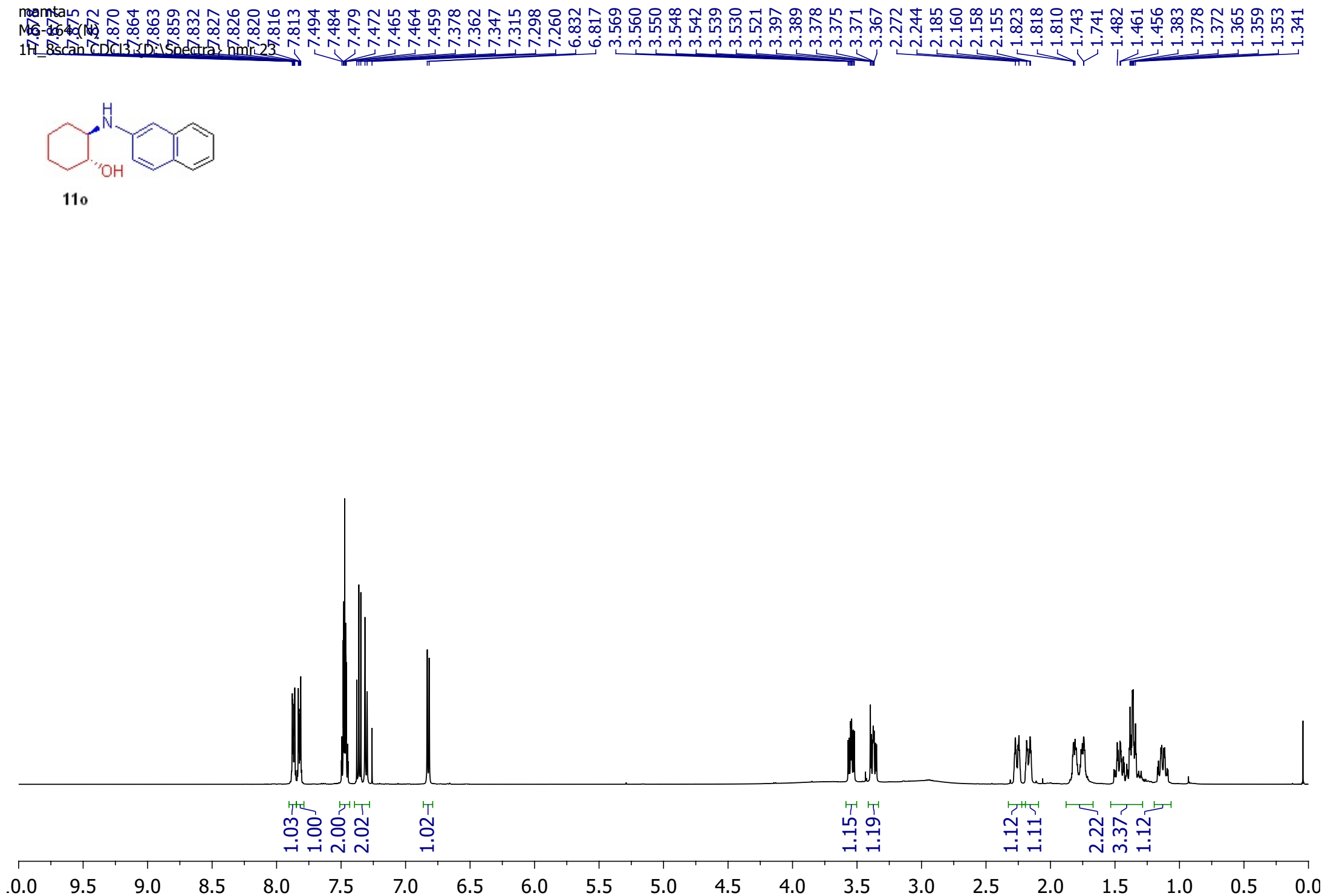
33.263
31.819
25.079
24.351
20.371
17.690



name: 11o
Mg-64016
1H-8scan CDCl3 (D:\Spectra\hmr_23



11o



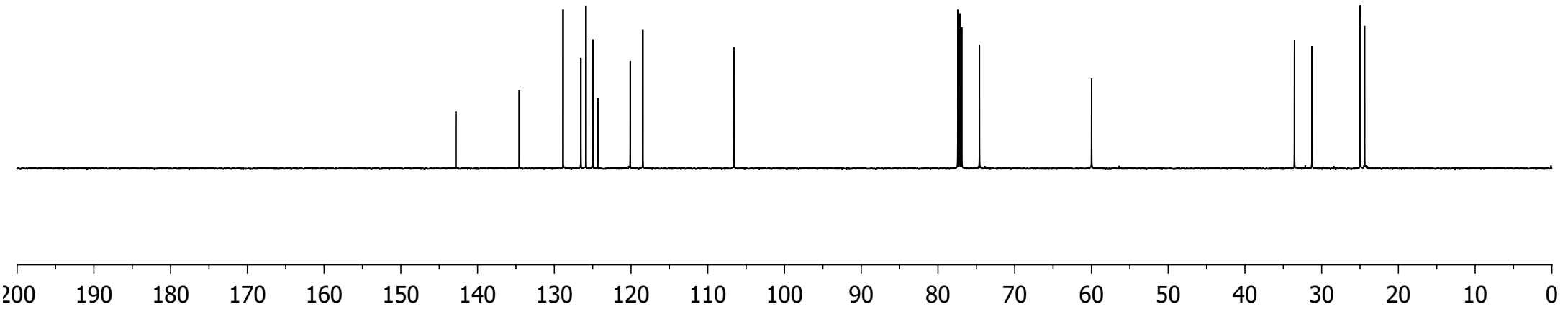
mamta
MG-164 (N)
C13CPD CDCl3 {D:\Spectra} nmr 23



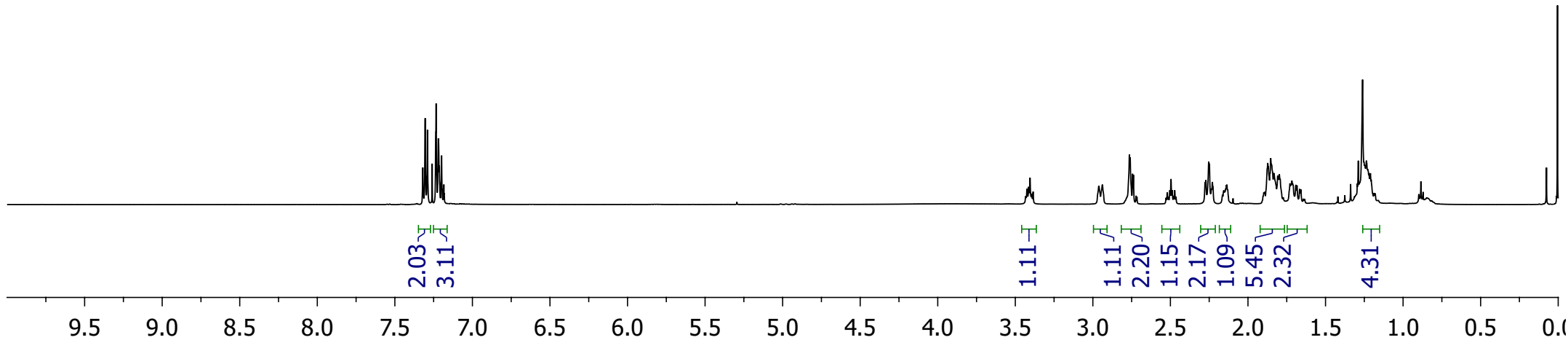
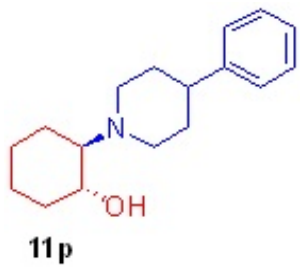
—142.825
134.573
128.857
126.533
125.864
124.976
124.345
120.108
118.458
—106.596

77.413
77.160
76.905
74.591
—59.983

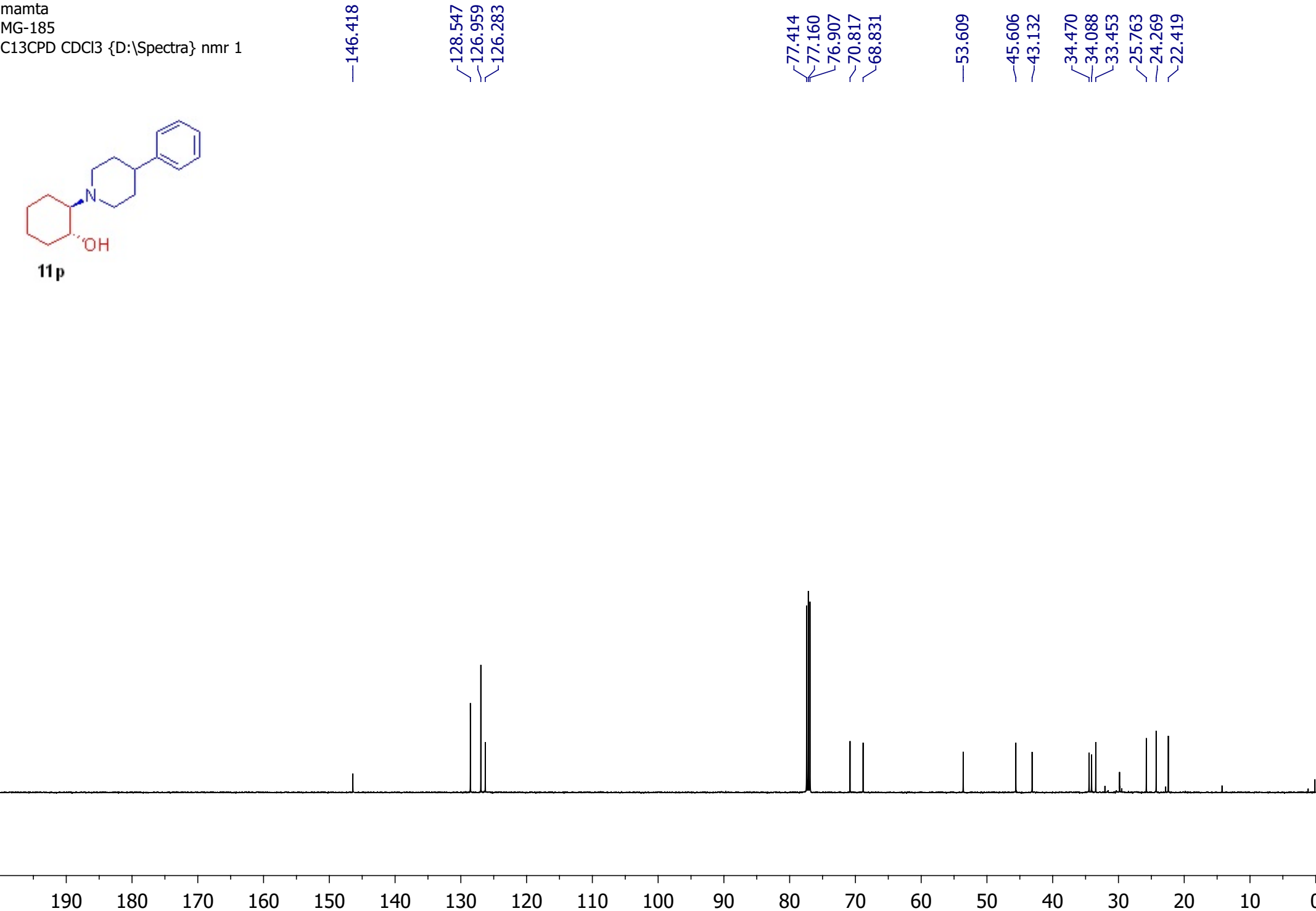
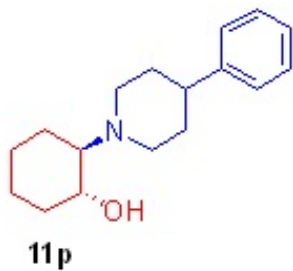
33.548
31.262
24.972
24.405

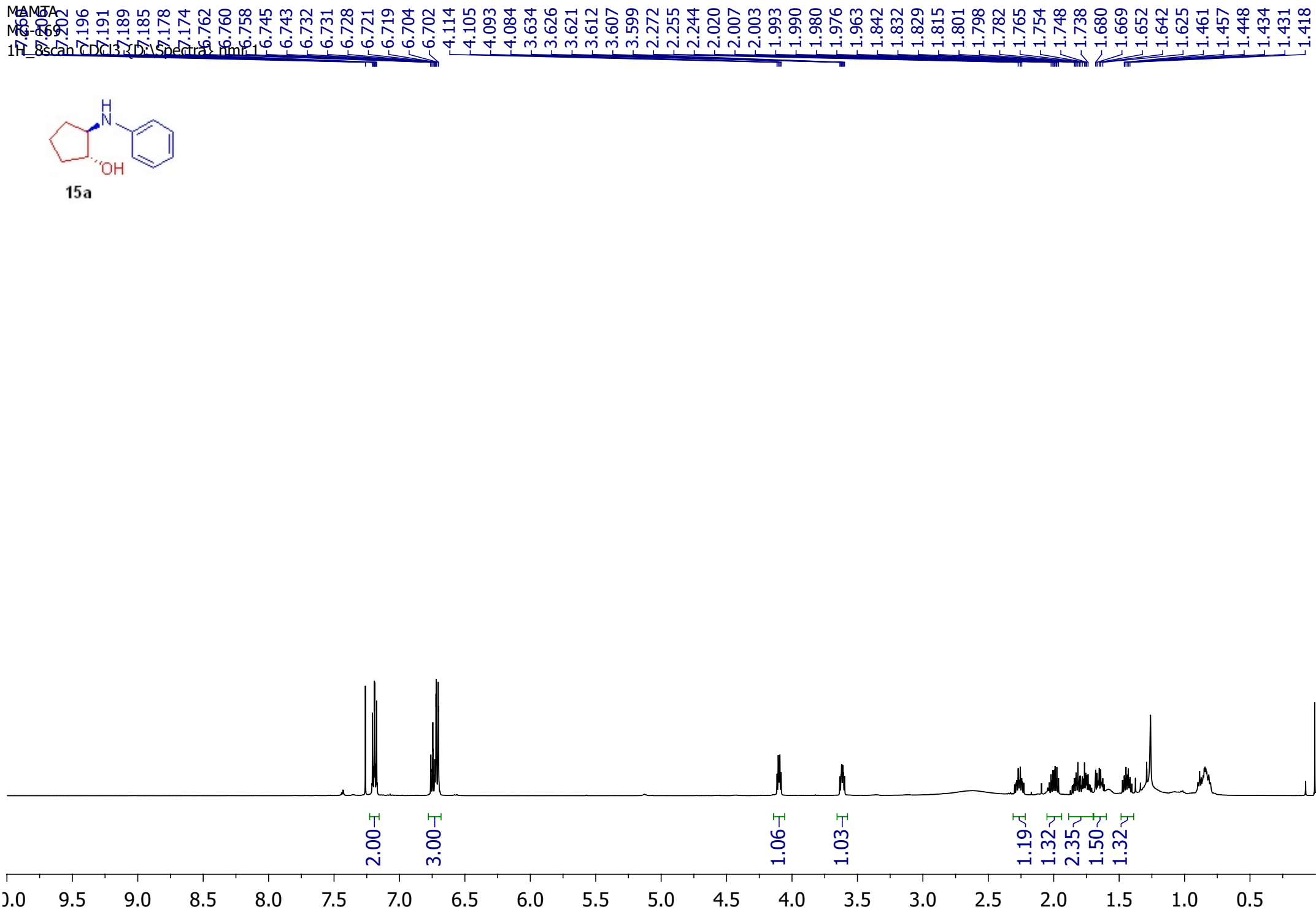


name: 11p
 Mol. Weight: 248.35
 1H NMR scan CDCl3 (DMSO-d6) Spectra: nmr1

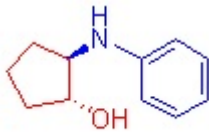


mamta
MG-185
C13CPD CDCl3 {D:\Spectra} nmr 1

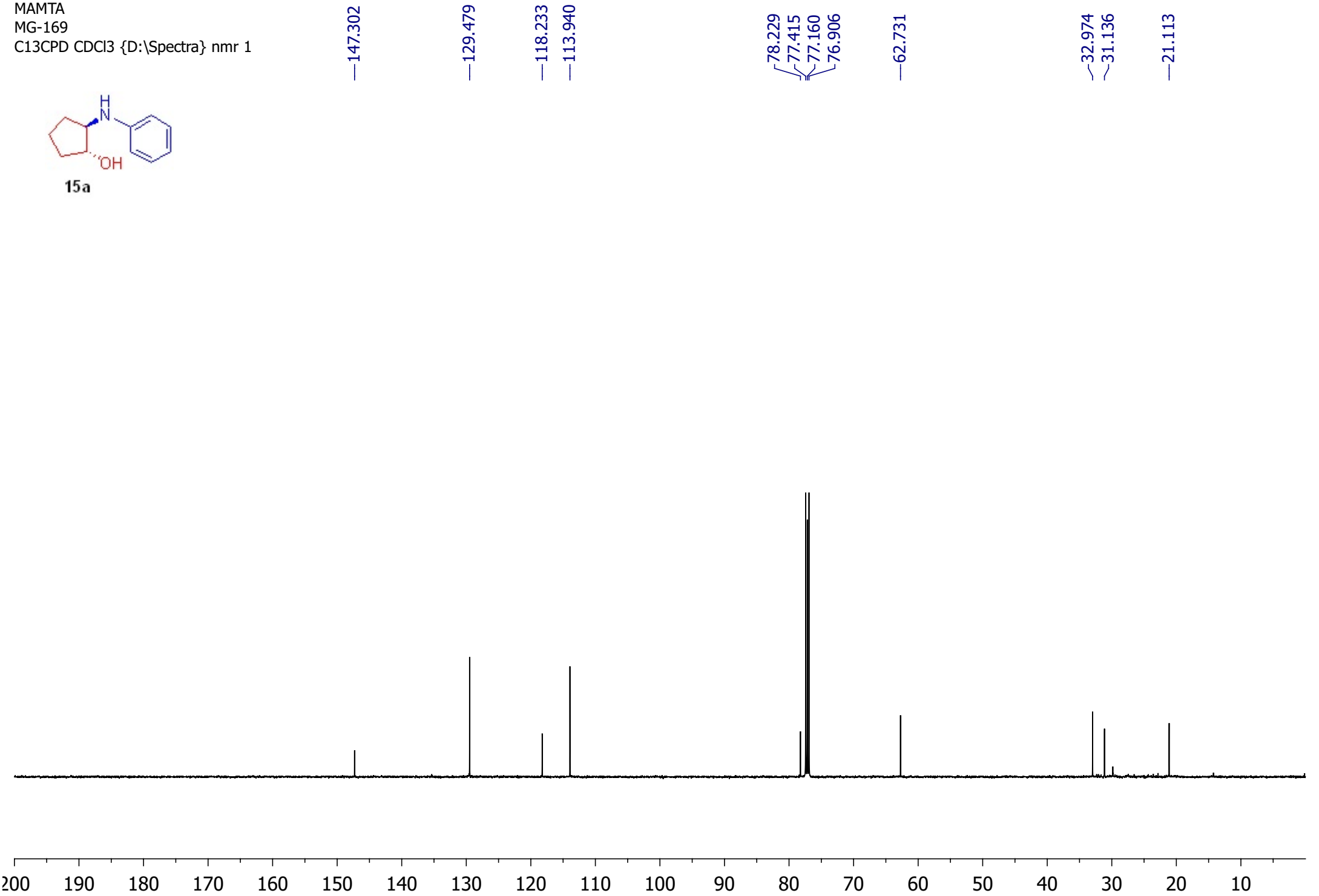




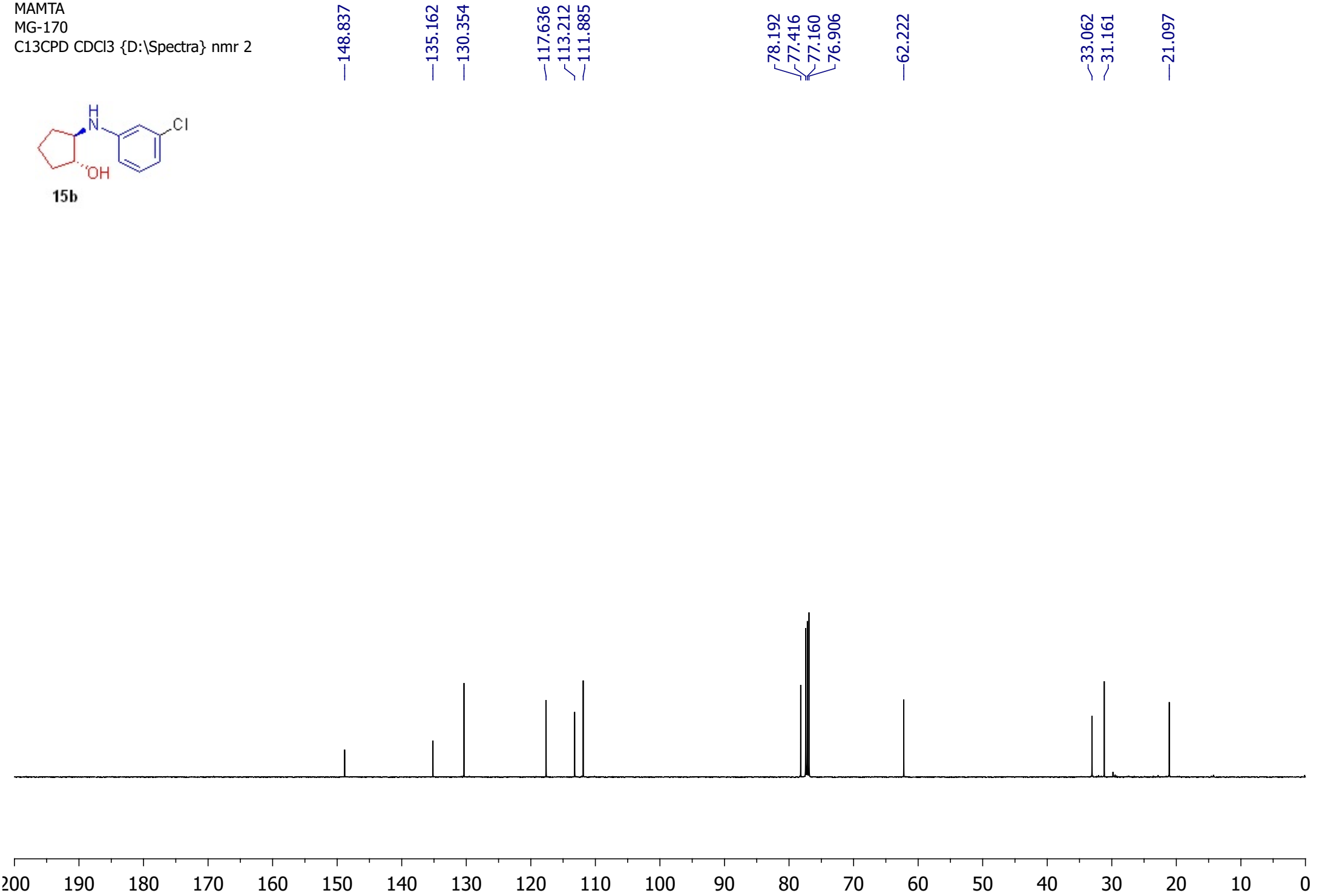
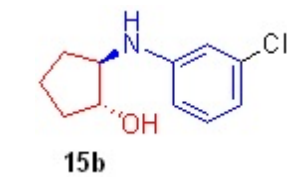
MAMTA
MG-169
C13CPD CDCl3 {D:\Spectra} nmr 1



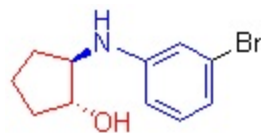
15a



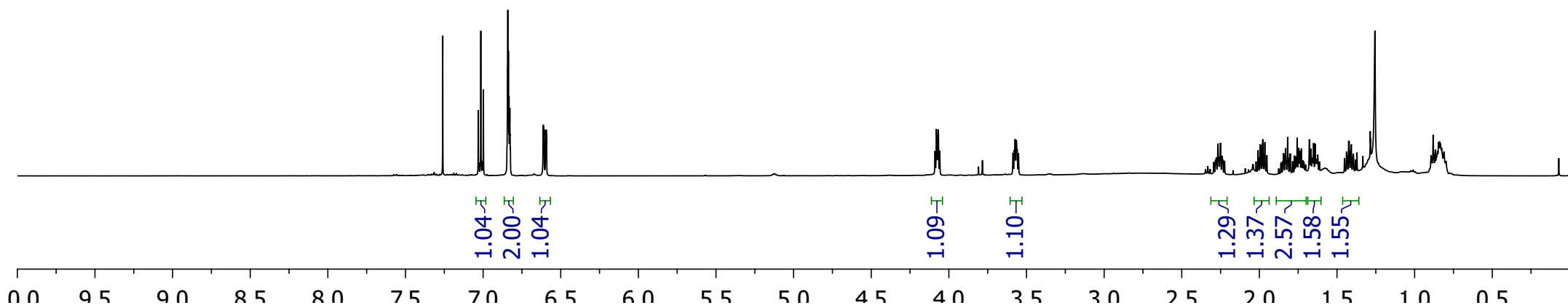
MAMTA
MG-170
C13CPD CDCl3 {D:\Spectra} nmr 2



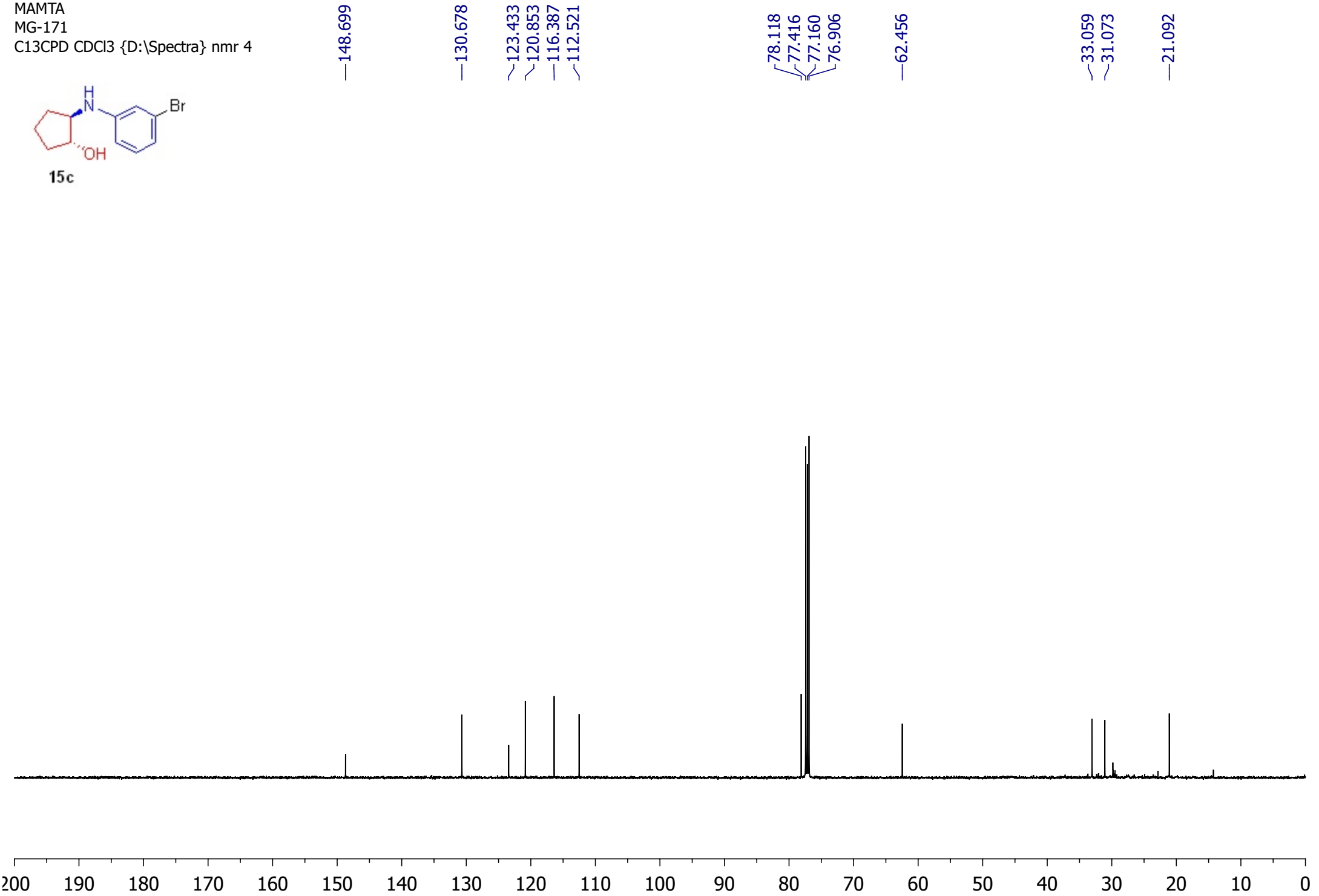
MAMTA
NMR
1H
8scan CDCl3
15c
ppm

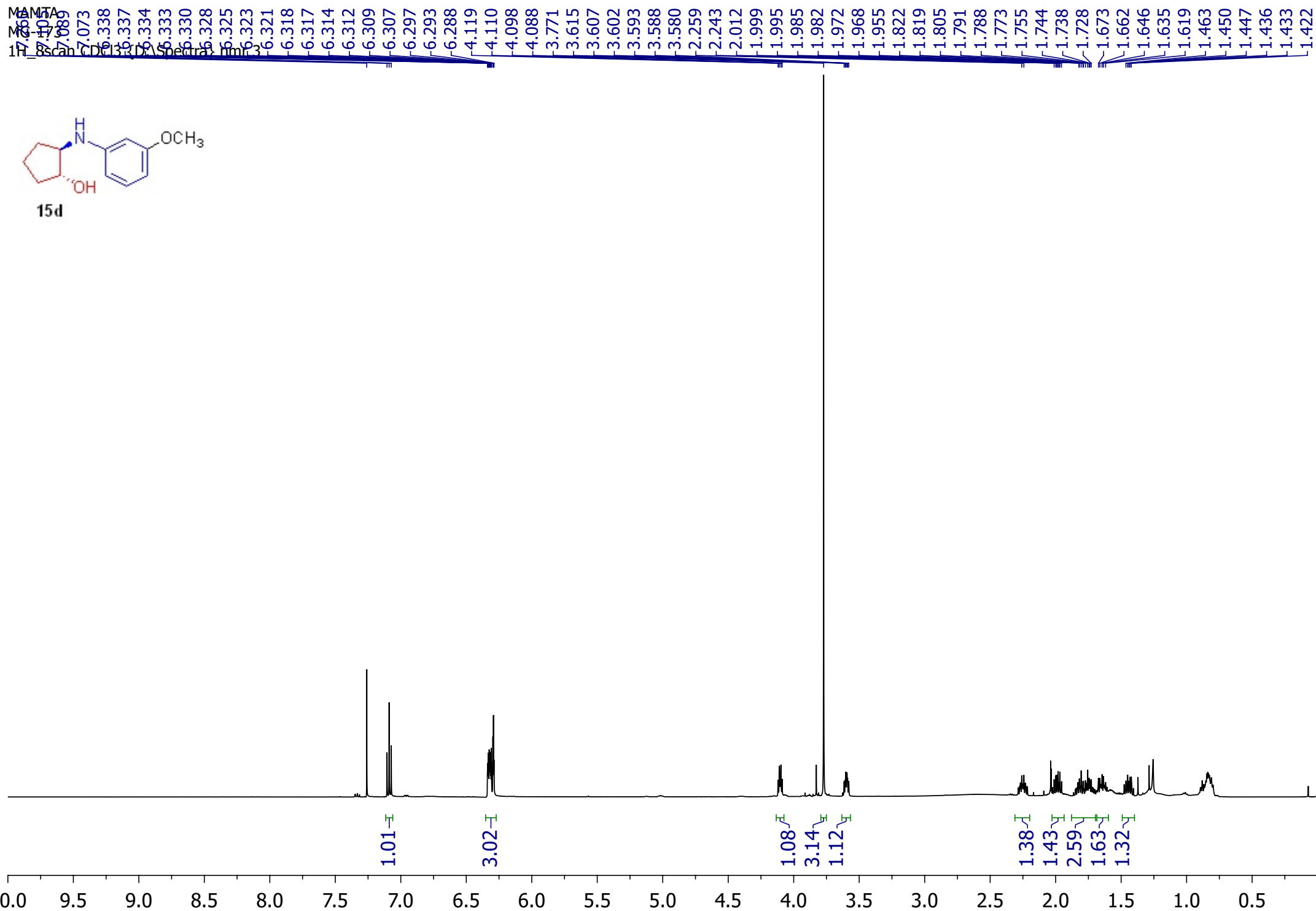


15c

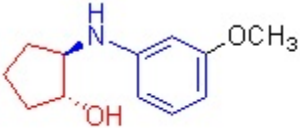


MAMTA
MG-171
C13CPD CDCl3 {D:\Spectra} nmr 4

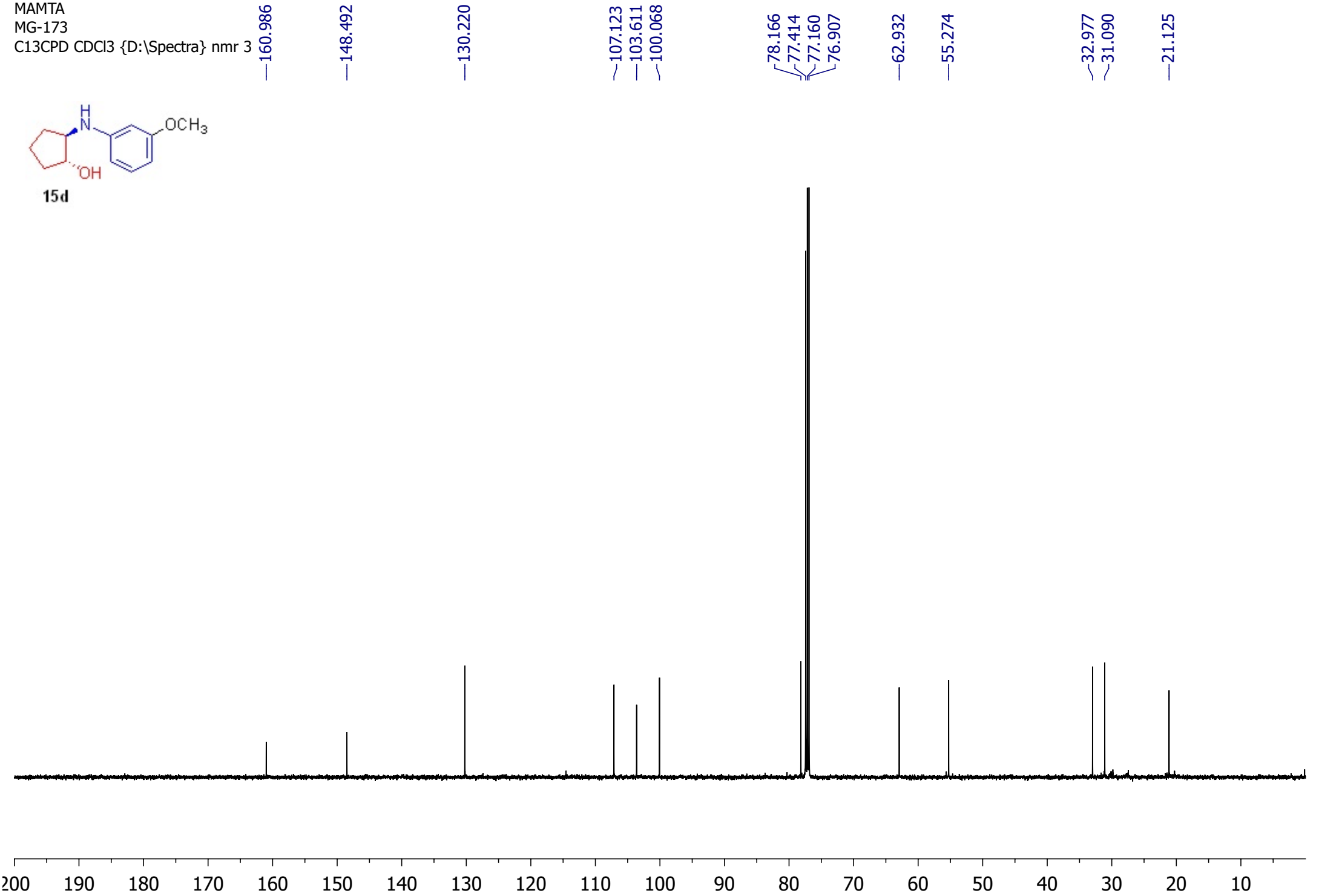




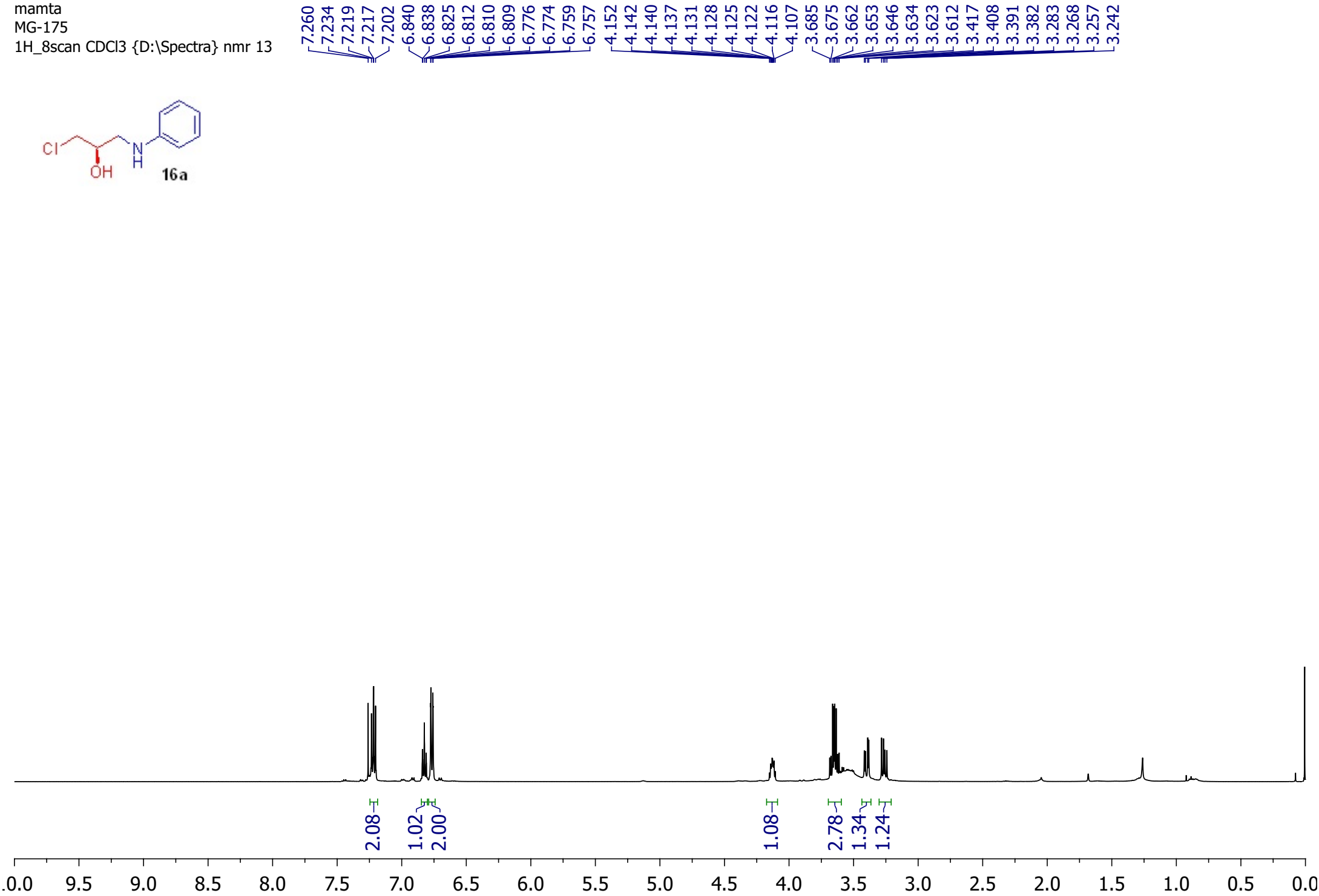
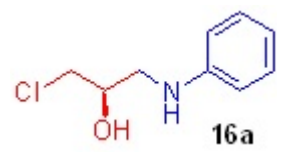
MAMTA
MG-173
C13CPD CDCl3 {D:\Spectra} nmr 3



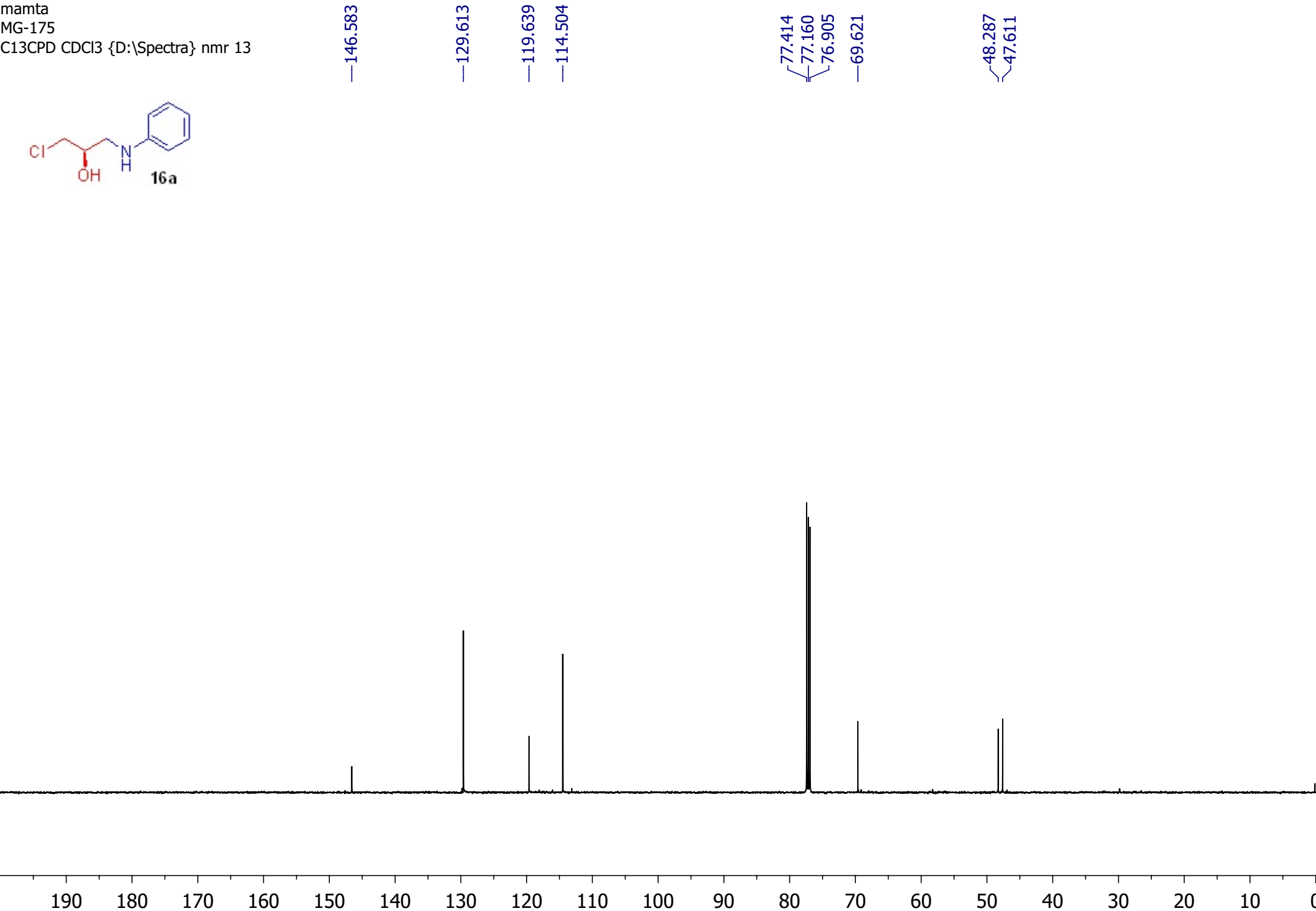
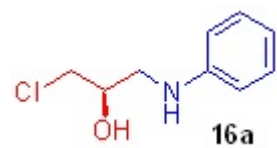
15d



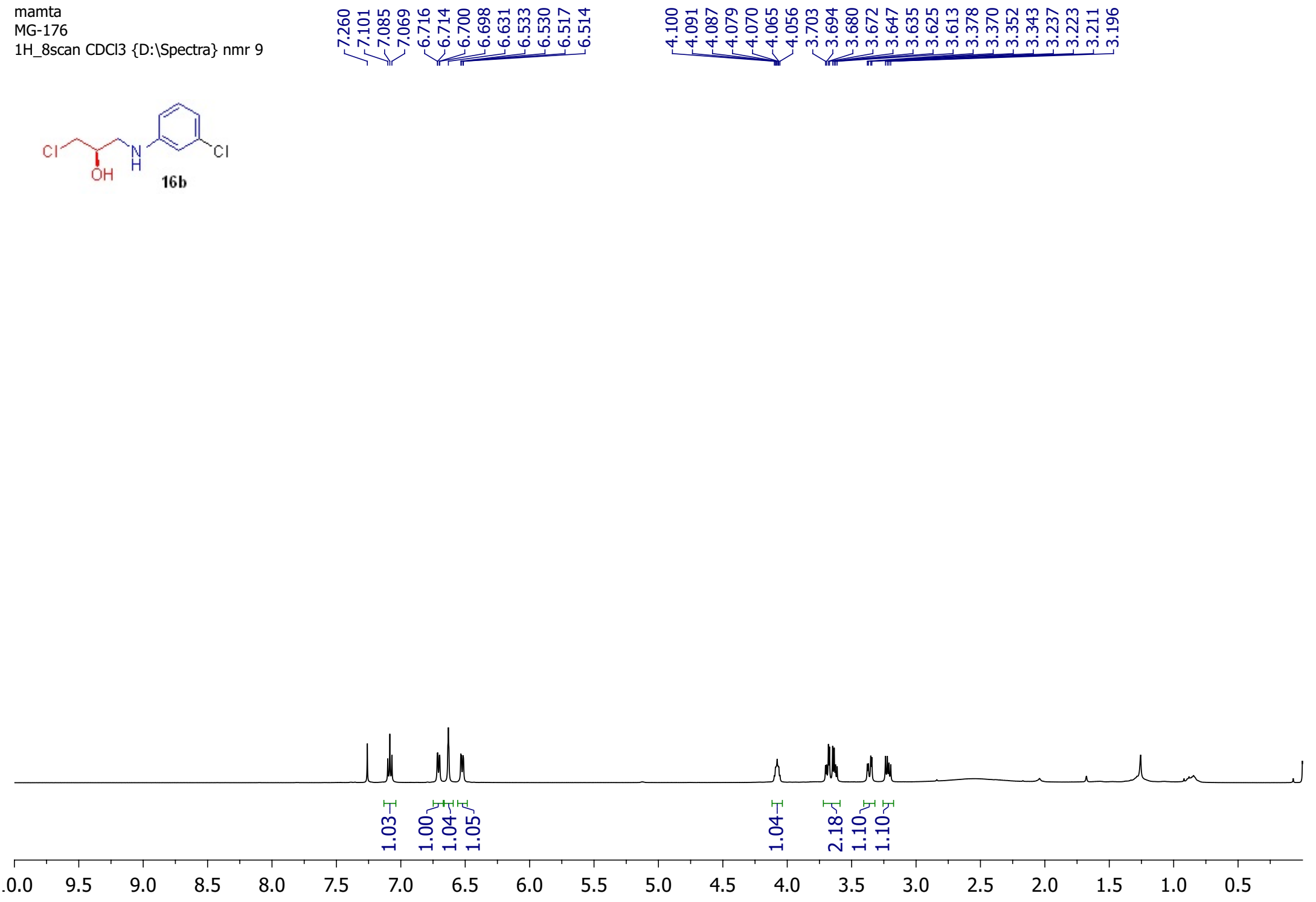
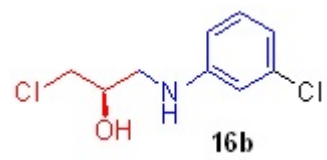
mamta
MG-175
1H_8scan CDCl3 {D:\Spectra} nmr 13



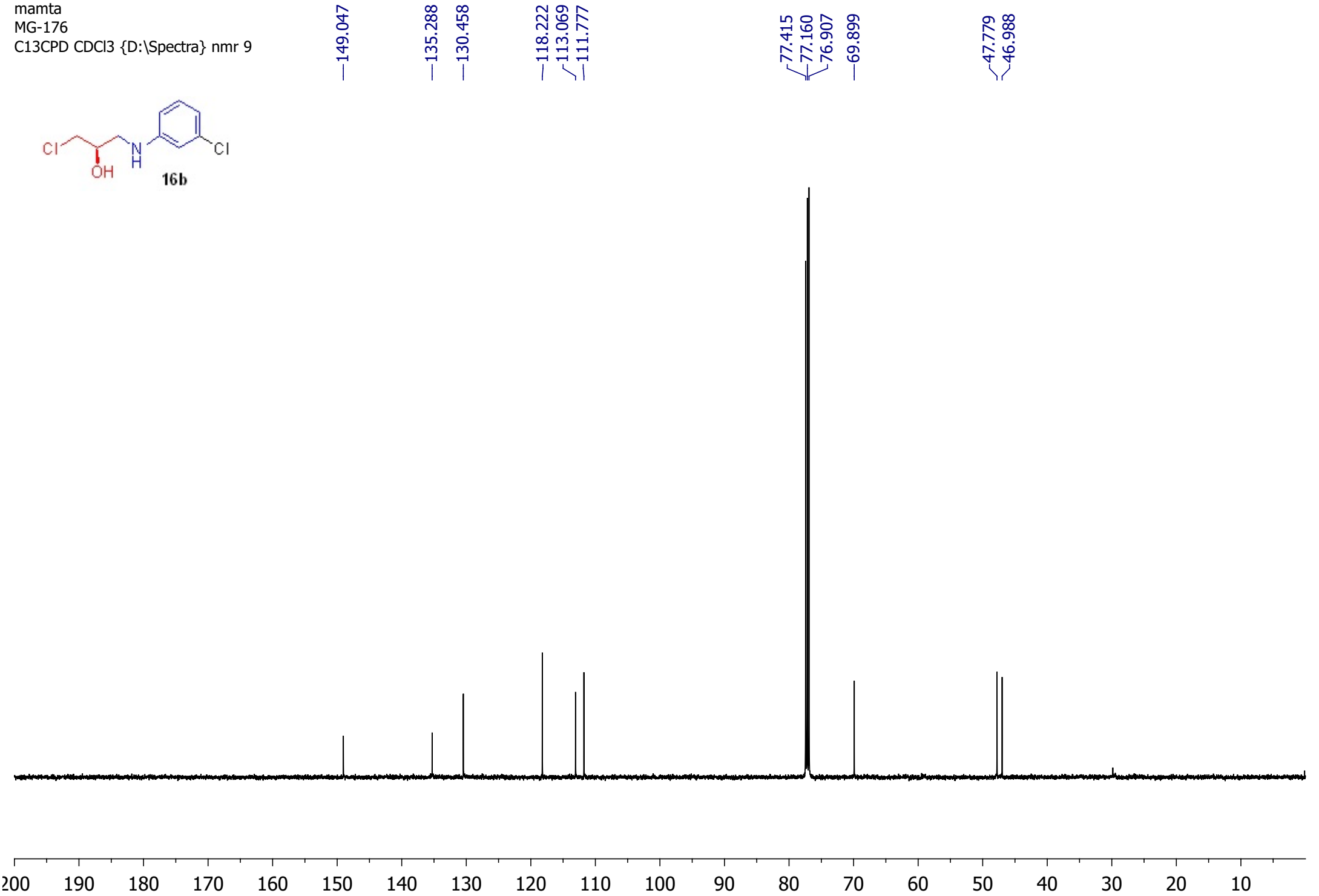
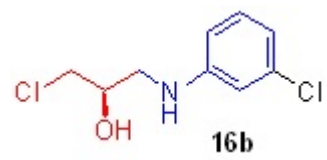
mamta
MG-175
C13CPD CDCl3 {D:\Spectra} nmr 13



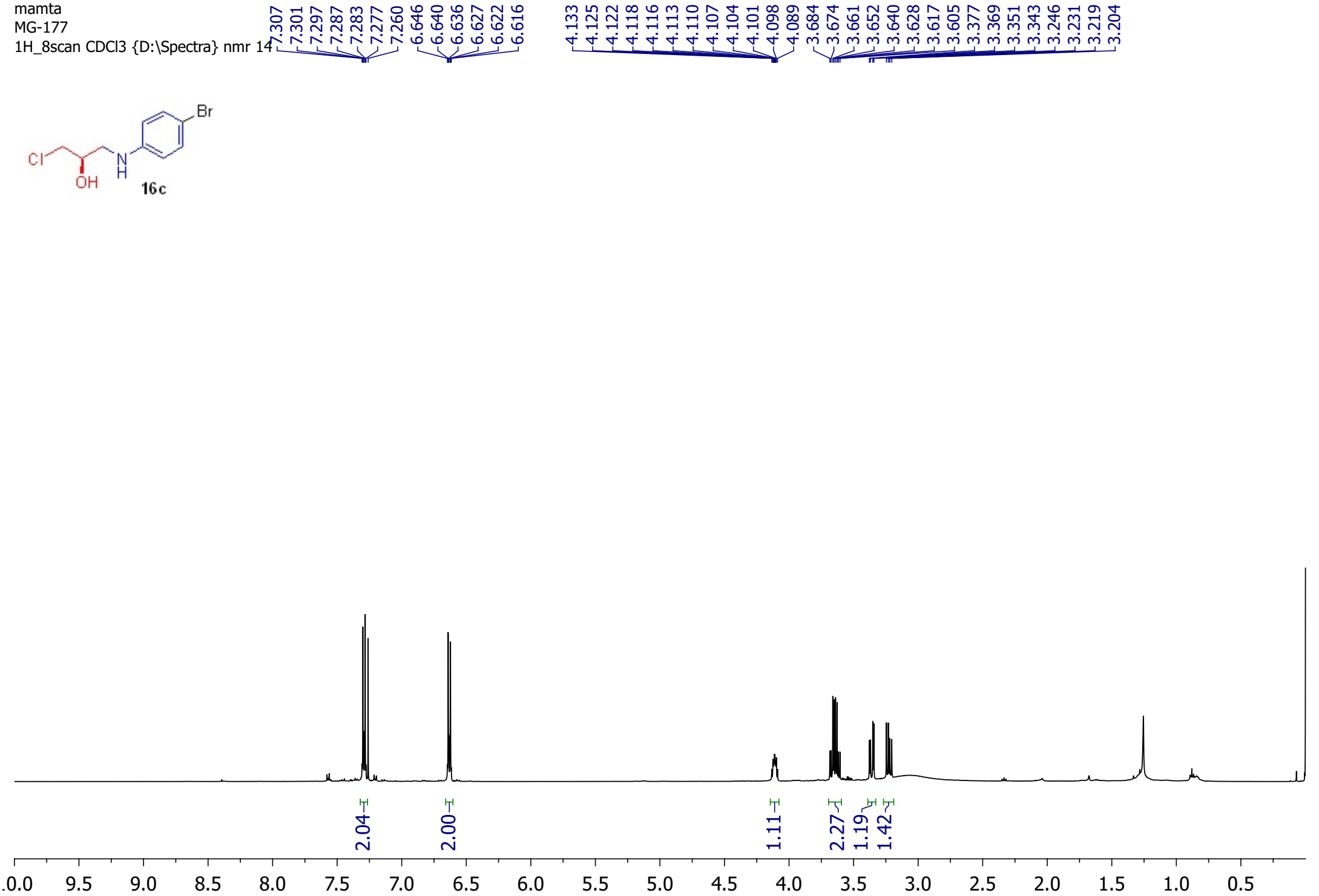
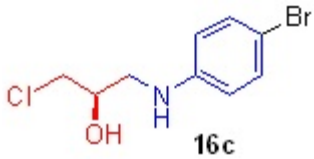
mamta
MG-176
1H_8scan CDCl3 {D:\Spectra} nmr 9



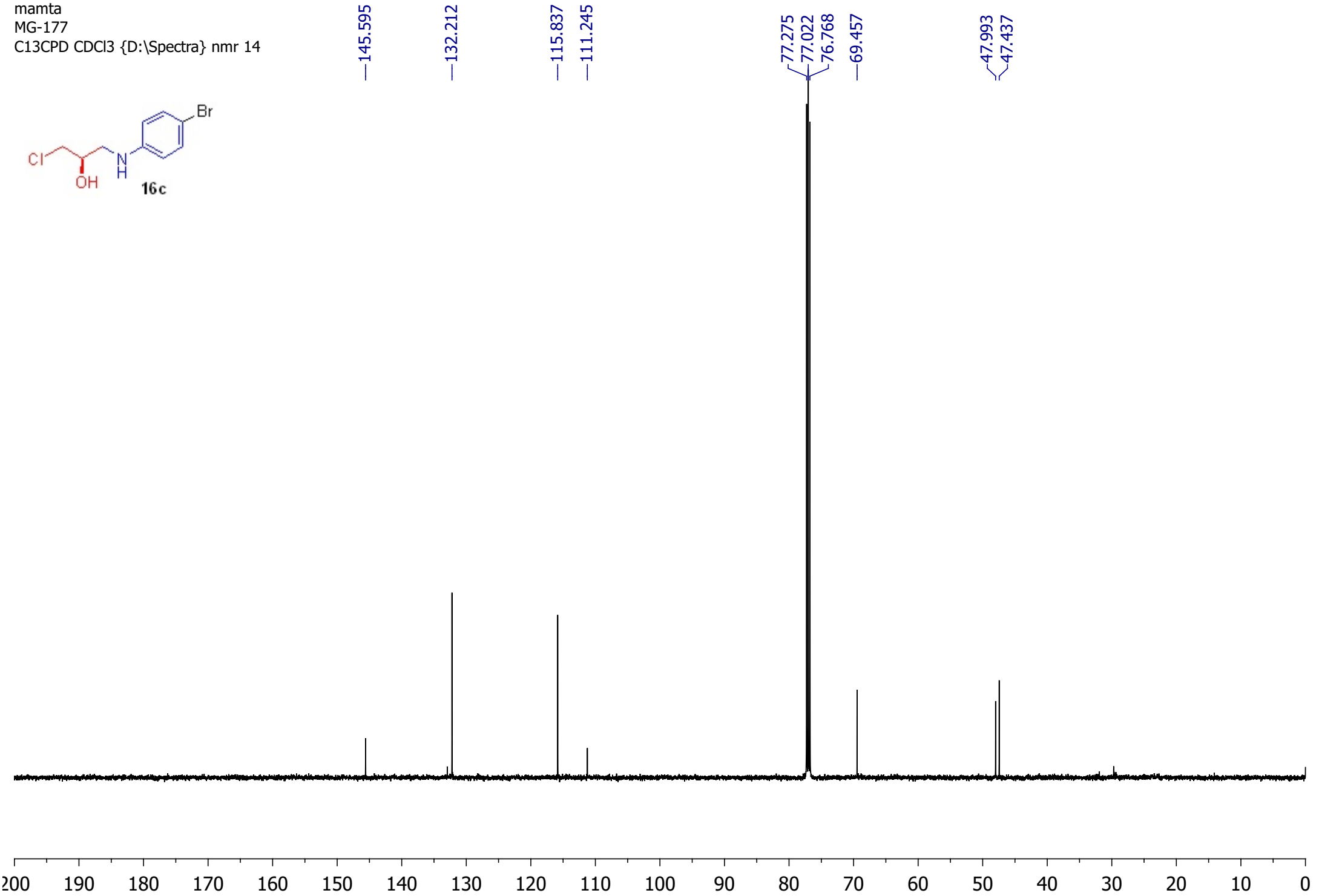
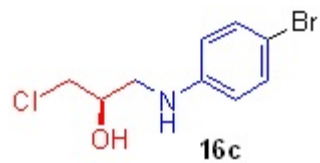
mamta
MG-176
C13CPD CDCl3 {D:\Spectra} nmr 9



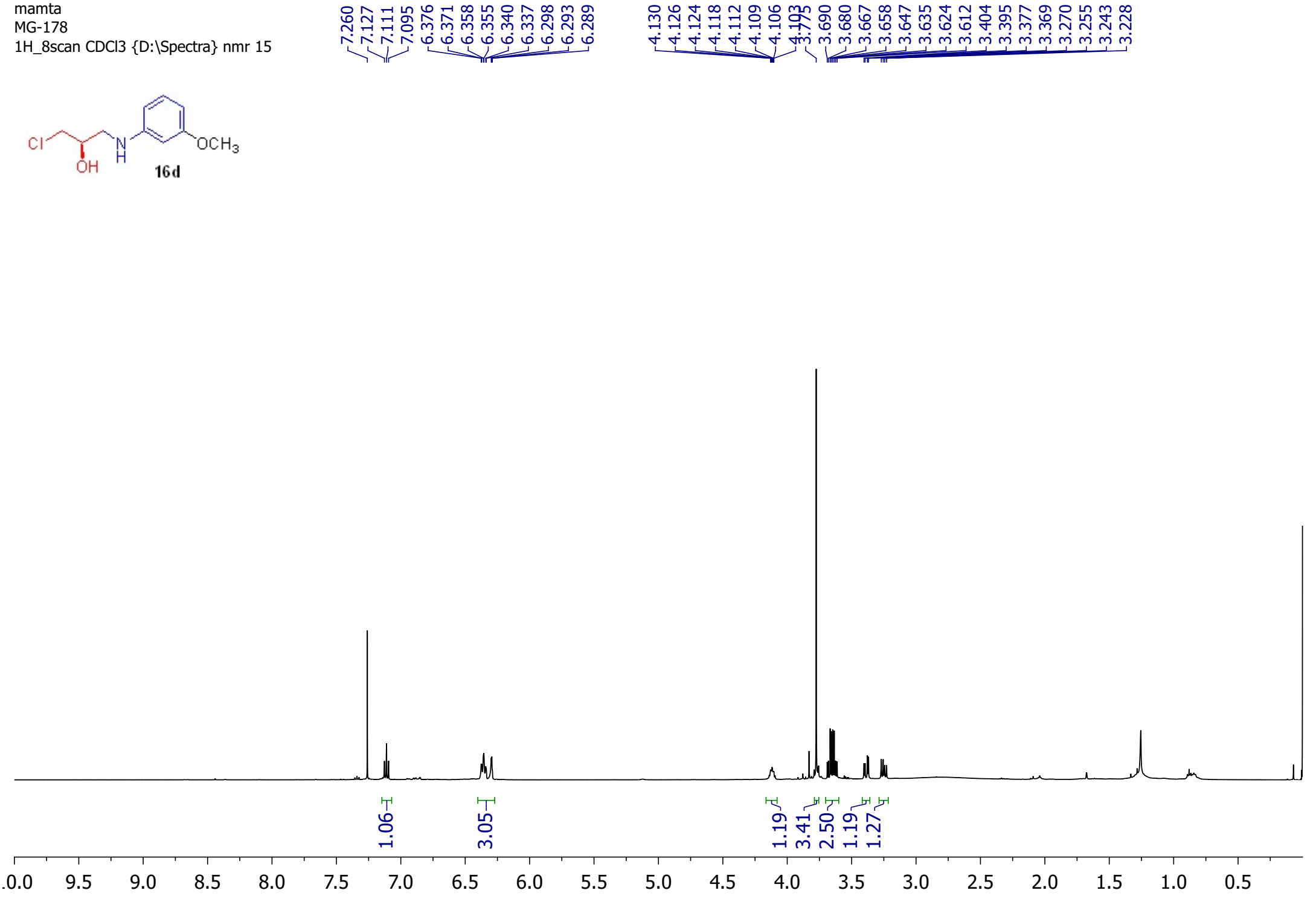
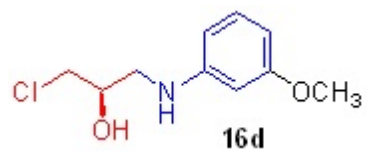
mamta
MG-177
1H_8scan CDCl3 {D:\Spectra} nmr 14



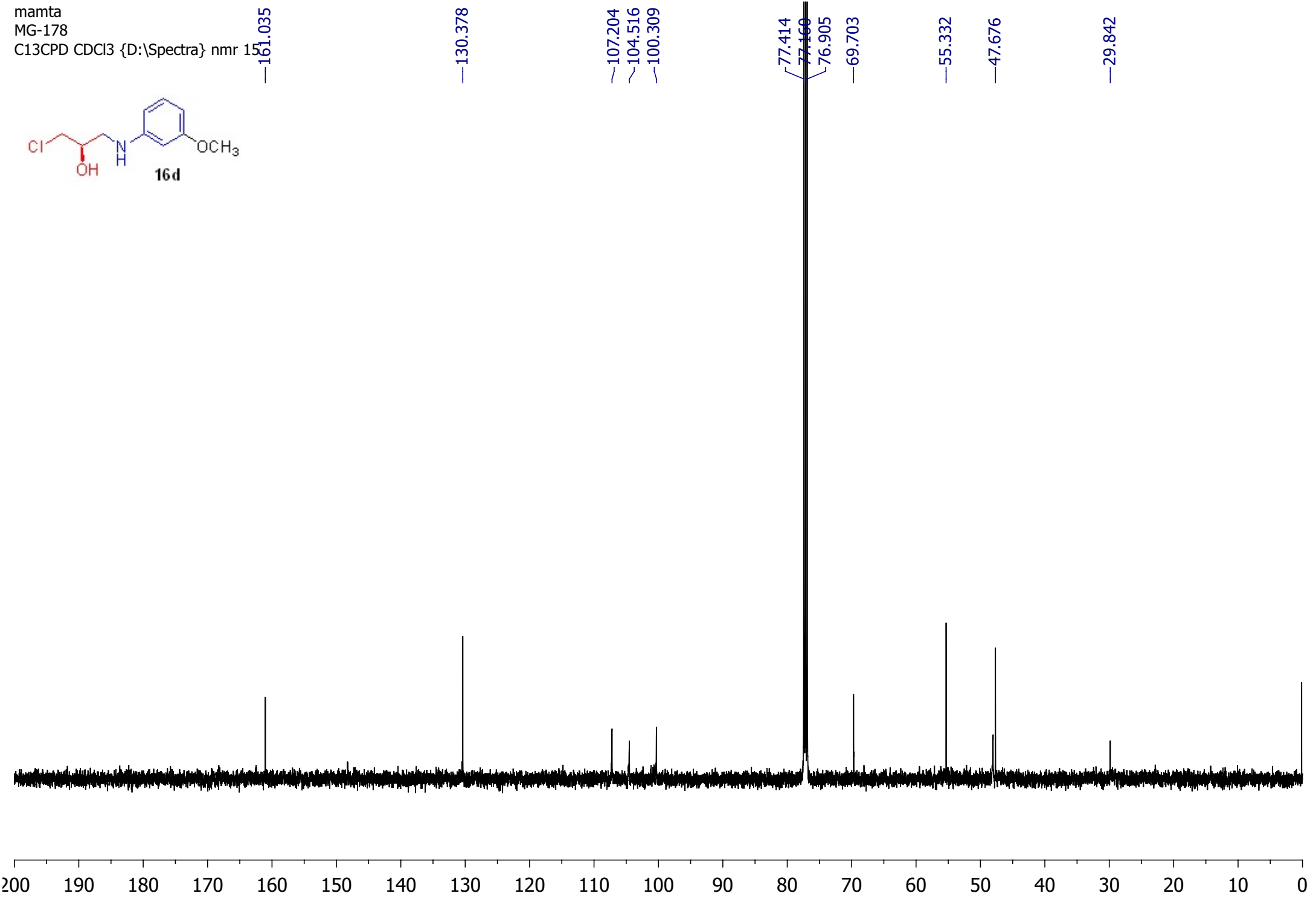
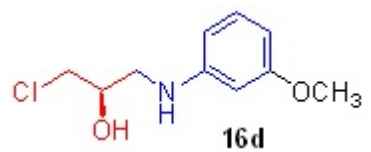
mamta
MG-177
C13CPD CDCl3 {D:\Spectra} nmr 14



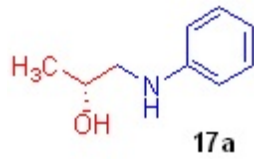
mamta
MG-178
1H_8scan CDCl3 {D:\Spectra} nmr 15



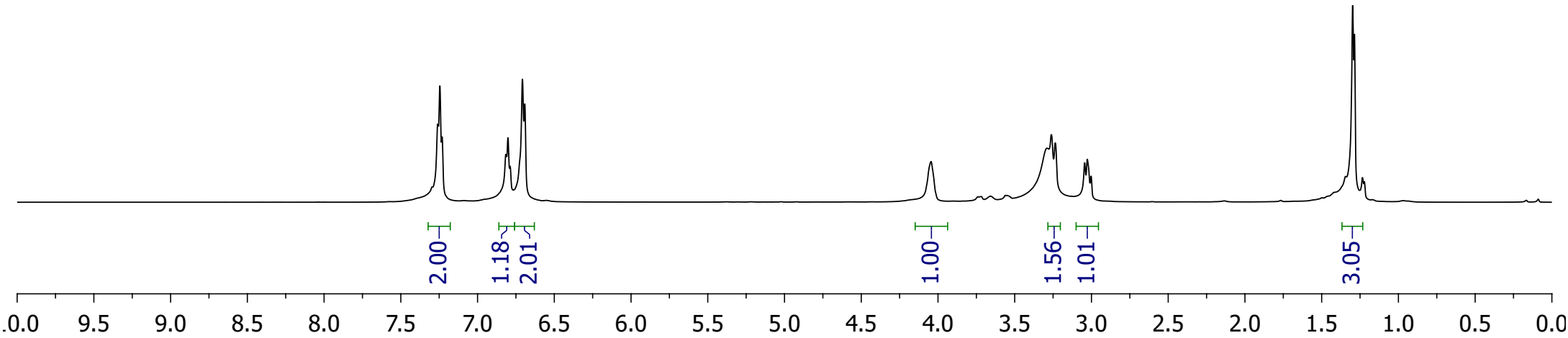
mamta
MG-178
C13CPD CDCl3 {D:\Spectra} nmr 15



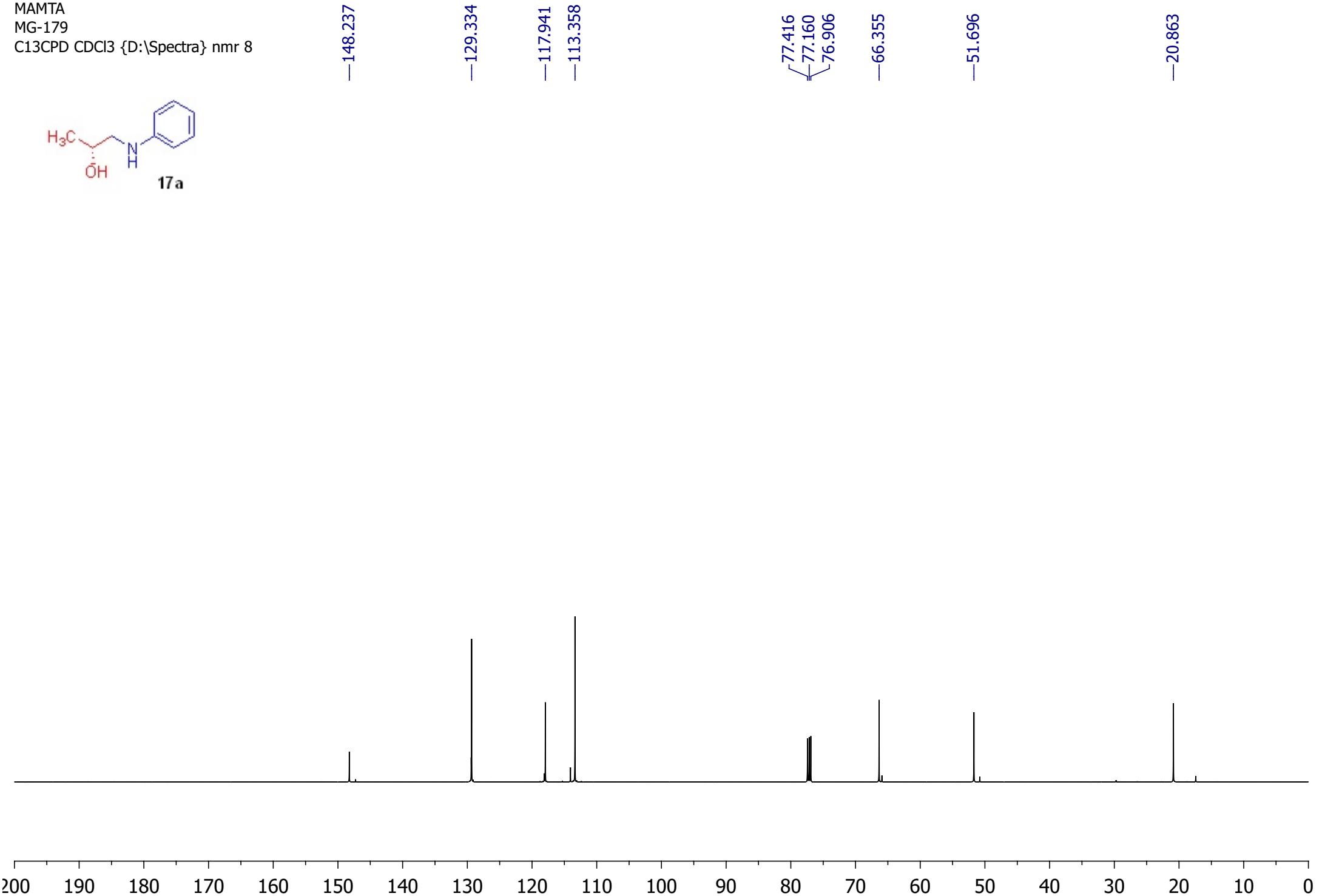
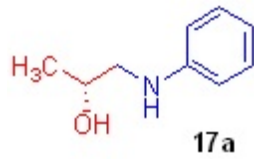
MAMTA
MG-179
1H_8scan CDCl3 {D:\Spectra} nmr 8



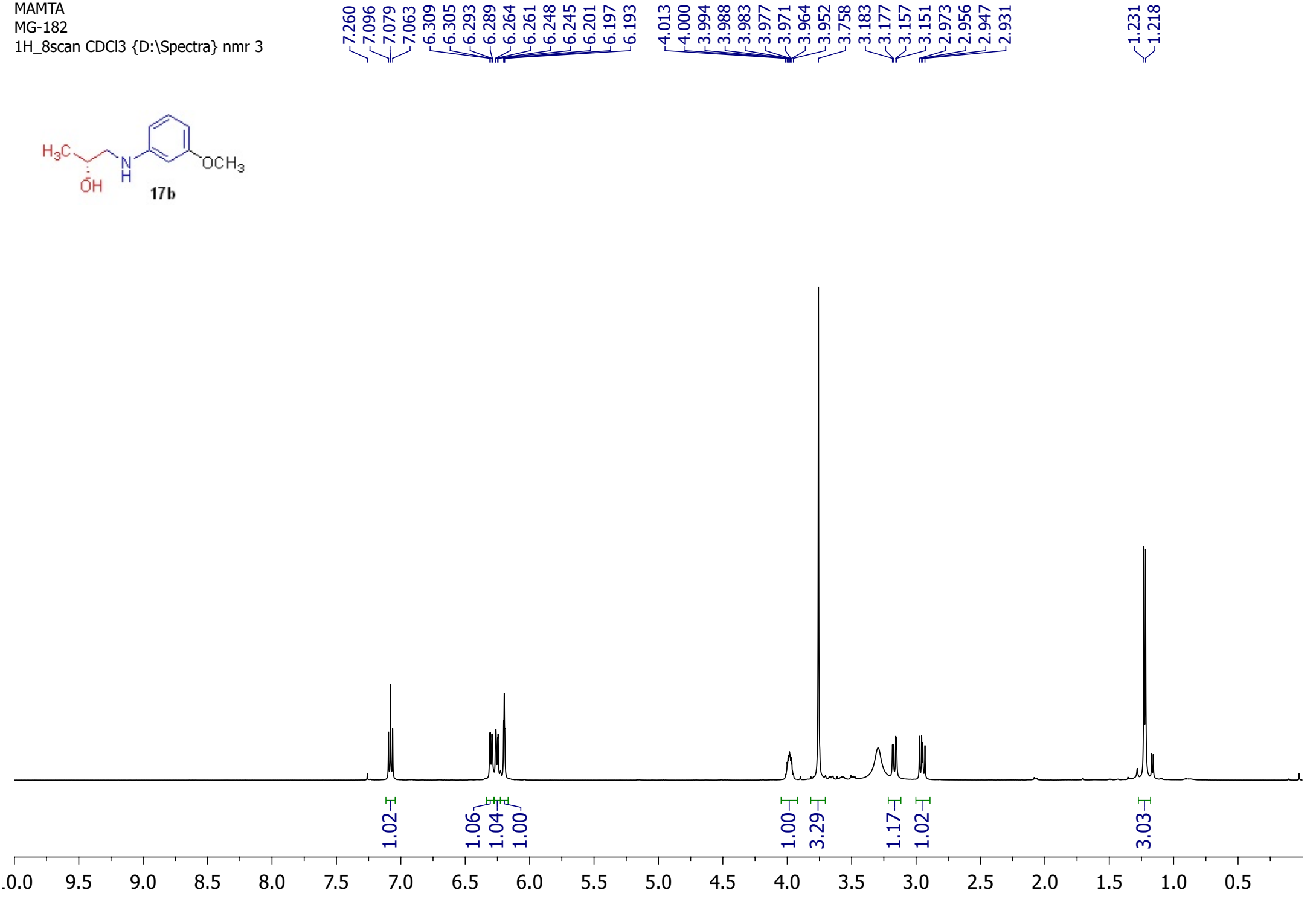
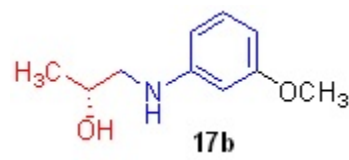
7.260
7.246
7.231
6.815
6.802
6.788
6.708
6.692
— 4.045
3.290
3.262
3.236
3.044
3.027
3.002
1.298
1.286



MAMTA
MG-179
C13CPD CDCl3 {D:\Spectra} nmr 8



MAMTA
MG-182
1H_8scan CDCl3 {D:\Spectra} nmr 3



mamta
MG-182
C13CPD CDCl3 {D:\Spectra} nmr 12

