

Supporting Information

Organoselenium-Based Schiff Bases and Amidic Acid Derivatives as Promising Anticancer Agents Targeting Breast Cancer by Downregulating BCL-2: Design, Synthesis, and Biological Evaluation

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Methods and Materials

Melting points (MP) in °C were recorded on the Gallenkamp instrument. The IR spectra (KBr, cm^{-1}) were recorded at King Faisal University on a Mattson 5000 FTIR Spectrophotometer. The ^1H NMR (400 MHz) and the ^{13}C NMR (100.6 MHz) spectra were recorded at Martin-Luther-University, Halle, Germany, using a Varian VXR spectrometer employing the TMS internal reference and DMSO- d_6 and pyridine- d_6 solvents. The Mass spectra were measured at Martin Luther University, Halle, Germany, on an Agilent 6530 LC Q-TOF instrument.

Chemistry

4-Aminophenylselenocyanate (**2**)¹, bis(4-aminophenyl)diselenide (**3**)², methyl 2-amino-5-selenocyanatobenzoate (**6**)^{3, 4}, and dimethyl 5,5-diselanediyldis(2-aminobenzoate) (**7**)^{3, 4} were synthesized as per the reported protocols.

4-Selenocyanatoaniline (**2**)¹

Selenium dioxide (6 mmol) was added under stirring to a solution of malononitrile (3 mmol) in DMSO (15 mL). The mixture was stirred at room temperature for 15 min in order to obtain triselenium dicyanide. When the exothermic reaction had finished, aniline (5 mmol) was added. The mixture was stirred for 20 min. Water (150 mL) was added to the reaction mixture, and the resulting precipitate (4-selenocyanatobenzenamine) was filtered off, dried, and used without further purifications.

4-Aminophenylselenocyanate (**2**) is a yellow solid, melting point = 73–74 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.44 (d, J = 8.4 Hz, 2H, Ar-H), 6.64 (d, J = 8.4 Hz, 2H, Ar-H), 3.95 (s, 2H, NH_2).

Synthesis of 4,4'-diselanediyldianiline (**3**)¹

Under argon, NaBH_4 (3 mmol) was added in small portions with caution to a solution of 4-selenocyanatobenzenamine (1 mmol) in absolute ethanol (40 mL). The mixture was stirred at room temperature for 2 h. The solvent was removed under reduced pressure, and the remaining residue was dissolved in dichloromethane, washed with water (350 mL). The organic layer was separated, dried with anhydrous Na_2SO_4 , and removed under vacuum. The residue was purified by chromatography on silica gel (petroleum ether: ethyl acetate 4:2).

4,4'-Diselanediyldianiline (**3**) was obtained as pale-yellow crystals (82 % yield. Mp: 78–80 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.3 (m, 4H, Ar-H), 6.5 (m, 4H, Ar-H), 3.7 ppm (s, 4H, NH_2).

Synthesis of methyl 2-amino-5-selenocyanatobenzoate (**6**)^{3, 4}

Methyl 2-amino-5-selenocyanatobenzoate (**6**) was synthesized from the reaction of methyl 2-aminobenzoate with TSD prepared *in situ* from $\text{CH}_2(\text{CN})_2$ and SeO_2 in 96% yield. Briefly, SeO_2 (30 mmol,

3300 mg) was added to $\text{CH}_2(\text{CN})_2$ (15 mmol, 1000 mg) in 10 mL DMSO- D_6 , and the mixture was stirred for 20 min at RT. Next, the mixture was filtered off to get rid of any formed black selenium, and methyl 2-aminobenzoate (12.5 mmol, 1800 mg) was then added, and the reaction mixture was stirred for a further 2 hrs. Finally, adding 10 gm of ice terminated the reaction, and the formed precipitate was filtered, washed several times with H_2O and Na_2CO_3 solution, dried, and recrystallized from petroleum ether.

2-Amino-5-selenocyanatobenzoate was isolated as reddish solid; yield: 3072 mg (96%); MP = 118-119 °C; Rf: 0.4 (petroleum ether/ ethyl acetate 4:2). IR(KBr): $\lambda_{\text{max}} \cdot \text{cm}^{-1}$: 3475, 3366, 2946, 2147, 1691. ^1H NMR (400 MHz, DMSO- D_6 - d_6) δ 8.02 (s, 1H, Ar-H), 7.57 (d, J = 8.8 Hz, 1H, Ar-H), 7.08 (d, 1H, Ar-H), 6.84 (s, 2H, NH_2), 3.82 (s, 3H, OCH_3). ^{13}C NMR (101 MHz, DMSO- D_6 - d_6) δ 166, 152, 140, 137, 118, 109, 105, 105, 51. MS (EI, 70 ev) m/z (%) = 259.35 (M+3H, 2.39), 117 (29.02), 87 (26.6), 75 (2.70), 59 (100.0, base peak).

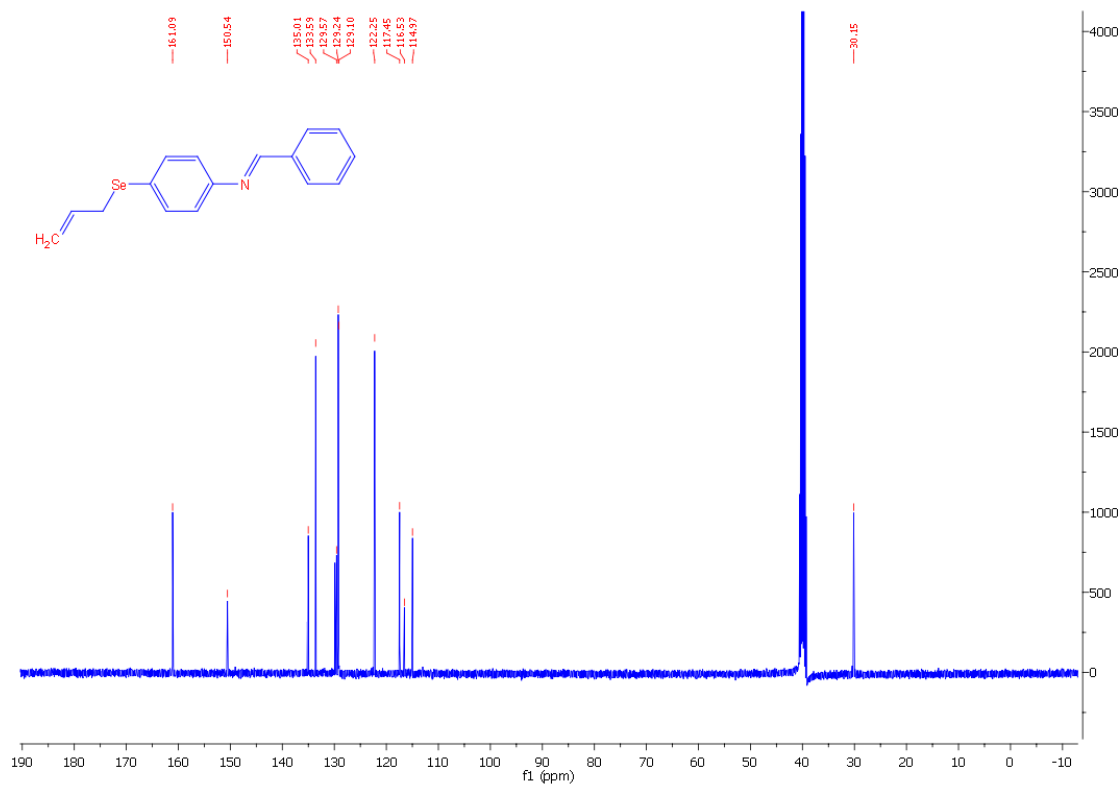
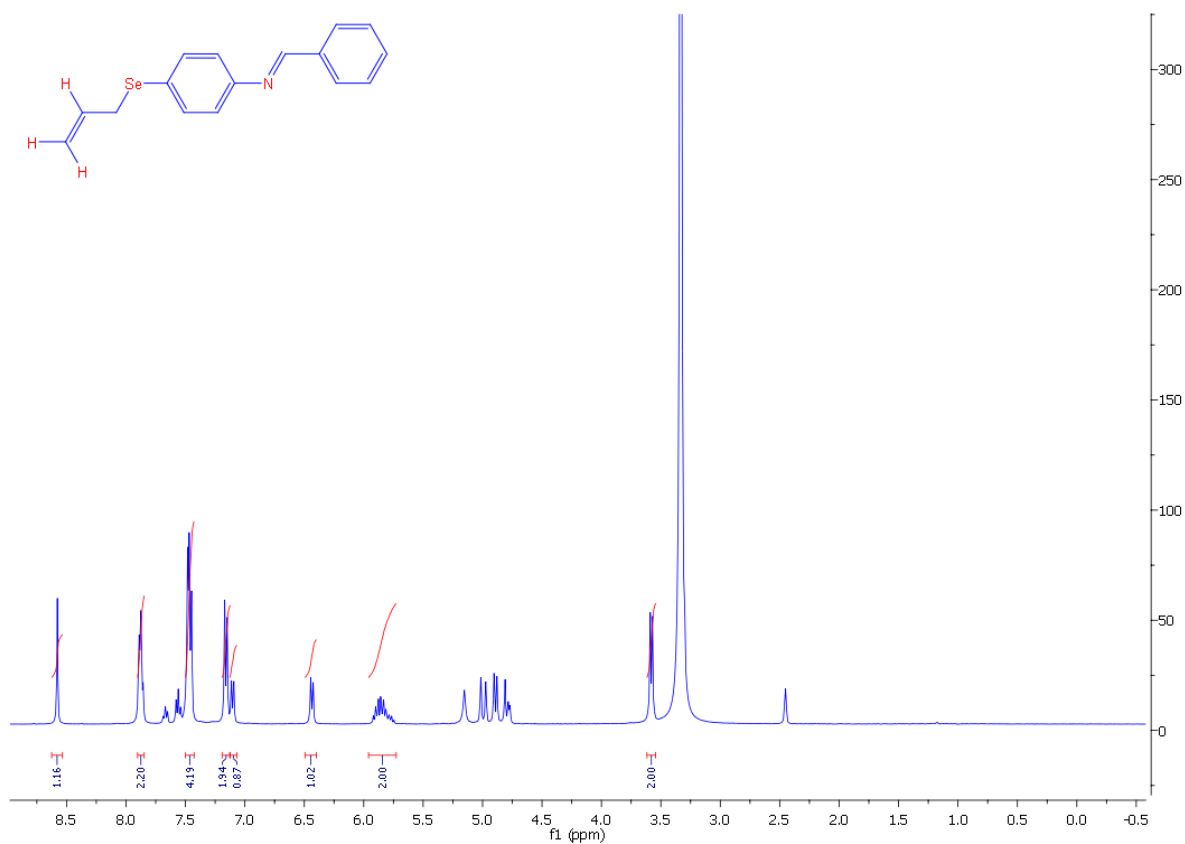
Synthesis of dimethyl 5,5-diselanediyldis(2-aminobenzoate) (7) ^{3, 4}

Dimethyl 5,5-diselanediyldis(2-aminobenzoate) (7) was synthesized from the reaction of **6** and sodium hydroxide in 92% yield. Briefly, compound **6** (4 mmol, 1000 mg) was dissolved in EtOH (20 mL), and then sodium hydroxide (4 mmol, 160 mg) was added. The reaction mixture was stirred for 2 hours at room temperature, and the resulting precipitate was filtered, washed several times with H_2O , and recrystallized from chloroform.

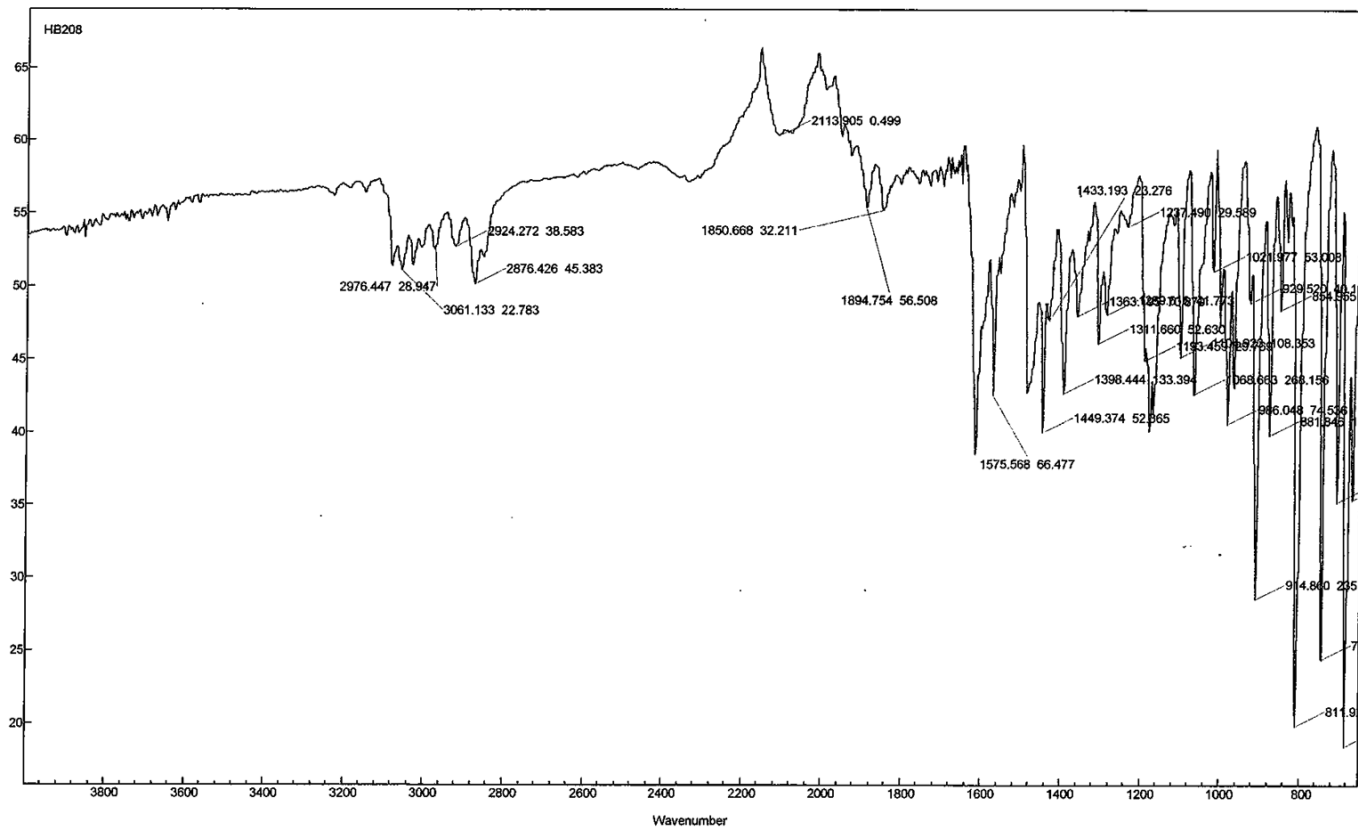
Dimethyl 5,5-diselanediyldis(2-aminobenzoate) was isolated as a yellow solid; yield: 1692.43 mg (92%); MP = 138- 139 °C; Rf = 0.5 (petroleum ether/ ethyl acetate 4:3). IR(KBr): $\lambda_{\text{max}} \cdot \text{cm}^{-1}$: 3455, 3344, 2931, 1684. ^1H NMR (400 MHz, DMSO- D_6) δ 7.70 (s, 2H, Ar-H), 7.44 (d, J = 8.6 Hz, 2H, Ar-H), 7.00 (s, 4H, 2 NH_2), 6.77 (d, J = 8.7 Hz, 2H, Ar-H), 3.74 (s, 6H, 2 OCH_3). ^{13}C NMR (101 MHz, DMSO- D_6 - d_6) δ 167, 151, 140, 138, 117, 113, 108, 51. MS (EI, 70 ev) m/z (%) = 460.15 (M+H, 20.76), 459.15 (M, 5.20) or 230 (24.42), 119 (9.45), 91 (100.0, base peak), 65 (8.88).

Supporting Information

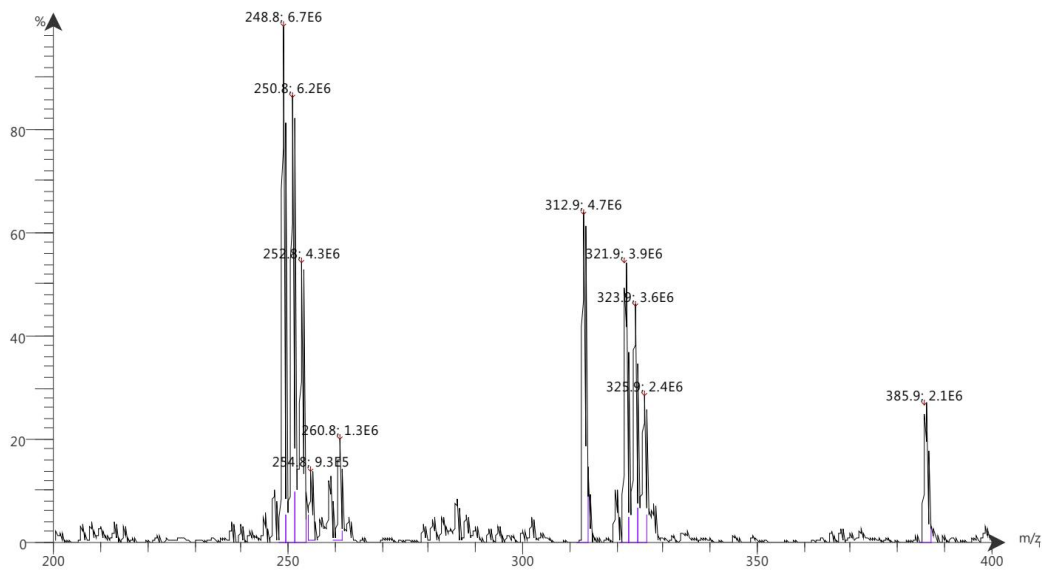
N-(4-(Allylselanyl)phenyl)-1-phenylmethanimine (HB208)



Supporting Information

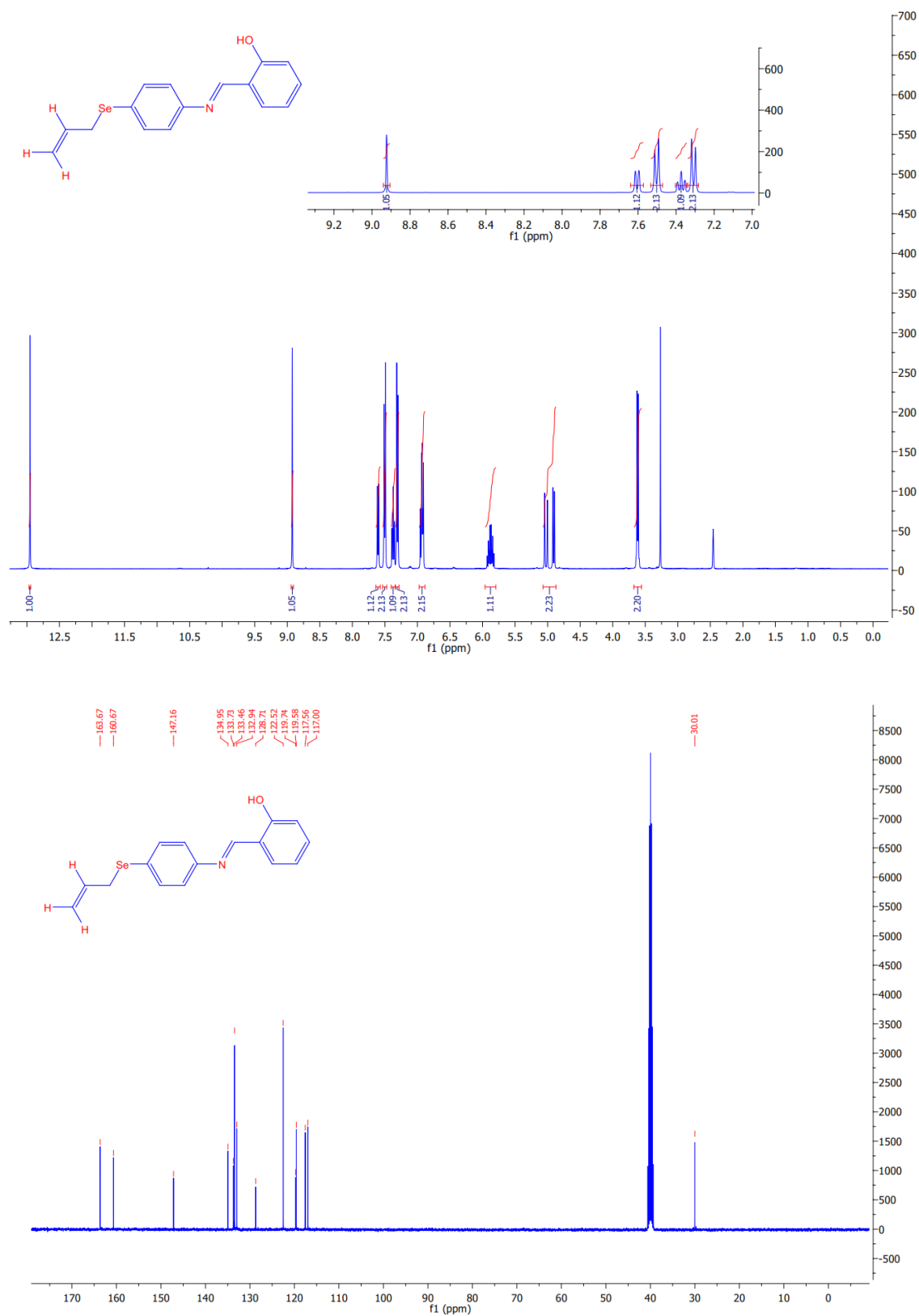


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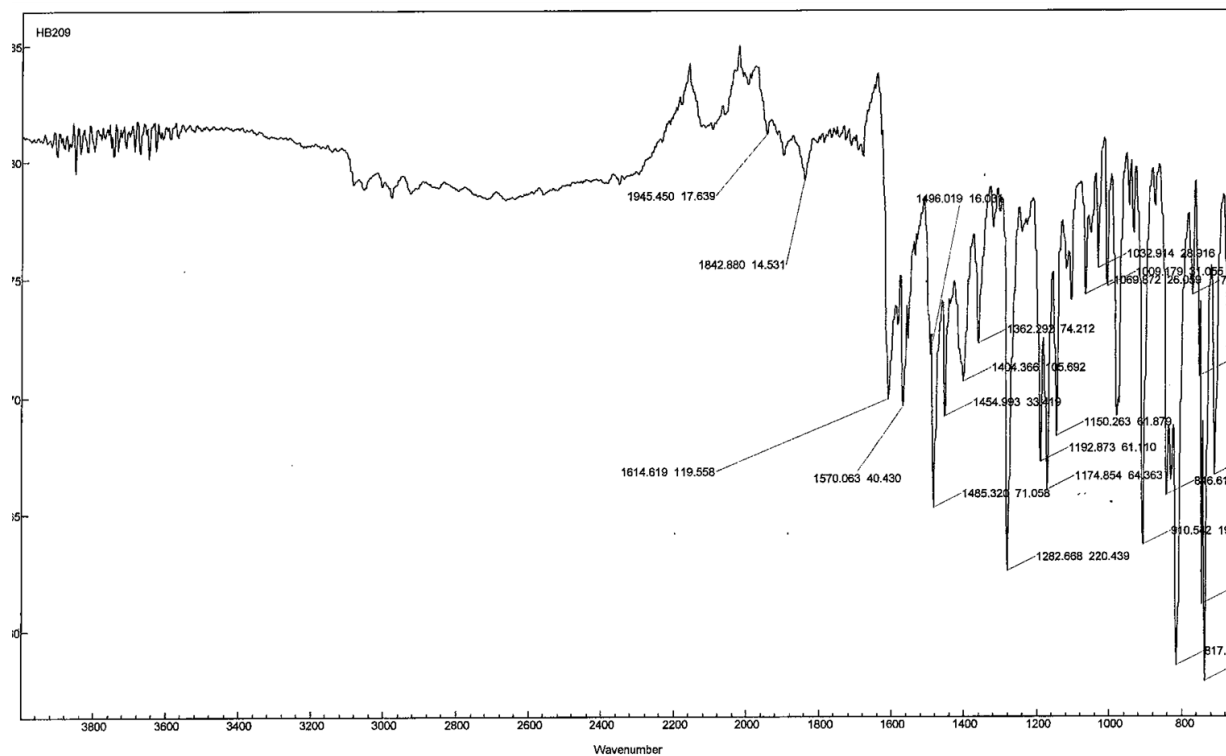


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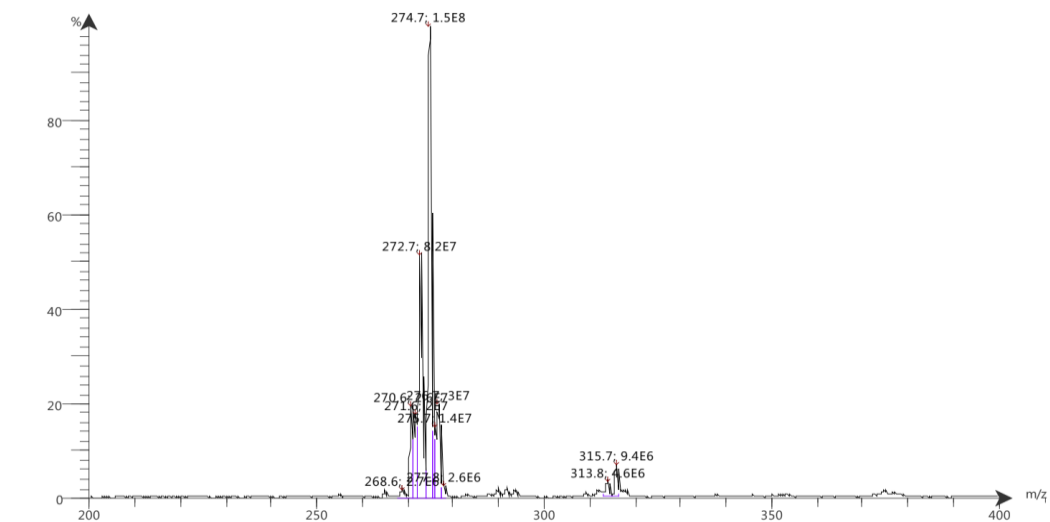
2-(((4-(Allylselanyl)phenyl)imino)methyl)phenol (HB209)



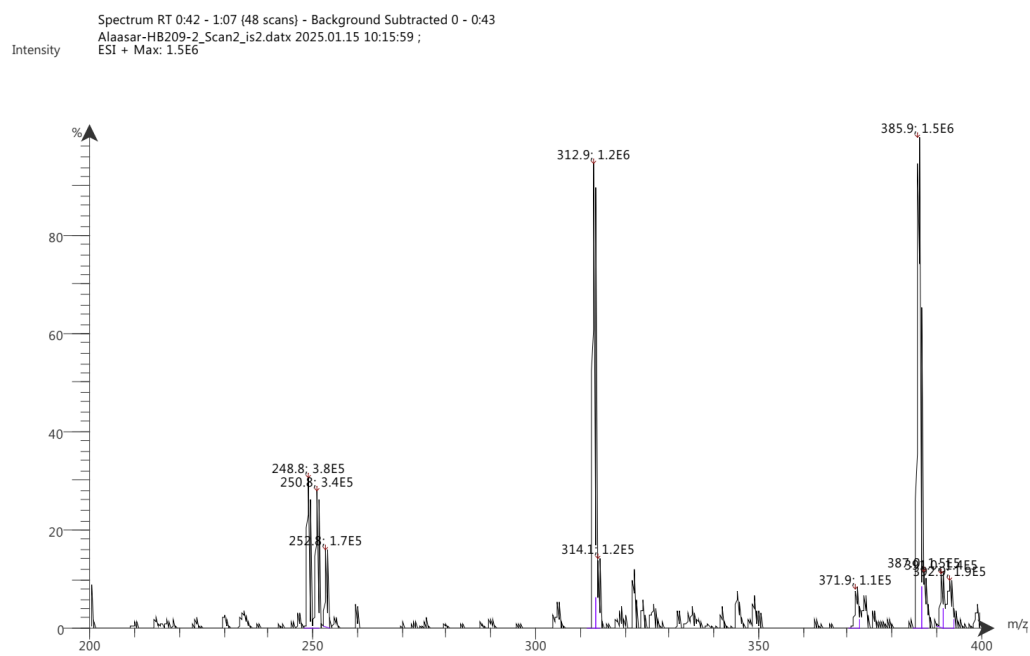
Supporting Information



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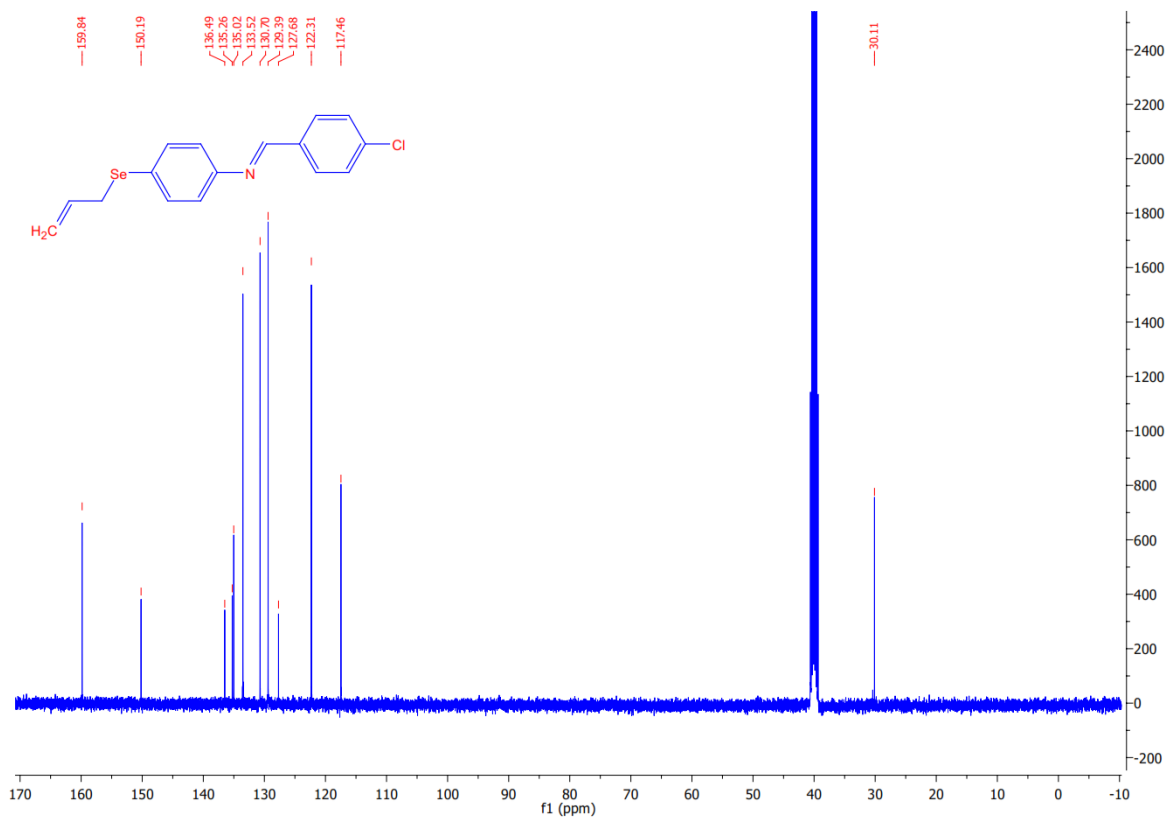
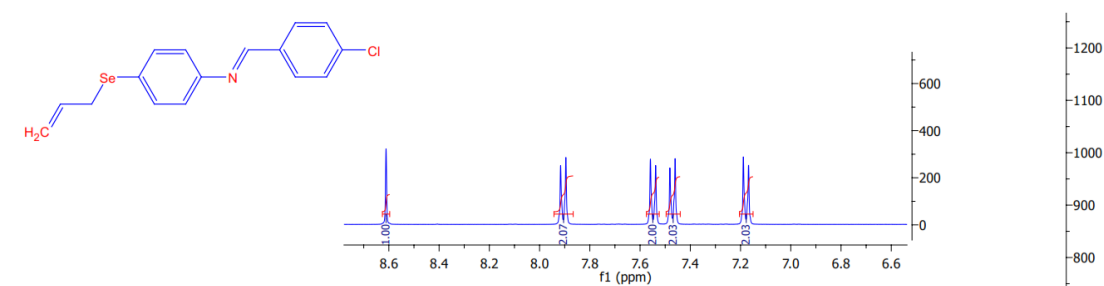


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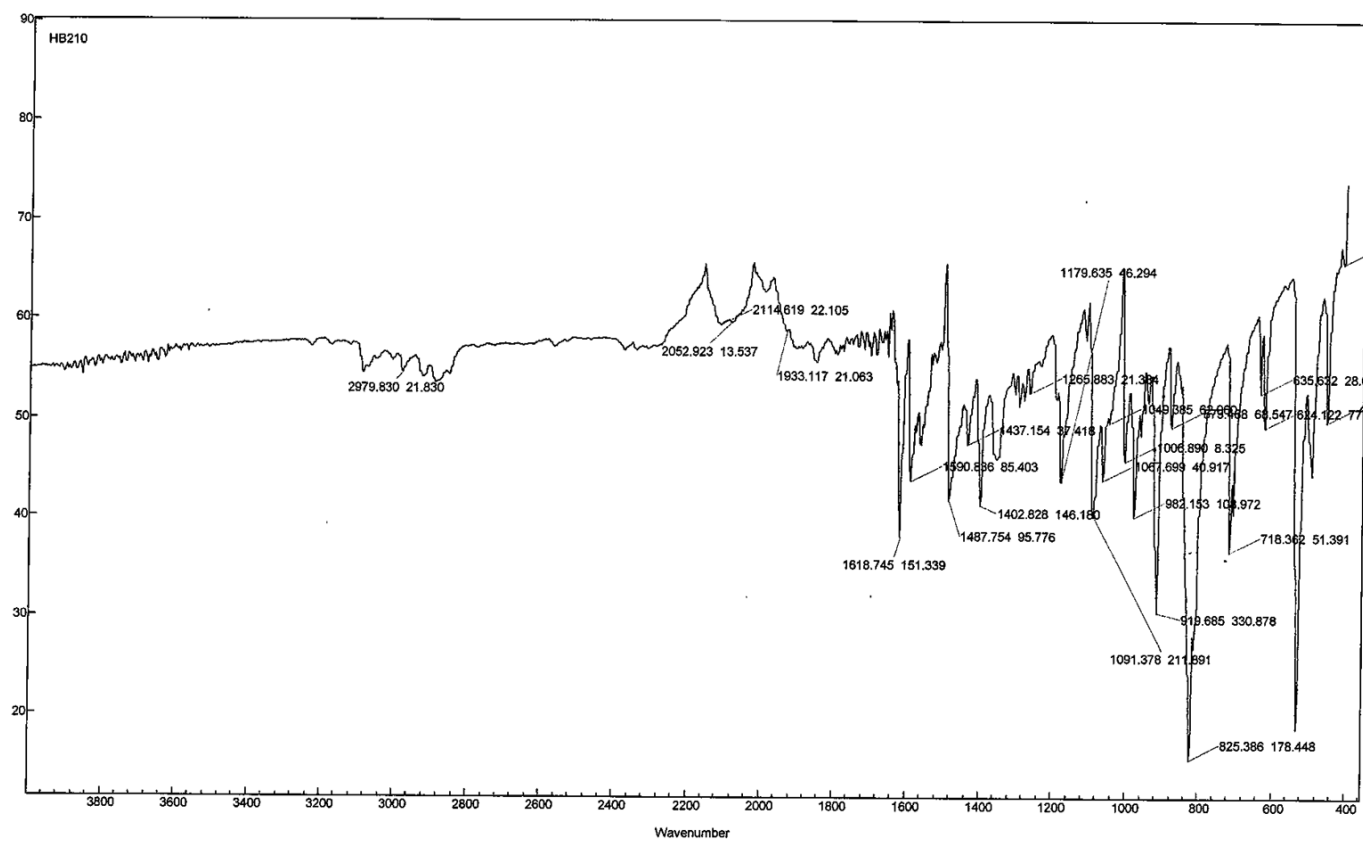


Supporting Information

N-(4-(Allylselanyl)phenyl)-1-(4-chlorophenyl)methanimine (HB210)

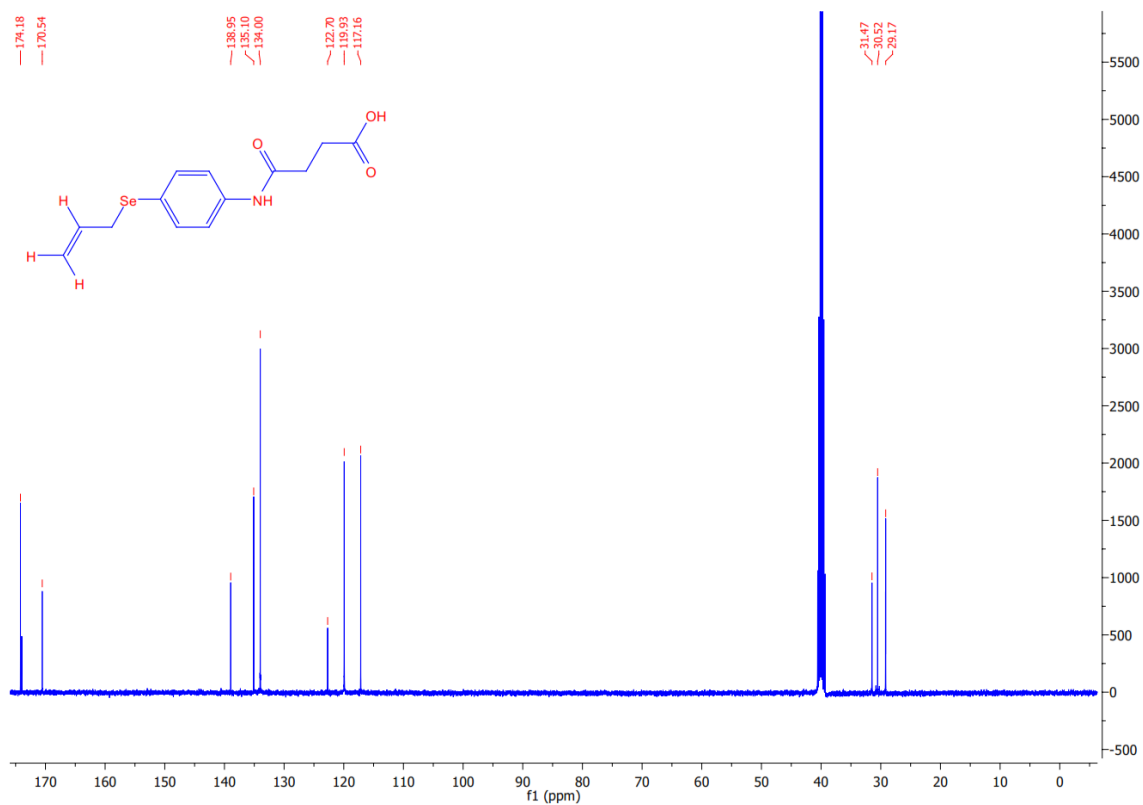
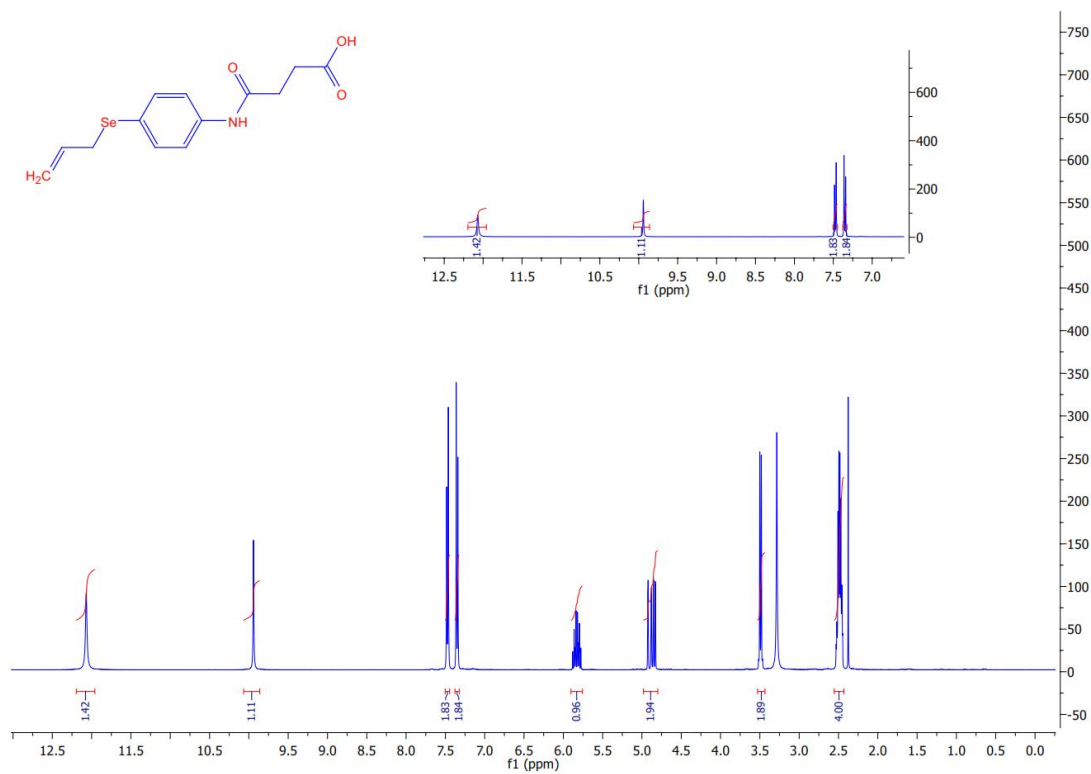


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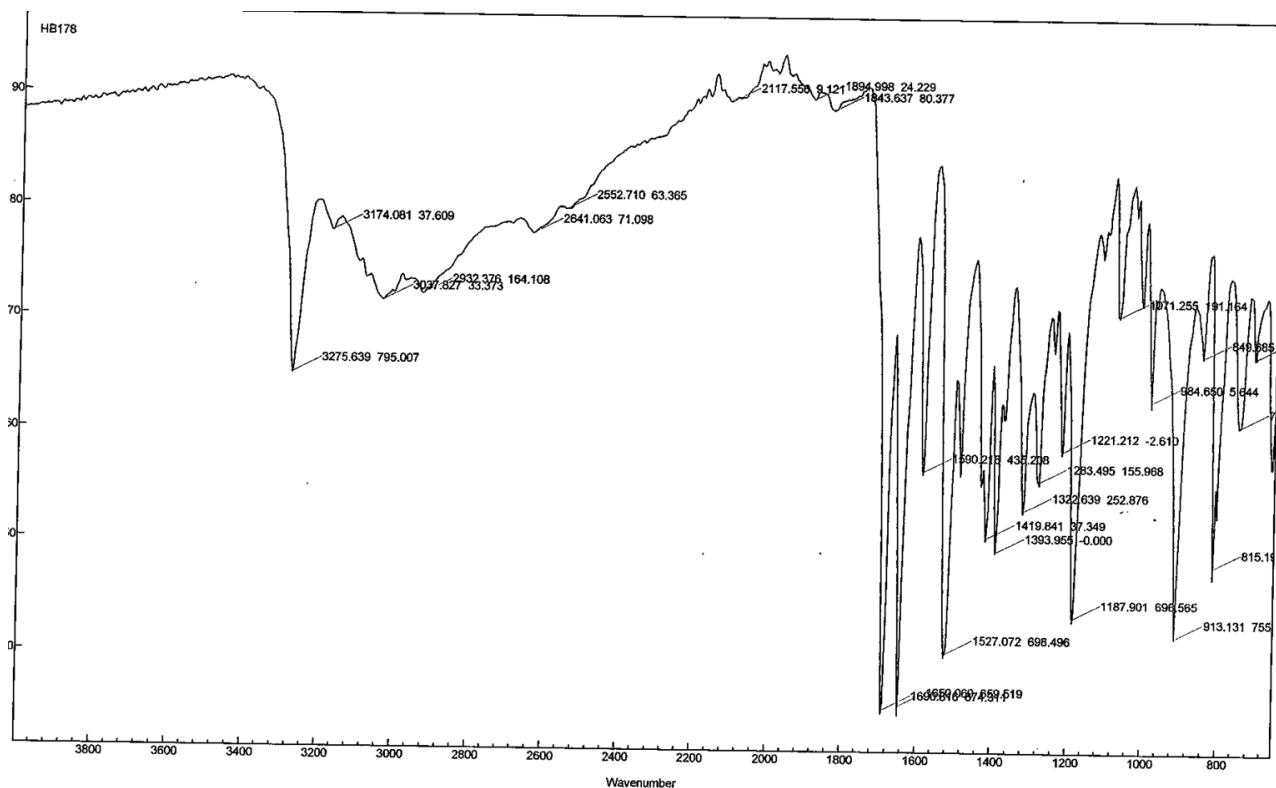


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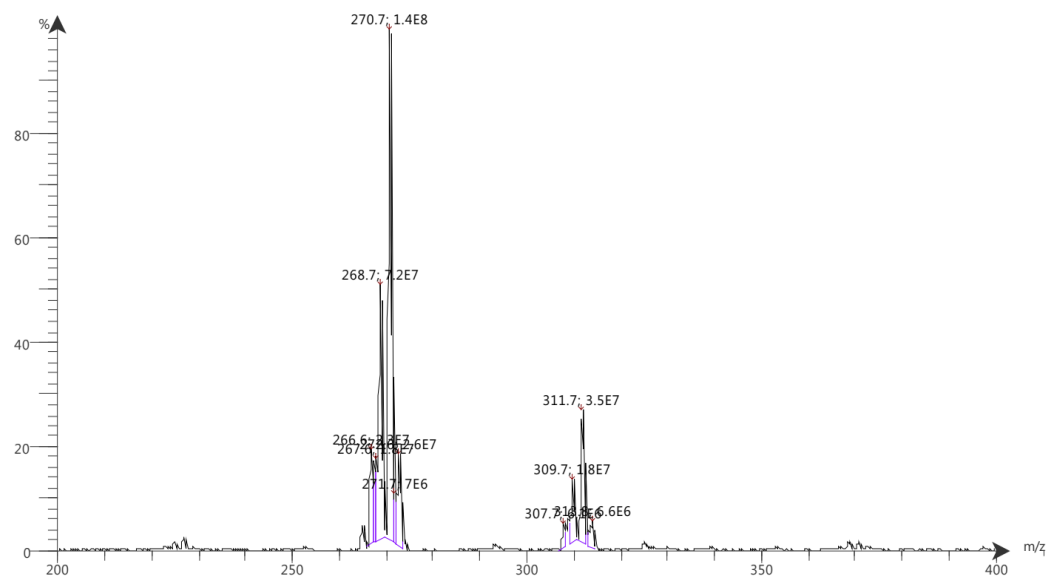
4-((4-(Allylselanyl)phenyl)amino)-4-oxobutanoic acid (HB178)



Supporting Information

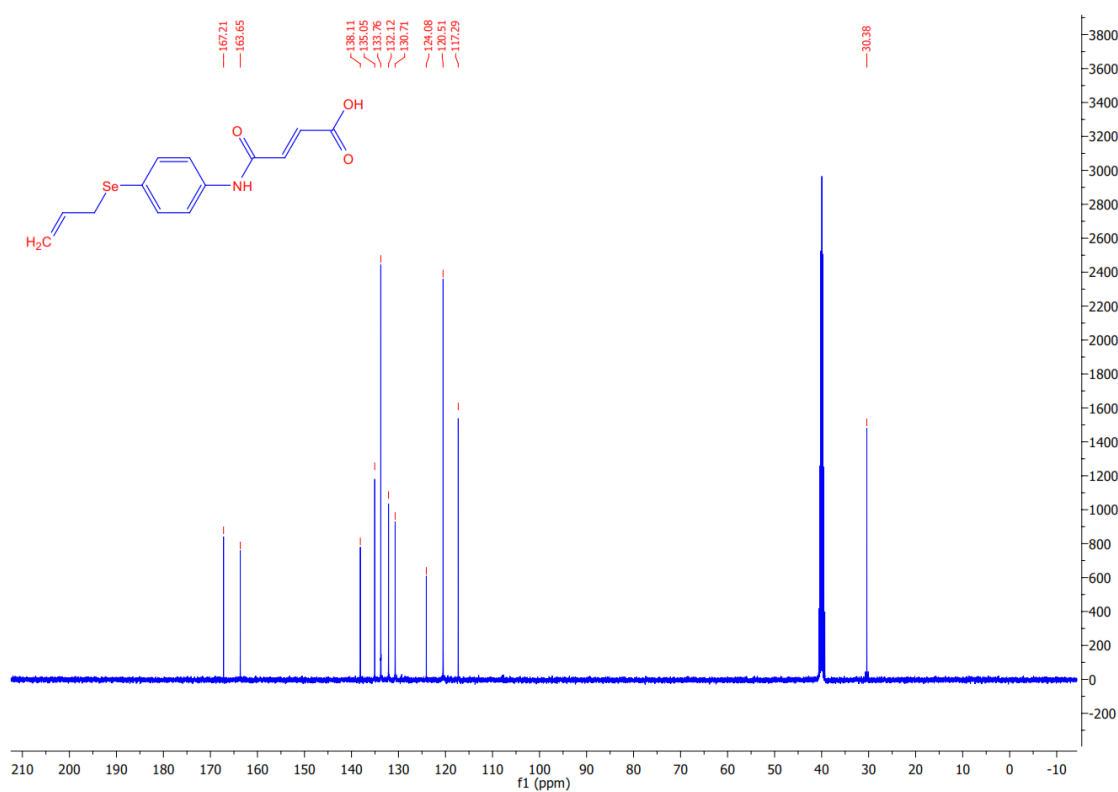
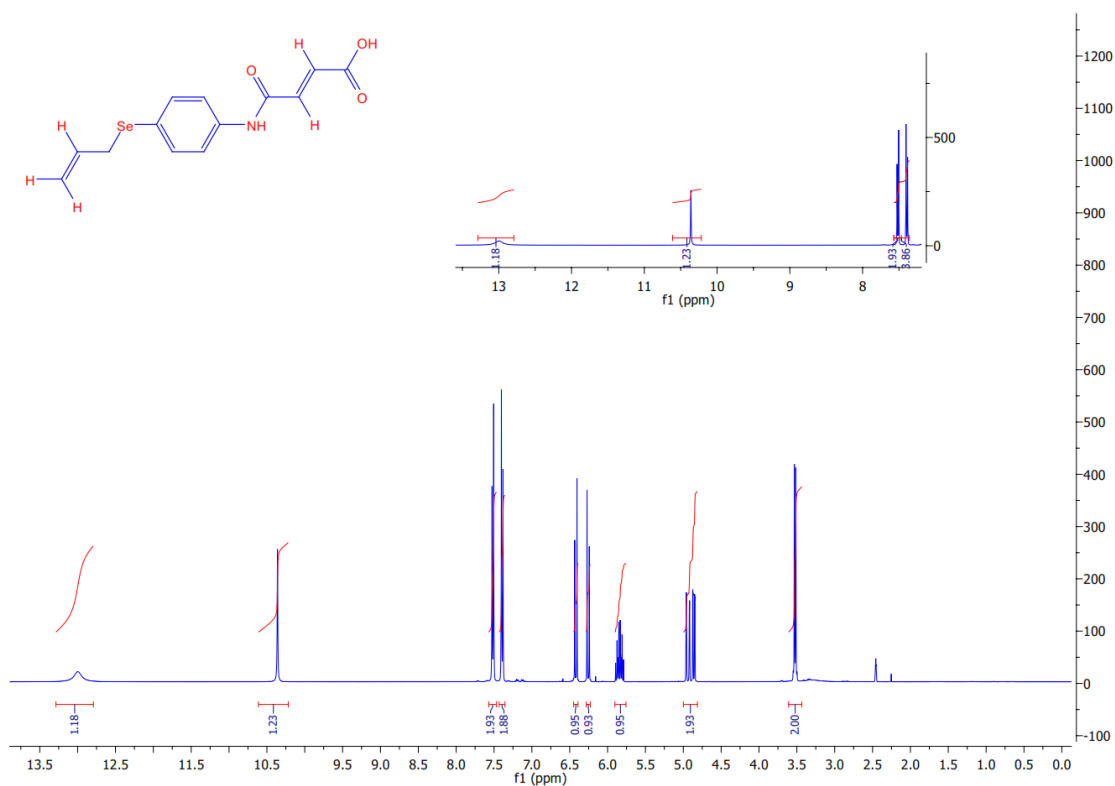


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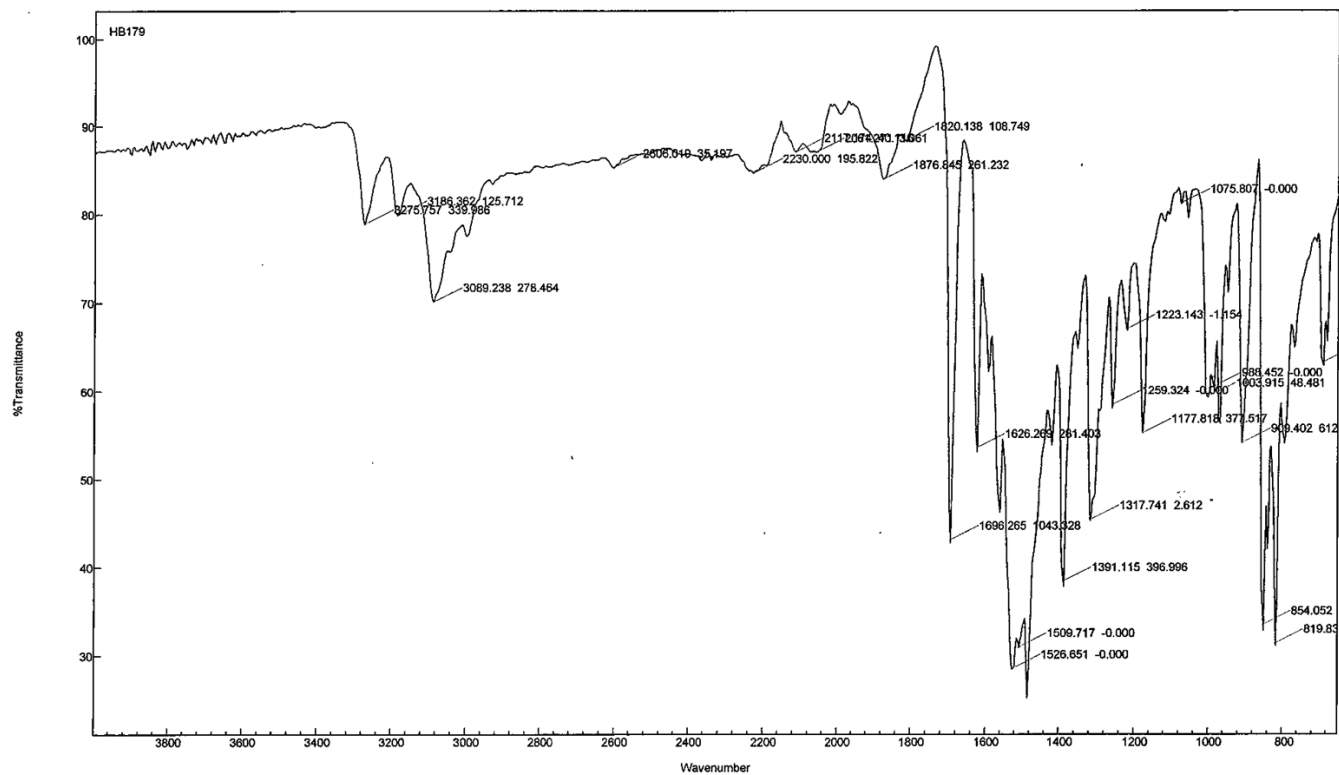


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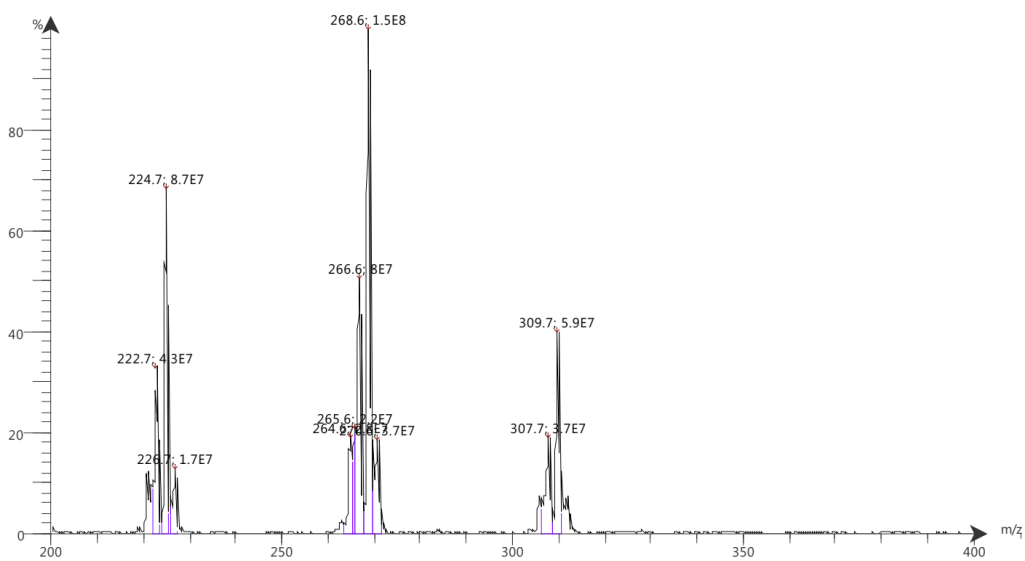
4-((4-(Allylselanyl)phenyl)amino)-4-oxobut-2-enoic acid (HB179)



Supporting Information

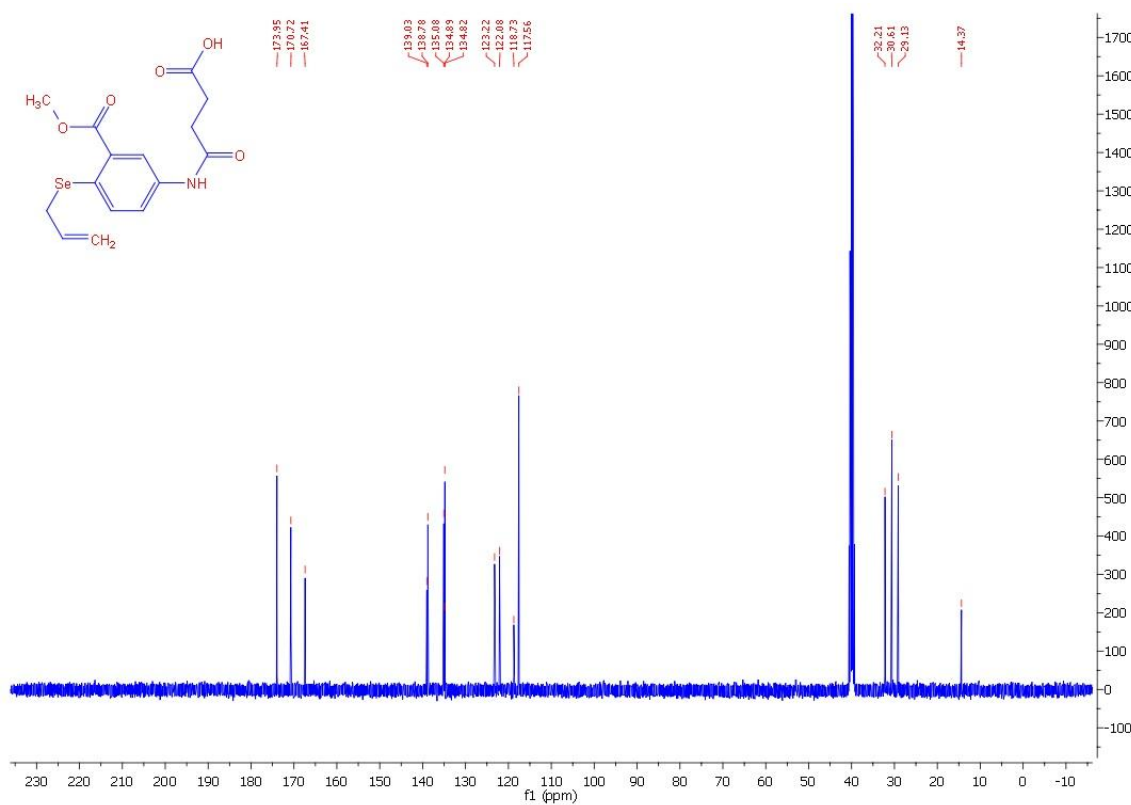
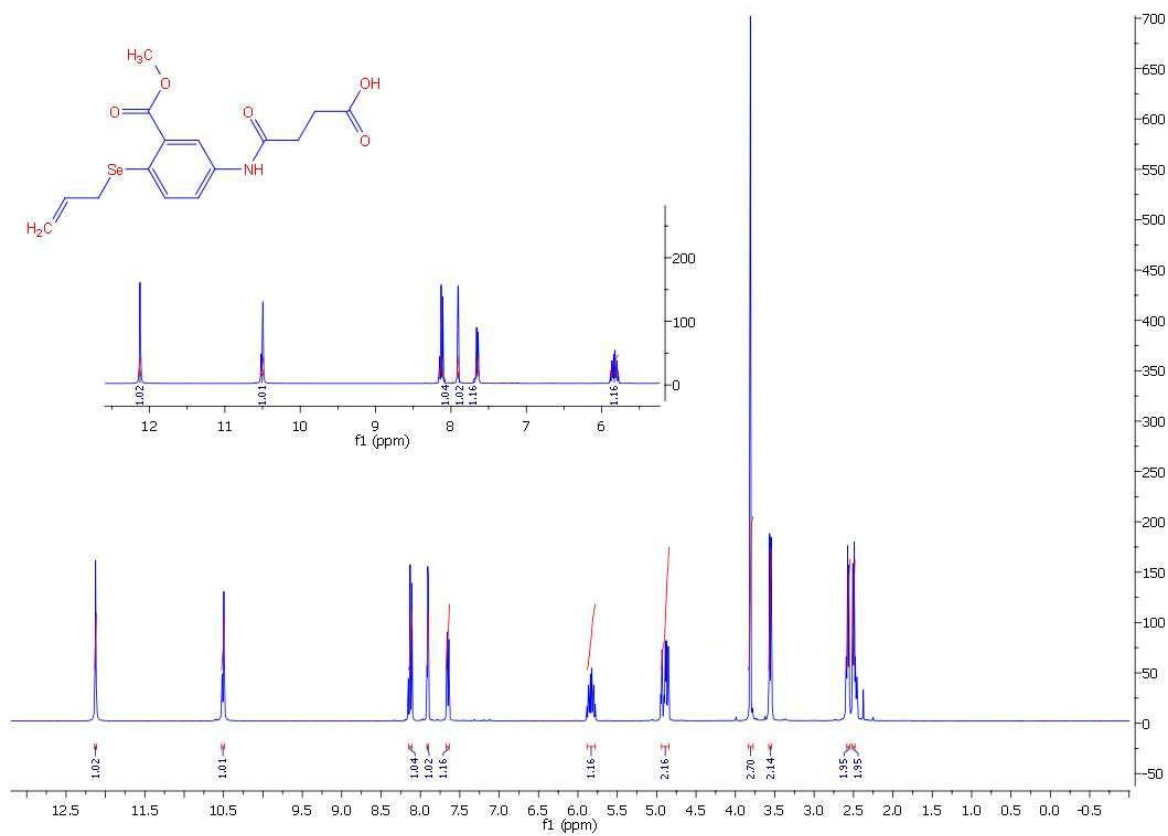


Spectrum RT 0:33 - 1:08 (68 scans) - Background Subtracted 0:03 - 0:34
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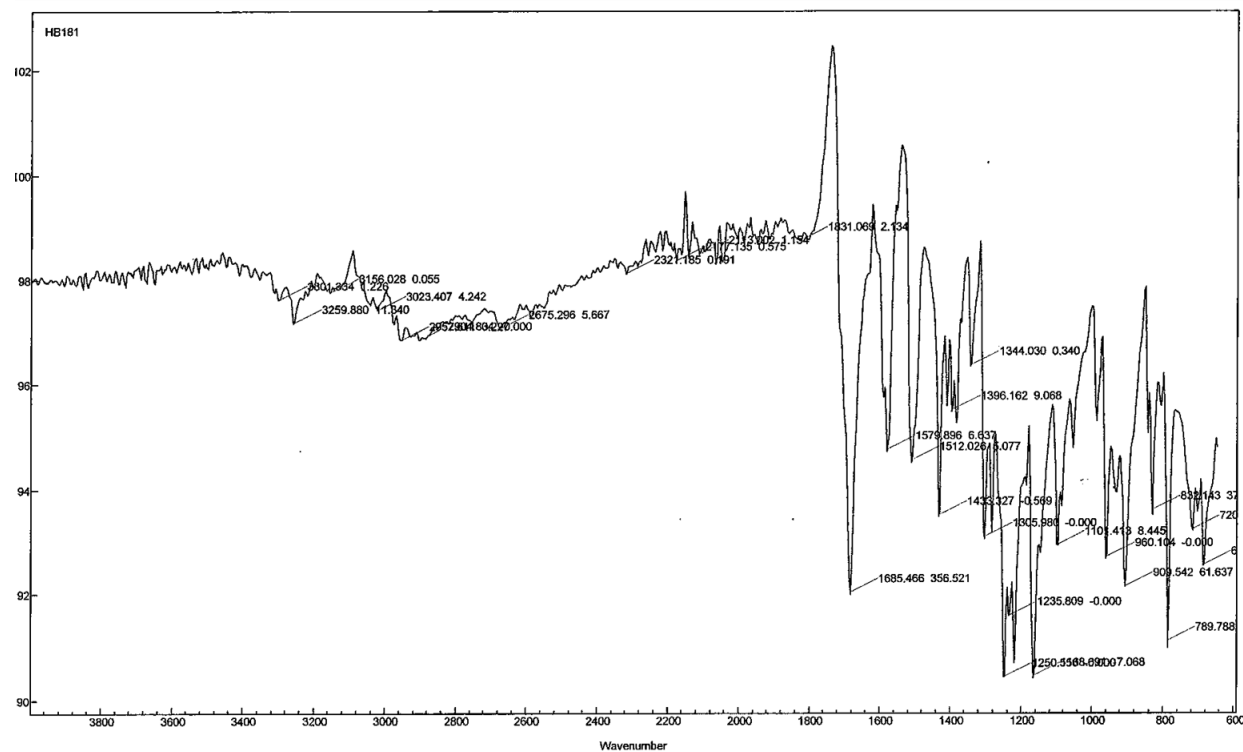


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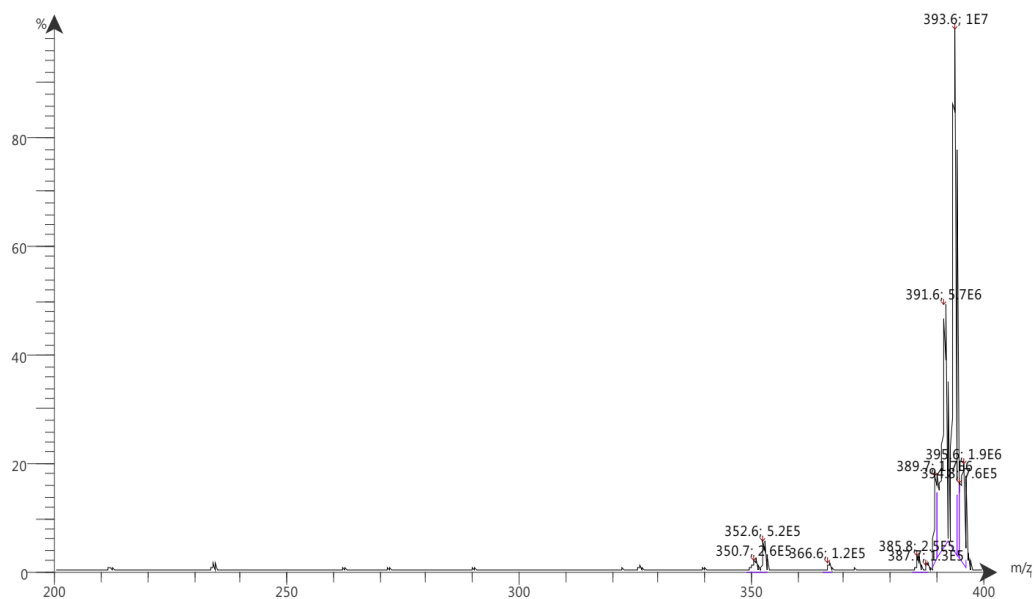
4-((4-(Allylselanyl)-3-(methoxycarbonyl)phenyl)amino)-4-oxobutanoic acid (HB181)



Supporting Information

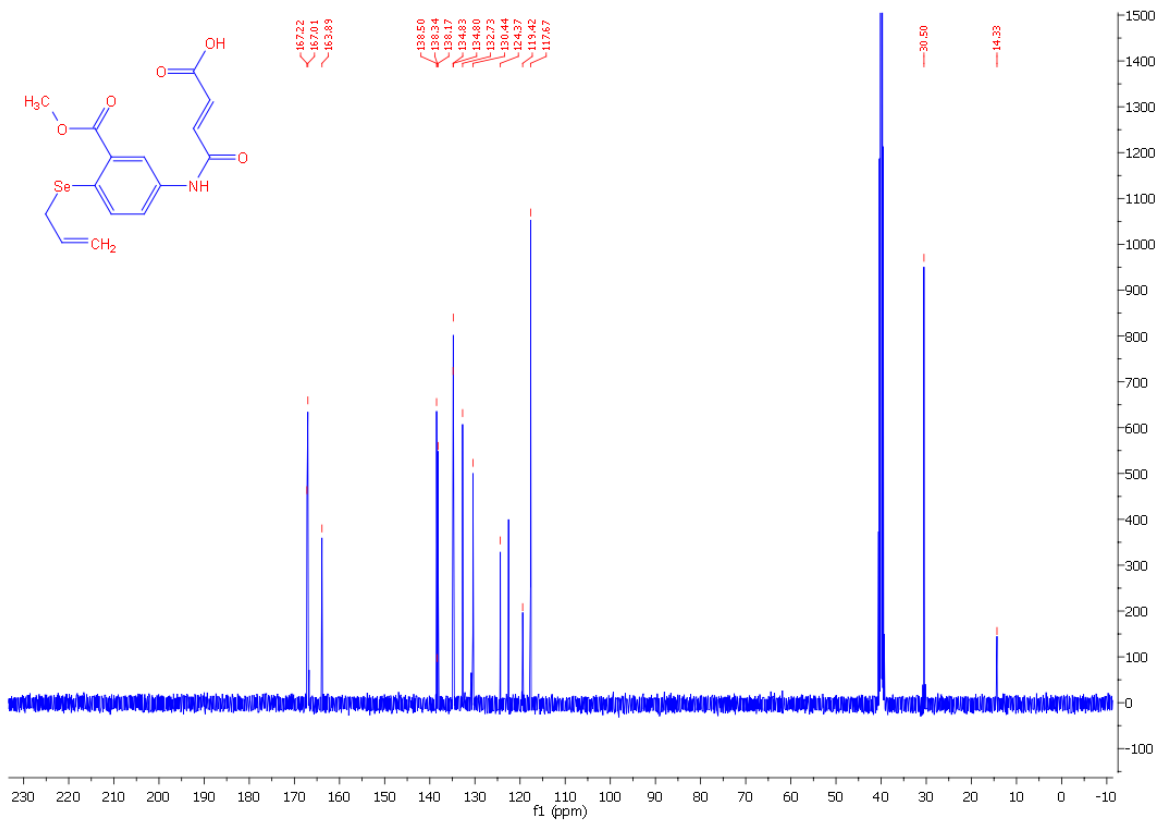
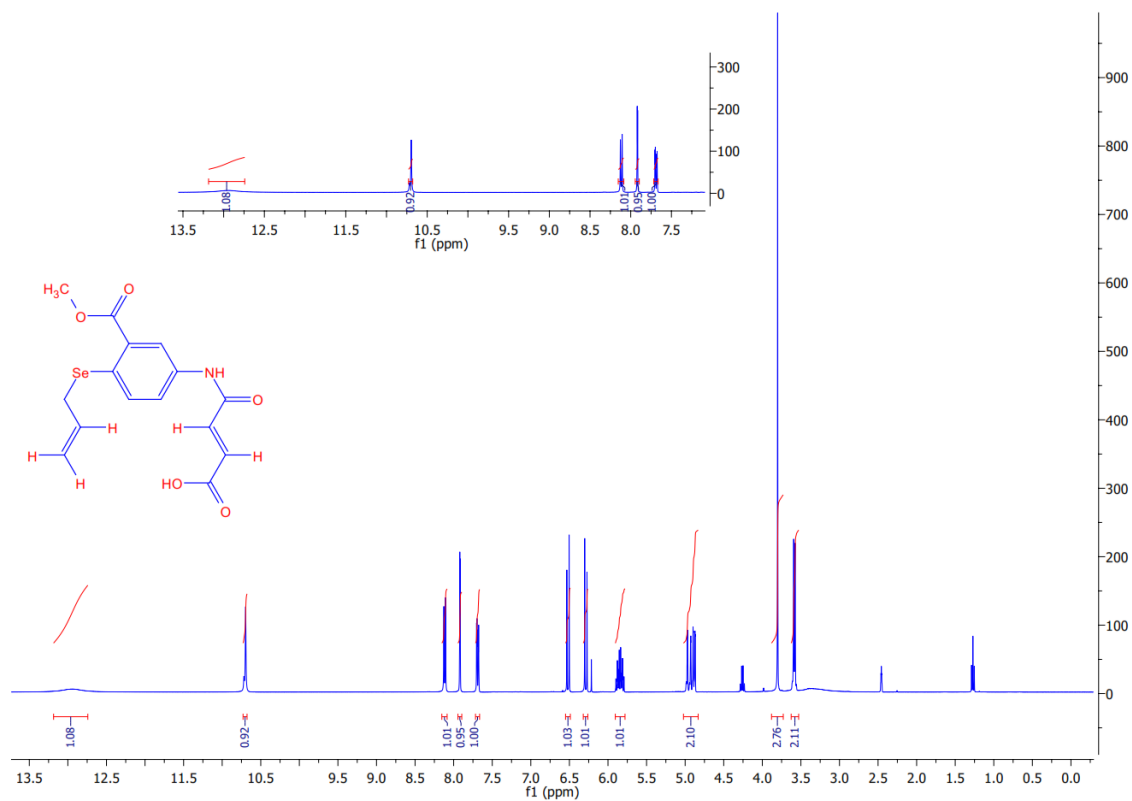


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 ESI + Max: 1.1E7

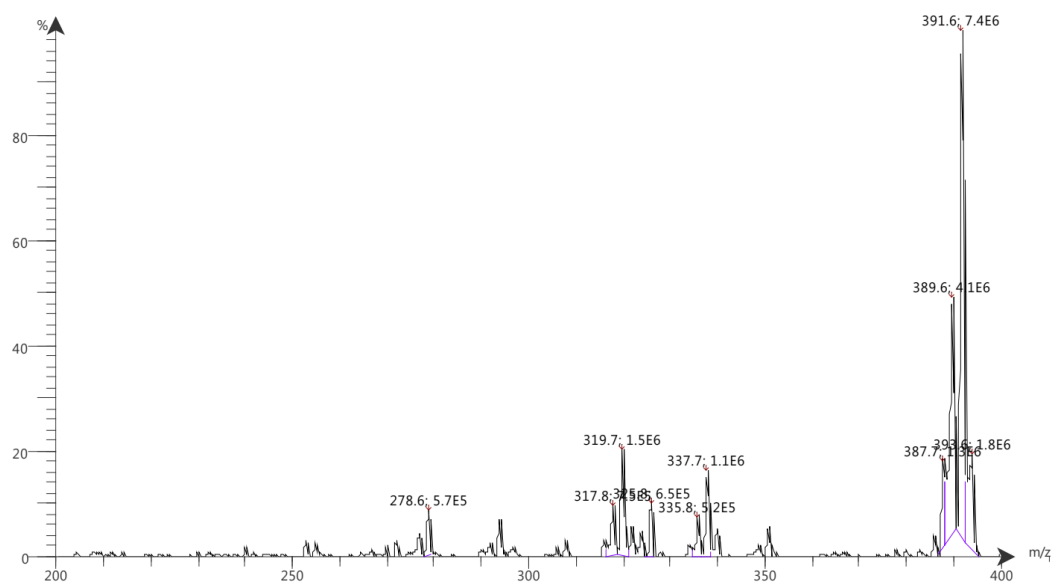
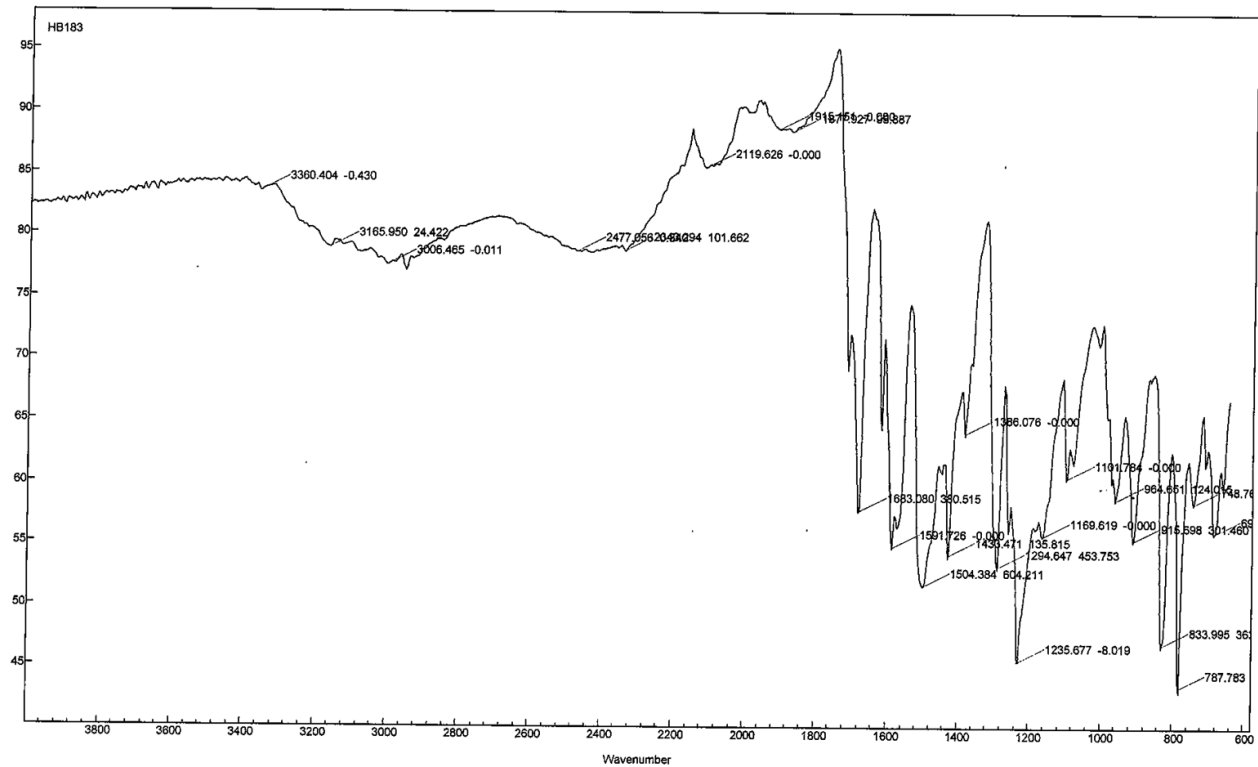


Supporting Information

4-((4-(Allylselanyl)-3-(methoxycarbonyl)phenyl)amino)-4-oxobut-2-enoic acid (HB183)



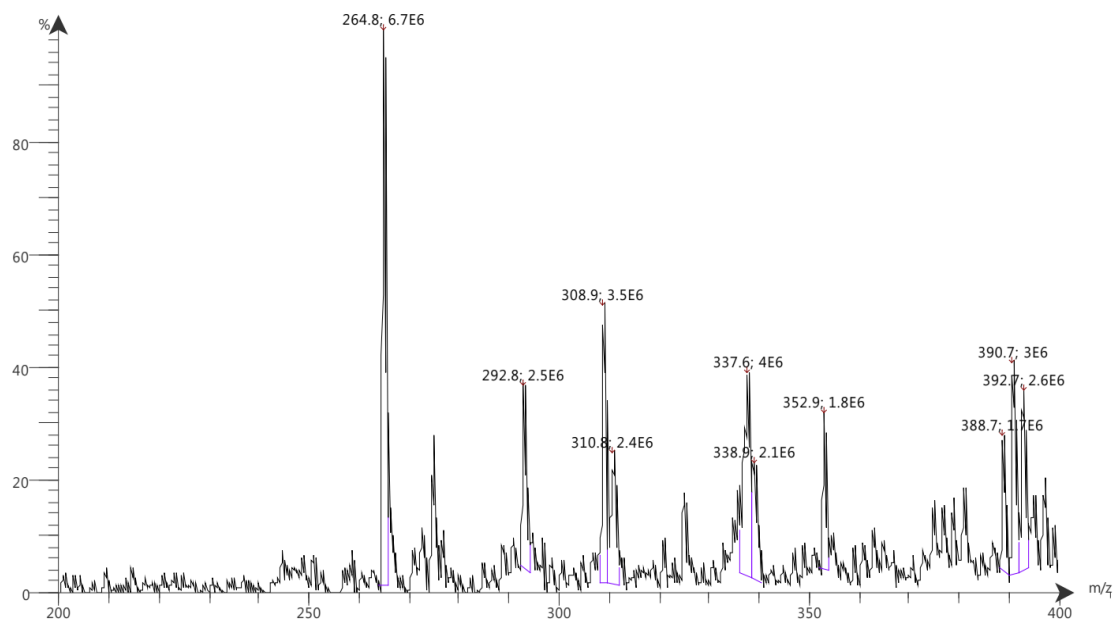
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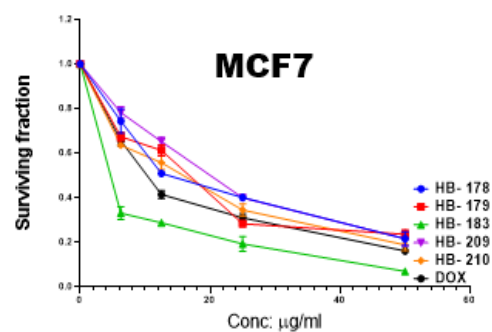
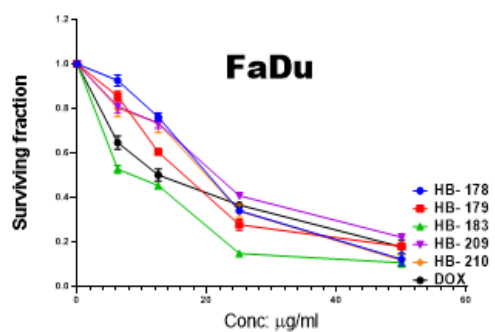
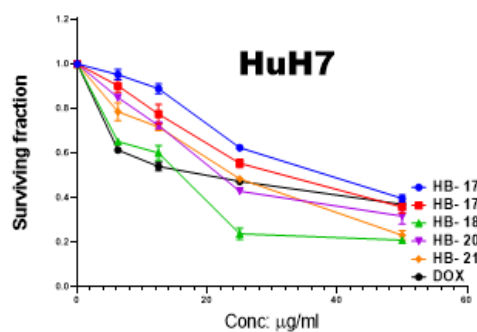
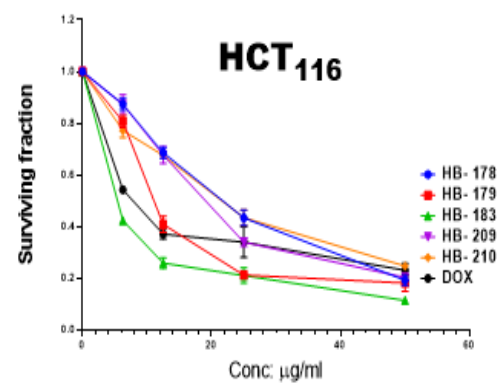
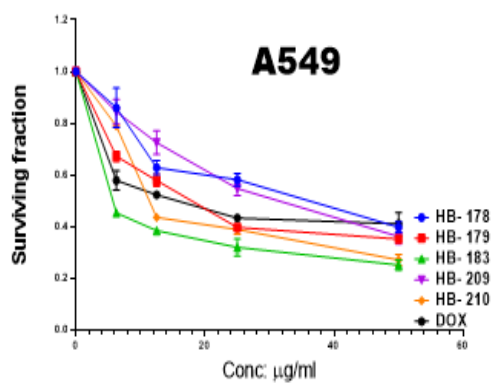
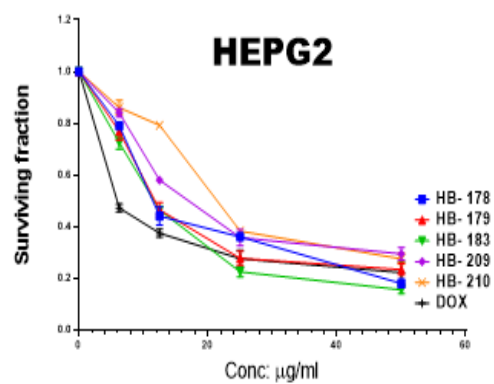


Supporting Information

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ESI - Max: 8E6

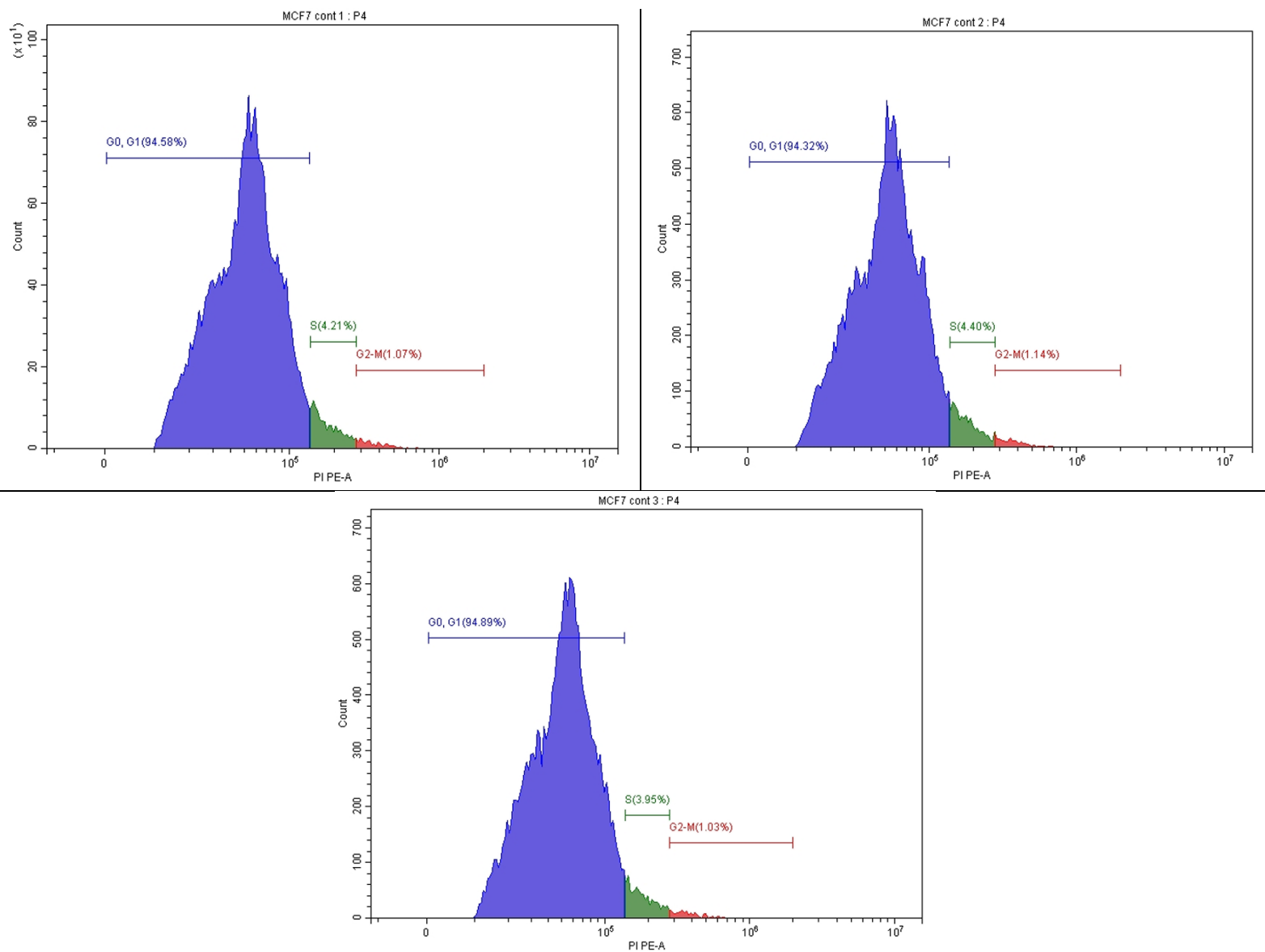
Intensity



Biological Evaluation

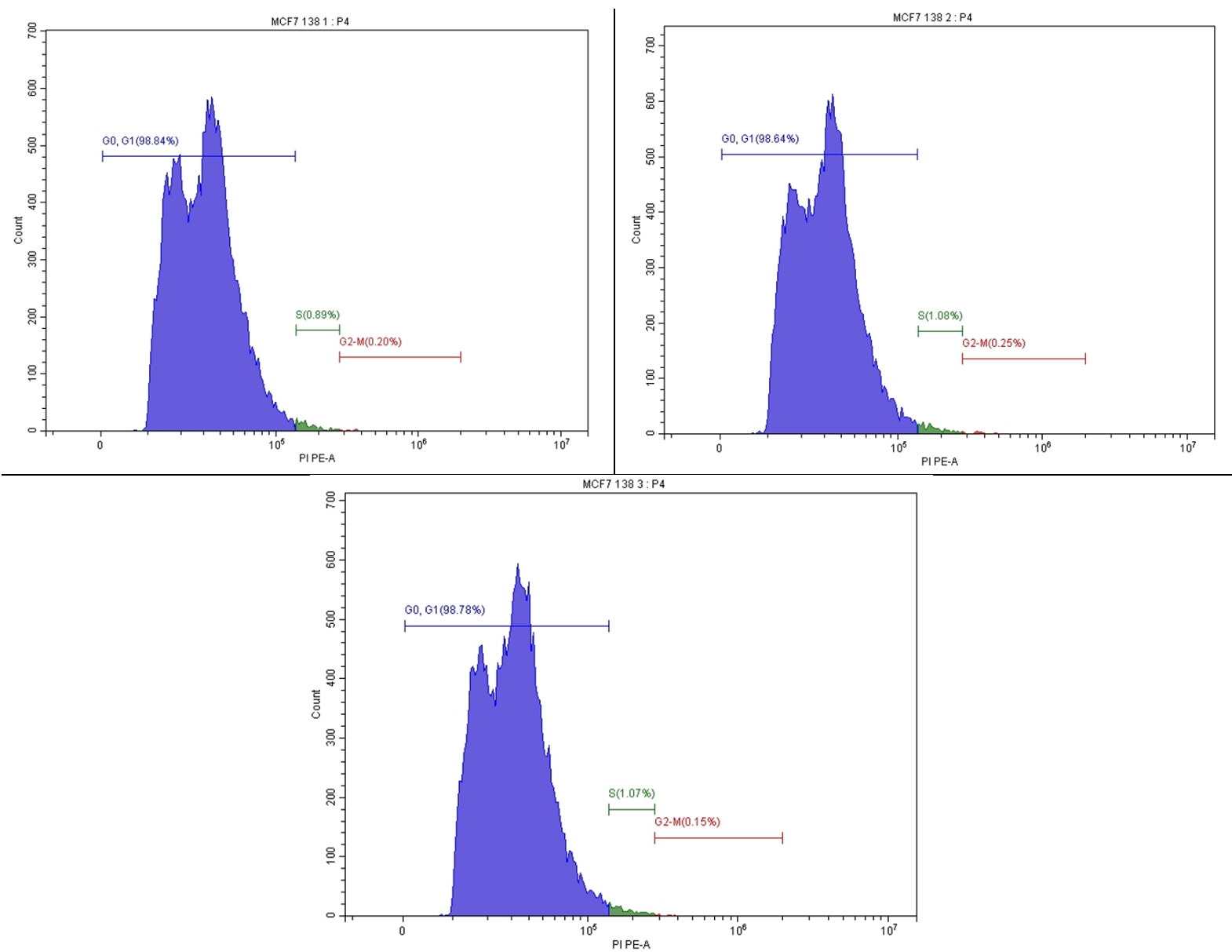
Detailed representation of the IC₅₀ of each tested cell line with the evaluated compound.

Supporting Information



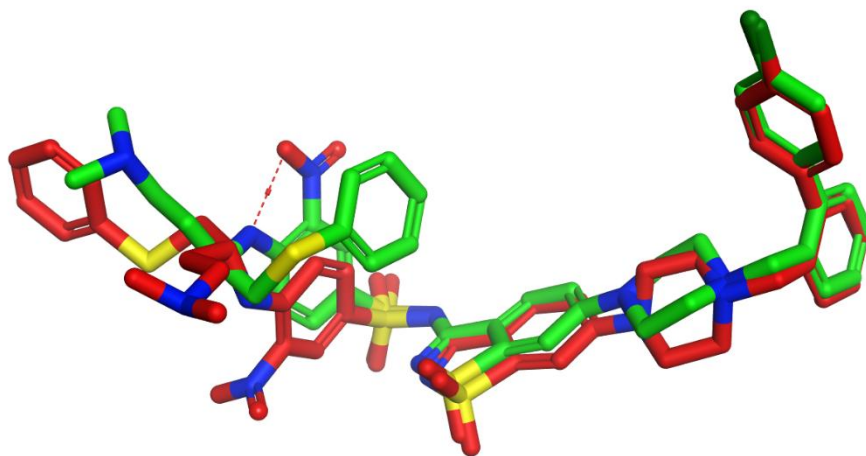
The triplicate experiments of the cell cycle analysis histogram of the untreated control MCF7 cancer cell line.

Supporting Information



The triplicate experiments of the cell cycle analysis histogram of the HB183-treated MCF7 cancer cell line.

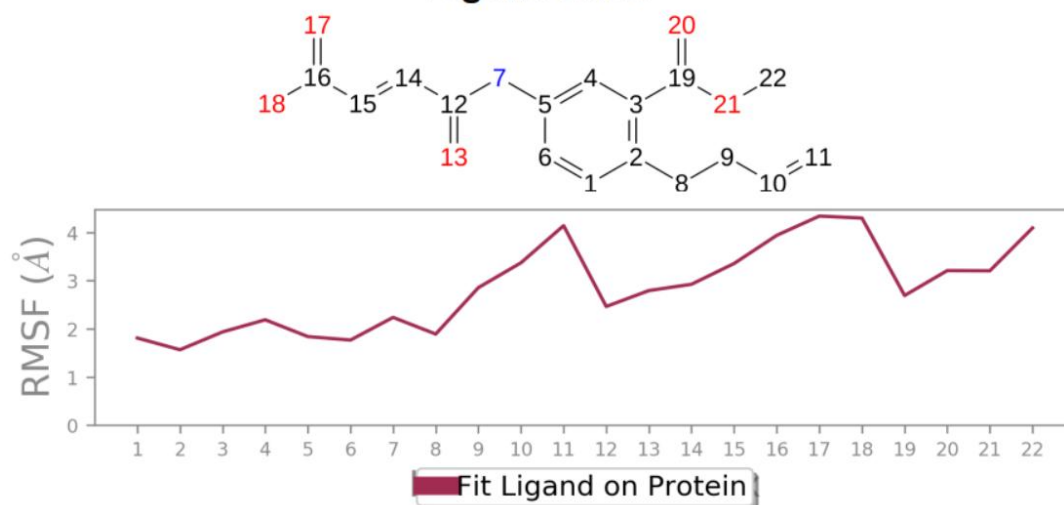
Docking Validation



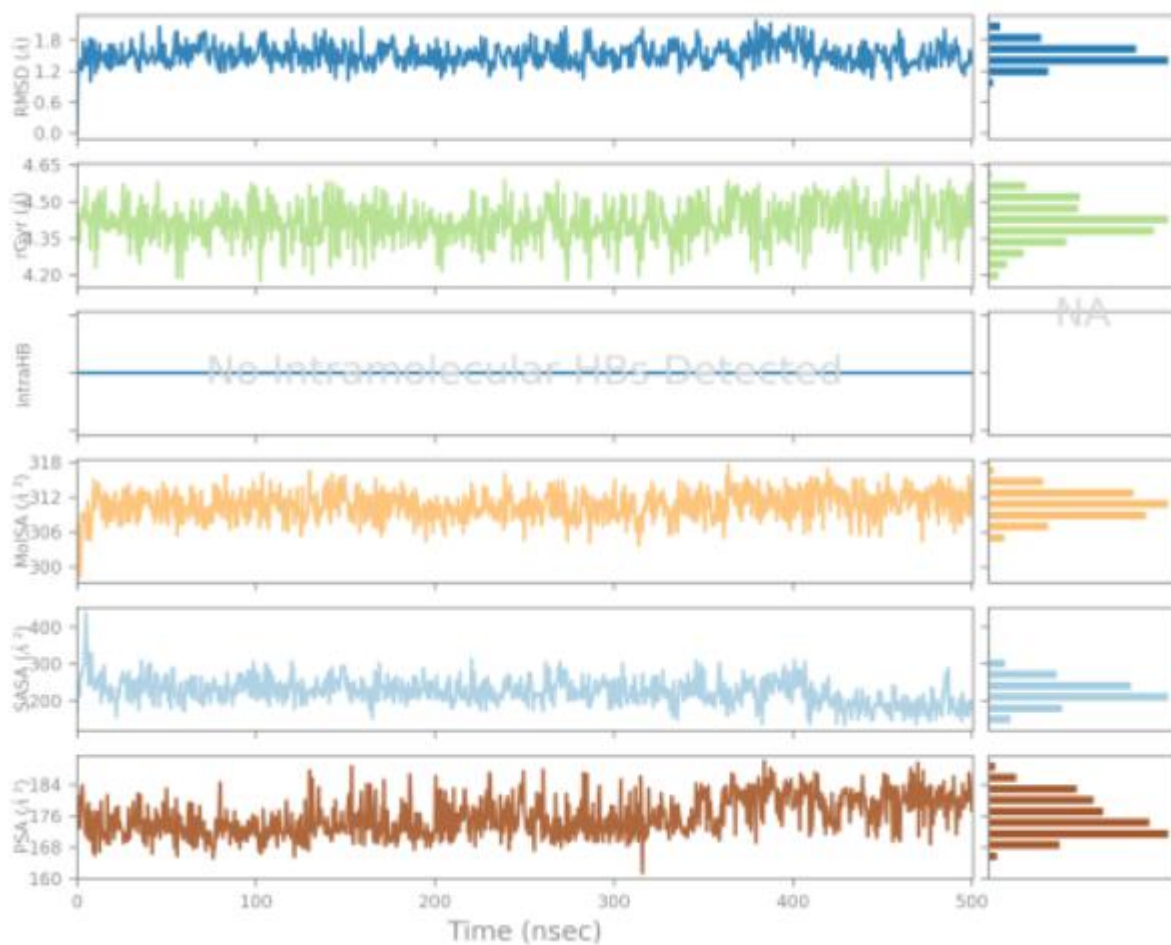
3D Superimposition of the redocked co-crystal (green) of the BCL-2 receptor (PDB ID: 4IEH) over the native one (red) with an RMSD value of 1.68 Å.

Molecular Dynamics Simulation

Ligand RMSF



Ligand Properties



Molecular dynamics (MD) simulation

The molecular dynamics simulation was carried out using the Desmond simulation package of Schrödinger LLC.⁵⁻⁷ The NPT ensemble with the temperature 300 K and a pressure of 1.01 bar was applied in all runs. The simulation length was 500 ns with a relaxation time of 1 ps. The OPLS4 force field parameters were used in all simulations.⁸ The cutoff radius in Coulomb interactions was 9.0 Å. The orthorhombic periodic box boundaries were set 10 Å away from the protein atoms. The water molecules were explicitly described using the transferable intermolecular potential with the three-point (TIP3P) model.⁹ Salt concentration was set to 0.15 M NaCl and was built using the System Builder utility of Desmond. The Martyna–Tuckerman–Klein chain coupling scheme with a coupling constant of 2.0 ps was used for the pressure control, and the Nosé–Hoover chain coupling scheme for the temperature control.^{10, 11} Nonbonded forces were calculated using a RESPA integrator where the short-range forces were updated every step, and the long-range forces were updated every three steps. The trajectories were saved at 300 ps intervals for analysis. The behavior and interactions between the ligands and protein were analyzed using the Simulation Interaction Diagram tool implemented in the Desmond MD package. The stability of MD simulations was monitored by looking at the RMSD of the ligand and protein atom positions as a function of simulation time.

MD trajectory analysis and prime MM-GBSA calculations

The simulation interactions diagram panel of the Maestro software was used to monitor interactions contribution in the ligand-protein stability. The molecular mechanics generalized born/solvent accessibility (MM – GBSA) was performed to calculate the ligand binding free energies and ligand strain energies for docked compounds over the last 50 ns with `thermal_mmgb.py` python script provided by Schrodinger which takes a Desmond trajectory file, splits it into individual snapshots, runs the MM-GBSA calculations on each frame, and outputs the average computed binding energy.

References

1. S. Shaaban, A. Negm, M. A. Sobh and L. A. Wessjohann, *European journal of medicinal chemistry*, 2015, **97**, 190-201.
2. C. Nitsche, V. N. Schreier, M. A. Behnam, A. Kumar, R. Bartenschlager and C. D. Klein, *Journal of Medicinal Chemistry*, 2013, **56**, 8389-8403.
3. B. Al-Abdallah, Y. S. Al-Faiyz and S. Shaaban, *Biomolecules*, 2022, **12**, 1765.
4. B. Al-Abdallah, Y. S. Al-Faiyz and S. Shaaban, *Inorganics*, 2022, **10**, 246.
5. K. J. Bowers, D. E. Chow, H. Xu, R. O. Dror, M. P. Eastwood, B. A. Gregersen, J. L. Klepeis, I. Kolossvary, M. A. Moraes, F. D. Sacerdoti, J. K. Salmon, Y. Shan and D. E. Shaw, 2006.
6. M. H. El-Shershaby, A. Ghiaty, A. H. Bayoumi, A. A. Al-Karmalawy, E. M. Husseiny, M. S. El-Zoghbi and H. S. Abulkhair, *Bioorganic & Medicinal Chemistry*, 2021, **42**, 116266.
7. D. E. S. Research, *Journal*, 2021.
8. E. Harder, W. Damm, J. Maple, C. Wu, M. Reboul, J. Y. Xiang, L. Wang, D. Lupyan, M. K. Dahlgren, J. L. Knight, J. W. Kaus, D. S. Cerutti, G. Krilov, W. L. Jorgensen, R. Abel and R. A. Friesner, *Journal of Chemical Theory and Computation*, 2016, **12**, 281-296.
9. W. L. Jorgensen, J. Chandrasekhar, J. D. Madura, R. W. Impey and M. L. Klein, *Journal of Chemical Physics*, 1983, **79**, 926-935.
10. G. J. Martyna, M. L. Klein and M. Tuckerman, *Journal of Chemical Physics*, 1992, **97**, 2635-2643.
11. G. J. Martyna, D. J. Tobias and M. L. Klein, *Journal of Chemical Physics*, 1994, **101**, 4177-4189.