

SUPPORTING INFORMATION

A general aqueous synthetic strategy towards 1-benzylTHIQs enabled by HOME chemistry

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1. General Experimental Information.

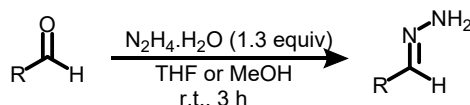
All solvents and reagents were purchased from Ambeed, TCI chemicals, Combi-blocks, and Oakwood and were used without further purification unless otherwise specified. Reactions in organic solvents were carried out in oven-dried vials sealed with aluminum caps and PTFE-faced silicone septa. Procedures involving organic solvents were conducted under a nitrogen atmosphere, with vials prepared and sealed in a glovebox unless otherwise stated. In contrast, reactions in water were performed in 10 mL oven-dried Schlenk tubes sealed under an argon atmosphere. Air-sensitive ligands, catalysts, and reagents were stored in an MBRAUN UNIlab Pro Glove Box Workstation unless otherwise stated.

All NMR experiments (^1H : 500 MHz; ^{13}C : 100, 125 MHz) were recorded on either a Bruker AV500 or Varian MERCURY plus-500 spectrometer at room temperature. ^1H and ^{13}C NMR chemical shifts (δ) were reported in parts per million (ppm) using the residual solvent signal of CDCl_3 as internal standard (CDCl_3 : δ 7.28 for ^1H ; δ 77.00 for ^{13}C). Data are described by δ , multiplicity (s = singlet; d = doublet; dd = doublet of doublets; t = triplet; td = triplet of doublets; q = quartet; quin = quintet; sep = septet; m = multiplet; br = broad), J (Hz), and integration. High-resolution mass spectrometry (HRMS) was performed at McGill University using a maXis ImpactTM QTOF mass spectrometer equipped with electrospray ionization (ESI) and atmospheric Pressure Chemical Ionization (APCI). All solvents were purified and dried using established laboratory protocols or were used directly from a solvent purification system without further treatment.

Reagent-grade solvents were used for all work-up and purification procedures. Short-packed column chromatography was performed using Silicycle SiliaFlash silica gel F60 (230–400 mesh) or Biotage Sfär silica HC D 20 μm and a hand-packed neutral alumina column. The Isolera One Prime advanced automatic flash purification system was used for Flash column Chromatography. Thin layer chromatography (TLC) was performed using Silicycle 60Å 250 μm glass-backed plates. Visualization was accomplished with UV light.

2. Experimental Procedures

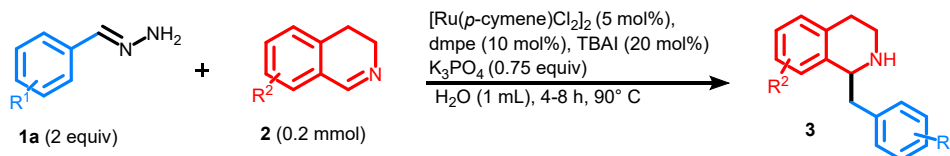
2.1 General procedure for the preparation of hydrazone



Based on reported literature¹, hydrazones were prepared: to a 25 mL round-bottom flask equipped with a magnetic stir bar, hydrazine monohydrate (3.12 mmol, 1.3 equiv.) and solvent (THF or MeOH, 0.5 mL) were added. For polar aldehydes, methanol (MeOH) was used as a solvent. While stirring, the aldehyde (2.40 mmol, 1.0 equiv) was dissolved in solvent (THF or MeOH, 10 mL) and added dropwise to the solution of hydrazine over 10 minutes. An appropriate amount of

anhydrous Na₂SO₄ was added to remove water. After stirring for another 2-3 h, Na₂SO₄ was filtered off, and the organic layer was extracted with H₂O (15 mL) and CH₂Cl₂ (15 mL). The organic layer was washed with brine (15 mL) and dried over Na₂SO₄. Under reduced pressure, the solvent was removed at room temperature to isolate the hydrazone. For polar aldehydes, no extraction was performed. The solvent was removed under reduced pressure at room temperature, and the hydrazone was isolated directly.

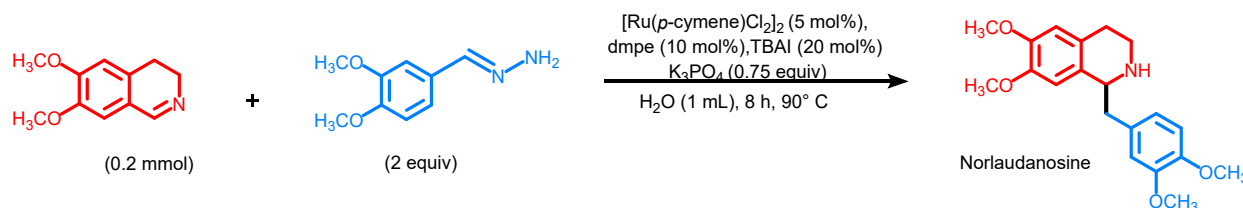
2.2 General procedure for the synthesis of 1-benzyl substituted THIQ



To an oven-dried 10 mL Schlenk tube with a magnetic stir bar, [Ru(*p*-cymene)Cl₂]₂ (6.12 mg, 0.01 mmol), K₃PO₄ (31.8 mg, 0.75 equiv, 0.15 mmol), 3,4-DHIQ (26.2 mg, 0.2 mmol), and TBAI (14.77 mg, 0.04 mmol, 0.2 equiv) were added. The mixture was transferred into a glovebox, where dmpe (3 μL, 0.02 mmol) was added, and the tube was removed from the glovebox. Afterwards, hydrazone (0.4 mmol, 2 equiv) was added, followed by 1 mL of degassed water. For 5 min, air was exchanged with an argon atmosphere, the lid was tightened, and the reaction was sealed. Then the reaction mixture was sonicated for 3-4 min and then stirred at 90 °C for 4 h. Upon completion, the reaction mixture was extracted with ethyl acetate (2 x 15 mL) and concentrated under reduced pressure. The residue was purified by flash chromatography using a hand-packed neutral alumina (Al₂O₃) column with a co-solvent system (ethyl acetate and hexane, 1:1 and then 2–4% methanol/DCM gradient).

2.3- Procedure for synthesis of natural products.

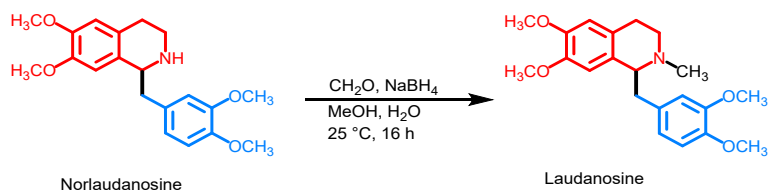
Synthesis of Norlaudanosine (3bo)



To an oven-dried 10 mL Schlenk tube with a magnetic stir bar, [Ru(*p*-cymene)Cl₂]₂ (6.12 mg, 0.01 mmol), TBAI (14.77 mg, 0.04 mmol), K₃PO₄ (31.8 mg, 0.75 equiv), and 6,7-dimethoxy-3,4-DHIQ (38.24 mg, 0.2 mmol) were introduced. The mixture was transferred into a glovebox, where dmpe (3 μL, 0.02 mmol) was added, and the tube was removed from the glovebox. Afterwards, hydrazone (2 equiv) was added, followed by 1 mL of degassed water. For 5 min, air was exchanged with an argon atmosphere, the lid was tightened, and the tube was sealed. The reaction mixture was sonicated for 3-4 min and then stirred at 90 °C for 8 h. Upon completion, the mixture was

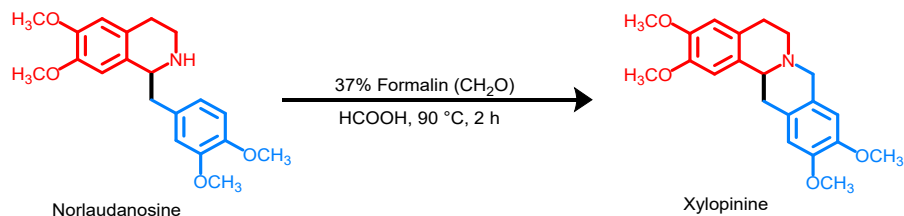
extracted with ethyl acetate (2 x 15 mL) and concentrated under vacuum. The residue was purified by Flash chromatography over a hand-packed neutral alumina column (Al_2O_3) using a cosolvent system (ethyl acetate: hexane, 1:1 and 2-4% methanol/DCM gradient), reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 6.84 – 6.80 (m, 1H), 6.80 – 6.74 (m, 2H), 6.62 (s, 1H), 6.58 (s, 1H), 4.16 (dd, J = 8.9, 4.7 Hz, 1H), 3.86 (s, 3H), 3.85 (s, 3H), 3.84 (s, 3H), 3.81 (s, 3H), 3.26 – 3.12 (m, 2H), 2.97 – 2.86 (m, 2H), 2.74 (tdd, J = 16.1, 11.3, 5.8 Hz, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 148.9, 147.7, 147.5, 147.0, 131.2, 129.9, 127.2, 121.4, 112.4, 111.8, 111.3, 109.4, 56.7, 55.9, 55.9, 55.8, 55.8, 42.1, 40.8, 29.0.

Synthesis of Laudanosine (3bp)



To a solution of Norlaudanosine (70 mg, 0.2 mmol) in MeOH (1.28 mL), aqueous CH_2O (0.42 mL, c = 37 %, 0.0056 mmol) was added. After stirring the reaction for 3 h at 25 °C, NaBH_4 (85 mg, 2.24 mmol) was added slowly. The reaction mixture was stirred for an additional 16 h at 25 °C. Saturated aqueous NH_4Cl (5.3 mL) was added, and the mixture was extracted with CH_2Cl_2 (3×7 mL). The combined organic layers were dried over Na_2SO_4 , and the solvent was concentrated under reduced pressure. The crude product was purified by flash chromatography over neutral Al_2O_3 as a cream-colored solid, giving 85% yield. ^1H NMR (500 MHz, CDCl_3) δ 6.76 (d, J = 8.1 Hz, 1H), 6.66 – 6.59 (m, 2H), 6.56 (s, 1H), 6.03 (s, 1H), 3.84 (s, 3H), 3.84 (s, 3H), 3.79 (s, 3H), 3.75 – 3.71 (m, 1H), 3.56 (s, 3H), 3.26 – 3.15 (m, 2H), 2.88 – 2.74 (m, 3H), 2.65 – 2.58 (m, 1H), 2.56 (s, 3H). $\delta^{13}\text{C}$ NMR (126 MHz, CDCl_3) δ 148.6, 147.4, 147.4, 146.3, 132.1, 128.6, 125.6, 121.9, 113.0, 111.1, 111.1, 111.0, 64.8, 55.9, 55.8, 55.7, 55.5, 46.7, 42.4, 40.8, 25.2. Analytical data were consistent with literature values.⁶

Synthesis of Xylopinine (3bq)



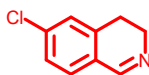
In a 10 mL Schlenk tube, Norlaudanosine (80 mg, 0.23 mmol), 0.33 mL of 88% formic acid, and 0.22 mL of 37% formalin solution were combined and heated at 90 °C for 2 h under a nitrogen atmosphere. After the reaction mixture was cooled to room temperature, it was basified with saturated NaHCO_3 and extracted with dichloromethane (3×10 mL). The combined organic layers

were dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by column chromatography on neutral alumina. (Al_2O_3), yielding 70%, yellow solid. ^1H NMR (500 MHz, CDCl_3) δ 6.73 (s, 1H), 6.66 (s, 1H), 6.61 (s, 1H), 6.57 (s, 1H), 3.94 (d, $J = 14.5$ Hz, 1H), 3.88 (s, 3H), 3.86 (s, 3H), 3.85 (s, 3H), 3.84 (s, 3H), 3.67 (d, $J = 14.5$ Hz, 1H), 3.58 (dd, $J = 11.3, 3.9$ Hz, 1H), 3.24 (dd, $J = 15.7, 3.9$ Hz, 1H), 3.19 – 3.09 (m, 2H), 2.83 (dd, $J = 15.9, 12.0$ Hz, 1H), 2.69 – 2.57 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 147.6, 147.4, 147.4, 147.4, 129.7, 126.7, 126.3, 126.2, 111.3, 111.3, 109.0, 108.5, 59.6, 58.2, 56.0, 55.9, 55.9, 55.8, 51.3, 36.4, 29.0. Analytical data were consistent with literature values.^{5,6}

3. General Procedure and Characterization Data for Starting Compounds 2a-2f

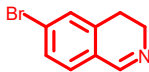
General procedure: *N*-bromosuccinimide (NBS) (4.0 mmol, 4.0 equiv) was added to a solution of 1,2,3,4-tetrahydroisoquinoline (1mmol, 1.0 equiv) in anhydrous CH_2Cl_2 at room temperature under a nitrogen atmosphere. The reaction mixture was stirred at room temperature for 1 h. A 30% v/v NaOH aqueous solution was then added, and the mixture was stirred for an additional hour at room temperature. Then, the aqueous phase was extracted with CH_2Cl_2 (3 x 15 mL), and the combined organic layers were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure.¹

6-Chloro-3,4-dihydroisoquinoline (2a)



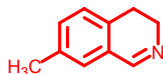
6-Chloro-1,2,3,4-tetrahydroisoquinoline (75%) was synthesized according to general procedure 3 as a yellow oil without further purification. : ^1H NMR (500 MHz, CDCl_3): δ 8.29 (1H, t, $J=2.1$ Hz), 7.25 (1H, m), 7.18 (1H, d, $J=8.0$ Hz), 7.14 (1H, s), 3.75 (2H, ddd, $J=2.3, 7.3, 8.2$ Hz), 2.71 (2H, dd, $J=8.0, 7.6$ Hz). ^{13}C NMR (126 MHz, CDCl_3): δ 159.1, 138.1, 136.6, 128.3, 127.6, 127.2, 126.8, 46.9, 24.8. Data is consistent with the literature.^{2,3}

6-Bromo-3,4-dihydroisoquinoline (2b)



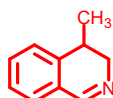
6-Bromo-1,2,3,4-tetrahydroisoquinoline (80%) was synthesized according to general procedure 3 as a yellow oil without further purification. : ^1H NMR (400 MHz, CDCl_3) δ 8.29 (s, 1H), 7.43 (dd, $J = 8.0, 1.6$ Hz, 1H), 7.31 (s, 1H), 7.13 (d, $J = 8.0$ Hz, 1H), 3.77–3.73 (m, 2H), 2.72 (t, $J = 8.0$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 159.3, 138.3, 130.5, 130.2, 128.5, 127.1, 125.1, 46.9, 24.7, Data is consistent with literature.²

7-Methyl-3,4-dihydroisoquinoline (2c)



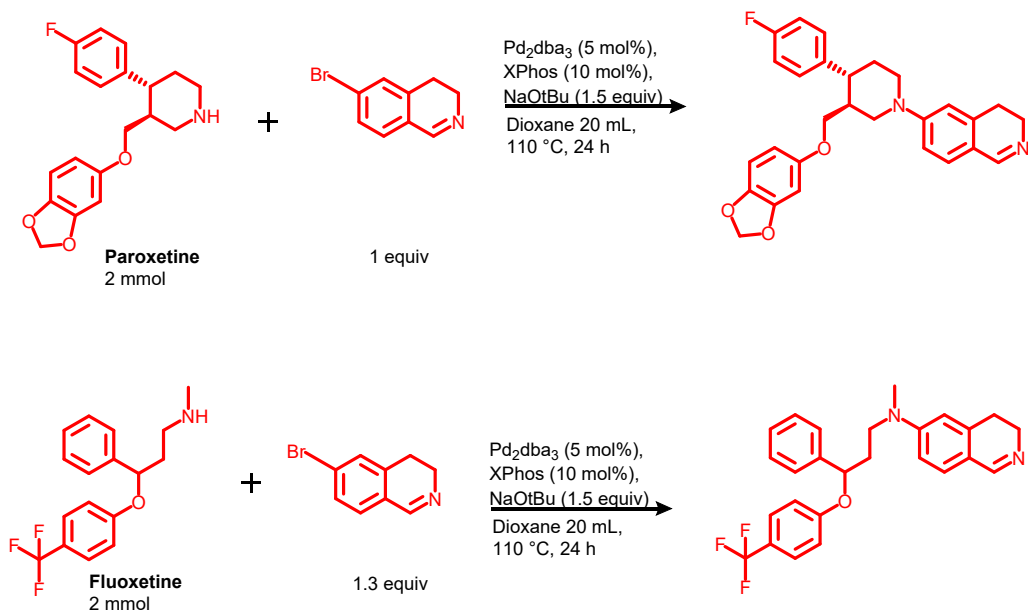
7-Methyl-3,4-dihydroisoquinoline (60%) was synthesized according to General Procedure 3 as a yellow oil and purified using a mixture of ethyl acetate and hexane. ¹H-NMR (400 MHz, CDCl₃): δ 8.27 (s, 1H), 7.14 (d, *J* = 7.7 Hz; 1H), 7.05 (s, 1H), 7.01 (d, *J* = 7.7 Hz; 1H), 3.72 (t, *J* = 7.8 Hz; 2H), 2.67 (t, *J* = 7.8 Hz; 2H), 2.33 (s, 3H); ¹³C-NMR (100 MHz, CDCl₃): δ 160.3, 136.5, 133.1, 131.5, 128.2, 127.7, 127.1, 47.4, 24.5, 20.8. Data is consistent with literature.⁴

4-methyl-3,4-dihydroisoquinoline (2d)



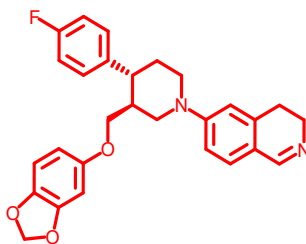
4-methyl-3,4-dihydroisoquinoline (65%) was synthesized according to general procedure 3 as a yellow solid and purified by using ethyl acetate and hexane. ¹H NMR (500 MHz, CDCl₃) δ 8.37 (s, 1H), 7.41 (td, *J* = 7.3, 1.8 Hz, 1H), 7.34 – 7.27 (m, 2H), 7.27 – 7.23 (m, 1H), 3.84 (ddd, *J* = 16.1, 6.2, 2.2 Hz, 1H), 3.55 (ddd, *J* = 16.2, 8.8, 2.3 Hz, 1H), 2.94 – 2.86 (m, 1H), 1.26 (d, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 160.3, 141.3, 131.4, 127.6, 127.3, 126.8, 125.5, 54.5, 29.1, 17.5. HRMS (ESI) *m/z* calculated for C₁₀H₁₂N([M+H]⁺): 146.0964, found: 146.0964.

General procedure for the synthesis of 2e and 2f



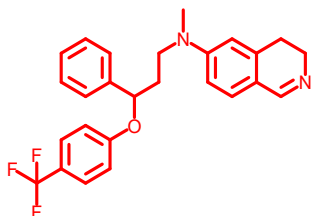
In a 25 mL Schlenk tube equipped with a magnetic stir bar, Pd₂dba₃ (5 mol%), Xphos (10 mol%), NaOtBu (1.5 equiv), Fluoxetine or Paroxetine (2.0 mmol), and 6-bromoisoquinoline (2.0 mmol, 1.0 equiv for Paroxetine or 2.6 mmol, 1.3 equiv for Fluoxetine) were taken in 20 mL dioxane. The reaction mixture was stirred at 110 °C under an argon atmosphere for 24 h. After completion, the product was isolated as a yellow solid by silica gel column chromatography using 10-20 % methanol and DCM.

2-((3S,4R)-3-((benzo[d][1,3]dioxol-5-yloxy)methyl)-4-(4-fluorophenyl)piperidin-1-yl)-3,4-dihydroisoquinoline (2e)



2-((3S,4R)-3-((benzo[d][1,3]dioxol-5-yloxy)methyl)-4-(4-fluorophenyl)piperidin-1-yl)-3,4-dihydroisoquinoline (2e) was prepared according to general procedure. Yield, 60%, yellowish solid. ¹H NMR (500 MHz, CDCl₃) δ 8.22 (s, 1H), 7.18 (d, *J* = 8.4 Hz, 1H), 7.16 – 7.09 (m, 2H), 7.00 – 6.92 (m, 2H), 6.81 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.69 (d, *J* = 2.6 Hz, 1H), 6.61 (d, *J* = 8.5 Hz, 1H), 6.37 (d, *J* = 2.5 Hz, 1H), 6.15 (dd, *J* = 8.5, 2.5 Hz, 1H), 5.84 (s, 2H), 4.17 (dd, *J* = 12.9, 2.1 Hz, 1H), 3.96 (dd, *J* = 14.7, 2.1 Hz, 1H), 3.71 (td, *J* = 7.9, 2.0 Hz, 2H), 3.64 (dd, *J* = 9.5, 3.0 Hz, 1H), 3.57 – 3.48 (m, 1H), 2.99 – 2.93 (m, 1H), 2.89 (t, 1H), 2.72 (t, 2H), 2.66 (td, 1H), 2.28 – 2.15 (m, 1H), 1.90 – 1.81 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 161.6 (d, *J* = 244.8 Hz), 159.9, 154.1, 153.1, 148.2, 141.7, 138.9 (d, *J* = 3.2 Hz), 138.3, 129.8, 128.7 (d, *J* = 7.7 Hz), 119.3, 115.5 (d, *J* = 21.0 Hz), 113.5, 113.0, 107.9, 105.6, 101.1, 98.0, 69.1, 67.0, 49.0, 46.5, 44.1, 41.4, 33.6, 26.0. ¹⁹F NMR (471 MHz, CDCl₃) δ -115.64. HRMS (ESI) *m/z* calculated for C₂₈H₂₈FN₂O₃ ([M+H]⁺): 459.2078, found: 459.2080.

N-methyl-N-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-3,4-dihydroisoquinolin-6-amine (2f)

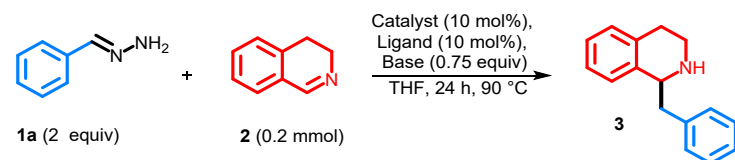


N-methyl-N-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-3,4-dihydroisoquinolin-6-amine (2f) was prepared according to the general procedure. Yield, 51%, Yellowish solid. ¹H NMR (500 MHz, CDCl₃) δ 8.19 (s, 1H), 7.44 (d, *J* = 8.7 Hz, 2H), 7.36 – 7.30 (m, 4H), 7.30 – 7.26

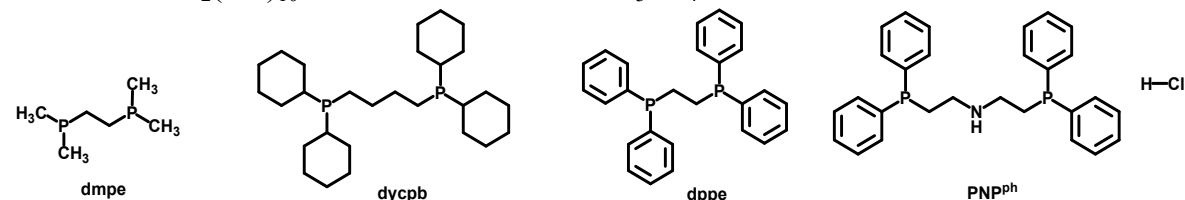
(m, 1H), 7.13 (d, $J = 8.5$ Hz, 1H), 6.90 (d, $J = 8.3$ Hz, 2H), 6.52 (dd, $J = 8.5, 2.6$ Hz, 1H), 6.36 (d, $J = 2.5$ Hz, 1H), 5.20 (dd, $J = 8.3, 4.2$ Hz, 1H), 3.75 – 3.61 (m, 3H), 3.60 – 3.51 (m, 1H), 3.00 (s, 3H), 2.67 – 2.57 (m, 1H), 2.57 – 2.42 (m, 1H), 2.27 – 2.13 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 160.1, 160.0, 151.7, 140.3, 138.5, 130.2, 128.9, 128.1, 126.8 (q, $J = 3.6$ Hz), 125.6, 124.3 (q, $J = 272.2$ Hz), 123.1 (q, $J = 32.5$ Hz), 117.2, 115.7, 110.1, 109.6, 77.5, 48.6, 46.0, 38.4, 36.2, 26.0. ^{19}F NMR (471 MHz, CDCl_3) δ -61.56. HRMS (ESI) m/z calculated for $\text{C}_{26}\text{H}_{26}\text{F}_3\text{N}_2\text{O}([\text{M}+\text{H}]^+)$: 439.1992, found: 439.1992.

4. Optimization Data of Reaction Conditions

Table S1. Optimization of pre-catalyst

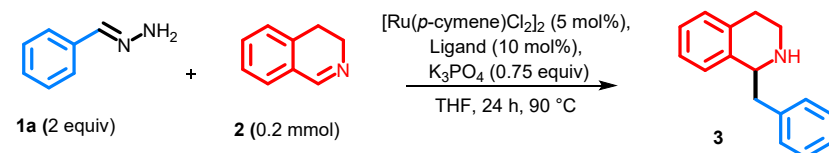


Entry	Catalyst	Ligand	Base	Yield 3 (%)
1	$[\text{Ru}(p\text{-cymene})\text{Cl}_2]_2$	dmpe	K_3PO_4	20
2	$[\text{Ru}(\text{PPh}_3)_3\text{Cl}_2]$	dycpb	K_3PO_4	7
3	$[\text{Ru}(\text{COD})\text{Cl}_2]_n$	dmpe	K_3PO_4	9
4	$\text{Ru}(\text{PNP}^{\text{ph}})(\text{PMe}_3)\text{Cl}_2$		LiOtBu	5
5	Mesityl copper(I)	dppe	LiOtBu	0
6	$[\text{Ir}(\text{OCH}_3)(\text{C}_8\text{H}_{12})_2]$	dmpe	K_3PO_4	0
7	FeCl_2	dmpe	K_3PO_4	0
8	$\text{Mn}_2(\text{CO})_{10}$	PNP^{ph}	K_3PO_4	0

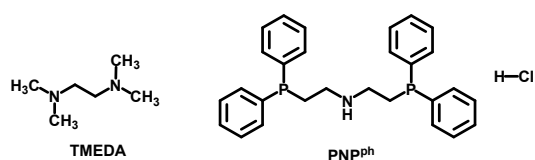


a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), catalyst (10 mol%), ligand (monodentate, 20 mol%, bidentate, 10 mol%), and base (0.15 mmol, 0.75 equiv) in 1.0 mL THF at 90 °C for 24 h under N_2 . ^1H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

Table S2. Optimization of ligand

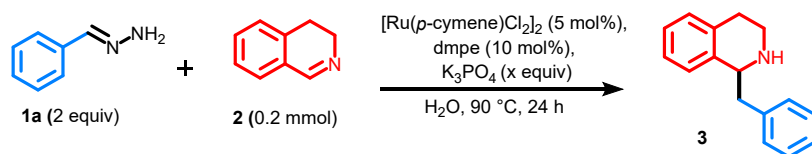


Entry	Ligand	Yield 3 (%)
1	dmpe	20
2	dppe	0
3	PMe ₃	23
4	PCy ₃	20
5	PNP	0
6	dppf	0
7	H ₂ N(CH ₂) ₂ NH ₂	20
8	TMEDA	16



a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (10 mol%), ligand (monodentate, 20 mol%, bidentate, 10 mol%), and base (0.15 mmol, 0.75 equiv) in 1.0 mL THF at 90 °C for 24 h under N₂. ¹H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

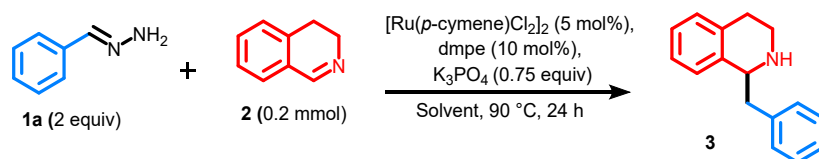
Table S3. Optimization of base loading



Entry	x equiv	Yield (%)
1	1.00	57
2	0.75	70
3	0.50	61

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (10 mol%), dmpe (10 mol%) and K₃PO₄ (x equiv) in 1.0 mL H₂O at 90 °C for 24 h under Ar. ¹H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

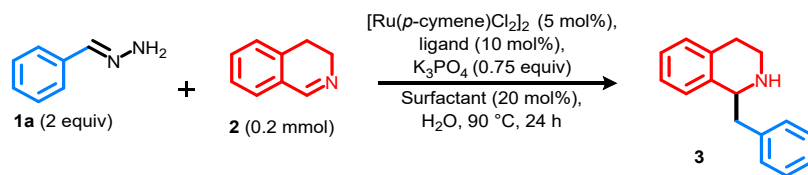
Table S4. Optimization of solvent



Entry	Solvent	Yield (%)
1	THF	20
2	1,4-dioxane	20
3	Toulene	18
4	H ₂ O	70

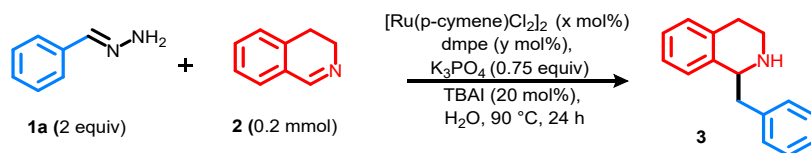
a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (10 mol%), dmpe (10 mol%), and K₃PO₄ (0.15 mmol, 0.75 equiv) in 1.0 mL solvent at 90 °C for 24 h under Ar. ¹H NMR-determined yields using 1,3,5-trimethoxybenzene as an internal standard.

Table S5. Optimization of surfactant and ligand



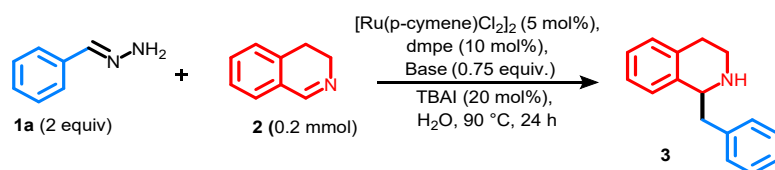
Entry	Surfactant	Ligand	Yield 3 (%)
1	18-crown-6	dmpe	60
2	TPGS-750M	TMEDA	75
3	TPGS-750M	dmpe	71
4	Triton-X-100	dmpe	61
5	Brij C10	dmpe	62
6	TBAI	dmpe	86(80)
7	TBACl	PMe ₃	64
8	TBACl	dmpe	67
9	TBACl	dycpb	54
10	TBACl	TMEDA	62
11	TBACl	H ₂ N(CH ₂) ₆ NH ₂	70
12	TBAH	dmpe	28
13	TBAB	dmpe	74

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (10 mol%), ligand (10 mol%), K₃PO₄ (0.15 mmol, 0.75 equiv), and surfactant (20 mol%) in 1.0 mL H₂O at 90 °C for 24 h under Ar. ¹H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

Table S6. Optimization of catalyst and ligand loading

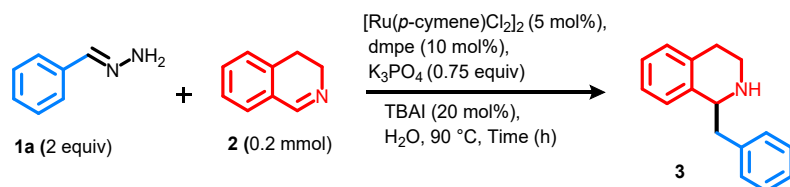
Entry	Catalyst (mol%)	Ligand (mol%)	Yield 3 (%)
1	1.5	3	57
2	4	8	63
3	5	10	86

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), $[\text{Ru}(\text{p-cymene})\text{Cl}_2]_2$ (x mol%), dmpe (y mol%), K_3PO_4 (0.15 mmol, 0.75 equiv), and surfactant (20 mol%) in 1.0 mL H_2O at 90 °C for 24 h under Ar. ^1H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

Table S7. Optimization of Base

Entry	Base	Yield 3 (%)
1	K_3PO_4	86
2	LiO^tBu	80
3	Cs_2CO_3	n.d
4	K_2CO_3	53
5	DBU	n.d

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), $[\text{Ru}(\text{p-cymene})\text{Cl}_2]_2$ (x mol%), dmpe (y mol%), K_3PO_4 (0.15 mmol, 0.75 equiv), and surfactant (20 mol%) in 1.0 mL H_2O at 90 °C for 24 h under Ar. ^1H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

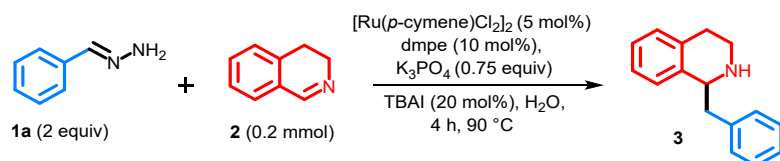
Table S8. Optimization of reaction time

Entry	Time (h)	Yield 3 (%)
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1	48 h	81
2	12 h	80
3	6 h	83
4	4 h	86

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (x mol%), dmpe (y mol%), K₃PO₄ (0.15 mmol, 0.75 equiv), and TBAI (20 mol%) in 1.0 mL H₂O at 90 °C for time (h) under Ar. ¹H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

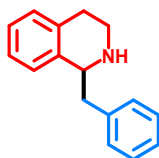
Table S9. Control Experiments



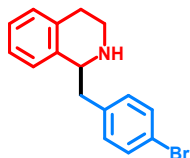
Entry	Condition	Yield 3 (%)
1	Without catalyst	3
2	Without ligand	0
3	Without catalyst +ligand	2.5
4	Without surfactant	70
5	Under air	73

a) Reaction conditions: Hydrazone **1a** (0.4 mmol, 2 equiv), 3,4-DHIQ **2** (0.2 mmol), [Ru(*p*-cymene)Cl₂]₂ (5 mol%), dmpe (10 mol%), K₃PO₄ (0.15 mmol, 0.75 equiv), and TBAI (20 mol%) in 1.0 mL H₂O at 90 °C for 4 h under Ar. ¹H NMR determined yields with 1,3,5-trimethoxybenzene as an internal standard.

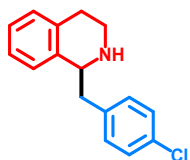
Characterization data of compounds **3aa-3bn**



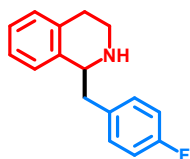
1-benzyl-1,2,3,4-tetrahydroisoquinoline (3aa) was prepared according to General Procedure (2.2), and the reaction was heated for 4 h. Yield, 80%, reddish oil. ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.29 (m, 2H), 7.28 – 7.19 (m, 4H), 7.19 – 7.11 (m, 2H), 7.11 – 7.07 (m, 1H), 4.20 (dd, *J* = 10.2, 3.8 Hz, 1H), 3.30 – 3.16 (m, 2H), 2.96 – 2.73 (m, 4H), 2.04 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 139.2, 138.6, 135.3, 129.4, 128.7, 126.6, 126.2, 125.8, 57.3, 42.6, 40.7, 30.0. HRMS (APCI) *m/z* calculated for C₁₆H₁₈N([M+H]⁺): 224.1434, found: 224.1441.



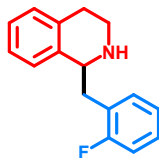
1-(4-bromobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ab) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 79%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.47 – 7.42 (m, 2H), 7.23 – 7.09 (m, 6H), 4.22 (dd, J = 9.8, 4.0 Hz, 1H), 3.25 – 3.17 (m, 2H), 2.99 – 2.74 (m, 4H), 2.22 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.0, 137.9, 135.1, 131.6, 131.1, 129.4, 126.3, 126.1, 125.8, 120.4, 57.0, 41.9, 40.6, 29.7. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{BrN}$ ($[\text{M}+\text{H}]^+$): 302.0539, found: 302.0534.



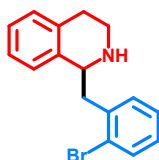
1-(4-chlorobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ac) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 76%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.33 – 7.28 (m, 2H), 7.24 – 7.14 (m, 5H), 7.14 – 7.08 (m, 1H), 4.20 (dd, 1H), 3.28 – 3.17 (m, 2H), 3.00 – 2.71 (m, 4H), 1.98 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.3, 137.6, 135.3, 132.3, 130.7, 129.4, 128.7, 126.3, 126.1, 125.7, 57.1, 41.9, 40.6, 29.9. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{ClN}$ ($[\text{M}+\text{H}]^+$): 258.1044, found: 258.1039.



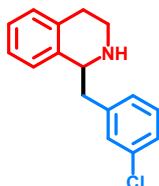
1-(4-fluorobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ad) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 65%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.25 – 7.20 (m, 3H), 7.20 – 7.14 (m, 2H), 7.14 – 7.10 (m, 1H), 7.05 – 6.99 (m, 2H), 4.20 (dd, J = 10.0, 3.9 Hz, 1H), 3.27 – 3.17 (m, 2H), 2.97 – 2.74 (m, 4H), 1.89 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 161. (d, J = 244.6 Hz), 138.2, 135.2, 134.7 (d, J = 3.2 Hz), 130.7 (d, J = 7.8 Hz), 129.4, 126.2, 126.1, 125.7, 115.4 (d, J = 21.1 Hz), 57.2, 41.7, 40.6, 29.8. ^{19}F NMR (471 MHz, CDCl_3) δ -116.63. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{FN}$ ($[\text{M}+\text{H}]^+$): 242.1339, found: 242.1338.



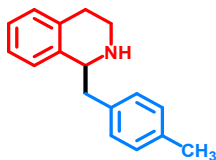
1-(2-fluorobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ae) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h, reddish oil. Yield, 63%. ^1H NMR (500 MHz, CDCl_3) δ 7.32 – 7.21 (m, 3H), 7.22 – 7.14 (m, 2H), 7.14 – 7.05 (m, 3H), 4.27 (dd, $J = 10.4, 3.6$ Hz, 1H), 3.34 – 3.18 (m, 2H), 3.03 – 2.69 (m, 4H), 1.83 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 161.5 (d, $J = 245.0$ Hz), 138.6, 135.1, 131.7 (d, $J = 5.0$ Hz), 129.3, 128.2 (d, $J = 8.2$ Hz), 126.5, 126.4 (d, $J = 15.7$ Hz), 126.2, 125.7, 124.1 (d, $J = 3.7$ Hz), 115.4 (d, $J = 22.4$ Hz), 56.0, 40.0, 36.2, 29.8. ^{19}F NMR (471 MHz, CDCl_3) δ -117.79. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{FN}([\text{M}+\text{H}]^+)$: 242.1340, found: 242.1350.



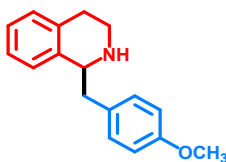
1-(2-bromobenzyl)-1,2,3,4-tetrahydroisoquinoline (3af) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 70%, reddish solid. ^1H NMR (500 MHz, CDCl_3) δ 7.61 (d, $J = 7.5$ Hz, 1H), 7.37 – 7.33 (m, 1H), 7.31 – 7.26 (m, 2H), 7.22 – 7.09 (m, 4H), 4.36 (dd, $J = 10.5, 3.5$ Hz, 1H), 3.50 – 3.36 (m, 1H), 3.35 – 3.23 (m, 1H), 3.08 – 2.77 (m, 4H), 1.87 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.7, 138.6, 135.1, 133.1, 131.9, 129.3, 128.2, 127.4, 126.5, 126.2, 125.8, 124.9, 55.1, 42.9, 40.1, 29.9. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{BrN}([\text{M}+\text{H}]^+)$: 302.0539, Found: 302.0536.



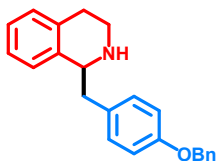
1-(3-chlorobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ag) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 76%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.29 – 7.27 (m, 1H), 7.26 – 7.14 (m, 6H), 7.13 – 7.10 (m, 1H), 4.22 (dd, $J = 10.1, 3.9$ Hz, 1H), 3.31 – 3.17 (m, 2H), 3.00 – 2.71 (m, 4H), 2.05 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 141.2, 138.1, 135.2, 134.4, 129.8, 129.4, 129.4, 127.5, 126.7, 126.3, 126.1, 125.8, 57.0, 42.3, 40.6, 29.8. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{ClN}([\text{M}+\text{H}]^+)$: 258.1044, found: 258.1037.



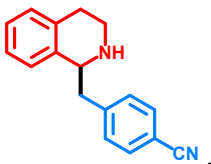
1-(4-methylbenzyl)-1,2,3,4-tetrahydroisoquinoline (3ah) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 78%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.29 – 7.27 (m, 1H), 7.22 – 7.09 (m, 7H), 4.20 (dd, J = 10.4, 3.7 Hz, 1H), 3.28 – 3.17 (m, 2H), 2.97 – 2.75 (m, 4H), 2.36 (s, 3H), 1.93 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.7, 136.1, 136.0, 135.3, 129.4, 129.3, 129.2, 126.2, 126.1, 125.7, 57.3, 42.0, 40.7, 30.0, 21.0. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{N}$ ($[\text{M}+\text{H}]^+$): 238.1590, found 238.1596.



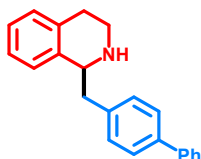
1-(4-methoxybenzyl)-1,2,3,4-tetrahydroisoquinoline (3ai) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 76%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.25 – 7.09 (m, 6H), 6.91 – 6.85 (m, 2H), 4.19 (dd, J = 10.0, 3.9 Hz, 1H), 3.81 (s, 3H), 3.27 – 3.18 (m, 2H), 2.98 – 2.75 (m, 4H), 2.13 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 158.3, 138.4, 135.1, 130.9, 130.3, 129.3, 128.6, 126.2, 125.7, 114.0, 57.3, 55.3, 41.4, 40.6, 29.8. HRMS (APCI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{ON}$ ($[\text{M}+\text{H}]^+$): 254.1539, found: 254.1536.



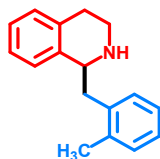
1-(4-(benzyloxy)benzyl)-1,2,3,4-tetrahydroisoquinoline (3aj) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 57%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.48 – 7.36 (m, 4H), 7.34 (d, J = 7.2 Hz, 1H), 7.26 – 7.22 (m, 1H), 7.21 – 7.14 (m, 4H), 7.13 – 7.09 (m, 1H), 6.95 (d, J = 8.8 Hz, 2H), 5.06 (s, 2H), 4.18 (dd, J = 10.1, 4.0 Hz, 1H), 3.26 – 3.16 (m, 2H), 2.98 – 2.73 (m, 4H), 1.83 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 157.6, 137.0, 134.7, 132.8, 130.7, 130.4, 129.3, 128.6, 127.9, 127.5, 126.4, 126.3, 125.8, 115.0, 70.0, 57.0, 41.3, 40.4, 29.3. HRMS (APCI) m/z calculated for $\text{C}_{23}\text{H}_{24}\text{ON}$ ($[\text{M}+\text{H}]^+$): 330.1852, found: 330.1858.



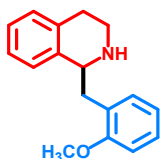
4-((1,2,3,4-tetrahydroisoquinolin-1-yl)methyl)benzonitrile (3ak) was prepared according to General Procedure (2.2), and the reaction was heated for 24 h. Yield, 65 %, reddish solid. ^1H NMR (500 MHz, CDCl_3) δ 7.62 (d, J = 8.4 Hz, 2H), 7.38 (d, J = 8.3 Hz, 2H), 7.22 – 7.15 (m, 3H), 7.14 – 7.09 (m, 1H), 4.25 (dd, J = 10.0, 4.0 Hz, 1H), 3.32 – 3.17 (m, 2H), 3.04 – 2.90 (m, 2H), 2.87 – 2.69 (m, 2H), 1.58 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 145.1, 138.0, 135.3, 132.3, 130.2, 129.5, 126.4, 126.1, 125.8, 118.9, 110.3, 56.9, 42.9, 40.6, 29.8. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{17}\text{N}_2$ ($[\text{M}+\text{H}]^+$): 249.1386, found: 249.1384.



1-([1,1'-biphenyl]-4-ylmethyl)-1,2,3,4-tetrahydroisoquinoline (3al) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 82%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.63 – 7.54 (m, 4H), 7.44 (t, J = 7.8 Hz, 2H), 7.39 – 7.31 (m, 3H), 7.25 – 7.21 (m, 1H), 7.21 – 7.16 (m, 2H), 7.15 – 7.10 (m, 1H), 4.34 (dd, J = 9.7, 4.2 Hz, 1H), 3.37 – 3.23 (m, 2H), 3.08 – 2.95 (m, 2H), 2.96 – 2.79 (m, 2H), 2.51 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 140.8, 139.5, 137.8, 137.7, 134.8, 129.8, 129.3, 128.7, 127.3, 127.2, 127.0, 126.4, 126.3, 125.8, 57.0, 42.0, 40.5, 29.4. HRMS (ESI) m/z calculated for $\text{C}_{22}\text{H}_{22}\text{N}$ ($[\text{M}+\text{H}]^+$): 300.1747, found: 300.1737.

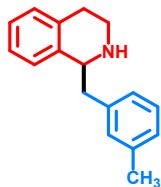


1-(2-methylbenzyl)-1,2,3,4-tetrahydroisoquinoline (3am) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 75%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.34 – 7.26 (m, 5H), 7.26 – 7.19 (m, 3H), 4.30 (dd, J = 10.5, 3.7 Hz, 1H), 3.39 – 3.32 (m, 2H), 3.08 – 2.88 (m, 4H), 2.50 (s, 3H), 2.28 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.8, 137.5, 136.6, 135.2, 130.6, 130.2, 129.4, 126.6, 126.3, 126.2, 126.1, 125.7, 55.8, 40.4, 39.9, 29.9, 19.7. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{N}$ ($[\text{M}+\text{H}]^+$): 238.1590, found: 238.1583.

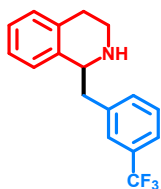


1-(2-methoxybenzyl)-1,2,3,4-tetrahydroisoquinoline (3an) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 91%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.33 – 7.28 (m, 1H), 7.28 – 7.24 (m, 1H), 7.22 – 7.14 (m, 3H), 7.14 – 7.08 (m, 1H), 6.96

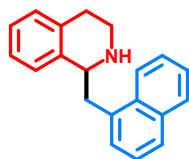
– 6.90 (m, 2H), 4.30 (dd, $J = 10.3, 3.5$ Hz, 1H), 3.88 (s, 3H), 3.39 – 3.32 (m, 1H), 3.31 – 3.25 (m, 1H), 2.99 – 2.92 (m, 1H), 2.91 – 2.80 (m, 3H), 2.29 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 157.9, 139.2, 135.1, 131.3, 129.2, 127.9, 127.8, 126.8, 126.1, 125.7, 120.6, 110.5, 55.5, 55.4, 40.1, 37.6, 29.9. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{NO}([\text{M}+\text{H}]^+)$: 254.1539, found: 254.1533.



1-(3-methylbenzyl)-1,2,3,4-tetrahydroisoquinoline (3ao) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 76%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.25 – 7.21 (m, 2H), 7.20 – 7.15 (m, 2H), 7.14 – 7.06 (m, 4H), 4.25 (dd, $J = 10.1, 3.9$ Hz, 1H), 3.29 – 3.20 (m, 2H), 2.99 – 2.77 (m, 4H), 2.46 (s, 1H), 2.36 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.6, 138.1, 138.0, 134.8, 130.0, 129.1, 128.4, 127.1, 126.2, 126.1, 126.1, 125.6, 57.0, 42.2, 40.4, 29.5, 21.3. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{N}([\text{M}+\text{H}]^+)$: 238.1590, found: 238.1579.

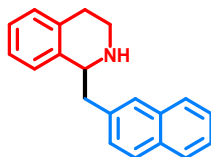


1-(3-(trifluoromethyl)benzyl)-1,2,3,4-tetrahydroisoquinoline (3ap) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 68%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.55 – 7.41 (m, 4H), 7.25 – 7.08 (m, 4H), 4.25 (dd, $J = 10.1, 3.8$ Hz, 1H), 3.36 – 3.18 (m, 2H), 3.06 – 2.74 (m, 4H), 1.79 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 140.2, 138.1, 135.3, 132.8, 130.8 (q, $J = 32.0$ Hz), 129.4, 128.9, 126.3, 126.1, 126.0 (q, $J = 3.8$ Hz), 125.8, 125.2 (q, $J = 272.3$ Hz), 123.4 (q, $J = 3.9$ Hz), 57.0, 42.5, 40.6, 29.8. ^{19}F NMR (471 MHz, CDCl_3) δ -62.51. HRMS (APCI) m/z calculated for $\text{C}_{17}\text{H}_{17}\text{NF}_3([\text{M}+\text{H}]^+)$: 292.1307, found: 292.1304.

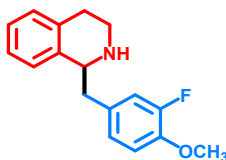


1-(naphthalen-1-ylmethyl)-1,2,3,4-tetrahydroisoquinoline (3as) was prepared according to General Procedure (2.2), and the reaction was heated for 24 h. Yield, 59%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 8.06 (d, $J = 8.9$ Hz, 1H), 7.73 (dd, $J = 8.1, 1.6$ Hz, 1H), 7.62 (d, $J = 7.8$ Hz, 1H), 7.42 – 7.34 (m, 2H), 7.34 – 7.27 (m, 1H), 7.27 – 7.22 (m, 2H), 7.10 – 7.02 (m, 2H), 7.01 – 6.97 (m, 1H), 4.23 (dd, $J = 10.4, 3.5$ Hz, 1H), 3.69 (dd, $J = 13.9, 3.6$ Hz, 1H), 3.11 – 3.03 (m, 2H),

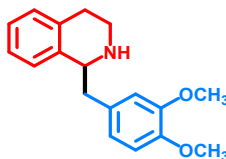
2.80 – 2.61 (m, 3H), 1.57 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.8, 135.2, 135.0, 134.1, 132.0, 129.4, 128.9, 127.8, 127.4, 126.3, 126.2, 126.0, 125.7, 125.7, 125.5, 123.7, 56.1, 40.6, 39.7, 29.9. HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{20}\text{N}([\text{M}+\text{H}]^+)$: 274.1430, found: 274.1584.



1-(naphthalen-2-ylmethyl)-1,2,3,4-tetrahydroisoquinoline (3at) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 91%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.88 – 7.77 (m, 3H), 7.74 (s, 1H), 7.51 – 7.41 (m, 3H), 7.31 – 7.27 (m, 1H), 7.24 – 7.16 (m, 2H), 7.16 – 7.12 (m, 1H), 4.37 (dd, J = 10.0, 4.0 Hz, 1H), 3.47 (dd, J = 13.7, 4.0 Hz, 1H), 3.29 – 3.20 (m, 1H), 3.16 – 3.07 (m, 1H), 3.00 – 2.80 (m, 3H), 2.74 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.1, 136.4, 135.1, 133.6, 132.3, 129.4, 128.3, 128.0, 127.7, 127.6, 127.5, 126.3, 126.2, 126.1, 125.8, 125.5, 57.0, 42.6, 40.7, 29.6. HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{20}\text{N}([\text{M}+\text{H}]^+)$: 274.1590, found: 274.1584.

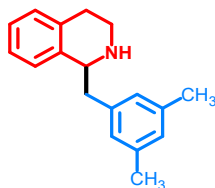


1-(3-fluoro-4-methoxybenzyl)-1,2,3,4-tetrahydroisoquinoline (3au) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 55%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.20 – 7.09 (m, 4H), 7.01 – 6.95 (m, 2H), 6.94 – 6.89 (m, 1H), 4.26 (dd, 1H), 3.88 (s, 3H), 3.27 – 3.17 (m, 2H), 3.06 – 2.91 (m, 2H), 2.90 – 2.78 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 152.3 (d, J = 245.8 Hz), 146.3 (d, J = 10.5 Hz), 137.1, 134.6, 131.4 (d, J = 6.0 Hz), 129.4, 126.5, 126.2, 125.9, 125.1 (d, J = 3.4 Hz), 117.0 (d, J = 18.0 Hz), 113.5 (d, J = 2.3 Hz), 56.8, 56.3, 41.2, 40.41, 29.1. ^{19}F NMR (471 MHz, CDCl_3) δ -134.95 – -135.01 (m, 1F). HRMS (APCI) m/z calculated for $\text{C}_{17}\text{H}_{19}\text{FNO}([\text{M}+\text{H}]^+)$: 272.1445, found: 272.1442.

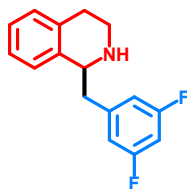


1-(3,4-dimethoxybenzyl)-1,2,3,4-tetrahydroisoquinoline (3av) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 41%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.28 – 7.22 (m, 1H), 7.21 – 7.13 (m, 2H), 7.13 – 7.08 (m, 1H), 6.85 – 6.79 (m, 2H), 6.73 (d, J = 1.8 Hz, 1H), 4.23 (dd, J = 9.6, 3.9 Hz, 1H), 3.87 (s, 3H), 3.84 (s, 3H), 3.28 – 3.18 (m, 2H), 3.00 – 2.85 (m, 2H), 2.87 – 2.73 (m, 2H), 1.87 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3)

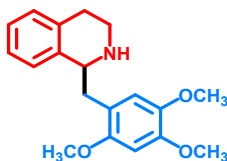
δ 148.9, 147.7, 138.0, 135.2, 131.1, 129.3, 126.2, 126.2, 125.7, 121.4, 112.4, 111.3, 57.1, 55.9, 55.9, 41.9, 40.8, 29.7. HRMS (APCI) m/z calculated for $C_{18}H_{22}O_2N([M+H]^+)$: 284.1645, found: 284.1642.



1-(2,4-dimethylbenzyl)-1,2,3,4-tetrahydroisoquinoline (3aw) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h, reddish oil. Yield, 72%. 1H NMR (500 MHz, $CDCl_3$) δ 7.29 – 7.26 (m, 1H), 7.22 – 7.10 (m, 3H), 6.93 – 6.87 (m, 3H), 4.20 (dd, J = 10.4, 3.8 Hz, 1H), 3.28 – 3.18 (m, 2H), 2.98 – 2.74 (m, 4H), 2.32 (s, 6H), 1.92 (s, 1H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 139.0, 138.7, 138.1, 135.2, 129.3, 128.1, 127.1, 126.1, 126.1, 125.7, 57.3, 42.3, 40.7, 29.9, 21.3. HRMS (APCI) m/z calculated for $C_{18}H_{22}N([M+H]^+)$: 252.1746, found: 252.1743.

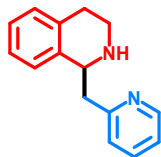


1-(3,5-difluorobenzyl)-1,2,3,4-tetrahydroisoquinoline (3ax) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 60%, reddish solid. 1H NMR (500 MHz, $CDCl_3$) δ 7.20 – 7.14 (m, 3H), 7.15 – 7.10 (m, 1H), 6.85 – 6.78 (m, 2H), 6.74 – 6.66 (m, 1H), 4.23 (dd, J = 10.1, 3.8 Hz, 1H), 3.27 – 3.16 (m, 2H), 3.02 – 2.72 (m, 4H), 1.70 (s, 1H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 163.0 (dd, J = 248.4, 13.0 Hz), 143.2 (t, J = 9.0 Hz), 137.9, 135.3, 129.5, 126.4, 126.1, 125.8, 112.12 (dd, J = 13.3, 5.6 Hz), 102.0 (t, J = 25.3 Hz), 56.8, 42.4 (t, J = 2.1 Hz), 40.5, 29.8. ^{19}F NMR (471 MHz, $CDCl_3$) δ -62.51 (s, 2F). HRMS (APCI) m/z calculated for $C_{16}H_{16}F_2N([M+H]^+)$: 260.1245, found: 260.1253.

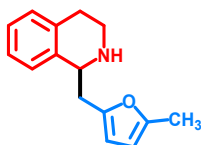


1-(2,4,5-trimethoxybenzyl)-1,2,3,4-tetrahydroisoquinoline (3ay) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 60%, reddish oil. 1H NMR (500 MHz, $CDCl_3$) δ 7.26 – 7.22 (m, 1H), 7.20 – 7.11 (m, 2H), 7.12 – 7.06 (m, 1H), 6.72 (s, 1H), 6.55 (s, 1H), 4.33 (dd, J = 9.8, 3.8 Hz, 1H), 3.89 (s, 3H), 3.83 (s, 3H), 3.81 (s, 3H), 3.32 – 3.25 (m, 2H), 3.02 – 2.93 (m, 1H), 2.88 – 2.81 (m, 3H), 2.47 (s, 1H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 151.9,

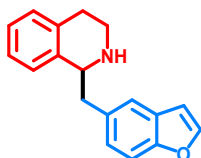
148.3, 142.8, 134.7, 132.1, 129.1, 126.6, 126.2, 125.8, 115.2, 97.7, 56.6, 56.3, 56.2, 55.8, 40.3, 36.8, 29.3. HRMS (ESI) m/z calculated for $C_{19}H_{24}O_3N([M+H]^+)$: 314.1750, found: 314.1749.



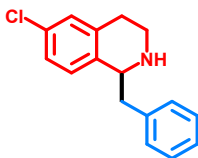
1-(pyridin-2-ylmethyl)-1,2,3,4-tetrahydroisoquinoline (3az) was prepared according to General Procedure (2.2), and the reaction was heated for 24 h. Yield, 56%, reddish oil. 1H NMR (500 MHz, $CDCl_3$) δ 8.56 (d, J = 3.9 Hz, 1H), 7.62 (td, J = 7.6, 1.8 Hz, 1H), 7.24 – 7.21 (m, 1H), 7.21 – 7.13 (m, 4H), 7.12 – 7.08 (m, 1H), 4.63 (dd, J = 9.7, 3.5 Hz, 1H), 3.48 – 3.40 (m, 1H), 3.35 – 3.27 (m, 1H), 3.24 – 3.13 (m, 1H), 3.10 – 3.01 (m, 1H), 2.95 (s, 1H), 2.91 – 2.78 (m, 2H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 159.2, 149.2, 137.3, 136.7, 134.8, 129.3, 126.4, 126.2, 126.0, 124.2, 121.7, 55.6, 43.4, 40.3, 29.2. HRMS (ESI) m/z calculated for $C_{15}H_{17}N_2([M+H]^+)$: 225.1386, found: 225.1376.



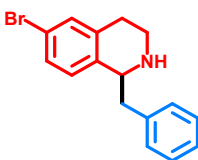
1-((5-methylfuran-2-yl)methyl)-1,2,3,4-tetrahydroisoquinoline (3ba) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 57%, reddish solid. 1H NMR (500 MHz, $CDCl_3$) δ 7.18 – 7.07 (m, 4H), 5.98 (d, J = 3.0 Hz, 1H), 5.88 (d, J = 1.1 Hz, 1H), 4.31 (dd, J = 9.8, 3.9 Hz, 1H), 3.28 – 3.20 (m, 1H), 3.20 – 3.13 (m, 1H), 3.04 – 2.93 (m, 2H), 2.92 – 2.76 (m, 2H), 2.40 (s, 1H), 2.28 (s, 3H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 151.1, 151.1, 137.7, 135.1, 129.2, 126.3, 126.2, 125.7, 108.0, 106.1, 54.8, 40.3, 35.1, 29.6, 13.6. HRMS (APCI) m/z calculated for $C_{15}H_{18}NO([M+H]^+)$: 228.1383, found; 228.1377.



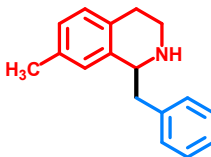
1-(benzofuran-5-ylmethyl)-1,2,3,4-tetrahydroisoquinoline (3bb) was prepared according to General Procedure (2.2), and the reaction was heated for 8 h. Yield, 60%, reddish oil. 1H NMR (500 MHz, $CDCl_3$) δ 7.62 (d, J = 2.1 Hz, 1H), 7.52 – 7.49 (m, 1H), 7.47 (d, J = 8.4 Hz, 1H), 7.32 – 7.27 (m, 1H), 7.23 – 7.15 (m, 3H), 7.15 – 7.10 (m, 1H), 6.74 (dd, J = 2.2, 0.9 Hz, 1H), 4.25 (dd, J = 10.3, 3.8 Hz, 1H), 3.54 – 3.32 (m, 1H), 3.29 – 3.19 (m, 1H), 3.06 – 2.72 (m, 4H), 1.92 (s, 1H). ^{13}C NMR (126 MHz, $CDCl_3$) δ 153.9, 145.3, 138.6, 135.2, 133.4, 129.3, 127.7, 126.1, 126.1, 125.7, 125.5, 121.6, 111.4, 106.4, 57.6, 42.3, 40.7, 29.9. HRMS (APCI) m/z calculated for $C_{18}H_{18}ON([M+H]^+)$: 264.1382, found: 264.1381.



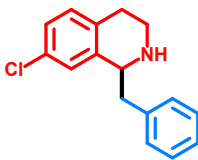
1-benzyl-6-chloro-1,2,3,4-tetrahydroisoquinoline (3be) was prepared according to General Procedure (2.2), and the reaction was heated for 6 h. Yield, 70%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.37 – 7.30 (m, 2H), 7.28 – 7.24 (m, 3H), 7.18 – 7.12 (m, 2H), 7.11 (s, 1H), 4.17 (dd, J = 10.2, 3.9 Hz, 1H), 3.29 – 3.12 (m, 2H), 2.99 – 2.67 (m, 4H), 1.66 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.7, 137.3, 137.1, 131.6, 129.3, 129.0, 128.7, 127.6, 126.6, 125.8, 56.9, 42.4, 40.3, 29.9. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{ClN}$ ($[\text{M}+\text{H}]^+$): 258.1044, found: 258.1046.



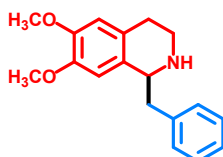
1-benzyl-6-bromo-1,2,3,4-tetrahydroisoquinoline (3bf) was prepared according to General Procedure (2.2), and the reaction was heated for 6 h. Yield, 78%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.31 (m, 2H), 7.30 – 7.27 (m, 2H), 7.27 – 7.23 (m, 3H), 7.04 (d, J = 8.7 Hz, 1H), 4.22 (dd, J = 9.4, 4.4 Hz, 1H), 3.26 – 3.16 (m, 2H), 3.00 – 2.91 (m, 2H), 2.91 – 2.77 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 138.4, 137.4, 137.1, 132.0, 129.3, 128.8, 128.7, 128.0, 126.7, 119.9, 56.8, 42.2, 40.2, 29.5. HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{NBr}$ ($[\text{M}+\text{H}]^+$): 302.0538, found: 302.0536.



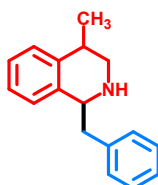
1-benzyl-7-methyl-1,2,3,4-tetrahydroisoquinoline (3bg) was prepared according to General Procedure (2.2), and the reaction was heated to 6 h. Yield, 75%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.32 (m, 2H), 7.30 – 7.22 (m, 3H), 7.06 (s, 1H), 7.03 – 6.97 (m, 2H), 4.19 (dd, J = 10.3, 3.7 Hz, 1H), 3.33 – 3.11 (m, 2H), 2.98 – 2.69 (m, 4H), 2.33 (s, 3H), 2.01 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 139.2, 135.1, 132.1, 129.3, 129.2, 128.6, 128.6, 127.0, 126.7, 126.4, 57.2, 42.5, 40.7, 29.5, 21.2. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{N}$ ($[\text{M}+\text{H}]^+$): 238.1590, found: 238.1583.



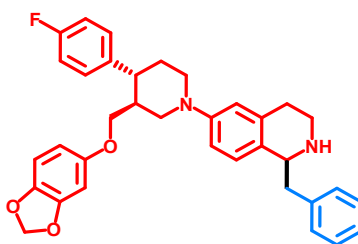
1-benzyl-7-chloro-1,2,3,4-tetrahydroisoquinoline (3bh) was prepared according to General Procedure (2.2), and the reaction was heated for 6 h. Yield, 67%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.32 (m, 2H), 7.28 – 7.25 (m, 2H), 7.24 – 7.21 (m, 2H), 7.15 – 7.11 (m, 1H), 7.05 (d, J = 8.1 Hz, 1H), 4.17 (dd, J = 10.2, 3.9 Hz, 1H), 3.27 – 3.17 (m, 2H), 2.93 – 2.69 (m, 4H), 1.77 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 140.3, 138.6, 133.7, 131.2, 130.7, 129.3, 128.7, 126.6, 126.3, 126.1, 57.1, 42.3, 40.5, 29.4. HRMS (APCI) m/z calculated for $\text{C}_{16}\text{H}_{17}\text{ClN}$ ($[\text{M}+\text{H}]^+$): 258.1044, found: 258.1051.



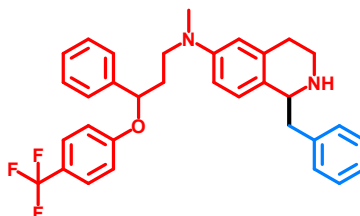
1-benzyl-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline (3bk) was prepared according to General Procedure (2.2), and the reaction was heated for 6 h. Yield, 90%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.31 (m, 2H), 7.29 – 7.22 (m, 3H), 6.59 (s, 2H), 4.18 (dd, J = 9.3, 4.7 Hz, 1H), 3.86 (s, 3H), 3.79 (s, 3H), 3.27 – 3.17 (m, 2H), 2.99 – 2.90 (m, 2H), 2.84 – 2.66 (m, 2H), 1.81 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 147.7, 147.0, 138.5, 129.5, 128.9, 128.6, 126.6, 126.5, 111.7, 109.5, 56.6, 55.8, 42.4, 40.2, 28.6. HRMS (ESI) m/z calculated for $\text{C}_{18}\text{H}_{22}\text{NO}_2$ ($[\text{M}+\text{H}]^+$): 284.1645, found: 284.1648.



1-benzyl-4-methyl-1,2,3,4-tetrahydroisoquinoline (3bl) was prepared according to General Procedure (2.2), and the reaction was heated for 6 h. Yield, 73%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.34 – 7.30 (m, 2H), 7.29 – 7.22 (m, 5H), 7.19 (ddd, J = 8.7, 4.4, 2.3 Hz, 2H), 4.23 (dd, J = 9.4, 4.4 Hz, 1H), 3.32 – 3.16 (m, 1H), 3.06 – 2.96 (m, 1H), 2.97 – 2.82 (m, 2H), 2.67 – 2.55 (m, 1H), 1.79 (s, 1H), 1.29 – 1.21 (m, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 140.7, 139.0, 138.3, 129.4, 128.5, 128.2, 126.4, 126.2, 126.0, 125.6, 57.4, 47.4, 42.5, 32.9, 20.5. HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{20}\text{N}$ ($[\text{M}+\text{H}]^+$): 238.1583, found: 238.1590.



6-((3S,4R)-3-((benzo[d][1,3]dioxol-5-yloxy)methyl)-4-(4-fluorophenyl)piperidin-1-yl)-1-benzyl-1,2,3,4-tetrahydroisoquinoline (Paroxetine derivative) 3bm was prepared according to General Procedure (2.2), and the reaction was heated for 8 h, and 3 equiv of benzaldehyde hydrazone was added. Yield, 64%, reddish oil, dr value is 1:1 which is calculated by LCMS. ^1H NMR (500 MHz, CDCl_3) δ 7.33 (t, $J = 7.5$ Hz, 2H), 7.30 – 7.26 (m, 3H), 7.23 – 7.14 (m, 3H), 7.00 (t, $J = 8.5$ Hz, 2H), 6.88 (d, $J = 9.5$ Hz, 1H), 6.74 (s, 1H), 6.64 (d, $J = 8.4$ Hz, 1H), 6.38 (d, $J = 2.5$ Hz, 1H), 6.17 (dd, $J = 8.6, 2.5$ Hz, 1H), 5.89 (s, 2H), 4.17 (dd, $J = 10.2, 3.7$ Hz, 1H), 4.00 (dd, $J = 12.3$ Hz, 1H), 3.79 (d, $J = 12.4$ Hz, 1H), 3.70 – 3.62 (m, 1H), 3.57 – 3.50 (m, 1H), 3.32 – 3.17 (m, 2H), 2.97 – 2.81 (m, 4H), 2.81 – 2.72 (m, 1H), 2.61 (dt, $J = 11.6, 4.3$ Hz, 1H), 2.37 – 2.27 (m, 1H), 2.06 – 1.90 (m, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 161.6(d, $J = 244.4$ Hz), 154.3, 149.8, 148.2, 141.6, 139.4(d, $J = 3.10$ Hz), 139.2, 135.9, 129.4, 128.8(d, $J = 7.6$ Hz), 128.8, 128.6, 126.8, 126.4, 115.5 (d, $J = 21.0$ Hz), 115.1, 115.0, 107.8, 105.6, 101.1, 98.0, 69.4, 56.8, 53.8, 50.8, 44.1, 42.5, 41.8, 41.0, 34.1, 30.3. ^{19}F NMR (471 MHz, CDCl_3) δ -116.28. HRMS (ESI) m/z calculated for $\text{C}_{35}\text{H}_{36}\text{FN}_2\text{O}_3$ ($[\text{M}+\text{H}]^+$): 551.2704, found: 551.2716.



1-benzyl-N-methyl-N-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-1,2,3,4-tetrahydroisoquinolin-6-amine (Fluoxetine derivative) 3bn was prepared according to General Procedure (2.2), and the reaction was heated for 8 h, and 3 equiv of benzaldehyde hydrazone was added. Yield, 67%, reddish oil. ^1H NMR (500 MHz, CDCl_3) δ 7.44 (d, $J = 8.5$ Hz, 2H), 7.33 (d, $J = 6.7$ Hz, 6H), 7.27 (d, $J = 6.8$ Hz, 2H), 7.25 – 7.22 (m, 2H), 7.05 (dd, $J = 8.7, 4.4$ Hz, 1H), 6.90 (d, $J = 8.5$ Hz, 2H), 6.56 (dd, $J = 8.6, 2.7$ Hz, 1H), 6.37 (s, 1H), 5.22 (dd, $J = 8.5, 4.2$ Hz, 1H), 4.15 – 4.09 (m, 1H), 3.65 – 3.56 (m, 1H), 3.51 – 3.38 (m, 1H), 3.23 (dt, $J = 13.6, 3.7$ Hz, 1H), 3.19 – 3.09 (m, 1H), 3.05 – 2.95 (m, 1H), 2.90 (s, 3H), 2.89 – 2.69 (m, 2H), 2.64 (t, $J = 5.9$ Hz, 1H), 2.22 – 2.11 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 160.4, 147.7, 140.8, 139.1, 135.8, 129.4, 128.9, 128.8, 128.5, 127.9, 126.9, 126.8 (q, $J = 2.95$ Hz), 126.4, 125.7, 125.6 (q, $J = 270.1$ Hz), 123.0 (q, $J = 32.62$ Hz), 115.7, 112.5, 110.5, 77.8, 56.7, 49.1, 42.5, 41.0, 38.3, 36.2, 30.0. ^{19}F NMR (471 MHz, CDCl_3) δ -61.61. HRMS (ESI) m/z calculated for $\text{C}_{33}\text{H}_{34}\text{F}_3\text{N}_2\text{O}$ ($[\text{M}+\text{H}]^+$): 531.2618, found: 531.2611.

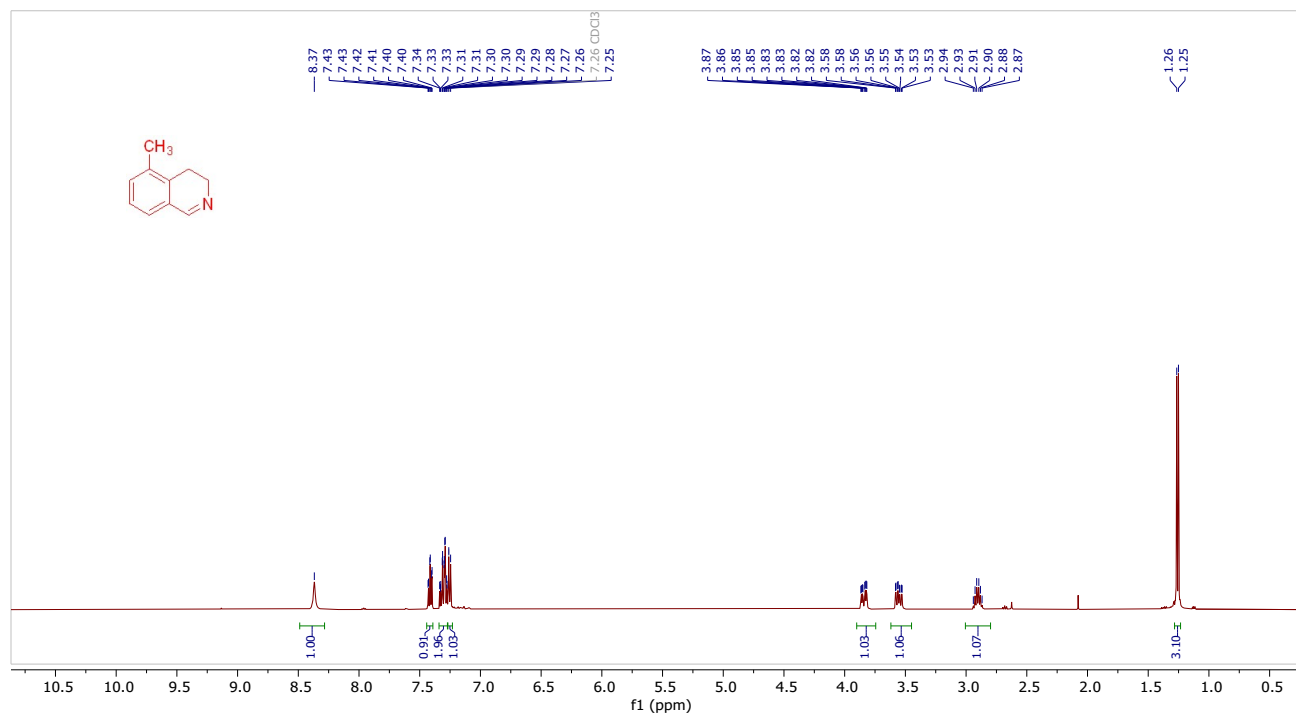
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2. Sangster, J. J.; Ruscoe, R. E.; Cosgrove, S. C.; Mangas-Sanchez, J.; Turner, N. J., *J. Am. Chem. Soc.* **2023**, *145* (8), 4431–4437.

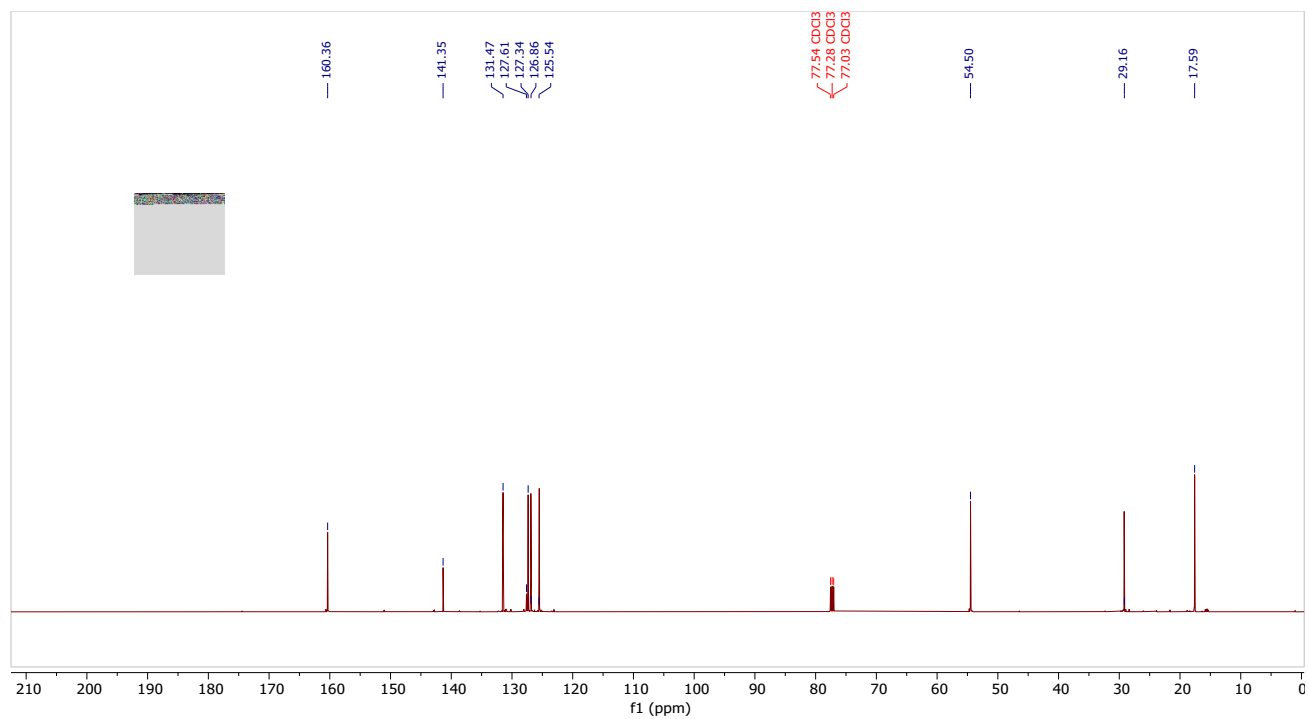
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NMR spectra collection for compounds

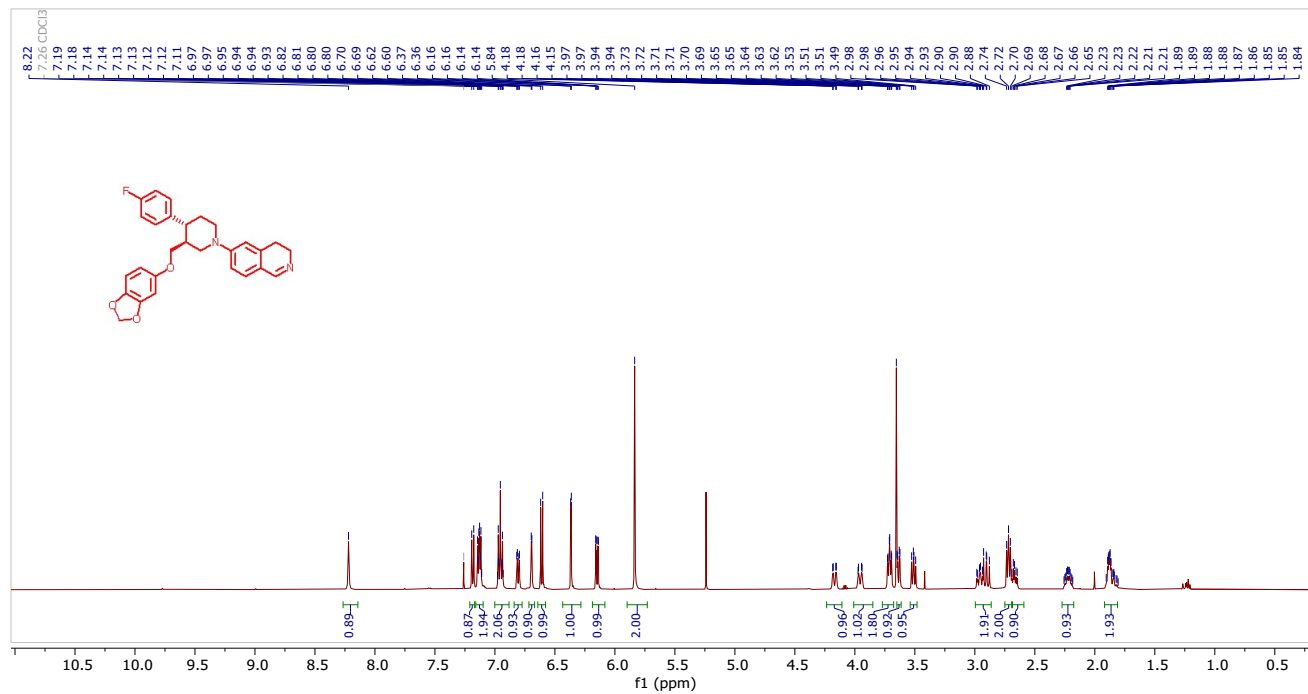
^1H NMR (500 MHz, CDCl_3) spectrum of **2d**



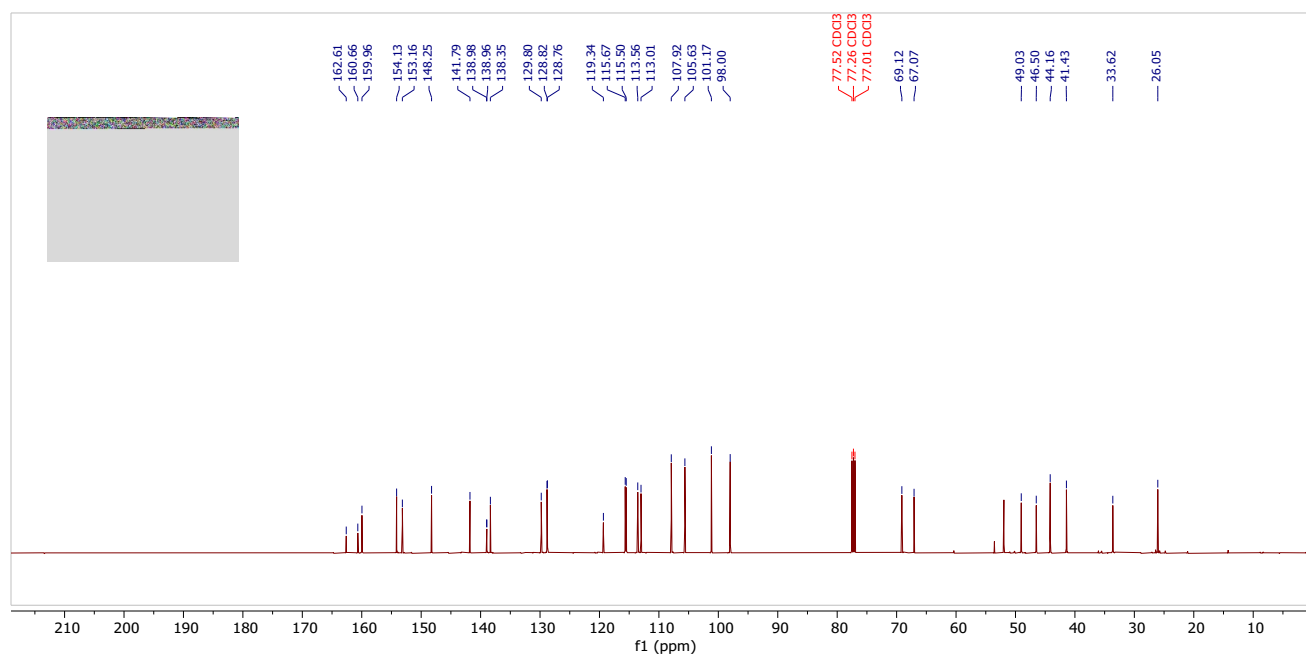
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **2d**



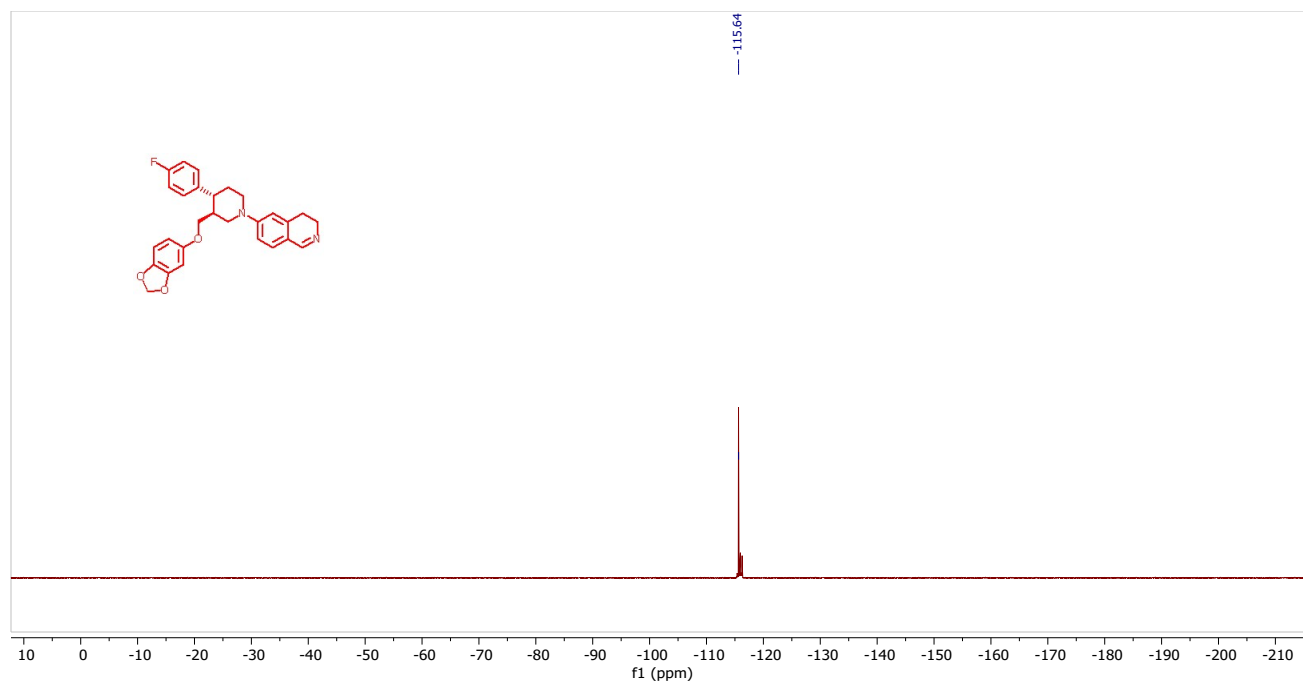
¹H NMR (500 MHz, CDCl₃) spectrum of **2e**



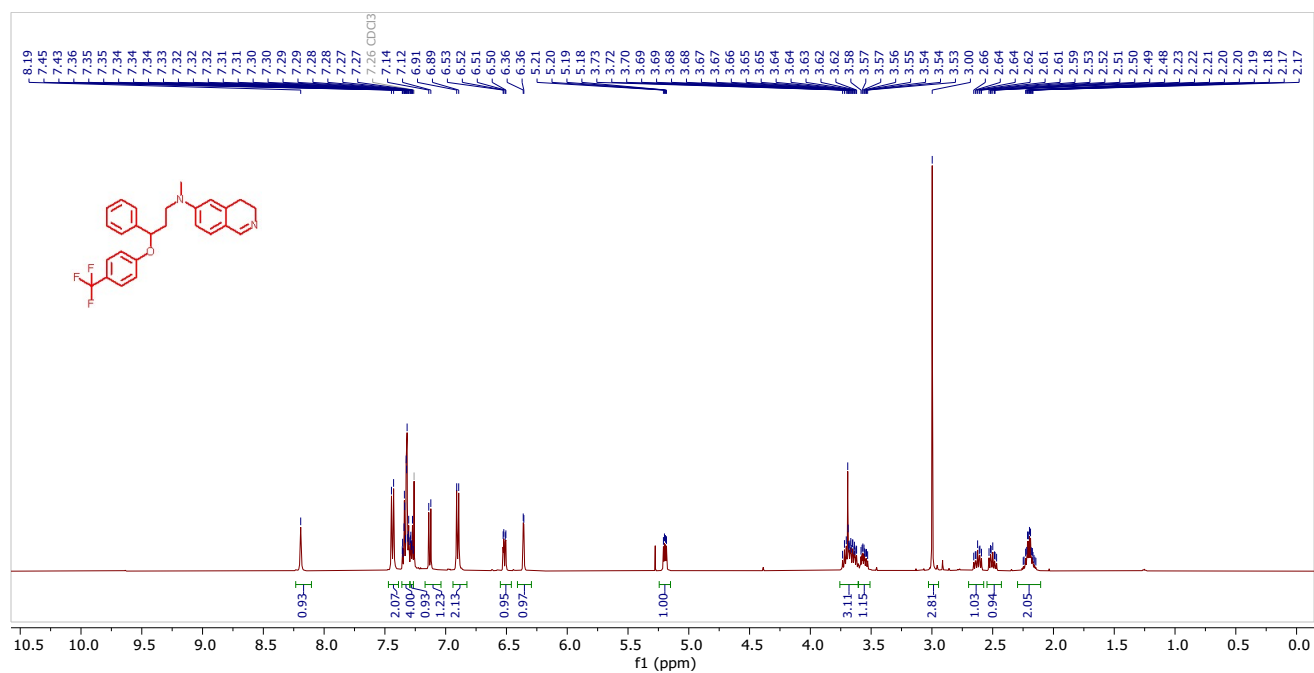
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **2e**



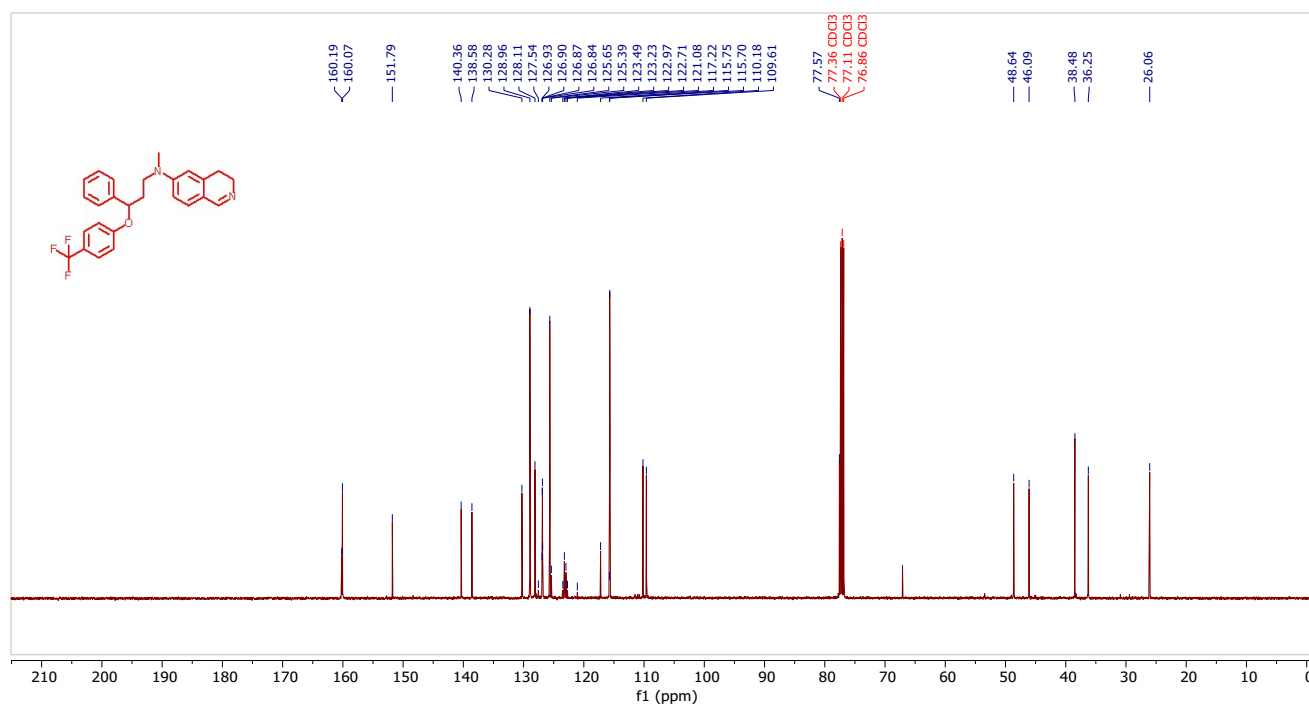
^{19}F NMR (471 MHz, CDCl_3) spectrum of **2e**



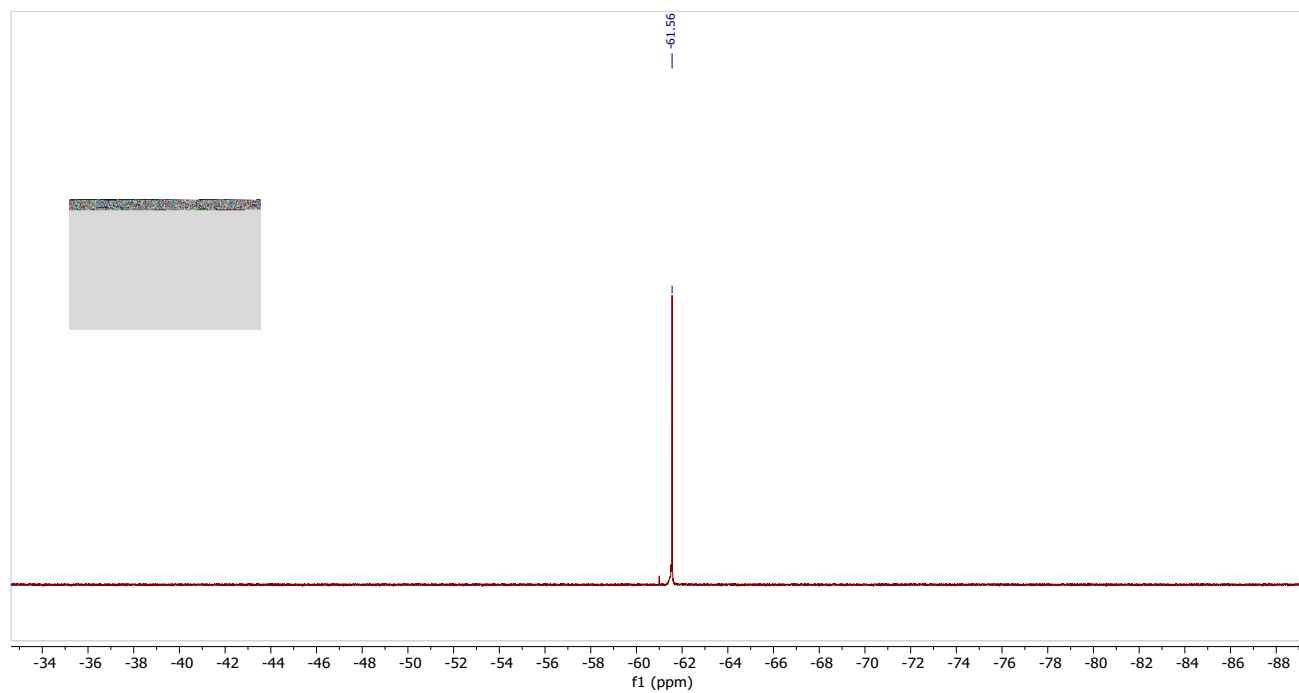
¹H NMR (500 MHz, CDCl₃) spectrum of **2f**



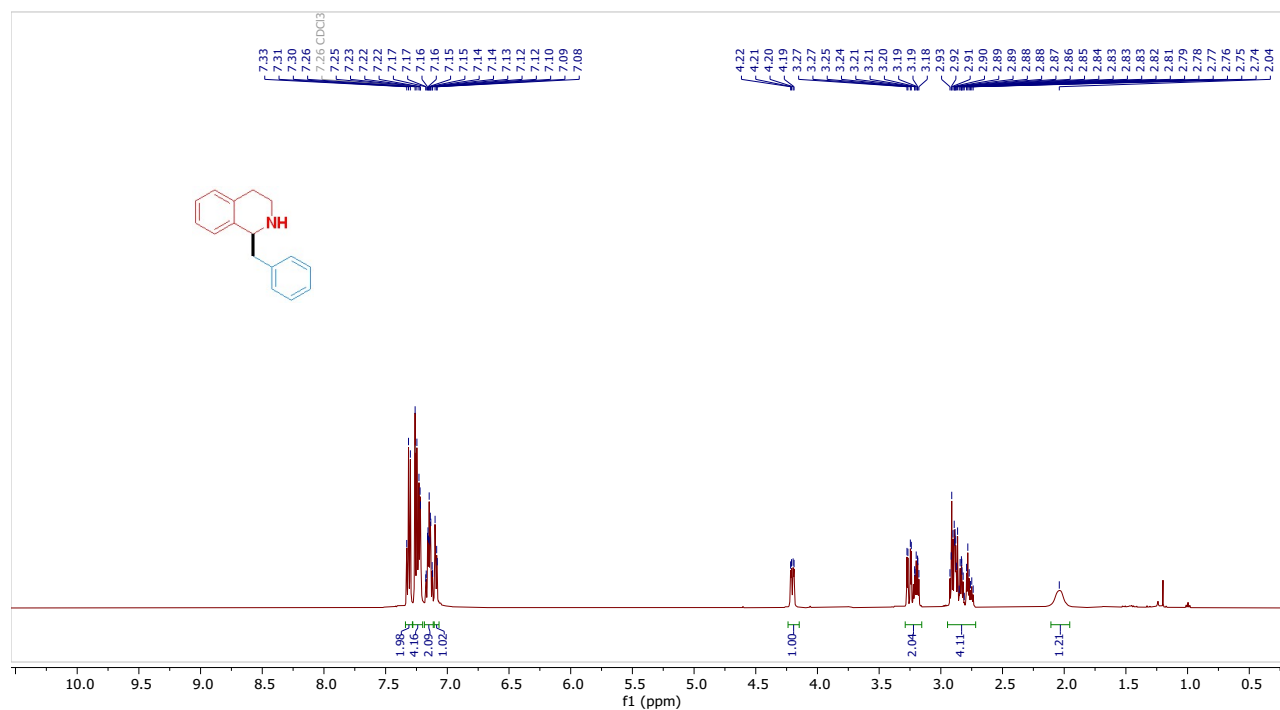
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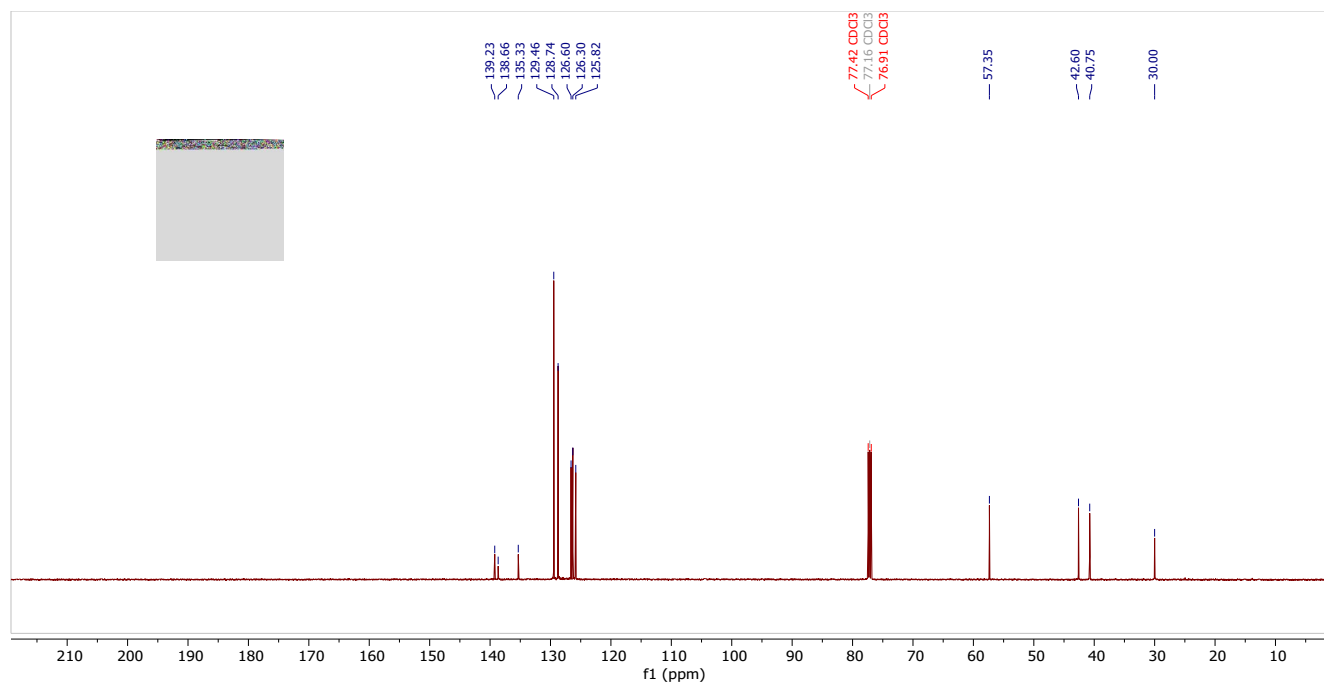
^{19}F NMR (471 MHz, CDCl_3) spectrum of **2f**



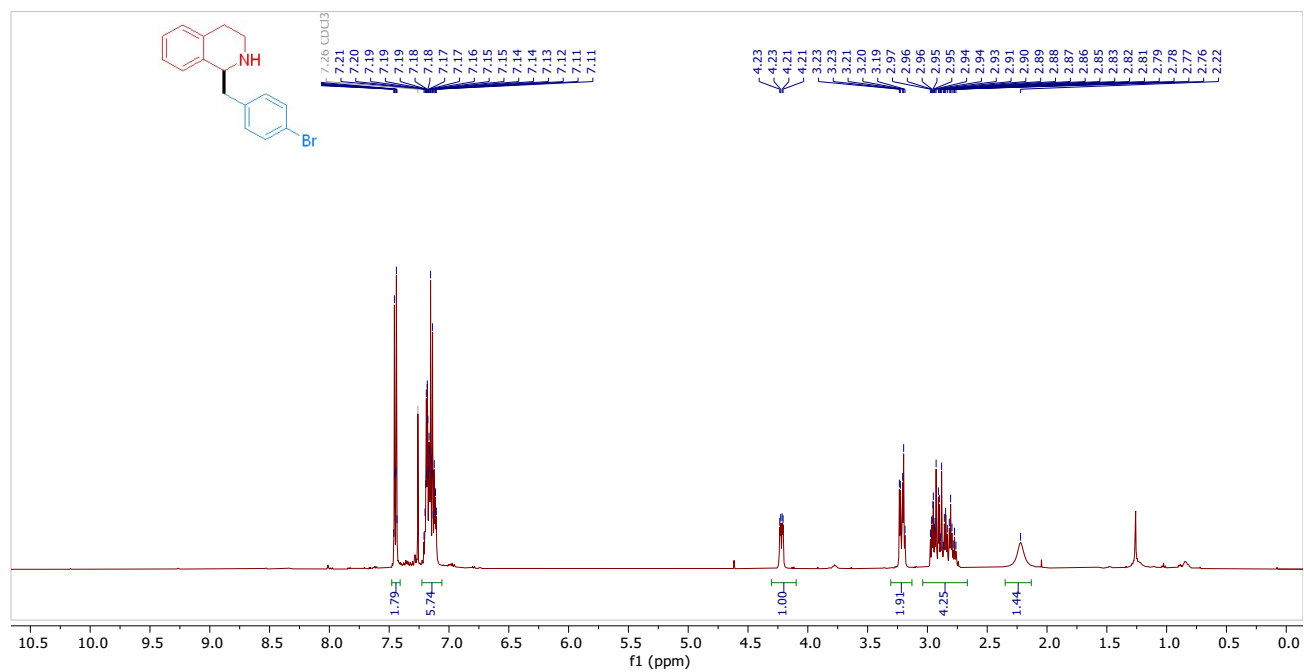
^1H NMR (500 MHz, CDCl_3) spectrum of **3aa**



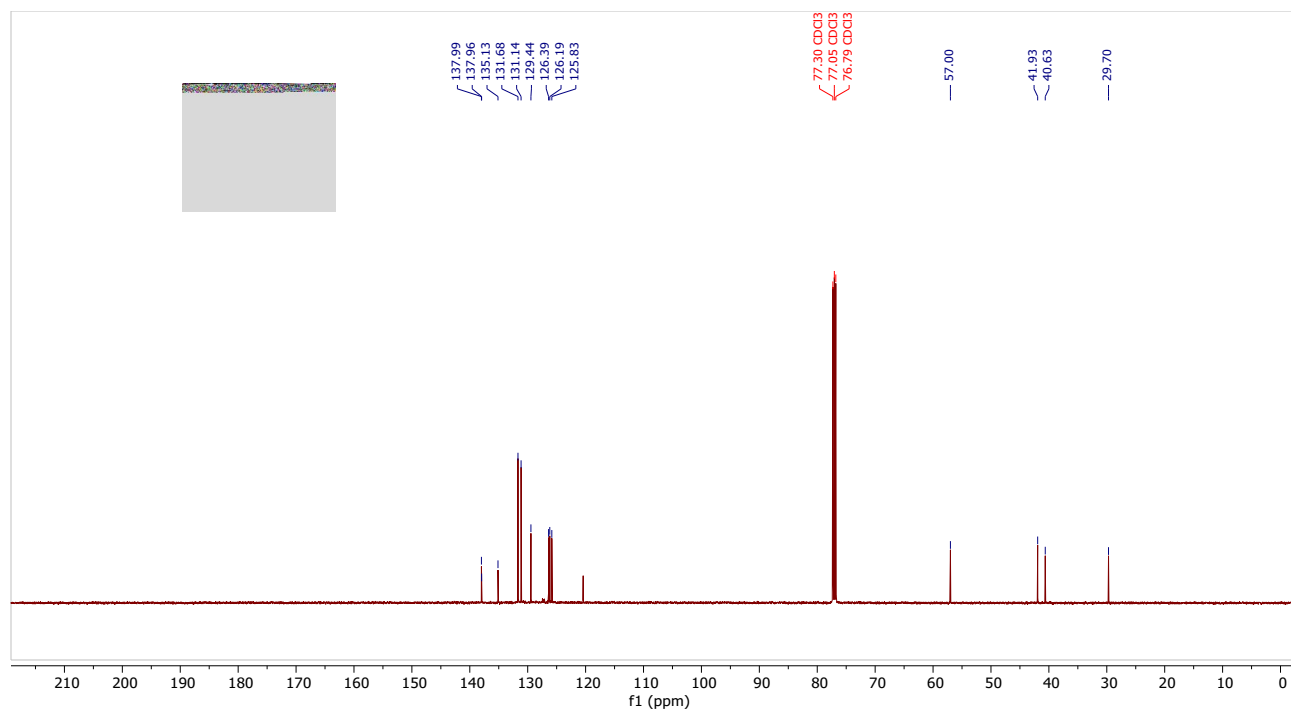
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3aa**



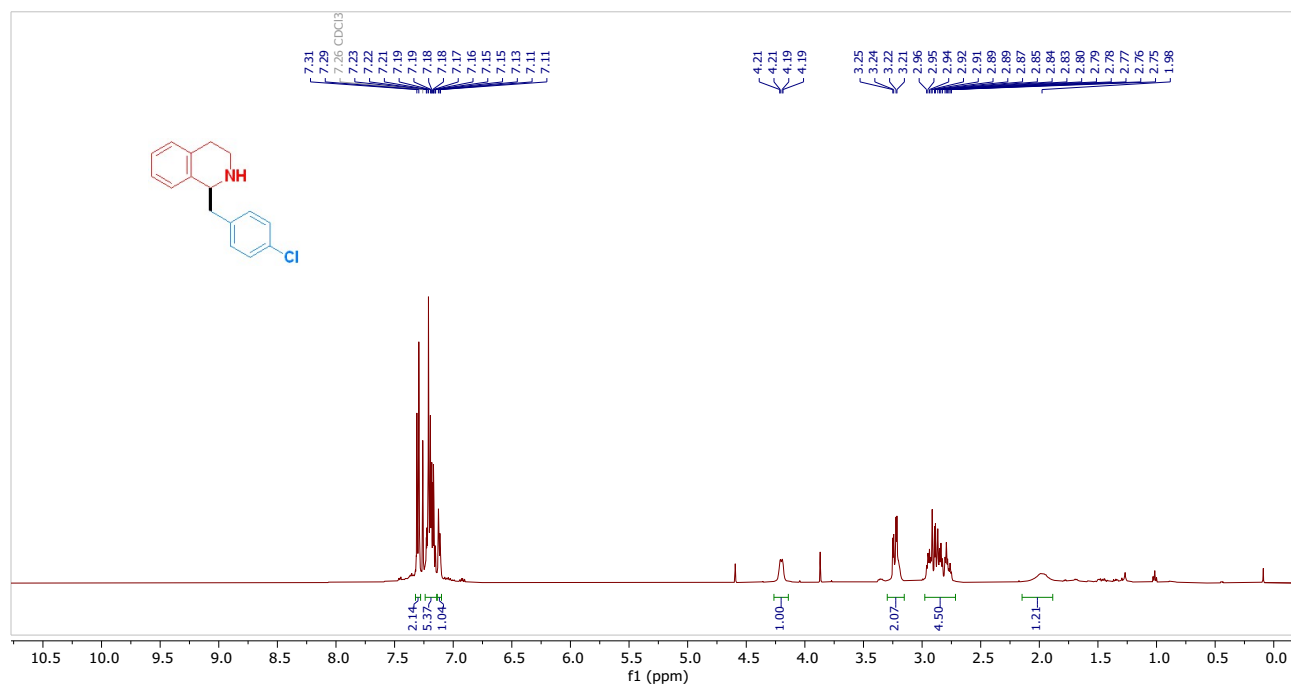
¹H NMR (500 MHz, CDCl₃) spectrum of **3ab**



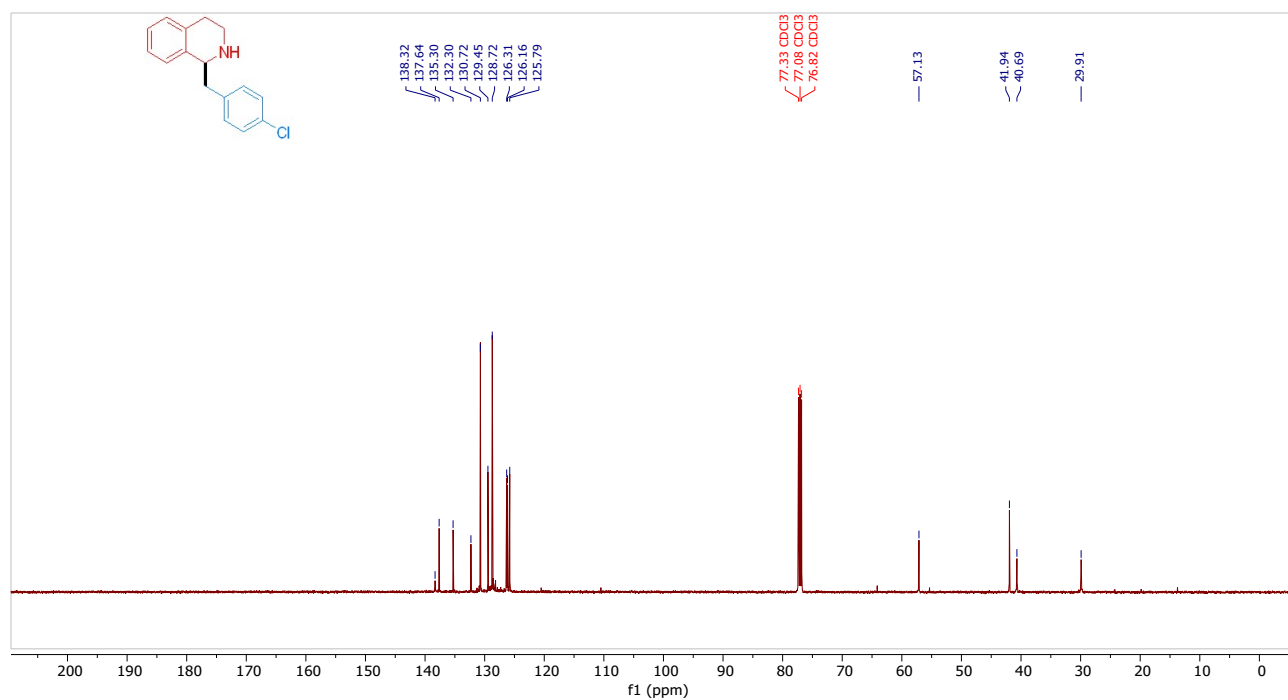
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of **3ab**



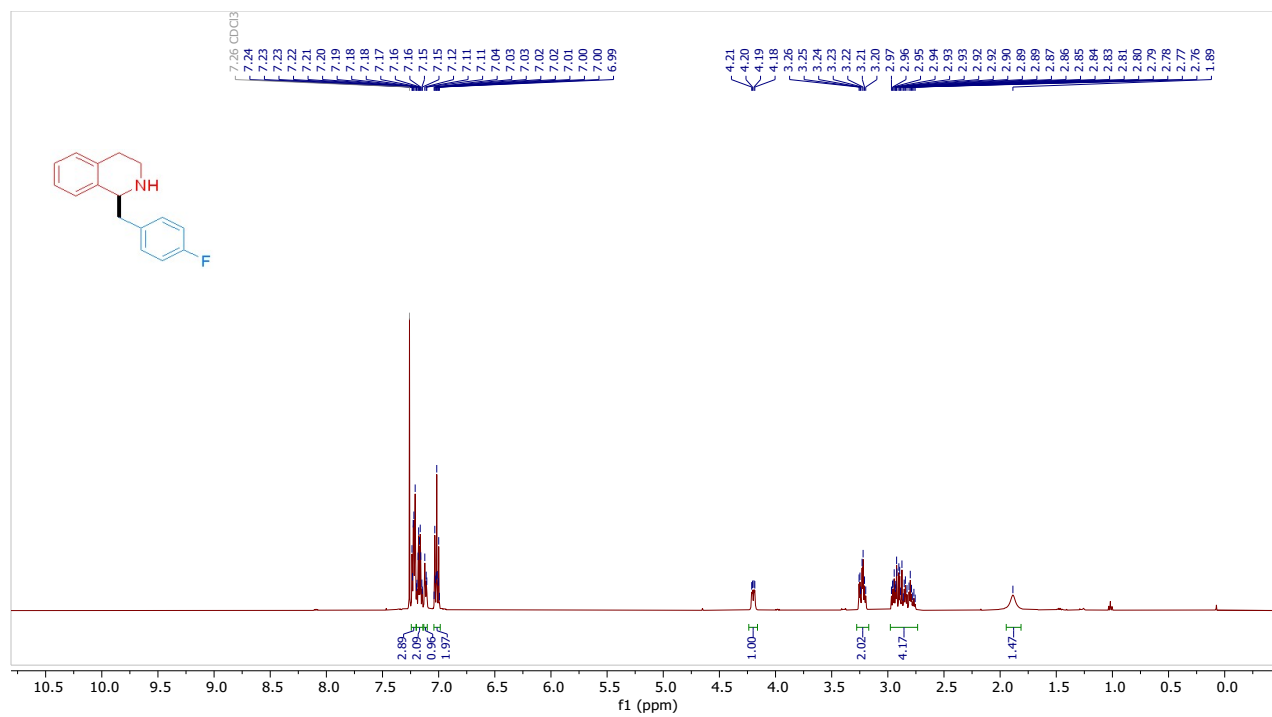
¹H NMR (500 MHz, CDCl₃) spectrum of **3ac**



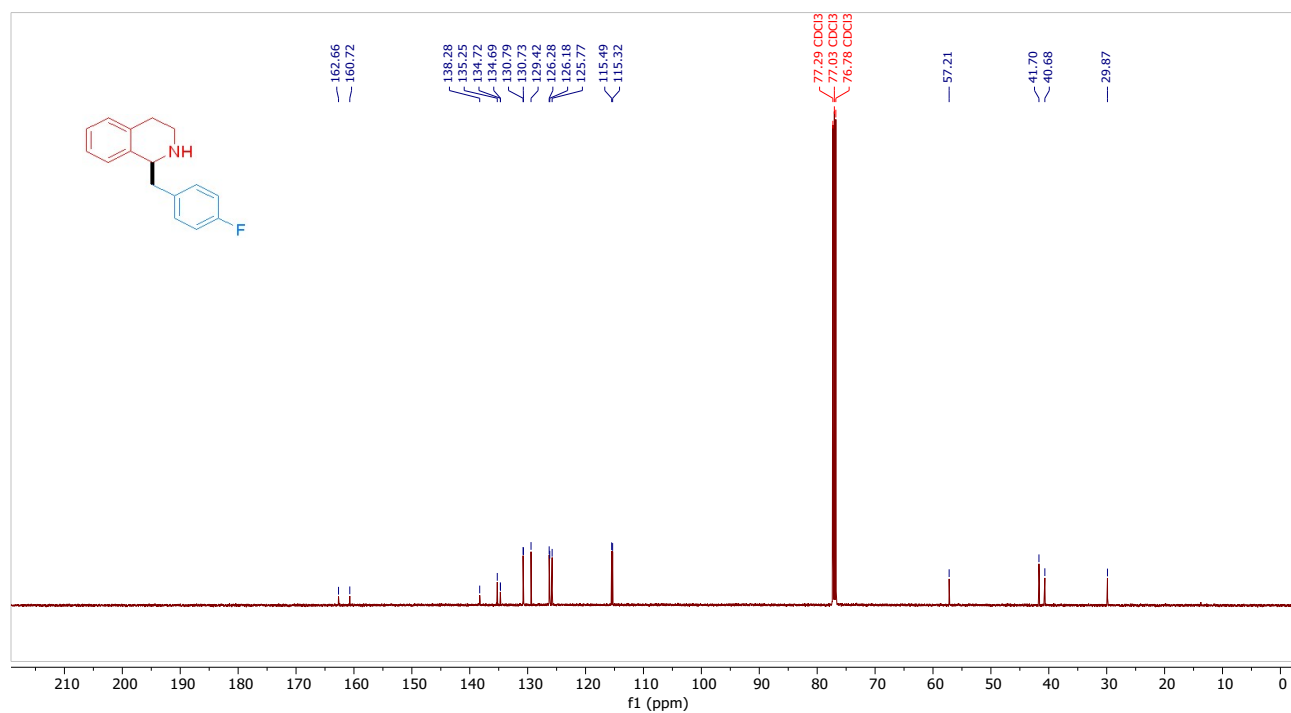
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ac**



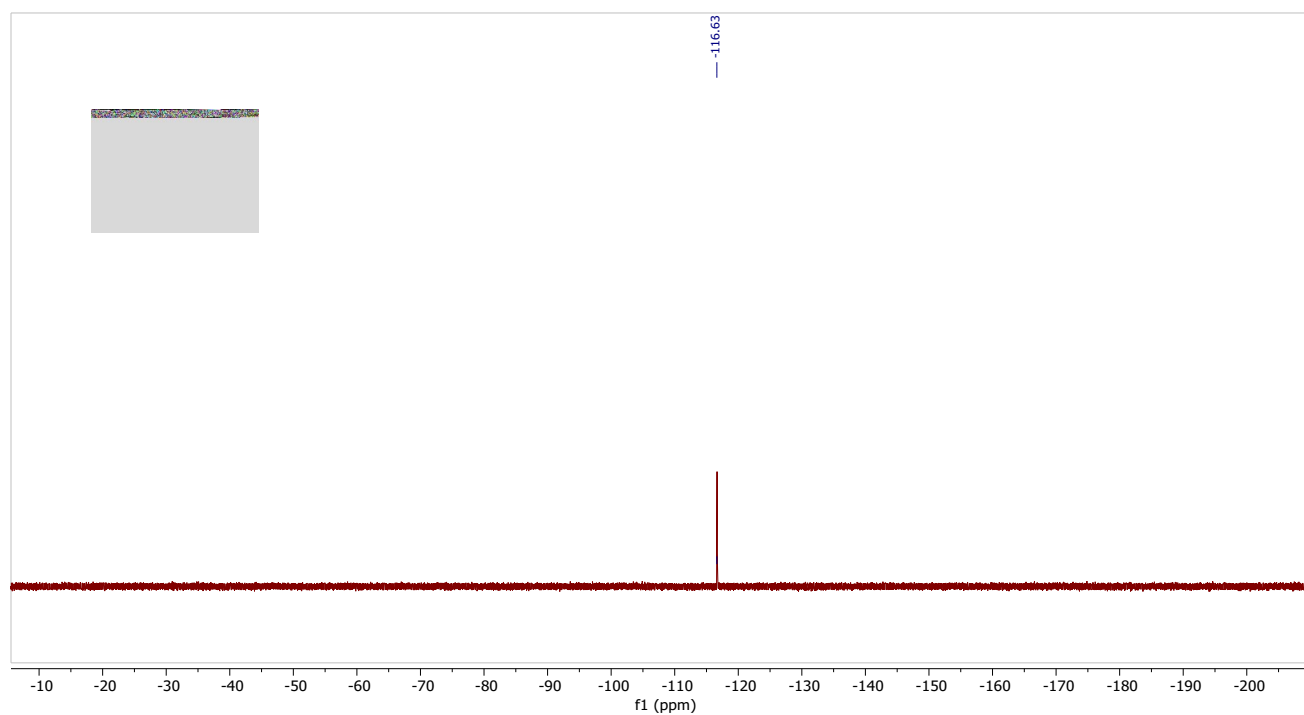
^1H NMR (500 MHz, CDCl_3) spectrum of **3ad**



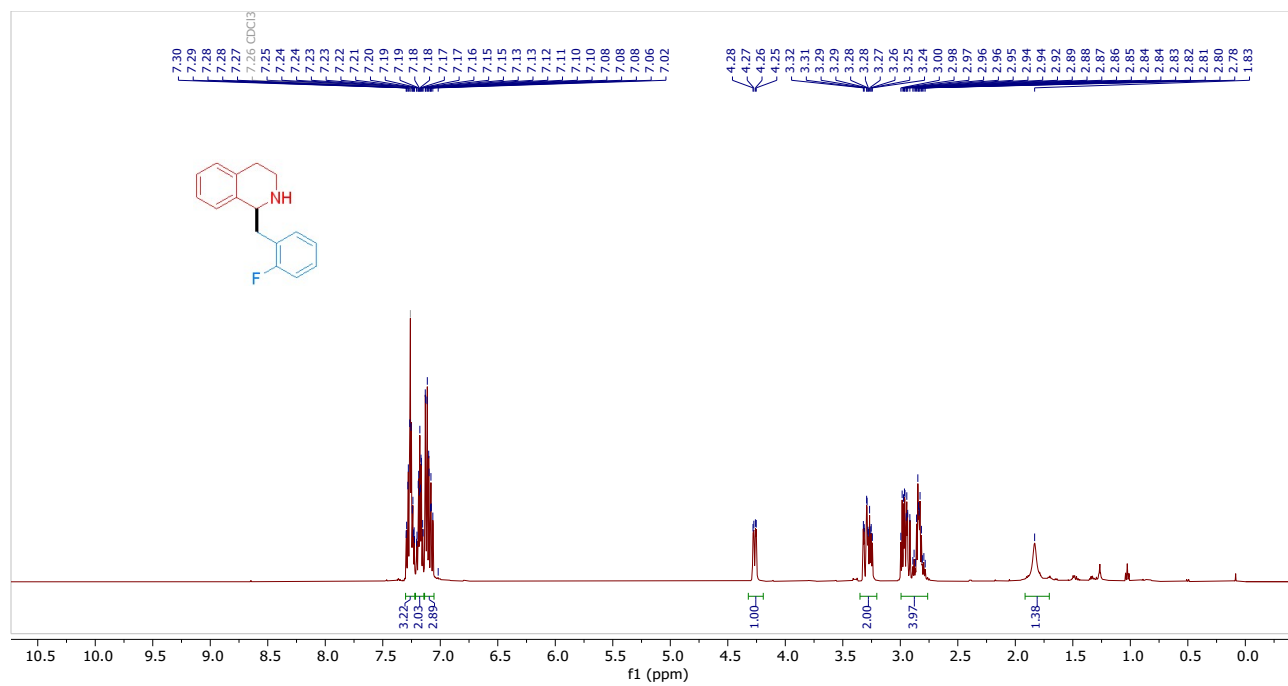
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of **3ad**



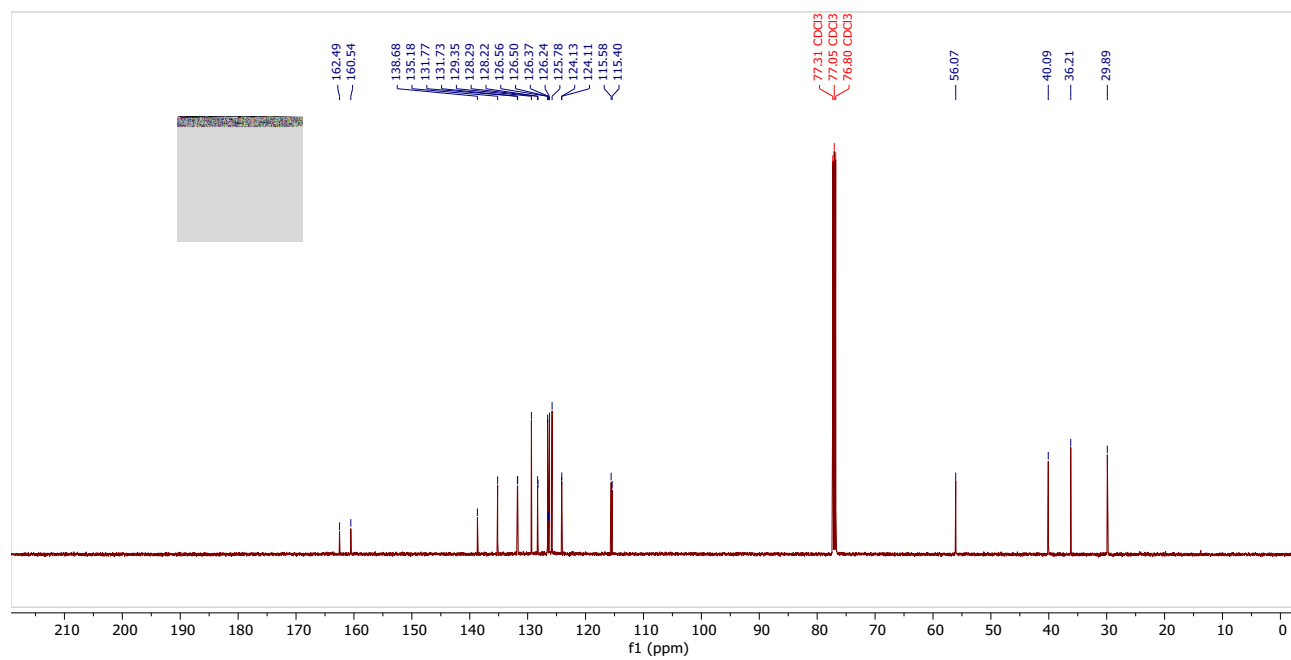
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3ad**



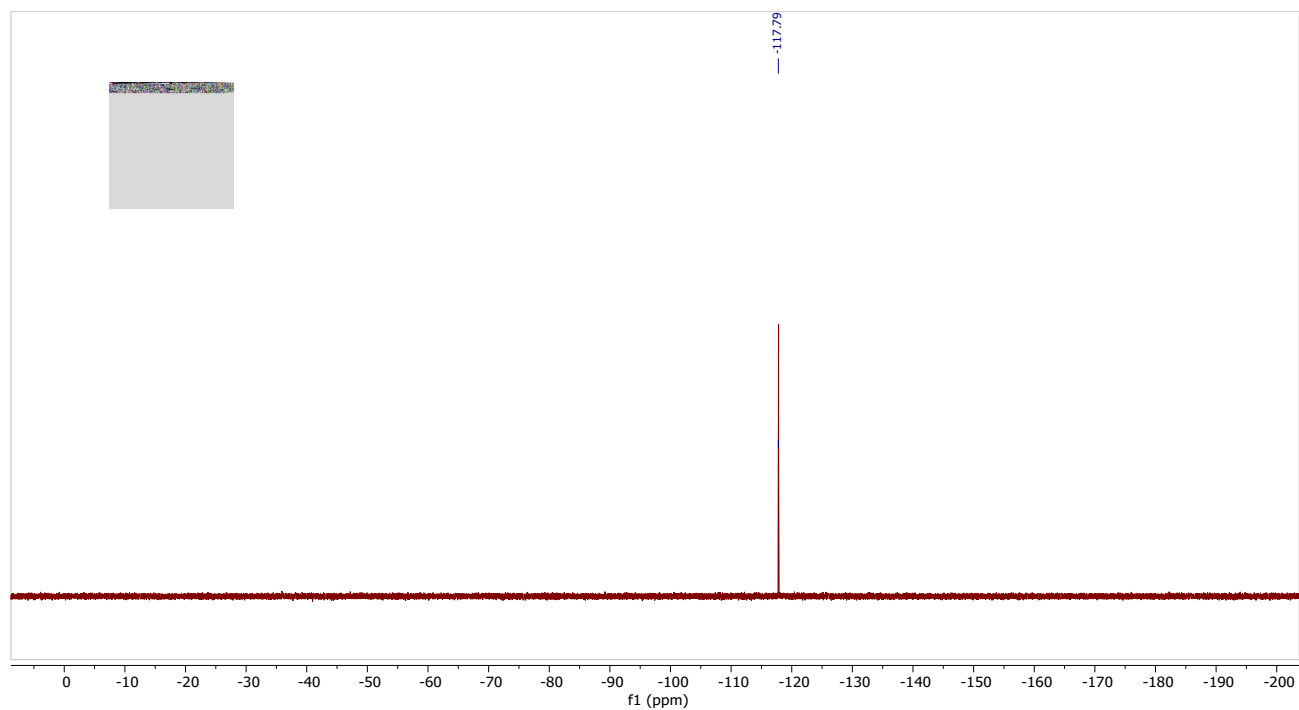
^1H NMR (500 MHz, CDCl_3) spectrum of **3ae**



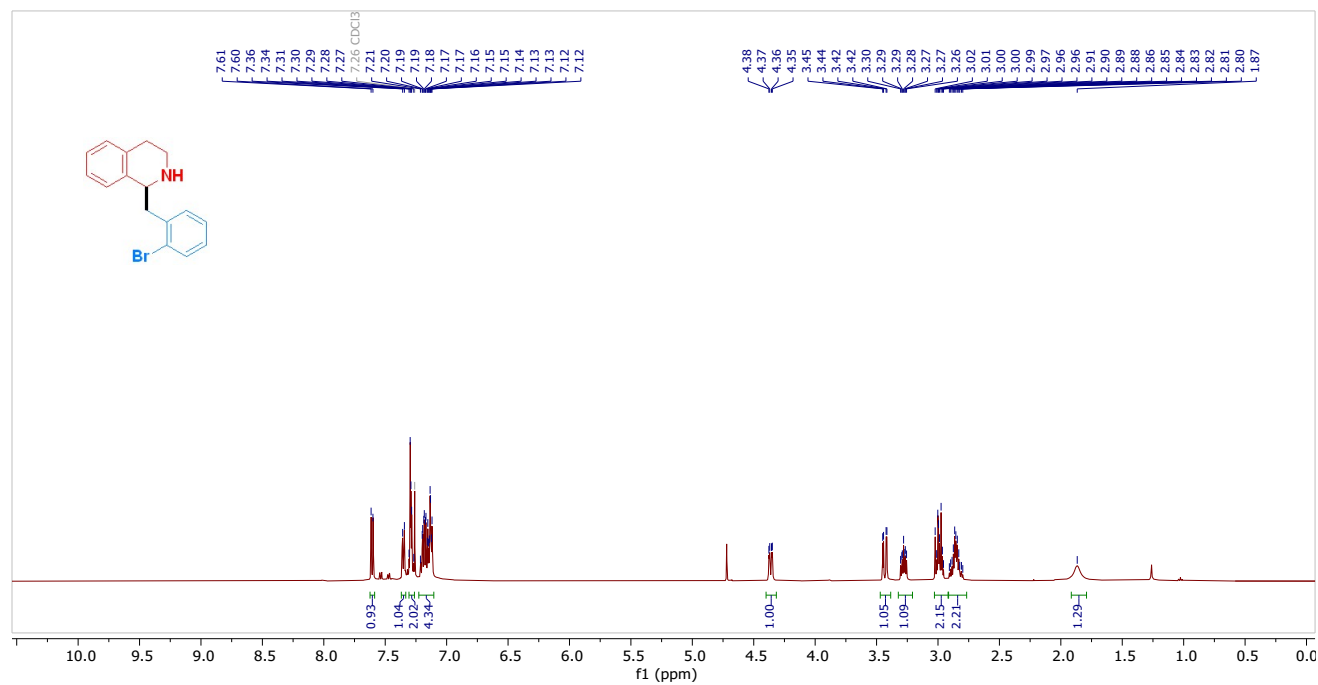
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ae**



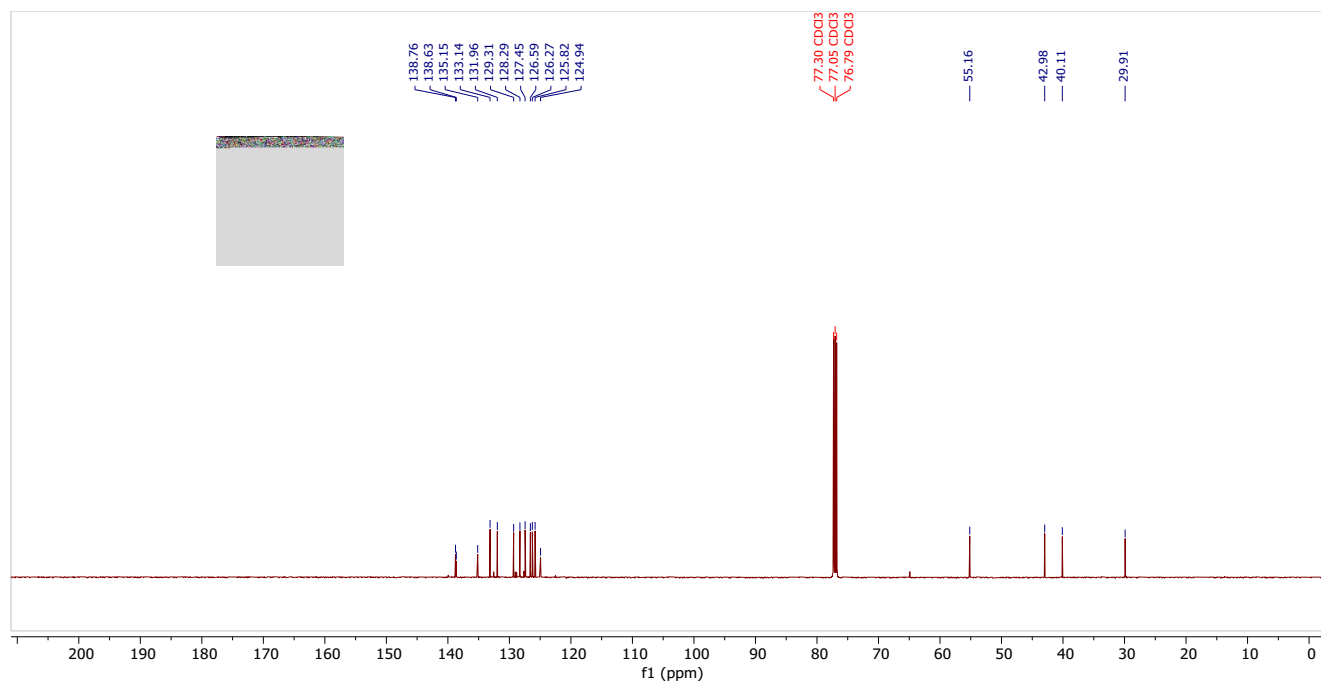
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3ae**



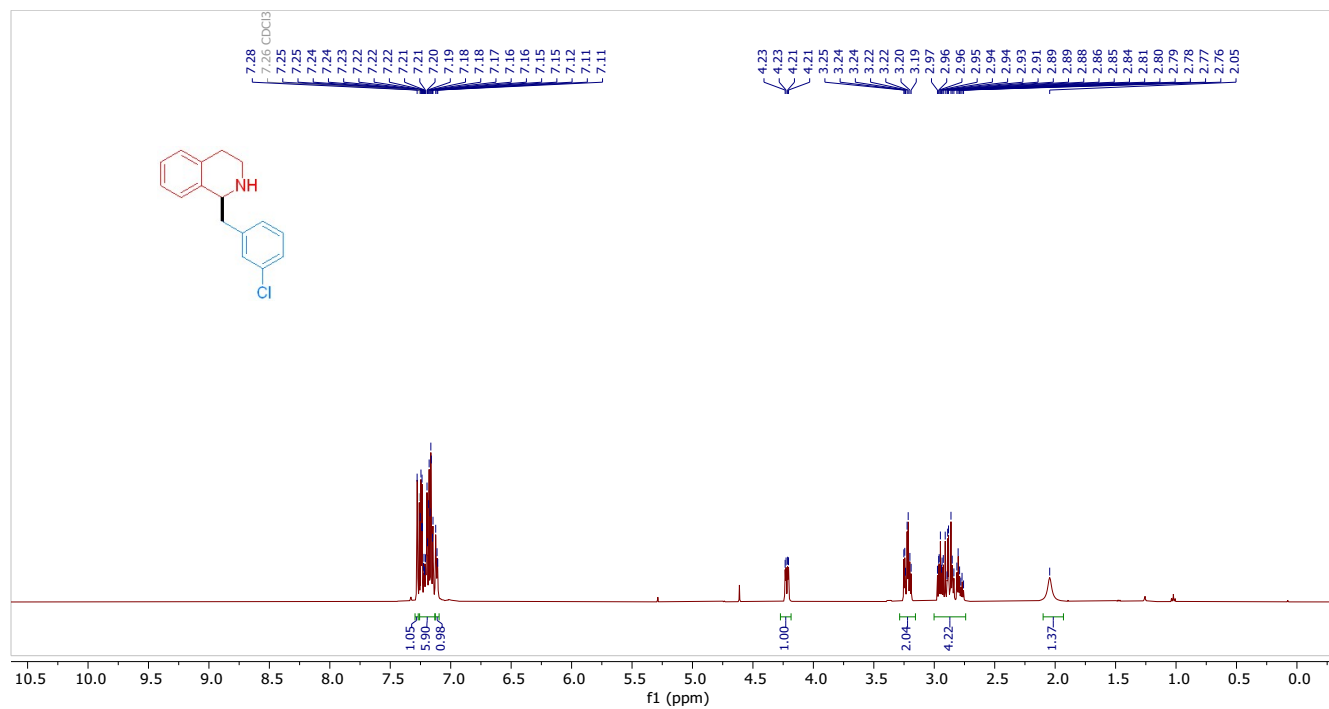
^1H NMR (500 MHz, CDCl_3) spectrum of **3af**



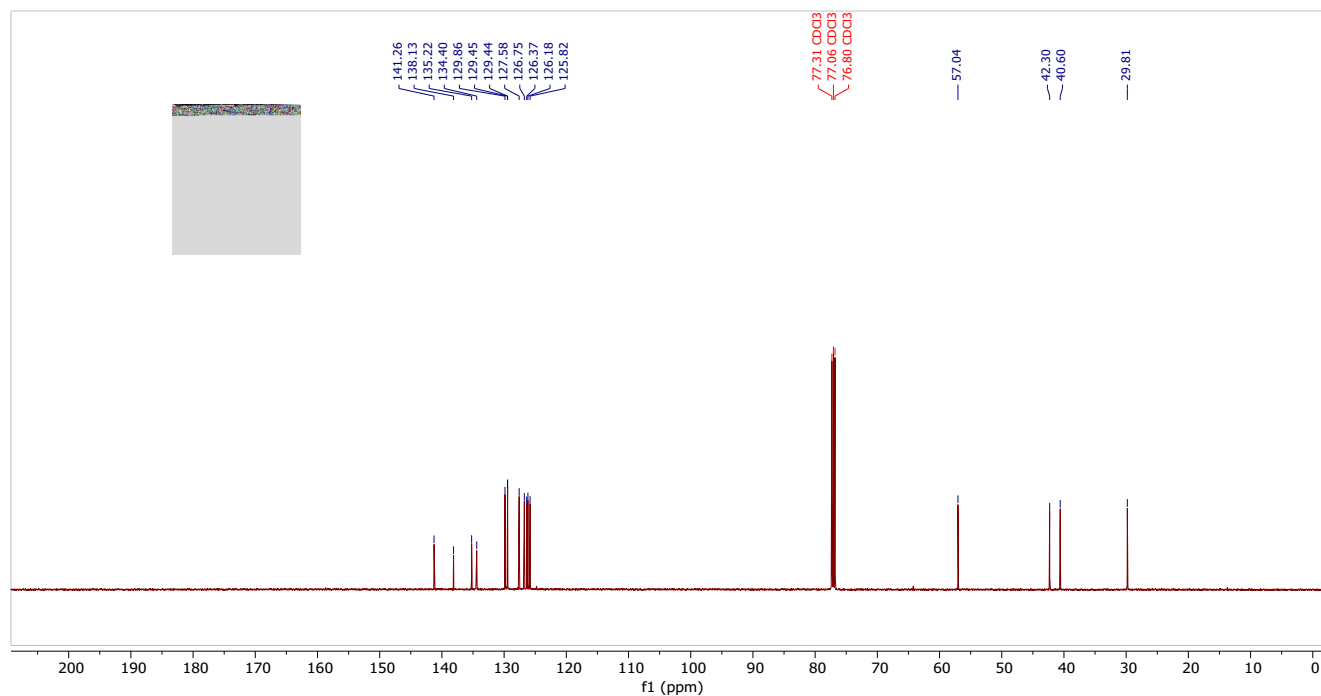
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3af**



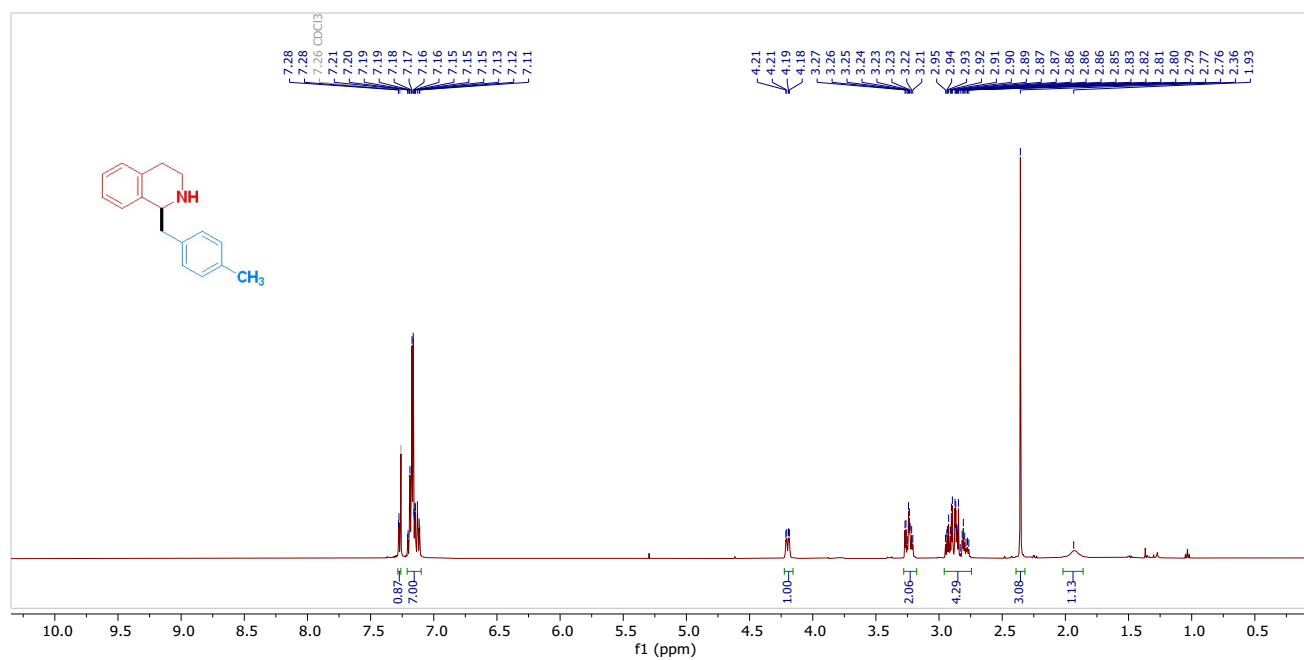
¹H NMR (500 MHz, CDCl₃) spectrum of **3ag**



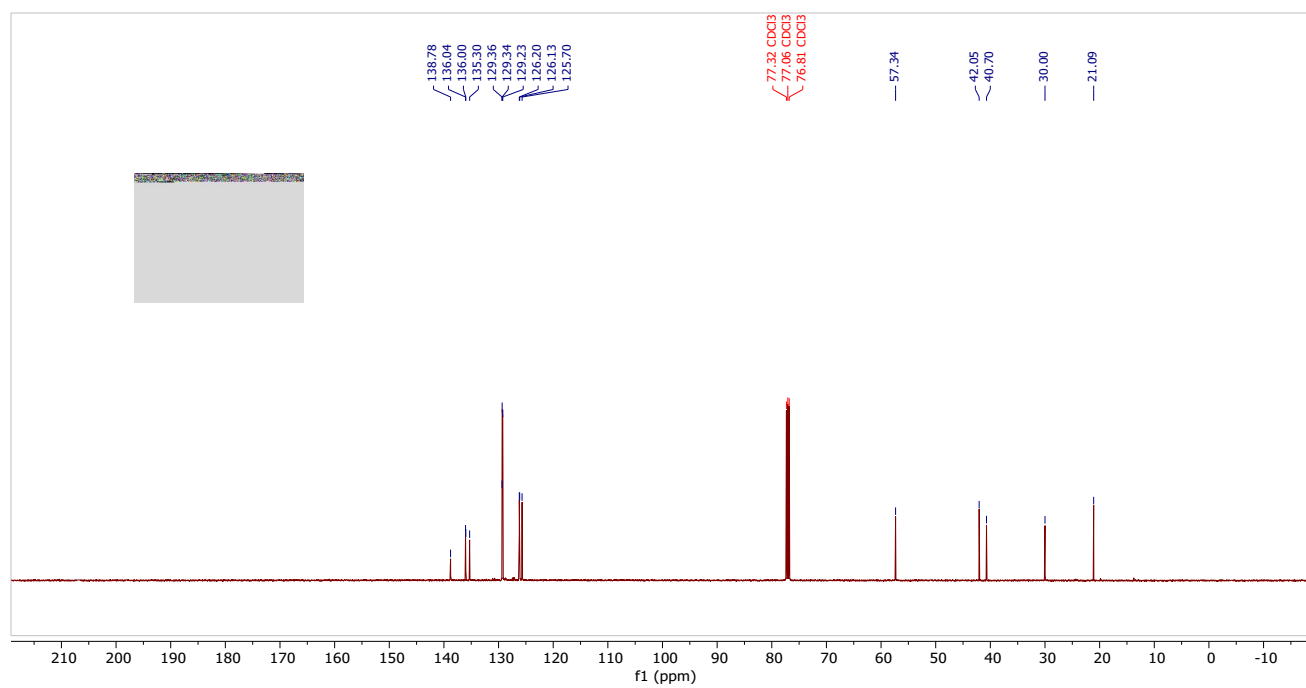
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ag**



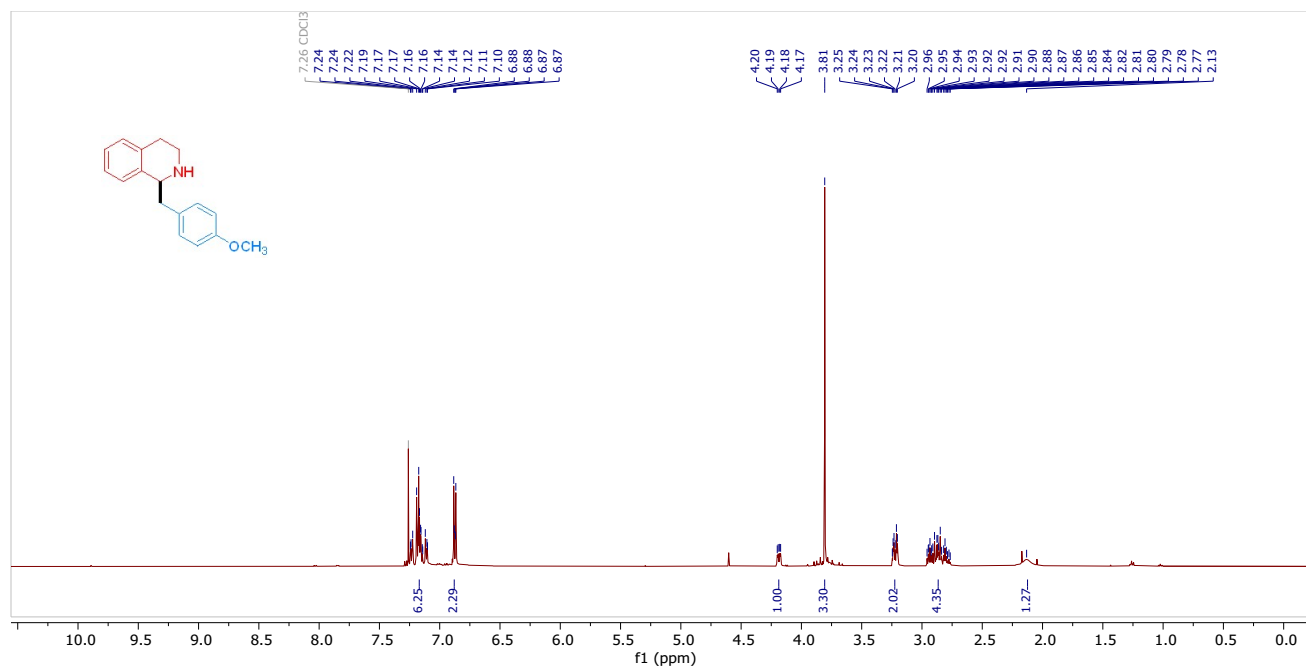
^1H NMR (500 MHz, CDCl_3) spectrum of **3ah**



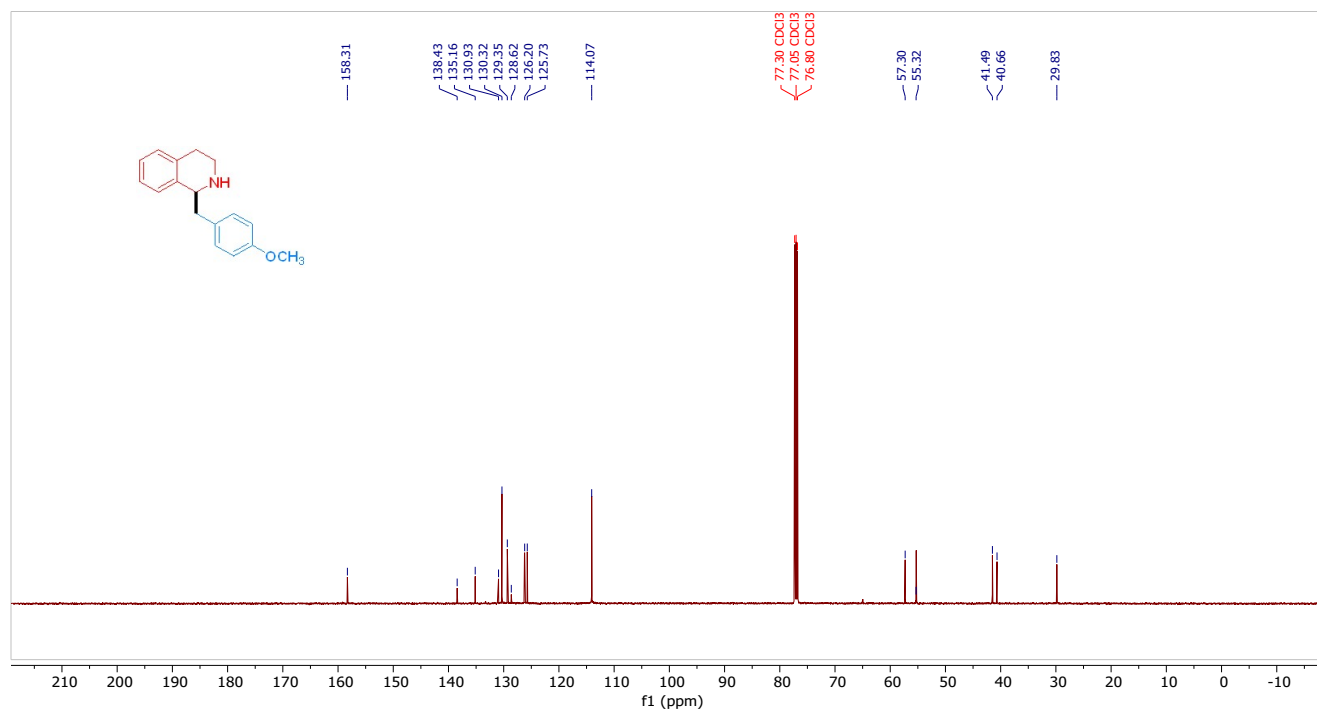
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ah**



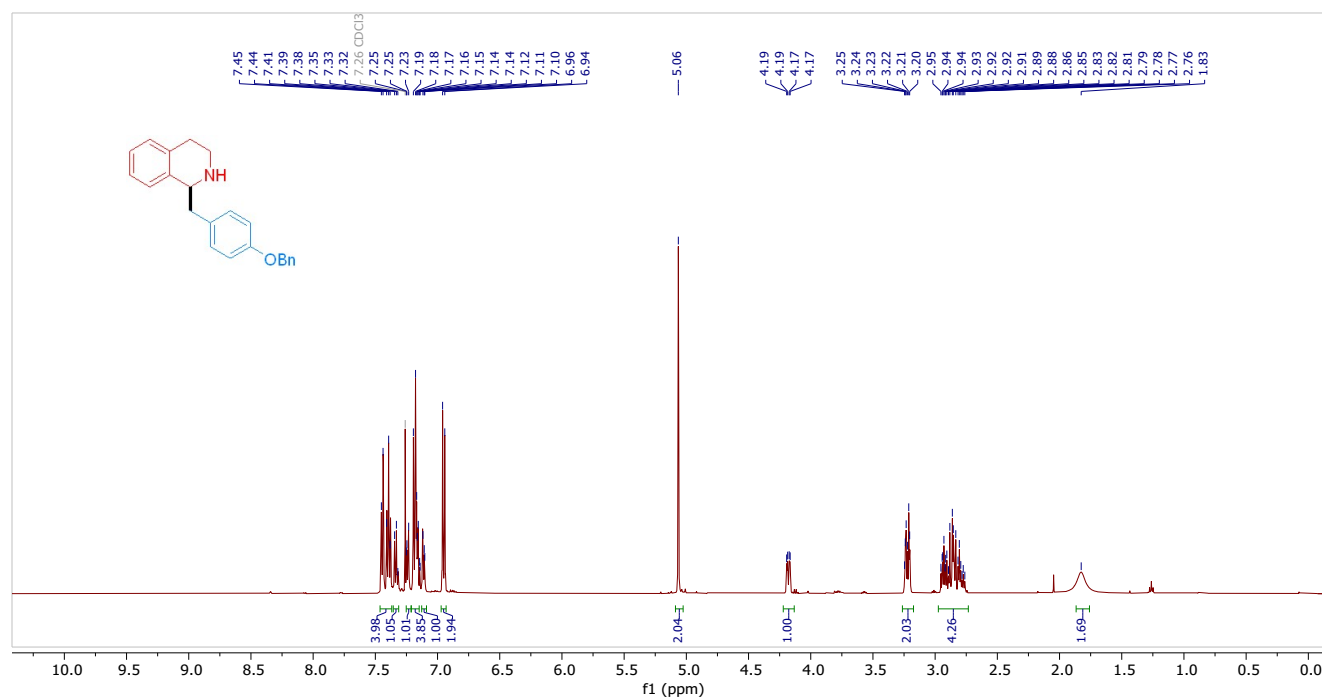
^1H NMR (500 MHz, CDCl_3) spectrum of **3ai**



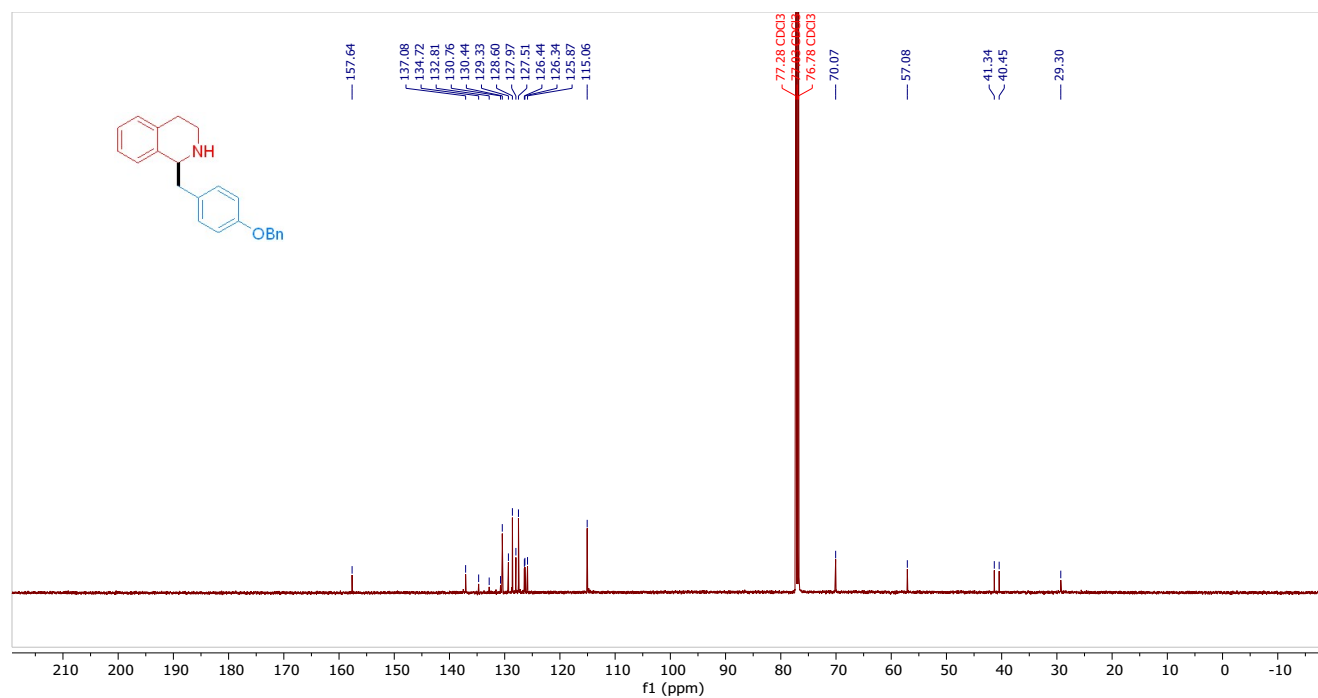
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of 3ai



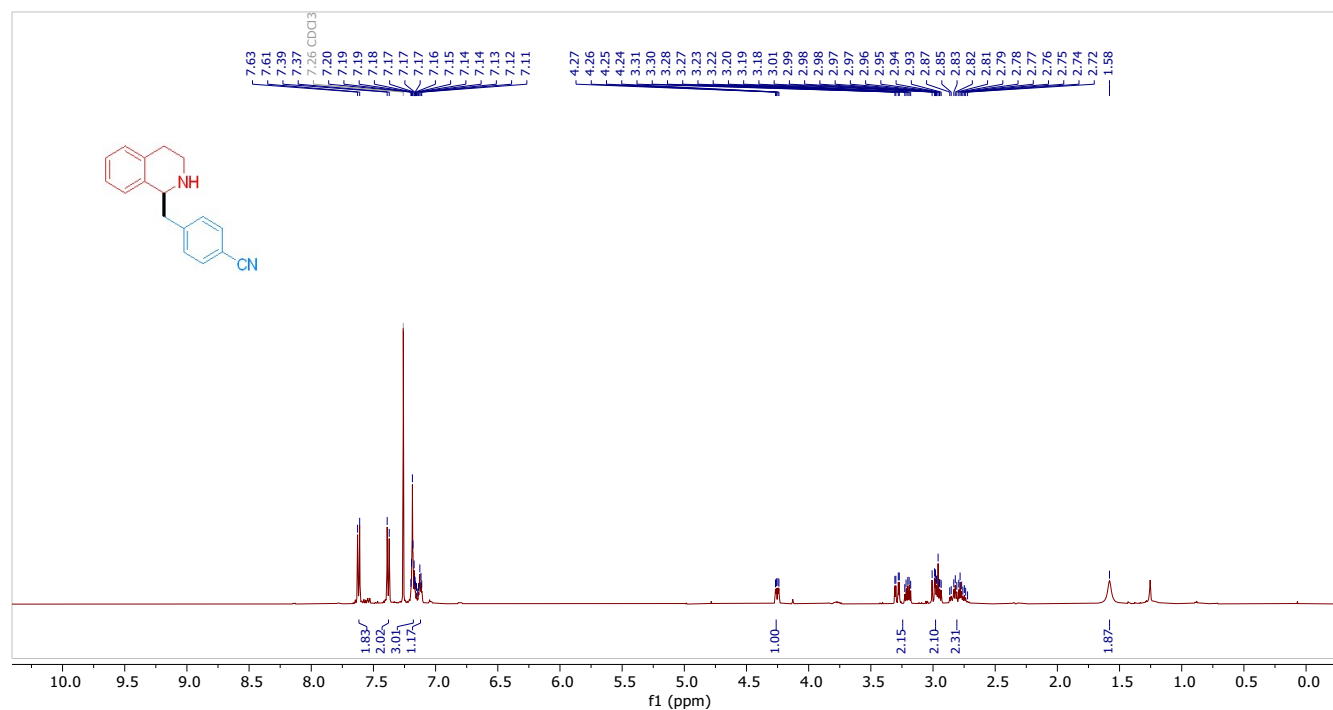
^1H NMR (500 MHz, CDCl_3) spectrum of **3aj**



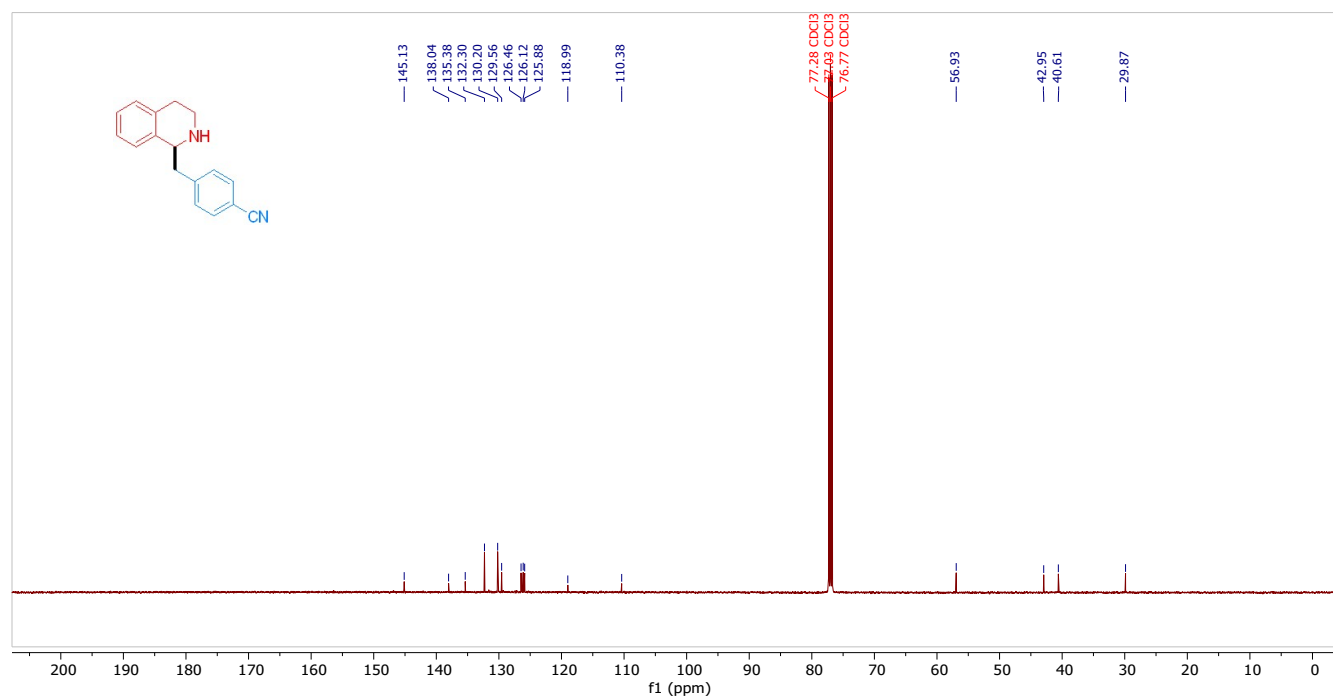
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, C spectrum of **3aj**)



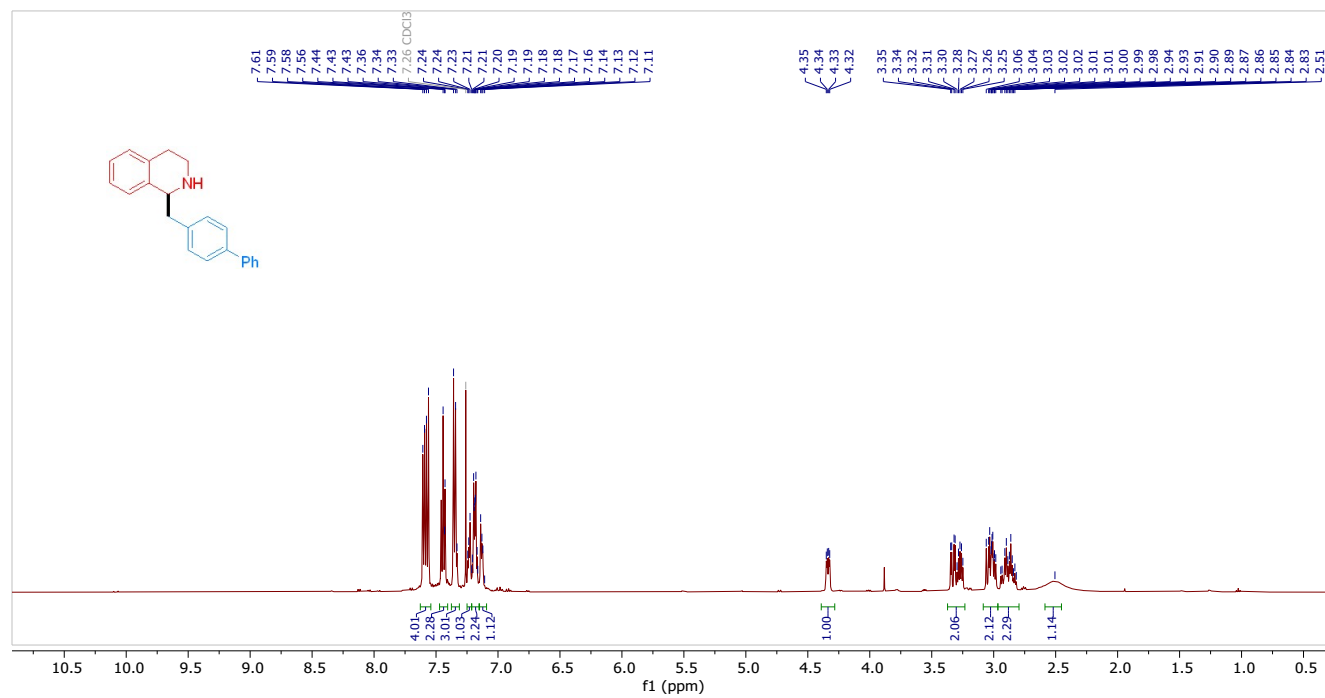
^1H NMR (500 MHz, CDCl_3) spectrum of **3ak**



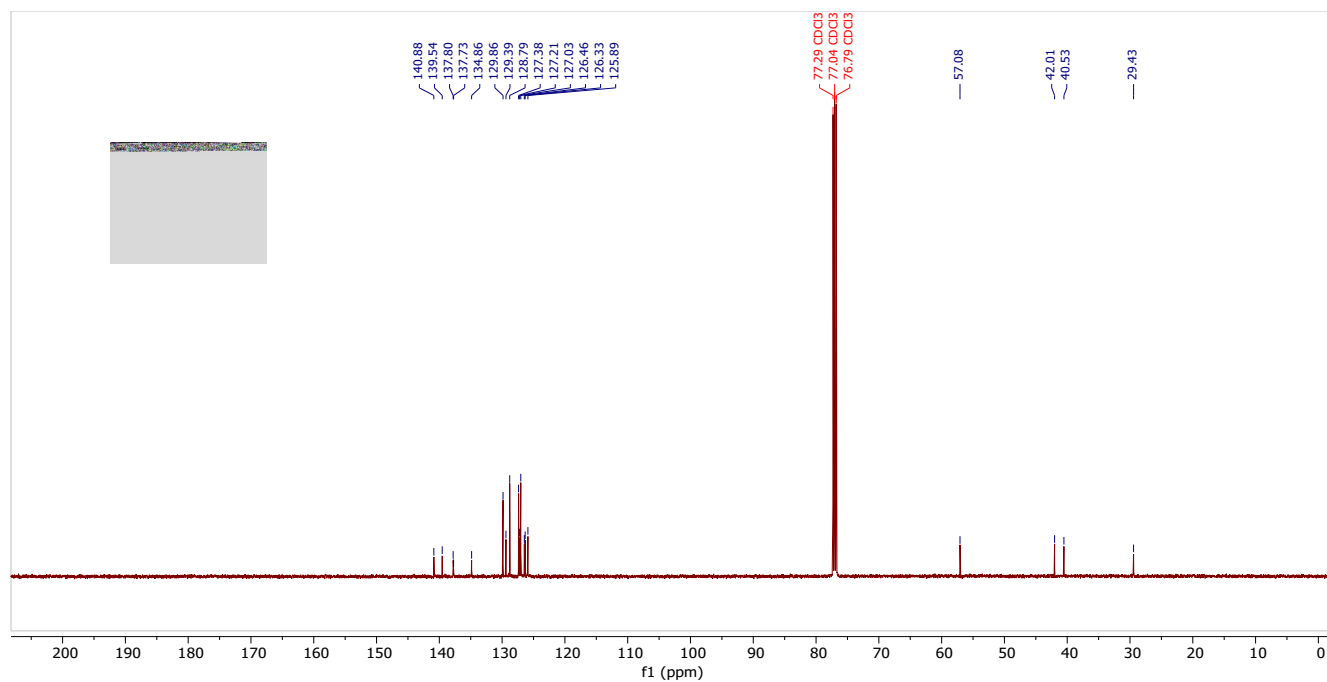
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ak**



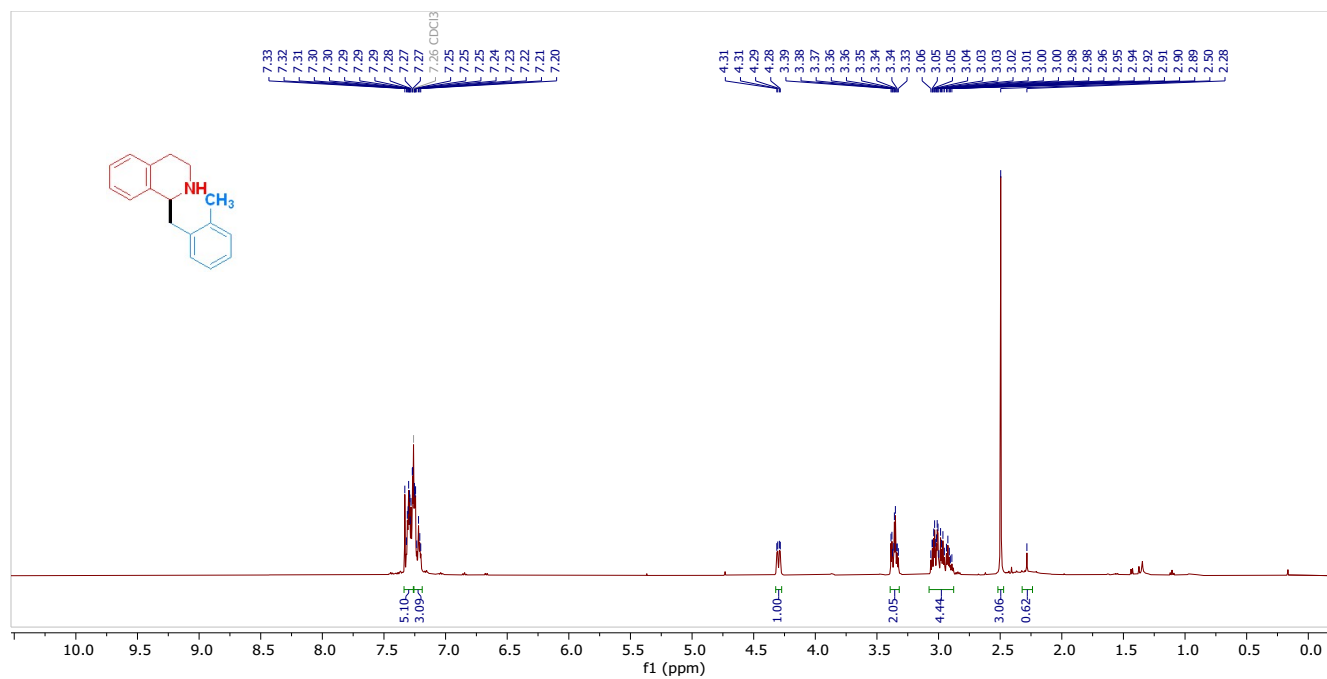
^1H NMR (500 MHz, CDCl_3) spectrum of **3al**



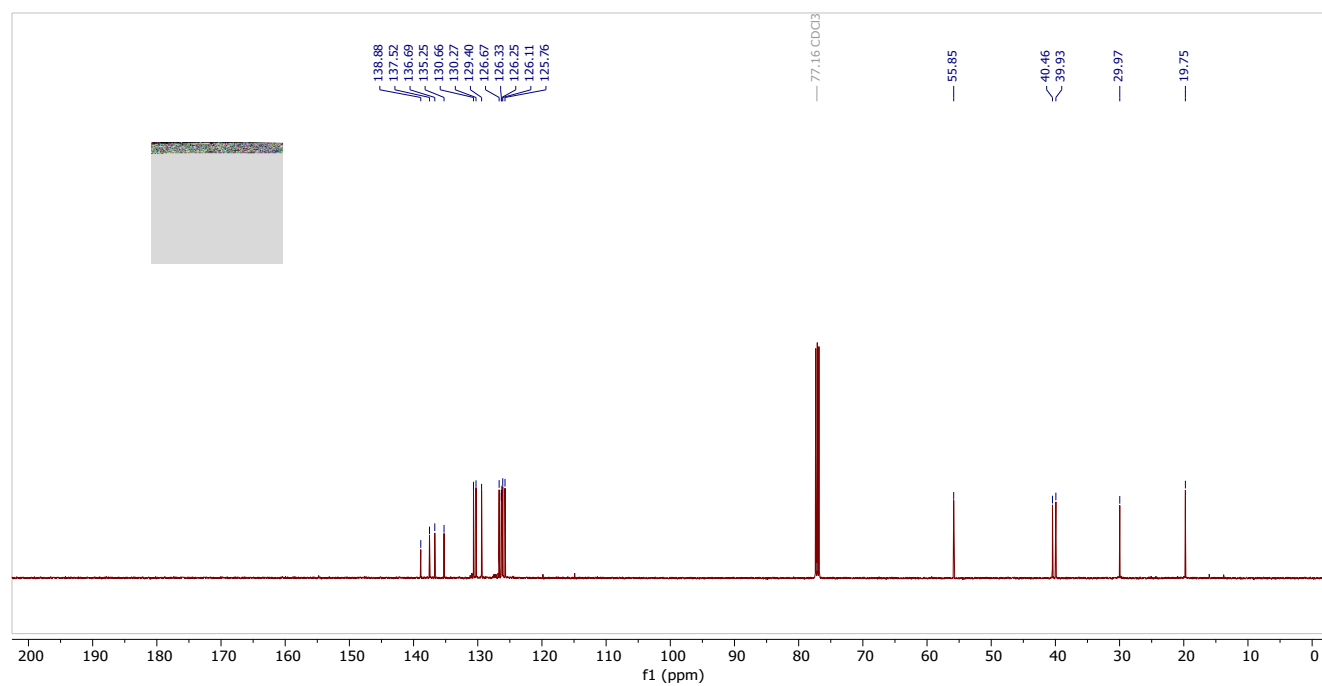
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3al**



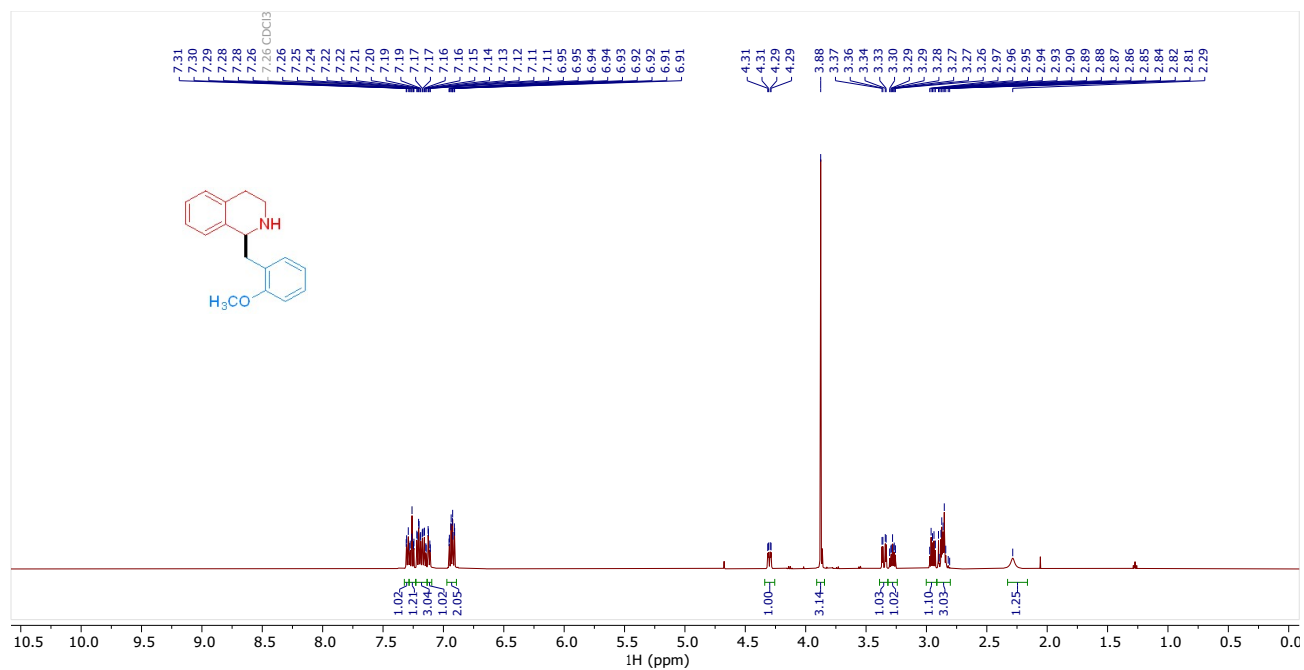
¹H NMR (500 M Hz, CDCl₃) spectrum of **3am**



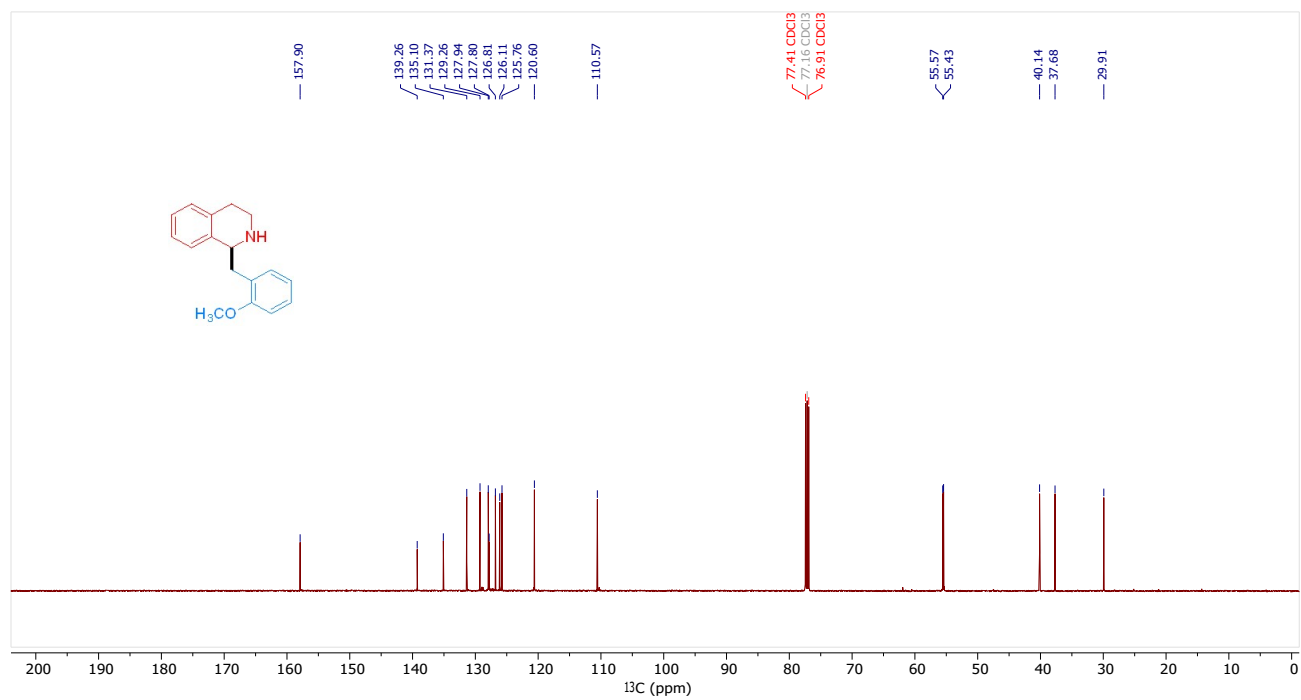
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3am**



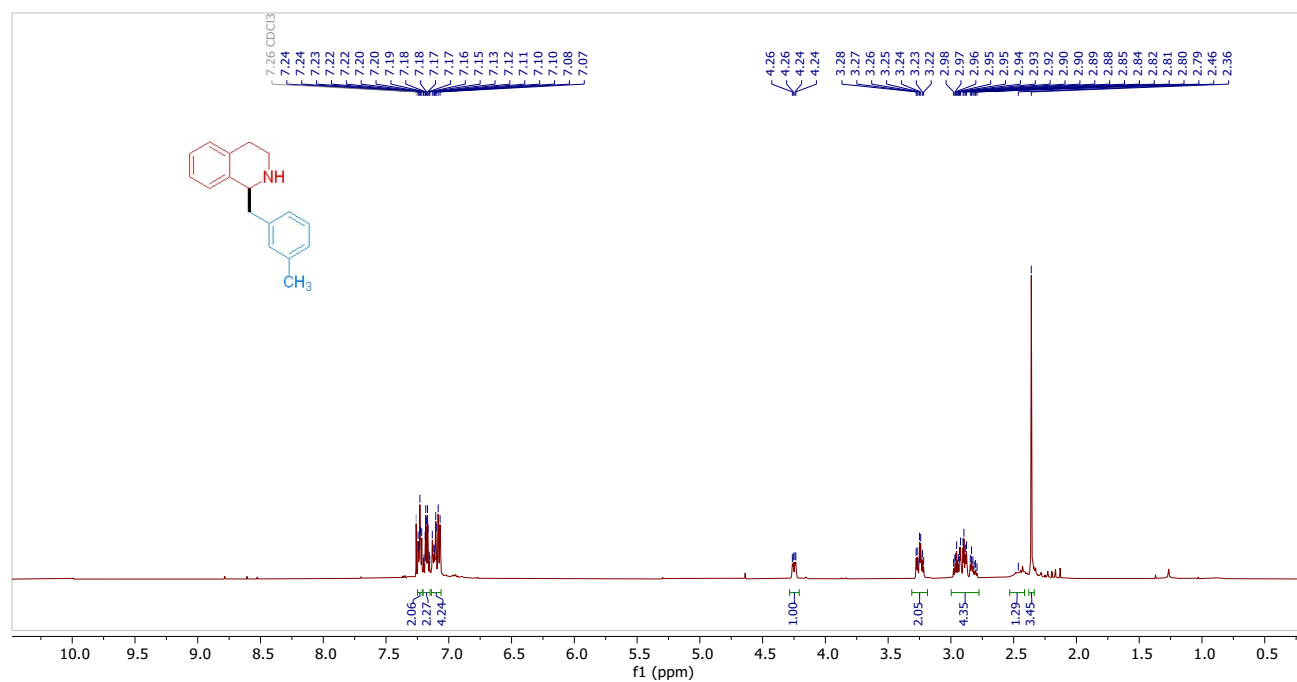
^1H NMR (500 MHz, CDCl_3) spectrum of **3an**



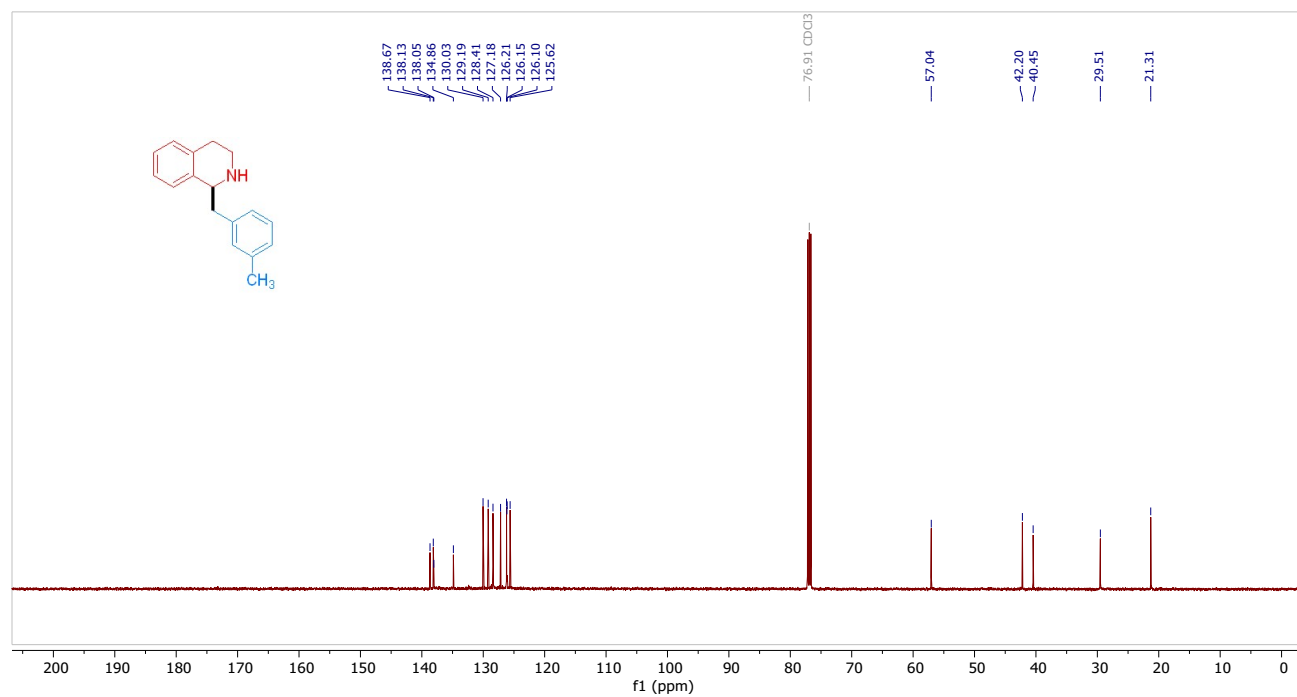
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of 3an



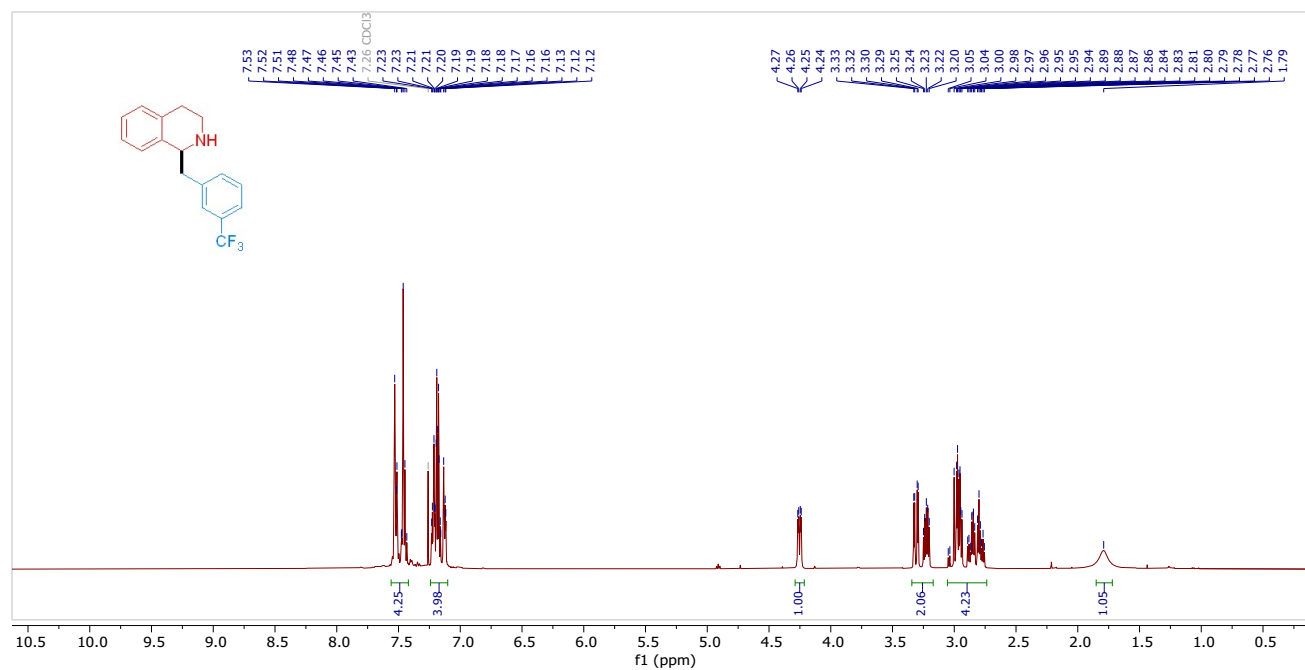
^1H NMR (500 MHz, CDCl_3) spectrum of **3ao**



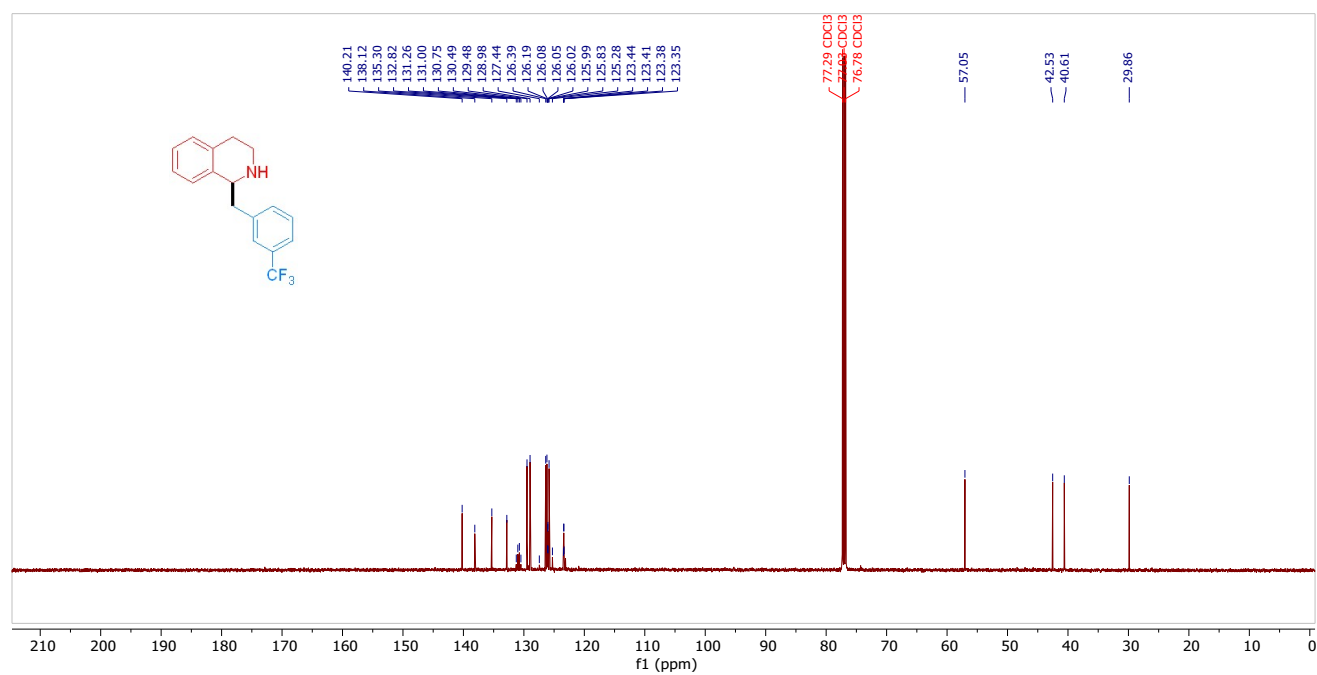
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ao**



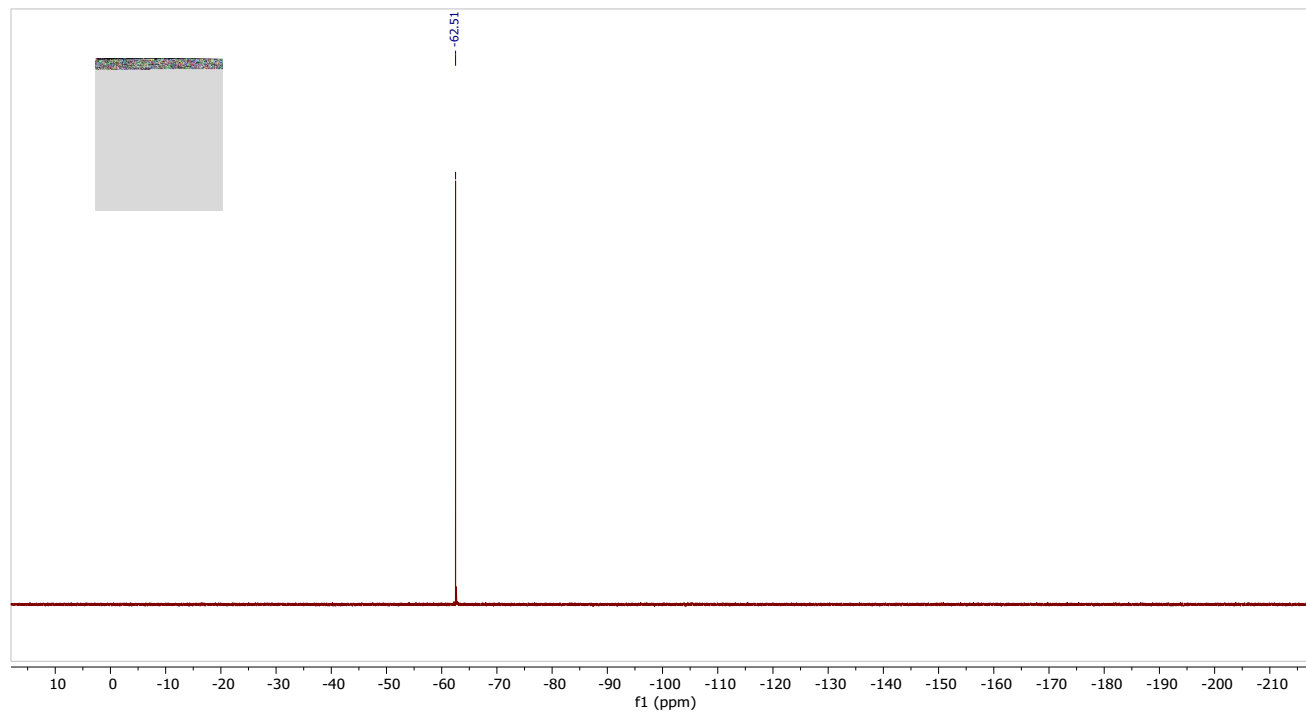
^1H NMR (500 MHz, CDCl_3) spectrum of **3ap**



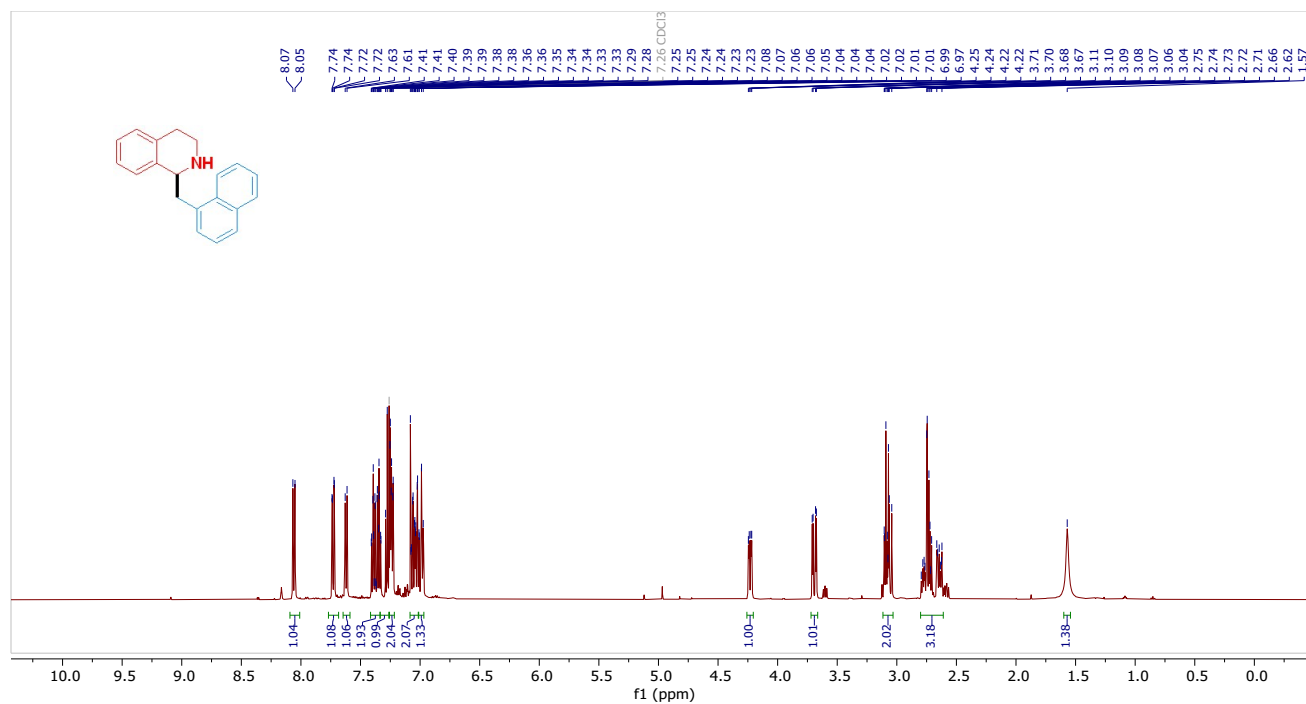
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ap**



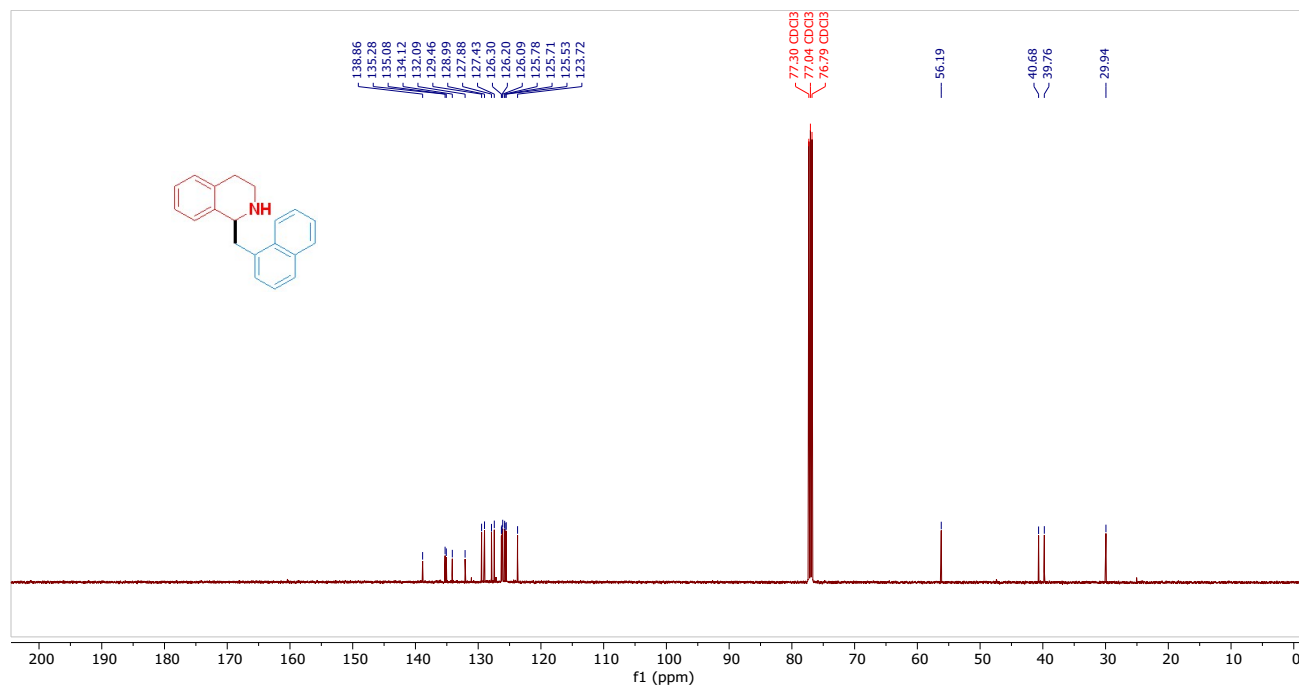
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3ap**



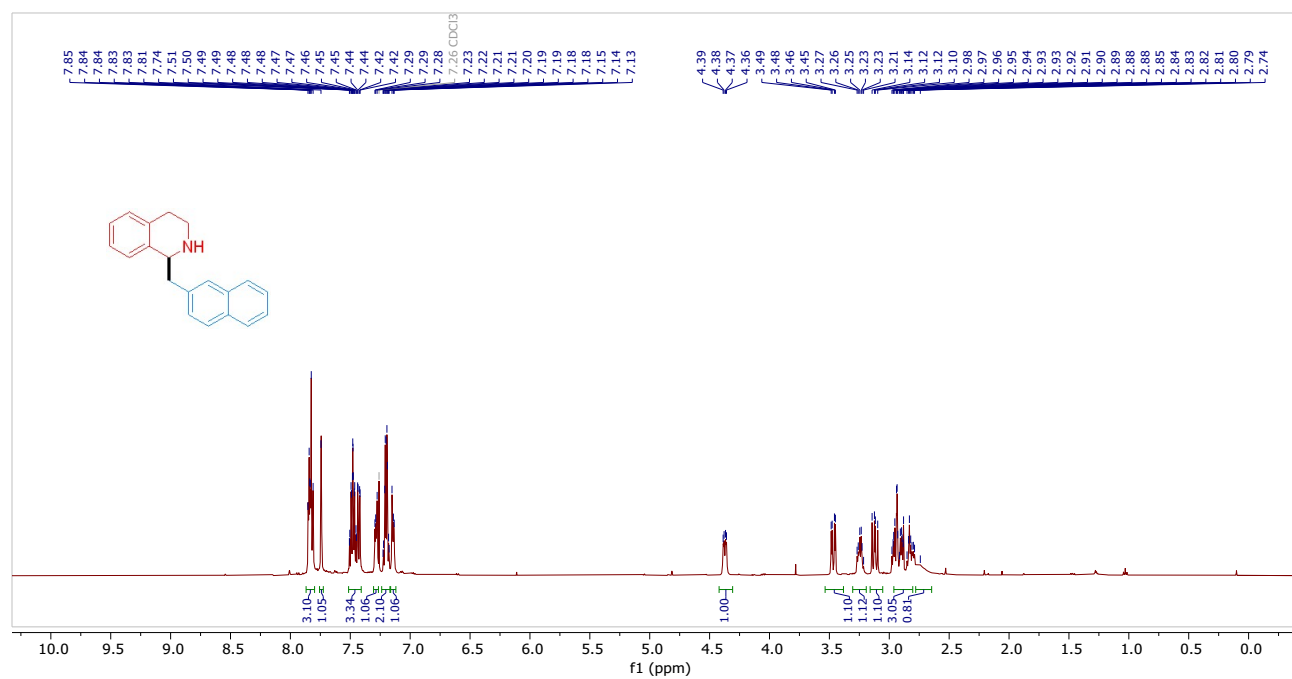
^1H NMR (500 MHz, CDCl_3) spectrum of **3as**



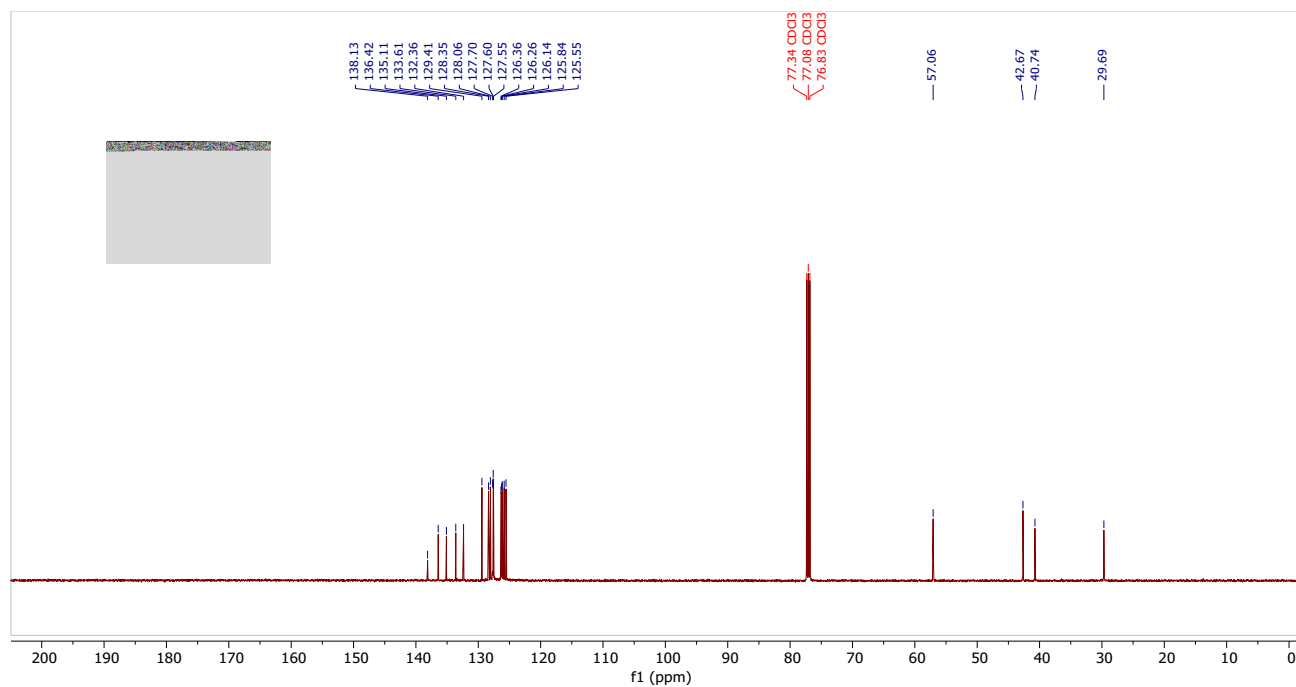
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3as**



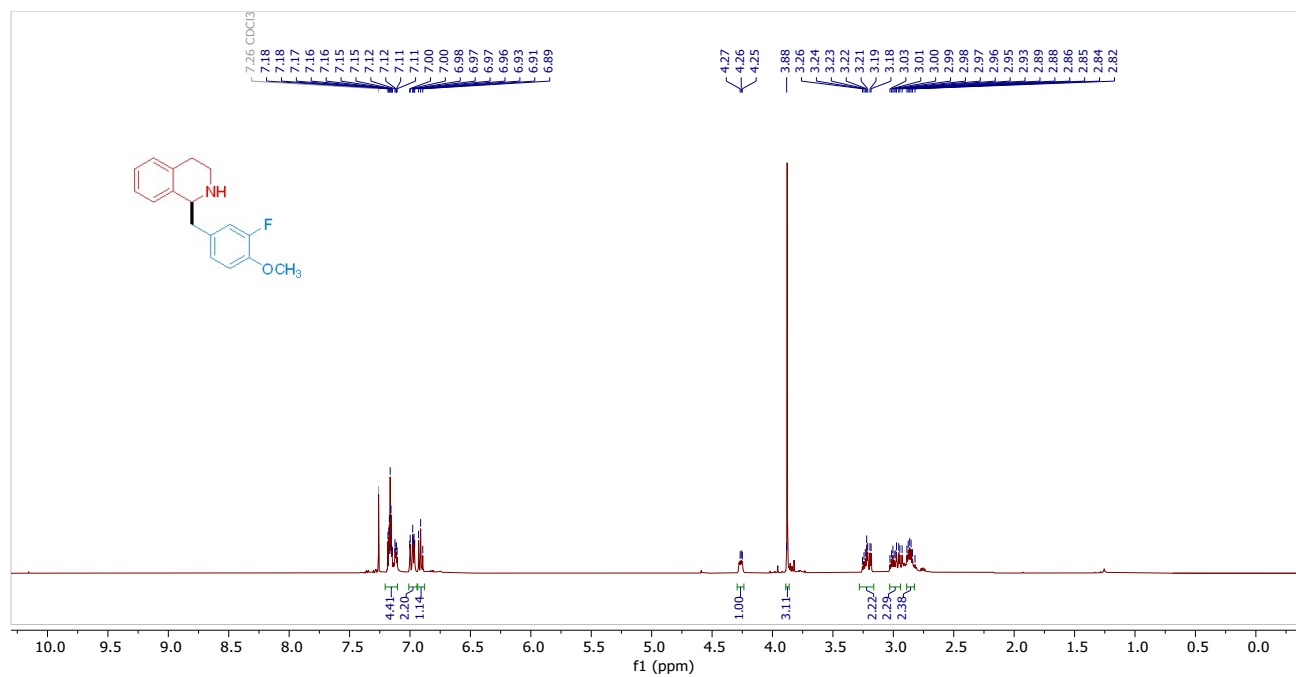
^1H NMR (500 MHz, CDCl_3) spectrum of **3at**



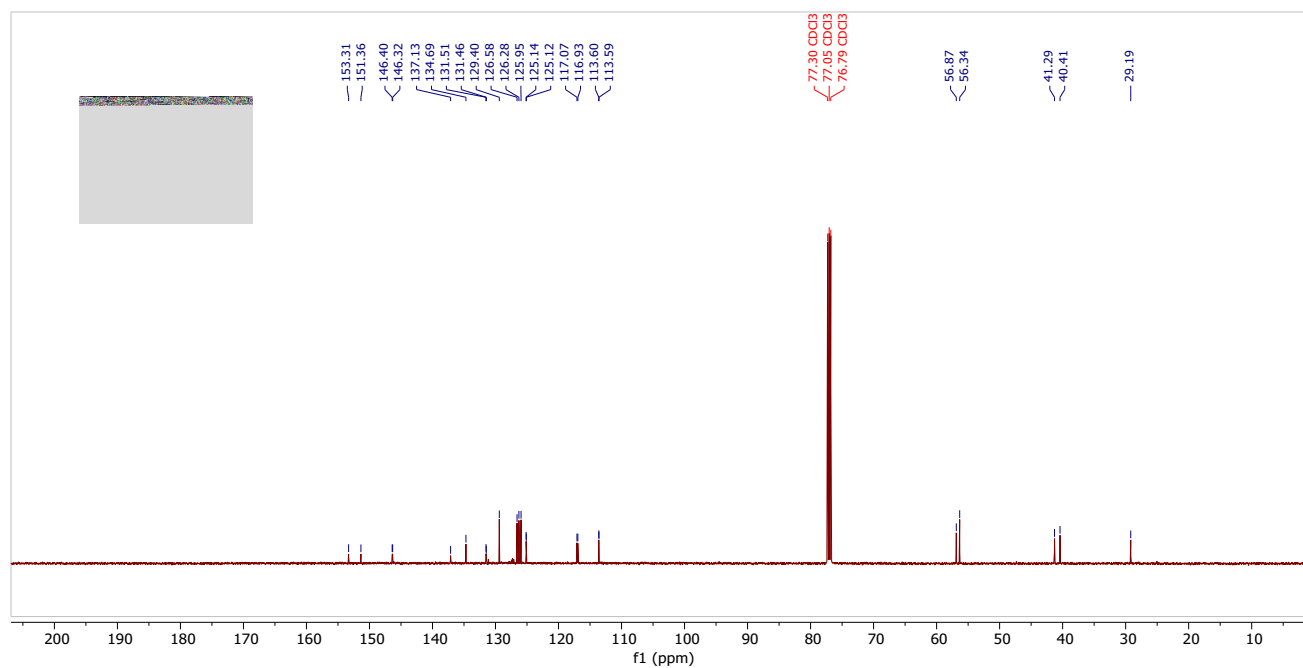
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3at**



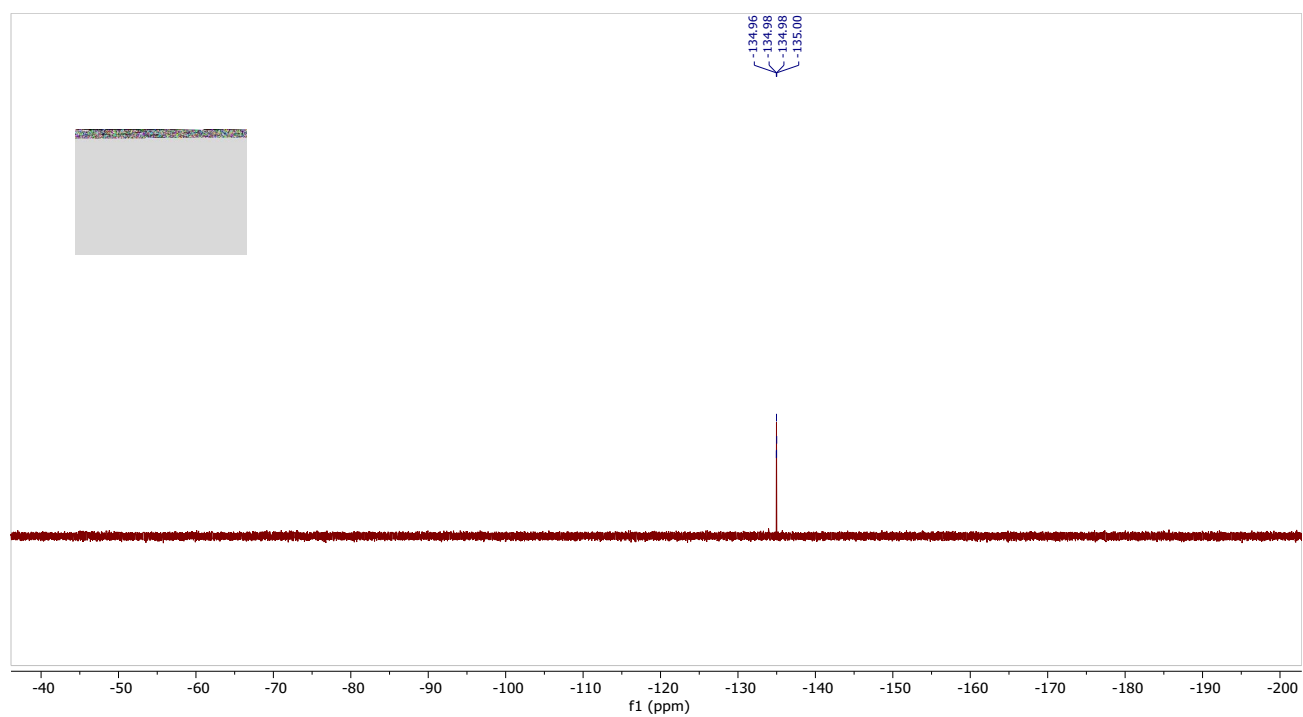
¹H NMR (500 MHz, CDCl₃) spectrum of **3au**



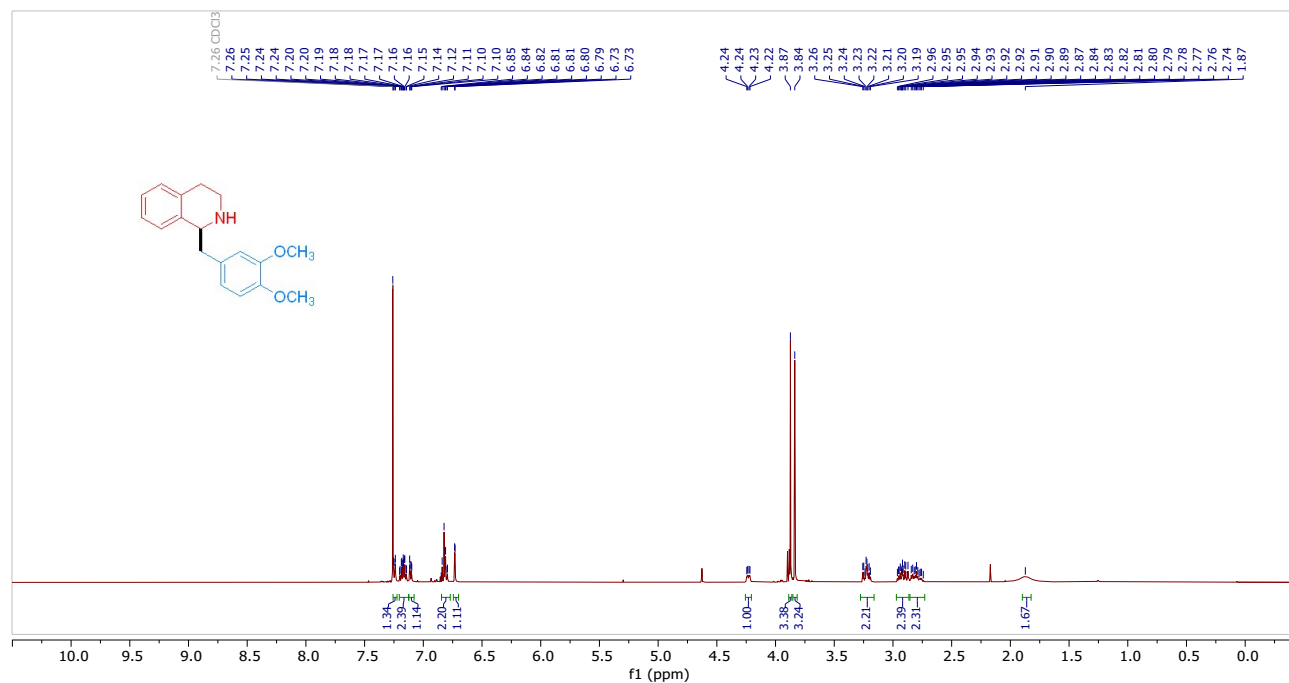
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3au**



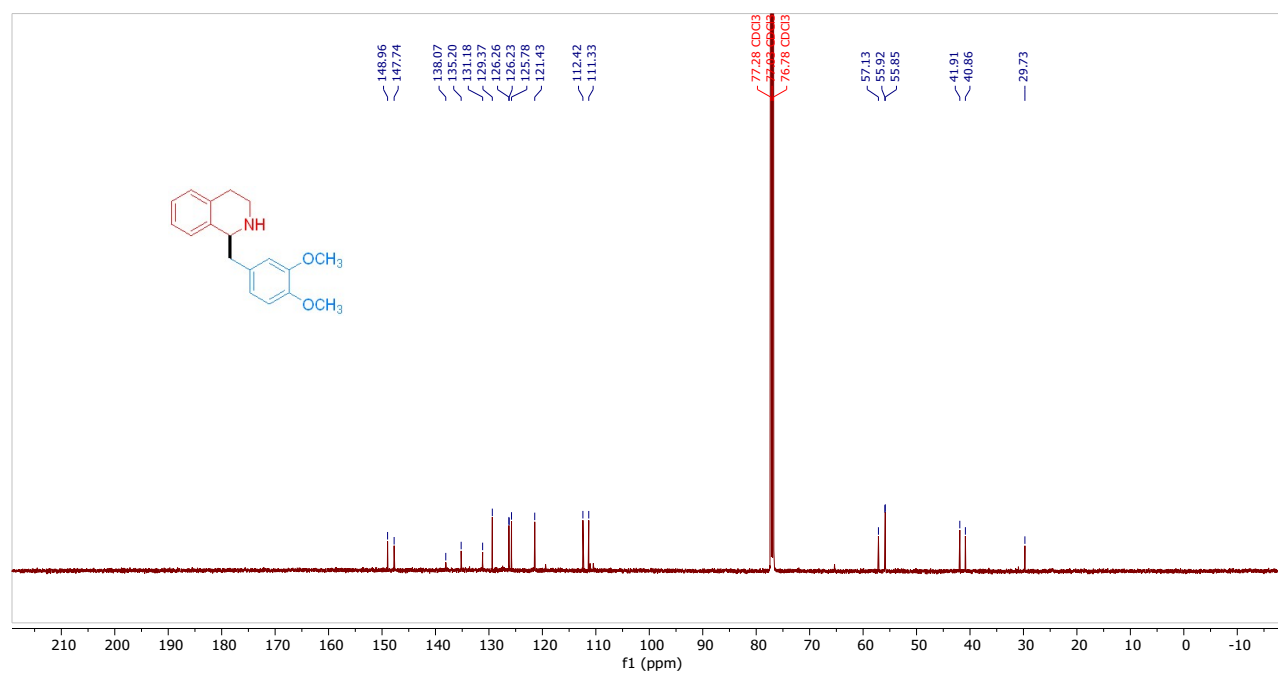
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3au**



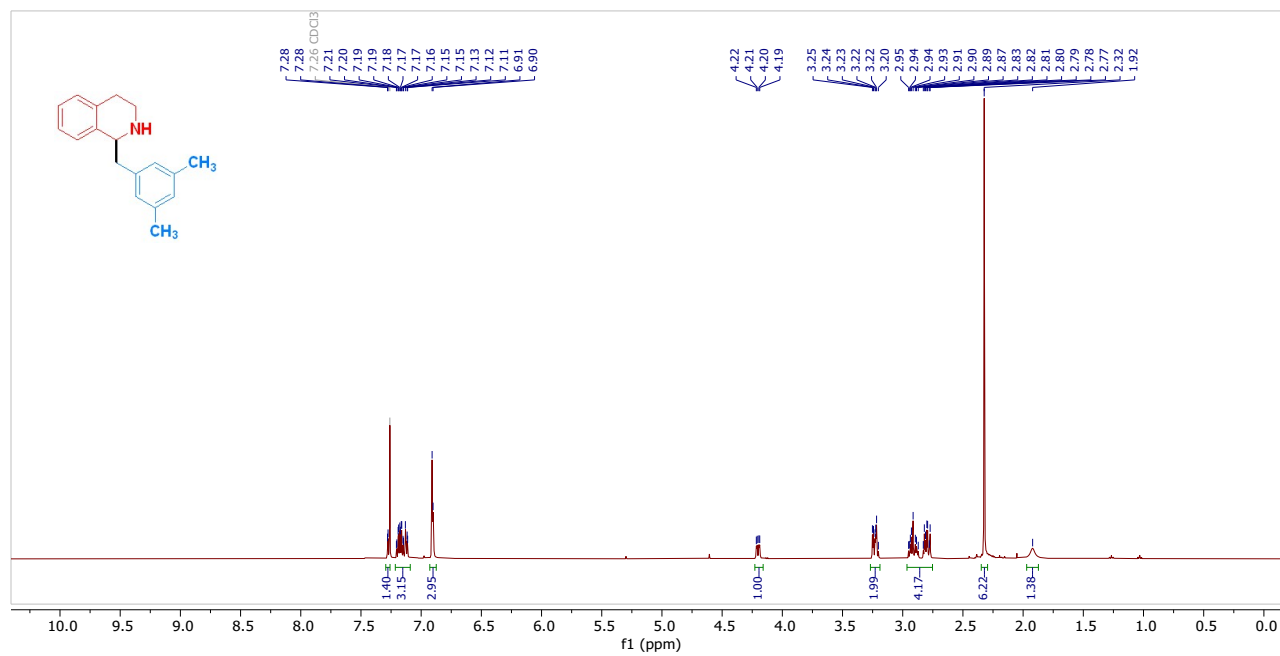
^1H NMR (500 MHz, CDCl_3) spectrum of **3av**



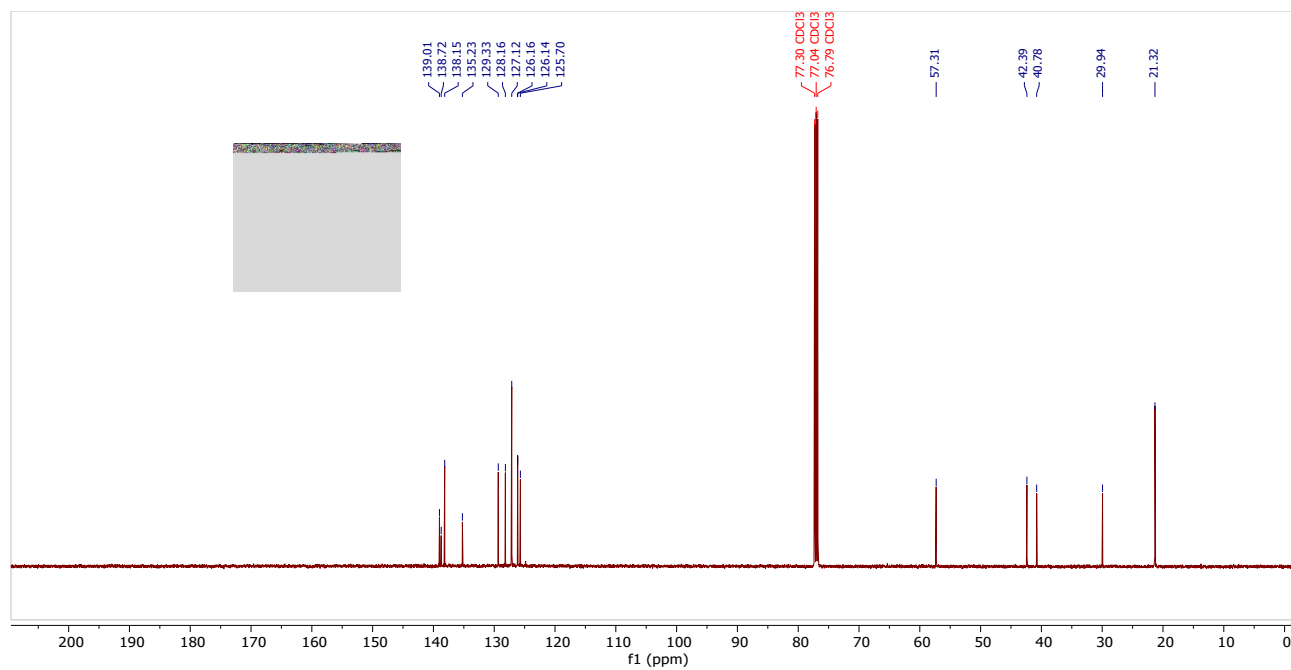
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3av**



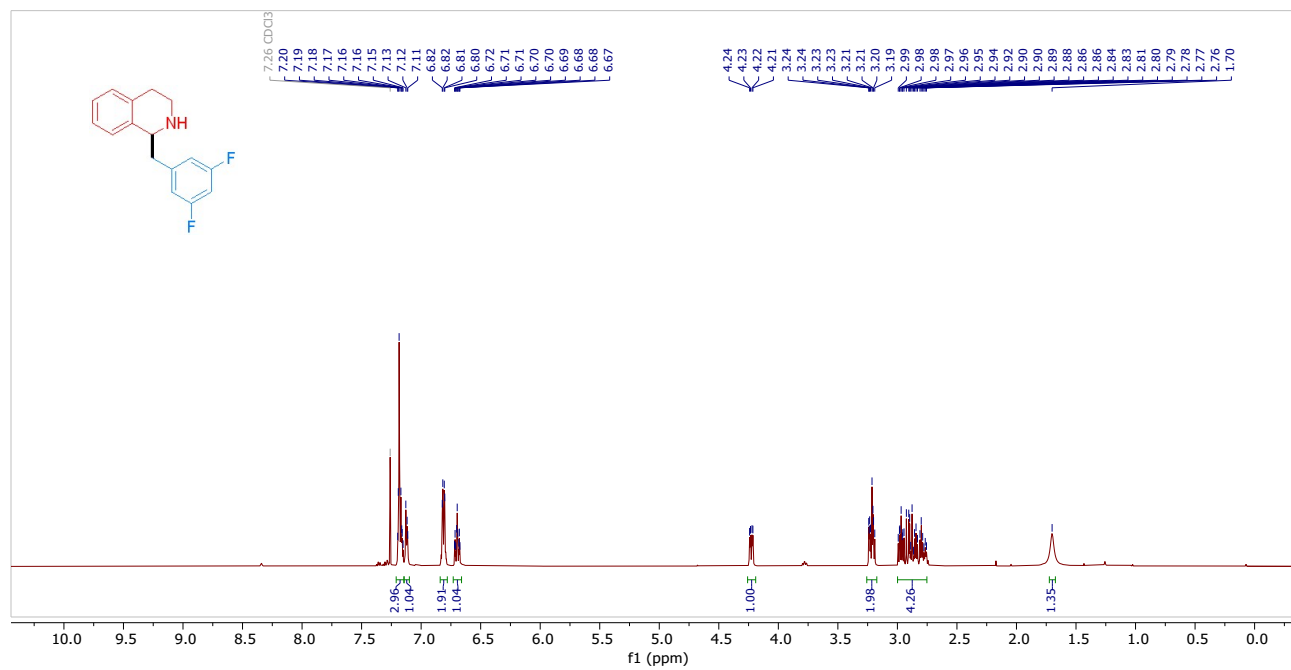
^1H NMR (500 MHz, CDCl_3) spectrum of **3aw**



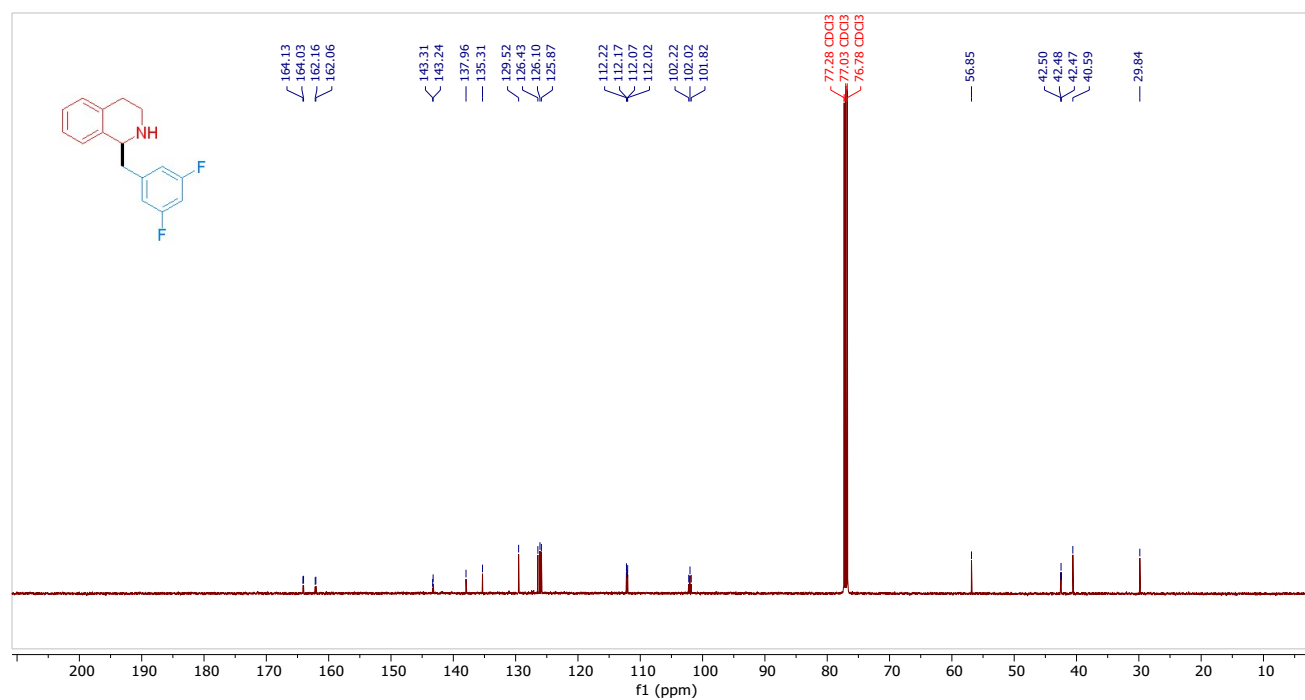
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3aw**



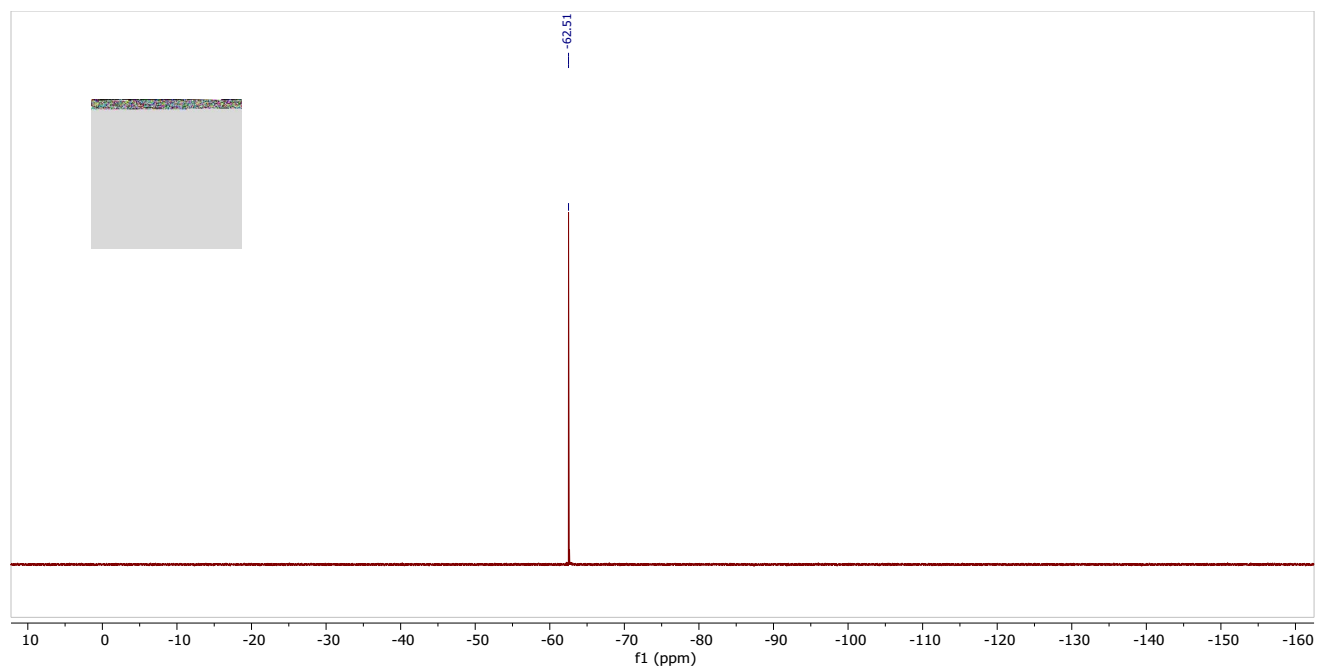
¹H NMR (500 MHz, CDCl₃) spectrum of **3ax**



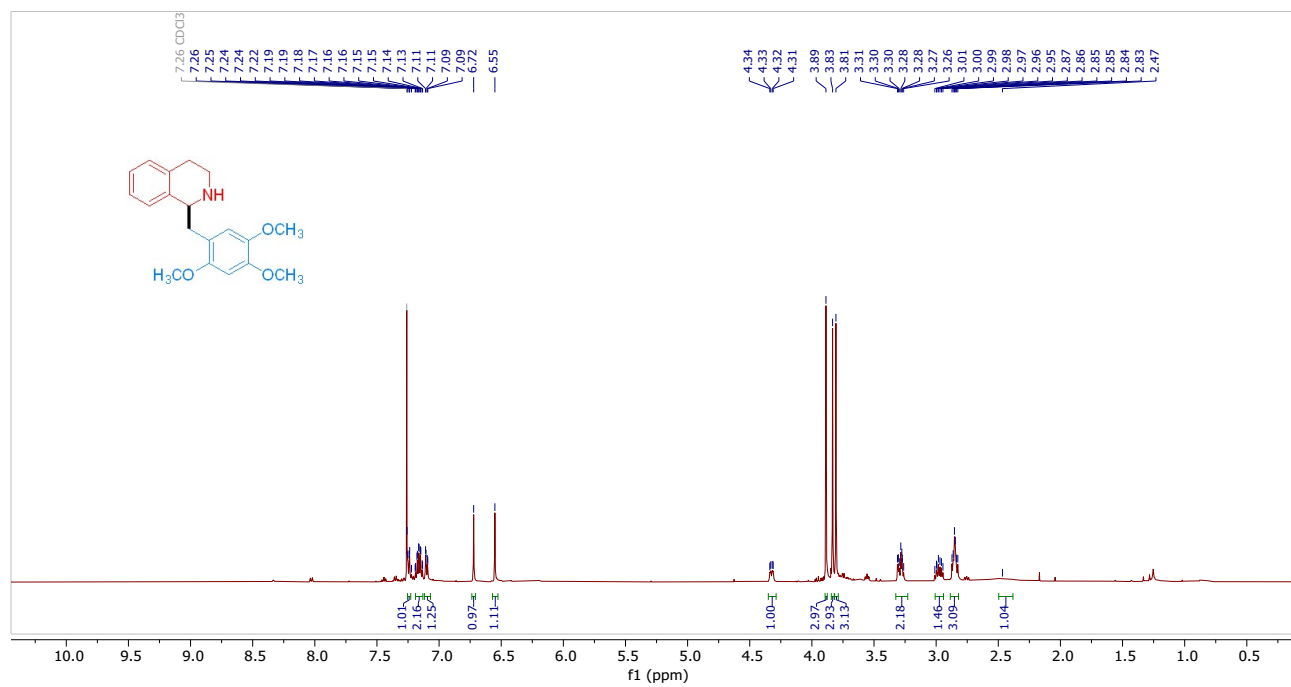
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ax**



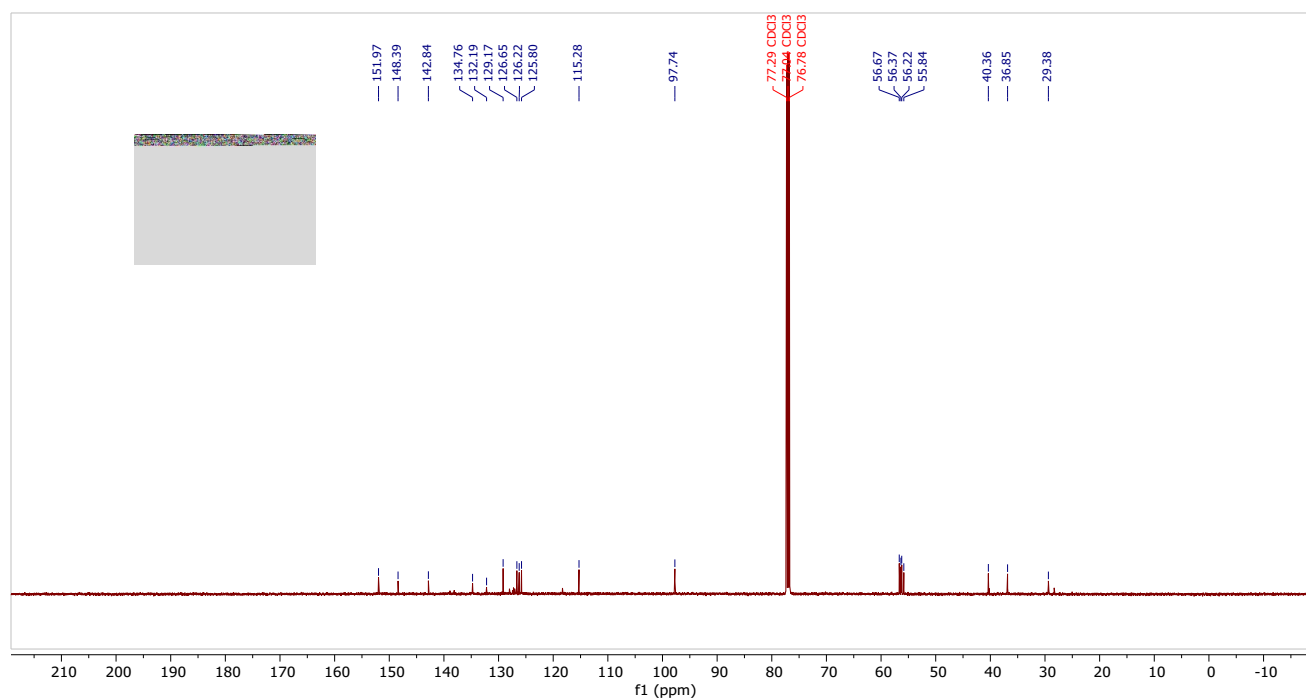
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3ax**



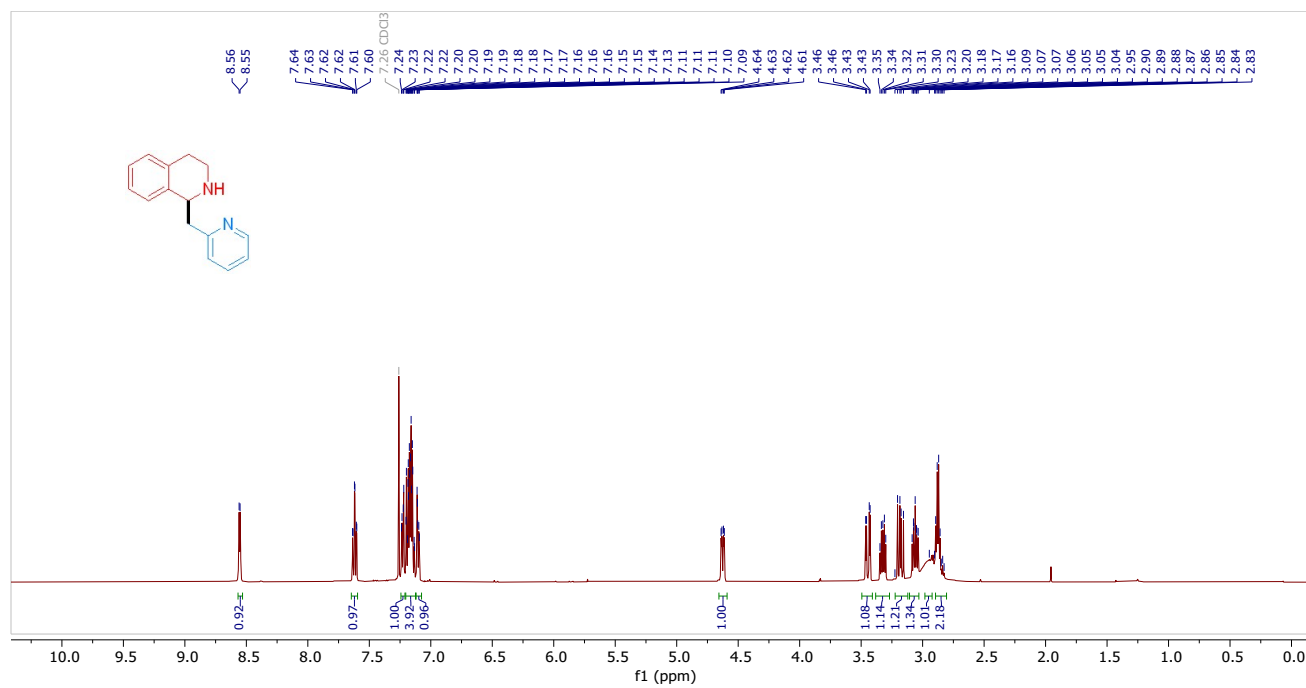
^1H NMR (500 MHz, CDCl_3) spectrum of **3ay**



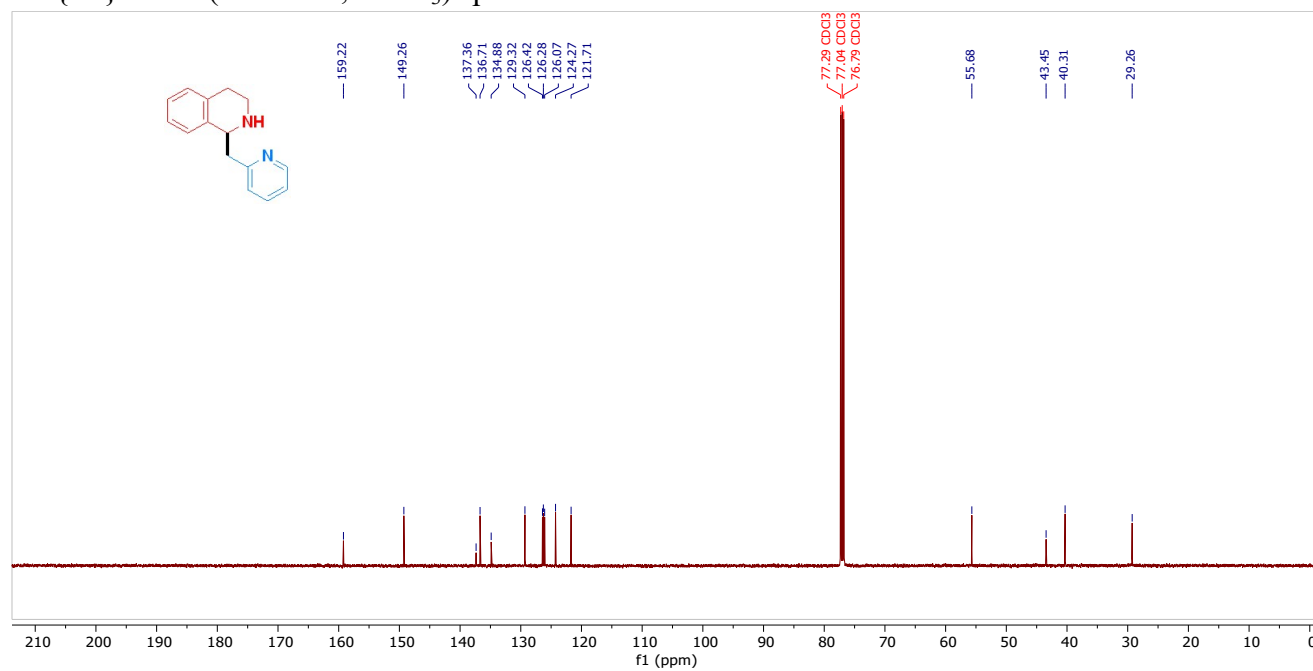
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ay**



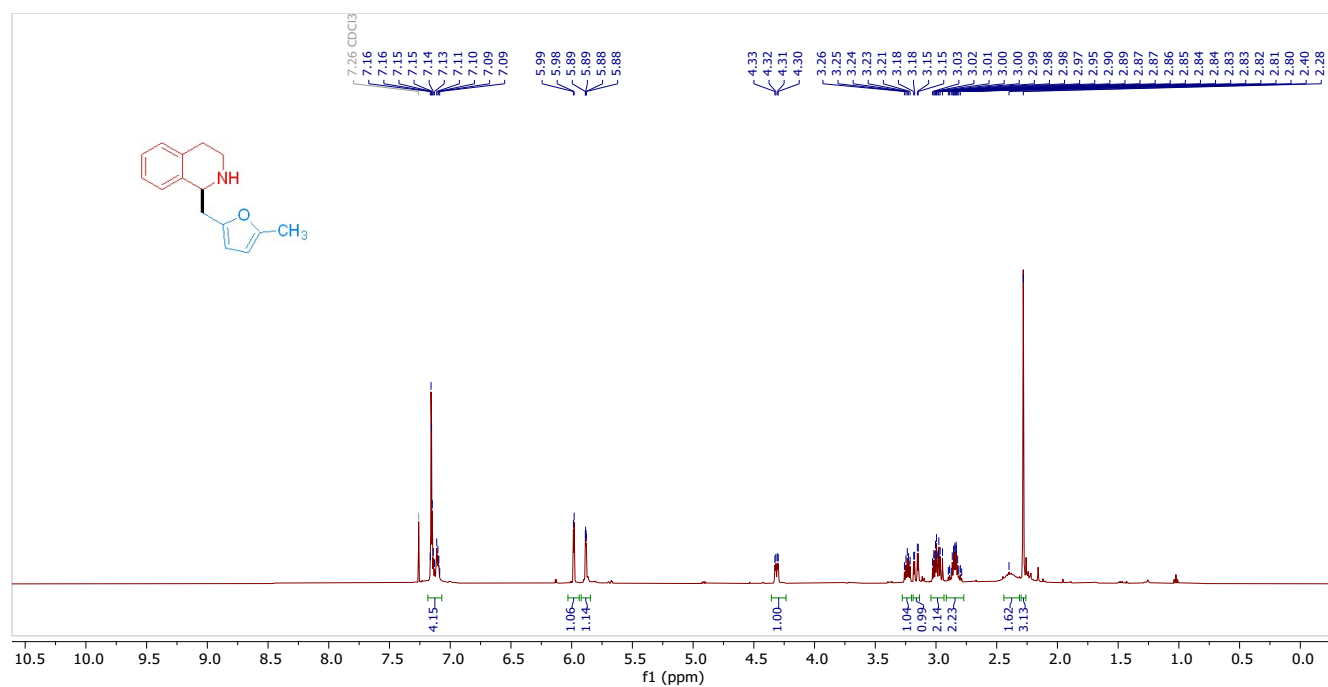
^1H NMR (500 MHz, CDCl_3) spectrum of **3az**



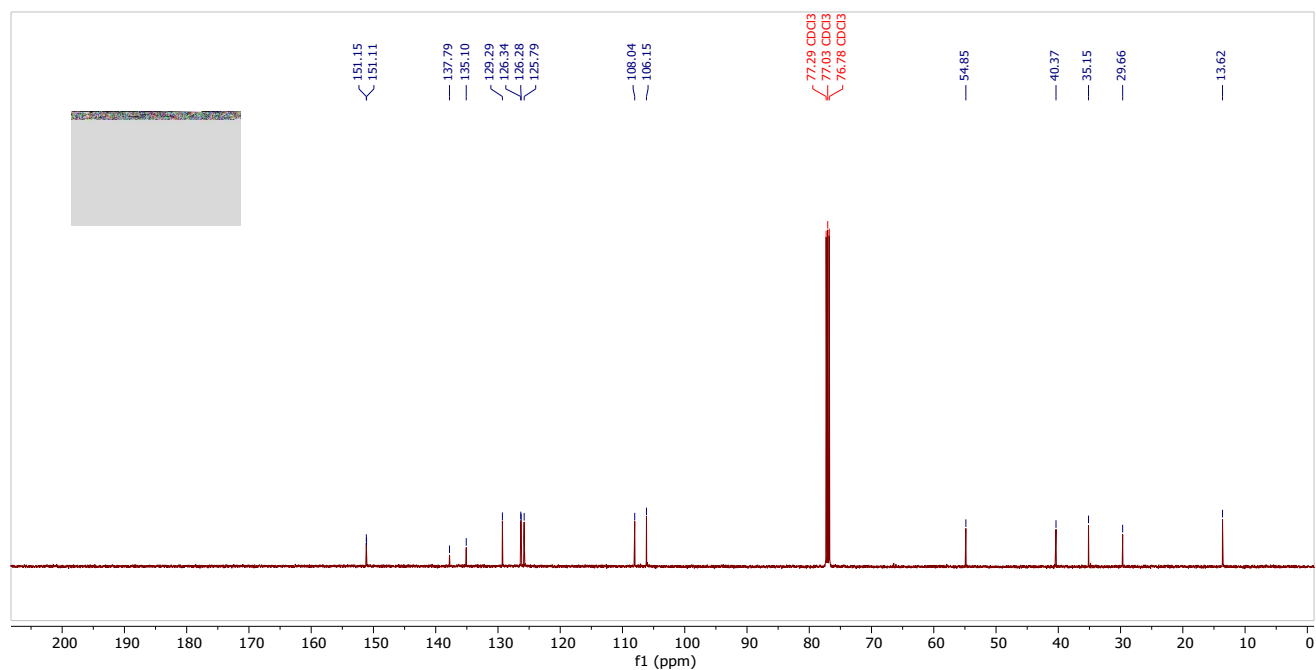
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3az**



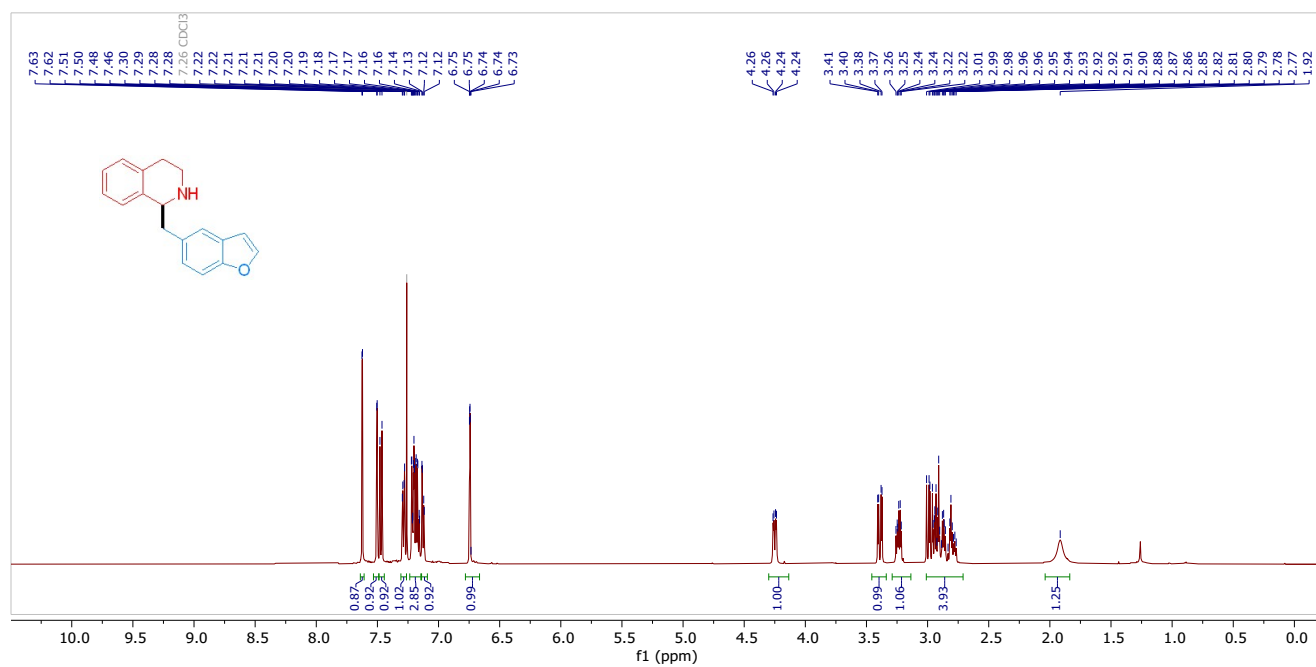
^1H NMR (500 MHz, CDCl_3) spectrum of **3ba**



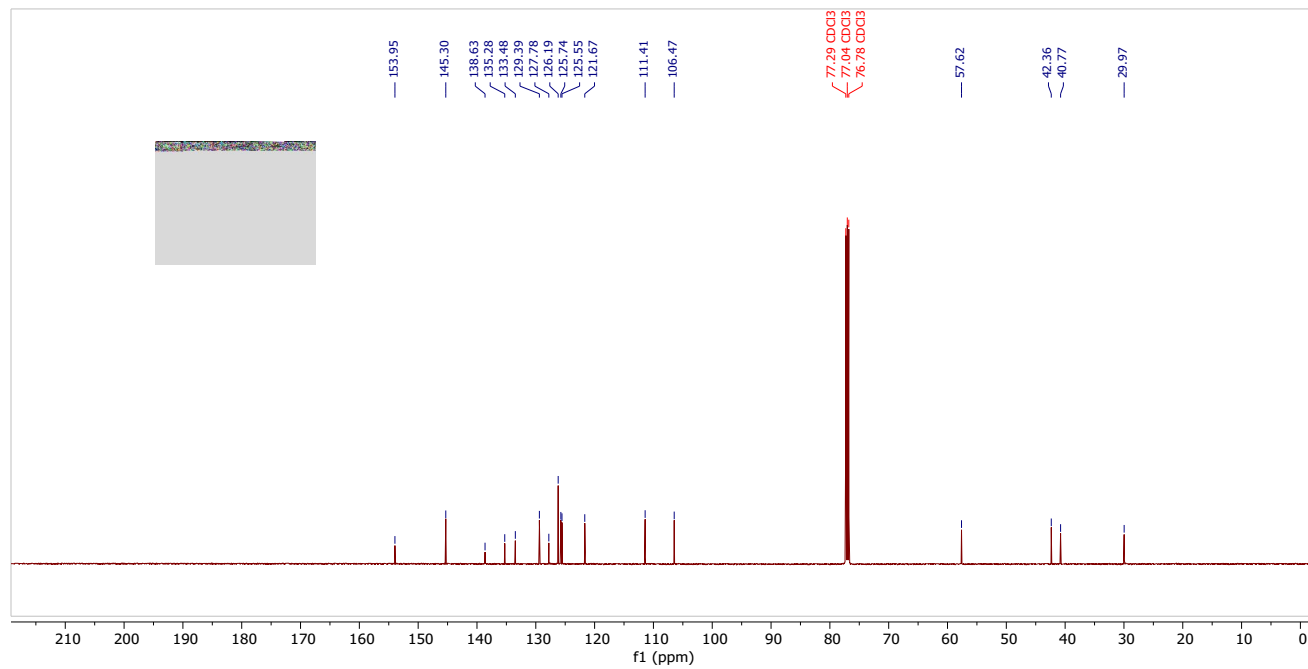
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3ba**



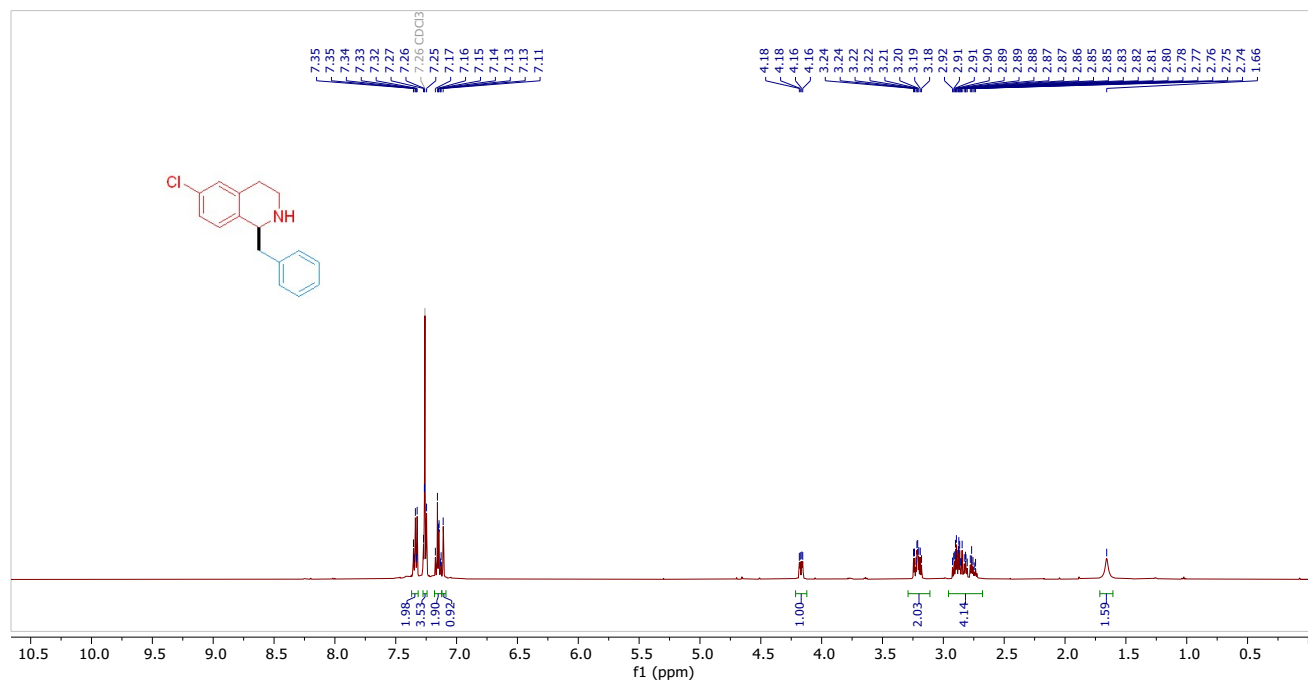
^1H NMR (500 MHz, CDCl_3) spectrum of **3bb**



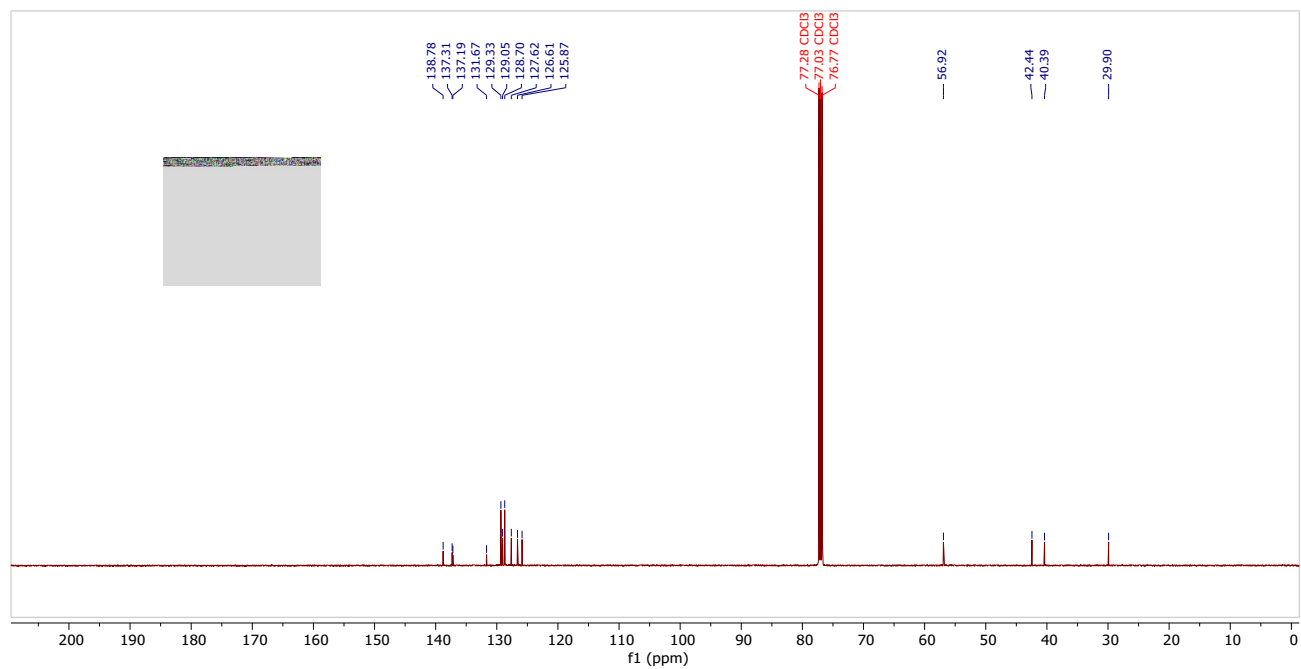
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3bb**



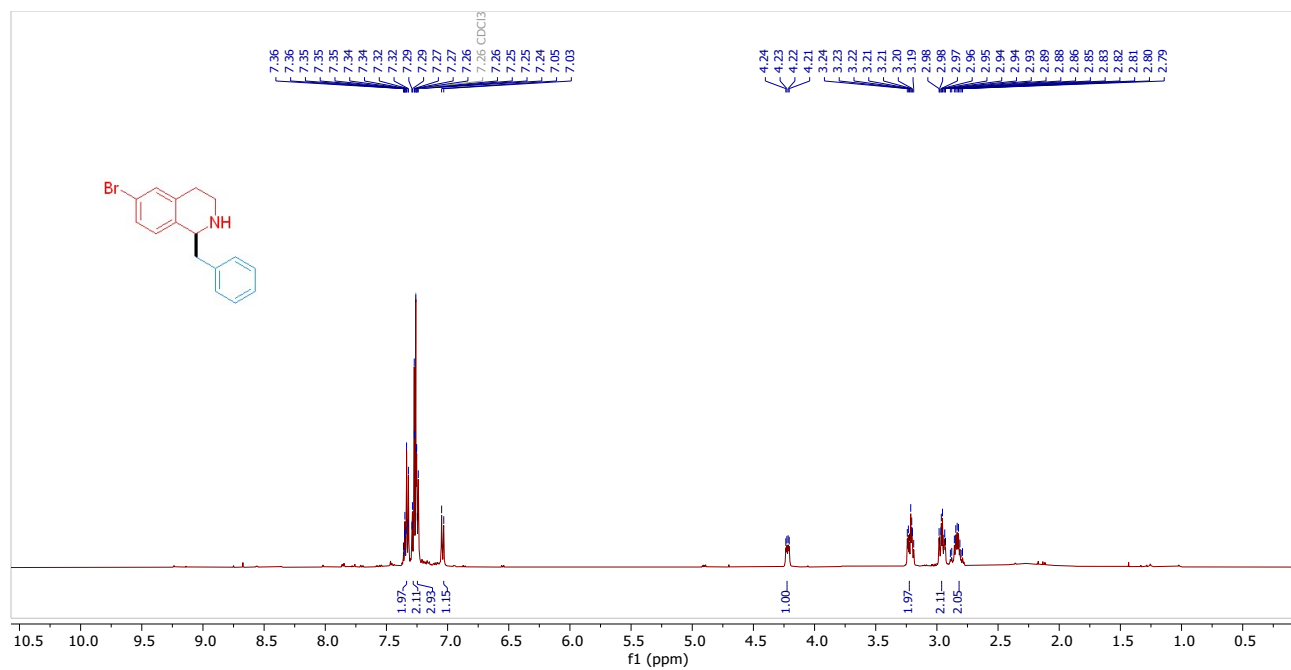
^1H NMR (500 MHz, CDCl_3) spectrum of **3be**



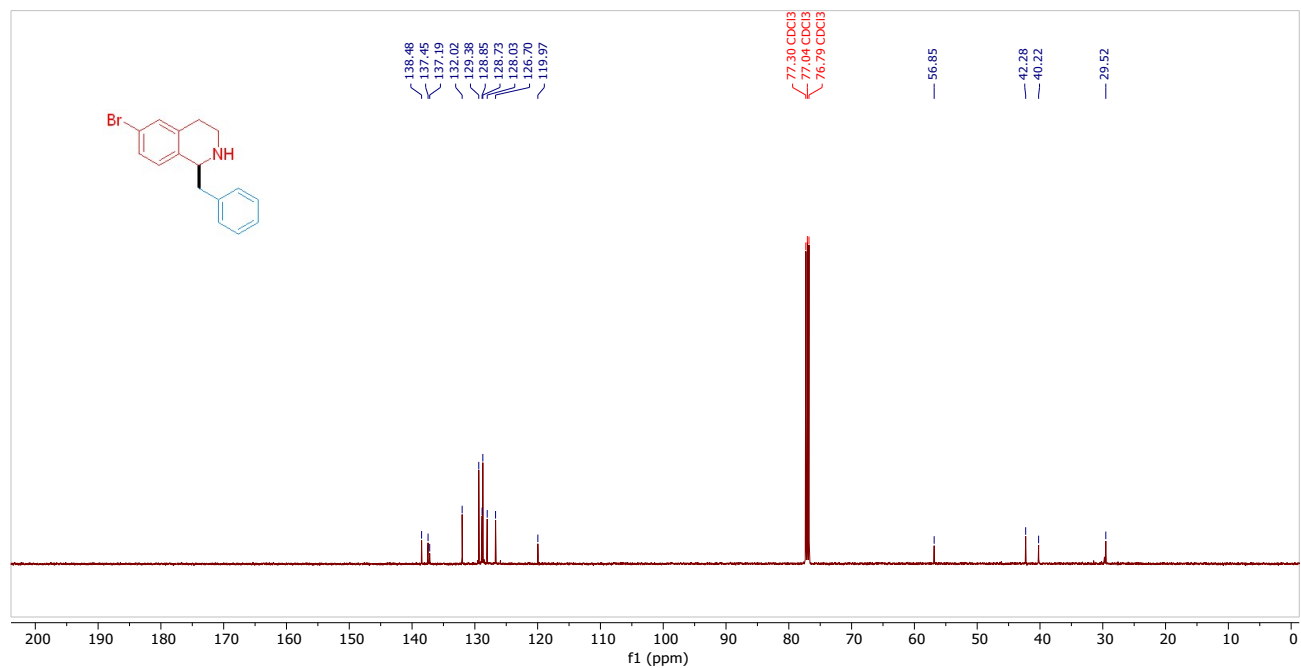
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3be**



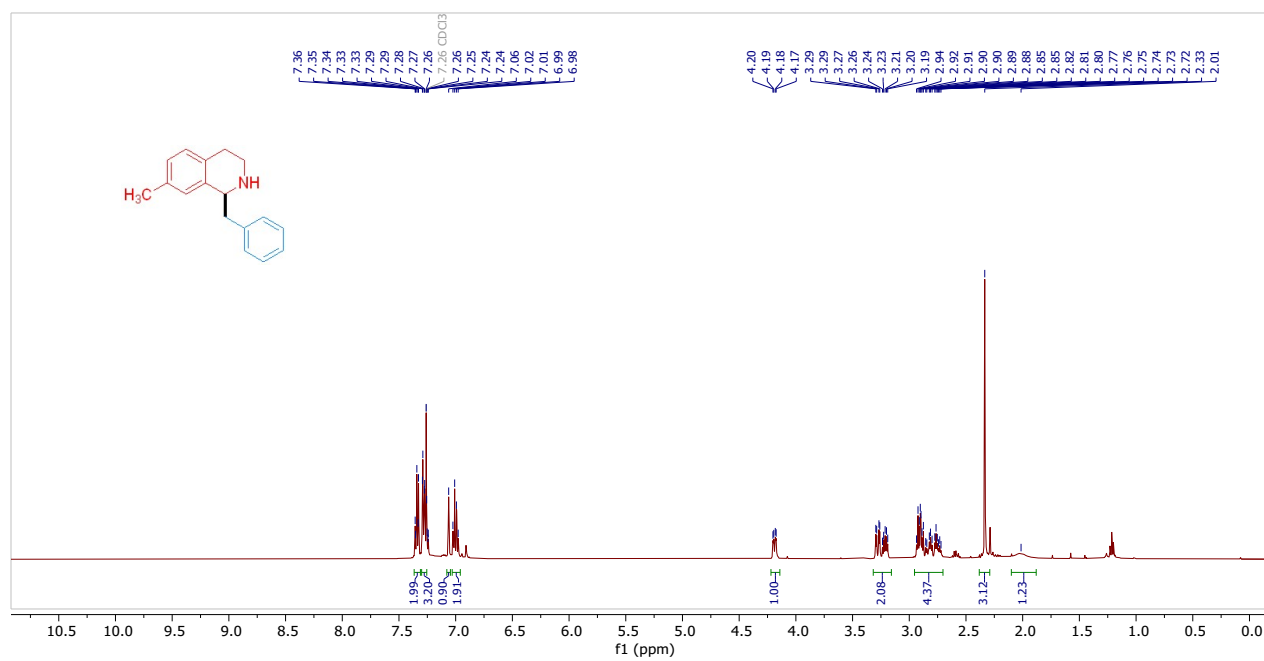
^1H NMR (500 MHz, CDCl_3) spectrum of **3bf**



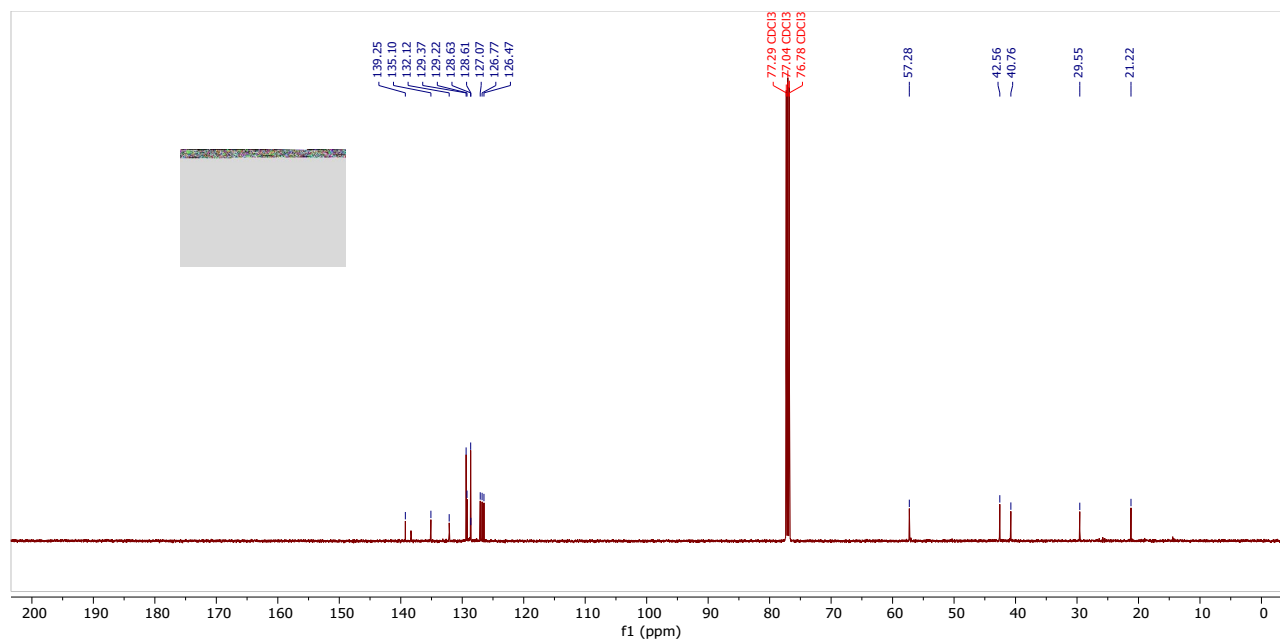
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of **3bf**



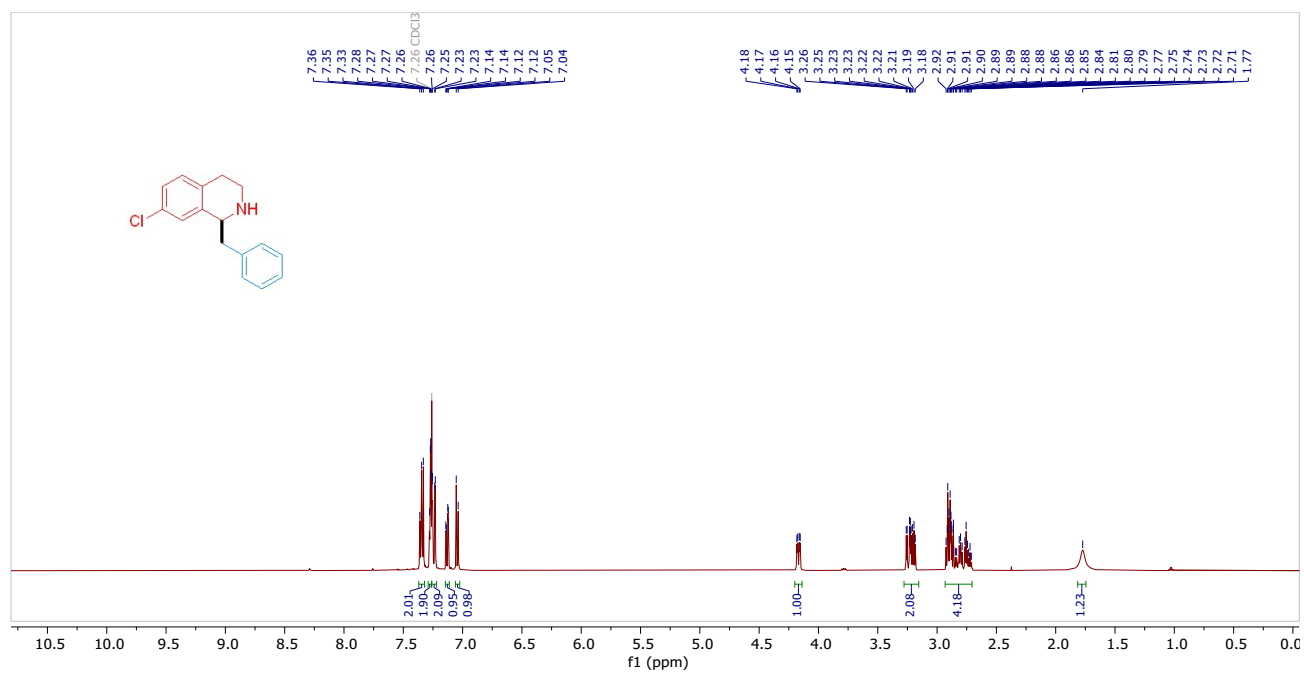
^1H NMR (500 MHz, CDCl_3) spectrum of **3bg**



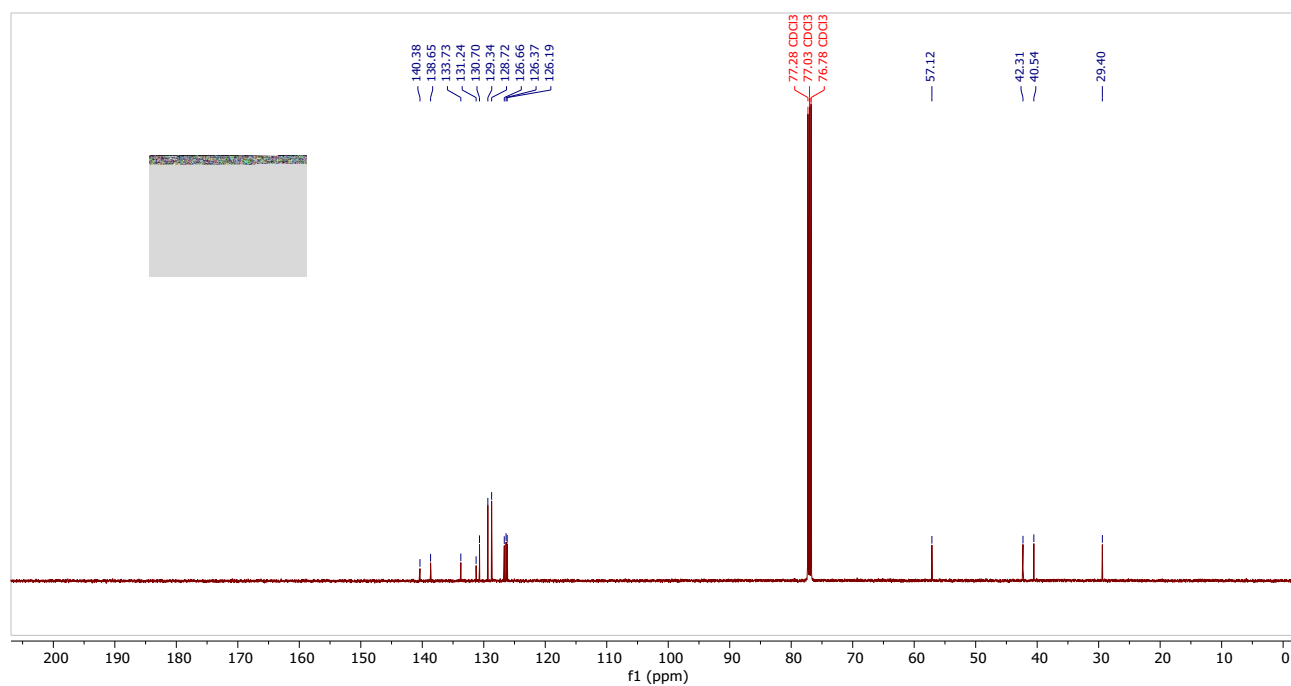
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3bg**



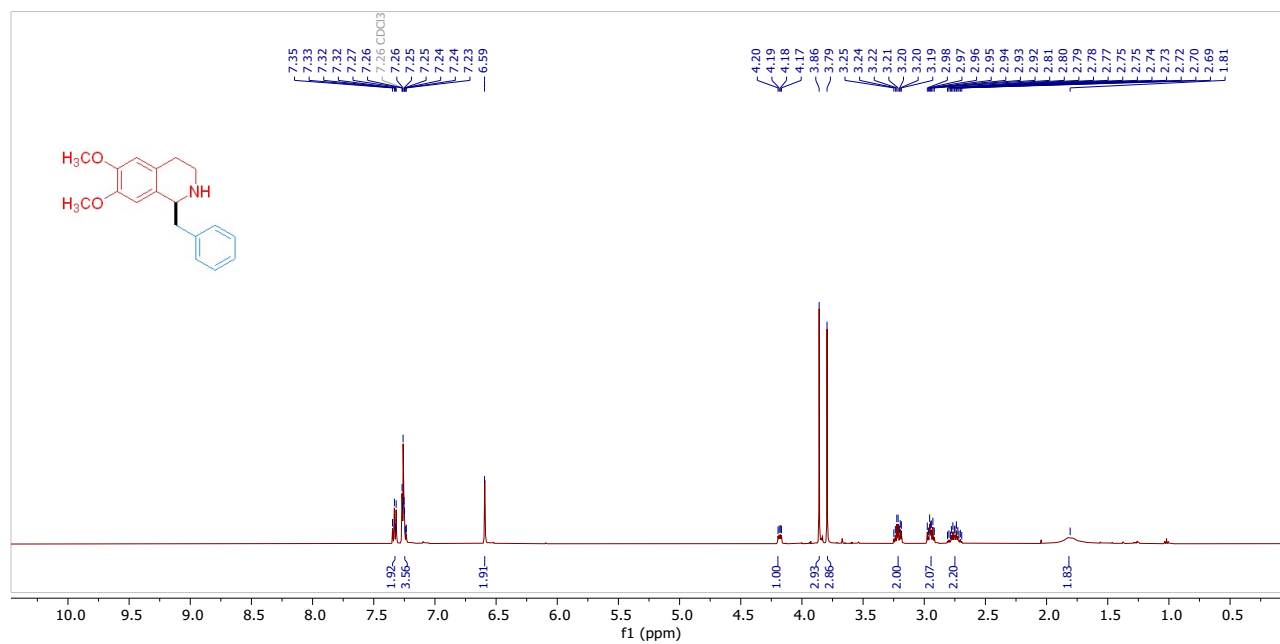
¹H NMR (500 MHz, CDCl₃) spectrum of **3bh**



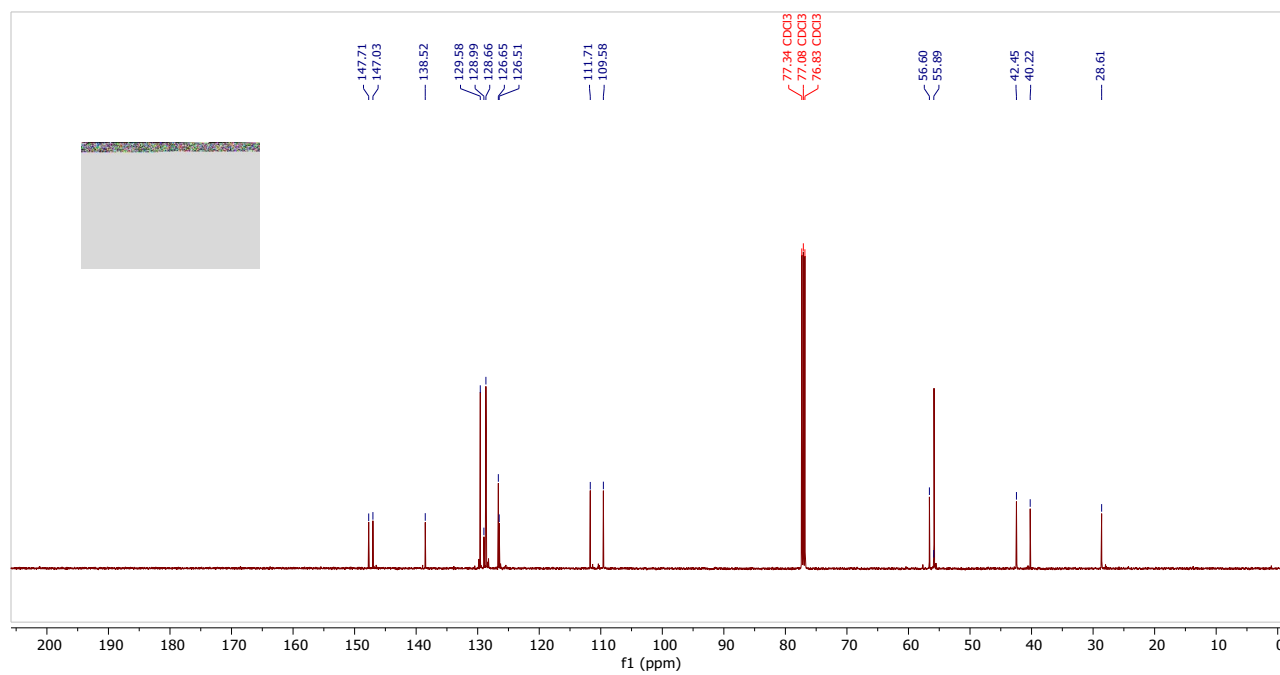
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3bh**



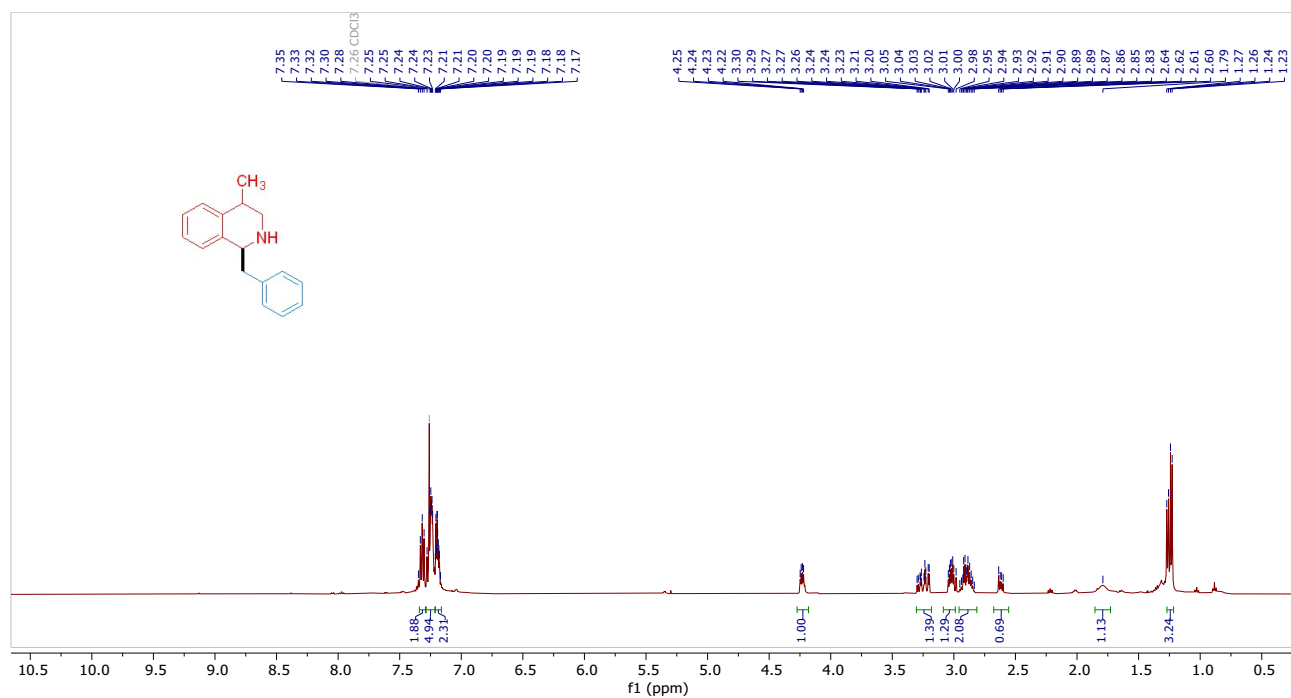
^1H NMR (500 MHz, CDCl_3) spectrum of **3bk**



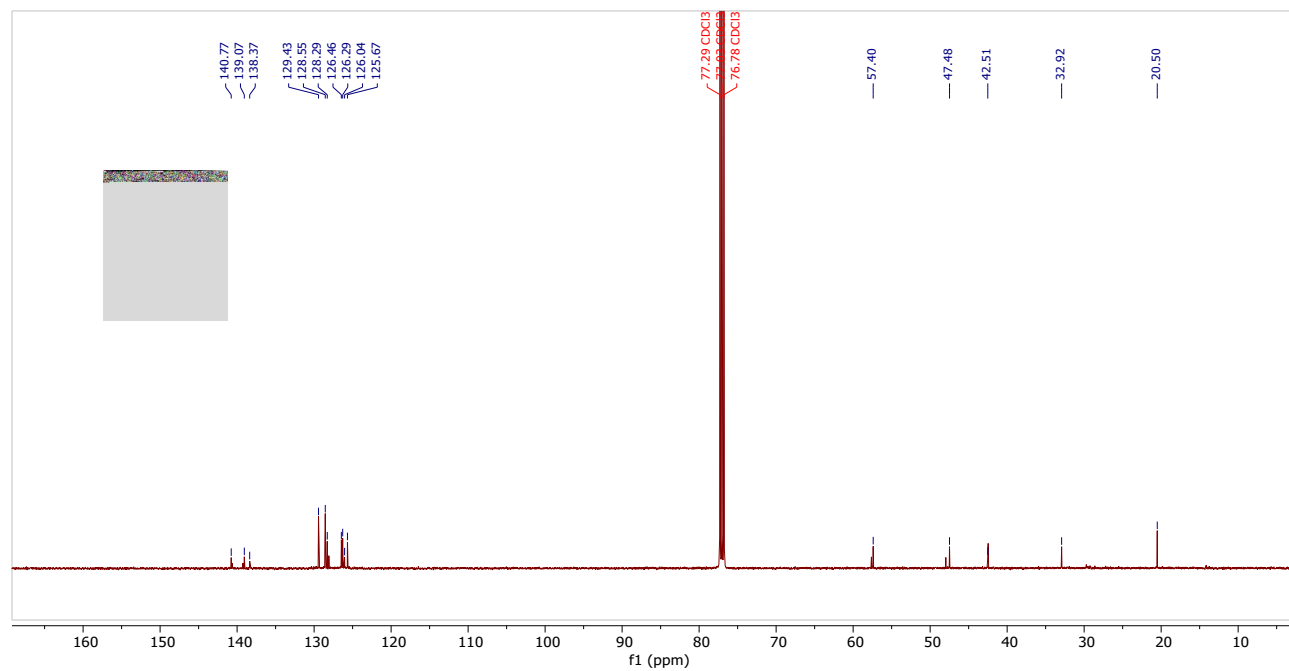
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3bk**



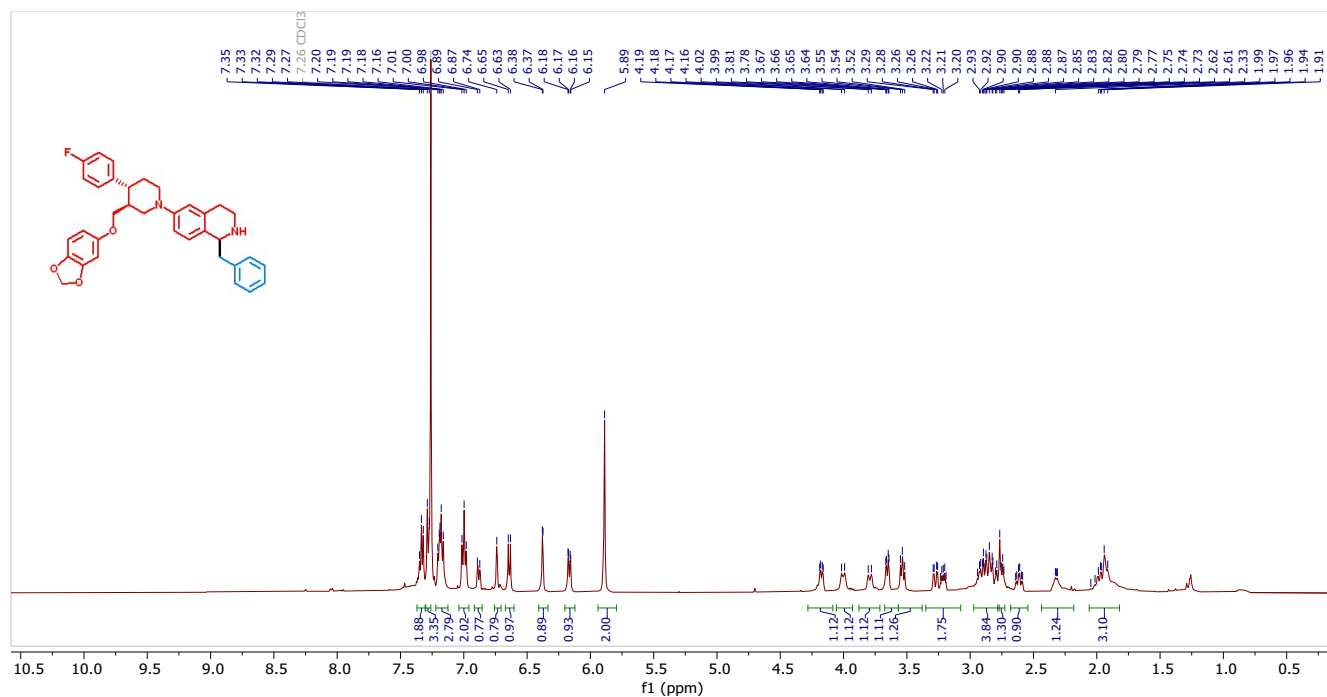
^1H NMR (500 MHz, CDCl_3) spectrum of **3bl**



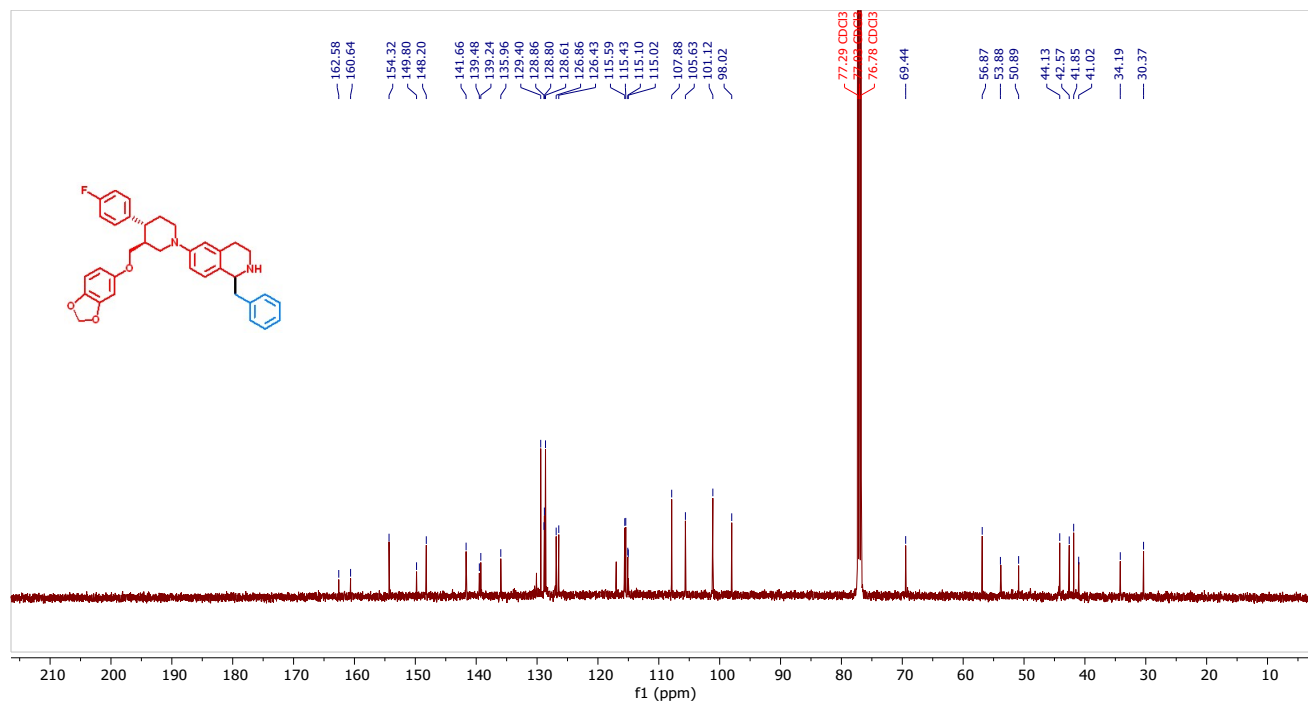
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **3bl**



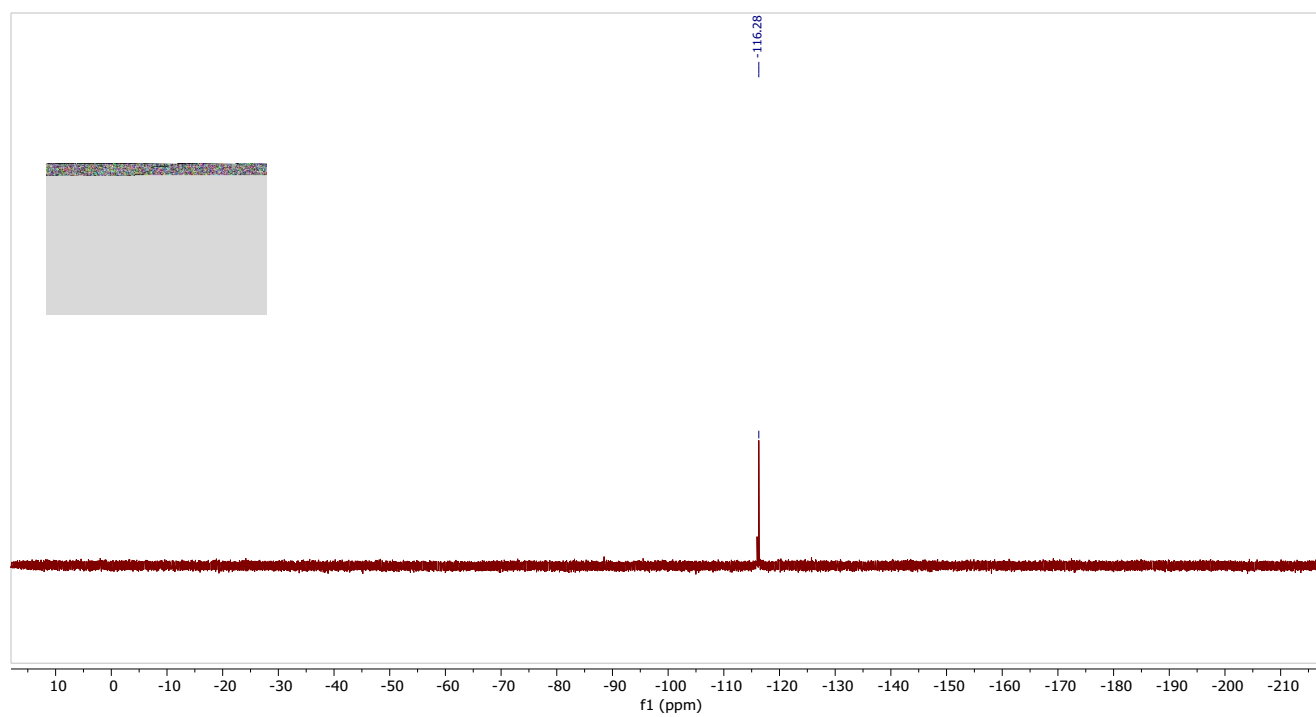
^1H NMR (500 MHz, CDCl_3) spectrum of **3bm**



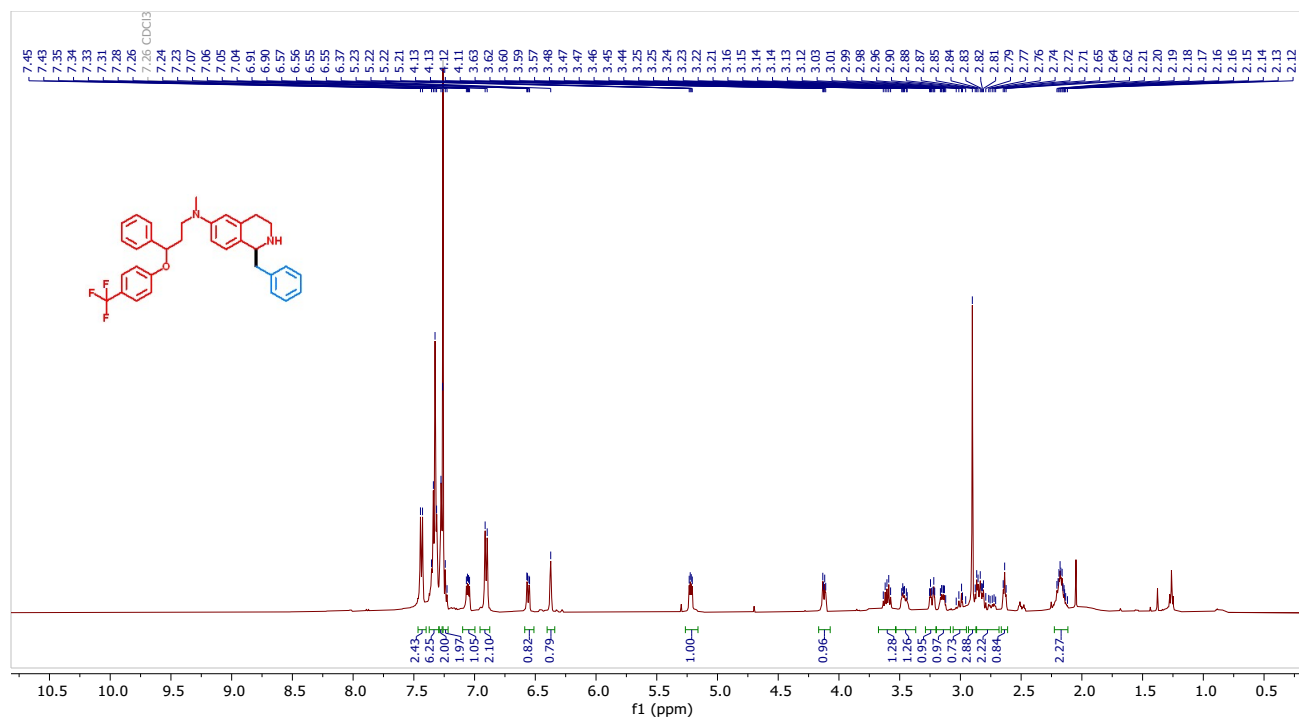
¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of **3bm**



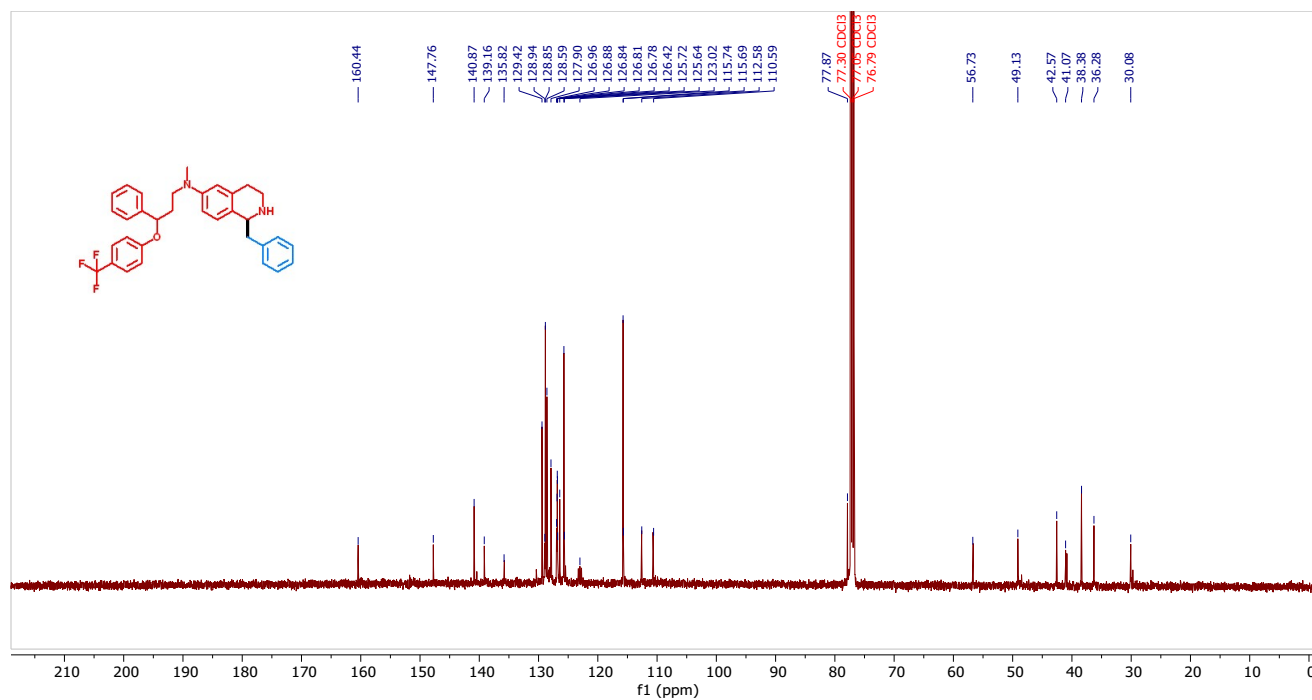
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3bm**



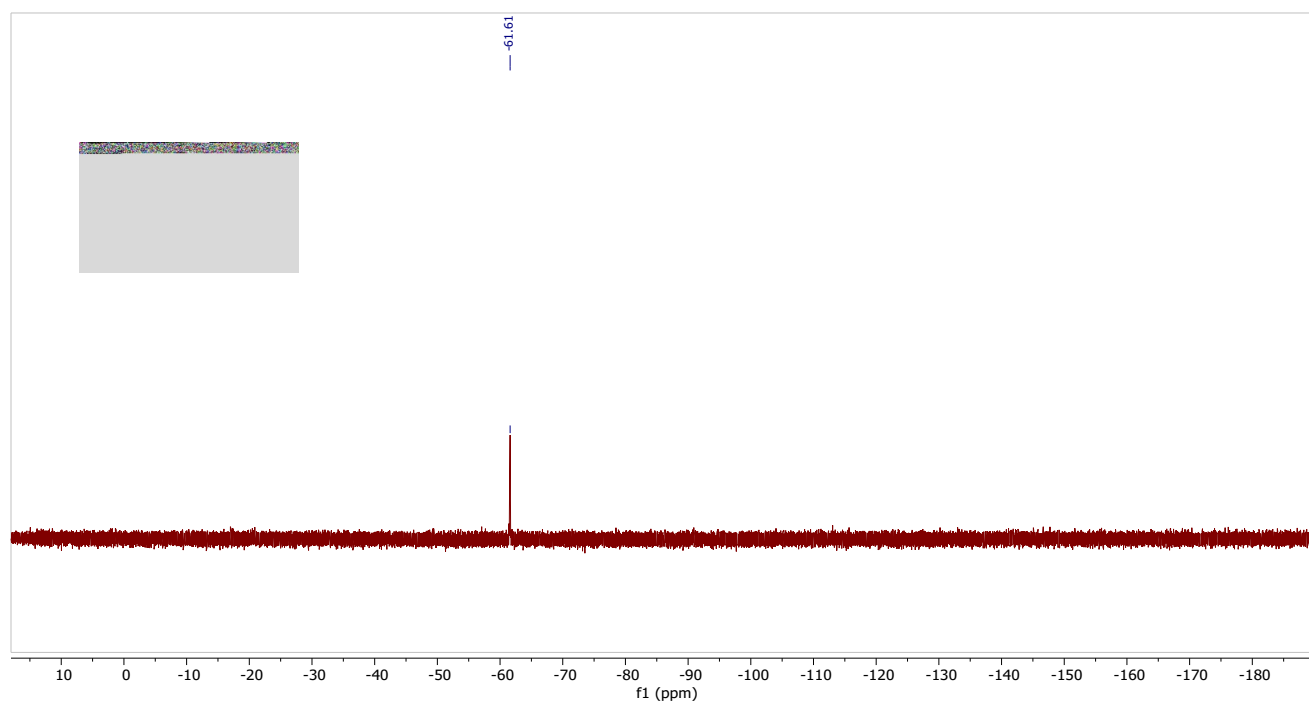
^1H NMR (500 MHz, CDCl_3) spectrum of **3bn**



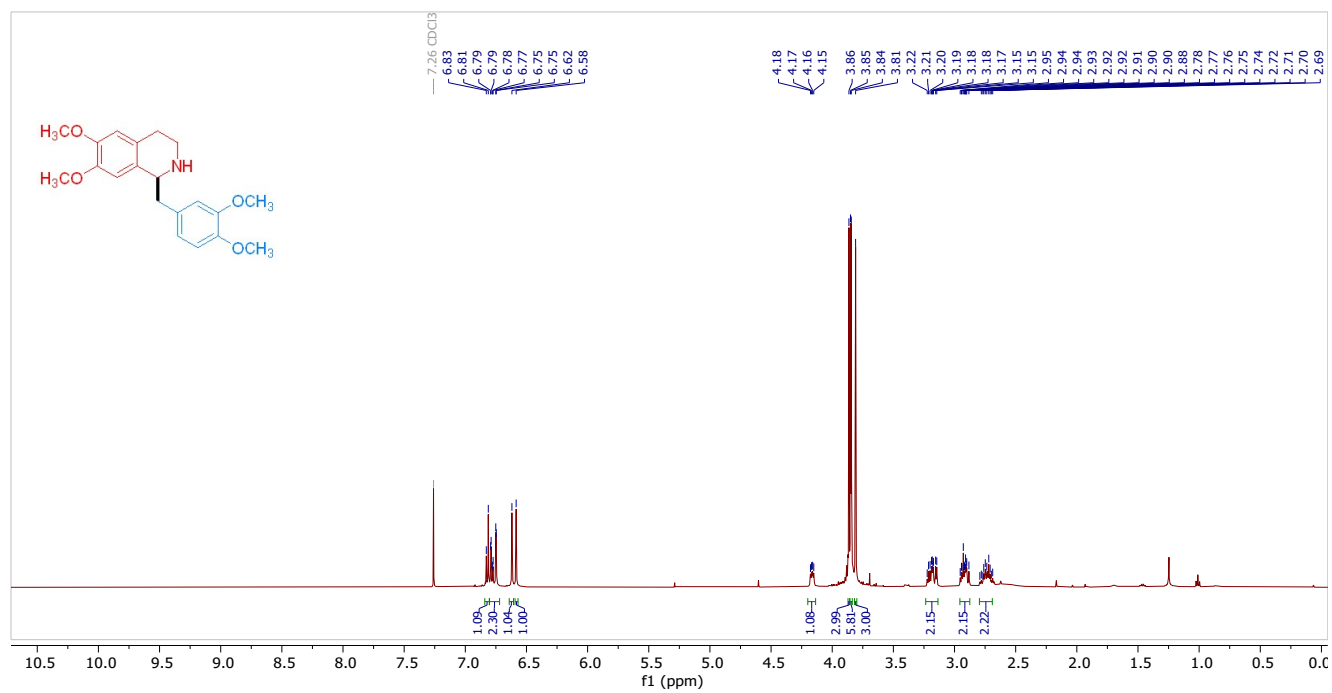
¹H NMR (500 MHz, CDCl₃) spectrum of **3bn**



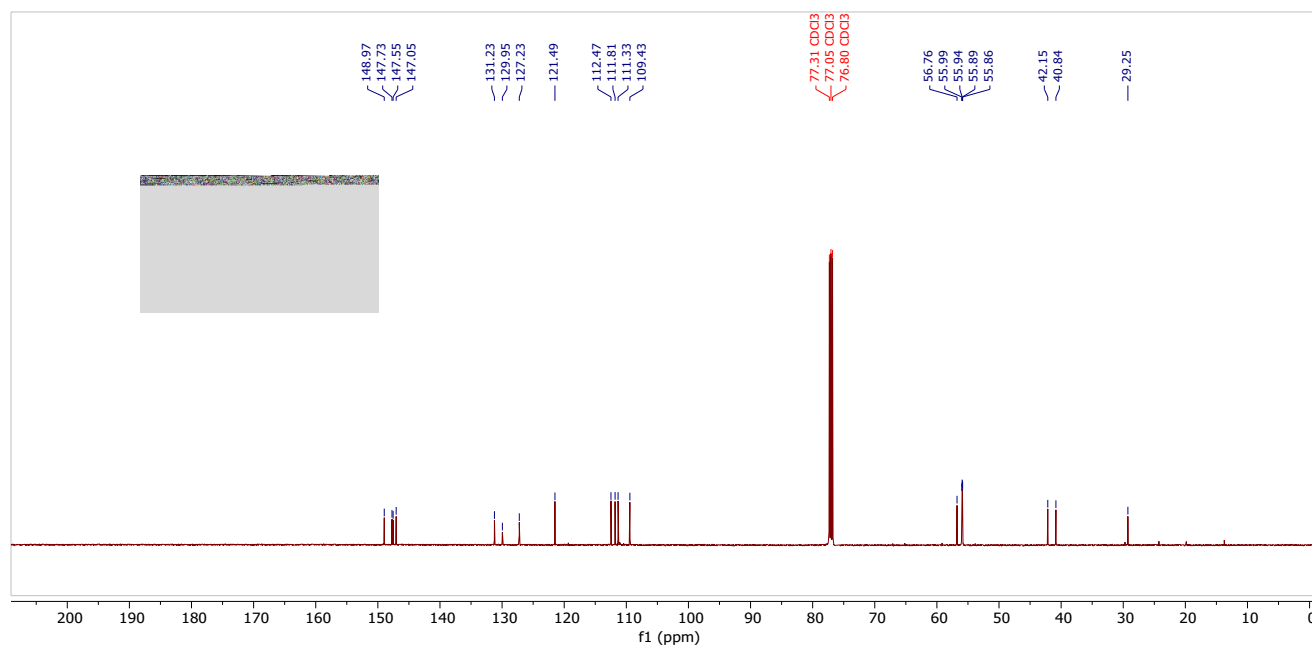
^{19}F NMR (471 MHz, CDCl_3) spectrum of **3bn**



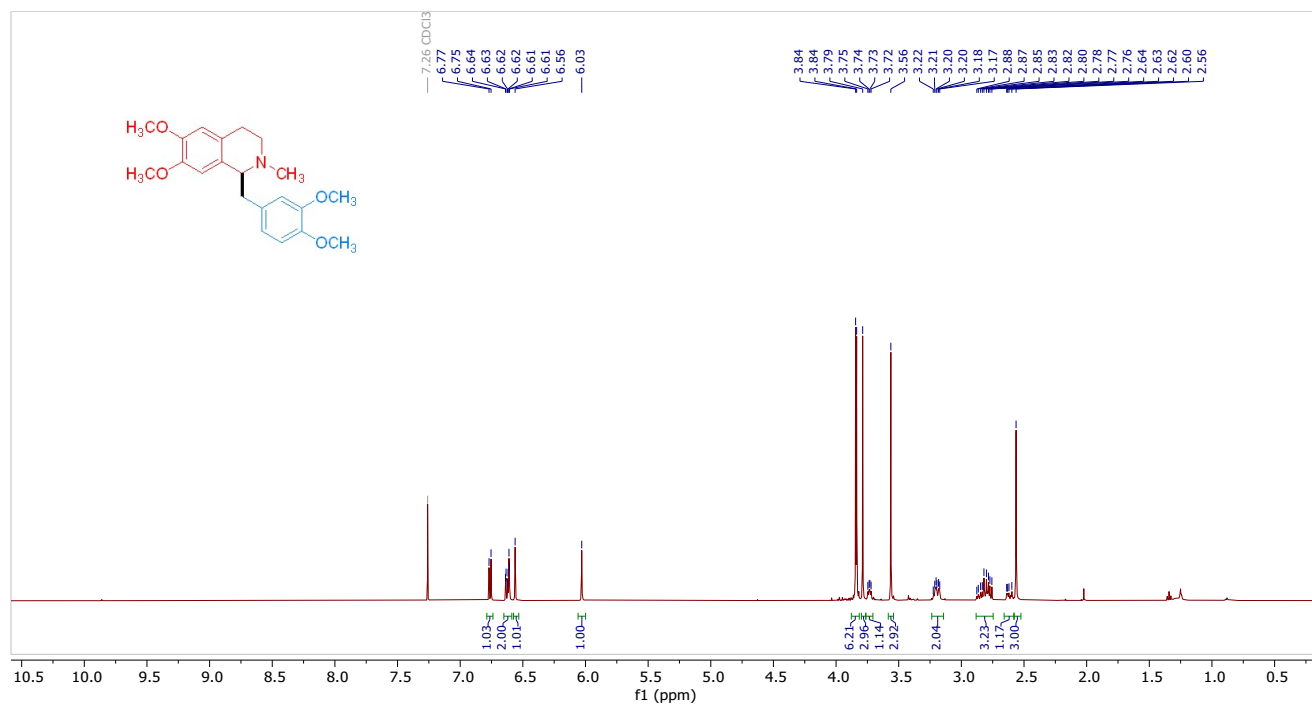
^1H NMR (500 MHz, CDCl_3) spectrum of norlaudanosine (**3bo**)



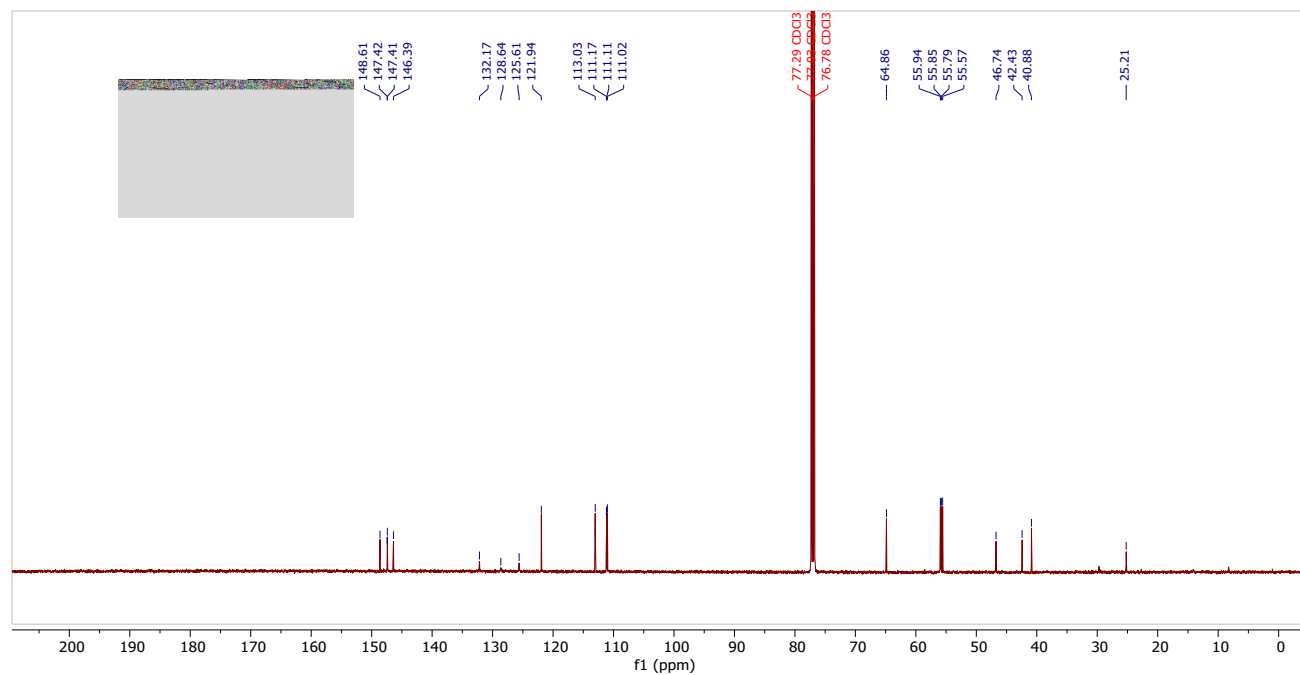
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **norlaudanosine (3bo)**



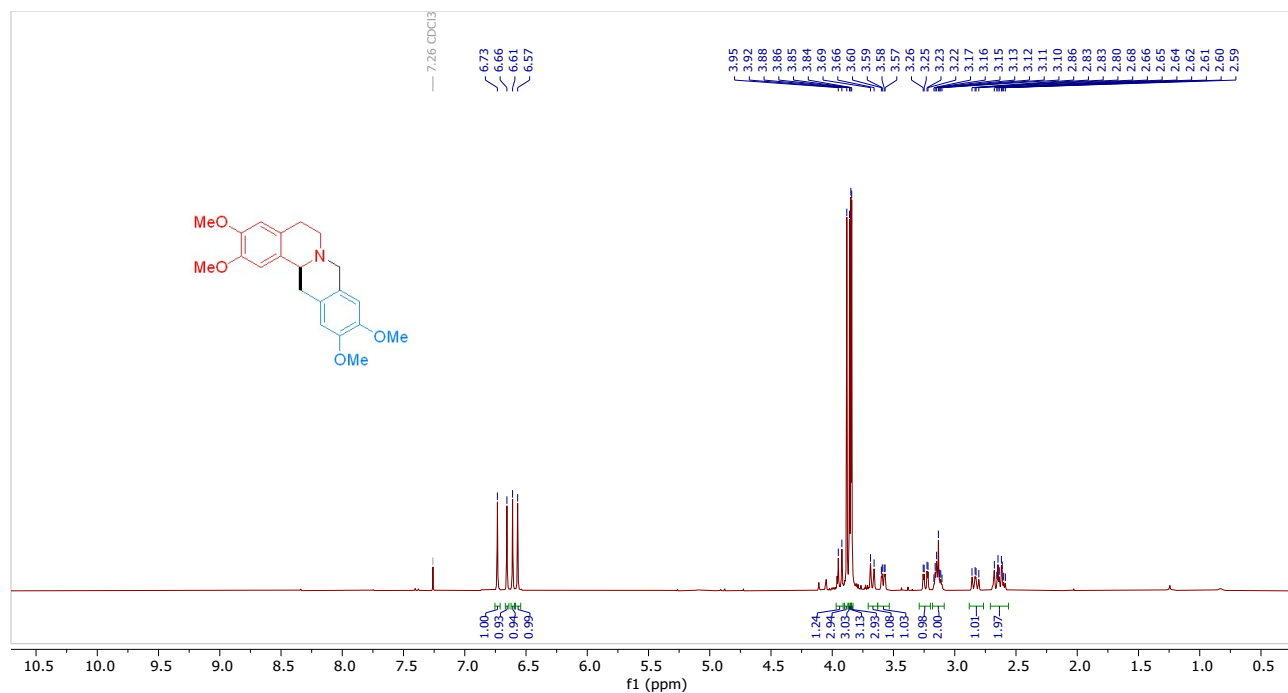
^1H NMR (500 MHz, CDCl_3) spectrum of **laudanosine (3bp)**



$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) spectrum of **laudanoline (3bp)**



^1H NMR (500 MHz, CDCl_3) spectrum of **xylopinine (3bq)**



¹³C{¹H} NMR (126 MHz, CDCl₃) spectrum of xylopinine (3bq)

