## **Supporting Information for**

# Polyphenol-mediated assembly of peptide for engineering functional materials

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#### **Experimental Section**

#### **Materials and Reagents**

Tannic acid (TA), propyl gallate (PG), ellagic acid (EA), gallic acid (GA), epigallocatechin gallate (EGCG), and catechin (CAT) were purchased from Macklin Biochemical Technology Co. Ltd (Shanghai, China). All peptides (purity  $\geq 95\%$ ) were chemically synthesized as lyophilized powders using the standard solid phase method. The peptide sequences are listed in Table S1. Branch peptide BR4G5 was synthesized by Hefei Scierbio Co., Ltd (China). All other peptides were synthesized by GenScript Biotech Co. Ltd (Nanjing China). Horseradish Peroxidase (HRP), Cytochrome C (CYC), 3,3',5,5'-tetramethylbenzidine (TMB) · streptavidin (SA) were purchased from Aladdin Reagent (Shanghai) Co., Ltd. SA-RBITC, human immunoglobulin G (IgG), antifade mounting medium (with DAPI) and biotin-labeled HRP (HRP-Bio) were purchased from Solarbio Science & Technology Co. Ltd. (Beijing, China). Amplex Red (AR), bovine serum albumin (BSA), Tween-20, fetal bovine serum (FBS), 100×penicillin-streptomycin solution, and 4% paraformaldehyde fix solution were purchased from Shanghai Beyotime Biotechnology Co., Ltd (China). AntiCD44-FITC and mAb-HRP were purchased from Sino Biological Technology Co., Ltd. (Beijing, China). Glucose oxidase (GOx) and glucose were purchased from Sigma-Aldrich (Shanghai, China). BSA-FITC, IgG-FITC, FITC coupling kit, dimethyl sulfoxide (DMSO), and DMEM medium were purchased from Shanghai Sangon Biotechnology Co., Ltd (China). NaOH, NaCl, H<sub>2</sub>O<sub>2</sub>, HCl, Na<sub>2</sub>CO<sub>3</sub>, and NaHCO<sub>3</sub> were purchased from Nanjing Chemical Reagent Co., Ltd (China). All chemicals and reagents are analytical grade and used without further purification. Ultrapure water used in all experiments was prepared by the Milli-Q system (18.2 M $\Omega$ ·cm).

#### Apparatus

Transmission electron microscopy (TEM) images were taken from JEM 1200 EX (JEOL, Japan). The UV-vis spectra were recorded using a UV-2450 spectrophotometer (SHIMADZU, Japan). The fluorescence spectra were recorded using an FL-7000 fluorescence spectrophotometer (HITACHI, Japan). Dynamic light scattering (DLS)

and zeta potential of samples were conducted on Zetasizer Nano ZSE (Malvern, UK). Optical and fluorescence microscope images were pictured by inverted fluorescence microscope IX73 (Olympus, Japan). Confocal laser scanning microscopy (CLSM) images were acquired with Zeiss LSM880.

### **Results and Discussion**



Fig. S1. Optical photographs of RR and TA alone versus a mixture of RR and TA.



Fig. S2. UV-vis absorption spectra of RR, TA, and mixture of RR and TA.



Fig. S3. AFM images of TRC and TKC.



Fig. S4. Optical microscope images of various peptides mixed with TA in PBS (scale bars are  $50 \ \mu m$ ).



Fig. S5. UV-Vis absorption spectra of TA, TA+RR at different pH conditions.



Fig. S6. Optical microscopy images (scale bars are 50  $\mu$ m) (a) and TEM images of PPAs at different pH conditions (scale bars are 200 nm).



Fig. S7. a) Optical microscopy images of TRC after resuspension in PBS, Urea, NaCl, and Tween 20 (scale bars are 50  $\mu$ m). b) UV-Vis absorption spectra of TRC, TKC, THC, and TWC after resuspension in PBS, Urea, NaCl, and Tween 20.



Fig. S8. Tyndall phenomenon of PG, EA, GA, EGCG, CAT mixed with RR at different pH conditions.



Fig. S9. UV-Vis absorption spectra of PG, EA, GA, EGCG, CAT, and their mixture with RR at different pH conditions.



Fig. S10. Optical microscope images of PG, EA, GA, EGCG, and CAT mixed with RR at different pH conditions (scale bars are  $100 \ \mu m$ ).



Fig. S11. Tyndall phenomenon of various peptides mixed with TA at various pH conditions.



Fig. S12. Optical microscope images of assemblies of various peptides and TA (scale bars are  $100 \ \mu m$ ).



Fig. S13. Optical microscope image of TA mixed with RR in each ratio (scale bars are  $100 \ \mu m$ ).



Fig. S14. Fluorescence microscopy images of PPAs@IgG-FITC (scale bars are  $100 \ \mu m$ ).



Fig. S15. Size distribution of PPAs@IgG.



Fig. S16. EDS mapping of PPA@IgG (scale bars are 50 nm).



Fig. S17. Potential of PPAs and PPAs@IgG.



Fig. S18. a) Optimization of time for PPAs loading proteins. b) Optimization of the number of washes for PPAs loading proteins.



Fig. S19. Stability of proteins loaded on PPAs.



Fig. S20. The ability of PPAs to load IgG-FITC.



Fig. S21. Fluorescence microscopy images of various proteins loaded on PPAs (scale bars are  $100 \ \mu m$ ).



Fig. S22. Enzyme kinetic curve of PPAs@HRP.



Fig. S23. Stability of mAb-HRP bound on PPAs@IgG.



Fig. S24. Relationship between the amount of Bio-HRP bound on PPA@SA and its concentration.



Fig. S25. UV-Vis absorption spectra of TA and P1, P2 mixing with TA. Inset: Tyndall phenomenon of P1, P2 mixed with TA.



Fig. S26. Fluorescence microscopy images and fluorescence intensities of MDA-MB-231 after incubation 3 h with PPAs@AntiCD44-FITC@BSA, PPAs@IgG-FITC@BSA (scale bars are 50 μm).



Fig. S27. Cell viability of MDA-MB-231 cells treated with PPAs@BSA for different times.

Name	Sequence (N-terminal to C-terminal)		
RR/R2	RRGGGRR		
KK	KKGGGKK		
HH	HHGGGHH		
DD	DDGGGDD		
PP	PPGGGPP		
VV	VVGGGVV		
WW	WWGGGWW		
YY	YYGGGYY		
PP	PPGGGPP		
VV	VVGGGVV		
AA	AAGGGAA		
R1	RGGGR		
R3	RRRGGGRRR		
G5	GGGGG		
R2G5	RRGGGGG		
R3G5	RRRGGGGG		
R4G5	RRRRGGGGG		
BR4G5	(RR) <sub>2</sub> KGGGGG		
P1	KTCENLADTY		
P2	KTCENLADTYRRR		
P3	ACSAG		
P4	ACSAGRRR		

Table S1. Sequences of all peptides used in this work.

Name	Molecular Weight	Isoelectric Point (pI)	Aliphatic Index
	(kDa)		
GOx	131.28	4.2	86.12
Insulin	5.741	5.3	80.2
GFP	26.886	5.8	74.87
HRP	38.825	7.2	90.34
IgG	144.263	8.5	67.64
CYC	11.703	9.4	58.57
BSA	69.293	4.6	77.46
SA	65	6	63.4

Table S2. Molecular weight, isoelectric point, and aliphatic index of different proteins