

Supplementary Information

“A perfect pair”: Extracellular vesicle-based novel strategies for cancer precise diagnosis and effective treatment

Hongxin Zhao, ‡ Fangxue Du, ‡ Ruiqian Guo, Li Qiu*

Department of Department of Ultrasound, West China Hospital, Sichuan University,
Chengdu 610041, China

Corresponding author: qiulihx@scu.edu.cn (Li Qiu).

‡ Contributed equally.

Table. S1 Pros and cons of extracellular-vesicle extraction/isolation

Separation methods	Principle	Pros	cons	References
Differential ultracentrifugation	Steps of differential centrifugation	Simple operation; Suitable for vesicle collection of nature large samples	Low degree of purity and time-consuming	¹
Density-gradient centrifugation	Employ prefabricated density gradient	The operation is simple; The purity is high, cross-contamination is avoided and the integrity of the sample is maintained	The operation is complex; Has a low yield	²
Size exclusion chromatography	Comprises ultrafiltration and SEC	Large and rapid collections;	Time-consuming; With low	³

		Low cost; yield and low purity approaches	
Affinity capture	Utilize antibodies for specific seizure	High specificity along with high purity	The yield is low; The cost is high; The operation is complex; And only EVs with specific markers can be separated
Ultrafiltration	The resulting EV particle size	Simple to operate; Economical distribution is and determined by the pore size of the membrane	The structure of EVs is prone to change; With uniform low purity; And the ultrafiltration membrane is liable to be blocked
Precipitation	Employ PEG to lower the solubility of EVs	Simple; Rapid; to lower the and highly purity	Low level of productivity

Table. S2 Advantages and disadvantages of drug loading approaches for EVs.

Drug loading	Suitable	Advantages	Disadvantage	Reference
--------------	----------	------------	--------------	-----------

methods	cargoes	s	s	
Electroporation	Small molecule medications; siRNAs; Nanoparticles	Universality; High loading efficacy; Simple and convenient	High voltage pulses undermine the integrity of EV membranes; Low output	⁸
Sonication	Small-molecule medications; Nanoparticulate s	Simple and convenient; And suitable for loading hydrophilic drug molecules	Damage to the integrity of EVs; It could undermine the integrity of EV membranes or macromolecular proteins	⁹
Freeze and thaw cycles	Liposomes loaded with drugs; Proteins; RNAs	High yield; High loading efficiency; High encapsulation efficiency	Complex operation; Damage to EVs	¹⁰
Saponin treatment	Drugs with hydrophilic properties	Mild; efficiency of drug loading is enhanced	The Toxicity; Additional purification	¹¹
Transfection and transduction	Proteins; RNAs	Preserve the integrity of EV membranes;	High technical complexity; High expense	¹²

Facilitate access					
Incubation	Small molecule drugs; Nanomaterials	Easy accessibility; Universality; Simple and convenient,	Time-consuming; Cytotoxicity exists for the parent cells		13

Table. S3 Clinical Trials of EVs in Cancer Diagnosis and Monitoring

Cancer type	Year	Enrollment	Sample Type	Study Design	Purpose	Treatment	Detection	Detection	Status
							targ	meth	
							et	od	
Retinoblastoma (NCT04164134)	2018	378	Blood	Observational	Monitoring	N/A	EV RNA ,	N/A	Completed
Lymphomas and Lymphoproliferative Disorders (NCT06782854)	2022	102	Plasma and mononuclear cells	N/A	N/A	N/A	N/A	N/A	Completed

References

1. N. Zarovni, A. Corrado, P. Guazzi, D. Zocco, E. Lari, G. Radano, J. Muhhina, C. Fondelli, J. Gavrilova and A. Chiesi, *Methods*, 2015, **87**, 46-58.
2. L. Paolini, A. Zendrini, G. D. Noto, S. Busatto, E. Lottini, A. Radeghieri, A. Dossi, A. Caneschi, D. Ricotta and P. Bergese, *Scientific Reports*, 2016, **6**, 23550.
3. R. Stranska, L. Gysbrechts, J. Wouters, P. Vermeersch, K. Bloch, D. Dierickx, G. Andrei and R. Snoeck, *Journal of Translational Medicine*, 2018, **16**, 1.
4. Y. Zhang, Y. Liu, H. Liu and W. H. Tang, *Cell & Bioscience*, 2019, **9**, 19.

5. D. Yang, W. Zhang, H. Zhang, F. Zhang, L. Chen, L. Ma, L. M. Larcher, S. Chen, N. Liu, Q. Zhao, P. H. L. Tran, C. Chen, R. N. Veedu and T. Wang, *Theranostics*, 2020, **10**, 3684-3707.
6. K. Kumar, E. Kim, M. Alhammadi, U. Reddicherla, S. Aliya, J. N. Tiwari, H. S. Park, J. H. Choi, C. Y. Son, A. T. E. Vilian, Y.-K. Han, J. Bu and Y. S. Huh, *TrAC Trends in Analytical Chemistry*, 2023, **159**, 116912.
7. T. Soares Martins, J. Catita, I. Martins Rosa, O. A. B. da Cruz e Silva and A. G. Henriques, *PLOS ONE*, 2018, **13**, e0198820.
8. R. Tenchov, J. M. Sasso, X. Wang, W.-S. Liaw, C.-A. Chen and Q. A. Zhou, *ACS Nano*, 2022, **16**, 17802-17846.
9. R. Kar, R. Dhar, S. Mukherjee, S. Nag, S. Gorai, N. Mukerjee, D. Mukherjee, R. Vatsa, M. Chandrakanth Jadhav, A. Ghosh, A. Devi, A. Krishnan and N. D. Thorat, *ACS Biomaterials Science & Engineering*, 2023, **9**, 577-594.
10. J. Wang, D. Chen and E. A. Ho, *Journal of Controlled Release*, 2021, **329**, 894-906.
11. M. J. Haney, N. L. Klyachko, Y. Zhao, R. Gupta, E. G. Plotnikova, Z. He, T. Patel, A. Piroyan, M. Sokolsky, A. V. Kabanov and E. V. Batrakova, *Journal of Controlled Release*, 2015, **207**, 18-30.
12. L. Cheng, K. Zhang, S. Wu, M. Cui and T. Xu, *Stem Cells International*, 2017, **2017**, 6305295.
13. I. K. Herrmann, M. J. A. Wood and G. Fuhrmann, *Nature Nanotechnology*, 2021, **16**, 748-759.