

Next-Generation Fluorescent Chemosensor for Dual-Mode Detection of Aluminium Ions and DCP: A Bridge Between Sensing and Bio-imaging

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Materials and methods

Binding constant and limit of detection

The binding constants were determined with Benesi-Hildebrand equation, and the limit of detections were calculated with the help of a standardized IUPAC equation.

$$\frac{1}{I-I_o} = \frac{1}{K_a (I_{\max} - I_o)[C]^n} + \frac{1}{I_{\max} - I_o}$$

Where,

I_o = Intensity of probe in the absence of analyte

I = Intensity of probe at an intermediate analyte concentration

I_{\max} = Intensity of probe at a saturation

C = Concentration of analyte,

K_a = Binding constant

The detection limit (DL) was calculated from the following equation:

$$DL = \frac{3 \times \text{standard deviation of blank solution}}{\text{slope of calibration curve}}$$

Quantum Yield

We have calculated the fluorescence quantum yield of **BINNA** in the absence and presence of Al^{3+} ions and DCP using fluorescein as a standard, with a quantum yield of 0.54 (0.1M H_2SO_4) and phenanthrene with a quantum yield of 0.13 (cyclohexane). The formula used is as follows

$$\Phi_S = \Phi_R \times \frac{A_S}{A_R} \times \frac{(Abs)_R}{(Abs)_S} \times \frac{\eta_S}{\eta_R}$$

Where Φ_S is the quantum yield of sample to be tested, Φ_R is the quantum yield of the fluorescein; A_S and A_R are the emission band areas of tested sample and fluorescein, $(Abs)_S$ and $(Abs)_R$ are the maximum absorbance values of the tested sample and fluorescein; η_S and η_R are the refractive index of the solvent used for the sample and fluorescein to record emission band.

Sample preparation for DLS studies

The prepared stock solution was filtered to remove the suspended particles with the help of 0.02 μ M filter. Zeta potential analyser (ZEN 3600) was used for conducting DLS experiment. The solution having a concentration of 20 μ M was prepared to determine the hydrodynamic size of the particles.

Field Emission-Scanning Electron Microscopy (FE-SEM) measurement

The effect of aggregation has been explored by measuring the changes in particle size and surface morphology using FE-SEM imaging. The **BINNA** solution was placed on a cover slip by drop cast method and allowed to dry for 24 h. The dried sample was subsequently plated with gold spray and FE-SEM analysis was performed. ZEISS MERLIN Compact FE-SEM was used to perform the experiment.

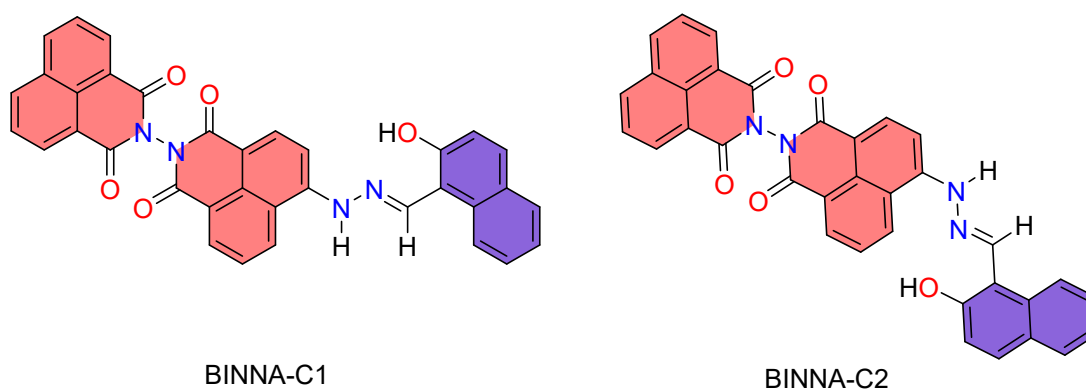


Figure S1: Possible conformations of probe **BINNA**

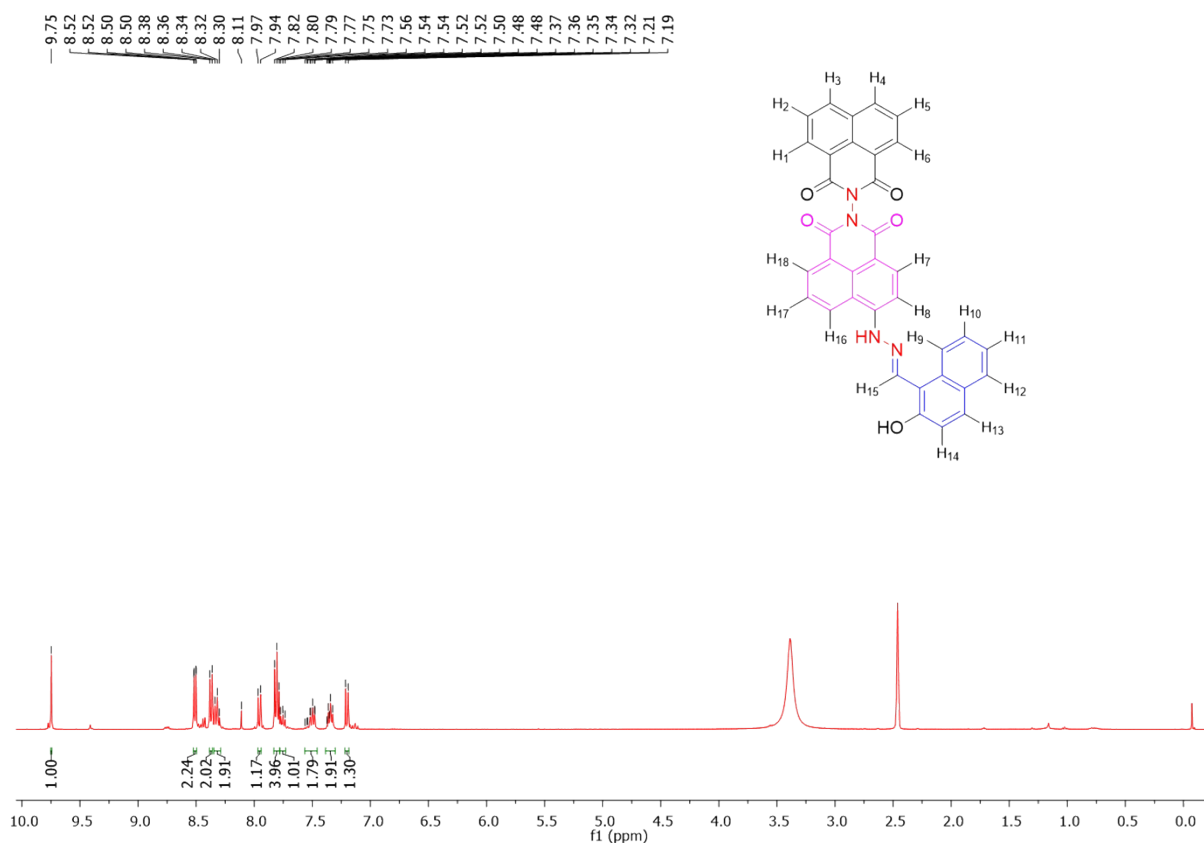


Figure S2: ¹H NMR spectrum (DMSO-*d*₆) of (*E*)-6-(2-((2-hydroxynaphthalen-1-yl)methylene)hydrazinyl)-1*H*,1'*H*,3*H*,3'*H*-[2,2'-bibenzo[*de*]isoquinoline]-1,1',3,3'-tetraone

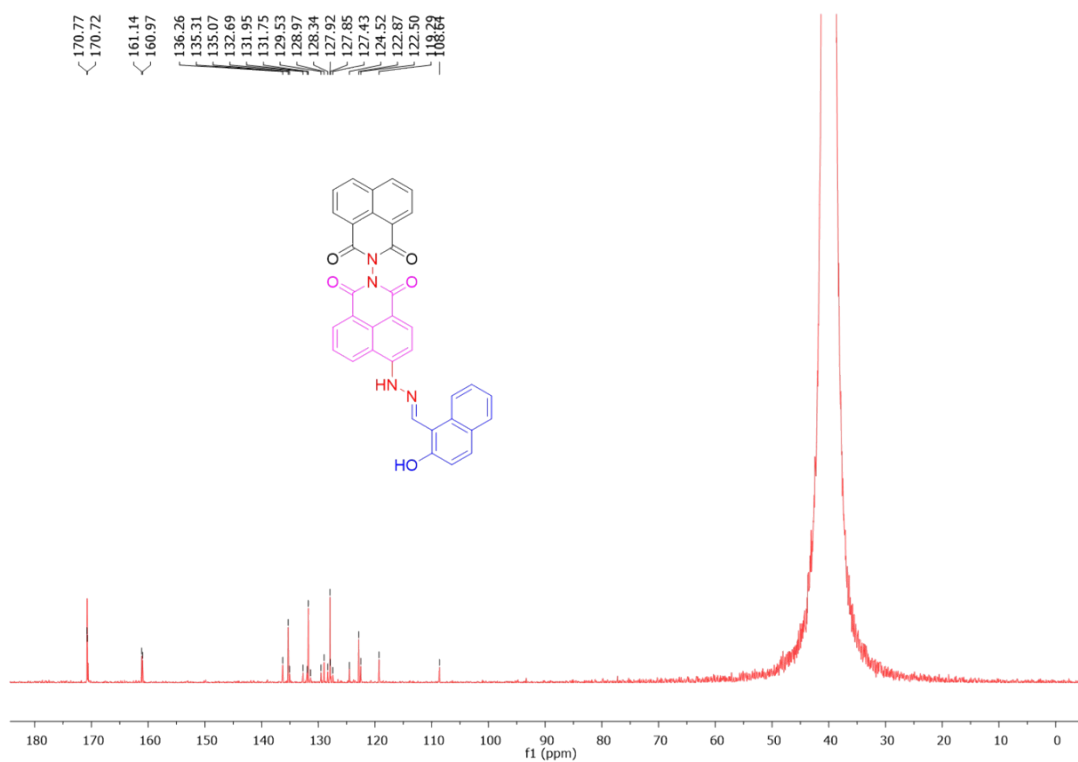


Figure S3: ¹³C NMR spectrum (DMSO-*d*₆) of (*E*)-6-(2-((2-hydroxynaphthalen-1-yl)methylene)hydrazinyl)-1*H*,1'*H*,3*H*,3'*H*-[2,2'-bibenzo[*de*]isoquinoline]-1,1',3,3'-tetraone

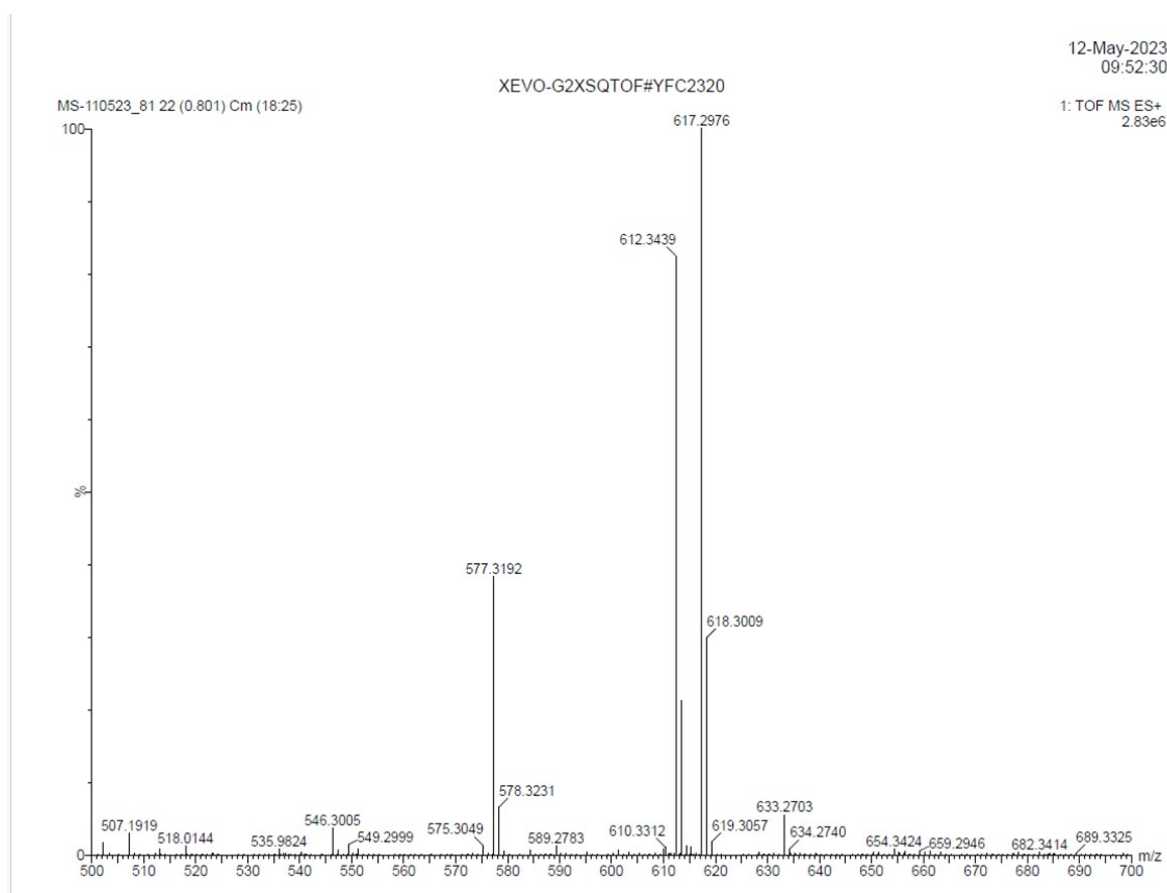
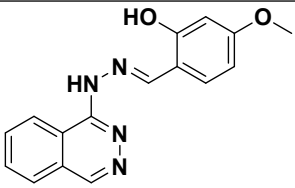
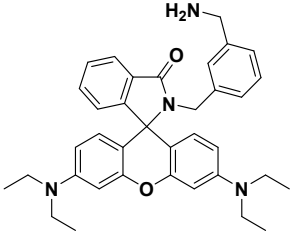
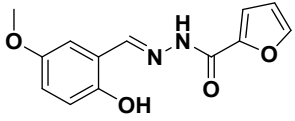
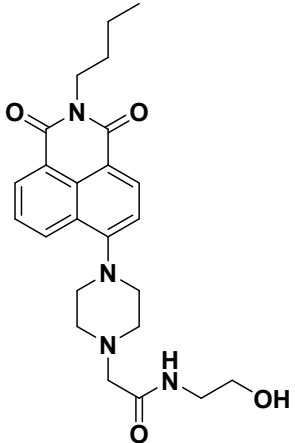


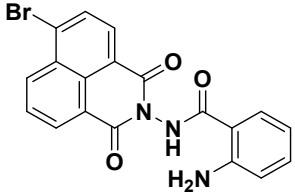
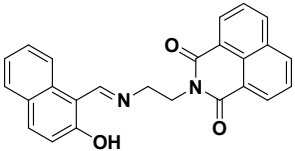
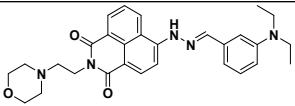
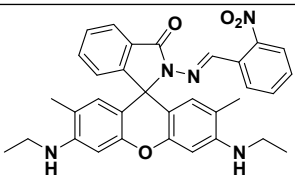
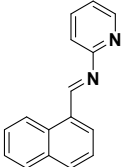
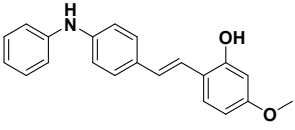
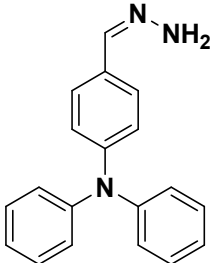
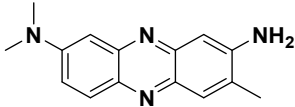
Figure S4: HRMS spectrum of BINNA

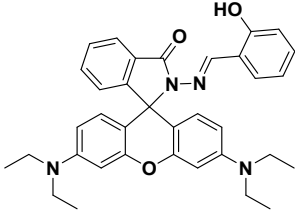
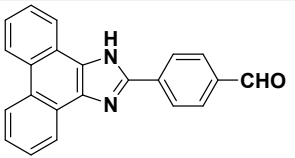
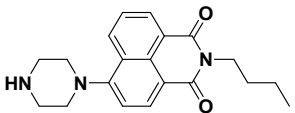
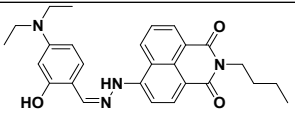
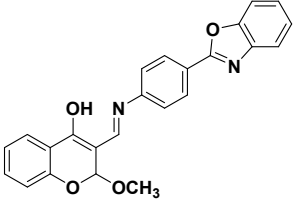
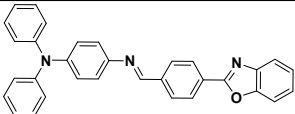
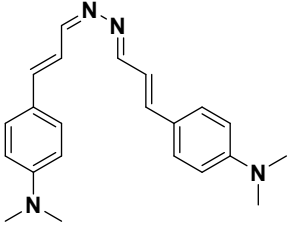
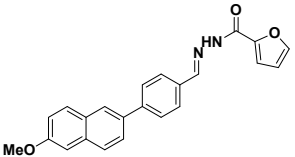
Efficiency of BINNA

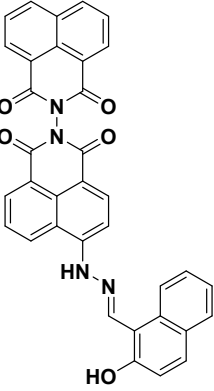
BINNA effectiveness and efficacy in detecting Al^{3+} ions and DCP were contrasted with those of previously published Al^{3+} and DCP sensors (**Table S1**). Based on this data, it is evident that **BINNA** offers a number of benefits over the reported sensors, as it can detect both Al^{3+} and DCP, including a "Turn-On" sensor with better detection limits compared to other reported chemosensors.

Table S1: Comparison of the present sensor with known sensors

S. No.	Sensor	Solvent System	Ion Detected	LOD	Application	Ref
1		H_2O	Al^{3+}	0.022 μM	Flexible and foldable film	1
2		$\text{C}_2\text{H}_5\text{OH}:\text{H}_2\text{O}$ (7:3)	Al^{3+}	0.098 μM	Test strip and Logic gates	2
3		HEPES buffer	Al^{3+}	0.082 μM	Test strip	3
4		MeOH	Al^{3+}	0.15 μM	Test strip and logic gates	4

5		DMSO:H ₂ O (1:1 v/v)	Al ³⁺	15.2 μM	Cell imaging	5
6		DMSO:H ₂ O (1:1 v/v)	Al ³⁺	0.033 μM	Pharmaceutical samples	6
7		MeOH	Al ³⁺	0.79 μM	Cell imaging and smartphone	7
8		CH ₃ CN:HEPES (1:1 v/v)	Al ³⁺	1.88 μM	Cell imaging	8
9		THF:H ₂ O (1:1 v/v)	Al ³⁺	3.15 μM	Real Sample analysis and bioimaging	9
10		MeOH: HEPES (6:4)	Al ³⁺	12.6 μM	Smartphone and test kit	10
11		CHCl ₃	DCP	23.31 nM	Test strip and Bioimaging	11
12		CH ₃ CN	DCP	0.36 μM	Real Sample analysis	12

13		DMSO:H ₂ O (6:4)	DCP	0.16 μM	Paper strip, smartphone analysis	13
14		DMSO	DCP	7.4 μM	Smartphone and paper strip	14
15		DMF	DCP	5.5 nM	Paper strip	15
16		CH ₃ CN	DCP	1.54 μM	Paper strip	16
17		DMSO:H ₂ O (1:1)	DCP	0.36 μM	Paper strip	17
18		DMSO	DCP	0.42 μM	Paper strip and Real sample analysis	18
19		CH ₃ CN	DCP	0.3 μM	Test strip and real sample analysis	19
20		CH ₃ CN:H ₂ O (9:1)	DCP	12.2 nM	Paper Strip	20

21		MeOH	Al ³⁺ , DCP	18.2 nM (0.0182 μM) and 8.5 nM (0.008 μM)	Cell Imaging, Logic gate, Real sample analysis	This work
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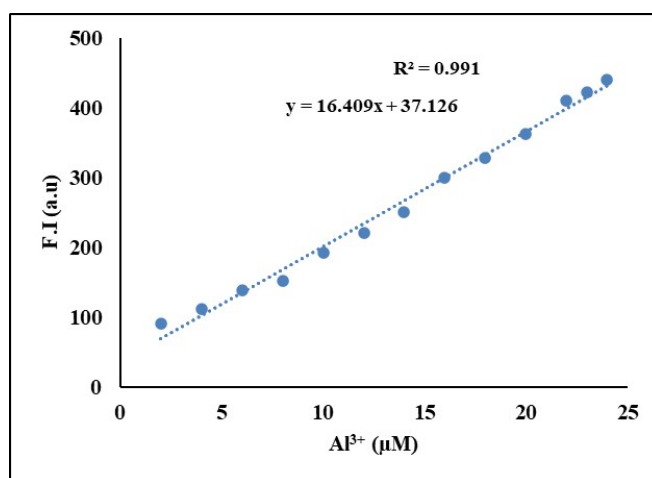


Figure S5: Determination of limit of detection for **BINNA** towards Al³⁺ ions.

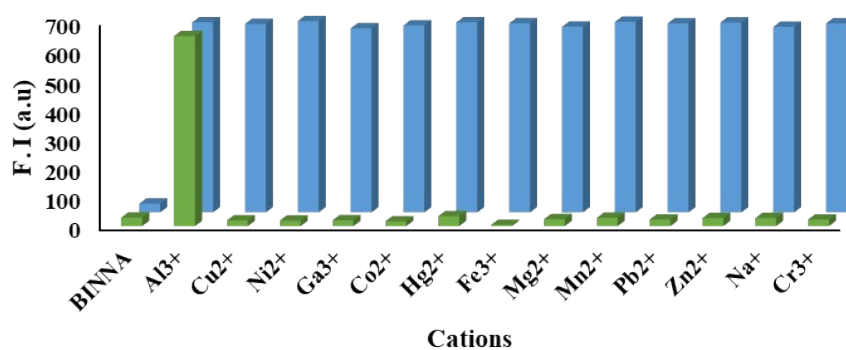


Figure S6: The relative intensity of **BINNA** at 450 nm in the presence and absence of Al³⁺ ions (10 μM), while considering various competing metal ions (50 μM). The change in emission intensity of **BINNA** with different metal ions was represented by green bars, whereas blue bars represent the changes in emission intensity of **BINNA** in the presence of Al³⁺ ions and different competing ions

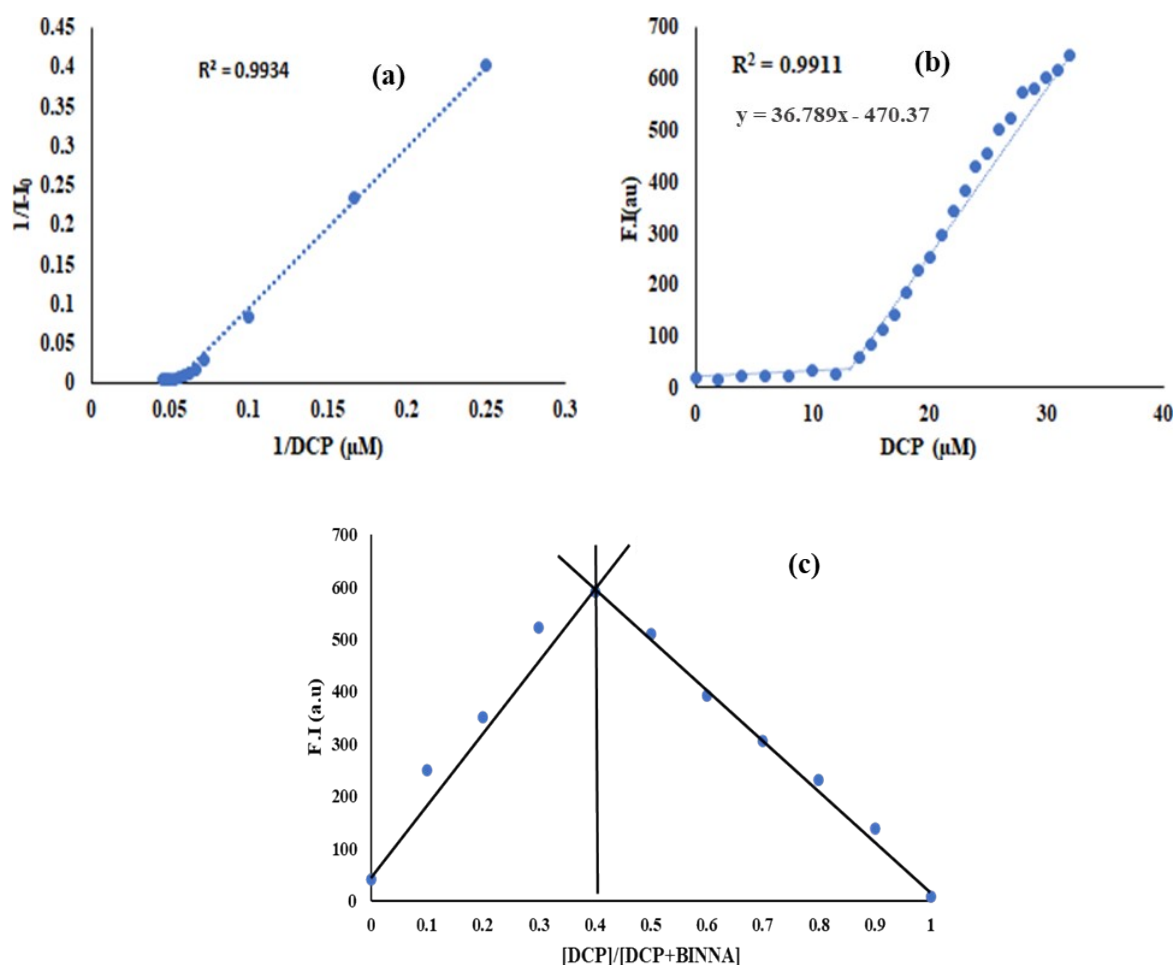


Figure S7: (a) Calculation of binding constant for **BINNA** towards DCP using the Benesi-Hildebrand equation, (b) Determination of limit of detection for **BINNA** towards DCP ions. (c) Job's plot for calculating the stoichiometry of **BINNA** and DCP using fluorescence spectra.

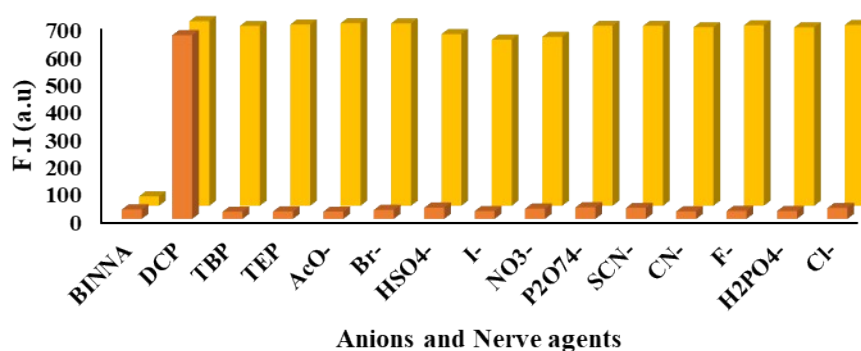


Figure S8: Relative intensity of **BINNA** at 371 nm in the presence and absence of DCP (10 μM) with several anions (50 μM). The fluorescence intensity change of **BINNA** with different anions and nerve agents is represented by orange bars, and yellow bars represent **BINNA**-DCP in the presence of different competing ions and nerve agents.

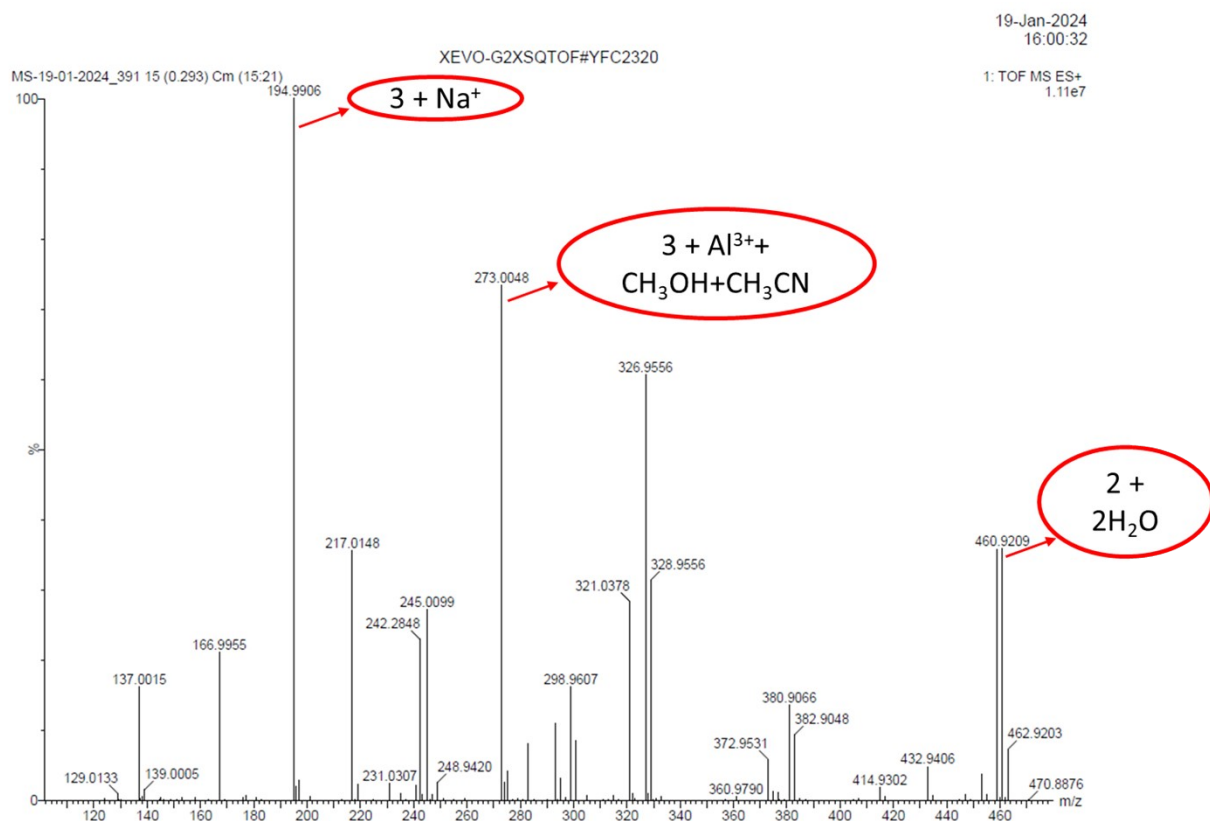


Figure S9: HRMS spectrum of **BINNA** in the presence of Al^{3+} ions

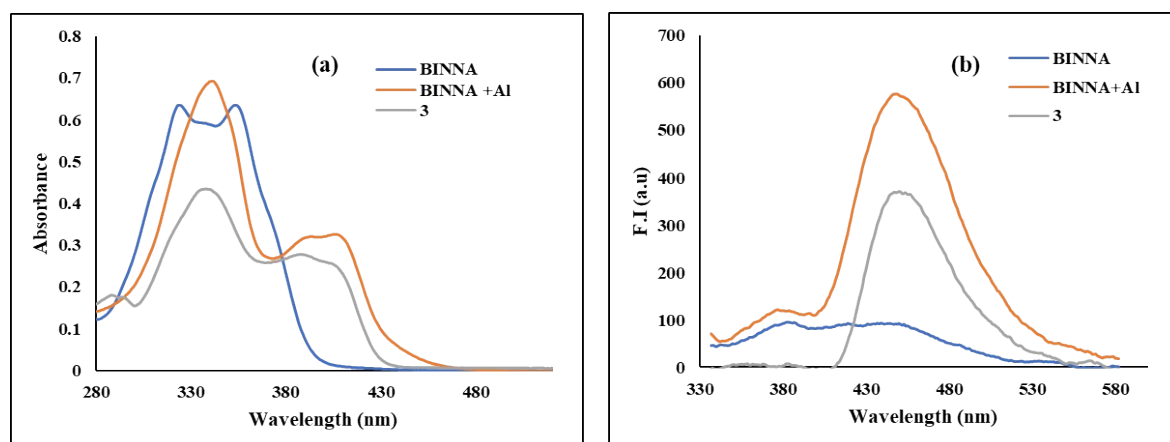


Figure S10: (a) Absorption and (b) emission spectra of **BINNA**, **BINNA**+ Al^{3+} and **3**

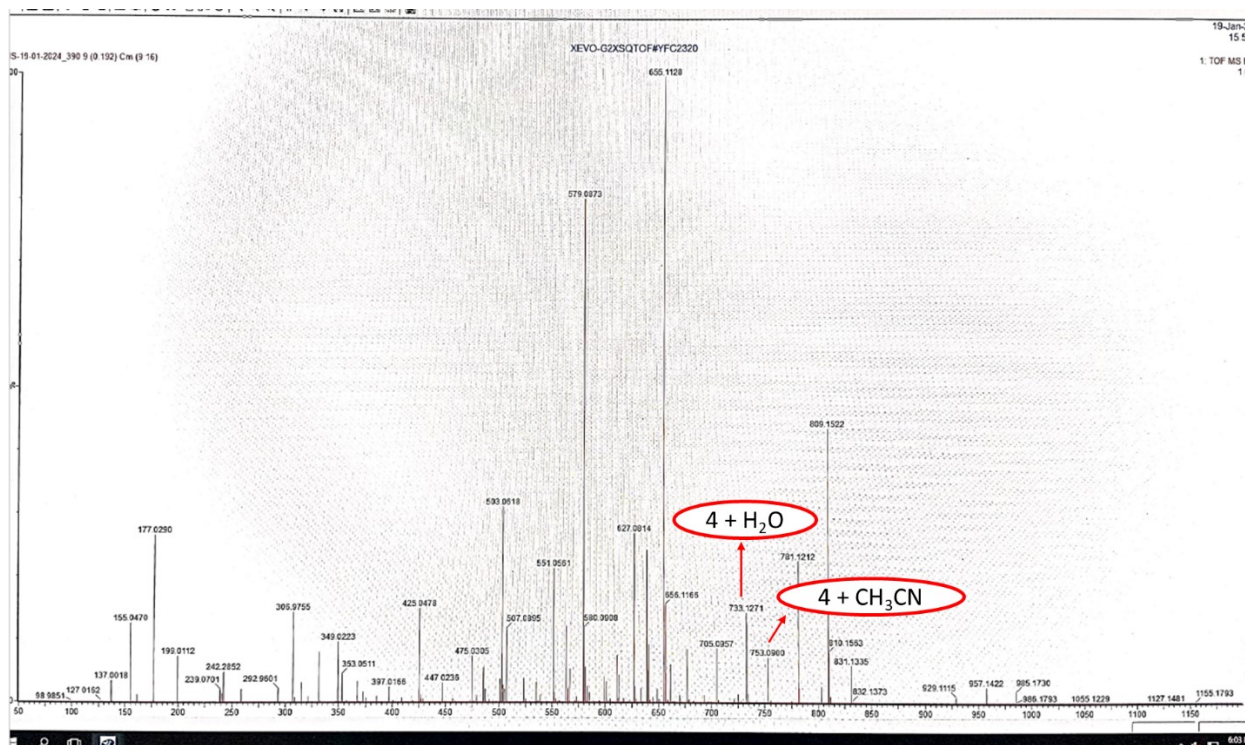


Figure S11: HRMS spectrum of **BINNA** in the presence of DCP

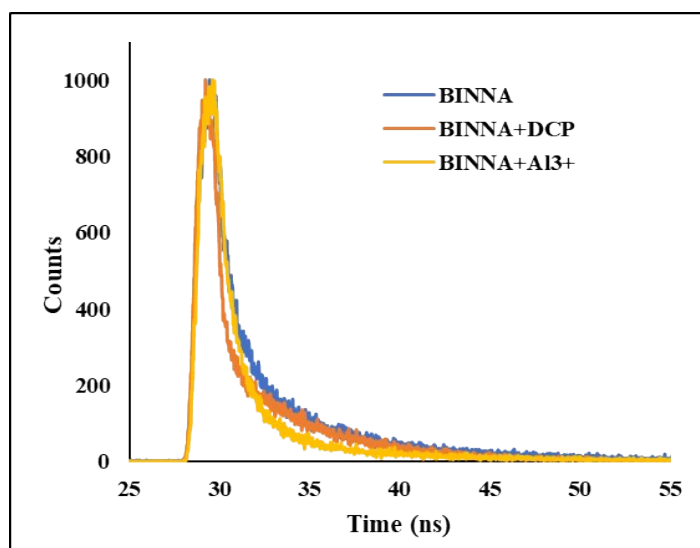


Figure S12: Time-resolved spectra of **BINNA** (CH_3OH) in the absence and presence of DCP and Al^{3+} ions.

Table S2: Calculated electronic spectra of **BINNA** in different solvents

		Absorption peak (λ ; nm)	Oscillation strength (f)	Dipole moment (a.u.)
Solvents	Chloroform	392	1.1576	4.52
	MeOH	397	1.1692	5.33
	DMSO	399	1.1854	5.38
	Acetone	393	1.1418	4.96

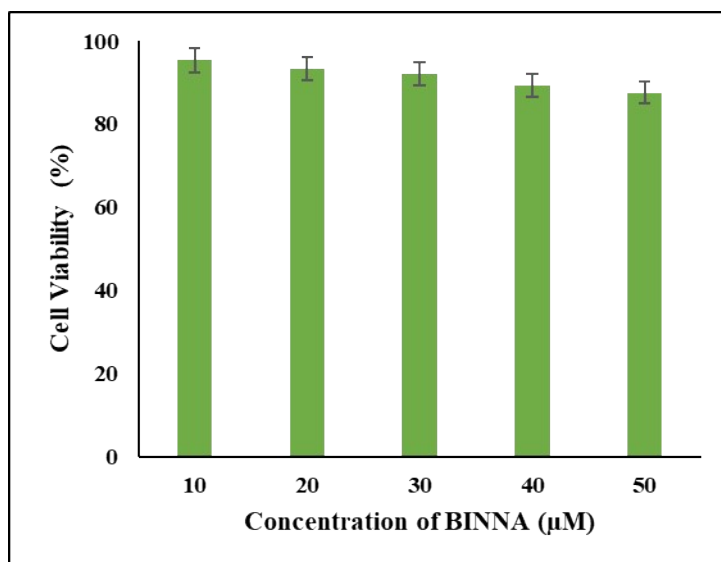


Figure S13: Cell viability of *E. coli* cells incubated with **BINNA** with different concentration (10, 20, 30, 40 and 50 μM) for 24h

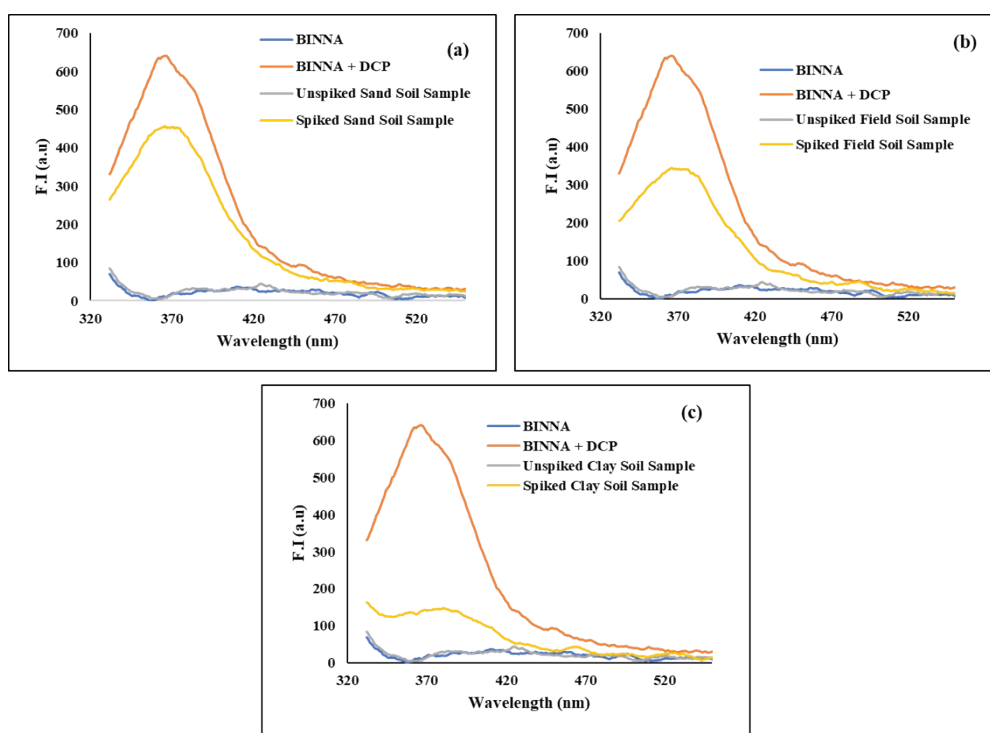


Figure S14: Emission spectra of **BINNA** solution in different spiked and unspiked DCP soil samples such as (a) sand, (b) field, and (c) clay soil.

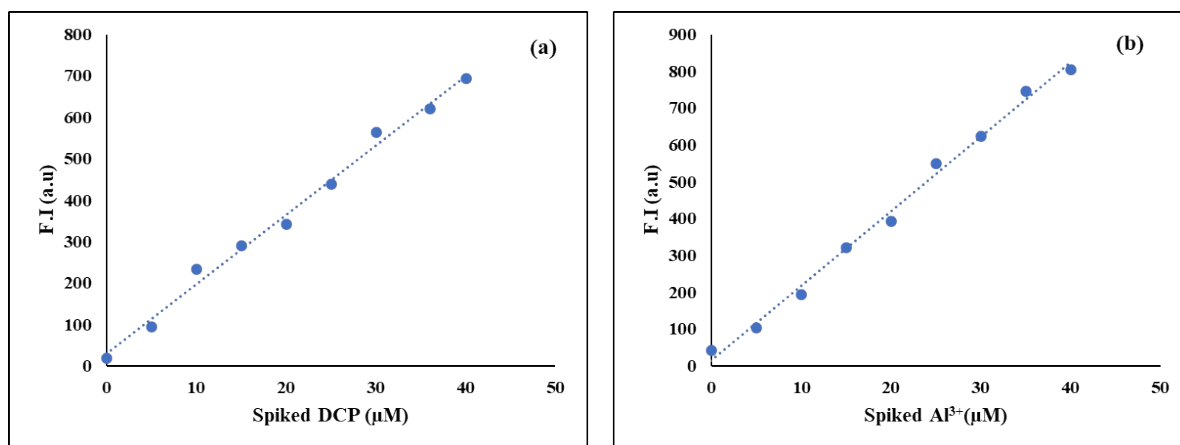


Figure S15: Calibration curve for (a) DCP and (b) Al³⁺ ions.

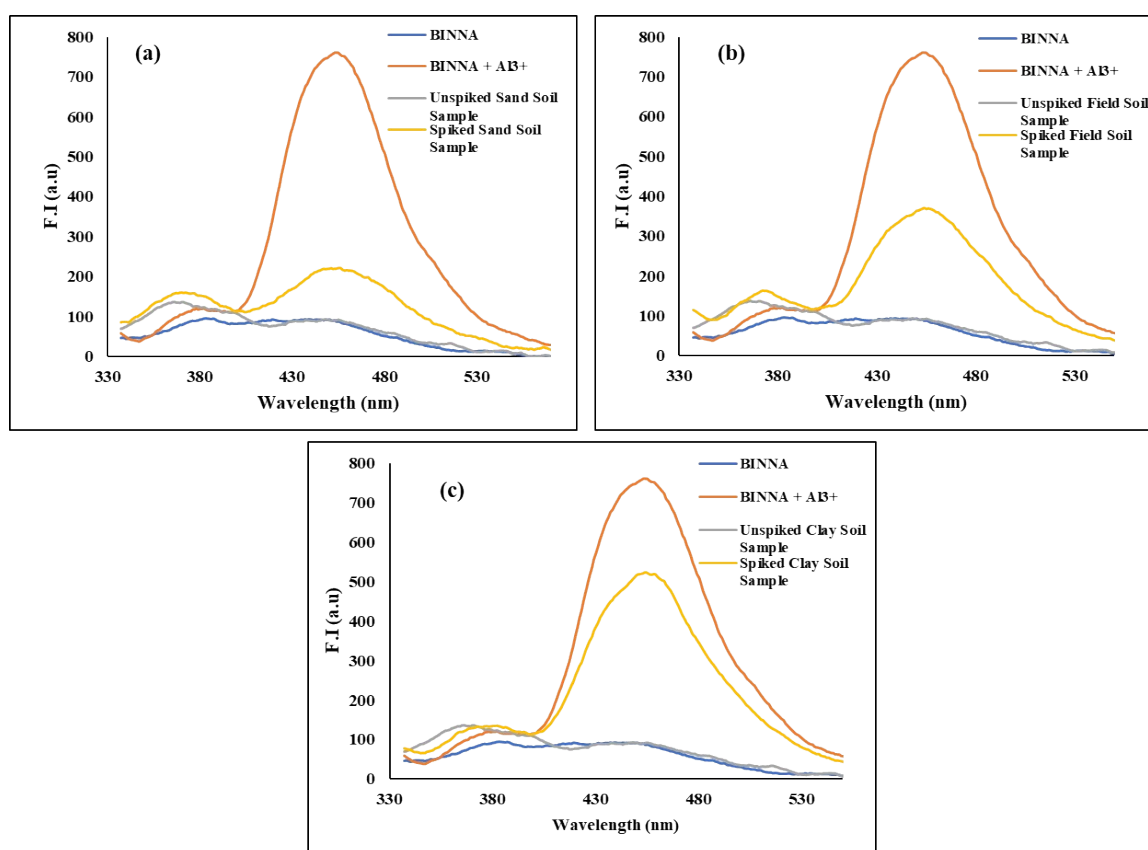


Figure S16: Emission spectra of BINNA solution in different Al³⁺ spiked and unspiked soil samples such as (a) sand, (b) field, and (c) clay soil

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