

Supporting Information

Ultrasensitive Triple-Mode Nile Red-L64 Niosome Nanothermometers for Real-Time Cellular Thermogenesis Mapping

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								(ns)
20	510	618	0.101	2.189	0.955	0.044	1.013	0.194
25	510	618	0.110	2.176	0.953	0.046	0.957	0.207
30	510	618	0.126	2.208	0.946	0.053	0.925	0.238
35	510	618	0.138	2.207	0.941	0.058	0.955	0.260
36	510	618	0.138	2.203	0.942	0.057	0.901	0.257
37	510	618	0.140	2.206	0.940	0.059	0.908	0.263
40	510	618	0.151	2.199	0.938	0.061	0.966	0.278

Table S2: the lifetime decay values of NRL64 in cell media at different temperatures

Temperature (°C)	λ_{ex} (nm)	λ_{em} (nm)	τ_1 (ns)	τ_2 (ns)	α_1	α_2	χ^2	Average lifetime (ns)
37	510	618	0.122	1.920	0.940	0.059	0.961	0.229
38	510	618	0.165	2.107	0.911	0.088	1.200	0.336
39	510	618	0.177	2.210	0.889	0.110	1.085	0.402
40	510	618	0.249	2.570	0.779	0.220	1.185	0.760

Table S3: The lifetime decay values of NR-L64 in live cells at different temperatures.

Temperature (°C)	λ_{ex} (nm)	λ_{em} (nm)	τ_1 (ns)	τ_2 (ns)	α_1	α_2	χ^2	Average lifetime (ns)
37	510	618	0.263	4.260	0.485	0.514	1.07	2.318
37.4	510	618	0.247	4.269	0.500	0.499	0.996	2.256
37.8	510	618	0.248	4.250	0.509	0.490	0.992	2.211
38.2	510	618	0.231	4.257	0.502	0.479	1.092	2.162
38.6	510	618	0.231	4.181	0.521	0.478	1.059	2.119
39	510	618	0.230	4.181	0.558	0.441	1.075	1.972
39.4	510	618	0.234	4.143	0.550	0.449	1.120	1.990
39.8	510	618	0.255	4.266	0.569	0.430	0.939	1.980

Table S4: the lifetime decay values of NR with glucose at different time intervals

Temperature (°C)	λ_{ex} (nm)	λ_{em} (nm)	τ_1 (ns)	τ_2 (ns)	α_1	α_2	χ^2	Average lifetime (ns)
5	510	618	0.1	2.119	0.969	0.030	0.859	0.16

10	510	618	0.122	2.189	0.962	0.037	0.853	0.199
15	510	618	0.118	2.097	0.964	0.035	0.938	0.188
20	510	618	0.126	2.214	0.965	0.034	0.866	0.199
25	510	618	0.115	2.196	0.964	0.035	0.864	0.188
30	510	618	0.140	2.247	0.960	0.039	0.833	0.222
35	510	618	0.127	2.145	0.963	0.036	1.124	0.200
40	510	618	0.128	2.301	0.966	0.033	0.990	0.200
45	510	618	0.142	2.236	0.959	0.040	0.925	0.226
50	510	618	0.125	2.216	0.963	0.036	0.973	0.202
55	510	618	0.126	2.144	0.963	0.036	1.008	0.199
60	510	618	0.114	2.126	0.965	0.034	0.928	0.183

Method S1. Two-photon power-dependent intensity of NR-L64 loaded in cells.

To confirm the nonlinear (quadratic) nature of two-photon excitation, the relationship between fluorescence intensity and excitation laser power was investigated. NR-L64 was loaded in cells and their fluorescence images were acquired at varying laser powers, and the total fluorescence intensity in each image was measured by taking the region of interest (ROI) of the probe in the cells at different laser powers. Both the fluorescence intensity and laser power values were plotted on a log-log scale (\log_{10} of fluorescence intensity versus \log_{10} of laser power). Linear regression of the data yielded a slope of approximately 1.74, consistent with the expected near-quadratic dependence for two-photon excitation. This provides strong evidence that the observed fluorescence of NR-L64 in FaDu cells arises predominantly from a two-photon absorption process.

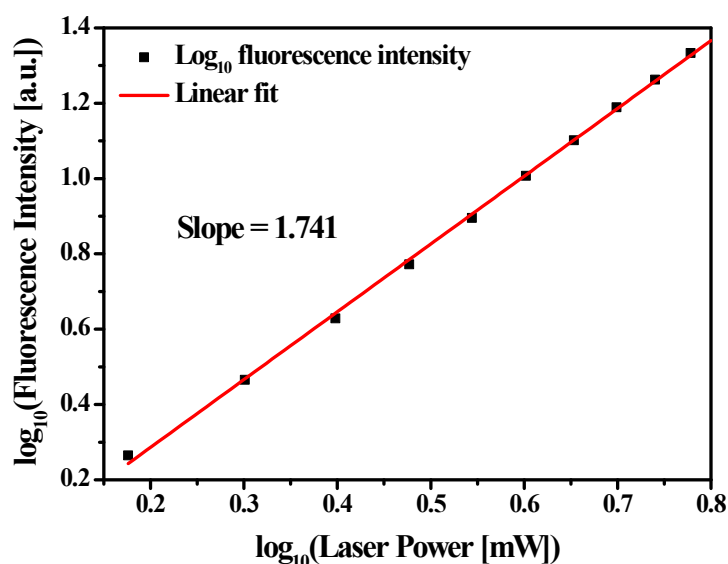


Figure S3. Two-photon power-dependent intensity of NR-L64 loaded in cells.

Table S5: Mean fluorescence intensity of NR-L64 niosomes loaded in FaDu cells irradiated under a two-photon fluorescence microscope at 37 °C and 39 °C for 70 min.

Time (min)	Average fluorescence	Average fluorescence
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	intensity at 37 °C	intensity at 39 °C
0	18.65	54.58
5	18.7	54.96
10	18.95	53.99
15	19.03	55.16
20	18.59	54.73
25	18.49	55.52
30	18.71	55.4
35	18.72	54.56
40	18.43	54.63
45	18.5	53.98
50	18.57	53.97
55	18.84	54.05
60	18.59	54.96
65	19.1	55.18
70	19.05	55.07

Method S2. Thermal monitoring of cell metabolism using confocal and multiphoton imaging

FaDu cells were seeded in confocal petri dishes (50×10^3 cells/dish) and cultured for 24 h at 37 °C under 95% relative humidity and 5% CO₂. The cells were then incubated for 2 h with NR-L64 (1.25×10^{-5} M NR), suspended in EMEM supplemented with 10% FBS, based on our previous cell viability studies. After incubation, the cells were washed and resuspended in fresh 10% FBS-containing EMEM.

Cellular metabolism was stimulated by adding 1 mL of glucose solution (5 mg mL^{-1}) to the culture medium, mimicking hyperglycemic conditions in the cell. Once taken up by the cells, glucose undergoes glycolysis to generate pyruvate, which either enters the mitochondria to fuel oxidative phosphorylation or is converted into lactate, with both pathways contributing to energy production¹. A parallel group containing only NR-L64 was used to obtain the calibration curve. Fluorescence intensity changes of NR-L64 in response to cellular metabolism were monitored using confocal laser scanning microscopy (LSCM) and two-photon fluorescence and lifetime microscopy. For confocal imaging, NR-l64 in live FaDu was excited at 514 nm, and emission was collected between 610–670 nm. For two-photon imaging, 825 nm excitation was used, and emission was recorded using a 580–632 nm band pass filter. Calibration was achieved by recording fluorescence and lifetime images between 37–39 °C in 0.2 °C increments. For experimental measurements, glucose-treated cells were observed for 60 min, with fluorescence images captured every 5 min at 20× magnification. The fluorescence

intensity and lifetime values were obtained from the images by taking a region of interest (ROI) from the cells (as described in methods S3 and S4). Temperature was precisely maintained using a controller (H301-EC-M, OKO Laboratories) with an accuracy of ± 0.1 °C.

Table S6: Calculation of relative sensitivity from average fluorescence intensity obtained from the confocal calibration experiment.

Temperature (°C)	Average Fluorescence Intensity (Q)	d(Q)	Relative sensitivity (Sr)
37.2	0.05035	0.00185	18.37
37.4	0.05251	0.00216	20.56
37.6	0.06252	0.01001	80.05
37.8	0.06739	0.01001	74.26
38	0.07685	0.01446	94.07
38.2	0.09639	0.01954	101.35
38.4	0.10658	0.01019	47.80
38.6	0.11148	0.0049	21.97
38.8	0.12171	0.01023	42.02
39	0.14171	0.02	70.56

Table S7: Calculation of relative sensitivity from average fluorescence intensity obtained from the Two-photon calibration experiment.

Temperature (°C)	Average Fluorescence Intensity (Q)	d(Q)	Relative sensitivity (Sr)
37.2	0.15506	0.00891	28.73
37.4	0.15685	0.00179	5.70
37.6	0.16566	0.00881	26.59
37.8	0.17081	0.00515	15.07
38	0.1847	0.01389	37.60
38.2	0.23525	0.05055	107.43
38.4	0.27959	0.04434	79.29
38.6	0.30678	0.02719	44.31
38.8	0.32419	0.01741	26.85
39	0.35419	0.03	42.35

Table S8: Calculation of relative sensitivity from the average fluorescence lifetime obtained from the fluorescence lifetime calibration experiment.

Temperature (°C)	Average Fluorescence Lifetime (Q)	d(Q)	Relative sensitivity (Sr)
37.2	3.278689	0.02	3.278689
37.4	3.30033	0.02	3.30033
37.6	5	0.03	5
37.8	1.672241	0.01	1.672241
38	1.677852	0.01	1.677852
38.2	6.802721	0.04	6.802721
38.4	3.424658	0.02	3.424658
38.6	1.706485	0.01	1.70648
38.8	1.70068	0.01	1.70068

Method S3. Average Fluorescence intensity calculation from confocal and Two- photon microscope images using LAS X software

The confocal images of NR-L64 in cells were taken using leica confocal microscope (TCS SP8, Leica Microsystems, Germany) and the Two-photon images in cells were taken using Leica SP8 microscope coupled with a Chameleon Vision II femtosecond Ti: Sapphire oscillator. To calculate the average fluorescence intensity from the images, LAS X software from Leica was used. The images were opened in the quantification window of the software and from the tools tab “Histogram” was selected which determines the distribution of gray-scale values in images and in the individual images of image series based on regions of interest (ROI). The region of interest (ROI) was drawn around different cells present in the image using the “draw polygon” feature present in the tool bar. The mean intensity of each ROI was obtained from the statistics window of the software which corresponds to the fluorescence intensity obtained from that particular cell as highlighted in figures (S6, S7, S8 and S9), to calculate the average fluorescence intensity the mean intensity of each cell was divided by the ROI area of that particular cell to get the average fluorescence intensity per area. The average fluorescence intensity per area of multiple cells were taken in a image and their mean value was used as the average fluorescence intensity of that particular image as given in table S5 and S6.

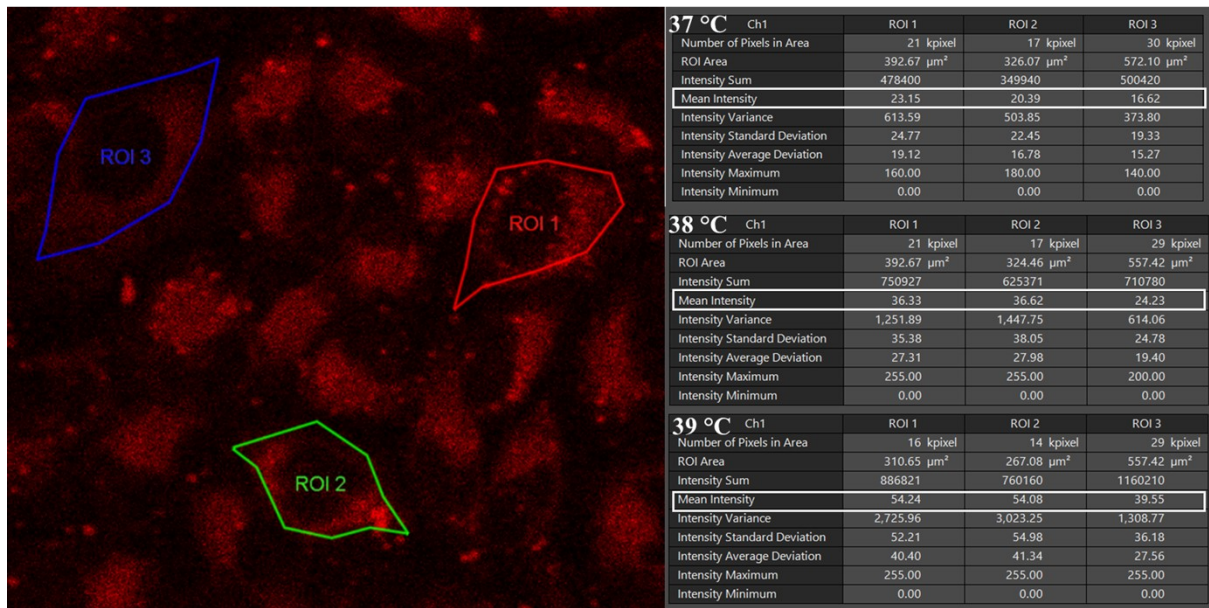


Figure S4: Confocal image (only at 37°C is shown) showing the ROI of cells taken for calibration calculations and their corresponding fluorescence mean intensity values at different temperatures.

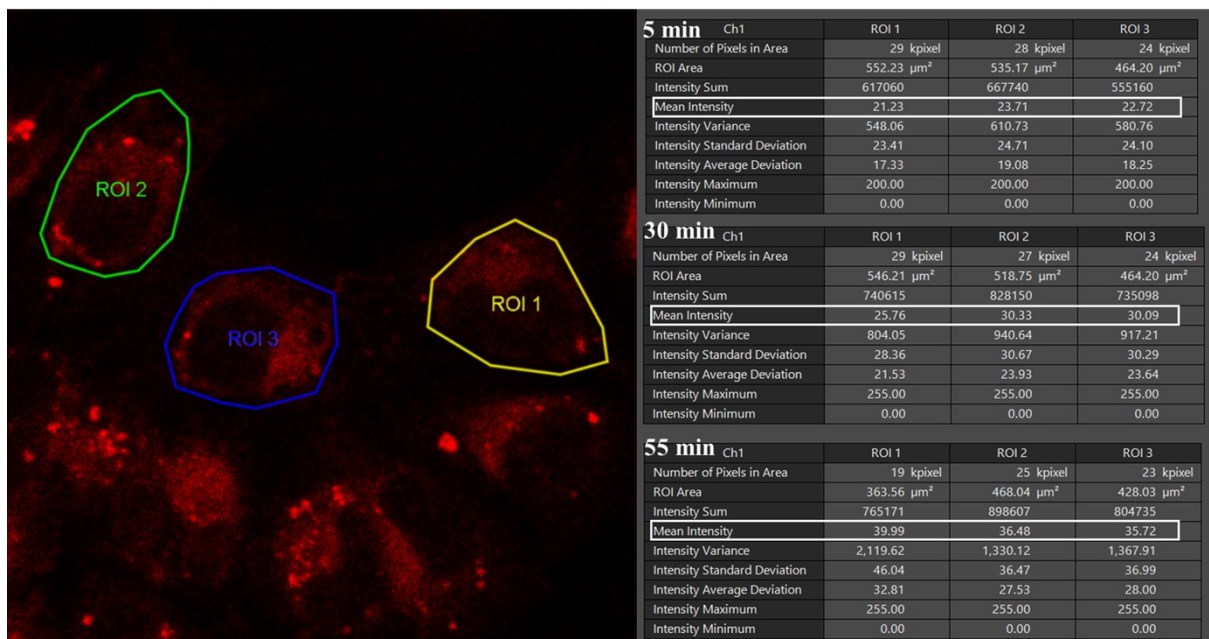


Figure S5: Confocal image (after 55 min is shown) showing the ROI of the cells after glucose addition and their corresponding fluorescence mean intensity values at different time intervals.

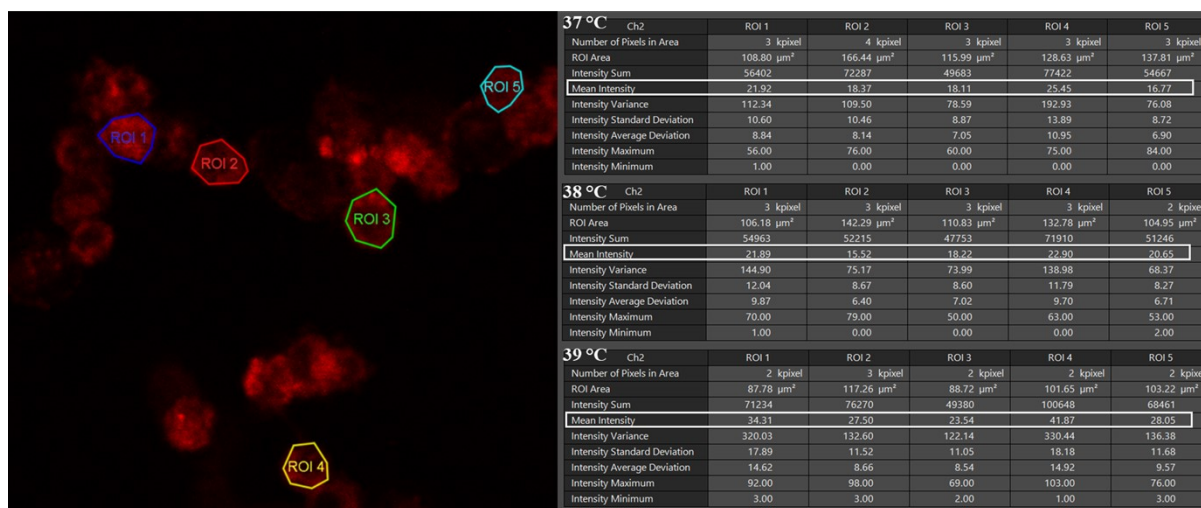


Figure S6: TPFM image (only at 37°C is shown) showing the ROI of cells taken for calibration calculations and their corresponding fluorescence mean intensity values at different temperatures.

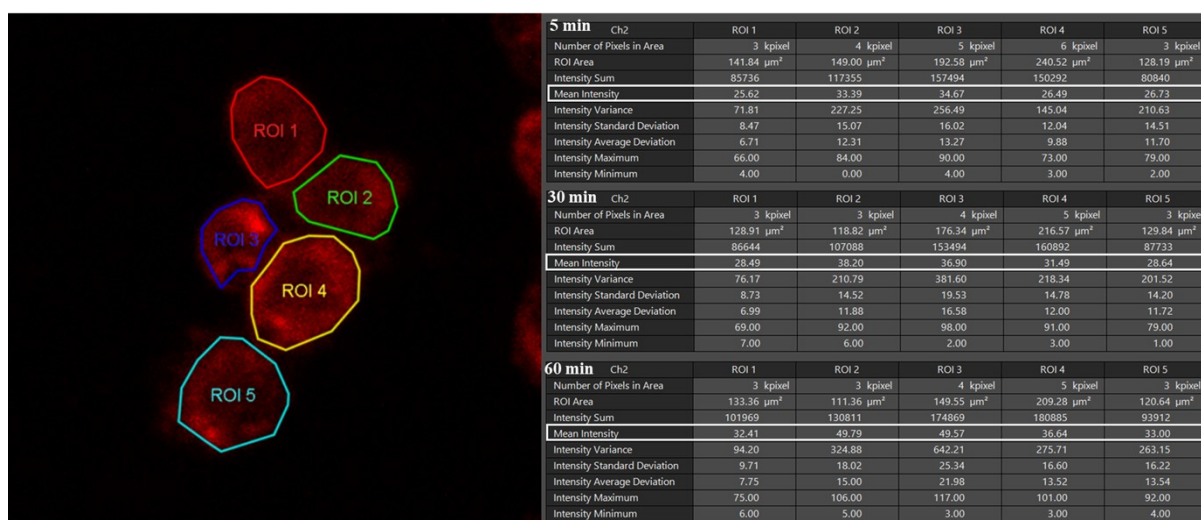


Figure S7: TPFM image (after 60 min is shown) showing the ROI of the cells after glucose addition and their corresponding fluorescence mean intensity values at different time intervals.

Method S4: TauModes for the quantification of the average fluorescence lifetime from the Two- photon microscope images using LAS X software

The Fluorescence lifetime images in cells were taken using a Leica SP8 microscope coupled with a Chameleon Vision II femtosecond Ti: Sapphire oscillator. To visualize the fluorescence lifetime in images taken with the Leica microscope, the Leica system provides different

TauModes and a TauContrast function. The TauContrast provides the average photon arrival time per pixel in the detector and displays it as a color-lifetime image (figures S10 and S11). In the TauModes feature, the Tau Intensity feature was selected, which gives the full photon arrival time intensity image. To calculate the average fluorescence lifetime from the images, a process similar to that described in Method S3 was used. The region of interest (ROI) was drawn around different cells present in the image using the “draw polygon” feature present in the toolbar. The “mean weighted arrival time” of each ROI was obtained from the statistics window of the software which corresponds to the average fluorescence lifetime obtained from that particular cell as highlighted in figures (S10, and S11), to calculate the average fluorescence lifetime the “mean weighted arrival time” of multiple cells were taken in an image and their mean value was calculated as the average fluorescence lifetime of that particular image as given in Table S7.

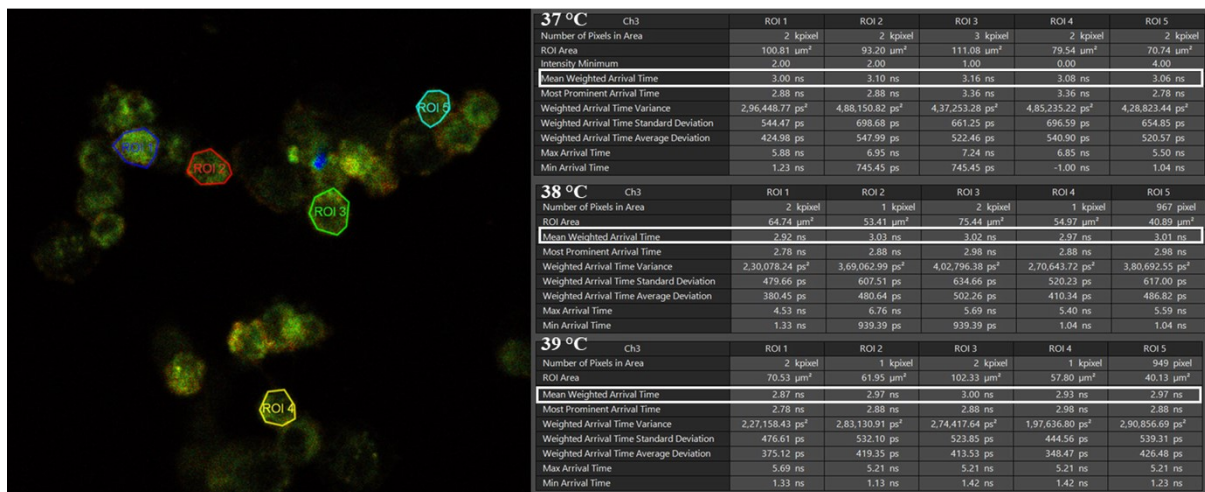


Figure S8: Two-photon Fluorescence lifetime image (only at 37°C is shown) showing the ROI of cells taken for calibration calculations and their corresponding fluorescence lifetime values at different temperatures.

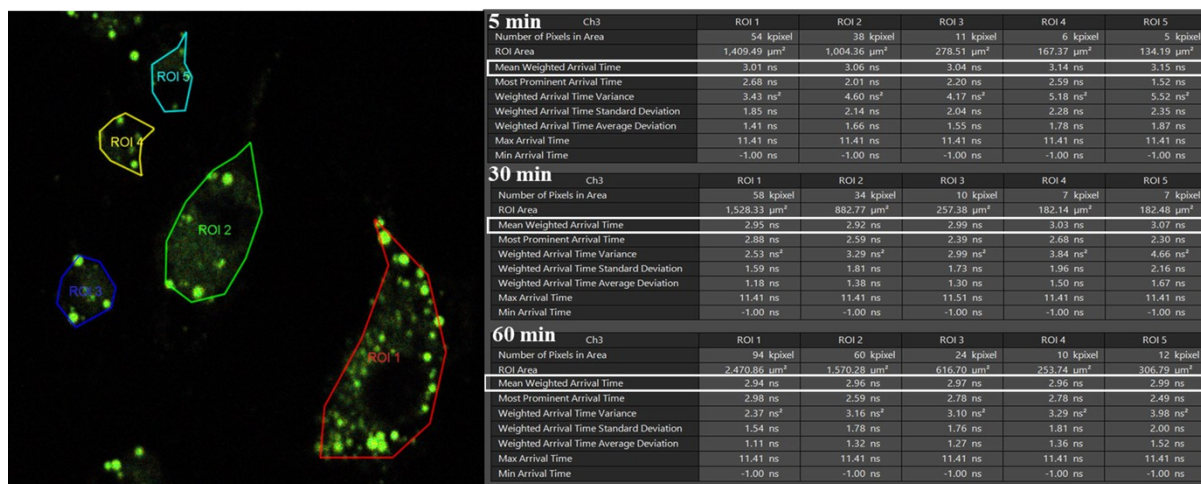


Figure S9: Two-photon Fluorescence lifetime image (after 60 min is shown) showing the ROI of the cells after glucose addition and their corresponding fluorescence lifetime values at different time intervals.

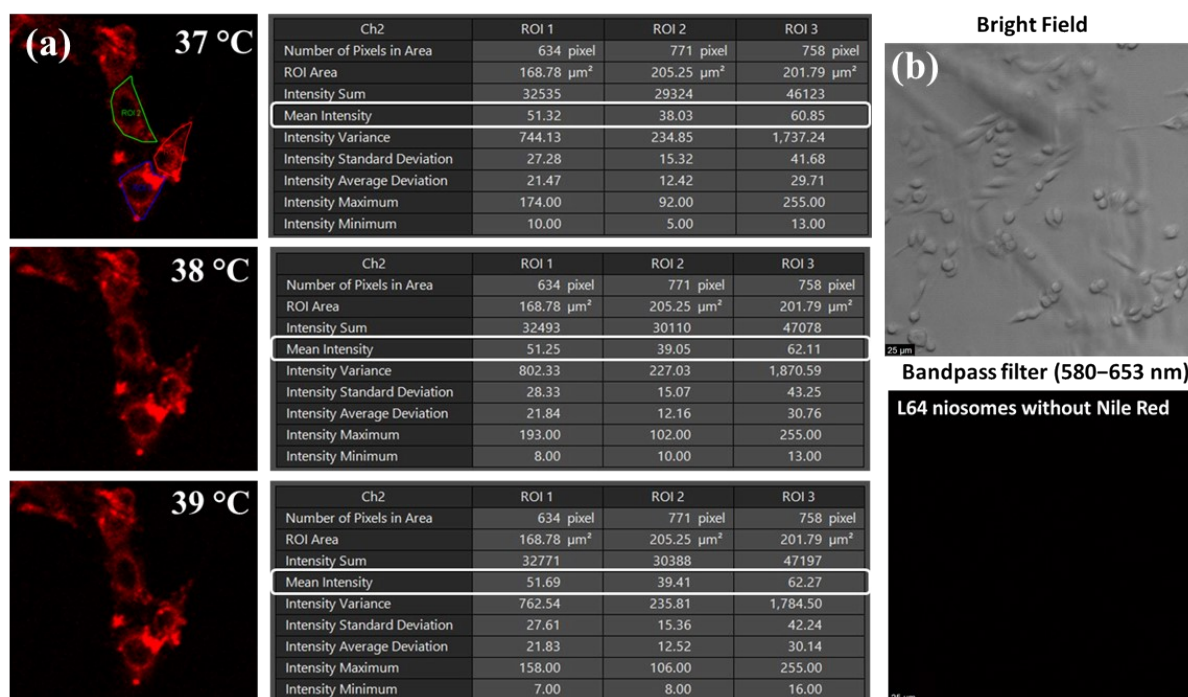


Figure S10: TPFM images of (a) only Nile red in cells at different temperatures, and (b) only L64 niosomes in cells at 37°C.

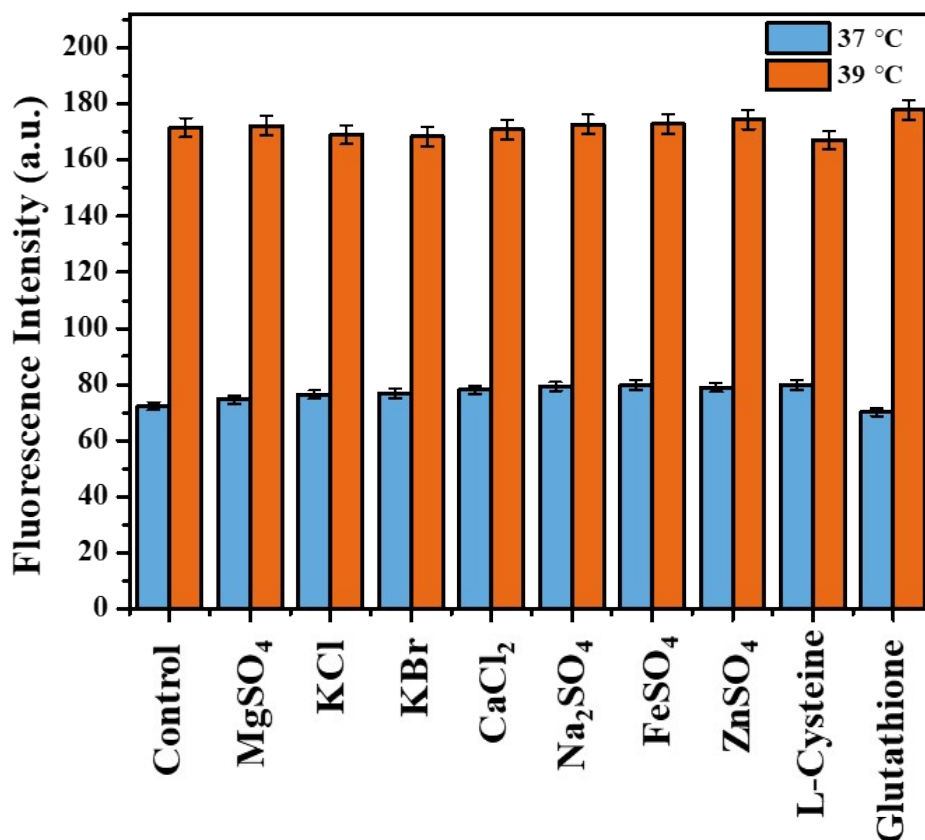


Figure S11. Fluorescence response of Nile Red-L64 (at 37 °C and 39 °C) in the presence of common intracellular interferents (metal ions, anions, and biothiols), each at 10 mM. Control: Without any interferents.

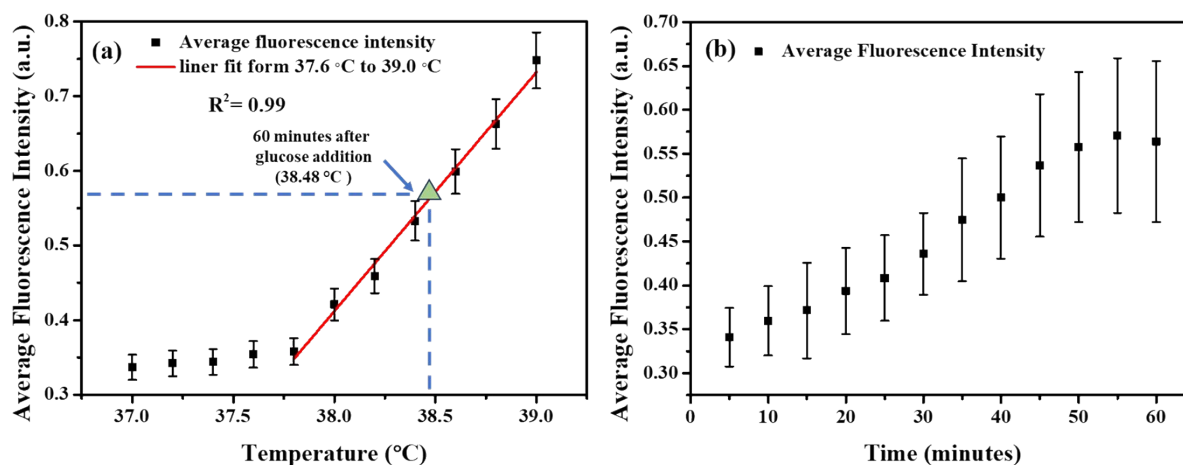


Figure S12: (a) Two-photon fluorescence calibration curve obtained from NR-L64 in live FaDu cells, and (b) Two-photon average fluorescence intensity obtained after glucose addition in live FaDu cells at lower laser power (1 mW).

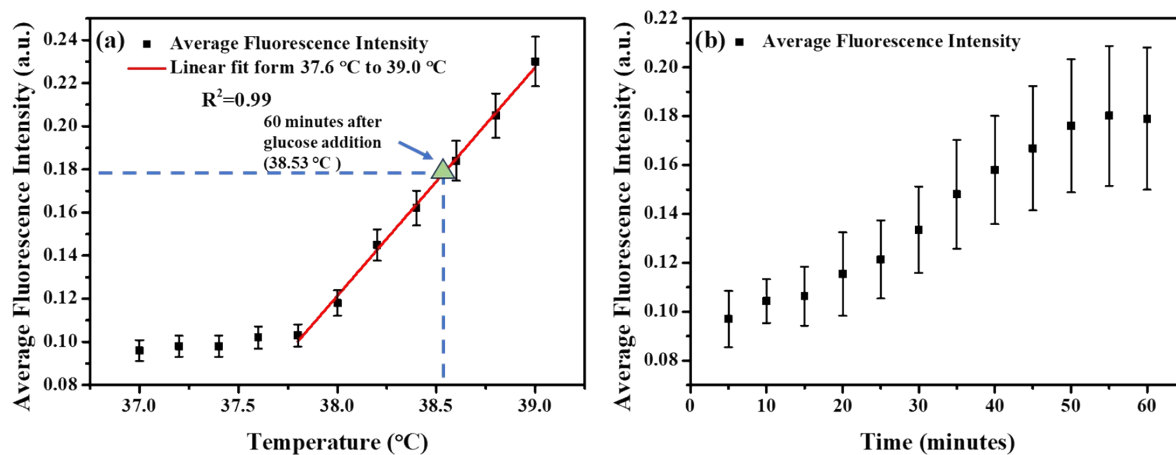


Figure S13: (a) Two-photon fluorescence calibration curve obtained from NR-L64 in live FaDu cells, and (b) Two-photon average fluorescence intensity obtained after glucose addition in live FaDu cells at higher laser power (4 mW).

References:

1. S. J. Kierans and C. T Taylor, *J. Biol. Chem.*, 2024, **300**, 107906.