

Supplementary Information

Microwave-assisted hydrolysis for the physicochemical characterization of functional methacrylic polymers and their bioconjugates

Ilaria Porello¹, Paola Nastri¹, Marta Bozzi¹, Filippo Moncalvo¹, Philippe Gonzalez², Alessandro Sacchetti¹, Francesco Cellesi^{1}*

¹Department of Chemistry, Materials and Chemical Engineering “Giulio Natta”, Politecnico di Milano, Via Luigi Mancinelli 7, 20131, Milan, Italy.

²Department of Polymers for Health and Biomaterials, IBMM, Univ Montpellier, CNRS, ENSCM, 34090, Montpellier, France.

*Corresponding author.

Email address: francesco.cellesi@polimi.it (F. Cellesi).

Experimental

Lysozyme functionalization with AROMA-1 ATRP initiator. Lysozyme from hen egg white (20 mg, 0.0014 mmol, 1 eq.) was inserted in a glass vial together with 2-methylpyridine borane complex (4.49 mg, 0.042 mmol, 30 eq.) and dissolved in 10 mL of PO_4^{3-} 0.05 M NaCl 0.1 M buffer. 4-Formylphenyl 2-bromo-2-methylpropanoate (AROMA-1, A1) ATRP initiator was synthesized according to the procedure previously reported.^{34,35} 21 μL of AROMA-1 initiator solution (0.0069 mmol, 5 eq.) were added to the reaction mixture withdrawing an aliquot from a previously prepared stock solution in DMSO ($[\text{A1}] = 90.33 \text{ mg/mL}$). The reaction mixture was stirred at 25°C for 4 h. The final product (LYS-A1) was purified via dialysis against deionized water for 48 h (3.5 kDa MWCO membranes) then freeze-dried.

Synthesis of LYS-PGMA_m. 20 mL of PBS 100 mM were inserted in a two-necks round bottom flask, 3 cycles of vacuum/ N_2 of 5 min each were performed followed by 15 min of N_2 purge. GMA monomer (14.23 mg, 0.0889 mmol, m eq., where m = 125) was inserted in a two-necks round bottom flask together with LYS-A1 (10.4 mg, 0.000711 mmol, 1 eq.) and 3 vacuum/ N_2 cycles of 5 min each were performed. 548 mL of degassed PBS 100 mM were added to the Schlenk vial and the reaction mixture was left under continuous flow of N_2 . CuCl_2 (0.00717 mg, 0.053 mmol, 0.075 eq) and TPMA (0.1238 mg, 0.0443 mmol, 0.6 eq) were added to the reaction mixture from a stock solution in PBS 100 mM ($[\text{CuCl}_2] = 25 \text{ mM}$, $[\text{TPMA}] = 200 \text{ mM}$). A stock solution of AA was prepared in a two-necks round bottom flask by firstly introducing dry AA (degassed with 3 vacuum/ N_2 cycles), then adding degassed PBS 100 mM ($[\text{AA}] = 5.28 \text{ mg/mL}$). An aliquot of AA stock solution (0.0976 mg, 0.0554 mmol, 0.779 eq.) was inserted in the reaction flask to start the polymerization. The reaction mixture was stirred at 30°C for 16 h. The resulting product was purified by ultrafiltration with centrifugal filters (Amicon® Ultra 15, 30 kDa MWCO, Merck Scientific) at 2500 RCF for 10 min, washing four times with 1 mL of PBS and lyophilized. Yield 70–80%.

Results and discussion

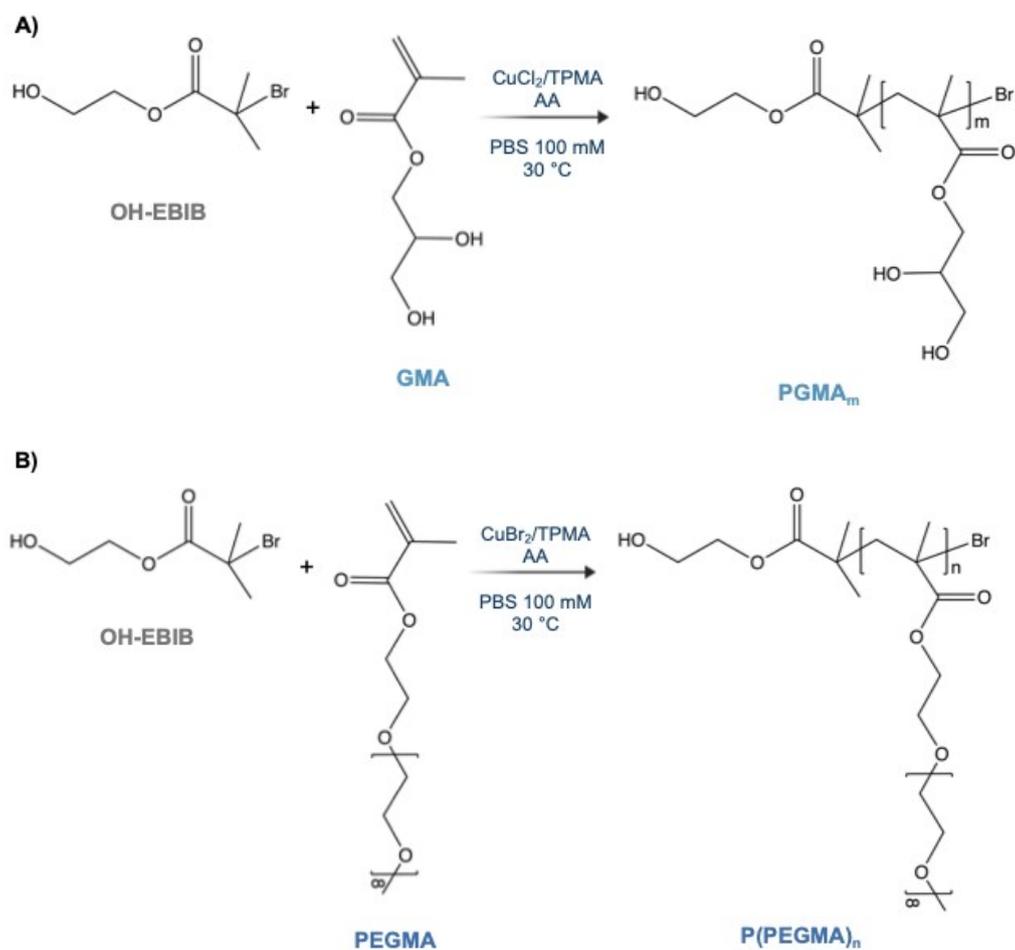


Figure S1. Synthetic pathway followed for the polymerization of: **A)** PGMA_m **B)** P(PEGMA)_n from commercial initiator via AGET ATRP.

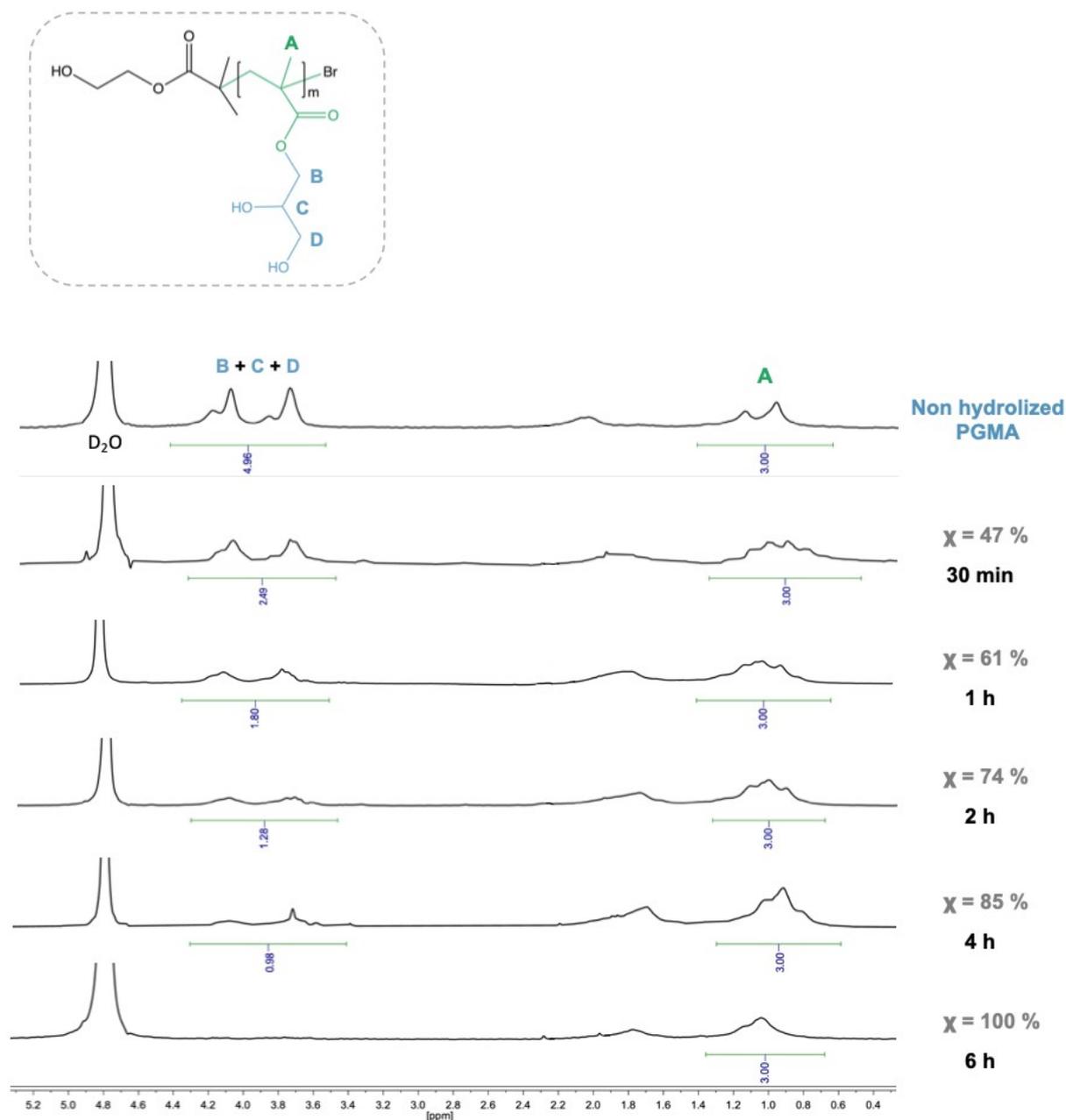


Figure S2. Sequence of ¹H-NMR spectra recorded before the hydrolysis of PGMA, using an oil bath-based heating system, and after various reaction time points.

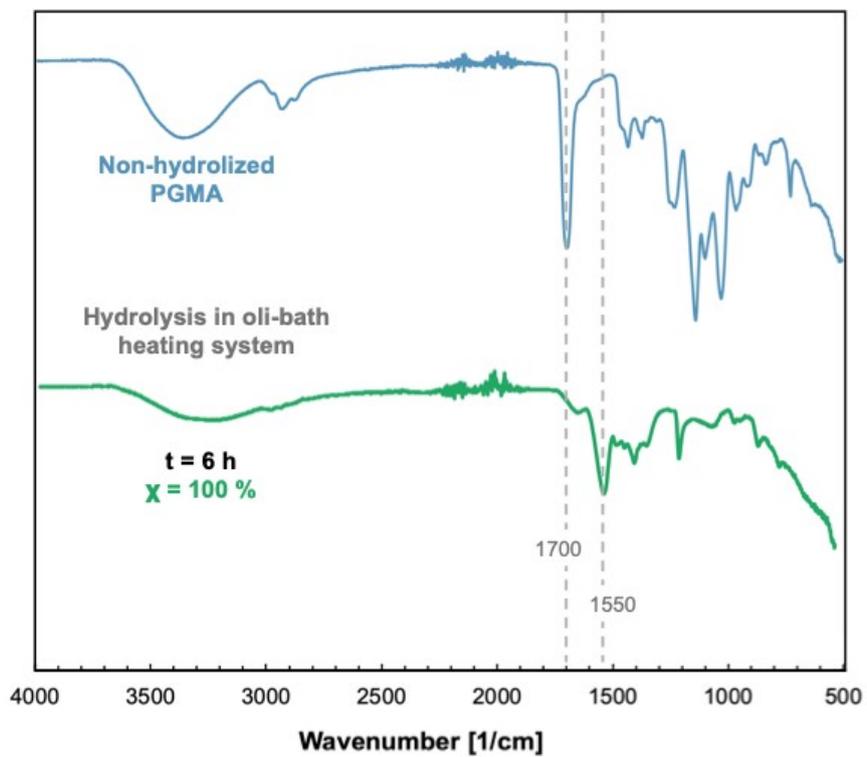


Figure S3. FTIR spectra recorded before PGMA hydrolysis (light blue) and after 6 h-hydrolysis leading 100% of conversion employing an oil-bath based heating system.

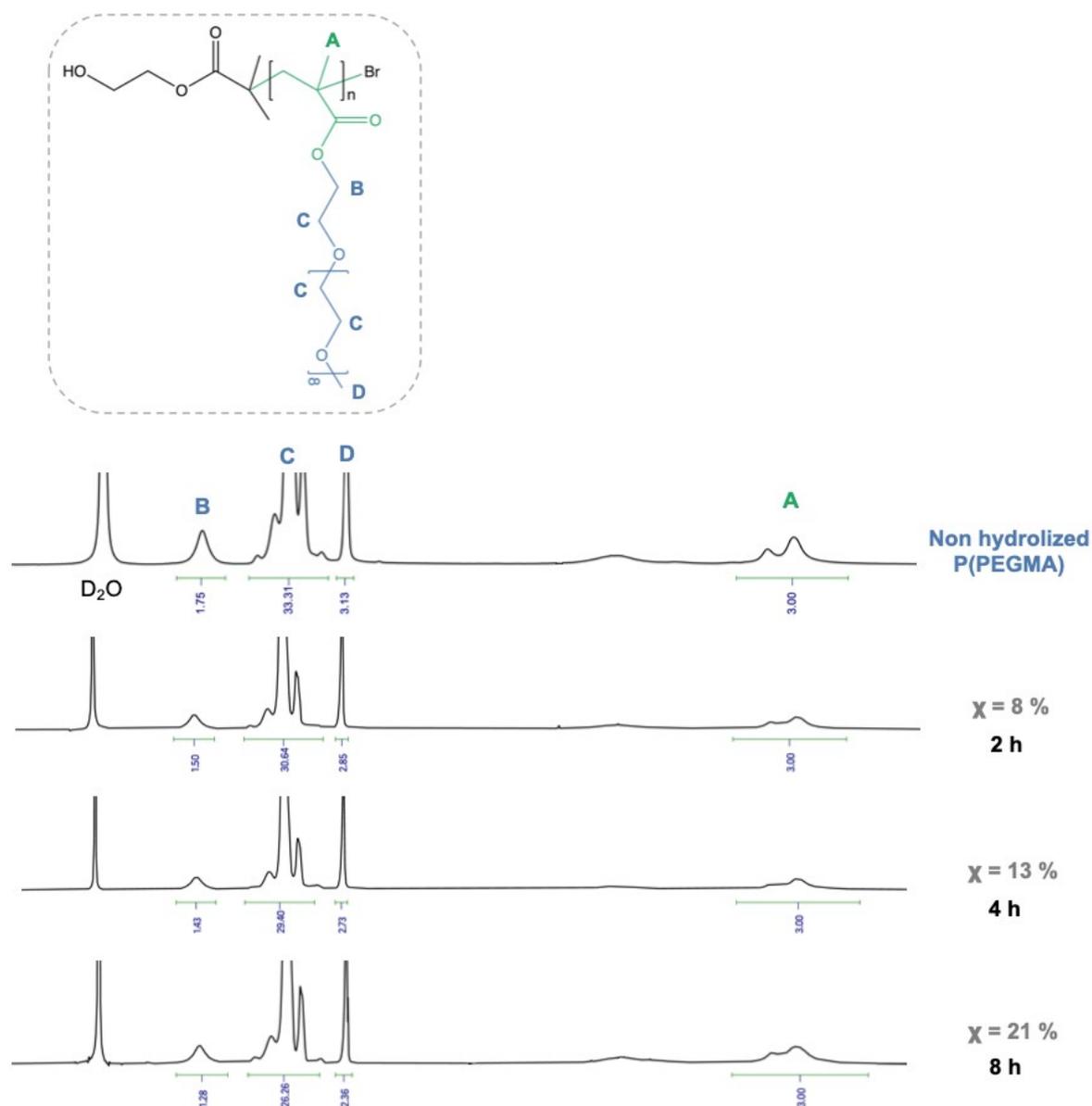


Figure S4. Sequence of $^1\text{H-NMR}$ spectra recorded before the hydrolysis of P(PEGMA), using an oil bath-based heating system, and after various reaction time points.