

## A Ligation DNzyme-Driven Catalytic Hairpin Assembly Strategy for miRNA Detection

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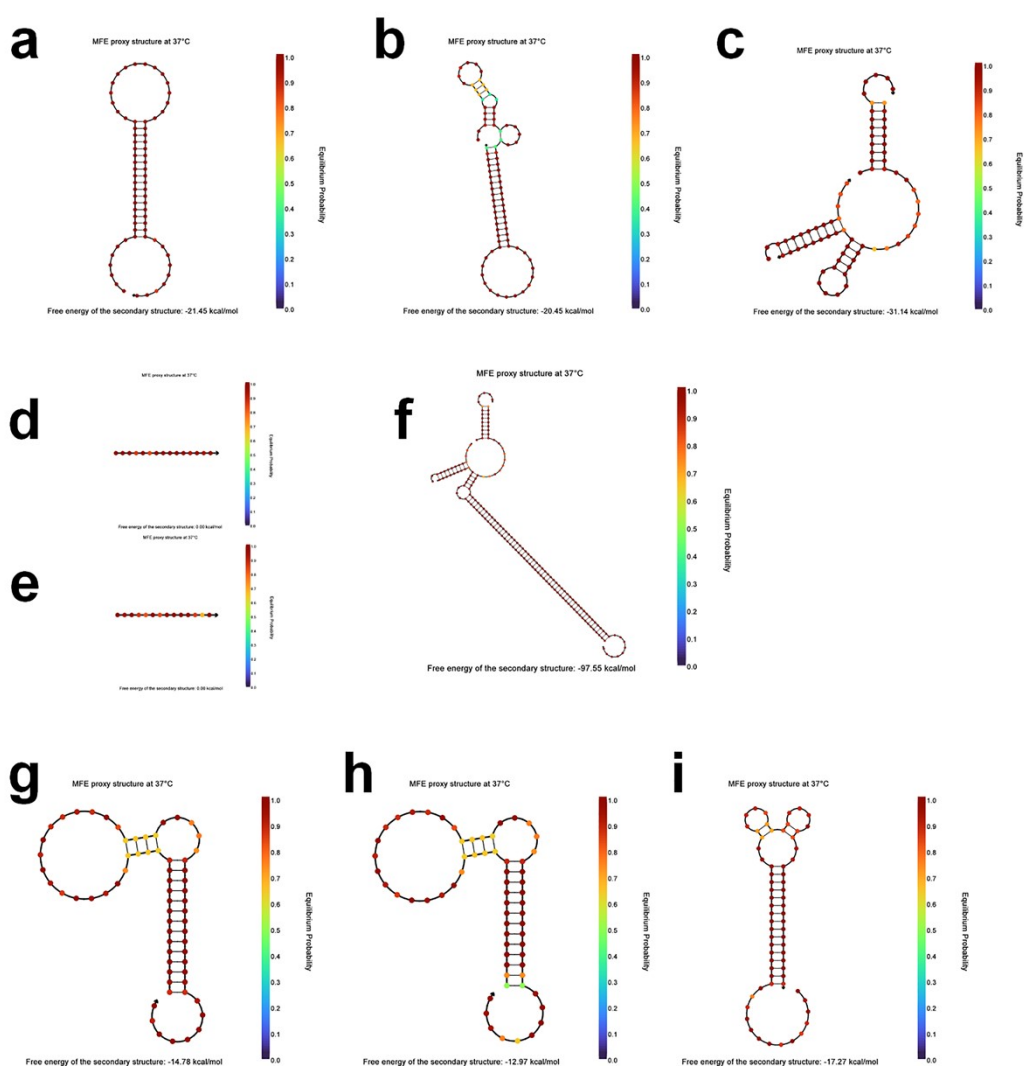
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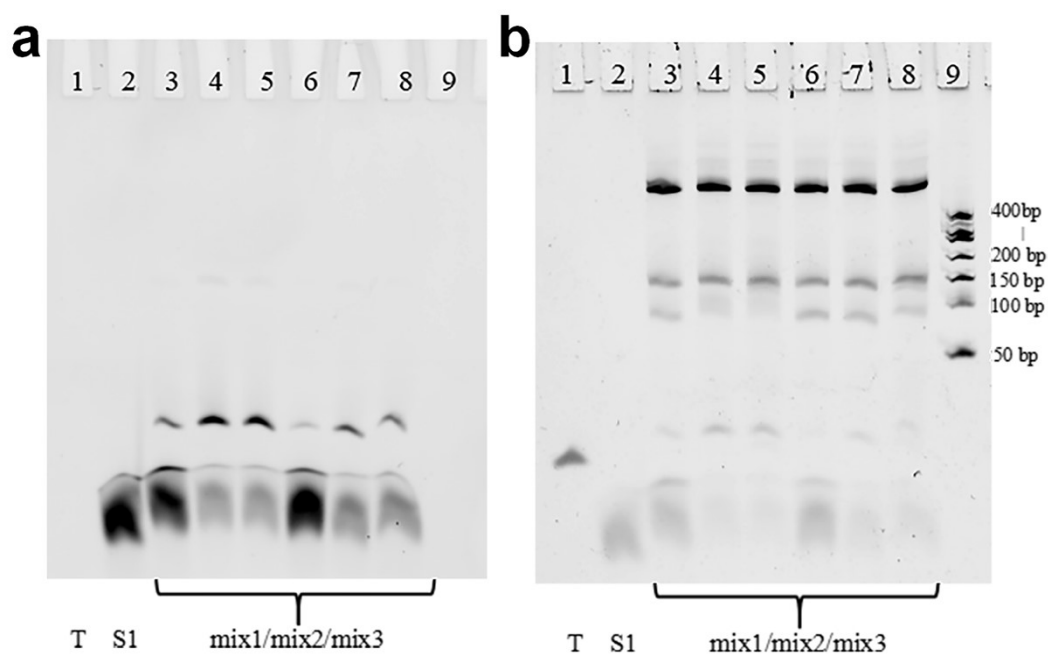
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Fig. S1



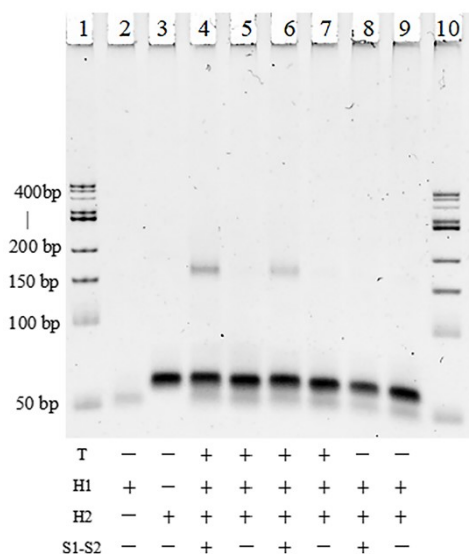
**Fig. S1.** NUPACK-predicted secondary structures of oligonucleotide components used in the LiD-CHA system. (a) H1. (b) H2. (c) E47-A in the presence of substrates S1 and S2. (d) S1. (e) S2. (f) Mixture of H1, H2, S1, and S2. (g) H0-1. (h) H0-2. (i) H0-3.

Fig. S2



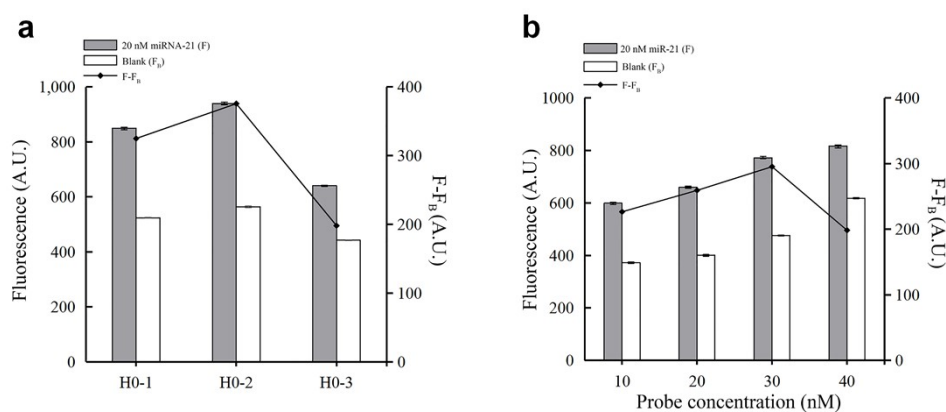
**Fig. S2 A 20% denaturing PAGE analysis of the ligation activity of the LiD-CHA system. (a)** Unstained gel image. **(b)** SYBR Green I-stained gel image. Lane 1, trigger (T); lane 2, FAM-labeled S1; lanes 3–5, reaction mixtures (mix-1 to mix-3) incubated at room temperature overnight; lanes 6–8, reaction mixtures (mix-1 to mix-3) incubated at room temperature for 4 h; lane 9, DNA marker. Mix-1 and mix-2 contain 1  $\mu$ M and 2  $\mu$ M of the CHA-generated H1-H2 complex, respectively, together with FAM-labeled S1 and S2. Mix-3 contains T, H1, H2, FAM-labeled S1, and S2.

**Fig. S3**



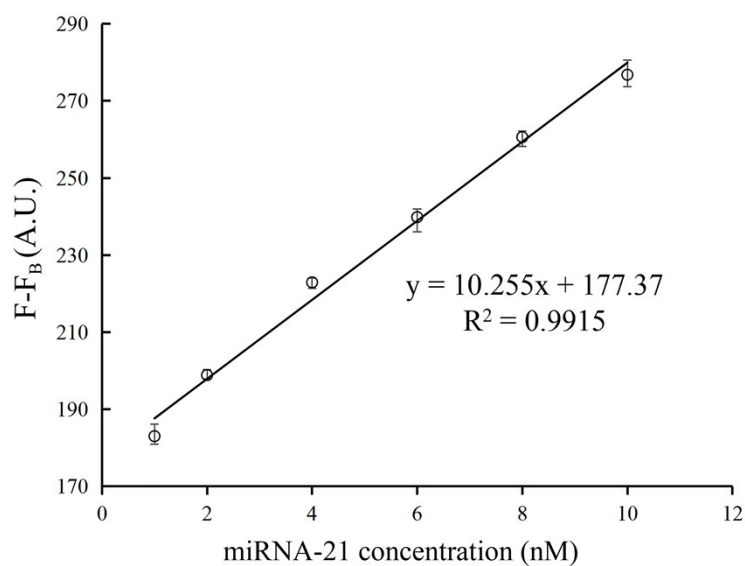
**Fig. S3. Evaluation of background interference from substrates S1 and S2 in the LiD-CHA system.** A 12% native PAGE analysis of different reaction mixtures. Lane 1, DNA marker; lane 2, H1; lane 3, H2; lane 4, 10 nM trigger (T) + H1 + H2 + S1 + S2; lane 5, 10 nM T + H1 + H2; lane 6, 5 nM T + H1 + H2 + S1 + S2; lane 7, 5 nM T + H1 + H2; lane 8, H1 + H2 + S1 + S2; lane 9, H1 + H2; lane 10, DNA marker.

**Fig. S4**



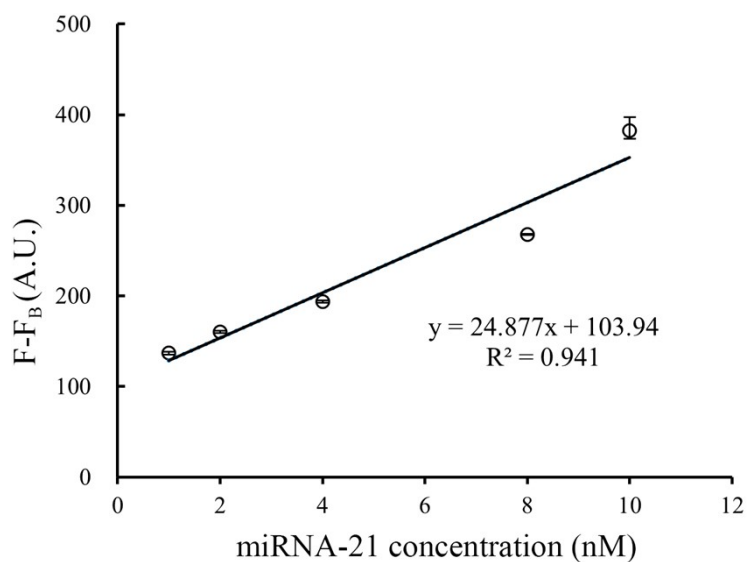
**Fig. S4. Optimization of the H0 hairpin probe for miRNA-21 recognition in the LiD-CHA system.** (a) Fluorescence responses of LiD-CHA systems constructed with different H0 sequences (H0-1, H0-2, and H0-3) in the presence of 20 nM miRNA-21 and in the absence of the target (blank). (b) Effect of H0 probe concentration on the fluorescence response of the LiD-CHA system in the presence of 20 nM miRNA-21 and under blank conditions. Gray bars represent fluorescence signals obtained in the presence of miRNA-21 (F), white bars represent background fluorescence signals (F<sub>B</sub>), and black lines correspond to the net fluorescence intensity (F - F<sub>B</sub>). Error bars represent the standard deviation of three independent experiments.

**Fig. S5**



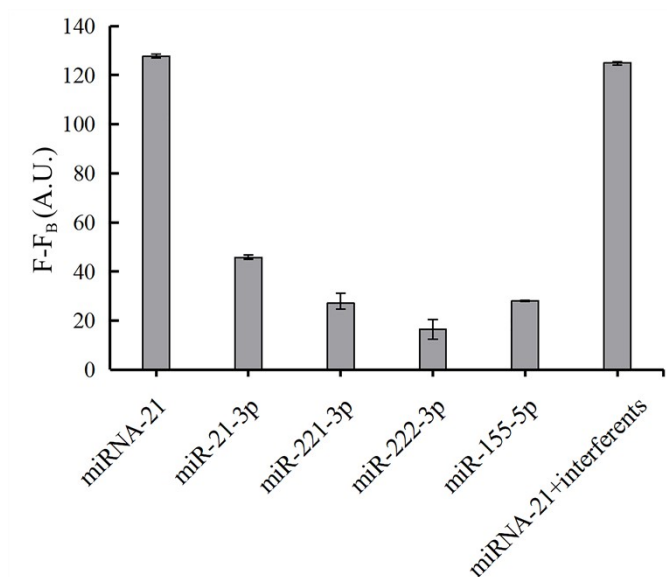
**Fig. S5 Calibration curve of fluorescence response for the LiD-CHA system in the concentration range of 1-10 nM.** Net fluorescence intensity ( $F-F_B$ ) plotted as a function of miRNA-21 concentration. The solid line represents the linear fitting of the experimental data. Error bars represent the standard deviation of three independent experiments.

**Fig. S6**



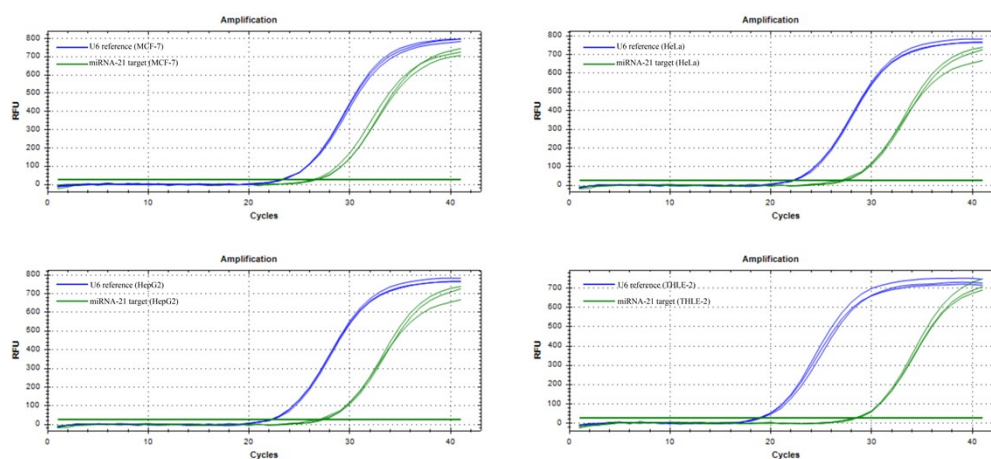
**Fig. S6. Calibration curve of fluorescence response for the conventional CHA system lacking S1 and S2.** Net fluorescence intensity ( $F-F_B$ ) plotted as a function of miRNA-21 concentration. The solid line represents the linear fitting of the experimental data. Error bars represent the standard deviation of three independent experiments.

**Fig. S7**



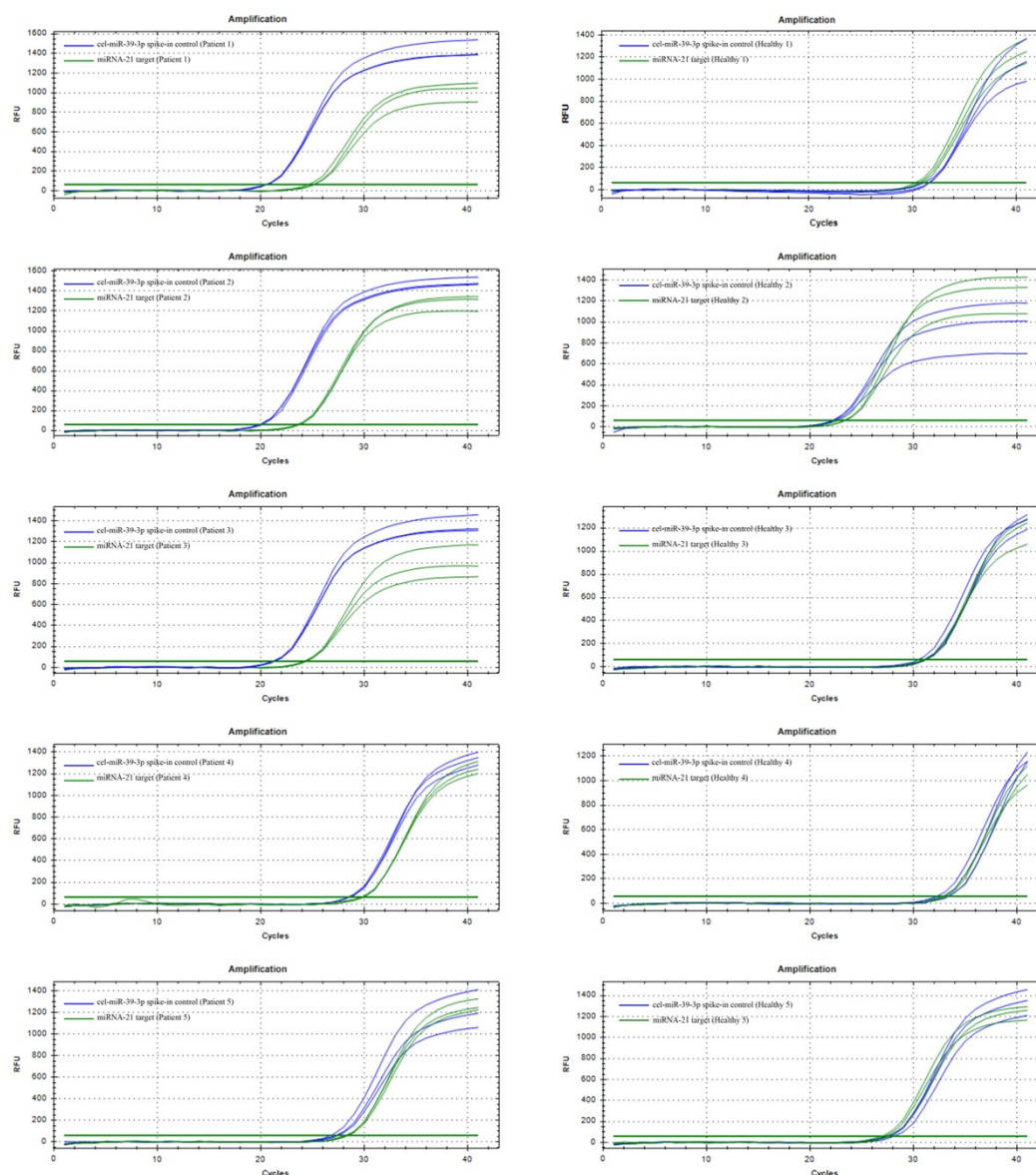
**Fig. S7 Specificity and anti-interference evaluation of the LiD-CHA system toward representative non-target miRNAs.** Net fluorescence intensities (F-F<sub>B</sub>) were measured for miRNA-21, miR-21-3p, miR-221-3p, miR-222-3p, miR-155-5p, and a mixed sample containing miRNA-21 together with miR-21-3p, miR-221-3p, miR-222-3p, and miR-155-5p. Error bars represent the standard deviation of three independent experiments.

**Fig. S8**



**Fi. S8 Raw RT-qPCR amplification curves of miRNA-21 in different cell lines.** Amplification curves obtained from RT-qPCR analysis of miRNA-21 in MCF-7, HeLa, HepG2 and THLE-2 cells. Blue curves represent the amplification of the internal reference gene U6, while green curves represent the amplification of miRNA-21.

**Fig. S9**



**Fig. S9 Raw RT-qPCR amplification curves of miRNA-21 in blood samples from cancer patients and healthy volunteers.** Amplification curves obtained from RT-qPCR analysis of miRNA-21 in blood samples from cancer patients and healthy volunteers. Blue curves represent the amplification of the spike-in control cel-miR-39-3p, while green curves represent the amplification of miRNA-21.

**Table S1. Recovery of miRNA-21 in spiked samples using the LiD-CHA system. (n=3)**

Spiked (pM)	Found (pM)	Recovery (%)	RSD (%)
80	78.7 ± 1.1	98.4	1.3
50	49.6 ± 0.3	99.2	0.7
30	31.6 ± 0.2	105.2	0.6
15	15.5 ± 0.4	103.4	2.3

