

## Supplementary Information

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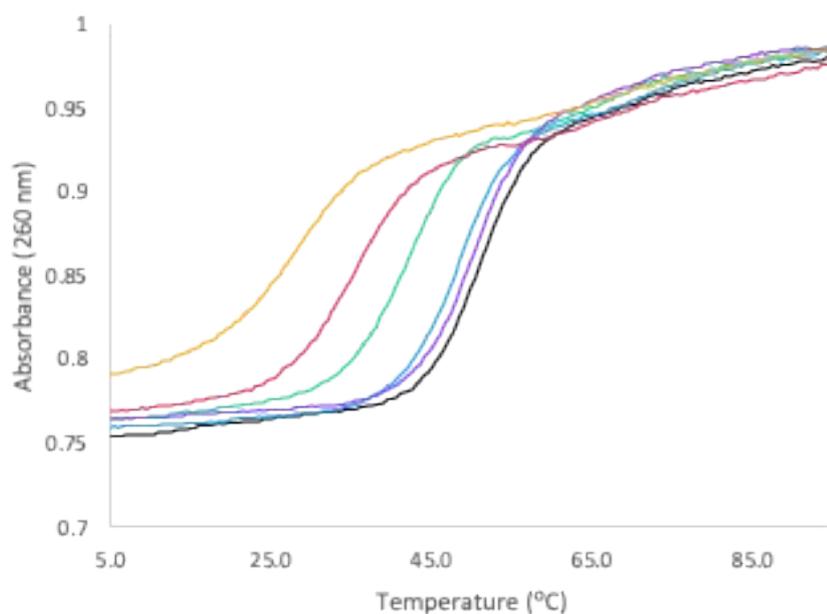
1. Supplementary data
2.  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and  $^{31}\text{P}$  NMR spectra of compounds
3. Characterization data (HPLC and mass data) of synthesized oligonucleotides

## 1. Supplementary data

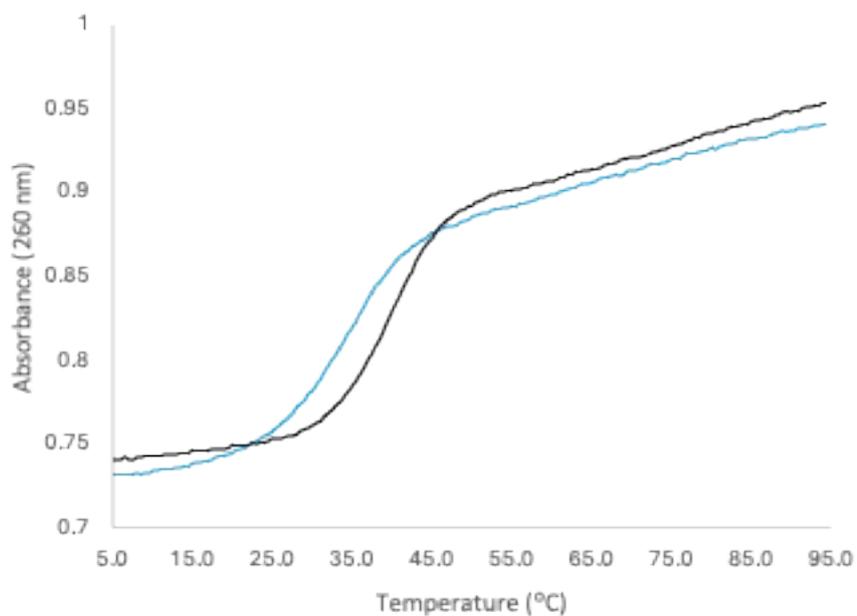
**Table S1.** Isolated yields of oligonucleotides

	Sequences (5'-3') <sup>a</sup>	Yield <sup>b</sup>
<b>ON1</b>	GCG TTT TTT GCT	24%
<b>ON2</b>	GCG TTT TTT GCT	39%
<b>ON3</b>	GCG TTT TTT GCT	33%
<b>ON4</b>	GCG TTT TTT GCT	34%
<b>ON5</b>	GCG <b>TTT</b> TTT GCT	21%
<b>ON6</b>	GCG TTT <b>TTT</b> GCT	24%
<b>ON7</b>	GCG TTC TTT GCT	44%
<b>ON8</b>	GCG TTC <b>TTT</b> GCT	16%
<b>ON9</b>	TTT TTT TTT T	23%
<b>ON10</b>	TTT TTT TTT <b>T</b>	32%
<b>ON11</b>	TTT TTT TTT <b>T</b>	23%
<b>ON12</b>	TTT TTT TTT T	14%

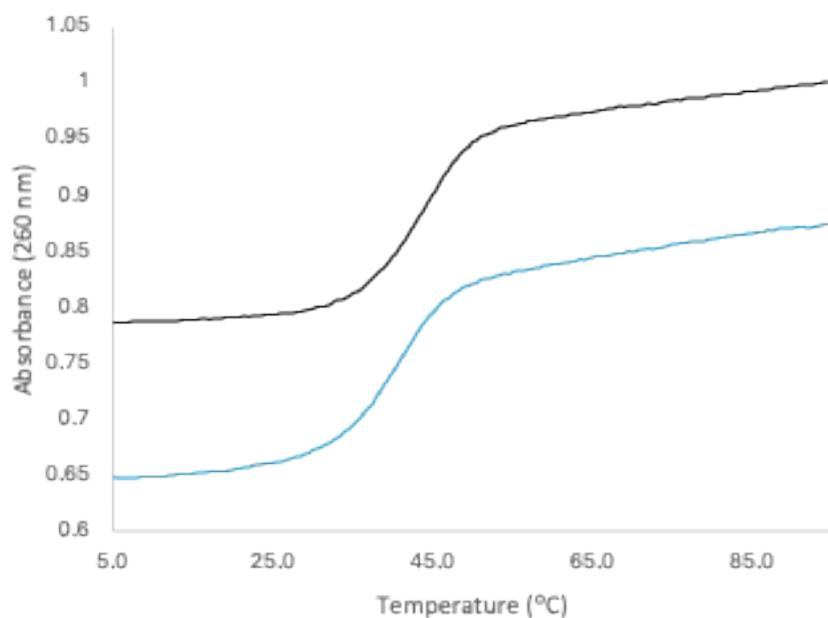
<sup>a</sup> Capital letters and Bold letters indicate DNA and L-lyxo-thioBsNA, respectively. Mark “^” indicate phosphorothioate. <sup>b</sup> The isolated yield for **ON1–ON12** were calculated from the UV absorbance at 260 nm.



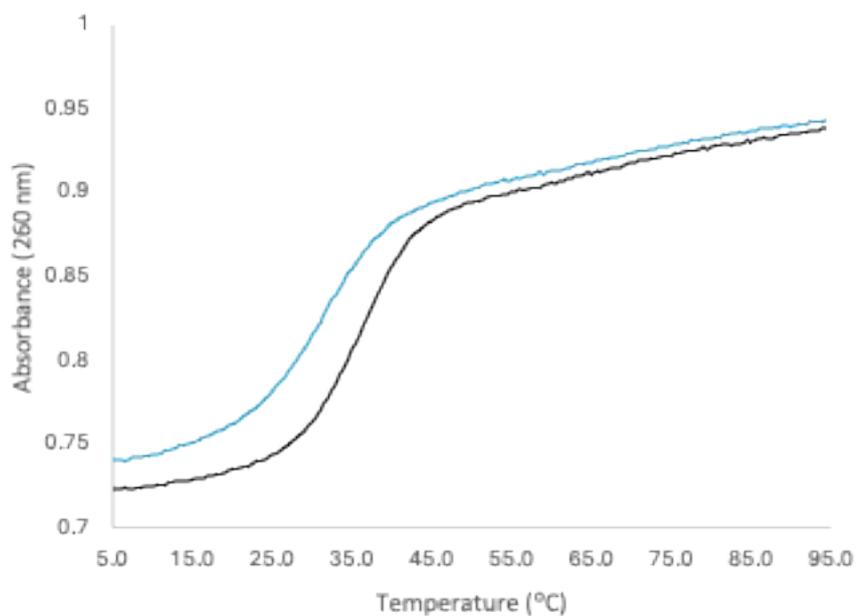
**Figure S1.** UV melting studies for ONs against complementary DNA. **ON1** black, **ON2** blue, **ON3** green, **ON4** violet, **ON5** pink, and **ON6** orange. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



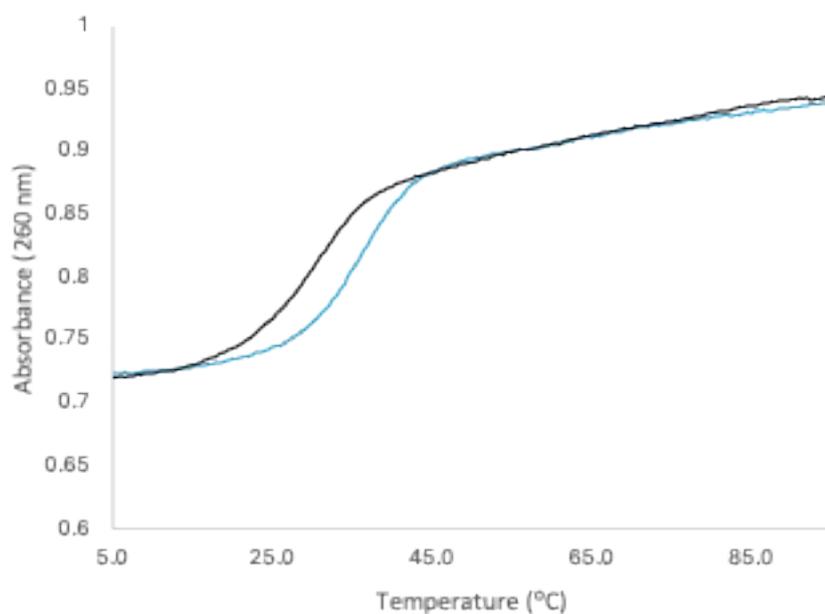
**Figure S2.** UV melting studies for ONs against G mismatch DNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



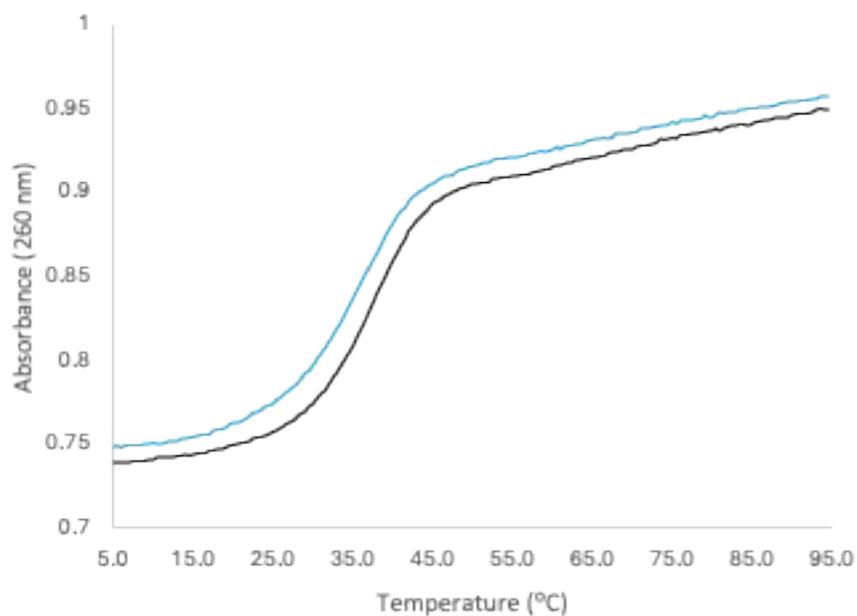
**Figure S3.** UV melting studies for ONs against G mismatch RNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



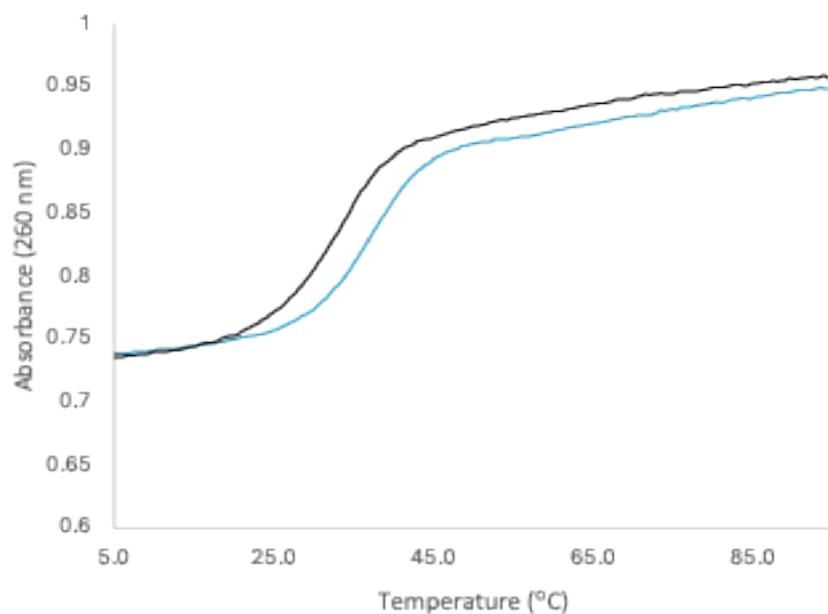
**Figure S4.** UV melting studies for ONs against C mismatch DNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



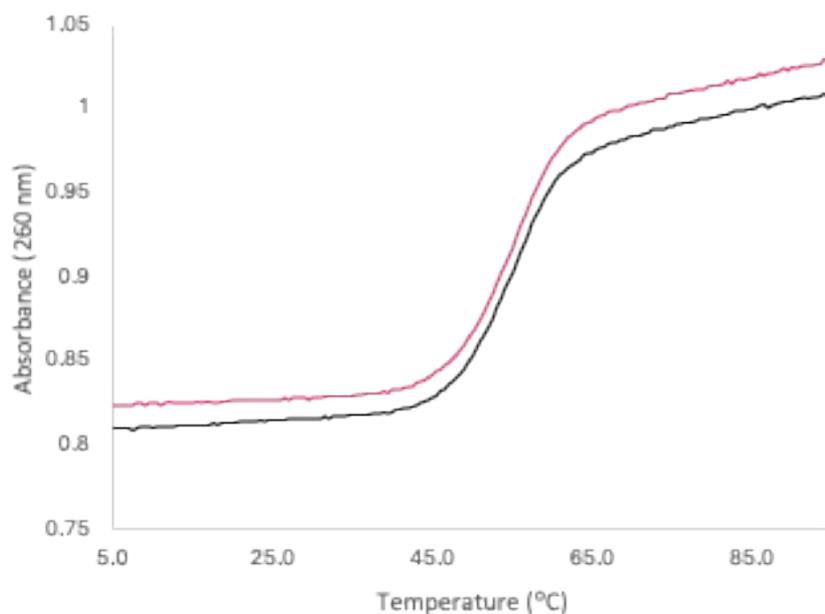
**Figure S5.** UV melting studies for ONs against C mismatch RNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



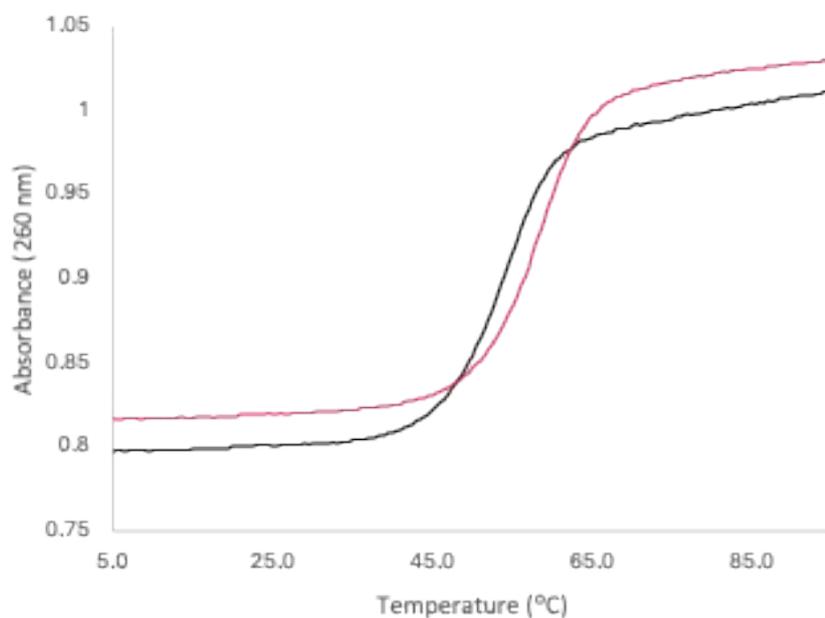
**Figure S6.** UV melting studies for ONs against T mismatch DNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



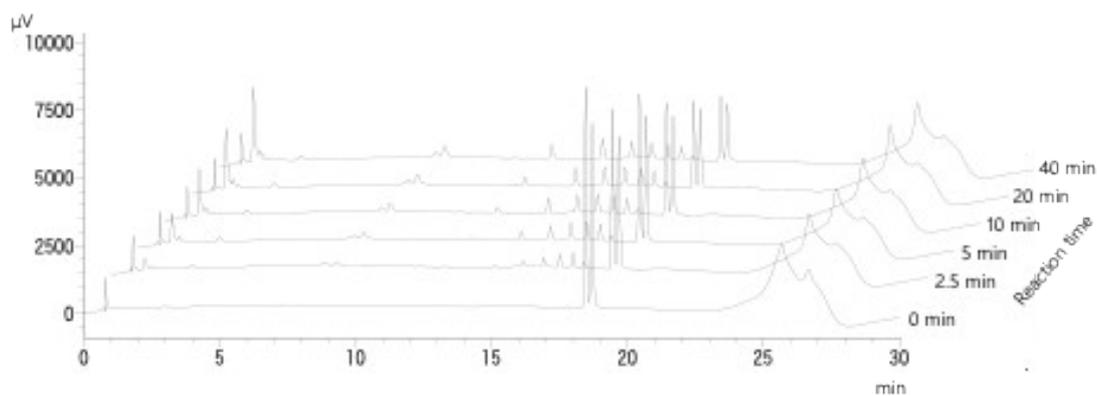
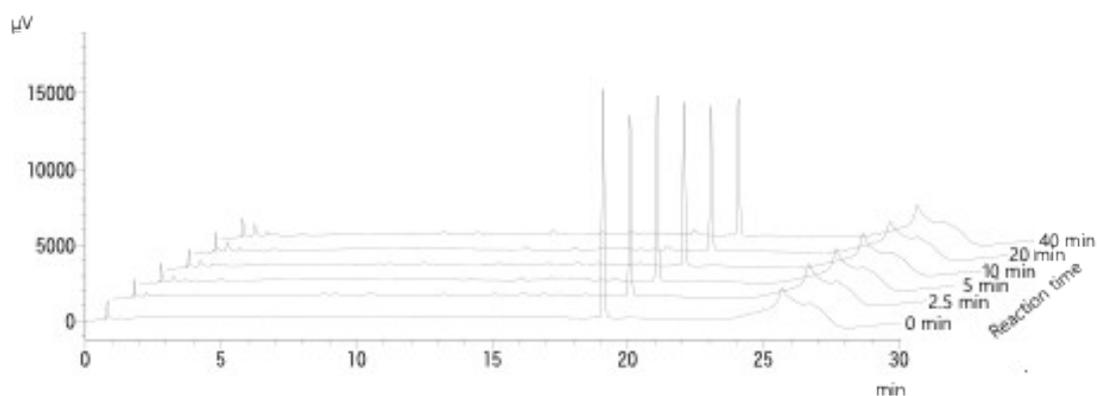
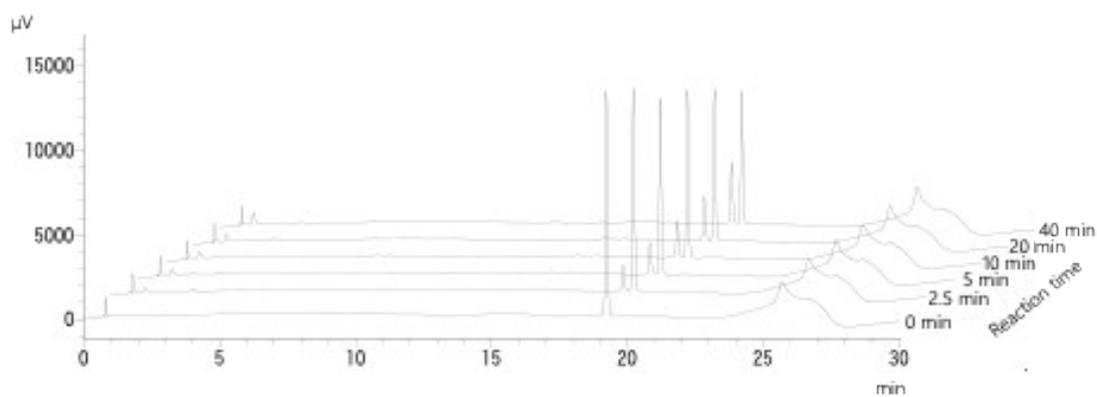
**Figure S7.** UV melting studies for ONs against U mismatch RNA. **ON1** black, and **ON2** blue. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



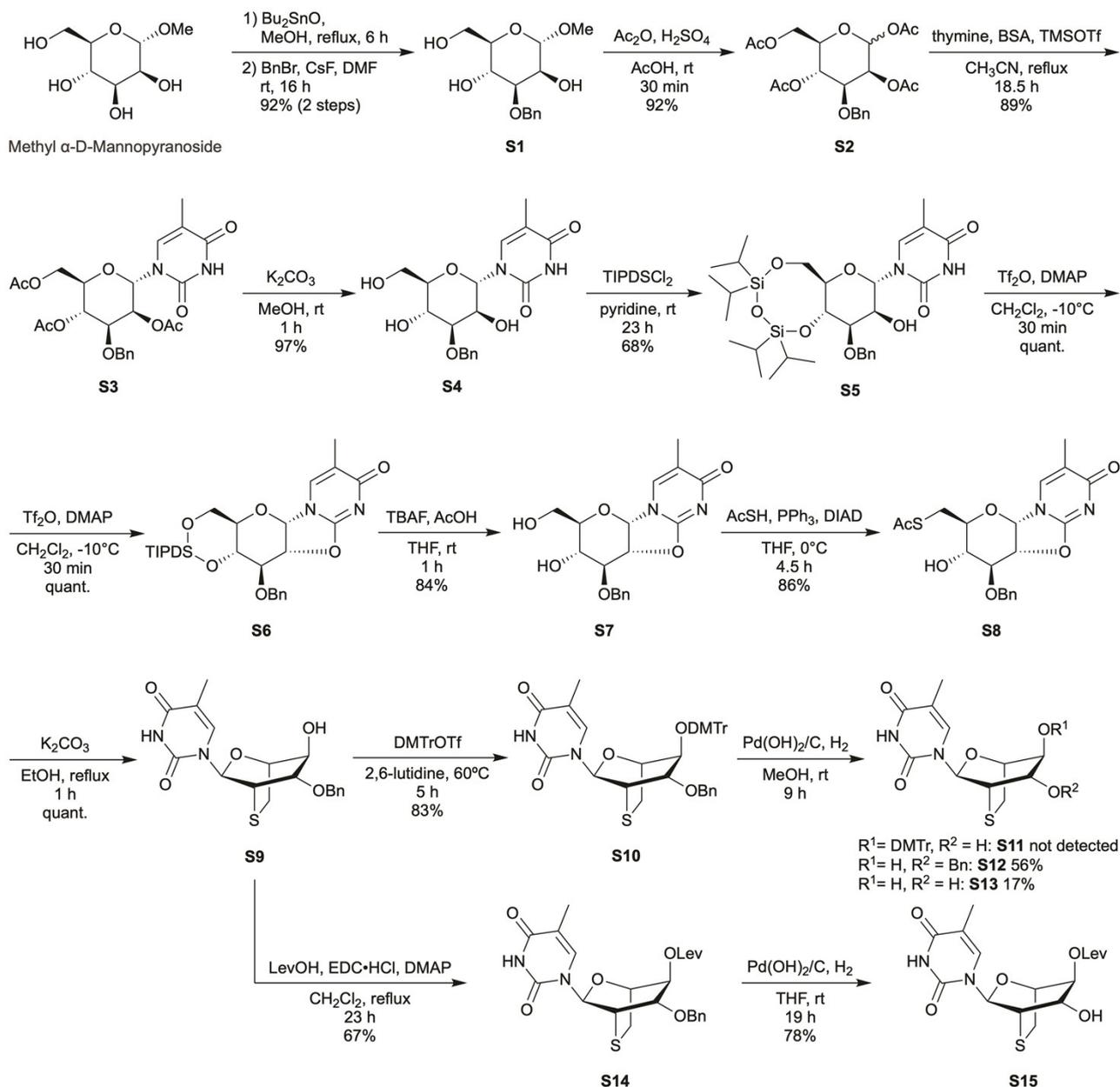
**Figure S8.** UV melting studies for ONs against complementary DNA. **ON7** black, and **ON8** pink. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.



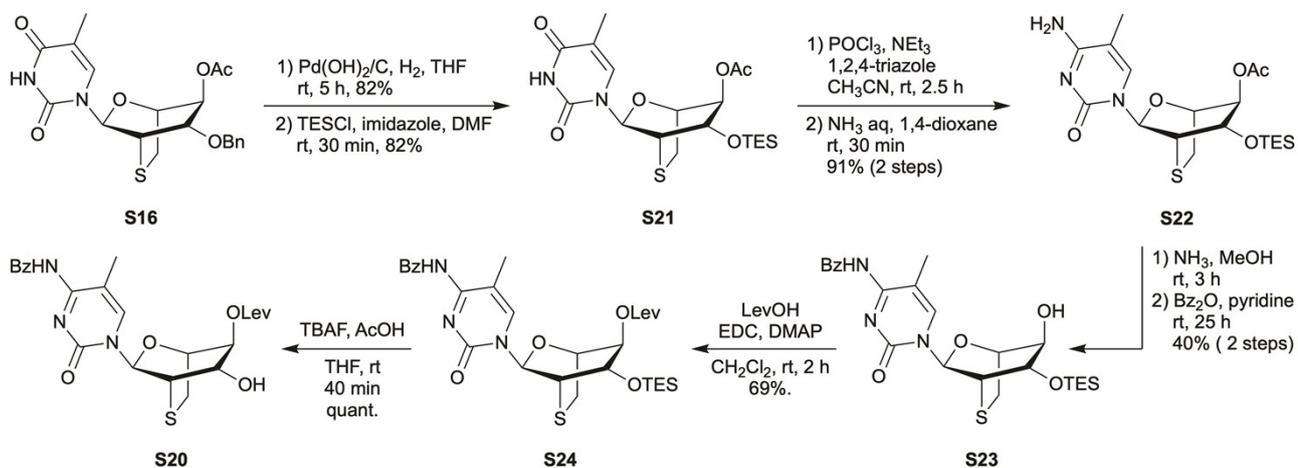
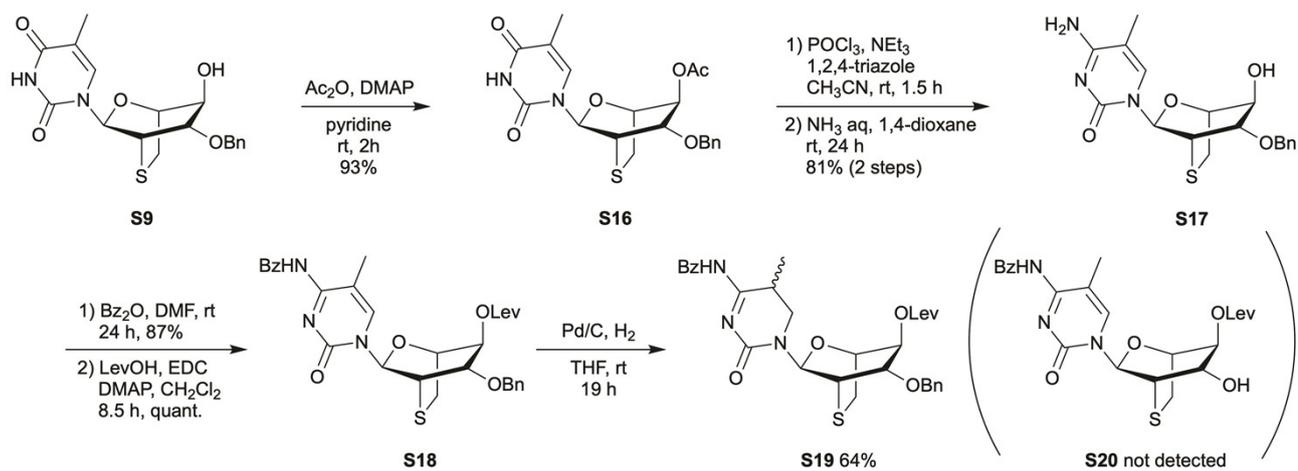
**Figure S9.** UV melting studies for ONs against complementary RNA. **ON7** black, and **ON8** pink. Conditions: 10 mM sodium phosphate (pH 7.2), 100 mM NaCl, 4.0  $\mu$ M of each oligonucleotide, and 4.0  $\mu$ M of complementary RNA. The curves shown are representative of three independent repeats, each consisting of at least three technical repeats.

**ON10****ON11****ON12**

**Figure S10.** Reverse-phase HPLC analysis of the nuclease stability of **ON10**, **ON11**, and **ON12** against svPDE.. Conditions: 50 mM Tris-HCl (pH 8.0), 10 mM  $\text{MgCl}_2$ , 4.0  $\mu\text{M}$  of each oligonucleotide, and 25  $\mu\text{M}$  svPDE at 37 °C. The samples were analyzed using a reverse-phase HPLC column (Waters XBridge Oligonucleotide BEH C18 column 2.5  $\mu\text{m}$ , 4.6 x 50 mm; eluent A (0.1 M TEAA buffer) and eluent B (MeOH), eluted with a linear gradient (5%–25% of eluent B in 20 min) and UV detection at 260 nm.



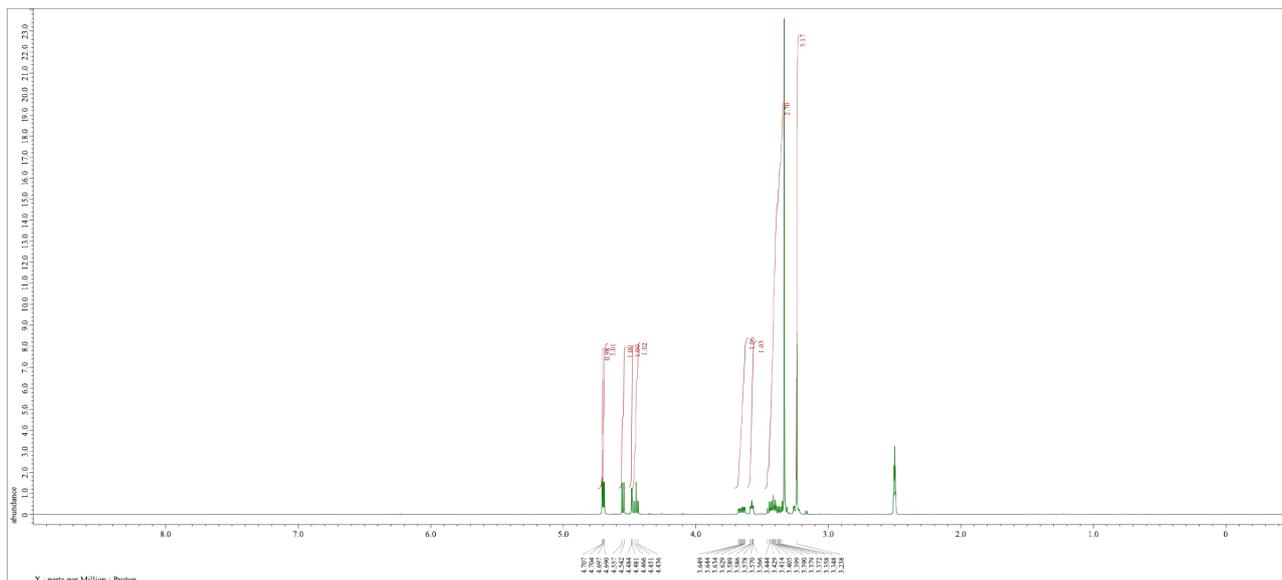
**Scheme S1.** Synthesis of *D*-lyxo-thioBsNA-T phosphoroamidite precursor



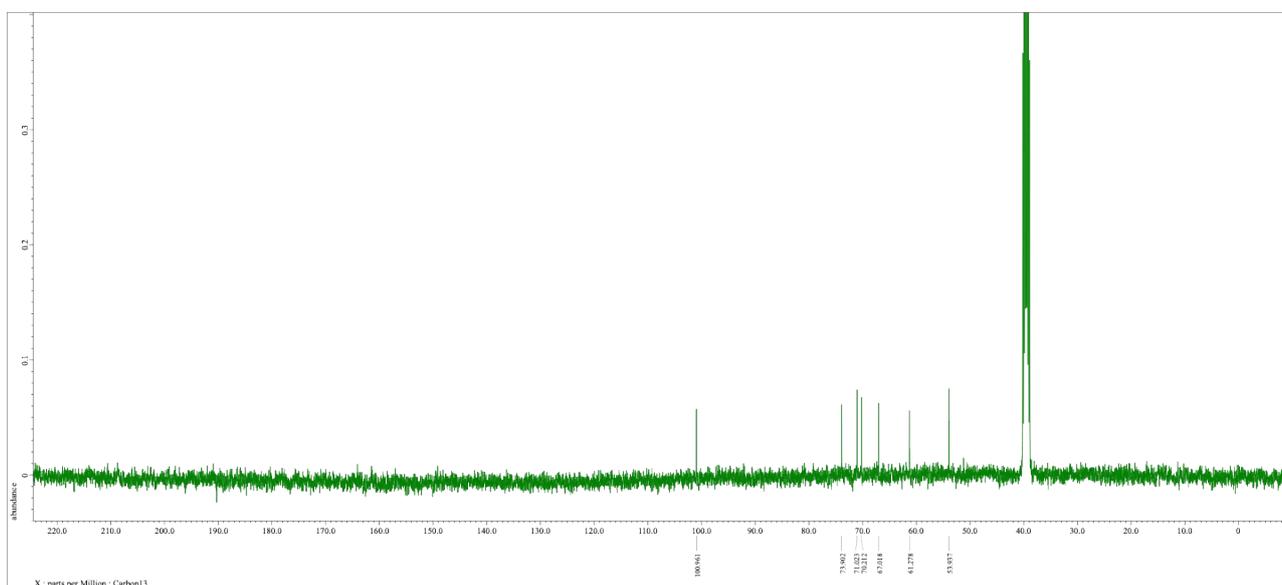
**Scheme S2.** Synthesis of *D*-lyxo-thioBsNA-meC phosphoroamidite precursor

## 2. $^1\text{H}$ NMR, $^{13}\text{C}$ NMR and $^{31}\text{P}$ NMR spectra of compounds

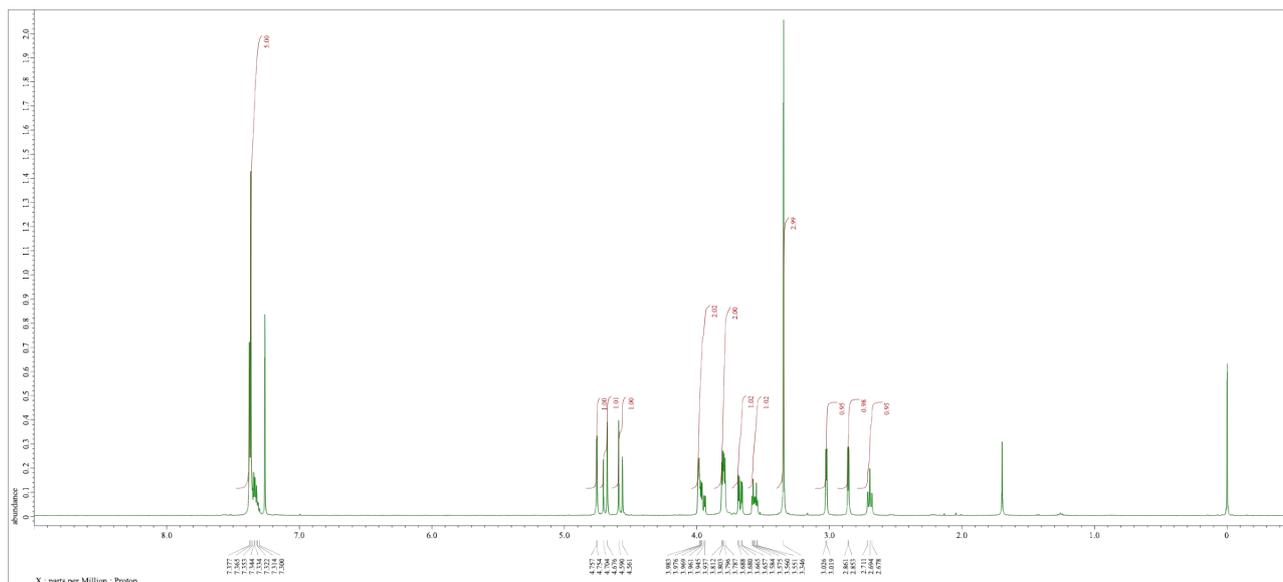
Compound 1 ( $^1\text{H}$ -NMR, DMSO- $\text{D}_6$ , 400 MHz)



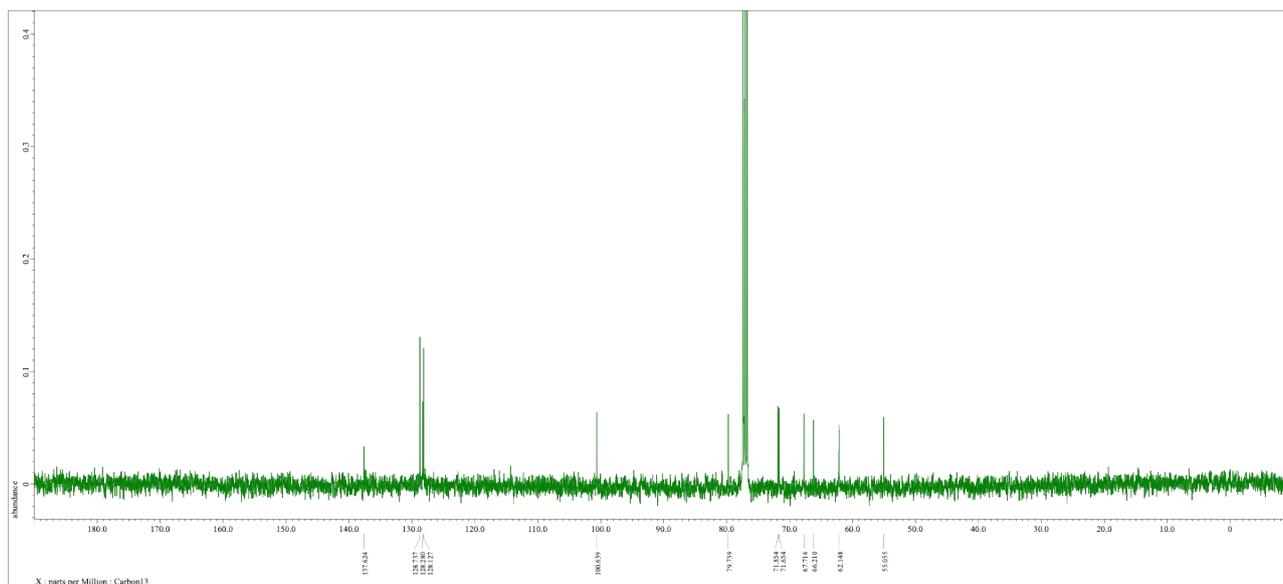
Compound 1 ( $^{13}\text{C}$ -NMR, DMSO- $\text{D}_6$ , 101 MHz)



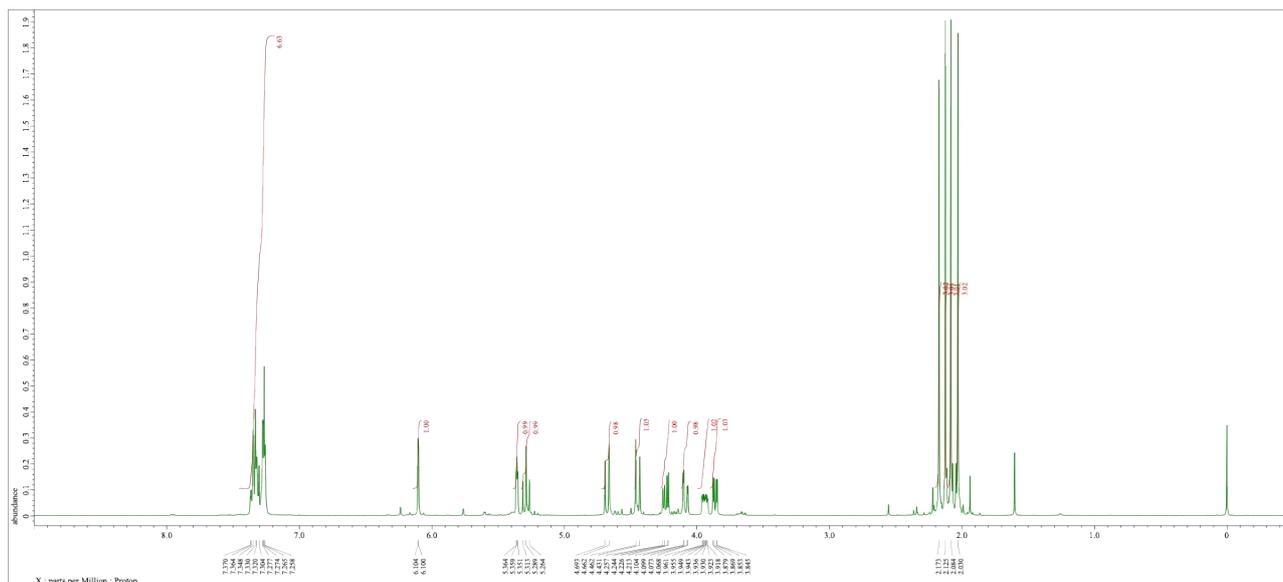
Compound 2 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



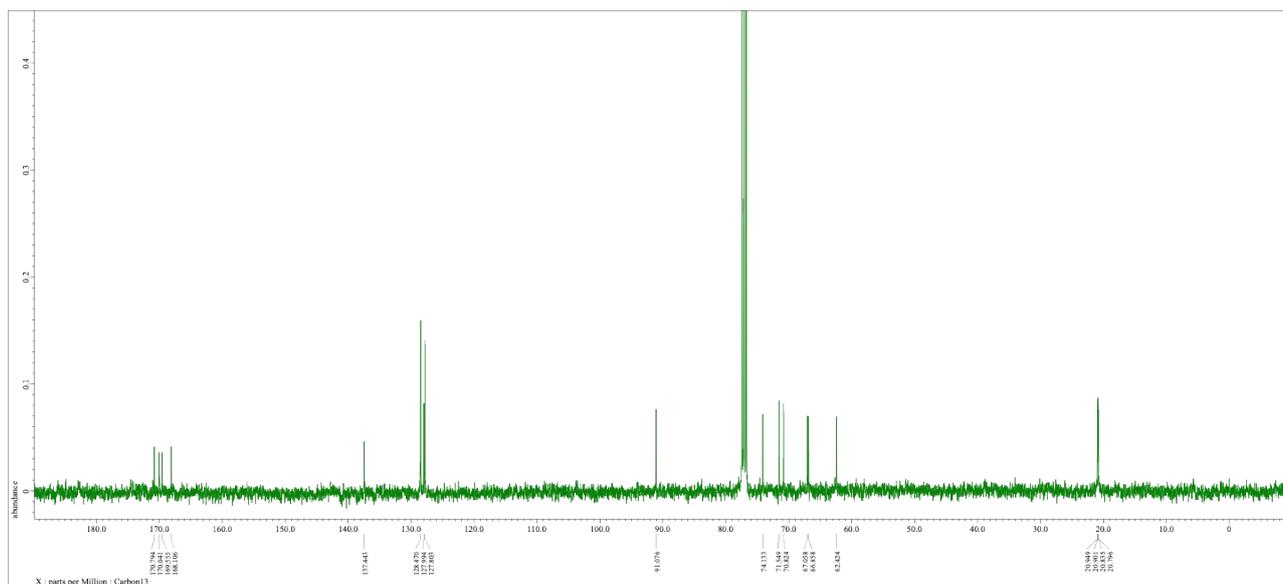
Compound 2 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



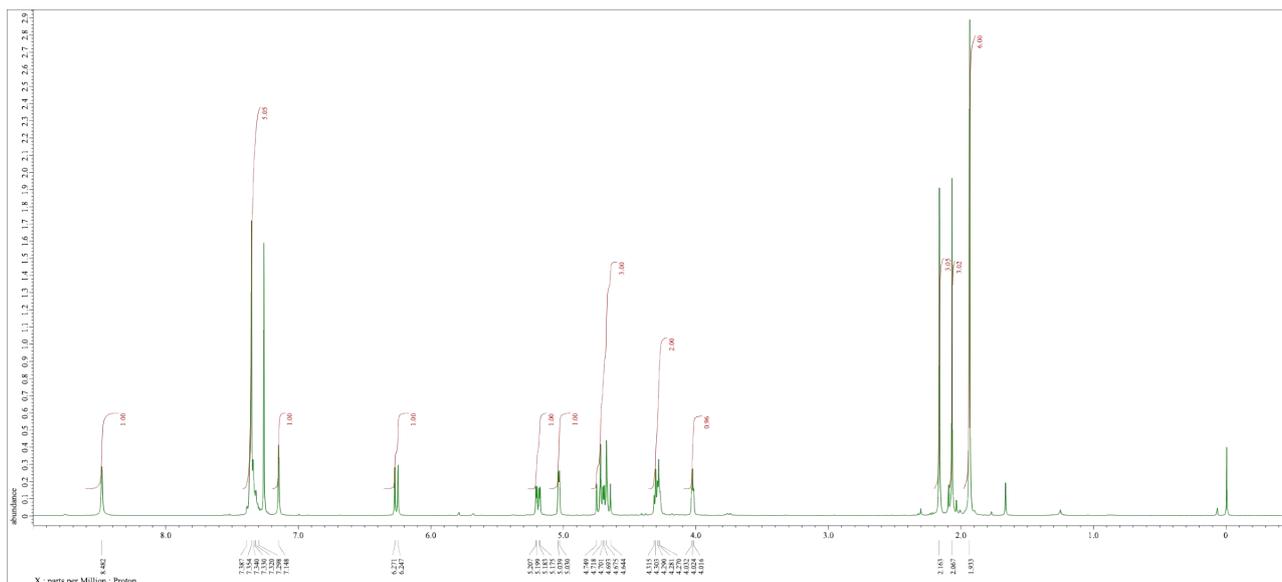
Compound **3** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



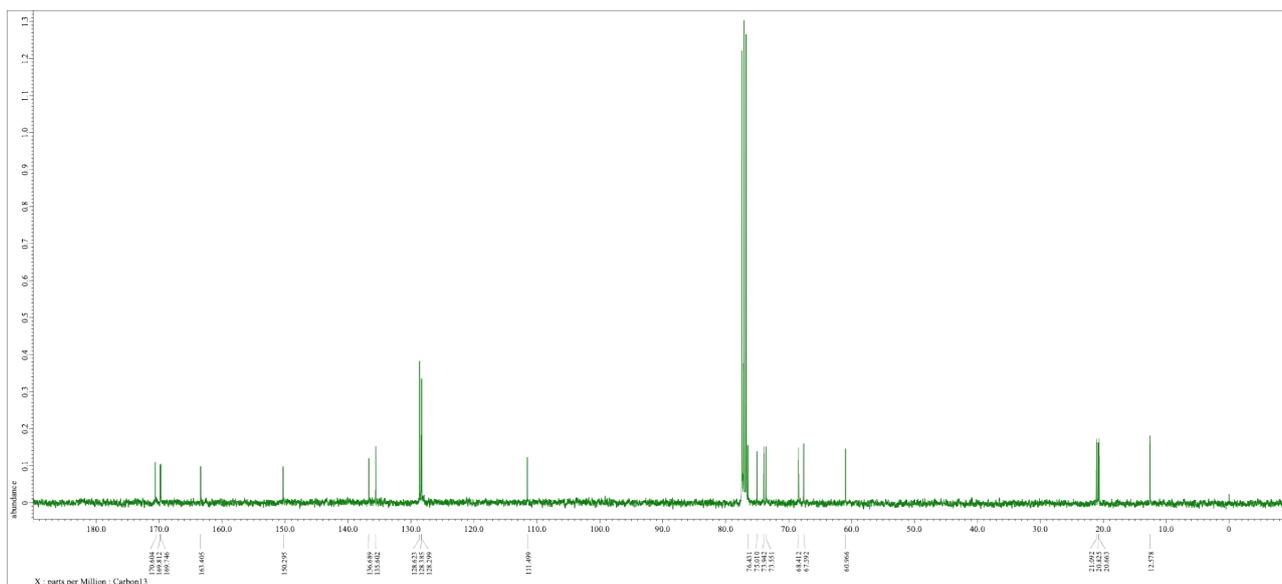
Compound **3** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



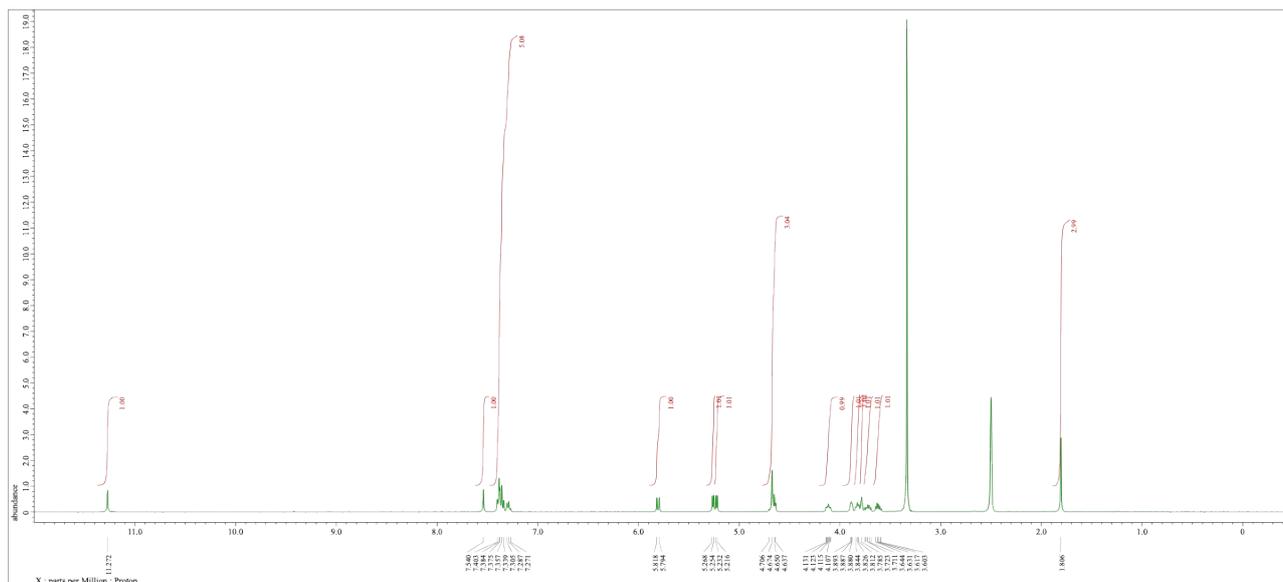
Compound 4 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



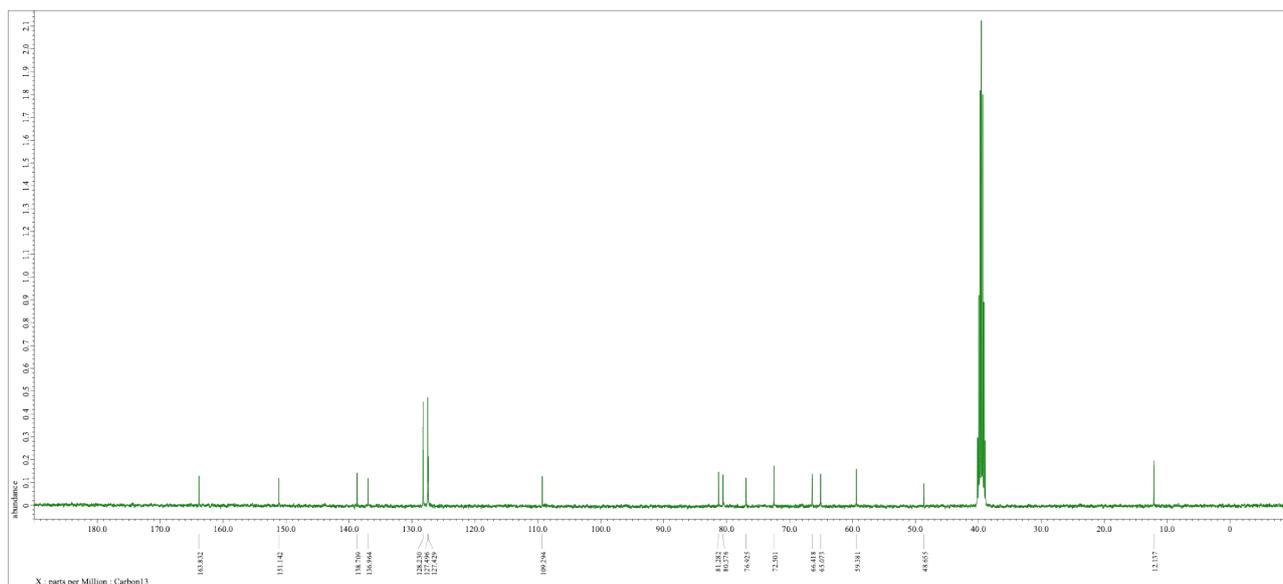
Compound 4 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



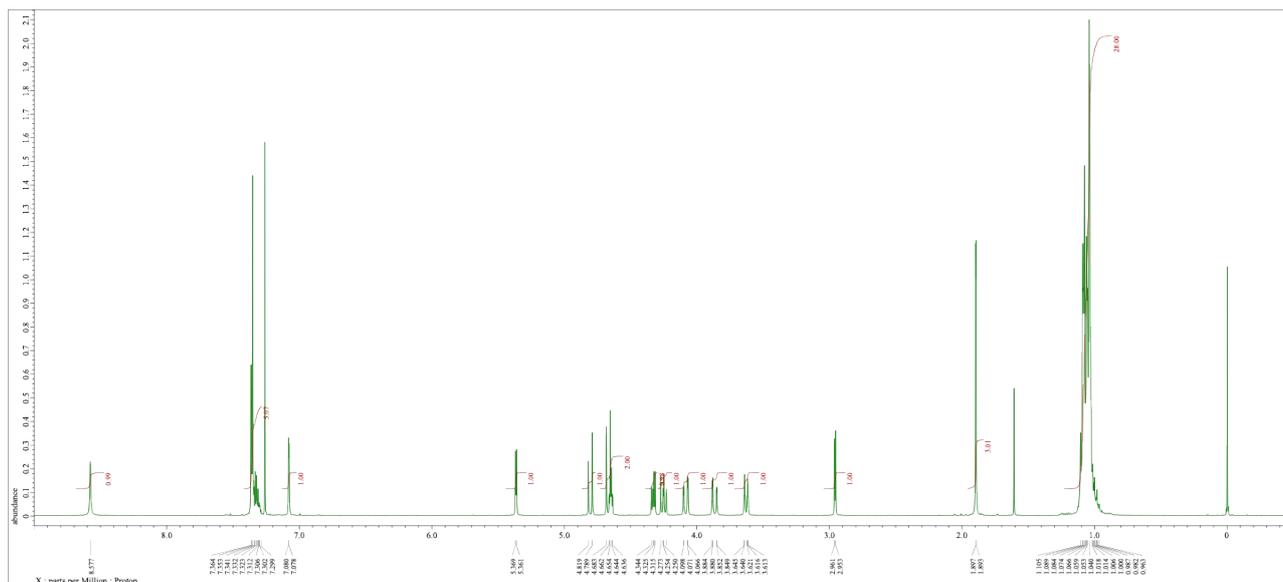
Compound 5 (<sup>1</sup>H-NMR, DMSO-D<sub>6</sub>, 400 MHz)



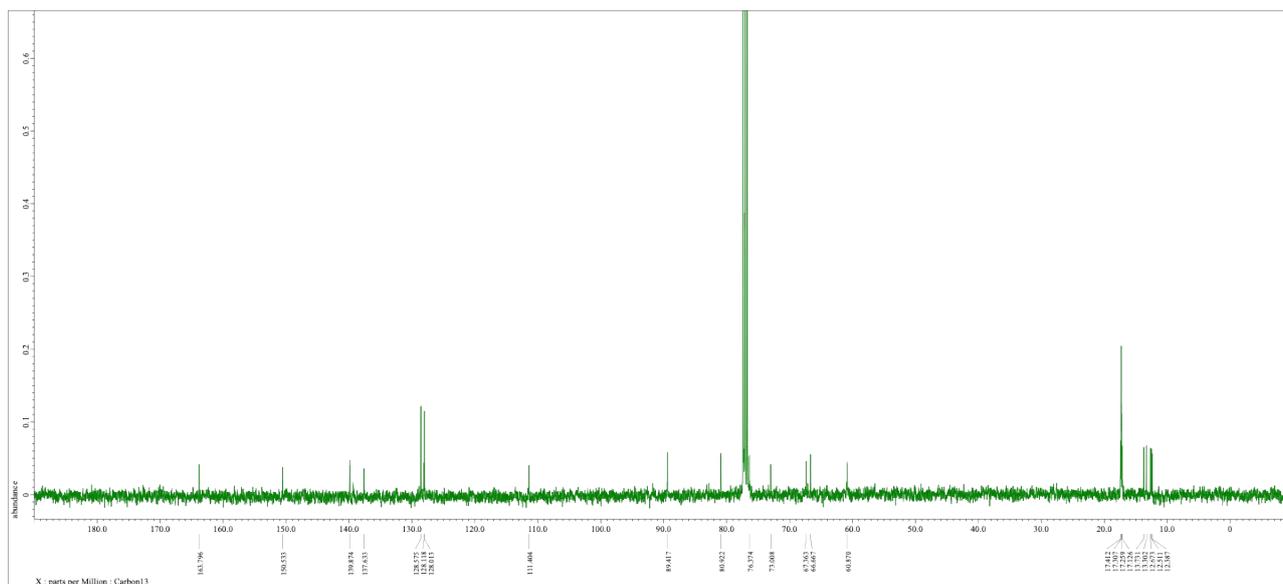
Compound 5 (<sup>13</sup>C-NMR, DMSO-D<sub>6</sub>, 101 MHz)



Compound 6 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)

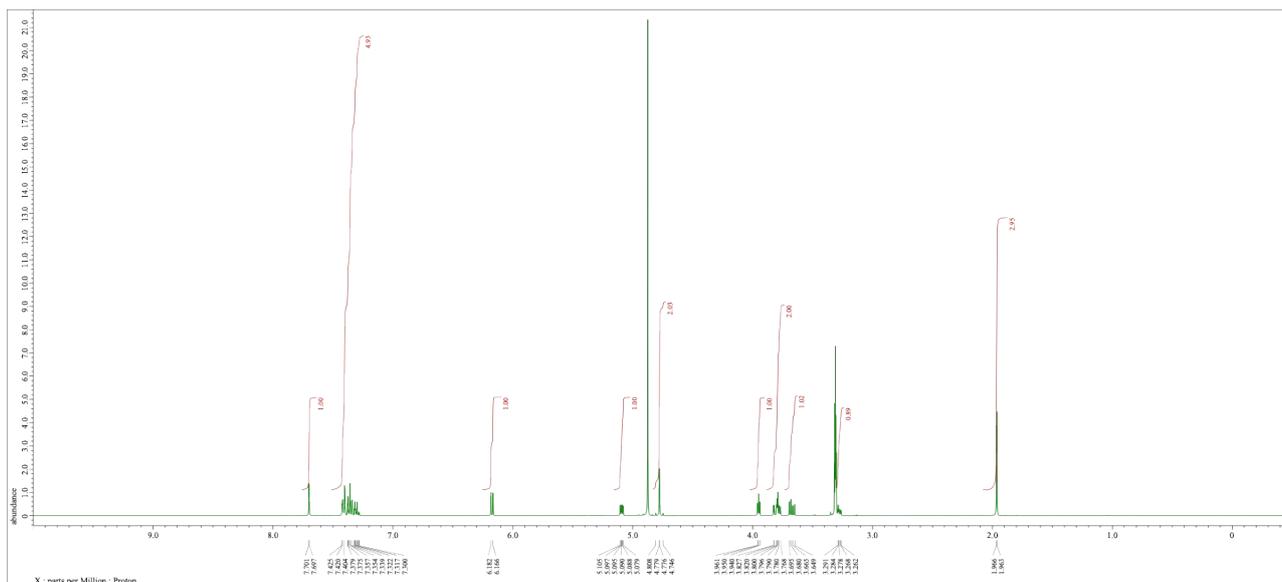


Compound 6 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)

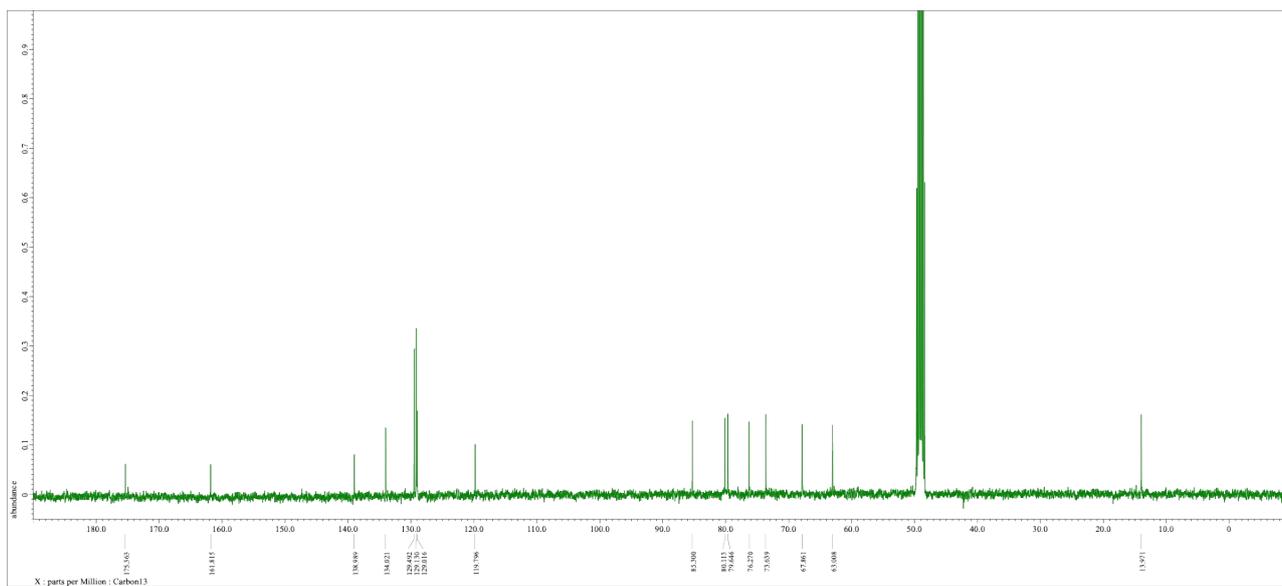




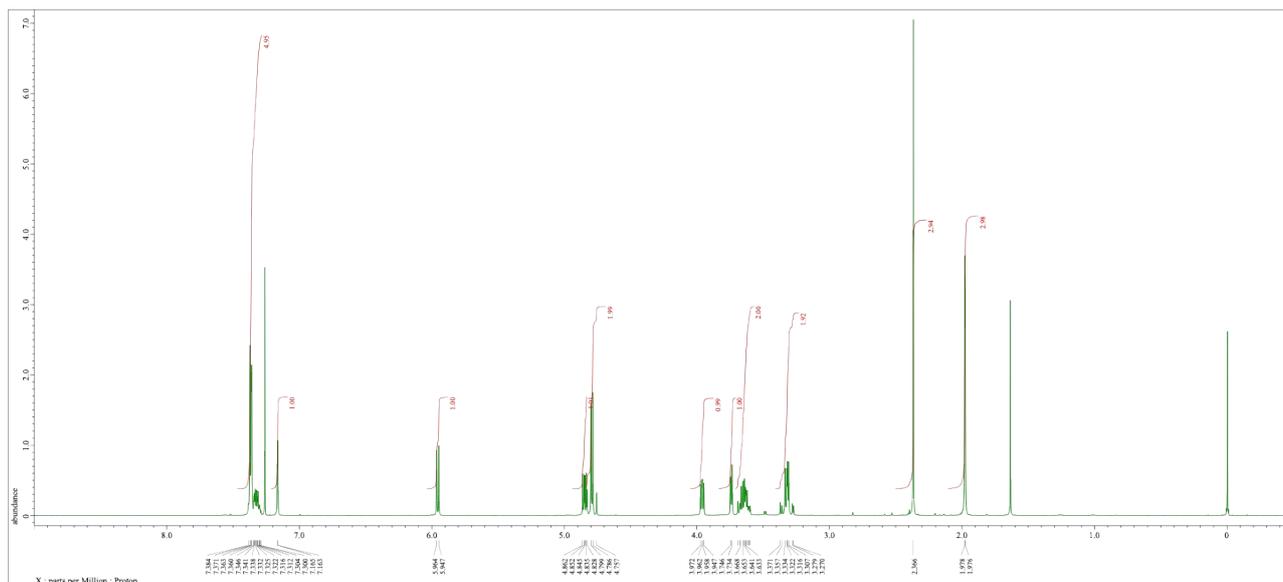
Compound **8** ( $^1\text{H-NMR}$ , MeOD-D<sub>4</sub>, 400 MHz)



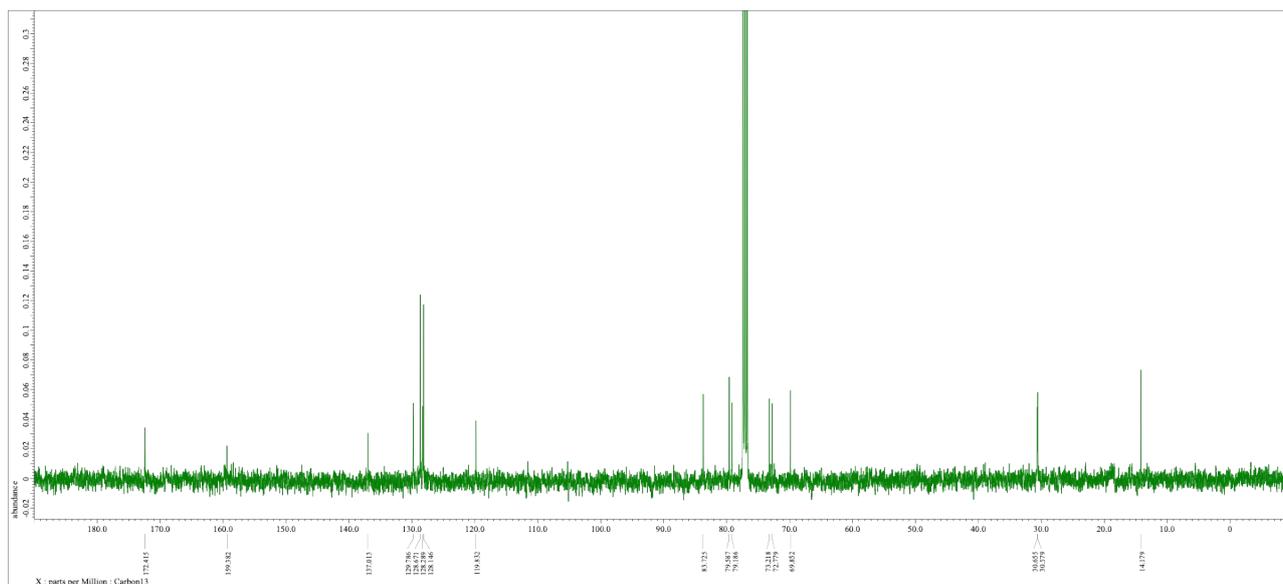
Compound **8** ( $^{13}\text{C-NMR}$ , MeOD-D<sub>4</sub>, 101 MHz)



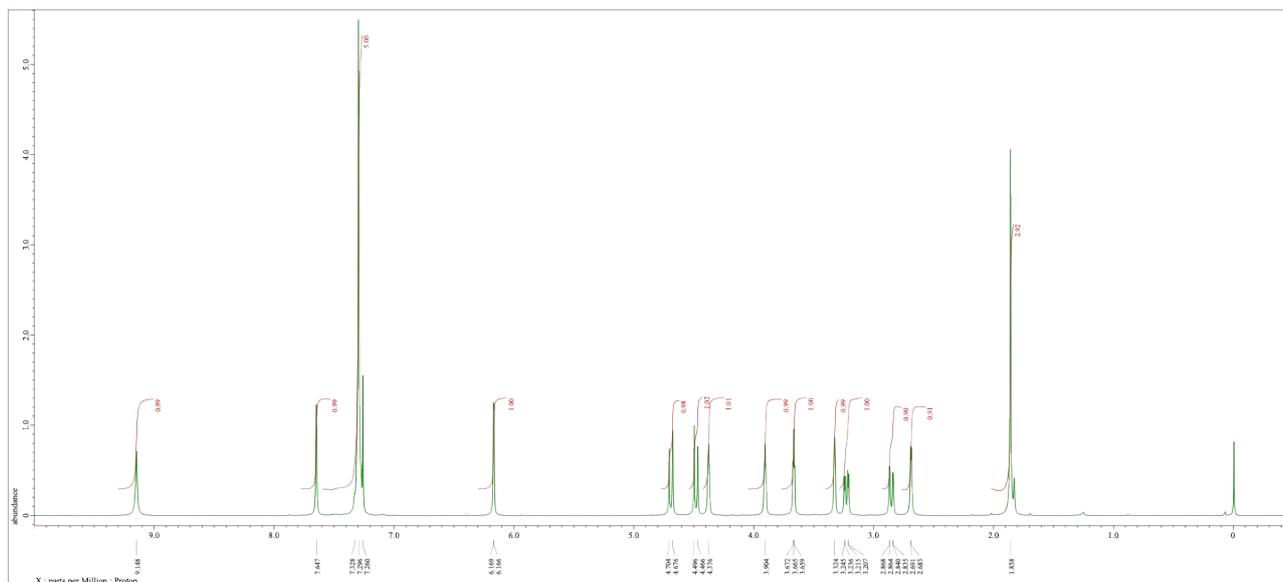
Compound 9 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



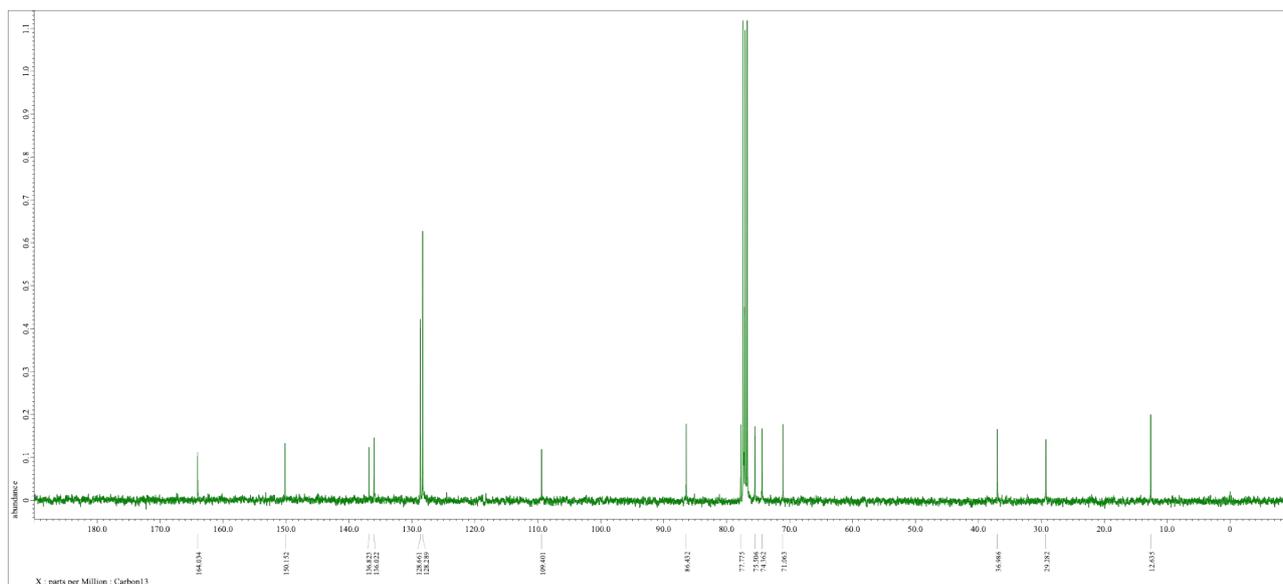
Compound 9 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



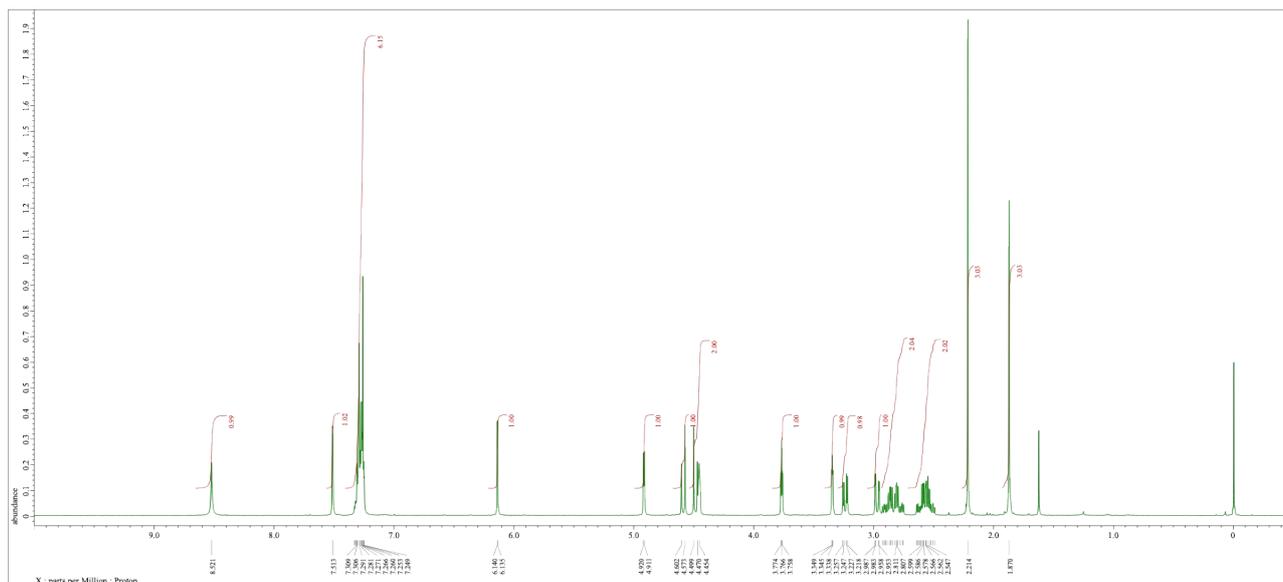
Compound **10** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



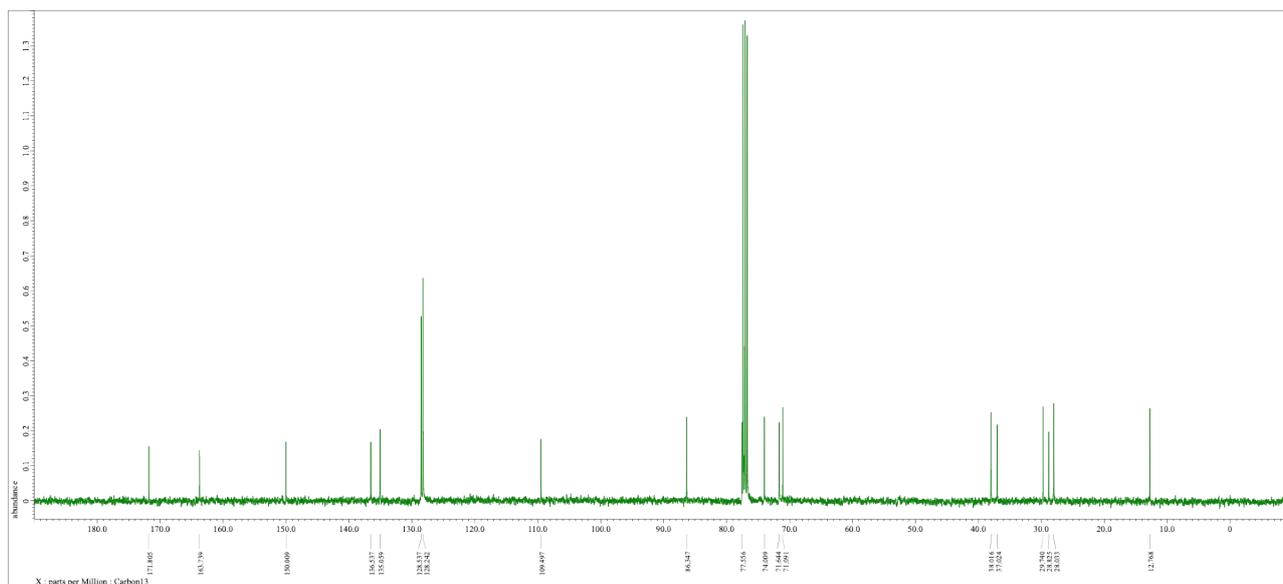
Compound **10** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



Compound **11** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)

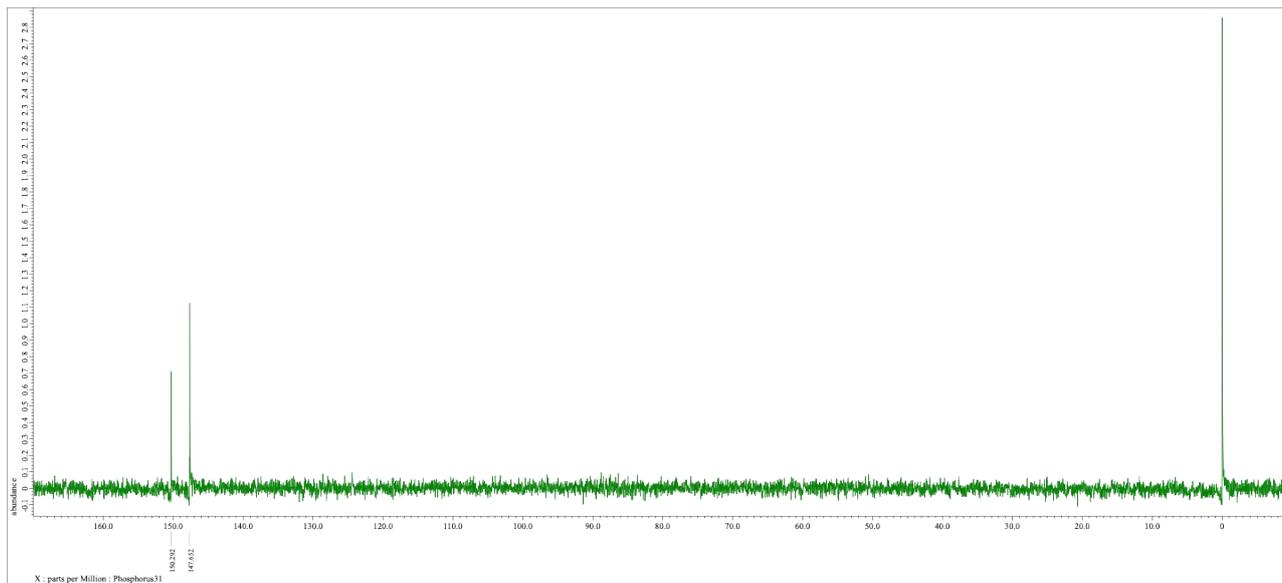


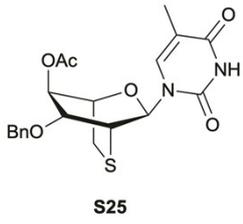
Compound **11** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



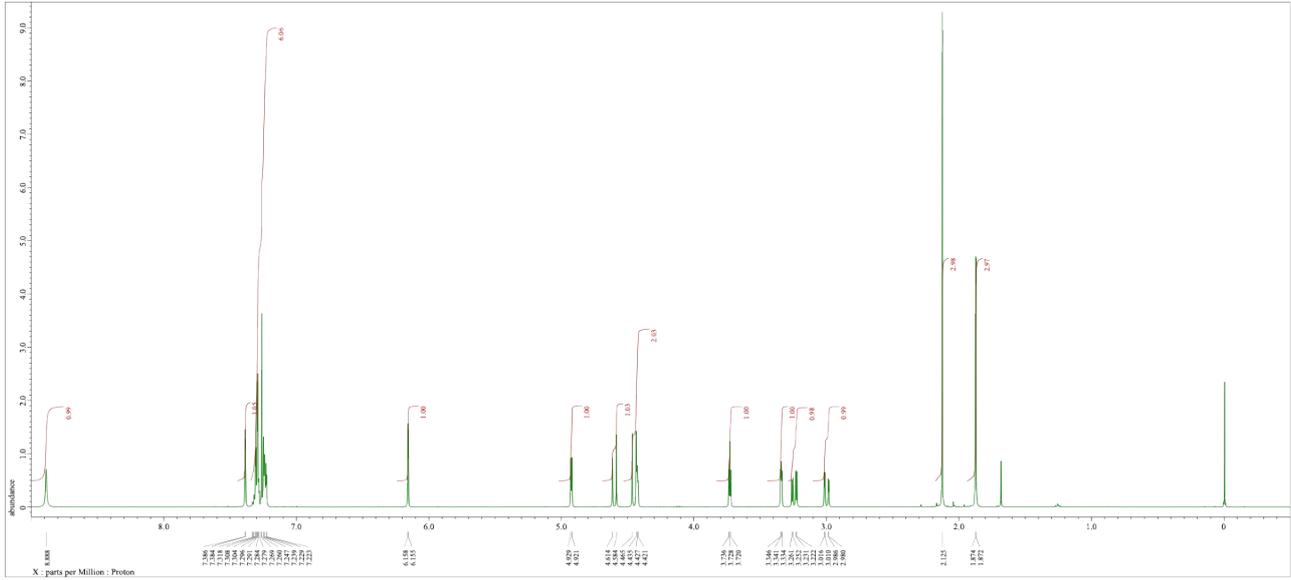


Compound **13** ( $^{31}\text{P}$ -NMR,  $\text{CDCl}_3$ , 162 MHz)

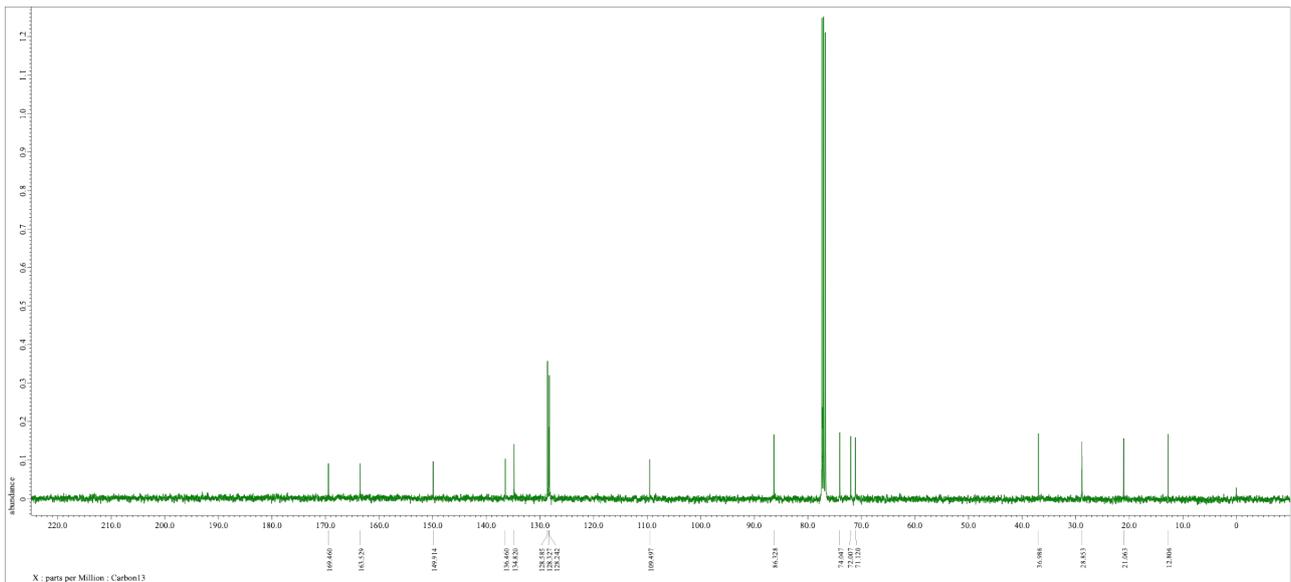


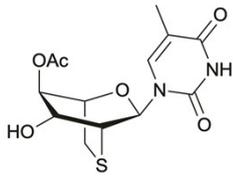


Compound **S25** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



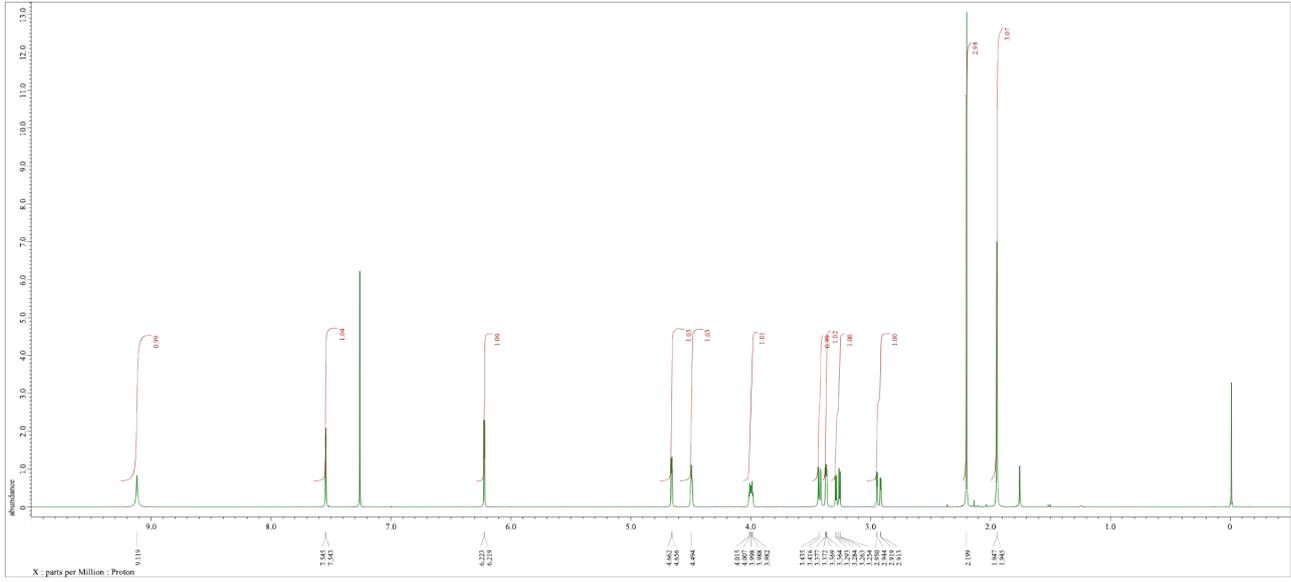
Compound **S25** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



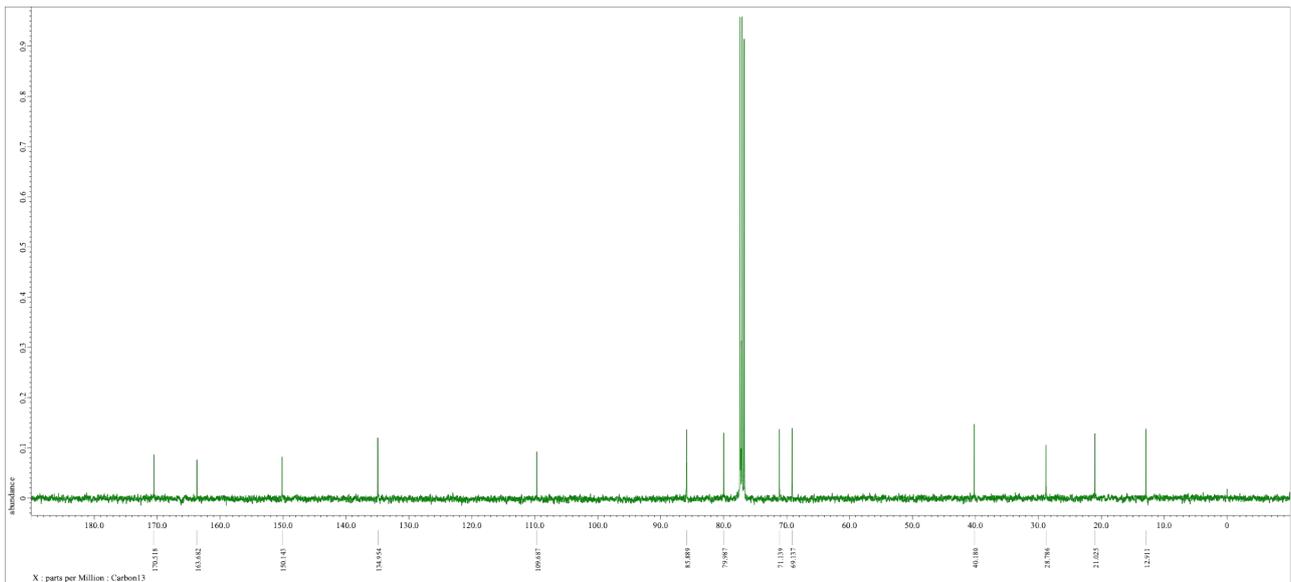


**S26**

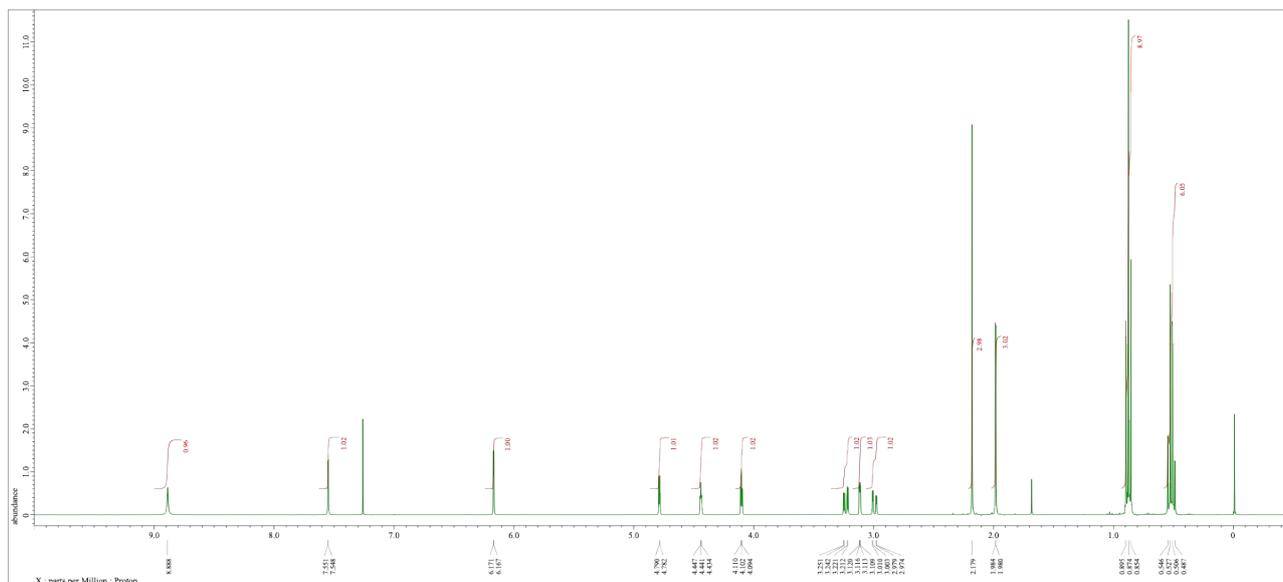
Compound **S26** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



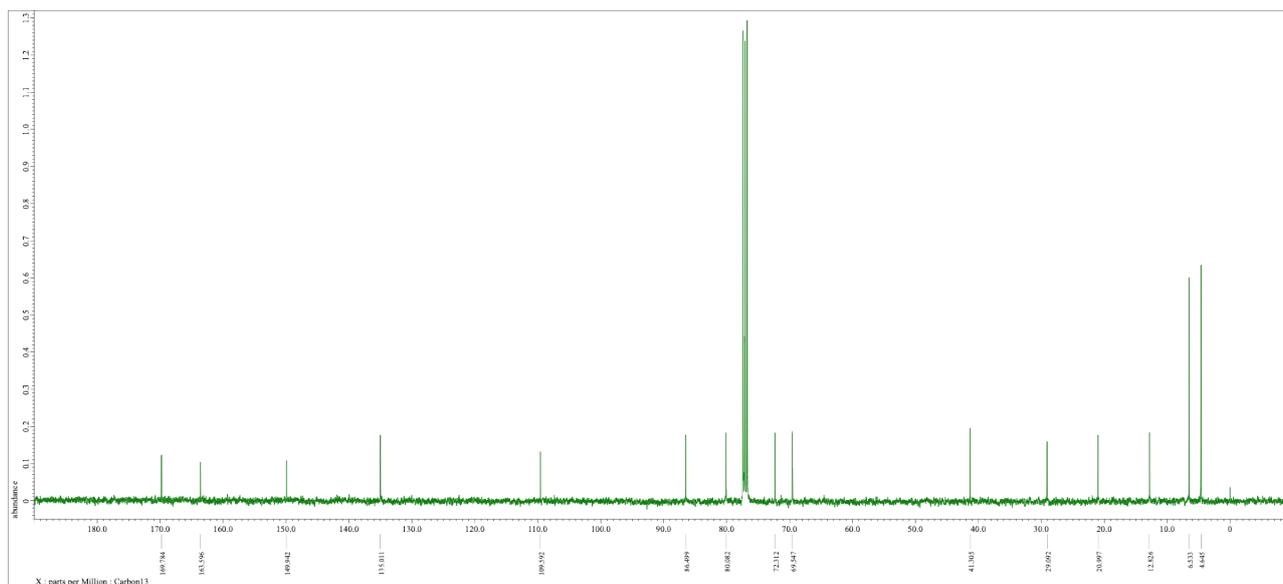
Compound **S26** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



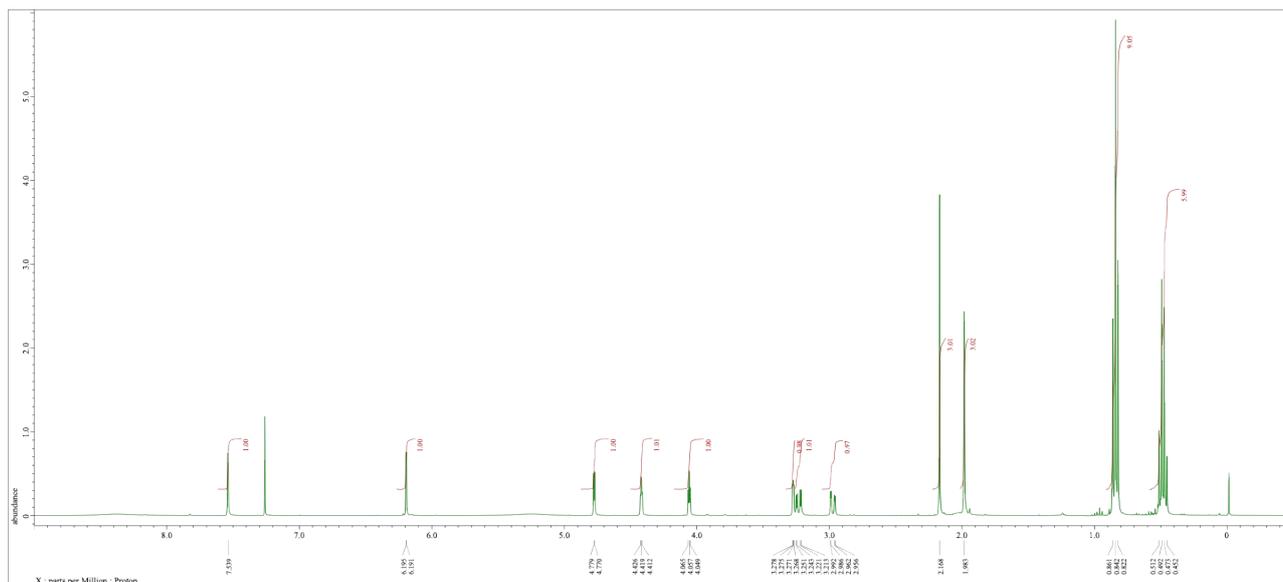
Compound **14** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



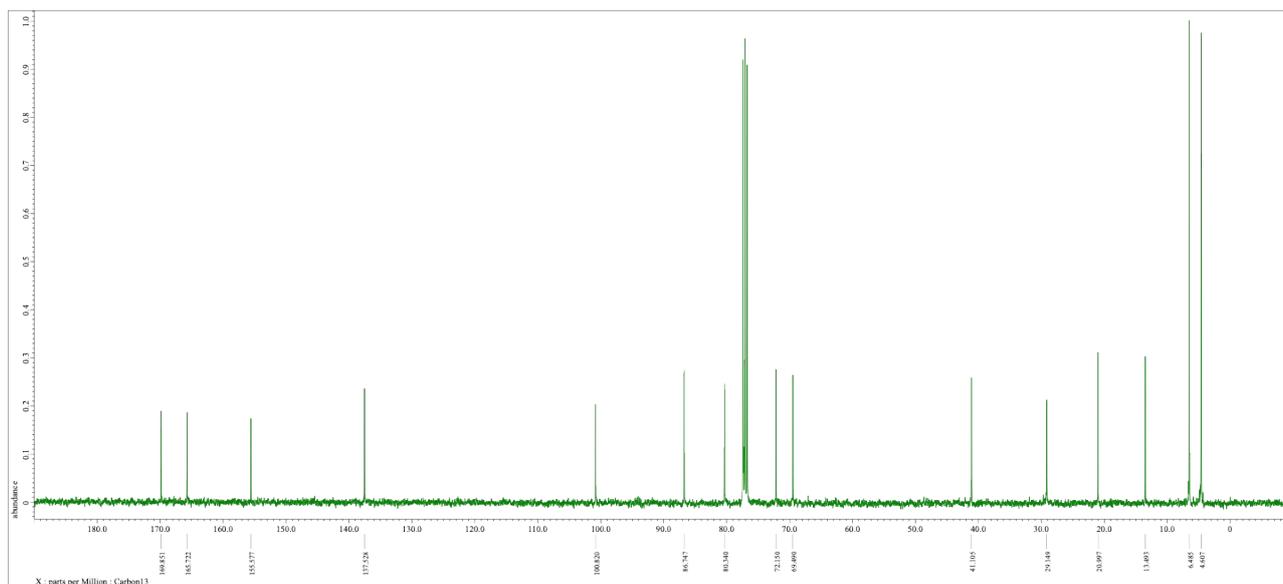
Compound **14** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



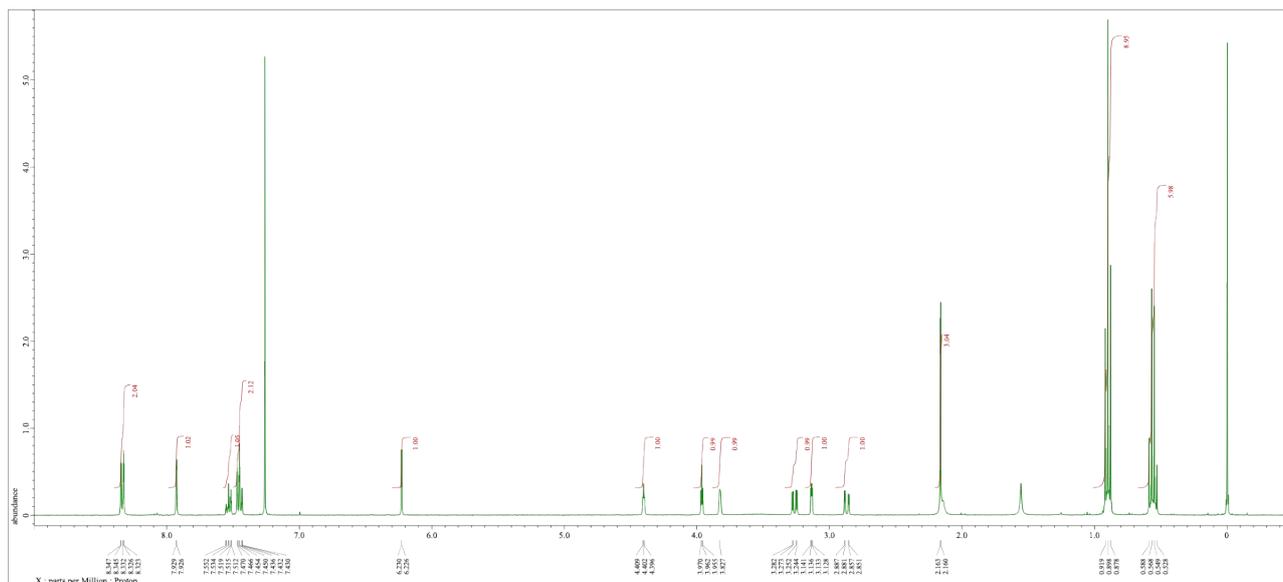
Compound 15 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



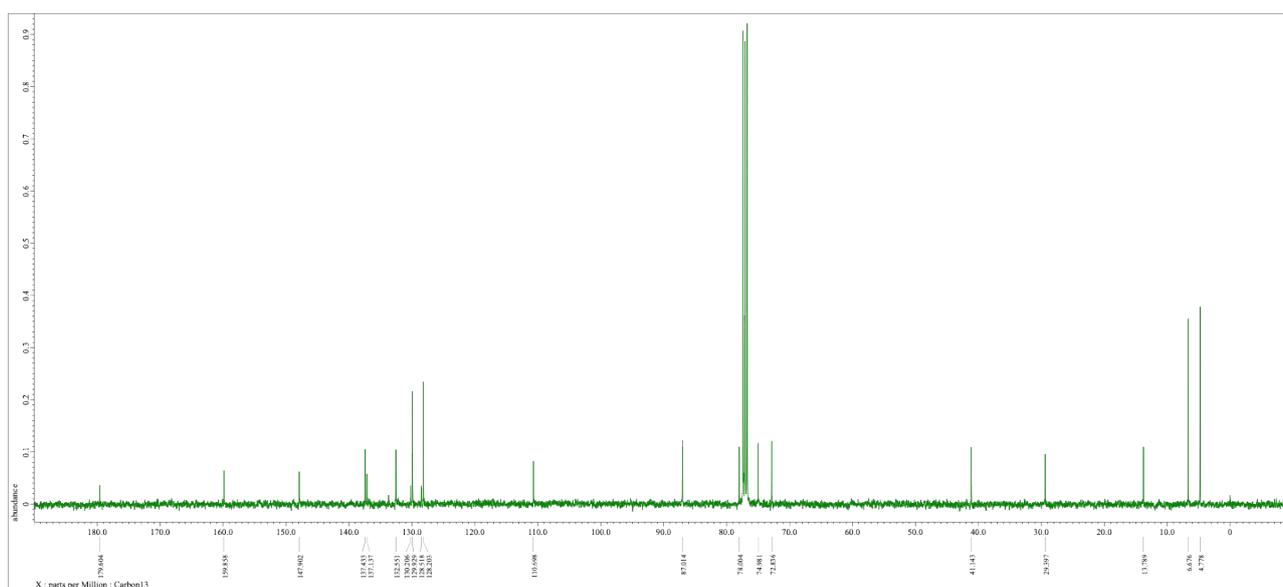
Compound 15 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)

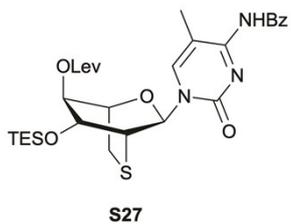


Compound 16 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)

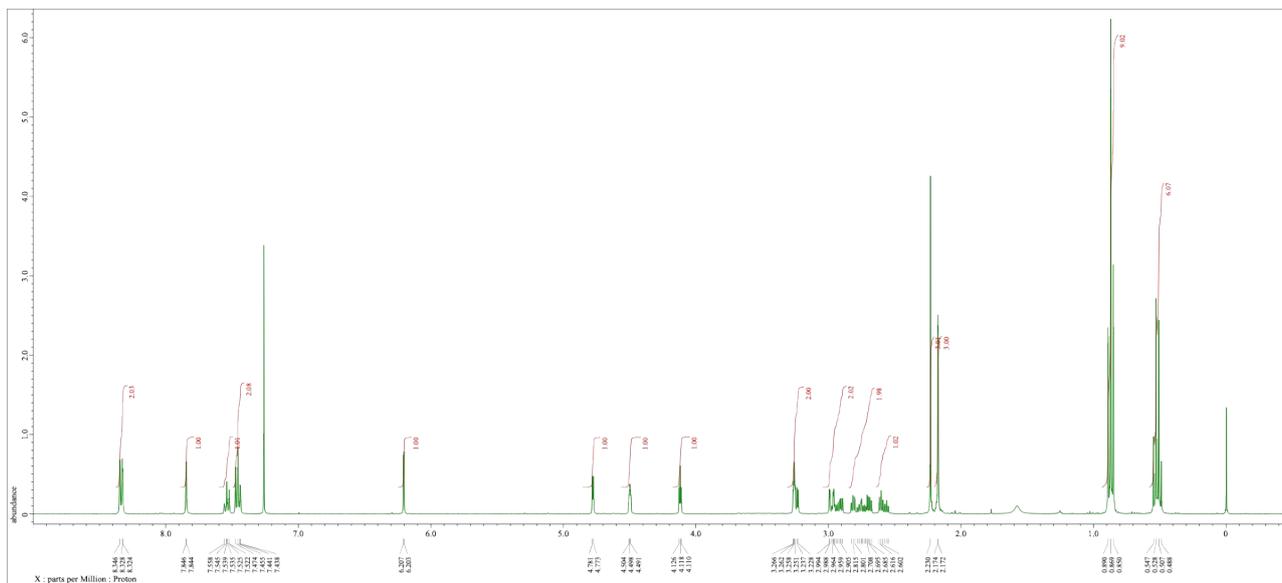


Compound 16 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)

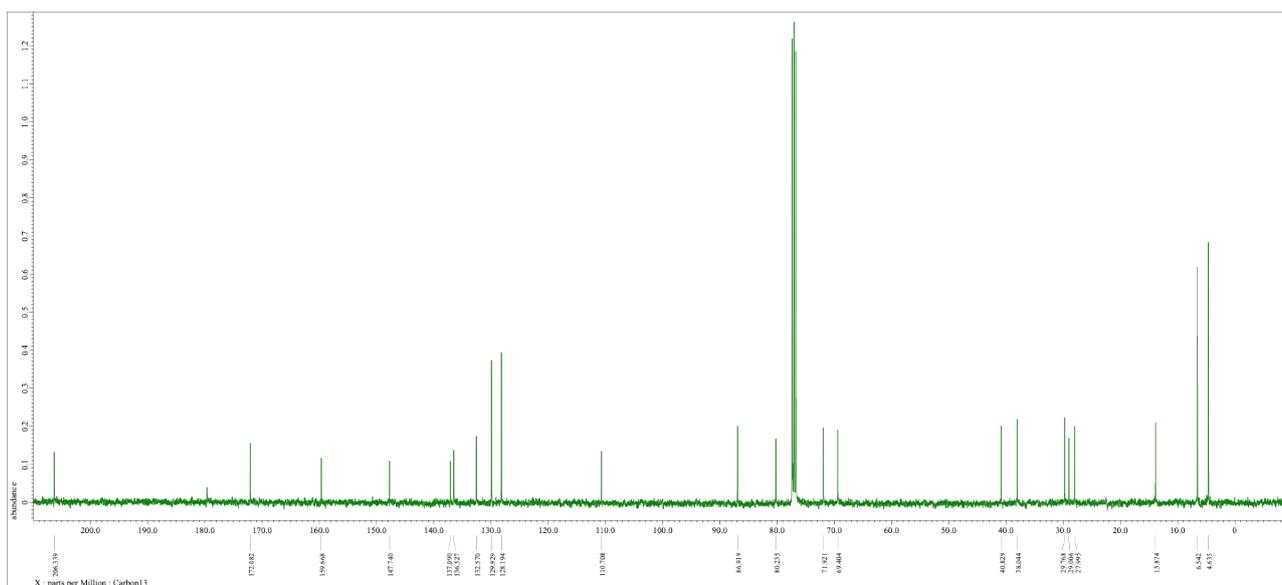




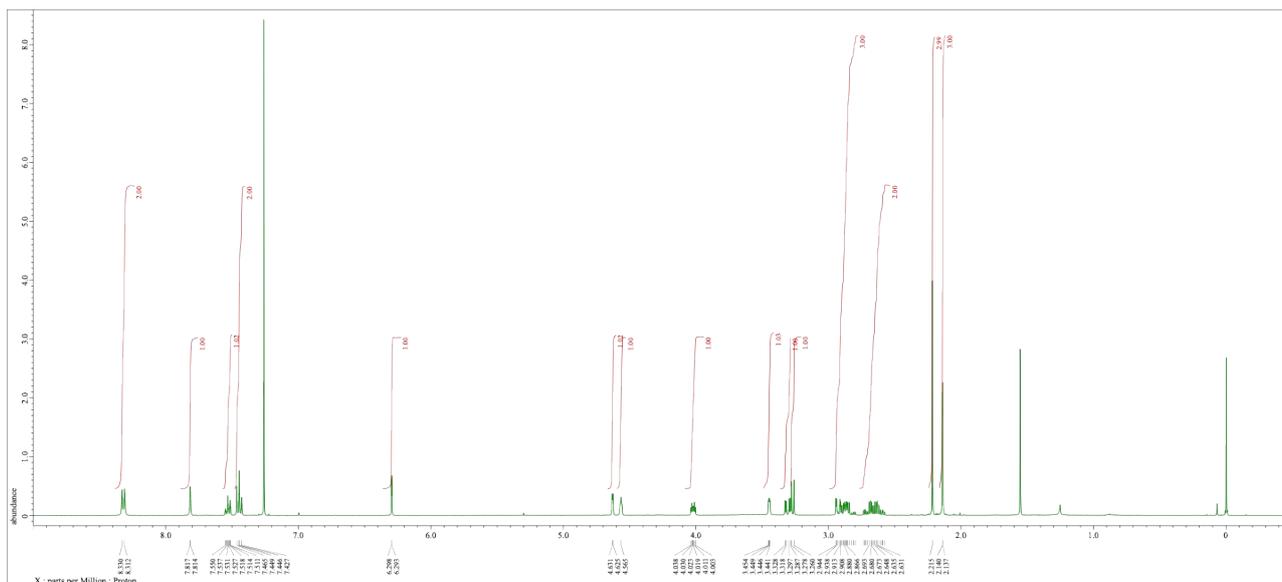
Compound **S27** ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



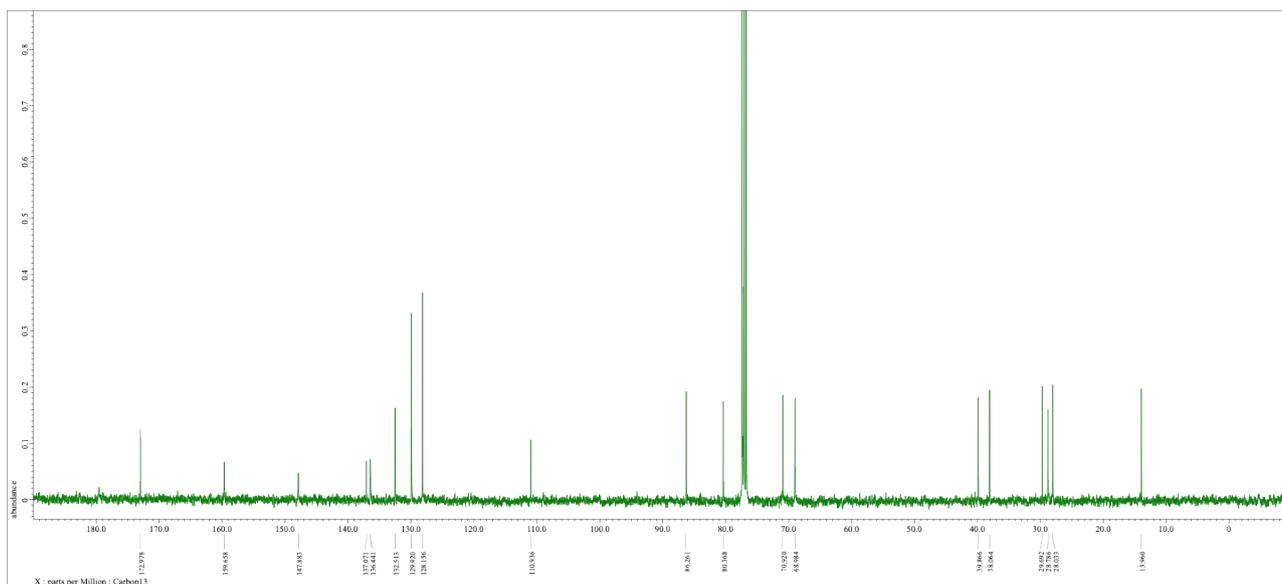
Compound **S27** ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)



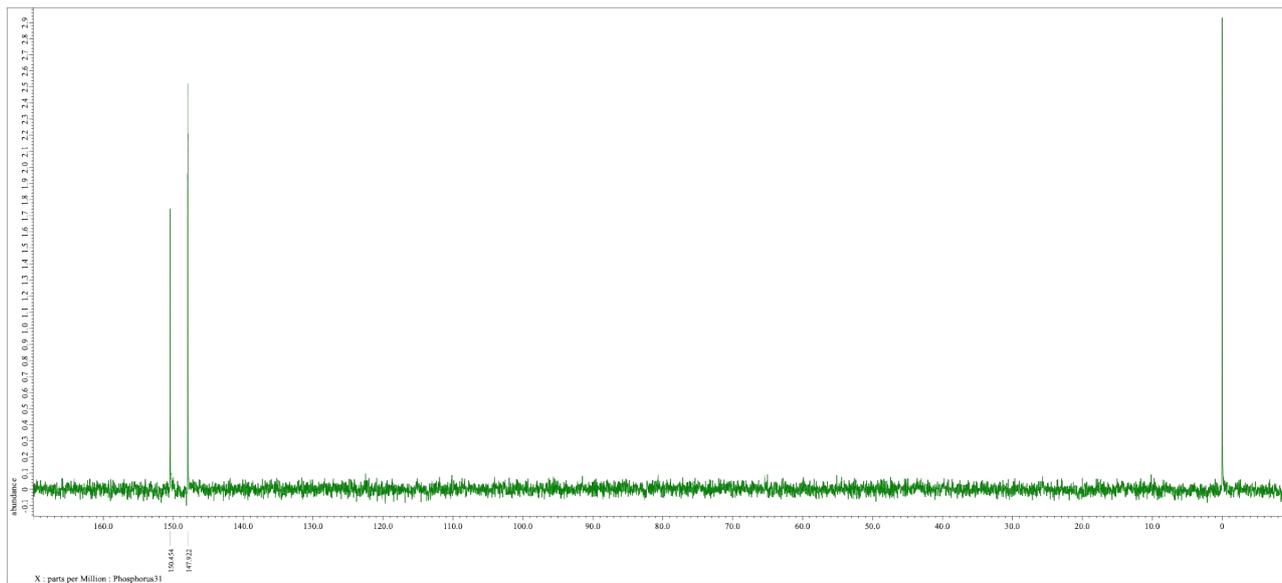
Compound 17 ( $^1\text{H-NMR}$ ,  $\text{CDCl}_3$ , 400 MHz)



Compound 17 ( $^{13}\text{C-NMR}$ ,  $\text{CDCl}_3$ , 101 MHz)

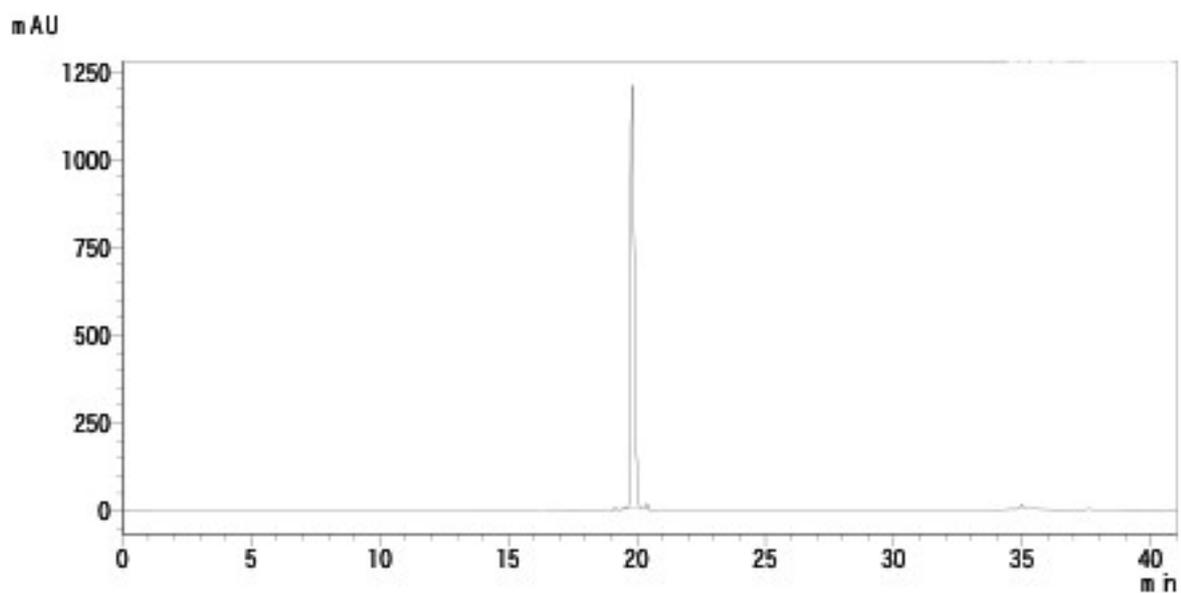


Compound **18** ( $^{31}\text{P}$ -NMR,  $\text{CDCl}_3$ , 162 MHz)

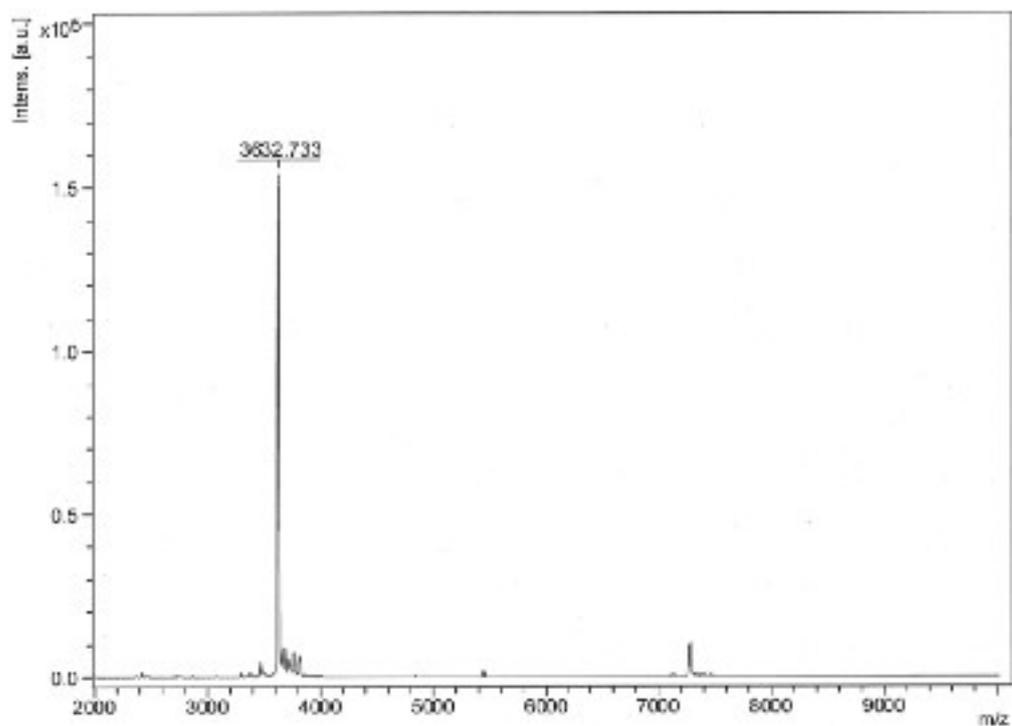


### 3. Characterization data (HPLC and mass data) of synthesized oligonucleotide

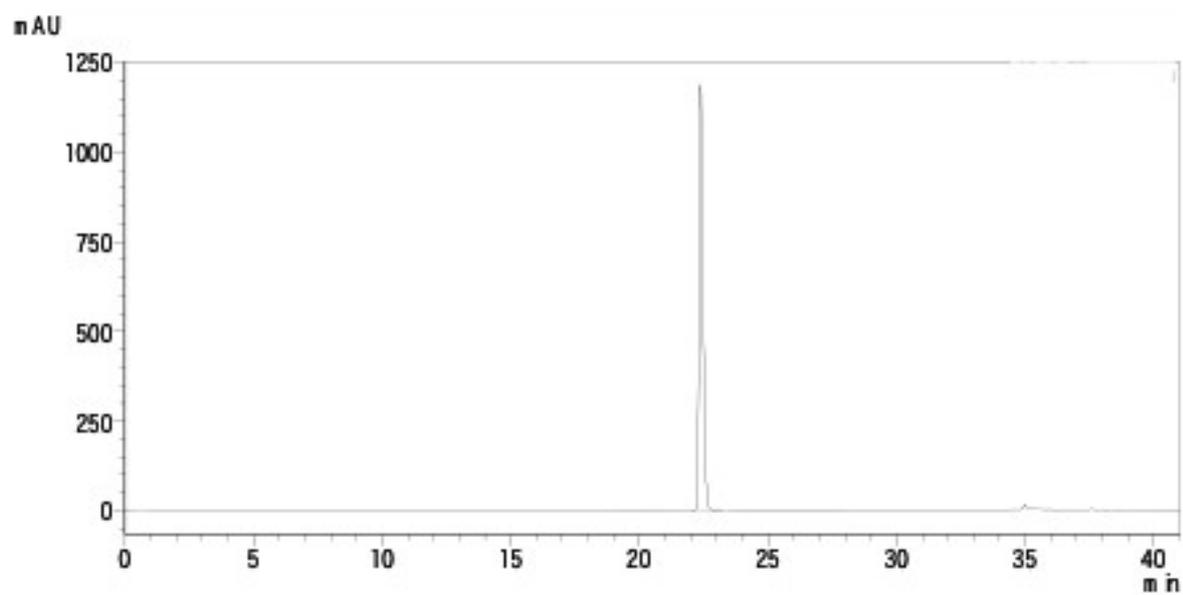
#### HPLC (ON1)



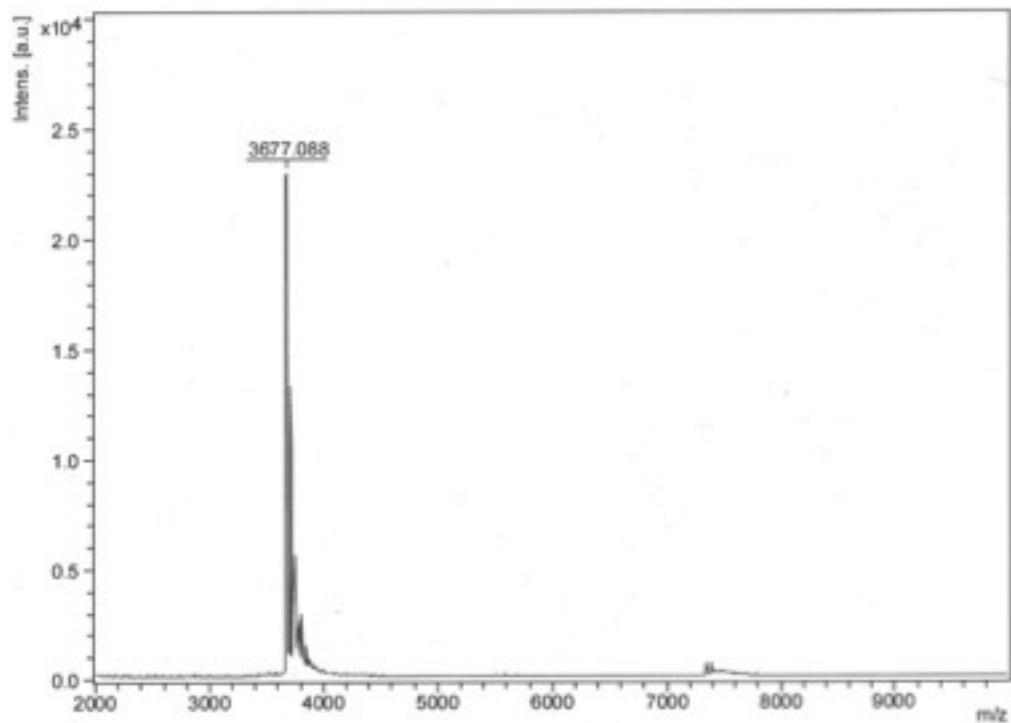
#### MALDI-TOF MS (ON1)



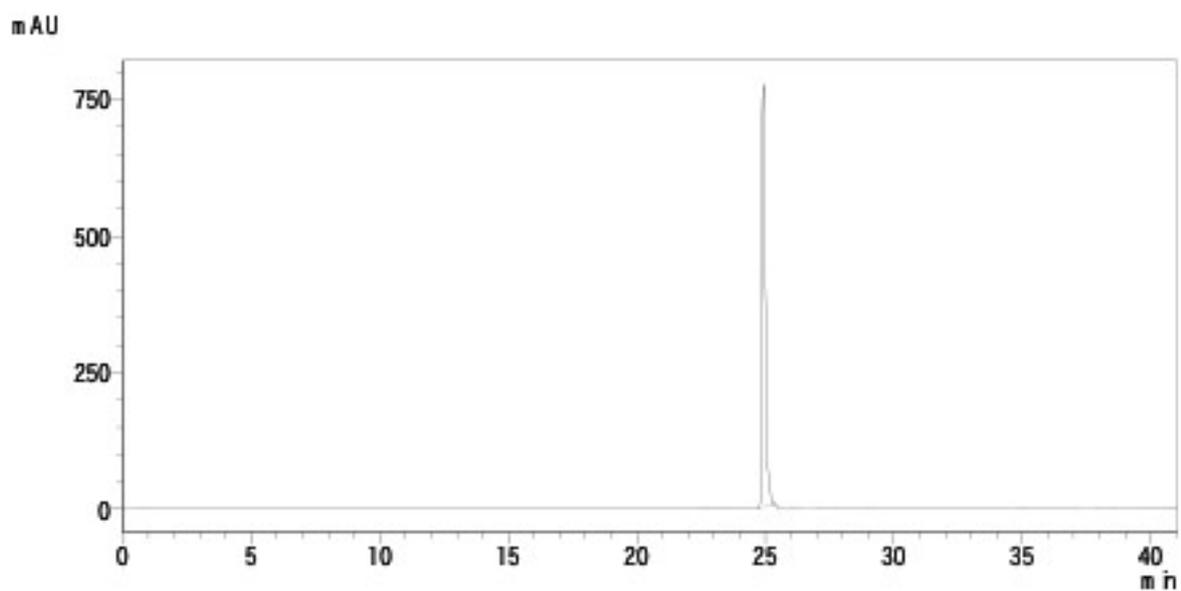
### HPLC (ON2)



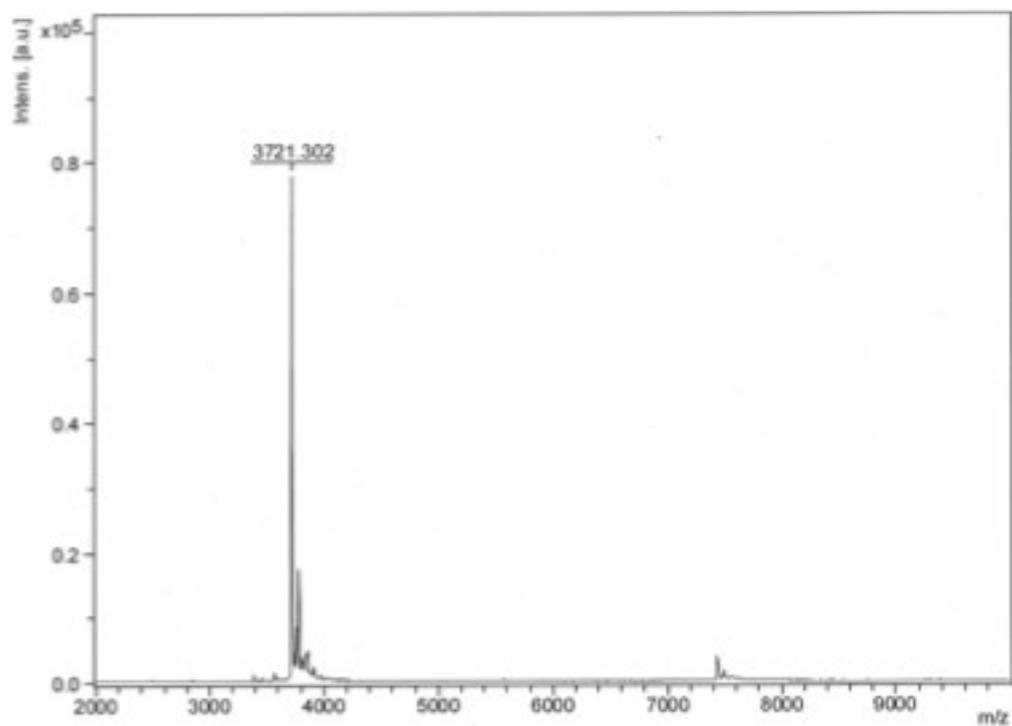
### MALDI-TOF MS (ON2)



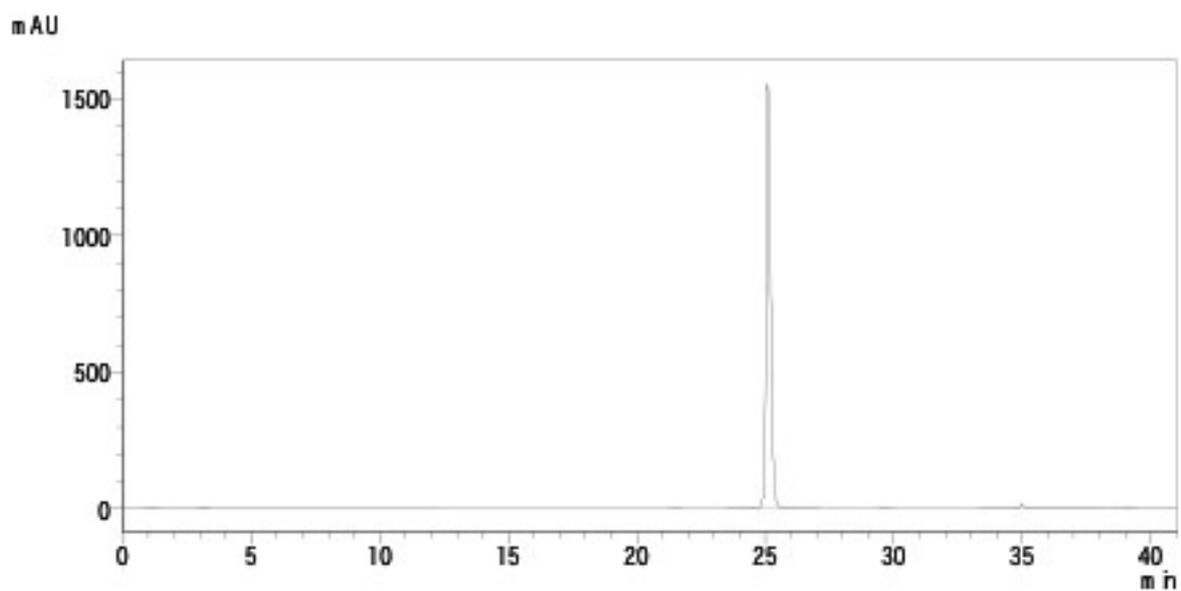
### HPLC (ON3)



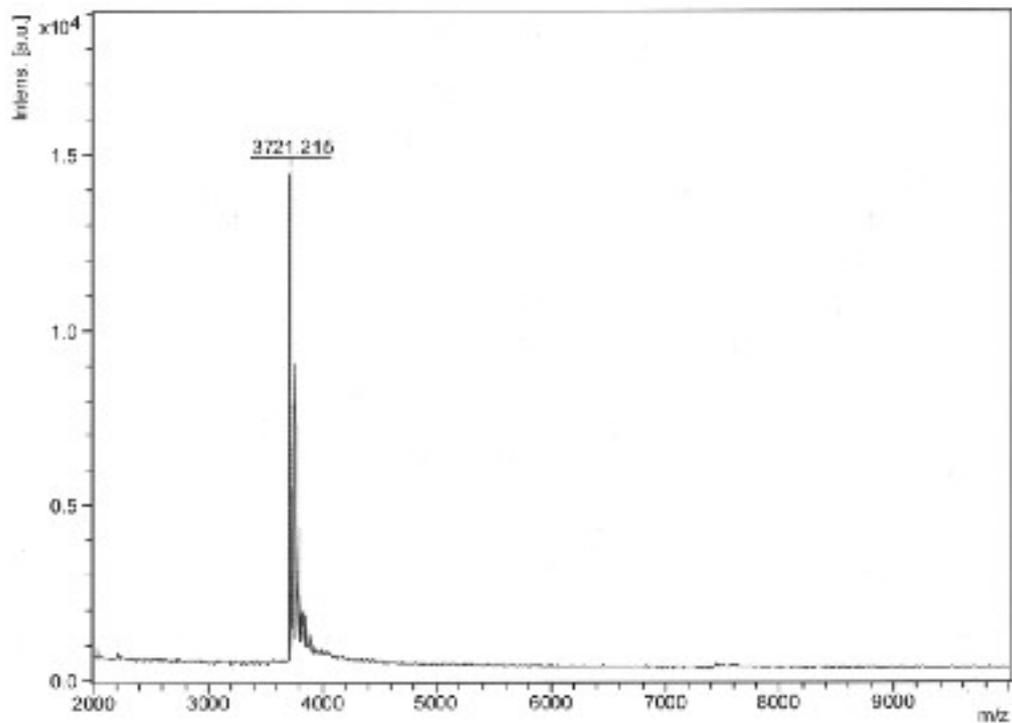
### MALDI-TOF MS (ON3)



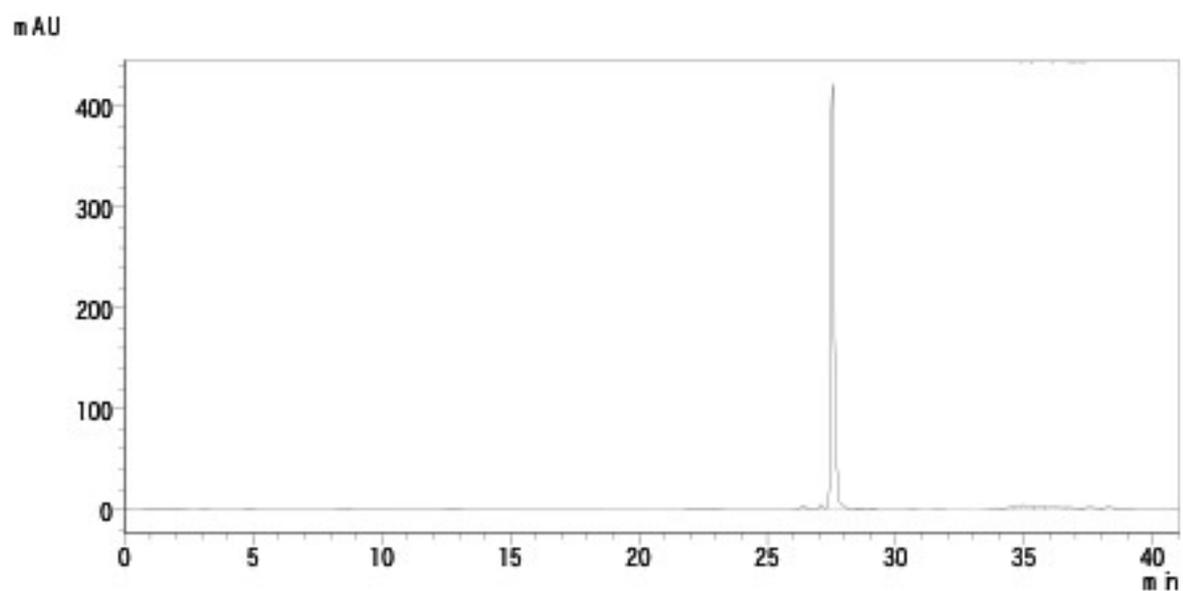
### HPLC (ON4)



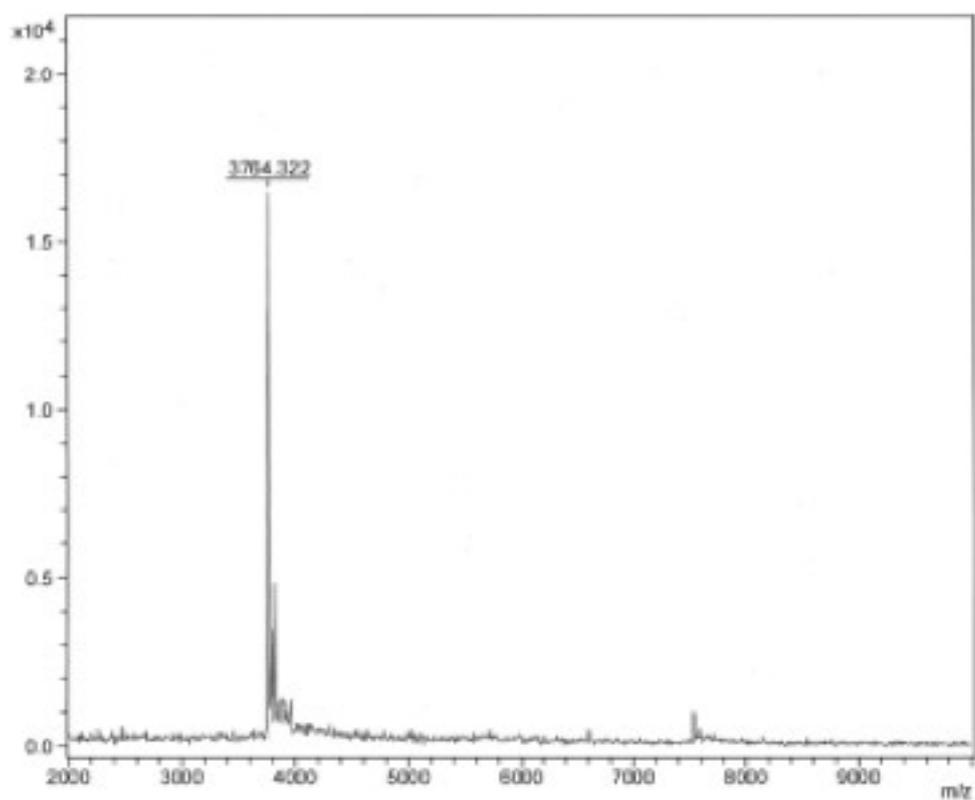
### MALDI-TOF MS (ON4)



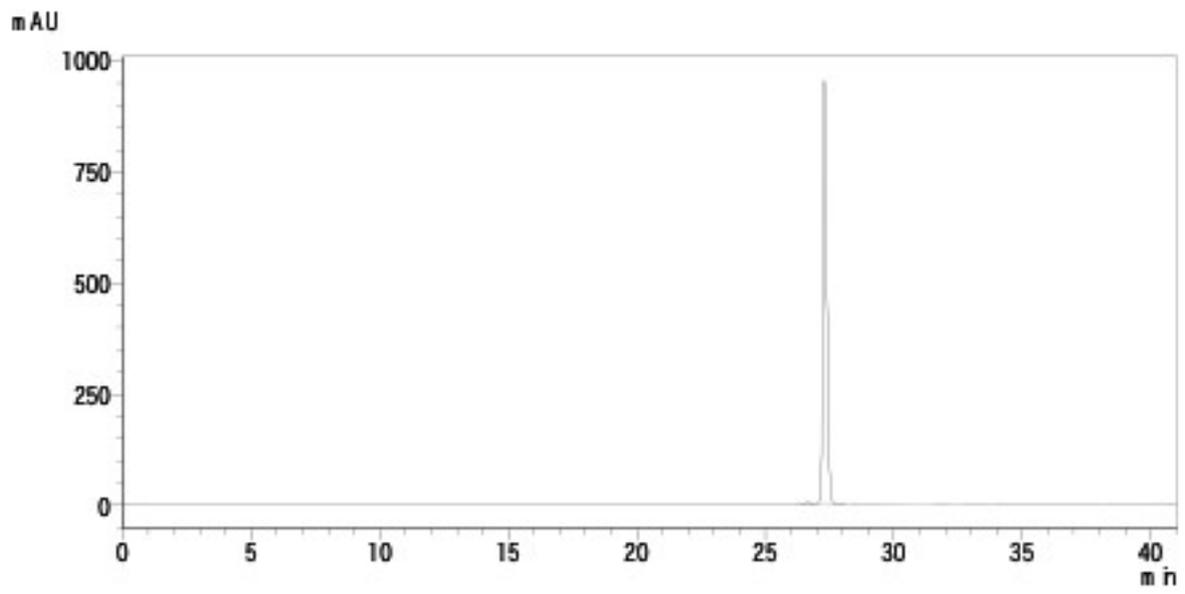
### HPLC (ON5)



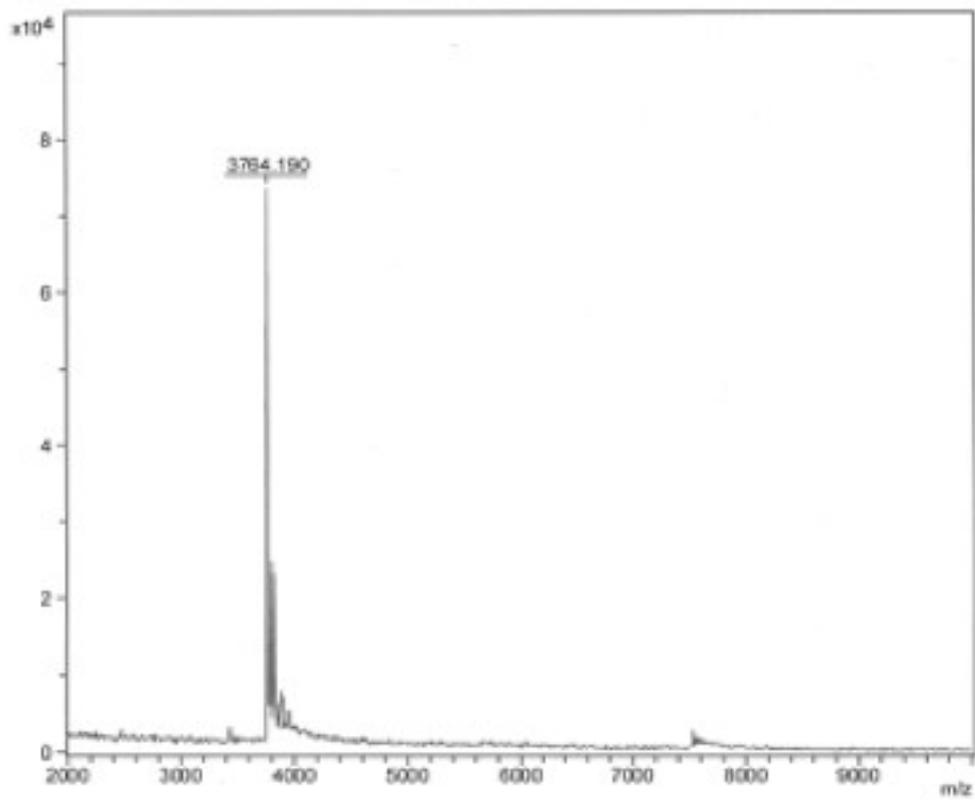
### MALDI-TOF MS (ON5)



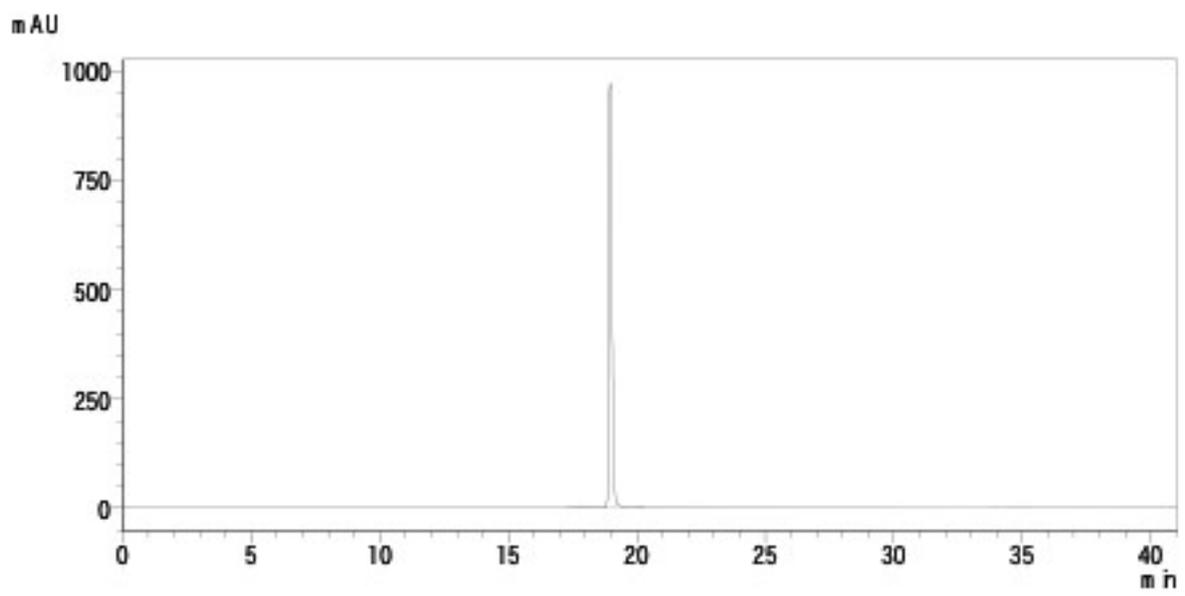
HPLC (ON6)



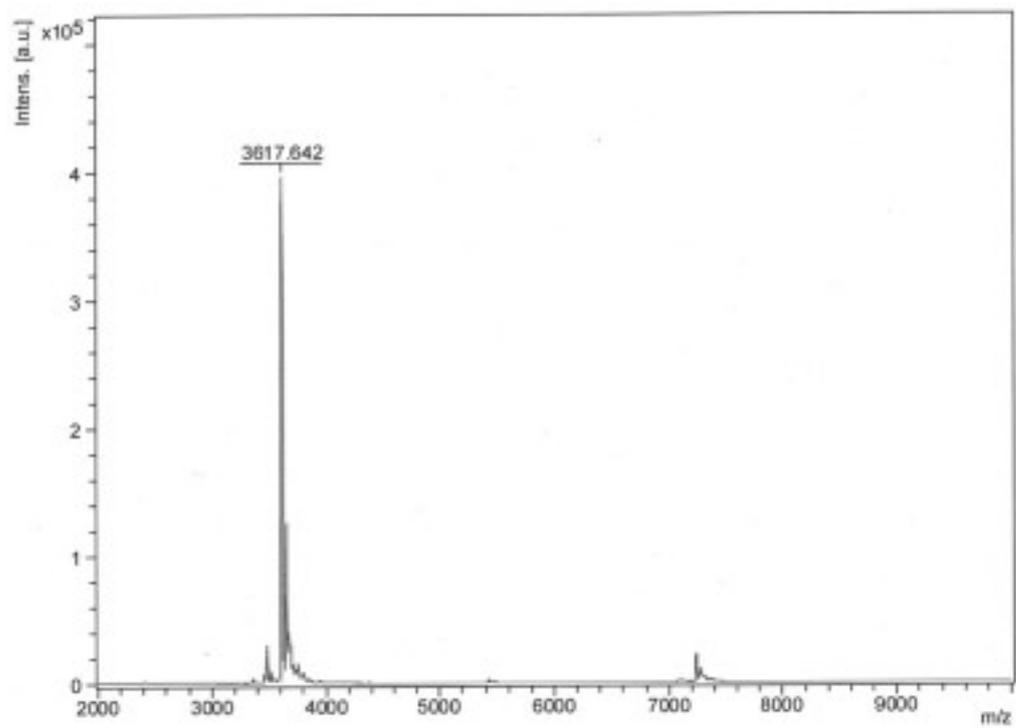
MALDI-TOF MS (ON6)



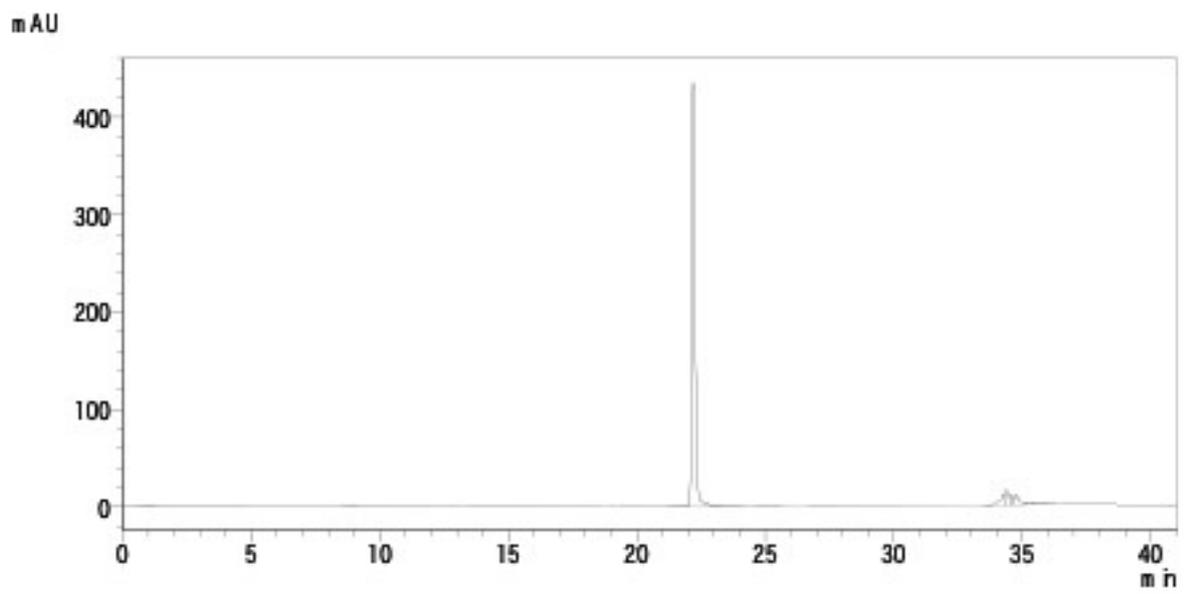
### HPLC (ON7)



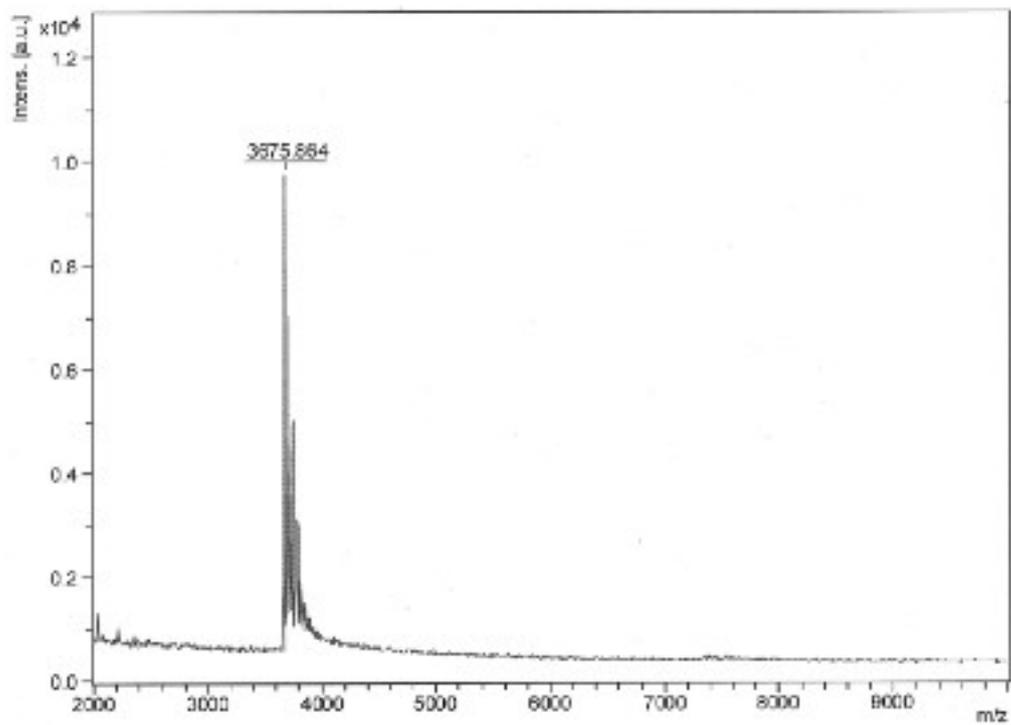
### MALDI-TOF MS (ON7)



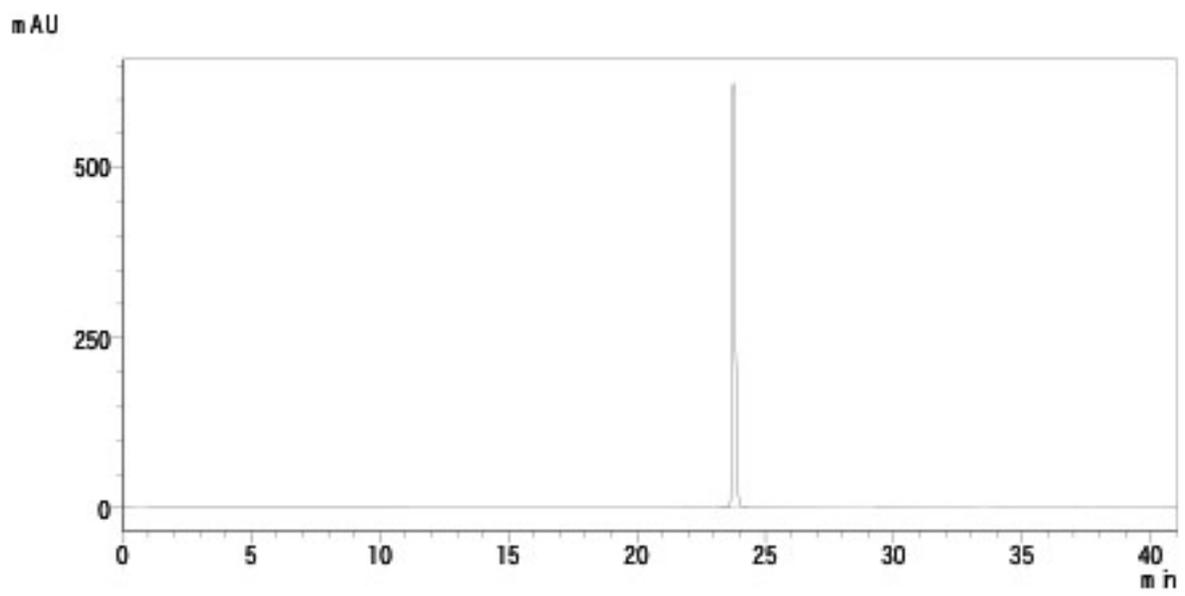
### HPLC (ON8)



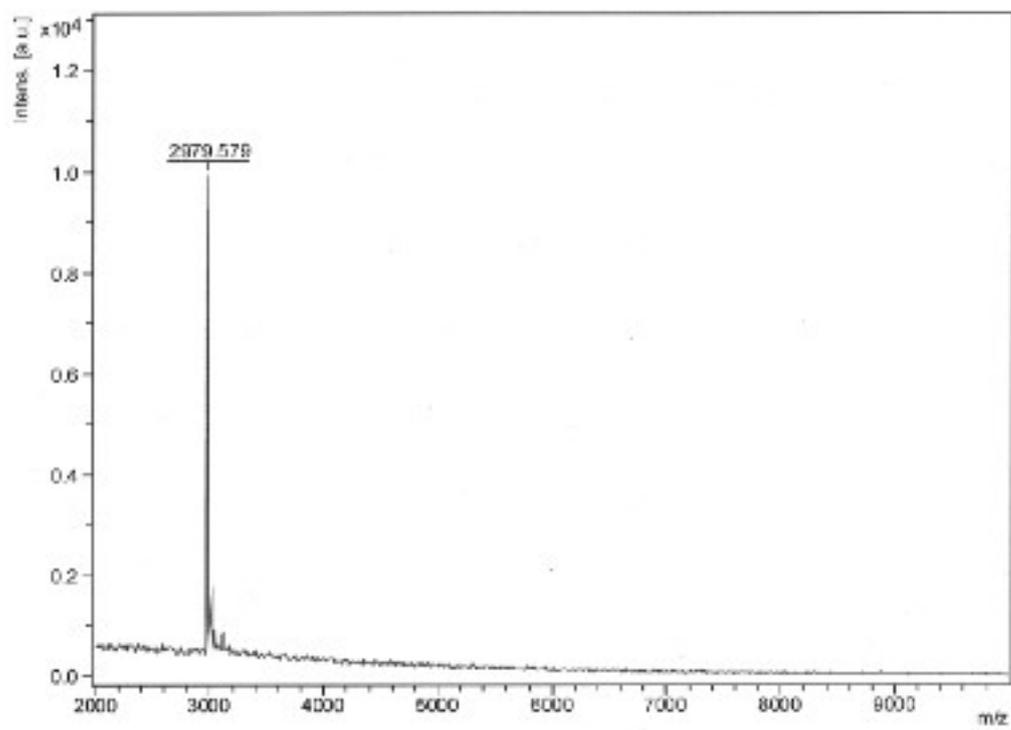
### MALDI-TOF MS (ON8)



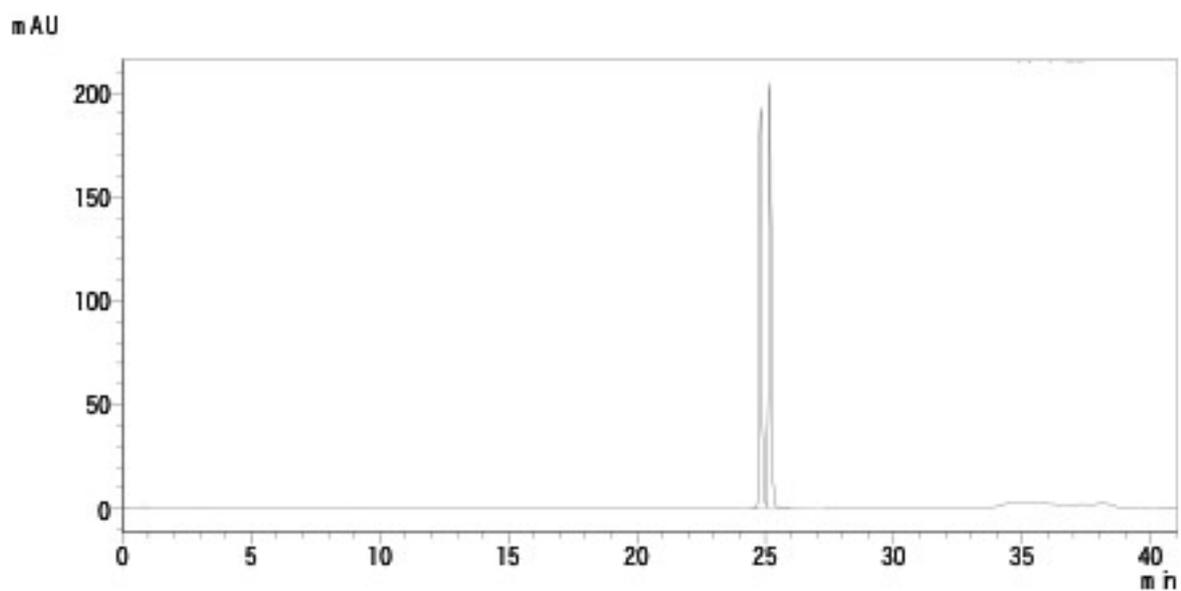
### HPLC (ON9)



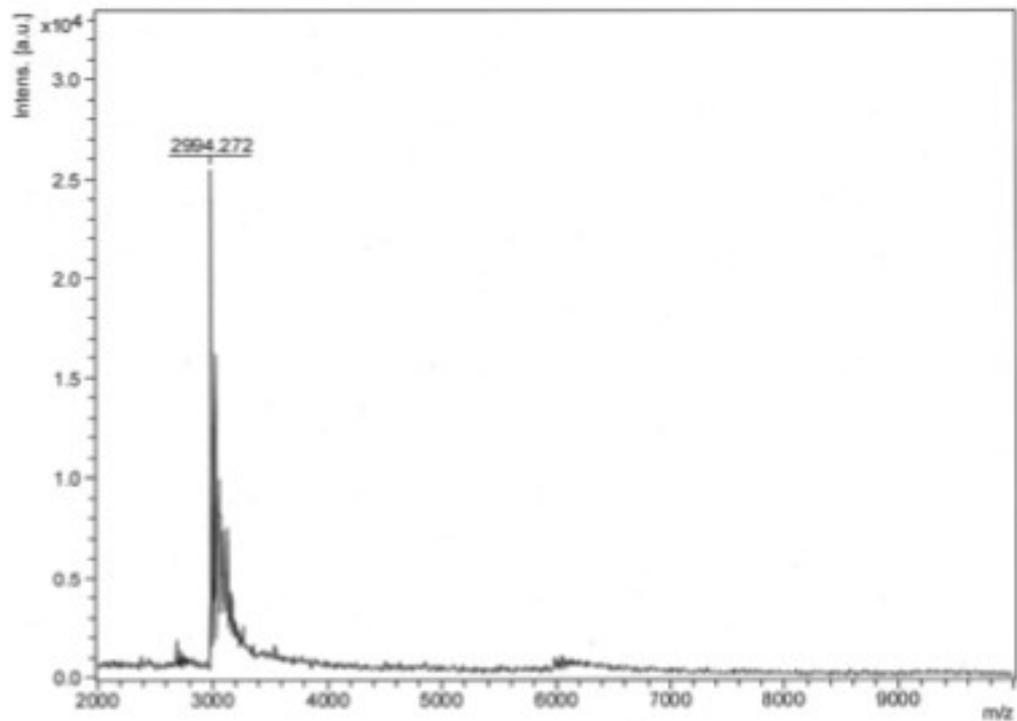
### MALDI-TOF MS (ON9)



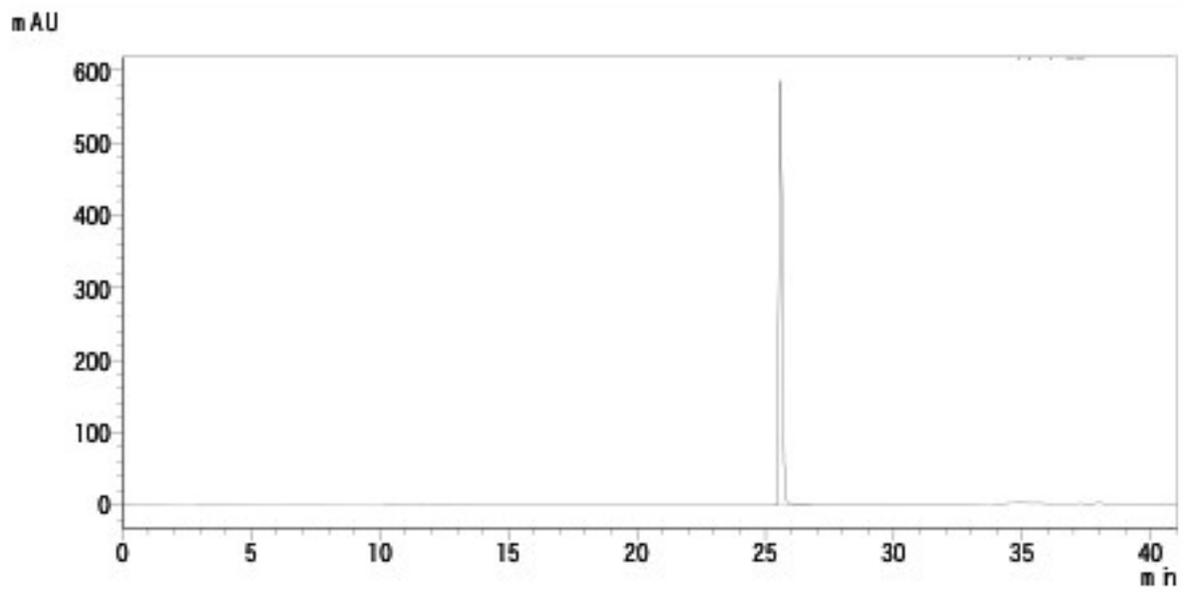
### HPLC (ON10)



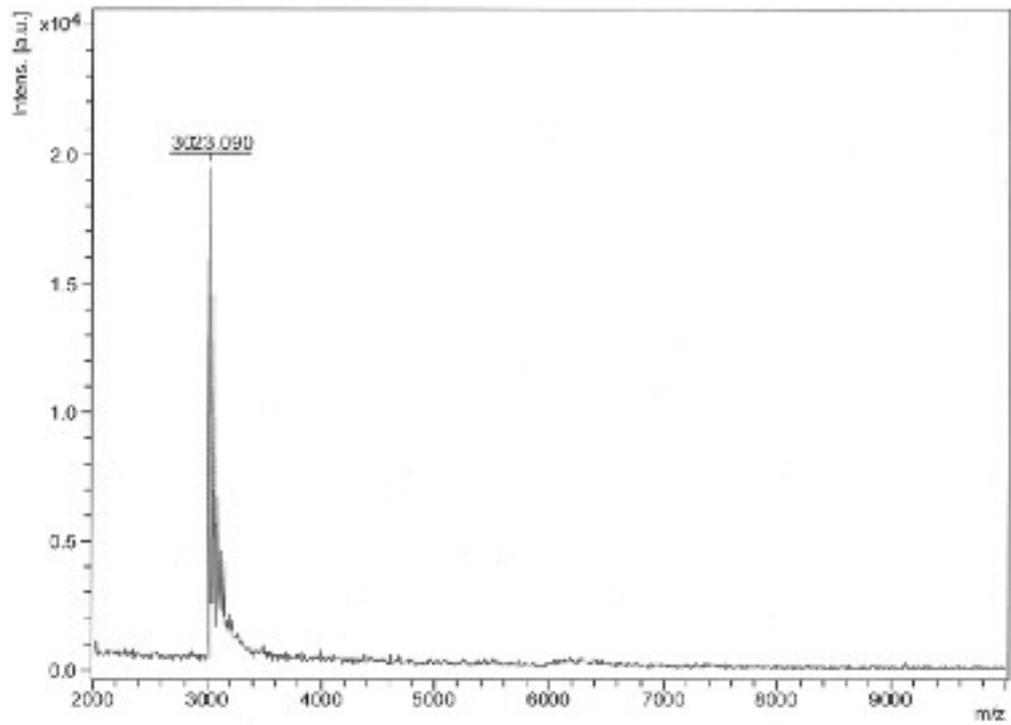
### MALDI-TOF MS (ON10)



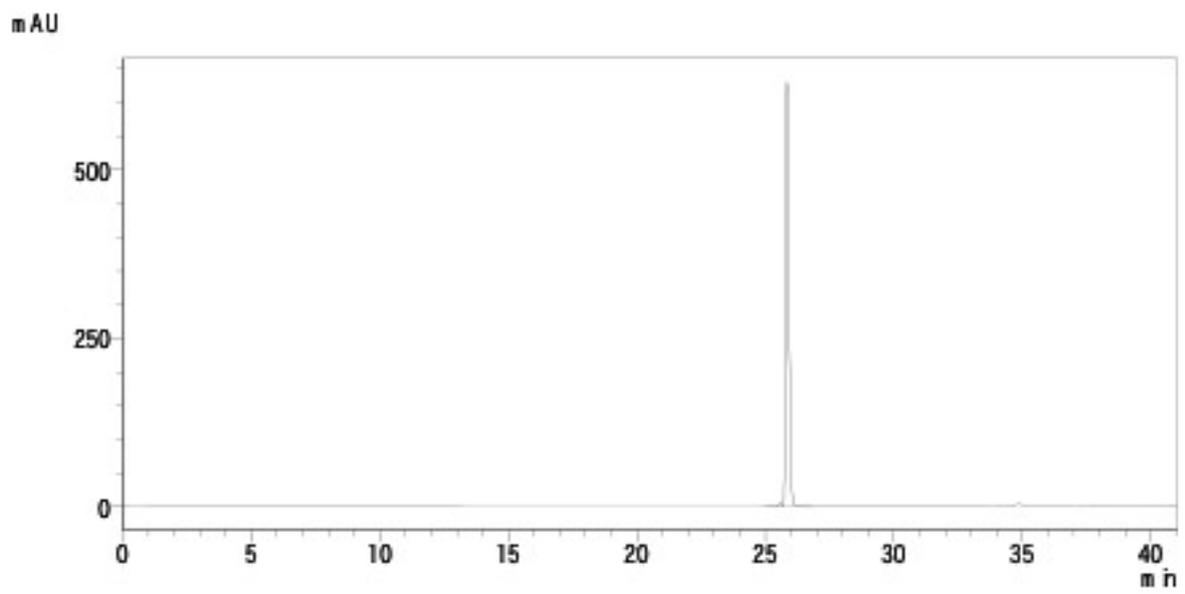
### HPLC (ON11)



### MALDI-TOF MS (ON11)



HPLC (ON12)



MALDI-TOF MS (ON12)

